

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

**Occurrence of ascaridoid nematodes in *Illex coindetii*, a commercially relevant cephalopod species from the Ligurian Sea (Northwest Mediterranean Sea)**

**This is the author's manuscript**

*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/1737057> since 2020-06-17T22:26:58Z

*Published version:*

DOI:10.1016/j.foodcont.2020.107311

*Terms of use:*

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

**Occurrence of ascaridoid nematodes in *Illex coindetii*, a commercially relevant cephalopod species from the Ligurian Sea (Northwest Mediterranean Sea)**

Vasco Menconi<sup>1</sup>, Paolo Pastorino<sup>1,2,\*</sup>, Erika Astrid Virginie Burioli<sup>1,3</sup>, Marzia Righetti<sup>1</sup>, Tommaso Scanzio<sup>1</sup>, Livio Favaro<sup>1,4</sup>, Maria Cristina Bona<sup>1</sup>, Elena Pavoletti<sup>5</sup>, Alessandro Dondo<sup>1</sup>, Marino Prearo<sup>1</sup>

<sup>1</sup> Istituto Zooprofilattico Sperimentale del Piemonte, Liguria e Valle d'Aosta, via Bologna 148, 10154 Torino, Italy

<sup>2</sup> Dipartimento di Scienze della Vita, Università degli Studi di Trieste, via Giorgieri 10, 34127 Trieste, Italy

<sup>3</sup> LABÉO Frank Duncombe, 1 route de Rosel – Saint Contest – 14053 Caen, France;

<sup>4</sup> Dipartimento di Scienze della Vita e Biologia dei Sistemi, Università degli Studi di Torino, via Accademia Albertina 13, 10123 Torino, Italy

<sup>5</sup> Servizio Veterinario ASL Vercelli, Area Igiene della produzione, trasformazione, commercializzazione, conservazione e trasporto degli alimenti di origine animale e loro derivati, Largo Giusti 13, 13100 Vercelli, Italy.

\*corresponding author: Istituto Zooprofilattico Sperimentale del Piemonte, Liguria e Valle d'Aosta, via Bologna 148, 10154 Torino, Italy. E-mail: paolo.pastorino@izsto.it (P.P.); telephone: +39 0112686295

**Keywords:** *Anisakis pegreffii*; **Anisakidae**; food safety; *Hysterothylacium* sp.; food-borne parasite

**Abstract**

In this study we investigated the occurrence of larval ascaridoid nematodes in *Illex coindetii*, a commercially relevant cephalopod, via a cross-sectional prevalence study (January 2015 to February 2018) in two fishing areas (Savona and Piombino) in the Ligurian Sea (northwest Mediterranean Sea). A total of 745 specimens of *I. coindetii* were caught by local fisherman through bottom trawling. Parasitological examination revealed ascaridoid nematodes (L3) larvae (n=9): *Hysterothylacium* sp. (n=6) and *Anisakis* spp. (n=3). All larvae morphologically referred to the genus *Anisakis* were identified by PCR-RFLP as *A. pegreffii*. The prevalence of *Hysterothylacium* sp. infestation was 0.88% for the Savona and 0.56% for the Piombino sampling site; the prevalence of *A. pegreffii* was 0.18% for Savona and 1.11% for Piombino and did not differ

between the two sampling sites (chi-square test;  $p=0.0848$ ). Better knowledge of the distribution of ascaridoid nematodes, especially of *Anisakis* species is essential for defining their epidemiological role and the potential risk for consumer health.

## 1. Introduction

Anisakid nematodes have a complex life cycle that progresses through four larval stages (L1–L4) and involves a broad spectrum of hosts (crustaceans, cephalopods, fish, marine mammals, and sea birds). The heteroxenous life cycle involves marine mammals as definitive hosts, crustaceans as first intermediate hosts, and fish, squid and other invertebrates as intermediate or paratenic hosts (Køie 1993; Mattiucci et al., 2018). Infection is transmitted through the marine food web (Klimpel and Palm 2011).

Humans become accidental hosts after consuming raw or semi-cooked fish or cephalopods infected with third-stage larvae (Audicana and Kennedy, 2008). The consumption of improperly processed or uncooked seafood (i.e., sushi and sashimi, salted or smoked herring, Nordic gravlax, Lomi Lomi, salmon, American cebiche and marinated anchovies) is a key factor in human infection (Audicana et al., 2002). The most common symptoms of human anisakiasis are abdominal pain, nausea, emesis, and mild fever (Arizono et al., 2012). In most clinical cases, the larvae have penetrated the stomach mucosa; larvae have rarely been found in the small intestine and even more rarely in the colonic mucosa (Arizono et al., 2012). Several episodes of allergic reactions have been described as a possible consequence of *Anisakis* spp. infection, even with the ingestion of well-cooked fishery products (Daschner et al., 2000; Mattiucci et al., 2013).

Public health authorities and the seafood industry are aware that fish-borne zoonotic nematode infection can affect both consumer health and the economy of fish production. In response to these issues, the European Food Safety Authority (EFSA) published a Scientific Opinion on risk assessment of parasites in fishery products (EFSA, 2010). Nonetheless, the zoonotic implications and the distribution and epidemiology of anisakid nematodes in the Mediterranean Sea remain poorly understood. Furthermore, the bulk of published data refer to the occurrence of *Anisakis* spp. larvae in teleost species. Few studies have described the occurrence of ascaridoid larvae in cephalopods from the Mediterranean Sea (Gestal et al., 1999; Farjallah et al., 2008; Angelucci et al., 2011; Petrić et al., 2011; Serracca et al., 2013; Picó-Durán et al., 2016; Goffredo et al., 2019).

Cephalopods are important predators in the marine trophic chain (Boyle and Pierce 1994; Clarke, 1996). Furthermore, the global landings of cephalopods have increased with growing consumer appreciation. Broadtail shortfin (*Illex coindetii*) is a benthic ommastrephid squid commonly fished in the Mediterranean Sea by bottom trawl (Lefkaditou et al., 2008; Picó-Durán et al., 2016). It is

widely distributed throughout the Mediterranean, the eastern Atlantic (from south of Great Britain to Namibia), and the western Atlantic (from the Caribbean to the Gulf of Mexico and the Straits of Florida) (Sanchez et al., 1998). *I. coindetii* has been recorded up to 1100 m depth, but commercial catches are normally obtained at an average depth of 60-400 m (Guerra 1992; Jereb and Roper, 2010). Records of cephalopods caught at depths greater than 500 m are few and come mainly from the western Mediterranean (Quetglas et al., 2000). With this study we wanted to investigate the prevalence and the anatomical location of ascaridoid nematodes infestation, as well as the geographical distribution of ascaridoid nematodes in *I. coindetii* from the Ligurian Sea. Our findings may inform the creation of risk maps of cephalopod consumption.

## 2. Material and Methods

### 2.1 Study area and squid sampling

Between January 2015 and February 2018 (3-year monitoring period), a cross-sectional prevalence study was carried out on ascaridoid nematodes recovered from broadtail shortfin (*Illex coindetii*). A total of 745 specimens of *I. coindetii* were caught by local fisherman through bottom trawling in two fishing areas: Savona (n=565) and Piombino (n=180) in the Ligurian Sea (Fig. 1).

Squid samples were put in cold boxes at 4°C and transferred to the Fish Diseases Laboratory of the Istituto Zooprofilattico Sperimentale del Piemonte, Liguria e Valle d'Aosta for analysis. For each specimen, species identification and measurement of mantle length (cm) were performed. Anatomopathological and parasitological examination focused on the occurrence of larval ascaridoid nematodes. The internal organs were placed in Petri dishes with physiological saline solution and left for 30 min under a heat source to allow mobilization of the anisakid larvae (Serracca et al., 2013). The visceral surfaces were visually inspected and observed under a stereomicroscope (Zeiss Stemi V8, Zeiss, Oberkochen, Germany) to detect the presence of larval ascaridoid nematodes. The muscle tissue was sectioned in thick slices (< 5 mm) and analysed by transillumination (UVP White Light Transilluminators, TW-43, Analytik Jena AG, Jena, Germany). The muscle tissue and visceral organs were digested separately using an enzymatic digestion method based on the Codex Alimentarius Commission (Codex Alimentarius Commission, 2004) and EU Regulation (EC) No. 2075/2005 (European Commission, 2005). The larvae were rinsed in deionized water and counted. Morphological identification was performed by light microscopy (Olympus BX40, Olympus, Tokyo, Japan). The anterior and posterior parts from each nematode were cleared using Amman's lactophenol to allow morphological identification. Morphological classification was performed as described in Berland (1961). A small piece of the mid-part of the larvae morphologically identified as *Anisakis* spp. underwent molecular analysis by polymerase

chain reaction-restriction fragment length polymorphism (PCR-RFLP) according to the protocol reported by Serracca et al. (2013).

105

## 2.2 Statistical analysis

Normality of data was checked using the Shapiro-Wilk test. The non-parametric Mann-Whitney U test was used to check for differences in mantle length (the biometric parameter used to measure squid size) of *I. coindetii* samples from the two sampling sites (Arkhipkin, 1996). The prevalence of nematode infestation was calculated for the two sampling sites (Bush et al., 1997) and the differences between them were tested using the chi-square test. The 95% confidence intervals (95% CI) were also presented. The criterion for significance was set at  $p < 0.05$ . Statistical analyses were performed using GraphPad Prism version 8.0.2.

114

## 3. Results

The range in mantle length was 3-15 cm ( $8.65 \pm 2.11$  cm mean  $\pm$  standard deviation [ $\pm$ SD]) for the squid from the Savona sampling site and 7-20 cm ( $10.67 \pm 2.69$  cm) for the squid from the Piombino site. A significant difference in mantle length between the two sites was noted (Mann-Whitney U test;  $p=0.0001$ ).

Parasitological examination revealed ascaridoid nematodes larvae (n=9) (Table 1): *Hysterothylacium* sp. (n=6, 5 of which from the Savona and 1 from the Piombino site) and *Anisakis* spp. (n=3, 1 of which from the Savona and 2 from the Piombino site). We found third larval stage (L3) of *Anisakis* larvae type I (*sensu* Berland, 1961) and third larval stage (L3) of *Hysterothylacium* sp. All larvae morphologically referred to the genus *Anisakis* were identified as *A. pegreffii* by PCR-RFLP.

None of the ascaridoid nematodes had penetrated the musculature but were found encapsulated on the surface of the peritoneal membrane lining the mantle cavity. The prevalence of *A. pegreffii* infestation was 0.18% (95% confidence interval [CI] 0 - 0.52) for the Savona site and 1.11% (95% CI 0 - 2.64) for the Piombino site; the prevalence of *Hysterothylacium* sp. was 0.88% (95% CI 0 - 2.64) for the Savona site and 0.56% (95% CI 0.01 - 3.06) for the Piombino site. There was no difference in the prevalence of *A. pegreffii* between the two sites ( $\chi^2=2.97$ ;  $p=0.0848$ ).

132

## 4. Discussion

With this study we investigated the occurrence of *Anisakis pegreffii* and *Hysterothylacium* spp. in *I. coindetii* from the Ligurian Sea. Few studies to date have described the occurrence of ascaridoid

136 **nematodes** in *I. coindetii* from the Mediterranean Sea. This knowledge gap poses a potential health  
 137 risk for squid consumption.

138 Published data show a higher prevalence of **ascaridoid nematodes** infestation in *I. coindetii* than in  
 139 our samples: Gestal et al. (1999) found a prevalence of 4.8 % for *A. simplex* and of 28 % for  
 140 *Hysterothylacium* spp. (n=42; Tyrrhenian Sea); Angelucci et al. (2011) reported a prevalence of 50  
 141 % for *Anisakis* spp. but only in 4 samples from the Sardinian Sea; Picó-Durán et al. (2016) reported  
 142 a prevalence of 11.4 % for *A. pegreffii*, 16.6 % for *A. physeteris*, 0.8 % for *A. simplex* x *A. pegreffii*  
 143 hybrid, and 4.1 % for *Hysterothylacium* spp. in samples from Spanish Mediterranean areas (n=123).  
 144 All specimens of *I. coindetii* (n=137) from Apulia (Italy) analysed by Goffredo et al. (2019) tested  
 145 negative for anisakid nematode larvae. Finally, Serracca et al. (2013) found only one larva of  
 146 *Hysterothylacium* spp. (prevalence 1.6 %; n=60) in samples from the Ligurian Sea.

147 Cipriani et al. (2019) also investigated the occurrence of **ascaridoids nematodes in squids but**  
 148 **belonged to the species *Illex argentinus* (n=70) caught off the Falkland Islands (South Atlantic**  
 149 **Ocean). Prevalence values of *Anisakis pegreffii* (15.7 %) and *Hysterothylacium aduncum* (2.9 %)**  
 150 **were higher in comparison to our results, probably due to the different environmental conditions of**  
 151 **ocean waters that affect the biological cycle of ascaridoids nematodes.**

152 In our sample, the average mantle length (8.65 cm for the Savona and 10.67 cm for the Piombino  
 153 sampling site) **was smaller than** in the study samples mentioned above, in which the mean mantle  
 154 length reached up to 36.5 cm (Gestal et al., 1999; Petrić et al., 2011; Serracca et al., 2013; Picó-  
 155 Durán et al., 2016). The higher prevalence the previous studies reported may stem from the fact that  
 156 older *I. coindetii* host more nematodes than young ones since squids change their food habits as  
 157 they grow. *I. coindetii* change diet during the life cycle: juveniles generally prey on crustaceans  
 158 (Euphausiacea, Penaeidae and Phrosinidae) before switching to fish and cephalopods as adults  
 159 (Boyle and Rodhouse, 2005; Rosas-Luis et al., 2014). Broadtail shortfin most likely acquires  
 160 anisakid larvae through the predation of infected euphausiids or from small teleosts (Pascual et al.,  
 161 1996; Abollo et al., 2001). Based on the results of this study and previous findings, it may be  
 162 assumed that the consumption of smaller *I. coindetii* carries a lower health risk for nematode  
 163 zoonoses.

164 This study is the first to reveal *A. pegreffii* identified at the species level by PCR-RFLP in *I.*  
 165 *coindetii* from the Ligurian Sea. This finding is consistent with published data documenting it as the  
 166 most widespread species in the Mediterranean Sea (Mattiucci and Nascetti, 2006). Furthermore, our  
 167 observation of visceral infestation of **ascaridoid nematodes** larvae is shared by several previous  
 168 studies (Smith, 1984; Gestal et al., 1999; Petrić et al., 2011) but not by Picó-Durán et al. (2016)  
 169 who found 10 larvae of *Hysterothylacium* sp. in the muscle tissue.

170 The role of *I. coindetii* in the *Anisakis* spp. life cycle may be limited by their shorter lifespan  
171 compared to fish (Packard, 1972). Nevertheless, they make up an important part of the diet of  
172 odontocete cetaceans (definitive hosts) (Smith, 1984). The Ligurian Sea, also designated as the  
173 Pelagos Sanctuary for Cetaceans, is a protected area where cetaceans are more abundant than in the  
174 rest of the western Mediterranean Sea (Di Sciara et al., 1993; Azzellino et al., 2008; Serracca et al.,  
175 2013). Anisakidae transmission pathways are strictly habitat-dependent and involve a wide  
176 spectrum of invertebrates and intermediate or paratenic fish hosts (Klimpel and Rückert 2005;  
177 McClelland 2005; Palm and Klimpel, 2007). Despite predisposing factors present in both sampling  
178 sites (Pelagos Sanctuary for Cetaceans), the survey data show a very low prevalence of *Anisakis* and  
179 *Histerothylacium* species.

180 Further investigations on cephalopods, as well as parasite hosts, may improve our knowledge of the  
181 food web structure and better define their role in the **ascaridoid nematodes** life cycle.

182

## 183 **5. Conclusions**

184 Cephalopods are intermediate or paratenic hosts in the biological cycles and transmission of several  
185 parasites (Petricić et al., 2011). This study provides new information on the distribution of **ascaridoid**  
186 **nematodes** larvae in broadtail shortfin from the Ligurian Sea. Improving knowledge of zoonotic  
187 food-borne parasites is a milestone in health risk assessment of fish consumption (Menconi et al.,  
188 2020). The survey data suggest that the risk of fish-borne zoonotic nematode infection after the  
189 consumption of broadtail shortfin from the Ligurian Sea is very low. Prevalence rates, low  
190 consumption of raw and/or undercooked cephalopods, and the tendency of the larvae to encapsulate  
191 in the visceral compartment rather than in the muscle tissue of the host bear this out. Further studies  
192 are needed for detailed risk assessment of **ascaridoid nematodes** larvae in other commercially  
193 relevant cephalopod species to define consumer health risk.

194

## 195 **Acknowledgments**

196 This research was partially funded by the Italian Ministry of Health (**project IZS PLV 12/18 RC**).

197

## 198 **Declaration of competing interest**

199 None.

200

## 201 **References**



202

- 203 Abollo, E., Gestal, C., & Pascual, S. (2001). *Anisakis* infestation in marine fish and cephalopods  
204 from Galician waters: an updated perspective. *Parasitology Research*, 87(6), 492-499.  
205 <https://doi.org/10.1007/s004360100389>.
- 206 Angelucci, G., Meloni, M., Merella, P., Sardu, F., Madeddu, S., Marrosu, R., Petza, F., & Salati, F.  
207 (2011). Prevalence of *Anisakis* spp. and *Hysterothylacium* spp. larvae in teleosts and cephalopods  
208 sampled from waters off Sardinia. *Journal of Food Protection*, 74(10), 1769-1775.  
209 <https://doi.org/10.4315/0362-028X.JFP-10-482>.
- 210 Arkhipkin, A. (1996). Geographical variation in growth and maturation of the squid *Illex coindetii*  
211 (Oegopsida, Ommastrephidae) off the north-west African coast. *Journal of the Marine Biological*  
212 *Association of the United Kingdom*, 76(4), 1091-1106. [https://doi.org/10.1017/](https://doi.org/10.1017/S0025315400040984)  
213 [S0025315400040984](https://doi.org/10.1017/S0025315400040984).
- 214 Arizono, N., Yamada, M., Tegoshi, T., & Yoshikawa, M. (2012). *Anisakis simplex* sensu stricto and  
215 *Anisakis pegreffii*: biological characteristics and pathogenetic potential in human anisakiasis.  
216 *Foodborne pathogens and disease*, 9(6), 517-521. <https://doi.org/10.1089/fpd.2011.1076>.
- 217 Audicana, M. T., Ansotegui, I. J., de Corres, L. F., & Kennedy, M. W. (2002). *Anisakis simplex*:  
218 dangerous dead and alive? *Trends in Parasitology*, 18(1), 20-25. [https://doi.org/10.1016/S1471-](https://doi.org/10.1016/S1471-4922(01)02152-3)  
219 [4922\(01\)02152-3](https://doi.org/10.1016/S1471-4922(01)02152-3).
- 220 Audicana, M. T., & Kennedy, M. W. (2008). *Anisakis simplex*: from obscure infectious worm to  
221 inducer of immune hypersensitivity. *Clinical Microbiology Reviews*, 21(2), 360-379.  
222 <https://doi.org/10.1128/CMR.00012-07>
- 223 Azzellino, A., Gaspari, S., Airoidi, S., & Nani, B. (2008). Habitat use and preferences of cetaceans  
224 along the continental slope and the adjacent pelagic waters in the western Ligurian Sea. *Deep Sea*  
225 *Research Part I: Oceanographic Research Papers*, 55(3), 296-323. [https://doi.org/10.1016/](https://doi.org/10.1016/j.dsr.2007.11.006)  
226 [j.dsr.2007.11.006](https://doi.org/10.1016/j.dsr.2007.11.006).
- 227 Berland, B. (1961). Nematodes from some Norwegian marine fishes. *Sarsia*, 2(1), 1-50.  
228 <https://doi.org/10.1080/00364827.1961.10410245>.
- 229 Boyle, P., & Rodhouse, P. (2005). Cephalopods: ecology and fisheries. Blackwell Publishing,  
230 London, pp. 464.
- 231 Boyle, P. R., & Pierce, G. J. (1994). Fishery biology of northeast Atlantic squid: an overview.  
232 *Fisheries Research*, 21(1-2), 1-15. [https://doi.org/10.1016/0165-7836\(94\)90093-0](https://doi.org/10.1016/0165-7836(94)90093-0).
- 233 Bush, A. O., Lafferty, K. D., Lotz, M., Shostak, A.W. (1997). Parasitology meets ecology. *Journal*  
234 *of Parasitology*, 83(4), 575-583.



Clarke, M. R. (1996). The role of cephalopods in the world's oceans: an introduction. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 351, 977–1112. <https://doi.org/10.1098/rstb.1996.0088>.

Codex Alimentarius Commission (2004). Standard for salted Atlantic herring and salted sprat. *Codex Stan*, 244-2004, 1–8.

Daschner, A., Alonso-Gómez, A., Cabañas, R., Suarez-de-Parga, J. M., & López-Serrano, M. C. (2000). Gastroallergic anisakiasis: borderline between food allergy and parasitic disease - Clinical and allergologic evaluation of 20 patients with confirmed acute parasitism by *Anisakis simplex*. *Journal of Allergy and Clinical Immunology*, 105(1), 176-181. [https://doi.org/10.1016/S0091-6749\(00\)90194-5](https://doi.org/10.1016/S0091-6749(00)90194-5).

Di Sciara, G. N., Venturino, M. C., Zanardelli, M., Bearzi, G., Borsani, F. J., & Cavalloni, B. (1993). Cetaceans in the central Mediterranean Sea: distribution and sighting frequencies. *Italian Journal of Zoology*, 60(1), 131-138. <https://doi.org/10.1080/11250009309355800>.

EFSA (2010). Panel on Biological Hazards (BIOHAZ). Scientific opinion on risk assessment of parasites in fishery products. *EFSA Journal*, 8(4), 1543. <https://doi.org/10.2903/j.efsa.2010.1543>.

European Commission (2005). Commission Regulation (EC) No. 2075/2005 of 5 December 2005 laying down specific rules on official controls for *Trichinella* in meat. *Official Journal of the European Union*, 338, 75-76.

Farjallah, S., Slimane, B. B., Busi, M., Paggi, L., Amor, N., Blel Said, K., & D'Amelio, S. (2008). Occurrence and molecular identification of *Anisakis* spp. from the North African coasts of Mediterranean Sea. *Parasitology Research*, 102(3), 371. <https://doi.org/10.1007/s00436-007-0771-9>.

Gestal, C., Belcari, P., Abollo, E., & Pascual, S. (1999). Parasites of cephalopods in the northern Tyrrhenian Sea (western Mediterranean): new host records and host specificity. *Scientia Marina*, 63(1), 39-43. <https://doi.org/10.3989/scimar.1999.63n139>.

Goffredo, E., Azzarito, L., Di Taranto, P., Mancini, M.E., Normanno, G., Didonna, A., Faleo, S., Occhiochiuso, G., D'Attoli, L., Pedarra, C., Pinto, P., Cammilleri, G., Graci, S., Sciortino, S., & Costa, A. (2019). Prevalence of anisakid parasites in fish collected from Apulia region (Italy) and quantification of nematode larvae in flesh. *International Journal of Food Microbiology*, 292, 159-170. <https://doi.org/10.1016/j.ijfoodmicro.2018.12.025>.

Guerra, A. (1992). Mollusca, Cephalopoda. In: Ramos, M. A. (ed.), "Fauna Ibérica". Museo Nacional de Ciencias Naturales, CSIC Madrid: 327.

Jereb, P., & Roper, C. F. (2010). Cephalopods of the world. An annotated and illustrated catalogue of cephalopod species known to date. *FAO Species Catalogue for Fishery Purposes*, 4(2), 262.

269 Klimpel, S., & Palm, H. W. (2011). Anisakid nematode (Ascaridoidea) life cycles and distribution:  
 270 increasing zoonotic potential in the time of climate change? In: Progress in parasitology, Springer,  
 271 Berlin, Heidelberg, 201-222.

272 Klimpel, S., & Rückert, S. (2005). Life cycle strategy of *Hysterothylacium aduncum* to become the  
 273 most abundant anisakid fish nematode in the North Sea. *Parasitology Research*, 97(2), 141-149.  
 274 <https://doi.org/10.1007/s00436-005-1407-6>.

275 Køie, M. (1993). Aspects of the life cycle and morphology of *Hysterothylacium aduncum*  
 276 (Rudolphi, 1802) (Nematoda, Ascaridoidea, Anisakidae). *Canadian Journal of Zoology*, 71(7),  
 277 1289-1296. <https://doi.org/10.1139/z93-178>.

278 Lefkaditou, E., Politou, C. Y., Palialexis, A., Dokos, J., Cosmopoulos, P., & Valavanis, V. D.  
 279 (2008). Influences of environmental variability on the population structure and distribution patterns  
 280 of the short-fin squid *Illex coindetii* (Cephalopoda: Ommastrephidae) in the Eastern Ionian Sea.  
 281 *Hydrobiologia*, 612, 71-90. <https://doi.org/10.1007/s10750-008-9490-1>.

282 Mattiucci, S., Cipriani, P., Levsen, A., Paoletti, M., & Nascetti, G. (2018). Molecular epidemiology  
 283 of *Anisakis* and anisakiasis: an ecological and evolutionary road map. In D. Rollinson, & J. R.  
 284 Stothard (Eds.), *Advances in Parasitology* (pp. 93-263) Academic Press.  
 285 <https://doi.org/10.1016/bs.apar.2017.12.001>

286 Mattiucci, S., Fazii, P., Paoletti, M., De Rosa, A., Salomone Megna, A., Glielmo, A., Bruschi, F.,  
 287 De Angelis, M., Costa, A., Meucci, C., Sorrentini, L., Calvaruso, V., & Nascetti G. (2013)  
 288 Anisakiasis and gastroallergic reactions associated with *Anisakis pegreffii* infection, Italy .  
 289 *Emerging Infectious Disease*, 19(3): 496-499. <https://doi.org/10.3201/eid1903.121017>

290 Mattiucci, S., & Nascetti, G. (2006) Molecular systematics, phylogeny and ecology of anisakid  
 291 nematodes of the genus *Anisakis* Dujardin, 1845: an update. *Parasite*, 13(2): 99-113.  
 292 <https://doi.org/10.1051/parasite/2006132099>

293 McClelland, G. (2005) Nematoda (roundworms). In: Rohde K (ed) “Marine parasitology”. CABI  
 294 Publishing, Wallingford, 104-115.

295 Menconi, V., Manfrin, C., Pastorino, P., Mugetti, D., Cortinovis, L., Pizzul, E., Pallavicini, A., &  
 296 Prearo, M. (2020). First Report of *Clinostomum complanatum* (Trematoda: Digenea) in European  
 297 Perch (*Perca fluviatilis*) from an Italian Subalpine Lake: A Risk for Public Health? *International*  
 298 *journal of environmental research and public health*, 17, 1389.  
 299 <https://doi.org/10.3390/ijerph17041389>

300 Packard, A. (1972). Cephalopods and fish: the limits of convergence. *Biological Reviews*, 47(2),  
 301 241-307. <https://doi.org/10.1111/j.1469-185X.1972.tb00975.x>.

302 Palm, H. W., & Klimpel, S. (2007). Evolution of parasitic life in the ocean. *Trends in Parasitology*,  
303 23(1), 10-12. <https://doi.org/10.1016/j.pt.2006.11.001>

304 Pascual, S., Gonzalez, A., Arias, C., & Guerra, A. (1996). Biotic relationships of *Illex coindetii* and  
305 *Todaropsis eblanae* (Cephalopoda, Ommastrephidae) in the northeast Atlantic: evidence from  
306 parasites. *Sarsia*, 81(3), 265-274. <https://doi.org/10.1080/00364827.1996.10413624>.

307 Petrić, M., Mladineo, I., & Šifner, S. K. (2011). Insight into the short-finned squid *Illex coindetii*  
308 (Cephalopoda: Ommastrephidae) feeding ecology: is there a link between helminth parasites and  
309 food composition? *Journal of Parasitology*, 97(1), 55-62. <https://doi.org/10.1645/GE-2562.1>.

310 Picó-Durán, G., Pulleiro-Potel, L., Abollo, E., Pascual, S., & Muñoz, P. (2016). Molecular  
311 identification of *Anisakis* and *Hysterothylacium* larvae in commercial cephalopods from the Spanish  
312 Mediterranean coast. *Veterinary Parasitology*, 220, 47-53. <https://doi.org/10.1016/j.vetpar.2016.02.020>.

314 Quetglas, A., Carbonell, A., & Sánchez, P. (2000). Demersal continental shelf and upper slope  
315 cephalopod assemblages from the Balearic Sea (north-western Mediterranean). Biological aspects  
316 of some deep-sea species. *Estuarine, Coastal and Shelf Science*, 50(6), 739-749.  
317 <https://doi.org/10.1006/ecss.1999.0603>.

318 Rosas-Luis, R., Villanueva, R., & Sánchez, P. (2014). Trophic habits of the Ommastrephid squid  
319 *Illex coindetii* and *Todarodes sagittatus* in the northwestern Mediterranean Sea. *Fisheries Research*,  
320 152, 21-28. <https://doi.org/10.1016/j.fishres.2013.10.009>.

321 Sanchez, P., Gonzalez F., Jereb P., Laptikhovsky V. V., Mangold K., Nigmatullin C., Ragonese S.  
322 (1998). *Illex coindetii*. In: "Squid recruitment dynamics: The commercial *Illex* species", P. G.  
323 Rodhouse, E. G. Dawe, and R. K. O'Dor (eds.). FAO Fisheries Technical Paper 376, FAO, Rome,  
324 Italy, 59-76.

325 Serracca, L., Cencetti, E., Battistini, R., Rossini, I., Prearo, M., Pavoletti, E., Fioravanti, M. L.,  
326 Righetti, M., Di Donfrancesco, B., & Ercolini, C. (2013). Survey on the presence of *Anisakis* and  
327 *Hysterothylacium* larvae in fishes and squids caught in Ligurian Sea. *Veterinary Parasitology*,  
328 196(3-4), 547-551. <https://doi.org/10.1016/j.vetpar.2013.02.024>.

329 Smith, J. W. (1984). Larval ascaridoid nematodes in myopsid and oegopsid cephalopods from  
330 around Scotland and in the northern North Sea. *Journal of the Marine Biological Association of the*  
331 *United Kingdom*, 64(3), 563-572. <https://doi.org/10.1017/S0025315400030253>.

332

333

334

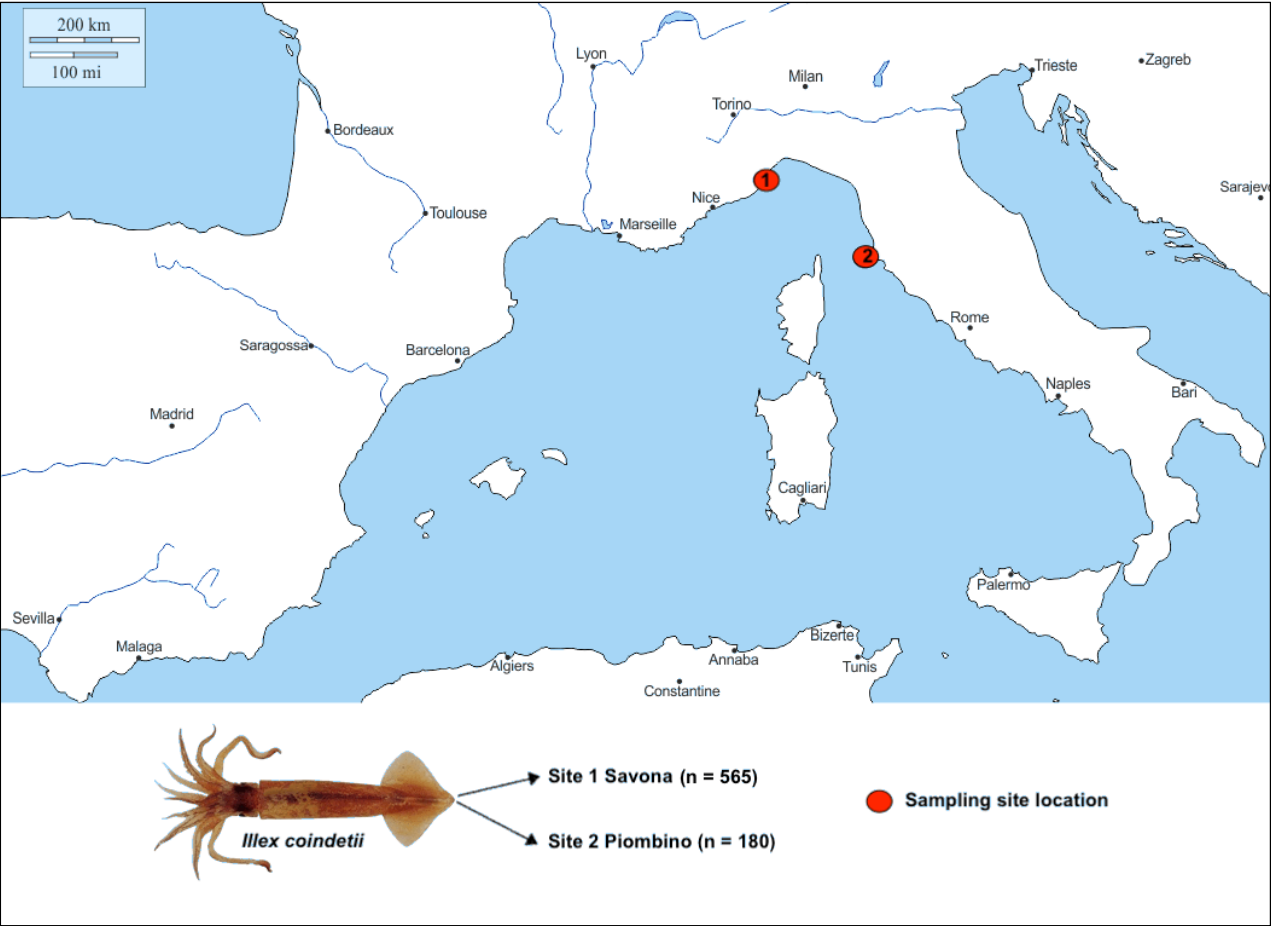
335

**Table 1.** Mantel length (cm) and number (N) of *Illex coindetii* caught from the two sites with the number and mean intensity of infestation by *Hysterothylacium* sp. and *Anisakis pegreffii*.

Sampling site	<i>Illex coindetii</i>	Mantel length	<i>Hysterothylacium</i> sp.		<i>Anisakis pegreffii</i>	
	N	cm (mean $\pm$ SD)	N	mean intensity	N	mean intensity
Savona	565	8.65 $\pm$ 2.11	5	1	1	1
Piombino	180	10.67 $\pm$ 2.69	1	1	2	1

370

371 **Figure 1.** Western Mediterranean Sea and location of sampling sites (in red). Site 1 (Savona):  
372 44°17'25.58"N 8°39'6.23"E; site 2 (Piombino): 42°56'34.164"N 10°25'37.713"E.

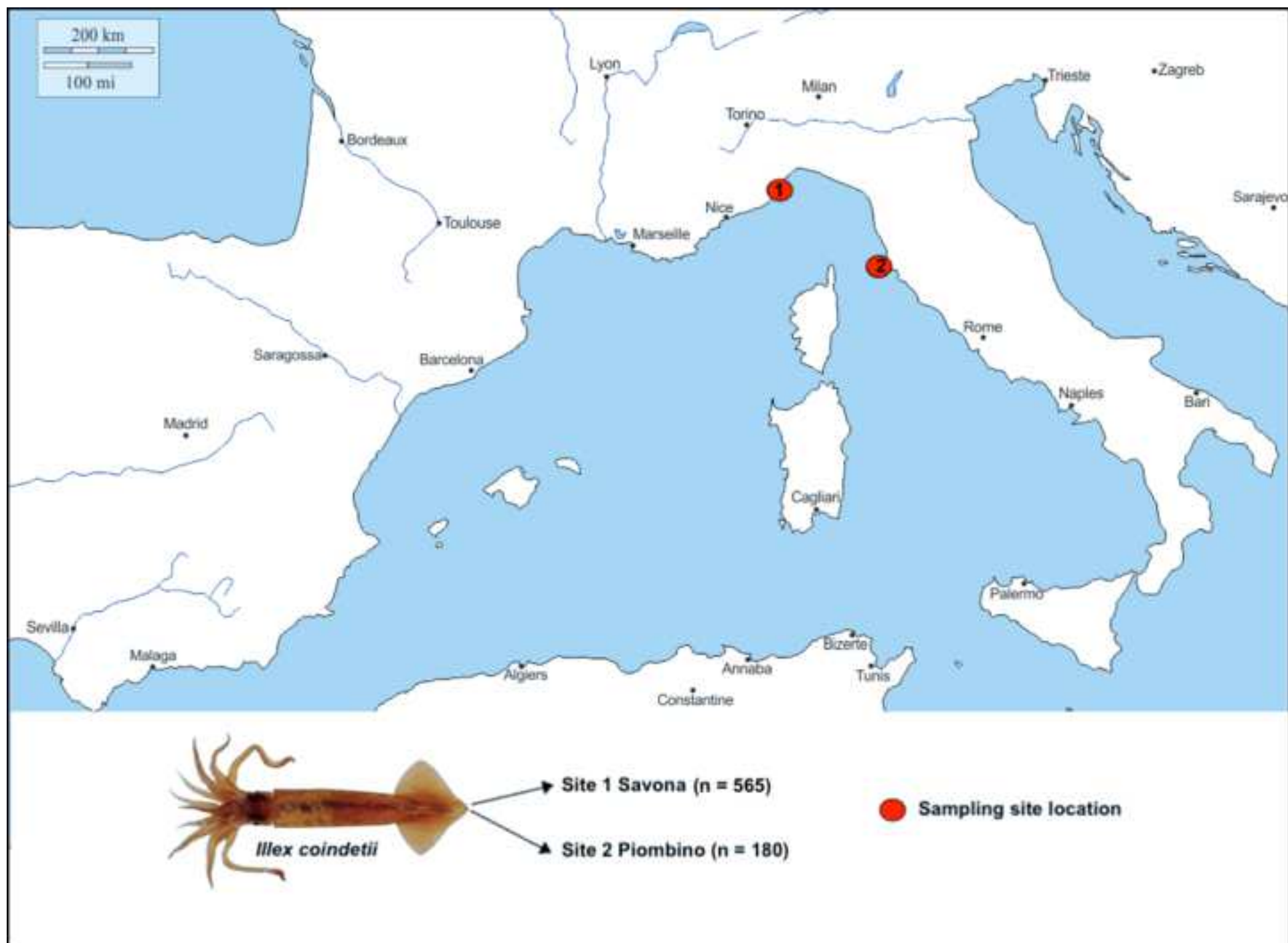


373

374

375

Figure 1



## **Credit Author Statement**

Conceptualization: **Vasco Menconi, Paolo Pastorino, Maria Cristina Bona, and Marino Prearo**;  
Data curation: **Vasco Menconi, Paolo Pastorino and Maria Cristina Bona**; Investigation: **Vasco Menconi, Paolo Pastorino, Erika Astrid Virginie Burioli, Marzia Righetti, Tommaso Scanzio, Livio Favaro, Maria Cristina Bona, Elena Pavoletti, Alessandro Dondo and Marino Prearo**;  
Methodology: **Vasco Menconi, Paolo Pastorino and Marino Prearo**; Supervision: **Marino Prearo**;  
Writing – original draft: **Vasco Menconi and Paolo Pastorino**; Writing – review & editing: **Vasco Menconi, Paolo Pastorino and Marino Prearo**. All authors have read and agreed to the published version of the manuscript.



**Declaration of interests**

☒ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

☐ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: