

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

**miR-214 as a Key Hub that Controls Cancer
Networks: Small Player, Multiple Functions**

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/152450> since 2016-06-20T17:06:12Z

Published version:

DOI:10.1038/jid.2014.479

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

miR-214 as a Key Hub that Controls Cancer Networks: Small Player, Multiple Functions

Elisa Penna^{1,2}, Francesca Orso^{1,2,3} and Daniela Taverna^{1,2,3}

MicroRNAs are short regulatory RNAs that are able to post-transcriptionally modulate gene expression and that have crucial roles in the control of physiological and pathological processes including cancer onset, growth, and progression. miR-214, located inside the sequence of the long noncoding *Dnm3os* transcript, contributes to the regulation of normal and cancer cell biology, even if it operates in a context-dependent and sometimes contradictory manner. miR-214 is deregulated in several human tumors including melanoma, breast, ovarian, gastric, and hepatocellular carcinomas. miR-214's pleiotropic and tumor-specific contribution to various cancer formation and progression hallmarks is achieved via its several target genes. In fact, miR-214 behaves as a key hub by coordinating fundamental signaling networks such as PTEN/AKT, β -catenin, and tyrosine kinase receptor pathways. Interestingly, miR-214 also regulates the levels of crucial gene expression modulators: the epigenetic repressor Ezh2, "genome guardian" p53, transcription factors TFAP2, and another microRNA, miR-148b. Thus, miR-214 seems to have essential roles in coordinating tumor proliferation, stemness, angiogenesis, invasiveness, extravasation, metastasis, resistance to chemotherapy, and microenvironment. The sum of current literature reports suggests that miR-214 is a molecular hub involved in the control of cancer networks and, as such, could be a potential diagnostic/prognostic biomarker and target for therapeutic intervention.

Journal of Investigative Dermatology (2015) **135**, 960–969;
doi:10.1038/jid.2014.479; published online 11 December 2014

INTRODUCTION

MicroRNAs (miRs) are a family of small, 20–25-nucleotide-long noncoding RNAs with the ability to post-transcriptionally downregulate the expression of specific target genes. At least a thousand miRs are predicted to operate in humans, regulating

multiple physiological and pathological cellular processes. Bioinformatic predictions indicate that miRs are expected to regulate the expression of around 50% of all protein-coding genes, thus controlling the activity of a large fraction of the entire transcriptome (Inui *et al.*, 2010). miR biogenesis and action mechanisms have been extensively reviewed in the study by Bartel, (2009). miR-mediated gene expression control is achieved by specific base pair binding to the 3'UTRs of their target mRNAs, causing either mRNA degradation or translation inhibition (Filipowicz *et al.*, 2008; Bartel, 2009; Inui *et al.*, 2010). A single 3'UTR can contain numerous different miR binding sites, and therefore the crucial impact miRs have on biological processes is mainly because of the fact that each miR can simultaneously regulate several targets with disparate functions, making them fundamental regulators of tumor onset, growth, and progression (Inui *et al.*, 2010).

Cancer cells are characterized by peculiar hallmarks acquired during the multisteps of tumor progression. These include sustained proliferation, resistance to cell death, metabolism reprogramming, host microenvironment interactions, angiogenesis, invasion, and seeding to distant organs (Hanahan and Weinberg, 2011). miR deregulation has a crucial role in cancer as it interferes with these specific hallmarks meaning miRs can either act as oncogenes or tumor suppressors in the early disease stages (Croce, 2009) or control the invasive and metastatic progression of cancer in the latter. Examples of miRs that control malignancy are miR-10b, miR-373/520c, miR-9 (pro-metastatic miRs), miR-335, the miR-200 family, and miR-31 (anti-metastatic miRs; Ma *et al.*, 2007; Huang *et al.*, 2008; Korpál *et al.*, 2008; Tavazoie *et al.*, 2008; Valastyan *et al.*, 2009; Ma *et al.*, 2010b). Relevantly, certain miRs are involved in specific tumor types, for instance the let-7 family, miR-34 family, miR-137, miR-182, miR-221/222, and miR-214 control melanoma progression (Mueller and Bosserhoff, 2009; Penna *et al.*, 2011).

This review focuses on the role of miR-214 in tumor progression. miR-214 is involved in numerous physiological and pathological processes including several cancers. The human miR-214 gene is located in the chromosomal region 1q24.3, in intron 14 of the Dynamin-3 gene (*DNM3*). As shown in Figure 1, an almost 8kb-long noncoding RNA, named *DNM3os*, originates from intron 14 of *DNM3* in humans and mice. This transcript contains the sequences for miR-214 and miR-199a-2, two clustered miRs that are approximately 6 kb apart. Another small RNA sequence, that of miR-3120, is also present in this genomic region; however, it is on the reverse strand (Loebel *et al.*, 2005; Watanabe *et al.*, 2008; Lee *et al.*, 2009; Yin *et al.*, 2010; Scott *et al.*, 2012).

¹Molecular Biotechnology Center (MBC), Torino, Italy; ²Department of Molecular Biotechnology and Health Sciences, Torino, Italy and ³Center for Molecular Systems Biology, University of Torino, Torino, Italy

Correspondence: Daniela Taverna, MBC and Department of Molecular Biotechnology and Health Sciences, University of Torino, Via Nizza 52, 10126 Torino, Italy.

E-mail: daniela.taverna@unito.it

Abbreviations: *DNM3*, Dynamin-3 gene; miR, microRNA

Received 15 May 2014; revised 29 October 2014; accepted 29 October 2014; published online 11 December 2014

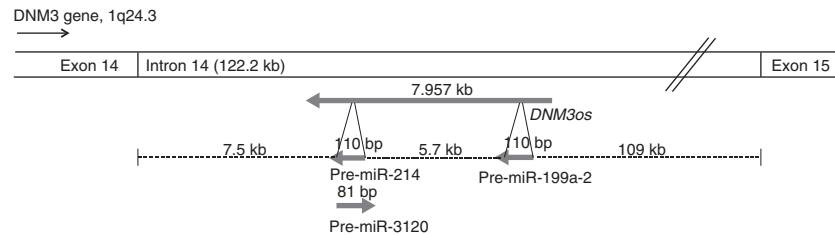


Figure 1. Genomic location of human miR-214. miR-214 and miR-199a-2 are 6 kb apart, located inside the 7.9 kb-long noncoding RNA precursor, *DNMT3os*, in intron 14 of the Dynamin-3 (*DNMT3*) gene (1q24.3). miR-3120 overlaps the miR-214 locus but in its complementary strand.

Twist-1, ZEB1, Ezh2, NFkB, and SMAD4 have been reported to control miR-214 and/or *DNMT3os* transcription (Kong *et al.*, 2008; Juan *et al.*, 2009; Lee *et al.*, 2009; Yin *et al.*, 2010; Duan *et al.*, 2012; Williams *et al.*, 2012). miR-214 is highly conserved among species, which indicates that it is involved in broad physiological functions. In fact, it coordinates the cell fate, differentiation and morphogenesis of muscles, the skeleton, nervous system, retina, and pancreas (Flynt *et al.*, 2007; Joglekar *et al.*, 2007; Decembrini *et al.*, 2009; Chen *et al.*, 2010a; Shi *et al.*, 2013). High miR-214 expression is visible early in mouse embryogenesis in the heart, cerebellum, midbrain, nasal process, and limb buds and reaches a peak at E12.5. It is downregulated after E15.5 and further decreases in postnatal tissues, although its expression is detected in some adult organs, including the lungs, heart, and kidneys (Watanabe *et al.*, 2008; Aurora *et al.*, 2012). Despite the fact that miR-214 and miR-199a-2 belong to the same cluster and are mostly co-expressed during development (Watanabe *et al.*, 2008), the large distance between them in the locus and their different seed sequences suggest distinct regulatory mechanisms and biological functions. Disruption of the entire mouse *DNMT3os* locus leads to skeletal abnormalities and death within 1 month after birth (Watanabe *et al.*, 2008). However, ubiquitous miR-214-specific knockout mice are viable and fertile, while displaying heart defects and cardiac contractility loss following ischemia-reperfusion injury (Aurora *et al.*, 2012), whereas bone-specific miR-214-overexpressing mice show defective osteogenic differentiation and bone formation (Wang *et al.*, 2013c). A specific miR-199a knockout mouse model is, however, necessary to clarify the contributions of miR-214 and miR-199a and to determine whether the differences between *DNMT3os* and miR-214-null mice are due to the combination of these two miRs or due to the specific functions of the long noncoding RNA.

If we consider the fact that embryonic stemness and differentiation programs are often reactivated during tumor progression, the hypothesis that miR-214 has a role in tumorigenesis is well-grounded.

MIR-214 DOUBLE-FACED DEREGULATION IN CANCER

miR-214 is deregulated in several human cancers and displays contrasting behavior, thus suggesting that cell context is crucial for miR-214 function (Table 1).

Cutaneous cancers

There are three main types of skin cancers; basal cell carcinoma, Squamous Cell Carcinoma (SCC), and malignant

melanoma. miR-214 is often altered in these, but particularly in melanoma, which is the least common but often most fatal of dermatologic tumors.

Although normal melanocytes display low miR-214 levels, the expression of this small RNA is high in nevi, which present abnormal, although benign, melanocyte proliferation and frequently contain genetic mutations (Grichnik, 2008; Glud *et al.*, 2009; Philippidou *et al.*, 2010; Chen *et al.*, 2010b; Penna *et al.*, 2011). Importantly, miR-214 is highly expressed in primary malignant cutaneous melanomas (Molnar *et al.*, 2008; Segura *et al.*, 2010; Penna *et al.*, 2011, 2013), although showing low expression levels in *in situ* noninvasive melanomas (Penna *et al.*, 2011). Moreover, miR-214 correlates significantly more with acral than with non-acral melanoma subtypes (Chan *et al.*, 2011) and is heavily expressed and associated with poor prognosis and elevated metastatic risk in ocular melanomas (Worley *et al.*, 2008). However, miR-214 levels may be slightly lower in metastases than in primary tumors, thus suggesting that this miR-214 increase has an important role at the time of radial to vertical/invasive melanoma growth (Penna *et al.*, 2011). Nevertheless, miR-214 upregulation in melanoma metastases is also associated with longer post-recurrence survival (Segura *et al.*, 2010). It is also important to note that miR-214 resides in a genomic region that is frequently amplified in melanoma, as demonstrated by Comparative Genomic Hybridization (CGH) in the study by Zhang *et al.*, (2006), whereas no reports indicate any correlation between miR-214 expression and typical melanoma genetic lesions, such as BRAF or KRAS mutations, to our knowledge. miR-199a-2, located in the same cluster as miR-214, is also amplified (Zhang *et al.*, 2006) and deregulated (Pencheva *et al.*, 2012) in melanoma. Patients with high miR-199a-3p and miR-199a-5p levels in primary melanomas show shorter metastasis-free survival compared with patients expressing low levels of these miRs (Pencheva *et al.*, 2012). Accordingly, high miR-199a levels in ocular melanomas correlate with metastatic risk (Worley *et al.*, 2008). As described in the following sections, miR-214 and miR-199a's involvement in driving melanoma metastasis has been clearly demonstrated (Penna *et al.*, 2011; Pencheva *et al.*, 2012; Penna *et al.*, 2013).

Although miR-214 is well expressed in epidermal keratinocytes and hair follicles and is relevant in skin morphogenesis (Ahmed *et al.*, 2012), contrasting observations have been reported regarding SCC and basal cell carcinoma. In fact, miR-214 is upregulated in oral and tongue SCC (Scapoli *et al.*, 2010; Yu *et al.*, 2010), whereas it was found to be significantly

Table 1. MicroRNA-214 alterations in cancer

Tumor	miR-214 expression in tumor vs. normal tissue	miR-214 correlation with metastasis or poor prognosis	Validated miR-214 targets	References
Melanoma	Up Down	Yes Yes	TFAP2, ITGA3	Penna <i>et al.</i> , 2011 Worley <i>et al.</i> , 2008 Segura <i>et al.</i> , 2010
Pancreas	Up	—	ING4	Zhang <i>et al.</i> , 2010 Volinia <i>et al.</i> , 2006
Stomach	Up	Yes	PTEN	Volinia <i>et al.</i> , 2006 Ueda <i>et al.</i> , 2010 Yang <i>et al.</i> , 2013
Prostate	Up	—	—	Volinia <i>et al.</i> , 2006
Nasopharyngeal	Up	—	LTF, BIM	Deng <i>et al.</i> , 2013 Zhang <i>et al.</i> , 2014
Oral/tongue squamous	Up	—	—	Yu <i>et al.</i> , 2010 Scapoli <i>et al.</i> , 2010
Osteosarcoma	Up	Yes	LZTS1	Wang <i>et al.</i> , 2014c Xu and Wang, 2014
T-cell lymphoma	Up	Yes	—	Narducci <i>et al.</i> , 2011
Lung	Up	—	—	Yanaihara <i>et al.</i> , 2006
Hepatocellular	Down	Yes	HDGF, XBP1, EZH2, FGFR1, β -catenin	Shih <i>et al.</i> , 2012 Xia <i>et al.</i> , 2012 Wang <i>et al.</i> , 2013b
Uterus cervix	Down	Yes	PLXNB1, GALNT7, MEK3, JNK1	Yang <i>et al.</i> , 2009 Qiang <i>et al.</i> , 2011
Adrenocortical	Down	—	—	Tombol <i>et al.</i> , 2009
Esophageal	Down	—	—	Huang <i>et al.</i> , 2012
Cutaneous squamous	Down	—	ERK1	Yamane <i>et al.</i> , 2013
Myeloma	Down	—	ASF1B, GANKYRIN	Gutierrez <i>et al.</i> , 2010
Bladder	Down	Yes	—	Ratert <i>et al.</i> , 2013
Glioma	Down	Yes	UBC9	Wang <i>et al.</i> , 2014b
Colorectal	Down	Yes	FGFR1	Chen <i>et al.</i> , 2014
Rhabdomyosarcoma	Down	—	NRAS	Huang <i>et al.</i> , 2014
Ovary	Up Down	Yes Yes	PTEN, p53	Yang <i>et al.</i> , 2008 Marchini <i>et al.</i> , 2011 Iorio <i>et al.</i> , 2007
Breast	Up Down	Yes Yes	EZH2	Derfoul <i>et al.</i> , 2011 Schwarzenbach <i>et al.</i> , 2012 Blenkiron <i>et al.</i> , 2007

MicroRNA-214 (miR-214) altered expression (up- or downregulation) in different types of human cancers. Where possible, the correlation with metastasis/poor prognosis and malignancy-involved validated targets is indicated.

downregulated in esophageal and cutaneous SCC, leading to abnormal keratinocyte proliferation (Huang *et al.*, 2012; Yamane *et al.*, 2013). miR-214 expression seems not to be involved in basal cell carcinoma, however (Sand *et al.*, 2012).

Various non-cutaneous tumors

miR-214 is upregulated in pancreatic (Volinia *et al.*, 2006; Zhang *et al.*, 2010), prostate (Volinia *et al.*, 2006), stomach (Volinia *et al.*, 2006; Ueda *et al.*, 2010; Li *et al.*, 2011; Yang *et al.*, 2013; Wang *et al.*, 2014a), nasopharyngeal (Deng *et al.*, 2013; Zhang *et al.*, 2014), and lung (Yanaihara *et al.*, 2006) tumors, as well as in osteosarcomas (Xu and Wang, 2014;

Wang *et al.*, 2014c) and in a leukemic variant of cutaneous T-cell lymphomas, the *Sezary syndrome* (Narducci *et al.*, 2011), as compared with healthy tissues. High miR-214 expression has been associated with unfavorable prognosis in overall survival and clinical staging, invasiveness, metastasis, and poor response to therapy (Ueda *et al.*, 2010; Narducci *et al.*, 2011; Yang *et al.*, 2013; Wang *et al.*, 2014c). Conversely, noteworthy miR-214 downregulation occurs in hepatocellular (Duan *et al.*, 2012; Shih *et al.*, 2012; Wang *et al.*, 2012a, 2013b), uterus cervix (Yang *et al.*, 2009), adrenocortical (Tombol *et al.*, 2009), bladder (Ratert *et al.*, 2013), colorectal (Chen *et al.*, 2014), and cholangio (Li *et al.*,

2012) carcinomas, as compared with healthy tissues, as well as in multiple myelomas (Gutierrez *et al.*, 2010), gliomas (Wang *et al.*, 2014b), and in lung adenocarcinoma-derived brain metastases (Zhao *et al.*, 2013). In these neoplasias, miR-214 reduction is often associated with malignancy, metastasis, or poor survival/recurrence (Qiang *et al.*, 2011; Li *et al.*, 2012; Xia *et al.*, 2012; Ratert *et al.*, 2013; Wang *et al.*, 2013b; Chen *et al.*, 2014).

miR-214 displays more complex behavior in breast and ovary cancers, possibly due to high tumor heterogeneity and hormone responsiveness. The miR-214 locus is frequently amplified in these tumors, as shown by CGH (Zhang *et al.*, 2006); however, it has been found to have been deleted in 24% of a small group of breast cancer cases (Derfoul *et al.*, 2011). miR-214 is less expressed in large cohorts (Dvinge *et al.*, 2013) of breast tumor samples than in normal tissues, whereas its upregulation has been found in Luminal A, Normal-like and ER-,PR-,HER2 +/– breast cancer subtypes (Blenkiron *et al.*, 2007; Sempere *et al.*, 2007). Similarly, miR-214 is overexpressed in ovary cancers and significantly associated with high grade and late/metastatic tumor stages, as well as with overall and progression-free survival and post-surgical/chemotherapy recurrence (Yang *et al.*, 2008; Marchini *et al.*, 2011). However, miR-214 downregulation has also been described in neoplastic, as compared with normal, ovaries, as well as in ovary cancer–derived effusions as compared with primary tumors (Iorio *et al.*, 2007; Vaksman *et al.*, 2011).

These data, taken as a whole, suggest that miR-214 deregulation could have a relevant impact on malignancy, even if it can operate in opposite manners in different tumors. Therefore, it should be evaluated for diagnostic/therapeutic purposes.

CIRCULATING MIR-214 AS A DIAGNOSTIC MARKER

Interestingly, miR-214 has been observed to be a circulating miR, and its serum levels have shown diagnostic potential. In fact, analyses of miR-214 levels in the serum of over 100 breast cancer patients were able to discriminate between malignant and benign tumors, as well as healthy controls, whereas levels were significantly decreased in post-operative sera but increased in association with metastatic spread to regional lymph nodes (Schwarzenbach *et al.*, 2012). miR-214 blood serum expression was also able to identify patients carrying malignant peripheral nerve sheath tumors (Weng *et al.*, 2013). Similarly, the analysis of circulating exosomes isolated from ovarian carcinoma patients revealed the presence of miR-214 (overexpressed in the primary tumor mass), together with 7 other miRs, which strongly supports the case for miR-214's diagnostic power (Taylor and Gercel-Taylor, 2008). The same situation was observed when exosomes from lung cancer patients were analyzed (Rabinowits *et al.*, 2009). Furthermore, urinary miR-214 has been proven to be a diagnostic biomarker for bladder and prostate cancers (Kim *et al.*, 2013; Srivastava *et al.*, 2013). These observations reinforce the potential of extracellular miR-214 use as a diagnostic or prognostic biomarker.

MIR-214 FUNCTIONS AND TARGETS IN TUMOR PROGRESSION

Altered miR-214 expression in cancer has led numerous researchers to investigate its tumor-specific functions and to identify its targets in the various tumor formation and progression steps. miR-214's pleiotropic roles and direct targets are summarized in Figure 2 and discussed below.

Proliferation, cell cycle, and survival

Altered levels of miR-214 in tumors seem to hint at a possible influence on proliferation and cell cycle control; however, existing data are controversial. No positive miR-214 cell viability or growth regulation was observed in melanomas (Penna *et al.*, 2011), lung (Fei *et al.*, 2008), or pancreatic (Zhang *et al.*, 2010) carcinomas. By contrast, miR-214 was found to promote AKT signaling, proliferation, and survival in gastric tumors via PTEN and in nasopharyngeal carcinomas via lactoferrin and Bim targeting (Deng *et al.*, 2013; Yang *et al.*, 2013; Zhang *et al.*, 2014), and to favor osteosarcoma growth via LZTS1 repression (Xu and Wang, 2014). On the other hand, miR-214 inhibits proliferation in breast or hepatic carcinomas via Ezh2, β -catenin, and XBP1 (an ER-stress-induced pro-survival factor) targeting; miR-214 expression also reduces hepatoma xenograft growth (Derfoul *et al.*, 2011; Duan *et al.*, 2012; Shih *et al.*, 2012; Xia *et al.*, 2012; Wang *et al.*, 2012a). Furthermore, proliferation is decreased by miR-214 in rhabdomyosarcomas via N-Ras oncogene suppression (Huang *et al.*, 2014), in cervical cancers via MEK3 and JNK1, GALNT7 and Plexin-B1 downregulation (Cheng *et al.*, 2005; Yang *et al.*, 2009; Qiang *et al.*, 2011; Peng *et al.*, 2012), and in squamous cell carcinoma cells via ERK1 inhibition (Yamane *et al.*, 2013). In myelomas, miR-214 targets ASF1B and gankirin, thus inducing p53, p21, and Bax upregulation and resulting in the inhibition of DNA replication and apoptosis (Misiewicz-Krzeminska *et al.*, 2013). A link between miR-214 and Receptor Tyrosine Kinases (RTKs) was also observed. In fact, miR-214 targets FGFR1 in hepatocellular and colorectal carcinomas, thus influencing cell growth (Wang *et al.*, 2013b; Chen *et al.*, 2014), and the ErbB3 cis-interactor Necl-2, leading to enhanced ErbB2/3 signaling in colon carcinomas (Momose *et al.*, 2013).

Angiogenesis

Tumor vascularisation has a central role to play in cancer progression. Indeed, neo-angiogenesis within the tumor mass provides cancer cells with the blood vessels necessary for growth and, in the meanwhile, facilitates the intravasation and dissemination of metastatic cells. On the other hand, hypoxic conditions, caused by insufficient or aberrant vascularization, often contribute to the selection of more malignant tumor cell subpopulations (Carmeliet and Jain, 2011). Interestingly, miR-214 has a key role in angiogenesis. It is expressed in endothelial cells (Wurdinger *et al.*, 2008; Rippe *et al.*, 2012) and inhibits HUVEC angiogenesis and vascular network formation by targeting Quaking and, consequently, reduces the secretion of pro-angiogenic growth factors and cytokines, including VEGF, CCL5, and CXCL5 (van Mil *et al.*, 2012). In addition, miR-214 downregulates the eNOS (nitric oxide

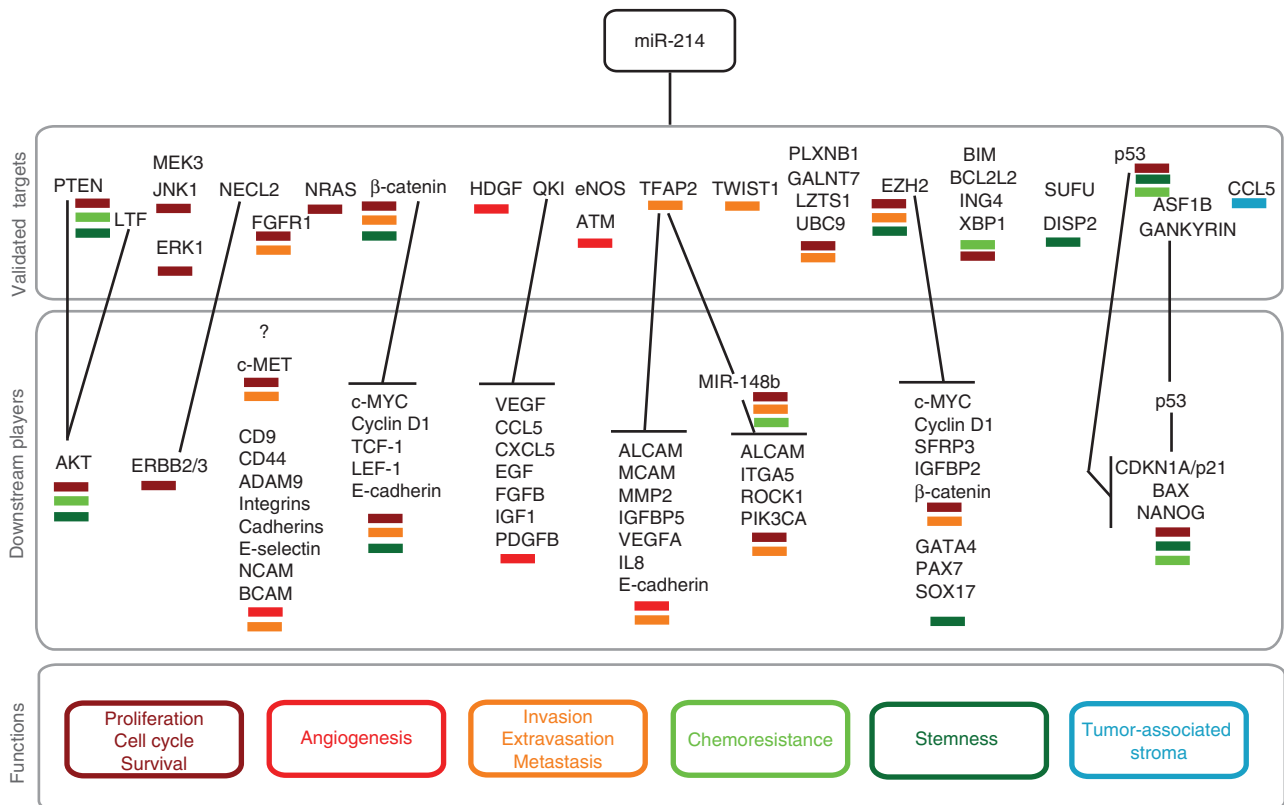


Figure 2. miR-214 controls cancer networks. miR-214 direct target genes and indirect downstream players are shown, together with their multiple functions in tumor cells (different colors).

synthase) enzyme, which is essential for endothelial cell migration (Chan *et al.*, 2009). By contrast, adult endothelial cells (HMEC-1) secrete miR-214-containing exosomes which, in turn, stimulate a pro-angiogenic program in neighboring cells via ATM downregulation (van Balkom *et al.*, 2013). Considering the cross talk between tumor and endothelial cells within a tumor mass, the existence of a pivotal role for miR-214 can be envisaged in tumor vascularization. Indeed, miR-214 downregulation in hepatocellular carcinoma contributes to tumor angiogenesis, whereas its ectopic overexpression reduces tumor growth and vascularity. This occurs via its direct target, hepatoma-derived growth factor (HDGF; Shih *et al.*, 2012), thus demonstrating that miR-214 has an anti-angiogenic role in hepatocarcinomas, in which miR-214 downregulation is associated with worse clinical outcome.

Migration, invasion, extravasation, and metastasis

miR-214 upregulation in melanoma cell lines significantly promotes tumor cell migration, invasion, adhesion to extracellular matrices, transendothelial migration, and survival to anoikis *in vitro*, as well as extravasation from blood vessels and metastasis formation in mice. This occurs without affecting cell proliferation and tumor growth via the simultaneous coordination of a network including over 70 genes (Penna *et al.*, 2011). In particular, miR-214 acts by directly targeting transcription factors TFAP2, well-established melanoma tumor suppressors, and upregulating ALCAM, an adhesion receptor

known as a melanoma progression marker. ALCAM upregulation occurs via TFAP2 and miR-148b, a small RNA that is downregulated by miR-214. Rescuing experiments have demonstrated that TFAP2 or miR-148b downmodulation and ALCAM upregulation control miR-214-dependent melanoma metastatic traits and metastasis formation (Penna *et al.*, 2011, 2013), thus highlighting the importance of a critical pathway that involves these players. Importantly, several miR-148b target genes, including ALCAM, ROCK1, PIK3CA, and ITGA5, are upregulated by miR-214, because of miR-148b downmodulation, which could account for additional miR-214 pro-malignant functions (Penna *et al.*, 2013; Orso *et al.*, unpublished data). Moreover, the miR-214-dependent modulation of other genes in this network may influence melanoma progression, as well. These include genes that regulate adhesion, movement, invasion, and matrix degradation (integrins, cadherins, MCAM, NCAM, BCAM, PAK2, MMP2, ADAMs, TIMPs and the MET oncogene), others that are implicated in interactions with endothelial cells and extravasation (E-Selectin, CD9, IL-8, SEMA3A, VEGFR2, VEGFA and ALCAM itself), and still others that are members of the TGFβ, Wnt, and Notch pathways and that are potentially involved in the epithelial-to-mesenchymal transition. These findings appear to indicate a clear pro-invasive and pro-metastatic role played by miR-214 in melanoma. Nevertheless, other authors have observed, in a similar context, the existence of a significant role for the small RNA that is clustered

with miR-214, miR-199a-2. Indeed, both miR-199a-5p and miR-199a-3p have been demonstrated to strongly drive melanoma invasion and metastatic colonization and to enhance angiogenesis by synergistically targeting ApoE and DNAJA4 and thus controlling the LPR1 and LPR8 pathways (Pencheva *et al.*, 2012). It is possible that miR-214 and miR-199a operate in an alternative or synergistic manner in melanoma progression, and detailed experiments need to be performed to unravel the precise functions of these two clustered miRs.

miR-214 significantly encourages migration and invasion in gastric cancer by targeting PTEN (Yang *et al.*, 2013), whereas it favors osteosarcoma invasion by silencing LZTS1 (Xu and Wang, 2014), and, furthermore, nasopharyngeal carcinoma metastasis formation has been found to be promoted via lactoferrin repression in a mouse xenograft model (Deng *et al.*, 2013). miR-214 has been shown to display pro-invasive or pro-metastatic properties in mammary tumor cells (Penna *et al.*, 2011), although a deeper analysis of miR-214 functions in breast cancer is still pending. Indeed, Derfoul and colleagues have reported opposite data: decreased miR-214 levels in mammary carcinoma cells correlate with increased cell invasion via the accumulation of miR-214 target, Ezh2, a known marker of malignancy that coincides with the histone H3K27 trimethylation catalytic subunit of the Polycomb repressive complex 2, which epigenetically regulates chromatin and thereby influences gene expression (Derfoul *et al.*, 2011; Crea *et al.*, 2012). Similarly, miR-214 suppresses cell invasion by downregulating Ezh2, β -catenin, and FGFR1 in hepatocellular carcinoma cells; the high expression of these genes correlates with low miR-214 levels and increased malignancy (Xia *et al.*, 2012; Wang *et al.*, 2013b). Importantly, sustained β -catenin and/or Ezh2 expression in breast or hepatic tumors leads to malignancy as the levels of their downstream genes, such as c-Myc, cyclin D1, TCF-1, LEF-1, E-cadherin, SFRP3, IGFBP2, and even β -catenin itself, are affected (Derfoul *et al.*, 2011; Xia *et al.*, 2012; Wang *et al.*, 2012a). Interestingly, Ezh2 repression by miR-214 in esophageal squamous carcinoma cells also results in migration and invasion inhibition (Huang *et al.*, 2012). Furthermore, miR-214 has been shown to inhibit invasion via Plexin-B1 and GALNT7 targeting in cervical cancers (Qiang *et al.*, 2011; Peng *et al.*, 2012), via FGFR1 repression in colorectal tumors (Chen *et al.*, 2014) and via Twist-1 (a master regulator of metastasis) downregulation in intrahepatic cholangiocarcinomas (Li *et al.*, 2012).

Chemoresistance

Despite the controversial findings concerning the role of miR-214 in proliferation, one of the first miR-214 functions described was its ability to reduce apoptotic cell death, contributing to decreased chemotherapy sensitivity, the main cause of treatment failure and disease progression. In particular, miR-214 has been shown to inhibit cisplatin-induced apoptosis in human ovarian cancers primarily by targeting PTEN and enhancing the AKT pathway and survival (Yang *et al.*, 2008). Moreover, miR-214 has been proven to contribute to cisplatin chemoresistance in tongue squamous cell carcinomas (Yu *et al.*, 2010). Accordingly, miR-214 is

significantly upregulated in gefitinib-resistant EGFR-mutant non-small cell-lung cancer cells. PTEN targeting also has a major role in this case. In fact, miR-214 knockdown is sufficient to normalize AKT pathway activation and to resensitize cells to the therapy (Wang *et al.*, 2012b). Furthermore, miR-214 overexpression in pancreatic cancer cells decreases sensitivity to gemcitabine, the first line treatment for advanced pancreatic cancers, probably via its direct target ING4, a gene involved in cell cycle arrest, DNA repair, and apoptosis (Zhang *et al.*, 2010). Moreover, high miR-214 levels have been associated with poor chemotherapy response in pediatric osteosarcomas (Wang *et al.*, 2014c) and with increased T-cell lymphoma cell survival and apoptotic resistance (Narducci *et al.*, 2011). Finally, miR-214 has been predicted to control paclitaxel sensitivity in laryngeal cancer cells (Xu *et al.*, 2013). The only study that has published contrary results was performed in cervical cancer cells where miR-214 enhances cisplatin-induced cytotoxicity, via the downregulation of Bcl2l2 (Wang *et al.*, 2013a). In melanomas, miR-214 reduces apoptosis and facilitates survival in adhesion-lacking (anoikis) conditions (Penna *et al.*, 2011), which would seem to suggest a potential role in chemoresistance; however, the details of its actual involvement are still to be explored.

Cancer stemness

miR-214 has a key role in normal cell differentiation by targeting Ezh2 (see above), which leads to the consequent derepression of the master developmental regulators Gata-4, Pax7, and Sox17 (Juan *et al.*, 2009). As developmental pathways are often reactivated during tumorigenesis and stemness markers are observed in tumor stem cells (Nguyen *et al.*, 2012), the involvement of miR-214 in cancer stem cells has been investigated and confirmed. In fact, miR-214 has been shown to regulate ovarian cancer stem cell properties by directly repressing p53 and indirectly upregulating Nanog, leading to increased stem cell population and self-renewal (Xu *et al.*, 2012). However, lower miR-214 expression has been reported in ovarian cancer stem cells than in mature cells, and here the miR-214-controlled PTEN-AKT pathway is seen to be a major regulator of stemness (Yin *et al.*, 2010). Interestingly, miR-214 was found to be the most highly expressed of a list of 37 differentially expressed small noncoding RNAs in an analysis of miR expression in purified CD44⁺CD24[−]lin[−] breast cancer stem cells (Shimono *et al.*, 2009). miR-214 expression has also been found to be significantly increased in pancreatic cancer cell-derived holoclones, which are capable of long-term survival, tumor initiation, and chemoresistance (Tan *et al.*, 2011). In contrast, miR-214 has been demonstrated to suppress stem-like traits in human hepatocellular carcinoma cells by directly targeting Ezh2 and β -catenin signaling pathways (Xia *et al.*, 2012). These findings indicate that miR-214 has contrasting functions in controlling cancer stemness, depending on its role in specific malignancies.

Tumor-associated stroma

Cancer-associated fibroblasts (CAFs) have a major role in the tumor microenvironment; however, the conversion of quiescent fibroblasts into CAFs is still quite a mystery. miR-214 is

relevant in this process as ovarian cancer cells, in fact, induce miR-214 downregulation in normal tumor-adjacent fibroblasts, thus reprogramming them into CAFs with increased chemokine secretion. A particular example is CCL5, which is a direct miR-214 target (Mitra *et al.*, 2012). On the other hand, miR-214 is upregulated in gastric cancer-derived mesenchymal stem cells, which are involved in invasion (Wang *et al.*, 2014a).

CONCLUDING REMARKS

In conclusion, miR-214 appears to be a pleiotropic hub that participates in the control of cancer cell signaling networks. In fact, it contributes to the coordination of essential signaling pathways, such as the PTEN/AKT, β -catenin, or receptor tyrosine kinase pathways, as well as crucial gene expression modulators, including Ezh2, p53, TFAP2, and miR-148b. The fact that miR-214 displays specific and even contrasting functions in different tumor types is most likely due to differential mRNA targeting and/or target gene expression in different cancer cell contexts. In particular, as miR-214 broadly controls gene expression by regulating transcription factors and other miRs, disparities in the expression of subsets of these downstream players in different tumors, or tumor cell populations, may account for the versatility of miR-214 actions. Thus, miR-214 may have contrasting roles according to co-expression with its own direct or indirect target genes or with other miRs (for instance miR-148b or the clustered miR-199a-2) and their targets. This may also explain the switch between proliferation/survival phenotypes in certain cancers to movement/invasion traits in others. Importantly, some known miR-214-regulated genes (e.g., TFAP2 and ALCAM) display contrasting behavior in different tumor types in which miR-214 also has contrasting functions. For instance, although TFAP2 factors are well-defined tumor suppressors in melanoma and are often lost during disease progression (Bar-Eli, 2001), their role in breast tumors is more complex. In fact, TFAP2A is frequently upregulated in ErbB2/HER2-positive breast carcinomas and induces ErbB2/HER2 transcription (Bosher *et al.*, 1996; Turner *et al.*, 1998; Shiu *et al.*, 2014). However, other studies have observed reduced TFAP2 levels in high grade and metastatic breast cancers (Pellikainen *et al.*, 2002; Sotiriou *et al.*, 2006). Moreover TFAP2C overexpression in a MMTV/Neu transgenic mouse model resulted in reduced tumorigenesis (Jager *et al.*, 2005). Similarly, high ALCAM positivity is a clinical marker for melanoma thickness and invasiveness (van Kempen *et al.*, 2000). ALCAM upregulation, as opposed to normal mammary tissue, has furthermore been observed in intraductal/invasive breast carcinomas (Burkhardt *et al.*, 2006) and correlates with metastasis, as well (Piao *et al.*, 2012). However, low ALCAM levels have also been associated with mammary nodal infiltration, metastatic dissemination, and worse clinical outcome (King *et al.*, 2004; Jezierska *et al.*, 2006).

Tumor mouse models are essential to an improved definition of miR-214 involvement in cancer, and as such other tissue-specific miR-214-deleted or miR-214-overexpressing mice need to be established and analyzed for this purpose, in addition to existing animal models (Aurora *et al.*, 2012; Wang *et al.*, 2013c). Additional investigations must carefully

evaluate miR-214 genomic locus and transcriptional/post-transcriptional events if we want to understand how miR-214 deregulation occurs in cancer. Furthermore, circulating miR-214 has been proven to be a potential biomarker (Schwarzenbach *et al.*, 2012), and therefore its diagnostic/prognostic value should be explored in depth and in various tumors. Finally and more importantly, as miR-based *in vivo* therapy attempts have already been successful (Care *et al.*, 2007; Elyakim *et al.*, 2010; Wiggins *et al.*, 2010; Ma *et al.*, 2010a), the potential use of miR-214 as a therapy target merits careful investigation.

CONFLICT OF INTEREST

The authors state no conflict of interest.

REFERENCES

- Ahmed M, Emelianov V, Sharov A *et al.* (2012) MicroRNA-214 controls skin and hair follicle development via modulating the activity of Wnt, Edar and Bmp signalling pathways. *J Invest Dermatol* 132:S104–10
- Aurora AB, Mahmoud AI, Luo X *et al.* (2012) MicroRNA-214 protects the mouse heart from ischemic injury by controlling Ca(2)(+) overload and cell death. *J Clin Invest* 122:1222–32
- Bar-Eli M (2001) Gene regulation in melanoma progression by the AP-2 transcription factor. *Pigment Cell Res* 14:78–85
- Bartel DP (2009) MicroRNAs: target recognition and regulatory functions. *Cell* 136:215–33
- Blenkiron C, Goldstein LD, Thorne NP *et al.* (2007) MicroRNA expression profiling of human breast cancer identifies new markers of tumor subtype. *Genome Biol* 8:R214
- Bosher JM, Totty NF, Hsuan JJ *et al.* (1996) A family of AP-2 proteins regulates c-erbB-2 expression in mammary carcinoma. *Oncogene* 13:1701–7
- Burkhardt M, Mayordomo E, Winzer KJ *et al.* (2006) Cytoplasmic overexpression of ALCAM is prognostic of disease progression in breast cancer. *J Clin Pathol* 59:403–9
- Care A, Catalucci D, Felicetti F *et al.* (2007) MicroRNA-133 controls cardiac hypertrophy. *Nat Med* 13:613–8
- Carmeliet P, Jain RK (2011) Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat Rev Drug Discov* 10:417–27
- Chan E, Patel R, Nallur S *et al.* (2011) MicroRNA signatures differentiate melanoma subtypes. *Cell Cycle* 10:1845–52
- Chan LS, Yue PY, Mak NK *et al.* (2009) Role of microRNA-214 in ginsenoside-Rg1-induced angiogenesis. *Eur J Pharm Sci* 38:370–7
- Chen DL, Wang ZQ, Zeng ZL *et al.* (2014) Identification of miR-214 as a negative regulator of colorectal cancer liver metastasis via regulation of FGFR1 expression. *Hepatology* 60:598–609
- Chen H, Shalom-Feuerstein R, Riley J *et al.* (2010a) miR-7 and miR-214 are specifically expressed during neuroblastoma differentiation, cortical development and embryonic stem cells differentiation, and control neurite outgrowth in vitro. *Biochem Biophys Res Commun* 394:921–7
- Chen J, Feilott HE, Pare GC *et al.* (2010b) MicroRNA-193b represses cell proliferation and regulates cyclin D1 in melanoma. *Am J Pathol* 176:2520–9
- Cheng AM, Byrom MW, Shelton J *et al.* (2005) Antisense inhibition of human miRNAs and indications for an involvement of miRNA in cell growth and apoptosis. *Nucleic Acids Res* 33:1290–7
- Crea F, Fornaro L, Bocci G *et al.* (2012) EZH2 inhibition: targeting the crossroad of tumor invasion and angiogenesis. *Cancer Metastasis Rev* 31:753–61
- Croce CM (2009) Causes and consequences of microRNA dysregulation in cancer. *Nat Rev Genet* 10:704–14
- Decembrini S, Bressan D, Vignali R *et al.* (2009) MicroRNAs couple cell fate and developmental timing in retina. *Proc Natl Acad Sci USA* 106:21179–84

- Deng M, Ye Q, Qin Z *et al.* (2013) miR-214 promotes tumorigenesis by targeting lactotransferrin in nasopharyngeal carcinoma. *Tumour Biol* 34: 1793–800
- Derfoul A, Juan AH, Difilippantonio MJ *et al.* (2011) Decreased microRNA-214 levels in breast cancer cells coincides with increased cell proliferation, invasion and accumulation of the Polycomb Ezh2 methyltransferase. *Carcinogenesis* 32:1607–14
- Duan Q, Wang X, Gong W *et al.* (2012) ER stress negatively modulates the expression of the miR-199a/214 cluster to regulates tumor survival and progression in human hepatocellular cancer. *PLoS One* 7:e31518
- Dvinge H, Git A, Graf S *et al.* (2013) The shaping and functional consequences of the microRNA landscape in breast cancer. *Nature* 497:378–82
- Elyakim E, Sitbon E, Faerman A *et al.* (2010) hsa-miR-191 is a candidate oncogene target for hepatocellular carcinoma therapy. *Cancer Res* 70:8077–87
- Fei J, Lan F, Guo M *et al.* (2008) Inhibitory effects of anti-miRNA oligonucleotides (AMOs) on A549 cell growth. *J Drug Target* 16:688–93
- Filipowicz W, Bhattacharyya SN, Sonenberg N (2008) Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? *Nat Rev Genet* 9:102–14
- Flynt AS, Li N, Thatcher EJ *et al.* (2007) Zebrafish miR-214 modulates Hedgehog signaling to specify muscle cell fate. *Nat Genet* 39:259–63
- Glud M, Klausen M, Gniadecki R *et al.* (2009) MicroRNA expression in melanocytic nevi: the usefulness of formalin-fixed, paraffin-embedded material for miRNA microarray profiling. *J Invest Dermatol* 129:1219–24
- Grichnik JM (2008) Melanoma, neovogenesis, and stem cell biology. *J Invest Dermatol* 128:2365–80
- Gutierrez NC, Sarasquete ME, Misiewicz-Krzeminska I *et al.* (2010) Deregulation of microRNA expression in the different genetic subtypes of multiple myeloma and correlation with gene expression profiling. *Leukemia* 24:629–37
- Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. *Cell* 144:646–74
- Huang HJ, Liu J, Hua H *et al.* (2014) MiR-214 and N-ras regulatory loop suppresses rhabdomyosarcoma cell growth and xenograft tumorigenesis. *Oncotarget* 5:2161–75
- Huang Q, Gumireddy K, Schrier M *et al.* (2008) The microRNAs miR-373 and miR-520c promote tumour invasion and metastasis. *Nat Cell Biol* 10: 202–10
- Huang SD, Yuan Y, Zhuang CW *et al.* (2012) MicroRNA-98 and microRNA-214 post-transcriptionally regulate enhancer of zeste homolog 2 and inhibit migration and invasion in human esophageal squamous cell carcinoma. *Mol Cancer* 11:51
- Inui M, Martello G, Piccolo S (2010) MicroRNA control of signal transduction. *Nat Rev Mol Cell Biol* 11:252–63
- Iorio MV, Visone R, Di Leva G *et al.* (2007) MicroRNA signatures in human ovarian cancer. *Cancer Res* 67:8699–707
- Jager R, Friedrichs N, Heim I *et al.* (2005) Dual role of AP-2gamma in ErbB-2-induced mammary tumorigenesis. *Breast Cancer Res Treat* 90:273–80
- Jezierska A, Olszewski WP, Pietruszkiewicz J *et al.* (2006) Activated leukocyte cell adhesion molecule (ALCAM) is associated with suppression of breast cancer cells invasion. *Med Sci Monit* 12:BR245–56
- Joglekar MV, Parekh VS, Hardikar AA (2007) New pancreas from old: micro-regulators of pancreas regeneration. *Trends Endocrinol Metab* 18:393–400
- Juan AH, Kumar RM, Marx JG *et al.* (2009) Mir-214-dependent regulation of the polycomb protein Ezh2 in skeletal muscle and embryonic stem cells. *Mol Cell* 36:61–74
- Kim SM, Kang HW, Kim WT *et al.* (2013) Cell-free microRNA-214 from urine as a biomarker for non-muscle-invasive bladder cancer. *Korean J Urol* 54:791–6
- King JA, Ofori-Acquah SF, Stevens T *et al.* (2004) Activated leukocyte cell adhesion molecule in breast cancer: prognostic indicator. *Breast Cancer Res* 6:R478–87
- Kong W, Yang H, He L *et al.* (2008) MicroRNA-155 is regulated by the transforming growth factor beta/Smad pathway and contributes to epithelial cell plasticity by targeting RhoA. *Mol Cell Biol* 28:6773–84
- Korpai M, Lee ES, Hu G *et al.* (2008) The miR-200 family inhibits epithelial-mesenchymal transition and cancer cell migration by direct targeting of E-cadherin transcriptional repressors ZEB1 and ZEB2. *J Biol Chem* 283:14910–4
- Lee YB, Bantounas I, Lee DY *et al.* (2009) Twist-1 regulates the miR-199a/214 cluster during development. *Nucleic Acids Res* 37:123–8
- Li B, Han Q, Zhu Y *et al.* (2012) Down-regulation of miR-214 contributes to intrahepatic cholangiocarcinoma metastasis by targeting Twist. *FEBS J* 279:2393–8
- Li X, Zhang Y, Zhang H *et al.* (2011) miRNA-223 promotes gastric cancer invasion and metastasis by targeting tumor suppressor EPB41L3. *Mol Cancer Res* 9:824–33
- Loebel DA, Tsoi B, Wong N *et al.* (2005) A conserved noncoding intronic transcript at the mouse Dnm3 locus. *Genomics* 85:782–9
- Ma L, Reinhardt F, Pan E *et al.* (2010a) Therapeutic silencing of miR-10b inhibits metastasis in a mouse mammary tumor model. *Nat Biotechnol* 28:341–7
- Ma L, Teruya-Feldstein J, Weinberg RA (2007) Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. *Nature* 449:682–8
- Ma L, Young J, Prabhala H *et al.* (2010b) miR-9, a MYC/MYCN-activated microRNA, regulates E-cadherin and cancer metastasis. *Nat Cell Biol* 12:247–56
- Marchini S, Cavalieri D, Fruscio R *et al.* (2011) Association between miR-200c and the survival of patients with stage I epithelial ovarian cancer: a retrospective study of two independent tumour tissue collections. *Lancet Oncol* 12:273–85
- Misiewicz-Krzeminska I, Sarasquete ME, Quwaider D *et al.* (2013) Restoration of microRNA-214 expression reduces growth of myeloma cells through positive regulation of P53 and inhibition of DNA replication. *Haematologica* 98:640–8
- Mitra AK, Zillhardt M, Hua Y *et al.* (2012) MicroRNAs reprogram normal fibroblasts into cancer-associated fibroblasts in ovarian cancer. *Cancer Discov* 2:1100–8
- Molnar V, Tamasi V, Bakos B *et al.* (2008) Changes in miRNA expression in solid tumors: an miRNA profiling in melanomas. *Semin Cancer Biol* 18:111–22
- Momose K, Minami A, Shimono Y *et al.* (2013) miR-214 and hypoxia down-regulate Necl-2/CADM1 and enhance ErbB2/ErbB3 signaling. *Genes Cells* 18:195–202
- Mueller DW, Bosserhoff AK (2009) Role of miRNAs in the progression of malignant melanoma. *Br J Cancer* 101:551–6
- Narducci MG, Arcelli D, Picchio MC *et al.* (2011) MicroRNA profiling reveals that miR-21, miR486 and miR-214 are upregulated and involved in cell survival in Sezary syndrome. *Cell Death Dis* 2:e151
- Nguyen LV, Vanner R, Dirks P *et al.* (2012) Cancer stem cells: an evolving concept. *Nat Rev Cancer* 12:133–43
- Pellikainen J, Kataja V, Ropponen K *et al.* (2002) Reduced nuclear expression of transcription factor AP-2 associates with aggressive breast cancer. *Clin Cancer Res* 8:3487–95
- Pencheva N, Tran H, Buss C *et al.* (2012) Convergent multi-miRNA targeting of ApoE drives LRP1/LRP8-dependent melanoma metastasis and angiogenesis. *Cell* 151:1068–82
- Peng RQ, Wan HY, Li HF *et al.* (2012) MicroRNA-214 suppresses growth and invasiveness of cervical cancer cells by targeting UDP-N-acetyl-alpha-D-galactosamine:polypeptide N-acetyltransferase 7. *J Biol Chem* 287:14301–9
- Penna E, Orso F, Cimino D *et al.* (2011) microRNA-214 contributes to melanoma tumour progression through suppression of TFAP2C. *EMBO J* 30:1990–2007
- Penna E, Orso F, Cimino D *et al.* (2013) miR-214 coordinates melanoma progression by upregulating ALCAM through TFAP2 and miR-148b downmodulation. *Cancer Res* 73:4098–111
- Philippidou D, Schmitt M, Moser D *et al.* (2010) Signatures of microRNAs and selected microRNA target genes in human melanoma. *Cancer Res* 70:4163–73
- Piao D, Jiang T, Liu G *et al.* (2012) Clinical implications of activated leukocyte cell adhesion molecule expression in breast cancer. *Mol Biol Rep* 39:661–8

- Qiang R, Wang F, Shi LY *et al.* (2011) Plexin-B1 is a target of miR-214 in cervical cancer and promotes the growth and invasion of HeLa cells. *Int J Biochem Cell Biol* 43:632–41
- Rabinowitz G, Gercel-Taylor C, Day JM *et al.* (2009) Exosomal microRNA: a diagnostic marker for lung cancer. *Clin Lung Cancer* 10:42–6
- Ratert N, Meyer HA, Jung M *et al.* (2013) miRNA profiling identifies candidate miRNAs for bladder cancer diagnosis and clinical outcome. *J Mol Diagn* 15:695–705
- Rippe C, Blimline M, Magerko KA *et al.* (2012) MicroRNA changes in human arterial endothelial cells with senescence: relation to apoptosis, eNOS and inflammation. *Exp Gerontol* 47:45–51
- Sand M, Skrygan M, Sand D *et al.* (2012) Expression of microRNAs in basal cell carcinoma. *Br J Dermatol* 167:847–55
- Scapoli L, Palmieri A, Lo Muzio L *et al.* (2010) MicroRNA expression profiling of oral carcinoma identifies new markers of tumor progression. *Int J Immunopathol Pharmacol* 23:1229–34
- Schwarzenbach H, Milde-Langosch K, Steinbach B *et al.* (2012) Diagnostic potential of PTEN-targeting miR-214 in the blood of breast cancer patients. *Breast Cancer Res Treat* 134:933–41
- Scott H, Howarth J, Lee YB *et al.* (2012) MiR-3120 is a mirror microRNA that targets heat shock cognate protein 70 and auxilin messenger RNAs and regulates clathrin vesicle uncoating. *J Biol Chem* 287:14726–33
- Segura MF, Belitskaya-Levy I, Rose AE *et al.* (2010) Melanoma MicroRNA signature predicts post-recurrence survival. *Clin Cancer Res* 16:1577–86
- Sempere LF, Christensen M, Silaharoglu A *et al.* (2007) Altered MicroRNA expression confined to specific epithelial cell subpopulations in breast cancer. *Cancer Res* 67:11612–20
- Shi K, Lu J, Zhao Y *et al.* (2013) MicroRNA-214 suppresses osteogenic differentiation of C2C12 myoblast cells by targeting Osterix. *Bone* 55:487–94
- Shih TC, Tien YJ, Wen CJ *et al.* (2012) MicroRNA-214 downregulation contributes to tumor angiogenesis by inducing secretion of the hepatoma-derived growth factor in human hepatoma. *J Hepatol* 57:584–91
- Shimono Y, Zabala M, Cho RW *et al.* (2009) Downregulation of miRNA-200c links breast cancer stem cells with normal stem cells. *Cell* 138:592–603
- Shiu KK, Wetterskog D, Mackay A *et al.* (2014) Integrative molecular and functional profiling of ERBB2-amplified breast cancers identifies new genetic dependencies. *Oncogene* 33:619–31
- Sotiriou C, Wipatipat P, Loi S *et al.* (2006) Gene expression profiling in breast cancer: understanding the molecular basis of histologic grade to improve prognosis. *J Natl Cancer Inst* 98:262–72
- Srivastava A, Goldberger H, Dimtchev A *et al.* (2013) MicroRNA profiling in prostate cancer—the diagnostic potential of urinary miR-205 and miR-214. *PLoS One* 8:e76994
- Tan L, Sui X, Deng H *et al.* (2011) Holoclone forming cells from pancreatic cancer cells enrich tumor initiating cells and represent a novel model for study of cancer stem cells. *PLoS One* 6:e23383
- Tavazoie SF, Alarcon C, Oskarsson T *et al.* (2008) Endogenous human microRNAs that suppress breast cancer metastasis. *Nature* 451:147–52
- Taylor DD, Gercel-Taylor C (2008) MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol Oncol* 110:13–21
- Tombol Z, Szabo PM, Molnar V *et al.* (2009) Integrative molecular bioinformatics study of human adrenocortical tumors: microRNA, tissue-specific target prediction, and pathway analysis. *Endocr Relat Cancer* 16:895–906
- Turner BC, Zhang J, Gumbs AA *et al.* (1998) Expression of AP-2 transcription factors in human breast cancer correlates with the regulation of multiple growth factor signalling pathways. *Cancer Res* 58:5466–72
- Ueda T, Volinia S, Okumura H *et al.* (2010) Relation between microRNA expression and progression and prognosis of gastric cancer: a microRNA expression analysis. *Lancet Oncol* 11:136–46
- Vaksman O, Stavnes HT, Kaern J *et al.* (2011) miRNA profiling along tumour progression in ovarian carcinoma. *J Cell Mol Med* 15:1593–602
- Valastyan S, Reinhardt F, Benaich N *et al.* (2009) A pleiotropically acting microRNA, miR-31, inhibits breast cancer metastasis. *Cell* 137:1032–46
- van Balkom BW, de Jong OG, Smits M *et al.* (2013) Endothelial cells require miR-214 to secrete exosomes that suppress senescence and induce angiogenesis in human and mouse endothelial cells. *Blood* 121:3997–4006. S1-15
- van Kempen LC, van den Oord JJ, van Muijen GN *et al.* (2000) Activated leukocyte cell adhesion molecule/CD166, a marker of tumor progression in primary malignant melanoma of the skin. *Am J Pathol* 156:769–74
- van Mil A, Grundmann S, Goumans MJ *et al.* (2012) MicroRNA-214 inhibits angiogenesis by targeting Quaking and reducing angiogenic growth factor release. *Cardiovasc Res* 93:655–65
- Volinia S, Calin GA, Liu CG *et al.* (2006) A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci USA* 103:2257–61
- Wang F, Liu M, Li X *et al.* (2013a) MiR-214 reduces cell survival and enhances cisplatin-induced cytotoxicity via down-regulation of Bcl2l2 in cervical cancer cells. *FEBS Lett* 587:488–95
- Wang J, Li J, Wang X *et al.* (2013b) Downregulation of microRNA-214 and overexpression of FGFR-1 contribute to hepatocellular carcinoma metastasis. *Biochem Biophys Res Commun* 439:47–53
- Wang M, Zhao C, Shi H *et al.* (2014a) Deregulated microRNAs in gastric cancer tissue-derived mesenchymal stem cells: novel biomarkers and a mechanism for gastric cancer. *Br J Cancer* 110:1199–210
- Wang S, Jiao B, Geng S *et al.* (2014b) Combined aberrant expression of microRNA-214 and UBC9 is an independent unfavorable prognostic factor for patients with gliomas. *Med Oncol* 31:767
- Wang X, Chen J, Li F *et al.* (2012a) MiR-214 inhibits cell growth in hepatocellular carcinoma through suppression of beta-catenin. *Biochem Biophys Res Commun* 428:525–31
- Wang X, Guo B, Li Q *et al.* (2013c) miR-214 targets ATF4 to inhibit bone formation. *Nat Med* 19:93–100
- Wang YS, Wang YH, Xia HP *et al.* (2012b) MicroRNA-214 regulates the acquired resistance to gefitinib via the PTEN/AKT pathway in EGFR-mutant cell lines. *Asian Pac J Cancer Prev* 13:255–60
- Wang Z, Cai H, Lin L *et al.* (2014c) Upregulated expression of microRNA-214 is linked to tumor progression and adverse prognosis in pediatric osteosarcoma. *Pediatr Blood Cancer* 61:206–10
- Watanabe T, Sato T, Amano T *et al.* (2008) Dnm3os, a non-coding RNA, is required for normal growth and skeletal development in mice. *Dev Dyn* 237:3738–48
- Weng Y, Chen Y, Chen J *et al.* (2013) Identification of serum microRNAs in genome-wide serum microRNA expression profiles as novel noninvasive biomarkers for malignant peripheral nerve sheath tumor diagnosis. *Med Oncol* 30:531
- Wiggins JF, Ruffino L, Kelnar K *et al.* (2010) Development of a lung cancer therapeutic based on the tumor suppressor microRNA-34. *Cancer Res* 70:5923–30
- Williams KC, Renthal NE, Gerard RD *et al.* (2012) The microRNA (miR)-199a/214 cluster mediates opposing effects of progesterone and estrogen on uterine contractility during pregnancy and labor. *Mol Endocrinol* 26:1857–67
- Worley LA, Long MD, Onken MD *et al.* (2008) Micro-RNAs associated with metastasis in uveal melanoma identified by multiplexed microarray profiling. *Melanoma Res* 18:184–90
- Wurdinger T, Tannous BA, Saydam O *et al.* (2008) miR-296 regulates growth factor receptor overexpression in angiogenic endothelial cells. *Cancer Cell* 14:382–93
- Xia H, Ooi LL, Hui KM (2012) MiR-214 targets beta-catenin pathway to suppress invasion, stem-like traits and recurrence of human hepatocellular carcinoma. *PLoS One* 7:e44206
- Xu CX, Xu M, Tan L *et al.* (2012) MicroRNA miR-214 regulates ovarian cancer cell stemness by targeting p53/Nanog. *J Biol Chem* 287:34970–8
- Xu CZ, Xie J, Jin B *et al.* (2013) Gene and microRNA expression reveals sensitivity to paclitaxel in laryngeal cancer cell line. *Int J Clin Exp Pathol* 6:1351–61

- Xu Z, Wang T (2014) miR-214 promotes the proliferation and invasion of osteosarcoma cells through direct suppression of LZTS1. *Biochem Biophys Res Commun* 449:190–5
- Yamane K, Jinnin M, Etoh T *et al.* (2013) Down-regulation of miR-124/-214 in cutaneous squamous cell carcinoma mediates abnormal cell proliferation via the induction of ERK. *J Mol Med* 91:69–81
- Yanaihara N, Caplen N, Bowman E *et al.* (2006) Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 9:189–98
- Yang H, Kong W, He L *et al.* (2008) MicroRNA expression profiling in human ovarian cancer: miR-214 induces cell survival and cisplatin resistance by targeting PTEN. *Cancer Res* 68:425–33
- Yang TS, Yang XH, Wang XD *et al.* (2013) MiR-214 regulate gastric cancer cell proliferation, migration and invasion by targeting PTEN. *Cancer Cell Int* 13:68
- Yang Z, Chen S, Luan X *et al.* (2009) MicroRNA-214 is aberrantly expressed in cervical cancers and inhibits the growth of HeLa cells. *IUBMB Life* 61:1075–82
- Yin G, Chen R, Alvero AB *et al.* (2010) TWISTing stemness, inflammation and proliferation of epithelial ovarian cancer cells through MIR199A2/214. *Oncogene* 29:3545–53
- Yu ZW, Zhong LP, Ji T *et al.* (2010) MicroRNAs contribute to the chemoresistance of cisplatin in tongue squamous cell carcinoma lines. *Oral Oncol* 46:317–22
- Zhang L, Huang J, Yang N *et al.* (2006) microRNAs exhibit high frequency genomic alterations in human cancer. *Proc Natl Acad Sci USA* 103: 9136–41
- Zhang XJ, Ye H, Zeng CW *et al.* (2010) Dysregulation of miR-15a and miR-214 in human pancreatic cancer. *J Hematol Oncol* 3:46
- Zhang ZC, Li YY, Wang HY *et al.* (2014) Knockdown of miR-214 promotes apoptosis and inhibits cell proliferation in nasopharyngeal carcinoma. *PLoS One* 9:e86149
- Zhao C, Xu Y, Zhang Y *et al.* (2013) Downregulation of miR-145 contributes to lung adenocarcinoma cell growth to form brain metastases. *Oncol Rep* 30:2027–34