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Cancer stem cell antigens as targets for new combined anti-cancer therapies

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Abbreviations: CSC: cancer stem cells; CT: cancer testis; DC: dendritic cells; DNA

methyltransferase, DNMT; EMT: epithelial-to-mesenchymal transition; FDA: Food and Drug

Administration; head and neck squamous cell carcinomas, HNSCC; immune checkpoint inhibitors,

ICI; major histocompatibility complex class I, MHC I; melanoma-associated antigen gene, MAGE; myeloid-derived suppressor cells, MDSCs; mucin 1, MUC1; The Cancer Genome Atlas, TCGA; tumor growth factor, TGF-β; tumor microenvironment, TME; T-regulatory cells, Tregs

Highlights

- Cancer stem cells (CSC) are involved in tumor resistance, recurrence and metastasis
- CSCs are an obstacle to the success of immune checkpoint inhibitors (ICIs)
- Vaccination is a possible approach to CSC elimination
- Antigens crucial for CSC maintenance and function are ideal targets
- Combining CSC-targeting vaccines, ICIs and other drugs may improve patient outcomes

Abstract

The introduction of immune checkpoint inhibitors (ICIs) has ushered in a new, golden age for cancer immunotherapy. However, their clinical success remains limited in several solid cancer types because of the low intrinsic immunogenicity of tumors and the development of immune escape mechanisms. Cancer stem cells (CSCs), a small population of cancer cells that are responsible for tumor onset, metastatic spread and relapse after treatment, play a pivotal role in resistance to ICIs. The development of novel therapies that can target CSCs would thus improve the outcomes of current immunotherapy regimens. In this light, vaccines that target CSCs are a promising strategy. This paper briefly describes the immunologic properties of CSCs and their antigenic profile, and reviews current preclinical and clinical approaches that combine CSC-targeting vaccines with different synergistic therapies for the development of more effective antineoplastic treatments.

INTRODUCTION

Cancer immunotherapy is revolutionizing the treatment of oncological patients, as immune checkpoint inhibitors (ICIs) ameliorate the prognosis of many patients suffering from tumors such as melanoma, non-small-cell lung cancer, colorectal and renal carcinoma (1). However, ICIs fail to induce clinical responses in a significant proportion of patients, with single-agent response rates of between 10 and 35% (2).

ICI effectiveness is dependent on a pre-existing antitumor immune response, and vaccination is a good strategy with which to induce one (1). Although research on cancer vaccines has led to disappointing results, with many clinical trials failing to improve patient survival (3), and only one FDA-approved vaccine (4), the possible combination of vaccines and ICIs has renewed interest in the topic.

The eradication of a tumor entails the elimination of CSCs, which are a small population of cells with stem-like features that are responsible for tumor initiation and metastasis and, being resistant to current therapies, for its recurrence (5,6). The efficacy of a cancer vaccine might therefore depend on its ability to target CSCs (7). Interestingly, most vaccines tested so far have targeted antigens expressed by differentiated cancer cells, sparing CSCs, which display particular antigenic and immune-modulating properties (8).

1. Cancer stem cell immunology

CSCs are slowly dividing cells endowed with unlimited self-renewal potential that initiate tumors and that, thanks to their high degree of phenotypic plasticity, undergo epithelial-to-mesenchymal transition (EMT) and disseminate to distant organs, generating metastases (5). Being resistant to chemo- and radiotherapy, CSCs are responsible for cancer relapse (9). Moreover, CSCs possess immune-evasive properties that allow them to escape T-cell killing. Indeed, defects in antigen presentation, such as the downregulation of major histocompatibility complex class I (MHC I) and

molecules involved in antigen presentation, are common in CSCs from several cancers, including breast (10), colon (11) and melanoma (12). Moreover, CSCs exert immunosuppressive effects in the tumor microenvironment (TME) through intense cross-talk with immune cells (13). A machine-learning-based analysis of The Cancer Genome Atlas (TCGA) has revealed a negative association between CSC frequency and tumor leukocyte infiltration, which correlates with high tumor grade (14). CSCs impair effector T-cell recruitment and activation by releasing a wide variety of cytokines and growth factors that attract immunosuppressor cells. Indeed, when high amounts of tumor growth factor (TGF)- β are secreted by CSCs (12,15), T-regulatory cells (Tregs), which inhibit effector T-cell proliferation, are activated, and fibrosis, which is a physical barrier that hampers T-lymphocyte infiltration, is promoted (16). Similarly, CSCs inhibit cytotoxic T-cell function and induce the recruitment of myeloid-derived suppressor cells (MDSCs), as well as the polarization of Th2 cells and M2 pro-tumoral macrophages (17-19). Interestingly, the cytokines and growth factors that are released by MDSCs, and other immunosuppressive cells recruited in the TME, support CSC survival and self-renewal, generating a vicious circle that promotes cancer progression (20).

The link between stemness and immunosuppression has been further confirmed by a TCGA analysis on 21 solid cancer types, and a negative association between stemness and interferon- α/β signaling, and a striking positive association with several immunosuppressive genes, including *TGFB1* and *CD276* and *CD155*, two inhibitors of T and natural killer cells, were observed (21). Indeed, CSCs also inhibit immune effector cells thanks to the overexpression of several ICs, which, moreover, exert cell-intrinsic pro-tumoral mechanisms, favoring CSC survival and self-renewal (22). For instance, CSCs from melanoma, ovarian, colorectal and breast cancers express high levels of PD-L1 (23,24), which induces the AKT-dependent expression of the stem-cell markers OCT-4A, Nanog and BMI1 (25). Similarly, CD47 is overexpressed on CSCs from liver, pancreatic, breast and other tumors, and, besides inhibiting CSC phagocytosis by macrophages, directly protects CSCs from apoptosis (26). CTLA-4, which promotes melanoma-CSC proliferation and survival, has shown a similar effect (27). Despite the numerous immune-evasive properties of CSCs, preclinical studies that target them with immunotherapy have provided encouraging results, demonstrating that vaccines based on CSC lysates or dendritic cells (DC) that are loaded with CSCs are more effective than non-CSC-based equivalents in preventing tumor onset (28-34). Clinical trials using CSC-loaded DCs or CSC lysates have been performed on patients affected by different solid cancers, demonstrating that vaccination is safe and effectively induces a specific immune response (35-37). Lasting protection, with about 70% of patients still alive after 20 years, has been reported for AGI-101H, a vaccine that is comprised of irradiated melanoma cell lines engineered with hyper IL-6 (a fusion form of IL-6 and its soluble receptor) that grants it stem-like features (38).

The superiority exhibited by CSC- over non-CSC-based vaccines suggests that the antigenic profile of these two cancer cell populations differs, and that the targeting of CSC antigens is required to achieve durable and effective antitumor responses.

2. Cancer stem cell oncoantigens as targets for vaccination

There is still no clear antigen profile for CSCs. However, it is well known that some antigens are preferentially expressed by CSCs and others by differentiated cancer cells, while a third group is expressed by both CSCs and non-CSCs (5). Tumor cells are characterized by a high degree of plasticity that allows them to dynamically switch between CSCs and differentiated cells (39). It is therefore crucial that vaccines specific for antigens expressed by differentiated cancer cells and overexpressed in CSCs are developed to eliminate both current and *de-novo* generated CSCs for complete tumor eradication (6). Several cancer testis (CT) antigens, such as NY-ESO-1 and some melanoma-associated antigen gene (MAGE) family members, which are expressed by differentiated cells and overexpressed in CSCs, are immunogenic and are good targets for anti-CSC vaccination (40). However, many CSC-specific antigens are self-proteins that are also expressed on normal stem cells, which significantly limits their use as targets for vaccination, as central tolerance would

lead to poor immune responses. Moreover, if vaccination were able to break tolerance, severe side effects and autoimmunity would be induced (41). This occurs for vaccines against stem-cell markers, such as CD44, aldehyde dehydrogenase, and CD133, whose use in patients is limited by safety concerns (42-46).

The recently demonstrated positive association between stemness and cancer-mutational burden in numerous solid tumors (21), has brought forwards the idea of developing neoantigen-based CSCtargeting vaccines. Although this is an interesting strategy, it should be noted that the correlation between tumor mutational load and response to immunotherapy is far from perfect, especially as only a minority of mutations lead to the generation of neoantigens (47). Moreover, mutations often occur in genes that do not play a pivotal role in either carcinogenesis or CSC maintenance, meaning that vaccination against many neoantigens may lead to the expansion of resistant clones that lack their expression, favoring cancer recurrence after initial shrinkage (48). Oncoantigens, i.e. antigens that play a pivotal role in cancer progression and CSC self-renewal, are a valid means with which to overcome this issue (49,50). In particular, oncoantigens expressed on cell surfaces are the most promising for the development of CSC-directed vaccines as they can be targeted by both T and B cell-mediated responses, thus enabling CSC elimination despite their downregulation of MHC I (10,51). Some oncoantigens expressed on CSC plasma membranes are currently giving promising results in preclinical models. For example, one DC-based vaccine targets integrin β 4, which is overexpressed in breast and colon cancers and plays a role in CSC self-renewal. The humoral and cellular immune responses elicited by this vaccine are effective in inhibiting tumor growth and spontaneous pulmonary metastases as they kill both CSCs and differentiated cancer cells (52). Cripto-1, a GPI-anchored membrane oncofetal protein that promotes CSC self-renewal, EMT and migration in melanoma and breast cancer, is another promising CSC oncoantigen (53). DNA vaccination against Cripto-1 decreased both tumor growth and lung metastases in preclinical models (53,54). Similarly, the cystine/glutamate antiporter xCT, which is overexpressed in CSCs from different solid tumors and plays a key role in the maintenance of their redox balance and

metabolism (55), is a good candidate for anti-cancer vaccination. Indeed, immunotargeting xCT using DNA-, viral vector- or virus-like particle-based vaccines induced a strong antibody response and, in some cases, a cytotoxic T-cell response that significantly protected mice from breast-cancer growth and metastasis (56-59).

EMT-associated altered glycosylation is another mechanism of oncoantigen generation, in addition to protein overexpression, in CSCs (60). Clinical trials with vaccines that target some aberrantly glycosylated CSC oncoantigens, such as mucin 1 (MUC1), are currently ongoing (61). Although a lack of response was observed in patients with a history of premalignant lesions and who displayed elevated levels of circulating MDSCs, encouraging results were obtained in less immunosuppressed patients (62).

3. Combination therapies

The results obtained from the MUC1-vaccine trials suggest that, like all monotherapies, single immunotherapy is insufficient for cancer treatment, and that combinatorial approaches are needed (**Figure 1**). Several studies on preclinical models of melanoma, breast, colon and head and neck squamous cell carcinomas (HNSCC) have shown that combining CSC-targeting vaccines with ICIs induces increased activation of tumor-specific CD8⁺ T cells, decreased CSC frequency and better tumor-progression control than single treatments, paving the way for clinical experimentation (33,52,63,64).

Besides the use of ICIs, strategies to revert TME immunosuppressive activity may improve the efficacy of anti-CSC vaccination. Tadalafil is an inhibitor of phosphodiesterase-5, which alters the TME by reducing the number of MDSCs and Tregs and thus promotes tumor immunity, and is a possible candidate (65). Interim results from a phase I clinical trial (NCT02544880) in patients with primary HNSCC treated with tadalafil and a MUC1 vaccine indicate that the combination therapy was well tolerated and able to decrease the number of PD-L1⁺ macrophages and increase that of

activated tumor-infiltrating T cells. However, PD-L1 upregulation was observed on tumor cells, suggesting that a combination of more than two therapies may be needed (62).

The administration of multiple vaccines that target oncoantigens that are involved in different cell processes is a good strategy with which to attack cancer on multiple fronts. We have recently demonstrated that a combination of two vaccines that target HER2 and xCT exerts a synergistic effect in preclinical models of HER2⁺ breast cancer, with HER2 immunotargeting slowing primary-tumor growth and xCT immunotargeting mainly affecting CSCs and inhibiting metastases (66). Combining CSC-targeting vaccines with non-CSC-targeted treatments is an additional approach that can potentially induce tumor shrinkage and prevent metastasis and recurrence. CSC-vaccination may be successfully combined with chemo- and radiotherapy or with oncolytic viruses, which target differentiated cancer cells and further activate the immune response through the induction of immunogenic cell death, inducing the release of other tumor antigens (67-69). A phase I clinical trial (NCT00179309) has demonstrated that the combination of a MUC1-targeting vaccine and chemotherapy significantly improved progression-free survival, compared to chemotherapy alone, in breast-cancer patients (70).

Combining CSC immunotargeting with epigenetic drugs, which augment CSC immunogenicity by upregulating MHC and antigen-processing machinery (71), and decrease MDSC and Treg numbers (72), is a further way to improve treatment efficacy. Histone deacetylases and DNA methyltransferase (DNMT) inhibitors are currently used in clinical trials as they hinder tumor-cell growth and induce cell differentiation (73). Clinical trials performed on patients with ovarian (NCT01673217) and pediatric brain tumors (NCT02332889, NCT01241162) that were treated with the DNMT inhibitor decitabine, in association with a NY-ESO-1 peptide vaccine or a DC-based vaccine that targeted MAGE-A1, MAGE-A3 and NY-ESO-1, demonstrated that these regimens are safe (74,75). In 6 out of 10 patients treated with decitabine plus the NY-ESO-1 peptide, specific antibodies and T-cell responses either led to disease stabilization or partial clinical response (74).

Conclusions

Although ICIs have founded a new golden age for cancer immunotherapy, not all tumors are immunogenic, and several resistance mechanisms hinder ICI efficacy. As CSCs are an obstacle to ICI success, vaccines directed towards CSC oncoantigens may improve immunotherapy efficacy. Deeper genomic, biological and immunological characterization of CSCs, and their crosstalk with the immune system, is crucial if we are to address the difficulties associated with CSC heterogeneity and plasticity, and will lay the foundations for the development of novel combination therapies for the eradication of cancer. Acknowledgements: We thank Dr. Dale Lawson for his revision and editing of the manuscript.

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References

- 1. Maeng HM, Berzofsky JA. Strategies for developing and optimizing cancer vaccines. F1000Research **2019**;8 doi 10.12688/f1000research.18693.1.
- 2. Anandappa AJ, Wu CJ, Ott PA. Directing Traffic: How to Effectively Drive T Cells into Tumors. Cancer discovery **2020**;10(2):185-97 doi 10.1158/2159-8290.CD-19-0790.
- 3. Kissick HT, Sanda MG. The role of active vaccination in cancer immunotherapy: lessons from clinical trials. Current opinion in immunology **2015**;35:15-22 doi 10.1016/j.coi.2015.05.004.
- 4. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, *et al.* Sipuleucel-T immunotherapy for castration-resistant prostate cancer. The New England journal of medicine **2010**;363(5):411-22 doi 10.1056/NEJMoa1001294.
- 5. Ruiu R, Tarone L, Rolih V, Barutello G, Bolli E, Riccardo F, *et al.* Cancer stem cell immunology and immunotherapy: Harnessing the immune system against cancer's source. Progress in molecular biology and translational science **2019**;164:119-88 doi 10.1016/bs.pmbts.2019.03.008.
- 6. Quaglino E, Conti L, Cavallo F. Breast cancer stem cell antigens as targets for immunotherapy. Seminars in immunology **2020**:101386 doi 10.1016/j.smim.2020.101386.
- 7. Cordero F, Beccuti M, Fornari C, Lanzardo S, Conti L, Cavallo F, *et al.* Multi-level model for the investigation of oncoantigen-driven vaccination effect. BMC bioinformatics **2013**;14 Suppl 6:S11 doi 10.1186/1471-2105-14-S6-S11.
- 8. Schatton T, Frank MH. Antitumor immunity and cancer stem cells. Annals of the New York Academy of Sciences **2009**;1176:154-69 doi 10.1111/j.1749-6632.2009.04568.x.
- 9. Najafi M, Mortezaee K, Majidpoor J. Cancer stem cell (CSC) resistance drivers. Life sciences **2019**;234:116781 doi 10.1016/j.lfs.2019.116781.
- Tallerico R, Conti L, Lanzardo S, Sottile R, Garofalo C, Wagner AK, *et al.* NK cells control breast cancer and related cancer stem cell hematological spread. Oncoimmunology 2017;6(3):e1284718 doi 10.1080/2162402X.2017.1284718.
- Tallerico R, Todaro M, Di Franco S, Maccalli C, Garofalo C, Sottile R, *et al.* Human NK cells selective targeting of colon cancer-initiating cells: a role for natural cytotoxicity receptors and MHC class I molecules. J Immunol **2013**;190(5):2381-90 doi 10.4049/jimmunol.1201542.
- 12. Schatton T, Schutte U, Frank NY, Zhan Q, Hoerning A, Robles SC, *et al.* Modulation of T-cell activation by malignant melanoma initiating cells. Cancer Res **2010**;70(2):697-708 doi 10.1158/0008-5472.CAN-09-1592.

- 13. Conti L, Ruiu R, Barutello G, Macagno M, Bandini S, Cavallo F, *et al.* Microenvironment, oncoantigens, and antitumor vaccination: lessons learned from BALB-neuT mice. Biomed Res Int **2014**;2014:534969 doi 10.1155/2014/534969.
- Malta TM, Sokolov A, Gentles AJ, Burzykowski T, Poisson L, Weinstein JN, *et al.* Machine Learning Identifies Stemness Features Associated with Oncogenic Dedifferentiation. Cell 2018;173(2):338-54 e15 doi 10.1016/j.cell.2018.03.034.
- 15. Conti L, Lanzardo S, Arigoni M, Antonazzo R, Radaelli E, Cantarella D, *et al.* The noninflammatory role of high mobility group box 1/Toll-like receptor 2 axis in the self-renewal of mammary cancer stem cells. FASEB J **2013**;27(12):4731-44 doi 10.1096/fj.13-230201.
- 16. Lodyga M, Hinz B. TGF-beta1 A truly transforming growth factor in fibrosis and immunity. Seminars in cell & developmental biology **2020**;101:123-39 doi 10.1016/j.semcdb.2019.12.010.
- 17. Levina V, Marrangoni AM, Demarco R, Gorelik E, Lokshin AE. Drug-Selected Human Lung Cancer Stem Cells : Cytokine Network , Tumorigenic and Metastatic Properties. PLoS ONE **2008**;3 doi 10.1371/journal.pone.0003077.
- Di Tomaso T, Mazzoleni S, Wang E, Sovena G, Clavenna D, Franzin A, *et al.* Immunobiological characterization of cancer stem cells isolated from glioblastoma patients. Clin Cancer Res 2010;16(3):800-13 doi 10.1158/1078-0432.CCR-09-2730.
- 19. Wei J, Barr J, Kong LY, Wang Y, Wu A, Sharma AK, *et al.* Glioblastoma cancer-initiating cells inhibit T-cell proliferation and effector responses by the signal transducers and activators of transcription 3 pathway. Mol Cancer Ther **2010**;9(1):67-78 doi 10.1158/1535-7163.MCT-09-0734.
- 20. Plaks V, Kong N, Werb Z. The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? Cell Stem Cell **2015**;16(3):225-38 doi 10.1016/j.stem.2015.02.015.
- 21. Miranda A, Hamilton PT, Zhang AW, Pattnaik S, Becht E, Mezheyeuski A, *et al.* Cancer stemness, intratumoral heterogeneity, and immune response across cancers. Proc Natl Acad Sci U S A **2019**;116(18):9020-9 doi 10.1073/pnas.1818210116.
- 22. Castagnoli L, De Santis F, Volpari T, Vernieri C, Tagliabue E, Di Nicola M, *et al.* Cancer Stem Cells: Devil or Savior-Looking behind the Scenes of Immunotherapy Failure. Cells **2020**;9(3) doi 10.3390/cells9030555.
- 23. Wu Y, Chen M, Wu P, Chen C, Xu ZP, Gu W. Increased PD-L1 expression in breast and colon cancer stem cells. Clin Exp Pharmacol Physiol **2017**;44(5):602-4 doi 10.1111/1440-1681.12732.
- 24. Gupta HB, Clark CA, Yuan B, Sareddy G, Pandeswara S, Padron AS, *et al.* Tumor cellintrinsic PD-L1 promotes tumor-initiating cell generation and functions in melanoma and ovarian cancer. Signal transduction and targeted therapy **2016**;1 doi 10.1038/sigtrans.2016.30.
- 25. Almozyan S, Colak D, Mansour F, Alaiya A, Al-Harazi O, Qattan A, *et al.* PD-L1 promotes OCT4 and Nanog expression in breast cancer stem cells by sustaining PI3K/AKT pathway activation. Int J Cancer **2017**;141(7):1402-12 doi 10.1002/ijc.30834.
- 26. Cioffi M, Trabulo S, Hidalgo M, Costello E, Greenhalf W, Erkan M, *et al.* Inhibition of CD47 Effectively Targets Pancreatic Cancer Stem Cells via Dual Mechanisms. Clin Cancer Res **2015**;21(10):2325-37 doi 10.1158/1078-0432.CCR-14-1399.
- 27. Zhang B, Dang J, Ba D, Wang C, Han J, Zheng F. Potential function of CTLA-4 in the tumourigenic capacity of melanoma stem cells. Oncology letters **2018**;16:6163-70 doi 10.3892/ol.2018.9354.
- 28. Guo M, Luo B, Pan M, Li M, Zhao F, Dou J. MUC1 plays an essential role in tumor immunity of colorectal cancer stem cell vaccine. International immunopharmacology **2020**;85:106631 doi 10.1016/j.intimp.2020.106631.

- 29. Jachetti E, Mazzoleni S, Grioni M, Ricupito A, Brambillasca C, Generoso L, *et al.* Prostate cancer stem cells are targets of both innate and adaptive immunity and elicit tumor-specific immune responses. Oncoimmunology **2013**:1-12.
- 30. Li X, Zhang Z, Lin G, Gao Y, Yan Z, Yin H, *et al.* Antigen-specific T cell response from dendritic cell vaccination using side population cell-associated antigens targets hepatocellular carcinoma. Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine **2016**;37(8):11267-78 doi 10.1007/s13277-016-4935-z.
- 31. Dashti A, Ebrahimi M, Hadjati J, Memarnejadian A, Moazzeni SM. Dendritic cell based immunotherapy using tumor stem cells mediates potent antitumor immune responses. Cancer Lett **2016**;374(1):175-85 doi 10.1016/j.canlet.2016.01.021.
- 32. Ning N, Pan Q, Zheng F, Teitz-Tennenbaum S, Egenti M, Yet J, *et al.* Cancer stem cell vaccination confers significant antitumor immunity. Cancer Res **2012**;72(7):1853-64 doi 10.1158/0008-5472.CAN-11-1400.
- 33. Hu Y, Lu L, Xia Y, Chen X, Chang AE, Hollingsworth RE, *et al.* Therapeutic Efficacy of Cancer Stem Cell Vaccines in the Adjuvant Setting. Cancer Res **2016**;76(16):4661-72 doi 10.1158/0008-5472.CAN-15-2664.
- 34. Lu L, Tao H, Chang AE, Hu Y, Shu G, Chen Q, *et al.* Cancer stem cell vaccine inhibits metastases of primary tumors and induces humoral immune responses against cancer stem cells. Oncoimmunology **2015**;4(3):e990767 doi 10.4161/2162402X.2014.990767.
- 35. Lin M, Xu S-yLK-c, Feng Z-pL, Yuan MY-y. Safety and efficacy study of lung cancer stem cell vaccine. Immunologic Research **2015**:16-22 doi 10.1007/s12026-015-8631-7.
- 36. Lin M, Ying Y, Shu Y, Liu P, Juan J, Xin S, *et al.* Prospective study of the safety and efficacy of a pancreatic cancer stem cell vaccine. Journal of cancer research and clinical oncology **2015**:1827-33 doi 10.1007/s00432-015-1968-4.
- 37. Vik-Mo E, Nyakas M, Mikkelsen B, Moe M, Due-Tonnesen P, Langmoen I. Therapeutic vaccination against autologous cancer stem cells with mRNA-transfected dendritic cells in patients with glioblastoma. Cancer Immunology Immunotherapy **2013**:1499-509 doi 10.1007/s00262-013-1453-3.
- 38. Czerwinska P, Rucinski M, Włodarczyk N, Jaworska A, Grzadzielewska I, Gryska K, et al. Therapeutic melanoma vaccine with cancer stem cell phenotype represses exhaustion and maintains antigen-specific T cell stemness by up-regulating BCL6. Oncoimmunology 2020;9(1):1710063 doi 10.1080/2162402X.2019.1710063.
- 39. Chen W, Dong J, Haiech J, Kilhoffer MC, Zeniou M. Cancer stem cell quiescence and plasticity as major challenges in cancer therapy. Stem cells international **2016**;2016 doi 10.1155/2016/1740936.
- 40. Hirohashi Y, Torigoe T, Tsujahara T, Kanaseki T, Kochin V, Sato N. Immune responses to human cancer stem-like cells/cancer-initiating cells. Cancer science **2016** doi 10.1111/cas.12830.
- 41. Maccalli C, Parmiani G, Ferrone S. Immunomodulating and Immunoresistance Properties of Cancer-Initiating Cells: Implications for the Clinical Success of Immunotherapy. Immunological investigations **2017**;46(3):221-38 doi 10.1080/08820139.2017.1280051.
- 42. Wu X, Yin T, Tian J, Tang C, Huang J, Zhao Y, *et al.* Distinctive effects of CD34- and CD133-specific antibody-coated stents on re-endothelialization and in-stent restenosis at the early phase of vascular injury. Regenerative biomaterials **2015**;2(2):87-96 doi 10.1093/rb/rbv007.
- 43. Deng Z, Wu Y, Ma W, Zhang S, Zhang YQ. Adoptive T-cell therapy of prostate cancer targeting the cancer stem cell antigen EpCAM. BMC immunology **2015**;16:1 doi 10.1186/s12865-014-0064-x.

- 44. Zhang YH, Wang ZY, Hao FY, Zhang L. Cluster of differentiation 24 monoclonal antibody induces apoptosis in the osteosarcoma cells. European review for medical and pharmacological sciences **2014**;18(14):2038-41.
- 45. Wallach-Dayan SB, Rubinstein AM, Hand C, Breuer R, Naor D. DNA vaccination with CD44 variant isoform reduces mammary tumor local growth and lung metastasis. Mol Cancer Ther **2008**;7(6):1615-23 doi 10.1158/1535-7163.MCT-07-2383.
- 46. Parmiani G, Russo V, Maccalli C, Parolini D, Rizzo N, Maio M. Peptide-based vaccines for cancer therapy. Human vaccines & immunotherapeutics **2014**;10(11):3175-8 doi 10.4161/hv.29418.
- 47. Schumacher TN, Scheper W, Kvistborg P. Cancer Neoantigens. Annu Rev Immunol **2019**;37:173-200 doi 10.1146/annurev-immunol-042617-053402.
- 48. Jiang T, Shi T, Zhang H, Hu J, Song Y, Wei J, *et al.* Tumor neoantigens: from basic research to clinical applications. J Hematol Oncol **2019**;12(1):93 doi 10.1186/s13045-019-0787-5.
- 49. Cavallo F, Calogero RA, Forni G. Are oncoantigens suitable targets for anti-tumour therapy? Nature Reviews Cancer **2007**;7:707-13 doi 10.1038/nrc2208.
- 50. Lollini PL, Cavallo F, Nanni P, Forni G. Vaccines for tumour prevention. Nat Rev Cancer **2006**;6(3):204-16 doi 10.1038/nrc1815.
- 51. Iezzi M, Quaglino E, Amici A, Lollini PL, Forni G, Cavallo F. DNA vaccination against oncoantigens: A promise. Oncoimmunology **2012**;1(3):316-25 doi 10.4161/onci.19127.
- 52. Ruan S, Lin M, Zhu Y, Lum L, Thakur A, Jin R, *et al.* Integrin beta4-Targeted Cancer Immunotherapies Inhibit Tumor Growth and Decrease Metastasis. Cancer Res **2020**;80(4):771-83 doi 10.1158/0008-5472.CAN-19-1145.
- 53. Witt K, Ligtenberg MA, Conti L, Lanzardo S, Ruiu R, Wallmann T, *et al.* Cripto-1 plasmid DNA vaccination targets metastasis and cancer stem cells in murine mammary carcinoma. Cancer immunology research **2018**;1:canimm.0572.2017 doi 10.1158/2326-6066.CIR-17-0572.
- 54. Ligtenberg MA, Witt K, Galvez-Cancino F, Sette A, Lundqvist A, Lladser A, *et al.* Cripto-1 vaccination elicits protective immunity against metastatic melanoma. Oncoimmunology **2016**;5(5):e1128613 doi 10.1080/2162402X.2015.1128613.
- 55. Ruiu R, Rolih V, Bolli E, Barutello G, Riccardo F, Quaglino E, *et al.* Fighting breast cancer stem cells through the immune-targeting of the xCT cystine–glutamate antiporter. Cancer Immunology, Immunotherapy **2018**;0:1-11 doi 10.1007/s00262-018-2185-1.
- 56. Lanzardo S, Conti L, Rooke R, Ruiu R, Accart N, Bolli E, *et al.* Immunotargeting of Antigen xCT Attenuates Stem-like Cell Behavior and Metastatic Progression in Breast Cancer. Cancer Res **2016**;76(1):62-72 doi 10.1158/0008-5472.CAN-15-1208.
- 57. Donofrio G, Tebaldi G, Lanzardo S, Ruiu R, Bolli E, Ballatore A, *et al.* Bovine herpesvirus 4-based vector delivering the full length xCT DNA efficiently protects mice from mammary cancer metastases by targeting cancer stem cells. Oncoimmunology **2018**;7(12):e1494108 doi 10.1080/2162402X.2018.1494108.
- 58. Bolli E, O'Rourke JP, Conti L, Lanzardo S, Rolih V, Christen JM, *et al.* A Virus-Like-Particle immunotherapy targeting Epitope-Specific anti-xCT expressed on cancer stem cell inhibits the progression of metastatic cancer in vivo. Oncoimmunology **2017**:e1408746 doi 10.1080/2162402X.2017.1408746.
- 59. Rolih V, Caldeira J, Bolli E, Salameh A, Conti L, Barutello G, *et al.* Development of a VLP-Based Vaccine Displaying an xCT Extracellular Domain for the Treatment of Metastatic Breast Cancer. Cancers **2020**;12(6) doi 10.3390/cancers12061492.
- 60. Barkeer S, Chugh S, Batra SK, Ponnusamy MP. Glycosylation of Cancer Stem Cells: Function in Stemness, Tumorigenesis, and Metastasis. Neoplasia **2018**;20(8):813-25 doi 10.1016/j.neo.2018.06.001.

- 61. Guo M, You C, Dou J. Role of transmembrane glycoprotein mucin 1 (MUC1) in various types of colorectal cancer and therapies: Current research status and updates. Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie **2018**;107:1318-25 doi 10.1016/j.biopha.2018.08.109.
- 62. Weed DT, Zilio S, Reis IM, Sargi Z, Abouyared M, Gomez-Fernandez CR, *et al.* The Reversal of Immune Exclusion Mediated by Tadalafil and an Anti-tumor Vaccine Also Induces PDL1 Upregulation in Recurrent Head and Neck Squamous Cell Carcinoma: Interim Analysis of a Phase I Clinical Trial. Frontiers in immunology **2019**;10:1206 doi 10.3389/fimmu.2019.01206.
- 63. Zheng F, Dang J, Zhang H, Xu F, Ba D, Zhang B, *et al.* Cancer Stem Cell Vaccination With PD-L1 and CTLA-4 Blockades Enhances the Eradication of Melanoma Stem Cells in a Mouse Tumor Model. Journal of Immunotherapy **2018** doi 10.1097/CJI.0000000000242.
- 64. Remy-Ziller C, Thioudellet C, Hortelano J, Gantzer M, Nourtier V, Claudepierre MC, *et al.* Sequential administration of MVA-based vaccines and PD-1/PD-L1-blocking antibodies confers measurable benefits on tumor growth and survival: Preclinical studies with MVAbetaGal and MVA-MUC1 (TG4010) in a murine tumor model. Human vaccines & immunotherapeutics **2018**;14(1):140-5 doi 10.1080/21645515.2017.1373921.
- 65. Weed DT, Vella JL, Reis IM, De la Fuente AC, Gomez C, Sargi Z, *et al.* Tadalafil reduces myeloid-derived suppressor cells and regulatory T cells and promotes tumor immunity in patients with head and neck squamous cell carcinoma. Clin Cancer Res **2015**;21(1):39-48 doi 10.1158/1078-0432.CCR-14-1711.
- 66. Conti L, Bolli E, Di Lorenzo A, Franceschi V, Macchi F, Riccardo F, *et al.* Immunotargeting of the xCT cystine/glutamate antiporter potentiates the efficacy of Her2targeted immunotherapies in breast cancer. Cancer immunology research **2020** doi 10.1158/2326-6066.CIR-20-0082.
- 67. Goto T. Radiation as an In Situ Auto-Vaccination: Current Perspectives and Challenges. Vaccines **2019**;7(3) doi 10.3390/vaccines7030100.
- 68. Bracci L, Schiavoni G, Sistigu A, Belardelli F. Immune-based mechanisms of cytotoxic chemotherapy: implications for the design of novel and rationale-based combined treatments against cancer. Cell death and differentiation **2014**;21:15-25 doi 10.1038/cdd.2013.67.
- Nguyen T, Avci NG, Shin DH, Martinez-Velez N, Jiang H. Tune Up In Situ Autovaccination against Solid Tumors with Oncolytic Viruses. Cancers 2018;10(6) doi 10.3390/cancers10060171.
- 70. Heery CR, Ibrahim NK, Arlen PM, Mohebtash M, Murray JL, Koenig K, *et al.* Docetaxel Alone or in Combination With a Therapeutic Cancer Vaccine (PANVAC) in Patients With Metastatic Breast Cancer: A Randomized Clinical Trial. JAMA oncology **2015**;1(8):1087-95 doi 10.1001/jamaoncol.2015.2736.
- 71. Roca MS, Di Gennaro E, Budillon A. Implication for Cancer Stem Cells in Solid Cancer Chemo-Resistance: Promising Therapeutic Strategies Based on the Use of HDAC Inhibitors. Journal of clinical medicine **2019**;8(7) doi 10.3390/jcm8070912.
- 72. Bae J, Hideshima T, Tai YT, Song Y, Richardson P, Raje N, *et al.* Histone deacetylase (HDAC) inhibitor ACY241 enhances anti-tumor activities of antigen-specific central memory cytotoxic T lymphocytes against multiple myeloma and solid tumors. Leukemia **2018**;32(9):1932-47 doi 10.1038/s41375-018-0062-8.
- 73. Eckschlager T, Plch J, Stiborova M, Hrabeta J. Histone Deacetylase Inhibitors as Anticancer Drugs. Int J Mol Sci **2017**;18(7) doi 10.3390/ijms18071414.
- 74. Odunsi K, Matsuzaki J, James SR, Mhawech-Fauceglia P, Tsuji T, Miller A, *et al.* Epigenetic potentiation of NY-ESO-1 vaccine therapy in human ovarian cancer. Cancer immunology research **2014**;2(1):37-49 doi 10.1158/2326-6066.CIR-13-0126.

75. Krishnadas DK, Shusterman S, Bai F, Diller L, Sullivan JE, Cheerva AC, *et al.* A phase I trial combining decitabine/dendritic cell vaccine targeting MAGE-A1, MAGE-A3 and NY-ESO-1 for children with relapsed or therapy-refractory neuroblastoma and sarcoma. Cancer Immunol Immunother **2015**;64(10):1251-60 doi 10.1007/s00262-015-1731-3.



Figure 1. Combination therapies based on anti-CSC vaccination. Schematic representation of the effects exerted by combining CSC-oncoantigen-directed vaccines with chemotherapy, radiotherapy and oncolytic therapies, ICIs, vaccines to other oncoantigens and epigenetic drugs. All these combination therapies have the potential to induce the elimination of both CSCs and differentiated cancer cells, finally leading to cancer eradication.