

# Genome sequence of potential psychobiotic *Lacticaseibacillus rhamnosus* A20T1A using long-read ONT technology

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**ABSTRACT** We present the genome sequence of *Lacticaseibacillus rhamnosus* isolated from human gut using Oxford Nanopore technology sequencing. The annotated draft genome of *L. rhamnosus* A20T1A showed the presence of probiotics and psychobiotics genomic traits relevant for gut colonization and human health.

**KEYWORDS** Oxford Nanopore technology, probiotics, psychobiotics

In the last years, lactic acid bacteria (LAB) have been increasingly reported to produce neuroactive molecules, able to modulate mood and cognition in humans (psychobiotics) (1). The strain was isolated from the human gut of healthy adult volunteers in Torino, Italy (Ethics statement Prot. no. 0676002) on da De Man, Rogosa e Sharpe agar (MRS, Neogen, USA; NCM0190A) incubated at 30°C for 48 h. Strain was purified on MRS broth under the same isolation condition and identified by Matrix-Assisted Laser Desorption Ionization-Time of Flight mass spectrometry (2). About 1 mL of the culture (on MRS broth at 30°C for 48 h) was then used for DNA extraction by the MasterPure Complete DNA and RNA Purification Kit (Biosearch Technologies). The strain was identified as *Lacticaseibacillus rhamnosus* sequencing was performed using the Rapid Sequencing Kit (SQK-RAD004, Oxford Nanopore, UK) and sequenced on a MinION Nanopore device R9.4.1 (FLO-MIN106, Oxford Nanopore, UK) according to the manufacturer instructions without DNA fragmenting, ligation, and size selection.

Default parameters were used for all software unless otherwise specified. 3.77 Gb of raw reads with a read length range from 1.38 to 26.88 kbp (average read length 3,360.92 bp) were subjected to base calling and low-quality base filtering with Guppy v1.6.10 (3) and assembled using Flye v2.9.2 (--asm-coverage 50) (4). Overlap of the initial and final genome sequences was checked with BLAST+2.15.0 (5) showing one single circularized genome of 2.92 Mbp with a G + C content of 46.83%, genome coverage 100%. Annotation was performed by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP v4.9) (6). The genome was then analyzed using *Tormes* v1.3.0 (7) that includes antibiotic resistance genes tools (ARG-ANNOT v.6, CARD v.2020, and ResFinder v.4) (8–10). The amino acidic sequences were analysed with e-mapper v2.1.12 (11) and Dfast v1.2.0 (12).

Genes annotation allowed the assignment of functional categories to 2,762 CDS, and the detection of 15 rRNA, 59 tRNA, and 2 CRISPR sequences. The most abundant genes were correlated to carbohydrate transport and metabolism, replication, recombination, and repair genes and amino acid transport and metabolism.

Part of the genes were putatively related to probiotic traits, that is, genes related to pilus structure (*spaE*, *spaD*, and *spaF*) and adhesion (*fbpA*). Beyond some antimicrobial peptides (AMP) gene related, that is, *ydel* correlated gene (OmpD-associated protein), which confer AMP resistance via an unknown mechanism (13), we identified ABC transporters associated to bacteriocin export (14), and CAAX protease involved in bacteriocin self-immunity (15). Psychobiotics traits have been detected

**Editor** David Rasko, University of Maryland School of Medicine, Baltimore, Maryland, USA

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The authors declare no conflict of interest.

**Received** 22 June 2024

**Accepted** 4 November 2024

**Published** 26 November 2024

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using BLASTn ( $-e$  value  $1e-05$ ) against psychobiotics genes selected from literature that identified correlations between these genes and probiotic and psychobiotic traits such as: tryptophan synthase (16), propionate synthase via fumarate hydratase (17, 18); indolelactate dehydrogenase (19), linoleate isomerase (20), genes involved in taurine ABC transport system (21), glutamate/gamma-amino butyrate antiporter (22) and genes involved in propionate synthase via malate dehydrogenase (18). This genome annotation allowed a better understanding of the psychobiotic traits of this strain.

### Nucleotide sequence accession number

*L. rhamnosus* A20T1A raw reads and assembly have been deposited to NCBI under accession numbers [SRX22451036](https://www.ncbi.nlm.nih.gov/nuclink/SRX22451036) and [CP151435](https://www.ncbi.nlm.nih.gov/nuclink/CP151435), respectively. FASTA file of psychobiotics genes has been deposited on ZENODO (<https://doi.org/10.5281/zenodo.13923846>).

### ACKNOWLEDGMENTS

This study was supported by the Italian Ministry of University and Research PRIN-PNRR 2022 program (exploiting autochthonous microbial resources from traditional Italian fermented foods for gut-brain axis modulation (HOLOGRAM) Prot. P2022X8A9M).

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Irene Franciosa, Formal analysis, Investigation, Methodology, Writing – original draft | Davide Buzzanca, Formal analysis, Investigation, Methodology, Software, Writing – original draft | Luca Cocolin, Supervision, Writing – review and editing | Francesca De Filippis, Funding acquisition, Supervision, Writing – review and editing | Ilario Ferrocino, Conceptualization, Funding acquisition, Supervision, Writing – review and editing

### REFERENCES

- Del Toro-Barbosa M, Hurtado-Romero A, Garcia-Amezquita LE, Garcia-Cayuela T. 2020. Psychobiotics: mechanisms of action, evaluation methods and effectiveness in applications with food products. *Nutrients* 12:1–31. <https://doi.org/10.3390/nu12123896>
- Gantzias C, Lappa IK, Aerts M, Georgalaki M, Manolopoulou E, Papadimitriou K, De Brandt E, Tsakalidou E, Vandamme P. 2020. MALDI-TOF MS profiling of non-starter lactic acid bacteria from artisanal cheeses of the Greek island of Naxos. *Int J Food Microbiol* 323:108586. <https://doi.org/10.1016/j.ijfoodmicro.2020.108586>
- Wick RR, Judd LM, Holt KE. 2019. Performance of neural network basecalling tools for Oxford Nanopore sequencing. *Genome Biol* 20:129. <https://doi.org/10.1186/s13059-019-1727-y>
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. *BLAST+*: architecture and applications. *BMC Bioinformatics* 10:1–9. <https://doi.org/10.1186/1471-2105-10-421>
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>
- Quijada NM, Rodríguez-Lázaro D, Eiros JM, Hernández M. 2019. TORMES: an automated pipeline for whole bacterial genome analysis. *Bioinformatics* 35:4207–4212. <https://doi.org/10.1093/bioinformatics/btz220>
- Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen A-LV, Cheng AA, Liu S, et al. 2020. CARD 2020: antibiotic resistome surveillance with the comprehensive antibiotic

- resistance database. *Nucleic Acids Res* 48:517–525. <https://doi.org/10.1093/nar/gkz935>
9. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>
  10. Gupta SK, Padmanabhan BR, Diene SM, Lopez-Rojas R, Kempf M, Landraud L, Rolain J-M. 2014. ARG-ANNOT, a new bioinformatic tool to discover antibiotic resistance genes in bacterial genomes. *Antimicrob Agents Chemother* 58:212–220. <https://doi.org/10.1128/AAC.01310-13>
  11. Huerta-Cepas J, Szklarczyk D, Heller D, Hernández-Plaza A, Forslund SK, Cook H, Mende DR, Letunic I, Rattei T, Jensen LJ, von Mering C, Bork P. 2019. eggNOG 5.0: a hierarchical, functionally and phylogenetically annotated orthology resource based on 5090 organisms and 2502 viruses. *Nucleic Acids Res* 47:D309–D314. <https://doi.org/10.1093/nar/gky1085>
  12. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. *Bioinformatics* 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>
  13. Pilonieta MC, Erickson KD, Ernst RK, Detweiler CS. 2009. A protein important for antimicrobial peptide resistance, Ydel/OmdA, is in the periplasm and interacts with OmpD/NmpC. *J Bacteriol* 191:7243–7252. <https://doi.org/10.1128/JB.00688-09>
  14. Beis K, Rebuffat S. 2019. Multifaceted ABC transporters associated to microcin and bacteriocin export. *Res Microbiol* 170:399–406. <https://doi.org/10.1016/j.resmic.2019.07.002>
  15. Kjos M, Snipen L, Salehian Z, Nes IF, Diep DB. 2010. The abi proteins and their involvement in bacteriocin self-immunity. *J Bacteriol* 192:2068–2076. <https://doi.org/10.1128/JB.01553-09>
  16. Parolisi S, Montanari C, Borghi E, Cazzorla C, Zuvadelli J, Tosi M, Barone R, Bensi G, Bonfanti C, Dionisi Vici C, Biasucci G, Burlina A, Carbone MT, Verduci E, SIMMESN Working Group for Gut Microbiota in Inborn Errors of Metabolism. 2023. Possible role of tryptophan metabolism along the microbiota-gut-brain axis on cognitive & behavioral aspects in Phenylketonuria. *Pharmacol Res* 197:106952. <https://doi.org/10.1016/j.phrs.2023.106952>
  17. Miri S, Yeo J, Abubaker S, Hammami R. 2023. Neuromicrobiology, an emerging neurometabolic facet of the gut microbiome? *Front Microbiol* 14. <https://doi.org/10.3389/fmicb.2023.1098412>
  18. Silva YP, Bernardi A, Frozza RL. 2020. The role of short-chain fatty acids from gut microbiota in gut-brain communication. *Front Endocrinol (Lausanne)* 11:1–14. <https://doi.org/10.3389/fendo.2020.00025>
  19. Jiang H, Chen C, Gao J. 2022. Extensive summary of the important roles of indole propionic acid, a gut microbial metabolite in host health and disease. *Nutrients* 15. <https://doi.org/10.3390/nu15010151>
  20. Murru E, Carta G, Manca C, Sogos V, Pistis M, Melis M, Banni S. 2020. Conjugated linoleic acid and brain metabolism: a possible anti-neuroinflammatory role mediated by PPAR $\alpha$  activation.. *Front Pharmacol* 11:1–12. <https://doi.org/10.3389/fphar.2020.587140>
  21. Qu F, ElOmari K, Wagner A, De Simone A, Beis K. 2019. Desolvation of the substrate-binding protein TauA dictates ligand specificity for the alkanesulfonate ABC importer *TauABC*. *Biochem J* 476:3649–3660. <https://doi.org/10.1042/BCJ20190779>
  22. Mazzoli R, Pessione E. 2016. The neuro-endocrinological role of microbial glutamate and GABA signaling. *Front Microbiol* 7:1–17. <https://doi.org/10.3389/fmicb.2016.01934>