

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

**CLINICAL IMPACT OF IMMUNOPHENOTYPIC REMISSION AFTER
ALLOGENEIC HEMATOPOIETIC CELL TRANSPLANTATION IN MULTIPLE MYELOMA**

This is a pre print version of the following article:

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1507616> since 2015-08-27T06:58:08Z

Published version:

DOI:10.1038/bmt.2014.319

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)



UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

Questa è la versione dell'autore dell'opera:

*[Bone Marrow Transplant. 2015 Apr;50(4):511-6. doi:
10.1038/bmt.2014.319. Epub 2015 Feb 9]*

The definitive version is available at:

La versione definitiva è disponibile alla URL:

[<http://www.nature.com/bmt/journal/v50/n4/full/bmt2014319a.html>]

**CLINICAL IMPACT OF IMMUNOPHENOTYPIC REMISSION AFTER
ALLOGENEIC HEMATOPOIETIC CELL TRANSPLANTATION IN MULTIPLE
MYELOMA**

Short title: Immunophenotypic remission post allografting

Luisa Giaccone MD,¹ Lucia Brunello MD,¹ Moreno Festuccia MD,¹ Milena Gilestro PhD,¹
Enrico Maffini MD,¹ Federica Ferrando MD,¹ Elisa Talamo, MD,¹ Roberto Passera PhD,²
Mario Boccadoro MD,¹ Paola Omedè PhD,¹ Benedetto Bruno MD, PhD.¹

¹ Division of Hematology, Azienda Ospedaliera Universitaria Città della Salute e della
Scienza and Dipartimento di Biotecnologie Molecolari e Scienze per la Salute, University of
Torino, Torino, Italy ² Division of Nuclear Medicine, Azienda Ospedaliera Universitaria Città
della Salute e della Scienza and University of Torino, Torino, Italy

Address correspondence to: Luisa Giaccone, M.D.

Division of Hematology

A.O. Città della Salute e della Scienza di Torino

Via Genova 3, 10126, TORINO, Italy

E-mail: luisa.giaccone@unito.it

Phone: +39-011-6334354

Fax: +39-011-6963737

Conflict of interest: The Authors declare no potential conflicts of interest relevant to the
article.

1

2 **ABSTRACT**

3 Immunophenotypic remission (IR) is a strong prognostic factor in myeloma patients. The
4 combination of IR and conventional complete remission (CR) was retrospectively evaluated in 66
5 patients after allografting. IR was defined as absence of monoclonal plasma cells in bone marrow
6 aspirates by multiparameter flow-cytometry. Conditioning was non-myeloablative in 55 patients;
7 reduced-intensity in 10 and myeloablative in 1 patient. The allograft was given upfront in 35/66
8 (53%) patients. After a median follow-up of 7.1 years, 24 patients achieved both CR and IR (CR/IR
9 group), 21 achieved IR but not CR with persistence of urine/serum M-component (noCR/IR group),
10 and 21 did not achieve either CR or IR (noCR/noIR group). Median overall survival (OS) and event
11 free survival (EFS) were “not reached” and 59 months in the CR/IR group; 64 and 16 months in the
12 noCR/IR; and 36 and 6 months in the noCR/noIR respectively ($p<0.001$). Cumulative incidence of
13 extra-medullary disease was 4,4 % in the CR/IR, 38,1% in the noCR/IR and 14,3% in the
14 noCR/noIR groups respectively at 4 years ($p<0.001$). IR was a valid tool to monitor residual disease
15 after allografting and allowed to define a cohort of patients at higher incidence of extra-medullary
16 relapse.

17

18
19
20

INTRODUCTION

The increment in response rates of recent years, longer life expectancy and several treatment options in patients with multiple myeloma (MM) have drawn particular attention to the importance of an in-depth evaluation of "complete remission" and the interest in the assessment of minimal residual disease (MRD) has been growing [1-3]. Two sensitive techniques are currently employed to evaluate MRD: qualitative and quantitative polymerase chain reaction (PCR)-based methods and multi-parameter flow-cytometry. PCR-based methods have been of great value in predicting clinical outcomes in MM patients following allografting [4-7], though expensive and labor-intensive, they are characterized by higher sensitivity. A patient-specific molecular marker is, however, detected in only 60% to 70% of patients. The evaluation of MRD through immunophenotyping is more broadly applicable in the MM patients population than PCR as it involves the identification of phenotypic aberrancies in myelomatous plasma cells, which are seen in more than 90% of MM patients. However, the antibody panels used for immunophenotype analysis consist of markers recommended by experts' opinions, and only recently attempts to validate and standardize them have been made [8].

MRD studies using flow-cytometry have so far been carried out on patients treated with autologous transplantation, conventional chemotherapy or new drugs [9-13]. MRD studies in the setting of allografting are however lacking. Here, we report an analysis on the achievement of immunophenotypic response (IR) after an allograft and its prognostic impact when combined with conventional complete remission (CR).

MATERIAL AND METHODS

Patients Between January 2000 and December 2011, 80 consecutive MM patients underwent an allograft at our Center. Sixty-nine out of 80 (median age 54 years, range 35-66), with a follow-up of at least 3 months were included in this study. Three were excluded from this analysis because of early

46 treatment related death at 4 months post-transplant (no. 2), and because of incomplete data (no. 1).
47 Overall, 66 patients were included and their characteristics are summarized in Table 1. Median time from
48 diagnosis to allogeneic transplant was 13.2 months (range 6.6-101 months). Thirty-five out of 66 (43%)
49 were treated at diagnosis according to a planned tandem “auto/allo” program and were also included in
50 previously published prospective clinical trials [14-17]. All patients provided written informed consent to
51 the proposed treatment and to the use of medical records for research purposes. The present study was
52 approved by the Institutional Review Board of our Center and conducted according to the Declaration of
53 Helsinki (NCT01440556).

54

55 **Graft-vs.-host disease** Acute graft-vs.-host disease (GVHD) was diagnosed according to the
56 recent indications of the National Institute of Health [18]. Chronic GVHD was graded as previously
57 described [19].

58

59 **Response Assessment** Disease response was assessed by urine and serum immune-fixation and
60 bone marrow aspirates at 3, 6, 12, 18, 24 months after allografting, and yearly thereafter. Whole-body
61 conventional radiography or magnetic resonance imaging were performed yearly or as clinically indicated
62 (overt relapse or complaints of bone pain). Disease response and disease relapse were defined according
63 to the European Bone Marrow Transplantation Group criteria [20]. Achievement of CR was defined as
64 the absence of monoclonal component by immunofixation on both serum and urine, disappearance of any
65 soft tissue plasmacytoma and less than 5% plasma cells in the bone marrow. The incidence of extra-
66 medullary disease (EMD) in first relapse post allografting was monitored, and EMD was defined as
67 previously described [21].

68 First pulls of bone marrow samples had to contain at least 13000 cells/uL for flow-cytometry
69 MRD studies. Plasma cells quantification was obtained by 4 to 6-colour staining with the following
70 monoclonal antibodies: CD38, CD138, CD56, CD19, CD45, cyKappa, cyLambda. A FACSCanto II

71 Flow-cytometer equipped with FACSDiva software (BD Biosciences, San José, CA) was used. A total of
72 1×10^6 events were acquired and analyzed for each sample, as previously reported [22]. Flow-cytometry
73 analysis had a sensitivity of 10^{-4} cells [23]. IR was defined as less than 0.01% monoclonal plasma cells in
74 the bone marrow sample.

75 Assessment of CR and IR was done at best response. According to the achievement of CR and/or
76 IR, patients were divided into 4 groups: those who achieved CR and IR (CR/IR), those who obtained CR
77 but not IR (CR/noIR), those not in CR but in IR (noCR/IR) and those who did not achieve either CR or IR
78 (noCR/noIR). Time to CR and IR was evaluated excluding patients who were in CR and IR at the time of
79 transplant, respectively.

80

81 **Genetic abnormalities** Although single evaluation of chromosome 13 deletion (del(13)) is no
82 more considered an optimal prognostic marker, it still has value as it is frequently associated with t(4;14),
83 del(17) or t(14;20). Thus patients presenting del(13) with/without other cytogenetic aberrations were
84 considered as at high risk [24].

85

86 **Statistical methods** Primary endpoints were overall survival (OS) and event free survival (EFS)
87 in the 4 patient cohorts defined by the achievement of CR and/or IR. OS was defined as time from
88 transplant to death by any cause, and EFS as the time from transplant to progression/relapse/death as a
89 result of any cause, whichever occurred first. Alive patients were censored as of October 1st, 2013. OS
90 and EFS curves were estimated by the Kaplan-Meier method and compared using the log-rank test. OS
91 and EFS were then analyzed by the univariate and multivariate Cox proportional hazards model,
92 comparing by the Wald test the following risk factors: age at diagnosis (>55 vs. ≤55 years), gender (male
93 vs. female), year of diagnosis (2008-2011 vs. 2004-2007 vs. 2000-2003), number of chemotherapy
94 regimens (≥ 2 vs. 1), ISS (stage III vs. I-II), Durie and Salmon stage (IIIA-IIIB vs. IA-IB-IIA-IIB), donor
95 gender (male vs. female), donor type (matched unrelated donor vs. sibling donor), cytogenetic profile

96 (high risk vs. standard risk), EMD [21] in the clinical course before allografting, occurrence of acute and
97 chronic GVHD (any vs. none) and disease response (CR/IR, CR/noIR, noCR/IR, noCR/noIR). Six- and
98 twelve-month landmark analyses were performed to estimate survival by disease response. The
99 occurrence of acute and chronic GVHD and post-transplant IR and CR were treated as time-dependent
100 variables. Cumulative incidences of developing acute GVHD, chronic GVHD, overall relapse and
101 extramedullary relapse were estimated by the Gray test to compare the cumulative incidence curves of the
102 main event, in the presence of a competing event (defined as death without acute or chronic GVHD or
103 relapse occurred before the development of acute or chronic GVHD for acute and chronic GVHD, as
104 death without previous relapse for overall relapse, as death without previous extramedullary relapse or
105 occurrence of bone relapse for extramedullary relapse) [25]. Non-relapse mortality (NRM) was defined
106 as death without previous relapse [25]. Patient characteristics were tested using the Fisher's exact test for
107 categorical variables and the Mann-Whitney test for continuous ones. All reported p-values were two-
108 sided, at the conventional 5% significance level. Data were analyzed as of January 2014 by IBM SPSS
109 21.0.0 (Chicago-IL, USA) and R 2.15.2 package cmprsk (The R Foundation for Statistical Computing,
110 Wien-A).

112 RESULTS

113 **Study population** At diagnosis all patients presented with measurable disease and 12 out of 66
114 (18%) with EMD. Thirty-five out of 66 received the allograft as part of their first line treatment, whereas
115 the remaining (31/66, 46%) were transplanted at relapse (Table 1). In 2/31 (6%) EMD presented at
116 relapse before the allograft. Conditionings are summarized in Table 1. Post-grafting immuno-suppression
117 consisted of calcineurine inhibitors (cyclosporine or tacrolimus) and mycophenolate mofetil in 60 (91%),
118 and cyclosporine and methotrexate in the remaining. Patients did not receive maintenance therapies or
119 donor lymphocyte infusion post allograft until relapse, with the exception of 6 recent patients who started
120 lenalidomide at six months post transplant as per protocol. Due to the rather long study period, fluorescent

121 in situ hybridization (FISH) was performed in only 20 (30%) patients: del(13) aberration was detectable
122 in 6 patients, 1 patient presented del(13) associated with del(17) and t(4;14) and 1 patient resulted positive
123 for t(4;14); the remaining 12 patients were negative for del(13).

124 All patients had suitable bone marrow aspirates for IR evaluation.

125
126 **Non-relapse mortality and GVHD** NRM of the overall population of 80 patients was 13.8% at 1
127 and 3 years, 15.3% at 5 years. In the 66 patients who survived at least 3 months and formed the study
128 population, NRM was 6.1%, 9.1% and 10.8% at 1, 3 and 5 years respectively. After a median follow-up
129 of 7.1 years (range 2.6-13.2), the incidence of acute and chronic GVHD was 44.6% and 52.4%. Patients
130 transplanted at relapse developed more acute GVHD ($p=0.03$), whereas those transplanted upfront
131 developed more chronic GVHD ($p=0.034$). Overall, main cause of death was disease relapse in both
132 patients transplanted upfront and at relapse.

133
134 **Disease response and relapse** At the time of the allograft, 9 (14%) patients were in CR and 21
135 (32%) in IR, 5 of these were both in CR and IR. After the allograft, all 21 IR patients remained in IR and
136 25 additional patients entered IR for a total of 45/66 (68%), whereas 24/66 (36%) patients achieved CR,
137 of whom only 7 were in CR pre-transplant. Median time to IR was 7 months (range 1-48, no. 23),
138 whereas median time to CR was 8 months (range 1-60, no. 17). Among the 45 patients who achieved IR,
139 26 performed Magnetic Resonance Imaging (MRI) and 2 Computerized Tomography (CT) scans of the
140 spine at the time of best response, 14 were in the CR/IR group and 14 in the noCR/IR group. Only 4 out
141 of 28 MRI/TC scans showed myeloma infiltration, and all in the noCR/IR group. Seventeen patients in IR
142 did not perform any MRI/CT scan. Twenty-one patients showed discrepant results with persistent serum
143 and/or urine monoclonal component despite the absence of monoclonal marrow plasma cells. Overall,
144 patients were divided into the following cohorts: 24 in CR and IR (CR/IR group); 21 in IR but not CR
145 (noCR/IR group); 21 in neither CR nor IR (noCR/noIR group). No patient was in CR but not in IR

146 (CR/noIF group) (Table 2). Among patients in the CR/IR group, 5/24 only achieved CR before IR. Given
147 the small cohort an analysis could not be carried out. Patients in the 3 cohorts were equally balanced for
148 age, year of transplant, disease stage, median β 2microglobulin, number of previous therapies,
149 conditioning, donor gender and type. Conditioning regimen and acute GVHD were not correlated with
150 disease response group ($p=0.703$ and $p=0.282$, respectively), whereas chronic GVHD ($p=0.047$) and
151 previous therapy lines ($p=0.015$) were.

152 Overall, at follow up, cumulative incidence of disease relapse was 32%, 50% and 62% at
153 1, 3 and 5 years, respectively. At the same time-points, it was higher in the noCR/noIR group (67%, 81%,
154 not applicable) as compared with the noCR/IR group (33%, 62%, 72%) and with the CR/IR group (0%,
155 13%, 30%, $p<0.001$). Among patients who achieved IR, median time to clinical relapse post-transplant
156 was 9.7 months in the noCR/IR group and 30 months in the CR/IR one. The overall incidence of extra-
157 medullary first relapse was 9%, 15% and 20% at 1, 3 and 5 years, respectively. At the same time-points, it
158 was 5%, 14%, not applicable in the noCR/noIR; 24%, 33%, 44% in the noCR/IR; and 0%, 0%, 4% in the
159 CR/IR group ($p<0.001$) (Figure 1). Sites of EMD are reported in Table 3. Fourteen (12 at diagnosis and 2
160 at pre-transplant relapse) out of 66 (21%) developed EMD before the allograft. However, only 3 of these
161 14 were among those who experienced EMD after the allograft.

162
163 **Clinical outcomes** Overall, after a median follow-up of 7.1 years (range 2.6-13.2), median OS
164 and EFS were 5.5 and 1.4 years respectively. In patients in CR, median OS and EFS were not reached and
165 59 months as compared with 40 and 9 months in those not in CR ($p<0.001$). Median OS and EFS in
166 patients who achieved IR were 96 and 41 months as compared with 36 and 6 months in those who did not
167 ($p<0.001$). Landmark analysis showed that being in IR at six months post-transplant was not statistically
168 associated with better OS and EFS (7.5 vs. 5.0 years. $p=0.132$ and 4.1 vs. 1.2 years $p=0.065$,
169 respectively), whereas IR at 12 months post-transplant conferred an advantage in OS (10.3 vs. 2.4
170 $p=0.018$) but not in EFS (3.6 vs. 1 year, $p=0.634$).

171 By patient cohort, median OS and EFS were not reached and 59 months in the CR/IR cohort, 64
172 and 16 months in the noCR/IR cohort, and 36 and 6 months in the noCR/noIR cohort respectively
173 ($p < 0.001$, both for OS and EFS) (Figure 1). Among patients not in CR, there was a significant advantage
174 in EFS and a trend for better OS for those who reached IR compared to the noCR/noIR group ($p = 0.001$
175 and $p = 0.063$, respectively).

176 With the limitations of the small sample size (only 20 patients evaluated), OS in high risk patients
177 by FISH analysis was 39 months as compared with “not reached” in standard risk patients ($p = 0.009$),
178 whereas EFS was not statistically significant (19 months vs. 64 months, $p = 0.097$).

179 All patients with EMD at first relapse (no.13) after the allograft eventually died of disease
180 progression. OS in patients first relapsed with EMD was significantly shorter than in those relapsed
181 without EMD (39 vs. 57 months, $p = 0.034$). By contrast, there was no difference in OS and EFS between
182 newly diagnosed patients with EMD and those without.

183 By univariate and multivariate analysis, belonging to the CR/IR cohort was the only significant
184 predictor for prolonged OS and EFS ($p < 0.001$) (Table 4, Table 5).

186 DISCUSSION

187 MRD analysis is currently used for evaluating treatment efficiency and patient risk stratification in
188 several hematological malignancies [26]. In MM, not only is MRD of primary importance to assess tumor
189 shrinkage, but it is now regarded as one of the strongest prognostic predictors, irrespective of any given
190 treatment. MRD analysis by multicolor flow-cytometry has been introduced in many clinical trials on
191 myeloma. The prognostic impact of achieving IR has been described after conventional chemotherapy,
192 autografting and, more recently, after new drugs [9-13].

193 Despite some limitations due to its retrospective nature, our study underlines the clinical
194 importance of achieving IR also after allografting (Figure 1). Post-transplant IR was associated with

195 significantly better OS and EFS. Landmark analyses suggested that IR at 12 months post allografting had
196 greater impact on OS than IR at 6 months. This might be explained by an ongoing and/or late occurrence
197 of graft-vs-myeloma effect. However, in patients in IR, clinical outcomes were different in the light of CR
198 status. OS and EFS were not reached and 59 months in the CR/IR group, and 64 and 16 months in the
199 noCR/IR group, respectively ($p<0.007$ and $p<0.014$, Figure 1). To stress the role of IR, we also observed
200 that patients in noCR/IR showed an intermediate clinical outcome compared with those in CR/IR and in
201 noCR/noIR (Figure 1). IR and CR status was the only variable significantly associated with improved OS
202 and EFS by multivariate analysis ($p=0.001$), whereas GVHD, the number of previous therapy lines,
203 conditioning regimen, and year of transplant were not (Table 4, Table 5). Other authors reported similar
204 outcomes between patients who were MRD negative but not in CR and those MRD positive [13]. Paiva et
205 al. [11] reported 21% of patients in IR with persistent positive immunofixation after autografting.
206 Moreover, progression free survival (PFS) was progressively shorter, 71, 65, and 37 months, in patients in
207 IR/CR, in IR/noCR and in noIR/CR respectively ($p=0.001$). This study clearly showed that the
208 achievement of remission by flow-cytometry had a higher prognostic value than remission by
209 immunofixation. In our study, we cannot draw such a definitive conclusion on the role of IR, given the
210 lack of patients in CR but not in IR.

211 The discrepancy between IR and not CR was observed in 32% of our transplant patients. This
212 finding may be explained by a number of reasons. It may partly be argued that bone marrow aspirates do
213 not systemically represent the marrow status and areas of marrow disease may persist. It may however be
214 more plausible that residual extra-medullary plasma cells continue secreting monoclonal
215 immunoglobulins in sanctuary sites where agents with anti-myeloma activity and/or a potential *graft-vs.-*
216 *myeloma* may have little or slower effect. This hypothesis is supported by a higher incidence of extra-
217 medullary relapse in the IR/noCR cohort: 44% in noCR/IR group vs. 4% in CR/IR group at 5 years
218 ($p<0.001$). A high incidence of EMD following allografting after reduced-intensity conditioning was
219 previously reported. In a series of 70 patients enrolled in a Spanish study, extra-medullary involvement
220 was documented in 10 out of the 27 patients at first relapse (37%) [27]. Interestingly, the incidence of

221 extra-medullary relapses was higher in patients who had developed chronic GVHD. Importantly, these
222 patients had no evidence of disease recurrence in the marrow at the time of relapse. The Authors
223 suggested that *graft-vs.-myeloma* effects may have been more efficient in the marrow or, alternatively,
224 that monoclonal plasma cells involved in extra-medullary relapse were more resistant to donor T-cells. In
225 another multi-center study, Minnema et al. reported an incidence of EMD of 20.4% in 54 relapsed MM
226 patients from a total group of 172 treated with sequential autologous-allogeneic non-myeloablative
227 transplantation [28]. Interestingly, no association with chronic GVHD and EMD at relapse was found. In
228 our experience, chronic GVHD did not impact on extra-medullary relapse. Overall, the association
229 between chronic GVHD and *graft-vs.-myeloma* effects, is still debated [29].

230 Finally, the recent observation of a possible increase of the occurrence of EMD, especially after
231 multiple relapses, may partly be explained by the current natural history of myeloma where patients
232 commonly live longer as compared to past decades [30]. A study on 1003 MM patients showed an
233 increase in EMD incidence in the period 2000-2007 as compared with previous years raising concerns,
234 despite a dramatic improvement in OS, about a correlation with the use of novel agents with potent anti-
235 myeloma activity and/or a greater use of high-dose therapy [30]. However, the observation that the
236 increase was evident both at diagnosis and at relapse suggests that other factors are contributory [31]. To
237 reduce the risk of bias when comparing the patient cohorts of our study, we particularly focused on the
238 presence of EMD at diagnosis and at first relapse post-transplant. In our series, the presence of EMD at
239 diagnosis did not correlate with a higher risk of EMD development post transplant and only the noCR/IR
240 status was significantly associated with extra-medullary relapse. Though EMD before allografting did not
241 impact on survival, post transplant extra-medullary relapse was associated with poorer outcome in
242 comparison with bone relapse (OS 39 vs. 57 months, $p=0.034$).

243 Although the potential role of positron emission tomography integrated with computed
244 tomography (PET/CT) in the assessment of MM continues to be a matter of debate [32], it may be
245 particularly informative to early diagnose EMD, together with other readily available laboratory assays

246 such as serum free light chains assay [33]. Patients in IR/noCR could be ideal candidates for a clinical
247 follow up that routinely includes PET/CT to possibly detect extra-medullary relapse before the occurrence
248 of symptoms.

249 In conclusion, evaluation of MRD by flow-cytometry is a sensitive prognostic tool after
250 allografting and should routinely be introduced in clinical practice. The achievement of IR is associated
251 with better clinical outcomes. Moreover, the combination of IR and CR is helpful to identify a subset of
252 patients (IR-noCR) at higher risk of developing extra-medullary relapse who may benefit from a more
253 stringent follow up and consolidation treatment with new agents [34,35].

254 255 256 **Acknowledgments:**

257 Our thanks to the patients who participated in this study, all physicians, nurses, and support
258 personnel for their care of patients, and to Jessica Mastrovito and Maria Josè Fornaro for manuscript
259 preparation,

260 **Conflict of interests:**

261 The Authors declare no potential conflicts of interest relevant to the article.

262 Supported in part by Progetti di Ricerca ex-60%, Ministero dell'Università e della Ricerca
263 Scientifica (MIUR); Regione Piemonte: Ricerca Finalizzata 2008, 2009; Fondazione Cassa di Risparmio
264 di Torino (CRT); Compagnia di San Paolo; Comitato Regionale Piemontese Gigi Ghirotti; and
265 Fondazione Neoplasie Sangue Onlus (FONESA).

266 **References:**

267

268 1.Durie BG, Harousseau JL. International uniform response criteria for multiple myeloma. *Leukemia*.
269 2006; **20**: 1467-1473.

270 2.San-Miguel JF, Mateos MV. Can multiple myeloma become a curable disease? *Haematologica*. 2011;
271 **96**: 1246–1248.

272 3.Stewart AK, Richardson PG, San-Miguel JF. How I treat multiple myeloma in younger patients. *Blood*.
273 2009; **114**: 5436–5443.

274 4.Corradini P, Voena C. Molecular and clinical remissions in multiple myeloma: role of autologous and
275 allogeneic transplantation of hematopoietic cells. *J Clin Oncol*. 1999; **17**: 208-215.

276 5.Martinelli G, Terragna C. Molecular remission after allogeneic or autologous transplantation of
277 hematopoietic stem cells for multiple myeloma. *J Clin Oncol*. 2000; **18**: 2273-2281.

278 6.Cavo M, Terragna C. Molecular monitoring of minimal residual disease in patients in long-term
279 complete remission after allogeneic stem cell transplantation for multiple myeloma. *Blood*. 2000; **96**: 355-
280 357.

281 7.Corradini P, Cavo M. Molecular remission after myeloablative allogeneic stem cell transplantation
282 predicts a better relapse-free survival in patients with multiple myeloma. *Blood*. 2003; **102**: 1927-1929.

283 8. Van Dongen JJ, Lhermitte L. EuroFlow antibody panels for standardized n-dimensional flow
284 cytometric immunophenotyping of normal, reactive and malignant leukocytes. *Leukemia*. 2012 Sep; **26**:
285 1908-75.

286 9.Rawstron AC, Davies FE. Flow cytometric disease monitoring in multiple myeloma: the relationship
287 between normal and neoplastic plasma cells predicts outcome after transplantation. *Blood*. 2002; **100**:
288 3095-3100.

289 10.Sarasquete ME, García-Sanz R. Minimal residual disease monitoring in multiple myeloma: a
290 comparison between allelic-specific oligonucleotide real-time quantitative polymerase chain reaction and
291 flow-cytometry . *Haematologica*. 2005; **90**: 1365-1372.

292 11.Paiva B, Vidriales MB. Multiparameter flow cytometric remission is the most relevant prognostic
293 factor for multiple myeloma patients who undergo autologous stem cell transplantation. *Blood*. 2008; **112**:
294 4017-4023.

295 12.Paiva B, Martinez-Lopez J. Comparison of immunofixation, serum free light chain, and
296 immunophenotyping for response evaluation and prognostication in multiple myeloma. *J Clin Oncol*.
297 2011; **29**: 1627-1633.

298 13.Rawstron AC, Child JA. Minimal residual disease assessed by multiparameter flow cytometry in
299 multiple myeloma: impact on outcome in the Medical Research Council Myeloma IX Study. *J Clin*
300 *Oncol*. 2013 Jul 10; **31**: 2540-7.

301 14.Bruno B, Rotta M. A comparison of allografting with autografting for newly diagnosed myeloma. *N*
302 *Engl J Med*. 2007; **356**: 1110-1120.

303 15.Giaccone L, Storer B. Long-term follow-up of a comparison of nonmyeloablative allografting with
304 autografting for newly diagnosed myeloma. *Blood*. 2011; **117**: 6721-6727.

305 16.Bruno B, Rotta M. Non-myeloablative allografting for newly diagnosed multiple myeloma: the
306 experience of the Gruppo Italiano Trapianti di Midollo. *Blood*. 2009; **113**: 3375-3382.

307 17.Rotta M, Storer BE. Long-term outcome of patients with multiple myeloma after autologous
308 hematopoietic cell transplantation and nonmyeloablative allografting. *Blood*. 2009; **113**: 3383-3391.

309 18.A.H. Filipovich, D. Weisdorf. National Institutes of Health consensus development project on criteria
310 for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging working group report. *Biol*
311 *Blood Marrow Transplant*. 2005; **11**: 945-956.

312 19.Sullivan KM, Agura E. Chronic graft-versus-host disease and other late complications of bone marrow
313 transplantation. *Semin Hematol*. 1991; **28**: 250-259.

314 20.Bladé J, Samson D. Criteria for evaluating disease response and progression in patients with multiple
315 myeloma treated by high-dose therapy and haemopoietic stem cell transplantation. Myeloma
316 Subcommittee of the EBMT. European Group for Blood and Marrow Transplant. *Br J Haematol*. 1998;
317 **102**: 1115-1123.

- 318 21. Bladé J, Fernández de Larrea C, Rosiñol L, Cibeira MT, Jimenez R, Powles R. Soft-Tissue
319 plasmacytomas in multiple myeloma: incidence, mechanisms of extramedullary spread, and treatment
320 approach. *J Clin Oncol*. 2011; **29**: 3805-3812.
- 321 22. Moreau P, San Miguel J. Multiple myeloma: clinical practice guidelines for diagnosis, treatment and
322 follow-up. *Annals of Oncology* 2013; **24**:vi133-vi137.
- 323 23. Paiva B, Almeida J. Utility of flow-cytometry immunophenotyping in multiple myeloma and other
324 clonal plasma cell-related disorders. *Cytometry B Clin Cytom*. 2010; **78**: 239-252.
- 325 24. Avet-Loiseau H, Attal M. Long-term analysis of the IFM 99 trials for myeloma: cytogenetic
326 abnormalities [t(4;14), del(17p), 1q gains] play a major role in defining long-term survival. *J Clin Oncol*.
327 2012; **30**: 1949-1952.
- 328 25. Gray RJ. A class of K-sample tests for comparing the cumulative incidence of a competing risk. *Ann*
329 *Stat*. 1988; **16**: 1141-1154.
- 330 26. Hauwel M, Matthes T. Minimal residual disease monitoring: the new standard for treatment evaluation
331 of haematological malignancies?. *Swiss Med Wkly*. 2014; **144**: w13907.
- 332 27. Pérez-Simón JA, Sureda A. Reduced-intensity conditioning allogeneic transplantation is associated
333 with a high incidence of extramedullary relapses in multiple myeloma patients. *Leukemia*. 2006; **20**: 542-
334 545.
- 335 28. Minnema MC, van de Donk NW. Extramedullary relapses after allogeneic non-myeloablative stem
336 cell transplantation in multiple myeloma patients do not negatively affect treatment outcome. *Bone*
337 *Marrow Transplant*. 2008; **41**: 779-784.
- 338 29. Passera R, Pollichieni S. Allogeneic Hematopoietic Cell Transplantation from Unrelated Donors in
339 Multiple Myeloma: Study from the Italian Bone Marrow Donor Registry. *Biol Blood Marrow Transplant*.
340 2013; **19**: 940-948.
- 341 30. Kumar SK, Rajkumar SV. Improved survival in multiple myeloma and the impact of novel therapies.
342 *Blood*. 2008; **111**: 2516-2520.
- 343 31. Varettoni M, Corso A. Incidence, presenting features and outcomes of extramedullary disease in

344 multiple myeloma: a longitudinal study on 1003 consecutive patients. *Ann Oncol.* 2010; **21**: 325-330.

345 32.Caers J, Withofs N. The role of positron emission tomography-computed tomography and magnetic
346 resonance imaging in diagnosis and follow up of multiple myeloma. *Haematologica.* 2014; **99**: 629-37.

347 33.Stawis AN, Maennle D. Recurrent plasmacytomas after allografting in a patient with multiple
348 myeloma. *Case Rep Med.* 2012; **2012**: 168785.

349 34.Bruno B, Giaccone L. Novel targeted drugs for the treatment of multiple myeloma: from bench to
350 bedside. *Leukemia.* 2005; **19**: 1729-1738.

351 35.Bruno B, Rotta M. New drugs for treatment of multiple myeloma. *Lancet Oncol.* 2004; **5**: 430-442.

352
353
354
355
356
357
358
359
360
361
362
363
364
365
366
367
368

FIGURE LEGEND

369 **Figure 1.** Clinical outcomes in three cohorts of patients defined by achievement of complete
370 clinical remission (CR) and immunophenotypic remission (IR): patients in CR and IR (CR/IR)
371 (green line); patients not in CR but in IR (blue line); patients not in CR and not in IR (pink line)