

Systematic Review

Frequency of Germline and Somatic *BRCA1* and *BRCA2* Mutations in Prostate Cancer: An Updated Systematic Review and Meta-Analysis

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Simple Summary: Our updated systematic review and meta-analysis investigates the frequency of germline and somatic *BRCA1* and *BRCA2* mutations in patients with prostate cancer (PC), with subgroup analysis according to the type of mutation (germline or somatic mutations; mutation of *BRCA1* and/or *BRCA2*) and according to the disease setting (any stage PC or metastatic PC or metastatic castration-resistant PC). As known, *BRCA* testing has recently become standard in clinical practice in prostate cancer because of new available target therapies. However, several open questions remain, in terms of the best time to perform it, the genes to look for (*BRCA* only or genes related to the DNA repair pathway of homologous recombination as well), and the optimal molecular analysis technique (somatic and/or germline testing or, in the future, liquid biopsy, which interestingly could assess both somatic and germline mutations simultaneously).

Abstract: In prostate cancer (PC), the presence of *BRCA* somatic and/or germline mutation provides prognostic and predictive information. Meta-analysis aims to estimate the frequency of *BRCA* mutations in patients with PC (PCp). In November 2022, we reviewed literature searching for all articles testing the proportion of *BRCA* mutations in PCp, without explicit enrichment for familial risk. The frequency of germline and somatic *BRCA1* and/or *BRCA2* mutations was described in three stage disease populations (any/metastatic/metastatic castration-resistant PC, mCRPC). Out of 2253 identified articles, 40 were eligible. Here, 0.73% and 1.20% of any stage PCp, 0.94% and 1.10% of metastatic PCp, and 1.21% and 1.10% of mCRPC patients carried germline and somatic *BRCA1* mutation, respectively; 3.25% and 6.29% of any stage PCp, 4.51% and 10.26% of metastatic PCp, and 3.90% and 10.52% of mCRPC patients carried germline and somatic *BRCA2* mutation, respectively; and 4.47% and 7.18% of any stage PCp, 5.84% and 10.94% of metastatic PCp, and 5.26% and 11.26% of mCRPC patients carried germline and somatic *BRCA1/2* mutation, respectively. Somatic mutations are more common than germline and *BRCA2* are more common than *BRCA1* mutations; the frequency of mutations is higher in the metastatic setting. Despite that *BRCA* testing in PC is now standard in clinical practice, several open questions remain.

Keywords: prostate cancer; *BRCA1*; *BRCA2*; germline mutation; somatic mutation; meta-analysis



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1. Introduction

In oncology, the demand for breast cancer gene (*BRCA*) genetic testing in various tumor types, such as ovarian, breast, pancreatic, and prostate cancer (PC), is rapidly and continuously increasing to predict the efficacy of cancer treatment, help physicians make decisions about therapeutic options, and assess individual and familial risk [1,2].

Regarding patients with PC, knowledge of the presence of *BRCA1/2* mutations in cancer tissue (somatic mutations) or in peripheral blood (germline mutations) provides useful information of prognostic and predictive value.

In particular, first, *BRCA* mutation identification allows the planning of an appropriate therapeutic algorithm. Indeed, *BRCA* testing is essential to determine whether patients are eligible for new targeted and effective therapeutic strategies, such as poly-ADP-ribose polymerase inhibitors (PARPis). While treatment of metastatic PC has historically consisted of hormonal therapy with androgen deprivation, chemotherapy, and various radiotherapy approaches, the recent approval of PARPis, such as rucaparib and olaparib, has revolutionized the therapeutic algorithm of metastatic castration-resistant PC (mCRPC) and led to a marked improvement in clinical outcomes for patients with *BRCA1/2* mutations [3–7].

Second, the identification of a pathogenetic germline variant in *BRCA* genes provides access to prevention programs, oncogenetic counseling of family members to identify high-risk carriers, special screening programs for early detection of *BRCA*-related heredo-familial tumors, and risk-reduction strategies [8].

BRCA testing requires standardized and harmonized procedures for germline and tumor DNA sequencing and for the interpretation of results; *BRCA* mutational status should be verified by a specialized laboratory using a validated analytical method [9,10].

According to the latest position paper of Italian Scientific Societies and the most recent European Society of Medical Oncology (ESMO) clinical practice guidelines, it is preferable to investigate pathogenetic *BRCA* variants in tumor tissue first, as the probability of detecting *BRCA* mutations is higher than with germline analysis. Patients who are found to have somatic pathogenetic *BRCA* mutations should be referred for germline testing to identify possible constitutional and hereditary variants. Somatic testing should also be proposed to patients who initially underwent germline testing that did not identify a pathogenetic variant and who are potential candidates for treatment with PARPis [9,10].

Data on the exact proportion of PC patients with *BRCA* mutations come from a 2018 systematic review and meta-analysis by Mok et al. They showed that the frequency of *BRCA1* and *BRCA2* carriers in PC patients was 0.9% and 2.2%, respectively [11].

As these data did not include more recent studies, and *BRCA* testing is now standard in clinical practice in metastatic PC thanks to the approval of specific treatments, we decided to conduct an updated systematic literature review and meta-analysis with the aim of evaluating the proportion of PC patients with *BRCA* mutations, dividing the data obtained into subgroups according to the type of mutation (germline or somatic mutations; mutation of *BRCA1* and/or *BRCA2*) and according to the disease setting (any stage or metastatic PC or mCRPC).

2. Materials and Methods

This systematic review was conducted according to the Preferred Reporting Items for Systematic Review and meta-analysis (PRISMA) guidelines, as reported in Figure 1.

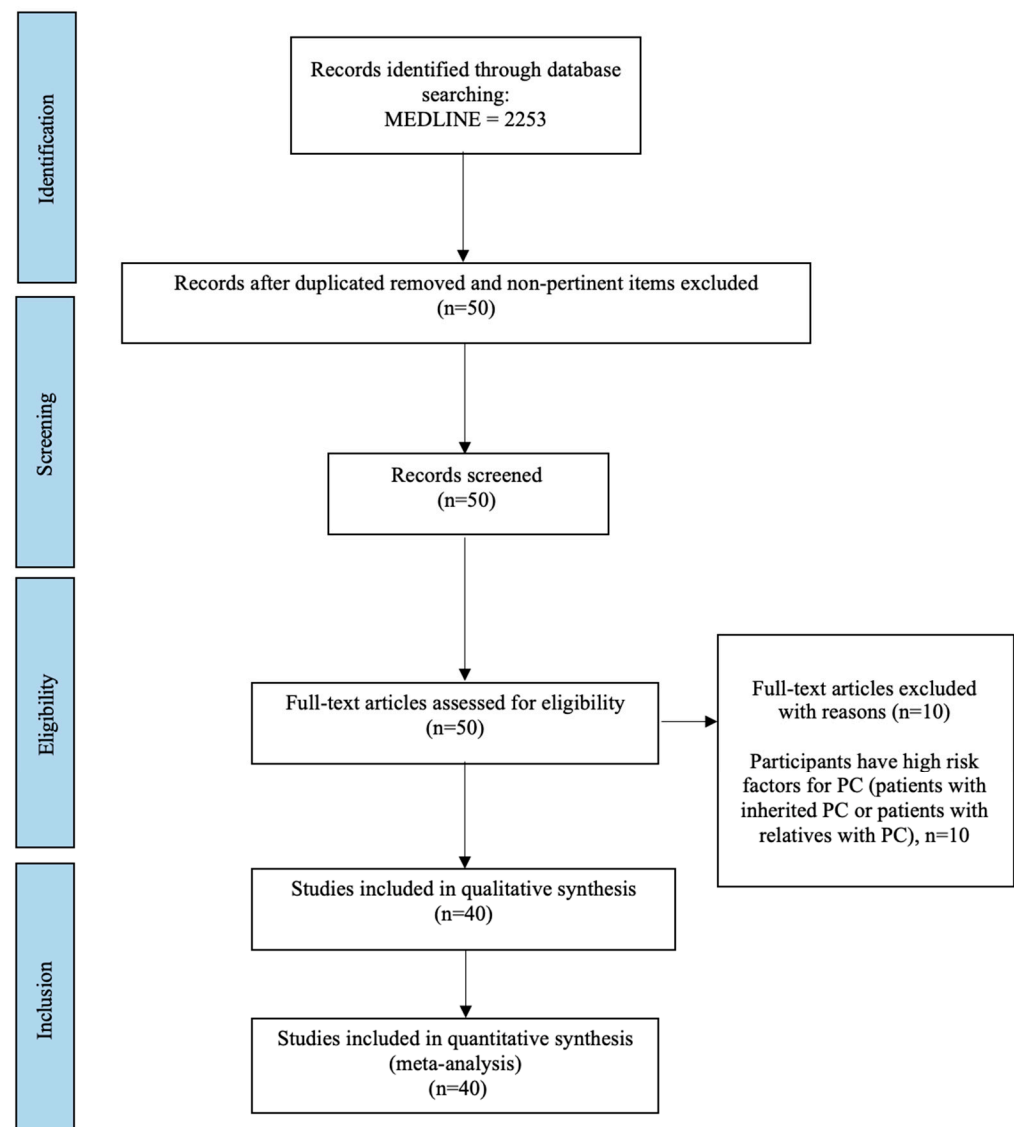


Figure 1. PRISMA diagram.

2.1. Search Strategy

An extensive literature search in PubMed, Web of Sciences, and Scopus databases was performed in November 2022 to identify all articles testing the proportion of BRCA1 and BRCA2 mutations in patients with PC.

The following keywords were used in our search strategy: “(prostate cancer) and (BRCA)”, “(prostate cancer) and (BRCA1 gene)”, “(prostate cancer) and (BRCA2 gene)”, “(prostate cancer) and (BRCA mutation)”, “(prostate cancer) and (BRCA testing)”, “(prostate cancer) and (germline BRCA)”, and “(prostate cancer) and (somatic BRCA)”. References of the identified articles were also checked manually to identify additional eligible items.

Initial screening was performed by one investigator (A.A.V.) and ineligible results were identified based on the titles and abstracts. If the study’s topic could not be ascertained from its title or abstract, the full-text version would be retrieved for evaluation. Disagreement was resolved by discussion or consensus with another co-author (M.D.M.).

2.2. Study Selection

To have sufficient data to calculate the number of BRCA mutation carriers among patients with PC, studies were screened for eligibility using the following inclusion criteria: (1) participants must be patients with PC, regardless of disease stage; (2) included studies

must report the proportion of patients with *BRCA* mutations tested by somatic and/or germline testing, regardless of the gene involved (*BRCA1* or *BRCA2* or any *BRCA*) and mutation variant; and (3) articles must be in English and published between 2000 and 2022.

The following criteria were used as exclusion criteria: (1) participants with established risk factors for PC such as patients with inherited PC or patients with relatives with PC and (2) case reports and reviews.

2.3. Data Collection

For each eligible article, the following data were collected: (1) first author's name; (2) year of publication; (3) total number of patients; (4) number of patients with or without *BRCA* mutations; (5) details of population disease setting: any stage PC, metastatic PC, and mCRPC; and (6) details of type of *BRCA* mutation: germline, somatic, *BRCA1*, and *BRCA2*.

2.4. Statistical Methods

The meta-analysis of the proportion of patients with PC with *BRCA* mutations was performed with MedCalc Statistical Software version 20.211 (MedCalc Software Ltd., Ostend, Belgium; <https://www.medcalc.org>; 2023). The software uses a Freeman–Tukey transformation (arcsine square root transformation) to calculate the weighted summary proportion under the fixed and random effects model. Heterogeneity is measured by Cochran's *Q*, calculated as the weighted sum of squared differences between the individual study proportion and the pooled proportion across studies. *Q* is distributed as a chi-square statistic with *k* (number of studies) minus 1 degrees of freedom. The I^2 statistic describes the percentage of variation across studies that is due to heterogeneity rather than chance. $I^2 = 100\% \times (Q - df)/Q$.

2.5. Role of Funding Source

There was no funding source for this systematic review and meta-analysis. All authors had full access to all data and the corresponding author (M.D.M.) had the final responsibility for the decision to submit for publication.

3. Results

Our research items led to the identification of 2253 titles. After removing duplicates, non-pertinent items and ineligible studies, 40 articles were included in this systematic review and meta-analysis (Figure 1; Table 1) [12–51].

See Appendix A for all detailed statistical results of the meta-analysis.

3.1. Meta-Analysis: Proportion of Patients with Prostate Cancer with *BRCA1* Mutation

3.1.1. Proportion of Patients with Any Stage PC with *BRCA1* Mutation

The proportion of germline *BRCA1* mutation carriers among patients with any stage PC was available from 31 articles, for a total of 32,525 patients, and was equal to 0.73% (95% confidence interval, CI: 0.51–1.00), with significant heterogeneity ($I^2 = 81.19\%$; $p < 0.0001$) (Figure 2a).

The proportion of somatic *BRCA1* mutation carriers among patients with any stage PC was available from 10 articles, for a total of 3229 patients, and was equal to 1.20% (95% CI: 0.85–1.60), without significant heterogeneity ($I^2 = 0.00\%$; $p = 0.7423$) (Figure 2b).

Table 1. Summary of the results of the meta-analysis.

	<i>BRCA1</i>		<i>BRCA2</i>		<i>BRCA1/2</i>	
	Germline	Somatic	Germline	Somatic	Germline	Somatic
Patients with any stage PC						
Number of studies	31	10	30	10	29	10
Number of patients	32,525	3229	29,813	3229	33,784	3229
% (fixed effect)	0.53 (95% CI: 0.45–0.62)	1.20 (95% CI: 0.85–1.64)	2.47 (95% CI: 2.30–2.66)	4.77 (95% CI: 4.06–5.56)	4.17 (95% CI: 3.96–4.39)	6.07 (95% CI: 5.27–6.95)
% (random effect)	0.73 (95% CI: 0.51–1.00)	1.20 (95% CI: 0.85–1.60)	3.25 (95% CI: 2.54–4.04)	6.29 (95% CI: 3.79–9.38)	4.47 (95% CI: 3.38–5.70)	7.18 (95% CI: 4.89–9.87)
Heterogeneity I^2 (p -value)	81.19% ($p < 0.0001$)	0.00% ($p = 0.7423$)	90.96% ($p < 0.0001$)	89.14% ($p < 0.0001$)	95.57% ($p < 0.0001$)	84.17% ($p < 0.0001$)
Metastatic PC patients						
Number of studies	10	6	10	6	11	6
Number of patients	3963	1384	3963	1384	11,670	1384
% (fixed effect)	0.58 (95% CI: 0.37–0.87)	1.10 (95% CI: 0.62–1.79)	3.44 (95% CI: 2.89–4.05)	9.16 (95% CI: 7.70–10.80)	6.56 (95% CI: 6.12–7.03)	10.12 (95% CI: 8.58–11.82)
% (random effect)	0.94 (95% CI: 0.19–2.23)	1.10 (95% CI: 0.62–1.71)	4.51 (95% CI: 2.93–6.42)	10.26 (95% CI: 7.92–12.85)	5.84 (95% CI: 3.72–8.41)	10.94 (95% CI: 8.73–13.36)
Heterogeneity I^2 (p -value)	88.85% ($p < 0.0001$)	0.00% ($p = 0.9224$)	81.54% ($p < 0.0001$)	38.42% ($p = 0.1498$)	93.61% ($p < 0.0001$)	29.07% ($p = 0.2170$)
mCRPC patients						
Number of studies	7	5	7	5	7	5
Number of patients	2571	1243	2571	1243	2571	1243
% (fixed effect)	0.56 (95% CI: 0.31–0.93)	1.10 (95% CI: 0.60–1.85)	2.69 (95% CI: 2.10–3.39)	9.05 (95% CI: 7.51–10.78)	3.50 (95% CI: 2.82–4.28)	10.03 (95% CI: 8.42–11.83)
% (random effect)	1.21 (95% CI: 0.05–3.84)	1.10 (95% CI: 0.60–1.76)	3.90 (95% CI: 2.13–6.16)	10.52 (95% CI: 7.64–13.81)	5.26 (95% CI: 2.18–9.57)	11.26 (95% CI: 8.49–14.38)
Heterogeneity I^2 (p -value)	92.36% ($p < 0.0001$)	0.00% ($p = 0.8425$)	76.71% ($p = 0.0002$)	49.50% ($p = 0.0945$)	91.57% ($p < 0.0001$)	42.38% ($p = 0.1390$)

PC: prostate cancer; mCRPC: metastatic castration-resistant prostate cancer; CI: confidence interval.

3.1.2. Proportion of Patients with Metastatic PC with BRCA1 Mutation

The proportion of germline BRCA1 mutation carriers among patients with metastatic PC was available from 10 articles, for a total of 3963 patients, and was equal to 0.94% (95% CI: 0.19–2.23), with significant heterogeneity ($I^2 = 88.85%$; $p < 0.0001$) (Figure 2c).

The proportion of somatic BRCA1 mutation carriers among patients with metastatic PC was available from six articles, for a total of 1384 patients, and was equal to 1.10% (95% CI: 0.62–1.71), without significant heterogeneity ($I^2 = 0.00%$; $p = 0.9224$) (Figure 2d).

3.1.3. Proportion of Patients with mCRPC with BRCA1 Mutation

The proportion of germline BRCA1 mutation carriers among patients with mCRPC was available from seven articles, for a total of 2571 patients, and was equal to 1.21% (95% CI: 0.053–3.84), with significant heterogeneity ($I^2 = 92.36%$; $p < 0.0001$) (Figure 2e).

The proportion of somatic BRCA1 mutation carriers among patients with mCRPC was available from five articles, for a total of 1243 patients, and was equal to 1.10% (95% CI: 0.60–1.76), without significant heterogeneity ($I^2 = 0.00%$; $p = 0.8425$) (Figure 2f).

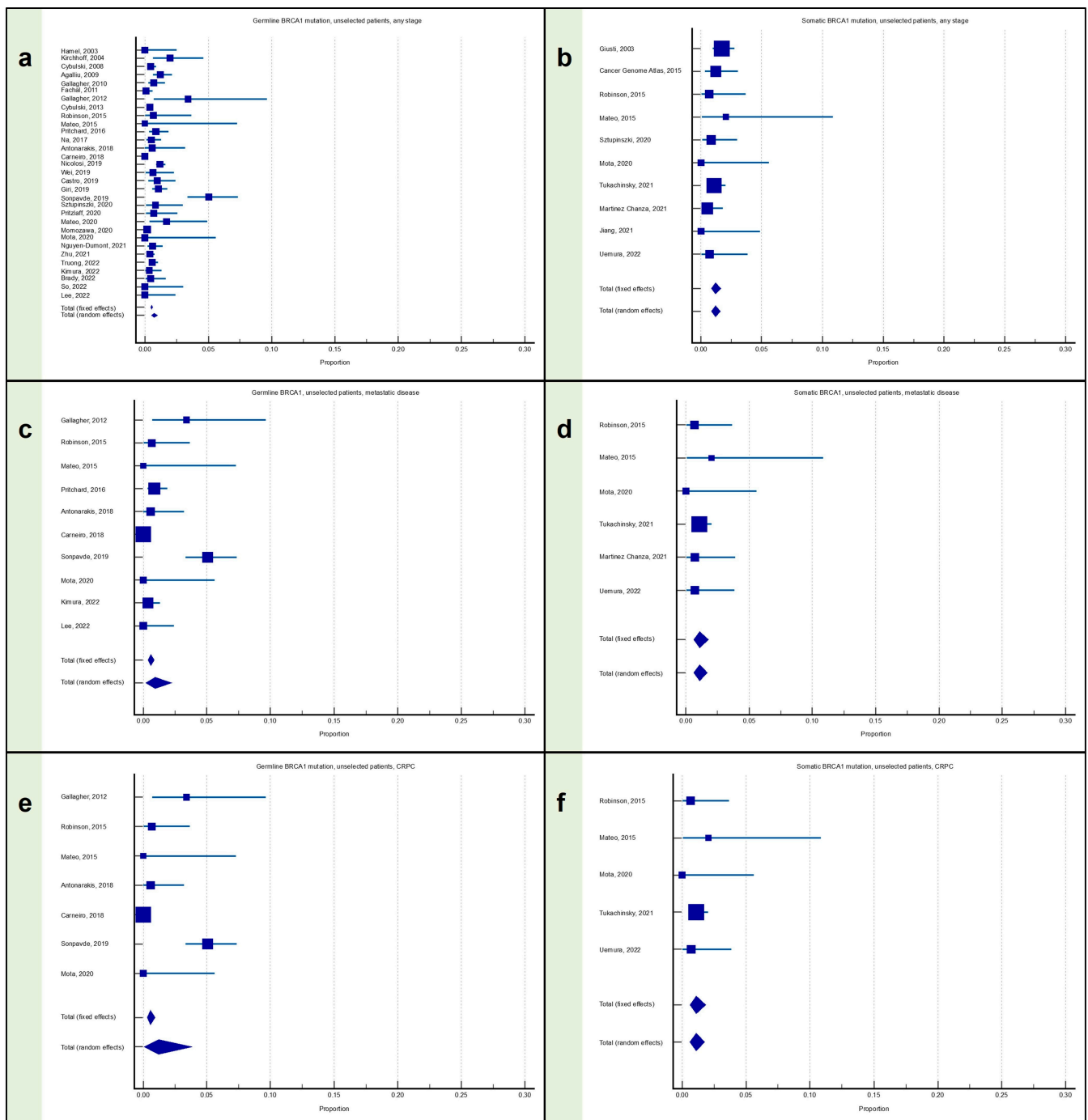


Figure 2. Proportion of patients with prostate cancer harboring the BRCA1 mutation. (a) Proportion of patients with any stage PC with the germline BRCA1 mutation; (b) proportion of patients with any stage PC with the somatic BRCA1 mutation; (c) proportion of patients with metastatic PC with the germline BRCA1 mutation; (d) proportion of patients with metastatic PC with the somatic BRCA1 mutation; (e) proportion of patients with mCRPC with the germline BRCA1 mutation; and (f) proportion of patients with mCRPC with the somatic BRCA1 mutation [12–16,18–20,22–34,37–42,44–51].

3.2. Meta-Analysis: Proportion of Patients with Prostate Cancer with BRCA2 Mutation

3.2.1. Proportion of Patients with Any Stage PC with BRCA2 Mutation

The proportion of germline BRCA2 mutation carriers among patients with any stage PC was available from 30 articles, for a total of 29,813 patients, and was equal to 3.25% (95% CI: 2.54–4.04), with significant heterogeneity ($I^2 = 90.96\%$; $p < 0.0001$) (Figure 3a).

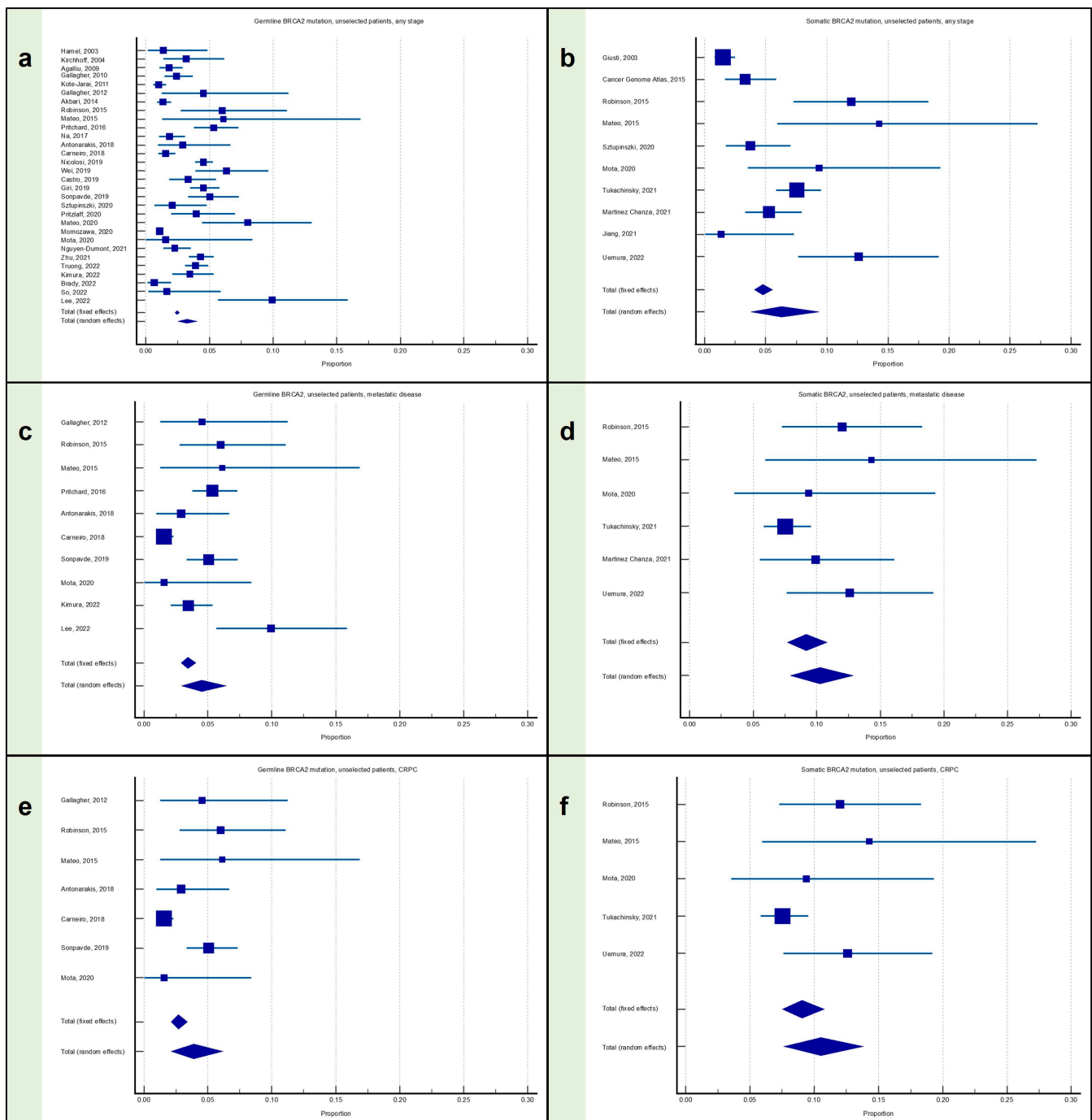


Figure 3. Proportion of patients with prostate cancer harboring the BRCA2 mutation. (a) Proportion of patients with any stage PC with the germline BRCA2 mutation; (b) proportion of patients with any stage PC with the somatic BRCA2 mutation; (c) proportion of patients with metastatic PC with the germline BRCA2 mutation; (d) proportion of patients with metastatic PC with the somatic BRCA2 mutation; (e) proportion of patients with mCRPC with the germline BRCA2 mutation; and (f) proportion of patients with mCRPC with the somatic BRCA2 mutation [12,13,15–19,21–35,37–51].

The proportion of somatic BRCA2 mutation carriers among patients with any stage PC was available from 10 articles, for a total of 3229 patients, and was equal to 6.29% (95% CI: 3.79–9.38), with significant heterogeneity ($I^2 = 89.14\%$; $p < 0.0001$) (Figure 3b).

3.2.2. Proportion of Patients with Metastatic PC with BRCA2 Mutation

The proportion of germline BRCA2 mutation carriers among patients with metastatic PC was available from 10 articles, for a total of 3963 patients, and was equal to 4.51% (95% CI: 2.93–6.42), with significant heterogeneity ($I^2 = 81.54\%$; $p < 0.0001$) (Figure 3c).

The proportion of somatic BRCA2 mutation carriers among patients with metastatic PC was available from six articles, for a total of 1384 patients, and was equal to 10.26% (95% CI: 7.92–12.85), without significant heterogeneity ($I^2 = 38.42\%$; $p = 0.1498$) (Figure 3d).

3.2.3. Proportion of Patients with mCRPC with BRCA2 Mutation

The proportion of germline BRCA2 mutation carriers among patients with mCRPC was available from seven articles, for a total of 2571 patients, and was equal to 3.90% (95% CI: 2.13–6.16), with significant heterogeneity ($I^2 = 76.71\%$; $p = 0.0002$) (Figure 3e).

The proportion of somatic BRCA2 mutation carriers among patients with mCRPC was available from five articles, for a total of 1243 patients, and was equal to 10.52% (95% CI: 7.64–13.81), without significant heterogeneity ($I^2 = 49.50\%$; $p = 0.0945$) (Figure 3f).

3.3. Meta-Analysis: Proportion of Patients with Prostate Cancer with Any BRCA Mutation

3.3.1. Proportion of Patients with Any Stage PC with Any BRCA Mutation

The proportion of germline BRCA1/2 mutation carriers among patients with any stage PC was available from 29 articles, for a total of 33,784 patients, and was equal to 4.47% (95% CI: 3.38–5.70), with significant heterogeneity ($I^2 = 95.57\%$; $p < 0.0001$) (Figure 4a).

The proportion of somatic BRCA1/2 mutation carriers among patients with any stage PC was available from 10 articles, for a total of 3229 patients, and was equal to 7.18% (95% CI: 4.89–9.87), with significant heterogeneity ($I^2 = 84.17\%$; $p < 0.0001$) (Figure 4b).

3.3.2. Proportion of Patients with Metastatic PC with Any BRCA Mutation

The proportion of germline BRCA1/2 mutation carriers among patients with metastatic PC was available from 11 articles, for a total of 11,670 patients, and was equal to 5.84% (95% CI: 3.72–8.41), with significant heterogeneity ($I^2 = 93.61\%$; $p < 0.0001$) (Figure 4c).

The proportion of somatic BRCA1/2 mutation carriers among patients with metastatic PC was available from six articles, for a total of 1384 patients, and was equal to 10.94% (95% CI: 8.73–13.36), without significant heterogeneity ($I^2 = 29.07\%$; $p = 0.2170$) (Figure 4d).

3.3.3. Proportion of Patients with mCRPC with Any BRCA Mutation

The proportion of germline BRCA1/2 mutation carriers among patients with mCRPC was available from seven articles, for a total of 2571 patients, and was equal to 5.26% (95% CI: 2.18–9.57), with significant heterogeneity ($I^2 = 91.57\%$; $p < 0.0001$) (Figure 4e).

The proportion of somatic BRCA1/2 mutation carriers among patients with mCRPC was available from five articles, for a total of 1243 patients, and was equal to 11.26% (95% CI: 8.49–14.38), without significant heterogeneity ($I^2 = 42.38\%$; $p = 0.1390$) (Figure 4f).

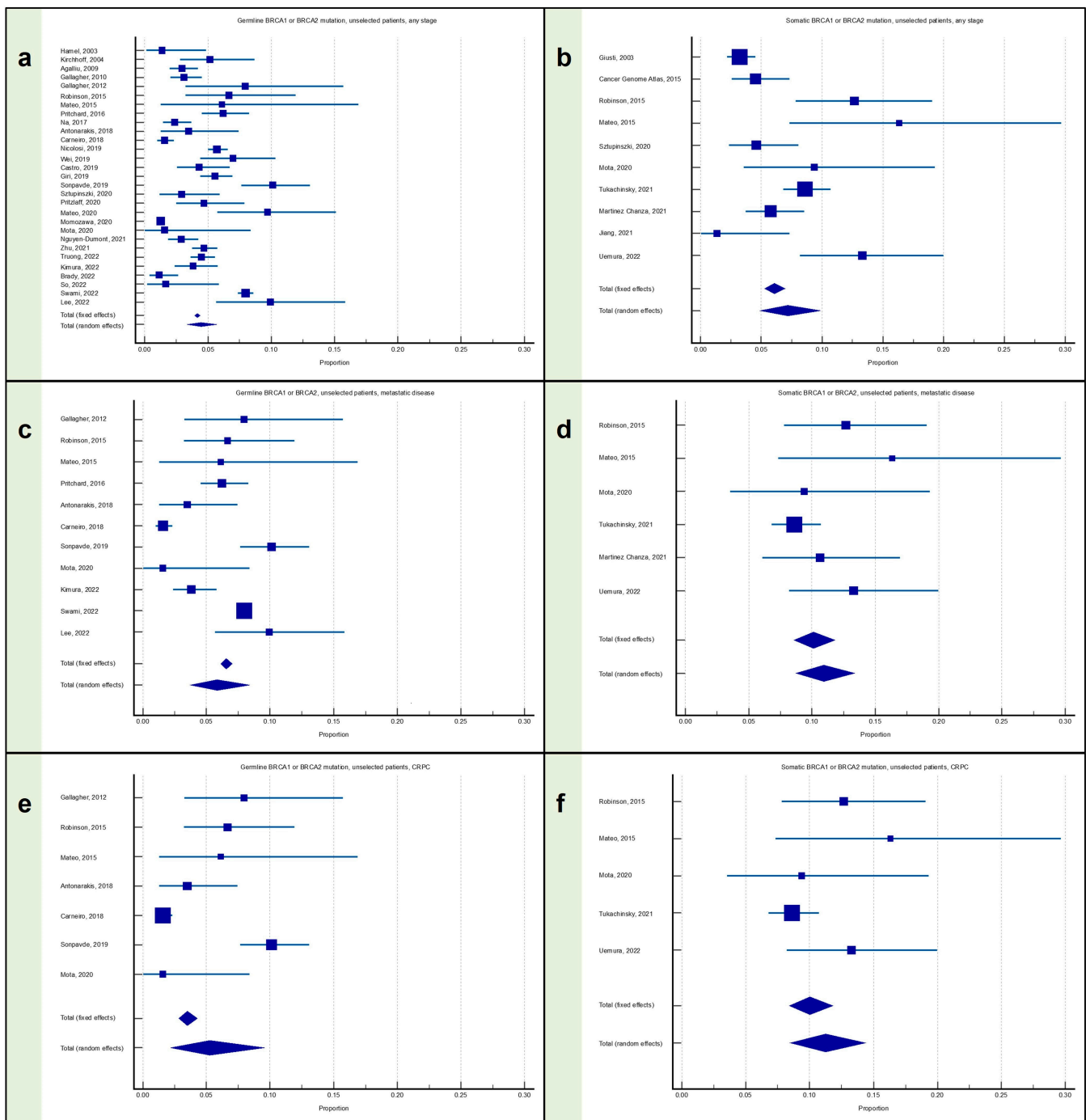


Figure 4. Proportion of patients with prostate cancer harboring any BRCA mutation. (a) Proportion of patients with any stage PC with the germline BRCA1/2 mutation; (b) proportion of patients with any stage PC with the somatic BRCA1/2 mutation; (c) proportion of patients with metastatic PC with the germline BRCA1/2 mutation; (d) proportion of patients with metastatic PC with the somatic BRCA1/2 mutation; (e) proportion of patients with mCRPC with the germline BRCA1/2 mutation; and (f) proportion of patients with mCRPC with the somatic BRCA1/2 mutation [12,13,15,16,19,22–51].

4. Discussion

In this systematic review and meta-analysis, we collected all papers describing the frequency of somatic and/or germline *BRCA1* and *BRCA2* mutations in patients with PC. We analyzed this frequency in three populations of patients: all PC patients regardless of the stage, patients with metastatic PC, and patients with mCRPC.

First, although the complete information about somatic and germline status was available only in a subset of studies, we confirmed that, overall, somatic *BRCA* mutations are markedly more frequent than germline mutations: 7.18% versus 4.47%, respectively, in all patients with PC regardless of the stage of PC disease; 10.94% versus 5.84%, respectively, in patients with metastatic disease; and 11.26% versus 5.26%, respectively, in patients with mCRPC. Data obtained considering mutations of *BRCA1* or *BRCA2* separately confirmed a higher frequency of somatic mutations than germline mutations for both genes.

Second, both germline and somatic *BRCA2* mutations are more common than *BRCA1* mutations in both metastatic and patients with any stage PC. Specifically, among metastatic patients, 10.26% and 4.51% of cases have somatic and germline *BRCA2* mutations, respectively, while 1.1% and 0.94% have somatic and germline *BRCA1* mutations, respectively. Among patients with any stage PC, 6.29% and 3.25% have somatic and germline *BRCA2* mutations, respectively, while 1.20% and 0.73% have somatic and germline *BRCA1* mutations, respectively.

Finally, the frequency of *BRCA* mutations is higher in the series including only patients with metastatic disease than in the whole population of all patients studied, regardless of stage. Namely, the frequency of somatic *BRCA1/2* mutations is 10.94% in patients with metastatic disease (11.26% when the analysis is limited to the castration-resistant setting) and 7.18% in all patients with any stage PC.

Similar to other solid tumors, including breast and ovarian cancer, in prostate cancer, the presence of *BRCA* mutation is an important clinical factor with prognostic and predictive value, especially owing to the recent introduction of target therapies such as PARPis into clinical practice. To date, these drugs have only been approved for mCRPC disease, although several studies are underway to predict their use in earlier stages of PC [52–54]. Therefore, molecular characterization of patients with PC is essential to avoid depriving them of a potential effective therapeutic option.

Our data confirm that many more cases can be identified with the somatic test than with the germline test alone. Therefore, the possibility of performing the somatic test must be guaranteed in all oncological centers. Until a few years ago, the only relevant determination in clinical practice was the search for germline mutations, in the context of genetic counseling for known or suspected hereditary cases. Nowadays, with the availability of target drugs, the determination of *BRCA* mutational status becomes relevant for therapeutic choices, and this implies a marked increase in the number of cases eligible for testing, as well as the need to obtain results more quickly in order to allow timely therapeutic decisions. This is a good example of the risk of disparities among different countries and different centers, owing to the asymmetry in reimbursement systems and in technical pathways for carrying out molecular tests; that is, patients could be at risk of unequal access not only to drugs, but also to tests.

In our meta-analysis, we focused on evaluating only the rate of *BRCA1* and *BRCA2* genes. Actually, although the real predictive value of other genes is controversial, mutations in genes related to the DNA repair pathway of homologous recombination (HR) (HRD-positive patients) have also been proposed and studied as predictive factors for PARPis. Therefore, in addition to *BRCA* mutations, other HRD-related gene aberrations may also serve as novel biomarkers for predicting the efficacy of PARPis [55].

However, in PC, the recommendations in the various international guidelines are not entirely congruent.

The 2022 Italian Association of Medical Oncology (AIOM) guidelines recommend *BRCA* testing for all patients with metastatic PC, without a recommendation about other genes. Namely, the indication to perform the test is also extended to patients who meet certain criteria regarding personal and family history, number of affected relatives, cancer type, multiple primary tumors, and age at diagnosis, as well as histologic, immunohistochemical, and molecular tumor characteristics [9].

Instead, European guidelines, issued by ESMO in 2020, recommend that tissue-based molecular assays may be used in conjunction with clinicopathological factors for treatment

decisions in localized prostate cancer; germline testing for *BRCA2* and other DDR genes associated with cancer predisposition syndromes is recommended in patients with a family history of cancer and should be considered in all patients with metastatic PC; tumor testing for HR genes can be considered in patients with mCRPC [10].

Still somewhat different are the recommendations of the National Comprehensive Cancer Network (NCCN) guidelines published in 2023: germline multigene testing that includes at least *BRCA1*, *BRCA2*, *ATM*, *PALB2*, *CHEK2*, *HOXB13*, *MLH1*, *MSH2*, *MSH6*, and *PMS2* is recommended if the patient is affected by metastatic, regional (node positive), very-high-risk localized, or high-risk localized PC (diagnosed at any age) and/or if certain criteria about family history and/or ancestry are met, while tumor testing for alterations in HR DNA repair genes, such as *BRCA1*, *BRCA2*, *ATM*, *PALB2*, *FANCA*, *RAD51D*, *CHEK2*, and *CDK12*, is recommended in patients with metastatic PC and can be considered in patients with regional PC [5].

The test has also recently acquired, in addition to the traditional implications for the management of hereditary–familial cases, implications for the therapeutic management of patients. At least in part, probably, this fact explains the heterogeneity between different recommendations and guidelines.

Our meta-analysis, although based on a systematic and updated review of the literature, has some important limitations.

First of all, the absence of individual patient data implies that the information on the characteristics of the patients in the studies is limited. Because the incidence of mutations according to individual characteristics for all patients, their ethnicity, or geographic origin are not uniformly available, clinical characteristics associated with the presence of a mutation could not be analyzed. As for the association with clinical stage, the higher frequency of mutations in patients with metastatic disease seems more relevant for somatic than for germline mutations. However, we are unable to establish the exact timing of the appearance of somatic mutations with respect to disease progression, also because most somatic tests are performed on tissue previously archived at the time of initial diagnosis. Only serial tests on tissue samples taken at different stages of the disease could establish whether, in cases of wildtype for germline mutations, the appearance of somatic mutations is an early event potentially associated with higher risk of metastases and a worse prognosis or simply a late event in the natural history of the disease. However, the execution of molecular testing on archived tissue is consistent with daily clinical practice. Thus, the collected data help to estimate the number of patients with PC with *BRCA* mutations that we can expect to see in clinical practice. In this scenario, it would be important for urologists to be aware at the time of a prostate biopsy that the tissue is not only needed for histologic analysis, but could also be useful for genetic analysis. The biopsy or tissue removed should be quantitatively sufficient for both analyses so that the patient is not biopsied again later.

Another limitation is the heterogeneity that characterizes the techniques of molecular analysis used in the studies included in the meta-analysis. Different techniques can be different for sensitivity and specificity, and this could contribute to the high heterogeneity found in the incidence of mutations among different studies. Furthermore, different *BRCA* mutations variants were not uniformly distinguished according to their predictive value.

Lastly, our meta-analysis did not include studies that investigated *BRCA* mutations using the liquid biopsy technique. According to recent studies, liquid biopsy seems to have a very interesting role for three main reasons.

First, a study by Tukachinsky et al. showed that there is a good agreement between data obtained from somatic testing and those obtained from liquid biopsy [46]. Thus, the liquid biopsy technique would allow to assess both somatic and germline mutations simultaneously using only one blood sample.

Second, a recent exploratory analysis of the PROfound study evaluated the efficacy of olaparib in patients with *BRCA/ATM* mutations investigated by liquid biopsy, showing that the clinical outcome endpoints were similar to those reached in the cohort in which mutations had been studied with somatic testing [3,56]. Therefore, this study highlights

that the liquid biopsy technique could be a test with the same prognostic and predictive value as somatic testing.

Third, somatic testing has failure rates for several reasons, such as a lack of quantitatively sufficient tumor tissue or other technical difficulties. For example, in the PROfound study, the success rate of somatic testing was 69% [3]. In this scenario, liquid biopsy could exceed the limitations of somatic testing and become a valid and useful alternative.

Therefore, the role of liquid biopsy will become increasingly intriguing in the future because it could offer our patients with mCRPC a less invasive technique than somatic testing that can overcome its limitations while maintaining the ability to provide predictive and prognostic information.

5. Conclusions

In prostate cancer, knowledge of the presence of somatic and/or germline mutations of *BRCA* provides useful information of prognostic and predictive value to plan an appropriate therapeutic algorithm thanks to the introduction of new therapeutic options and ensure access to prevention programs and oncogenetic counseling.

In summary, as *BRCA* testing is now well-established in clinical practice, this meta-analysis aimed to describe the rate of *BRCA* mutations that clinicians should expect to see on a daily basis.

Meta-analysis demonstrates that somatic mutations are more common than germline mutations, *BRCA2* mutations are more common than *BRCA1* mutations in both metastatic patients and patients with any stage PC, and that the frequency of *BRCA* mutations is higher in the series including only patients with metastatic disease than in the whole population of all patients studied regardless of stage.

Because the test has recently acquired implications for the therapeutic management of patients with PC, the recommendations in the various international guidelines are not entirely congruent and, in this scenario, several questions remain for the future, both in terms of the best time to perform *BRCA* testing based on ongoing studies of the use of PARPis at an earlier stage of PC disease, as well as in terms of the genes to look for (*BRCA* or HRD panel) and the optimal molecular analysis technique (somatic and/or germline testing or liquid biopsy).

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Conflicts of Interest: Massimo Di Maio received honoraria and had roles as consultant or advisor for AstraZeneca, Pfizer, Novartis, Roche, Takeda, Eisai, Merck Sharp & Dohme, Janssen, Astellas, Boehringer Ingelheim, Amgen, and Merck, outside this work; and received an institutional research grant by Tesaro—GlaxoSmithKline, outside this work. Other authors (A.A.V.; R.D.; O.P.; J.P.; F.V.; A.P.) declare no conflict of interest.

Appendix A

This appendix has been provided by the authors to give readers additional information about their work.

Table A1. Meta-analysis: proportion of unselected PC patients with somatic *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Giusti, 2003	940	1.702	0.976 to 2.749	29.05	29.05
Cancer Genome Atlas, 2015	333	1.201	0.328 to 3.047	10.31	10.31
Robinson, 2015	150	0.667	0.0169 to 3.658	4.66	4.66
Mateo, 2015	49	2.041	0.0517 to 10.854	1.54	1.54
Sztupinszki, 2020	240	0.833	0.101 to 2.978	7.44	7.44
Mota, 2020	64	0.000	0.000 to 5.601	2.01	2.01
Tukachinsky, 2021	837	1.075	0.493 to 2.031	25.87	25.87
Martinez Chanza, 2021	399	0.501	0.0608 to 1.799	12.35	12.35
Jiang, 2021	74	0.000	0.000 to 4.863	2.32	2.32
Uemura, 2022	143	0.699	0.0177 to 3.835	4.45	4.45
Total (fixed effects)	3229	1.199	0.853 to 1.637	100.00	100.00
Total (random effects)	3229	1.199	0.853 to 1.603	100.00	100.00

Test for heterogeneity

Q	5.9758
DF	9
Significance level	$p = 0.7423$
I ² (inconsistency)	0.00%
95% CI for I ²	0.00 to 43.62

Publication bias

Egger's test	
Intercept	-0.6710
95% CI	-1.9905 to 0.6484
Significance level	$p = 0.2746$
Begg's test	
Kendall's Tau	-0.2000
Significance level	$p = 0.4208$

Table A2. Meta-analysis: proportion of unselected PC patients with somatic *BRCA2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Giusti, 2003	940	1.489	0.817 to 2.486	29.05	11.84
Cancer Genome Atlas, 2015	333	3.303	1.660 to 5.834	10.31	11.11
Robinson, 2015	150	12.000	7.269 to 18.301	4.66	9.95
Mateo, 2015	49	14.286	5.942 to 27.242	1.54	7.18
Sztupinszki, 2020	240	3.750	1.729 to 6.999	7.44	10.71
Mota, 2020	64	9.375	3.519 to 19.297	2.01	7.94
Tukachinsky, 2021	837	7.527	5.832 to 9.528	25.87	11.79

Table A2. Cont.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Martinez Chanza, 2021	399	5.263	3.287 to 7.933	12.35	11.29
Jiang, 2021	74	1.351	0.0342 to 7.301	2.32	8.34
Uemura, 2022	143	12.587	7.634 to 19.162	4.45	9.86
Total (fixed effects)	3229	4.766	4.058 to 5.558	100.00	100.00
Total (random effects)	3229	6.294	3.792 to 9.375	100.00	100.00

Test for heterogeneity

Q	82.8977
DF	9
Significance level	$p < 0.0001$
I ² (inconsistency)	89.14%
95% CI for I ²	82.15 to 93.40

Publication bias

Egger's test	
Intercept	3.0217
95% CI	−1.6936 to 7.7370
Significance level	$p = 0.1777$
Begg's test	
Kendall's Tau	0.1556
Significance level	$p = 0.5312$

Table A3. Meta-analysis: proportion of unselected PC patients with somatic BRCA1/2 mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Giusti, 2003	940	3.191	2.163 to 4.525	29.05	12.62
Cancer Genome Atlas, 2015	333	4.505	2.543 to 7.321	10.31	11.47
Robinson, 2015	150	12.667	7.801 to 19.072	4.66	9.79
Mateo, 2015	49	16.327	7.322 to 29.657	1.54	6.36
Sztupinszki, 2020	240	4.583	2.310 to 8.053	7.44	10.88
Mota, 2020	64	9.375	3.519 to 19.297	2.01	7.24
Tukachinsky, 2021	837	8.602	6.791 to 10.710	25.87	12.53
Martinez Chanza, 2021	399	5.764	3.689 to 8.524	12.35	11.74
Jiang, 2021	74	1.351	0.0342 to 7.301	2.32	7.71
Uemura, 2022	143	13.287	8.193 to 19.969	4.45	9.67
Total (fixed effects)	3229	6.072	5.274 to 6.950	100.00	100.00
Total (random effects)	3229	7.183	4.892 to 9.874	100.00	100.00

Test for heterogeneity	
Q	56.8386
DF	9
Significance level	$p < 0.0001$
I ² (inconsistency)	84.17%
95% CI for I ²	72.49 to 90.89
Publication bias	
Egger's test	
Intercept	2.1709
95% CI	−1.8631 to 6.2049
Significance level	$p = 0.2498$
Begg's test	
Kendall's Tau	0.1556
Significance level	$p = 0.5312$

Table A4. Meta-analysis: proportion of unselected PC patients with germline *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Hamel, 2003	146	0.000	0.000 to 2.495	0.45	1.97
Kirchhoff, 2004	251	1.992	0.650 to 4.587	0.77	2.64
Cybulski, 2008	1793	0.446	0.193 to 0.877	5.51	4.45
Agalliu, 2009	979	1.226	0.635 to 2.131	3.01	4.07
Gallagher, 2010	832	0.721	0.265 to 1.563	2.56	3.94
Fachal, 2011	905	0.110	0.00280 to 0.614	2.78	4.01
Gallagher, 2012	88	3.409	0.709 to 9.641	0.27	1.41
Cybulski, 2013	3750	0.373	0.204 to 0.626	11.52	4.72
Robinson, 2015	150	0.667	0.0169 to 3.658	0.46	2.00
Mateo, 2015	49	0.000	0.000 to 7.252	0.15	0.91
Pritchard, 2016	692	0.867	0.319 to 1.878	2.13	3.78
Na, 2017	799	0.501	0.137 to 1.277	2.46	3.90
Antonarakis, 2018	172	0.581	0.0147 to 3.197	0.53	2.17
Carneiro, 2018	1534	0.000	0.000 to 0.240	4.71	4.37
Nicolosi, 2019	3607	1.192	0.864 to 1.602	11.08	4.71
Wei, 2019	316	0.633	0.0767 to 2.267	0.97	2.92
Castro, 2019	419	0.955	0.261 to 2.426	1.29	3.26
Giri, 2019	1328	1.054	0.578 to 1.762	4.08	4.28
Sonpavde, 2019	514	5.058	3.331 to 7.324	1.58	3.48
Sztupinszki, 2020	240	0.833	0.101 to 2.978	0.74	2.58
Pritzlaff, 2020	277	0.722	0.0876 to 2.584	0.85	2.76

Table A4. Cont.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Mateo, 2020	175	1.714	0.355 to 4.928	0.54	2.19
Momozawa, 2020	7636	0.183	0.100 to 0.307	23.46	4.87
Mota, 2020	64	0.000	0.000 to 5.601	0.20	1.12
Nguyen-Dumont, 2021	833	0.600	0.195 to 1.395	2.56	3.94
Zhu, 2021	1836	0.381	0.153 to 0.784	5.64	4.46
Truong, 2022	1883	0.584	0.292 to 1.043	5.79	4.47
Kimura, 2022	549	0.364	0.0441 to 1.310	1.69	3.55
Brady, 2022	437	0.458	0.0555 to 1.643	1.35	3.30
So, 2022	120	0.000	0.000 to 3.027	0.37	1.75
Lee, 2022	151	0.000	0.000 to 2.413	0.47	2.01
Total (fixed effects)	32,525	0.530	0.454 to 0.615	100.00	100.00
Total (random effects)	32,525	0.733	0.506 to 1.003	100.00	100.00

Test for heterogeneity

Q	159.5048
DF	30
Significance level	$p < 0.0001$
I ² (inconsistency)	81.19%
95% CI for I ²	74.05 to 86.37

Publication bias

Egger's test	
Intercept	1.4944
95% CI	0.03504 to 2.9537
Significance level	$p = 0.0451$
Begg's test	
Kendall's Tau	0.09247
Significance level	$p = 0.4649$

Table A5. Meta-analysis: proportion of unselected PC patients with germline *BRCA2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Hamel, 2003	146	1.370	0.166 to 4.861	0.49	2.62
Kirchhoff, 2004	251	3.187	1.386 to 6.183	0.84	3.12
Agalliu, 2009	979	1.839	1.093 to 2.890	3.28	3.89
Gallagher, 2010	832	2.404	1.474 to 3.688	2.79	3.83
Kote-Jarai, 2011	1832	1.037	0.626 to 1.615	6.14	4.05

Table A5. Cont.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	4.545	1.252 to 11.231	0.30	2.10
Akbari, 2014	1904	1.366	0.894 to 1.994	6.38	4.06
Robinson, 2015	150	6.000	2.780 to 11.084	0.51	2.65
Mateo, 2015	49	6.122	1.281 to 16.866	0.17	1.50
Pritchard, 2016	692	5.347	3.792 to 7.295	2.32	3.76
Na, 2017	799	1.877	1.054 to 3.078	2.68	3.82
Antonarakis, 2018	172	2.907	0.950 to 6.653	0.58	2.78
Carneiro, 2018	1534	1.565	1.005 to 2.319	5.14	4.02
Nicolosi, 2019	3607	4.547	3.890 to 5.278	12.09	4.15
Wei, 2019	316	6.329	3.908 to 9.606	1.06	3.30
Castro, 2019	419	3.341	1.839 to 5.543	1.41	3.49
Giri, 2019	1328	4.518	3.465 to 5.778	4.45	3.98
Sonpavde, 2019	514	5.058	3.331 to 7.324	1.73	3.61
Sztupinszki, 2020	240	2.083	0.680 to 4.795	0.81	3.08
Pritzlaff, 2020	277	3.971	1.999 to 6.994	0.93	3.20
Mateo, 2020	175	8.000	4.443 to 13.058	0.59	2.80
Momozawa, 2020	7636	1.087	0.867 to 1.346	25.59	4.20
Mota, 2020	64	1.562	0.0396 to 8.401	0.22	1.77
Nguyen-Dumont, 2021	833	2.281	1.379 to 3.539	2.79	3.83
Zhu, 2021	1836	4.303	3.421 to 5.334	6.16	4.05
Truong, 2022	1883	3.930	3.098 to 4.909	6.31	4.06
Kimura, 2022	549	3.461	2.096 to 5.352	1.84	3.65
Brady, 2022	437	0.686	0.142 to 1.993	1.47	3.52
So, 2022	120	1.667	0.202 to 5.891	0.41	2.42
Lee, 2022	151	9.934	5.667 to 15.855	0.51	2.66
Total (fixed effects)	29,813	2.473	2.300 to 2.656	100.00	100.00
Total (random effects)	29,813	3.246	2.539 to 4.037	100.00	100.00

Test for heterogeneity

Q	320.7889
DF	29
Significance level	$p < 0.0001$
I ² (inconsistency)	90.96%
95% CI for I ²	88.22 to 93.06

Publication bias	
Egger's test	
Intercept	2.4186
95% CI	0.3057 to 4.5316
Significance level	$p = 0.0264$
Begg's test	
Kendall's Tau	0.07126
Significance level	$p = 0.5802$

Table A6. Meta-analysis: proportion of unselected PC patients with germline *BRCA1/2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Hamel, 2003	146	1.370	0.166 to 4.861	0.43	3.02
Kirchhoff, 2004	251	5.179	2.786 to 8.694	0.75	3.37
Agalliu, 2009	979	2.962	1.993 to 4.227	2.90	3.83
Gallagher, 2010	832	3.125	2.051 to 4.545	2.46	3.80
Gallagher, 2012	88	7.955	3.258 to 15.705	0.26	2.60
Robinson, 2015	150	6.667	3.243 to 11.918	0.45	3.04
Mateo, 2015	49	6.122	1.281 to 16.866	0.15	2.04
Pritchard, 2016	692	6.214	4.533 to 8.279	2.05	3.76
Na, 2017	799	2.378	1.438 to 3.689	2.37	3.79
Antonarakis, 2018	172	3.488	1.291 to 7.438	0.51	3.14
Carneiro, 2018	1534	1.565	1.005 to 2.319	4.54	3.90
Nicolosi, 2019	3607	5.739	5.002 to 6.548	10.67	3.97
Wei, 2019	316	6.962	4.414 to 10.351	0.94	3.49
Castro, 2019	419	4.296	2.566 to 6.705	1.24	3.60
Giri, 2019	1328	5.572	4.400 to 6.945	3.93	3.88
Sonpavde, 2019	514	10.117	7.648 to 13.055	1.52	3.68
Sztupinszki, 2020	240	2.917	1.181 to 5.917	0.71	3.35
Pritzlaff, 2020	277	4.693	2.522 to 7.892	0.82	3.42
Mateo, 2020	175	9.714	5.761 to 15.098	0.52	3.15
Momozawa, 2020	7636	1.270	1.031 to 1.547	22.59	4.00
Mota, 2020	64	1.562	0.0396 to 8.401	0.19	2.30
Nguyen-Dumont, 2021	833	2.881	1.855 to 4.257	2.47	3.80
Zhu, 2021	1836	4.684	3.763 to 5.753	5.43	3.92
Truong, 2022	1883	4.514	3.621 to 5.552	5.57	3.92
Kimura, 2022	549	3.825	2.383 to 5.788	1.63	3.70
Brady, 2022	437	1.144	0.373 to 2.650	1.30	3.62
So, 2022	120	1.667	0.202 to 5.891	0.36	2.87
Swami, 2022	7707	7.967	7.372 to 8.594	22.80	4.00
Lee, 2022	151	9.934	5.667 to 15.855	0.45	3.05
Total (fixed effects)	33,784	4.174	3.964 to 4.393	100.00	100.00
Total (random effects)	33,784	4.466	3.376 to 5.700	100.00	100.00

Test for heterogeneity	
Q	632.0792
DF	28
Significance level	$p < 0.0001$
I ² (inconsistency)	95.57%
95% CI for I ²	94.50 to 96.43
Publication bias	
Egger's test	
Intercept	0.4449
95% CI	−2.6280 to 3.5179
Significance level	$p = 0.7687$
Begg's test	
Kendall's Tau	0.03941
Significance level	$p = 0.7641$

Table A7. Meta-analysis: proportion of metastatic PC patients with somatic *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	0.667	0.0169 to 3.658	10.86	10.86
Mateo, 2015	49	2.041	0.0517 to 10.854	3.60	3.60
Mota, 2020	64	0.000	0.000 to 5.601	4.68	4.68
Tukachinsky, 2021	837	1.075	0.493 to 2.031	60.29	60.29
Martinez Chanza, 2021	141	0.709	0.0180 to 3.888	10.22	10.22
Uemura, 2022	143	0.699	0.0177 to 3.835	10.36	10.36
Total (fixed effects)	1384	1.096	0.618 to 1.794	100.00	100.00
Total (random effects)	1384	1.096	0.616 to 1.710	100.00	100.00

Test for heterogeneity	
Q	1.4171
DF	5
Significance level	$p = 0.9224$
I ² (inconsistency)	0.00%
95% CI for I ²	0.00 to 13.04
Publication bias	
Egger's test	
Intercept	0.01503
95% CI	−1.4005 to 1.4305
Significance level	$p = 0.9779$
Begg's test	
Kendall's Tau	0.06667
Significance level	$p = 0.8510$

Table A8. Meta-analysis: proportion of metastatic PC patients with somatic *BRCA2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	12.000	7.269 to 18.301	10.86	16.64
Mateo, 2015	49	14.286	5.942 to 27.242	3.60	7.26
Mota, 2020	64	9.375	3.519 to 19.297	4.68	9.02
Tukachinsky, 2021	837	7.527	5.832 to 9.528	60.29	34.95
Martinez Chanza, 2021	141	9.929	5.535 to 16.098	10.22	15.99
Uemura, 2022	143	12.587	7.634 to 19.162	10.36	16.14
Total (fixed effects)	1384	9.161	7.696 to 10.802	100.00	100.00
Total (random effects)	1384	10.256	7.921 to 12.854	100.00	100.00

Test for heterogeneity

Q	8.1196
DF	5
Significance level	$p = 0.1498$
I ² (inconsistency)	38.42%
95% CI for I ²	0.00 to 75.53

Publication bias

Egger's test	
Intercept	2.1365
95% CI	0.4976 to 3.7754
Significance level	$p = 0.0224$
Begg's test	
Kendall's Tau	0.06667
Significance level	$p = 0.8510$

Table A9. Meta-analysis: proportion of metastatic PC patients with somatic *BRCA1/2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	12.667	7.801 to 19.072	10.86	15.82
Mateo, 2015	49	16.327	7.322 to 29.657	3.60	6.40
Mota, 2020	64	9.375	3.519 to 19.297	4.68	8.05
Tukachinsky, 2021	837	8.602	6.791 to 10.710	60.29	39.34
Martinez Chanza, 2021	141	10.638	6.078 to 16.939	10.22	15.12
Uemura, 2022	143	13.287	8.193 to 19.969	10.36	15.28
Total (fixed effects)	1384	10.115	8.580 to 11.822	100.00	100.00
Total (random effects)	1384	10.940	8.732 to 13.364	100.00	100.00

Test for heterogeneity	
Q	7.0496
DF	5
Significance level	$p = 0.2170$
I ² (inconsistency)	29.07%
95% CI for I ²	0.00 to 70.93
Publication bias	
Egger's test	
Intercept	1.9127
95% CI	0.2041 to 3.6213
Significance level	$p = 0.0359$
Begg's test	
Kendall's Tau	0.3333
Significance level	$p = 0.3476$

Table A10. Meta-analysis: proportion of metastatic PC patients with germline *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	3.409	0.709 to 9.641	2.24	8.49
Robinson, 2015	150	0.667	0.0169 to 3.658	3.80	9.81
Mateo, 2015	49	0.000	0.000 to 7.252	1.26	6.77
Pritchard, 2016	692	0.867	0.319 to 1.878	17.44	11.87
Antonarakis, 2018	172	0.581	0.0147 to 3.197	4.35	10.09
Carneiro, 2018	1534	0.000	0.000 to 0.240	38.64	12.26
Sonpavde, 2019	514	5.058	3.331 to 7.324	12.96	11.63
Mota, 2020	64	0.000	0.000 to 5.601	1.64	7.58
Kimura, 2022	549	0.364	0.0441 to 1.310	13.84	11.69
Lee, 2022	151	0.000	0.000 to 2.413	3.83	9.82
Total (fixed effects)	3963	0.583	0.371 to 0.873	100.00	100.00
Total (random effects)	3963	0.935	0.192 to 2.229	100.00	100.00

Test for heterogeneity	
Q	80.7401
DF	9
Significance level	$p < 0.0001$
I ² (inconsistency)	88.85%
95% CI for I ²	81.60 to 93.25

Publication bias	
Egger's test	
Intercept	1.9459
95% CI	−2.5292 to 6.4209
Significance level	$p = 0.3454$
Begg's test	
Kendall's Tau	0.1111
Significance level	$p = 0.6547$

Table A11. Meta-analysis: proportion of metastatic PC patients with germline *BRCA2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	4.545	1.252 to 11.231	2.24	7.64
Robinson, 2015	150	6.000	2.780 to 11.084	3.80	9.45
Mateo, 2015	49	6.122	1.281 to 16.866	1.26	5.60
Pritchard, 2016	692	5.347	3.792 to 7.295	17.44	12.87
Antonarakis, 2018	172	2.907	0.950 to 6.653	4.35	9.87
Carneiro, 2018	1534	1.565	1.005 to 2.319	38.64	13.62
Sonpavde, 2019	514	5.058	3.331 to 7.324	12.96	12.43
Mota, 2020	64	1.562	0.0396 to 8.401	1.64	6.51
Kimura, 2022	549	3.461	2.096 to 5.352	13.84	12.54
Lee, 2022	151	9.934	5.667 to 15.855	3.83	9.47
Total (fixed effects)	3963	3.439	2.894 to 4.054	100.00	100.00
Total (random effects)	3963	4.514	2.932 to 6.418	100.00	100.00

Test for heterogeneity	
Q	48.7604
DF	9
Significance level	$p < 0.0001$
I ² (inconsistency)	81.54%
95% CI for I ²	67.17 to 89.62
Publication bias	
Egger's test	
Intercept	2.5522
95% CI	−0.4949 to 5.5992
Significance level	$p = 0.0895$
Begg's test	
Kendall's Tau	−0.02222
Significance level	$p = 0.9287$

Table A12. Meta-analysis: proportion of metastatic PC patients with germline *BRCA1/2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	7.955	3.258 to 15.705	0.76	7.55
Robinson, 2015	150	6.667	3.243 to 11.918	1.29	8.71
Mateo, 2015	49	6.122	1.281 to 16.866	0.43	6.02
Pritchard, 2016	692	6.214	4.533 to 8.279	5.93	10.54
Antonarakis, 2018	172	3.488	1.291 to 7.438	1.48	8.97
Carneiro, 2018	1534	1.565	1.005 to 2.319	13.14	10.89
Sonpavde, 2019	514	10.117	7.648 to 13.055	4.41	10.33
Mota, 2020	64	1.562	0.0396 to 8.401	0.56	6.74
Kimura, 2022	549	3.825	2.383 to 5.788	4.71	10.38
Swami, 2022	7707	7.967	7.372 to 8.594	65.99	11.13
Lee, 2022	151	9.934	5.667 to 15.855	1.30	8.73
Total (fixed effects)	11,670	6.561	6.119 to 7.026	100.00	100.00
Total (random effects)	11,670	5.842	3.721 to 8.405	100.00	100.00

Test for heterogeneity

Q	156.4338
DF	10
Significance level	$p < 0.0001$
I ² (inconsistency)	93.61%
95% CI for I ²	90.42 to 95.73

Publication bias

Egger's test	
Intercept	-1.2792
95% CI	-5.3286 to 2.7702
Significance level	$p = 0.4930$
Begg's test	
Kendall's Tau	-0.01818
Significance level	$p = 0.9379$

Table A13. Meta-analysis: proportion of mCRPC patients with somatic *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	0.667	0.0169 to 3.658	12.10	12.10
Mateo, 2015	49	2.041	0.0517 to 10.854	4.01	4.01
Mota, 2020	64	0.000	0.000 to 5.601	5.21	5.21
Tukachinsky, 2021	837	1.075	0.493 to 2.031	67.15	67.15
Uemura, 2022	143	0.699	0.0177 to 3.835	11.54	11.54
Total (fixed effects)	1243	1.104	0.601 to 1.853	100.00	100.00
Total (random effects)	1243	1.104	0.600 to 1.758	100.00	100.00

Test for heterogeneity	
Q	1.4098
DF	4
Significance level	$p = 0.8425$
I ² (inconsistency)	0.00%
95% CI for I ²	0.00 to 44.46
Publication bias	
Egger's test	
Intercept	0.03864
95% CI	−1.9079 to 1.9852
Significance level	$p = 0.9536$
Begg's test	
Kendall's Tau	0.0000
Significance level	$p = 1.0000$

Table A14. Meta-analysis: proportion of mCRPC patients with somatic *BRCA2* mutation.

Study	Sample size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	12.000	7.269 to 18.301	12.10	20.83
Mateo, 2015	49	14.286	5.942 to 27.242	4.01	10.15
Mota, 2020	64	9.375	3.519 to 19.297	5.21	12.33
Tukachinsky, 2021	837	7.527	5.832 to 9.528	67.15	36.38
Uemura, 2022	143	12.587	7.634 to 19.162	11.54	20.32
Total (fixed effects)	1243	9.045	7.512 to 10.775	100.00	100.00
Total (random effects)	1243	10.523	7.635 to 13.812	100.00	100.00

Test for heterogeneity	
Q	7.9212
DF	4
Significance level	$p = 0.0945$
I ² (inconsistency)	49.50%
95% CI for I ²	0.00 to 81.49
Publication bias	
Egger's test	
Intercept	2.2040
95% CI	−0.01351 to 4.4216
Significance level	$p = 0.0508$
Begg's test	
Kendall's Tau	0.2000
Significance level	$p = 0.6242$

Table A15. Meta-analysis: proportion of mCRPC patients with somatic *BRCA1/2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Robinson, 2015	150	12.667	7.801 to 19.072	12.10	20.28
Mateo, 2015	49	16.327	7.322 to 29.657	4.01	9.23
Mota, 2020	64	9.375	3.519 to 19.297	5.21	11.37
Tukachinsky, 2021	837	8.602	6.791 to 10.710	67.15	39.41
Uemura, 2022	143	13.287	8.193 to 19.969	11.54	19.71
Total (fixed effects)	1243	10.026	8.415 to 11.828	100.00	100.00
Total (random effects)	1243	11.263	8.485 to 14.378	100.00	100.00

Test for heterogeneity

Q	6.9426
DF	4
Significance level	$p = 0.1390$
I ² (inconsistency)	42.38%
95% CI for I ²	0.00 to 78.81

Publication bias

Egger's test	
Intercept	1.9919
95% CI	-0.3078 to 4.2916
Significance level	$p = 0.0704$
Begg's test	
Kendall's Tau	0.6000
Significance level	$p = 0.1416$

Table A16. Meta-analysis: proportion of mCRPC patients with germline *BRCA1* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	3.409	0.709 to 9.641	3.45	13.51
Robinson, 2015	150	0.667	0.0169 to 3.658	5.86	14.67
Mateo, 2015	49	0.000	0.000 to 7.252	1.94	11.75
Antonarakis, 2018	172	0.581	0.0147 to 3.197	6.71	14.90
Carneiro, 2018	1534	0.000	0.000 to 0.240	59.54	16.49
Sonpavde, 2019	514	5.058	3.331 to 7.324	19.98	16.06
Mota, 2020	64	0.000	0.000 to 5.601	2.52	12.62
Total (fixed effects)	2571	0.562	0.311 to 0.934	100.00	100.00
Total (random effects)	2571	1.207	0.0526 to 3.839	100.00	100.00

Test for heterogeneity	
Q	78.5852
DF	6
Significance level	$p < 0.0001$
I ² (inconsistency)	92.36%
95% CI for I ²	86.81 to 95.58
Publication bias	
Egger's test	
Intercept	2.4198
95% CI	−3.9590 to 8.7987
Significance level	$p = 0.3743$
Begg's test	
Kendall's Tau	−0.04762
Significance level	$p = 0.8806$

Table A17. Meta-analysis: proportion of mCRPC patients with germline *BRCA2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	4.545	1.252 to 11.231	3.45	11.73
Robinson, 2015	150	6.000	2.780 to 11.084	5.86	14.50
Mateo, 2015	49	6.122	1.281 to 16.866	1.94	8.60
Antonarakis, 2018	172	2.907	0.950 to 6.653	6.71	15.16
Carneiro, 2018	1534	1.565	1.005 to 2.319	59.54	20.91
Sonpavde, 2019	514	5.058	3.331 to 7.324	19.98	19.09
Mota, 2020	64	1.562	0.0396 to 8.401	2.52	10.01
Total (fixed effects)	2571	2.691	2.101 to 3.391	100.00	100.00
Total (random effects)	2571	3.895	2.132 to 6.158	100.00	100.00

Test for heterogeneity	
Q	25.7604
DF	6
Significance level	$p = 0.0002$
I ² (inconsistency)	76.71%
95% CI for I ²	51.20 to 88.88

Publication bias	
Egger's test	
Intercept	2.2629
95% CI	−0.7550 to 5.2808
Significance level	$p = 0.1118$
Begg's test	
Kendall's Tau	0.04762
Significance level	$p = 0.8806$

Table A18. Meta-analysis: proportion of mCRPC patients with germline *BRCA1/2* mutation.

Study	Sample Size	Proportion (%)	95% CI	Weight (%)	
				Fixed	Random
Gallagher, 2012	88	7.955	3.258 to 15.705	3.45	13.42
Robinson, 2015	150	6.667	3.243 to 11.918	5.86	14.68
Mateo, 2015	49	6.122	1.281 to 16.866	1.94	11.54
Antonarakis, 2018	172	3.488	1.291 to 7.438	6.71	14.94
Carneiro, 2018	1534	1.565	1.005 to 2.319	59.54	16.72
Sonpavde, 2019	514	10.117	7.648 to 13.055	19.98	16.23
Mota, 2020	64	1.562	0.0396 to 8.401	2.52	12.46
Total (fixed effects)	2571	3.500	2.824 to 4.283	100.00	100.00
Total (random effects)	2571	5.255	2.177 to 9.569	100.00	100.00

Test for heterogeneity	
Q	71.1738
DF	6
Significance level	$p < 0.0001$
I ² (inconsistency)	91.57%
95% CI for I ²	85.20 to 95.20
Publication bias	
Egger's test	
Intercept	2.8014
95% CI	−2.9857 to 8.5884
Significance level	$p = 0.2685$
Begg's test	
Kendall's Tau	0.04762
Significance level	$p = 0.8806$

References

1. Kuchenbaecker, K.B.; Hopper, J.L.; Barnes, D.R.; Phillips, K.A.; Mooij, T.M.; Roos-Blom, M.J.; Jervis, S.; van Leeuwen, F.E.; Milne, R.L.; Andrieu, N.; et al. Risks of Breast, Ovarian, and Contralateral Breast Cancer for *BRCA1* and *BRCA2* Mutation Carriers. *JAMA* **2017**, *317*, 2402–2416. [[CrossRef](#)] [[PubMed](#)]
2. Shindo, K.; Yu, J.; Suenaga, M.; Fesharakizadeh, S.; Cho, C.; Macgregor-Das, A.; Siddiqui, A.; Witmer, P.D.; Tamura, K.; Song, T.J.; et al. Deleterious Germline Mutations in Patients with Apparently Sporadic Pancreatic Adenocarcinoma. *J. Clin. Oncol.* **2017**, *35*, 3382–3390. [[CrossRef](#)]

3. de Bono, J.; Mateo, J.; Fizazi, K.; Saad, F.; Shore, N.; Sandhu, S.; Chi, K.N.; Sartor, O.; Agarwal, N.; Olmos, D.; et al. Olaparib for Metastatic Castration-Resistant Prostate Cancer. *N. Engl. J. Med.* **2020**, *382*, 2091–2102. [[CrossRef](#)]
4. Mateo, J.; Porta, N.; Bianchini, D.; McGovern, U.; Elliott, T.; Jones, R.; Syndikus, I.; Ralph, C.; Jain, S.; Varughese, M.; et al. Olaparib in patients with metastatic castration-resistant prostate cancer with DNA repair gene aberrations (TOPARP-B): A multicentre, open-label, randomised, phase 2 trial. *Lancet Oncol.* **2020**, *21*, 162–174. [[CrossRef](#)] [[PubMed](#)]
5. Schaeffer, E.M.; Srinivas, S.; Adra, N.; An, Y.; Barocas, D.; Bitting, R.; Bryce, A.; Chapin, B.; Cheng, H.H.; D’Amico, A.V.; et al. NCCN Guidelines[®] Insights: Prostate Cancer, Version 1.2023. *J. Natl. Compr. Cancer Netw.* **2022**, *20*, 1288–1298.
6. Anscher, M.S.; Chang, E.; Gao, X.; Gong, Y.; Weinstock, C.; Bloomquist, E.; Adeniyi, O.; Charlab, R.; Zimmerman, S.; Serlemitsos-Day, M.; et al. FDA Approval Summary: Rucaparib for the Treatment of Patients with Deleterious BRCA-Mutated Metastatic Castrate-Resistant Prostate Cancer. *Oncologist* **2021**, *26*, 139–146. [[CrossRef](#)]
7. Abida, W.; Patnaik, A.; Campbell, D.; Shapiro, J.; Bryce, A.H.; McDermott, R.; Sautois, B.; Vogelzang, N.J.; Bambury, R.M.; Voog, E.; et al. Rucaparib in Men With Metastatic Castration-Resistant Prostate Cancer Harboring a BRCA1 or BRCA2 Gene Alteration. *J. Clin. Oncol.* **2020**, *38*, 3763–3772. [[CrossRef](#)]
8. Hemminki, K. Familial risk and familial survival in prostate cancer. *World J. Urol.* **2012**, *30*, 143–148. [[CrossRef](#)] [[PubMed](#)]
9. Russo, A.; Incorvaia, L.; Capoluongo, E.; Tagliaferri, P.; Gori, S.; Cortesi, L.; Genuardi, M.; Turchetti, D.; De Giorgi, U.; Di Maio, M.; et al. Implementation of preventive and predictive BRCA testing in patients with breast, ovarian, pancreatic, and prostate cancer: A position paper of Italian Scientific Societies. *ESMO Open* **2022**, *7*, 100459. [[CrossRef](#)] [[PubMed](#)]
10. Parker, C.; Castro, E.; Fizazi, K.; Heidenreich, A.; Ost, P.; Procopio, G.; Tombal, B.; Gillessen, S. Prostate cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **2020**, *31*, 1119–1134. [[CrossRef](#)]
11. Oh, M.; Alkushaym, N.; Fallatah, S.; Althagafi, A.; Aljadeed, R.; Alsowaida, Y.; Jeter, J.; Martin, J.R.; Babiker, H.M.; McBride, A.; et al. The association of BRCA1 and BRCA2 mutations with prostate cancer risk, frequency, and mortality: A meta-analysis. *Prostate* **2019**, *79*, 880–895. [[CrossRef](#)]
12. Giusti, R.M.; Rutter, J.L.; Duray, P.H.; Freedman, L.S.; Konichevsky, M.; Fisher-Fischbein, J.; Greene, M.H.; Maslansky, B.; Fischbein, A.; Gruber, S.B.; et al. A twofold increase in BRCA mutation related prostate cancer among Ashkenazi Israelis is not associated with distinctive histopathology. *J. Med. Genet.* **2003**, *40*, 787–792. [[CrossRef](#)] [[PubMed](#)]
13. Kirchoff, T.; Kauff, N.D.; Mitra, N.; Nafa, K.; Huang, H.; Palmer, C.; Gulati, T.; Wadsworth, E.; Donat, S.; Robson, M.E.; et al. BRCA mutations and risk of prostate cancer in Ashkenazi Jews. *Clin. Cancer Res.* **2004**, *10*, 2918–2921. [[CrossRef](#)]
14. Cybulski, C.; Górski, B.; Gronwald, J.; Huzarski, T.; Byrski, T.; Debniak, T.; Jakubowska, A.; Wokołarczyk, D.; Gliniewicz, B.; Sikorski, A.; et al. BRCA1 mutations and prostate cancer in Poland. *Eur. J. Cancer Prev.* **2008**, *17*, 62–66. [[CrossRef](#)] [[PubMed](#)]
15. Agalliu, I.; Gern, R.; Leanza, S.; Burk, R.D. Associations of high-grade prostate cancer with BRCA1 and BRCA2 founder mutations. *Clin. Cancer Res.* **2009**, *15*, 1112–1120. [[CrossRef](#)] [[PubMed](#)]
16. Gallagher, D.J.; Gaudet, M.M.; Pal, P.; Kirchoff, T.; Balistreri, L.; Vora, K.; Bhatia, J.; Stadler, Z.; Fine, S.W.; Reuter, V.; et al. Germline BRCA mutations denote a clinicopathologic subset of prostate cancer. *Clin. Cancer Res.* **2010**, *16*, 2115–2121. [[CrossRef](#)]
17. Kote-Jarai, Z.; Leongamornlert, D.; Saunders, E.; Tymrakiewicz, M.; Castro, E.; Mahmud, N.; Guy, M.; Edwards, S.; O’Brien, L.; Sawyer, E.; et al. BRCA2 is a moderate penetrance gene contributing to young-onset prostate cancer: Implications for genetic testing in prostate cancer patients. *Br. J. Cancer* **2011**, *105*, 1230–1234. [[CrossRef](#)] [[PubMed](#)]
18. Fachal, L.; Gómez-Caamaño, A.; Celeiro-Muñoz, C.; Peleteiro, P.; Blanco, A.; Carballo, A.; Forteza, J.; Carracedo, A.; Vega, A. BRCA1 mutations do not increase prostate cancer risk: Results from a meta-analysis including new data. *Prostate* **2011**, *71*, 1768–1779. [[CrossRef](#)] [[PubMed](#)]
19. Gallagher, D.J.; Cronin, A.M.; Milowsky, M.I.; Morris, M.J.; Bhatia, J.; Scardino, P.T.; Eastham, J.A.; Offit, K.; Robson, M.E. Germline BRCA mutation does not prevent response to taxane-based therapy for the treatment of castration-resistant prostate cancer. *BJU Int.* **2012**, *109*, 713–719. [[CrossRef](#)]
20. Cybulski, C.; Wokołarczyk, D.; Kluźniak, W.; Jakubowska, A.; Górski, B.; Gronwald, J.; Huzarski, T.; Kashyap, A.; Byrski, T.; Debniak, T.; et al. An inherited NBN mutation is associated with poor prognosis prostate cancer. *Br. J. Cancer* **2013**, *108*, 461–468. [[CrossRef](#)]
21. Akbari, M.R.; Wallis, C.J.; Toi, A.; Trachtenberg, J.; Sun, P.; Narod, S.A.; Nam, R.K. The impact of a BRCA2 mutation on mortality from screen-detected prostate cancer. *Br. J. Cancer* **2014**, *111*, 1238–1240. [[CrossRef](#)] [[PubMed](#)]
22. Abeshouse, A.; Ahn, J.; Akbani, R.; Ally, A.; Amin, S.; Andry, C.D.; Annala, M.; Aprikian, A.; Armenia, J.; Arora, A.; et al. The Molecular Taxonomy of Primary Prostate Cancer. *Cell* **2015**, *163*, 1011–1025. [[CrossRef](#)] [[PubMed](#)]
23. Robinson, D.; Van Allen, E.M.; Wu, Y.M.; Schultz, N.; Lonigro, R.J.; Mosquera, J.M.; Montgomery, B.; Taplin, M.E.; Pritchard, C.C.; Attard, G.; et al. Integrative clinical genomics of advanced prostate cancer. *Cell* **2015**, *161*, 1215–1228. [[CrossRef](#)] [[PubMed](#)]
24. Pritchard, C.C.; Mateo, J.; Walsh, M.F.; De Sarkar, N.; Abida, W.; Beltran, H.; Garofalo, A.; Gulati, R.; Carreira, S.; Eeles, R.; et al. Inherited DNA-Repair Gene Mutations in Men with Metastatic Prostate Cancer. *N. Engl. J. Med.* **2016**, *375*, 443–453. [[CrossRef](#)] [[PubMed](#)]
25. Na, R.; Zheng, S.L.; Han, M.; Yu, H.; Jiang, D.; Shah, S.; Ewing, C.M.; Zhang, L.; Novakovic, K.; Petkewicz, J.; et al. Germline Mutations in ATM and BRCA1/2 Distinguish Risk for Lethal and Indolent Prostate Cancer and are Associated with Early Age at Death. *Eur. Urol.* **2017**, *71*, 740–747. [[CrossRef](#)]

26. Antonarakis, E.S.; Lu, C.; Luber, B.; Liang, C.; Wang, H.; Chen, Y.; Silberstein, J.L.; Piana, D.; Lai, Z.; Isaacs, W.B.; et al. Germline DNA-repair Gene Mutations and Outcomes in Men with Metastatic Castration-resistant Prostate Cancer Receiving First-line Abiraterone and Enzalutamide. *Eur. Urol.* **2018**, *74*, 218–225. [[CrossRef](#)]
27. Carneiro, B.A.; Collier, K.A.; Nagy, R.J.; Pamarthy, S.; Sagar, V.; Fairclough, S.; Odegaard, J.; Lanman, R.B.; Costa, R.; Taxter, T.; et al. Acquired Resistance to Poly (ADP-ribose) Polymerase Inhibitor Olaparib in BRCA2-Associated Prostate Cancer Resulting from Biallelic BRCA2 Reversion Mutations Restores Both Germline and Somatic Loss-of-Function Mutations. *JCO Precis. Oncol.* **2018**, *2*, PO.17.00176. [[CrossRef](#)]
28. Nicolosi, P.; Ledet, E.; Yang, S.; Michalski, S.; Freschi, B.; O’Leary, E.; Esplin, E.D.; Nussbaum, R.L.; Sartor, O. Prevalence of Germline Variants in Prostate Cancer and Implications for Current Genetic Testing Guidelines. *JAMA Oncol.* **2019**, *5*, 523–528. [[CrossRef](#)]
29. Castro, E.; Romero-Laorden, N.; Del Pozo, A.; Lozano, R.; Medina, A.; Puente, J.; Piulats, J.M.; Lorente, D.; Saez, M.I.; Morales-Barrera, R.; et al. PROREPAIR-B: A Prospective Cohort Study of the Impact of Germline DNA Repair Mutations on the Outcomes of Patients With Metastatic Castration-Resistant Prostate Cancer. *J. Clin. Oncol.* **2019**, *37*, 490–503. [[CrossRef](#)]
30. Giri, V.N.; Hegarty, S.E.; Hyatt, C.; O’Leary, E.; Garcia, J.; Knudsen, K.E.; Kelly, W.K.; Gomella, L.G. Germline genetic testing for inherited prostate cancer in practice: Implications for genetic testing, precision therapy, and cascade testing. *Prostate* **2019**, *79*, 333–339. [[CrossRef](#)]
31. Sonpavde, G.; Agarwal, N.; Pond, G.R.; Nagy, R.J.; Nussenzveig, R.H.; Hahn, A.W.; Sartor, O.; Gourdin, T.S.; Nandagopal, L.; Ledet, E.M.; et al. Circulating tumor DNA alterations in patients with metastatic castration-resistant prostate cancer. *Cancer* **2019**, *125*, 1459–1469. [[CrossRef](#)]
32. Mateo, J.; Carreira, S.; Sandhu, S.; Miranda, S.; Mossop, H.; Perez-Lopez, R.; Nava Rodrigues, D.; Robinson, D.; Omlin, A.; Tunariu, N.; et al. DNA-Repair Defects and Olaparib in Metastatic Prostate Cancer. *N. Engl. J. Med.* **2015**, *373*, 1697–1708. [[CrossRef](#)] [[PubMed](#)]
33. Lee, A.M.; Saidian, A.; Shaya, J.; Nonato, T.; Cabal, A.; Randall, J.M.; Millard, F.; Stewart, T.; Rose, B.; Tamayo, P.; et al. The Prognostic Significance of Homologous Recombination Repair Pathway Alterations in Metastatic Hormone Sensitive Prostate Cancer. *Clin. Genitourin. Cancer* **2022**, *20*, 515–523. [[CrossRef](#)] [[PubMed](#)]
34. So, M.K.; Ahn, H.K.; Huh, J.; Kim, K.H. Germline pathogenic variants in unselected Korean men with prostate cancer. *Investig. Clin. Urol.* **2022**, *63*, 294–300. [[CrossRef](#)] [[PubMed](#)]
35. Brady, L.; Newcomb, L.F.; Zhu, K.; Zheng, Y.; Boyer, H.; Sarkar, N.; McKenney, J.K.; Brooks, J.D.; Carroll, P.R.; Dash, A.; et al. Germline mutations in penetrant cancer predisposition genes are rare in men with prostate cancer selecting active surveillance. *Cancer Med.* **2022**, *11*, 4332–4340. [[CrossRef](#)] [[PubMed](#)]
36. Swami, U.; Zimmerman, R.M.; Nussenzveig, R.H.; Hernandez, E.J.; Jo, Y.; Sayegh, N.; Wesolowski, S.; Kiedrowski, L.A.; Barata, P.C.; Lemmon, G.H.; et al. Genomic landscape of advanced prostate cancer patients with BRCA1 versus BRCA2 mutations as detected by comprehensive genomic profiling of cell-free DNA. *Front. Oncol.* **2022**, *12*, 966534. [[CrossRef](#)]
37. Hamel, N.; Kotar, K.; Foulkes, W.D. Founder mutations in BRCA1/2 are not frequent in Canadian Ashkenazi Jewish men with prostate cancer. *BMC Med. Genet.* **2003**, *4*, 7. [[CrossRef](#)]
38. Wei, Y.; Wu, J.; Gu, W.; Qin, X.; Dai, B.; Lin, G.; Gan, H.; Freedland, S.J.; Zhu, Y.; Ye, D. Germline DNA Repair Gene Mutation Landscape in Chinese Prostate Cancer Patients. *Eur. Urol.* **2019**, *76*, 280–283. [[CrossRef](#)]
39. Kimura, H.; Mizuno, K.; Shiota, M.; Narita, S.; Terada, N.; Fujimoto, N.; Ogura, K.; Hatano, S.; Iwasaki, Y.; Hakozaiki, N.; et al. Prognostic significance of pathogenic variants in BRCA1, BRCA2, ATM and PALB2 genes in men undergoing hormonal therapy for advanced prostate cancer. *Br. J. Cancer* **2022**, *127*, 1680–1690. [[CrossRef](#)]
40. Truong, H.; Breen, K.; Nandakumar, S.; Sjoberg, D.D.; Kemel, Y.; Mehta, N.; Lenis, A.T.; Reisz, P.A.; Carruthers, J.; Benfante, N.; et al. Gene-based Confirmatory Germline Testing Following Tumor-only Sequencing of Prostate Cancer. *Eur. Urol.* **2023**, *83*, 29–38. [[CrossRef](#)]
41. Uemura, H.; Oya, M.; Kamoto, T.; Sugimoto, M.; Shinozaki, K.; Morita, K.; Koto, R.; Takahashi, M.; Nii, M.; Shin, E.; et al. The prevalence of gene mutations in homologous recombination repair pathways in Japanese patients with metastatic castration-resistant prostate cancer in real-world clinical practice: The multi-institutional observational ZENSHIN study. *Cancer Med.* **2022**, *12*, 5265–5274. [[CrossRef](#)]
42. Jiang, X.; Hu, X.; Gu, Y.; Li, Y.; Jin, M.; Zhao, H.; Gao, R.; Huang, Z.; Lu, J. Homologous recombination repair gene mutations in Chinese localized and locally advanced prostate cancer patients. *Pathol. Res. Pract.* **2021**, *224*, 153507. [[CrossRef](#)]
43. Zhu, Y.; Wei, Y.; Zeng, H.; Li, Y.; Ng, C.F.; Zhou, F.; He, C.; Sun, G.; Ni, Y.; Chiu, P.K.F.; et al. Inherited Mutations in Chinese Men with Prostate Cancer. *J. Natl. Compr. Cancer Netw.* **2021**, *20*, 54–62. [[CrossRef](#)] [[PubMed](#)]
44. Nguyen-Dumont, T.; Dowty, J.G.; MacInnis, R.J.; Steen, J.A.; Riaz, M.; Dugué, P.A.; Renault, A.L.; Hammet, F.; Mahmoodi, M.; Theys, D.; et al. Rare Germline Pathogenic Variants Identified by Multigene Panel Testing and the Risk of Aggressive Prostate Cancer. *Cancers* **2021**, *13*, 1495. [[CrossRef](#)]
45. Martinez Chanza, N.; Bernard, B.; Barthelemy, P.; Accarain, A.; Paesmans, M.; Desmyter, L.; T’Kint de Roodenbeke, D.; Gil, T.; Sideris, S.; Roumeguere, T.; et al. Prevalence and clinical impact of tumor BRCA1 and BRCA2 mutations in patients presenting with localized or metastatic hormone-sensitive prostate cancer. *Prostate Cancer Prostatic. Dis.* **2022**, *25*, 199–207. [[CrossRef](#)]

46. Tukachinsky, H.; Madison, R.W.; Chung, J.H.; Gjoerup, O.V.; Severson, E.A.; Dennis, L.; Fendler, B.J.; Morley, S.; Zhong, L.; Graf, R.P.; et al. Genomic Analysis of Circulating Tumor DNA in 3,334 Patients with Advanced Prostate Cancer Identifies Targetable BRCA Alterations and AR Resistance Mechanisms. *Clin. Cancer Res.* **2021**, *27*, 3094–3105. [[CrossRef](#)] [[PubMed](#)]
47. Momozawa, Y.; Iwasaki, Y.; Hirata, M.; Liu, X.; Kamatani, Y.; Takahashi, A.; Sugano, K.; Yoshida, T.; Murakami, Y.; Matsuda, K.; et al. Germline Pathogenic Variants in 7636 Japanese Patients with Prostate Cancer and 12366 Controls. *J. Natl. Cancer Inst.* **2020**, *112*, 369–376. [[CrossRef](#)] [[PubMed](#)]
48. Mateo, J.; Seed, G.; Bertan, C.; Rescigno, P.; Dolling, D.; Figueiredo, I.; Miranda, S.; Nava Rodrigues, D.; Gurel, B.; Clarke, M.; et al. Genomics of lethal prostate cancer at diagnosis and castration resistance. *J. Clin. Investig.* **2020**, *130*, 1743–1751. [[CrossRef](#)]
49. Pritzlaff, M.; Tian, Y.; Reineke, P.; Stuenkel, A.J.; Allen, K.; Gutierrez, S.; Jackson, M.; Dolinsky, J.S.; LaDuca, H.; Xu, J.; et al. Diagnosing hereditary cancer predisposition in men with prostate cancer. *Genet. Med.* **2020**, *22*, 1517–1523. [[CrossRef](#)]
50. Mota, J.M.; Barnett, E.; Nauseef, J.T.; Nguyen, B.; Stopsack, K.H.; Wibmer, A.; Flynn, J.R.; Heller, G.; Danila, D.C.; Rathkopf, D.; et al. Platinum-Based Chemotherapy in Metastatic Prostate Cancer with DNA Repair Gene Alterations. *JCO Precis. Oncol.* **2020**, *4*, 355–366. [[CrossRef](#)]
51. Sztupinszki, Z.; Diossy, M.; Krzystanek, M.; Borcsok, J.; Pomerantz, M.M.; Tisza, V.; Spisak, S.; Rusz, O.; Csabai, I.; Freedman, M.L.; et al. Detection of Molecular Signatures of Homologous Recombination Deficiency in Prostate Cancer with or without BRCA1/2 Mutations. *Clin. Cancer Res.* **2020**, *26*, 2673–2680. [[CrossRef](#)]
52. Carr, T.H.; Adelman, C.; Barnicle, A.; Kozarewa, I.; Luke, S.; Lai, Z.; Hollis, S.; Dougherty, B.; Harrington, E.A.; Kang, J.; et al. Homologous Recombination Repair Gene Mutation Characterization by Liquid Biopsy: A Phase II Trial of Olaparib and Abiraterone in Metastatic Castrate-Resistant Prostate Cancer. *Cancers* **2021**, *13*, 5830. [[CrossRef](#)]
53. Rathkopf, D.; Chi, K.; Olmos, D.; Cheng, H.; Agarwal, N.; Graff, J.; Sandhu, S.; Hayreh, V.; Lopez-Gitlitz, A.; Francis, P.; et al. AMPLITUDE: A study of niraparib in combination with abiraterone acetate plus prednisone (AAP) versus AAP for the treatment of patients with deleterious germline or somatic homologous recombination repair (HRR) gene-altered metastatic castration-sensitive prostate cancer (mCSPC). *J. Clin. Oncol.* **2021**, *39*, TPS176. [[CrossRef](#)]
54. Chi, K.N.; Rathkopf, D.E.; Smith, M.R.; Efstathiou, E.; Attard, G.; Olmos, D.; Lee, J.Y.; Small, E.J.; Gomes, A.J.; Roubaud, G.; et al. Phase 3 MAGNITUDE study: First results of niraparib (NIRA) with abiraterone acetate and prednisone (AAP) as first-line therapy in patients (pts) with metastatic castration-resistant prostate cancer (mCRPC) with and without homologous recombination repair (HRR) gene alterations. *J. Clin. Oncol.* **2022**, *40*, 12. [[CrossRef](#)]
55. Hussain, M.; Mateo, J.; Fizazi, K.; Saad, F.; Shore, N.; Sandhu, S.; Chi, K.N.; Sartor, O.; Agarwal, N.; Olmos, D.; et al. Survival with Olaparib in Metastatic Castration-Resistant Prostate Cancer. *N. Engl. J. Med.* **2020**, *383*, 2345–2357. [[CrossRef](#)] [[PubMed](#)]
56. Matsubara, N.; de Bono, J.; Olmos, D.; Procopio, G.; Kawakami, S.; Ürün, Y.; van Alphen, R.; Flechon, A.; Carducci, M.A.; Choi, Y.D.; et al. Olaparib Efficacy in Patients with Metastatic Castration-resistant Prostate Cancer and BRCA1, BRCA2, or ATM Alterations Identified by Testing Circulating Tumor DNA. *Clin. Cancer Res.* **2023**, *29*, 92–99. [[CrossRef](#)]

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