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Applications of cyclodextrins in food science. A review

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(Article begins on next page)

1 **APPLICATIONS OF CYCLODEXTRINS IN FOOD**
2 **SCIENCE. A REVIEW**

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24 **ABSTRACT**

25 Background: The food industry is constantly attempting to develop better
26 products that will have a positive effect on health (commonly known as functional
27 foods). In this respect, cyclodextrins (CDs) could be of interest because they are
28 tasteless, non-caloric and odourless molecules with several valuable
29 characteristics, such as a capacity to separate chiral compounds and solubilize
30 or stabilize bioactive compounds (BaC).

31 Scope and Approach: This review represents a revision of the state-of-the-art of
32 CDs and their uses in the food industry.

33 Key Findings and Conclusions: We analysed their metabolism and regulatory
34 aspects of current applications of CDs: as carriers, for removing components, to
35 produce or extract BaC, their use as nanosensors or in food packaging. We study
36 how inclusion complexes are formed referring to the most common techniques
37 and parameters. Moreover, how inclusion complexes are formed will be studied
38 with reference to the most common techniques and parameters. In conclusion,
39 their applications in the food and other industries will increase in the coming years
40 without a doubt.

41

42 **Keywords**: Cyclodextrins; Food industry; nutraceuticals; packaging; “free
43 products”; nanosensor.

44

45

46 **HIGHLIGHTS**

47

- 48 1. The paper reviews the state-of-the-art of CDs as used in the food
49 industry.
50 2. The mechanism, metabolism, pharmacokinetics and regulatory aspects
51 were discussed.
52 3. Different applications are reviewed, including formulations, production
53 and packaging.
54 4. Products containing CDs that are already on the market are described.

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81 1. INTRODUCTION

82 Antoine Villiers would not have suspected the number of applications of
83 their cyclodextrins (CDs) that would be discovered in just over one hundred years
84 (Antoine Villiers, 1891). CDs are a torus-shaped oligosaccharides made up of α -
85 (1,4) linked glucose units, obtained from the degradation of starch by the enzyme
86 Cyclodextrin Glucosyltransferase (CGTase). Although the most common CDs
87 are the natural α , β and γ - forms (**Fig. 1A**, which contain six, seven and eight
88 glucose units, respectively), CDs containing up to nineteen units have been
89 characterized (French et al., 1965; Pulley, 1961). However, these novel CDs are
90 not used commonly because of their propensity to collapse. Even smaller CDs
91 with 3 or 4 glucose units have recently been synthesized with the same type of
92 linkage of natural CDs (Ikuta et al., 2019).

93 The CD ring is a conical cylinder of an amphiphilic nature, with a
94 hydrophilic outer part (formed by the hydroxyl groups) and a predominantly
95 lipophilic cavity, which can contain water (Pereva et al., 2019). Although both
96 inorganic and organic salts and neutral molecules can form complexes with CDs
97 (Fourmentin, Crini, & Lichtfouse, 2018), they are more generally used to complex
98 (**Fig. 1B**) poorly-soluble drugs or bioactive compounds (BaC), creating so-called
99 "inclusion complexes", novel BaC/CD nanoparticles. In solution, the equilibrium
100 between free and complexed BaC is dynamic, meaning that when the solution is
101 diluted or another BaC is added, the first BaC is easily released.

102 In general, the generation of CD/BaC nanoparticles improves the apparent
103 solubility of the drug, which increases its final concentration and possibly its
104 bioactivities (Shieh & Hedges, 1996). Among previously cited benefits of using
105 CDs are:

- 106 1. Increased stability under abiotic (light, oxidants...) or biotic (enzymatic
107 degradation) conditions.
- 108 2. The ability to control the release of molecules (e.g. volatiles).
- 109 3. The elimination of undesirable tastes and odours.
- 110 4. Chiral separation.
- 111 5. Controlling chemical reactions.
- 112 6. The selective purification of molecules.
- 113 7. Dietary fibre(EFSA, 2012) .

114 To improve the properties of CDs, different chemically obtained derivatives
115 (e.g. Hydroxypropyl- β -CD or Methyl- β -CD) and materials containing CDs (e.g.
116 CD-nanosponges or CD-based nanoparticles) have been obtained. Although
117 these have improved capacities compared with natural CDs(Salazar et al., 2018;
118 Sherje et al., 2017) even Hydroxypropyl- β -CD is considered *orphan drug* for
119 Niemann Pick type C (Matencio, Navarro-Orcajada, González-Ramón, et al.,
120 2020), only natural CDs (Szente&Szejtli, 2004) are permitted as food additives
121 (E-457, E-458 and E-459).

122 Due to their interesting applications in the pharmaceutical and
123 nutraceutical industries and similar fields, the number of publications on CDs has
124 rapidly increased, for example only in 2018, 1530 research articles were
125 published with CDs in the title and the word “cyclodextrin” appears in 4800
126 patents. Furthermore, Jansook et al. (2018) have published that the CD annual
127 production is over 10,000 metric tonnes (information from Roquette, France), of
128 which 20% is used in food applications (Jansook, Ogawa, &Loftsson, 2018). Such
129 data attest to the importance of CDs in the food industry.

130 In spite of the importance of CDs in food, the last exhaustive review on
131 their use in food science was published in 2009 (Astray et al., 2009), with partial
132 updates (dos Santos et al., 2017; Fenyvesi et al., 2016a). Although, a review
133 focusing on beverages was published in 2020(Astray et al., 2020), there is a clear
134 need for a fresh review of the uses and applications of CDs in food science. For
135 this reason, the purpose of the present contribution is to provide, a general
136 overview of the use of CDs and modified CDs in the food and nutraceutical
137 industries, with special attention paid to functional food production.

138 **2. METABOLISM, PHARMACOKINETICS AND TOXICOLOGY**

139 In nature, only CGTase is able to convert starch into CDs, although other
140 enzymes can help in their industrial production (Biber, Antranikian, &Heinzle,
141 2002; Tonkova, 1998).However, various enzymes or processes in our body may
142 degrade CDs into glucose derivatives: Starting in the mouth (when you eat
143 something containing CDs),salivary α -amylase can rapidly hydrolyse dextrans,
144 although rapid transport to the stomach means that the degree of degradation is
145 insignificant. Of the three natural CDs (α -, β - and γ -CD), the first two are
146 essentially stable towards α -amylase, and γ -CD is rapidly digested(John Marshall
147 & Miwa, 1981; Kurkov & Loftsson, 2013).

148 Unspecific pH-dependent degradation can occur in the stomach; however,
149 inclusion complexes present lower degradation (Singh et al., 2010). After the
150 stomach, in the neutral pH environment of the small intestine pancreatic amylase
151 continues the hydrolysis reaction. While α - and β -CDs are mainly digested by
152 bacteria in the colon (where α -CD is degraded more rapidly than β -CD), γ -CD is
153 almost completely digested in the gastrointestinal tract. Finally, undigested CDs
154 are metabolized by microbiota in the lower section of the digestive system, where

155 they are degraded almost completely, a fact that has led to their being used as
156 prebiotics (Fenyvesi et al., 2016a). The rest is eliminated in the faeces. In general,
157 the bioavailability of natural and common CDs derivatives is very low, which
158 makes them safe when administered orally (Arima et al., 2011; Jansook et al.,
159 2018; Kurkov & Loftsson, 2013).

160 Dextrins can also be administered parenterally, and study of their
161 pharmacokinetics has led to the publication of monographs such as the European
162 Pharmacopoeia (*European Pharmacopoeia (Ph. Eur.) 9th Edition | EDQM*). The
163 urinary clearance of linear dextrin and CDs decreases with increasing molecular
164 weight. Indeed, molecules of less than 15 KDa are almost totally excreted (≥ 90
165 %) by urine without much modification (Kurkov&Loftsson, 2013). The
166 pharmacokinetics of i) HP β -CD, ii) sulfobutylether β -CD sodium salt and iii)
167 Sugammadex sodium salt has been studied in rats, in which $t_{1/2}$ values of 1.9, 1.6
168 and 1.7, respectively were found. More than 90% of CD is excreted by urine within
169 24h; although the kidneys of affected subjects may maintain CD levels for longer
170 periods (Frijlink et al., 1990; Hamilton et al., 2018; Jansook et al., 2018; Kurkov
171 & Loftsson, 2013; Luke et al., 2012; Matencio, Alcaráz-Gómez, et al., 2018).

172 As regards the toxicity of CDs, most studies are concerned with their
173 medical application. A high orally administered dose of CDs could generate
174 diarrhoea and caecum enlargement, and even affect to the bioavailability of some
175 substances, to prevent which, the European Commission has published a guide
176 to help scientists in drug development (EMA, 2017). On the other hand, the
177 toxicology of HP β -CD has been studied in greater depth due to its classical use
178 as a medical excipient (Gould & Scott, 2005), and even its degree of substitution
179 has been studied (P. Li et al., 2016). The results of the last study showed that the

180 best option to lower toxicity levels would be to use at lower degrees of
181 substitution, although more studies are necessary in this respect.

182 **3. REGULATORY OF CDS IN FOOD SCIENCE.**

183 Before we approach our last objective using CDs- for human consumption
184 and as nutraceuticals - we should first take a look at the current regulations that
185 govern their use, including the quantity and type of CDs permitted. When used
186 as supplements in food products, natural CDs are considered food additives (E-
187 457, E-458 and E-459) and recognized as “Generally recognized as safe”
188 (GRAS). The recommendation of the Joint FAO/WHO Expert Committee on Food
189 Additives (JECFA) established the maximum advisable level of β -CD in foods at
190 5 mg/kg/day. On the other hand, there is no Acceptable Daily Intake (ADI) for α -
191 and γ -CD due to their favourable toxicological profiles. The European Food
192 Safety Authority (EFSA) permitted the health status claimed for α -CD dietary
193 fibre, and its ability to reduce of post-prandial glycaemic responses (EFSA, 2012).
194 In addition, the dosage of β -CD was re-evaluated in 2016 with no modifications
195 recommended (Mortensen et al., 2016). Furthermore, a recent review focused on
196 the safety and recommended levels of CDs in foods (Fenyvesi et al., 2016a) also
197 suggested no modification to the above levels was necessary. However, no
198 regulations apply to BaC/CD nanoparticles in food products. In these cases, the
199 regulation of both ingredients, BaC and CD, is considered. However, we think
200 that this should be re-considered.

201 When a drug is administrated orally, the excipient (in this case CD), must
202 be considered as an “active compound” and its safety must be evaluated
203 (Jansook et al., 2018). Although they are not directly tested, the formulation of
204 new drugs must be evaluated in the context of the application. Generally, GRAS

205 molecules are directly approved for use as excipient (in this case, natural CDs).
206 In this respect, the American Food and Drug administration (FDA) has published
207 a list of inactive pharmaceutical ingredients for downloading
208 (<https://www.fda.gov/Drugs/InformationOnDrugs/ucm113978.htm>). The list
209 indicates the route, dosage form and maximum concentration recommended. In
210 addition, the European Medicines Agency (EMA) has published several reports
211 on the use of CDs in pharmaceutical products
212 (<https://www.ema.europa.eu/en/cyclodextrins>), among others. A question and
213 answer document on CDs and their uses (EMA, 2017), provides information
214 about safety: for example, in general around 200 mg/kg/day of CD is
215 recommended for oral administration. Finally, The Japanese Pharmaceutical
216 Codex (JPC) has published several applications of CDs in its Pharmacopoeia (
217 医薬品医療機器レギュラトリーサイエンス財団, 2017).

218 4. FORMATION OF INCLUSION COMPLEXES

219 When a molecule enters the inner cavity of a CD a nanoparticle, known as
220 “inclusion complex”, is formed. This type of non-covalent interaction involves a
221 *host* and a *guest*, the latter of which is totally or partially included by the host. The
222 cavity volumes of α -, β - and γ -CD measured 0.174, 0.262 or 0.427 nm³,
223 respectively (Astray et al., 2009). The stability of the inclusion complex can be
224 described in terms of its complexation constant (K_F) or dissociation constant (K_D):



227 This is not easy to evaluate and sometimes requires a combination of
228 several techniques, as mentioned in two recent reviews (Mura, 2014, 2015), the

229 information they contained helping us to select the most interesting CDs for use
230 in our work.

231 The methods are generally based on the variations in any physical or
232 chemical property of the guest molecule that may be considered useful for
233 following complex formation (Table 1).

234 In recent years of chemoinformatic techniques such as molecular docking
235 or molecular dynamics have increased for their capacity to predict inclusion
236 complexes (Brocos et al., 2010; F. Garrido et al., 2020; Khuntawee et al., 2015;
237 Matencio, Hernández-García, et al., 2017; Matencio, Bermejo-Gimeno, et al.,
238 2017; Matencio et al., 2019; R. Wang et al., 2015), and interfaces that work with
239 CDs have been developed to facilitate input and output analysis (Montenegro
240 Rabello et al., 2019), and there are even libraries of computational molecular
241 dynamics simulations of cyclodextrins (Mixcoha et al., 2016).

242 In detail, molecular docking is a method which predicts the preferred
243 orientation of one molecule (host) towards a second molecule (guest), when they
244 bind to each other to form a stable complex. In turn, knowledge of the preferred
245 orientation may be used to predict the strength of association or binding affinity
246 between two molecules using, for example, scoring functions (Tao et al., 2020).
247 On the other hand, molecular dynamics is a computer simulation method for
248 analysing the physical movements of atoms and molecules. The atoms and
249 molecules can interact for a fixed period of time, revealing the dynamic "evolution"
250 of the system. In the most common version, the trajectories of atoms and
251 molecules are determined by numerically solving Newton's equations of motion
252 for a system of interacting particles, where forces between the particles and their

253 potential energies are often calculated using interatomic potentials or molecular
254 mechanics force fields (Durrant& McCammon, 2011).

255 **4.1 Cyclodextrin/guest mechanism, thermodynamic and structural** 256 **interactions**

257 The complexation generally induces the release of water from the cavity,
258 and any solvents or salts present might affect the complexation. Among the
259 different processes, steric hindrance, and the substitution of unfavourable
260 interactions by favourable new ones usually determine the energy state of the
261 nanoparticle. Interactions that may contribute to nanoparticle formation include
262 electrostatic/coulomb forces, van der Waals interactions, hydrogen bonding, and
263 hydrophobic effects, among others, although their impact depends on several
264 factors such as type of guest, the solvent used, etc. (Jansook et al., 2018;
265 Schönbeck & Holm, 2019).

266 Such changes are related to the classical thermodynamic parameters,
267 enthalpy, entropy and binding free energy. In general, attractive electrostatic and
268 van der Waals forces are assumed to result in enthalpy-driven binding, while
269 hydrophobic attraction seems to be characterized by a large gain in entropy.

270 Several CDs can be used to complex a guest, and so, the different cavities
271 of CDs and any modifications must be analysed. In general, β -CD and derivatives
272 are the most useful (Jansook et al., 2018). However, for the food industry only
273 the three natural CDs can be used, the cavity constituting the main difference
274 between them, although β -CD is usually the most used.

275 As regards the effect of polarity, the most important factors to consider are
276 structure, solvent and temperature. In general terms, hydrophobic molecules
277 form complexes with CDs, although neutral or polar molecules, ions or gases are

278 also able to form such complexes (Fourmentin et al., 2018b). The solvent is
279 important, since the presence of organic solvents might decrease the
280 complexation constant due to the higher solubility of the molecule (Charumanee
281 et al., 2016). On the other hand, the presence of a minimal quantity of water is
282 necessary to form the inclusion complex (Kamihira et al., 1990).

283 **5. APPLICATIONS OF CDS IN FOODS**

284 As mentioned above, the need for an update concerning the application of
285 CDs in food science has led us to provide some guidance for the community
286 involved. A selection of the most interesting applications is presented below.

287 **5.1 Production or extraction of bioactive compounds using CDs.**

288 The capacity of CDs to complex bioactive compounds can be used to
289 displace the Le Châtelier equilibrium or even to protect the freshly synthesized
290 compound.

291 It is common to use CDs in the production of bioactive compounds (**Fig.**
292 **2**) from plant cell cultures to act as elicitors and so obtain higher yields (García-
293 Pérez et al., 2019; Pinho et al., 2014). For example CDs are used to produce
294 polyphenols, such as stilbenes (Komaikul et al., 2019) or flavonoids (García-
295 Pérez et al., 2019), alone or, to take advantage of the resulting synergetic effects,
296 in combination with elicitors such as methyl jasmonate or salicylic acid (Perassolo
297 et al., 2016; Xu et al., 2015); indeed, the complex formed between methyl
298 jasmonate and salicylic acid with CDs have been evaluated (López-Nicolás et al.,
299 2013; Matencio, Bermejo-Gimeno, et al., 2017; Nishioka et al., 1984) and in
300 combination with the bioactive compounds produced in the cell culture (Oliva et
301 al., 2018). Additionally, the use of CDs has been extended to other types of
302 organism such as cyanobacteria in order to increase the production of antifungal

303 products (Shishido et al., 2015). The use of CDs in combination with enzymes
304 (Bonnet et al., 2010) has also been studied in recent years. For example, in one
305 study, using CDs, the biotechnological production of phenylethylamine-
306 betaxanthin and indoline-betacyanin, two of the most promising betalain
307 derivatives was studied obtaining higher production, around 51 and 26% of both
308 derivatives, respectively (Matencio, Guerrero-Rubio, Gandía-Herrero, et al.,
309 2020); while, CDs can also be used directly for chemical synthesis (Kokkiralala et
310 al., 2011), this is not the case with food products.

311 The use of CDs for the extraction of both hydrophobic and hydrophilic
312 substances has been studied (Gao et al., 2016). For example, polyphenols can
313 be extracted from many plants, including pomegranate or green pepper
314 (Athanasiadis et al., 2018; Diamanti et al., 2017; Favre et al., 2020) or catechins
315 (López-Miranda et al., 2016).

316 **5.2 Using CDs to produce “free” products and induce organoleptic** 317 **modifications.**

318 The growing tendency for consumers to look for “-free” products (sugar-
319 free, gluten-free, etc.) have highlighted the ability of CDs to remove some
320 compounds from a given food matrix (**Fig. 3**). For example, advantage is taken
321 of the capacity of CDs to complex cholesterol in order to extract it from food
322 products, thus creating low cholesterol products (Ahn&Kwak, 1999; Alonso et al.,
323 2019a, 2019b; Lee et al., 1999).

324 In addition, unpleasant flavours can be reduced by adding CDs: for
325 example the beany flavour of soy milk, and bitter components, and fish or goaty
326 flavours (Hadi et al., 2015; Serfert et al., 2010; Suratman et al., 2004; Ünlüsayin
327 et al., 2016; C. Wang et al., 2018). In addition, volatile fragrance complexes may

328 change the organoleptic profile of the product (Abril-Sánchez et al., 2019;
329 Ciobanu et al., 2013) because, although complexation prolongs the half-life of
330 volatiles in the matrix, the odour becomes less intense. Some allergens or toxins
331 could also be complexed – a potential that could be more widely used in industry
332 but which is not (Ayesh& Essa, 2002; Hou et al., 2015).

333 Due to the physicochemical properties of CDs, their use as emulsifiers is
334 a very promising application in the food industry. In particular, CDs have proved
335 to be effective in the formation of particle-stabilized emulsions, also known as
336 Pickering emulsions. In this sense, it has been described that oil/water Pickering
337 emulsions with natural CDs can provide less oxidized oil and fewer undesired
338 flavours (Moriyama et al., 2013). Soybean oil and kenaf seed oil are examples of
339 common oils that have been tested in this kind of emulsions with CDs (Cheong &
340 Nyam, 2016; Inoue et al., 2010). In a recent study (Xi et al., 2018), the amphiphilic
341 properties of β -CD were enhanced by esterification with octadecenyl succinic
342 anhydride leading to more stable emulsions. Furthermore, Pickering emulsions
343 made with CDs have been suggested for producing special foods for elderly
344 people with dysphagia (Moriyama et al., 2013).

345 **5.3 Formulation of CD nanoparticles as carriers of bioactive compounds for** 346 **functional foods.**

347 Every month studies appear on the subject of novel nanoparticles formed
348 by CDs containing drugs or bioactive compounds for possible use in functional
349 food. Since 2018, 1750 articles and patents have been published with the word
350 “cyclodextrin” in the title, pointing to the high potential of these molecules.

351 In general terms, CDs interact with bioactive compounds, thereby
352 enhancing their solubility or stability: for example, complexes of natural CDs with

353 antioxidants, vitamins, saponins, fatty acids or carotenoids (dos Santos et al.,
354 2017; dos Santos Lima et al., 2019; Fenyvesi et al., 2016a; López-Nicolás et al.,
355 2014; Matencio, Hernández-Gil, et al., 2017; Matencio, García-Carmona, et al.,
356 2017a). Of the most recently, formed nanoparticles, the most interesting
357 complexes and their possible applications will be discussed below.

358 In the case of vitamins, an interesting combinatorial use of the hydrophobic
359 Vitamin E and cholesterol complexed with CDs has been successfully tested to
360 improve sperm motility, cryopreservation and quality (Benhenia et al., 2016,
361 2018)and for reducing lipid peroxidation. Complexes with soluble vitamins
362 (vitamin C, nicotinic acid and vitamin L1) have also been evaluated (Saha et al.,
363 2016; Vázquez et al., 2019).An interesting application is the use of pH-responsive
364 CD nanoparticles to release compounds like polyunsaturated fatty acids
365 according to their pH(Xi et al., 2019). The pH-induced protonation/deprotonation
366 of carboxyl groups ensured that the emulsions remained steady at $\text{pH} \leq 4$ but were
367 unsteady under neutral conditions.

368 As regards antioxidants, a large number of publications have appeared in
369 recent years about nanoparticles formed by several antioxidants with CDs from
370 natural extracts (Rakmai et al., 2018) or complexed molecules from sources such
371 as polyphenols or essentials oils (Matencio et al., 2016; Matencio, García-
372 Carmona, et al., 2017b; Yildiz et al., 2018; Zhang et al., 2016). In general, the
373 complexation of antioxidants with CDs increases the total concentration of the
374 substance and, consequently, their antioxidant capacities (Celebioglu et al.,
375 2018; Das et al., 2019; López-Nicolás et al., 2014), which can be used by the
376 food industry to fortify products (Ho et al., 2017; Matencio, Navarro-Orcajada,
377 Conesa, et al., 2020). On the other hand, CDs can also be used as *secondary*

378 *antioxidants*, whereby an oxidising substrate is complexed by a CD, preventing
379 its oxidation (Crini et al., 2018; López-Nicolás et al., 2014), a quality widely used
380 to reduce the browning of some juices (Andreu-Sevilla et al., 2011).

381 Although only the three natural CDs α -, β - and γ -CD are authorized as food
382 additive, several derivatives such as hydroxypropyl- β -CD (HP β -CD) and methyl-
383 β -CD (M β -CD) are commonly used as excipients because of their lower toxicity
384 and wider applicability in drugs complexes (Jansook et al., 2018). A variety of
385 novel CD-based matrixes with BaCs have been synthesized, for example
386 polymeric CD based materials (Dhakar et al., 2019; Matencio, Dhakar, Bessone,
387 et al., 2020; Swaminathan et al., 2016; Yao et al., 2019), or nano-derivatives such
388 as amphiphilic CDs (Varan et al., 2017).

389 **5.4 Cyclodextrin effect on hydrocolloid properties.**

390 The use of CDs in different hydrocolloids such as carrageen or starch not
391 only improves the properties of the substance, but also changes the properties of
392 the hydrocolloid. Some interesting examples use approved food additives like
393 carrageen and, include the study of the influence of CDs on carrageen hydrogel
394 properties (Y. Wang et al., 2019; Yuan et al., 2016, 2018, 2019). It is highlighted
395 the case of κ -carrageenan/Konjac glucomannan compound gel, which hardness
396 was strengthened in the presence of 0.5 – 1.5 % (w/w) of CDs and then, gradually
397 decreased at higher CDs concentrations. In this study, Methyl- β CD (M β -CD) was
398 the most significant among all the selected CDs on the formation and
399 characteristics of κ -carrageenan/Konjac glucomannan compound gel (Yuan et
400 al., 2019). In another study, the influence of CDs on the rheological and structural
401 properties of κ -carrageenan gel was investigated. The gelling temperature (T_g)
402 and the plasticity were improved. The authors showed that the influence of CDs

403 on κ -carrageenan gelation was mainly through (i) the exclusion of CDs from κ -
404 CA in the sol state, (ii) the regular rearrangement of κ -carrageenan random coils
405 influenced by CDs in the sol state, (iii) the binding of CDs to the κ -carrageenan
406 surface by hydrogen bonds in the gel (Yuan et al., 2018).

407 The influence of β -CD on the retrogradation of rice starch was
408 evaluated. Retrogradation of normal rice starch was reduced more by β -CD than
409 by glycerol monostearate by the amylose- β -CD complex formation. β -CD
410 significantly lowered the crystallizing rate and increased the Avrami exponent of
411 amylose recrystallisation (Tian et al., 2009).

412 Finally, when CDs are used with guar or xanthan gum they are able to
413 enhance the viscosity of both hydrocolloids (Rao et al., 2014). In other study, a
414 novel self-assembling biopolymer was created between xanthan gum and β -CD,
415 this system exhibits superior mechanical and thermal stability, and also tolerance
416 to elevated brine salinity and hardness (Wei et al., 2015)

417 **5.5 Cyclodextrins for developing novel nanosensors.**

418 In the food industry it is important to know the quantity of a nutrients or
419 bioactive compounds in products, and, for this purpose novel measurement
420 methods with increased sensitivity have been developed. In this respect, the
421 many properties of CDs such as their capacity of chiral recognition of isomers or
422 to amplify their signals, means that they can be used as a platform for developing
423 novel nanosensors (**Fig. 4**).

424 For example, modified CDs have recently been used with grapheme
425 quantum dots to separate tyrosine enantiomers (Dong et al., 2017), after
426 obtaining β -CD quantum dot derivatives for inclusion in an electrode.

427 Not only hydrophobic molecules can form complexes with CDs, but ions
428 too, can be encapsulated, a property that can be used to create sensors. A recent
429 work analysed Zn^{2+} and CN^- in a solution using magnetic CDs (Q. Li et al., 2015).
430 A modified fluorescein was used to selectively bind the Zn^{2+} ion in preference to
431 others. The magnetic CD used showed high selectivity and an increased
432 fluorescent signal. Moreover, the CDs completely recovered their activity for at
433 least four more cycles. In this sense, a novel cholesterol nanosensor has been
434 studied (Y. Li et al., 2019), and natural CDs can be used to lay the foundations of
435 nanosensors to measure ellagic acid in a ternary complex with borax (Matencio,
436 Navarro-Orcajada, et al., 2018).

437 The use of CDs in water treatment involves using natural, chemically
438 modified or polymeric CDs to detect contaminants (Fourmentin et al., 2018a;
439 Salazar et al., 2018). The most common strategy is to create different
440 chemosensors focus on coloured or fluorescent CDs (modified with a
441 chromophore or fluorophore). CDs have demonstrated their capacity to recognize
442 and enantio-separate different water contaminants such as pesticides or 1-
443 adamantanol. Indeed, masks can be also modified with CDs to catch
444 contaminants (Alzate-Sánchez et al., 2016).

445 CDs can be used in chiral chromatography to separate isomers, a
446 technique widely used in the gas chromatography of essential oils (Fourmentin
447 et al., 2018a; Pragadheesh et al., 2015). CD columns are commercially available,
448 in which the degree of substitution or the type of CD contributes to the separation
449 of isomers.

450 **5.6 CDs in food packaging**

451 The food industry is constantly searching for ways to make food products
452 safer and more long-lasting, while maintaining quality and minimising any
453 environmental impact. One of the results in this respect is the development of
454 *active* and *smart* packaging, in which compounds are added to the packages in
455 order to extend the shelf life of the food or provide information on the state of the
456 food in real time. Applications of CDs in active packaging are abundant (**Fig. 5**),
457 although they are less used in smart packaging. Despite that, some promising
458 studies have been published, such as using p-methyl red in α -CD, which
459 produces a colour change in the event of leakage (Kuwabara, 2007). Additionally,
460 as CDs are starch-based oligosaccharides and therefore biodegradable
461 (Marshall & Miwa, 1981), if the material used is also easy-to-degrade (e.g.
462 chitosan, cellulose or polylactic acid) (Mohamed et al., 2020), the whole package
463 could be considered biodegradable.

464 CDs can be incorporated in food packages alone (the so-called “empty
465 cyclodextrins”) or complexed with a BaC acting as delivery systems
466 (Szente&Fenyvesi, 2018). Empty CDs in packaging can be used to encapsulate
467 hydrophobic compounds either from inside the package or from the outside. They
468 have been used to reduce the migration of plasticizers (Kwak et al., 2011) and
469 trichloroanisol (a contaminant from bottle corks (Angermaier, 2006)), to capture
470 undesirable volatile molecules, such as hexanal from fried peanuts (López-de-
471 Dicastillo et al., 2011) or sulphur off-flavours (Shin et al., 2018), and even to
472 reduce cholesterol in milk (López-de-Dicastillo et al., 2011). Meanwhile, inclusion
473 complexes in packaging can keep and controlled release a BaC in the inner
474 cavity, acting as preservatives and hence reduce the use of food additives, or in
475 the outer surface of the package to provide protection against environmental

476 factors. It has been described how an increase in humidity levels in the
477 headspace of packaged fresh-cut vegetables or fruits, which is associated with
478 spoilage, may trigger the release of BaC when using CDs as carriers (Ayala-
479 Zavala et al., 2008). In this sense, the quality and shelf life of fresh-cut pineapple
480 and papaya have been increased with the use of edible coatings that incorporate
481 *trans*-cinnamaldehyde in β -CD (Brasil et al., 2012; Mantilla et al., 2013), and the
482 same applies to fresh-cut onion using filter paper containing allyl isothiocyanate
483 in β -CD (Piercey et al., 2012).

484 Among such inclusion complexes used for food packaging, the most
485 studied are those that incorporate antimicrobial BaCs to increase the useful life
486 of the packaged foods by decreasing the growth of bacteria and/or fungi. Most of
487 the reviewed research on this matter involves the encapsulation of essential oils
488 or their major bioactive ingredients in HP β -CD or natural CDs, with β -CD being
489 the most predominant. Such complexes have demonstrated their ability to extend
490 shelf life and maintain the quality of packaged meat (Aytac, Ipek, et al., 2017;
491 Chen et al., 2019; Higuera et al., 2014; Lin et al., 2017, 2018; XiaoYun et al.,
492 2018), mushrooms (Cheng et al., 2019; Pan et al., 2019), and fruits
493 (Buendía–Moreno et al., 2020; da Rocha Neto et al., 2019; Wen et al., 2016). In
494 packaged fruits, some authors associated the improvement of shelf life with the
495 dual effect of BaC release and ethylene adsorption (Buendía–Moreno et al.,
496 2020).

497 The ripening of packaged fresh fruits and vegetables can be controlled by
498 encapsulating ripening agents or inhibitors. For instance, mango ripening was
499 modulated by packaging them with ethylene and α -CD complexes (Capozzi et

500 al., 2018), while, similar effects were observed using 1-methylcyclopropene in α -
501 CD (Wood et al., 2017) and hexanal in γ -CD (Lang, 2019).

502 Antioxidant compounds are also commonly used for active packaging with
503 CDs: for example, HP β -CD complexes with curcumin(Aytac&Uyar, 2017) or
504 ferulic acid (Sharif et al., 2018), β -CD with gallic acid (Rezaee et al., 2018) or
505 retinyl acetate (Lemma et al., 2015), and γ -CD with quercetin(Aytac et al., 2018)
506 or α -tocopherol (Aytac, Keskin, et al., 2017). Only the last mentioned authors
507 tested the package with a real food (meat), while Rezaee et al. (2018) and Sharif
508 et al. (2018) checked the release of the BaCin acid, alcoholic and fatty food
509 simulants, which consisted of 3 % acetic acid, 10 % ethanol or 95 - 96 % ethanol
510 in water, respectively.

511 Volatile fragrances such as vanillin (Kayaci&Uyar, 2012), geraniol(Kayaci
512 et al., 2014), D-limonene (Mallardo et al., 2016) and 2-phenyl ethanol (Zarandona
513 et al., 2020) have been successfully integrated into polymers with natural CDs.

514 Moreover, CDs in packaging can also provide thermal protection to volatile
515 or thermo-sensitive molecules during polymer processing (Fenyvesi et al., 2016b;
516 Sharif et al., 2018), increasing the amount of added BaC (Poverenov et al., 2013)
517 and, enhance the UV-light barrier property of packages (Cheng et al., 2019; Ye
518 et al., 2017). In some cases, there is no other way that can be used to retain the
519 BaC in the matrix (Chen et al., 2019; Zarandona et al., 2020).

520 Empty CDs or inclusion complexes can be embedded homogeneously in
521 the packaging material or be applied in a separate coating to limit their effect to
522 one direction through the package (inwards or outwards) (Szente&Fenyvesi,
523 2018). A wide variety of carrier materials (natural or synthetic) and techniques
524 can be applied to create the packaging. For example, the use of electrospun

525 nanofibres is increasing because of their appealing characteristics, such as high
526 surface-to-volume ratio, nanoporosity, and safety (Noruzi, 2016): Mixed with
527 inclusion complexes they can improve the thermal stability and shelf life of foods,
528 and ensure the slow release of some BaCs, such as eugenol (Kayaci et al., 2013).

529 **5.7 Market products with CDs.**

530 Although most applications of CDs are connected with the pharmaceutical
531 industry, where they are used as drug excipients (Jansook et al., 2018), some
532 food products also contain them: for example, α -CD is used for direct weight
533 control purposes and some products such as Calorease™, a natural supplement
534 that has been clinically researched to help control cholesterol and triglyceride
535 levels and maintain a healthy weight through diet and exercise (Comerford et al.,
536 2011; Fenyvesi et al., 2016a; Jarosz et al., 2013). In New Zealand Cyclopower™,
537 which is based on encapsulating the bioactive compounds found in manuka
538 honey and in propolis inside cyclodextrins, allow the controlled release of the
539 bioactive compounds where they are needed most in the body. Although the
540 current cost of CDs prevents the use of all the potential applications mentioned
541 in the literature, production costs are continuously falling, and they will surely be
542 more widely used in the near future.

543 **6. CONCLUSIONS**

544 Although CDs have been used for several years, the way in which they are
545 currently used in the food industry is changing: the classical (although equally
546 important) role as of a “carrier” to stabilize bioactive compounds is giving way to
547 novel applications: i) CDs as nutritional supplement in the form of prebiotic, and
548 for weight and lipid control, ii) in active and smart food packaging, enabling the
549 controlled release of antimicrobials, antioxidants, etc., iii) as nanosensor to

550 measure bioactive compounds and, iv) forming novel nanoparticles for functional
551 food uses, v) its use with hydrocolloids and vi) as enhancers of the extraction or
552 production enhancers of bioactive compounds.

553 As an evident prospect from this review, the versatility of cyclodextrins is
554 wide and promising. Traditional and chemically derived CDs find novel
555 applications and uses every day. Several research groups are increasing the
556 number of CDs and CD-based materials available for use in fresh applications.
557 As the costs of this technology are decreasing (CD production, protocols...),and
558 novel applications are constantly being found, their application in the food,
559 pharmaceutical and other industries (not only in scientific research) will
560 undoubtedly increase in forthcoming years.

561

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576 **BIBLIOGRAPHY**

577

- 578 Abril-Sánchez, C., Matencio, A., Navarro-Orcajada, S., García-Carmona, F., &
579 López-Nicolás, J. M. (2019). Evaluation of the properties of the essential
580 oil citronellal nanoencapsulated by cyclodextrins. *Chemistry and Physics
581 of Lipids*, 219, 72–78. <https://doi.org/10.1016/j.chemphyslip.2019.02.001>
- 582 Ahn, J., & Kwak, H. S. (1999). Optimizing Cholesterol Removal in Cream Using
583 β -Cyclodextrin and Response Surface Methodology. *Journal of Food
584 Science*, 64(4), 629–632. [https://doi.org/10.1111/j.1365-
585 2621.1999.tb15098.x](https://doi.org/10.1111/j.1365-2621.1999.tb15098.x)
- 586 Alonso, L., Calvo, M. V., & Fontecha, J. (2019a). A scale-up process for the
587 manufacture of reduced-cholesterol butter using beta-cyclodextrin.
588 *Journal of Food Process Engineering*, 42(3), e13009.
589 <https://doi.org/10.1111/jfpe.13009>
- 590 Alonso, L., Calvo, M. V., & Fontecha, J. (2019b). The Influence of β -
591 Cyclodextrin on the Reduction of Cholesterol Content in Egg and Duck
592 Liver Pâté. *Foods*, 8(7), 241. <https://doi.org/10.3390/foods8070241>
- 593 Alzate-Sánchez, D. M., Smith, B. J., Alsaiee, A., Hinestroza, J. P., & Dichtel,
594 W. R. (2016). Cotton Fabric Functionalized with a β -Cyclodextrin
595 Polymer Captures Organic Pollutants from Contaminated Air and Water.
596 *Chemistry of Materials*, 28(22), 8340–8346.
597 <https://doi.org/10.1021/acs.chemmater.6b03624>
- 598 Andreu-Sevilla, A. J., López-Nicolás, J. M., Carbonell-Barrachina, Á. A., &
599 García-Carmona, F. (2011). Comparative Effect of the Addition of α -, β -,
600 or γ -Cyclodextrin on Main Sensory and Physico–Chemical Parameters.

601 *Journal of Food Science*, 76(5), S347–S353.
602 <https://doi.org/10.1111/j.1750-3841.2011.02190.x>

603 Angermaier, K. (2006). *Bottle cork with reduced trichloroanisole release and*
604 *method for its production* (United States Patent No. US20060073295A1).
605 <https://patents.google.com/patent/US20060073295A1/en>

606 Antoine Villiers. (1891). Sur la fermentation de la fécule par l'action du ferment
607 butyrique. *Comptes Rendus de l'Académie Des Sciences*, 112, 536–538.

608 Arima, H., Motoyama, K., & Irie, T. (2011). Recent Findings on Safety Profiles of
609 Cyclodextrins, Cyclodextrin Conjugates, and Polypseudorotaxanes. In
610 *Cyclodextrins in Pharmaceuticals, Cosmetics, and Biomedicine* (pp. 91–
611 122). John Wiley & Sons, Ltd.
612 <https://doi.org/10.1002/9780470926819.ch5>

613 Astray, G., Gonzalez-Barreiro, C., Mejuto, J. C., Rial-Otero, R., & Simal-
614 Gándara, J. (2009). A review on the use of cyclodextrins in foods. *Food*
615 *Hydrocolloids*, 23(7), 1631–1640.
616 <https://doi.org/10.1016/j.foodhyd.2009.01.001>

617 Astray, G., Mejuto, J. C., & Simal-Gandara, J. (2020). Latest developments in
618 the application of cyclodextrin host-guest complexes in beverage
619 technology processes. *Food Hydrocolloids*, 106, 105882.
620 <https://doi.org/10.1016/j.foodhyd.2020.105882>

621 Athanasiadis, V., Grigorakis, S., Lalas, S., & Makris, D. P. (2018). Methyl β -
622 cyclodextrin as a booster for the extraction for *Olea europaea* leaf
623 polyphenols with a bio-based deep eutectic solvent. *Biomass Conversion*
624 *and Biorefinery*, 8(2), 345–355. [https://doi.org/10.1007/s13399-017-](https://doi.org/10.1007/s13399-017-0283-5)
625 0283-5

626 Ayala-Zavala, J. F., Del-Toro-Sánchez, L., Alvarez-Parrilla, E., & González-
627 Aguilar, G. A. (2008). High relative humidity in-package of fresh-cut fruits
628 and vegetables: advantage or disadvantage considering microbiological
629 problems and antimicrobial delivering systems? *Journal of Food Science*,
630 73(4), R41-47. <https://doi.org/10.1111/j.1750-3841.2008.00705.x>

631 Ayesh, A. M., & Essa, H. A. (2002). mycotoxins reduction and inhibition of
632 enzymatic browning during apple juice processing. *Egyptian Journal of*
633 *Food Science*, 30(1).
634 [https://www.kau.edu.sa/Show_Res.aspx?Site_ID=857&LNG=EN&RN=5](https://www.kau.edu.sa/Show_Res.aspx?Site_ID=857&LNG=EN&RN=58183)
635 8183

636 Aytac, Z., Ipek, S., Durgun, E., Tekinay, T., & Uyar, T. (2017). Antibacterial
637 electrospun zein nanofibrous web encapsulating thymol/cyclodextrin-
638 inclusion complex for food packaging. *Food Chemistry*, 233, 117–124.
639 <https://doi.org/10.1016/j.foodchem.2017.04.095>

640 Aytac, Z., Ipek, S., Durgun, E., & Uyar, T. (2018). Antioxidant electrospun zein
641 nanofibrous web encapsulating quercetin/cyclodextrin inclusion complex.
642 *Journal of Materials Science*, 53(2), 1527–1539.
643 <https://doi.org/10.1007/s10853-017-1580-x>

644 Aytac, Z., Keskin, N. O. S., Tekinay, T., & Uyar, T. (2017). Antioxidant α -
645 tocopherol/ γ -cyclodextrin–inclusion complex encapsulated poly(lactic
646 acid) electrospun nanofibrous web for food packaging. *Journal of Applied*
647 *Polymer Science*, 134(21). <https://doi.org/10.1002/app.44858>

648 Aytac, Z., & Uyar, T. (2017). Core-shell nanofibers of curcumin/cyclodextrin
649 inclusion complex and polylactic acid: Enhanced water solubility and

650 slow release of curcumin. *International Journal of Pharmaceutics*, 518(1),
651 177–184. <https://doi.org/10.1016/j.ijpharm.2016.12.061>

652 Benhenia, K., Lamara, A., Fatmi, S., & Iguer-Ouada, M. (2016). Effect of
653 cyclodextrins, cholesterol and vitamin E and their complexation on
654 cryopreserved epididymal ram semen. *Small Ruminant Research*, 141,
655 29–35. <https://doi.org/10.1016/j.smallrumres.2016.06.009>

656 Benhenia, K., Rahab, H., Smadi, M.-A., Benmakhlouf, H., Lamara, A., Idres, T.,
657 & Iguer-Ouada, M. (2018). Beneficial and harmful effects of cyclodextrin-
658 vitamin E complex on cryopreserved ram sperm. *Animal Reproduction
659 Science*, 195, 266–273. <https://doi.org/10.1016/j.anireprosci.2018.06.004>

660 Biber, A., Antranikian, G., & Heinzle, E. (2002). Enzymatic production of
661 cyclodextrins. *Applied Microbiology and Biotechnology*, 59(6), 609–617.
662 <https://doi.org/10.1007/s00253-002-1057-x>

663 Bonnet, V., Gervaise, C., Favrelle, A., Sarazin, C., & Djedaini-Pilard, F. (2010).
664 *Enzymatic Catalysis in Presence of Cyclodextrins* [Text].
665 <https://doi.org/info:doi/10.2174/138527210791616849>

666 Brasil, I. M., Gomes, C., Puerta-Gomez, A., Castell-Perez, M. E., & Moreira, R.
667 G. (2012). Polysaccharide-based multilayered antimicrobial edible
668 coating enhances quality of fresh-cut papaya. *LWT - Food Science and
669 Technology*, 47(1), 39–45. <https://doi.org/10.1016/j.lwt.2012.01.005>

670 Brocos, P., Díaz-Vergara, N., Banquy, X., Pérez-Casas, S., Costas, M., &
671 Piñeiro, Á. (2010). Similarities and Differences Between
672 Cyclodextrin–Sodium Dodecyl Sulfate Host–Guest Complexes of
673 Different Stoichiometries: Molecular Dynamics Simulations at Several

674 Temperatures. *The Journal of Physical Chemistry B*, 114(39), 12455–
675 12467. <https://doi.org/10.1021/jp103223u>

676 Buendía–Moreno, L., Sánchez–Martínez, M. J., Antolinos, V., Ros–Chumillas,
677 M., Navarro–Segura, L., Soto–Jover, S., Martínez–Hernández, G. B., &
678 López–Gómez, A. (2020). Active cardboard box with a coating including
679 essential oils entrapped within cyclodextrins and/or halloysite nanotubes.
680 A case study for fresh tomato storage. *Food Control*, 107, 106763.
681 <https://doi.org/10.1016/j.foodcont.2019.106763>

682 Capozzi, L. C., Bazzano, M., Sangermano, M., & Pisano, R. (2018). Inclusion
683 complexes dispersed in polystyrene-based labels for fruit ripening on
684 demand. *International Journal of Food Science & Technology*, 53(2),
685 389–394. <https://doi.org/10.1111/ijfs.13596>

686 Celebioglu, A., Yildiz, Z. I., & Uyar, T. (2018). Thymol/cyclodextrin inclusion
687 complex nanofibrous webs: Enhanced water solubility, high thermal
688 stability and antioxidant property of thymol. *Food Research International*,
689 106, 280–290. <https://doi.org/10.1016/j.foodres.2017.12.062>

690 Charumanee, S., Okonogi, S., Sirithunyalug, J., Wolschann, P., & Viernstein, H.
691 (2016). Effect of Cyclodextrin Types and Co-Solvent on Solubility of a
692 Poorly Water Soluble Drug. *Scientia Pharmaceutica*, 84(4), 694–705.
693 <https://doi.org/10.3390/scipharm84040694>

694 Chen, H., Li, L., Ma, Y., Mcdonald, T. P., & Wang, Y. (2019). Development of
695 active packaging film containing bioactive components encapsulated in
696 β-cyclodextrin and its application. *Food Hydrocolloids*, 90, 360–366.
697 <https://doi.org/10.1016/j.foodhyd.2018.12.043>

698 Cheng, M., Wang, J., Zhang, R., Kong, R., Lu, W., & Wang, X. (2019).
699 Characterization and application of the microencapsulated
700 carvacrol/sodium alginate films as food packaging materials.
701 *International Journal of Biological Macromolecules*, 141, 259–267.
702 <https://doi.org/10.1016/j.ijbiomac.2019.08.215>

703 Cheong, A. M., & Nyam, K. L. (2016). Improvement of physical stability of kenaf
704 seed oil-in-water nanoemulsions by addition of β -cyclodextrin to primary
705 emulsion containing sodium caseinate and Tween 20. *Journal of Food*
706 *Engineering*, 183, 24–31. <https://doi.org/10.1016/j.jfoodeng.2016.03.012>

707 Ciobanu, A., Landy, D., & Fourmentin, S. (2013). Complexation efficiency of
708 cyclodextrins for volatile flavor compounds. *Food Research International*,
709 53(1), 110–114. <https://doi.org/10.1016/j.foodres.2013.03.048>

710 Comerford, K. B., Artiss, J. D., Jen, K. L. C., & Karakas, S. E. (2011). The
711 beneficial effects of α -cyclodextrin on blood lipids and weight loss in
712 healthy humans. *Obesity (Silver Spring, Md.)*, 19(6), 1200–1204.
713 <https://doi.org/10.1038/oby.2010.280>

714 Crini, G., Fourmentin, S., Fenyvesi, É., Torri, G., Fourmentin, M., & Morin-Crini,
715 N. (2018). Cyclodextrins, from molecules to applications. *Environmental*
716 *Chemistry Letters*, 16(4), 1361–1375. [https://doi.org/10.1007/s10311-](https://doi.org/10.1007/s10311-018-0763-2)
717 [018-0763-2](https://doi.org/10.1007/s10311-018-0763-2)

718 da Rocha Neto, A. C., Beaudry, R., Maraschin, M., Di Piero, R. M., & Almenar,
719 E. (2019). Double-bottom antimicrobial packaging for apple shelf-life
720 extension. *Food Chemistry*, 279, 379–388.
721 <https://doi.org/10.1016/j.foodchem.2018.12.021>

722 Das, S., Gazdag, Z., Szente, L., Meggyes, M., Horváth, G., Lemli, B., Kunsági-
723 Máté, S., Kuzma, M., & Kőszegi, T. (2019). Antioxidant and antimicrobial
724 properties of randomly methylated β cyclodextrin – captured essential
725 oils. *Food Chemistry*, 278, 305–313.
726 <https://doi.org/10.1016/j.foodchem.2018.11.047>

727 Dhakar, N. K., Matencio, A., Caldera, F., Argenziano, M., Cavalli, R., Dianzani,
728 C., Zanetti, M., López-Nicolás, J. M., & Trotta, F. (2019). Comparative
729 Evaluation of Solubility, Cytotoxicity and Photostability Studies of
730 Resveratrol and Oxyresveratrol Loaded Nanosponges. *Pharmaceutics*,
731 11(10), 545. <https://doi.org/10.3390/pharmaceutics11100545>

732 Diamanti, A. C., Igoumenidis, P. E., Mourtzinou, I., Yannakopoulou, K., &
733 Karathanos, V. T. (2017). Green extraction of polyphenols from whole
734 pomegranate fruit using cyclodextrins. *Food Chemistry*, 214, 61–66.
735 <https://doi.org/10.1016/j.foodchem.2016.07.072>

736 Dong, S., Bi, Q., Qiao, C., Sun, Y., Zhang, X., Lu, X., & Zhao, L. (2017).
737 Electrochemical sensor for discrimination tyrosine enantiomers using
738 graphene quantum dots and β -cyclodextrins composites. *Talanta*, 173,
739 94–100. <https://doi.org/10.1016/j.talanta.2017.05.045>

740 dos Santos, C., Buera, P., & Mazzobre, F. (2017). Novel trends in cyclodextrins
741 encapsulation. Applications in food science. *Current Opinion in Food*
742 *Science*, 16, 106–113. <https://doi.org/10.1016/j.cofs.2017.09.002>

743 dos Santos Lima, B., Shanmugam, S., de Souza Siqueira Quintans, J.,
744 Quintans-Júnior, L. J., & de Souza Araújo, A. A. (2019). Inclusion
745 complex with cyclodextrins enhances the bioavailability of flavonoid

746 compounds: a systematic review. *Phytochemistry Reviews*, 18(5), 1337–
747 1359. <https://doi.org/10.1007/s11101-019-09650-y>

748 Durrant, J. D., & McCammon, J. A. (2011). Molecular dynamics simulations and
749 drug discovery. *BMC Biology*, 9(1), 71. [https://doi.org/10.1186/1741-
750 7007-9-71](https://doi.org/10.1186/1741-7007-9-71)

751 EFSA. (2012). Scientific Opinion on the substantiation of health claims related
752 to alpha cyclodextrin and reduction of post prandial glycaemic responses
753 (ID 2926, further assessment) pursuant to Article 13(1) of Regulation
754 (EC) No 1924/2006. *EFSA Journal*, 10(6), 2713.
755 <https://doi.org/10.2903/j.efsa.2012.2713>

756 EMA. (2017). Questions and answers on cyclodextrins used as excipients in
757 medicinal products for human use. *EMA*, 9.
758 *European Pharmacopoeia (Ph. Eur.) 9th Edition | EDQM*. (n.d.). Retrieved April
759 3, 2019, from [https://www.edqm.eu/en/european-pharmacopoeia-ph-eur-
760 9th-edition](https://www.edqm.eu/en/european-pharmacopoeia-ph-eur-9th-edition)

761 F. Garrido, P., Calvelo, M., Garcia-Fandiño, R., & Piñeiro, Á. (2020). Rings,
762 Hexagons, Petals, and Dipolar Moment Sink-Sources: The Fanciful
763 Behavior of Water around Cyclodextrin Complexes. *Biomolecules*, 10(3),
764 431. <https://doi.org/10.3390/biom10030431>

765 Favre, L. C., Rolandelli, G., Mshicileli, N., Vhangani, L. N., dos Santos Ferreira,
766 C., van Wyk, J., & Buera, M. del P. (2020). Antioxidant and anti-glycation
767 potential of green pepper (*Piper nigrum*): Optimization of β -cyclodextrin-
768 based extraction by response surface methodology. *Food Chemistry*,
769 316, 126280. <https://doi.org/10.1016/j.foodchem.2020.126280>

770 Fenyvesi, É., Vikmon, M., & Szente, L. (2016a). Cyclodextrins in Food
771 Technology and Human Nutrition: Benefits and Limitations. *Critical*
772 *Reviews in Food Science and Nutrition*, 56(12), 1981–2004.
773 <https://doi.org/10.1080/10408398.2013.809513>

774 Fenyvesi, É., Vikmon, M., & Szente, L. (2016b). Cyclodextrins in Food
775 Technology and Human Nutrition: Benefits and Limitations. *Critical*
776 *Reviews in Food Science and Nutrition*, 56(12), 1981–2004.
777 <https://doi.org/10.1080/10408398.2013.809513>

778 Fourmentin, S., Crini, G., & Lichtfouse, E. (Eds.). (2018a). *Cyclodextrin*
779 *Applications in Medicine, Food, Environment and Liquid Crystals*.
780 Springer International Publishing.
781 <https://www.springer.com/la/book/9783319761619>

782 Fourmentin, S., Crini, G., & Lichtfouse, E. (Eds.). (2018b). *Cyclodextrin*
783 *Fundamentals, Reactivity and Analysis*. Springer International
784 Publishing. <https://www.springer.com/la/book/9783319761589>

785 French, D., Pulley, A. O., Effenberger, J. A., Rougvie, M. A., & Abdullah, M.
786 (1965). Studies on the Schardinger dextrans: XII. The molecular size and
787 structure of the δ -, ϵ -, ζ -, and η -dextrans. *Archives of Biochemistry and*
788 *Biophysics*, 111(1), 153–160. [https://doi.org/10.1016/0003-](https://doi.org/10.1016/0003-9861(65)90334-6)
789 [9861\(65\)90334-6](https://doi.org/10.1016/0003-9861(65)90334-6)

790 Frijlink, H. W., Visser, J., Hefting, N. R., Oosting, R., Meijer, D. K. F., & Lerk, C.
791 F. (1990). The Pharmacokinetics of β -Cyclodextrin and Hydroxypropyl- β -
792 cyclodextrin in the Rat. *Pharmaceutical Research*, 7(12), 1248–1252.
793 <https://doi.org/10.1023/A:1015929720063>

- 794 Gao, F., Zhou, T., Hu, Y., Lan, L., Heyden, Y. V., Crommen, J., Lu, G., & Fan,
795 G. (2016). Cyclodextrin-based ultrasonic-assisted microwave extraction
796 and HPLC-PDA-ESI-ITMSn separation and identification of hydrophilic
797 and hydrophobic components of *Polygonum cuspidatum*: A green, rapid
798 and effective process. *Industrial Crops and Products*, 80, 59–69.
799 <https://doi.org/10.1016/j.indcrop.2015.10.039>
- 800 García-Pérez, P., Losada-Barreiro, S., Gallego, P. P., & Bravo-Díaz, C. (2019).
801 Cyclodextrin-Elicited Bryophyllum Suspension Cultured Cells:
802 Enhancement of the Production of Bioactive Compounds. *International*
803 *Journal of Molecular Sciences*, 20(20), 5180.
804 <https://doi.org/10.3390/ijms20205180>
- 805 Gould, S., & Scott, R. C. (2005). 2-Hydroxypropyl-beta-cyclodextrin (HP-beta-
806 CD): a toxicology review. *Food and Chemical Toxicology: An*
807 *International Journal Published for the British Industrial Biological*
808 *Research Association*, 43(10), 1451–1459.
809 <https://doi.org/10.1016/j.fct.2005.03.007>
- 810 Hadi, B. J., Sanagi, M. M., Aboul-Enein, H. Y., Ibrahim, W. A. W., Jamil, S., &
811 Mu'azu, M. A. (2015). Microwave-Assisted Extraction of Methyl β -
812 Cyclodextrin-Complexed Curcumin from Turmeric Rhizome Oleoresin.
813 *Food Analytical Methods*, 8(10), 2447–2456.
814 <https://doi.org/10.1007/s12161-015-0137-3>
- 815 Hamilton, D. A., Ernst, C. C., Kramer, W. G., Madden, D., Lang, E., Liao, E.,
816 Lacouture, P. G., Ramaiya, A., & Carr, D. B. (2018). Pharmacokinetics of
817 Diclofenac and Hydroxypropyl- β -Cyclodextrin (HP β CD) Following
818 Administration of Injectable HP β CD-Diclofenac in Subjects With Mild to

819 Moderate Renal Insufficiency or Mild Hepatic Impairment. *Clinical*
820 *Pharmacology in Drug Development*, 7(2), 110–122.
821 <https://doi.org/10.1002/cpdd.417>

822 Higuera, L., López-Carballo, G., Hernández-Muñoz, P., Catalá, R., & Gavara,
823 R. (2014). Antimicrobial packaging of chicken fillets based on the release
824 of carvacrol from chitosan/cyclodextrin films. *International Journal of*
825 *Food Microbiology*, 188, 53–59.
826 <https://doi.org/10.1016/j.ijfoodmicro.2014.07.018>

827 Ho, S., Thoo, Y. Y., Young, D. J., & Siow, L. F. (2017). Cyclodextrin
828 encapsulated catechin: Effect of pH, relative humidity and various food
829 models on antioxidant stability. *LWT - Food Science and Technology*, 85,
830 232–239. <https://doi.org/10.1016/j.lwt.2017.07.028>

831 Hou, X., Wang, L., Tang, X., Xiong, C., Guo, Y., & Liu, X. (2015). Application of
832 a β -cyclodextrin/graphene oxide-modified fiber for solid-phase
833 microextraction of six fragrance allergens in personal products. *Analyst*,
834 140(19), 6727–6735. <https://doi.org/10.1039/C5AN01030F>

835 Ikuta, D., Hirata, Y., Wakamori, S., Shimada, H., Tomabechi, Y., Kawasaki, Y.,
836 Ikeuchi, K., Hagimori, T., Matsumoto, S., & Yamada, H. (2019).
837 Conformationally supplemented glucose monomers enable synthesis of the
838 smallest cyclodextrins. *Science*, 364(6441), 674–677.
839 <https://doi.org/10.1126/science.aaw3053>

840 Inoue, M., Hashizaki, K., Taguchi, H., & Saito, Y. (2010). Emulsifying Ability of
841 β -Cyclodextrins for Common Oils. *Journal of Dispersion Science and*
842 *Technology*, 31(12), 1648–1651.
843 <https://doi.org/10.1080/01932690903297058>

- 844 Jansook, P., Ogawa, N., & Loftsson, T. (2018). Cyclodextrins: structure,
845 physicochemical properties and pharmaceutical applications.
846 *International Journal of Pharmaceutics*, 535(1), 272–284.
847 <https://doi.org/10.1016/j.ijpharm.2017.11.018>
- 848 Jarosz, P. A., Fletcher, E., Elserafy, E., Artiss, J. D., & Jen, K.-L. C. (2013). The
849 effect of α -cyclodextrin on postprandial lipid and glycemic responses to a
850 fat-containing meal. *Metabolism: Clinical and Experimental*, 62(10),
851 1443–1447. <https://doi.org/10.1016/j.metabol.2013.05.015>
- 852 John Marshall, J., & Miwa, I. (1981). Kinetic difference between hydrolyses of γ -
853 cyclodextrin by human salivary and pancreatic α -amylases. *Biochimica et*
854 *Biophysica Acta (BBA) - Enzymology*, 661(1), 142–147.
855 [https://doi.org/10.1016/0005-2744\(81\)90093-0](https://doi.org/10.1016/0005-2744(81)90093-0)
- 856 Kamihira, M., Asai, T., Yamagata, Y., Taniguchi, M., & Kobayashi, T. (1990).
857 Formation of inclusion complexes between cyclodextrins and aromatic
858 compounds under pressurized carbon dioxide. *Journal of Fermentation*
859 *and Bioengineering*, 69(6), 350–353. [https://doi.org/10.1016/0922-](https://doi.org/10.1016/0922-338X(90)90242-O)
860 [338X\(90\)90242-O](https://doi.org/10.1016/0922-338X(90)90242-O)
- 861 Kayaci, F., Ertas, Y., & Uyar, T. (2013). Enhanced Thermal Stability of Eugenol
862 by Cyclodextrin Inclusion Complex Encapsulated in Electrospun
863 Polymeric Nanofibers. *Journal of Agricultural and Food Chemistry*,
864 61(34), 8156–8165. <https://doi.org/10.1021/jf402923c>
- 865 Kayaci, F., Sen, H. S., Durgun, E., & Uyar, T. (2014). Functional electrospun
866 polymeric nanofibers incorporating geraniol–cyclodextrin inclusion
867 complexes: High thermal stability and enhanced durability of geraniol.

868 *Food Research International*, 62, 424–431.
869 <https://doi.org/10.1016/j.foodres.2014.03.033>

870 Kayaci, F., & Uyar, T. (2012). Encapsulation of vanillin/cyclodextrin inclusion
871 complex in electrospun polyvinyl alcohol (PVA) nanowebs: Prolonged
872 shelf-life and high temperature stability of vanillin. *Food Chemistry*,
873 133(3), 641–649. <https://doi.org/10.1016/j.foodchem.2012.01.040>

874 Khuntawee, W., Wolschann, P., Rungrotmongkol, T., Wong-ekkabut, J., &
875 Hannongbua, S. (2015). Molecular Dynamics Simulations of the
876 Interaction of Beta Cyclodextrin with a Lipid Bilayer. *Journal of Chemical*
877 *Information and Modeling*, 55(9), 1894–1902.
878 <https://doi.org/10.1021/acs.jcim.5b00152>

879 Kokkiralala, S., Sabbavarapu, N. M., & Yadavalli, V. D. N. (2011). β -Cyclodextrin
880 mediated synthesis of 1,8-dioxooctahydroxanthenes in water. *European*
881 *Journal of Chemistry*, 2(2), 272–275.
882 <https://doi.org/10.5155/eurjchem.2.2.272-275.359>

883 Komaikul, J., Kitisripanya, T., Likhitwitayawuid, K., Sritularak, B., Tanaka, H., &
884 Putalun, W. (2019). Improvement of stilbenoid production by 2-
885 hydroxypropyl- β -cyclodextrin in white mulberry (*Morus alba* L.) callus
886 cultures. *Natural Product Research*, 33(19), 2762–2769.
887 <https://doi.org/10.1080/14786419.2018.1499643>

888 Kurkov, S. V., & Loftsson, T. (2013). Cyclodextrins. *International Journal of*
889 *Pharmaceutics*, 453(1), 167–180.
890 <https://doi.org/10.1016/j.ijpharm.2012.06.055>

891 Kuwabara, T. (2007). *Modified Cyclodextrin Film Or Fiber, And Method For*
892 *Producing The Same* (United States Patent No. US20070212400A1).
893 <https://patents.google.com/patent/US20070212400A1/en>

894 Kwak, S.-Y., Jung, S.-J., & Chung, J.-W. (2011). *Poly(vinyl chloride) product*
895 *containing cyclodextrin derivatives with suppression of the migration of*
896 *plasticizer and manufacturing method thereof* (United States Patent No.
897 US8008375B2). <https://patents.google.com/patent/US8008375B2/en>

898 Lang, T. (2019). Investigation of the Encapsulation Efficiency of Hexanal in γ -
899 Cyclodextrin Metal Organic Frameworks. *Materials Engineering*.
900 <https://digitalcommons.calpoly.edu/matesp/201>

901 Lee, D. K., Ahn, J., & Kwak, H. S. (1999). Cholesterol Removal from
902 Homogenized Milk with β -Cyclodextrin. *Journal of Dairy Science*, 82(11),
903 2327–2330. [https://doi.org/10.3168/jds.S0022-0302\(99\)75481-0](https://doi.org/10.3168/jds.S0022-0302(99)75481-0)

904 Lemma, S. M., Scampicchio, M., Mahon, P. J., Sbarski, I., Wang, J., &
905 Kingshott, P. (2015). Controlled Release of Retinyl Acetate from β -
906 Cyclodextrin Functionalized Poly(vinyl alcohol) Electrospun Nanofibers.
907 *Journal of Agricultural and Food Chemistry*, 63(13), 3481–3488.
908 <https://doi.org/10.1021/acs.jafc.5b00103>

909 Li, P., Song, J., Ni, X., Guo, Q., Wen, H., Zhou, Q., Shen, Y., Huang, Y., Qiu,
910 P., Lin, S., & Hu, H. (2016). Comparison in toxicity and solubilizing
911 capacity of hydroxypropyl- β -cyclodextrin with different degree of
912 substitution. *International Journal of Pharmaceutics*, 513(1), 347–356.
913 <https://doi.org/10.1016/j.ijpharm.2016.09.036>

914 Li, Q., Zhang, Y., Jin, Y., Yang, Q., Du, J., & Li, Y. (2015). Fluorescent magnetic
915 nanosensors for Zn²⁺ and CN⁻ in aqueous solution prepared from

916 adamantane-modified fluorescein and β -cyclodextrin-modified
917 Fe₃O₄@SiO₂ via host–guest interactions. *RSC Advances*, 5(84),
918 68815–68821. <https://doi.org/10.1039/C5RA12258A>

919 Li, Y., Cai, J., Liu, F., Yang, H., Lin, Y., Li, S., Huang, X., & Lin, L. (2019).
920 Construction of a turn off-on fluorescent nanosensor for cholesterol
921 based on fluorescence resonance energy transfer and competitive host-
922 guest recognition. *Talanta*, 201, 82–89.
923 <https://doi.org/10.1016/j.talanta.2019.03.110>

924 Lin, L., Dai, Y., & Cui, H. (2017). Antibacterial poly(ethylene oxide) electrospun
925 nanofibers containing cinnamon essential oil/beta-cyclodextrin
926 proteoliposomes. *Carbohydrate Polymers*, 178, 131–140.
927 <https://doi.org/10.1016/j.carbpol.2017.09.043>

928 Lin, L., Zhu, Y., & Cui, H. (2018). Electrospun thyme essential oil/gelatin
929 nanofibers for active packaging against *Campylobacter jejuni* in chicken.
930 *LWT*, 97, 711–718. <https://doi.org/10.1016/j.lwt.2018.08.015>

931 López-de-Dicastillo, C., Catalá, R., Gavara, R., & Hernández-Muñoz, P. (2011).
932 Food applications of active packaging EVOH films containing
933 cyclodextrins for the preferential scavenging of undesirable compounds.
934 *Journal of Food Engineering*, 104(3), 380–386.
935 <https://doi.org/10.1016/j.jfoodeng.2010.12.033>

936 López-Miranda, S., Serrano-Martínez, A., Hernández-Sánchez, P., Guardiola,
937 L., Pérez-Sánchez, H., Fortea, I., Gabaldón, J. A., & Núñez-Delicado, E.
938 (2016). Use of cyclodextrins to recover catechin and epicatechin from red
939 grape pomace. *Food Chemistry*, 203, 379–385.
940 <https://doi.org/10.1016/j.foodchem.2016.02.100>

941 López-Nicolás, J. M., Escorial Camps, M., Pérez-Sánchez, H., & García-
942 Carmona, F. (2013). Physicochemical and Thermodynamic
943 Characterization of the Encapsulation of Methyl Jasmonate by Natural
944 and Modified Cyclodextrins Using Reversed-Phase High-Pressure Liquid
945 Chromatography. *Journal of Agricultural and Food Chemistry*, 61(47),
946 11347–11354. <https://doi.org/10.1021/jf402920p>

947 López-Nicolás, J. M., Rodríguez-Bonilla, P., & García-Carmona, F. (2014).
948 Cyclodextrins and Antioxidants. *Critical Reviews in Food Science and*
949 *Nutrition*, 54(2), 251–276. <https://doi.org/10.1080/10408398.2011.582544>

950 Luke, D. R., Wood, N. D., Tomaszewski, K. E., & Damle, B. (2012).
951 Pharmacokinetics of sulfobutylether- β -cyclodextrin (SBECD) in subjects
952 on hemodialysis. *Nephrology, Dialysis, Transplantation: Official*
953 *Publication of the European Dialysis and Transplant Association -*
954 *European Renal Association*, 27(3), 1207–1212.
955 <https://doi.org/10.1093/ndt/gfr472>

956 Mallardo, S., De Vito, V., Malinconico, M., Volpe, M. G., Santagata, G., & Di
957 Lorenzo, M. L. (2016). Poly(butylene succinate)-based composites
958 containing β -cyclodextrin/d-limonene inclusion complex. *European*
959 *Polymer Journal*, 79, 82–96.
960 <https://doi.org/10.1016/j.eurpolymj.2016.04.024>

961 Mantilla, N., Castell-Perez, M. E., Gomes, C., & Moreira, R. G. (2013).
962 Multilayered antimicrobial edible coating and its effect on quality and
963 shelf-life of fresh-cut pineapple (*Ananas comosus*). *LWT - Food Science*
964 *and Technology*, 51(1), 37–43. <https://doi.org/10.1016/j.lwt.2012.10.010>

965 Marshall, J. J., & Miwa, I. (1981). Kinetic difference between hydrolyses of γ -
966 cyclodextrin by human salivary and pancreatic α -amylases. *Biochimica et*
967 *Biophysica Acta (BBA) - Enzymology*, 661(1), 142–147.
968 [https://doi.org/10.1016/0005-2744\(81\)90093-0](https://doi.org/10.1016/0005-2744(81)90093-0)

969 Matencio, A., Alcaráz-Gómez, M. A., García-Carmona, F., Arias, B., & López-
970 Nicolás, J. M. (2018). Application of a simple methodology to analyze
971 Hydroxypropyl- β -Cyclodextrin in urine using HPLC–LS in early Niemann–
972 Pick disease type C patient. *Journal of Chromatography B*, 1093–1094,
973 47–51. <https://doi.org/10.1016/j.jchromb.2018.06.051>

974 Matencio, A., Bermejo-Gimeno, M. J., García-Carmona, F., & López-Nicolás, J.
975 M. (2017). Separating and Identifying the Four Stereoisomers of Methyl
976 Jasmonate by RP-HPLC and using Cyclodextrins in a Novel Way.
977 *Phytochemical Analysis*, 28(3), 151–158.
978 <https://doi.org/10.1002/pca.2654>

979 Matencio, A., Dhakar, N. K., Bessone, F., Musso, G., Cavalli, R., Dianzani, C.,
980 García-Carmona, F., López-Nicolás, J. M., & Trotta, F. (2020). Study of
981 oxyresveratrol complexes with insoluble cyclodextrin based
982 nanosponges: Developing a novel way to obtain their complexation
983 constants and application in an anticancer study. *Carbohydrate*
984 *Polymers*, 231, 115763. <https://doi.org/10.1016/j.carbpol.2019.115763>

985 Matencio, A., García-Carmona, F., & López-Nicolás, J. M. (2016).
986 Encapsulation of piceatannol, a naturally occurring hydroxylated
987 analogue of resveratrol, by natural and modified cyclodextrins. *Food &*
988 *Function*, 7(5), 2367–2373. <https://doi.org/10.1039/c6fo00557h>

- 989 Matencio, A., García-Carmona, F., & López-Nicolás, J. M. (2017a). Aggregation
990 of t10,c12 conjugated linoleic Acid in presence of natural and modified
991 cyclodextrins. A physicochemical, thermal and computational analysis.
992 *Chemistry and Physics of Lipids*, 204, 57–64.
993 <https://doi.org/10.1016/j.chemphyslip.2017.03.008>
- 994 Matencio, A., García-Carmona, F., & López-Nicolás, J. M. (2017b). The
995 inclusion complex of oxyresveratrol in modified cyclodextrins: A
996 thermodynamic, structural, physicochemical, fluorescent and
997 computational study. *Food Chemistry*, 232, 177–184.
998 <https://doi.org/10.1016/j.foodchem.2017.04.027>
- 999 Matencio, A., Guerrero-Rubio, M. A., Gandía-Herrero, F., García-Carmona, F.,
1000 & López-Nicolás, J. M. (2020). Nanoparticles of betalamic acid
1001 derivatives with cyclodextrins. Physicochemistry, production
1002 characterization and stability. *Food Hydrocolloids*, 106176.
1003 <https://doi.org/10.1016/j.foodhyd.2020.106176>
- 1004 Matencio, A., Hernández-García, S., García-Carmona, F., & López-Nicolás, J.
1005 M. (2019). A Way to Increase the Bioaccessibility and Photostability of
1006 Roflumilast, a COPD Treatment, by Cyclodextrin Monomers. *Polymers*,
1007 11(5). <https://doi.org/10.3390/polym11050801>
- 1008 Matencio, A., Hernández-García, S., García-Carmona, F., & Manuel López-
1009 Nicolás, J. (2017). An integral study of cyclodextrins as solubility
1010 enhancers of α -methylstilbene, a resveratrol analogue. *Food & Function*,
1011 8(1), 270–277. <https://doi.org/10.1039/C6FO01677D>
- 1012 Matencio, A., Hernández-Gil, C. J. G., García-Carmona, F., & López-Nicolás, J.
1013 M. (2017). Physicochemical, thermal and computational study of the

1014 encapsulation of ruminic acid by natural and modified cyclodextrins.
1015 *Food Chemistry*, 216, 289–295.
1016 <https://doi.org/10.1016/j.foodchem.2016.08.023>

1017 Matencio, A., Navarro-Orcajada, S., Conesa, I., Muñoz-Sánchez, I., Laveda-
1018 Cano, L., Cano-Yelo, D., García-Carmona, F., & López-Nicolás, J. M.
1019 (2020). Evaluation of juice and milk “food models” fortified with
1020 oxyresveratrol and β -Cyclodextrin. *Food Hydrocolloids*, 98, 105250.
1021 <https://doi.org/10.1016/j.foodhyd.2019.105250>

1022 Matencio, A., Navarro-Orcajada, S., García-Carmona, F., & Manuel López-
1023 Nicolás, J. (2018). Ellagic acid–borax fluorescence interaction:
1024 application for novel cyclodextrin-borax nanosensors for analyzing ellagic
1025 acid in food samples. *Food & Function*, 9(7), 3683–3687.
1026 <https://doi.org/10.1039/C8FO00906F>

1027 Matencio, A., Navarro-Orcajada, S., González-Ramón, A., García-Carmona, F.,
1028 & López-Nicolás, J. M. (2020). Recent advances in the treatment of
1029 Niemann pick disease type C: A mini-review. *International Journal of*
1030 *Pharmaceutics*, 584, 119440.
1031 <https://doi.org/10.1016/j.ijpharm.2020.119440>

1032 Mixcoha, E., Rosende, R., Garcia-Fandino, R., & Piñeiro, Á. (2016). Cyclo-lib: a
1033 database of computational molecular dynamics simulations of
1034 cyclodextrins. *Bioinformatics*, 32(21), 3371–3373.
1035 <https://doi.org/10.1093/bioinformatics/btw289>

1036 Mohamed, S. A. A., El-Sakhawy, M., & El-Sakhawy, M. A.-M. (2020).
1037 Polysaccharides, Protein and Lipid -Based Natural Edible Films in Food

1038 Packaging: A Review. *Carbohydrate Polymers*, 116178.
1039 <https://doi.org/10.1016/j.carbpol.2020.116178>

1040 Montenegro Rabello, M., Rolim, L. A., Rolim Neto, P. J., & Hernandez, M. Z.
1041 (2019). CycloMolder software: building theoretical cyclodextrin
1042 derivatives models and evaluating their host:guest interactions. *Journal*
1043 *of Inclusion Phenomena and Macrocyclic Chemistry*, 93(3), 301–308.
1044 <https://doi.org/10.1007/s10847-019-00880-3>

1045 Moriyama, H., Saito, Y., & Bagchi, D. (2013). Characterization of Cyclodextrin
1046 Nanoparticles as Emulsifiers. In *Bio-Nanotechnology* (pp. 476–486).
1047 John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118451915.ch27>

1048 Mortensen, A., Aguilar, F., Crebelli, R., Domenico, A. D., Dusemund, B., Frutos,
1049 M. J., Galtier, P., Gott, D., Gundert-Remy, U., Leblanc, J.-C., Lindtner,
1050 O., Moldeus, P., Mosesso, P., Parent-Massin, D., Oskarsson, A.,
1051 Stankovic, I., Waalkens-Berendsen, I., Woutersen, R. A., Wright, M., ...
1052 Lambré, C. (2016). Re-evaluation of β -cyclodextrin (E 459) as a food
1053 additive. *EFSA Journal*, 14(12), e04628.
1054 <https://doi.org/10.2903/j.efsa.2016.4628>

1055 Mura, P. (2014). Analytical techniques for characterization of cyclodextrin
1056 complexes in aqueous solution: A review. *Journal of Pharmaceutical and*
1057 *Biomedical Analysis*, 101, 238–250.
1058 <https://doi.org/10.1016/j.jpba.2014.02.022>

1059 Mura, P. (2015). Analytical techniques for characterization of cyclodextrin
1060 complexes in the solid state: A review. *Journal of Pharmaceutical and*
1061 *Biomedical Analysis*, 113, 226–238.
1062 <https://doi.org/10.1016/j.jpba.2015.01.058>

1063 Nishioka, F., Nakanishi, I., Fujiwara, T., & Tomita, K. (1984). The crystal and
1064 molecular structure of the β -cyclodextrin inclusion complex with aspirin
1065 and salicylic acid. *Journal of Inclusion Phenomena*, 2(3), 701–714.
1066 <https://doi.org/10.1007/BF00662238>

1067 Noruzi, M. (2016). Electrospun nanofibres in agriculture and the food industry: a
1068 review. *Journal of the Science of Food and Agriculture*, 96(14), 4663–
1069 4678. <https://doi.org/10.1002/jsfa.7737>

1070 Oliva, E., Mathiron, D., Bertaut, E., Landy, D., Cailleu, D., Pilard, S., Clément,
1071 C., Courot, E., Bonnet, V., & Djedaïni-Pilard, F. (2018). Physico-chemical
1072 studies of resveratrol, methyl-jasmonate and cyclodextrin interactions: an
1073 approach to resveratrol bioproduction optimization. *RSC Advances*, 8(3),
1074 1528–1538. <https://doi.org/10.1039/C7RA11619E>

1075 Pan, J., Ai, F., Shao, P., Chen, H., & Gao, H. (2019). Development of polyvinyl
1076 alcohol/ β -cyclodextrin antimicrobial nanofibers for fresh mushroom
1077 packaging. *Food Chemistry*, 300, 125249.
1078 <https://doi.org/10.1016/j.foodchem.2019.125249>

1079 Perassolo, M., Smith, M. E., Giulietti, A. M., & Rodríguez Talou, J. (2016).
1080 Synergistic effect of methyl jasmonate and cyclodextrins on
1081 anthraquinone accumulation in cell suspension cultures of *Morinda*
1082 *citrifolia* and *Rubia tinctorum*. *Plant Cell, Tissue, and Organ Culture*.
1083 <https://agris.fao.org/agris-search/search.do?recordID=US201600101960>

1084 Pereva, S., Nikolova, V., Angelova, S., Spasov, T., & Dudev, T. (2019). Water
1085 inside β -cyclodextrin cavity: amount, stability and mechanism of binding.
1086 *Beilstein Journal of Organic Chemistry*, 15, 1592–1600.
1087 <https://doi.org/10.3762/bjoc.15.163>

- 1088 Piercey, M. J., Mazzanti, G., Budge, S. M., Delaquis, P. J., Paulson, A. T., &
1089 Truelstrup Hansen, L. (2012). Antimicrobial activity of cyclodextrin
1090 entrapped allyl isothiocyanate in a model system and packaged fresh-cut
1091 onions. *Food Microbiology*, *30*(1), 213–218.
1092 <https://doi.org/10.1016/j.fm.2011.10.015>
- 1093 Pinho, E., Grootveld, M., Soares, G., & Henriques, M. (2014). Cyclodextrins as
1094 encapsulation agents for plant bioactive compounds. *Carbohydrate*
1095 *Polymers*, *101*, 121–135. <https://doi.org/10.1016/j.carbpol.2013.08.078>
- 1096 Poverenov, E., Granit, R., & Gabai, S. (2013). Encapsulation and controlled
1097 release of antifungal propionic acid utilizing biodegradable active films
1098 based on natural polymers. *European Food Research and Technology*,
1099 *237*(1), 19–26. <https://doi.org/10.1007/s00217-013-2011-0>
- 1100 Pragadheesh, V. S., Yadav, A., & Chanotiya, C. S. (2015). Role of substituents
1101 in cyclodextrin derivatives for enantioselective gas chromatographic
1102 separation of chiral terpenoids in the essential oils of *Mentha spicata*.
1103 *Journal of Chromatography B*, *1002*, 30–41.
1104 <https://doi.org/10.1016/j.jchromb.2015.07.034>
- 1105 Pulley, A. O. (1961). Studies on the Schardinger dextrans. XI. The isolation of
1106 new Schardinger dextrans. *Biochem. Biophys. Res. Commun.*, *5*, 11–15.
- 1107 Rakmai, J., Cheirsilp, B., Mejuto, J. C., Simal-Gándara, J., & Torrado-Agrasar,
1108 A. (2018). Antioxidant and antimicrobial properties of encapsulated
1109 guava leaf oil in hydroxypropyl-beta-cyclodextrin. *Industrial Crops and*
1110 *Products*, *111*, 219–225. <https://doi.org/10.1016/j.indcrop.2017.10.027>

- 1111 Rao, G. C. S., Ramadevi, K., & Sirisha, K. (2014). Effect of β -cyclodextrin on
1112 Rheological Properties of some Viscosity Modifiers. *Indian Journal of*
1113 *Pharmaceutical Sciences*, 76(6), 545–548.
- 1114 Rezaee, M., Askari, G., EmamDjomeh, Z., & Salami, M. (2018). Effect of
1115 organic additives on physiochemical properties and anti-oxidant release
1116 from chitosan-gelatin composite films to fatty food simulant. *International*
1117 *Journal of Biological Macromolecules*, 114, 844–850.
1118 <https://doi.org/10.1016/j.ijbiomac.2018.03.122>
- 1119 Saha, S., Roy, A., Roy, K., & Roy, M. N. (2016). Study to explore the
1120 mechanism to form inclusion complexes of β -cyclodextrin with vitamin
1121 molecules. *Scientific Reports*, 6(1), 35764.
1122 <https://doi.org/10.1038/srep35764>
- 1123 Salazar, S., Guerra, D., Yutronic, N., & Jara, P. (2018). Removal of Aromatic
1124 Chlorinated Pesticides from Aqueous Solution Using β -Cyclodextrin
1125 Polymers Decorated with Fe₃O₄ Nanoparticles. *Polymers*, 10(9), 1038.
1126 <https://doi.org/10.3390/polym10091038>
- 1127 Schönbeck, C., & Holm, R. (2019). Exploring the Origins of Enthalpy–Entropy
1128 Compensation by Calorimetric Studies of Cyclodextrin Complexes. *The*
1129 *Journal of Physical Chemistry B*, 123(31), 6686–6693.
1130 <https://doi.org/10.1021/acs.jpccb.9b03393>
- 1131 Serfert, Y., Drusch, S., & Schwarz, K. (2010). Sensory odour profiling and lipid
1132 oxidation status of fish oil and microencapsulated fish oil. *Food*
1133 *Chemistry*, 123(4), 968–975.
1134 <https://doi.org/10.1016/j.foodchem.2010.05.047>

- 1135 Sharif, N., Golmakani, M.-T., Niakousari, M., Hosseini, S. M. H., Ghorani, B., &
1136 Lopez-Rubio, A. (2018). Active Food Packaging Coatings Based on
1137 Hybrid Electrospun Gliadin Nanofibers Containing Ferulic
1138 Acid/Hydroxypropyl-Beta-Cyclodextrin Inclusion Complexes.
1139 *Nanomaterials*, 8(11), 919. <https://doi.org/10.3390/nano8110919>
- 1140 Sherje, A. P., Dravyakar, B. R., Kadam, D., & Jadhav, M. (2017). Cyclodextrin-
1141 based nanosponges: A critical review. *Carbohydrate Polymers*,
1142 173(Supplement C), 37–49. <https://doi.org/10.1016/j.carbpol.2017.05.086>
- 1143 Shieh, W. J., & Hedges, A. R. (1996). Properties and Applications of
1144 Cyclodextrins. *Journal of Macromolecular Science, Part A*, 33(5), 673–
1145 683. <https://doi.org/10.1080/10601329608010886>
- 1146 Shin, J., Lee, E. J., & Ahn, D. U. (2018). Electrospinning of tri-acetyl- β -
1147 cyclodextrin (TA- β -CD) functionalized low-density polyethylene to
1148 minimize sulfur odor volatile compounds. *Food Packaging and Shelf Life*,
1149 18, 107–114. <https://doi.org/10.1016/j.fpsl.2018.10.005>
- 1150 Shishido, T. K., Jokela, J., Kolehmainen, C.-T., Fewer, D. P., Wahlsten, M.,
1151 Wang, H., Rouhiainen, L., Rizzi, E., De Bellis, G., Permi, P., & Sivonen,
1152 K. (2015). Antifungal activity improved by coproduction of cyclodextrins
1153 and anabaenolysins in Cyanobacteria. *Proceedings of the National
1154 Academy of Sciences of the United States of America*, 112(44), 13669–
1155 13674. <https://doi.org/10.1073/pnas.1510432112>
- 1156 Singh, J., Dartois, A., & Kaur, L. (2010). Starch digestibility in food matrix: a
1157 review. *Trends in Food Science & Technology*, 21(4), 168–180.
1158 <https://doi.org/10.1016/j.tifs.2009.12.001>

- 1159 Suratman, L. L. I., Jeon, I. J., & Schmidt, K. A. (2004). Ability of Cyclodextrins to
1160 Entrap Volatile Beany Flavor Compounds in Soymilk. *Journal of Food*
1161 *Science*, 69(2), fct109–fct113. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2621.2004.tb15499.x)
1162 [2621.2004.tb15499.x](https://doi.org/10.1111/j.1365-2621.2004.tb15499.x)
- 1163 Swaminathan, S., Cavalli, R., & Trotta, F. (2016). Cyclodextrin-based
1164 nanosponges: a versatile platform for cancer nanotherapeutics
1165 development. *Wiley Interdisciplinary Reviews: Nanomedicine and*
1166 *Nanobiotechnology*, 8(4), 579–601. <https://doi.org/10.1002/wnan.1384>
- 1167 Szente, L., & Fenyvesi, É. (2018). Cyclodextrin-Enabled Polymer Composites
1168 for Packaging. *Molecules*, 23(7), 1556.
1169 <https://doi.org/10.3390/molecules23071556>
- 1170 Szente, L., & Szejtli, J. (2004). Cyclodextrins as food ingredients. *Trends in*
1171 *Food Science & Technology*, 15(3–4), 137–142.
1172 <https://doi.org/10.1016/j.tifs.2003.09.019>
- 1173 Tao, X., Huang, Y., Wang, C., Chen, F., Yang, L., Ling, L., Che, Z., & Chen, X.
1174 (2020). Recent developments in molecular docking technology applied in
1175 food science: a review. *International Journal of Food Science &*
1176 *Technology*, 55(1), 33–45. <https://doi.org/10.1111/ijfs.14325>
- 1177 Tian, Y., Li, Y., Manthey, F. A., Xu, X., Jin, Z., & Deng, L. (2009). Influence of β -
1178 cyclodextrin on the short-term retrogradation of rice starch. *Food*
1179 *Chemistry*, 116(1), 54–58.
1180 <https://doi.org/10.1016/j.foodchem.2009.02.003>
- 1181 Tonkova, A. (1998). Bacterial cyclodextrin glucanotransferase. *Enzyme and*
1182 *Microbial Technology*, 8, 678–686. [https://doi.org/10.1016/S0141-](https://doi.org/10.1016/S0141-0229(97)00263-9)
1183 [0229\(97\)00263-9](https://doi.org/10.1016/S0141-0229(97)00263-9)

- 1184 Ünlüsayın, M., Hădărugă, N. G., Rusu, G., Gruia, A. T., Păunescu, V., &
1185 Hădărugă, D. I. (2016). Nano-encapsulation competitiveness of omega-3
1186 fatty acids and correlations of thermal analysis and Karl Fischer water
1187 titration for European anchovy (*Engraulis encrasicolus* L.) oil/ β -
1188 cyclodextrin complexes. *LWT - Food Science and Technology*, 68, 135–
1189 144. <https://doi.org/10.1016/j.lwt.2015.12.017>
- 1190 Varan, G., Varan, C., Erdoğar, N., Hincal, A. A., & Bilensoy, E. (2017).
1191 Amphiphilic cyclodextrin nanoparticles. *International Journal of*
1192 *Pharmaceutics*, 531(2), 457–469.
1193 <https://doi.org/10.1016/j.ijpharm.2017.06.010>
- 1194 Vázquez, M. B., Matencio, A., García-Carmona, F., & López-Nicolás, J. M.
1195 (2019). Nanoencapsulation as fluorescence enhancer of vitamin L1
1196 (anthranilic acid). An exhaustive study. *Food Hydrocolloids*, 91, 198–203.
1197 <https://doi.org/10.1016/j.foodhyd.2019.01.008>
- 1198 Wang, C., Wang, C., Gao, F., Xu, Y., & Guo, M. (2018). Effects of polymerized
1199 whey protein on goaty flavor and texture properties of fermented goat
1200 milk in comparison with β -cyclodextrin. *Journal of Dairy Research*, 85(4),
1201 465–471. <https://doi.org/10.1017/S0022029918000742>
- 1202 Wang, R., Zhou, H., Siu, S. W. I., Gan, Y., Wang, Y., & Ouyang, D. (2015).
1203 *Comparison of Three Molecular Simulation Approaches for Cyclodextrin-*
1204 *Ibuprofen Complexation* [Research article]. *Journal of Nanomaterials*.
1205 <https://doi.org/10.1155/2015/193049>
- 1206 Wang, Y., Yuan, C., Liu, Y., Xu, D., & Cui, B. (2019). The influence of a
1207 hydroxypropyl-beta-cyclodextrin composite on the gelation of kappa-

- 1208 carrageenan. *Food Hydrocolloids*, 90, 276–284.
- 1209 <https://doi.org/10.1016/j.foodhyd.2018.12.037>
- 1210 Wei, B., Romero-Zerón, L., & Rodrigue, D. (2015). Improved viscoelasticity of
1211 xanthan gum through self-association with surfactant: β -cyclodextrin
1212 inclusion complexes for applications in enhanced oil recovery. *Polymer*
1213 *Engineering & Science*, 55(3), 523–532.
- 1214 <https://doi.org/10.1002/pen.23912>
- 1215 Wen, P., Zhu, D.-H., Wu, H., Zong, M.-H., Jing, Y.-R., & Han, S.-Y. (2016).
1216 Encapsulation of cinnamon essential oil in electrospun nanofibrous film
1217 for active food packaging. *Food Control*, 59, 366–376.
- 1218 <https://doi.org/10.1016/j.foodcont.2015.06.005>
- 1219 Wood, W. E., Beaverson, N. J., & Kuduk, W. J. (2017). *Maturation or ripening*
1220 *inhibitor release from polymer, fiber, film, sheet or packaging* (United
1221 States Patent No. US9642356B2).
- 1222 <https://patents.google.com/patent/US9642356B2/en>
- 1223 Xi, Y., Luo, Z., Lu, X., & Peng, X. (2018). Modulation of Cyclodextrin Particle
1224 Amphiphilic Properties to Stabilize Pickering Emulsion. *Journal of*
1225 *Agricultural and Food Chemistry*, 66(1), 228–237.
- 1226 <https://doi.org/10.1021/acs.jafc.7b03940>
- 1227 Xi, Y., Zou, Y., Luo, Z., Qi, L., & Lu, X. (2019). pH-Responsive Emulsions with
1228 β -Cyclodextrin/Vitamin E Assembled Shells for Controlled Delivery of
1229 Polyunsaturated Fatty Acids. *Journal of Agricultural and Food Chemistry*,
1230 67(43), 11931–11941. <https://doi.org/10.1021/acs.jafc.9b04168>
- 1231 XiaoYun, J., ShunLiang, Z., WenYing, L., YingNan, L., Yan, Z., Kai, Y., Le, W.,
1232 Xiang, L., ShouWei, W., & XiaoYu, C. (2018). Effect of incorporation of

1233 spearmint extract microencapsulated with β -cyclodextrin on properties of
1234 polypropylene packaging films and emulsion sausage shelf life. *Shipin*
1235 *Kexue / Food Science*, 39(11), 241–246.

1236 Xu, A., Zhan, J.-C., & Huang, W.-D. (2015). Effects of ultraviolet C, methyl
1237 jasmonate and salicylic acid, alone or in combination, on stilbene
1238 biosynthesis in cell suspension cultures of *Vitis vinifera* L. cv. Cabernet
1239 Sauvignon. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 122(1), 197–
1240 211. <https://doi.org/10.1007/s11240-015-0761-z>

1241 Yao, X., Huang, P., & Nie, Z. (2019). Cyclodextrin-Based Polymer Materials:
1242 from Controlled Synthesis to Applications. *Progress in Polymer Science*.
1243 <https://doi.org/10.1016/j.progpolymsci.2019.03.004>

1244 Ye, Y., Zhu, M., Miao, K., Li, X., Li, D., & Mu, C. (2017). Development of
1245 Antimicrobial Gelatin-Based Edible Films by Incorporation of Trans-
1246 Anethole/ β -Cyclodextrin Inclusion Complex. *Food and Bioprocess*
1247 *Technology*, 10(10), 1844–1853. [https://doi.org/10.1007/s11947-017-](https://doi.org/10.1007/s11947-017-1954-8)
1248 1954-8

1249 Yildiz, Z. I., Celebioglu, A., Kilic, M. E., Durgun, E., & Uyar, T. (2018). Fast-
1250 dissolving carvacrol/cyclodextrin inclusion complex electrospun fibers
1251 with enhanced thermal stability, water solubility, and antioxidant activity.
1252 *Journal of Materials Science*, 53(23), 15837–15849.
1253 <https://doi.org/10.1007/s10853-018-2750-1>

1254 Yuan, C., Du, L., Zhang, G., Jin, Z., & Liu, H. (2016). Influence of cyclodextrins
1255 on texture behavior and freeze-thaw stability of kappa-carrageenan gel.
1256 *Food Chemistry*, 210, 600–605.
1257 <https://doi.org/10.1016/j.foodchem.2016.05.014>

- 1258 Yuan, C., Sang, L., Wang, Y., & Cui, B. (2018). Influence of cyclodextrins on the
1259 gel properties of kappa-carrageenan. *Food Chemistry*, 266, 545–550.
1260 <https://doi.org/10.1016/j.foodchem.2018.06.060>
- 1261 Yuan, C., Xu, D., Cui, B., & Wang, Y. (2019). Gelation of κ -carrageenan/Konjac
1262 glucomannan compound gel: Effect of cyclodextrins. *Food*
1263 *Hydrocolloids*, 87, 158–164.
1264 <https://doi.org/10.1016/j.foodhyd.2018.07.037>
- 1265 Zarandona, I., Barba, C., Guerrero, P., de la Caba, K., & Maté, J. (2020).
1266 Development of chitosan films containing β -cyclodextrin inclusion
1267 complex for controlled release of bioactives. *Food Hydrocolloids*, 104,
1268 105720. <https://doi.org/10.1016/j.foodhyd.2020.105720>
- 1269 Zhang, L., Man, S., Qiu, H., Liu, Z., Zhang, M., Ma, L., & Gao, W. (2016).
1270 Curcumin-cyclodextrin complexes enhanced the anti-cancer effects of
1271 curcumin. *Environmental Toxicology and Pharmacology*, 48, 31–38.
1272 <https://doi.org/10.1016/j.etap.2016.09.021>
- 1273 医薬品医療機器レギュラトリーサイエンス財団. (2017). *Japanese*
1274 *Pharmacopoeia*. Stationery Office Books (TSO).
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1278 FIGURE CAPTIONS

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1280 **Fig. 1:** (A) Structure of β -CD. (B) Schematic illustration of an inclusion
1281 complex between Piceatannol (a bioactive compound) and β -CD. Molecular
1282 docking image was obtained from published data(Matencio et al., 2016).

1283 **Fig.2:** Graphical representation of the production or extraction of bioactive
1284 compounds using CDs.

1285 **Fig.3:** Graphical representation of the use of CDs to produce “free”
1286 products and induce organoleptic modifications.

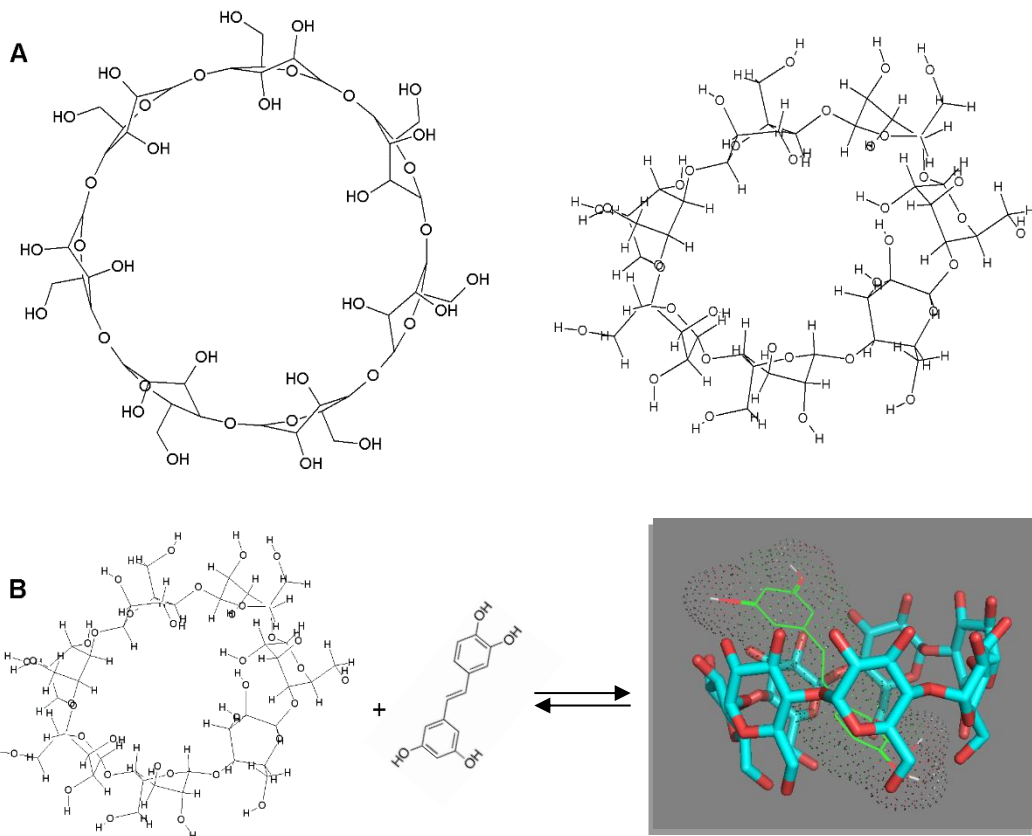
1287 **Fig.4:** Graphical representation of cyclodextrins for developing novel
1288 nanosensors.

1289 **Fig. 5:** Graphical representation of the applications of CDs in food
1290 packaging

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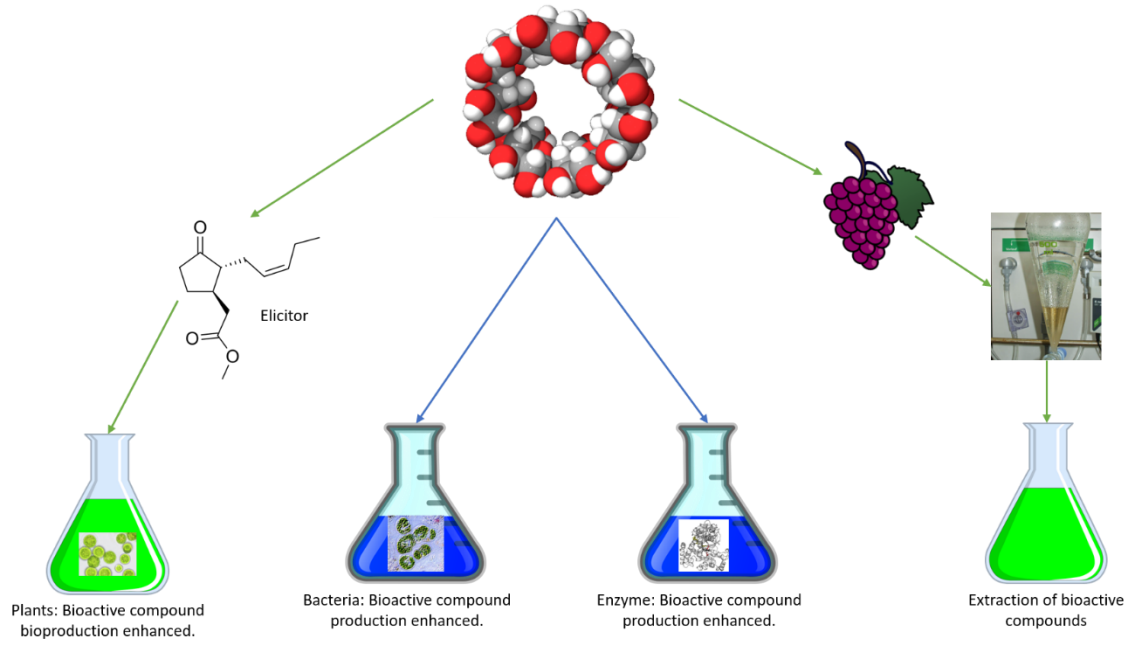


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Fig 1.

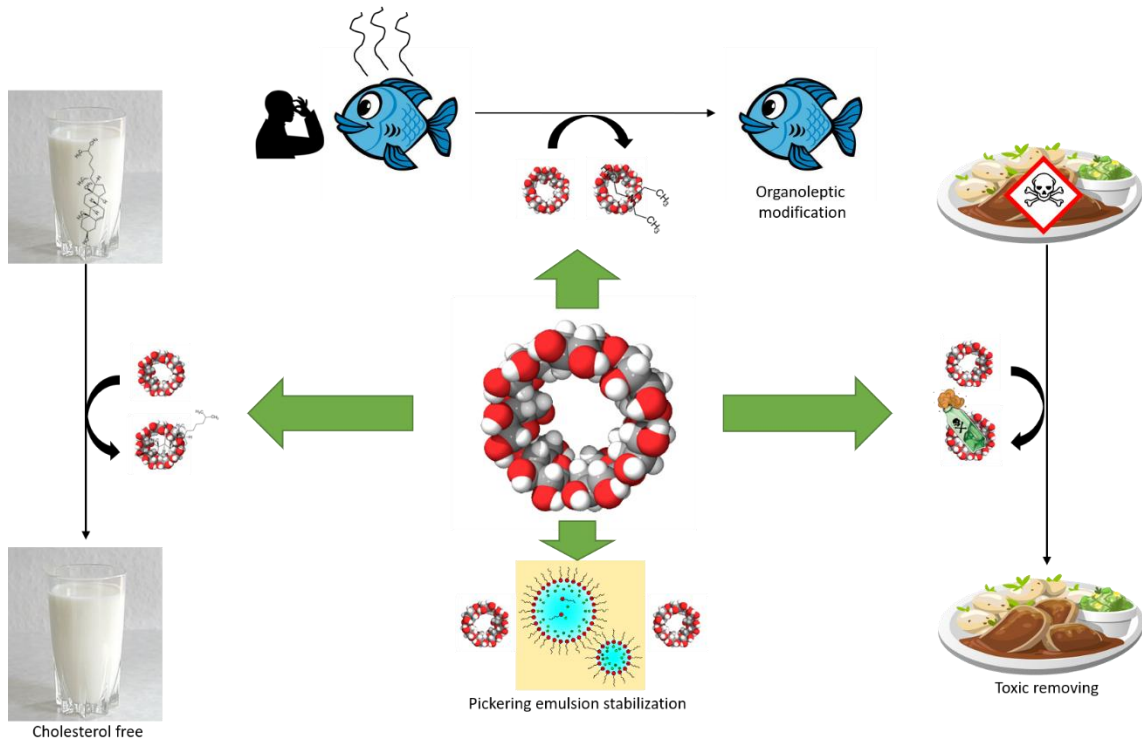


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Fig. 2



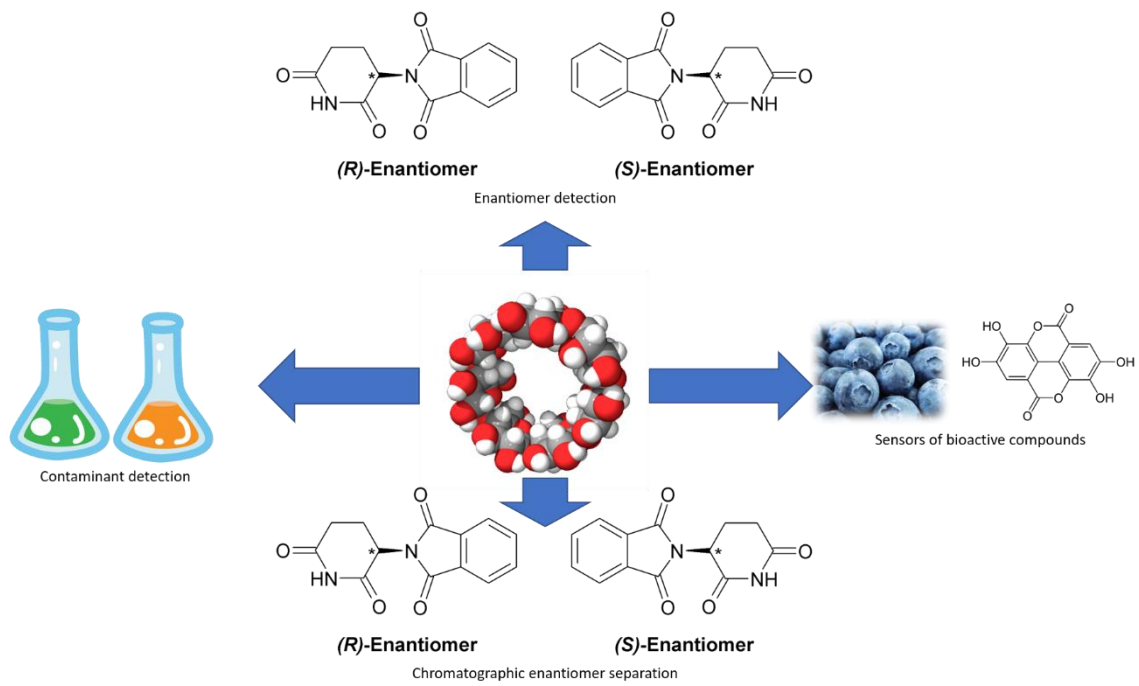
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Fig. 3



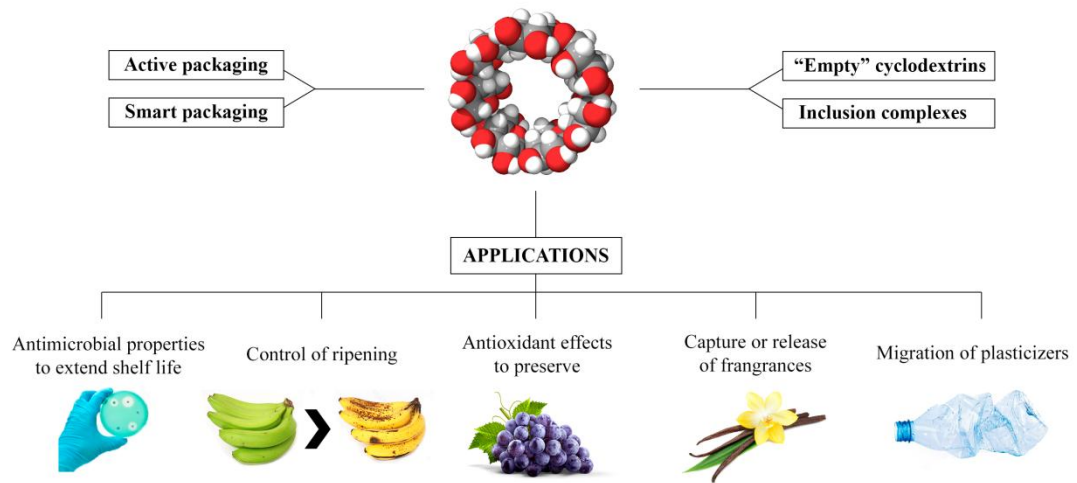
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Fig. 4



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Fig. 5

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TABLES

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Table 1: Different methods to characterize CD inclusion complexes.

Technique basis	Technique
Spectroscopic	Ultraviolet/Visible (UV)
	Circular dichroism
	Fluorescence
	Nuclear magnetic resonance (NMR)
	Electrons spin resonance (ESR)
	Fourier-transform infrared (FTIR)
Electroanalytical	Polarography
	Voltammetry
	Potentiometry
	Conductimetry
Separative	High performance liquid chromatography (HPLC)
	Capillary electrophoresis (CE)
	Gas chromatography
Thermal	Differential scanning calorimetry (DSC)
	Thermal gravimetric analysis (TGA)
	Isothermal titration calorimetry
Microscopy	Hot Stage microscopy (HSM)
X-Ray	Single crystal X-ray diffraction (SCXRD)
	Powder X-ray diffraction (PXRD)
	Polarimetry

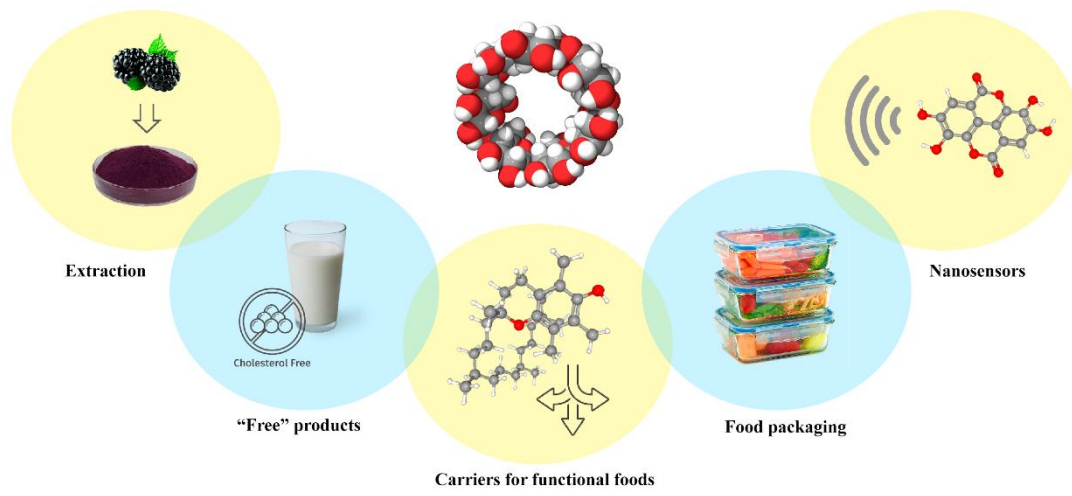
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Graphical Abstract

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