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# UNIVERSITÀ DEGLI STUDI DI TORINO

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# Targeting the MET gene for the treatment of non-small-cell lung cancer

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### **Abstract**

Recently, a better understanding of the specific mechanisms of oncogene addiction has led to the development of antitumor strategies aimed at blocking these abnormalities in different malignancies, including lung cancer. These abnormalities trigger constitutive activation of tyrosine kinase receptors (RTKs) involved in fundamental cell mechanisms such as proliferation, survival, differentiation and migration, and consequently the aberrant signaling of RTKs leads to cancer growth and survival. The inhibition of aberrant RTKs and downstream signaling pathways has opened the door to the targeted therapy era.

In non-small-cell lung cancer (NSCLC), molecular research has allowed the discrimination of different aberrant RTKs in lung cancer tumorigenesis and progression, and thus the identification of several targetable oncogenic drivers. Following the development of small molecules (gefitinib/erlotinib and crizotinib) able to reversibly inhibit the epidermal growth factor receptor (EGFR) and signaling pathways mediated by anaplastic lymphoma kinase (ALK), respectively, the MET signaling pathway has also been recognized as a potential target. Moreover, according to current knowledge, MET could be considered both as a secondary oncogenic mechanism and as a prognostic factor. Several therapeutic strategies for inhibiting activated hepatocyte growth factor receptor (HGFR) and the subsequent downstream signaling transduction have been improved in order to block tumor growth. This review will focus on the MET pathway and its role in resistance to EGFR TK (tyrosine kinase) inhibitors, the different strategies of its inhibition, and the potential approaches to overcoming acquired resistance.

## **Keywords**

Hepatocyte growth factor; MET; Kinase inhibitors; Targeted therapies; NSCLC

## **Abbreviations**

RTKs, tyrosine kinase receptors; NSCLC, non small cell lung cancer; EGFR, epidermal growth factor receptor; ALK, anaplastic lymphoma kinase; MET, hepatocyte growth factor receptor; HGF, hepatocyte growth factor; HGFR, hepatocyte growth factor receptor; Sema, semaphorin; PSI, plexin-semaphorin-integrin; SCLC, small cell lung cancer; MTD, maximum tolerated dose; AEs, adverse events; PFS,progression free survival; OS, overall survival; ITT, intent to treat; IHC, immunohistochemistry; FISH,fluorescent in situ hybridization; HR, hazard ratio

## 1. Introduction

Recently, improved understanding of the molecular mechanisms underlying oncogene addiction has led to the classification of non-small-cell lung cancer (NSCLC) into different molecular types and to the introduction of new targeted agents which have dramatically changed the natural history of this disease. Excellent examples of these agents

comprise erlotinib and gefitinib for NSCLC patients whose tumors harbor drug-sensitizing mutations in the EGFR TK domain [1], [2] and [3] and, more recently, crizotinib for NSCLCs carrying ALK translocations [4], [5], [6], [7] and [8].

Among different targets, the proto-oncogene *c-MET*, its product (HGFR, hepatocyte growth factor receptor) and its ligand (hepatocyte growth factor, HGF) have also been recognized as molecular targets. Indeed, aberrant MET pathway activation has been identified as an important oncogene addiction mechanism in different solid tumors [9] and [10] and it seems to correlate with poor clinical outcome and metastatic progression [11], [12], [13] and [14].

HGFR/HGF signaling has therefore become a new potential target for anticancer therapy through different blocking strategies, and new compounds have been tested to overcome resistance mechanisms.

### 2. HGFR/HGF structure and normal functions

#### 2.1. HGFR

The MET gene is located on chromosome 7q21-31 and encodes a protein product called the HGF receptor (HGFR). The HGFR is a single-chain heterodimer consisting of an extracellular  $\alpha$ -chain linked by a disulfide bond to a transmembrane  $\beta$ -chain possessing intracellular catalytic activity (Fig. 1 A).

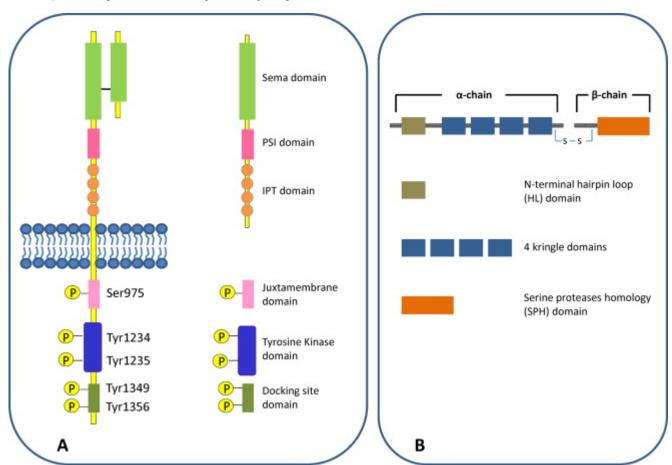


Fig. 1.
Structures of the c-MET receptor (A) and HGF (B).

The extracellular domain is composed of several domains. The semaphorin (Sema) domain, a key site for ligand binding and receptor dimerization, includes the entire  $\alpha$ -chain and the N-terminal part of the  $\beta$ -chain; a plexin–semaphorin–integrin (PSI) domain and four immunoglobulin-like (IPT) domains follow, which in turn are connected to the intracellular part of the  $\beta$ -chain (Fig. 1A).

The intracellular domain (Fig. 1A) is composed of: (1) a juxtamembrane region which contains both a serine residue (Ser975) and a tyrosine residue (Tyr1003); these are responsible for inhibiting the MET kinase activity and for degrading the receptor, respectively; (2) a catalytic region which contains two tyrosine residues (Y1234 and Y1235) that modulate the enzyme activity; and (3) a C-terminal tail, the so-called docking site [15], which contains two other tyrosine residues (Y1349 and Y1356) capable of recruiting many intracellular effectors, such as the p85 regulatory subunit of PI3K, Src and GRB2[15] and [16], and adaptors such as SHP2, PLC $\gamma$ 1 and GAB1 [15], [16] and [17], as well as STAT3[18] and [19] (Fig. 2).

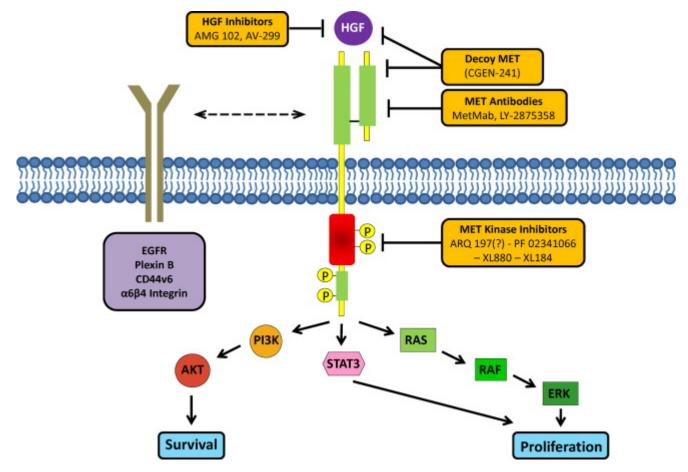


Fig. 2. MET signaling pathways and different blocking mechanisms. In addition to ligand–receptor interaction, a network of signaling coreceptors – such as the EGFR/ERB family, CD44v6, plexinB and  $\alpha6\beta4$  integrin – can interact with MET, even in an HGF-independent manner, promoting cell proliferation, invasive growth and survival. HGF inhibitors interfere with HGF binding to MET; MET antibodies prevent receptor dimerization; decoy MET prevents both HGF binding to receptor and MET dimerization; MET kinase inhibitors block MET kinase activity. ARQ 197 (?): question mark indicates that this compound is not a bona fide c-MET inhibitor.

# 2.2. HGF

HGF, the unique ligand of HGFR, belongs to the plasminogen family. It has a high affinity for HGFR and leads to activation of its receptor only in its cleaved mature form: a disulfide-bonded heterodimer which consists of an N-terminal domain, four domains known as kringle domains, and a C-terminal domain (Fig. 1B). The HGF residues that form the receptor binding site are unknown, although several studies have pointed out different roles for the  $\alpha$ - and  $\beta$ -chains [20], [21] and [22]. In particular, it would seem that a high-affinity site in the  $\alpha$ -chain is able to bind to the receptor in a manner independent of HGF maturation, even though the binding with a low-affinity site, accessible only when HGF is fully active, is necessary for receptor dimerization.

# 2.3. MET signaling activation and its normal function

Upon Sema domain-HGF binding, the MET receptor dimerizes and phosphorylation of its TK domain leads to the activation of different important pathways, such as PI3K-Akt signaling [15] and [16], Ras-MAP kinase cascades [16] and [23], STAT and the nuclear factor- $\kappa$ B complex [24] and [25], which promote cell proliferation, angiogenesis, morphogenesis, survival, cell scattering, migration and invasiveness (Fig. 2).

Moreover, HGFR can crosstalk with other pathways, such as the EGFR/ERBB family of receptors [26], even through its ligand TGF-α and K-RAS signaling [27], and it has been shown that these activated signaling pathways may be sensitive to MET inhibition both in *vitro* [28] and in *vivo* [29] (Fig. 2).

Nevertheless, it is noteworthy that a hypoxic status in tissues enhances both HGF levels and HGFR expression [30]. MET activation can also occur by semaphorins after HGFR-plexins interaction[31] and [32] (Fig. 2). In fact, HGFR and some classes of plexins share a highly homologous Sema domain, so that HGFR can be transactivated after oligomerization with plexins in response to their semaphorin ligands, also in the absence of its ligand. Moreover, MET signaling activity can be maintained even after receptor internalization [33], [34], [35], [36], [37], [38], [39] and [40]. After being internalized from the cellular membrane, the MET receptor continues to be active through its recruitment into early endosomes, favoring either HGF-induced cell migration via protein kinase C (PKC) and extracellular signal-regulated kinase (ERK), or phosphorylation and subsequent translocation into the nucleus of STAT3.

HGFR and its ligand are widely expressed in a variety of normal cells and tissues of epithelial and mesenchymal origin, respectively, and MET signaling activation has a key role in both embryonic and adult life. In particular, during embryonic development MET signaling transduction plays a crucial role in several processes, such as motogenesis [41], [42], [43] and [44], angiogenesis [45] and [46], mitogenesis [47] and morphogenesis [48] and [49]. In adult life, its signaling transduction has an important role in tissue repair and organ regeneration following acute tissue injury [50].

## 3. MET signaling in NSCLC

Several studies have demonstrated that an aberrant MET signaling pathway plays an important role in promoting tumor growth, progression and invasion in many cancers [51] and [52].

## 3.1. The different mechanisms of aberrant MET signaling activation

receptor overexpression has been reported in both small-cell lung cancer (SCLC) NSCLC [53],[54] and [55] and, in particular, has been identified in up to 40% of lung cancer tissues [56]. A significant correlation between MET receptor overexpression/hyperactivation and poor outcome has been demonstrated in different solid tumors [11], [13] and [14]. HGFR overexpression, high MET gene copy number and MET gene amplification, as well **HGF** high levels. also have а negative prognostic significance NSCLC [12], [57], [58], [59], [60], [61], [62], [63] and [64]. Besides being a consequence of HGF or HGFR overexpression – usually due to transcriptional upregulation – aberrant MET signaling can be also caused by gene amplification, activating gene mutations, or alternative splicing [53], [56] and [65].

## 3.1.1. HGF/HGFR overexpression

High HGF levels secreted by both primary and metastatic tumors (autocrine mechanism) and mesenchymal cells (paracrine mechanism) have been reported as ligand-dependent mechanism of aberrant MET signaling activation [66], [67], [68], [69] and [70]. Others studies have pointed out the importance of other tissue and transcriptional factors such as cytokines, growth factors and ETS (transcriptional factors involved in tumor invasion program), in

inducing MET upregulation and ultimately receptor overexpression, regardless of HGF stimulation [30], [71], [72], [73], [74], [75] and [76].

Hypoxic conditions can also stimulate both higher HGF levels and HGFR transcriptional levels via hypoxia-inducible factor- $1\alpha$  (HIF- $1\alpha$ ), which renders the cells more sensitive to HGF stimulation in the tumor invasion process; therefore, MET overexpression by itself nourishes this hypoxia-dependent invasion mechanism [30].

# 3.1.2. MET gene mutations

Activating point mutations in the MET coding sequence have been reported as somatic and germline variants in many solid tumors, albeit infrequently. In NSCLC, MET gene mutations can occur in the semaphorin extracellular domain, in the juxtamembrane region and also in the kinase domain. Missense mutations found in the Sema domain, encoded by exon 2 and necessary for MET dimerization, have been reported as germline mutations [77]. The mutations found in the juxtamembrane domain, encoded by exons 14–15, seem to be involved in tumorigenesis [78], [79], [80] and [81]. Nevertheless, the MET juxtamembrane region, necessary for receptor downregulation, contains a tyrosine residue (Tyr1003) which is able to bind to the c-Cbl TK domain; this complex in turn promotes MET polyubiquination and receptor degradation. A point mutation in this tyrosine residue does not permit the formation of the complex and polyubiquination, leading conversely to MET oncogenic activity [82].

Mutations in the MET tyrosine kinase domain [53] and [83], primarily described in patients with hereditary and sporadic papillary renal cell carcinoma [84] and head/neck squamous-cell carcinoma [85], have been found rarely in NSCLC as a secondary event resulting from exposure to prior therapies, such as tyrosine kinase inhibitors [86] and [87].

# 3.1.3. MET gene amplification

*MET* gene amplification has been reported in many primary human tumors and acts as a primary "oncogenic driver" in 2–21% TKI-naïve lung adenocarcinomas [53]. *MET* amplification has also been detected as a secondary event, both in preclinical and clinical studies, in EGFR–TKI-resistant NSCLC after exposure to gefitinib or erlotinib [88] and [89] with a frequency ranging from 5% to 25% [88], [89], [90], [91] and [92]. In this NSCLC population, TKI treatment specifically selects preexisting *MET*-amplified clones in which the ERBB3/PI3K/AKT signaling pathway is active, thus suggesting the potential impact of a concomitant blockade of MET for overcoming EGFR–TKI resistance [93] and [94]. The combination treatment strategy has proved capable of also overcoming primary EGFR–TKI resistance, as recently shown in xenograft models [95]. In another study, the use of golvatinib, a multitarget small-molecule inhibitor, has been shown to restore sensitivity to EGFR–TK inhibition and to prevent the emergence of resistant cell clones after continuous HGF exposure in vitro [96]. Indeed, another mechanism of both primary and acquired resistance to EGFR–TKIs is represented by HGF overexpression [97]. Furthermore, gefitinib-resistant MET-amplified NSCLC (HCC827 GR) cells showed an increased activation of the tyrosine kinase Src [98], and the use of Src inhibitors resulted in tumor-cell inhibition and apoptosis [99].

## 3.2. Mechanisms of acquired resistance to MET inhibition

As is known with EGFR and ALK inhibition, MET-driven NSCLC patients treated with a specific targeted strategy also invariably develop secondary resistance mechanisms which lead to tumor progression. To our knowledge, two mechanisms are involved in acquired resistance to MET inhibition, and both of them are simultaneously present: a point mutation in the MET tyrosine kinase domain at the tyrosine residue Y1230, and TGF- $\alpha$  overexpression, a condition that can activate alternative EGFR pathways [100]. In these cases, a treatment strategy combining both MET and EGFR inhibitors might allow to overcome resistance [53]. Other acquired mechanisms of resistance to MET inhibition have been identified in different solid tumors, such as papillary renal-cell carcinoma; these include an increased representation of a preexisting sensitive de novo somatic M1268T mutation associated with a copy number gain and

expression of genomic duplication [101]. On the other hand, aberrant MET signaling activation can represent a mechanism of oncogene expedience as a secondary event owing to the interference of other oncogenes (K-RAS), proinflammatory cytokines, HGF itself, and microenvironmental conditions (hypoxia) that can enhance invasiveness and metastatic properties of neoplastic cells.

# 4. Current treatment strategies targeting MET in NSCLC

Different targeting strategies to inhibit the aberrant MET signaling in NSCLC have been developed. In particular, MET signaling can be blocked at the ligand–receptor level by new drugs directed against HGF (HGF antagonists) or HGFR, such as anti-MET receptor monoclonal antibodies and decoy MET (the latter has so far been tested only in preclinical phase studies), or at the tyrosine kinase domain level by small-molecule MET kinase inhibitors (Fig. 1).

Currently, many molecules, either alone or in combination with other drugs, are under investigation in clinical trials on NSCLC in order to prevent or overcome possible mechanisms of resistance. A summary of these trials is provided in Table 1 and Table 2.

 Table 1.

 Ongoing studies with MET inhibitors in non-small-cell lung cancer (NSCLC)

Experimental drug	Combined treatment drug(s)	Study design	Description and primary end-points					
HGF inhibitors								
AMG 102 (rilotumumab)	Erlotinib	Phase I/II trial	AMG 102 + erlotinib in previously treated advanced NSCLC					
			Primary: safe dose to combine with erlotinib					
AV-299 (ficlatuzumab)	Gefitinib	Phase I/II trial	AV-299 + gefitinib AND gefitinib alone in NSCLC Asian patients					
			Primary: DLTs and RP2D; ORR					
Anti-MET monoc	lonal antibodies							
MetMab (onartuzumab)	Bevacizumab	4-arm phase II trial	MetMab + different chemotherapy treatments versus placebo + the same chemotherapy regimen in untreated advanced non-squamous NSCLC patients.					
	Platinum-based chemotherapy doublets		Primary: PFS in both the entire population and patients with MET/IHC+tumors					

Experimental drug	Combined treatment drug(s)	Study design	Description and primary end-points
	Placebo		
MetMab	Platinum-based chemotherapy doublet	2-arm phase II trial	MetMab + platinum-based chemotherapy doublet versus placebo + platinum-based chemotherapy doublet in untreated advanced squamous NSCLC patients
	Placebo		Primary: PFS both in ITT population and in patients with MET-positive squamous tumors, as assessed by IHC
MetMab	Erlotinib	2-arm phase III trial	MetMab + erlotinib versus placebo plus erlotinib in pretreated NSCLC patients with MET-positive tumors
	Placebo		Primary: OS
MET tyrosine kin	ase inhibitors		
PF 02341066 (crizotinib)	PF 00299804	2-arm phase I trial	PF 02341066 + PF 00299804 versus PF 00299804 followed by PF 02341066 plus PF 00299804 in advanced NSCLC patients
			Primary: safety profile
PF 02341066	PF 00299804	Phase I trial	PF 02341066 + PF 00299804 in pretreated advanced NSCLC patients.
			Primary: safety profile
ARQ 197	Sorafenib	Phase I trial	ARQ 197 + sorafenib in advanced solid tumors
			Primary: MTD and/or RP2D
			Secondary: PK; antitumor activity; changes of HGF, VEGF and c-MET in peripheral blood
ARQ 197	Erlotinib	2-arm phase II trial	ARQ 197 + erlotinib versus single-agent chemotherapy in previously treated advanced NSCLC carrying K-RAS mutation

Experimental drug	Combined treatment drug(s)	Study design	Description and primary end-points
	Pemetrexed		Primary: PFS
	Docetaxel		
	Gemcitabine		
ARQ 197	Erlotinib	Phase II trial	ARQ 197 + erlotinib in advanced NSCLC patients with tumors harboring EGFR mutation progressing on EGFR TKI monotherapy
			Primary: ORR
ARQ 197	Erlotinib	2-arm phase III trial	ARQ 197 + erlotinib versus placebo + erlotinib in pretreated advanced non-squamous EGFR wild-type NSCLC Asian patients
	Placebo		Primary: OS
XL184 (cabozantinib)	Erlotinib	Phase I/II trial	XL184 + erlotinib (Part 1) AND XL184 + erlotinib compared to XL184 alone (Part 2) in NSCLC patients beyond erlotinib progression
			Primary: safety, tolerability and MTD; PK; PD; ORR
XL184	Placebo	Phase II discontinuation trial	Responding patients will continue treatment, those with stable disease will be randomized to continue XL184 until disease progression versus placebo; a non-randomized expansion cohort is expected for patients receiving placebo. Patients progressing on XL184 will discontinue study treatment.
			Primary: efficacy
			Secondary: safety and tolerability; PK and PD; correlation between MET expression and clinical outcome
XL184	Erlotinib	3-arm phase II trial	Erlotinib versus XL184 versus erlotinib plus XL184 as 2nd or 3rd line therapy in metastatic EGFR wild-type NSCLC patients.

Experimental drug	Combined treatment drug(s)	Study design	Description and primary end-points
			Primary: PFS
GSK 1363089 (foretinib)	Erlotinib	2-arm phase I/II trial	GSK 1363089 + erlotinib versus erlotinib alone in previously treated NSCLC patients
			Primary: RP2D of GSK 1363089 in combination with erlotinib; safety, tolerability and DLTs; PK; toxicity; ORR
INC280	Gefitinib	Phase lb/II	INC280 + gefitinib in EGFR-mutated and c-MET amplified NSCLC patients who have progressed after EGFR inhibitor treatment
			Primary: DLTs (Phase I) and ORR (Phase II)
			Secondary: OS, safety, inhibition of c-MET signaling, PK

PK, pharmacokinetics; PD, pharmacodynamics; MTD, maximum tolerated dose; DLTs, dose limiting toxicities; RP2D, recommended phase 2 dose; HGF, hepatocyte growth factor; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor; DCR, disease control rate; ORR, objective response rate; PFS, progression free survival; OS, overall survival; ITT, intent to treat; IHC, immunohistochemistry.

**Table 2.**Ongoing phase I/II studies with new small molecules targeting MET

Trials (ClinicalTrials.gov Identifier)	Experimental drug(s)	Mechanism of action	Description and End-points
NCT01014936	EMD 1214063	MET kinase inhibition	Open-label, dose-escalation, first-in-man, non-randomized trial in patients with advanced solid tumors under different regimens.
			Primary: MTD
			Secondary: safety profile and tolerability; PK; PD; safety profile; anti-tumor effects, and PK/PD in subjects with and without specific c-Met alterations, anti-tumor effects and others
NCT01253707	AMG 337	MET kinase	Open-label, first-in-human, sequential dose-

Trials (ClinicalTrials.gov Identifier)	Experimental drug(s)	Mechanism of action	Description and End-points
		inhibition	escalation and expansion trial in adults with advanced solid tumors.
			Primary: safety and tolerability; PK and MTD
			Secondary: clinical response and tumor anti- proliferative response
NCT00813384	AMG 208	MET kinase inhibition	Open-label, first-in-human, sequential dose- escalation and expansion trial in adults with advanced solid tumors.
			Primary: MTD; clinical response; PK; safety and tolerability
			Secondary: decrease in tumor cell proliferation according to PET scanning; tumor volume changes according to CT/MRI scanning; biomarkers on skin specimens; correlation between c-MET expression, amplification or mutation and response
NCT01721148	BMS 777607	MET kinase inhibition	Phase I multiple ascending dose study in patients with advanced/metastatic solid tumors
			Primary: MTD
			Secondary: PK and antitumor activity
NCT01428141	E7050	MET kinase inhibition (and VEGFR2)	Phase I study in adults with advanced solid tumors (and gastric cancer).
			Primary: MTD
NCT01433991	E7050	MET kinase inhibition	Open-label phase Ib/II study to assess the combination of E7050 and E7080 in adults with advanced tumors (dose-escalation part) and with recurrent glioblastoma or unresectable stage III/IV melanoma (expansion cohort and phase II)

Trials (ClinicalTrials.gov Identifier)	Experimental drug(s)	Mechanism of action	Description and End-points
	E7080	Multi-target inhibition	Primary: DLTs and MTD (phase lb), antitumor activity (Phase II)
			Secondary: PK and PD of the two drugs, administered alone or in combination; antitumor activity
NCT00697632	MGCD265	Multi-target inhibition (including MET Kinase)	Open-label, phase I dose-escalation study to evaluate MGCD265 administered without interruption in patients with advanced tumors
			Primary: safety and tolerability
			Secondary: PK, PD and clinical response
NCT00975767	MGCD265	Multi-targets inhibition (including MET kinase)	Phase I/II study combining MGCD265 with erlotinib or docetaxel in patients with advanced tumors and advanced NSCLC
	Erlotinib		Primary: safety profile (phase I); antitumor activity (Phase II)
	Docetaxel		Secondary: PK, PD and antitumor activity (Phase I); safety profile (Phase II)
NCT01588821	Cabozantinib	Multi-target kinases inhibition (including MET and VEGFR2)	Phase II trial in patients with advanced solid (except breast and prostate) tumors and bone metastases
			Primary: effect on bone biomarkers, such as NTx, CTx and others
			Secondary: rate of SRE; QoL; ORR; correlation between response and MET amplification; response in metastatic bone sites; time to SRE

MTD, maximum tolerated dose; PK, pharmacokinetics; PD, pharmacodynamics; PET, positron emission tomography; CT, computed tomography; MRI, magnetic resonance imaging; VEGFR2, vascular endothelial growth factor receptor 2; DLTs, dose-limiting

toxicities; NTx, N-terminal telopeptide; CTx, C-terminal telopeptide; SRE, skeletal-related events; QoL, quality of life; ORR, objective response rate.

# 4.1. HGF antagonists

AMG-102 (rilotumumab) and AV-299 (ficlatuzumab) are the two anti-HGF monoclonal antibodies currently in the most advanced phases of clinical investigation.

#### 4.1.1. AMG-102

AMG-102, a fully human anti-HGF IgG2 monoclonal antibody, has shown preclinical activity both in vitro and in vivo by enhancing temozolomide and docetaxel efficacy in xenograft models overexpressing the HGF/MET pathway [102]. In the ensuing phase I trial, the maximum tolerated dose (MTD) was set at 20 mg/kg and was administered according to a 2-week schedule; the main adverse events (AEs) were fatigue, nausea, constipation, peripheral edema and hypertension of low to moderate grade [103]. This experimental drug has also shown good tolerability in association with antivascular agents [104]. AMG-102in association with standard chemotherapy in patients with advanced or metastatic gastric or gastroesophageal junction cancers has shown encouraging results, particularly in those with MET-positive tumors [105]. This compound is currently under evaluation in MET-positive gastric and gastroesophageal junction cancers and in pretreated NSCLC in combination with erlotinib.

## 4.1.2. AV-299

AV-299, a human anti-HGF IgG1 monoclonal antibody, showed a good tolerability profile and additive activity when combined with chemotherapy and an anti-EGFR drugs [106]. In a phase I trial AV-299 was administered at the previous MTD (20 mg/kg every 2 weeks) in combination with gefitinib (250 mg once daily); five out of 15 Asian NSCLC patients achieved a response and treatment was well tolerated. The most frequent AEs were rash, cough, diarrhea, decreased appetite, edema, drug hypersensitivity reactions and fatigue [107]. A randomized phase II study is evaluating AV-299 in combination with gefitinib in never smokers or light smokers and untreated Asian patients.

#### 4.2. Anti-HGFR monoclonal antibodies

MetMab (onartuzumab) is a humanized, monovalent, monoclonal antibody directed against the MET receptor which inhibits HGF/MET binding without exerting agonistic activity and inducing MET dimerization. It demonstrated significant antitumor activity both in glioblastoma featuring an HGF-dependent autocrine loop [108] and in pancreatic [109] xenograft models. In the following phase I trial MetMab was well tolerated both as a single agent and in combination with bevacizumab, and the phase II recommended dose was set at 15 mg/kg every 3 weeks [110].

More recently, a global randomized phase II study (OAM4558g) comparing MetMab in combination with erlotinib or placebo as second- or third-line treatment for NSCLC has been completed and the efficacy data presented ( Table 3) [111]. This study showed no significant improvement in progression-free survival (PFS) or overall survival (OS) in the intent-to-treat (ITT) population (137 patients). However, according to MET status determined by immunohistochemistry (IHC), patients with MET-positive (IHC score: 2+ or 3+) tumors benefited significantly from the addition of MetMab to erlotinib in terms of both median PFS (HR = 0.53, P = 0.04) and OS (HR = 0.37, P = 0.002). A significant improvement in OS was also observed in MET fluorescence in situ hybridization- (FISH-) positive (HR = 0.60) and MET FISH-negative/IHC-positive (HR 0.37, P = 0.01) NSCLC patients, which suggests that IHC might be a more sensitive tool for predicting a clinical benefit from MetMab. A significant improvement in OS (HR 0.45) was also observed in MET FISH-negative/IHC-positive/EGFR wild-type NSCLCs. Conversely, the addition of MetMab resulted in a detrimental effect in

patients with low MET expression (0 and 1+) tumors. *MetMab* in combination with erlotinib was well tolerated. Adverse events – such as rash, diarrhea, fatigue and nausea/vomiting – were comparable between the two treatment arms; peripheral edema was the only adverse event which occurred with a significantly higher frequency in the combination drug group (22.9% versus 6.5%).

Table 3.
Randomized phase II/III trials with MetMab [111] and Tivantinib [122] and [123].

OAM4558g - Randomized phase II study to evaluate MetMab or placebo in combination with erlotinib in advanced NSCLC[111]

		Median				Median PFS (months)					
	n			HR	CI 95%	p			HR	CI 95%	Р
		PE	ME				PE	ME			
ITT population	137	7.4	8.9	0.80	0.50– 1.28	0.34	2.6	2.2	1.09	0.73– 1.62	0.69
c-MET IHC+	66	4.6	12.6	0.37	0.19– 0.72	0.002	1.5	2.9	0.53	0.28- 0.99	0.04
c-MET IHC-a	56	15.3	8.1	1.78	0.79– 3.99	0.16	2.7	1.4	1.82	0.99– 3.32	0.05

Randomized phase II study to evaluate erlotinib plus tivantinib versus erlotinib plus placebo in previously treated, EGFR inhibitor-naive NSCLC patients [122]

			Median OS (months)					Median PFS (months)				
	n			HR	CI 95%	p			HR	CI 95%	p	
		EP	ET				EP	ET				
ITT population	167	6.8	8.5	0.87	0.59– 1.27	0.47	2.3	3.8	0.81	0.57– 1.16	0.24	
Non-SQCC histology	117	6.8	9.9	0.72	0.44– 1.17	0.18	2.2	4.3	0.71	0.46– 1.10	0.12	

Randomized phase III study to evaluate erlotinib plus tivantinib versus erlotinib plus placebo in previously treated, EGFR inhibitor-naive non-squamous NSCLC patients [123]

	n	Median OS (months)	HR	CI 95%	р	Median PFS (months)	HR	CI 95%	p	
ITT population										
Erlotinib + Tivantinib	526	8.5	0.98	0.84– 1.15	0.81	3.6	0.74	0.64– 0.85	<0.001	
Erlotinib + Placebo	522	7.8				1.9				
MET high IHC bsubgro	ир рор	ulation								
Erlotinib + Tivantinib	104	9.3	0.70	0.49– 1.01	0.03	3.6	0.72	0.52– 0.98	0.014	
Erlotinib + Placebo	107	5.9				1.9				
MET low IHC subgroup population										
Erlotinib + Tivantinib	107	8.5	0.90	0.64– 1.26	0.53	3.7	0.66	0.49– 0.89	0.006	
Erlotinib + Placebo	127	7.7				1.9				

EP: erlotinib plus placebo; ET: erlotinib plus tivantinib; SQCC: squamous cell carcinoma.

ITT: intent to treat; OS: overall survival; PFS; progression free survival; HR: hazard ratio; CI: confidence interval; IHC: immunohistochemistry.

а

Initial data cut; OS: overall survival; PFS; progression free survival; HR: hazard ratio; CI: confidence interval; ITT: intent to treat; PE: placebo plus erlotinib; ME: MetMab plus erlotinib; NA: not available; IHC: immunohistochemistry.

b

MET high IHC: ≥50% of tumor tissue stained with an intensity of 2+ and/or 3+.

*MetMab* also induced a 2-year complete response in a young patient with pretreated metastatic gastric cancer expressing high MET gene polysomy [112].

A randomized phase III study in pretreated NSCLC patients is currently evaluating *MetMab* in combination with erlotinib versus erlotinib alone in MET 2+/3+ tumors [113].

Another specific monoclonal antibody, LY-2875358, has been tested so far only in a phase I trial [114].

# 4.3. MET tyrosine kinase inhibitors

The inhibition of MET tyrosine kinase activity represents the last strategy developed to block the aberrant MET signaling pathway. A myriad new small molecules targeting the MET tyrosine kinase domain are currently under investigation in phase I–II trials, such as INC-280, EMD 1214063, EMD 1204831, PF-04217903, AMG 337, AMG 208, BMS-777607, LY-2801653, E-7050, MK2461, JNJ-38877605, MP470, SGX523, and MGCD-265 [53] and [114] (Table 2).

More recently, EMD 1214063 and EMD 1204831 showed a highly potent and reversible c-MET phosphorylation inhibition both in vitro and in vivo, which resulted in tumor regression in xenograft models in both an HGF-dependent and -independent manner [115].

Tivantinib (ARQ197), cabozantinib (XL184), foretinib (XL880) and crizotinib (PF 02341066) are in the more advanced phases of clinical development (Table 1).

# 4.3.1. Tivantinib (ARQ 197)

Until recently *tivantinib* (ARQ 197) has been considered a non-competitive ATP small molecule that inhibits in a very highly selective manner the inactive form of the MET tyrosine kinase domain. In fact, it demonstrated cytotoxic effects both in vitro and in vivo by inhibiting MET phosphorylation and its downstream effectors [116]. Conversely, as recently demonstrated in different cell models and in contrast to our previous understanding, the stabilization of microtubules represents the real mechanism of action of this compound [117]. Tivantinib showed its activity against both c-MET-dependent and -independent cell lines. In particular, in addition to inhibition of c-MET signaling, its potent activity primarily involved disruption of the microtubule dynamics via inhibition of tubulin polymerization, a mechanism similar to that of vinca alkaloids [118].

This compound has been studied in different tumor types, including NSCLC, both alone and in combination with erlotinib [119], [120] and [121]. In a phase I trial enrolling unselected patients with advanced solid tumors, six out of eight NSCLC patients treated with tivantinib plus erlotinib achieved prolonged disease stability, and the recommended phase Il study dose was set at 360 mg twice daily. Combination treatment was well tolerated, and fatigue, nausea/vomiting and diarrhea were the most common treatment-related adverse events [119]. Following these results, a randomized, doubleblind, placebo-controlled phase II trial, having PFS as the primary endpoint, was designed to evaluate erlotinib plus tivantinib in previously treated, EGFR-TKI-naïve, advanced NSCLC patients [122]; 167 patients were randomized in a 1:1 fashion to receive erlotinib (E) plus tivantinib (T) or placebo (P) (Table 3). For patients randomized to receive EP, crossover to ET was allowed on disease progression. Median PFS in the ITT population did not differ significantly between the two treatment arms (HR = 0.81, P = 0.24). After adjusting for key prognostic factors, PFS in the ITT population was significantly better in the experimental arm (HR = 0.68, P = 0.04). Notably, a significant advantage in terms of PFS (adjusted HR = 0.61; P = 0.04) and OS (adjusted HR = 0.58; P = 0.04) was observed in the non-squamous NSCLC subgroup treated with ET, with an impressive delay of 7.4 months in the development of new metastases. Nevertheless, the K-RAS-mutated NSCLC subgroup significantly benefited from ET treatment (median PFS 2.9 versus 1.0 months; HR = 0.18; Cl95% 0.05–0.70; P = 0.01); also EGFR wild-type NSCLC population had a longer median PFS when treated with combined drugs (3.2 versus 1.9 months; HR 0.70; CI95% 0.44–1.10), P = 0.12). The most common AEs of all grades were rash, diarrhea, fatique, anorexia, and nausea/vomiting. There were no significant differences in either overall or serious AEs between the two arms.

A phase II trial to evaluate tivantinib plus erlotinib compared to single-agent chemotherapy in previously treated advanced non-squamous KRAS-positive NSCLCs is currently ongoing.

A phase III study (MARQUEE trial) [123] in previously treated, but EGFR- and MET-inhibitors-naïve, non-squamous NSCLC patients was discontinued early because the preplanned interim analysis failed to demonstrate a significant advantage in terms of OS, its primary end-point. Final results have recently been given at the 38th ESMO Annual Meeting (Table 3). The study confirmed that in the ITT population there was a significant improvement in PFS (HR 0.74, P < 0.001) and ORR in favor of the combination arm, but this advantage did not translate into a significant improvement in OS (HR 0.98, P = 0.81). The combined treatment was generally well tolerated; patients treated with erlotinib plus tivantinib developed a higher incidence of neutropenia of grade  $\geq$ 3 (10% versus 1.0%). However, in the subgroup of patients whose tumors had higher MET IHC expression (2+ or 3+), the addition of tivantinib did improve OS significantly (HR = 0.70, P = 0.03), suggesting the potential for efficacy in a biomarker-selected population.

CYP2C19 is known to be the main enzyme system involved in tivantinib metabolism. Specific CYP2C19 polymorphisms, expressed mainly in Asian populations (20% versus 3% in Caucasian populations), define a subgroup of patients as poor metabolizers; for these patients, the tivantinib dose has been set at 240 mg twice daily [124]. Another phase III trial (ARQ197-006) in advanced non-squamous EGFR wild-type NSCLC Asian patients is currently ongoing to evaluate the combination of tivantinib and erlotinib versus erlotinib plus placebo. Tivantinib dose administration is based on CYP2C19 polymorphism (ATTENTION, NCT01377376).

# 4.3.2. Cabozantinib (XL184)

Cabozantinib (XL184) is a multitarget ATP competitive inhibitor of MET, VEGFR2 and RET, and it has shown antitumor activity in experimental MET-driven and not-MET-driven models [125]. Cabozantinib has been studied either alone or in association with erlotinib in NSCLC with acquired resistance to erlotinib; the results were encouraging, with several partial responses and prolonged disease stabilization, especially in NSCLC patients with MET-amplified tumors and harboring EGFR T790M mutation [126]. In the following phase II randomized discontinuation trial, which enrolled 483 patients with different solid tumors, cabozantinib demonstrated good activity [127]. In particular, in the NSCLC subgroup ORR was 13% (6/47 patients), with a disease control rate of 40%. In the overall population, the most common AEs of grade ≥3 were fatigue (9%), hand–foot syndrome (8%) and hypertension (5%). Interestingly, the presence of EGFR/K-RAS mutations was a predictor of clinical benefit; soft tissue, visceral and bone lesions were identified as the tumor sites where the drug was more active.

In other phase II discontinuation trials, cabozantinib has been shown to be active also in advanced ovarian[128] and prostate [129] cancers.

# 4.3.3. Foretinib (XL880)

Foretinib (XL880, GSK1363089) is a multitarget inhibitor directed against MET/VEGFR, RON, AXL, PDGFR-β, KIT, FLT3 and TIE-2. Foretinib has shown antitumor activity in vivo [130]. Results of a phase I study enrolling 40 patients with chemorefractory advanced solid tumors [131] showed partial responses in one medullary thyroid carcinoma and in two papillary renal-cell carcinomas. MTD was set at 3.6 mg/kg administered for 5 consecutive days every 2 weeks; the investigational drug was well tolerated. Hypertension, fatigue, diarrhea, vomiting, proteinuria and hematuria were the most common adverse events. At maximum administered dose (4.5 mg/kg), grade 3 elevations in aminotransferase (AST) and lipase levels were reported.

Recently, the results of a phase II study of foretinib in 74 patients with papillary renal-cell carcinoma confirmed the antitumor activity of the compound, especially in tumors with germline MET mutations [132]. At present, two phase II studies of foretinib in other cancers have been completed and results are awaited; moreover, the inhibitor is currently

under investigation in a phase I/II study in patients with previously treated, advanced or metastatic NSCLC (erlotinib versus erlotinib plus foretinib).

# 4.3.4. Crizotinib (PF 02341066)

Crizotinib (PF 02341066), a strong ALK inhibitor approved in August 2011 by the FDA for the treatment of ALK-rearranged NSCLC, was initially investigated as a c-MET inhibitor. Crizotinib showed a good antitumor activity in both preclinical [133] and [134] and phase I [135] studies, demonstrating that its activity correlated with its ability to inhibit MET phosphorylation. More recently, crizotinib was highly effective in a patient with NSCLC harboring a de novo MET amplification [136] and in MET-amplified gastroesophageal adenocarcinomas and glioblastoma [137] and [138]. Therefore, it is possible that crizotinib might possess a different antitumor activity according to MET gene alterations [139]. In a more recent study, Zhang et al. investigated crizotinib efficacy in different brain tumor cells. Interestingly, the authors demonstrated a heterogeneous antitumor activity and identified high HGF expression as a key determinant of major responsiveness to crizotinib through inhibition of the ERK/JAK/p53 pathway [140]. Surprisingly, a short-term pretreatment with exogenous HGF resulted in a greater cell apoptosis and tumor growth inhibition after crizotinib exposure than in non-pretreated cells both in vitro and in vivo.

Two phase I trials combining crizotinib with a pan-HER inhibitor in advanced NSCLC are currently ongoing.

# 4.4. On-target-based toxic effects of MET inhibition etc.

On-target toxicity refers to the exaggerated and adverse class effects due to inhibition of the direct target in normal cells and tissues.

Regarding MET inhibitors, particularly monoclonal antibodies and small molecules, these compounds have to date demonstrated few adverse effects in normal tissues. In fact, several MET inhibitors have been investigated in full doses both in combination with chemotherapy and other targeted drugs (EGFR-TK and VEGF inhibitors), and any combination treatment was well tolerated without enhanced known toxic effects. In particular, peripheral edema seems to be a peculiar on-target toxicity. Other adverse events seen in the trials with MET inhibitors – such as fatigue, anorexia, nausea/vomiting, fever and hypersensitivity reactions – arose regardless of their target.

Off-target toxicity refers to adverse effects related to inhibition activity on other targets. It occurs when a TK inhibitor causes a toxic effect by the inhibition of a kinase not known to be the target of a specific compound.

Regarding MET inhibitors, the majority of off-target adverse effects, such as proteinuria, hematuria, hypertension and bleeding, seem to be correlated with inhibition of other targets (VEGFR and others).

Drug-related toxicity refers to the physicochemical properties of a specific compound and their effects on cell compartments and metabolism.

A selective MET inhibitor, SGX523, was found to be safe and very effective in preclinical studies, but was discontinued early in a phase I trial because of the appearance of renal failure. This unexpected drug-related toxicity in humans was due to the presence of two insoluble drug metabolites that crystallized in renal tubules, leading to tubulointerstitial nephritis [141].

Although several toxicities are driven by plasma free drug levels, on-target tissue toxicities seem to be also correlated with drug concentrations in the target tissue. A recent pharmacokinetic study demonstrated that the significant liver toxicity of GEN-203, a small molecule MET inhibitor, was correlated with its lipophilic characteristics which enhance its distribution into liver tissue. By modifying the chemical properties of the compound, the authors were able to reduce its basicity and tissue concentrations, improving the safety profile of the drug [142].

## 5. Conclusions

Many preclinical and clinical studies have demonstrated the important role of the MET pathway in tumors, and also in NSCLC. The MET pathway can be aberrantly activated as a consequence of HGF or HGFR transcriptional upregulation, MET gene amplification and, rarely, as the final event of a MET gene mutation. Other signaling co-receptors can crosstalk with MET, even in an HGF-independent manner, providing an alternative way to induce proliferation, survival and invasive growth. On the other hand, aberrant MET signaling activation can represent an oncogene expedience to enhance invasiveness and metastatic properties of pretreated neoplastic cells.

Different compounds targeting MET have been developed and have shown antitumor activity; some of them, such as *MetMab* and tivantinib, are currently under advanced clinical development in NSCLC. A plethora of new molecules targeting MET or acting as multitarget inhibitors is emerging, but the increasing availability of new compounds might represent a double-edged sword. In fact, whereas ever-growing pharmaceutic efforts have led to the introduction of new molecules into preclinical and clinical studies, this race might generate confusion about the exact mechanisms through which these new drugs block tumor growth and those through which tumor cells are able to escape their effects. In this regard, we hope that the lesson learned from tivantinib will serve as a warning.

According to the safety profile of these drugs, overall selective MET inhibitors seem to have no particular toxicity issues, and the main expected toxicities are manageable. However, we think that our experience of the toxicity profile of these molecules is very limited, in particular regarding the late toxic effects.

Several questions remain to be answered to optimize MET-targeted therapy. First, we need to understand comprehensively if and when genetic alterations involving MET induce "oncogene addiction" or "oncogene expedience". This is a relevant issue because of its therapeutic implications; the mechanism underlying MET oncogene addiction is not yet fully elucidated, whereas a role for MET is now emerging in oncogene expedience which can potentiate other oncogenes and accelerate tumor progression. Therefore, we need to identify definitively the subgroup of tumors that can benefit from MET inhibition. Second, the best treatment strategy has yet to be determined; in particular, we have to clarify the best drug combination treatment (MET and EGFR inhibitors, MET and other inhibitors) and if and when a MET monotarget approach might have a role. Third, we have to determine the best sequence to use and the most appropriate setting in which we should consider its use. Finally, we need to identify specific tools as predictive factors of better outcome of MET inhibition. In this regard, at present MET IHC overexpression seems to have a promising role, but other efforts are required to detect new circulating predictive biomarkers.

Understanding the specific mechanisms of drug resistance is critical in selecting and evaluating subsequent therapeutic approaches. The strategy of combining treatments targeting several molecules, or molecules blocking downstream signaling transducers, might be used in order to address the common problem of crosstalk between signaling pathways and thus the development of resistance.

With the efforts of translational and clinical research, in a few years MET-targeted therapies will surely impact on lung cancer outcome, and they must be welcomed as one of the possible therapeutic options for patients with NSCLC.

# Conflict of interest statement

The authors declare they have no conflict of interest.

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## References

- [1] T.J. Lynch, D.W. Bell, R. Sordella, *et al.* Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. N Engl J Med, 350 (2004), pp. 2129–2139
- [2] J.G. Paez, P.A. Janne, J.C. Lee, *et al.* EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. Science, 304 (2004), pp. 1497–1500
- [3] L.V. Sequist, D.W. Bell, T.J. Lynch, Haber DA: Molecular predictors of response to epidermal growth factor receptor antagonists in non-small-cell lung cancer. J Clin Oncol, 25 (2007), pp. 587–595
- [4] E.L. Kwak, Y.J. Bang, D.R. Camidge, *et al.* Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. N Engl J Med, 363 (2010), pp. 1693–1703
- [5] A.T. Shaw, B.Y. Yeap, B.J. Solomon, *et al.* Effect of crizotinib on overall survival in patients with advanced non-small-cell lung cancer harbouring ALK gene rearrangement: a retrospective analysis. Lancet Oncol, 12 (2011), pp. 1004–1012
- [6] D.R. Camidge, Y.J. Bang, E.L. Kwak, *et al.* Activity and safety of crizotinib in patients with ALK-positive non-small-cell lung cancer: updated results from a phase 1 study. Lancet Oncol, 13 (2012), pp. 1011–1019
- [7] Shaw AT: Phase III study of crizotinib versus pemetrexed or docetaxel chemotherapy in patients with advanced ALK-positive NSCLC (PROFILE 1007). Proceedings of ESMO congress: 28 September–02 October Vienna (2012) [Abstract 2862]
- [8] D.-W. Kim, M.-J. Ahn, Y. Shi, et al. Results of a global phase II study with crizotinib in advanced ALK-positive non-small cell lung cancer (NSCLC). J Clin Oncol, 30 (2012), p. s7533
- [9] S. Giordano, C. Ponzetto, M.F. Di Renzo, C.S. Cooper, Comoglio PM. Tyrosine kinase receptor indistinguishable from the c-met protein. Nature, 339 (1989), pp. 155–156
- [10] M. Prat, R.P. Narsimhan, T. Crepaldi, M.R. Nicotra, P.G. Natali, Comoglio PM. The receptor encoded by the human c-MET oncogene is expressed in hepatocytes, epithelial cells and solid tumors. Int J Cancer, 49 (1991), pp. 323–328
- [11] D.L. Shattuck, J.K. Miller, K.L. Carraway 3rd, C. Sweeney. Met receptor contributes to trastuzumab resistance of Her2-overexpressing breast cancer cells. Cancer Res, 68 (2008), pp. 1471–1477
- [12] J.M. Siegfried, L.A. Weissfeld, P. Singh-Kaw, R.J. Weyant, J.R. Testa, Landreneau RJ. Association of immunoreactive hepatocyte growth factor with poor survival in resectable non-small cell lung cancer. Cancer Res, 57 (1997), pp. 433–439
- [13] K. Sawada, A.R. Radjabi, N. Shinomiya, *et al.* c-Met overexpression is a prognostic factor in ovarian cancer and an effective target for inhibition of peritoneal dissemination and invasion. Cancer Res, 67 (2007), pp. 1670–1679
- [14] L. Lo Muzio, A. Farina, C. Rubini, *et al.* Effect of c-Met expression on survival in head and neck squamous cell carcinoma. Tumor Biol, 27 (2006), pp. 115–121
- [15] C. Ponzetto, A. Bardelli, Z. Zhen, et al. A multifunctional docking site mediates signaling and transformation by the hepatocyte growth factor/scatter factor receptor family. Cell, 77 (1994), pp. 261–271

- [16] K.A. Furge, Y.W. Zhang, G.F. Vande Woude. Met receptor tyrosine kinase: enhanced signaling through adapter proteins. Oncogene, 19 (2000), pp. 5582–5589
- [17] G. Pelicci, S. Giordano, Z. Zhen, *et al.* The motogenic and mitogenic responses to HGF are amplified by the Shc adaptor protein. Oncogene, 10 (1995), pp. 1631–1638
- [18] C. Boccaccio, M. Ando, L. Tamagnone, *et al.* Induction of epithelial tubules by growth factor HGF depends on the STAT pathway. Nature, 391 (1998), pp. 285–288
- [19] Y.W. Zhang, L.M. Wang, R. Jove, G.F. Vande Woude. Requirement of Stat3 signaling for HGF/SF-Met mediated tumorigenesis. Oncogene, 21 (2002), pp. 217–226
- [20] E. Sonnenberg, D. Meyer, K.M. Weidner, Birchmeier C. Scatter factor/hepatocyte growth factor and its receptor, the c-met tyrosine kinase, can mediate a signal exchange between mesenchyme and ephitelia during mouse development. J Cell Biol, 123 (1993), pp. 223–235
- [21] E. Gherardi, S. Sandin, M.V. Petoukhov, *et al.* Structural basis of hepatocyte growth factor/scatter factor and MET signaling. Proc Natl Acad Sci U S A, 103 (2006), pp. 4046–4051
- [22] J. Stamos, R.A. Lazarus, X. Yao, D. Kirchhofer, Wiesmann C. Crystal structure of the HGF beta-chain in complex with the sema domain of the Met receptor. EMBO J, 23 (2004), pp. 2325–2335
- [23] G.L. Johnson, Lapadat R. Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. Science, 298 (2002), pp. 1911–1912
- [24] S. Fan, M. Gao, Q. Meng, *et al.* Role of NF-kappaB signaling in hepatocyte growth factor/scatter factor-mediated cell protection. Oncogene, 24 (2005), pp. 1749–1766
- [25] M. Muller, A. Morotti, Ponzetto C. Activation of NF-kappaB is essential for hepatocyte growth factor-mediated proliferation and tubulogenesis. Mol Cell Biol, 22 (2002), pp. 1060–1072
- [26] A. Guo, J. Villen, J. Kornhauser, *et al.* Signaling networks assembled by oncogenic EGFR and c-MET. Proc Natl Acad Sci U S A, 105 (2008), pp. 692–697
- [27] I.S. Long, K. Han, M. Li, *et al.* Met receptor overexpression and oncogenic Ki-ras mutation cooperate to enhance tumorigenicity of colon cancer cells in vivo. Mol Cancer Res, 1 (2003), pp. 393–401
- [28] D. Matsubara, S. Ishikawa, S. Oguni, H. Aburatani, M. Fukayama, T. Niki Molecular predictors of sensitivity to the MET inhibitor PHA665752 in lung carcinoma cells. J Thorac Oncol, 5 (2010), pp. 1317–1324
- [29] Y. Yang, M. Wislez, N. Fujimoto, *et al.* A selective small molecule inhibitor of c-Met, PHA-665752, reverses lung premalignancy induced by mutant K-ras. Mol Cancer Ther, 7 (2008), pp. 952–960
- [30] S. Pennacchietti, P. Michieli, M. Galluzzo, M. Mazzone, S. Giordano, P.M. Comoglio. Hypoxia promotes invasive growth by transcriptional activation of the met protooncogene. Cancer Cell, 3 (2003), pp. 347–361
- [31] P. Conrotto, S. Corso, S. Gamberini, P.M. Comoglio, S. Giordano. Interplay between scatter factor receptors and B plexins controls invasive growth. Oncogene, 23 (2004), pp. 5131–5137
- [32] L. Capparuccia, L. Tamagnone. Semaphorin signaling in cancer cells and in cells of the tumor microenvironment—two sides of a coin. J Cell Sci, 122 (2009), pp. 1723–1736
- [33] S. Kermorgant, P.J. Parker. c-Met signalling: spatio-temporal decisions. Cell Cycle, 4 (2005), pp. 352–355
- [34] S. Kermorgant, D. Zicha, Parker PJ. PKC controls HGF-dependent c-Met traffic, signalling and cell migration. EMBO J, 23 (2004), pp. 3721–3734

- [35] S. Kermorgant, P.J. Parker. Receptor trafficking controls weak signal delivery: a strategy used by c-Met for STAT3 nuclear accumulation. J Cell Biol, 182 (2008), pp. 855–863
- [36] A. Sorkin, M. von Zastrow. Endocytosis and signalling: intertwining molecular networks. Nat Rev Mol Cell Biol, 10 (2009), pp. 609–622
- [37] D.E. Hammond, S. Urbe, G.F. Vande Woude, M.J. Clague. Down-regulation of MET, the receptor for hepatocyte growth factor. Oncogene, 20 (2001), pp. 2761–2770
- [38] B. Foveau, F. Ancot, C. Leroy, *et al.* Down-regulation of the met receptor tyrosine kinase by presenilin-dependent regulated intramembrane proteolysis. Mol Biol Cell, 20 (2009), pp. 2495–2507
- [39] D. Nath, N.J. Williamson, R. Jarvis, G. Murphy. Shedding of c-Met is regulated by crosstalk between a G-protein coupled receptor and the EGF receptor and is mediated by a TIMP-3 sensitive metalloproteinase. J Cell Sci, 114 (2001), pp. 1213–1220
- [40] P. Michieli, M. Mazzone, C. Basilico, et al. Targeting the tumor and its microenvironment by a dual-function decoy Met receptor. Cancer Cell, 6 (2004), pp. 61–73
- [41] F. Bladt, D. Riethmacher, S. Isenmann, A. Aguzzi, C. Birchmeier. Essential role for the c-met receptor in the migration of myogenic precursor cells into the limb bud. Nature, 376 (1995), pp. 768–771
- [42] A. Streit, C.D. Stern, C. Thery, et al. A role for HGF/SF in neural induction and its expression in Hensen's node during gastrulation. Development, 121 (1995), pp. 813–824
- [43] E. Andermarcher, M.A. Surani, E. Gherardi. Co-expression of the HGF/SF and c-met genes during early mouse embryogenesis precedes reciprocal expression in adjacent tissues during organogenesis. Dev Genet, 18 (1996), pp. 254–266
- [44] A. Gentile, C. Trusolino, P.M. Comoglio. The Met tyrosine kinase receptor in development and cancer. Cancer Metastasis Rev, 27 (2008), pp. 85–94
- [45] H. Takayama, W.J. La Rochelle, M. Anver, D.E. Bockman, G. Merlino. Scatter factor/hepatocyte growth factor as a regulator of skeletal muscle and neural crest development. Proc Natl Acad Sci U S A, 93 (1996), pp. 5866–5871
- [46] F. Bussolino, M.F. Di Renzo, M. Ziche, *et al.* Hepatocyte growth factor is a potent angiogenic factor which stimulates endothelial cell motility and growth. J Cell Biol, 119 (1992), pp. 629–641
- [47] C. Birchmeier, E. Gherardi. Developmental roles of HGF/SF and its receptor, the c-Met tyrosine kinase. Trends Cell Biol, 8 (1998), pp. 404–410
- [48] C. Schmidt, F. Bladt, S. Goedecke, *et al.* Scatter factor/hepatocyte growth factor is essential for liver development. Nature, 373 (1995), pp. 699–702
- [49] Y. Uehara, O. Minowa, C. Mori, et al. Placental defect and embryonic lethality in mice lacking hepatocyte growth factor/scatter factor. Nature, 373 (1995), pp. 702–705
- [50] C.G. Huh, V.M. Factor, A. Sanchez, K. Uchida, E.A. Conner, S.S. Thorgeirsson. Hepatocyte growth factor/c-met signaling pathway is required for efficient liver regeneration and repair. Proc Natl Acad Sci USA, 101 (2004), pp. 4477–4482
- [51] C. Boccaccio, P.M. Comoglio. Invasive growth: a MET-driven genetic programme for cancer and stem cells. Nat Rev Cancer, 6 (2006), pp. 637–645
- [52] A. Danilkovitch-Miagkova, B. Zbar. Dysregulation of Met receptor tyrosine kinase activity in invasive tumors. J Clin Invest, 109 (2002), pp. 863–867

- [53] Y. Feng, P.S. Thiagarajan, P.C. Ma. MET signaling: novel targeted inhibition and its clinical development in lung cancer. J Thorac Oncol, 7 (2012), pp. 459–467
- [54] M. Olivero, M. Rizzo, R. Madeddu, *et al.* Overexpression and activation of hepatocyte growth factor/scatter factor in human non-small-cell lung carcinomas. Br J Cancer, 74 (1996), pp. 1862–1868
- [55] G. Maulik, T. Kijima, P.C. Ma, et al. Modulation of the c-Met/hepatocyte growth factor pathway in small cell lung cancer. Clin Cancer Res, 8 (2002), pp. 620–627
- [56] P.C. Ma, M.S. Tretiakova, A.C. MacKinnon, *et al.* Expression and mutational analysis of MET in human solid cancers. Genes Chromosomes Cancer, 47 (2008), pp. 1025–1037
- [57] F. Cappuzzo, A. Marchetti, M. Skokan, *et al.* Increased MET gene copy number negatively affects survival of surgically resected non-small-cell lung cancer patients. J Clin Oncol, 27 (2009), pp. 1667–1674
- [58] K. Okuda, H. Sasaki, H. Yukiue, M. Yano, Y. Fujii. MET gene copy number predicts the prognosis for completely resected non-small cell lung cancer. Cancer Sci, 99 (2008), pp. 2280–2285
- [59] M. Beau-Faller, A.M. Ruppert, A.C. Voegeli, *et al.* MET gene copy number in non-small cell lung cancer: molecular analysis in a targeted tyrosine kinase inhibitor naïve cohort. J Thorac Oncol, 3 (2008), pp. 331–339
- [60] S. Park, Y.L. Choi, C.O. Sung, et al. High MET copy number and MET overexpression: poor outcome in non-small cell lung cancer patients. Histol Histopathol, 27 (2012), pp. 197–207
- [61] E. Ichimura, A. Maeshima, T. Nakajima, T. Nakamura. Expression of c-met/HGF receptor in human non-small cell lung carcinomas in vitro and in vivo and its prognostic significance. Jpn J Cancer Res, 87 (1996), pp. 1063–1069
- [62] I. Takanami, F. Tanana, T. Hashizume, *et al.* Hepatocyte growth factor and c-Met/hepatocyte growth factor receptor in pulmonary adenocarcinomas: an evaluation of their expression as prognostic markers. Oncology, 53 (1996), pp. 392–397
- [63] D. Masuya, C. Huang, D. Liu, *et al.* The tumor-stromal interaction between intratumoral c-Met and stromal hepatocyte growth factor associated with tumor growth and prognosis in non-small-cell lung cancer patients. Br J Cancer, 90 (2004), pp. 1555–1562
- [64] M. Gumustekin, A. Kargi, G. Bulut, et al. HGF/c-Met overexpression, but not Met mutation, correlates with progression of non-small cell lung cancer. Pathol Oncol Res, 18 (2012), pp. 209–218
- [65] G.A. Smolen, R. Sordella, B. Muir, *et al.* Amplification of MET may identify a subset of cancers with extreme sensitivity to the selective tyrosine kinase inhibitor PHA-665752. Proc Natl Acad Sci U S A, 103 (2006), pp. 2316–2321
- [66] C. Baykal, E. Demirtas, A. Al, et al. Comparison of hepatocyte growth factor levels of epithelial ovarian cancer cyst fluids with benign ovarian cysts. Int J Gynecol Cancer, 14 (2004), pp. 152–156
- [67] A.B. Tuck, M. Park, E.E. Sterns, A. Boag, Elliott BE. Coexpression of hepatocyte growth factor and receptor (Met) in human breast carcinoma. Am J Pathol, 148 (1996), pp. 225–232
- [68] S. Koochekpour, M. Jeffers, S. Rulong, *et al.* Met and hepatocyte growth factor/scatter factor expression in human gliomas. Cancer Res, 57 (1997), pp. 5391–5398
- [69] G. Li, H. Schaider, K. Satyamoorthy, Y. Hanakawa, K. Hashimoto, M. Herlyn. Downregulation of E-cadherin and Desmoglein 1 by autocrine hepatocyte growth factor during melanoma development. Oncogene, 20 (2001), pp. 8125–8135
- [70] R. Ferracini, M.F. Di Renzo, K. Scotlandi, *et al.* The Met/HGF receptor is over-expressed in human osteosarcomas and is activated by either a paracrine or an autocrine circuit. Oncogene, 10 (1995), pp. 739–749

- [71] E.M. Boon, R. van der Neut, M. van de Wetering, H. Clevers, S.T. Pals. Wnt signaling regulates expression of the receptor tyrosine kinase met in colorectal cancer. Cancer Res, 62 (2002), pp. 5126–5128
- [72] F. Relaix, D. Rocancourt, A. Mansouri, M. Buckingham. Divergent functions of murine Pax3 and Pax7 in limb muscle development. Genes Dev, 18 (2004), pp. 1088–1105
- [73] R. Kanteti, V. Nallasura, S. Loganathan, *et al.* PAX5 is expressed in small-cell lung cancer and positively regulates c-Met transcription. Lab Invest, 89 (2009), pp. 301–314
- [74] M. Ivan, J.A. Bond, M. Prat, P.M. Comoglio, D. Wynford-Thomas. Activated ras and ret oncogenes induce over-expression of c-met (hepatocyte growth factor receptor) in human thyroid epithelial cells. Oncogene, 14 (1997), pp. 2417–2423
- [75] F. Shirasaki, H.A. Makhluf, C. LeRoy, D.K. Watson, Trojanowska M. Ets transcription factors cooperate with Sp1 to activate the human tenascin-C promoter. Oncogene, 18 (1999), pp. 7755–7764
- [76] G. Gambarotta, C. Boccaccio, S. Giordano, M. Ando, M.C. Stella, P.M. Comoglio. Ets up-regulates MET transcription. Oncogene, 13 (1996), pp. 1911–1917
- [77] S. Krishnaswamy, R. Kanteti, J.S. Duke-Cohan, *et al.* Ethnic differences and functional analysis of MET mutations in lung cancer. Clin Cancer Res, 15 (2009), pp. 5714–5723
- [78] C. Yun, J.H. Lee, H. Park, *et al.* Chemotherapeutic drug, adriamycin, restores the function of p53 protein in hepatitis B virus X (HBx) protein-expressing liver cells. Oncogene, 19 (2000), pp. 5163–5172
- [79] D. Zaffaroni, M. Spinola, A. Galvan, et al. Met proto-oncogene juxtamembrane rare variations in mouse and humans: differential effects of Arg and Cys alleles on mouse lung tumorigenesis. Oncogene, 24 (2005), pp. 1084–1090
- [80] R. Jagadeeswaran, H. Surawska, S. Krishnaswamy, *et al.* Paxillin is a target for somatic mutations in lung cancer: implications for cell growth and invasion. Cancer Res, 68 (2008), pp. 132–142
- [81] P.C. Ma, T. Kijima, G. Maulik, *et al.* c-MET mutational analysis in small cell lung cancer: novel juxtamembrane domain mutations regulating cytoskeletal functions. Cancer Res, 63 (2003), pp. 6272–6281
- [82] P. Peschard, T.M. Fournier, L. Lamorte, *et al.* Mutation of the c-Cbl TKB domain binding site on the Met receptor tyrosine kinase converts it into a transforming protein. Mol Cell, 8 (2001), pp. 995–1004
- [83] P.C. Ma, R. Jagadeeswaran, S. Jagadeesh, *et al.* Functional expression and mutations of c-Met and its therapeutic inhibition with SU11274 and small interfering RNA in non-small cell lung cancer. Cancer Res, 65 (2005), pp. 1479–1488
- [84] L. Schmidt, F.M. Duh, F. Chen, et al. Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. Nat Genet, 16 (1997), pp. 68–73
- [85] G. Cortesina, P. Rosso, P. Mola, *et al.* Molecular markers study in pTNM of squamous carcinoma of the head and neck. Acta Otorhinolaryngol Ital, 20 (2000), pp. 380–382
- [86] C. Walz, M. Sattler. Novel targeted therapies to overcome imatinib mesylate resistance in chronic myeloid leukemia (CML). Crit Rev Oncol Hematol, 57 (2006), pp. 145–164
- [87] J.A. Engelman, P.A. Janne. Mechanisms of acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small cell lung cancer. Clin Cancer Res, 14 (2008), pp. 2895–2899
- [88] J.A. Engelman, K. Zejnullahu, T. Mitsudomi, *et al.* MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. Science, 316 (2007), pp. 1039–1043
- [89] J. Bean, C. Brennan, J.Y. Shih, *et al.* MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. Proc Natl Acad Sci U S A, 104 (2007), pp. 20932–20937

- [90] C.L. Arteaga. HER3 and mutant EGFR meet MET. Nat Med, 13 (2007), pp. 675-677
- [91] M.E. Arcila, G.R. Oxnard, K. Nafa, *et al.* Rebiopsy of lung cancer patients with acquired resistance to EGFR inhibitors and enhanced detection of the T790M mutation using a locked nucleic acid-based assay. Clin Cancer Res, 17 (2011), pp. 1169–1180
- [92] L.V. Sequist, B.A. Waltman, D. Dias-Santagata, et al. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. Sci Transl Med, 3 (75) (2011), p. ra26
- [93] A.B. Turke, K. Zejnullahu, Y.L. Wu, *et al.* Preexistence and clonal selection of MET amplification in EGFR mutant NSCLC. Cancer Cell, 17 (2010), pp. 77–88
- [94] F. Cappuzzo, P.A. Janne, M. Skokan, *et al.* MET increased gene copy number and primary resistance to gefitinib therapy in non-small-cell lung cancer patients. Ann Oncol, 20 (2009), pp. 298–304
- [95] L. Xu, E. Kikuchi, C. Xu, et al. Combined EGFR/MET or EGFR/HSP90 inhibition is effective in the treatment of lung cancers codriven by mutant EGFR containing T790M and MET. Cancer Res, 72 (2012), pp. 3302–3311
- [96] W. Wang, Q. Li, S. Takeuchi, et al. Met kinase inhibitor E7050 reverses three different mechanisms of hepatocyte growth factor-induced tyrosine kinase inhibitor resistance in EGFR mutant lung cancer. Clin Cancer Res, 18 (2012), pp. 1663–1671
- [97] S. Yano, T. Yamada, S. Takeuchi, *et al.* Hepatocyte growth factor expression in EGFR mutant lung cancer with intrinsic and acquired resistance to tyrosine kinase inhibitors in a Japanese cohort. J Thorac Oncol, 6 (2011), pp. 2011–2017
- [98] G. Daniele, M. Ranson, M. Blanco-Codesido, *et al.* Phase I dose-finding study of golvatinib (E7050), a c-Met and Eph receptor targeted multi-kinase inhibitor, administered orally QD to patients with advanced solid tumors. J Clin Oncol, 30 (2012) [suppl; abstr 3030]
- [99] R.H. Alvarez, H.M. Kantarjian, J.E. Cortes. The role of Src in solid and hematologic malignancies: development of new-generation Src inhibitors. Cancer, 107 (2006), pp. 1918–1929
- [100] J. Qi, M.A. McTigue, A. Rogers, *et al.* Multiple mutations and bypass mechanisms can contribute to development of acquired resistance to MET inhibitors. Cancer Res, 71 (2011), pp. 1081–1091
- [101] J.R. Diamond, R. Salgia, M. Varella-Garcia, *et al.* Initial clinical sensitivity and acquired resistance to MET inhibition in MET-mutated papillary renal cell carcinoma. J Clin Oncol, 31 (2013), pp. e254–e258
- [102] H.T. Jun, J. Sun, K. Rex, *et al.* AMG 102, a fully human anti-hepatocyte growth factor/scatter factor neutralizing antibody, enhances the efficacy of temozolomide or docetaxel in U-87 MG cells and xenografts. Clin Cancer Res, 13 (2007), pp. 6735–6742
- [103] M.S. Gordon, C.S. Sweeney, D.S. Mendelson, *et al.* Safety, pharmacokinetics, and pharmacodynamics of AMG 102, a fully human hepatocyte growth factor neutralizing monoclonal antibody, in a first-in-human study of patients with advanced solid tumors. Clin Cancer Res, 16 (2010), pp. 699–710
- [104] P.J. Rosen, C.J. Sweeney, D.J. Park, *et al.* A phase Ib study of AMG 102 in combination with bevacizumab or motesanib in patients with advanced solid tumors. Clin Cancer Res, 16 (2010), pp. 2677–2687
- [105] K.S. Oliner, R. Tang, A. Anderson, *et al.* Evaluation of MET pathway biomarkers in a phase II study of rilotumumab (R AMG 102) or placebo (P) in combination with epirubicin, cisplatin, and capecitabine (ECX) in patients (pts) with locally advanced or metastatic gastric (G) or esophagogastric junction (EGJ) cancer. J Clin Oncol, 30 (2012) [suppl; abstr 4005]

- [106] A. Patnaik, G.J. Weiss, K. Papadopoulos, *et al.* Phase I study of SCH900105 (SC), an anti-hepatocyte growth factor (HGF) monoclonal antibody (MAb), as a single agent and in combination with erlotinib (E) in patients (pts) with advanced solid tumors. J Clin Oncol, 28 (2010) [suppl; abstr 2525]
- [107] E. Tan, K. Park, W.T. Lim, *et al.* Phase Ib study of ficiatuzumab (formerly AV-299), an anti-hepatocyte growth factor (HGF) monoclonal antibody (MAb) in combination with gefitinib (G) in Asian patients (pts) with NSCLC. J Clin Oncol, 29 (2011) [suppl; abstr 7571]
- [108] T. Martens, N.O. Schmidt, C. Eckerich, *et al.* A novel one-armed anti-c-Met antibody inhibits glioblastoma growth in vivo. Clin Cancer Res, 12 (2006), pp. 6144–6152
- [109] H. Jin, R. Yang, Z. Zheng, et al. MetMab, the one-armed 5D5 anti-c-Met antibody, inhibits orthotopic pancreatic tumor growth and improves survival. Cancer Res, 68 (2008), pp. 4360–4368
- [110] R.A. Moss, P. Patel, J. Bothos. Complete results from phase I dose escalation study of MetMab, a monovalent antagonist antibody to the receptor MET, doses as single agent and in combination with bevacizumab in patients with advanced solid malignancies. Ann Oncol, 21 (viii 165. Suppl. 8) (2010) abstract 504P
- [111] D.R. Spigel, T.J. Ervin, R. Ramlau, *et al.* Final efficacy results from OAM4558g, a randomized phase II study evaluating MetMAb or placebo in combination with erlotinib in advanced NSCLC. J Clin Oncol, 29 (2011) [suppl: Abstr 7505]
- [112] D.V. Catenacci, L. Henderson, S.Y. Xiao, *et al.* Durable complete response of metastatic gastric cancer with anti-Met therapy followed by resistance at recurrence. Cancer Discov, 1 (2011), pp. 573–579
- [113] D.R. Spigel, M.J. Edelman, T. Mok, *et al.* Treatment rationale study design for the MetLung trial: a randomized, double-blind phase III study of onartuzumab (MetMab) in combination with erlotinib versus erlotinib alone in patients who have received standard chemotherapy for stage IIIB or IV Met-positive non-small-cell lung cancer. Clin Lung Cancer, 13 (2012), pp. 500–504
- [114] G.R. Blumenschein Jr., G.B. Mills, A.M. Gonzales-Angulo. Targeting the hepatocyte growth factor cMet axis in cancer therapy. J Clin Oncol, 30 (2012), pp. 3287–3290
- [115] F. Bladt, B. Faden, M. Friese-Hamim, et al. EMD 1214063 and EMD 1204831 constitute a new class of potent and highly selective c-Met inhibitors. Clin Cancer Res, 19 (2013), pp. 2941–2951
- [116] N. Munshi, S. Jeay, Y. Li, *et al.* ARQ 197, a novel and selective inhibitor of the human c-Met receptor tyrosine kinase with antitumor activity. Mol Cancer Ther, 9 (2010), pp. 1544–1550
- [117] C. Basilico, S. Pennacchietti, E. Vigna, et al. Tivantinib (ARQ197) displays cytotoxic activity that is independent of its ability to bind MET. Clin Cancer Res, 19 (2013), pp. 2381–2390
- [118] R. Katayama, A. Aoyama, T. Yamori, *et al.* Cytotoxic activity of tivantinib (ARQ 197) is not due solely to c-MET inhibition. Clin Cancer Res, 73 (2013), pp. 3087–3090
- [119] J.W. Goldman, I. Laux, F. Chai, *et al.* Phase 1 dose-escalation trial evaluating the combination of the selective MET inhibitor tivantinib (ARQ 197) plus erlotinib. Cancer, 118 (2012), pp. 5903–5910
- [120] L.S. Rosen, N. Senzer, T. Mekhail, *et al.* A phase I dose-escalation study of tivantinib (ARQ 197) in adult patients with metastatic solid tumors. Clin Cancer Res, 17 (2011), pp. 7754–7760
- [121] T.A. Yap, D. Olmos, A.T. Brunetto, *et al.* Phase I trial of a selective c-MET inhibitor ARQ 197 incorporating proof of mechanism pharmacodynamic studies. J Clin Oncol, 29 (2011), pp. 1271–1279
- [122] L.V. Sequist, J. von Pawel, E.G. Garmey, et al. Randomized phase II study of erlotinib plus tivantinib versus erlotinib plus placebo in previously treated non-small-cell lung cancer. J Clin Oncol, 29 (2011), pp. 3307–3315

- [123] G.V. Scagliotti, S. Novello, R. Ramlau, *et al.* Results of the phase 3 MARQUEE study: MET Inhibitor Tivantinib (ARQ 197) Plus Erlotinib vs Erlotinib Plus Placebo in NSCLC. Proceedings of 17th ECCO–38th ESMO–32nd ESTRO European cancer congress: 27 September–01 October, Amsterdam (2013) [abstract E17-1821]
- [124] T. Nishina, T. Hirashima, K. Sugio, *et al.* The effect of CYP2C19 polymorphism on the tolerability of ARQ 197, results from phase I trial in Japanese patients with metastatic solid tumors. J Clin Oncol, 29 (2011) [suppl: abstr 2516]
- [125] F.M. Yakes, J. Chen, J. Tan, et al. Cabozantinib (XL184), a novel MET and VEGFR2 inhibitor, simultaneously suppresses metastasis, angiogenesis and tumor growth. Mol Cancer Ther, 10 (2011), pp. 2298–2308
- [126] H.A. Wakelee, S.N. Gettinger, J.A. Engelman, et al. A phase lb/II study of XL184 (BMS 907351) with and without erlotinib (E) in patients (pts) with non-small cell lung cancer (NSCLC). J Clin Oncol, 28 (2010) [15S: abstr 3017]
- [127] M.S. Gordon, N.J. Vogelzang, P. Schoffski, *et al.* Activity of cabozantinib (XL184) in soft tissue and bone: results of a phase II randomized discontinuation trial (RDT) in patients (pts) with advanced solid tumors. J Clin Oncol, 29 (2011) [suppl: abstr 3010]
- [128] R.J. Buckanovich, R. Berger, A. Sella, *et al.* Activity of cabozantinib (XL184) in advanced ovarian cancer patients (pts): results from a phase II randomized discontinuation trial (RDT). J Clin Oncol, 29 (2011) [suppl: abstr 5008]
- [129] M. Hussain, M.R. Smith, C. Sweeney, *et al.* Cabozantinib (XL184) in metastatic castration-resistant prostate cancer (mCRPC): results from a phase II randomized discontinuation trial. J Clin Oncol, 29 (2011) [suppl: abstr 4516]
- [130] F. Qian, S. Engst, K. Yamaguchi, *et al.* Inhibition of tumor cell growth, invasion and metastasis by EXEL-2880 (XL880, GSK1363089), a novel inhibitor of HGF and VEGF receptor tyrosine kinases. Cancer Res, 69 (2009), pp. 8009–8016
- [131] J.P. Eder, G.I. Shapiro, L.J. Appleman, *et al.* A phase I study of foretinib, a multi target inhibitor of c-MET and vascular endothelial growth factor receptor 2. Clin Cancer Res, 16 (2010), pp. 3507–3510
- [132] T.K. Choueiri, U. Vaishampayan, J.E. Rosenberg, *et al.* Phase II and biomarker study of the dual MET/VEGFR inhibitor foretinib in patients with papillary renal cell carcinoma. J Clin Oncol, 31 (2013), pp. 181–186
- [133] H.Y. Zou, Q. Li, J.H. Lee, *et al.* An orally available small molecule inhibitor of c-Met (PF-02341066) exhibits cytoreductive antitumor efficacy through antiproliferative and antiangiogenic mechanisms. Cancer Res, 67 (2007), pp. 4408–4417
- [134] J.J. Cui, M. Tran-Dubé, H. Shen, et al. Structure based drug design of crizotinib (PF-02341066), a potent and selective dual inhibitor of mesenchymal-epithelial transition factor (c-Met) kinase and anaplastic lymphoma kinase (ALK)
- [135] E.L. Kwak, D.R. Camidge, J. Clark, *et al.* Clinical activity observed in a phase I dose escalation trial of an oral cmet and ALK inhibitor. J Clin Oncol, 27 (2009), p. 3509 [suppl: abstr]
- [136] S.H. Ou, E.L. Kwak, C. Siwak-Tapp, *et al.* Activity of crizotinib (PF02341066), a dual mesenchymal-epithelial transition (MET) and anaplastic lymphoma kinase (ALK) inhibitor, in a non-small cell lung cancer patient with de novo MET amplification. J Thorac Oncol, 6 (2011), pp. 942–946
- [137] J.K. Lennerz, E.L. Kwak, A. Ackerman, et al. MET amplification identifies a small and aggressive subgroup of esophagogastric adenocarcinoma with evidence of responsiveness to crizotinib. J Clin Oncol, 29 (2011), pp. 4803–4810
- [138] A. Chi, E. Kwak, J. Clark, *et al.* Clinical improvement and rapid radiographic regression induced by a MET inhibitor in a patient with MET amplified glioblastoma. J Clin Oncol, 29 (2011), p. 2072 [suppl: abstr]
- [139] J. Tanizaki, I. Okamoto, K. Okamoto, et al. MET tyrosine kinase inhibitor crizotinib (PF-02341066) shows differential antitumor effects in non-small cell lung cancer according to MET alterations. J Thorac Oncol, 6 (2011), pp. 1624–1631

[140] Y. Zhang, K. Farenholtz, Y. Yang, *et al.* Hepatocyte growth factor sensitizes brain tumors to c-Met kinase inhibition. Clin Cancer Res, 19 (2013), pp. 1433–1440

[141] J.R. Infante, T. Rugg, M. Gordon, *et al.* Unexpected renal toxicity associated with SGX523, a small molecule inhibitor of MET. Invest New Drugs, 31 (2013), pp. 363–369

[142] D. Diaz, K.A. Ford, D.P. Hartley, *et al.* Pharmacokinetic drivers of toxicity for basic molecules: Strategy to lower pKa results in decreased tissue exposure and toxicity for a small molecule Met inhibitor. Toxicol Appl Pharmacol, 266 (2013), pp. 86–94

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