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Studies of Cysteine 298 Role in [FeFe]-hydrogenases

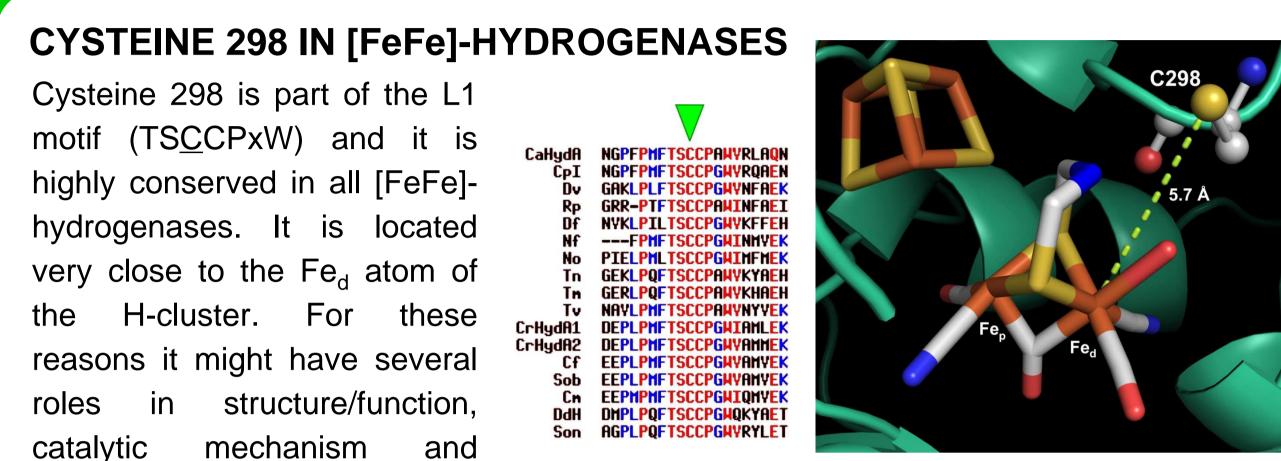
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INTRODUCTION

Cysteine 298 in Clostridium acetobutylicum HydA hydrogenase (CaHydA) is a highly conserved residue in all [FeFe]-hydrogenases and it is one of the residues that are closer to the Fe_d atom of the H-cluster. It has previously been proposed to be involved in the catalytic cycle during proton transfer and to be an important structural element of the cavity that hosts the active site H-cluster.

In order to obtain a comprehensive study of its role, a semi-random approach was used. C298 was studied by site saturation mutagenesis, to simultaneously evaluate the functional effect of cysteine replacement with all the other 19 aminoacids. A specific activity screening protocol was developed in order to test the effect of the mutations directly in libraries composed of a large number of *E. coli* clones and the most relevant variants have been characterised.



PURIFICATION AND CHARACTERISATION OF SELECTED VARIANTS

Five variants were selected on the basis of the screening results and structural similarity and compared to the *wild type* enzyme. Recombinant expression in *E. coli* and affinity purification were accomplished under strict anaerobic conditions taking special care to decrease variability.

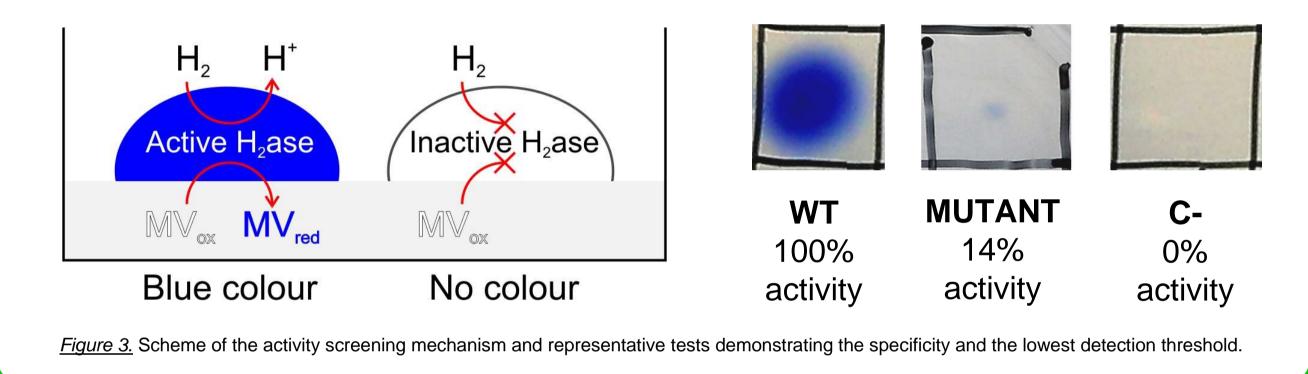
catalytic mechanism oxygen sensitivity.

Figure 1. Sequence alignment of various [FeFe]-hydrogenases.

Figure 2. Structure model of CaHydA with the H-cluster and cysteine 298.

ACTIVITY SCREENING PROTOCOL

A specific screening protocol was set up to directly assay [FeFe]-hydrogenase activity in *E. coli* colonies. The screening takes advantage of methyl viologen (MV) reduction coupled to H_2 uptake by the enzyme. The specificity and the lowest detection threshold were also tested.



RANDOM LIBRARY GENERATION AND SCREENING

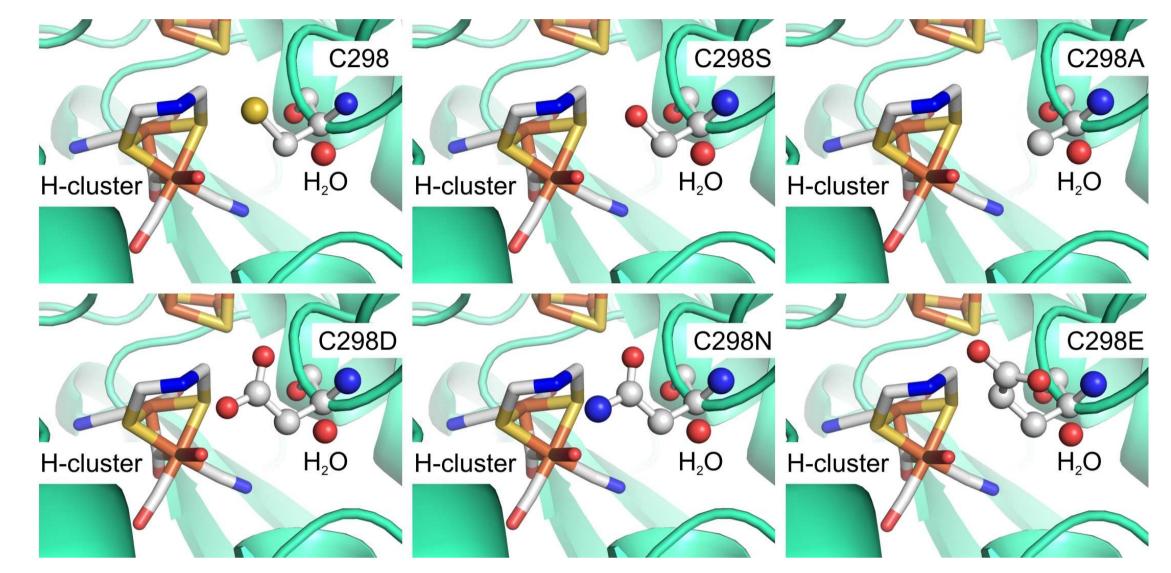
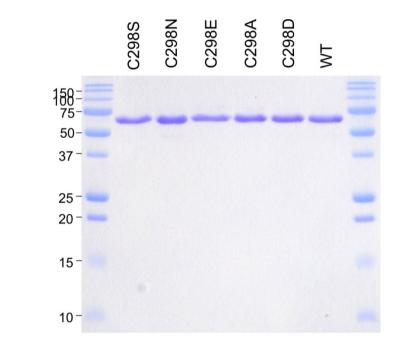


Figure 7. Summary of the 5 variants selected and the wild type enzyme. Models of the structures were obtained using PyMol.



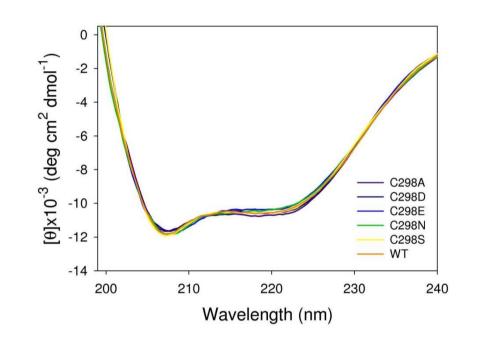


Figure 8. Coomassie stained SDS-PAGE of CaHydA WT and the 5 selected variants (65.5 kDa).

Figure 9. Far UV circular dichroism spectra, showing no major structural effect due to the mutation in the selected variants.

The secondary structure was not importantly affected, as determined by circular dichroism spectroscopy in the far UV region.

The site saturation library was generated using an NNK degenerated codon, thus allowing the generation of 32 possible DNA variants, encoding for all the 20 aminoacids and requiring a reasonably low screening effort. Screening of 315 clones allowed to cover all the possible variants with a probability of completeness of 99.79%.

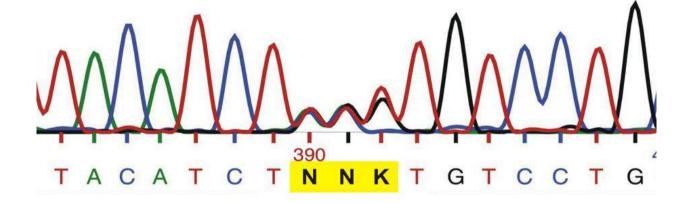


Figure 4. DNA sequencing of the plasmid library showing the expected randomisation

The screening revealed that most CaHydA variants did not show any detectable hydrogenase activity, while some showed *wild type*-like signals. No clones with an intermediate signal could be observed. DNA sequencing revealed that the active clones were variants encoding for cysteine (codon TGT) or aspartic acid (codon GAT) only.

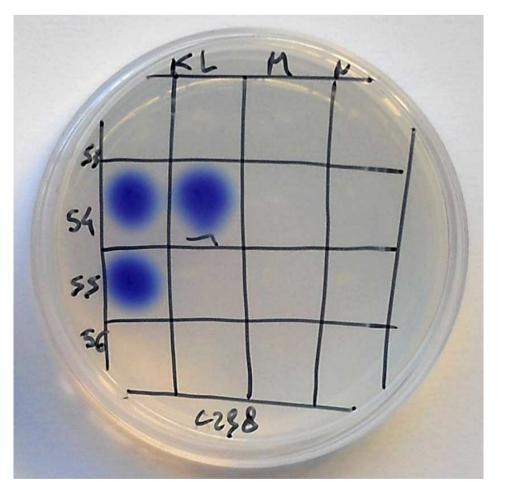


Figure 5. Representative set of the screened clones including CaHydA WT as a control (L54).

Codon	Amino acid	Clones	Frequency	Screen	Sequence	Codon	Amino acid	Clones	Frequency	Screen
GAT	Aspartic acid	8	2.5%	Active		CAT	Histidine	-	-	Negative
TGT	Cysteine	9	2.9%	Active		AAG	Lysine	-	-	Negative
AGT	Serine	-	-	Negative	Č Ğ Ť	CGT	Arginine	a 	-	Negative
TCG	Serine	-	-	Negative	A t G	ATG	Methionine	-	-	Negative
TCT	Serine	-	-	Negative	Λ_{\uparrow}	TTT	Phenylalanine	-	-	Negative
AAT	Asparagine	-	-	Negative		GCT	Alanine	-	-	Negative
	GAT TGT AGT TCG TCT	GATAspartic acidTGTCysteineAGTSerineTCGSerineTCTSerine	GATAspartic acid8TGTCysteine9AGTSerine-TCGSerine-TCTSerine-	GATAspartic acid82.5%TGTCysteine92.9%AGTSerineTCGSerineTCTSerine	GATAspartic acid82.5%ActiveTGTCysteine92.9%ActiveAGTSerineNegativeTCGSerineNegativeTCTSerineNegative	GATAspartic acid82.5%ActiveTGTCysteine92.9%ActiveAGTSerineNegativeTCGSerineNegativeTCTSerineNegative	GATAspartic acid82.5%Active \bigwedge_{CAT} CATTGTCysteine92.9%Active \bigwedge_{AAG} AAGAGTSerineNegative \bigwedge_{CGT} CGTTCGSerineNegative \bigwedge_{ATG} ATGTCTSerineNegative \bigwedge_{TTG} TTT	GATAspartic acid82.5%Active $\bigwedge_{C A T}$ CATHistidineTGTCysteine92.9%Active $\bigwedge_{A A G}$ AAGLysineAGTSerineNegative $\bigwedge_{A A G}$ CGTArginineTCGSerineNegative $\bigwedge_{A T G}$ ATGMethionineTCTSerineNegative $\bigwedge_{A T G}$ TTTPhenylalanine	GATAspartic acid82.5%Active $\bigwedge_{C A \uparrow}$ CATHistidine-TGTCysteine92.9%Active $\bigwedge_{A \downarrow A \downarrow G}$ AAGLysine-AGTSerineNegative $\bigwedge_{C \downarrow G \uparrow}$ CGTArginine-TCGSerineNegative $\bigwedge_{A \uparrow J \downarrow G}$ ATGMethionine-TCTSerineNegative $\bigwedge_{A \uparrow J \downarrow G}$ ATGMethionine- $\bigwedge_{A \uparrow J \downarrow G}$ SerineNegative $\bigwedge_{A \uparrow J \downarrow G}$ TTTPhenylalanine-	GATAspartic acid82.5%Active \bigwedge_{CAT} HistidineTGTCysteine92.9%Active \bigwedge_{AAG} AAGLysineAGTSerineNegative \bigwedge_{CGT} CGTArginineTCGSerineNegative \bigwedge_{ATG} ATGMethionineTCTSerineNegative \bigwedge_{TTT} TTTPhenylalanine

The enzymatic activity assays confirmed the results obtained during the library screening: only C298D had a remarkable activity (50% in comparison to the WT). C298E and C298S showed very little residual activity, while C298N and C298A did not show any detectable activity. Also, C298D caused a shift in the pH activity profile towards acidic conditions.

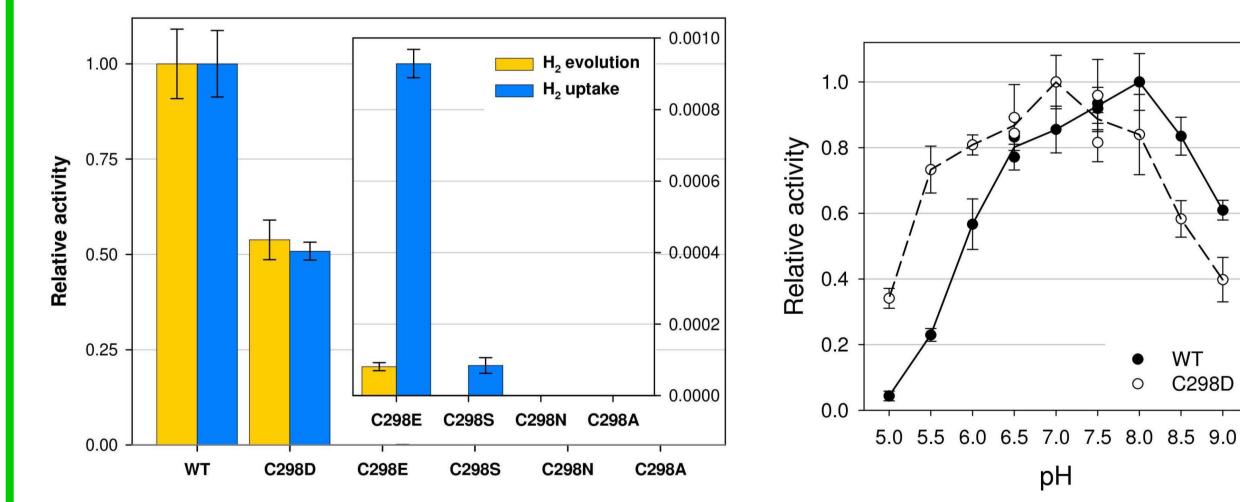


Figure 10. Relative hydrogenase activity with methyl viologen as artificial electron mediator. H₂ evolution was assayed by gas chromatography; H₂ uptake was assayed spectrophotometrically. Relative activity was calculated as the ratio with the WT activity.

Figure 11. Hydrogen evolution activity dependence of the pH.

CONCLUSIONS AND FUTURE PERSPECTIVES

The exclusive functional replacement of cysteine with aspartic acid, an ionisable residue, and the shift in the pH activity profile demonstrate experimentally the central role of C298 in the proton transfer pathway to the active site during [FeFe]-hydrogenase catalysis. Since C298 is strongly conserved among all the known functional [FeFe]hydrogenases, it is reasonable that the results obtained here for CaHydA have broader implications regarding its functional significance in the entire class.

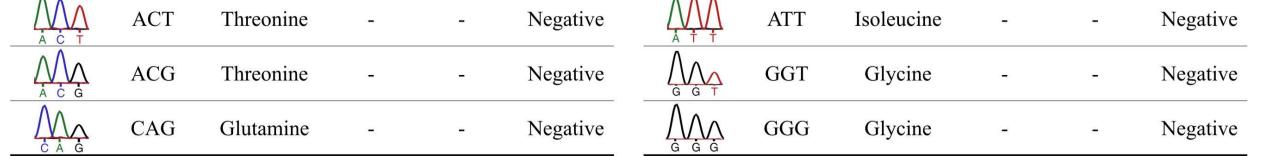


Figure 6. DNA sequencing of all the active clones and of a representative subset of negative clones, demonstrating the diversity of screened clones.

Future work will aim at a more detailed characterisation of the most relevant variants, application of site saturation mutagenesis to other residues within the enzyme core and study of other [FeFe]-hydrogenases.

REFERENCES

Cornish AJ et al., 2011, J. Biol. Chem., 286, 38341-38347. Knörzer P et al., 2012, J. Biol. Chem., 287, 1489-1499. Morra S et al., 2012, PLoS ONE, 7, e48400. Peters JW et al., 1998, Science, 282, 1853-1858. Vignais PM et al., 2007, Chem. Rev., 107, 4206-4272.

King PW et al., 2006, J. Bacteriol., 188, 2163-2172. Lautier T et al., 2011, Faraday Discuss., 148, 385-407. Patrick WM et al., 2003, Protein Eng., 16, 451-457. Reetz MT et al., 2007, Nat. Protoc., 2, 891-903.

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