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Anisakis spp. larvae in different kinds of ready to eat products made of anchovies (Engraulis encrasicolus) sold in Italian supermarkets.

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(Article begins on next page)

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Abstract: In this study the occurrence of visible anisakid larvae in semi-preserved anchovy products sold on the Italian market was investigated. Totally, 107 ready to eat products (33 salted-ripened, 49 in oil and 25 marinated) were sampled. Each sample was digested, then the digested material was observed under natural and UV light. Parasites were counted, collected and microscopically identified to genus level. A representative subset was molecularly identified using the cox2 gene. At least one visible Anisakis sp. larva was found in 54.2% of the total 107 products analysed. Totally 1283 dead larvae were collected. Anisakis sp. larvae were found in all the 33 salted products and 1139 (88.8%) larvae were collected, with a range of 1-105 parasites per product. Larval density per gram was 0.13. Anisakis sp. larvae were found in 49.0% of the products in oil and 143 (11.1%) larvae were isolated, with a range of 0-28 and a density of 0.03. Only 1 larva was found in the 25 marinated products (4.0%), the density was 0.00. A highly significant difference between all the product categories in respect of number of larvae per product, frequency of products contaminated by at least one larva and larval density per gram was found. Within the subset of larvae molecularly analysed (n=122), 92 larvae (75.4%) were identified as A. pegreffii and 30 (24.6%) as A. simplex. This study showed that semipreserved anchovy products heavily contaminated with Anisakis spp. larvae reach the market. Beyond the negligible risk for anisakidosis, the presence of dead visible parasites may cause immediate rejection in consumers. In addition, the potential risk related to allergic reactions in sensitized individuals needs to be further assessed. In order to avoid commercialization of obviously contaminated products , fresh anchovies' batches intended for the production of such products should be accurately selected by the processing industry applying inspection methods.

Dear Editor,

Please find enclosed the manuscript entitled "*Anisakis* **spp. larvae in different kinds of ready to eat products made of anchovies: a defect or a hazard?**" to be considered for publication in the International Journal of Food Microbiology.

The European anchovy (*Engraulis encrasicolus*) is one of the most important fish resources of Mediterranean countries, where it is commonly used to produce traditional semi-preserved products, such as salted-ripened, in oil, marinated/pickled anchovies. Among the most important biohazards related to the consumption of raw anchovies is the presence of viable zoonotic nematode larvae belonging to the *Anisakis* genus. The parasitological risk associated to the presence of viable larvae in semi-processed seafood products can be prevented applying a freezing treatment or an appropriate brining or pickling process for a sufficient time. Nevertheless, the presence of dead visible parasites in processed products represents a defect that alters the overall quality, causes immediate consumers' rejection and may damage the reputation of the brand. In addition, although it is generally believed that sensitization with live *Anisakis* spp. larvae is required prior to the development of a clinical allergic responses, the allergenic potential of dead larvae is still debated.

Taking into account the increasing market request of ready to eat semi-preserved anchovies, their high prices and the scarcity of data on the presence of anisakid parasites in these preparations, the aim of this study was to assess the presence of visible anisakid larvae in different commercial categories of the most appreciated types of these semi-preserves on the Italian market.

Totally, 107 ready to eat products (33 salted-ripened, 49 in oil and 25 marinated) were sampled and separately submitted to artificial digestion. Parasites were counted, collected and microscopically identified to genus level. A subset was molecularly identified using the *cox2* gene.

Of the total 107 products analysed, 54.2% were positive for the presence of at least one visible *Anisakis* sp. larva and a total of 1283 larvae were collected. All the parasites found were dead. All the 33 salted products were positive and 1139 (88.8%) *Anisakis* spp. larvae were collected, with a range of 1-105 parasites per product. Among products in oil, 48.9% were positive with 143 (11.1%) *Anisakis* spp. larvae isolated and a range of 0-28. Only 1 out of the 25 marinated products (4%) was positive, with the presence of one larva. Within the larvae subset (n=122), 92 larvae (75.4%) were molecularly identified as *A. pegreffii* and 30 (24.6%) as *A. simplex*.

The present results showed that semi preserved anchovy products heavily contaminated with *Anisakis* spp. larvae can reach the market. In particular, the level of contamination was different

depending on the products typology, being linked to the processing procedure and to the preliminary preparation of the fish, especially depending on the removal of the viscera. Beyond the negligible risk for anisakidosis, the presence of dead parasites may cause immediate rejection in consumers. In addition, the risk related to allergic reactions in sensitized individuals is still an open issue.

The manuscript has not been published elsewhere nor is it being considered for publication elsewhere. All authors have approved this manuscript, agree to the order in which their names are listed, declare that no conflict of interests exists and disclose any commercial affiliation.

Yours sincerely, Andrea Armani

Dear editor,

thank you very much for considering our manuscript for publication in International Journal of Food Microbiology.

Please find below our answers to the reviewers' comments. We would like to thank both reviewers for their constructive suggestions. We are sorry they were not completely satisfied by the first revision. Some of the issues were probably due to the fact that the reviewers' comments were partly contrasting.

We hope that we have now succeeded in making all the requested modifications and clarifications.

Reviewer #1:

Line 231. Sorry but this is still unclear to me. Does it mean that the MA threshold is calculated as 3/N where N is 10% of the batch, 10% of the samples collected for analysis or a maximum of 3 larvae in all the samples within 10% of the sample size? It is unclear what is the value of the MA threshold and how the MA is calculated at individual level to define the samples exceeding this threshold.

The mean abundance threshold was calculated on the basis of the Liguria Region Circular n. 1 of 1997. This regulation states that: "if when opening the coelomic cavity numerous viable larvae appear, giving a repellent aspect to the product, the batch is withdrawn from the market; ii) if the number of visible parasites is higher than 3 per anchovy in the 10 % of the examined specimens, or the number of parasitized specimens is higher than 10 % of the total, the batch should be submitted to decontamination by means of freezing, according on the existing law; iii) if the number of larvae is \leq 3 per anchovy in maximum 10 % of the examined specimens, the batch is intended to free consumption.

As regards the number of examined specimens, we referred to the Lombardy Region circular (Circular Letter VS8/C790/94) which states that: "knowing the total weight of the fish lot, it is possible to calculate the total number of specimens and then, by means of conversion rates and using an appropriate table, the number of subjects to be examined in each case. In the case of fish species caught in large batches (>600 specimens, such as anchovies), <u>the number of subjects to collect is, at least, 29</u>".

Therefore:

Number of examined specimens = 29

10% of 29 specimens = 2,9

3 (maximum number of tolerated parasites) * 2,9 = 8,7 maximum (theoretical) number of parasites in 29 specimens \rightarrow 9 maximum (real) number of parasites in 29 specimens

9/29 = 0.3 mean abundance threshold

It derives that a MA of 0.3 corresponds to the threshold that allows to divide the batches in "non-marketable" (MA>0.3) or "marketable" (MA≤0.3).

The combination of the described sampling plan and mean abundance threshold is the most applied approach in Italy (D'Amico et al., 2014 Food research international, 64, 348-362).

The proposed MA threshold was first used in a work comparing the performance of three different techniques (visual inspection, UV press method and digestion) (Guardone et al. 2016 Food Analytical Methods 9.5, 1418-1427) and subsequently in a work aimed at assessing the reliability of the digestion of a subsample of 150 g (\pm 30 g) of viscera and adjacent muscles, randomly collected from 29 specimens, in estimating the marketability of fresh anchovies' batch.

We added more details also in the text (lines 222-227). We really hope it is clear now. If you think it is necessary, we can add further explanations.

Finally, we would like to point out that, although we have used the MA threshold also in this publication, it was only applied to a part of the samples (those composed by whole specimens for which the MA could be calculated). The paper focuses on the number of larvae per product and on the contamination (presence of at least one larva). Following the suggestion of reviewer 2 we have also introduced another epidemiological index, the larval density per gram.

Discussion

In the discussion section, aspects related to batch selection and practical meaning of this study for food

industry has been removed from the previous version. I really think it would be important to discuss this and demonstrate how this work could be used in practice to inform decision making in fishery industry in long term.

Except for the above aspect of the MA, which I am sure will be clarified, the manuscript is well written but as presented, it remains a descriptive study. Considering the level of the journal, including some strong final remarks on the practical utility of the findings, would noticeably increase the quality of the work.

Therefore, I propose again what I wrote in the first revision:

"what 'a precautionary approach' would mean in practice for decision makers and the food industry? is it related to the batch selection reported in the conclusion? what options the food industry would have in practice to select the batches (e.g. fishing area?)? furthermore, from the results it appears that many samples are above the threshold in terms of MA. In the light of your results, it would be good to discuss the role of this threshold on the economic impact on the food industry if this value is used, as suggested, for batch selection."

We thank the reviewer for the suggestion. These points have been added to the discussion (lines 432-451).

Reviewer #2:

1. I think the title should be considered for revision; the title 'Anisakis spp~ a defect or a hazard ?' sounds like the authors investigated whether the Anisakis species in anchovy products are hazardous to humans. But the authors assessed the prevalence of anisakid larvae in different kinds of anchovy products, as mentioned in the end of the INTRODUCTION. Of course, the authors can mention the dead larvae have the potential risks of allergic reactions to humans in DISCUSSION. But this is not the main focus (aim) of this study (if the authors want to keep the aim of this study as described in the INTRODUCTION). So if the authors also want to emphasize the potential risk of the larvae (or want to keep the title), I suggest that they should at least raise enough evidence that dead Anisakis larvae are allergic to humans, particularly dead Anisakis larvae in ANCHOVIES can cause allergic reactions to humans, not in pink salmon.

The title has been changed, we preferred to maintain the aim as it is.

2. Please include the scientific name of anchovy in the title. **done**

3. Please make all percentage data round off to one decimal place (e.g., 88.8%, not 88.98%; 4.0%, not 4%) throughout the whole MS including Tables. **done**

4. 'positivity' and 'number of larvae' do not seem to be the terms generally used in parasitology. The authors should refer Bush et al. (1997)'s reference and select appropriate terms which fit into the definitions.

We are aware of the terminology defined in Bush et al., 1997. However, most of these terms refer to animal hosts, while in this study we have analysed seafood products which were composed of many individual hosts. This is the reason why we used the Mean Abundance, in the case of products composed of whole anchovies for which the number of individuals was countable. Now, we also added the larval density per gram as a further epidemiological index.

We have removed the term "positivity" and, where possible, we have replaced it with the term "contaminated", in accordance with the EU regulations, EFSA (<u>https://www.efsa.europa.eu/en/topics/topic/parasites-food</u>) and with previous works (Audicana & Kennedy 2008 *Clinical microbiology reviews*, *21*(2), 360-379; Fæste et al., 2015 *Food Analytical Methods*, *8*(6), 1390-1402; Llarena-Reino et al., 2012 *Food Control*, 23(1), 54-58).

The term "number of larvae" was kept since it is a measure and not a parasitological index and therefor it cannot be replaced. Besides, it has been used in similar recent studies:

- Cipriani, P., Acerra, V., Bellisario, B., Sbaraglia, G. L., Cheleschi, R., Nascetti, G., & Mattiucci, S. (2016). Larval migration of the zoonotic parasite Anisakis pegreffii (Nematoda: Anisakidae) in European anchovy, Engraulis encrasicolus: Implications to seafood safety. Food Control, 59, 148-157.
- Pierce, G. J., Bao, M., MacKenzie, K., Dunser, A., Giulietti, L., Cipriani, P., ... & Hastie, L.
 C. (2017). Ascaridoid nematode infection in haddock (*Melanogrammus aeglefinus*) and whiting (*Merlangius merlangus*) in Northeast Atlantic waters. *Fisheries Research*, in press

5. Keywords in the MS should be selected for being detected by DB (e.g., PUBMED) as much as possible. I think the keywords in this MS are not useful for being detected in PUBMED or other DBs. I suggest that the keywords should be carefully selected again. **done**

6. Line 114~115: it has been supposed that also ingestion \sim -> it has been also suspected that ingestion \sim **done**

7. Line $121 \sim$: Taking into ~EU. the high prices of ~ -> Taking into ~ EU and the high prices of ~ .

It is not possible to change the sentence, as we cannot substitute the comma with "and" because there is another element listed and there would be two "and" in the sentence

8. Line 144 : ~ University of Pisa, Department ~ -> Department of ~ University of Pisa done

9. Line 149: ~ their number -> their number of what? The sentence has been modified

10. Line 151: \sim rinsed in a glass beaker. -> rinsed with what ? with tap water, the indication has been added

11. Line 152: The oil was carefully removed also ~ -> The oil was also carefully removed **done**

12. Line 154: With the aim to test the recovery \sim -> To test the recovery \sim **done**

13. Line 159~162 : Considering that ~ per time -> I do not understand what the authors want to describe. Please revise this sentence. **Done**

14. Line 196~197: by the Experimental ~ (Turin, Italy) -> Delete done

15. Line 252: trials ->What trials ? Those mentioned at line 156. However, the sentence has benne modified to make it more clear.

16. Line 261~264: All the parasites ~dead. In fact, even though ~and might be ~ treatment -> All the parasites ~dead. Although In fact, even though the emission of fluorescence are known to not always discriminate between live and dead larvae, might be related to ~ by the treatment spontaneous and stimulated movements of the larvae were absent in this study. **We found very hard to understand this point. However, we have modified the sentence hoping to have correctly interpreted the meaning.**

17. Among those ~ analyzed (n=122) 92 (75.4%) -> Among these ~ analyzed larvae (n=122), 92(75.4%)~ we have modified the sentence trying to clarify it. However, we cannot use "these", instead of "those", because not all the larvae were moleculary analysed. 18. Line 296~297: see section 3.2 -> delete **done**

19. Line 299: ~A. simplex (22.6%) -> A. simplex (22.6%) (Table 2). Done

20. Line 305: The MA \sim 3.92 -> Delete. The sentence was wrong and did not make sense. However, this sentence cannot be deleted because this information is not present elsewhere in the text. Therefore, the sentence was corrected.

21. Line 313: corresponding to 11.1% of the $\sim ->$ corresponding to 11.1% (00/00) of the \sim **Done**

22. Line 323: ~ for 15 product, in fact although other ~ -> ~ for 15 products. in fact although Other 3 products ~ **done**

23. Line 351: Analogously -> Similarly **done**

24. Line 353: As known, most anisakid larvae are located ~ -> As known, most anisakid larvae are known to be located ~? The sentence has been modified assuming you wanted to move "known" from the beginning to the middle of the sentence. Otherwise it would be repeated.

25. Line 357: ~ after the capture especially ~ -> ~ after the capture, especially ~ done

26. Line 364: \sim were positive. -> positive for what? For the presence of at least one larva. The sentence has been modified.

27. Line 389: ~ inactivates Anisakidae larvae -> inactivates anisakid larvae. done

28. Line 392~395: In addition, salted-ripened anchovies ~ for E. encrasicolus -> This sentence dose not make sense; The scientific name of European anchovy is E. encrasicolus. Delete 'for E. encrasicolus'. **Done**

29. Line 417~418: ~ live larvae that can actively ~ the external surface. -> ~ live larvae which can actively move and become evident also on the external surface. **done**

30. Line 419: However, also the presence \sim -> However, also the presence \sim we don't understand this point.

31. Line 467~476: I think this paragraph is not helpful for discussion and better to be deleted. As mentioned in the INTRODUCTION, the aim of this study is that the assessment of the presence of visible anisakid larvae in different commercial categories of products. But this paragraph contains how the consumers and authorities should react with the presence of the larvae in fish products. And what is FBO? I suggest the authors should keep the MS clear, concise, readable, not too wordy.

Conclusions have been modified and shortened. FBO stands for Food Business Operators, as stated at line 96.

32. Line 478~479: I think the authors should make it clear what the aim of this study is. The authors described that 'The present work highlighted how semi-preserved anchovy products heavily ~ can reach the market'. I think the authors should discuss the distribution structure, marketing system regarding the fisheries products, to highlight the issue written in this sentence. But there is no mention regarding 'how ~ can reach the market' in this MS, and as in the INTRODUCTION, the authors assessed the prevalence of anisakid larvae in different kinds of anchovy products. Furthermore, the title also includes the debate about allergic issue of dead larvae. All of these make the readers confused and the MS difficult to understand.

We think the word "how" was probably misleading. We did not intend to describe the products' distribution on the market but to point out that highly contaminated products are already sold in Italian supermarkets. We have change the sentence hoping to clarify our idea and to better agree with the aim of our work.

33. I strongly suggest that the authors should replace the references which cannot be read nor accessed by international readers. If the references cannot be accessed or cannot be obtained by the readers who want to have a look, those references should not be referred. Unfortunately, there are so many references which I cannot read nor access in this MS.

The references have been revised. All the documents are now accessible to international readers.

- Different kinds of anchovy ready to eat products were analysed by digestion
- 54.2% of the products were positive for at least one visible *Anisakis* spp. larva
- A total of 1283 dead larvae were collected
- The product category influenced the number of larvae and positivity rate
- Salted products were found to be the most contaminated (positivity 100%)

1	Anisakis spp. larvae in different kinds of ready to eat products made of anchovies
2	(Engraulis encrasicolus) sold in Italian supermarkets: a defect or a hazard?
3	
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The aim of In this study was to assess the presence occurrence of visible anisakid larvae in 27 semi-preserved anchovy products sold on the Italian market was investigated. Totally, 107 28 29 ready to eat products (33 salted-ripened, 49 in oil and 25 marinated) were sampled. Each sample was digested, then the digested material was observed under natural and UV light. 30 Parasites were counted, collected and microscopically identified to genus level. A 31 representative subset was molecularly identified using the cox2 gene. At least one visible 32 Anisakis sp. larva was found in 54.2% of the total 107 products analysed. Of the total 107 33 products analysed, 54.2% were positive for the presence of at least one visible Anisakis sp. 34 larva and a total of and tTotally 1283 dead larvae were collected. All Anisakis sp. larvae were 35 found in all the 33 salted products were positive and 1139 (88.8%) Anisakis sp. larvae were 36 37 collected, with a range of 1-105 parasites per product. Larval density per gram was 0.13. Anisakis sp. larvae were found in 49.0% of the products in oil Among products in oil, 38 49.08.98% were positive and 143 (11.1%) Anisakis spp. larvae were isolated, with a range of 39 0-28 and a density of 0.03. Only 1 larva was found in-out of the 25 marinated products 40 (4.0%), the density was 0.00 was positive, with the presence of one larva. A highly 41 significant difference between all the product categories in respect of both-number of larvae 42 per product, frequency of products contaminated by at least one larva and larval density per 43 gram was found and positivity was found. Within the subset of larvae molecularly identified 44 45 analysed (n=122), 92 larvae (75.4%) were identified as A. pegreffii and 30 (24.6%) as A. *simplex*. This study highlighted how showed that semi-preserved anchovy products heavily 46 contaminated with Anisakis spp. larvae can reach the market. Beyond the negligible risk for 47 48 anisakidosis, the presence of dead visible parasites may cause immediate rejection in consumers. In addition, the potential risk related to allergic reactions in sensitized individuals 49 50 needs to be further assessed. In order to avoid commercialization of obviously contaminated 51 products corrective measures on the final products, fresh anchovies' batches intended for the
52 production of such products should be accurately selected by the processing industry
53 applying inspection methods.

54 Keywords

Salted ripened <u>Processed seafood products</u>, anchovies, anchovies in oil, marinated
anchovies, visible parasite, <u>anisakid dead larvae</u>, product quality, defect, <u>risk assessment</u>
Anisakid larvae, anchovies, *Engraulis encrasicolus*, artificial digestion, contamination,
semi-preserved seafood products, Italy

59

60 **1. Introduction**

The European anchovy (*Engraulis encrasicolus*) is an economically important fish species particularly appreciated in Mediterranean countries, where it is commonly used to produce traditional salted-ripened, in oil and marinated/pickled products (Anastasio et al., 2016; Felix et al., 2016; Triqui and Reineccius, 1995).

In the presence of salt, anchovies undergo physicochemical modifications giving origin to 65 a product called "ripened" or "matured" (Codex Alimentarius, 2012). Usually, salting-66 ripening involves a preliminary operation of brining, where the whole fish is immersed in 67 saturated brine. Following this, anchovies are beheaded and gutted, placed in barrels, 68 69 alternating layers of fish and salt, and pressed (Czerner et al., 2011; Felix et al., 2016). In 70 some cases, fish are beheaded and gutted immediately at the beginning of the process (Granata et al., 2012). The curing process takes several months and the final product is 71 characterized by firm consistency, reddish colour, juicy texture and characteristic odour and 72 flavour (Felix et al., 2016; Granata et al., 2012; Sospedra et al., 2015). Salted-ripened 73 anchovies may be packed in brine or preserved in oil. For preservation in oil, fish are 74 generally skinned, washed, dried and filleted (Mohamed et al., 2016). 75

76 The term "marinades" or "marinated fish" is used to define products consisting of fish processed with an edible organic acid, usually acetic acid, and salt, which gives them a 77 characteristic white colour of the flesh, and put into brines, sauces, or oil (McLay, 1972). 78 79 Pickled anchovies are very popular in Spain as *boquerones en vinaigre* and in Italy as *alici* marinate. Traditionally, homemade marinated anchovies are prepared with fresh fish 80 eviscerated and de-boned by hand, then pickled in lemon juice or vinegar and salt for less 81 than 24h before consumption. Although the Italian and Spanish legislation requires 82 preventive freezing treatment also in case of domestic preparation of raw, marinated or not 83 84 fully cooked fish (D'Amico et al., 2014Decreto Legislativo 17 Luglio 2013; Real Decreto 1420/2006), this is frequently not applied (Serracca et al., 2014), because it alters the texture 85 and the taste of fish meat (Sánchez-Monsalvez et al., 2005; Vidaček et al., 2009). 86

Among the most important biohazards related to the consumption of raw anchovies is the presence of viable zoonotic nematode larvae belonging to the genus *Anisakis*, as their ingestion is responsible for a zoonotic disease known as anisakiasis (Mattiucci et al., 2013). Of the nine genetically characterized species of the genus *Anisakis*, only *A. pegreffii* and *A. simplex* (*s. s.*) have been reported as causative agents of human gastric, intestinal and gastroallergic anisakiasis (Cipriani et al., 2017). *A. simplex s.l.* and *A. pegreffii* are frequently found in European anchovies (Bao et al., 2017; Costa et al., 2016).

The presence-occurrence of anisakid larvae in fish is a natural condition throughout the supply chain and their complete elimination from fishery products is not feasible (EFSA, 2010). Food Business Operators (FBOs) must ensure that fishery products obviously contaminated with visible parasites are not placed on the market for human consumption, by conducting a visual inspection of fresh fish products (Commission Reg. EC No 2074/2005). In addition, the parasitological risk associated to the presence of viable larvae in semiprocessed seafood products can be prevented by applying a freezing treatment or an 101 appropriate brining or pickling process for a sufficient time (AESAN, 2007; Anastasio et al., 2016; Sánchez-Monsalvez et al. 2005). Nevertheless, the presence of dead visible parasites in 102 processed products represents a defect that alters the overall quality (Codex Alimentarius, 103 104 2012; Council Reg. EC No 2406/1996) making them unfit for human consumption (Reg. EC No 178/2002). In fact, the finding of parasites in fish products causes immediate consumers' 105 rejection and may damage the reputation of the brand. Moreover, although it is generally 106 107 believed that sensitization with live Anisakis spp. larvae is required prior to the development of a clinical allergic responses, it has been also suspected supposed that also ingestion (and 108 109 inhalation) of dead larvae or their allergens might induce allergic reactions (Bao et al., 2017; EFSA, 2010; Mattiucci et al., 2017). 110

In a preliminary phase of this study 44 ready to eat products made of anchovies, herrings, 111 mackerel and sardines were analysed (Guardone et al., 2016a). Considering that all the 112 samples made of mackerel and sardines were negative, while larvae were found in 80.0% of 113 the products made of anchovies, the present study specifically addressed this type of product. 114 Taking into account the increasing request of ready to eat seafood products from the EU 115 (EUMOFA, 2017), the high prices of semi-preserved anchovies and the scarcity of data on on 116 the presence of anisakid parasites associated toin these kind of preparations (Fraulo et al., 117 2014; Sospedra et al., 2015), the aim of this study was to assess the presence occurrence of 118 visible anisakid larvae in different commercial categories of products sold in Italian 119 120 supermarkets. The most appreciated types of semi-preserves on the national market, such as salted-ripened, in oil and marinated anchovies, were collected and analyzed. 121

122 **2.** Materials and methods

123 *2.1 Sampling*

A total of 107 ready to eat products made of anchovies, belonging to 17 different brands and to different lots were sampled between April 2015 and May 2017 in Tuscany (Northern 126 Italy), at different points of sale of a large national purchasing consortium. A convenience, non-probabilistic sampling was conducted, structured to include a proportional number of 127 products per type and brand. Three different types of commercial products were collected: 128 salted-ripened, in oil and marinated (Fig. 1). In 51 products, the fishes were only beheaded 129 and (partially) gutted, but the bones were not removed and the structure of the body was 130 maintained unaltered ("whole" anchovies) (Fig. 1a), while in the remaining 56 products the 131 anchovies were deboned and opened to become flat ("fillets") (Fig. 1b-c). Thirty-three 132 products were salted-ripened anchovies (all whole fishes), 49 products were in oil (18 whole 133 134 fishes and 31 fillets) and 25 products were marinated anchovies (all fillets). The samples were then transferred to the FishLab, University of Pisa, Department of Veterinary Sciences, 135 University of Pisa, and analysed. 136

137

2.2 Parasitological analysis

2.2.1 Digestion procedure. Each sample was registered with an internal unique code. 138 Photos of the external packaging with the labelling information and of the internal content 139 were taken. In the case of whole specimens anchovies, their number of specimens wasere 140 counted, and the Mean Abundance (MA) was calculated (see Section 2.4). Salt, brine and oil 141 were carefully removed from the products. Salted products were also lightly rinsed with tap 142 water in a glass beaker. The oil was also carefully removed also with the aid of absorbent 143 paper. Then, the edible part was weighted. Considering that the whole content of the 144 145 collected products is edible, the full weight of each sample was digested. With the aim tT o test the recovery rate of parasites from semi-preserved anchovy products, preliminary trials 146 were performed. Larvae collected from products analysed in the preliminary phase of this 147 study (Guardone et al., 2016a) were submitted to artificial digestion using the Trichineasy[®], 148 according to the manufacturer's instructions (CTSV, 2007). according to the procedure 149 described in Guardone et al., (2017). Considering that aAll the larvae were recovered with 150

thisset procedure, which was then applied to all the samples. , digesting. A <u>aA</u> maximum of
 200 g of tissue <u>was digested</u> per time. <u>was digested according to the manufacturer's</u>
 instructions (CTSV, 2007http://www.ctsv.biz/image-ctsv/PDF/TrichinEasy-anisakis.pdf).

At the end of the digestion the material retained in the filter was rinsed with water and 154 divided in Petri dishes to create a thin layer of a few mm. The Petri dishes were observed 155 under natural and UV light (UltraBright UV Transilluminator, 302/365 nm, Maestrogen, Las 156 Vegas, USA) for the detection of anisakid larvae. During this step, spontaneous and 157 stimulated movements of the larvae were assessed to evaluate viability. In consideration of 158 the provisions of the Regulation EC No 853/2004 and subsequent amendments, only the 159 visible larvae (non-encapsulated nematodes longer than 1 cm or parasites with a capsular 160 diameter of at least 3 mm according to the definition given by the Codex Alimentarius 161 162 Commission, 1971) were counted and collected. The residual salt and oil and the water used to rinse the anchovies were inspected as described above. The larvae found during this step 163 were collected and summed to those found after the complete digestion. All the larvae were 164 identified to genus level following Sakanari and McKerrow (1989) and Berland (1989) by 165 observation under a microscope (Nikon Eclipse E200) and then stored in 70% alcohol for 166 molecular analysis. 167

2.2.2 Molecular identification. A subset of Anisakis larvae (from 1 to 4 larvae per product)
was submitted to molecular identification. Total DNA extraction was performed according to
the protocol used in Guardone et al., (2016b). DNA concentration and purity were determined
by a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Wilmington, DE,
USA).

A 629-bp fragment of the mitochondrial cytochrome *c* oxidase subunit II (*cox2*) gene was amplified using the primers 211F (5'-TTT TCT AGT TAT ATA GAT TGR TTY AT-3') and 210R (5'-CAC CAA CTC TTA AAA TTA TC-3') (Nadler & Hudspeth, 2000). PCR 176 amplifications were set up in a 20 μ l reaction volume containing 2 μ l of a 10× buffer (biotechrabbit GmbH, Hennigsdorf, Germany), 200 µM of each dNTP (dNTPmix, 177 EurocloneS.p.A-Life Sciences Division, Pavia, Italy), 200 nM primers, 1.25 U PerfectTaq 178 DNA Polymerase (biotechrabbit GmbH, Hennigsdorf, Germany), and 50-100 ng of DNA and 179 DNase free water (Water Mol. Bio. Grade, DNase-RNase and Protease free, 5Prime GmbH, 180 Hamburg, Germany) with the following cycling program: initial denaturation at 94 °C for 3 181 min; 40 cycles at 94 °C for 20 s, 45 °C for 20 s, 72 °C for 25 s; final extension at 72 °C for 10 182 min, as in Guardone et al., (2016b). 183

184 PCR products were checked by gel electrophoresis and the presence of fragments of the expected length was assessed by comparison with the marker SharpMass[™]50-DNA ladder 185 (Euroclone, Wetherby, UK). PCR products were purified with EuroSAP PCR Enzymatic 186 187 Clean-up kit (EuroClone Spa, Milano) and stored at -80°C prior to the sequencing. The sequencing of the PCR products were sequenced was carried out by the Experimental 188 Institute of Zooprophylaxis of Piedmont, Liguria and Aosta Valley (Turin, Italy) to obtain 189 forward and reverse direction sequences for each PCR product. The sequencing reaction was 190 performed by the use of a 4-capillary 3130 Genetic Analyzer (Applied Biosystems) and the 191 BigDye® Terminator v3.1 Cycle Sequencing kit (Life Technology, Thermo Fisher Scientific 192 Inc.). 193

All the obtained sequences were analyzed using Bioedit version 7.0.9 (Hall, 1999).
Adjustments were made after visual checking and the sequences were analysed on GenBank
by using the Basic Local Alignment Search Tool (BLAST) (Altschul et al., 1990).

197 2.4 Statistical analysis

2.4.1 Comparison of the three product categories. Salted, in oil and marinated products
 were compared in respect to: positivity (presence of at least one larva, (nominal variable) and
 number of total larvae in each analysed sample (counting variable). presence of at least one

201 larva (nominal variable), number of larvae per product and density (larvae/gram) (quantitative variables). To assess differences among groups two tests were applied: the γ^2 202 test for the nominal variable and the Kruskal-Wallis test for the counting variable. The non-203 parametric tests were chosen given the unequal sample size, the presence of categories with 204 less than 30 products and, not least, the violation of the ANOVA assumptions, mostly the 205 homogeneity of variance. For all the analyses, significant results were those associated with 206 p<0.05. If overall significance was observed, pair-wise comparisons were analysed using χ^2 207 (for nominal variables) and Mann-Whitney (for quantitative data) tests. In these comparisons, 208 209 in order to protect for type I error increase, a threshold of α =0.01 was chosen for the interpretation of the results. Analyses were performed using SPSS v 15^(R). 210

211 2.4.2 Comparison between products made of fillets and whole anchovies. Differences in 212 positive samples the number of larvae per product, frequency of products contaminated by at 213 least one larva and larval density per gram were also analysed the occurrence, in the number 214 of larvaedetected wereas also analyzed in respect to the product being composed by whole 215 anchovies or by fillets. The analyses were carried out using χ^2 (for nominal variables) and 216 Mann-Whitney (for quantitative data) tests. These comparisons were performed only for 217 products preserved in oil, the only category containing both fillets and whole fishes.

2.4.3 Mean abundance (MA). The mean abundance (MA) (total number of individuals of a 218 particular parasite species in a sample of a particular host species divided by the total number 219 of hosts of that species examined, Bush et al., 1997) was calculated after the complete 220 digestion of products made of whole specimens whole products and the value obtained was 221 used to issue a marketability judgement. The MA threshold was calculated by applying an 222 approach <u>regional law</u> widely used throughout Italy (D'Amico et al., 2014) which defines the 223 maximum number of tolerated larvae in fresh batches of anchovies (three-3 larvae in 10% of 224 the sampled fish). Considering that in the case of fish species caught in large batches, such as 225

anchovies, the number of subjects to collect for a significant sampling is, at least, 29, the
 maximum number of parasites tolerated is 9 and therefore the MA threshold is 0.3 (Guardone
 et al., 2016b, Guardone et al., 2017).

3. Results and discussion

230 *3.1 Parasitological analysis*

The official method for the detection of parasites in fish is the visual inspection 231 (Commission Reg. EC 2074/2005). The pressing method of frozen fillets followed by the 232 examination under ultraviolet light is also frequently used (Gómez-Morales et al., 2017). 233 234 Moreover, the artificial digestion may also be applied to isolate larvae from fish and it is considered the gold standard for its higher sensitivity (Guardone et al., 2016b; Llarena-Reino 235 et al., 2013). The cuticle of parasitic nematodes has been reported as highly resistant to strong 236 237 acids and digestive enzymes, regardless of whether the nematodes are live or have been killed by freezing or conventional heating (Tejada et al., 2006). However, damages to the cuticle 238 occurring during processing (Anastasio et al., 2016; Tejada et al., 2006; Vidacek et al., 2009) 239 can affect the resistance of the larvae to the artificial digestion. For this reasonAs mentioned, 240 trials were performed using dead Anisakis spp. larvae-collected from products (salted and in 241 oil) analysed in a preliminary phase of this study (Guardone et al., 2016a). Since Aall the 242 larvae were recovered after the digestion, therefore, the procedure applied to fresh anchovies 243 (Guardone et al., 2017), already proven to be able to recover live larvae, the procedure was 244 245 considered suitable also for semi-preserved products.

At least one visible larva was found in 58 (54.2%) of Of the total 107 products analysed,
58 (54.2%) were positive for the presence of at least one visible larva. A total of 1283 visible
larvae were collected, which were all morphologically identified as *Anisakis* sp.Overall a
total of 1283 anisakid larvae were collected. Strong differences were observed between the
various categories of products and also between whole and filleted products (Table 1). All the

251 parasites found during the analysis were dead. In fact, even Aalthough emission of fluorescence is known didto not always allow to discriminate between live and dead larvae 252 and might be related to the stress produced in the larvae by the treatment (Tejada et al., 2006; 253 Vidaček et al., 2009), spontaneous and stimulated movements of the larvae were absent in 254 this study. All the visible parasites were morphologically identified as Anisakis sp. Among 255 the subset of ose molecularly analysed larvae (n=122), 92 (75.4%) were identified as A. 256 pegreffii and 30 (24.6%) as A. simplex (Table 2). Larvae of the genus Hysterothylacium were 257 found very rarely (4 samples) and were always shorter than 1 cm,-and thus they were not 258 259 counted as visible larvae. The low prevalence of Hysterothylacium spp. may be due to the fact that these parasites are generally smaller and thinner than Anisakis spp. and might be less 260 resistant to processing techniques. 261

262 The complete elimination of parasites from fishery products is not feasible (EFSA, 2010), therefore it is necessary to establish a threshold to discriminate between fit and unfit products 263 (Reg. EC 178/2002). In particular, it is essential to identify the number of larvae that can be 264 tolerated in a product and to adopt a criterion for taking decisions on the marketability of 265 fishery products. According to the "Guidance document on the implementation of certain 266 provisions of Regulation (EC) No 853/2004 on the hygiene of food of animal origin" 267 (European Commission, 2014) a fishery product is considered obviously contaminated if 268 visible parasites are detected in edible portions. However, such document does not define a 269 270 maximum number of parasites. Therefore, in a previous work (Guardone et al., 2016b), a MA threshold calculated as described above, was used to assess the marketability of fresh batches 271 of anchovies. Especially in the case of small fish, which are not sold individually, the MA 272 273 could be used to estimate the degree of infestation.

3.1.1 Salted anchovies. <u>At least one visible *Anisakis* sp. larva was found in all All</u>the 33
products (100%) were found positive for the presence of at least one larvae. Totally, 1139

larvae of *Anisakis* sp. were collected in this category, corresponding to 88.8% of the total
collected larvae. The mean number of larvae per product was 34.5² (±29.3³ standard
deviation), with great variability (range: 1- 105). The mean density (larvae per gram) was
0.13 (Table 1). The highest number of larvae (439) was found in the products belonging to
brand 5 (Table 2). The results show that salted anchovies are the most contaminated type of
products, which is likely due to the type of processing (see Section 3.2).

Parasites recovered from these products were molecularly identified as *A. pegreffii* [77.4%] and *A. simplex* (22.6%) (Table 2). The majority of the larvae of *A. simplex* found in these samples was collected from anchovies declared to be fished in the Cantabrian sea (FAO area 27), while *A. pegreffii* was the dominant species in samples declared as fished in the Mediterranean Sea, confirming previous epidemiological data (Costa et al., 2016 and references therein).

<u>T</u>the MA_, which varied from 0.04 to 3.92. Twenty-nine products (87.98%) exceeded the
<u>MA</u> threshold of positivity previously set for fresh anchovies. No differences in MA values
were observed in the distribution of the positivity to the MA threshold in relation to the
different brands (Table 2).

3.1.2 Products in oil. Among the 49 products, 18 were made of whole anchovies and the remaining 31 of fillets. The 18 whole products belonged to 4 different brands. Two of them consisted of previously salted anchovies (red flesh, brand 2 and 7), while the other two presented a white meat (brand 1 and 8) (Table 2).

At least one *Anisakis* sp. larva was found inOverall 24 (49.08.98%) products in oil were found positive for the presence of at least one larva and a total of 143 larvae of *Anisakis* sp. were collected, corresponding to 11.1% (143/1283) of the total larvae collected. A mean number of 2.9 larvae per product was detected (± 5.80 standard deviation) with a great variability (range: 0-28 larvae). The larval density per gram was 0.03 (Table 1 and 2). Parasites recovered from products in oil were molecularly identified as *A. pegreffii* (70.3%) and *A. simplex* (29.7%). The geographical origin is not compulsory for fishery products in oil (D'Amico et al., 2016) and it was not reported for 5 of the 14 brands. All the larvae molecularly identified from these products were *A. pegreffii*. Most of the remaining indicated FAO 37 or FAO 37.2.1 and the dominant species was *A. pegreffii*. Only the products of one brand were claimed to originate from FAO area 27. In these samples the majority of the identified larvae were *A. simplex*.

It was possible to calculate the MA for 15 products₁₇ <u>J</u>in fact₁ although other 3 products (brand 8) were originally prepared with whole anchovies it was not possible to count them due to the loss of integrity of the specimens induced by the processing (Table 2). Of these 15 samples, all the 10 products made of salted anchovies (brand 2 and 7) exceeded the set MA threshold. On the contrary, <u>no larvae were found in all</u>-the 5 products of brand 1-were <u>negative</u>. The high <u>positivity-contamination level</u> in whole salted in oil anchovies confirms the results obtained for salted-ripened anchovies. The MA varied from 0.0 to 2.8.

Different levels of contamination ces in positivity were observed in relation to the whole and filleted products (at least one larva was found in 61.14% of the whole products and 41.94% of the fillets was positive for at least one larvae). Within whole products, differences were also observed between red and white fish: 83.92% of the parasites (n=120) were found in the 2 products made of red whole anchovies.

3.1.3 Marinated anchovies. Only 1 visible *Anisakis* spp. larva was found in out of the 25
marinated products (4.0%). The larva was found positive with the presence of one *Anisakis*spp. larva that was subsequently molecularly identified as *A. pegreffii*. The larval density per
gram was 0.00 (Table 1 and Table 2). Considering that all these products consisted in filleted
anchovies it was not possible to calculate the MA.

13

325 *3.2 Comparison between product categories: influence of the processing technology on* 326 *the presence-occurrence and viability of anisakid larvae*

The processing technology can influence the presence of parasites in the final products. The present study showed a significant difference between all the product categories in respect of both number of larvae_per_product (Kruskal-Wallis' $\chi^2=69_{1.7}95$; p<0.001), __and positivity <u>number of contaminated products ($\chi^2=50.34$; p<0.001). frequency of contaminated</u> products ($\chi^2=50.34$; p<0.001) and density of larvae per gram ($\chi^2=58.89$; p<0.001).

The average number of larvae <u>per product</u> was <u>about around 35</u>, 3 and 0 for salted, in oil and marinated products, respectively (Table 1). <u>Analogously Similarly, the</u> frequency of <u>contaminated products being positive in each category was 100.0%</u>, 49.0% and 4.0% (Table 1). <u>In addition the density was different across products: mean density of 0.13 (s.d.= 0.09) in</u> <u>salted products, 0.03 (s.d=0.06) in products conserved in oil and 0.0 (s.d.=0.001) in marinated</u> <u>products.</u>

As known, for other fish species, also in the case of anchovies most aAnisakid larvae are 338 known to be located in the fish visceral cavity and/or embedded in the visceral organs and in 339 the adjacent muscles (belly flap) (EFSA, 2010). Larval migration to the muscles may occur 340 after the capture, especially in the case of an inappropriate refrigeration (Cipriani et al., 341 2016). When visible parasites are only found in non-edible parts of the fishery product, 342 processing procedures, such as gutting, ensure that the raw materials are not obviously 343 344 contaminated (European Commission, 2014). On the countrary, when the viscera removal is not complete, the final product may harbour a high number of parasites. This is the case of 345 salted-ripened anchovies, where the gut is not completely removed as intestinal enzymes 346 347 seem to play an essential role in ripening (Czerner et al., 2011). In fact, at least one larva was found in all theeach analysed salted products were positive. Similarly, all the whole salted 348 anchovies in oil were positive contaminated withfor a high number of larvae and exceeded all 349

14

350 thresholds. Overall 1259 larvae were found in whole salted anchovies in brine and in oil these products (1139 in 33 salted products and 120 in 2 salted whole anchovies in oil). Another 351 larva was found in one of the "white" whole anchovies in oil (Table 1 and 2). As concerns the 352 fillets in oil, these are generally previously treated as whole salted anchovies for the 353 maturation process, and only after this phase they are filleted and put under oil. The lower 354 presence of parasites in this kind of products can be explained by the fact that parasites are 355 removed together with the gut residual during filleting. Statistical analyses revealed the 356 significance of the differences (Z=-2.98; p<0.01) observed between whole and filleted 357 358 anchovies in oil. The same differences were found when the larval density was evaluated (Z=-2.98; p<0.02), with a value of 0.07 (s.d.=0.08) in whole fish compared to 0.01 (s.d=0.02) 359 in fish fillets. The analyses were performed only in products preserved in oil considering that 360 361 the salted products were all whole fish and the marinated ones were all filleted.- The presence of very low positivity found for marinated products (only one larva in the 25 marinated 362 products) analysed may be explained by the fact that this kind of products are usually filleted 363 364 as fresh, hampering the parasitic migration from the viscera to the muscle. The very low contamination of industrially marinated anchovies sampled in this study agrees with the 365 results of Sospedra et al., (2015) who analysed the same products from Spanish restaurants, 366 while it is well known that domestically prepared marinated anchovies are one of the 367 products most at risk for human anisakiasis (Bao et al., 2017; Mattiucci et al., 2013). 368

As concerns the viability of the larvae in semi processed anchovy products, it is known that salting may reduce the parasite hazard by killing anisakid larvae if salt content and time are adequate (Codex Alimentarius, 2012; Karl et al., 1994). Recently, the opinion No. 2007-SA-0379 of the French Food Safety Agency (AFSSA, 2007), reported that salting inactivates A<u>a</u>nisakidae larvae within 21 or 28 days depending on the final salt concentration in fish. In a recent work, all the larvae collected from anchovies salted according to a traditional Italian procedure (final salt concentration of 24.5%) were found dead after 15 days (Anastasio et al.,
2016). In addition, sSalted-ripened anchovies undergo a ripening process after salting that
takes at least 2-3 months for *E. encrasicolus* (Anastasio et al., 2016). Therefore, the
processing time in this kind of products is much longer than the one required to effectively
kill the larvae.

Nematodes have been reported as highly resistant to the conditions created by traditional marinating methods, being able to survive for periods of a few days up to several weeks, depending on the concentration of salt, acetic acid and marinating times (AESAN 2007; Anastasio et al., 2016; Karl et al., 1994). In the traditional marinating process, the fish is left in a solution of vinegar and salt for less than 24 h. However, in a study the death of all larvae in fillets exposed to vinegar did not occur until day 13 (Sánchez -Monsalvez et al., 2005).

Considering that all the larvae found were dead, the processing technologies (including the preventive freezing treatment applied by FBOs according to the European legislation) for the production of semi preserved anchovy products analysed in this study seem to be effective to nullify the risk of contracting human gastrointestinal anisakiasis.

390 3.3 Dead anisakid larvae in semi-preserved anchovies: <u>a potentially hazardous</u> defect 391 and hazard?

Dead visible larvae can be considered a defect according to the definition of the Codex 392 Alimentarius: "A condition found in a product that fails to meet essential quality, composition 393 and/or labelling provisions of the appropriate Codex product standards" (Codex 394 Alimentarius, 2012). Anisakis sp. larvae are whitish to transparent and are not easily detected 395 by the naked eye when they reside deeply embedded in fish muscles. On the contrary, they 396 397 are evident when they infect in high number the celomatic cavity of fish species. This is particularly true in case of fresh fish containing live larvae that which can actively move and 398 become evident also on the external surface (Guardone et al., 2016b). However, dead visible 399

larvae can also be considered a defect according to the definition of the Codex Alimentarius:
"A condition found in a product that fails to meet essential quality, composition and/or *Labelling provisions of the appropriate Codex product standards*" (Codex Alimentarius,
2012). also In fact, the presence of dead larvae can-represents a reason to disqualify the fish
product (Council Reg. EC No 2406/1996) and to consider it not fit for human consumption
according to(-Reg. (EC) No 178/2002).

406 The finding of parasitized products on the European market has elicited numerous RASFF (Rapid Alert System for Food and Feed) notifications over the years. Between 2010 and 407 408 2016, 409 notifications for the presence of anisakid larvae in fishery products were issued. Among these, the state of the product was indicated in 327 cases: besides fresh or chilled 409 products (n=254), 81 referred to non-fresh products (frozen, smoked, salted, marinated and in 410 411 oil) and thus probably involving dead larvae. In some of the heavily contaminated products found in this study, visible parasites were evident at visual inspection even before opening 412 the packet or simply observing the fish edible tissue (Fig. 2). The observation of a similar 413 414 contamination by consumers might result in disgust and rejection of the product and may also damage the brand reputation. 415

The ingestion of live Anisakis spp. worms may cause hazardous allergic reactions, 416 including anaphylaxis, generally in association with gastrointestinal forms (EFSA, 2010; 417 Daschner et al., 2012; Mattiucci et al., 2013). On the contrary, Tthe potential of dead larvae 418 419 to induce allergies in sensitized subjects is still debated (Daschner et al., 2012). Oral challenges performed in clearly allergic subjects with non-infective frozen or lyophilized 420 larvae (Alonso-Gòmez et al., 2004; Sastre et al., 2000) and parasitic antigens (Baeza et al., 421 422 2004; Daschner et al., 2000) did not elicit any adverse effect. However, according to different authors, allergic reactions may also occur after ingestion of processed fish or parasite proteins 423 alone (Audicana and Kennedy, 2008; Nieuwenhuizen et al. 2006) and it has been supposed 424

that no-viable larvae or related antigens could be involved in chronic urticarial reactions
(Mattiucci et al., 2017). Accordingly, the high prevalence (72.5%) of *Anisakis* larvae in
frozen fillets of pink salmon was considered a public health issue due to the potential risk for
allergic reactions in sensitized persons (Bilska-Zajac et al., 2016). The issue of allergic
reactions is also related to different fish-eating habits, which probably account for different
sensitization rates or the frequency of allergic symptoms in the different regions of the world
(Mattiucci et al., 2017).

Therefore, even though it is not possible, on the basis of the current knowledge, to 432 433 consider dead larvae as a proven hazard, appropriate measures should be implemented to avoid commercialization of obviously contaminated products. This would require FBOs 434 involved in processing of salted, in oil or marinated anchovies, at industrial or artisanal level, 435 436 to include appropriate risk management measures in theirs self-checking programs. In practice, FBOs should implement a system, based on the sampling method associated with a 437 visual inspection as usually applied in Italy (D'Amico et al. 2014), or others of similar 438 efficiency, to inspect batches of fresh anchovies. This would allow to select the most 439 appropriate kind of processing (salting, preparation in oil or marinating) on the basis of the 440 level of contamination detected. In fact, in this study, the level of contamination depended on 441 the products' typology, being high in salted-ripened, medium in fillets in oil and very low for 442 industrially marinated anchovies. The observed differences are linked to the preliminary 443 444 preparation of the fish, in particular to the complete or incomplete removal of the viscera. Batches with a higher level of contamination should be destined to the production of 445 marinated products. This would be economically advantageous for industries to reduce the 446 costs arising from the discard of heavily contaminated batches of fresh anchovies and from 447 the withdrawal of unfit product from the market. The continuously growing awareness of 448 consumers and food authorities as to the occurrence of parasites in seafood, emphasises the 449

450 importance of providing the fish processing industries with procedures able to reduce hazards
451 and defects.

452 <u>In fact, an incorrect risk communication can influence consumers' trust and even lead to a</u> 453 significant reduction of fish consumption.

A study conducted in Italy showed that the highest prevalence was detected along the
Adriatic and Tyrrhenian coasts where marinated anchovies are a frequently consumed
traditional food, often prepared at home. In seaside areas of Southern Italy, where anchovies
are generally eaten fried rather than marinated, *Anisakis* hypersensitivity was much less
commonly found (AAITO IFIACI, 2011). The same association between *Anisakis*hypersensitivity and marinated seafood was observed in studies in Spain (Garcia et al., 1997;
Valinas et al., 2001).

461 Therefore, even though it is not possible, on the basis of the current knowledge to consider
462 the dead larvae as a proven hazard, a precautionary approach should be adopted. In practice,
463 this would mean to adopt appropriate measures to reduce the risk of ingestion of dead larvae.
464 ... In fact, when the available supporting information and data are not sufficiently complete
465 to enable a comprehensive risk assessment, official authorities may take measures based on
466 the precautionary principle, while seeking more complete scientific and other data (Reg. EC
467 178/2002).

Incorrect risk communication can influence consumers' trust and even lead to a significant
reduction of fish consumption. Therefore, the finding of contaminated products by FBOs
within their self-control programs requires corrective actions to avoid that products heavily
contaminated with dead larvae reach the market. The HACCP approach, usually aimed at
ensuring food safety and preventing risks, can also be applied to cover food quality aspects, if
instead of identifying the hazards of the process, potential defects are considered. The
continuously growing awareness of consumers and food authorities as to the possible

475 presence of parasite or parasite-related quality defects in seafood emphasises the importance
476 of providing the fish processing industries with feasible procedures able to monitor hazard
477 and defect.

478 Conclusion

The present work highlighted how showed that semi preserved anchovy products heavily 479 contaminated with Anisakis spp. larvae can reach the market and that the processing 480 481 technology can influence the occurrence of parasites in semi-preserved products. Therefore, the batches intended for the production of these products (whole or filleted) should be 482 483 accurately selected by industries, at the initial phases of the fish supply chain, according to the industrial fate of the raw material. Beyond the negligible risk for anisakidosis, due to the 484 inactivation of larvae by freezing and processing technologies, the occurrence of dead 485 486 parasites may cause immediate rejection in consumers. In addition, the risk related to allergic reactions in sensitized individuals is still an open issue. Providing the fish processing 487 industries with procedures able to reduce hazards and defects is particularly important in the 488 light of the continuously growing awareness of consumers and food authorities as to the 489 occurrence of parasites in seafood. and that the processing technology can influence the 490 occurrence of parasites in semi-preserved products. Therefore, the batches intended for the 491 production of these products (whole or filleted) should be accurately selected by industries, at 492 the initial phases of the fish supply chain, according to the industrial fate of the raw material. 493 Providing the fish processing industries with procedures able to reduce hazards and defects is 494 particularly important in the light of the continuously growing awareness of consumers and 495 food authorities as to the occurrence of parasites in seafood. In particular, the level of 496 497 contamination depended on the products' typology, being high in salted-ripened, medium in fillets in oil and very low for industrially marinated anchovies. The observed differences are 498 499 strictly linked to the processing procedure and to the preliminary preparation of the fish, in

500 particular to the complete or incomplete removal of the viscera. Beyond the negligible risk for anisakidosis, due to the inactivation of larvae by freezing and processing technologies, the 501 presenceoccurrence of dead parasites may cause immediate rejection in consumers. In 502 addition, the risk related to allergic reactions in sensitized individuals is still an open issue. 503 This study demonstratedshowed that the processing technology can influence the presence 504 of parasites in the final semi-preserved products. Therefore, the batches intended for the 505 506 production of these products (whole or filleted) should be accurately selected by industries, at the initial phases of the fish supply chain, according to the industrial fate of the raw material. 507

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514 Captions

515 Figure 1 Presentation of the most part of the products analysed in the present study: whole 516 salted anchovy (left), salted fillet preserved in oil (centre), marinated fillet (right).

Figure 2 From left to right: (a) salted anchovies heavily contaminated, one of the larvae was already visible from outside the glass jar before opening; (b) detail of another heavily contaminated salted product, the larva was visible from the external of the package; (c) larva in the muscle (edible part) of a salted anchovy; (d-e) parasites collected from the one of the most contaminated products: (d) natural light, (e) UV light.

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(n)	n (% of the total of<u>contaminated</u> products for each	(% of the total- of collected	Range	larvae per product	(larvae/gram)	
	category)	larvae <mark>products</mark>)				
Salted (33)	33 (100.0%)	1139 (88.8%)	1-105	34.5 (± 29.3 SD)	0.13	
In oil (49)	24 (49.0%)	143 (11.1%)	0-28	2.9 (± 5.8 SD)	0.03	
Marinated (25)	1 (4.0%)	1 (0.1%)	0-1	0.0 (± 0.2 SD)	0.00	
Total (107)	58 (54.2%)	1283	0-105	12.0 (± 22.5 SD)	0.05	

SD: Standard Deviation

Table 2 Summary of the results obtained analysing the 107 products, sub-divided per product category and per brand. NA: Not Available

Commercial name of the product	Product codes	Geographical origin	Product presentation	Mean net weight/ product (g)	N analysed products	Total n L3 <i>Anisakis</i> spp.	Density (larvae/gram)	Range	N positive <u>contaminated</u> products ^a	N products exceeding MA threshold ^b	Molecular identification (n analyzed)
Salted anchovies Brand 1	RTE25, RTE33, RTE72, RTE73, RTE74	Atlantic Ocean NE FAO 27.VIII.C, Cantabrian Sea	whole	153.8	5	105	0.1	5-51	5	4	A. pegreffii (1) A. simplex (11)
Salted anchovies Brand 2	RTE12, RTE36, RTE42, RTE63, RTE69, RTE70	South Gulf of Biscay, Cantabrian Sea	whole	195.8	6	165	<mark>0.1</mark>	1-42	6	5	A. pegreffii (11) A. simplex (2)
Salted anchovies Brand 3	RTE 46, RTE48, RTE49, RTE87, RTE88, RTE89	Mediterranean Sea FAO 37.2.1	whole	100.0	6	55	<mark>0.1</mark>	3-15	6	5	A. pegreffii (12) A. simplex (2)
Salted anchovies Brand 4	RTE53, RTE75, RTE76, RTE93, RTE94	FAO 37	whole	221.4	5	146	<mark>0.1</mark>	20-48	5	5	A. pegreffii (13) A. simplex (1)
Salted anchovies Brand 5	RTE5, RTE6, RTE7, RTE24, RTE144, RTE145	FAO 37.2.1	whole	427.9	6	439	<mark>0.2</mark>	39-105	6	6	A. pegreffii (16) A. simplex (2)
Salted anchovies Brand 6	RTE32, RTE110, RTE121, RTE126, RTE149	FAO 37.2	whole	571.8	5	229	<mark>0.1</mark>	2-87	5	4	A. pegreffii (12) A. simplex (1)
Total salted anchovies (%)					33	1139		1-105	33 (100.0)	29 (87.9)	A. pegreffii (65) A. simplex (19)
Anchovies in oil Brand 7	RTE23, RTE40, RTE65, RTE67, RTE71	FAO 37	whole (red)	73.2	5	40	<mark>0.1</mark>	3-13	5	5	A. pegreffii (5) A. simplex (4)
Anchovies in oil Brand 2	RTE37, RTE58, RTE64, RTE66, RTE95	Not reported	whole (red)	109.5	5	80	<mark>0.1</mark>	8-28	5	5	A. pegreffii (13)
Anchovies in oil Brand 1	RTE54, RTE59, RTE61, RTE68, RTE92	Not reported	whole (whitish)	88.4	5	0	<mark>0.0</mark>	0	0	0	
Anchovies in oil Brand 8	RTE127, RTE128, RTE129	Not reported	whole ^c (whitish)	285.0	3	1	0.0	0-1	1	NA ^c	
Anchovies in oil Brand 9	RTE103, RTE104, RTE105, RTE106	Mediterranean Sea FAO 37	fillets	88.0	4	1	<mark>0.0</mark>	0-1	1	NA	A. simplex (1)
Anchovies in oil Brand 5	RTE8, RTE142, RTE143	FAO 37	fillets	140.0	3	2	<mark>0.0</mark>	0-1	2	NA	A. pegreffii (1)
Anchovies in oil Brand 10	RTE34, RTE108, RTE109	FAO 37	fillets	60.0	3	2	<mark>0.0</mark>	0-1	2	NA	A. simplex (1)
Anchovies in oil Brand 7	RTE116, RTE117, RTE118	FAO 37	fillets	50.7	3	0	<mark>0.0</mark>	0	0	NA	
Anchovies in oil Brand 11	RTE136, RTE137, RTE138	Not reported	fillets	82.3	3	0	<mark>0.0</mark>	0	0	NA	

Total (%)					107	1283		0-105	58 (54.2)		A. pegreffii (92) A. simplex (30)
Total marinated anchovies (%)					25	1		0-1	1 (4 <u>.0</u>)		A. pegreffii (1)
Marinated anchovies Brand 17	RTE9, RTE10, RTE146, RTE147, RTE148	Not reported	fillets	121.9	5	0	<mark>0.0</mark>	0	0	NA	
Marinated anchovies Brand 16	RTE113, RTE114, RET115, RTE152, RTE153	Adriatic Sea	fillets	135.6	5	1	<mark>0.0</mark>	0-1	1	NA	A. pegreffii (1)
Marinated anchovies Brand 2	RTE55, RTE56, RTE57, RTE96, RTE97	Not reported	fillets	108.0	5	0	<mark>0.0</mark>	0	0	NA	
Marinated anchovies Brand 12	RTE52, RTE60, RTE62, RTE98, RTE99	Adriatic Sea	fillets	124.4	5	0	0.0	0	0	NA	
Marinated anchovies Brand 9	RTE39, RTE102, RTE107, RTE150, RTE151	Mediterranean Sea FAO 37	fillets	70.1	5	0	<mark>0.0</mark>	0	0	NA	
Total anchovies in oil (%)					49	143		0-28	24 (49.0)		A. pegreffii (26) A. simplex (11)
Anchovies in oil Brand 6	RTE120, RTE122, RTE123	FAO 37.2.1	fillets	22.3	3	1	<mark>0.0</mark>	0-1	1	NA	A. pegreffii (1)
Anchovies in oil Brand 15	RTE139, RTE140, RTE141	Not reported	fillets	59.3	3	0	<mark>0.0</mark>	0	0	NA	
Anchovies in oil Brand 14	RTE119, RTE124, RTE125	FAO 37.2	fillets	112.4	3	2	<mark>0.0</mark>	0-1	2	NA	A. pegreffii (1) A. simplex (1)
Anchovies in oil Brand 13	RTE130, RTE131, RTE132	FAO 37.2.1	fillets	51.2	3	4	<mark>0.0</mark>	0-1	2	NA	A. pegreffii (3)
Anchovies in oil Brand 12	RTE133, RTE134, RTE135	FAO 27	fillets	64.7	3	10	<mark>0.0</mark>	2-5	3	NA	A. pegreffii (2) A. simplex (4)

^apresence of at least 1 larva; MA: mean abundance; L3: third stage larvae; ^bMA thresholdLpG 1 proposed in Guardone et al., <u>2016b2017</u>; ^cdespite the fact that the product was originally prepared with whole anchovies it was not possible to count their number of specimens due to the loss of <u>anatomical</u> integrity of the specimens induced by processing.



