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**Chemical characterisation and cytotoxic effects in A549 cells of urban-air PM10 collected in Torino, Italy.**

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(Article begins on next page)



# UNIVERSITÀ DEGLI STUDI DI TORINO

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CHARACTERIZATION OF URBAN-AIR PM10 COLLECTED IN TORINO (ITALY).

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Gilli

Abstract: Human type II alveolar cell line (A549) was exposed to aqueous and organic solvent PM10 extracts in order to evaluate cell proliferation, proinflammatory cytokines and cytotoxicity (lactate dehydrogenase, LDH). PM10 samples, collected in Torino (north-west Italy), were analyzed for inorganic chemical species (bioavailable iron and secondary particulates) and endotoxins, potentially inflammatory promoters to human airways.

Meanly, in the considered period, PM10 concentration was  $55.4 \pm 39.1 \mu\text{g}/\text{m}^3$ , iron concentration was  $0.078 \pm 0.095$  and secondary particles constituted  $42 \pm 9 \%$  of the PM10 total mass. PM10 inhibits cell proliferation and induces both IL-6 and LDH release in a dose and time dependent manner, with a seasonal trend. The different roles of aqueous and organic solvent PM10 extracts demonstrate the importance of particle composition for the biological effects. Nevertheless few significant correlations were found between the biological effects and PM10 components evaluated in this methods development study.

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Great interests in the monitoring and risk assessment of the environmental pollutants.

Opposed Reviewers:

Dear Editor,

We are sending the manuscript "*Cytotoxic effects induced on A549 cells and chemical characterization of urban-air PM10 collected in Torino (Italy)*" that we submit for possible publication on *Environmental Toxicology and Pharmacology*.

PM10 samples were collected in an urban site of Torino, an industrial north-western Italian city, with high levels of particulate matter among European cities. PM10 samples were analyzed for the amount of inorganic chemical species (bioavailable iron and secondary particulates: sulfates and nitrates) and endotoxins, described as potentially inflammatory promoters to human airways. In order to better understand how ambient particulate matter affects lung health, A549 cells were exposed to aqueous and organic solvent PM10 extracts. Effects on cell proliferation, on the release of proinflammatory cytokines and cytotoxicity, measuring lactate dehydrogenase (LDH) release in the extracellular medium, were evaluated.

In general PM10 inhibits cell proliferation and induces both IL-6 and LDH release in a dose and time dependent manner and these biological effects have a seasonal trend. Few significant correlations were found between the biological effects and PM10 components evaluated in this methods development study. Nevertheless the different roles of aqueous and organic solvent PM10 extracts demonstrate the importance of particle composition for the biological effects.

Best regards

Tiziana Schilirò

1     1     **CYTOTOXIC EFFECTS INDUCED ON A549 CELLS AND CHEMICAL**  
2  
3     2     **CHARACTERIZATION OF URBAN-AIR PM10 COLLECTED IN TORINO (ITALY).**

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7     4     Tiziana Schilirò<sup>1\*</sup>, Luca Alessandria<sup>1</sup>, Raffaella Degan<sup>1</sup>, Deborah Traversi<sup>1</sup>, Giorgio  
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41     19     **Running head:** Biological effects of PM10 induced on A549 cells.

1 28 **Abstract**

2  
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5 30 PM10 extracts in order to evaluate cell proliferation, proinflammatory cytokines and  
6  
7 31 cytotoxicity (lactate dehydrogenase , LDH). PM10 samples, collected in Torino (north-  
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9 32 west Italy), were analyzed for inorganic chemical species (bioavailable iron and  
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13 34 airways.

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22 38 release in a dose and time dependent manner, with a seasonal trend. The different  
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## 1. Introduction

The health burden due to particulate matter (PM) air pollution is one of the biggest environmental health concerns in the World Health Organization (WHO) European Region and all around the world. PM is a complex and heterogeneous mixture, whose composition (particle size distribution and chemical characteristics) changes in time, space and depends on emissions from various sources, atmospheric chemistry and weather conditions (WHO, 2007).

The breathable fraction of ambient PM is often referred to PM<sub>10</sub>, defined as particles with a median aerodynamic diameter less than 10  $\mu\text{m}$ , that is a quality air indicator widely measured (WHO, 2006). PM<sub>10</sub> may further be divided into two main size fractions, a “coarse” (2.5–10  $\mu\text{m}$ ) and a “fine” (0.1–2.5  $\mu\text{m}$ ). The *coarse* fraction is dominated by natural sources (geological material: fugitive and resuspended dust; biological material: pollen, endotoxin) and its composition changes depending on the geology of the site considered. The *fine* fraction is dominated by anthropogenic emissions: mixture of carbon particles from combustion processes and secondary particles produced by photochemical reactions in the atmosphere (sulfates and nitrates). The carbonaceous fraction consists of aggregates of organic and inorganic carbon on which are adsorbed transition metals (Pb, Cd, V, Ni, Cu, Zn, Mn, Fe), organic compounds and biological constituents (US EPA, 1996).

Epidemiological and experimental studies have shown that particulate air pollution may induce and aggravate respiratory and cardiovascular diseases. Significant associations between exposure to ambient air particles and increased morbidity and mortality have been demonstrated (Englert 2004; Katsouyanni et al. 2001; Krewski and Rainham 2007; Pope 2007). Most of the available studies do not attribute the observed health effects to a particular characteristic of PM (WHO, 2007). Moreover the exact physiochemical mechanism by which PM produces adverse effects is still poorly



1 81 known; one of the hypotheses considered on PM's mechanisms of action is the  
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3 82 oxidative potential of the particles or specific components.  
4  
5 83 Several *in vitro* studies with collected particulate matter fractions indicate that particles  
6  
7 84 with a higher oxidative potential have a greater ability to deplete antioxidant defences  
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9 85 and to induce airway inflammation (Ghio et al. 2002; Moller et al. 2008; Schins et al.  
10  
11 86 2004). In this process transition metals such as Fe (Carter et al. 1997; Hutchison et al.  
12  
13 87 2005; Schins et al. 2004), organics such as polycyclic aromatic hydrocarbons (PAHs)  
14  
15 88 (Billet et al. 2008) and endotoxins (Becker et al. 2003; Oberdorster et al. 2000) in PM  
16  
17 89 seem to be involved (Donaldson et al. 2003; Schwarze et al. 2006; Sorensen et al.  
18  
19 90 2003). Moreover, although secondary inorganic aerosols have less toxic activity when  
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21 91 tested under controlled laboratory conditions, epidemiological studies show significant  
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23 92 associations between sulfates and nitrates and various health outcomes. In ambient  
24  
25 93 air, this fraction may act as a carrier for other components or as a surrogate for PM  
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27 94 emitted from the combustion of sulfur-containing fuels (Schwarze et al. 2006).  
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30 95 Recent studies have emphasised the ability of particles to induce cellular responses in  
31  
32 96 different types of lung cells, as well as inflammation and toxic effects in the whole lung  
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34 97 (Calcabrini et al. 2004; Churg and Brauer 2000; de Kok et al. 2006; Gualtieri et al.  
35  
36 98 2008; Hetland et al. 2004; Monn and Becker 1999; Shi et al. 2006). *In vitro* studies on  
37  
38 99 the toxicity of PM10 and its fractions were made by exposing cultures of macrophages  
39  
40 100 and pulmonary epithelial cells to suspension of particles (Becker et al. 2005; Churg and  
41  
42 101 Brauer 2000). The results of these studies show that exposure to environmental  
43  
44 102 particles induces cytotoxicity, increase in transcription factors expression (NFkB) and  
45  
46 103 release in the culture medium of cytokines (such as IL-1, IL- 6, IL-8, MIP-2, GM CSF,  
47  
48 104 TNF) and reactive oxygen species (Becker et al. 2005; Huang et al. 2002; Monn and  
49  
50 105 Becker 1999; Ning et al. 2000). Other *in vitro* studies show that particles can induce  
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52 106 damage in plasma membrane and release of cytosolic enzyme lactate dehydrogenase  
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(LDH) (Billet et al. 2007; Garcon et al. 2006; Geng et al. 2006; Molinelli et al. 2002); LDH is considered a marker for acute cytotoxicity (Seagrave and Nikula 2000). The aim of the present study was to investigate the biological *in vitro* effect of urban-air PM10 collected in Torino, a north-western Italian city, during two years (2005-2006). PM10 samples were analyzed for the amount of inorganic species (bioavailable iron and secondary particulates: sulfates and nitrates) and endotoxins, described as potentially toxic to human airways. Adherent A549 cells were exposed to PM10 aqueous and organic solvent extracts at different concentrations. A549 cells constituted a useful *in vitro* model for studying human respiratory epithelial cell biology as they present characteristics similar to human alveolar type II cells (Hatch 1992; Li et al. 2003; Shi et al. 2006; Veronesi et al. 2002) . This study investigated whether different PM10 extracts could 1) affect the cell proliferation, 2) increase the release of proinflammatory cytokines e.g. IL-6, and 3) induce cytotoxicity by measuring LDH release in culture medium. The association of the chemical and *in vitro* toxicological characterization of urban-air PM10 in one of the most industrialized areas of Italy, with high levels of particulate matter among European cities (Hazenkamp-Von Arx et al. 2004; Marcazzan et al. 2003) may provide scientific evidences for environmental and sanitary purposes.

## 2. Materials and Methods

### 2.1 PM10 sampling

PM10 (PM passing through a size-selective inlet with a 50% efficiency cut-off at 10  $\mu$ m aerodynamic diameter) was sampled on glass microfiber filters (Type A/E, 8" x 10", Gelman Sciences, Michigan, USA), with a Sierra Andersen High Volume Sampler 1200/VFC (Andersen Samplers, Atlanta, Georgia, USA) using a flow of 1160 L/min. Sample duration was controlled by a timer accurate to  $\pm$  15 min over a 24 hr sample period. The exact flow was calculated daily, corrected for variation in atmospheric

1 134 pressure and actual differential pressure across the filter. The filters were pre- and  
2  
3 135 post- conditioned by moving them to a dry and dark environment for 48h, and they  
4  
5 136 were weighed in a room with controlled temperature and humidity. Procedures were  
6  
7 137 conducted according to the European Committee for Standardization (CEN, 1998). The  
8  
9 138 PM10 concentration,  $C$  ( $\mu\text{g}/\text{m}^3$ ), in the air volume sampled,  $V$  (L), was calculated as  
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11  
12 139 follows:

$$14 140 C = [(W_2 - W_1) - (B_2 - B_1)] * 10^3 / V$$

16 141 where  $W_1$  is the mean of three tare weights of the same filter before sampling (mg),  $W_2$   
17  
18 142 is mean of three post-sampling weights of the same sample-containing filter (mg),  $B_1$  is  
19  
20 143 mean tare weight of blank filters (mg).  $B_2$  is mean post-sampling weight of blank filters  
21  
22 144 (mg) and  $V$  is volume as sampled at the nominal flow rate.

25 145 Samplings were carried out from January 2005 to December 2006 (2 filters every  
26  
27 146 month, one sampled at the half and one at the end of every month, without considering  
28  
29 147 week-ends) in a meteorological-chemical background station located in the urban  
30  
31 148 centre of Torino, an industrial north-western Italian city (Gilli et al. 2007b).

## 34 149 *2.2 Particles extractions*

36 150 Each PM10 filter (typically 20.32 cm by 25.40 cm) was treated individually: two different  
37  
38 151 6.35 cm by 20.32 cm wide strips were cut from the same filters and each portion has  
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40 152 been used with different extraction medium: acetone and the Dulbecco's Modified  
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42 153 Eagle's Medium (DMEM) culture medium without fetal calf serum (FCS). Acetone was  
43  
44 154 chosen as solvent to extract organic-extractable compounds (Claxton et al. 2004).  
45  
46 155 DMEM without FCS was chosen to extract water-soluble components, hypothetically  
47  
48 156 comparable to the extraction at the lung cells (Calcabrini et al. 2004; Hetland et al.  
49  
50 157 2004). Each portion of the filter has been cut into little strips and placed in 50 mL  
51  
52 158 polypropylene sterile tube with 15 mL of the extraction medium. The tubes were placed  
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54 159 in an ultrasonic water bath for 10 min followed by 1 min of vortexing. This procedure  
55  
56 160 was repeated 3 times. The samples were centrifugated at 5000 rpm for 10 min to  
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1 161 remove the filter material and the supernatant was collected. Acetone extracts were  
2  
3 162 evaporated with a rotary evaporator and re-suspended in culture medium. Aqueous  
4  
5 163 extracts were directly assayed. It was assumed that particles were totally removed from  
6  
7 164 the filters through extraction procedures (Hutchison et al. 2005). Unless specified  
8  
9 165 otherwise, all chemicals were purchased from Sigma, USA.

### 12 166 *2.3 Iron and bioavailable iron determinations*

14 167 Iron determination was performed according to the procedure of Gilli et al. (2007b).  
15  
16 168 Briefly, the metals were extracted from filter strips (3.18 cm by 20.32 cm) by a nitric  
17  
18 169 acid solution. After cooling, the sample was mixed and centrifuged and the trace  
19  
20 170 element concentrations in each sample were determined by atomic absorption  
21  
22 171 spectrometry, Varian GTA-96.

25 172 The bioavailable iron from urban particulates was determined as previously described  
26  
27 173 (Lund and Aust 1990), with some modifications as reported by (Smith and Aust 1997)  
28  
29 174 and Gilli et al. (2007b). Briefly, filter strips (3.18 cm by 20.32 cm) were suspended in  
30  
31 175 NaCl 50nM, mixed and pH adjusted to 7.5. Citrate was then added to samples to obtain  
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33 176 a final concentration of 1 mM and all samples were placed on a wrist-action shaker in  
34  
35 177 the dark for 24 h. One mL samples were withdrawn and centrifuged to remove the  
36  
37 178 particulates. The amount of iron mobilized as the citrate: Fe complex in the supernatant  
38  
39 179 was determined using a spectrophotometric total non-heme iron assay (Brumby and  
40  
41 180 Massey, 1967). The concentration of iron mobilized by citrate was expressed as  $\mu\text{g}$  of  
42  
43 181  $\text{Fe}/\text{m}^3$  and as  $\text{ng}$  of  $\text{Fe}/\mu\text{g}$  of particulate.

### 48 182 *2.4 Sulfates and nitrates determination*

50 183 Sulfates and nitrates determination was performed according to the procedure of Gilli  
51  
52 184 et al. (2007a, 2007b). Briefly each filter strip (3.18 cm by 20.32 cm) was extracted in 15  
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54 185 mL in distilled deionized waters, via 30 min sonication, 30 min agitation and an  
55  
56 186 overnight refrigeration at 4°C. Prior to the analysis, the samples were centrifuged to  
57  
58 187 remove particles. Ion chromatography (IC) was used to determine the soluble ion

1 188 content sulfates ( $\text{SO}_4^-$ ) and nitrates ( $\text{NO}_3^-$ ), applying a Dionex DX-100 ion  
2  
3 189 chromatograph with  $\text{NaHCO}_3$  0.3 mM and  $\text{Na}_2\text{CO}_3$  2.7 mM for eluent and IonPac  
4  
5 190 analytical column S12A for anions. Both the utilized standards, sodium sulfate and  
6  
7 191 sodium nitrate (71959 and 71759 FLUKA respectively) ranged from 0.1  $\mu\text{g}/\text{mL}$  to 100  
8  
9 192  $\mu\text{g}/\text{mL}$ . The ions were identified by their elution/retention times (about 8.75 min and  
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11 193 12.5 min for nitrates and sulfates respectively) and quantified by the conductivity peak  
12  
13 194 area or peak height (300A method, US EPA, 1996).

### 16 195 *2.5 Endotoxins determination*

18 196 Endotoxin was assayed with an end-point chromogenic Limulus ameocyte lysate  
19  
20 197 (LAL) method (QLC-1000 n° 50-648U, Cambrex, Walkersville, MD, USA) at 37°C with  
21  
22 198 an automated micro-plate reader (ELX 800 UV, Bio-Tek Instruments, Inc) following  
23  
24 199 manufacturer's instructions. *Escherichia coli* 0111:B4 endotoxin was used as standard  
25  
26 200 endotoxin. Sample concentration were reported as endotoxin units (EU) per millilitre of  
27  
28 201 eluant, EU per milligram of PM10, and EU per cubic meter of air collected. The limit of  
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30 202 detection (LOD) was 0.01 EU/mL. To control for the potential interference of samples  
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32 203 substances with the LAL assay, all samples were spiked with a known amount of  
33  
34 204 endotoxin (0.4 EU/mL). The spiked solution was assayed along with the unspiked  
35  
36 205 samples and their respective endotoxin concentrations are determined. The difference  
37  
38 206 between these two calculated endotoxin values should equal the known concentration  
39  
40 207 of the spike  $\pm 25\%$  to accept the results of the determinations.

### 45 208 *2.6 Cell culture*

47 209 The human lung epithelial cell line A549 (non-small cell lung cancer) from Interlab Cell  
48  
49 210 Line Collection (Genova, IT) was used as a model for human epithelial lung cells. Cells  
50  
51 211 were grown as a monolayer, maintained and treated in DMEM supplemented with 10%  
52  
53 212 FCS, 2% L-glutamine 200mM, 2% HEPES 1M, 1% sodium pyruvate 100 mM and 1%  
54  
55 213 penicillin/streptomycin 10mg/mL, at 37°C in an humidified atmosphere containing 5%  
56  
57 214  $\text{CO}_2$ .

## 2.7 Cell proliferation

Cell proliferation was evaluated using crystal violet method (Kueng et al. 1989). Cells were seeded in 24-well plates at a density of  $4 \times 10^4$  cell/well and exposed to particle concentration of 50 and 100  $\mu\text{g/mL}$  (30 and 60  $\mu\text{g/cm}^2$ ), (Alfaro-Moreno et al. 2002; Calcabrini et al. 2004; Veronesi et al. 2002). Cell proliferation was determined at 24h, 48h and 72h of exposure measuring the residual cell number by crystal violet by determining the absorbance at 595 nm with a micro-plate reader (ELX 800 UV, Bio-Tek Instruments, Inc). At the same time blank filters were treated in the same way. All experiments were performed in triplicate. Percentage cell proliferation was calculated comparing the absorbance of exposed cultures with the absorbance of non-exposed cultures.

## 2.8 Proinflammatory cytokine in cell supernatant

IL-6 is a multifactorial cytokine protein that plays a major role in the mediation of airways inflammation associated with infection or toxic insult. A549 cells were exposed to particle concentration of 50 and 100  $\mu\text{g/mL}$  (30 and 60  $\mu\text{g/cm}^2$ ) and after 24, 48 and 72h of treatment, supernatant aliquots were removed for the cytokine assay. IL-6 was measured in the culture supernatant by enzyme-linked immunoabsorbent assay (ELISA) kits (PeliPair reagent set, Sanquin, Netherlands) with an automated micro-plate reader (ELX 800 UV, Bio-Tek Instruments, Inc) according to the manufacturer's recommendations. Results were calculated by expressing samples IL-6 increment as a percentage of IL-6 expression from non exposed cells.

## 2.9 LDH assay

To evaluate PM10 cytotoxicity, released lactate dehydrogenase (LDH) activities, from damaged cells, were measured in cell-free culture supernatants. Enzymatic activities, measured spectrophotometrically as absorbance variation at 340 nm (37°C), were expressed as nmol NADH oxidized/min in the conversion of pyruvate to lactate (Golladay et al. 1997; Kinnula et al. 1994; Riganti et al. 2002). A549 cells were seeded

1 242 in 6 well plates at a density of  $1 \times 10^6$  cell/well and exposed to PM10 extracts containing  
2  
3 243 50, 100 and 200  $\mu\text{g/mL}$  of particles (30, 60 and  $110 \mu\text{g/cm}^2$ ), (Billet et al. 2007; Geng et  
4  
5 244 al. 2006; Hetland et al. 2000). At the same time blank filters were treated in the same  
6  
7 245 manner. At 24h, 48h and 72h LDH activity was measured in supernatant and cell  
8  
9 246 lysate. LDH activity was calculated as ratio between the extracellular LDH (measured  
10  
11 247 in the supernatant) and total LDH (expressed as sum of LDH measured in supernatant  
12  
13 248 and cell lysate). To obtain cell lysate, cells were washed with PBS, detached with  
14  
15 249 trypsin-EDTA, resuspended with 1 mL of TRAP (82.3 mM triethanolamine  
16  
17 250 hydrochloride, pH 7.6) and sonicated for 10". Then LDH was measured by adding 250  
18  
19 251  $\mu\text{l}$  of mix containing NADH 0.25 mM and pyruvate 0.5 mM and consumption of NADH  
20  
21 252 was measured as absorbance in a microplate reader (Benchmark Plus Microplate  
22  
23 253 Reader, Biorad) at 340 nm. All experiments were performed in triplicate. Results were  
24  
25 254 calculated by expressing samples LDH activity as a percentage of LDH activity from  
26  
27 255 non exposed cells.

### 256 2.10 Statistical analysis

257 Statistical analyses were performed using the SPSS Package, version 14.0 for  
258 Windows. T-test analysis was used to compare means and Spearman correlation  
259 coefficient was used to assess relationship between variables. The mean difference  
260 and correlation were considered significant at  $p < 0.05$ .

## 261 3. Results

### 262 3.1 PM10 concentration

263 A total of 46 PM10 filters were analyzed during 2005 and 2006 without considering  
264 week-ends. The mean PM10 concentration during the whole sampling period was  $55.4$   
265  $\pm 39.1 \mu\text{g/m}^3$ . The highest value was observed in winter ( $198.5 \mu\text{g/m}^3$ ), while the lowest  
266 was observed in summer ( $12.4 \mu\text{g/m}^3$ ); the T-test analysis show significant ( $p < 0.01$ )  
267

1 268 difference for the seasonal (autumn/winter vs spring/summer) PM10 concentrations  
2  
3 269 (Table 1).

### 4 5 270 *3.2 Iron and bioavailable iron concentrations*

6  
7 271 The total iron mean concentration was  $1.035 \pm 0.811 \mu\text{g}/\text{m}^3$  and, in relation to PM10,  
8  
9 272  $19.43 \pm 11.64 \text{ ng}/\mu\text{g}$  particles. The highest value was observed in winter ( $3.751 \mu\text{g}/\text{m}^3$ ),  
10  
11 273 while the lowest was observed in summer ( $0.099 \mu\text{g}/\text{m}^3$ ). A statistically significant  
12  
13 274 difference was found for the seasonal concentrations (autumn/winter vs  
14  
15 275 spring/summer) expressed as  $\mu\text{g}/\text{m}^3$  (t-test,  $p < 0.01$ ) but not as  $\text{ng}/\mu\text{g}$  particles (Table  
16  
17 276 2). A significant positive correlation was found with PM10 concentrations (Spearman  
18  
19 277 correlation:  $r = 0.800$ ,  $p < 0.01$ ).

20  
21 278 Table 2 shows also the mean concentrations of bioavailable iron,  $6.9 \pm 4.5\%$  of total  
22  
23 279 iron was bioavailable. No statistically significant differences were found for the mean  
24  
25 280 seasonal concentrations expressed both as  $\mu\text{g}/\text{m}^3$  (t-test,  $p < 0.01$ ) and as  $\text{ng}/\mu\text{g}$   
26  
27 281 particles. A positive significant correlation was found with PM10 concentrations  
28  
29 282 (Spearman correlation:  $r = 0.670$ ,  $p < 0.01$ ).

### 30 31 283 *3.3 Sulfates and nitrates concentrations*

32  
33 284 The sulfates mean concentration was  $10.5 \pm 5.7 \mu\text{g}/\text{m}^3$  and, in relation to PM10,  $216.7$   
34  
35 285  $\pm 72.7 \text{ ng}/\mu\text{g}$  particles, such that sulfates represented  $21.2 \pm 6.9\%$  of total PM10 mass.  
36  
37 286 The nitrates mean concentration was  $11.7 \pm 9.2 \mu\text{g}/\text{m}^3$  and, in relation to PM10,  $216.3$   
38  
39 287  $\pm 80.5 \text{ ng}/\mu\text{g}$  particles, such that nitrates represented  $20.6 \pm 6.1\%$  of total PM10 mass.  
40  
41 288 Considering the sum of sulfates and nitrates as principal components of secondary  
42  
43 289 particulate, it was found that they meanly constituted  $42 \pm 9\%$  of the PM10 total mass  
44  
45 290 (Table 1). A statistically significant difference was found for the seasonal  
46  
47 291 concentrations for both sulfates and nitrates expressed as  $\mu\text{g}/\text{m}^3$  (t-test,  $p < 0.05$  and  $p$   
48  
49 292  $< 0.01$ , respectively). Considering the concentrations of the two species in relation to  
50  
51 293 PM10 (percentage), it was found that in the hot season the sulphate amount ( $23.6 \pm$   
52  
53 294  $5.6\%$ ) was more elevated than in the cold season ( $19.2 \pm 7.3\%$ ) and this trend was



1 295 significant (t-test,  $p < 0.01$ ), while for nitrates the amount was similar in the two  
2  
3 296 seasons. A significant positive correlation was found with PM10 concentrations both for  
4  
5 297 sulfates (Spearman correlation:  $r=0.740$ ,  $p<0.01$ ) and for nitrates (Spearman  
6  
7 298 correlation:  $r=0.900$ ,  $p<0.01$ ) and also a significant positive correlation between sulfates  
8  
9 299 and bioavailable iron (Spearman correlation:  $r=0.490$ ,  $p<0.01$ ).

### 11 300 *3.4 Endotoxins concentrations*

12 301 Endotoxin mean concentration during the whole sampling period was  $0.19 \pm 0.07$   
13  
14 302 EU/m<sup>3</sup> and in relation to PM10  $3.70 \pm 0.95$  EU/mg. A statistically significant difference  
15  
16 303 was found for the seasonal concentrations (autumn/winter vs spring/summer)  
17  
18 304 expressed as EU/m<sup>3</sup> ( $0.24 \pm 0.07$  vs  $0.14 \pm 0.01$  respectively) (t-test,  $p<0.05$ ) but not as  
19  
20 305 EU/mg particles. Endotoxins, EU/mg, were inversely correlated with PM10  
21  
22 306 concentrations (Spearman correlation:  $r = - 0.710$ ,  $p < 0.05$ ).

### 27 307 *3.5 A549 cell proliferation*

28 308 Mean absorbance values at 595 nm of the negative control at 24, 48 and 72 h were  
29  
30 309 respectively  $0.120 \pm 0.010$ ,  $0.225 \pm 0.010$  and  $0.410 \pm 0.015$ . Extracts from blank filters  
31  
32 310 had no significant effect on cell proliferation. Figure 1 shows effects of organic solvent  
33  
34 311 (1a) and aqueous (1b) PM10 extracts on cell proliferation. PM10 inhibits cell  
35  
36 312 proliferation in a dose and time dependent manner. Aqueous extracts seem to have  
37  
38 313 stronger effects than organic solvent extracts on cell proliferation, this trend was  
39  
40 314 confirmed by T-tests for inhibition at 48 and 72 h ( $p < 0.05$ ) but not at 24 h, for both the  
41  
42 315 tested concentrations. The maximum inhibition of cell proliferation,  $25.3 \pm 5.8$  %, was  
43  
44 316 achieved at 72h by the 100  $\mu\text{g/mL}$  aqueous extract.

45 317 Proliferation inhibition of A549 cells due to organic solvent extract, in some cases (50  
46  
47 318  $\mu\text{g/mL}$  at 72h, 100  $\mu\text{g/mL}$  at 48 and 72h) was correlated with PM10 concentrations  
48  
49 319 (Spearman correlations:  $r=0.860$ ,  $p<0.01$ ;  $r=0.810$ ,  $p<0.01$  and  $r=0.620$ ,  $p<0.05$

1 320 respectively), no correlations were found with any other parameters or for the aqueous  
2  
3 321 extracts.

4  
5 322 Figure 3(a) shows different seasonal (autumn/winter vs spring/summer) proliferation  
6  
7 323 inhibition of A549 cells at 72h after their incubation in the continuous presence of two  
8  
9 324 concentrations of particle matter (50 and 100 µg/mL). Despite PM10 organic solvent  
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11 325 extracts and PM10 aqueous extracts always had higher effect in the cold months these  
12  
13 326 differences were not significant (T-tests  $p > 0.05$ ).

### 14 327 *3.6 IL-6 release in A549 cells*

15  
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18 328 In order to investigate the proinflammatory potential of the particulate, the production of  
19  
20 329 IL-6 in the presence of two different extracts of PM10 was analyzed. Mean IL-6 release  
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22 330 in the negative control at 24, 48 and 72h range from 0.2 to 1.0 pg/mL. PM10 extracts  
23  
24 331 induce IL-6 release in a dose and time dependent manner. As shown in figure 2a, the  
25  
26 332 organic solvent extract induce significant release of IL-6 in relation to the control at 48h  
27  
28 333 and 72h of exposure (T-test  $p < 0.05$ ) for both the concentrations, the lowest  
29  
30 334 concentration at 24h of exposure has no significant increase of IL-6 release in relation  
31  
32 335 to the control ( $6.2 \pm 3.5\%$ ). The two tested concentrations exhibited similar expression  
33  
34 336 of cytokine without significant difference (t-test  $p > 0.05$ ).

35  
36  
37  
38 337 In figure 2b, the aqueous extract induce significant release of IL-6 in relation to the  
39  
40 338 control at 24h, 48h and 72h of exposure (T-test  $p < 0.05$ ) for cells exposed to 100  
41  
42 339 µg/mL while for cells exposed to 50 µg/ml only at 72h.

43  
44  
45 340 Despite the aqueous extract exhibit higher mean levels of IL-6 expression the  
46  
47 341 difference between the production of IL-6 in the presence of organic solvent extract or  
48  
49 342 aqueous extract was not statistically significant (T-test  $p > 0.05$ ) for any concentrations  
50  
51 343 or times. The maximum increment of IL-6 release,  $108.3 \pm 39.1\%$ , was obtained at 72h  
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53 344 by the 100 µg/mL aqueous extract.  
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1 345 IL-6 release by A549 due to organic solvent extracts (50 and 100 µg/mL at 72h) was  
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3 346 correlated with PM10 concentrations (Spearman correlations:  $r=0.680$ ,  $p<0.05$  and  
4  
5 347  $r=0.730$ ,  $p<0.05$  respectively). No correlations were found with any other parameters.  
6  
7 348 IL-6 release by A549 due to aqueous extracts was always correlated with PM10,  
8  
9 349 sulfate, nitrate and Fe concentrations for both the extract concentrations and at each  
10  
11 350 time of exposure in a range of Spearman correlation between  $r = 0.720$  to  $r = 0.920$ ,  
12  
13 351  $p<0.05$ .

14  
15  
16 352 Figure 3(b) shows different seasonal (autumn/winter vs spring/summer) IL-6 release at  
17  
18 353 72h after their incubation in the continuous presence of two concentrations of particle  
19  
20 354 matter (50 and 100 µg/mL). PM10 organic solvent extracts and PM10 aqueous extracts  
21  
22 355 always had higher effect in the cold months and these differences were statistically  
23  
24 356 significant (t-test,  $p < 0.01$ ) except for one case (50µg/mL organic solvent extract).

### 27 357 *3.7 LDH release in A549 cells*

28  
29 358 An increase of extracellular LDH enzyme activity reflects an increase in the amount of  
30  
31 359 membrane-damage cells (Lobner 2000). The release of the cytoplasmatic enzyme LDH  
32  
33 360 into the culture supernatant was used to measure cell cytotoxicity. Extracts from blank  
34  
35 361 filters had no significant effect on LDH release. Incubation of A549 cells with particles  
36  
37 362 at a concentration of 50 µg/mL had no effect on LDH release as compared to control  
38  
39 363 cells (data not shown). Results are shown in table 3: for organic solvent extract and for  
40  
41 364 both the concentrations the highest and significant release of LDH was obtained at 72h  
42  
43 365 of exposure (T-tests,  $p < 0.05$ ), while at 24h and 48h no significant effects were  
44  
45 366 observed in relation to the control (T-tests,  $p > 0.05$ ). For aqueous extracts exposure to  
46  
47 367 100 µg/mL of particles increase LDH release by approximately 30-35% for all  
48  
49 368 incubation time. Statistically significant increase of extracellular LDH activities were  
50  
51 369 observed at 24h, 48h and 72h after cell exposure at 200 µg/mL of particles  
52  
53 370 concentration (T-tests,  $p < 0.05$ ). The maximum increment of LDH release,  $87.6 \pm 41.9$   
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1 371 % and  $83.7 \pm 44.1$  %, was respectively obtained at 48h by the 200  $\mu\text{g}/\text{mL}$  aqueous  
2  
3 372 extract and at 72h by the 200  $\mu\text{g}/\text{mL}$  organic solvent extract .  
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5 373 LDH increase by A549 due to both organic solvent extracts and aqueous extracts was  
6  
7 374 never correlated with PM10 concentrations or with any other parameters, with the  
8  
9 375 exception of the aqueous 200  $\mu\text{g}/\text{mL}$  extract at 72h vs PM10 concentrations.  
10  
11 376 Figure 3(c) shows different seasonal (autumn/winter vs spring/summer) release of LDH  
12  
13 377 activity from A549 cells at 72h after their incubation in the continuous presence of two  
14  
15 378 concentrations of particle matter (100 and 200  $\mu\text{g}/\text{mL}$ ). PM10 organic solvent extracts  
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17 379 had similar effect in the two seasons while PM10 aqueous extracts always had higher  
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19 380 effect in the cold months, this difference was significant only for the lowest extract  
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21 381 concentration.  
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#### 27 383 **4. Discussion and Conclusions**

28 384 The PM10 monitoring in Torino has showed concentrations of PM10 often over the  
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30 385 daily ( $50 \mu\text{g}/\text{m}^3$ ) and yearly ( $40 \mu\text{g}/\text{m}^3$ ) quality targets (Air Quality Directive,  
31  
32 386 2008/50/CE). High pollution episodes are frequent during winter time in Northern Italy.  
33  
34 387 This is due to the regional unique climate and topography which hamper particles  
35  
36 388 dispersion. Human alveolar epithelial cells were exposed to concentration of particles  
37  
38 389 comparable to  $100 \mu\text{g}/\text{m}^3$  such as those found in winter season in Torino, indeed the  
39  
40 390 averages were  $93.5 \pm 54.1 \mu\text{g}/\text{m}^3$  for 2005 and  $78.4 \pm 53.1 \mu\text{g}/\text{m}^3$  for 2006 (Oberdörster  
41  
42 391 and Yu 1999).  
43  
44 392 In urban sites like that utilized in this study, significant seasonal differences in total Fe  
45  
46 393 concentrations occur probably due to minor presence of anthropogenic sources (for  
47  
48 394 example domestic and industrial heating) in the warm season (Gilli et al. 2007b;  
49  
50 395 Ziemacki et al. 2003). A small amount of iron is presumably bioavailable (about 6% of  
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52 396 total Fe) and this form is capable of generating ROS and induces inflammation in  
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1 397 cellular systems (Aust et al. 2002). In this study this bioavailable Fe has not a  
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3 398 significant seasonal trend and it is less correlated with PM10 concentration than total  
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5 399 Fe.  
6  
7 400 In relation to secondary PM10 components, the seasonal trend of the two species is  
8  
9 401 normally due to the photochemical reactions that, in warm season occur more  
10  
11 402 frequently. It was found that the secondary airborne particulate matter (as sum of  
12  
13 403 sulfates and nitrates) mass concentration was about 42% of the total PM10, and this  
14  
15 404 value was confirmed by other studies conducted in the Po valley of Northern Italy (Gilli  
16  
17 405 et al. 2007b; Marcazzan et al. 2003).  
18  
19 406 Among relevant air pollution components in causing toxicity there are also endotoxins;  
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21 407 in this study the mean endotoxins concentration was low and quite comparable with  
22  
23 408 those present in literature (Heinrich et al. 2003; Morgenstern et al. 2005; Mueller-  
24  
25 409 Anneling et al. 2004; Solomon et al. 2006).  
26  
27 410 Many studies have investigated the toxicity and mechanism of PM in the airway  
28  
29 411 epithelial cells (Scapellato and Lotti 2007); PM induces different biologic effects  
30  
31 412 depending on sampling site (Rosas-Pérez et al., 2007), size fraction (Alfaro-Moreno et  
32  
33 413 al. 2002; Hetland et al. 2004; Osornio-Vargas et al. 2003), sampling time (Frampton et  
34  
35 414 al. 1999) and contaminants adsorbed on the particles (Baulig et al. 2003; Billet et al.  
36  
37 415 2007; Calcabrini et al. 2004; Muller et al. 2006). Moreover all in vitro studies with PM10  
38  
39 416 usually used high concentration of particles so that a potential confounding effect of  
40  
41 417 high doses need to be taken into account when extrapolating to *in vivo* conditions  
42  
43 418 (Oberdorster and Yu 1999).  
44  
45 419 In this study PM10 inhibits cell proliferation and induces both IL-6 and LDH release in a  
46  
47 420 dose and time dependent manner according to other studies on epithelial cells (Alfaro-  
48  
49 421 Moreno et al. 2002; Osornio-Vargas et al. 2003). These biological effects have a  
50  
51 422 seasonal trend as well as other biological effects such as mutagenicity evaluated in a  
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53 423 study conducted in the Torino city (Gilli et al. 2007a).  
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1 424 In general the maximum biological effect on A549 cells was obtained in term of IL-6  
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3 425 release ( $108.3 \pm 39.1$  %), then LDH production ( $87.6 \pm 41.9$  %) and in the end inhibition  
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5 426 of cell proliferation ( $25.3 \pm 5.8$  %). Indeed other studies have shown that PM10 induce  
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7  
8 427 - first the production of reactive oxygen species and inflammatory mediators such as  
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10 428 interleukin IL-6 - than acute cytotoxicity as release of LDH and in the end - an alteration  
11  
12 429 of the cellular function highlighted by inhibition of the cellular proliferation (Jalava et al.  
13  
14 430 2006). It is also important to consider that these outcomes might occur independently  
15  
16 431 or in association with each others (Seagrave and Nikula 2000).  
17  
18 432 In our investigation aqueous extracts seem to induce a greater IL-6 release and a  
19  
20 433 greater proliferation inhibition than organic solvent (acetone) ones, while, on the  
21  
22 434 contrary, organic solvent extract induce a greater release of LDH. Probably ,although  
23  
24 435 both the two PM10 extracts produce effects on cells, water-soluble components, such  
25  
26 436 as transition metals, could rapidly cross the plasma membrane and induce the  
27  
28 437 inflammatory response, while polar compounds, from organic solvent extracts, cross  
29  
30 438 the plasma membrane more slowly and are able to directly interact with DNA and  
31  
32 439 induce mutations (Gilli et al. 2007a; Gilli et al. 2007b).  
33  
34 440 In general the biological effects are frequently attributed to the intracellular generation  
35  
36 441 of ROS, usually correlated with aqueous PM extracts, that may damage protein  
37  
38 442 structure, plasmatic membrane associated lipids and other cellular compartments  
39  
40 443 (Knaapen et al. 2004; Sevanian and Ursini 2000; Sorensen et al. 2003). Soukup and  
41  
42 444 Becker (Soukup and Becker 2001) found that expression of cytokines by human  
43  
44 445 alveolar macrophages incubated with ambient insoluble PM10 was more than 50 times  
45  
46 446 higher than that caused by the water-soluble fraction. Another study, conducted by  
47  
48 447 Javala et al. (2008), is focused on the stronger cytotoxic effects of insoluble (organic)  
49  
50 448 than water-soluble fraction of PM. In another investigation, mRNA levels of IL-1 $\alpha$  and  
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52 449 IL-8 were significantly more expressed by human bronchial epithelial cells when  
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1 450 incubated with the water-soluble fraction of ambient PM10 than with the corresponding  
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3 451 suspensions (10–500 µg/mL, up to 24 h) (Fujii et al. 2001). To our knowledge, few  
4  
5 452 studies in literature compare the different effects of urban PM10 extracts.  
6  
7 453 Few significant correlations were found between the biological effects and PM10  
8  
9 454 components (bioavailable iron, secondary particulates and endotoxins) evaluated in  
10  
11 455 this methods development study and it is important to note that the correlations have  
12  
13 456 become significant only after some days of exposure (72h). These results confirmed  
14  
15 457 other recent studies in which, for *in vitro* lung cells exposure, there were inconsistent  
16  
17 458 associations between both cell proliferation and IL-6 release and secondary  
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19 459 particulates or metals (Happo et al. 2008). One possible explanation for the presence  
20  
21 460 of few correlations could be that the biological end-points and the PM10 components  
22  
23 461 were quantified in chemically separate fractions. Furthermore the aqueous extracts did  
24  
25 462 not reflected the activity of any particular PM10 chemicals components but only  
26  
27 463 highlighted the activity of physiologic-soluble components.  
28  
29 464 Nevertheless the significant difference in biological effects induced by the two different  
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31 465 extracts (aqueous and organic solvent) and the observed seasonal trend, support the  
32  
33 466 need of further studies, in order to determine particle components responsible for  
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35 467 biological effects associated both to PM10 and fine particulate matter.  
36  
37 468 The identification of PM components carrying the relevant burden in causing  
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39 469 inflammation remains unsettled, and it was said to represent one of the biggest gaps in  
40  
41 470 our knowledge (Pope and Dockery 2006).  
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43 471 In the future, better understanding of the relative toxicity and health effects of particles  
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45 472 from various sources could facilitate targeted abatement policies and more effective  
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47 473 control measures to reduce the burden of disease due to air pollution.  
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1 504 **5. References**

2  
3 505 Ambient air quality and cleaner air for Europe. Directive 2008/50/CE. European  
4  
5 506 Parliament and Council, 2008  
6  
7 507 Alfaro-Moreno E, Martinez L, Garcia-Cuellar C, Bonner JC, Murray JC, Rosas I,  
8  
9 508 Rosales SP, Osornio-Vargas AR. 2002. Biologic effects induced in vitro by  
10  
11 509 PM10 from three different zones of Mexico City. Environ Health Perspect  
12  
13 510 110(7):715-20.  
14  
15  
16 511 Aust AE, Ball JC, Hu AA, Lighty JS, Smith KR, Straccia AM, Veranth JM, Young WC.  
17  
18 512 2002. Particle characteristics responsible for effects on human lung epithelial  
19  
20 513 cells. Res Rep Health Eff Inst(110):1-65; discussion 67-76.  
21  
22  
23 514 Baulig A, Sourdeval M, Meyer M, Marano F, Baeza-Squiban A. 2003. Biological effects  
24  
25 515 of atmospheric particles on human bronchial epithelial cells. Comparison with  
26  
27 516 diesel exhaust particles. Toxicol In Vitro 17(5-6):567-73.  
28  
29  
30 517 Becker S, Mundandhara S, Devlin RB, Madden M. 2005. Regulation of cytokine  
31  
32 518 production in human alveolar macrophages and airway epithelial cells in  
33  
34 519 response to ambient air pollution particles: Further mechanistic studies. Toxicol  
35  
36 520 Appl Pharmacol 207(2 Suppl):269-75.  
37  
38  
39 521 Becker S, Soukup JM, Sioutas C, Cassee FR. 2003. Response of human alveolar  
40  
41 522 macrophages to ultrafine, fine, and coarse urban air pollution particles. Exp  
42  
43 523 Lung Res 29(1):29-44.  
44  
45  
46 524 Billet S, Abbas I, Goff JL, Verdin A, Andre V, Lafargue PE, Hachimi A, Cazier F, Sichel  
47  
48 525 F, Shirali P and others. 2008. Genotoxic potential of Polycyclic Aromatic  
49  
50 526 Hydrocarbons-coated onto airborne Particulate Matter (PM(2.5)) in human lung  
51  
52 527 epithelial A549 cells. Cancer Lett.  
53  
54  
55 528 Billet S, Garcon G, Dagher Z, Verdin A, Ledoux F, Cazier F, Courcot D, Aboukais A,  
56  
57 529 Shirali P. 2007. Ambient particulate matter (PM(2.5)): Physicochemical  
58  
59  
60  
61  
62  
63  
64  
65

1 530 characterization and metabolic activation of the organic fraction in human lung  
2  
3 531 epithelial cells (A549). *Environ Res* 105(2):212-223.  
4  
5 532 Brumby, P.E.; Massey, V.; 1967. Determination of nonheme iron, total iron, and  
6  
7 533 copper. In: Estabrook, R.W.; Pullman, M.E.; eds. *Methods in Enzymology*, vol.  
8  
9 534 10. New York: AcademicPress;. p. 463–74.  
10  
11 535 Calcabrini A, Meschini S, Marra M, Falzano L, Colone M, De Berardis B, Paoletti L,  
12  
13 536 Arancia G, Fiorentini C. 2004. Fine environmental particulate engenders  
14  
15 537 alterations in human lung epithelial A549 cells. *Environ Res* 95(1):82-91.  
16  
17 538 Carter JD, Ghio AJ, Samet JM, Devlin RB. 1997. Cytokine production by human airway  
18  
19 539 epithelial cells after exposure to an air pollution particle is metal-dependent.  
20  
21 540 *Toxicol Appl Pharmacol* 146(2):180-8.  
22  
23 541 CEN. Air quality - determination of the PM10 fraction of suspended particulate matter –  
24  
25 542 reference method and field test procedure to demonstrate reference  
26  
27 543 equivalence of measurement methods. Brussels: European Committee for  
28  
29 544 Standardization; (European Standard EN 12341) 1998.  
30  
31 545 Churg A, Brauer M. 2000. Ambient atmospheric particles in the airways of human  
32  
33 546 lungs. *Ultrastruct Pathol* 24(6):353-61.  
34  
35 547 Claxton LD, Matthews PP, Warren SH. 2004. The genotoxicity of ambient outdoor air, a  
36  
37 548 review: *Salmonella* mutagenicity. *Mutat Res* 567(2-3):347-99.  
38  
39 549 de Kok TM, Drieece HA, Hogervorst JG, Briede JJ. 2006. Toxicological assessment of  
40  
41 550 ambient and traffic-related particulate matter: a review of recent studies. *Mutat*  
42  
43 551 *Res* 613(2-3):103-22.  
44  
45 552 Donaldson K, Stone V, Borm PJ, Jimenez LA, Gilmour PS, Schins RP, Knaapen AM,  
46  
47 553 Rahman I, Faux SP, Brown DM and others. 2003. Oxidative stress and calcium  
48  
49 554 signaling in the adverse effects of environmental particles (PM10). *Free Radic*  
50  
51 555 *Biol Med* 34(11):1369-82.  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 556 Englert N. 2004. Fine particles and human health--a review of epidemiological studies.  
2  
3 557 Toxicol Lett 149(1-3):235-42.  
4  
5 558 Frampton MW, Ghio AJ, Samet JM, Carson JL, Carter JD, Devlin RB. 1999. Effects of  
6  
7 559 aqueous extracts of PM(10) filters from the Utah valley on human airway  
8  
9 560 epithelial cells. Am J Physiol 277(5 Pt 1):L960-7.  
10  
11 561 Fujii T, Hayashi S, Hogg JC, Vincent R, Van Eeden SF. 2001. Particulate matter  
12  
13 562 induces cytokine expression in human bronchial epithelial cells. Am J Respir  
14  
15 563 Cell Mol Biol 25(3):265-71.  
16  
17 564 Garcon G, Dagher Z, Zerimech F, Ledoux F, Courcot D, Aboukais A, Puskaric E,  
18  
19 565 Shirali P. 2006. Dunkerque City air pollution particulate matter-induced  
20  
21 566 cytotoxicity, oxidative stress and inflammation in human epithelial lung cells  
22  
23 567 (L132) in culture. Toxicol In Vitro 20(4):519-28.  
24  
25 568 Geng H, Meng Z, Zhang Q. 2006. In vitro responses of rat alveolar macrophages to  
26  
27 569 particle suspensions and water-soluble components of dust storm PM(2.5).  
28  
29 570 Toxicol In Vitro 20(5):575-84.  
30  
31 571 Ghio AJ, Silbajoris R, Carson JL, Samet JM. 2002. Biologic effects of oil fly ash.  
32  
33 572 Environ Health Perspect 110 Suppl 1:89-94.  
34  
35 573 Gilli G, Pignata C, Schiliro T, Bono R, La Rosa A, Traversi D. 2007a. The mutagenic  
36  
37 574 hazards of environmental PM2.5 in Turin. Environ Res 103(2):168-75.  
38  
39 575 Gilli G, Traversi D, Rovere R, Pignata C, Schiliro T. 2007b. Chemical characteristics  
40  
41 576 and mutagenic activity of PM10 in Torino, a northern Italian city. Sci Total  
42  
43 577 Environ 385(1-3):97-107.  
44  
45 578 Golladay SA, Park SH, Aust AE. 1997. Efflux of reduced glutathione after exposure of  
46  
47 579 human lung epithelial cells to crocidolite asbestos. Environ Health Perspect 105  
48  
49 580 Suppl 5:1273-7.  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 581 Gualtieri M, Mantecca P, Cetta F, Camatini M. 2008. Organic compounds in tire particle  
2  
3 582 induce reactive oxygen species and heat-shock proteins in the human alveolar  
4  
5 583 cell line A549. *Environ Int* 34(4):437-42.  
6  
7 584 Happonen MS, Hirvonen MR, Halinen AI, Jalava PI, Pennanen AS, Sillanpaa M, Hillamo R,  
8  
9 585 Salonen RO. 2008. Chemical compositions responsible for inflammation and  
10  
11 586 tissue damage in the mouse lung by coarse and fine particulate samples from  
12  
13 587 contrasting air pollution in Europe. *Inhal Toxicol* 20(14):1215-31.  
14  
15  
16 588 Hatch G. 1992. Comparative biochemistry of airway lining fluid. In: Parent RA, editor.  
17  
18 589 Comparative Biology of the Normal Lung. Boca Raton, FL: CRC Press. p 617-  
19  
20 590 632.  
21  
22  
23 591 Hazenkamp-Von Arx ME, Gotschi T, Ackermann-Liebrich U, Bono R, Burney P, Cyrus  
24  
25 592 J, Jarvis D, Lillienberg L, Luczynska C, Maldonado JA and others. 2004. PM2.5  
26  
27 593 and NO2 assessment in 21 European study centres of ECRHS II: annual  
28  
29 594 means and seasonal differences. *Atmospheric Environment* 38(13):1943-1953.  
30  
31  
32 595 Heinrich J, Pitz M, Bischof W, Krug N, Borm PJA. 2003. Endotoxin in fine (PM2.5) and  
33  
34 596 coarse (PM2.5-10) particle mass of ambient aerosols. A temporo-spatial  
35  
36 597 analysis. *Epidemiology* 14(5):S62-S62.  
37  
38  
39 598 Hetland RB, Cassee FR, Refsnes M, Schwarze PE, Lag M, Boere AJ, Dybing E. 2004.  
40  
41 599 Release of inflammatory cytokines, cell toxicity and apoptosis in epithelial lung  
42  
43 600 cells after exposure to ambient air particles of different size fractions. *Toxicol In*  
44  
45 601 *Vitro* 18(2):203-12.  
46  
47  
48 602 Hetland RB, Refsnes M, Myran T, Johansen BV, Uthus N, Schwarze PE. 2000. Mineral  
49  
50 603 and/or metal content as critical determinants of particle-induced release of IL-6  
51  
52 604 and IL-8 from A549 cells. *J Toxicol Environ Health A* 60(1):47-65.  
53  
54  
55 605 Huang SL, Cheng WL, Lee CT, Huang HC, Chan CC. 2002. Contribution of endotoxin  
56  
57 606 in macrophage cytokine response to ambient particles in vitro. *J Toxicol Environ*  
58  
59 607 *Health A* 65(17):1261-72.  
60  
61  
62  
63  
64  
65

1 608 Hutchison GR, Brown DM, Hibbs LR, Heal MR, Donaldson K, Maynard RL, Monaghan  
2  
3 609 M, Nicholl A, Stone V. 2005. The effect of refurbishing a UK steel plant on  
4  
5 610 PM10 metal composition and ability to induce inflammation. *Respir Res* 6:43.  
6  
7 611 Jalava PI, Salonen RO, Halinen AI, Penttinen P, Pennanen AS, Sillanpaa M, Sandell E,  
8  
9 612 Hillamo R, Hirvonen MR. 2006. In vitro inflammatory and cytotoxic effects of  
10  
11 613 size-segregated particulate samples collected during long-range transport of  
12  
13 614 wildfire smoke to Helsinki. *Toxicol Appl Pharmacol* 215(3):341-53.  
14  
15 615 Katsouyanni K, Touloumi G, Samoli E, Gryparis A, Le Tertre A, Monopolis Y, Rossi G,  
16  
17 616 Zmirou D, Ballester F, Boumghar A and others. 2001. Confounding and effect  
18  
19 617 modification in the short-term effects of ambient particles on total mortality:  
20  
21 618 results from 29 European cities within the APHEA2 project. *Epidemiology*  
22  
23 619 12(5):521-31.  
24  
25 620 Kinnula VL, Aalto K, Raivio KO, Walles S, Linnainmaa K. 1994. Cytotoxicity of oxidants  
26  
27 621 and asbestos fibers in cultured human mesothelial cells. *Free Radic Biol Med*  
28  
29 622 16(2):169-76.  
30  
31 623 Knaapen AM, Borm PJ, Albrecht C, Schins RP. 2004. Inhaled particles and lung  
32  
33 624 cancer. Part A: Mechanisms. *Int J Cancer* 109(6):799-809.  
34  
35 625 Krewski D, Rainham D. 2007. Ambient air pollution and population health: overview. *J*  
36  
37 626 *Toxicol Environ Health A* 70(3-4):275-83.  
38  
39 627 Kueng W, Silber E, Eppenberger U. 1989. Quantification of cells cultured on 96-well  
40  
41 628 plates. *Anal Biochem* 182(1):16-9.  
42  
43 629 Li N, Hao M, Phalen RF, Hinds WC, Nel AE. 2003. Particulate air pollutants and  
44  
45 630 asthma. A paradigm for the role of oxidative stress in PM-induced adverse  
46  
47 631 health effects. *Clin Immunol* 109(3):250-65.  
48  
49 632 Lobner D. 2000. Comparison of the LDH and MTT assays for quantifying cell death:  
50  
51 633 validity for neuronal apoptosis? *J Neurosci Methods* 96(2):147-52.  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 634 Lund LG, Aust AE. 1990. Iron mobilization from asbestos by chelators and ascorbic  
2  
3 635 acid. Arch Biochem Biophys 278(1):61-4.  
4  
5 636 Marcazzan GM, Ceriani M, Valli G, Vecchi R. 2003. Source apportionment of PM10  
6  
7 637 and PM2.5 in Milan (Italy) using receptor modelling. Sci Total Environ 317(1-  
8  
9 638 3):137-47.  
10  
11 639 Molinelli AR, Madden MC, McGee JK, Stonehuerner JG, Ghio AJ. 2002. Effect of metal  
12  
13 640 removal on the toxicity of airborne particulate matter from the Utah Valley. Inhal  
14  
15 641 Toxicol 14(10):1069-86.  
16  
17 642 Moller P, Folkmann JK, Forchhammer L, Brauner EV, Danielsen PH, Risom L, Loft S.  
18  
19 643 2008. Air pollution, oxidative damage to DNA, and carcinogenesis. Cancer Lett  
20  
21 644 266(1):84-97.  
22  
23 645 Monn C, Becker S. 1999. Cytotoxicity and induction of proinflammatory cytokines from  
24  
25 646 human monocytes exposed to fine (PM2.5) and coarse particles (PM10-2.5) in  
26  
27 647 outdoor and indoor air. Toxicol Appl Pharmacol 155(3):245-52.  
28  
29 648 Morgenstern V, Carty CL, Gehring U, Cyrus J, Bischof W, Heinrich J. 2005. Lack of  
30  
31 649 spatial variation of endotoxin in ambient particulate matter across a German  
32  
33 650 metropolitan area. Atmospheric Environment 39(36):6931-6941.  
34  
35 651 Mueller-Anneling L, Avol E, Peters JM, Thorne PS. 2004. Ambient endotoxin  
36  
37 652 concentrations in PM10 from Southern California. Environ Health Perspect  
38  
39 653 112(5):583-8.  
40  
41 654 Muller A, Wichmann G, Massolo L, Rehwagen M, Grabsch C, Loffhagen N, Herbarth  
42  
43 655 O, Ronco A. 2006. Cytotoxicity and oxidative stress caused by chemicals  
44  
45 656 adsorbed on particulate matter. Environ Toxicol 21(5):457-63.  
46  
47 657 Ning Y, Imrich A, Goldsmith CA, Qin G, Kobzik L. 2000. Alveolar macrophage cytokine  
48  
49 658 production in response to air particles in vitro: role of endotoxin. J Toxicol  
50  
51 659 Environ Health A 59(3):165-80.  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 660 Oberdorster G, Finkelstein JN, Johnston C, Gelein R, Cox C, Baggs R, Elder AC. 2000.  
2  
3 661 Acute pulmonary effects of ultrafine particles in rats and mice. *Res Rep Health*  
4  
5 662 *Eff Inst(96):5-74; disc 75-86.*  
6  
7 663 Oberdorster G, Yu CP. 1999. Lung dosimetry--considerations for noninhalation studies.  
8  
9 664 *Exp Lung Res 25(1):1-6.*  
10  
11 665 Osornio-Vargas AR, Bonner JC, Alfaro-Moreno E, Martinez L, Garcia-Cuellar C,  
12  
13 666 Ponce-de-Leon Rosales S, Miranda J, Rosas I. 2003. Proinflammatory and  
14  
15 667 cytotoxic effects of Mexico City air pollution particulate matter in vitro are  
16  
17 668 dependent on particle size and composition. *Environ Health Perspect*  
18  
19 669 *111(10):1289-93.*  
20  
21 670 Pope CA, 3rd. 2007. Mortality effects of longer term exposures to fine particulate air  
22  
23 671 pollution: review of recent epidemiological evidence. *Inhal Toxicol 19 Suppl*  
24  
25 672 *1:33-8.*  
26  
27 673 Pope CA, 3rd, Dockery DW. 2006. Health effects of fine particulate air pollution: lines  
28  
29 674 that connect. *J Air Waste Manag Assoc 56(6):709-42.*  
30  
31 675 Riganti C, Aldieri E, Bergandi L, Fenoglio I, Costamagna C, Fubini B, Bosia A, Ghigo  
32  
33 676 D. 2002. Crocidolite asbestos inhibits pentose phosphate oxidative pathway  
34  
35 677 and glucose 6-phosphate dehydrogenase activity in human lung epithelial cells.  
36  
37 678 *Free Radic Biol Med 32(9):938-49.*  
38  
39 679 Scapellato ML, Lotti M. 2007. Short-term effects of particulate matter: an inflammatory  
40  
41 680 mechanism? *Crit Rev Toxicol 37(6):461-87.*  
42  
43 681 Schins RP, Lightbody JH, Borm PJ, Shi T, Donaldson K, Stone V. 2004. Inflammatory  
44  
45 682 effects of coarse and fine particulate matter in relation to chemical and  
46  
47 683 biological constituents. *Toxicol Appl Pharmacol 195(1):1-11.*  
48  
49 684 Schwarze PE, Ovreivik J, Lag M, Refsnes M, Nafstad P, Hetland RB, Dybing E. 2006.  
50  
51 685 Particulate matter properties and health effects: consistency of epidemiological  
52  
53 686 and toxicological studies. *Hum Exp Toxicol 25(10):559-79.*  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 687 Seagrave JC, Nikula KJ. 2000. Multiple modes of responses to air pollution particulate  
2  
3 688 materials in A549 alveolar type II cells. *Inhal Toxicol* 12 Suppl 4:247-60.  
4  
5 689 Sevanian A, Ursini F. 2000. Lipid peroxidation in membranes and low-density  
6  
7 690 lipoproteins: similarities and differences. *Free Radic Biol Med* 29(3-4):306-11.  
8  
9  
10 691 Shi T, Duffin R, Borm PJ, Li H, Weishaupt C, Schins RP. 2006. Hydroxyl-radical-  
11  
12 692 dependent DNA damage by ambient particulate matter from contrasting  
13  
14 693 sampling locations. *Environ Res* 101(1):18-24.  
15  
16 694 Smith KR, Aust AE. 1997. Mobilization of iron from urban particulates leads to  
17  
18 695 generation of reactive oxygen species in vitro and induction of ferritin synthesis  
19  
20 696 in human lung epithelial cells. *Chem Res Toxicol* 10(7):828-34.  
21  
22  
23 697 Solomon GM, Hjelmroos-Koski M, Rotkin-Ellman M, Hammond SK. 2006. Airborne  
24  
25 698 mold and endotoxin concentrations in New Orleans, Louisiana, after flooding,  
26  
27 699 October through November 2005. *Environmental Health Perspectives*  
28  
29 700 114(9):1381-1386.  
30  
31  
32 701 Sorensen M, Autrup H, Moller P, Hertel O, Jensen SS, Vinzents P, Knudsen LE, Loft S.  
33  
34 702 2003. Linking exposure to environmental pollutants with biological effects. *Mutat*  
35  
36 703 *Res* 544(2-3):255-71.  
37  
38  
39 704 Soukup JM, Becker S. 2001. Human alveolar macrophage responses to air pollution  
40  
41 705 particulates are associated with insoluble components of coarse material,  
42  
43 706 including particulate endotoxin. *Toxicol Appl Pharmacol* 171(1):20-6.  
44  
45 707 U.S. EPA. Air quality criteria for particulate matter. vol 1-3. EPA/600/P-95/001a. 1996.  
46  
47  
48 708 Veronesi B, de Haar C, Lee L, Oortgiesen M. 2002. The surface charge of visible  
49  
50 709 particulate matter predicts biological activation in human bronchial epithelial  
51  
52 710 cells. *Toxicol Appl Pharmacol* 178(3):144-54.  
53  
54 711 World Health Organization Europe. 2006. Air Quality Guidelines — Global Update  
55  
56 712 2005.  
57  
58  
59  
60  
61  
62  
63  
64  
65



1 713 World Health Organization Regional Office for Europe, 2007. Health relevance of  
2  
3 714 particulate matter from various sources. Report on a WHO Workshop, Bonn,  
4  
5 715 Germany, 26-27 March 2007.

6  
7 716 Ziemacki G, Cattani G, Cusano MC, Stacchini G, Marconi A. 2003. [Occurrence of  
8  
9 717 metals in various size fractions of particulate matter]. Ann Ist Super Sanita  
10  
11 718 39(3):371-9.

12  
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14 719

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16 720 **Figure Captions**

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21 722 **Figure 1.** Proliferation inhibition of A549 cells after their incubation in the continuous  
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23 723 presence of increasing concentrations of particulate matter (50 and 100 µg/mL) from  
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25 724 PM10 organic solvent extracts (a) and PM10 aqueous extracts (b). Inhibition of cell  
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27 725 proliferation was calculated comparing the absorbance of exposed cultures with the  
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29 726 absorbance of non-exposed cultures. Asterisks indicate statistically significant  
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31 727 differences from the control ,  $p < 0.05$  (T-test).

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36 729 **Figure 2.** IL-6 release by A549 cells after their incubation in the continuous presence of  
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38 730 increasing concentrations of particulate matter (50 and 100 µg/mL) from PM10 organic  
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40 731 solvent extracts (a) and PM10 aqueous extracts (b). Results were calculated by  
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42 732 expressing samples IL-6 relies as a percentage of IL-6 release from non exposed cells.  
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44 733 Asterisks indicate statistically significant differences from the control ,  $p < 0.05$  (T-test).

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50 735 **Figure 3.** Different seasonal (autumn/winter vs spring/summer) proliferation inhibitions  
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52 736 3(a), IL-6 releases 3(b) and LDH releases 3(c) from A549 cells at 72h after their  
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54 737 incubation in the continuous presence of PM10 organic solvent extracts and PM10  
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56 738 aqueous extracts (50, 100 µg/mL or 200 µg/mL). Asterisks indicates statistically  
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58 739 significant differences for the two seasons,  $p < 0.05$  (T-test).



*Environmental Toxicology and Pharmacology*

Conflict of Interest Policy

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**Declarations**

*Environmental Toxicology and Pharmacology* requires that all authors sign a declaration of conflicting interests. If you have nothing to declare in any of these categories then this should be stated.

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**Signature** (a scanned signature is acceptable, but each author must sign)

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**Table 1.** Means and standard deviations of PM10, sulfates and nitrates concentrations ( $\mu\text{g}/\text{m}^3$ ) of the whole sampling period (2005/2006) and divided by seasons.

|               | <b>PM10</b><br>( $\mu\text{g}/\text{m}^3$ ) | <b>SO<sub>4</sub><sup>=</sup></b><br>( $\mu\text{g}/\text{m}^3$ ) | <b>NO<sub>3</sub><sup>-</sup></b><br>( $\mu\text{g}/\text{m}^3$ ) | <b>SO<sub>4</sub><sup>=</sup></b><br>(% of PM10<br>mass) | <b>NO<sub>3</sub><sup>-</sup></b><br>(% of PM10<br>mass) |
|---------------|---|---|---|--|--|
| Whole period  | 55.4 ± 39.1                                 | 10.5 ± 5.7  | 11.7 ± 9.2  | 21.2 ± 6.9   | 20.6 ± 6.1   |
| Winter/Autumn | 71.3 ± 43.1**                               | 12.4 ± 6.4*   | 14.9 ± 10.1**   | 19.2 ± 7.3*  | 20.8 ± 6.0   |
| Summer/Spring | 37.4 ± 24.1**                               | 8.3 ± 5.7*  | 8.0 ± 6.3**   | 23.6 ± 5.6*  | 20.3 ± 6.3   |

\*\* statistically significant differences, (autumn/winter vs spring/summer)  $p < 0.01$  (T-test).

\* statistically significant differences, (autumn/winter vs spring/summer)  $p < 0.05$  (T-test).

**Table 2.** Means and standard deviations of Fe concentrations ( $\mu\text{g}/\text{m}^3$ ) of the whole sampling period (2005/2006) and divided by seasons.

|                       | <b>Fe total</b><br><b>(<math>\mu\text{g}/\text{m}^3</math>)</b> | <b>Fe bio</b><br><b>(<math>\mu\text{g}/\text{m}^3</math>)</b> | <b>% Fe bio</b><br><b>(of total)</b> |
|-----------------------|---|---|--------------------------------------|
| Whole sampling period | 1.035 $\pm$ 0.811   | 0.078 $\pm$ 0.095   | 6.9 $\pm$ 4.5                        |
| Winter/Autumn         | 1.355 $\pm$ 0.919**   | 0.097 $\pm$ 0.089   | 7.5 $\pm$ 4.8                        |
| Summer/Spring         | 0.672 $\pm$ 0.463**   | 0.056 $\pm$ 0.099   | 6.2 $\pm$ 4.1                        |

\*\* statistically significant differences, (autumn/winter vs spring/summer)  $p < 0.01$  (T-test).

**Table 3.** Released lactate dehydrogenase (LDH) activity from A549 cell 24h, 48h and 72h after their incubation in the continuous presence of increasing concentrations of particle matter (100 and 200 µg/mL) from PM10 organic solvent extracts and from PM10 aqueous extracts. Results were calculated by expressing samples LDH activity as a percentage of LDH activity from non exposed cells.

| LDH increase %                |           | 24 h         | 48 h         | 72 h         |
|-------------------------------|-----------|--------------|--------------|--------------|
| PM10 organic solvent extracts | 100 µg/mL | -2.7 ± 7.9   | 3.9 ± 7.1    | 64.0 ± 22.5* |
|                               | 200 µg/mL | -7.3 ± 3.1   | 2.9 ± 5.5    | 83.7 ± 44.1* |
| PM10 aqueous extracts         | 100 µg/mL | 30.2 ± 24.3  | 33.5 ± 23.0* | 35.6 ± 18.3* |
|                               | 200 µg/mL | 78.1 ± 34.1* | 87.6 ± 41.9* | 53.6 ± 24.6* |

\* statistically significant differences in relation to the control,  $p < 0.05$  (T-test).

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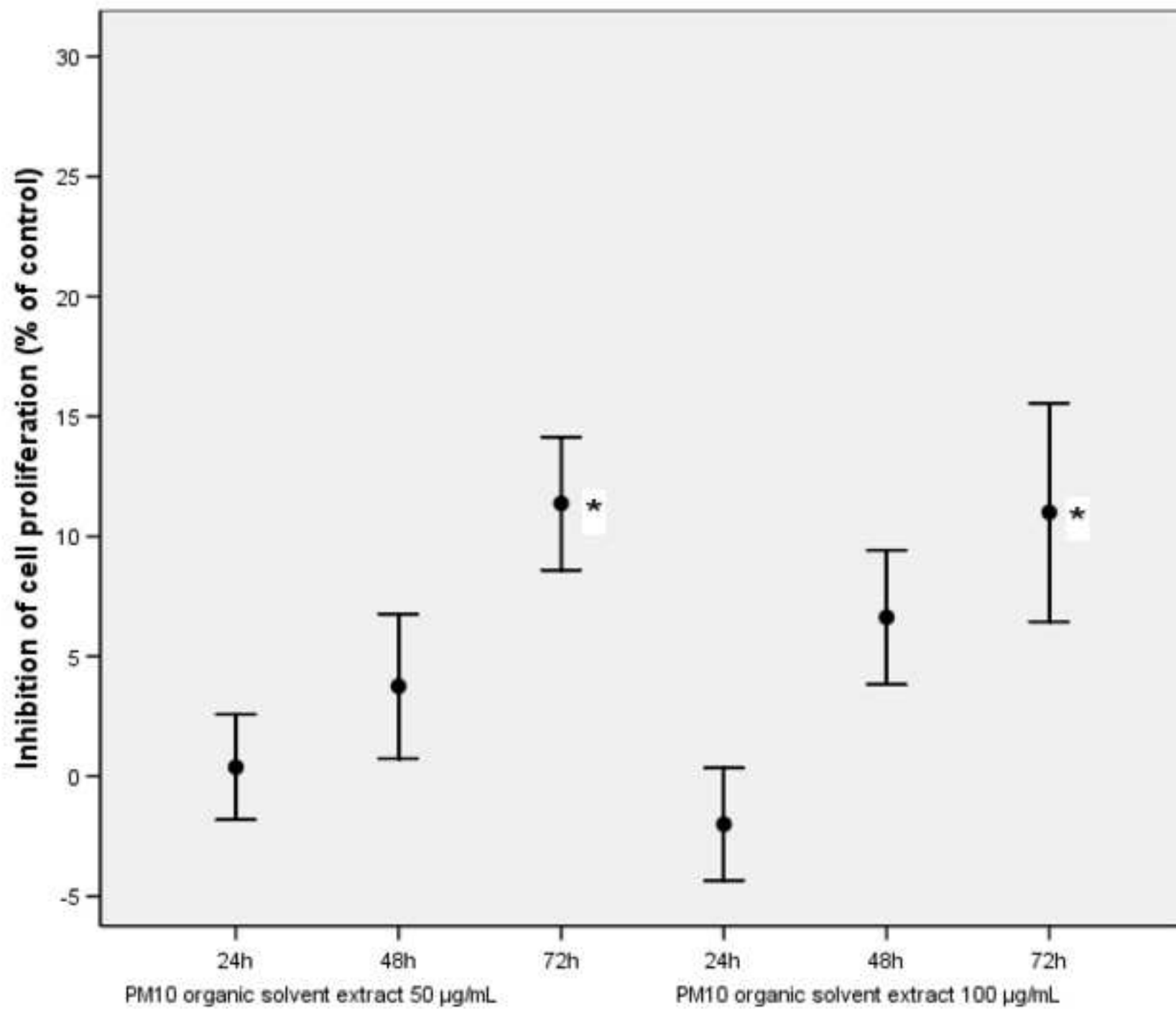


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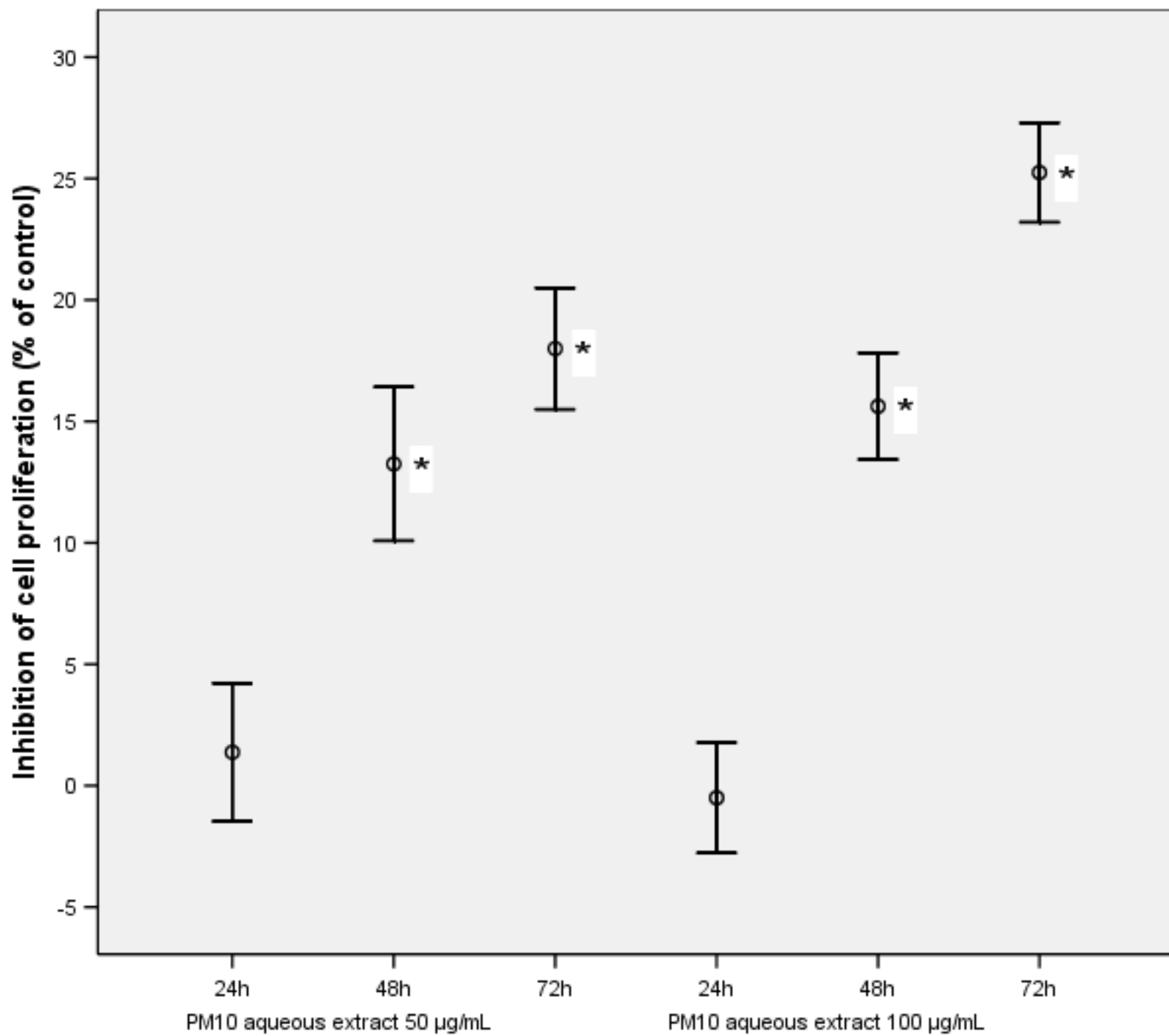


Figure 2a

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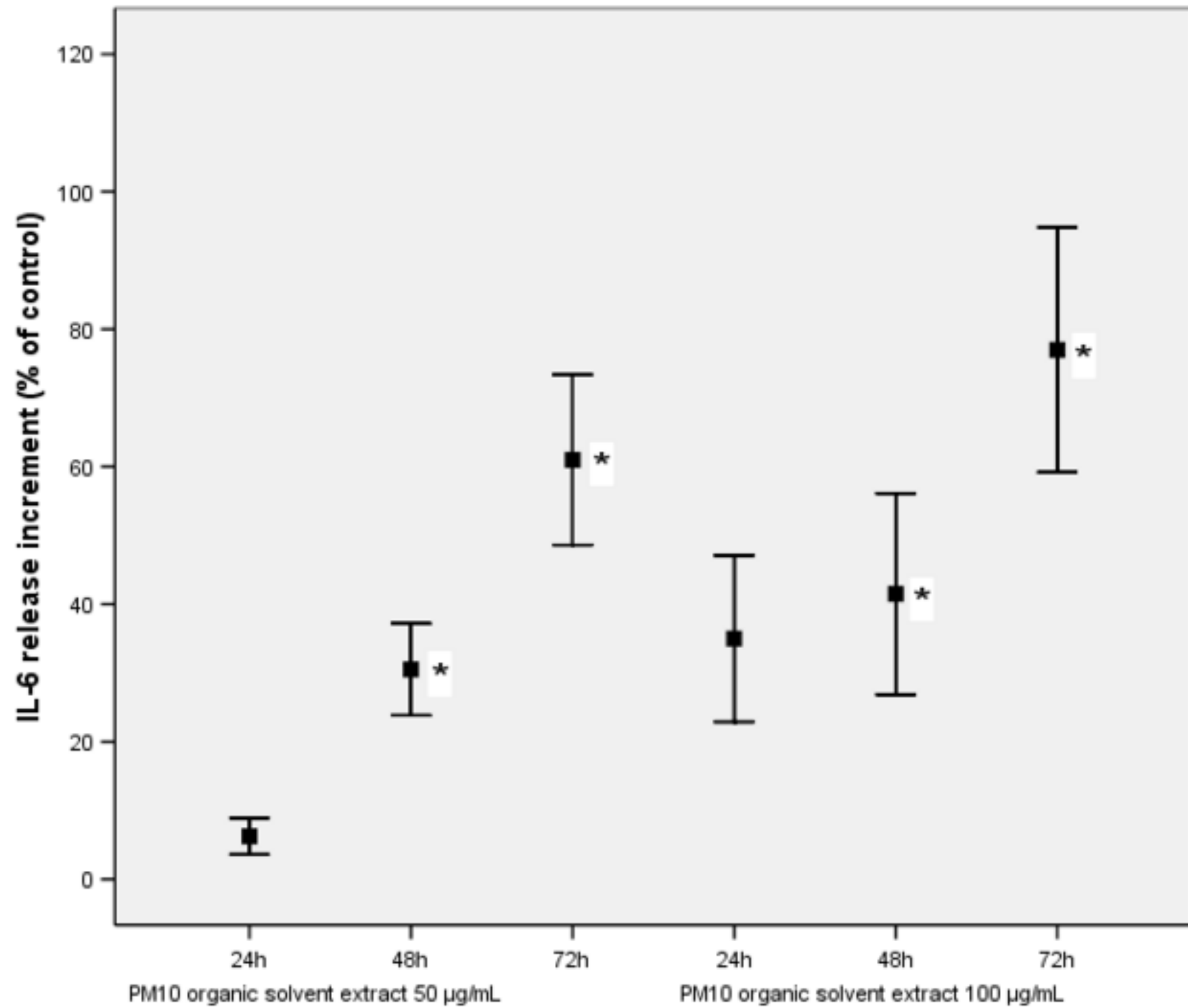




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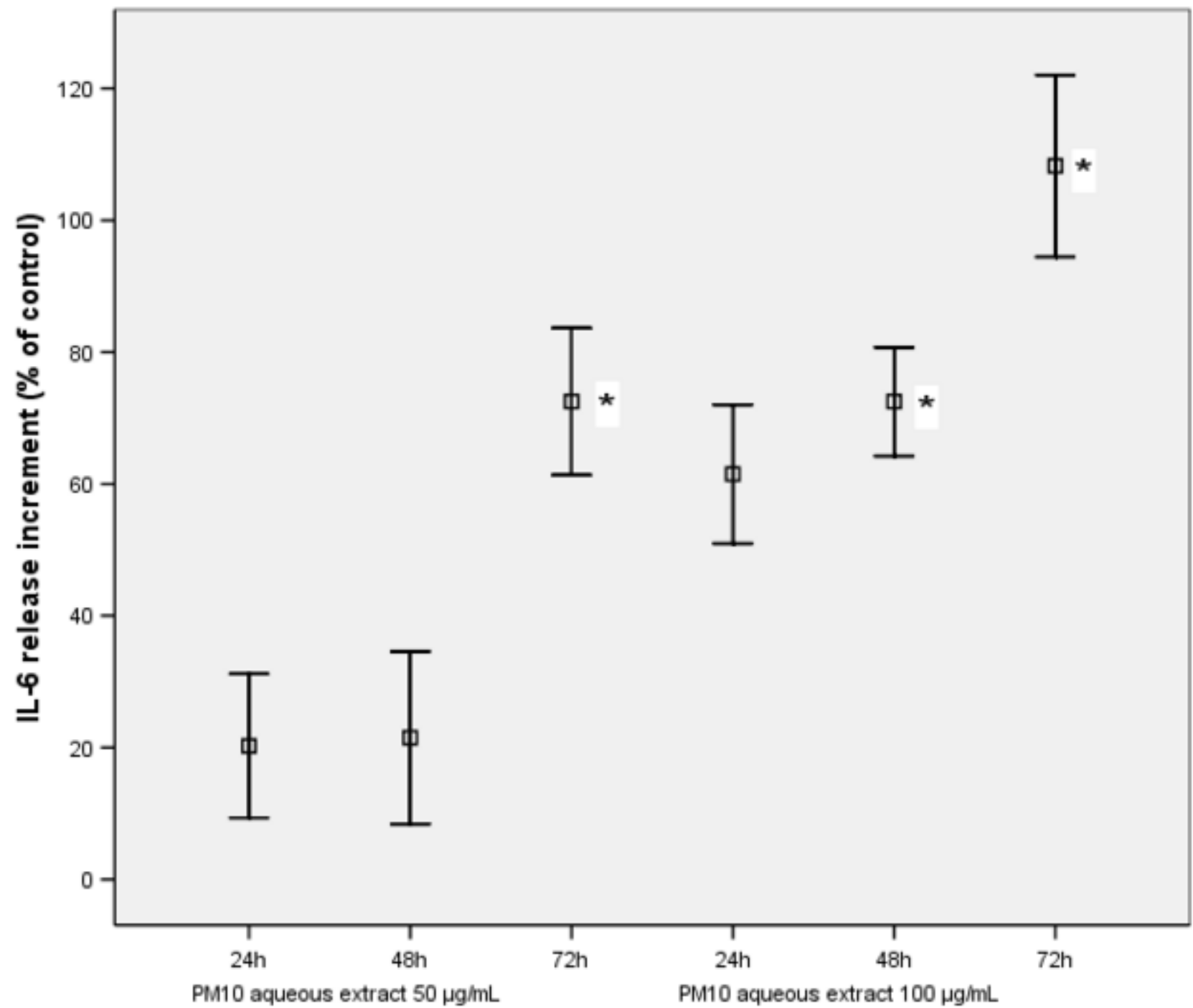


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