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*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/100878> since

*Published version:*

DOI:10.1007/s00374-012-0683-4

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This is an author version of the contribution published on:

R. Borriello; E. Lumini; M. Girlanda; P. Bonfante; V. Bianciotto  
Effects of different management practices on arbuscular mycorrhizal fungal  
diversity in maize fields by a molecular approach  
BIOLOGY AND FERTILITY OF SOILS (2012) 48(8)  
DOI: 10.1007/s00374-012-0683-4

The definitive version is available at:

<http://www.springerlink.com/index/pdf/10.1007/s00374-012-0683-4>

# Effects of different management practices on arbuscular mycorrhizal fungal diversity in maize fields by a molecular approach

Roberto Borriello, Erica Lumini, Mariangela Girlanda, Paola Bonfante and Valeria Bianciotto

## Abstract

As obligate mutualistic symbionts, arbuscular mycorrhizal fungi (AMF) colonize the roots of many agricultural crops, and it is often claimed that agricultural practices are detrimental to AMF. As a result, agroecosystems impoverished in AMF may not get the fully expected range of benefits from these fungi. Using molecular markers on DNA extracted directly from soil and roots, we studied the effects of different management practices (tillage and N fertilization) on the AMF communities colonizing an experimental maize field in Central Italy. Our molecular analysis based on three different nuclear rRNA regions (18S, 28S and ITS) allowed us to assess AMF biodiversity. Glomeraceae members were the main colonizer, and they co-occurred with Gigasporaceae and *Paraglomus* regardless of the management practices applied. Diversisporaceae and Entrophosporaceae members were instead detected in the N-fertilized soils and in the untreated soil, respectively. The results obtained indicated that the general AMF assemblages structure and composition in the maize field plots appear to be primarily influenced by N fertilization and, to a lesser extent, by tillage. This study also validates the usefulness of multiple molecular markers to consolidate and refine the assessment of the environmental AMF diversity.

## Introduction

The symbiotic fungi that form arbuscular mycorrhizae (all belonging to the phylum Glomeromycota) are among the most important soil microorganisms (Schüssler et al. 2001). They are thought to be the oldest group of organisms living in symbiosis with land plants (Redecker et al. 2000). Arbuscular mycorrhizal fungi (AMF) are generalist and colonize the roots of most land plants, where they facilitate mineral nutrient uptake from the soil in exchange of plant-assimilated C (Smith and Read 2008). In addition to improved plant nutrition, AM fungi protect their hosts from pathogens (Pozo and Azcón-Aguilar 2007) and affect plant growth traits (Artursson et al. 2006; Barret et al. 2011). Although their spores can germinate in the absence of host plants, AMF are obligate biotrophs, and they depend on a living photoautotrophic partner to complete their life cycle.

About 240 AMF species have been described so far (Oehl et al. 2011), primarily on the basis of spore morphology, but increasingly combined with DNA sequence analyses (Öpik et al. 2010). The use of DNA sequencing methods and high-throughput molecular tools has improved our understanding of the diversity of AMF, in particular because they have allowed AMF to be directly detected from environmental samples (Öpik et al. 2009; Lumini et al. 2010; Dumbrell et al. 2011). Apparently, natural communities of AM fungi are biologically diverse, with large numbers of undescribed taxa. Large numbers of plant–fungus combinations occur naturally, but not all fungus–plant combinations behave similarly, implying that the cost/benefit ratio differs with different fungal–plant cohorts (Helgason and Fitter 2009). There is evidence that experimental (van der Heijden et al. 1998) and natural (Moora et al. 2004) AMF communities, with different taxa compositions, may induce a different growth response in plants and may thus influence the structure and composition of vascular plant communities. Like natural host plants, many important crop plants (sorghum, maize, wheat, rice) belonging to worldwide agroecosystems establish symbioses with AMF (Öpik et al. 2006), having the possibility to exploit the entire range of benefit granted by this interaction.

However, modern intensive farming practices seem to be a threat to AMF, affecting and shaping AMF communities to a great extent. Many studies indicated that AMF abundance and effectiveness, with respect to root colonisation and plant growth promotion, are declining as agricultural practices are intensified (Jansa et al. 2003; Oehl et al. 2005; Alguacil et al. 2008; Verbruggen et al. 2010).

Until now, most surveys described AMF communities present in natural or in agricultural fields using molecular and morphological approaches, on either mycorrhizal roots (Alguacil et al. 2009; Verbruggen et al. 2010) or spores (Moreira-Souza et al. 2003; Oehl et al. 2003), but just a handful of studies have considered the entire soil, or at least the entire topsoil (Hempel et al. 2007; Pivato et al. 2007; Cesaro et al. 2008).

In this work, we have undertaken a study of the impact of different management practices on AMF diversity in a maize crop agroecosystem located in the coastal hills of the Marche region (Central Italy) mainly using a molecular approach based on “environmental” DNA directly extracted from soil and roots. Samples were collected during the maize-growing season in conventional and no-tillage systems in fertilized and unfertilized soils, and the composition of the arbuscular mycorrhizal fungal community was then characterized.

The innovative aspect of this work lies in the application of three different primer sets to molecularly assess the AMF biodiversity. The simultaneous analysis of three different rRNA gene regions allowed obtaining a wide coverage of the AMF taxa, either as spores, extraradical and intraradical mycelia, present in maize fields subjected to different agricultural practices.

Our final goal was to identify which of the aforementioned agricultural practices may have impacted AMF communities and therefore potentially affected soil quality and the sustainability of this agricultural ecosystem.

## **Materials and methods**

### **Experimental sites**

The “P. Rosati” Experimental Agriculture Station, established in 1994 and located in Agugliano (43°32'20.62" N, 13°22'07.51" E, 100 m a.s.l.), Ancona, Italy, was selected for this study as a sampling site. The mean annual temperature is 15.3°C and the mean annual precipitation 700 mm. The field experiment was set up on a hillside (20 % slope) where the soil is classified as Calcaric Gleyic Cambisol (FAO, 2006), which in the first 30 cm has an Ap horizon, silt–clay texture (sand 10 %, clay 47 %, silt 43 %), 7.9 g kg<sup>-1</sup> organic C, 33 % carbonates and a pH of 8.3. Between 30 and 50 cm, the soil has a Bw horizon, pH of 8.5, 6.40 g kg<sup>-1</sup> organic C, as well as a texture and carbonate percentages similar to those of the upper layers. The site is managed under a *Triticum durum* and *Zea mays* rotation. *Z. mays* (hybrid DK440) was planted on 20th April and harvested on 24th September 2007. The experimental field was organized as two main plots to compare two tillage techniques (T, conventional 40-cm-deep ploughing; S, no-till with chemical desiccation and chopping) with two N fertilizer treatments as the subplots (1, 90 Kg N ha<sup>-1</sup> year<sup>-1</sup> as ammonium nitrate; 0, no fertilization—320 m<sup>2</sup> each). The T plots were ploughed every year by moldboard at 40-cm depth in autumn (October) for wheat and at the end of summer (from 30th August until 30th September) for maize. The seedbed was prepared with double harrowing before the sowing date. The S plots were left undisturbed, except for sod seeding, crop residuals and weed chopping, and total herbicide spraying prior to seeding. Tilling at 40-cm depth represents the traditional agronomic practice adopted by local farmers (Marche region) for agricultural fields placed on a hillside and characterized by heavy textured soil. Ammonium nitrate or urea was applied at a rate of

90 kg N ha<sup>-1</sup>, split equally in February and April to wheat; a single application of 90 kg N ha<sup>-1</sup> was applied to maize. Triple superphosphate was always applied before sowing at a rate of 50 kg ha<sup>-1</sup> of P<sub>2</sub>O<sub>5</sub> per year. The four considered treatments were abbreviated as follows for ease of labelling: tilled with N fertilizer (T1), tilled without N fertilizer (T0), no-tilled with N fertilizer (S1) and no-tilled without N fertilizer (S0). In winter, no cover crops were present in the tilled plots (T0, T1), whilst the no-tilled plots (S0, S1) were covered by weeds (*Setaria* sp.).

#### Soil and root sampling

Soil samples (three replicates) were collected on 11th June 2007, during maize rotation, in the four considered treatments, for a total of 12 samples. The soil cores, measuring 5 cm in diameter and 20 cm in length, were taken at 10- to 30-cm depth. The soil samples were sieved immediately at 2 mm, frozen and stored until analysed.

Two full maize plants were collected during the anthesis development stage from each treatment plot and taken to the laboratory (a total of eight plants). Root fragments from each plant (0.5 g each sample) were washed free of soil, air-dried at room temperature and immediately used for morphological analyses. The roots were stored at -20°C until used for molecular analyses.

#### Assessment of root colonisation by AMF

Maize roots were stained with 0.01 % cotton blue in lactic acid for about 20 h and then destained four times with lactic acid. Roots were cut into small fragments (1 cm each) and mounted onto microscope slides with lactic acid. Forty-five fragments were observed for each root apparatus, for a total of 360 root fragments. AMF colonisation intensity in the root cortex and arbuscule presence were determined, as described in Trouvelot et al. (1986). All the root fragments analysed showed a good mycorrhization level with a mean value of 78.5 %.

#### DNA extractions from soils and roots

Two independent DNA extractions (0.5 g of soil each) from the 12 soil samples were performed using a FastDNA Kit (MP Biomedicals) according to the manufacturer's recommendations. An additional step, consisting of six washes with guanidine thiocyanate (6 M), was introduced after the addition of the binding matrix suspension due to the high levels of clay in the soil samples. DNA extractions from the eight root samples were performed using a DNeasy Plant Mini Kit (Qiagen, Crawley, UK), according to the protocol for frozen samples.

#### PCR, cloning and sequencing of the fungal ribosomal (rRNA) genes

Three sets of primers were independently used to amplify three different regions of Glomeromycota ribosomal DNA. The universal eukaryotic primer NS31 (Simon et al. 1992) was used as the forward primer for the first primer set. A mixture of three different primers (AMmix) was used as the reverse primer—AM1 (Helgason et al. 1998), AM2 and AM3 (Santos-González et al. 2007) primers—designed to amplify ~550-bp fragments of partial SSU rDNA. The second primer set (28G1–28G2) was designed to amplify a ~600-bp fragment of partial LSU rDNA (da Silva et al. 2006). In addition, a nested PCR approach (Millner et al. 2001) was applied to both the soil and root DNA first with the primer set LSU-Glom1/SSUGlom1 (Renker et al. 2003) and then a PCR round with the Paraglomeraceae family-specific primer pair GOCC56/GOCC427 designed to amplify a fragment of rDNA gene including part of the ITS1, the complete 5.8S and part of ITS2 regions. The annealing temperature was raised to 58°C when these primers were used to increase specificity, as suggested by Hempel et al. (2007).

### *Amplification of genomic DNA from soil and root*

PCR on DNA extracted from soil were performed using 0.1  $\mu\text{g } \mu\text{l}^{-1}$  bovine serum albumin (BSA), 0.2 mM dNTPs, 3.5 mM of  $\text{MgCl}_2$ , 0.5  $\mu\text{M}$  of each primer (28G1–28G2 and NS31-AMmix) and the supplied reaction buffer, with 2 U of High Fidelity Taq (Roche) to obtain a final volume of 40  $\mu\text{l}$ . Amplifications were carried out in 0.2-ml PCR tubes using a Biometra T Gradient thermocycler in the following steps: 5 min initial denaturation at 94°C; 35 cycles of 1 min at 94°C, 1 min at 55°C and 1 min at 72°C; and a final elongation of 7 min at 72°C. A negative control was included in the PCR to check for contamination. The PCR on DNA extracted from root were carried out as described above, except for the annealing temperature that was 58°C. All PCR products were checked using 1.5 % agarose gel with SYBR®safe DNA gel stain (Invitrogen).

### *Cloning and sequencing*

The PCR products were purified using QIAquick (Qiagen, Hilden, Germany), cloned in a pGEM-T Easy vector (Promega, Madison, Wisconsin, USA) and transformed into *Escherichia coli* (X11 blue). Putative positive transformants were screened in each resulting SSU, LSU, ITS rRNA gene library. The reaction mixture for the PCR amplification of the mycorrhizal root DNA (25  $\mu\text{l}$ ) contained: 10 mM Tris–HCl, 50 mM KCl, 1.5 mM  $\text{MgCl}_2$ , 0.1 % gelatin, 0.2 mM of each dNTP, 0.1  $\mu\text{g } \mu\text{l}^{-1}$  BSA, 0.5  $\mu\text{M}$  of each primer and 1 U of RedTaq DNA polymerase (Sigma). Amplifications were carried out as described above, except for the cycling parameters which instead were: 35 cycles of 1 min at 94°C, 1 min at 58°C, 1 min at 72°C for 35 cycles and a final elongation of 5 min at 72°C after the last cycle. All PCR products were checked using 1.5 % agarose gel with SYBR®safe DNA gel stain (Invitrogen). All clones having inserts of the correct size in each library were sequenced using T7 vector primers by Macrogen sequencing services (Macrogen, Seoul, Korea).

Sequences generated in this study were registered in GenBank under the following accession numbers HM215625–HM216180.

### *Sequence analyses and phylogenetic inference*

Sequence editing was done using Sequencer v4.2.2 (Gene Codes Corporation). Possible chimera sequences were identified using the Chimera Detection programme (Cole et al. 2003). A search for similar sequences to the ones from this study was conducted with the BLAST tool (Zhang et al. 2000) provided by GenBank. The sequences were with high similarity and a wide selection of AMF taxa, including representatives of the major clades described by Oehl et al. (2011). All the sequences were aligned, separately for the SSU and LSU groups, using the multiple sequence comparison by log-expectation, MUSCLE v3.7, programme (Edgar2004).

The alignments obtained using MUSCLE 3.7 were manually edited and the distance matrices constructed using DNAdist from the PHYLIP suite of programmes, version 3.6, with default parameters (Felsenstein2005). These pairwise distances were used as input for DOTUR (Schloss and Handelsman 2005) in order to group the sequences into operational taxonomic units (OTUs) of a defined sequence identity. A threshold of 97 % identity, corresponding to 0.03 dissimilarity ( $\text{OTU}_{0.03}$ ), was used to define OTUs, and it was chosen on the basis of previous studies on AMF biodiversity (Pivato et al. 2007; Santos-González et al. 2007). Neighbour-joining (NJ) phylogenetic analyses (1,000 bootstrap replicates) were performed with MEGA version 4 programme (Tamura et al. 2007) using the default parameters. Sequences of *Mortierella verticillata* (AF157145), *Chaetomium globosum* (AB048285), *Trichoderma* sp. (EF672340) and *Fusarium oxysporum* (DQ916150) were used as outgroups in the SSU phylogenetic tree. The

outgroup for the LSU phylogenetic tree was a *Claroideoglossus claroideum* (AF235007) sequence. The same alignments were used to perform a Bayesian inference (BI) approach. BI of phylogeny using Monte Carlo Markov chains (MCMC) was carried out with MrBayes 3.1.2 (Huelsenbeck et al. 2001). For each alignment, four incrementally heated simultaneous MCMC were run over 1,000,000 generations, under model assumption, using random starting trees and default starting values of the models. Trees were sampled every 100 generations, resulting in an overall sampling of 10,001 trees. The first 2,500 trees were discarded as “burn-in” (25 %). For the remaining trees, a majority rule consensus tree showing all compatible partitions was computed to obtain estimates for Bayesian posterior probabilities. Branch lengths were estimated as the mean values over the sampled trees.

GLO\_Sx (*Glomerales*), GLO\_Lx (*Glomerales*), GIG\_Sx (*Gigasporales*) and DIV\_Sx (*Diversisporales*) OTUs were named according to their position in the major AMF clades, followed by the letter S (for SSU sequences) or L (for LSU sequences) and a numerical index (x) identifying the OTU number.

#### *Diversity and structure of the AMF communities*

In order to analyse the richness and diversity of the AMF communities in the four considered conditions, only sequences belonging to Glomeromycota were selected for the subsequent analyses. The use of DOTUR allowed us to obtain a variety of diversity and richness estimators: the Shannon–Weaver diversity index, a rarefaction curve, bias-corrected Chao1 richness and ACE abundance-based coverage estimators.

A comparison of the AMF spectra was carried out in the collected samples by means of canonical variates analysis (CVA), a multivariate statistical method. CVA was performed on OTU abundances (number of sequences comprising each OTU). The analysis was carried out using the “Canonical Variates” SYN-TAX 2000 package subroutine with the “Spherized scores of objects” (normalization of eigenvectors) option. Correlations with the original variables were also analysed. Only OTUs comprising more than ten sequences were considered for the analysis since the contribution of rare OTUs to community structure is limited. Due to the low number of Glomeromycota sequences obtained from the root samples, it was decided to consider them as subsamples of each soil treatment in the CVA analysis.

## **Results**

### AMF assemblage identification in soils and roots

The three primer combinations (NS31/AMmix, 28G1/28G2 and GOCC56/GOCC427) yielded three clone libraries, carrying SSU, LSU and ITS fragments, respectively, from each sampling point. We screened and sequenced 1,209 clones in total from soil and roots: 724 clones from the libraries carrying SSU fragments, 389 clones from the LSU libraries and 96 clones from the ITS libraries.

Apart from sequences showing high similarity (98–99 % identity) with AM fungi (Glomeromycota phylum), sequences corresponding to non-target organisms were also obtained. BLAST searches revealed indeed that out of the 724 SSU sequences obtained using the NS31/AMmix primers, 427 sequences (59 % of the total sequences) featured high similarity to sequences from taxa belonging to the phylum Glomeromycota, whilst the remaining 297 sequences (41 % total sequences) were related to sequences from Ascomycota. The high number of Ascomycota sequences detected analysing the SSU library is due to a not complete specificity of the primers (NS31/AMmix) for the Glomeromycota group, as already shown in other studies (Santos-González et al. 2007; Toljander

et al. 2008). Out of the 389 sequences obtained with the 28G1/28G2 primer set, 364 sequences (94 % total sequences) showed high similarity to sequences from Glomeromycota, whilst the remaining 25 sequences (6 % total sequences) showed BLAST similarity with other eukaryotes. All the 96 ITS sequences obtained with the GOCC primers set were found to belong to Glomeromycota. These ITS sequences were obtained from the soils samples and can be referred to only two clades, related to an undescribed *Paraglomus* species and *Paraglomus brasilianum*, respectively (HM215625–HM215633).

The 427 SSU and 364 LSU Glomeromycota sequences were then grouped, using DOTUR software, into OTUs. Considering a similarity threshold of 97 %, the programme identified 27 SSU OTUs (seven of which were singletons) and 55 LSU OTUs (32 of which were singletons). OTUs containing only one sequence (singletons) were discarded, and only OTUs containing two or more sequences retrieved from two different clones were considered for further analyses. This choice allowed us to minimize the possibility to incorporate in the analysis PCR cloning and sequencing artefacts. In conclusion, 20 SSU and 23 LSU OTUs were considered for further analyses.

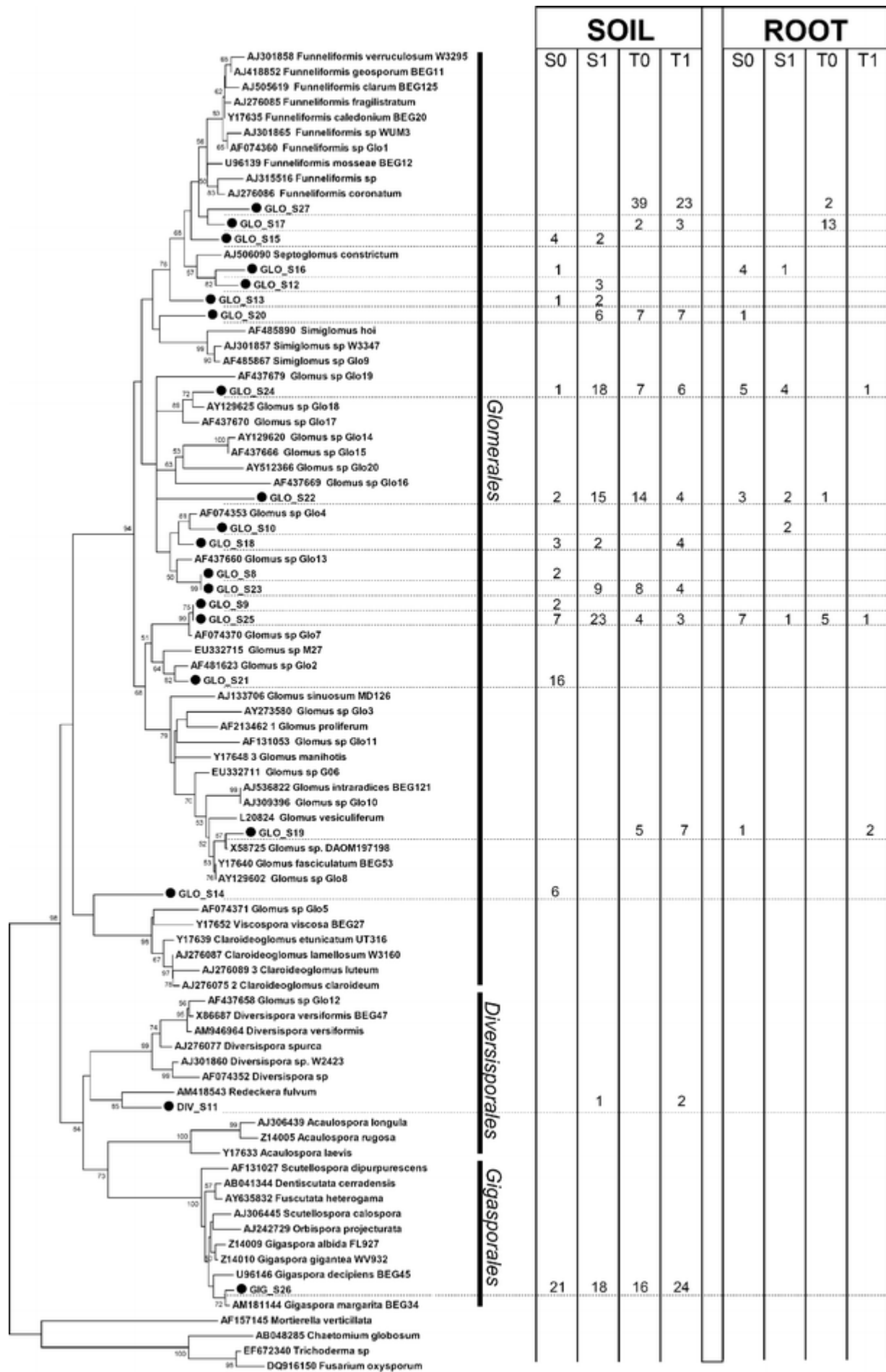
#### Phylogenetic analyses of AMF sequences from maize field

Separate phylogenetic analyses were performed for SSU and LSU sequences to assign the different OTUs to the cognate Glomeromycota groups.

##### *Small subunit*

The phylogenetic analysis of the SSU rRNA sequences is shown in Fig. 1. By far, the most abundant and diverse AMF taxon in soil samples was Glomeraceae, as defined by Oehl et al. (2011), with 276 sequences grouped in 17 OTUs. Three OTUs (GLO\_S25, GLO\_S22, GLO\_S24) were detected in soil under all management conditions. OTU GIG\_S26, which the phylogenetic tree showed to be related to *Gigasporales* clade, was also retrieved in the soil under all analysed conditions. OTU DIV\_S11, related to *Redeckera fulva* AM418543 (85 % bootstrap support), was only detected in soil subjected to N fertilization.





0.21

### Fig. 1

Neighbour-joining phylogenetic analysis of arbuscular mycorrhizal fungal SSU rDNA sequences (~500 bp) obtained from maize root and bulk soil samples. The scale represents substitutions per site. Sequences obtained in the present study are highlighted by a *dot*. Sequences from databases were labelled with the accession number. Bootstrap values from 1,000 replications larger than 50 % are indicated *above branches*. Brackets to the right indicate diverse taxonomic orders as described by Oehl et al. (2011). Phylogenetic tree is rooted with outgroup sequences of *M. verticillata* (AF157145), *C. globosum* (AB048285), *Trichoderma* sp. (EF672340) and *F. oxysporum* (DQ916150). The table beside the phylogenetic tree shows the distribution of the sequences of each OTU among the different management techniques and between soil and roots. The values represent the sampling point pooled together

Only OTUs related to the Glomeraceae clade were found in the root compartment. GLO\_S25 was the only OTU detected in roots under the four different management conditions. GLO\_S22 and GLO\_S24, which were retrieved in the soil for all the different management conditions, were instead present in the roots of plants grown under only three out of the four management conditions. The other six OTUs retrieved in the roots were associated with one or two management conditions. Phylogenetic analysis, Bayesian inference and neighbour-joining, resulted in the same main tree topology.

#### *Large subunit*

The phylogenetic analysis of the LSU rRNA sequences is shown in Fig. 2. The phylogenetic analysis confirmed the exclusive presence of Glomeraceae species, as already suggested by preliminary BLAST analyses. Seven OTU clades can be recognized in the phylogenetic tree receiving bootstrap support  $\geq 96$  %.



(S1 and S0) and in the tilled and N-fertilized plot (T1) roots. Both phylogenetic analysis, Bayesian inference and neighbour-joining, resulted in the same main tree topology.

#### Analysis of AMF diversity and community structure based on the SSU region

The rarefaction curves related to the 97 % sequence identity SSU OTUs detected in the four soils indicate that our sampling efforts yielded a large proportion of the soil AMF communities diversity, at least for the two tilled plots (T1 and T0), whereas the rarefaction curves relative to the non-tilled plots (S0 and S1) show a decreasing accumulation rate of OTUs, but do not reach the asymptote (Fig. 3). This finding was confirmed by the ACE and Chao1 non-parametric richness estimators, which indeed were in the range to the actually observed OTU numbers in the tilled plots (Table 1).

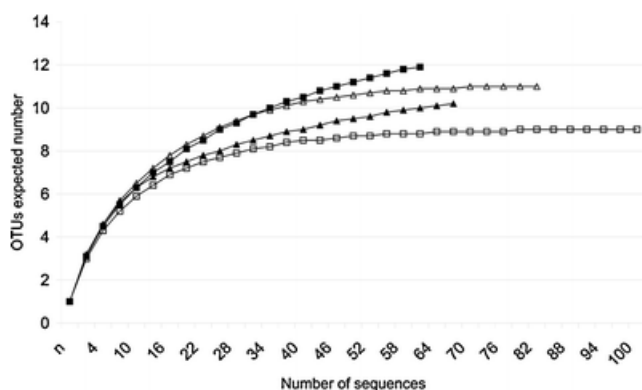


Fig. 3

Rarefaction curves for SSU 97 % sequence identity OTUs detected in S0 (closed square), S1 (closed triangle), T0 (open square) and T1 (open triangle) soils, as determined using DOTUR

Table 1

Observed versus estimated ( $\pm$ SE, based on ACE and Chao1 non-parametric richness estimators) 97 % sequence identity OTU numbers for the four soil management conditions

Soil management	Observed OTUs	Estimated OTUs	
		ACE	Chao1
S0	12	14.2 ( $\pm$ 2.6)	13.5 ( $\pm$ 2.3)
S1	11	11.5 ( $\pm$ 1)	11.2 ( $\pm$ 0.5)
T0	9	9 ( $\pm$ 0)	9 ( $\pm$ 0)
T1	11	11 ( $\pm$ 0)	11 ( $\pm$ 0)

The Shannon–Weaver index, calculated on the SSU data, was not significantly different ( $df=3$ ,  $F=3.205$ ,  $P=0.361$ ) among the four soils under analysis. The values detected were:  $1.59 \pm 0.45$  for S0,  $1.68 \pm 0.34$  for S1,  $1.10 \pm 0.39$  for T0 and  $1.54 \pm 0.24$  for T1.

The CVA analysis is shown in a biplot (Fig. 4), where Axis1 and Axis2 account for 94.72 and 3.47 % of the total variance, respectively. The CVA biplot (Fig. 4) clearly shows that replicates of the same cultural conditions are well grouped together. The analysis discriminates the different

groups mainly on the basis of N fertilization (mainly due to OTUs GLO\_S27, GLO\_S17, GLO\_S24 and GLO\_S23). In detail, OTUs GLO\_S23 and GLO\_S24, belonging to unidentified *Glomus*, seem to be enhanced by N fertilization, whilst GLO\_S27 and GLO\_S17, belonging to the *Funneliformis* clade, by the lack of fertilization. Both Axis1 and Axis2 discriminate between the AMF communities present in the tilled and the no-tilled plots for a given fertilization treatment.

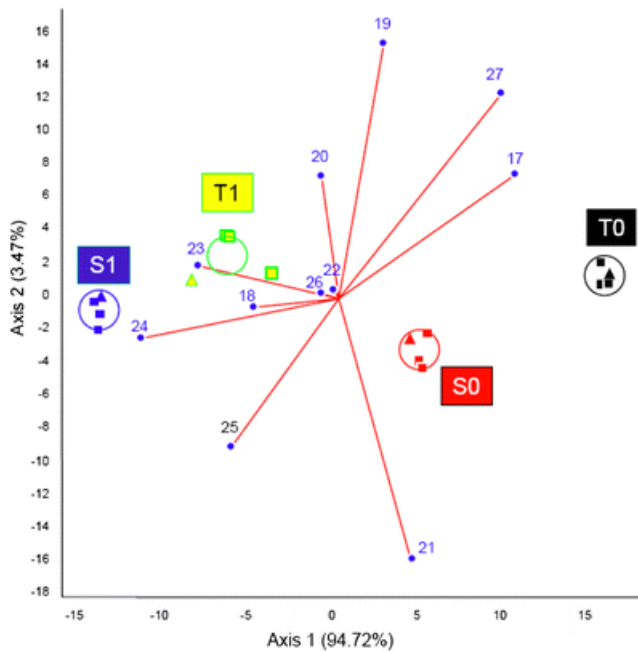


Fig. 4

CVA biplot of the AMF communities found in the soil (*square*) and maize roots (*triangle*) for the four different plots. The ellipses represent 90 % confidence intervals. The percentage of the total variance accounting for each canonical variate is reported. Treatments: tilled and fertilized (*T1*); tilled and non-fertilized (*T0*); no-tilled and fertilized (*S1*); no-tilled and non-fertilized (*S0*). The vectors represent the correlations of SSU OTUs with the canonical axes: GLO\_S17 (17), GLO\_S18 (18), GLO\_S19 (19), GLO\_S20 (20), GLO\_S21 (21), GLO\_S22 (22), GLO\_S23 (23), GLO\_S24 (24), GLO\_S25 (25), GLO\_S26 (26), GLO\_S27 (27)

## Discussion

In this work, the diversity of AMF communities occurring in a maize field subjected to different agricultural management practices was investigated using molecular markers based on rRNA gene regions. Since no primer combination designed until now is able to detect all Glomeromycota fungi in environmental conditions, we decided to use three primer combinations (NS31/AMmix, 28G1/28G2 and GOCC56/GOCC427) in order to detect a wide spectrum of AMF species difficult to cover with a single primer set. The molecular analysis of DNA directly extracted from soil allowed investigating which AMF were present as mycelia and spores in soil avoiding the selection effects due to trap plants or the tedious and time-expensive morphological observation of the spores. To our knowledge, only a few other papers (Pivato et al. 2007; Hempel et al. 2007; Cesaro et al. 2008; Balestrini et al. 2010) have dealt with the analysis of AMF assemblages detected using DNA extracted directly from soil.

Each primer combination contributes to AMF diversity description

Even if the comparison of the different primer set performances was not the main aim of this study, we provide some general considerations to better understand the results. The small

subunit rRNA gene was examined because it is the most widely used marker in field studies analysing AMF communities, also due to the availability of primer sets which are able to detect a wide range of glomeromycotan genera as well as the increasing number of available AMF sequences in GenBank following large-scale 454 pyrosequencing (Öpik et al. 2009; Lumini et al. 2010). In particular, the NS31-AMmix primer set should allow the detection of Glomeraceae, Entrophosporaceae, Diversisporaceae, Acaulosporaceae, Gigasporaceae, Scutellosporaceae, Dentiscutaceae, Racocetraceae and Archaeosporaceae (Santos-González et al. 2007; Schechter and Bruns 2008). As expected, the phylogenetic analyses of 427 SSU AMF sequences obtained from soil and maize roots using this primer set allowed us to identify sequences belonging to Glomeraceae (79 %), Entrophosporaceae (1.4 %), Diversisporaceae (0.8 %) and Gigasporaceae (18.8 %). Lack of detection of Archaeosporaceae in both samples (maize roots and soils) did not probably depend on the primers used since NS31/AM1 successfully amplified many *Archaeospora* sequences from *Collinsia sparsiflora* roots in Californian soils (Schechter and Bruns 2008). On the other hand, whilst *Archaeospora* species have frequently been found in trap cultures, they have never been detected in arable soils (Hijri et al. 2006), which is in agreement with the results of other molecular studies on AMF communities in Central Europe (Daniell et al. 2001; Vandenkoornhuyse et al. 2002; Gollotte et al. 2004; Scheublin et al. 2004; Renker et al. 2005; Börstler et al. 2006).

The second primer set used (28G1/28G2), amplifying a region of the LSU rDNA, was chosen because this region is claimed to exhibit a higher phylogenetic resolution than the SSU, and it has recently been used to design new primer sets for AMF species-level identification (Krüger et al. 2009). Even though this primer set also amplifies *Diversisporales* (da Silva et al. 2006), we only amplified sequences from fungi belonging to *Glomerales*, probably due to a major affinity of the primers towards this group of AMF. Despite the relatively restricted range of sequences detected, the phylogenetic analysis highlighted six well-supported clades inside the Glomeraceae showing different distributions across the different agricultural practices. However, the lack of LSU sequences of reliably identified species in public databases does not allow affiliating these clades to other subgroups already described, with the exception for two of them (*Funneliformis/Septoglomus* subgroup and *Glomus* subgroup).

The third primer set targeting the ITS rDNA region of Paraglomeraceae was used since this family was not detected by the other two primer sets.

#### AMF biodiversity in maize field

AMF diversity, as expressed by Shannon–Weaver diversity indices, was almost consistent for the different soil management conditions (1.10–1.68) and definitely higher than those obtained from other fields studies based on plant root molecular analysis (Daniell et al. 2001; Hijri et al. 2006; Alguacil et al. 2009). Moreover, rarefaction curves, together with ACE and Chao1 indices, suggested that the number of observed OTUs is close to the estimated number, indicating good sampling efficiency for AMF diversity in soil. However, other field studies based on spore isolation and characterization from agricultural soils showed slightly higher Shannon–Weaver diversity indices (Oehl et al. 2010). Even though the results obtained with these two different approaches cannot be easily compared, it is possible that AMF biodiversity based on spore morphological characterization allows catching some morphospecies probably biased by molecular analysis. On the other hand, the molecular approaches allow detecting putative cryptic species not yet characterized as morphospecies because these are present in soil as mycelia and not as spores.

Our results show a predominance of OTUs belonging to the family Glomeraceae, demonstrating that the members of this genus were the main colonizers of the maize field. Of the 18 SSU OTUs belonging to Glomeraceae, only OTU GLO\_A<sub>5</sub>19 could be ascribed to a known isolate, *Glomus* sp. DAOM197198. The other 17 Glomeraceae OTUs found in both soil and maize roots were often closely related to other root-derived sequences from various ecosystems around the world, but none was closely related to any described AMF species in GenBank. The higher phylogenetic resolution power of the LSU region allowed us to recognize seven well-supported subgroups inside the Glomeraceae clade. Two subgroups out of seven were clearly affiliated with *Funneliformis/Septoglomus* and *Glomus*, respectively. Within such subgroups, only two phylotypes, GLO\_L52 and GLO\_L51, could be identified at the species level, as *F. mosseae* and *G. intraradices*, respectively. These OTUs were mainly retrieved from the soil and roots subjected to tillage. The absence, in public databases, of fully identified sequences clustering in the remaining five subgroups (GLOM1, GLOM2, GLOM3, GLOM4, GLOM5) did not allow us to ascribe such subgroups to known species. OTU GLO\_L55, which was comprised within such subgroups, was the most frequent OTU retrieved from the soil (24 % of the total sequences) as well as the main colonizer of maize roots analysed in the S0 treatment along with GLO\_L53 (9.7 % of the total sequence).

The SSU OTU comprising the Entrophosporaceae group (GLO\_S14) was only detected in S0, suggesting a low tolerance to perturbed conditions. Moreover, OTU DIV\_S11, related to *R. fulva*, only seems to be associated with the N-fertilized samples (S1 and T1), even though the low number of sequences comprising this OTU does not allow firm conclusions to be drawn. In addition, OTU GIG\_S26, grouping many sequences related to Gigasporales, was found in soil under all treatments. This finding was rather unexpected since members of this family are not usually detected (either as spores or sequences obtained from crop plants) in agricultural fields upon soil disturbance imposed either through agricultural use or by heavy landscaping machinery (Jansa et al. 2003; Mathimaran et al. 2007), but are in line with the results of Oehl et al. (2005,2010) based on spore identification tools from field samples. The results obtained with the GOCC56/GOCC427 primer pair highlights the difficulty of detecting Paraglomeraceae in root samples, as already reported by (Hijri et al. 2006). In addition, Hempel et al. (2007) observed that these fungi can be dominant in certain soils. In the current study, some sequences related to this taxon were found under all soil management conditions, in particular sequences with high similarity to an undescribed *Paraglomus* species, and to *P. brasilianum*, which has already been reported in maize roots (Millner et al. 2001; Morton and Redecker 2001).

The explorative analysis of the AMF community present in the roots highlighted a strong predominance of sequences belonging to Glomeraceae, ascribable to the most frequent OTUs retrieved in soil. The predominance of species belonging to Glomeraceae detected in soil and roots confirm the well-known characteristics of the member of this family, adaptability and stress tolerance, confirming that they can be retrieved across a wide range of habitats, either natural or agricultural (Öpik et al. 2006).

#### Agricultural practice impacts AMF distribution

The observed dominance of Glomeraceae sequences is in agreement with previous reports concerning agricultural lands (Öpik et al. 2006). For example, Helgason et al. (1998) found a dominance of *F. mosseae* or closely related species, whereas Daniell et al. (2001) mostly found *Funneliformis caledonium–Funneliformis geosporum* sequences. The dominance of Glomeraceae species has also been reported in studies based on spore morphology, *F. mosseae* being the most frequently observed species (Oehl et al. 2003; Cheng and

Baumgartner 2004; Sjöberg et al. 2004; Wang et al. 2008). In other studies, instead, *G. intraradices* was reported as the dominant species especially in heavy textured agricultural soils (Mathimaran et al. 2005; Hijri et al. 2006). The presence and abundance of sequences related to *G. intraradices* and *F. mosseae* in perturbed environments is not surprising in light of the many studies reporting a “ruderal” lifestyle for these species (Oehl et al. 2003, 2004; Hijri et al. 2006). The dominance of a few Glomeraceae species is probably a consequence of the strong selection pressure imposed by agricultural practices which leads to the predominance of fast root-colonizing species (Oehl et al. 2004) and species that are able to tolerate, among others, the repeated disruption of external hyphal networks, periods without mycorrhizal host plants, and the application of fertilizers and fungicides (Gosling et al. 2006). The analysis conducted using the 28G1–28G2 primer set highlighted several different Glomeraceae OTUs that show strong differences in their response to tillage and N fertilization. Although the effects of mineral N on AMF community composition at present are not well understood, a number of studies have indicated that N fertilization can have a negative impact on the functional structures of AMF and on the richness of AMF colonizing plant roots (Santos et al. 2006). As to the relative importance of the different agricultural practices (N fertilization and tillage), CVA indicated that the AMF assemblage structure and composition in the maize field plots examined are primarily influenced by N fertilization and, to a lesser extent, by tillage, as indicated by the shift in the dominant groups of AMF that occur in the different treatments.

In conclusion, the results obtained in the maize fields under study highlighted the stronger effect of N fertilization to shape the AMF communities with respect to tillage. High-input agricultural practices force the selection of AMF species showing generally non-beneficial traits, for example high investment in reproduction at the expense of nutrient scavenging and transfer process that are beneficial for hosts. Instead, a lower selection pressure imposed by a modulation of management practices could allow the presence of AMF species showing beneficial–complementary functional traits necessary to guarantee an improvement of the agroecosystem in which they are present (Verbruggen and Kiers 2010; Barret et al. 2011). This information is a prerequisite to achieve efficient use and manipulation of AMF communities for long-term agricultural stability and productivity in light of a desirable shift from conventional agricultural systems to more sustainable agricultural models.

#### **Note added in proof**

The authors would like to indicate that since the submission of this paper, Krüger M, et al (2012) *New Phytologist* 193:970–984, in March 2012, reported an extensive molecular phylogeny of arbuscular mycorrhizal fungi (AMF) commented on the same issue by P. Young (2012) *New Phytologist* 193:823–826.

#### **Acknowledgements**

The authors would like to thank P.P. Roggero and his team for their contribution to the collection of the soils in the frame of the Italian National Project SOILSINK (FISR). Thanks are also due to A. Berruti for his help in the root analysis, to S. Ghignone for his help in the sequence submission process and to R. Farina for her helpful comments. The P.B. research was funded by the SOILSINK Project (FISR). V.B.’s research was funded by IPP-CNR (Biodiversity Projects–Torino) and the ENDURE project (European Network of Excellence). R.B. was supported by SOILSINK Project grants. We are grateful to the anonymous referees and to the editor for their helpful comments.



## References

- Alguacil MM, Lumini E, Roldán A, Salinas-García JR, Bonfante P, Bianciotto V (2008) The impact of tillage practices on arbuscular mycorrhizal fungal diversity in subtropical crops. *Ecol Appl* 18:527–536
- Alguacil MM, Roldán A, Torres MP (2009) Complexity of semiarid gypsophilous shrub communities mediates the AMF biodiversity at the plant species level. *Microb Ecol* 57:718–727
- Artursson V, Finlay RD, Jansson JK (2006) Interactions between arbuscular mycorrhizal fungi and bacteria and their potential for stimulating plant growth. *Environ Microbiol* 8:1–10
- Balestrini R, Magurno F, Walker C, Lumini E, Bianciotto V (2010) Cohorts of arbuscular mycorrhizal fungi (AMF) in *Vitis vinifera*, a typical Mediterranean fruit crop. *Environ Microbiol Rep* 2:594–604
- Barret M, Morrissey JP, O’Gara F (2011) Functional genomics analysis of plant growth-promoting rhizobacterial traits involved in rhizosphere competence. *Biol Fert Soils* 47:729–743
- Börstler B, Renker C, Kahmen A, Buscot F (2006) Species composition of arbuscular mycorrhizal fungi in two mountain meadows with differing management types and levels of plant biodiversity. *Biol Fert Soils* 42:286–298
- Cesaro P, van Tuinen D, Copetta A, Chatagnier O, Berta G, Gianinazzi S, Lingua G (2008) Preferential colonization of *Solanum tuberosum* L. roots by the fungus *Glomus intraradices* in arable soil of a potato farming area. *Appl Environ Microbiol* 74:5776–5783
- Cheng X, Baumgartner K (2004) Survey of arbuscular mycorrhizal fungal communities in northern California vineyards and mycorrhizal colonization potential of grapevine nursery stock. *HortSci* 39:1702–1706
- Cole JR, Chai B, Marsh TL, Farris RJ, Wang Q, Kulam SA, Chandra S, McGarrell DM, Schmidt TM, Garrity GM, Tiedje JM (2003) The Ribosomal Database Project (RDP-II): previewing a new autoaligner that allows regular updates and the new prokaryotic taxonomy. *Nucleic Acids Res* 31:442–443
- da Silva GA, Lumini E, Maia LC, Bonfante P, Bianciotto V (2006) Phylogenetic analysis of Glomeromycota by partial LSU rDNA sequences. *Mycorrhiza* 16:183–189
- Daniell TJ, Husband R, Fitter AH, Young JPW (2001) Molecular diversity of arbuscular mycorrhizal fungi colonising arable crops. *FEMS Microbiol Ecol* 36:203–209
- Dumbrell AJ, Ashton PD, Aziz N, Feng G, Nelson M, Dytham C, Fitter AH, Helgason T (2011) Distinct seasonal assemblages of arbuscular mycorrhizal fungi revealed by massively parallel pyrosequencing. *New Phytol* 190:794–804
- Edgar RC (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* 32:1792–1797
- Felsenstein J (2005) Using the quantitative genetic threshold model for inferences between and within species. *Philos Trans R Soc Lond B Biol Sci* 360:1427–1434

- Golotte A, van Tuinen D, Atkinson D (2004) Diversity of arbuscular mycorrhizal fungi colonising roots of the grass species *Agrostis capillaris* and *Lolium perenne* in a field experiment. *Mycorrhiza* 14:111–117
- Gosling P, Hodge A, Goodlass G, Bending GD (2006) Arbuscular mycorrhizal fungi and organic farming. *Agric Ecosyst Environ* 113:17–35
- Helgason T, Fitter AH (2009) Natural selection and the evolutionary ecology of the arbuscular mycorrhizal fungi (phylum Glomeromycota). *J Exp Bot* 60:2465–2480
- Helgason T, Daniell T, Husband R, Fitter A, Young J (1998) Ploughing up the wood-wide web? *Nature* 394:431
- Hempel S, Renker C, Buscot F (2007) Differences in the species composition of arbuscular mycorrhizal fungi in spore, root and soil communities in a grassland ecosystem. *Environ Microbiol* 9:1930–1938
- Hijri I, Sykorová Z, Oehl F, Ineichen K, Mader P, Wiemken A, Redecker D (2006) Communities of arbuscular mycorrhizal fungi in arable soils are not necessarily low in diversity. *Mol Ecol* 15:2277–2289
- Huelsenbeck JP, Ronquist F, Nielsen R, Bollback JP (2001) Bayesian inference of phylogeny and its impact on evolutionary biology. *Science* 294:2310–2314
- Jansa J, Mozafar A, Kuhn G, Anken T, Ruh R, Sanders IR, Frossard E (2003) Soil tillage affects the community structure of mycorrhizal fungi in maize roots. *Ecol Appl* 13:1164–1176
- Krüger M, Stockinger H, Krüger C, Schüssler A (2009) DNA-based species level detection of Glomeromycota: one PCR primer set for all arbuscular mycorrhizal fungi. *New Phytol* 183:212–223
- Lumini E, Orgiazzi A, Borriello R, Bonfante P, Bianciotto V (2010) Disclosing arbuscular mycorrhizal fungal biodiversity in soil through a land-use gradient using a pyrosequencing approach. *Environ Microbiol* 12:2165–2179
- Mathimaran N, Ruh R, Vullioud P, Frossard E, Jansa J (2005) *Glomus intraradices* dominates arbuscular mycorrhizal communities in a heavy textured agricultural soil. *Mycorrhiza* 16:61–66
- Mathimaran N, Ruh R, Jama B, Verchot L, Frossard E, Jansa J (2007) Impact of agricultural management on arbuscular mycorrhizal fungal communities in Kenyan ferralsol. *Agric Ecosyst Environ* 119:22–32
- Millner PD, Mulbry WW, Reynolds SL (2001) Taxon-specific oligonucleotide primers for detection of two ancient endomycorrhizal fungi, *Glomus occultum* and *Glomus brasilianum*. *FEMS Microbiol Lett* 196:165–170
- Moora M, Öpik M, Sen R, Zobel M (2004) Native arbuscular mycorrhizal fungal communities differentially influence the seedling performance of rare and common *Pulsatilla* species. *Funct Ecol* 18:554–562
- Moreira-Souza M, Trufem SFB, Gomes-da-Costa SM, Cardoso EJBN (2003) Arbuscular mycorrhizal fungi associated with *Araucaria angustifolia*(Bert.) O. Ktze. *Mycorrhiza* 13:211–215

- Morton JB, Redecker D (2001) Two new families of Glomales, Archaeosporaceae and Paraglomaceae, with two new genera *Archaeospora* and *Paraglomus*, based on concordant molecular and morphological characters. *Mycologia* 93:181–195
- Oehl F, Sieverding E, Ineichen K, Mader P, Boller T, Wiemken A (2003) Impact of land use intensity on the species diversity of arbuscular mycorrhizal fungi in agroecosystems of Central Europe. *Appl Environ Microbiol* 69:2816–2824
- Oehl F, Sieverding E, Mäder P, Dubois D, Ineichen K, Boller T, Wiemken A (2004) Impact of long-term conventional and organic farming on the diversity of arbuscular mycorrhizal fungi. *Oecologia* 138:574–583
- Oehl F, Sieverding E, Ineichen K, Ris E-A, Boller T, Wiemken A (2005) Community structure of arbuscular mycorrhizal fungi at different soil depths in extensively and intensively managed agroecosystems. *New Phytol* 165:273–283
- Oehl F, Laczko E, Bogenrieder A et al (2010) Soil type and land use intensity determine the composition of arbuscular mycorrhizal fungal communities. *Soil Biol Biochem* 42:724–738
- Oehl F, Sieverding E, Palenzuela J et al (2011) Advances in Glomeromycota taxonomy and classification. *IMA FUNGUS* 2:191–199
- Öpik M, Moora M, Liira J, Zobel M (2006) Composition of root-colonizing arbuscular mycorrhizal fungal communities in different ecosystems around the globe. *J Ecol* 94:778–790
- Öpik M, Metsis M, Daniell TJ, Zobel M, Moora M (2009) Large-scale parallel 454 sequencing reveals host ecological group specificity of arbuscular mycorrhizal fungi in a boreonemoral forest. *New Phytol* 184:424–437
- Öpik M, Vanatoa A, Vanatoa E, Moora M, Davison J, Kalwij JM, Reier Ü, Zobel M (2010) The online database MaarjAM reveals global and ecosystemic distribution patterns in arbuscular mycorrhizal fungi (Glomeromycota). *New Phytol* 188:223–241
- Pivato B, Mazurier S, Lemanceau P, Siblot S, Berta G, Mougél C, van Tuinen D (2007) *Medicago* species affect the community composition of arbuscular mycorrhizal fungi associated with roots. *New Phytol* 176:197–210
- Pozo MJ, Azcón-Aguilar C (2007) Unraveling mycorrhiza-induced resistance. *Curr Opin Plant Biol* 10:393–398
- Redecker D, Kodner R, Graham LE (2000) Glomalean fungi from the Ordovician. *Science* 289:1920–1921
- Renker C, Heinrichs J, Kaldorf M, Buscot F (2003) Combining nested PCR and restriction digest of the internal transcribed spacer region to characterize arbuscular mycorrhizal fungi on roots from the field. *Mycorrhiza* 13:191–198
- Renker C, Blanke V, Buscot F (2005) Diversity of arbuscular mycorrhizal fungi in grassland spontaneously developed on area polluted by a fertilizer plant. *Environ Pollut* 135:255–266
- Santos JC, Finlay RD, Tehler A (2006) Molecular analysis of arbuscular mycorrhizal fungi colonising a semi-natural grassland along a fertilisation gradient. *New Phytol* 172:159–168

- Santos-González JC, Finlay RD, Tehler A (2007) Seasonal dynamics of arbuscular mycorrhizal fungal communities in roots in a seminatural grassland. *Appl Environ Microbiol* 73:5613–5623
- Schechter SP, Bruns TD (2008) Serpentine and non-serpentine ecotypes of *Collinsia sparsiflora* associate with distinct arbuscular mycorrhizal fungal assemblages. *Mol Ecol* 17:3198–3210
- Scheublin TR, Ridgway KP, Young JPW, van der Heijden MGA (2004) Nonlegumes, legumes, and root nodules harbor different arbuscular mycorrhizal fungal communities. *Appl Environ Microbiol* 70:6240–6246
- Schloss PD, Handelsman J (2005) Introducing DOTUR, a computer program for defining operational taxonomic units and estimating species richness. *Appl Environ Microbiol* 71:1501–1506
- Schüßler A, Schwarzott D, Walker C (2001) A new fungal phylum, the Glomeromycota: phylogeny and evolution. *Mycol Res* 105:1413–1421
- Simon L, Lalonde M, Bruns TD (1992) Specific amplification of 18S fungal ribosomal genes from vesicular–arbuscular endomycorrhizal fungi colonizing roots. *Appl Environ Microbiol* 58:291–295
- Sjöberg J, Persson P, Mårtensson A, Mattsson L, Adholeya A, Alström S (2004) Occurrence of Glomeromycota spores and some arbuscular mycorrhiza fungal species in arable fields in Sweden. *Acta Agr Scand B-S P* 54:202–212
- Smith SE, Read DJ (2008) *Mycorrhizal symbiosis*, 3rd edn. Academic, London
- Tamura K, Dudley J, Nei M, Kumar S (2007) MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol Biol Evol* 24:1596–1599
- Toljander JF, Santos-González JC, Tehler A, Finlay RD (2008) Community analysis of arbuscular mycorrhizal fungi and bacteria in the maize mycorrhizosphere in a long-term fertilization trial. *FEMS Microbiol Ecol* 65:323–338
- Trouvelot A, Kough JL, Gianinazzi-Pearson V (1986) Mesure du taux de mycorhization VA d'un système racinaire. Recherche de méthodes d'estimation ayant une signification fonctionnelle. In: Gianinazzi-Pearson V, Gianinazzi S (eds) *Physiological and genetical aspects of mycorrhizae*. INRA Press, Paris, pp 217–221
- van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, Wiemken A, Sanders IR (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* 396:69–72
- Vandenkoornhuyse P, Baldauf SL, Leyval C, Straczek J, Young JPW (2002) Extensive fungal diversity in plant roots. *Science* 295:2051
- Verbruggen E, Kiers ET (2010) Evolutionary ecology of mycorrhizal functional diversity in agricultural systems. *Evol Appl* 3:547–560
- Verbruggen E, Röling WFM, Gamper HA, Kowalchuk GA, Verhoef HA, Van der Heijden MGA (2010) Positive effects of organic farming on below-ground mutualists: large-scale comparison of mycorrhizal fungal communities in agricultural soils. *New Phytol* 186:968–979

Wang Y, Vestberg M, Walker C, Hurme T, Zhang X, Lindström K (2008) Diversity and infectivity of arbuscular mycorrhizal fungi in agricultural soils of the Sichuan Province of mainland China. *Mycorrhiza* 18:59–68

Zhang Z, Schwartz S, Wagner L, Miller W (2000) A greedy algorithm for aligning DNA sequences. *J Comput Biol* 7:203–214