

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

## Structural and environmental constraints on reduction of paired appendages among vertebrates

### This is the author's manuscript

*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/1704663> since 2019-06-18T16:29:36Z

*Published version:*

DOI:10.1093/biolinnean/blz097

*Terms of use:*

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

# **Structural and environmental constraints on paired appendages reduction among vertebrates**

LOREDANA MACALUSO<sup>1\*</sup>, GIORGIO CARNEVALE<sup>1</sup>, RAFFAELLO CASU<sup>2</sup>, DANIEL  
PIETROCOLA<sup>1</sup>, ANDREA VILLA<sup>3,1</sup> and MASSIMO DELFINO<sup>1,4</sup>

<sup>1</sup>Dipartimento di Scienze della Terra, Università degli Studi di Torino, Via Valperga Caluso 35, 10125, Torino, Italy

<sup>2</sup> Dipartimento di Fisica, Università degli Studi di Torino, Via Pietro Giuria 1, 10125, Torino, Italy

<sup>3</sup> Bayerische Staatssammlung für Paläontologie und Geologie, Richard-Wagner-Straße 10, 80333 München, Germany

<sup>4</sup> Institut Català de Paleontologia Miquel Crusafont, Universitat Autònoma de Barcelona, Edifici Z (ICTA-ICP), Carrer de les Columnes s/n, Campus de la UAB, E-08193 Cerdanyola del Valles, Barcelona, Spain

\*Corresponding author. E-mail: [loredana.macaluso@unito.it](mailto:loredana.macaluso@unito.it)

## ABSTRACT

Burrowing habits or complex environments have been generally considered as potential drivers acting on appendicular skeleton reduction and loss among vertebrates. Herein we suggest that this may be the case for lissamphibians and squamates, but that fin loss in fishes is usually prevented due to important structural constraints, as pectoral fins are commonly used to control rolling and pitching. We provide an overview of the distribution of paired appendage reduction across vertebrates while examining the ecological affinities of finless and limbless clades. We analysed the correlation between life style and fin or limb loss using the discrete comparative analysis. The resulting Bayesian Factors indicate a strong evidence of correlation between: (i) pectoral-fin loss and coexistence of anguilliform elongation and burrowing habits or complex habitat in teleost fishes, and (ii) limb loss and burrowing or grass-swimming life style in squamate reptiles and lissamphibians. These correlations suggest that a complex environment or a fossorial habit constitute driving forces leading to appendage loss. The only locomotion style that is functional even in absence of paired appendages is the undulatory one, which is typical of all elongated reptiles and lissamphibians, but certainly less common in teleost fishes.

**ADDITIONAL KEYWORDS:** limb loss - eel-like fishes - elongated tetrapods - comparative phylogenetic analyses - amphibians - reptiles

## INTRODUCTION

Reduction or loss of the appendicular skeleton occurred multiple times in representatives of several fish and tetrapods lineages. Among extant tetrapod classes (Moyle & Cech, 2003; Vaughan *et al.*, 2011; Pough *et al.*, 2015; Morrison *et al.*, 2018), extreme reduction or loss of limbs occurred in mammals (cetaceans and sirenians), birds (moa and kiwi), squamate reptiles (snakes, amphisbaenians and several groups of lizards) and “amphibians” (the Carboniferous-Permian aistopods and the lissamphibian caecilians plus a few groups of salamanders), whereas the loss of paired fins has been observed in several groups of teleost fishes. Even though in both mammals and birds, reduction and skeletal loss affect alternatively the forelimbs or hindlimbs only, there are several groups of squamate reptiles, amphibians and teleost fishes that exhibit a complete absence of paired limbs or fins. Despite these groups are phylogenetically distant, they all share some similarities. First of all, squamate reptiles, lissamphibians and teleosts characterized by a reduction of the appendicular skeleton usually exhibit an elongated body (Gans, 1975; Ward & Mehta, 2010 and references therein). According to Greer (1991), there are fifty-three squamate lineages that have undergone limb reduction, distributed among Agamidae, Amphisbaenia, Anguillidae, Anniellidae, Chamaleonidae, Cordylidae, Dibamidae, Diploglossidae, Gekkota, Gerrhosauridae, Helodermatidae, Lanthanotidae, Ophidia, Pygopodidae, Scincidae and Teiidae. The situation is different among lissamphibians as, although different groups exhibit an elongated body, only three lineages show appendages reduction or loss. Caecilians (Gymnophiona) are the only lissamphibians characterized by a complete loss of both fore- and hindlimbs. Members of Sirenidae, on the other hand, have lost the hindlimbs, but retain reduced forelimbs. In the end, the only genus included within Amphiumidae, *Amphiuma*, has very short and motionless limbs. Among extinct basal tetrapods, the Carboniferous-Permian Aistopoda are completely limbless, showing a remarkable convergence with the serpentine body plan of snakes and caecilians (Carroll *et al.*, 1998; Pardo & Mann, 2018). In general, all the elongated squamates and lissamphibians exhibit a similar lateral undulation even if using different types of locomotion (Gans, 1975). Conversely, elongated fishes exhibit carangiform or anguilliform locomotion (Sfakiotakis *et al.*, 1999, Horner & Jayne, 2008; Pfaff *et al.*, 2016). If the carangiform pattern is characterized by more or less rigid movements of the caudal portion of the body and tail, the anguilliform pattern is characterized by a sinuous wave that moves

through the body (e.g., eels) or only in its posterior part (e.g., catfishes). The anguilliform movement, which could be associated to the “serpent-like” movement of tetrapods, is typical of at least half of the highly elongated actinopterygians (Ward & Mehta, 2010; Reece & Mehta, 2013). In particular, anguilliform elongation (*sensu* Ward & Mehta, 2010) is typical of *Erpetoichthys calabaricus* among Polypteriformes (Suzuki *et al.*, 2010), some members of the clades Ophidiiformes, and Gobionellidae (genus *Luciogobius*; Yamada *et al.*, 2009), and most of the members of the clade Anguilliformes, Lampridiformes, Mastacembelidae, and Zoarcales (Ward & Mehta, 2010). Some elongated members of the clades Ateleopodiformes, Liparidae, and Siluriformes show a body plan with an enlarged, heavy head and their anguilliform movement is limited to the posterior part of the body (i.e., not including the pectoral area). The pelvic fins are lost in several, morphologically different groups of fishes, in more than 90 teleost lineages (Nelson, 1989; Kriwet & Pfaff, 2019), whereas the pectoral fins are primarily lost in elongated fishes with anguilliform locomotion (Mehta *et al.*, 2010). Eel-like clariid species represent a unique case among vertebrates, as their paired fins show a very high intraspecific level of morphological variability, preventing their absence to be used as a diagnostic feature at the species level (Devaere *et al.*, 2004). As in other vertebrates (O’Reilly *et al.*, 1997), the loss of fins in clariids was regarded as related to a highly specialized fossorial mode of life by Devaere *et al.* (2004), although this hypothesis has not been tested yet. More generally, the idea that the presence of appendages may represent an impediment for burrowing for both terrestrial and aquatic animals is widely accepted. Recently, Da Silva *et al.* (2018) demonstrated that fossoriality has been the evolutionary driver leading to the origin and development of the snake body plan. Extinct relatives of extant limbless forms (e.g., amphisbaenians and caecilians) indicate that fossoriality evolved before the limb loss since they have cranial adaptations for burrowing, but maintain variably developed appendages (Evans & Sigogneau-Russell, 2001; Jenkins *et al.*, 2007; Talanda, 2016). Nevertheless, fossoriality is not the only evolutionary driver for limb loss in reptiles, and limbless squamates are traditionally divided into short-tailed burrowers or long-tailed surface dwellers, moving through loose sand or vegetation (Evans, 1998; Wiens *et al.*, 2006).

As far as fishes are concerned, it is known that a number of fishes exhibiting an eel-like body morphology are either crevice-dwellers or burrowers (tail- or head-first; De Schepper *et al.*, 2007a, b; Herrel

*et al.*, 2011). Nevertheless, previous studies mainly focused on the correlation between elongation and habitat, or trophic adaptations (Ward & Mehta, 2010; Mehta *et al.*, 2010; Claverie & Wainwright, 2014), but did not find any apparent connection. Mehta *et al.* (2010), in particular, stated that, although it is generally true that terrestrial vertebrates evolved an elongated, limb-reduced body plan as an adaptation for the burrowing lifestyle, little is known about how much the elongate body form may be adapted for aquatic habits.

Herein we suggest that a distinction can be done in fishes between anguilliform and stiffer-body elongation (*sensu* Ward & Mehta, 2010) when studying correlations between habitat and body plan, as the constraints due to balance problem during swimming connected with these two body forms are different. Moreover, we attempt to address the question: once a fish has evolved an eel-like elongation, does the habitat have an influence on paired-fins loss? Is there a common trend in appendage loss in teleosts and tetrapods? Herein, we argue that the limbless body plan in vertebrates is affected by similar environmental constraints, provided by the habitat or life style, and we try to summarize how widespread this pattern (i.e. dense, complex environment acting on appendage reduction and loss) actually is.

#### GENETIC AND DEVELOPMENTAL CONTROL OF APPENDAGE LOSS

The development of appendages is polygenic, involving genes with pleiotropic effects (Lande, 1978; Hall, 2008). Therefore, genes involved in limbs and paired fins development also function on other developing systems, such as jaws or genitals (Rosa-Molinar & Burke, 2002). This is the reason why genes associated with limb buds are generally not lost, even in limbless forms (Bejder & Hall, 2002). The developmental mechanism of the formation of paired appendages is deeply conserved among gnathostomes (Dahn *et al.*, 2007; Letelier *et al.*, 2018) and it involves two signalling centers located in the fin/limb bud. The first of them is the apical ectodermal ridge (AER), which helps to maintain the second one, the zone of polarizing activity (ZPA), the cells of which express the Sonic hedgehog (Shh) gene, associated with the development of the fins or limbs (Cohn, 2001; Bejder & Hall, 2002; Thewissen *et al.*, 2006). At the same time Hox genes control the position of both girdles and appendages along the body. In particular, the anterior expression boundaries of HoxC-6 and HoxC-8-10 coincide with the localization of fore- and hindlimb buds respectively (Bejder & Hall, 2002). Reduction and loss of appendages can occur due to regression of

different phases in the conserved genetic pathway for appendage development. Tanaka *et al.* (2005), for example, reported that pelvic-fin loss can be achieved through different mechanisms in pufferfishes and sticklebacks. In the first case, the reduction is due to an altered expression of the gene *Hoxd9a* in lateral mesoderm, whereas in the second case *Pitx1*, a gene responsible for appendage initiation, fails to be expressed (Shapiro *et al.*, 2004). In pythons, limbs development is arrested in two different ways. Forelimb buds are not developed at all, because of the widespread expression of *HoxC-6* and *HoxC-8* genes throughout the lateral plate mesoderm, meaning that no boundary conditions are established for forelimbs to form and therefore there is no pectoral limb initiation (Cohn & Tickle, 1999; Cohn, 2001; Bejder & Hall, 2002). On the contrary, hindlimb buds are formed, but they have a very smooth ectodermal jacket forming a small AER, which causes a precocious interruption of the growth (Cohn, 2001). Serpentine lizards and urodeles have different levels of limb reduction that can vary considerably depending on the species (Greer, 1991). Nevertheless, in general, the formation of their limb buds starts and then regresses, or the expression of *Shh* can have a shortened duration, leading to loss of some digits or of a larger part of the limbs (Raynaud, 1990; Hinchliffe, 2002; Shapiro *et al.*, 2003; Tanaka *et al.*, 2005). As far as cetaceans are concerned, pelvic limb buds begin to form but fail to fully develop, in a similar way to the python hindlimbs (Bejder & Hall, 2002; Tanaka *et al.*, 2005). Nevertheless, it is likely that the mechanism is slightly different from that of snakes, as the dolphin *Stenella attenuata* shows a normal AER during development, but the absence of *Hand2* – one of the upstream regulators of the *Shh* transcription – causes a perturbed initial establishment of the ZPA and the consequent absence of *Shh* expression (Thewissen *et al.*, 2006).

Therefore, fin and limb formation has indeed very conserved genetic and ontogenetic pathways among gnathostomes, but developmental causes of appendage loss can be very diverse within and among different groups (Hall, 2008).

## MATERIAL AND METHODS

We compiled a database (Appendix 1 and 2) that includes 125 species of teleost fishes, 74 species of lissamphibians and 151 species of squamate reptiles. For the taxon sampling of teleost fishes, we selected all the groups characterized by pectoral fin loss: Anguilliformes, Clariidae, Gobionellidae (strong reduction of the pectoral fin in the genus *Luciogobius*), and Trichomycteridae. The groups of Zoarcales and

Mastacembelidae have some finless members, but we did not include them in our analysis, as their ecology and phylogenetic relationship are poorly known.

We combined different phylogenetic trees that contain the group included in the study (either the recent-most or the complete-most phylogenetic analyses), and in particular: Santini *et al.* (2013) for Anguilliformes, Baskin (1973), Datovo & Bockmann (2010), and DoNascimento (2015) for Trichomycteridae, Wright (2017) for Clariidae, and Yamada *et al.* (2009) and Thacker (2013) for Gobionellidae. We sampled part of the taxa contained in these phylogenies, several of them being used as outgroups, as they do not show any pectoral fin loss. Particular attention was paid on groups with finless members included in these phylogenies, among which at least one representative species for every family has been selected, but including all the taxa with complete information about fins and ecological habits that were close to the node where pectoral fins were lost. In fact, for studies concerning causal relationships of specific traits, the most important taxa to sample are the ones near to the node where the trait we want to study (e.g., fin or limb loss) first occurred and got fixed. Any potential evolutionary innovation that originated after the first occurrence and fixation of the trait should not be considered as a potential evolutionary driver and it is therefore not essential to insert many derived taxa in the sample (Macaluso & Tschopp, 2018). The same procedure was followed to compile the database of lissamphibians (complete sample of basal Gymnophiona, Sirenidae, and Amphiumidae), starting from the phylogeny published by Pyron and Wiens (2011), and squamate reptiles (complete sample of basal limbless Agamidae, Amphisbaenia, Anguidae, Anniellidae, Dibamidae, Diploglossidae, Ophidia, Pygopodidae, and Scincidae), using the phylogenies of Reeder *et al.* (2015), Da Silva *et al.* (2018), and Pyron *et al.* (2013).

The taxon sampling was of course limited by the information available for the taxa and we chose species for which the following is known: i) phylogenetic position; ii) presence or absence of appendages; iii) behaviour or ecology. Our limited knowledge of these data is particularly relevant in the case of extinct taxa, because of the difficulty in reconstructing their life habits. Moreover, it is not so common to find a complete articulated skeleton of terrestrial animals that can tell us if appendages were present or not in a certain taxon (see for example the case of the stem-group caecilian described by Evans & Sigogneau-Russell, 2001). For these reasons, the sample of extinct taxa is limited in this study to relatives of extant taxa



representing their stem, whenever information about them is available. We collected information about elongation of the body, presence or absence of the appendages, and lifestyle (see Appendix 1, 2, and 3 for the references). Elongation has only been scored for fishes, which display different kinds of elongation (anguilliform and stiffer-body; Ward & Metha, 2010; Maxwell & Wilson, 2013) and it is therefore important to consider this character in the analysis. In particular, fishes are herein considered as elongated if their length is more than five times the maximum body depth. The considered cases of fin loss in fishes are restricted to those taxa without pectoral fins because the pelvic fins are absent in a huge number of groups due to different factors (e.g., Nelson, 1989). *Luciogobius* has been scored as lacking its pectoral fins as it shows an extreme pectoral fin reduction compared to its close relatives (Hyun-Geun & Seung-Ho, 2014). Squamate reptiles and lissamphibians were scored as lacking their limbs (1) if both pairs of appendages were absent, and as intermediate (01) if only one pair of appendages has been lost. The only exceptions to this rule are snakes with underdeveloped hindlimbs (e.g., pythons, *Pachyrhachis*, *Haasiophis*), the genus *Dibamus* in which the presence of hindlimbs are a sexual dimorphic character (vestigial hindlimbs only present in males; Koppetsch *et al.*, 2019), and *Amphiuma*, which have been scored as limbless although they possess small vestigial hindlimbs useless for locomotion. Lifestyle has been divided in “fossorial” or “not fossorial”. Fishes are scored as 1 concerning “fossoriality” (i.e., column “fossoriality” in Appendix 1) when they have either burrowing habits or are crevices-dwellers, whereas squamate reptiles and lissamphibians were scored as “fossorial” (i.e., 1 in column “fossoriality” in Appendix 2) when they are either burrowers or grass-swimmers. Although burrowing and grass-swimming result in different constraints acting on the whole body, the lateral sides of the body (and consequently the appendages) of burrowing and grass-swimming animals, are constrained in a similar way, by the substrate in burrowers and crevices-dwellers and grass in grass-swimmers. Additional information and references about life style, phylogenetic position, and presence/absence of appendices are present in the supplementary material (Appendix 1, 2, and 3). After collecting these data, we compiled a composite phylogenetic tree in Mesquite (Maddison & Maddison, 2018) reporting all the included taxa, based on already existing phylogenies listed above. We performed the discrete comparative analysis available in the software BayesTraitsV3 (Meade & Pagel, 2017). Discrete comparative analysis is used to test if two binary traits are correlated and its significance is established by

comparing the likelihoods (derived using Markov chain Monte Carlo – MCMC) of two models, one assuming that the traits evolved independently and the other assuming that their evolution is correlated. Two binary traits can be described by four possible states, written as “0,0”, “0,1”, “1,0” and “1,1”. The independent model assumes that the two traits evolve independently, e.g. the transition from 0 to 1 in the first trait is independent from the state of the second trait, whereas the dependent model assumes that the traits are correlated and the rate of change in one trait is dependent from the state of the other. The test was performed structuring an input database as an Excel file of a table at two entrances (see Appendix 1, 2): species in the rows and characters in the columns. Concerning fishes, the two binary characters are pectoral fin loss and a character that is scored as 1 only if the taxon is both elongated (as defined above) with anguilliform locomotion and either fossorial or crevices-dweller. As all the fossorial (or grass-swimmers) lissamphibians and squamates have an extremely elongated body, elongation has not been considered as an essential character in their case and the two binary characters are therefore limb loss and fossoriality or grass-swimming. Given that arbitrary branch lengths are commonly used and well-supported in the literature using Comparative Methods (Grizante *et al.*, 2012, and references therein), we performed the statistical analyses using an arbitrary branch length of 1.0 and all branches were scaled to 0.1, as suggested for the software BayesTraitsV3 (Meade & Pagel, 2017). As is usual in this kind of analysis, we set all the priors to an exponential with a mean of 10 and use the stepping stone sampler with 100 stones and 1000 iterations per stone to estimate the marginal likelihood (see the manual for users of BayesTraitsV3). We performed two different analyses, one for teleost fishes and a separate one for lissamphibians and squamate reptiles, to make it easier to manipulate the large trees in Mesquite. The phylogenetic trees we built are reported in Figures 1 and 2. It is worth noting that in the discrete analysis of BayesTraitsV3 it is not necessary to infer the ancestral state of the characters and thus the coloured branches in the figures are just graphical representations. Here, we evidence the character states with different colours, referring to appendices 1 and 2. In particular, names depicted in red indicate species scored with 1 for appendage loss, whereas light blue species are the ones scored as 01. Colour of the branches refers to the second character, intended as the co-occurrence of eel-like elongation and burrowing behaviour (or crevices or sea-grass inhabitants) for teleost fishes and burrowing or grass-swimming habits for squamate reptiles and lissamphibians.

## RESULTS

The analysis resulted in two values of the marginal likelihood, one for the dependent model and one for the independent model (Table 1). Both are described in a logarithmic scale. To test whether the traits are correlated or not, we calculate a log Bayes Factor between the dependent and independent models. The calculations for Log Bayes factors are given below.

$$\text{Log BF} = 2 (\log \text{marginal likelihood dependent model} - \log \text{marginal likelihood independent model})$$

The Log BF of 41.467358 in one case and of 44.253558 in the other suggest that there is strong evidence for correlated evolution, as a “strong evidence” of correlation is considered when Log BF has values higher than 5 (Gilks *et al.*, 1996).

## DISCUSSION

The environmental conditions may represent relevant evolutionary drivers leading to the emergence of new body morphologies within clades. Nevertheless, very different environmental conditions may provide similar constraints on organisms living – and moving – in them, thereby leading to the development of convergent morphologies. Burrowing (or interstitial) animals, grass-swimmers, and marine crevices-dwellers represent an example of this, because the presence of appendages is not favoured in the environment where they live.

### SQUAMATES AND LISSAMPHIBIANS

Previous hypotheses about squamate reptiles and lissamphibians developing a limbless body plan as an adaptative response to burrowing or grass-swimming (Evans, 1998; Wiens *et al.*, 2006; Da Silva *et al.*, 2018) are confirmed by our results. Basal scolecophidian snakes and more derived fossil taxa (e.g., *Dinilysia*, *Wonambi*) are fossorial (or semifossorial as in the case of *Yurlunggur*; Palci *et al.*, 2018) and several lines of evidence support the hypothesis that the fossoriality of basal snakes is plesiomorphic (Miralles *et al.*, 2018). Moreover, comparative geometric morphometric studies on skulls demonstrated that lizards could not have transitioned to snakes by any other evolutionary path than through fossoriality (Da Silva *et al.*, 2018).

The phylogenetic relationships of the stem lineage of Ophidia are still highly controversial and it is therefore difficult to understand the ecology of the basal-most fossil snakes. For example, the life style of the Cretaceous snake *Coniophis* has been reconstructed as fossorial (Longrich *et al.*, 2012), but its phylogenetic

position is not resolved. In fact, Longrich *et al.* (2012) consider it as the basalmost stem ophidian, but Caldwell *et al.* (2015) place this taxon in a more derived position. The situation is similar as far as most of the stem taxa are concerned. In general, caution is warranted when using single fossil snakes to make broad extrapolations about early snake biology (Palci *et al.*, 2018). It is also still unresolved which one between the body- or head-first hypotheses is the most likely, with different evidence sustaining either the former or the latter (Longrich *et al.*, 2012; Caldwell *et al.*, 2015; Da Silva *et al.*, 2018). It is important, nevertheless, to remark that our results suggest a general evolutionary trend connecting fossoriality (and, more broadly, complex habitats) with limb-loss, but this does not mean that this same evolutionary force acted in every single group that evolved a reduction or loss of the limbs, as different constraints can act in different groups (Macaluso & Tschopp, 2018). There are, in fact, few groups of squamates that evolved a limbless body, which are generalist surface-dwellers. Two remarkable exceptions are, for example, the skinks and the pygopodids. Skinks evolved limblessness independently in several lineages, even within a single genus (e.g., *Lerista*; Skinner *et al.*, 2008; Fig. 3G), many of which are burrowers, whereas some others are also more generalist surface-dwellers (Weins *et al.*, 2006; Camaiti *et al.*, 2019). Pygopodidae is the only family of gekkotans that has members devoid of limbs. They are generally surface-dwellers, even if the basal-most forms live in the litter (Dorrough & Ash, 1999; Wall & Shine, 2013), a lifestyle that can be regarded as fossorial. The fact that limb loss is so common within squamates is not surprising, since elongated reptiles are characterized by an undulatory locomotion and appendages that primarily help to carry forward the body (Sfakiotakis *et al.*, 1999; Grillner, 2011) and limbs can therefore be lost without a relevant impact on their fitness.

Within Amphisbaenia (Fig. 3I), the stem taxon *Slavonia darevskii* apparently proves that fossoriality evolved before the limb loss, as it has clear cranial adaptations for burrowing, but also limbs (Tałanda, 2016). A different phylogenetic analysis placed *Cryptolacerta hassiaca* on the stem of this group (Müller *et al.*, 2011), but the situation is substantially unchanged, as it shows partially reduced limbs and cranial adaptations for burrowing. The same situation is found in the clade Gymnophiona, in which the fossil taxon *Eocaecilia micropodia* possesses fossorial adaptations and small limbs (Jenkins *et al.*, 2007). Salamanders of the family Sirenidae (Fig. 3E) lack hind limbs and show some digital reduction of the

forelimbs and Amphiumidae have very small and motionless limbs, but there is no information on the appendicular skeleton of stem forms referred to these groups (Lande, 1978).

Studies of developmental genetics clearly evidence that there are multiple ways to produce a limbless body plan (Kohlsdorf *et al.*, 2008). In a similar way, it is likely that fin or limb loss in different, unrelated groups may be originated through different evolutionary drivers. In any case, our analysis suggests that there is a general correlation between burrowing or grass-swimming habits and limb loss.

## TELEOSTS

As far as fishes are concerned, the situation is more complex, because of their locomotion style. Fishes generally use the caudal fin as a propulsor, whereas the paired fins are used to control lateral movement and to prevent rolling and pitching, although there are also fishes that use oscillatory or undulatory movements of the paired fins as thrust generation (see Sfakiotakis *et al.*, 1999 for an extensive review on fish swimming mode). Short-bodied fishes with ostraciiform swimming mode as well as elongated fishes with carangiform locomotion use their pectoral (and pelvic) fins to control their body and therefore they simply cannot lose their fins, even if this change would be advantageous in their environment. On the contrary, the anguilliform swimming mode may permit the loss of paired fins without a remarkable effect on locomotion capability. In this case, in fact, the paired fins are not useful to prevent rolling or pitching, even if they can help locomotion (Sfakiotakis *et al.*, 1999). This is also clearly demonstrated by the cetaceans and sirenians, the only mammal clades with representatives characterized by elongated bodies, which lose a pair of appendages. During the course of their evolutionary history, cetaceans and sirenians have lost pelvic fins, in a similar way to numerous fish clades (e.g., Nelson, 1989, Bejder & Hall, 2002). Their locomotion is undulatory, but differently from that of eel-like fishes, since waves are produced in a vertical plane, which do not prevent from the rolling and pitching problems. This is surely coupled with the fact that they did not lose their pectoral fins, which have acquired a stabilizing and steering function, not generating any propulsive movements (Bejder & Hall, 2002). Our results show that fin loss in fishes is restricted to taxa characterized by an eel-like morphology of the body and most likely related to the burrowing lifestyle or to cryptic life in reef ecosystems. This is clearly evidenced by the consistently eel-like morphology of finless taxa that are characterized by burrowing or crevice-dwelling habits (e.g., Muraenidae; Fig. 3A). The most diverse clade of

eel-like fishes is the elopomorph order Anguilliformes, in which pelvic fins are generally absent, but pectoral fins are present in some groups. Although the interpretation of the ecological preference of extinct fishes is not always easy, it is reasonable to hypothesize that basal anguilliforms (e.g., *Anguillavus* or *Luenchelys*; Belouze, 2002; Belouze *et al.*, 2003 a,b) were in some ways reef-associated, because the Cretaceous plattenkalk deposits in which they have been found originated on the outer part of the Lebanese carbonate platform, which was mostly occupied by oyster and rudist mounds and patch reefs (Hemleben & Swimburne, 1991). Members of the extant families Protanguillidae and Synaphobranchidae, regarded as the most basal lineages of crown Anguilliformes (Santini *et al.*, 2013), have small pectoral fins, and are characterized by a variety of ecological adaptations. For example, protoanguillids live in submarine caves (Johnson *et al.*, 2011) and the most basal synaphobranchid, *Simenchelys parasitica*, developed a peculiar parasitic lifestyle (e.g., Jaquet, 1920). The main anguilliform group of real burrowers is the Moringuidae, whose members are in general burrowers (head or tail-first) or crevice-dwellers with a marked reduction of paired fins (Castle, 1986; Allen & Steene, 1988; De Schepper *et al.*, 2005). The extant species of the genus *Anguilla* are demersal and do not show a clear reduction of the paired fins, a pattern also shared with three extinct species, *A. ignota*, *A. multiradiata* and *A. elegans* (Winckler, 1861; Micklich, 1985; Riede, 2004; Gaudant *et al.*, 2018). Some lineages within the family Congridae (i.e. the clade composed by *Ariosoma*, *Heteroconger* and *Paraconger*, see Santini *et al.*, 2013) includes burrowing fishes devoid of paired fins (Smith, 1981; Riede, 2004; Bacchet *et al.*, 2006). Pectoral fins are lost also in certain crevices-dwelling species belonging to the family Muraenidae (Robins *et al.*, 1991; Chen *et al.*, 1994; Lieske & Myers, 1994; McCosker, 2010; Reece *et al.*, 2010).

A unique case is the benthic gobionellid genus *Luciogobius*, because it is probably the only fish taxon adapted to an interstitial life in gravel beaches (Yamada *et al.*, 2009). The main adaptation consists in an anguilliform elongation of the body that confer it enough agility to move in a three-dimensional complex habitat, similar to that characteristic of terrestrial and aquatic burrowers (Gans, 1975; Yamada *et al.*, 2009). Interestingly, in parallel with the elongation of the body, interstitial species of *Luciogobius* (Fig. 3C) underwent fin reduction, whereas elongated species of Gobionellidae, which are not interstitial but have a benthic or nektonic lifestyle (e.g., genera *Inu* or *Clariger*), exhibit completely developed paired fins. A

similar condition is also characteristic of those species of the Gobionellidae that live in reefs or on muddy substrates but are not eel-shaped (e.g., *Periophthalmus barbarous*, *Scartelaos histiophorus*).

Within the Neotropical catfish family Trichomycteridae, the Glanapteryginae are interstitial fishes living in complex habitats and they are mostly eel-shaped with a clear reduction of paired fins, which are usually very thin becoming filiform and without any equilibrium function (De Pinna, 1988; Schaefer *et al.*, 2005; Villa-Verde & Costa, 2006). Catfishes belonging to Clariidae are generally elongated, with an anguilliform swimming mode that is often limited to the posterior part of the body, also extending to its anterior part in some species. All the species devoid of paired fins live in complex or highly vegetated habitats, thereby confirming the hypothesis of fin reduction driven by environmental complexity (Fig. 3B; see Appendix 1 for references).

Finally, the Mastacembelidae represent another very peculiar case, as they are eel-shaped fishes, also called spiny eels because of the long series of dorsal-fin spines (Vreven, 2005). It is worth mentioning them because of the particular case of the only two species of this group (*Mastacembelus apectoralis* (Fig. 3D) and its sister taxon, *M. micropectus*) that exhibit a considerable reduction in pectoral-fin size, which is considered to be related to the highly structured environments they live in (Brown *et al.*, 2011). Their phylogenetic relationships are poorly known, and thus they have not been sampled in our comparative analysis.

Our study reveals the existence of a correlation between the reduction and/or loss of pectoral fins and the coexistence of an eel-like body morphology, which makes unnecessary the use of pectoral fins to prevent rolling and pitching, and of an environmental constraint due to burrowing and crevice- or seagrass-dwelling. In fact, whereas being different habitat, the latter all have a similar effect on the appendages of fishes, because paired fins may hamper a free movement in these constrained environments. A similar correlation between burrowing or grass-swimming habits and limb loss is found also in squamate reptiles and lissamphibians, but the structural constraints are much less important in these groups as the locomotion is less constrained by appendages than in fishes due to the absence of balance problems.

## FUTURE PERSPECTIVES

A relation between long-tailed patterns and surface dwelling, and conversely pre-caudally elongated morphotypes and burrowers has been proposed for squamates reptiles (Bellairs & Underwood, 1951; Evans, 1998), even if this hypothesis has never been proved with rigorous analyses. An interesting future perspective is to expand this hypothesis to anguilliform fishes. In fact, crevice- and seagrass-dwellers could be somehow associated with terrestrial surface-dwellers. Mehta *et al.* (2010) reported that the elongation in muraenids (i.e., crevices-dwellers) results from the addition of caudal rather than precaudal vertebrae to their axial skeleton, whereas elongation of the body in ophichthids and congrid (i.e. burrowers) is achieved by adding a similar number of vertebrae to their precaudal and caudal regions; however, additional studies on this subject including a larger taxon sampling and much more comparative information would be desirable.

### ACKNOWLEDGEMENTS

We would like to show our gratitude to Marco Camaiti (Monash University, Clayton, Australia), for critical discussion on the topic of this paper, and Marcelo Sánchez-Villagra (Paläontologisches Institut und Museum, Zürich, Switzerland) and Phil Mannion (University College London), for their comments on an earlier draft of the manuscript. Many thanks go to Jürgen Kriwet (University of Vienna), Mateusz Tałanda (University of Warsaw), and anonymous reviewer and the Editor John A Allen (University of Southampton) for constructive advice on improving the manuscript. This work was supported by grants (ex-60% 2018) from the Università degli Studi di Torino.

### REFERENCES

- Allen GR, Steene RC. 1988.** *Fishes of Christmas Island Indian Ocean*. Australia: Christmas Island Natural History Association.
- Bacchet P, Zysman T, Lefèvre Y. 2006.** *Guide des poissons de Tahiti et ses îles*. Tahiti (Polynésie Française): Éditions Au Vent des Îles.
- Baskin JN. 1973.** Structure and relationships of the Trichomycteridae. PhD thesis, City University of New York.
- Bejder L, Hall BK. 2002.** Limbs in whales and limblessness in other vertebrates: mechanisms of evolutionary and developmental transformation and loss. *Evolution & Development* 4: 445-458.
- Bellairs ADA., Underwood G. 1951.** The origin of snakes. *Biological Reviews* 26: 193-237.



- Belouze A. 2002.** Compréhension morphologique et phylogénétique des taxons actuels et fossils rapportés aux Anguilliformes (“poisons”, Téléostéens). *Documents des Laboratoires de Géologie de Lyon* **158**: 1-401.
- Belouze A, Gayet M, Atallah C. 2003a.** Les premiers Anguilliformes: I. Révision des genres cénomaniens *Anguillavus* Hay, 1903 et *Luenchelys* nov. gen. *Geobios* **36**: 241-273.
- Belouze A, Gayet M, Atallah C. 2003b.** The first Anguilliformes: II. Paraphyly of the genus *Urenchelys* (Woodward 1900) and phylogenetic relationships. *Geobios* **36**: 351-78.
- Brown KJ, Britz R, Bills R, Rüber L, Day JJ. 2011.** Pectoral fin loss in the Mastacembelidae: a new species from Lake Tanganyika. *Journal of Zoology* **284**: 286-293.
- Caldwell MW, Nydam RL, Palci A, Apesteguía S. 2015.** The oldest known snakes from the Middle Jurassic-Lower Cretaceous provide insights on snake evolution. *Nature Communications* **6**: 5996.
- Camaiti M, Villa A, Wencker LCM, Bauer AM, Stanley EL, Delfino M. 2019.** Descriptive osteology and patterns of limb loss of the European limbless skink *Ophiomorus punctatissimus* (Squamata, Scincidae). *Journal of Anatomy* doi: 10.1111/joa.13017
- Carroll RL, Bossy KA, Milner AC, Andrews SM, Wellstead CF. 1998.** Lepospondyli. In: Wellnhofer P (ed.) *Encyclopedia of Paleoherpetology*. Gustav Fisher Verlag, Stuttgart: 1-216.
- Castle PHJ. 1986.** Moringuidae. In: M.M. Smith and P.C. Heemstra (eds.) *Smiths' sea fishes*. Springer-Verlag, Berlin: 187-188.
- Chen HM, Shao KT, Chen CT. 1994.** A review of the muraenid eels (Family Muraenidae) from Taiwan with descriptions of twelve new records. *Zoological Studies* **33**: 44-64.
- Claverie T, Wainwright PC. 2014.** A morphospace for reef fishes: elongation is the dominant axis of body shape evolution. *PLoS ONE* **9**: e112732.
- Cohn MJ. 2001.** Developmental mechanisms of vertebrate limb evolution. *Novartis Foundation Symposium* **232**: 47-57.
- Cohn MJ, Tickle C. 1999.** Developmental basis of limblessness and axial patterning in snakes. *Nature* **399**: 474-479.

- Da Silva FO, Fabre AC, Savriama Y, Ollonen J, Mahlow K, Herrel A, Müller J, Di-Poï N. 2018.** The ecological origins of snakes as revealed by skull evolution. *Nature Communications* **9**: 376.
- Dahn RD, Davis MC, Pappano WN, Shubin NH. 2007.** Sonic hedgehog function in chondrichthyan fins and the evolution of appendage patterning. *Nature* **445**: 311–314.
- Datovo A, Bockmann FA. 2010.** Dorsolateral head muscles of the catfish families Nematogenyidae and Trichomycteridae (Siluriformes: Loricarioidei): comparative anatomy and phylogenetic analysis. *Neotropical Ichthyology* **8**: 193-246.
- De Schepper N, Adriaens D, De Kegel B. 2005.** *Moringua edwardsi* (Moringuidae: Anguilliformes): Cranial specialization for head-first burrowing? *Journal of Morphology* **266**: 356-368.
- De Schepper N, De Kegel B, Adriaens D. 2007a.** *Pisodonophis boro* (Ophichthidae: Anguilliformes): specialization for head-first and tail-first burrowing? *Journal of Morphology* **268**: 112-126.
- De Schepper N, De Kegel B, Adriaens D. 2007b.** Morphological specializations in heterocongrinae (Anguilliformes: Congridae) related to burrowing and feeding. *Journal of Morphology* **268**: 343-356.
- De Pinna MCC. 1988.** A new genus of trichomycterid catfish (Siluroidei, Glanapteryginae), with comments on its phylogenetic relationships. *Revue Suisse de Zoologie* **95**: 113-128.
- Devaere S, Teugels GG, Adriaens D, Huysentruyt F, Verraes W. 2004.** Redescription of *Dolichallabes microphthalmus* (Poll, 1942) (Siluriformes, Clariidae). *Copeia* **2004**: 108-115.
- DoNascimento C. 2015.** Morphological evidence for the monophyly of the subfamily of parasitic catfishes Stegophilinae (Siluriformes, Trichomycteridae) and phylogenetic diagnoses of its genera. *Copeia* **2015**: 933-960.
- Dorrough J, Ash JE. 1999.** Using past and present habitat to predict the current distribution and abundance of a rare cryptic lizard, *Delma impar* (Pygopodidae). *Australian Journal of Ecology* **24**: 614-624.
- Evans S. 1998.** Crown group lizards (Reptilia, Squamata) from the middle Jurassic of the British Isles. *Palaeontographica Abteilung A* **250**: 123-154.
- Evans SE, Sigogneau-Russell D. 2001.** A stem-group caecilian (Lissamphibia: Gymnophiona) from the Lower Cretaceous of North Africa. *Palaeontology* **44**: 259-273.
- Gans C. 1975.** Tetrapod limblessness: evolution and functional corollaries. *American Zoologist* **15**: 455-67.

- Gaudant J, Nel A, Nury D, Véran M, Carnevale G. 2018.** The uppermost Oligocene of Aix-en-Provence (Bouches-du-Rhône, Southern France): A Cenozoic brackish subtropical Konservat-Lagerstätte, with fishes, insects and plants. *Comptes Rendus Palevol* **17**: 460-478.
- Gilks WR, Richardson S, Spiegelhalter DJ. 1996.** Introducing Markov chain Monte Carlo. In: Gilks WR, Richardson S, Spiegelhalter DJ (eds.). *Markov chain Monte Carlo in practice*. London, Chapman and Hall/CRC, 1-19.
- Greer AE. 1991.** Limb reduction in squamates: identification of the lineages and discussion of the trends. *Journal of Herpetology* **25**: 166-173.
- Grillner S. 2011.** Control of locomotion in bipeds, tetrapods, and fish. *Comprehensive Physiology* **2**: 1179-1236.
- Grizante MB, Brandt R, Kohlsdorf T. 2012.** Evolution of body elongation in gymnophthalmid lizards: relationships with climate. *PloS ONE* **7**: e49772.
- Hall BK. 2008.** *Fins into limbs: evolution, development, and transformation*. Chicago: University of Chicago Press.
- Hemleben CH, Swinburne NHM. 1991.** Cyclical deposition of the Plattenkalk Facies. In Einsele G., Ricken W, Seilacher A (Eds.) *Cycles and Events in Stratigraphy*. Springer Verlag, Berlin: 572-591.
- Herrel A, Choi HF, Dumont E, De Schepper N, Vanhooydonck B, Aerts P, Adriaens D. 2011.** Burrowing and subsurface locomotion in anguilliform fish: behavioral specializations and mechanical constraints. *Journal of Experimental Biology* **214**: 1379-1385.
- Hinchliffe JR. 2004.** Developmental basis of limb evolution. *International Journal of Developmental Biology* **46**: 835-845.
- Horner AM, Jayne BC. 2008.** The effects of viscosity on the axial motor pattern and kinematics of the African lungfish (*Protopterus annectens*) during lateral undulatory swimming. *Journal of Experimental Biology* **211**: 1612-1622.
- Hyun-Geun C, Seung-Ho C. 2014.** First Record of Two Gobiid Fishes, *Luciogobius elongatus*, *L. platycephalus* (Perciformes: Gobiidae) from Korea. *Animal Systematics, Evolution and Diversity* **30**: 22.

- Jaquet M. 1920.** Contribution à l'anatomie du *Simenchelis parasiticus* Gill. *Resultats des Campagnes Scientifiques accomplis par le Prince Albert I de Monaco* **56**: 1-77.
- Jenkins FA, Walsh DM, Carroll RL. 2007.** Anatomy of *Eocaecilia micropodia*, a limbed caecilian of the Early Jurassic. *Bulletin of the Museum of Comparative Zoology* **158**: 285-366.
- Johnson GD, Ida H, Sakaue J, Sado T, Asahida T, Miya M. 2011.** A 'living fossil' eels (Anguilliformes: Protanguillidae, fam. nov.) from an undersea cave in Palau. *Proceedings of the Royal Society B* **279**: 934-943.
- Kohlsdorf T, Cummings MP, Lynch VJ, Stopper GF, Takahashi K, Wagner GP. 2008.** A molecular footprint of limb loss: sequence variation of the autopodial identity gene *Hoxa-13*. *Journal of molecular evolution* **67**: 581-593.
- Koppetsch T, Boehme W, Koch A. 2019.** A new species of *Dibamus* Duméril & Bibron, 1839 (Squamata: Dibamidae) from Pulau Manado Tua, Northern Sulawesi, Indonesia. *Zootaxa* **4555**: 331-345.
- Kriwet J, Pfaff C. 2019.** Evolutionary Development of the Postcranial and Appendicular Skeleton in Fishes. In: Johanson Z, Underwood C, Richter M. *Evolution and Development of Fishes*, 188-206.
- Lande R. 1978.** Evolutionary mechanisms of limb loss in tetrapods. *Evolution* **32**: 73-92.
- Letelier J, Calle-Mustienes E, Pieretti J, Naranjo S, Maeso I, Nakamura T, Pasqual-Anaya J, Shubin, NH, Schneider I, Martinez.Morales JR., Gómez-Skarmeta JL. 2018.** A conserved *Shh* cis-regulatory module highlights a common developmental origin of unpaired and paired fins. *Nature genetics* **50**: 504.
- Lieske E, Myers R. 1994.** *Collins Pocket Guide. Coral reef fishes. Indo-Pacific & Caribbean including the Red Sea*. New York, Harper Collins Publishers.
- Longrich NR, Bhullar BAS, Gauthier JA. 2012.** A transitional snake from the Late Cretaceous period of North America. *Nature* **488**: 205.
- Macaluso L, Tschopp E. 2018.** Evolutionary changes in pubic orientation are more strongly correlated with the ventilation system in dinosaurs than with herbivory. *Palaeontology* **61**: 703-719.
- Maddison WP, Maddison DR. 2018.** Mesquite: a modular system for evolutionary analysis. Version 3.51 <http://www.mesquiteproject.org>

- Maxwell EE, Wilson LA. 2013.** Regionalization of the axial skeleton in the ‘ambush predator’ guild—are there developmental rules underlying body shape evolution in ray-finned fishes? *BMC Evolutionary Biology* **13**: 265.
- McCosker JE. 2010.** *Rhinomuraena quaesita* (errata version published in 2017). The IUCN Red List of Threatened Species 2010: e.T155301A115297865.
- Meade A, Pagel M. 2017.** BayesTraits V3. 0.
- Mehta RS, Ward AB, Alfaro ME, Wainwright PC. 2010.** Elongation of the body in eels. *Integrative and Comparative Biology* **50**: 1091-1105.
- Micklich N. 1985.** Biologisch-paläontologische Untersuchungen zur Fischfauna der Messeler Ölschiefer (Mittel-Eozän, Lutetium). *Andrias* **4**: 1-171.
- Miralles A, Marin J, Markus D, Herrel A, Hedges SB, Vidal N. 2018.** Molecular evidence for the paraphyly of Scolecophidia and its evolutionary implications. *Journal of Evolutionary Biology* **31**: 1782-1793.
- Morrison ML, Rodewald AD, Voelker G, Colón MR, Prather JF. 2018.** *Ornithology*. Baltimore: Johns Hopkins University Press.
- Moyle PB, Cech JJ. 2003.** *Fishes: An Introduction to Ichthyology*. Upper Saddle River: Prentice-Hall.
- Müller J, Hipsley CA, Head JJ, Kardjilov N, Hilger A, Wuttke M, Reisz RR. 2011.** Eocene lizard from Germany reveals amphisbaenian origins. *Nature* **473**: 364.
- Nelson JS. 1989.** Analysis of the multiple occurrence of pelvic fin absence in extant fishes. *Matsya* **15**: 21-38.
- O'Reilly JC, Ritter DA, Carrier DR. 1997.** Hydrostatic locomotion in a limbless tetrapod. *Nature* **386**: 269.
- Pagel M, Meade A. 2006.** Bayesian analysis of correlated evolution of discrete characters by reversible-jump Markov chain Monte Carlo. *The American Naturalist* **167**: 808-825.
- Palci A, Hutchinson MN, Caldwell MW, Scanlon JD, Lee MSY. 2018.** Palaeoecological inferences for the fossil Australian snakes *Yurlunggur* and *Wonambi* (Serpentes, Madtsoiidae). *Royal Society open science* **5**: 172012.

- Pardo JD, Mann A. 2018.** A basal aïstopod from the earliest Pennsylvanian of Canada and the antiquity of the first limbless tetrapod lineage. *Royal Society Open Science* **5**: 181096.
- Pfaff C, Zorzin R, Kriwet J. 2016.** Evolution of the locomotory system in eels (Teleostei: Elopomorpha). *BMC Evolutionary biology* **16**: 159.
- Pyron RA, Wiens JJ. 2011.** A large-scale phylogeny of Amphibia including over 2800 species, and a revised classification of extant frogs, salamanders, and caecilians. *Molecular Phylogenetics and Evolution* **61**: 543-583.
- Pyron RA, Burbrink FT, Wiens JJ. 2013.** A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC evolutionary biology* **13**: 93.
- Pough FH, Andrews RM, Crump ML, Savitzky AH, Wells KD, Brandley MC. 2015.** *Herpetology*. Oxford: Oxford University Press.
- Raynaud A. 1990.** Developmental mechanism involved in the embryonic reduction of limb in reptiles. *International Journal of Developmental Biology* **34**: 233-243
- Reece JS, Mehta RS. 2013.** Evolutionary history of elongation and maximum body length in moray eels (Anguilliformes: Muraenidae). *Biological Journal of the Linnean Society* **109**: 861-875.
- Reece JS, Smith DG, Holm E. 2010.** The moray eels of the *Anarchias cantonensis* group (Anguilliformes: Muraenidae), with description of two new species. *Copeia* **2010**: 421-430.
- Reeder TW, Townsend TM, Mulcahy DG, Noonan BP, Wood PL, Sites JW, Wiens JJ. 2015.** Integrated analyses resolve conflicts over squamate reptile phylogeny and reveal unexpected placements for fossil taxa. *PLoS ONE* **10**: e0118199.
- Riede K. 2004.** *Global register of migratory species - from global to regional scales*. Final Report of the R&D-Projekt 808 05 081. Federal Agency for Nature Conservation, Bonn, Germany.
- Robins CR, Bailey RM, Bond CE, Brooker JR, Lachner EA, Lea RN, Scott WB. 1991.** Common and scientific names of fishes from the United States and Canada. *American Fishery Society Special Publication* **20**: 1-183.
- Rosa-Molinar E, Burke AC. 2002.** Starting from fins: parallelism in the evolution of limbs and genitalia: the fin-to-genitalia transition. *Evolution & Development* **4**: 124–126.

- Santini F, Kong X, Sorenson L, Carnevale G, Mehta RS, Alfaro ME. 2013.** A multi-locus molecular timescale for the origin and diversification of eels (Order: Anguilliformes). *Molecular Phylogenetics and Evolution* **69**: 884-894.
- Schaefer SA, Provenzano F, Pinna MD, Baskin JN. 2005.** New and noteworthy Venezuelan glanapterygine catfishes (Siluriformes, Trichomycteridae), with discussion of their biogeography and psammophily. *American Museum Novitates* **3496**: 1-27.
- Sfakiotakis M, Lane DM, Davies JBC. 1999.** Review of fish swimming modes for aquatic locomotion. *IEEE Journal of Oceanic Engineering* **24**: 237-252.
- Shapiro MD, Hanken J, Rosenthal N. 2003.** Developmental basis of evolutionary digit loss in the Australian lizard *Hemiergis*. *Journal of Experimental Zoology Part B Molecular and Developmental Evolution* **297**: 48-56
- Shapiro MD, Marks ME, Peichel CL, Blackman BK, Nereng KS, Jónsson B, Schluter D, Kingsley DM. 2004.** Genetic and developmental basis of evolutionary pelvic reduction in threespine sticklebacks. *Nature* **428**: 717.
- Skinner A, Lee MS, Hutchinson MN. 2008.** Rapid and repeated limb loss in a clade of scincid lizards. *BMC Evolutionary Biology* **8**: 310.
- Smith DG. 1981.** Congridae. In: Fischer W, Bianchi G, Scott WB (Eds.) *FAO species identification sheets for fishery purposes. Eastern Central Atlantic; fishing areas 34, 47 (in part)*. Department of Fisheries and Oceans Canada and FAO, vol. 2.
- Suzuki D, Brandley MC, Tokita M. 2010.** The mitochondrial phylogeny of an ancient lineage of ray-finned fishes (Polypteridae) with implications for the evolution of body elongation, pelvic fin loss, and craniofacial morphology in Osteichthyes. *BMC evolutionary biology* **10**: 21.
- Talanda M. 2016.** Cretaceous roots of the amphisbaenian lizards. *Zoologica Scripta* **45**: 1-8.
- Tanaka M, Hale LA, Amores A, Yan YL, Cresko WA, Suzuki T, Postlethwait JH. 2005.** Developmental genetic basis for the evolution of pelvic fin loss in the pufferfish *Takifugu rubripes*. *Developmental biology* **281**: 227-239.

- Thacker CE. 2013.** Phylogenetic placement of the European sand gobies in Gobionellidae and characterization of gobionellid lineages (Gobiiformes: Gobioidae). *Zootaxa* **3619**: 369-382.
- Thewissen JGM, Cohn MJ, Stevens LS, Bajpai S, Heyning J, Horton WE. 2006.** Developmental basis for hind-limb loss in dolphins and origin of the cetacean bodyplan. *Proceedings of the National Academy of Sciences* **103**: 8414-8418.
- Vaughan TA, Ryan JM, Czaplewski NJ. 2011.** Mammalogy. 5th ed. Sudbury, Massachusetts: Jones and Bartlett Publishers.
- Villa-Verde L, Costa W. 2006.** A new glanapterygine catfish of the genus *Listrura* (Siluriformes: Trichomycteridae) from the southeastern Brazilian coastal plains. *Zootaxa* **1142**: 43-50.
- Vreven EJ. 2005.** Mastacembelidae (Teleostei; Synbranchiformes) subfamily division and African generic division: an evaluation. *Journal of Natural History* **39**: 351-370.
- Wall M, Shine R. 2013.** Ecology and behaviour of Burton's Legless Lizard (*Lialis burtonis*, Pygopodidae) in tropical Australia. *Asian Herpetological Research* **4**: 9-21.
- Ward AB, Mehta RS. 2010.** Axial elongation in fishes: using morphological approaches to elucidate developmental mechanisms in studying body shape. *Integrative and Comparative Biology* **50**: 1106-1119.
- Wiens JJ, Brandley MC, Reeder TW. 2006.** Why does a trait evolve multiple times within a clade? Repeated evolution of snake-like body form in squamate reptiles. *Evolution* **60**: 123-141.
- Winckler TC. 1861.** *Description de quelques nouvelles espèces de poissons fossiles des calcaires d'eau douce d'Oeningen* (Vol. 1). Loosjes: Mémoire couronné par la Société Hollandaise des Sciences à Harlem.
- Wright JJ. 2017.** A new diminutive genus and species of catfish from Lake Tanganyika (Siluriformes: Clariidae). *Journal of Fish biology* **91**: 789-805.
- Yamada T, Sugiyama T, Tamaki N, Kawakita A, Kato M. 2009.** Adaptive radiation of gobies in the interstitial habitats of gravel beaches accompanied by body elongation and excessive vertebral segmentation. *BMC Evolutionary Biology* **9**: 145.



## Figure and tables captions

**Figure 1.** Composite phylogenetic tree of teleost fishes used for the analysis. See material and methods for the references concerning phylogenetic position and character state reconstruction. The coloured branches in the figure are just graphical representations and they do not represent any ancestral state reconstruction, as it is not a necessary step for the discrete comparative analysis using BayesTraitsV3.

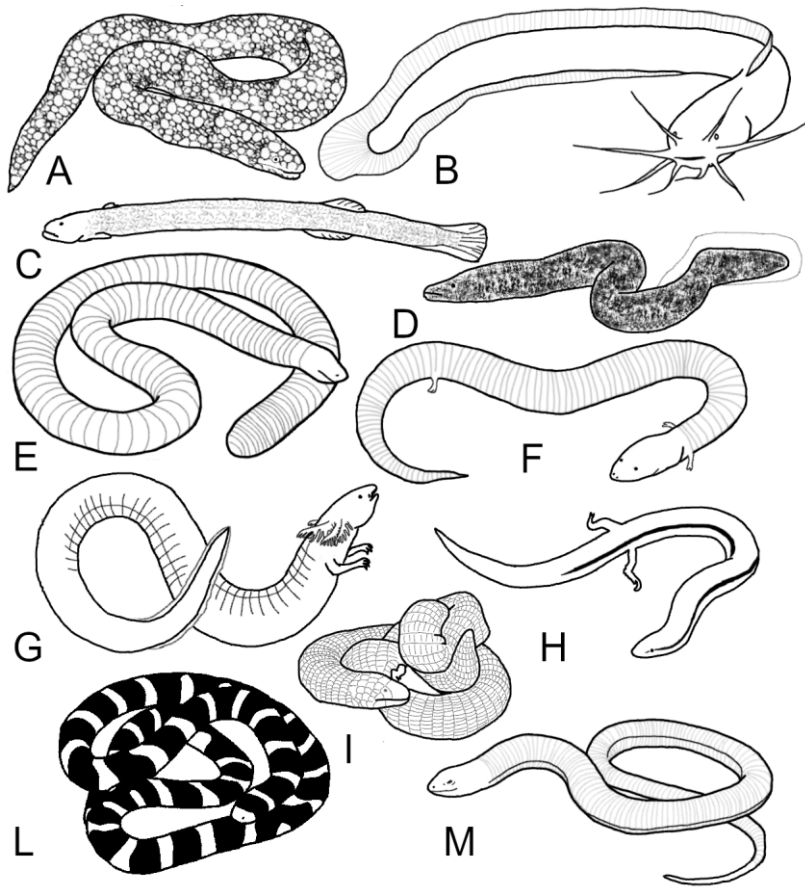
**Figure 2.** Composite phylogenetic tree of lissamphibians and squamates used for the analysis. See material and methods for the references concerning phylogenetic position and character state reconstruction. The coloured branches in the figure are just graphical representations and they do not represent any ancestral state reconstruction, as it is not a necessary step for the discrete comparative analysis using BayesTraitsV3.

**Figure 3.** Examples of elongated vertebrate with fin or limb reduction or loss. A, *Anarchias seychellensis*, crevice-dweller (fish); B, *Channalabes apus*, living among tree roots (fish); C, *Luciogobius elongatus*, interstitial (fish); D, *Mastacembelus apectoralis*, living among sea vegetation (fish); E, *Caecilia volceni*, burrower (amphibian); F, *Amphiuma means*, burrower (amphibian); G, *Siren lacertina*, burrower (amphibian); H, *Lerista bipes*, surface-dweller (reptile); I, *Blanus cinereus*, burrower (reptile); L, *Anilius scytale*, burrower (reptile); M, *Pseudopus apodus*, grass-swimmer (reptile).

**Table 1.** Values of Log of marginal likelihood of the dependent and independent models, and Bayes Factor (BF) obtained using BayesTraitsV3. In both teleost fishes and lissamphibians and reptiles the logarithmic values of the Bayes Factor indicate a strong evidence of correlation.







**Appendix 1.** Teleost fishes database. Grey columns are the two columns used for the discrete analysis.

Taxon	Group	Elongation	Anguilliform locomotion	Pectoral fins absent	Fossorial/complex habitat	Notes	Anguilliform + fossoriality	Data's reference	Phylogeny's reference
<i>Megalops cyprinoides</i>	Elopomorpha	0	0	0	0	reef-associated	0	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Albula vulpes</i>	Elopomorpha	0	0	0	0	reef-associated	0	Riede, 2004	Santini <i>et al.</i> , 2013
† <i>Anguillavus</i>	Elopomorpha	1	1	0	0	reef	0	Belouze, 2002	Pfaff <i>et al.</i> , 2016
† <i>Abisaadia</i>	Elopomorpha	1	1	0	0	reef	0	Belouze <i>et al.</i> , 2003 a	Pfaff <i>et al.</i> , 2016
† <i>Luenchelys</i>	Elopomorpha	1	1	0	0	reef	0	Belouze <i>et al.</i> , 2003 b	Pfaff <i>et al.</i> , 2016
† <i>Hayenchelys</i>	Elopomorpha	1	1	0	0	reef	0	Belouze <i>et al.</i> , 2003 a	Pfaff <i>et al.</i> , 2016
† <i>Urenchelys</i>	Elopomorpha	1	1	0	0	reef	0	Belouze, 2002	Pfaff <i>et al.</i> , 2016
† <i>Libanechelyidae</i>	Elopomorpha	1	1	0	0	reef	0	Taverne, 2004	Pfaff <i>et al.</i> , 2016

<i>Protoanguilla palau</i>	Anguilliformes	1	1	01	1	cave, reef associated	1	Belouze <i>et al.</i> , 2003 a	Santini <i>et al.</i> , 2013
<i>Simenchelys parasitica</i>	Synbranchidae Anguilliformes	1	1	1	0	parasitic	0	Shcherbachev, 1997	Santini <i>et al.</i> , 2013
† <i>Anguilloides</i>	Anguilliformes	1	1	0	0	reef burrower, head-first; small, vestigial	0	Blot, 1984	Pfaff <i>et al.</i> , 2016
<i>Moringua edwardsi</i>	Moringuidae	1	1	1	1	pectoral fins burrower in sand; small, vestigial	1	Smith, 1997	Santini <i>et al.</i> , 2013
<i>Moringua microchir</i>	Moringuidae	1	1	1	1	pectoral fins	1	Castle, 1986	Santini <i>et al.</i> , 2013
<i>Moringua javanica</i>	Moringuidae	1	1	1	1	reef	1	Steene, 1988	Santini <i>et al.</i> , 2013
† <i>Anguilla ignota</i>	Anguillidae	1	1	0	0	freshwater	0	Micklich, 1985	Pfaff <i>et al.</i> , 2016
† <i>Anguilla multiradiata</i>	Anguillidae	1	1	0	0	freshwater	0	Micklich, 1985	Pfaff <i>et al.</i> , 2016
† <i>Anguilla elegans</i>	Anguillidae	1	1	0	0	freshwater	0	Winckler, 1861	Pfaff <i>et al.</i> , 2016
<i>Anguilla mossambica</i>	Anguillidae	1	1	0	0	demersal under rocks in flowstreams	0	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Anguilla malgumora</i>	Anguillidae	1	1	0	0	demersal	0	Martin-Smith & Tan, 1998	Santini <i>et al.</i> , 2013
<i>Anguilla anguilla</i>	Anguillidae	1	1	0	0	demersal	0	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Anguilla australis</i>	Anguillidae	1	1	0	0	demersal	0	McDowall & Beumer, 1980	Santini <i>et al.</i> , 2013
<i>Anguilla reinhardtii</i>	Anguillidae	1	1	0	0	demersal	0	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Anguilla japonica</i>	Anguillidae	1	1	0	0	demersal	0	Yamada <i>et al.</i> , 1995	Santini <i>et al.</i> , 2013
<i>Anguilla marmorata</i>	Anguillidae	1	1	0	0	demersal	0	Rainboth, 1996	Santini <i>et al.</i> , 2013
<i>Anguilla</i>	Anguillidae	1	1	0	0	demersal	0	Allen <i>et al.</i>	Santini <i>et al.</i>

<i>obscura</i>					al		<i>al.</i> , 2002	<i>al.</i> , 2013
<i>Anguilla interioris</i>	Anguillidae	1	1	0	0 al demers	0	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Anguilla nebulosa</i>	Anguillidae	1	1	0	0 al demers	0	Riede, 2004	Santini <i>et al.</i> , 2013
† <i>Voltaconger</i>	Congroidea	1	1	0	0 ?	0	Blot, 1984	Pfaff <i>et al.</i> , 2016
† <i>Bolcyrus</i>	Congroidea	1	1	0	0 ?	0	Blot, 1984	<i>al.</i> , 2016
<i>Derichthys serpentinus</i>	Derichthyidae	1	1	0	0 mesopelagic	0	Mundy, 2005	Santini <i>et al.</i> , 2013
<i>Nettastoma parviceps</i>	Congridae	1	1	0	0 abyssal	0	Smith <i>et al.</i> , 1981	Santini <i>et al.</i> , 2013
<i>Conger cinereus</i>	Congridae	1	1	0	0 reef	0	Myers, 1991	Santini <i>et al.</i> , 2013
<i>Venefica procera</i>	Congridae	1	1	0	0 abyssal	0	McEachron & Fechhelm, 1998	Santini <i>et al.</i> , 2013
<i>Rhynchoconger gracilior</i>	Congridae	1	1	0	0 abyssal	0	Robins & Ray, 1986	Santini <i>et al.</i> , 2013
<i>Facciolella equatorialis</i>	Congridae	1	1	0	0 abyssal	0	Love <i>et al.</i> , 2005	Santini <i>et al.</i> , 2013
<i>Hoplunnis punctata</i>	Congridae	1	1	0	0 abyssal burrow	0	Smith, 1990	Santini <i>et al.</i> , 2013
<i>Ariosoma balearicus</i>	Congridae	1	1	01	1 reef burrow	1	Riede, 2004	Santini <i>et al.</i> , 2013
<i>Heteroconger hassi</i>	Congridae	1	1	1	1 reef burrow	1	Bacchet <i>et al.</i> , 2006	Santini <i>et al.</i> , 2013
<i>Paraconger notialis</i>	Congridae	1	1	01	1 reef burrow, tail-first	1	Smith, 1981	Santini <i>et al.</i> , 2013
<i>Scolecenchelys australis</i>	Ophichthidae	1	1	1	1 seagrass	1	Hibino & Kimura, 2015	Santini <i>et al.</i> , 2013
<i>Ahlia egmontis</i>	Ophichthidae	1	1	1	1 seagrass and crevices benthic but not burrow	1	Robins & Ray, 1986	Santini <i>et al.</i> , 2013
<i>Myrophis vafer</i>	Ophichthidae	1	1	01	0 reef	0	Charter & Moser, 1996	Santini <i>et al.</i> , 2013
<i>Ophichthus triserialis</i>	Ophichthidae	1	1	01	0 reef burrow	1	McCosker & Rosenblatt, 1998	Santini <i>et al.</i> , 2013
<i>Ophichthus cruentifer</i>	Ophichthidae	1	1	01	0 offshore water	0	Robins & Ray, 1986	Santini <i>et al.</i> , 2013
<i>Kaupichthys hyoproroides</i>	Anguilliformes	1	1	01	0 reef	01	Claro, 1994	Santini <i>et al.</i> , 2013
† <i>Paranguilla</i>	Anguilliformes	1	1	01	0 reef burrow	0	Blot, 1984	<i>al.</i> , 2016
<i>Pythonichthys asodes</i>	Heterenchelidae	1	1	1	1 reef in	1	Rosenblatt &	Santini <i>et al.</i> , 2013



					pectoral fins			
					intertidal rocky shore			
<i>Inu koma</i>	Gobionellidae	1	1	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					intertidal rocky shore			
<i>Inu saikaiensis</i>	Gobionellidae	1	1	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					intertidal rocky shore, intertidal			
<i>Gen. Sp. 1</i>	Gobionellidae	1	1	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					intertidal rocky shore, subtidal			
<i>Clariger cosmurus</i>	Gobionellidae	1	1	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					rocky shore subtidal			
<i>Clariger exilis</i>	Gobionellidae	1	1	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					rocky shore subtidal			
<i>Astrabe flavimaculata</i>	Gobionellidae	0	0	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					rocky shore subtidal			
<i>Chaenogobius gulosus</i>	Gobionellidae	0	0	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					freshwater lake			
<i>Gymnogobius isaza</i>	Gobionellidae	0	0	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					freshwater lake			
<i>Gymnogobius urotaenia</i>	Gobionellidae	0	0	0	0	0	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					mudshrimp burrow			
<i>Typhlogobius californiensis</i>	Gobionellidae	0	0	0	0	1	Yamada <i>et al.</i> , 2009	Yamada <i>et al.</i> , 2009
					mudshrimp burrow			
<i>Eutaeniichthys gilli</i>	Gobionellidae	1	1	1	1	1	Henmi & Itani, 2014	Henmi & Itani, 2014
					reef, amphibious mud, amphibious			
<i>Stenogobius</i>	Gobionellidae	0	0	0	0	0	Watson, 1991	Thacker, 2013
					reef, amphibious mud, amphibious			
<i>Periophtalmus barbarus</i>	Gobionellidae	1	0	0	1	0	Miller, 1981	Thacker, 2013
					reef, amphibious mud, amphibious			
<i>Scartelaos histiophorus</i>	Gobionellidae	1	0	0	1	0	Riede, 2004	Thacker, 2013
					reef, amphibious mud, amphibious			
<i>Gobiopterus semivestitus</i>	Gobionellidae	0	0	0	0	0	Thacker, 2013	Thacker, 2013
					pelagic			
<i>Pandaka lidwilli</i>	Gobionellidae	0	0	0	0	0	Masuda <i>et al.</i> , 1984	Thacker, 2013
					reef			
<i>Barbulifer ceuthoecus</i>	Gobiidae	0	0	0	1	0	Cervigón, 1994	Thacker, 2013
					reef			
<i>Amblygobius phalena</i>	Gobiidae	0	0	0	1	0	Randall, 1995	Thacker, 2013
<i>Copionodon</i>	Copionodont	1	0	0	0	0	Zanata &	Datovo &



<i>pecten</i>	inae							Primitivo, 2014	Lindam, 2005
<i>Trichogenes longipinnis</i>	Trichogeninae	1	0	0	0			0 Britski & Ortega, 1982; Sazima, 2004	Datovo & Lindam, 2005
<i>Bullockia maldonadoi</i>	Trichomycterinae	1	0	0	0			0 Bockman & Sazima, 2004	Datovo & Lindam, 2005
<i>Hatcheria macraei</i>	Trichomycterinae	1	0	0	0			0 Bockman & Sazima, 2004	Datovo & Lindam, 2005
<i>Ituglanis proops</i>	Trichomycterinae	1	1	0	0			01 Bockman & Sazima, 2004	Datovo & Lindam, 2005
<i>Trichomycterus maracaya</i>	Trichomycterinae	1	0	0	0			0 Bockman & Sazima, 2004	Datovo & Lindam, 2005
<i>Listrura tetraradiata</i>	Glanapteryginae	1	1	0	1	interstitial		1 Villa-Verde & Costa, 2006	Datovo & Lindam, 2005; Villa-Verde <i>et al.</i> , 2012
<i>Listrura picinguabae</i>	Glanapteryginae	1	1	1	1	interstitial, filiform fins		1 Villa-Verde & Costa, 2006	Baskin, 1973; Datovo & Lindam, 2005; Villa-Verde <i>et al.</i> , 2012
<i>Listrura nematopteryx</i>	Glanapteryginae	1	1	1	1	interstitial, filiform fins		1 De Pinna, 1988; Villa-Verde & Costa, 2006	Datovo & Lindam, 2005; Villa-Verde <i>et al.</i> , 2012
<i>Listrura costai</i>	Glanapteryginae	1	1	1	1	interstitial, filiform fins		1 Villa-Verde & Costa, 2006; Villa-Verde <i>et al.</i> , 2012	Baskin, 1973; Datovo & Lindam, 2005; Villa-Verde <i>et al.</i>

<i>Glanapteryx anguilla</i>	Glanapteryginae	1	1	1	1	leaf litter	1	De Pinna, 1988; Schaefer <i>et al.</i> , 2005	Baskin, 1973; Datovo & Lindam, 2005
<i>Pygidianops cuao</i>	Glanapteryginae	1	1	1	1	small, filiform pectoral fins	1	Schaefer <i>et al.</i> , 2005	Baskin, 1973; Datovo & Lindam, 2005
<i>Pygidianops magoi</i>	Glanapteryginae	1	1	1	1	sand-bottom	1	Schaefer <i>et al.</i> , 2005	Baskin, 1973; Datovo & Lindam, 2005
<i>Typhlobelus hundbergi</i>	Glanapteryginae	1	1	1	1	sand-bottom	1	Schaefer <i>et al.</i> , 2005	Baskin, 1973; Datovo & Lindam, 2005
<i>Typhlobelus guacamaya</i>	Glanapteryginae	1	1	1	1	sand-bottom	1	Schaefer <i>et al.</i> , 2005	Baskin, 1973; Datovo & Lindam, 2005
<i>Stauroglanis gouldingi</i>	Sarcoglanidinae	0	0	0	0	rheophilic	0	Zuanon & Sazima, 2004	Baskin, 1973; Datovo & Lindam, 2005
<i>Paracanthopoma parva</i>	Vandelliinae	1	1	0	0	parasitic	0	Schmidt, 1993; Zuanon & Sazima, 2005	Baskin, 1973; DoNascimento, 2015
<i>Vandellia sanguinea</i>	Vandelliinae	1	1	0	0	blood parasite	0	Baskin <i>et al.</i> , 1980	Baskin, 1973; DoNascimento, 2015
<i>Vandellia cirrhosa</i>	Vandelliinae	1	1	0	0	blood parasite	0	Baskin <i>et al.</i> , 1980	Baskin, 1973; DoNascimento, 2015
<i>Haemomaster venezuelae</i>	Stegophilinae	1	1	0	0	parasitic	0	DoNascimento, 2015	DoNascimento, 2015
<i>Ochmacanthus flabelliferus</i>	Stegophilinae	1	1	0	0	parasitic	0	DoNascimento, 2015	DoNascimento, 2015

<i>Homodiaetus anisitsi</i>	Stegophilinae	1	1	0	0	parasitic	0	DoNascimento, 2015	DoNascimento, 2015
<i>Henonemus</i>	Stegophilinae	1	1	0	0	parasitic	0	DoNascimento, 2015	DoNascimento, 2015
<i>Pareiodon Clarotes laticeps</i>	Stegophilinae	1	1	0	0	parasitic	0	DoNascimento, 2015	DoNascimento, 2015
<i>Heteropneustes fossilis</i>	Siluroidea	0	0	0	0		0	Risch, 2003	Wright, 2017
<i>Clarias meladerma</i>	Siluroidea	1	0	0	0		0	Ali <i>et al.</i> , 2015	Wright, 2017
<i>Gymnallabes typus</i>	Clariidae	1	01	0	0		0	Vidhayanon, 2002	Wright, 2017
<i>Tanganikallabes mortiauxi</i>	Clariidae	1	1	0	0		0	Teugels, 2003;	Wright, 2017
<i>Tanganikallabes alboperca</i>	Clariidae	1	1	0	0		0	Ward <i>et al.</i> , 2015	Wright, 2017
<i>Tanganikallabes stewarti</i>	Clariidae	1	1	0	0		0	Seegers, 2008	Wright, 2017
<i>Channallabes apus</i>	Clariidae	1	1	1	1	among trees roots, testa piccola rispetto agli altri	1	Seegers, 2008	Wright, 2017
<i>Clarias agboyiensis</i>	Clariidae	1	1	0	1	grasslands shallow s in vegetated areas high variability in fins, but often very small	1	Ezenwaji & Inyang, 1998	Wright, 2017
<i>Clarias theodora</i>	Clariidae	1	1	0	1		1	Teugels, 1986	Wright, 2017
<i>Dolichallabes microphthalmus</i>	Clariidae	1	1	1	1		1	Devaere <i>et al.</i> , 2004;	Wright, 2017
<i>Heterobranchius isopterus</i>	Clariidae	1	01	0	0		0	Seegers, 2008	Wright, 2017
<i>Heterobranchius Boulengeri</i>	Clariidae	1	01	0	0		0	Teugels <i>et al.</i> , 1990	Wright, 2017
<i>Heterobranchius bidorsalis</i>	Clariidae	1	01	0	0		0	Teugels <i>et al.</i> , 1990	Wright, 2017
<i>Dinotopterus</i>	Clariidae	1	01	0	0	on	0	Poll, 1953	Wright,

<i>cunningtoni</i>						rocky substrates permanent swamps burrower rivers		2017
<i>Clarias ngamensis</i>	Clariidae	1	1	0	0		0	Teugels, 1986; Wright, 2017
<i>Clarias anguillaris</i>	Clariidae	1	1	0	1		1	Teugels, 1986; Wright, 2017
<i>Clarias gariepinus</i>	Clariidae	1	1	0	0		0	Teugels, 1986; Wright, 2017
<i>Pseudotangani kallabes</i>	Clariidae	1	01	0	0		0	Wright, 2017; Wright, 2017
<i>Clarias platycephalus</i>	Clariidae	1	1	0	0		0	Teugels, 1986; Wright, 2017
<i>Channallabes longicaudatus</i>	Clariidae	1	1	0	0		0	Devaere <i>et al.</i> , 2007; Wright, 2017
<i>Channallabes alvarezi</i>	Clariidae	1	1	0	0		0	Devaere <i>et al.</i> , 2007; Wright, 2017
<i>Clariallabes longicauda</i>	Clariidae	1	1	0	0		0	Teugels, 1986; Wright, 2017

**Appendix 2.** Database of squamate reptiles and lissamphibians. Grey columns are the two columns used for the discrete analysis.

Taxon	Group	Absence forelimb	Absence hindlimb	Absence limb	Burrowing or grass-swimmer	Notes	Data's reference	Phylogeny's reference
<i>Anelytropsis papillosus</i>	Squamata, Dibamidae	1	1	1	1	burrower	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Dibamus novaeguineae</i>	Squamata, Dibamidae	1	1	1	1	burrower	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Nephrurus vertebralis</i>	Gekkota, Carphodactylidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Underwoodisaurus seorsus</i>	Gekkota, Carphodactylidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Lialis burtonis</i>	Gekkota, Pygopodidae	1	1	1	0	litter	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Delma labialis</i>	Gekkota, Pygopodidae	1	1	1	0	litter	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015

<i>Pygopus nigriceps</i>	Gekkota, Pygopodidae	1	1	1	0	generalist generalist (burrowing, according to IUCN Red List)	2009; Uetz <i>et al.</i> , 2019 IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Aprasia aurita</i>	Gekkota, Pygopodidae	1	1	1	0		IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Rhacodactylus leachianus</i>	Gekkota, Diplodactylidae	0	0	0	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Coleonyx variegatus</i>	Gekkota, Eublepharidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Eublepharis hardwickii</i>	Gekkota, Eublepharidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Hemitheconyx caudicinctus</i>	Gekkota, Eublepharidae	0	0	0	0	savannah	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Hemidactylus turcicus</i>	Gekkota, Gekkonidae	0	0	0	0	generalist	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Mediodactylus kotschyi</i>	Gekkota, Gekkonidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Phelsuma lineata</i>	Gekkota, Gekkonidae	0	0	0	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Gonatodes albogularis</i>	Gekkota, Sphaerodactylidae	0	0	0	0	arboreal, generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Euleptes europaea</i>	Gekkota, Sphaerodactylidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Pristurus rupestris</i>	Gekkota, Sphaerodactylidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Sphaerodactylus schwartzi</i>	Gekkota, Sphaerodactylidae	0	0	0	0	litter	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015

	dae						<i>et al.</i> , 2019
<i>Asaccus platyrhynchus</i>	Gekkota, Phyllodactylidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019 Talanda, 2018
<i>Tarentola mauritanica</i>	Gekkota, Phyllodactylidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019 Talanda, 2018
<i>Phyllodactylus reissii</i>	Gekkota, Phyllodactylidae	0	0	0	0	arid areas	IUCN Red List; Uetz <i>et al.</i> , 2019 Talanda, 2018
† <i>Ardeosaurus brevipes</i>	Scincoidea	0	0	0	0	generalist	Talanda, 2018
† <i>Yabeinosaurus robustus</i>	Scincoidea	0	0	0	0	generalist	Talanda, 2018 Talanda, 2018
<i>Acontias meleagris</i>	Squamata, Scincidae, Acontinae	1	1	1	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Acontias gariensis</i>	Squamata, Scincidae, Acontinae	1	1	1	-	?	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Acontias lineatus</i>	Squamata, Scincidae, Acontinae	1	1	1	-	?	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Typhlosaurus braini</i>	Squamata, Scincidae, Acontinae	1	1	1	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Typhlosaurus caecus</i>	Squamata, Scincidae, Acontinae	1	1	1	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Typhlosaurus lomiae</i>	Squamata, Scincidae, Acontinae	1	1	1	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Egernia kingii</i>	Squamata, Scincidae, Egerniinae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Tiliqua scincoides</i>	Squamata, Scincidae, Egerniinae	0	0	0	0	generalist coastal	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Cyclodomorphus michaeli</i>	Squamata, Scincidae, Egerniinae	0	0	0	0	environment	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Liopholis whitii</i>	Squamata, Scincidae, Egerniinae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Ablepharus kitaibelii</i>	Squamata, Scincidae, Eugongylinae	0	0	0	0	arid areas	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019
<i>Ablepharus budaki</i>	Squamata, Scincidae, Eugongylinae	0	0	0	0	litter	IUCN Red List; Uetz <i>et al.</i> , 2019 IUCN Red List; Uetz <i>et al.</i> , 2019

<i>Feylinia currori</i>	Squamata, Scincidae, Scincinae	1	1	1	01	semi fossorial	IUCN Red List; Wagner & Schmitz, 2006; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Chalcides ocellatus</i>	Squamata, Scincidae, Scincinae	0	0	0	0	generalist	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Chalcides chalcides</i>	Squamata, Scincidae, Scincinae	0	0	0	0	grassy areas	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Melanoseps ater</i>	Squamata, Scincidae, Scincinae	1	1	1	0	in litter	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Mesoscincus schwartzei</i>	Squamata, Scincidae, Scincinae	0	0	0	0	generalist	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Mesoscincus managuae</i>	Squamata, Scincidae, Scincinae	0	0	0	0	generalist	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Ophiomorus latastii</i>	Squamata, Scincidae, Scincinae	1	1	1	1	burrowin g	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Ophiomorus punctatissimus</i>	Squamata, Scincidae, Scincinae	1	1	1	1	burrowin g	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Brachymeles apus</i>	Squamata, Scincidae, Scincinae	1	1	1	1	burrowin g	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Brachymeles miriamae</i>	Squamata, Scincidae, Scincinae	1	1	1	1	burrowin g	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Brachymeles talinis</i>	Squamata, Scincidae, Scincinae	0	0	0	-	?	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Plestiodon multilineatus</i>	Squamata, Scincidae, Scincinae	0	0	0	0	generalist	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Scincus mitranus</i>	Squamata, Scincidae, Scincinae	0	0	0	0	sandy dunes	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Hakaria simonyi</i>	Squamata, Scincidae, Scincinae	0	0	0	0	generalist	Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015

<i>Scelotes caffer</i>	Squamata, Scincidae, Scincinae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Scelotes bipes</i>	Squamata, Scincidae, Scincinae	1	0	0	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Ameiva bifrontata</i>	Squamata, Teiidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Teius teyou</i>	Squamata, Teiidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Pholidobolus montium</i>	Squamata, Gymnophthal midae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Rhineura floridana</i>	Amphisbaenia, Rhineuridae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Blanus cinereus</i>	Amphisbaenia, Blanidae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Cadea blanoides</i>	Amphisbaenia, Cadeidae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Bipes biporus</i>	Amphisbaenia, Bipedidae	0	1	0	1	burrowin g	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Trogonophis wiegmanni</i>	Amphisbaenia, Trogonophida e	1	1	1	1	burrowin g	IUCN Red List; Vitt & Caldwell, 2009; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>† Slavoia darevskii</i>	Amphisbaenia	0	0	0	0	burrowin g	Talanda, 2016	Talanda, 2016
<i>Amphisbaena alba</i>	Amphisbaenia, Amphisbaenid ae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Chirindia langi</i>	Amphisbaenia, Amphisbaenid ae	1	1	1	1	burrowin g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Anomalepis mexicanus</i>	Serpentes, Anomalepidid ae	1	1	1	0	under trunk	IUCN Red List; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Leptotyphlops</i>	Serpentes, Leptotyphlopi dae	1	1	1	1	burrowin g	IUCN Red List	Da Silva <i>et al.</i> , 2018
<i>Indotyphlops braminus</i>	Serpentes, Typhlopidae	1	1	1	1	burrowin g	Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Xerotyphlops</i>	Serpentes,	1	1	1	1	burrowin	IUCN Red	Da Silva <i>et</i>



<i>vermicularis</i>	Typhlopidae					g	List; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
† <i>Dinlysia</i>	Serpentes	1	1	1	1	g	Da Silva <i>et al.</i> , 2018	Da Silva <i>et al.</i> , 2018
† <i>Wonambi</i>	Serpentes	1	1	1	-	generalist ?	Palci <i>et al.</i> , 2018	Da Silva <i>et al.</i> , 2018
† <i>Yurlunggur</i>	Serpentes	1	1	1	1	semifossorial	Palci <i>et al.</i> , 2018	Da Silva <i>et al.</i> , 2018
<i>Anilius scytale</i>	Serpentes, Aniliidae	1	1	1	1	g	Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
† <i>Haasiophis</i>	Serpentes	1	1	1	1	g	Da Silva <i>et al.</i> , 2018	Da Silva <i>et al.</i> , 2018
† <i>Pachyrhachis</i>	Serpentes	1	1	1	1	g	Da Silva <i>et al.</i> , 2018	Da Silva <i>et al.</i> , 2018
<i>Tropidophis melanurus</i>	Serpentes, Tropidophiidae	1	1	1	1	g	Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Casarea dussumieri</i>	Serpentes, Bolyeriidae	1	1	1	0	arboreal	IUCN Red List	Da Silva <i>et al.</i> , 2018
<i>Eryx jaculus</i>	Serpentes, Boidae	1	1	1	1	g	Uetz <i>et al.</i> , 2019	Reynolds <i>et al.</i> , 2014; Da Silva <i>et al.</i> , 2018
<i>Boa constrictor</i>	Serpentes, Boidae	1	1	1	0	generalist	Uetz <i>et al.</i> , 2019	Reynolds <i>et al.</i> , 2014; Da Silva <i>et al.</i> , 2018
<i>Corallus hortulanus</i>	Serpentes, Boidae	1	1	1	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reynolds <i>et al.</i> , 2014; Da Silva <i>et al.</i> , 2018
<i>Eunectes murinus</i>	Serpentes, Boidae	1	1	1	0	shallow water	Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Uropeltis ceylanica</i>	Serpentes, Uropeltidae	1	1	1	1	g	Gower <i>et al.</i> , 2008; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Python sebae</i>	Serpentes, Pythonidae	1	1	1	0	generalist	Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Crotalus horridus</i>	Serpentes, Viperidae	1	1	1	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Wüster <i>et al.</i> , 2008; Da Silva <i>et al.</i> , 2018
<i>Bitis arietans</i>	Serpentes, Viperidae	1	1	1	0	generalist	Uetz <i>et al.</i> , 2019	Wüster <i>et al.</i> , 2008; Da Silva <i>et al.</i> , 2018



<i>Coronella girondica</i>	Serpentes, Colubridae	1	1	1	0	generalist	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Elaphe dione</i>	Serpentes, Colubridae	1	1	1	0	generalist	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Eirenis modestus</i>	Serpentes, Colubridae	1	1	1	0	rocky areas	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Hierophis viridiflavus</i>	Serpentes, Colubridae	1	1	1	0	open habitats	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Natrix natrix</i>	Serpentes, Colubridae	1	1	1	0	humid, open habitats	IUCN Red List; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Virginia valeriae</i>	Serpentes, Colubridae	1	1	1	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Da Silva <i>et al.</i> , 2018
<i>Cordylus cordylus</i>	Squamata, Cordylidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Ouroborus cataphractus</i>	Squamata, Cordylidae	0	0	0	0	rocky areas burrowing and grass-swimming	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Smaug giganteus</i>	Squamata, Cordylidae	0	0	0	1	g	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Cricosaura typica</i>	Squamata, Xantusiidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Xantusia vigilis</i>	Squamata, Xantusiidae	0	0	0	0	arid areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Lacerta viridis</i>	Squamata, Lacertidae	0	0	0	0	generalist	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015

<i>Podarcis muralis</i>	Squamata, Lacertidae	0	0	0	0	generalist	IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Takydromus sexlineatus</i>	Squamata, Lacertidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Shinisaurus crocodilurus</i>	Squamata, Shinisauridae	0	0	0	0	semiaquatic	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Heloderma horridum</i>	Squamata, Helodermatidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Varanus exanthematicus</i>	Squamata, Varanidae	0	0	0	0	agricultural areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Xenosaurus grandis</i>	Squamata, Xenosauridae Anguillidae (Anniellidae secondo Conrad <i>et al.</i> 2011)	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>†Ophisaurus quadrupes †</i>	Conrad <i>et al.</i> 2011)	0	0	0	1	burrowing	Sullivan <i>et al.</i> , 1999	Conrad <i>et al.</i> , 2011
<i>†Apodosaurus minutus</i>	Squamata, Anniellidae	1	1	1	1	burrowing	Gauthier, <i>et al.</i> , 1995	Conrad <i>et al.</i> , 2011
<i>Anniella pulchra Celestus enneagrammus</i>	Squamata, Anniellidae	1	1	1	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Diploglossus bilobatus</i>	Squamata, Diploglossidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Diploglossus pleii</i>	Squamata, Diploglossidae	0	0	0	0	semi-fossorial	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Diploglossus pleii</i>	Squamata, Diploglossidae	0	0	0	0	burrowing, in litter	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Ophiodes intermedius</i>	Squamata, Diploglossidae	1	1*	1	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Ophiodes striatus</i>	Squamata, Diploglossidae	1	1	1	-	?	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Celestus agasepsoides</i>	Squamata, Diploglossidae	01	01	01	1	burrowing	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Celestus haetianus</i>	Squamata, Diploglossidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Ophisaurus</i>	Squamata,	1	1	1	01	semi-	IUCN Red	Reeder <i>et al.</i> ,

<i>ventralis</i>	Anguillidae					fossorial	List; Uetz <i>et al.</i> , 2019 IUCN Red List;	2015
<i>Anguis fragilis</i>	Squamata, Anguillidae	1	1	1	1	burrowing and grass-swimmer	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List; Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Pseudopus apodus</i>	Squamata, Anguillidae	1	1	1	1	grass-swimmer	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Elgaria multicarinata</i>	Squamata, Anguillidae	0	0	0	0	generalist	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Abronia oaxaca</i>	Squamata, Anguillidae	0	0	0	0	arboreal	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Barisia levicollis</i>	Squamata, Anguillidae	0	0	0	0	wood	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Gerrhonotus parvus</i>	Squamata, Anguillidae	0	0	0	0	wood	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Chamaeleo chamaeleon</i>	Squamata, Chamaeleonidae	0	0	0	0	arboreal	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Uromastyx aegyptia</i>	Squamata, Agamidae	0	0	0	0	open habitats	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Agama spinosa</i>	Squamata, Agamidae	0	0	0	0	rocky areas	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Draco cornutus</i>	Squamata, Agamidae	0	0	0	0	arboreal	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Hydrosaurus amboinensis</i>	Squamata, Agamidae	0	0	0	0	semiaquatic	Speybroeck <i>et al.</i> , 2016; Uetz <i>et al.</i> , 2019 IUCN Red List;	Reeder <i>et al.</i> , 2015
<i>Basiliscus basiliscus</i>	Squamata, Iguanidae (s.l.), Corytophanidae	0	0	0	0	semiaquatic	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Anolis proboscis</i>	Squamata, Iguanidae (s.l.), Dactyloidae	0	0	0	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015

<i>Iguana delicatissima</i>	Squamata, Iguanidae (s.l.), Iguanidae (s.s.)	0	0	0	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Phrynosoma asio</i>	Squamata, Iguanidae (s.l.), Phrynosomatidae	0	0	0	0	generalist	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Leiocephalus carinatus</i>	Squamata, Iguanidae (s.l.), Leiocephalidae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Polychrus gutturosus</i>	Squamata, Iguanidae (s.l.), Polychrotidae	0	0	0	0	arboreal	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Enyalioides heterolepis</i>	Squamata, Iguanidae (s.l.), Hoplocercidae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Leiosaurus catamarcensis</i>	Squamata, Iguanidae (s.l.), Leiosauridae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Oplurus saxicola</i>	Squamata, Iguanidae (s.l.), Opluridae	0	0	0	0	rocky areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Liolaemus anomalus</i>	Squamata, Iguanidae (s.l.), Liolaemidae	0	0	0	0	arid, salty areas	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
<i>Tropidurus torquatus</i>	Squamata, Iguanidae (s.l.), Tropiduridae	0	0	0	0	wood	IUCN Red List; Uetz <i>et al.</i> , 2019	Reeder <i>et al.</i> , 2015
† <i>Eocaecilia micropodia</i>	Gymnophiona	0	0	0	1	burrowing	Jenkins <i>et al.</i> , 2007	Evans & Sigogneau-Russell, 2011
<i>Epicrionops marmoratus</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Epicrionops niger</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Rhinatrema bivittatum</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Uraeotyphlus natayani</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Ichthyophis bombayensis</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Ichthyophis tricolor</i>	Gymnophiona	1	1	1	1	burrowing	Amphibia Web, 2019	Pyron & Wiens, 2011

<i>Ichthyophis orthoplicatus</i>	Gymnophiona	1	1	1	1	burrowin g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Ichthyophis glutinosus</i>	Gymnophiona	1	1	1	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Ichthyophis bannanicus</i>	Gymnophiona	1	1	1	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Caecilia tentaculata</i>	Gymnophiona	1	1	1	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Cryptobranchius alleganiensis</i>	Cryptobranchiidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Onychodactylus fischeri</i>	Hynobiidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudobranchius striatus</i>	Sirenidae	0	1	01	1	burrowin g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudobranchius axanthus</i>	Sirenidae	0	1	01	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Siren intermedia</i>	Sirenidae	0	1	01	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Siren lacertina</i>	Sirenidae	0	1	01	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Dicamptodon tenebrosus</i>	Dicamptodontidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Ambystoma cingulatum</i>	Ambystomidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Salamandrina terdigitata</i>	Salamandridae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Chioglossa lusitanica</i>	Salamandridae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Salamandra lanzai</i>	Salamandridae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Proteus anguinus</i>	Proteidae	0	0	0	1	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Necturus lewisi</i>	Proteidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Necturus punctatus</i>	Proteidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Necturus beyeri</i>	Proteidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Necturus alabamensis</i>	Proteidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Necturus maculosus</i>	Proteidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Rhyacotriton kezeri</i>	Rhyacotritonidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Rhyacotriton variegatus</i>	Rhyacotritonidae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Amphiuma tridactylum</i>	Amphiumidae	1	1	1	1	burrowin g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Amphiuma pholeter</i>	Amphiumidae	1	1	1	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Amphiuma means</i>	Amphiumidae	1	1	1	1	g	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Hydromantes</i>	Plethodontinae	0	0	0	0	generalist	Amphibia	Pyron &

<i>shastae</i>								Web, 2019	Wiens, 2011
<i>Ensatina</i>								Amphibia	Pyron &
<i>eschschollzii</i>	Plethodontinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Desmognathus</i>								Amphibia	Pyron &
<i>s carolinensis</i>	Plethodontinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Plethodon</i>								Amphibia	Pyron &
<i>larselli</i>	Plethodontinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Hemidactylium</i>	Hemidactylinae							Amphibia	Pyron &
<i>m scutatatum</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Gyrinophilus</i>								Amphibia	Pyron &
<i>porphyriticus</i>	Spelerpinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Eurycea</i>								Amphibia	Pyron &
<i>multiplicata</i>	Spelerpinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Haideotriton</i>								Amphibia	Pyron &
<i>wallacei</i>	Spelerpinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Euricea</i>								Amphibia	Pyron &
<i>quadridigitata</i>	Spelerpinae	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>wrighti</i>	e	0	0	0	1	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>diabolicus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>gabrieli</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>pacificus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>attenuatus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Batrachoseps</i>	Bolitoglossinae							Amphibia	Pyron &
<i>nigriventris</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Thorius</i>	Bolitoglossinae							Amphibia	Pyron &
<i>minutissimus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Thorius</i>	Bolitoglossinae							Amphibia	Pyron &
<i>trogodytes</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Thorius</i>	Bolitoglossinae							Amphibia	Pyron &
<i>dubitus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Chiropterotriton</i>	Bolitoglossinae							Amphibia	Pyron &
<i>on dimidiatus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Dendrotriton</i>	Bolitoglossinae							Amphibia	Pyron &
<i>rabbi</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Cryptotriton</i>									
<i>alvarezdeltoro</i>	Bolitoglossinae							Amphibia	Pyron &
<i>i</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Nototriton</i>	Bolitoglossinae							Amphibia	Pyron &
<i>brodiei</i>	e	0	0	0	0	litter		Web, 2019	Wiens, 2011
<i>Bradytriton</i>	Bolitoglossinae							Amphibia	Pyron &
<i>silus</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Oedipina</i>	Bolitoglossinae							Amphibia	Pyron &
<i>quadra</i>	e	0	0	0	0	litter		Web, 2019	Wiens, 2011
<i>Oedipina</i>	Bolitoglossinae							Amphibia	Pyron &
<i>carablanca</i>	e	0	0	0	0	generalist		Web, 2019	Wiens, 2011
<i>Oedipina</i>	Bolitoglossinae							Amphibia	Pyron &
<i>elongata</i>	e	0	0	0	0	litter		Web, 2019	Wiens, 2011
<i>Oedipina</i>	Bolitoglossinae							Amphibia	Pyron &
<i>maritima</i>	e	0	0	0	0	litter		Web, 2019	Wiens, 2011



<i>Oedipina alleni</i>	Bolitoglossinae	0	0	0	0	litter	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Oedipina stenopodia</i>	Bolitoglossinae	0	0	0	0	litter	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Oedipina gracilis</i>	Bolitoglossinae	0	0	0	0	litter	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Parvimolge townsendi</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoeurycea a cephalica</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoeurycea a boneti</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoeurycea a gigantea</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Lineatriton lineolus</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoeurycea a firscheini</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoeurycea a obesa</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Pseudoerycea melanomolga</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Bolitoglossa hartwegi</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Bolitoglossa platydactyla</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Bolitoglossa subpalmata</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011
<i>Bolitoglossa alvaradoi</i>	Bolitoglossinae	0	0	0	0	generalist	Amphibia Web, 2019	Pyron & Wiens, 2011

### Appendix 3. References cited in Appendix 1 and 2.

Ali AS, Jawad LA, Saad AA. 2015. Confirmation of the presence of the Indian stinging catfish, *Heteropneustes fossilis* (Bloch, 1794) (Heteropneustidae) in Syrian inland waters. *Journal of Applied Ichthyology* 32(1): 1-3.

Allen GR, Steene RC. 1988. *Fishes of Christmas Island Indian Ocean*. Christmas Island Natural History Association, Christmas Island, Indian Ocean, 6798, Australia. 197 p.

Allen GR, Midgley SH, Allen M. 2002. *Field guide to the freshwater fishes of Australia*. Western Australian Museum, Perth, Western Australia.

AmphibiaWeb, 2019. <<http://amphibiaweb.org>> University of California, Berkeley, CA, USA. Accessed 17 Jan 2019.

Bacchet P, Zysman T, Lefèvre Y. 2006. *Guide des poissons de Tahiti et ses îles*. Tahiti (Polynésie Française): Éditions Au Vent des Îles.

Baskin JN. 1973. Structure and relationships of the Trichomycteridae. PhD thesis, City University of New York.

Baskin JN, Zaret TM, Mago-Leccia F. 1980. Feeding of reportedly parasitic catfishes (Trichomycteridae and Cetopsidae) in the Rio Portuguesa Basin, Venezuela. *Biotropica* 12(3): 182-186.

- Bell CJ, Mead JI, Fay LP. 1995. Neogene history of *Anniella* Gray, 1852 (Squamata, Anniellidae) with comments on postcranial osteology. *Copeia* 3: 719-726.
- Belouze A. 2002. Compréhension morphologique et phylogénétique des taxons actuels et fossils rapportés aux Anguilliformes ("poissons", Téléostéens). *Documents des Laboratoires de Géologie de Lyon* 158: 1-401.
- Belouze A, Gayet M, Atallah C. 2003a. Les premiers Anguilliformes: II. Paraphylie du genre *Urenchelys* WOODWARD, 1900 et relations phylogénétiques. *Geobios* 36(4): 351-378.
- Belouze A, Gayet M, Atallah C. 2003b. Les premiers Anguilliformes: I. Révision des genres cénomaniens *Anguillavus* HAY, 1903 et *Luenchelys* nov. gen. *Geobios* 36(3): 241-273.
- Blot J. 1984. Les Apodes fossiles du Monte Bolca. II: Actinopterygii ordre des Apodes (= Anguilliformes). *Studi e Ricerche sui giacimenti terziari di Bolca* 4: 61-264.
- Bockmann FA, Sazima I. 2004. *Trichomycterus maracaya*, a new catfish from the upper rio Paraná, southeastern Brazil (Siluriformes: Trichomycteridae), with notes on the *T. brasiliensis* species-complex. *Neotropical Ichthyology* 2(2): 61-74.
- Britski HA, Ortega H. 1982. *Trichogenes longipinnis*, novo gênero e espécie de Trichomycterinae do sudeste do Brasil (Pisces, Siluriformes). *Revista Brasileira de Zoologia* 1(3): 211-216.
- Castle PHJ. 1986. Moringuidae. p. 187-188. In: Smith MM & Heemstra PC (eds.) *Smiths' sea fishes*. Springer-Verlag, Berlin.
- Cervigón F. 1994. *Los peces marinos de Venezuela. Volume 3*. Fundación Científica Los Roques, Caracas, Venezuela.
- Charter SR, Moser HG. 1996. Muraenidae: morays. p. 88-91. In: Moser HG (ed.) *The early stages of fishes in the California Current region*. California Cooperative Oceanic Fisheries Investigations (CalCOFI) Atlas No. 33.
- Chen HM, Shao KT, Chen CT. 1994. A review of the muraenid eels (Family Muraenidae) from Taiwan with descriptions of twelve new records. *Zoological Studies* 33(1): 44-64.
- Claro R. 1994. Características generales de la ictiofauna. p. 55-70. In: Claro R (ed.) *Ecología de los peces marinos de Cuba*. Instituto de Oceanología Academia de Ciencias de Cuba and Centro de Investigaciones de Quintana Roo.
- Conrad JL, Ast JC, Montanari S, Norell MA. 2011. A combined evidence phylogenetic analysis of Anguimorpha (Reptilia: Squamata). *Cladistics* 27(3): 230-277.
- Da Silva FO, Fabre AC, Savriama Y, Ollonen J, Mahlow K, Herrel A, Müller J, Di-Poi N. 2018. The ecological origins of snakes as revealed by skull evolution. *Nature communications* 9(1): 376.
- Datovo A, Bockmann FA. 2010. Dorsolateral head muscles of the catfish families Nematogenyidae and Trichomycteridae (Siluriformes: Loricarioidei): comparative anatomy and phylogenetic analysis. *Neotropical Ichthyology* 8(2): 193-246.
- De Pinna MCC. 1988. A new genus of trichomycterid catfish (Siluroidei, Glanapteryginae), with comments on its phylogenetic relationships. *Revue suisse de Zoologie* 95(1): 113-128.
- Devaere S, Teugels GG, Adriaens D, Huysentruyt F, Verraes W. 2004. Redescription of *Dolichallabes microphthalmus* (Poll, 1942) (Siluriformes, Clariidae). *Copeia* 1: 108-115.
- Devaere S, Adriaens D, Verraes W. 2007. *Channallabes sanghaensis* sp. n., a new anguilliform catfish from the Congo River basin, with some comments on other anguilliform clariids (Teleostei, Siluriformes). *Belgian journal of zoology* 137(1): 17.

- DoNascimento C. 2015. Morphological evidence for the monophyly of the subfamily of parasitic catfishes Stegophilinae (Siluriformes, Trichomycteridae) and phylogenetic diagnoses of its genera. *Copeia* 103(4): 933-960.
- Evans SE, Sigogneau-Russell D. 2001. A stem-group caecilian (Lissamphibia: Gymnophiona) from the Lower Cretaceous of North Africa. *Palaeontology* 44(2): 259-273.
- Ezenwaji HMG, Inyang NM. 1998. Observations on the biology of *Clarias agboyiensis* Syndenham, 1980 (Osteichthyes: Clariidae) in the Anambra floodriver system, Nigeria. *Fisheries Research* 36(1): 47-60.
- Gauthier JA. 1982. Fossil xenosaurid and anguid lizards from the early Eocene Wasatch Formation, south-east Wyoming, and a revision of the Anguioidea. *Rocky Mountain Geology* 21(1): 7-54.
- Gower DJ, Captain A, Thakur SS. 2008. On the taxonomic status of *Uropeltis bicatenata* (Günther) (Reptilia: Serpentes: Uropeltidae). *Hamadryad* 33: 64-82.
- Henmi Y, Itani G. 2014. Burrow utilization in the goby *Eutaeniichthys gilli* associated with the mud shrimp *Upogebia yokoyai*. *Zoological science* 31(8): 523-528.
- Hibino Y, Kimura S. 2015. Revision of the *Scolecenchelys gymnota* species group with descriptions of two new species (Anguilliformes: Ophichthidae: Myrophinae). *Ichthyological research* 63(1): 1-22.
- Hyun-Geun C, Seung-Ho C. 2014. First Record of Two Gobiid Fishes, *Luciogobius elongatus*, *L. platycephalus* (Perciformes: Gobiidae) from Korea. *Animal Systematics, Evolution and Diversity* 30(1): 22.
- Insacco G, Spadola F, Russotto S, Scaravelli D. 2015. *Eryx jaculus* (Linnaeus, 1758): a new species for the Italian herpetofauna (Squamata: Erycidae). *Acta Herpetologica* 10: 149-153.
- IUCN Red List: <https://www.iucnredlist.org/>
- Jenkins FA, Walsh DM, Carroll RL. 2007. Anatomy of *Eocaecilia micropodia*, a limbed caecilian of the Early Jurassic. *Bulletin of the Museum of Comparative Zoology* 158(6): 285-366.
- Lieske E, Myers R. 1994. *Collins Pocket Guide. Coral reef fishes. Indo-Pacific & Caribbean including the Red Sea*. Haper Collins Publishers.
- Love MS, Mecklenburg CW, Mecklenburg TA, Thorsteinson LK. 2005. *Resource inventory of marine and estuarine fishes of the West Coast and Alaska: A checklist of North Pacific and Arctic Ocean species from Baja California to the Alaska-Yukon border*. U.S. Department of the Interior, U.S. Geological Survey, Biological Resources Division, Seattle, Washington, 98104.
- Martin-Smith KM, Tan HH. 1998. Diversity of freshwater fishes from eastern Sabah: annotated checklist for Danum valley and a consideration of inter- and intra-catchment variability. *Raffles Bulletin of Zoology* 46(2): 573-604.
- Maschio GF, Prudente ALC, Rodrigues FS, Hoogmoed MS. 2010. Food habits of *Anilius scytale* (Serpentes: Aniliidae) in the Brazilian Amazonia. *Zoologia* 27: 184-190.
- Masuda H, Amaoka K, Araga C, Uyeno T, Yoshino T. 1984. *The fishes of the Japanese Archipelago. Vol. 1*. Tokai University Press, Tokyo, Japan.
- McDowall RM, Beumer JP. 1980. Family Anguillidae: freshwater eels. p. 44-47. In: McDowall RM (ed.) *Freshwater fishes of south-eastern Australia*. A.H. & A.W. Reed Pty. Ltd. Sydney.
- McCosker JE. 2010. *Rhinomuraena quaesita* (errata version published in 2017). The IUCN Red List of Threatened Species 2010: e.T155301A115297865.
- McCosker JE, Rosenblatt RH. 1998. A revision of the Eastern Pacific snake-eel genus *Ophichthus* (Anguilliformes: Ophichthidae) with the description of six new species. *Proceedings of the California Academy of Science* 50(19): 397-432.

- McEachran JD, Fechhelm JD. 1998. *Fishes of the Gulf of Mexico. Volume 1: Myxiniiformes to Gasterosteiformes*. University of Texas Press, Austin.
- Micklich N. 1985. Biologisch-paläontologische Untersuchungen zur Fischfauna der Messeler Ölschiefer (Mittel-Eozän, Lutetium). *Andrias* 4: 1-171.
- Miller PJ. 1981. Periophthalmidae. In: Fischer W, Bianchi G, Scott WB (eds.) *FAO species identification sheets for fishery purposes. Eastern Central Atlantic; fishing areas 34, 47 (in part)*. Department of Fisheries and Oceans Canada and FAO. vols. 1-7.
- Myers RF. 1991. *Micronesian reef fishes. Second Edition*. Coral Graphics, Barrigada, Guam.
- Mundy BC. 2005. Checklist of the fishes of the Hawaiian Archipelago. *Bishop Museum Bulletin of Zoology* 6: 1-704.
- Pfaff C, Zorzin R, Kriwet J. 2016. Evolution of the locomotory system in eels (Teleostei: Elopomorpha). *BMC evolutionary biology* 16(1): 159.
- Poll M. 1953. *Exploration hydrobiologique du lac Tanganika (1946-1947). Poissons non Cichlidae*. Institut Royal des Sciences Naturelles de Belgique, Bruxelles.
- Pyron RA, Wiens JJ. 2011. A large-scale phylogeny of Amphibia including over 2800 species, and a revised classification of extant frogs, salamanders, and caecilians. *Molecular Phylogenetics and Evolution* 61(2): 543-583.
- Rainboth WJ. 1996. *Fishes of the Cambodian Mekong*. FAO species identification field guide for fishery purposes. FAO, Rome.
- Randall JE. 1995. *Coastal fishes of Oman*. University of Hawaii Press, Honolulu, Hawaii.
- Reece JS, Smith DG, Holm E. 2010. The moray eels of the *Anarchias cantonensis* group (Anguilliformes: Muraenidae), with description of two new species. *Copeia* 2010(3): 421-430.
- Reeder TW, Townsend TM, Mulcahy DG, Noonan BP, Wood PL, Sites JW, Wiens JJ. 2015. Integrated analyses resolve conflicts over squamate reptile phylogeny and reveal unexpected placements for fossil taxa. *PLOS one* 10(3): e0118199.
- Reynolds RG, Niemiller ML, Revell LJ. 2014. Toward a Tree-of-Life for the boas and pythons: Multilocus species-level phylogeny with unprecedented taxon sampling. *Molecular Phylogenetics and Evolution* 71: 201-213.
- Riede K. 2004. *Global register of migratory species - from global to regional scales. Final Report of the R&D-Projekt 808 05 081*. Federal Agency for Nature Conservation, Bonn, Germany.
- Risch LM. 2003. Claroteidae. p. 60-96 In: Lévêque C, Paugy D, Teugels GG (eds.) *Faune des poissons d'eaux douce et saumâtres de l'Afrique de l'Ouest, Tome 2*. Collection Faune et Flore tropicales 40. Musée Royal de l'Afrique Centrale, Tervuren, Belgique, Museum National d'Histoire Naturelle, Paris, France and Institut de Recherche pour le Développement, Paris, France.
- Robins CR, Ray GC. 1986. *A field guide to Atlantic coast fishes of North America*. Houghton Mifflin Company, Boston, U.S.A.
- Robins CR, Bailey RM, Bond CE, Brooker JR, Lachner EA, Lea RN, Scott WB. 1991. Common and scientific names of fishes from the United States and Canada. *American Fisheries Society Special Publications* 20: 1-183.
- Rosenblatt RH, Rubinoff I. 1972. *Pythonichthys asodes*, a new heterenchelyid eel from the Gulf of Panama. *Bulletin of Marine Science* 22(2): 355-364.

- Santini F, Kong X, Sorenson L, Carnevale G, Mehta RS, Alfaro ME. 2013. A multi-locus molecular timescale for the origin and diversification of eels (Order: Anguilliformes). *Molecular phylogenetics and evolution* 69(3): 884-894.
- Sazima I. 2004. Natural history of *Trichogenes longipinnis*, a threatened trichomycterid catfish endemic to Atlantic forest streams in southeast Brazil. *Ichthyological Exploration of Freshwaters* 15(1): 49-60.
- Schaefer SA, Provenzano F, Pinna MD, Baskin JN. 2005. New and noteworthy Venezuelan glanapterygine catfishes (Siluriformes, Trichomycteridae), with discussion of their biogeography and psammophily. *American Museum Novitates* 3496: 1-27.
- Schmidt RE. 1993. On the biology of *Paracanthopoma parva*. *Ichthyological Exploration of Freshwaters* 4(2): 185-191.
- Seegers L. 2008. *The catfishes of Africa: A handbook for identification and maintenance*. Aqualog Verlag A.C.S. GmbH, Germany.
- Smith DG. 1981. Congridae. In: Fischer W, Bianchi G, Scott WB (eds.) *FAO species identification sheets for fishery purposes. Eastern Central Atlantic; fishing areas 34, 47 (in part)*. Department of Fisheries and Oceans Canada and FAO. Vol. 2.
- Smith DG. 1990. Heterenchelyidae. p. 153-155. In: Quéro JC, Hureau JC, Karrer C, Post A, Saldanha L (eds.) *Check-list of the fishes of the eastern tropical Atlantic (CLOFETA)*. JNICT, Lisbon; SEI, Paris; and UNESCO, Paris. Vol. 1.
- Smith CL. 1997. National Audubon Society field guide to tropical marine fishes of the Caribbean, the Gulf of Mexico, Florida, the Bahamas, and Bermuda. Alfred A. Knopf, Inc., New York. 720 p.
- Smith DG, Böhlke JE, Castle PHJ. 1981. A revision of the nettastomatid eel genera *Nettastoma* and *Nettenchelys* (Pisces: Anguilliformes), with descriptions of six new species. *Proceedings of the Biological Society of Washington* 94(2): 535-560.
- Speybroeck J, Beukema W, Bok B, Van der Voort J. 2016. *Field Guide to the Amphibians and Reptiles of Britain and Europe*. London: Bloomsbury Publishing.
- Sulak K, Shcherbachev YN. 1997. Zoogeography and systematics of six deep-living genera of synphobranchid eels, with a key to taxa and description of two new species of *Ilyophis*. *Bulletin of marine Science* 60(3): 1158-1194.
- Sullivan RM, Keller T, Habersetzer J. 1999. Middle Eocene (Geiseltalian) anguid lizards from Geiseltal and Messel, Germany. I. *Ophisauriscus quadrupes* Kuhn 1940. *Courier-Forschungsinstitut Senckenberg*: 97-130.
- Tałanda M. 2016. Cretaceous roots of the amphisbaenian lizards. *Zoologica Scripta* 45: 1-8.
- Tałanda M. 2018. An exceptionally preserved Jurassic skink suggests lizard diversification preceded fragmentation of Pangaea. *Palaeontology* 61(5): 659-677.
- Taverne L. 2004. *Libanechelys bultyncki* gen. et sp. nov., une nouvelle anguille primitive (Teleostei, Anguilliformes) du Cénomaniens marin du Liban. *Bulletin de l'Institut Royal des Sciences Naturelles de Belgique* 74: 73-87.
- Teugels GG. 1986. A systematic revision of the African species of the genus *Clarias* (Pisces; Clariidae). *Ann. Mus. R. Afr. Centr., Sci. Zool.* 247: 1-199.
- Teugels GG. 2003. Clariidae. p. 144-173 In: Lévêque C, Paugy D, Teugels GG (eds.) *Faune des poissons d'eaux douce et saumâtres de l'Afrique de l'Ouest, Tome 2*. Collection Faune et Flore tropicales 40. Musée Royal de

l'Afrique Centrale, Tervuren, Belgique, Museum National d'Histoire Naturelle, Paris, France and Institut de Recherche pour le Développement, Paris, France.

Teugels GG, Denayer B, Legendre M. 1990. A systematic revision of the African catfish genus *Heterobranchus* Geoffroy-Saint-Hilaire, 1809 (Pisces: Clariidae). *Zoological journal of the Linnean Society* 98(3): 237-257.

Thacker CE. 2013. Phylogenetic placement of the European sand gobies in Gobionellidae and characterization of gobionellid lineages (Gobiiformes: Gobioidae). *Zootaxa* 3619(3): 369-382.

Uetz P, Freed P, Hošek J (eds.). 2019. The Reptile Database, <http://www.reptile-database.org> Accessed 15 May 2018

Vidthayanon C. 2002. *Peat swamp fishes of Thailand*. Office of Environmental Policy and Planning, Bangkok, Thailand.

Villa-Verde L, Costa W. 2006. A new glanapterygine catfish of the genus *Listrura* (Siluriformes: Trichomycteridae) from the southeastern Brazilian coastal plains. *Zootaxa* 1142: 43-50.

Villa-Verde L, Lazzarotto H, Lima SM. 2012. A new glanapterygine catfish of the genus *Listrura* (Siluriformes: Trichomycteridae) from southeastern Brazil, corroborated by morphological and molecular data. *Neotropical Ichthyology* 10(3): 527-538.

Vitt LJ, Caldwell JP. 2009. *Herpetology, 3rd Edition*. Burlington: Academic Press.

Wagner P, Schmitz A. 2006. *Feylinia currori* GRAY, 1845 (Squamata: Scincidae): new distribution records from Kenya. *Salamandra* 42: 183-187.

Ward AB, Costa A, Monroe SL, Aluck RJ, Mehta RS. 2015. Locomotion in elongate fishes: A contact sport. *Zoology* 118(5): 312-319.

Watson RE. 1991. A provisional review of the genus *Stenogobius* with descriptions of a new subgenus and thirteen new species (Pisces: Teleostei: Gobiidae). *Records of the Western Australian Museum* 15(3): 627-710.

Winckler TC. 1861. *Description de quelques nouvelles espèces de poissons fossiles des calcaires d'eau douce d'Oeningen (Vol. 1)*. Loosjes: Mémoire couronné par la Société Hollandaise des Sciences à Harlem.

Wright JJ. 2017. A new diminutive genus and species of catfish from Lake Tanganyika (Siluriformes: Clariidae). *Journal of fish biology* 91(3): 789-805.

Wüster W, Peppin L, Pook CE, Walker DE. 2008. A nesting of vipers: phylogeny and historical biogeography of the Viperidae (Squamata: Serpentes). *Molecular Phylogenetics and Evolution* 49(2): 445-459.

Yamada U, Shirai S, Irie T, Tokimura M, Deng S, Zheng Y, Li C, Kim YU, Kim YS. 1995. *Names and illustrations of fishes from the East China Sea and the Yellow Sea*. Overseas Fishery Cooperation Foundation, Tokyo, Japan.

Yamada T, Sugiyama T, Tamaki N, Kawakita A, Kato M. 2009. Adaptive radiation of gobies in the interstitial habitats of gravel beaches accompanied by body elongation and excessive vertebral segmentation. *BMC Evolutionary Biology* 9(1): 145.

Zanata AM, Primitivo C. 2014. Natural history of *Copionodon pecten*, an endemic trichomycterid catfish from Chapada Diamantina in northeastern Brazil. *Journal of natural history* 48(3-4): 203-228.

Zuanon J, Sazima I. 2004. Natural history of *Stauroglanis gouldingi* (Siluriformes: Trichomycteridae), a miniature sand-dwelling candiru from central Amazonia streamlets. *Ichthyological Exploration of Freshwaters* 15(3): 201-208.