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Occurrence of human pathogenic viruses in drinking water and in its sources: A review

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(Article begins on next page)

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Virus occurrence in sources for drinking water production and in drinking water: a review --Manuscript Draft--

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Abstract:	Microbiological quality of drinking water (DW) is crucial for Public Health. Many diseases linked to DW consumption are due to viruses. The aim of this review was to describe virus presence detected using molecular methods in sources for DW production and in DW. Four water types were considered: surface water used for DW production (SW-D), groundwater used for DW production (GW-D), water used for human consumption (DW) and bottled water (BW). The considered viruses were human pathogens; moreover plant pathogens proposed as novel viral indicators were presented. Studies published in the last 10 years were analysed and 79 articles were included in the review. Regarding virus occurrence in SW-D, GW-D, DW, high percentages of positive samples were reported for adenovirus, polyomavirus and pepper mild mottle virus. The most searched viruses were adenovirus, enterovirus, norovirus GI/GII and rotavirus. These viruses were frequently detected in SW-D, while they were rarely found in GW-D, suggesting that GW may be safer as a DW source. These viruses were detected also in DW, posing a possible threat for human health. Considering global occurrence, the lowest percentages of positive samples were found in Europe, while the highest percentages in Asia and South America. Only three articles assessed viruses in BW. Considering detection methods, filtration was the most applied concentration method, while nucleic acid extraction and molecular detection were generally performed using spin columns with silica membrane and quantitative PCR respectively. This review highlighted some critical issues such as method standardization lack and need for legislation updates.			
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February 22nd, 2022

Dear Editor,

We send the review "Virus occurrence in sources for drinking water production and in drinking water: a review" by Marco Panizzolo, Marta Gea, Elisabetta Carraro, Giorgio Gilli, Silvia Bonetta, Cristina Pignata on JOURNAL OF ENVIRONMENTAL SCIENCES.

Many studies showed that microbial water quality used for human consumption is critical for Public Health. Outbreaks linked to the consumption of contaminated or improperly treated water have been reported all over the world. Among all the waterborne pathogens, viruses are of major concern. Indeed, they can induce gastroenteritis through the faecal-oral route and, due to their peculiar characteristics compared to other pathogens, they are not efficiently removed by drinking water treatments. Virus presence in drinking water is among the main causes of death in developing countries and it induces a consistent percentage of drinking water cycle, from wastewater to drinking water, has a key role for human health. However, a complete overview of viral occurrence in sources for drinking water production and in drinking water from all over the world is still lacking. The aim of the submitted review was to describe available data about virus occurrence in sources

for drinking water production and in drinking water using molecular methods. Water types considered were: surface water used for drinking production, groundwater used for drinking water production, water used for human consumption (drinking water) and bottled water. Two virus types were considered: human pathogens and plant pathogens proposed as novel viral indicators. Scientific studies published in the last 10 years from all over the world were analyzed and 79 articles were finally included in the review.

We believe that the paper fits the aims and scope of the Journal, specifically, fits the following subjects:

- Aquatic environments
- Environmental microbiology

The study does not involve human subjects. All of the authors have read and approved the paper and it has not been published previously nor is it being considered by any other peer-reviewed journal. All authors are aware of and accept responsibility for the manuscript. All figures and tables were produced by the authors. Lastly, all authors declare no conflicting interests.

Hoping that the manuscript may fulfil the scientific standards of JOURNAL OF ENVIRONMENTAL SCIENCES, our best regards.

Marta Gea and Co-authors



UNIVERSITÁ DEGLI STUDI DI TORINO DIPARTIMENTO DI SCIENZE DELLA SANITA' PUBBLICA E PEDIATRICHE DEPARTMENT OF PUBLIC HEALTH AND PEDIATRICS DIRETTORE: PROF. SSA CARLA MARIA ZOTTI

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Virus occurrence in sources for drinking water production and in drinking water: a

review

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Highlights (max 5, max 85 characters, including spaces, per bullet point)

- 1. Adenovirus, enterovirus, norovirus, rotavirus were the most searched viruses
- 2. High % of positive samples for adenovirus, polyomavirus, pepper mild mottle virus
- 3. Viruses frequently detected in surface water, rarely in ground and drinking water
- 4. Viruses more detected in drinking water from Asia/South America than from Europe
- 5. Detection usually performed with filtration (negative filter) and quantitative PCR





UNIVERSITÁ DEGLI STUDI DI TORINO DIPARTIMENTO DI SCIENZE DELLA SANITA' PUBBLICA E PEDIATRICHE DEPARTMENT OF PUBLIC HEALTH AND PEDIATRICS

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Virus occurrence in sources for drinking water production and in drinking water: a review

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2	water: a review
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19	ABSTRACT (max 250 words)
20	Microbiological quality of drinking water (DW) is crucial for Public Health. Many diseases
21	linked to DW consumption are due to viruses. The aim of this review was to describe virus
22	presence detected using molecular methods in sources for DW production and in DW. Four
23	water types were considered: surface water used for DW production (SW-D), groundwater
24	used for DW production (GW-D), water used for human consumption (DW) and bottled water
25	(BW). The considered viruses were human pathogens; moreover plant pathogens proposed
26	as novel viral indicators were presented. Studies published in the last 10 years were
27	analysed and 79 articles were included in the review.

28 Regarding virus occurrence in SW-D, GW-D, DW, high percentages of positive samples were reported for adenovirus, polyomavirus and pepper mild mottle virus. The most searched 29 30 viruses were adenovirus, enterovirus, norovirus GI/GII and rotavirus. These viruses were 31 frequently detected in SW-D, while they were rarely found in GW-D, suggesting that GW may be safer as a DW source. These viruses were detected also in DW, posing a possible threat 32 33 for human health. Considering global occurrence, the lowest percentages of positive samples 34 were found in Europe, while the highest percentages in Asia and South America. Only three 35 articles assessed viruses in BW. 36 Considering detection methods, filtration was the most applied concentration method, while 37 nucleic acid extraction and molecular detection were generally performed using spin columns

- 38 with silica membrane and quantitative PCR respectively.
- This review highlighted some critical issues such as method standardization lack and needfor legislation updates.
- 41

42 Keywords (max 6): drinking water, enteric virus, human health, microbial water quality,

- 43 molecular methods, surface water.
- 44
- 45 **Abbreviations:**
- 46 AdV = adenovirus
- 47 AiV = aichivirus
- 48 AstV = astrovirus
- 49 BW = bottled water (water used for human consumption)
- 50 DW = drinking water (water used for human consumption, not bottled)
- 51 EPA = Environmental Protection Agency
- 52 EV = enterovirus
- 53 GW = groundwater
- 54 GW-D = groundwater used as a source for DW production

- 55 HAV = hepatitis A virus
- 56 HEV = hepatitis E virus
- 57 NoV = norovirus
- 58 PMMoV = pepper mild mottle virus
- 59 PyV = polyomavirus
- 60 RoV = rotavirus
- 61 SW = surface water
- 62 SW-D = surface water used as a source for DW production
- 63 SaV = sapovirus
- 64 TMV = tobacco mosaic virus
- 65 TTV = torque teno virus
- 66 PCR = polymerase chain reaction
- 67 qPCR = quantitative PCR
- 68 QMRA = Quantitative Microbial Risk Assessment
- 69 WSP = Water Safety Plan
- 70

71 **Contents:**

- 72 1. INTRODUCTION
- 73 2. SEARCH CRITERIA
- 3. VIRUS TYPES AND CHARACTERISTICS IN SOURCES FOR DW PRODUCTION
- 75 AND IN DW
- 76 3.1 Human pathogens
- 3.2 Plant pathogens proposed as novel viral indicators
- 4. VIRUS OCCURRENCE IN SOURCES FOR DW PRODUCTION AND IN DW
- 79 4.1 Virus occurrence in SW-D, GW-D, DW
- 80 4.2 Comparison of virus occurrence among the water types
- 4.3 Comparison of virus occurrence in DW among continents
- 4.4 Virus occurrence in BW

- 5. METHODS FOR VIRUS CONCENTRATION AND DETECTION IN SOURCES FOR
- 84 DW PRODUCTION AND IN DW
- 85 5.1 Virus concentration methods
- 86 5.2 Virus detection methods
- 6. CONCLUSIONS
- 88 7. FUNDING
- 89 8. ACKNOWLEDGEMENTS
- 90 9. DECLARATION OF INTERESTS
- 91 10. REFERENCES
- 92

93 **1. INTRODUCTION**

Fresh water is an essential resource for life on our planet and just a small part is accessible
because the most one is present in aquifers or in form of ice. Water scarcity increases and
so its reuse is essential (Cocoran et al., 2010). Moreover, water quality as well as water
quantity is important. Nowadays, 20% of world's population have no access to drinking water
(DW) and 40% suffer the consequences of improperly treated water (Cocoran et al., 2010;
Seelen et al., 2019).

100 Microbiological water quality used for human consumption is considered by many studies to

101 be critical for Public Health. Outbreaks linked to the DW consumption have been reported

102 worldwide and the main causes for contamination were identified as intrusion of animal

103 faeces or wastewaters due to heavy rain in groundwaters (GWs), wastewaters discharge into

the DW source, malfunctioning of the disinfection equipment at DW treatment plants and

105 cross-connections, pipe breaks and wastewater intrusion into the distribution system (Ligon

and Bartram, 2016; Moreira and Bondelind, 2017). Waterborne outbreaks were reported, for

107 examples, in China (Shang et al., 2017; Xue et al., 2014), Denmark (Van Alphen et al.,

108 2014), Albania (Donia et al., 2011), Spain (Blanco et al., 2017), Switzerland (Breitenmoser et

al., 2011), Italy (Giammanco et al., 2014), Philippines (Rebato et al., 2019), India (Tripathy et

al., 2019) and United States (Beer et al., 2015). In these studies, through retrospective

investigations and environmental analyses, pathogenic viruses were hypothesised aspossible causative agents.

Viruses can induce viral gastroenteritis through the faecal-oral route. In developing countries diarrhoeal diseases due to virus presence in DW are one of the main causes of death (Fayomi et al., 2019; WWAP, 2017). Similarly, in high income countries data concerning DW outbreaks show that in 2013-2014 7% of outbreaks in the United States were caused by viruses (CDC, 2021), while in the European Union Member States in 2019 most of DW outbreaks with strong-evidence were related to norovirus (NoV) and other calicivirus (ECDC, 2021).

Viruses are naturally present in environmental matrices such as in water where their 120 121 presence can be promoted by the discharge of not properly treated wastewater (Gibson et 122 al., 2011; Masciopinto et al., 2019; Okoh et al., 2010; Upfold et al., 2021). Moreover, several 123 studies have shown that DW treatments do not always succeed in removing viruses (Kato et al., 2018; Salvador et al., 2020; Ye et al., 2012); therefore, detection of viruses at all phases 124 125 of the integrated water cycle (from wastewater to DW) has a key role for human health. 126 In literature, virus presence within wastewaters have been investigated by many reviews 127 (Bhatt et al., 2020; Corpuz et al., 2020; Foladori et al., 2020; Sano et al., 2016), whereas virus occurrence in DW has been considered by just few reviews which were focused on DW 128 129 treatment systems and DW related outbreaks (Chen et al., 2021; Moreira and Bondelind, 130 2017). An overview of viral presence in water used as a source for DW production and in DW 131 is still lacking. Consequently, the aim of this review is to report the recent available knowledge about virus occurrence in sources for DW production and in DW. Water types 132 considered were surface water used for DW production (SW-D), GW used for DW production 133 134 (GW-D), DW and bottled water (BW). Moreover, two virus types were considered: human pathogens and plant pathogens proposed as novel viral indicators. Scientific studies 135 published in the last 10 years from all over the world were analysed and data of virus 136 137 presence assessed using molecular methods were summarized and discussed. In addition, 138 virus characteristics, concentration methods, nucleic acid extraction and molecular detection

- techniques reported in these studies were detailed. To the best of our knowledge, this is the
 first review that describes the most recent data on the worldwide virus occurrence in water
 used as sources for DW production and in water used for human consumption (DW).
- 142

143 **2. SEARCH CRITERIA**

144 In order to find information about the virus presence in sources for DW production and in 145 DW, a literature search was performed in PubMed, Scopus and Web of Science. These 146 databases were selected to be the most relevant and used for research on environmental topics. The search terms "virus" and "presence" or "detection" were combined with "drinking 147 148 water" or "bottled water" or "mineral water". Article search was set in the last 10 years and 149 were chosen only articles published between 2011 and 2021. The search gave 798 results in PubMed, 379 results in Scopus and 367 results in Web of Science (total = 1,544 results). 150 Two authors of the review independently screened the 1,544 publications. Using PRISMA 151

approach, 79 articles were finally included in this review (Fig. 1).

153 The search was limited to environmental monitoring articles that analysed human pathogens

154 or plant pathogens proposed as novel viral indicators. The articles were included when they

were written in English and met the following criteria: i) the analysed water type was water

used as a source for DW production or water used as DW, ii) origin of sources for DW

157 production was reported (surface water-SW or GW), iii) the detection of viruses was

158 performed using molecular methods, iv) viruses were not spiked intentionally into the

samples, v) data could be extrapolated for each viral agent and for each water type.

160

3. VIRUS TYPES AND CHARACTERISTICS IN SOURCES FOR DW

162 **PRODUCTION AND IN DW**

163 Viruses are obligatory intracellular parasites able to spread and be environmentally

transmitted through air, inert surfaces or waters. One of the main vehicles of viral

transmission is water through faecal-oral route. Inevitably, all water types can be subject to

166 contamination starting with SW (rivers, lakes), GW (wells, springs) and finally the seas and

oceans. Therefore, it is essential to study virus's resistance within these matrices (Pinon and
Vialette, 2019; Shoham et al., 2012).

169 Without host cells, viruses may decrease in number or remain stable. Their reduction can 170 occur depending on the water type (SW, GW, DW) and on various environmental conditions such as temperature, sunlight (UV) and disinfection products (chlorine and derivatives). 171 Water type influences the persistence of viral agents. In fact, GW, unlike SW, is a more 172 173 stable environment with few changes in chemical and physical parameters over time, and 174 thus constitutes a favourable matrix (Espinosa et al., 2008). Temperature is one of the most 175 studied environmental condition and has been recognised as the most influential factor that affect viral persistence (Espinosa et al., 2008; Pinon and Vialette, 2019). 176 177 In general, it has been shown that most viral agents can survive for years at low 178 temperatures, whereas at higher temperatures viruses are reduced within a few days. In 179 some studies, resistance to different temperature ranges was tested for certain viral agents 180 in different water matrices. A reduction of 5 log units of polioviruses and echoviruses was 181 found after one month in ocean water at around 21-26°C, while the same reduction was seen 182 after over 2 months when ocean water had temperatures between 4 and 16°C. Moreover, in mineral waters 1 log unit decrease of poliovirus and hepatitis A virus (HAV) was 183 184 demonstrated in about 11 months at 4°C, in contrast, the same reduction at 23°C took about 185 1 month (Pinon and Vialette, 2019). Finally, Ogorzaly et al. (2010) study, carried out on DW 186 and GW, showed a decrease of 2-1 log units of astrovirus (AstV) and adenovirus (AdV) with 187 increasing water temperature from 4 to 20°C in half to a third of the time, respectively. Another important factor that significantly influences viral viability is sunlight (UV). It is well 188 189 known that UV promotes a significant reduction in viral particles, indeed the study by 190 Flannery et al. (2013) showed a reduction of 1 log unit after 4 hours exposure to light 191 simulating winter conditions (10°C), while the same reduction was induced by 15 minutes exposure to light simulating summer conditions (17°C). UV effectiveness is also confirmed by 192 Garver et al. (2013) study that showed a reduction of 2-3 logs in 3 hours in deep water (less 193 UV) compared to 3-4 logs in 1.5 hours in superficial water (more UV). 194

Other factors responsible for viral reduction may be the presence of disinfectants, pH
extremes, or heavy metals (Pinon and Vialette, 2019).

197 In parallel to the environmental factors, some viral characteristics can affect viral survival 198 such as aggregation tendency, genome type and capsid composition. Actually, under 199 adverse conditions, viruses tend to aggregate with each other and with organic matter to 200 form aggregate suspension material, making their elimination difficult. Indeed, disinfectants 201 are unable to penetrate the viral capsid so merely bind to the biofilm of organic matter in the 202 aggregates (Espinosa et al., 2008). Moreover, viruses are characterized by a different 203 persistence according to their genome type. For example, due to its double-stranded DNA 204 genome, AdV showed greater stability in water than enterovirus (EV) which has RNA 205 genome (Mena and Gerba, 2008). Due to the higher persistence in the environment, AdV 206 has been suggested as a possible indicator of the viral contamination water (Verani et al., 207 2019). Finally, a comparison of rotavirus (RoV) and AstV, both of which are RNA viruses, 208 showed that RoVs are more persistent in GW, as they have a triple-layer capsid and a 209 double-stranded RNA genome, whereas AstV have a single-layer capsid and a single-210 stranded RNA genome (Espinosa et al., 2008). In the 79 included articles, virus analysis in sources for DW production and in DW was 211 212 mainly focused on the detection of two virus types namely human pathogens and indicators. 213 The first viruses are able to infect hosts causing diseases, while the second ones are used

as indicators of faecal contamination. The articles included in this review mainly investigated

the following viruses: AdV, HAV, EV, aichivirus (AiV), hepatitis E virus (HEV), sapovirus

216 (SaV), NoV, torque teno virus (TTV), RoV, AstV, polyomavirus (PyV), pepper mild mottle

217 virus (PMMoV) and tobacco mosaic virus (TMV). The characteristics of these viruses are

219

218

220 3.1 Human pathogens

described in Table 1.

The integrated water cycle plays a key role in reduction and elimination of pathogenic
viruses, decreasing their spread in environment. Wastewater treatments are often unable to

223 eliminate viruses and fail to prevent their release in the environment; moreover, pathogenic viruses could be also resistant to DW treatments threatening Public Health. Among the 224 225 human pathogens, enteric viruses are obligate parasites that infect and replicate within the 226 human gastrointestinal tract (Upfold et al., 2021). Depending on the type of viral agent 227 colonising the gastrointestinal tract, different possible diseases may occur. Enteric viruses 228 are one of the main causes of waterborne diseases transmissible via the faecal-oral route so 229 their detection in water used as DW source or in DW is of crucial importance in order to 230 assess the risk for human health.

231

3.2 Plant pathogens proposed as novel viral indicators

233 Currently the monitoring of microbial water quality is generally performed quantifying 234 bacterial indicators of faecal contamination. Since they are easy to identify, more present and 235 resistant than pathogens, bacterial indicators are used to assess the faecal contamination 236 and to estimate the presence of microbial pathogens. However, numerous studies showed 237 that the concentration of indicator bacteria is not related to the concentration of pathogenic 238 viruses, suggesting that these indicators are unsuitable to define the presence of human viral 239 pathogens in water (Liang et al., 2015). Indeed, environmental conditions affect differently 240 bacteria and viruses (Kitajima et al., 2018).

241 Bacteriophages (e.g. coliphages) have been proposed as alternative indicators instead of 242 bacteria. Coliphages are viruses that infect Escherichia coli and other coliforms (Leclerc et 243 al., 2000). Environmental transport and survival of coliphages is similar to enteric viruses. However, coliphages show a greater persistence than human enteric viruses in environment 244 245 since their replication in bacterial hosts can continue after being shed in faeces. In addition, 246 only a small percentage of human or animal faecal samples test positive for coliphages so these viruses may be too sparse to be detected in some environmental waters (Griffin et al., 247 248 2008). Therefore, other viruses were suggested by the scientific community as possible viral indicators of faecal contamination. In particular, two plant pathogens were proposed as 249

250 alternative viral faecal indicators: PMMoV and TMV (Kitajima et al., 2018; Tandukar et al., 2020a). These two viruses were analysed by some research articles included in this review. 251 252 PMMoV is a plant pathogen globally distributed that causes significant economic and crop 253 losses worldwide (e.g. in the United States, Japan and China). Its presence may be 254 indicative of faecal contamination because is the most abundant virus type in human faecal samples (Kitajima et al., 2018). However, its application as a viral indicator has limitations, 255 256 since studies show conflicting results on the correlation between concentrations of this virus 257 and concentrations of human enteric viruses (Kitajima et al., 2018; Tandukar et al., 2020a). As PMMoV, TMV is a plant pathogen. TMV was discovered in the 19th century when a new 258 infection was affecting tobacco plants causing characteristic patterns, such as mosaic-like 259 260 mottling and discoloration on the leaves (Tandukar et al., 2020a). Similarly to PMMoV, this 261 virus is excreted by a large proportion of healthy people.

262 PMMoV and TMV are widely distributed in SW, in GW and even in DW. They are used as

263 indicators of faecal contamination in wastewater, SW and also in DW because their presence

is high in human faeces and in sewage. In the analysed articles the presence of these

viruses was studied in sources for DW production and in DW (Haramoto et al., 2013; Kuroda

266 et al., 2015; Tandukar et al., 2020a, 2018).

267

4. VIRUS OCCURRENCE IN SOURCES FOR DW PRODUCTION AND IN DW

269 4.1 Virus occurrence in SW-D, GW-D, DW

270 In the 79 included articles, water samples coming from all over the world were analysed (Fig.

271 2). In particular, 24 articles analysed samples from Asia, 17 from South America, 13 from

- Africa, 13 from Europe, 11 from North America and 1 from Oceania (79 total articles). The
- articles assessed virus occurrence in three different water types.
- SW-D was analysed by 43 articles, SW-D was collected from rivers, estuarine bays,
 dams, lagoons, ponds, lakes and other reservoirs.
- GW-D was analysed by 17 articles. GW-D was collected from wells and springs.

- DW was analysed by 45 articles. These articles analysed different water types (e.g.
 tap water, DW treatment plant effluents, SW used as DW without any treatment).
- 279 The mean of sample volume analysed was significantly different according to the water types

280 (Kruskal-Wallis test followed by pairwise comparisons, SW-D vs GW-D, SW-D vs DW, GW-D

281 vs DW, p<0.05). Mean values were 43.58 ± 114.83 L ranging from 0.050 L to 2340 L for SW-

282 D, 321.91 ± 407.99 L ranging from 0.250 L to 1783 L for GW-D and 242.05 ± 467.26 L

ranging from 0.050 L to 3400 L for DW. In particular, volumes were higher for GW-D/DW

samples than SW-D probably because a lower viral presence was expected.

Table 2 presents the cumulative percentages of positive samples for each viral agent (total

286 positive samples/total samples). For NoV the cumulative percentages were calculated

287 dividing data according to the viral subtype (cumulative percentages were calculated

independently for NoV GI, NoV GII, NoV GIII, NoV GIV).

The percentages of positive samples were compared among virus types. For some viral

290 types a small number of samples was analysed, therefore the percentages could not reflect

the real occurrence of these viruses in sources for DW production and in DW. For this

reason, comparison was performed considering viruses that were analysed in at least 100

samples. The highest percentages of positive samples were found for PMMoV (85.31%),

AdV (52.61%), PyV (44.25%), AiV (43.86%), RoV (41.81%) in SW-D, for PMMoV (5.94%),

295 RoV (4.88%), AdV (2.21%), PyV (0.97%), EV (0.78%) in GW-D, for PMMoV (28.33%), AdV

296 (15.96%), PyV (12.94%), AiV (11.27%), EV (7.73%) in DW.

AdV and PyV were among the human pathogenic viruses that showed the highest

298 percentages of positive samples. This result can be explained considering that AdV and PyV

are characterized by a DNA genome which is generally more stable in the environment and

300 less affected by the physico-chemical treatments applied to obtain DW with respect to RNA

301 genome (Ye et al., 2012).

302 For the three water types, the percentages of PMMoV positive samples were the highest

303 compared to the percentages of the other viruses. This finding is interesting since PMMoV

has been proposed as a possible viral indicator of human faecal contamination in several

305 studies (Kitajima et al., 2018; Zhang et al., 2006). Indeed, the high percentages of positive 306 samples found in SW-D, GW-D and DW confirm that this virus is more persistent in water 307 than other enteric viruses, including AdV and PyV (Hamza et al., 2011; Haramoto et al., 308 2013), suggesting that it could be an excellent candidate as an indicator and it could be used 309 as a possible process control to measure the removal of enteric viruses during water 310 treatments (Symonds et al., 2018). The higher presence of this virus with respect to human 311 pathogenic viruses could also be due to the fact that, while other enteric viruses are more 312 abundant in water when there is an increase of infected individuals, PMMoV presence seems 313 not to be characterized by seasonal variations (Haramoto et al., 2013).

Virus detection in SW-D, GW-D, DW is reported divided by reference in Table S.1, S.2, S.3,
respectively.

316

4.2 Comparison of virus occurrence among the water types

318 The percentages of positive samples were compared among the water types. The

319 comparison was performed considering viruses that were searched in the highest number of

320 samples and that were analysed by most articles (i.e. the most searched viruses for number

321 of total samples and number of total articles). This choice was adopted to compare data that

322 could reflect the real virus occurrence and could be considered representative of the global

323 situation. As can be seen in Table 2, AdV, EV, NoV GI, NoV GII and RoV were most

324 searched viruses in all water types. These viruses were the most searched probably because

are important foodborne pathogens (Koopmans and Duizer, 2004).

In Fig. 3 are reported the percentages of positive samples in SW-D, GW-D and DW of these

327 viruses. As can be seen, all the five viruses were frequently detected in SW-D. This result

328 could be explained considering that these enteric viruses are excreted in large quantities in

the faeces of infected individuals (symptomatic and asymptomatic), which are conveyed to

330 sewage treatment plants. Since the water treatments of these plants can be not efficient to

remove all viruses, they may be released into SW (Bhatt et al., 2020). Moreover, the high

332 presence of enteric viruses in SW is not only due to municipal wastewaters but may also

333 result from livestock slurry from livestock farms, which are sometimes not conveyed to the wastewater treatment plants but directly discharged in SW (Haramoto et al., 2018). 334 335 Among the water types, the percentages of positive samples in GW-D samples were the lowest. This result suggests that GWs are more protected from possible sources of 336 contamination, making them safer when they are used to produce DW. Nevertheless, GW, if 337 338 not properly protected, are susceptible and can easily be polluted from some contamination 339 sources. After a period of heavy rainfall, GW located in proximity to livestock farms can be 340 contaminated by livestock slurry leaching into the ground or due to damage or deficiency of 341 pipes conveying wastewater effluents to the plants (Gibson and Schwab, 2011a; Gotkowitz et al., 2016). Percentages of positive samples in GW-D were also lower than in DW. This 342 343 result is not surprising considering that the DWs include not only treated GWs but also 344 treated SWs.

The percentages of positive samples were higher in SW-D than in DW. This may be

346 attributable to the fact that DW are generally treated with physico-chemical processes which

can reduce viral presence in this water type (Asami et al., 2016; Atabakhsh et al., 2019;

Jacob et al., 2015; Kato et al., 2018; Tandukar et al., 2020b; Ye et al., 2012).

Even if at lower percentages compared to SW-D, the five enteric viruses were detected also 349 350 in DW. Since high percentages of positive samples in water used for human consumption 351 may be a source of risk to the population, the presence of these viruses in DW might pose a 352 possible threat to human health. Indeed, the ingestion of water contaminated by enteric 353 viruses can lead to sporadic episodes of viral gastroenteritis, which, if not treated with appropriate care, could lead to death in children (Wang et al., 2016). It is important to 354 355 highlight that in this review were presented only data of virus presence analysed using 356 molecular methods; therefore, the percentages of positive samples do not necessarily mean 357 that these samples contain active and pathogenic viruses but only that in these samples the viral genomic material was detected (Rachmadi et al., 2016). Indeed, many studies 358

359 compared virus infectivity and virus detection using molecular methods in water samples and

found that samples in which viral genomes were detected did not always contain infectious
viral particles (laconelli et al., 2017; Salvador et al., 2020).

362

4.3 Comparison of virus occurrence in DW among the continents

Considering virus detection in DW samples, the percentages of positive samples were 364 365 compared among the continents. As for the comparison among the water types, the 366 comparison was performed considering the most searched viruses for number of total 367 samples and number of total articles (see paragraph 4.2). In Fig. 4 are reported the 368 percentages of positive sample in DW samples divided according to continents. It should be noted that the number of studies is not the same across continents. Indeed, there are fewer 369 370 studies in Europe than in the other continents. The global distribution of the samples is 371 probably not homogenous because in some continents such as Europe the risk associated 372 with water consumption is not considered a major health concern, so the research articles 373 focused on this topic are limited. On the contrary, in developing countries diseases 374 associated with water consumption are a major issue, thus this research topic is more 375 investigated. Comparing the percentages among the continents, except for RoV, the lowest 376 percentages of positive samples were found in Europe. In contrast, the highest percentages 377 of positive samples were found in Asia and South America.

378 The different virus occurrence in Europe with respect to Asia and South America could be 379 due to several factors. Indeed, in developing countries the quality of sources for DW 380 production could be lower due to a higher discharge of not properly treated wastewaters; moreover, technologies used for DW treatment could be less efficient in virus removal. 381 382 Finally, water distribution networks could be less monitored and more prone to breakdowns 383 that may cause the intrusion of contaminated water in DW distribution systems. Regarding 384 RoV, the unexpected percentage of positive samples in Europe could be explained considering that only one study carried out in Slovenia assessed RoV occurrence in 385 386 European DW (Steyer et al., 2011), so this percentage could be not representative of the 387 whole European occurrence of this virus.

388 The results obtained analysing African samples are interesting. Indeed, although the number of articles is higher than in Europe and the articles analysed samples coming from different 389 390 African countries (giving a complete picture of virus occurrence throughout the continent), the 391 percentages of positive samples were less than 11%. These percentages seem to be too low 392 if compared with the incidence of viral gastrointestinal diseases transmitted by water 393 consumption in this continent, so further studies are needed to clarify this discrepancy. 394 Finally, considering the occurrence of viruses in North America, the percentages of positive 395 samples were quite low suggesting that the DW quality is quite good in this continent.

396

397 **4.4 Virus occurrence in BW**

398 Three articles assessed the virus presence in BW (Da Silva Luz et al., 2020; Dos Santos et 399 al., 2015; Kuroda et al., 2015). The analysed BW was produced using GW as source (water 400 from wells/springs) and samples were collected in Brazil and Vietnam from bottles containing 401 different water volumes (0.5, 1.5, 19, 20 L). The analysed water volume ranged from 0.5 L to 402 100 L, while the percentages of positive samples ranged from 81.69% (AdV) to 0% (PMMoV, 403 NoV GII, AiV). It's important to highlight that these percentages of positive samples were 404 calculated considering only three articles which analysed few samples collected in Brazil (2 405 articles) and in Vietnam (1 article). The limited number of articles on virus occurrence in BW 406 is probably due to the low frequency of outbreaks linked to the consumption of BW. However, 407 since also in this water type genomes of some viruses were found (e.g. AdV, EV, NoV, RoV), 408 more studies to assess the real virus occurrence in BW are needed. Virus detection in BW is 409 reported in Table S.4.

410

411 5. METHODS FOR VIRUS CONCENTRATION AND DETECTION IN SOURCES

412 FOR DW PRODUCTION AND IN DW

413 **5.1. Virus concentration methods**

In the analysed studies, different methods/methodologies of concentration, extraction and
identification of viral particles were reported. Many of these concentration methods were

established in the 1980s and have not been changed. They include the use of negatively and
positively charged membranes, glass wool filters and ultrafiltration. These methods can be
applied alone or can be followed by a secondary concentration that allows for a higher
concentration of the treated water sample (Ikner et al., 2012).

420 Concentration methods used in the analysed articles are described below.

421 Adsorption-elution method, also known as filtration method, is based on the 422 absorption of organisms on a solid membrane utilising the ionic properties of the micro-organisms (bacteria/viruses) to be concentrated. The filters mainly used in this 423 technique can be membrane filters (cellulose) or glass filters; moreover, filters could 424 be with neutral charge or could have electropositive/electronegative charge using 425 electrostatic forces to concentrate viruses (Cashdollar and Wymer, 2013; Ikner et al., 426 427 2012). The adsorption phase (with filters) is followed by an elution phase using a specific fluid which is variable according to the analysed virus type (Cai et al., 2015; 428 Ruhanya, 2016). For instance, the Environmental Protection Agency (EPA) has 429 430 proposed a procedure to detect human enteric viruses in water whose first step is 431 based on adsorption-elution method (i.e. filtration through electropositive filters, 432 followed by elution using a solution of glycine and beef extract) (Fout et al., 2015).

Tangential flow filtration system consists in flowing the liquid parallel to the filtering
 medium to reduce the probability of clogging of the latter and thus enhance its filtering
 capacity. This method is still used today to concentrate micro-organisms present in a
 matrix (e.g. water). It is essential to adopt an appropriate membrane according to the
 type of microbial agent researched (Cai et al., 2015). In the analysed studies, 30 kDa
 and 100 kDa filter membranes were used.

Ultrafiltration is commonly used as water treatment technology for the removal of
 human pathogens and can be considered as a special form of filtration that uses
 positive pressure to promote the flow of water through a membrane (Reeve et al.,
 2016). This method allows to retain not only particles and macromolecules but also

443 micro-organisms such as viruses and bacteria. The membranes used in ultrafiltration 444 process have pores with diameters ranging from 1 to $10^{-3} \mu m$ (Shao et al., 2011).

Polyethylene glycol is a biocompatible polymer used for protein precipitation. Its
 properties promote virus precipitation sequestering water molecules from the outer
 layer of their pericapsids/capsids to promote virus-virus interactions and thus virus
 concentration (Corpuz et al., 2020).

Skimmed milk flocculation is based on three physical processes, i.e. adsorption,
 sedimentation and dissolution. The first two steps consist in the adsorption of viruses
 on pre-flocculated skimmed milk proteins and precipitation of flakes with adsorbed
 viruses. After sedimentation, sediment is dissolved using a buffer solution. This
 methodology does not require the use of special equipment and long processing
 steps, making its use advantageous (Corpuz et al., 2020).

455 In the 79 articles analysed, 7 different primary concentration methods were used, which were or were not followed by other 4 types of secondary concentration methods for a total of 17 456 different combinations of primary-secondary concentration methods (Table 3). The most 457 frequently used combinations of primary-secondary methods were filtration with negatively 458 459 charged membranes (applied in 17 articles), filtration with negatively charged membranes followed by a secondary concentration using ultrafiltration (applied in 11 articles), filtration 460 with positively charged membranes (applied in 10 articles), filtration with positively charged 461 membranes followed by a secondary concentration with polyethylene glycol (applied in 7 462 articles). The other methods were reported in less than 6 articles. The filtration with 463 464 negatively charged membranes was the most applied method probably because it has 465 numerous advantages. Indeed, this method is cheap and it allows high recoveries for 466 viruses. Moreover, since electronegative filters are less influenced by clogging, this method 467 is suitable also for turbid waters (Cashdollar and Wymer, 2013).

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471 **5.2 Virus detection methods**

472 Virus detection can be performed using both cell culture and molecular methods. Cell culture 473 methods are based on virus ability to grow in cell cultures causing morphological changes. 474 Viruses are inoculated on cell monolayers and the number of infectious viruses is quantified counting the plaque forming units (cell destruction areas caused by viruses) (Gerba et al., 475 476 2017). Cell culture methods are able to quantify viruses that potentially can replicate in 477 humans causing the disease. Before the development of molecular methods, virus detection 478 in environmental matrices was exclusively performed using these methods (Gerba and 479 Betancourt, 2019). However, the main disadvantage of cell culture methods is that they can not be applied to analyse all virus types because some of them, such as NoV, can not 480 replicate in cell culture (Fout et al., 2015); therefore, in order to assess the presence of these 481 482 viruses in environmental matrices, only molecular methods can be applied. Moreover, these 483 detection methods seem to underestimate virus concentration at least by 2-3 orders of 484 magnitude (Chen et al., 2021). The underestimation could be due to viral aggregation; 485 indeed, many aggregated viruses can form a single plaque forming unit so they can be 486 counted as one infectious viral particle. Moreover, one group of viruses may grow faster than 487 another or interfere with the replication of another group of viruses, causing an underestimation of the viral particle number (Gerba and Betancourt, 2019). 488 489 Therefore, nowadays, virus detection in waters is generally performed using molecular 490 methods, which are based on the detection of viral genomes. Due to the low environmental 491 stability of genomes, especially for RNA viruses a positive molecular result indicates that viral 492 particles are intact; however, this result can be obtained also for viruses that have been 493 inactivated by chemical disinfection, heat or proteases (Kopecka et al., 1993). As a 494 consequence, one of the most important disadvantages of molecular methods is that they 495 can not distinguish between infectious and inactivated viruses. On the contrary, these 496 methods have numerous advantages. They are able to detect low virus concentrations so they are more sensitive than cell culture methods. In addition, they are characterized by high 497 498 specificity and rapidity. Finally, in contrast to cell culture methods, they potentially allow the

detection of all virus types, detecting also viruses that are hardly propagated using cell
cultures such as RoV and NoV (Carducci et al., 2003; Corpuz et al., 2020). Detection of viral
genomes is performed through extraction of nucleic acids followed by amplification of specific
nucleic acid fragments using polymerase chain reaction (PCR).

In the 79 analysed articles, different methods for nucleic acid extraction were reported. In
some articles more than one extraction type was used. The different methods are described
below.

Nucleic acids can be purified through the binding with silica membrane. The principle
 of this method is the following. DNA binds specifically to the silica-gel membrane,
 while contaminants pass through. Then unwanted materials are generally removed
 with washing steps and finally the remaining nucleic acids are eluted in either water
 or a buffer. This extraction type can be performed using both spin columns or vacuum
 columns. This method was the most applied for nucleic acid extraction. Indeed 71
 articles applied it using spin columns, while one using vacuum columns.

Magnetic beads separation is a method based on specific interaction between nucleic
 acids and magnetizable particles. Briefly, after a lysis step to release the nucleic
 acids, viral genomes bind to magnetizable particles in the presence of a binding
 buffer. The other molecules are washed with a water-based wash buffer and finally
 the nucleic acids are eluted in an elution buffer (Nargessi and Ou, 2010). This
 extraction type was applied by 13 articles.

Nucleic acids can be purified through the binding with glass fibre or glass powder. For
 example, nucleic acids can be immobilized through the binding to the surface of the
 glass fibre fleece in the presence of a chaotropic salt. Sample is mixed with a
 chaotropic salt and applied to the glass fibre fleece. Nucleic acids bind to the glass
 fleece, while contaminating substances are removed through washing steps. Nucleic
 acids are finally eluted in a small volume of low-salt buffer or water. Among the
 analysed articles, 4 applied this extraction method.

Organic extraction is a method that uses organic solvents. According to this method,
 samples are mixed with a reagent composed by a monophasic solution of guanidine
 thiocyanate and phenol. Then chloroform is added and the homogenate is allowed to
 separate into different phases containing RNA, DNA and proteins. The phases are
 separated and finally the nucleic acids are isolated through precipitation with organic
 solvents (e.g. isopropanol, ethanol). 4 included articles reported this extraction
 method.

533 After extraction of nucleic acids, the molecular detection of viruses is performed through the 534 amplification of specific nucleic acid fragments using PCR. For RNA viruses, viral genome is reverse transcribed through a reverse transcriptase-PCR before PCR to obtain the cDNA. 535 536 Molecular detection methods can provide both qualitative and quantitative data depending on the PCR type. Qualitative data can be obtained using conventional PCR or nested/semi-537 538 nested PCR (performed by two successive conventional PCR), whose products are 539 subjected to agarose gel electrophoresis (Corpuz et al., 2020). On the contrary, quantitative 540 data can be obtained using quantitative PCR (qPCR). Virus quantification can be affected by 541 some factors that can cause data variability. For example the quantification can be influenced by recovery efficiency of the applied extraction method, by PCR inhibitory 542

substances within the samples or by PCR conditions (number of replicates, primer/probe

design, thermal cycling conditions) (Gerba et al., 2018). In addition to providing quantitative

545 data, another qPCR advantage is that it has a high sensitivity, therefore it can detect even

small amounts of nucleic acids (Corpuz et al., 2020).

547 Regarding molecular detection in the 79 analysed articles, conventional PCR, qPCR, nested

548 PCR or semi-nested PCR were applied for the identification of viral particles. Overall, the

549 most applied detection method was qPCR (60/79 articles, 75.9%), followed by nested/semi-

nested PCR and conventional PCR (17/79 articles, 21.5% and 10/79 articles, 12.7%,

respectively). The use of qPCR was frequent, probably because it has a higher sensitivity

than the other molecular methods. The higher sensitivity was confirmed by Assis et al. (2015)

and Dos Santos et al. (2015) studies. These studies applied conventional PCR and qPCR to

detect the same virus type; the results showed that a higher number of positive samples was found using qPCR than using conventional PCR. The qPCR was frequently applied also because it can provide quantitative data. However, it is important to highlight that even if using qPCR, the number of genomic copies/L can be quantified, many of the included articles did not report virus concentrations. In some articles virus concentrations were not reported because data were under the quantification limit.

Methods applied for virus concentration, nucleic acid extraction and molecular analyses are reported in Table S.5 divided by reference, while data on detection limit/quantification limit are shown in Tables S1-S4. One of the main problems related to the monitoring of viruses in water is linked to the fact that different methods can be applied to detect them. Furthermore, even if quality assurance/quality control is important to assure data quality, in environmental monitoring studies, this information is often not reported.

566

567 **6. CONCLUSIONS**

The microbiological water quality for human consumption is crucial for Public Health. As long 568 as viruses are one of the most important causative agents of waterborne diseases, their 569 detection in sources for DW production and in DW has a key role in healthcare. In this 570 571 review, scientific studies of the last 10 years from all over the world were analysed in order to summarize data of virus presence assessed using molecular methods in sources for DW 572 production and in DW. Water types considered were SW-D, GW-D, DW and BW. 573 In the 79 articles finally included in the review different virus types were searched. However, 574 575 only some of them are important for Public Health because they may cause waterborne 576 outbreaks. Therefore, in order to collect more data in short times, the authors of this review believe that it could be more appropriate to focus research on these viruses (i.e. AdV, EV, 577 578 NoV GI, NoV GII, RoV). 579 As highlighted by this review, data on virus presence in water for human consumption are 580 very heterogeneous. This finding could be related to the methods used for virus detection;

581 indeed, different combinations of primary-secondary concentration methods and different

nucleic acid extraction methods were carried out. This evidence raises an important question
about a lack of standardization of methodologies for virus detection. It is not easy to compare
data collected using different methodologies and it would be desirable to standardise
methodologies in order to make data more comparable.

586 The comparison of virus detection among the water types showed that in SW-D viruses were

587 frequently detected, while the percentages of positive samples in GW-D were the lowest. It is

588 crucial to investigate viral presence in sources for DW production (SW-D and GW-D),

589 because a higher presence in SW and GW could lead to a higher presence in DW. In

590 particular, the assessment of virus occurrence in SW is important because the use of this

591 water as DW source will probably increase in the next years. Indeed, climate change and

592 global population growth will lead to more DW demand and less water availability.

593 Consequently, to produce DW it will be necessary to increase the use of sources most

vulnerable to contamination, such as SW.

595 Even if at lower percentages compared to SW-D, viruses were detected also in DW, where 596 they might pose a possible threat to human health. Although these percentages do not 597 necessarily mean that these samples contain viable pathogenic viruses (because they were found through molecular detection methods), this evidence suggests the need for regulatory 598 599 updates. Indeed, the monitoring of enteric viruses together with coliphages and phages, is 600 considered important for the assessment of DW treatments effectiveness. However, the only 601 parameter proposed by WHO guidelines for verification of microbial quality of DW is the 602 monitoring of Escherichia coli or thermotolerant coliform bacteria, whereas for viruses no 603 guidelines values have been proposed yet (WHO, 2017). Even the new European legislation 604 (The European Parliament and the Council of the European Union, 2020) requires only the 605 search for *E. coli* and fecal enterococci to establish the DW requirement for water intended 606 for human consumption. Clostridium perfringens and Legionella spp. must only be analysed on the basis of the risk assessment. Finally, the legislation provides for the search of somatic 607 608 coliphages in untreated waters if specifically indicated in the risk assessment.

609 As had already been proposed by the WHO (2004), an excellent way to take into account the risk associated with the presence of viruses in DW could be the application of the Water 610 611 Safety Plans (WSP) which have also been introduced by the new European legislation (The European Parliament and the Council of the European Union, 2020) and will be mandatory 612 for all Member States since 2029. It is an approach based on the risk assessment and 613 614 management throughout all the water supply chain, from catchment to consumer. The main 615 limitation derives from the fact that very often water companies do not have data on the 616 presence of enteric viruses from source to tap (Masciopinto et al., 2019; van den Berg et al., 617 2019) and therefore the risk assessment and management are based on the presence of the microbiological indicators required by the regulations, even if the correlation between E. coli, 618 619 faecal enterococci and bacteriophages and the presence of viruses is often absent or very 620 low (Edge et al., 2013; Goh et al., 2019; Lee et al., 2014; Payment and Locas, 2011). This 621 further strengthens the need to include the search for enteric viruses or alternative indicators 622 in the monitoring programs, in order to obtain objective data for the application of WSPs. 623 Considering the comparison of virus detection in DW among the continents, this review 624 showed that the number of studies is not homogeneously distributed across the continents. 625 Indeed, few studies have assessed DW collected in Europe while, to our knowledge, there is 626 no study on Oceanian DW. This finding highlighted the need to analyse additional samples 627 from these geographical regions. Moreover, the results obtained analysing African samples 628 were unexpected, so the authors believe that they are worth of further studies. Finally, the 629 bibliographic research performed in this review demonstrated that only three articles assessed virus presence in BW, underlining another research gap. 630 631 In order to estimate the potential human health risk due to virus exposure through DW, 632 reliable data on virus occurrence in this matrix are needed. However, virus concentration data in DW are still limited, so future studies are needed to fulfil this research gap. Besides 633 studies focused on viral occurrence in DW, future research should also investigate virus 634 distribution in other environmental matrices, such as SW and GW. Indeed, these data 635

together with DW data could be used as inputs to perform Quantitative Microbial Risk

Assessment (QMRA), allowing a more precise estimate of human health risk.

638

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- 641

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- 645

646 9. DECLARATION OF INTERESTS

- 647 The authors declare that they have no known competing financial interests or personal
- relationships that could have appeared to influence the work reported in this paper.
- 649

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1209 **Table captions**

- 1210 **Table 1.** Virus characteristics. AdV= adenovirus, HAV=hepatitis A virus, EV=enterovirus,
- 1211 AiV=aichivirus, HEV=hepatitis E virus, SaV=sapovirus, NoV=norovirus, TTV=torque teno
- 1212 virus, RoV=rotavirus, AstV=astrovirus, PyV=polyomavirus, PMMoV=pepper mild mottle virus,
- 1213 TMV=tobacco mosaic virus.
- 1214 **Table 2.** Total number of articles that investigated each viral agent and cumulative
- 1215 percentages of positive samples divided by type of water and type of virus. = not assessed;
- 1216 AdV = adenovirus; AiV = aichivirus; ASFLV = asfarvirus-like virus; AstV = astrovirus; CosV =
- 1217 cosavirus; EV = enterovirus; HAV = hepatitis A virus; HCV = hepatitis C virus; HEV =
- 1218 hepatitis E virus; KV = klassevirus; NoV = norovirus; PMMoV = pepper mild mottle virus; PyV
- 1219 = polyomavirus; ReV = reovirus; RoV = rotavirus; SaliV = salivirus; SaV = sapovirus; TMV =
- 1220 tobacco mosaic virus; TTV = torque teno virus.
- 1221 **Table 3.** Primary and secondary concentration methods reported in the analysed articles
- 1222 (total articles= 79, two concentration methods were applied by the study of Kuroda et al.,
- 1223 2015). TFF = tangential flow filtration; UF = ultrafiltration; F = filtration; F- = filtration with
- 1224 electronegative charged membrane; F+ = filtration with electropositive charged membrane;
- 1225 PEG = precipitation with polyethylene glycol; SMF = skimmed milk flocculation.
- 1226

1227 Figure captions

1228 **Figure 1.** Flow chart of study selection process.

1229 **Figure 2.** Number of articles that analysed SW-D, GW-D, DW or BW performed in each

- 1230 continent (Africa, Asia, Europe, Oceania, North America, South America). Total articles = 79
- 1231 (58 analysed one water type, 14 analysed two water types, 6 analysed three water types, 1
- analysed four water types). Satellite image from European Space Agency.
- 1233 Figure 3. Percentages of positive samples in SW-D, GW-D and DW of the most searched
- 1234 viruses (AdV, EV, NoV GI, NoV GII, RoV).
- 1235 Figure 4. Percentages of positive samples in DW of the most searched viruses (AdV, EV,
- 1236 NoV GI, NoV GII, RoV) divided by continent.









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Figure 4

 Table 1. Virus characteristics.

	Human pathogens					
Virus type	Family	Genome	Persistence in water supply ^a	Disease	Vaccine	Reference
AdV	Adenoviridae	double- stranded DNA	Long	pharyngitis, cystitis, gastroenteritis	yes	Broderick et al., 2017; Okoh et al., 2010; Upfold et al., 2021; Vellinga et al., 2005
AiV	Picornaviridae	single- stranded and positive-sense RNA	single- nded and NA gastroenteritis no tive-sense RNA		Upfold et al., 2021	
AstV	Astroviridae	single- stranded and positive-sense RNA	Long	gastroenteritis, respiratory diseases, encephalitis, meningitis, acute flaccid paralysis	no	Bosch et al., 2014; Upfold et al., 2021; Vu et al., 2017
EV	Picornaviridae	single- stranded and positive-sense RNA	Long	gastroenteritis, myocarditis, pericarditis, encephalitis	yes	Baggen et al., 2018; Li et al., 2021; Lugo and Krogstad, 2016
HAV	Picornaviridae	single- stranded and positive-sense RNA	Long	hepatitis	yes	Okoh et al. 2010; Smith and Simmonds, 2018; Upfold et al., 2021
HEV	Hepeviridae	single- stranded and positive-sense RNA	Long	hepatitis	yes	Larrue et al., 2021; Upfold et al., 2021
NoV	Caliciviridae	single- stranded and positive-sense RNA	Long	gastroenteritis	no	Carter et al., 2005; Okoh et al., 2010; Upfold et al., 2021
PyV	Polyomavirida e	double- stranded DNA	NA	progressive multifocal leukoencephalo pathy, nephropathy, pulmonary infections, possible oncogenic viruses	no	Calgua et al., 2013
RoV	Reoviridae	double- stranded RNA	Long	gastroenteritis	yes	Crawford et al., 2017; Okoh et al., 2010; Steele et al., 2003; Upfold et al., 2021
SaV	Caliciviridae	single- stranded and positive-sense RNA	Long	diarrhoea, nausea, myalgia	no	Upfold et al., 2021
TTV	Anelloviridae	single- stranded and negative- sense DNA	NA	partially unknown, co- factor in several diseases	no	Charest et al., 2015; Jiménez-Melsiò et al., 2013; Shoeib et al., 2011

Indicators				
Virus type	Family	Genome	Host	Reference
PMMoV	Virgoviridae	single- stranded and positive-sense RNA genome	plant	Haramoto et al., 2018; Kitajima et al., 2018
TMV	Virgaviridae	single- stranded and positive-sense RNA	plant	Tandukar et al., 2020a

AdV= adenovirus, HAV=hepatitis A virus, EV=enterovirus, AiV=aichivirus, HEV=hepatitis E virus, SaV=sapovirus, NA = not available, NoV=norovirus, TTV=torque teno virus, RoV=rotavirus, AstV=astrovirus, PyV=polyomavirus, PMMoV=pepper mild mottle virus, TMV=tobacco mosaic virus. ^a = detection period for infective stage in water at 20° C: short, up to 1 week; moderate, 1 week to 1 month,; long, over 1 month (WHO, 2017).

		SW-D			GW-D			DW	
virus type	positive/total	%	n° articles	positive/total	%	n° articles	positive/total	%	n° articles
AdV	755/1435	52.61%	29	102/4607	2.21%	13	535/3352	15.96%	29
AiV	50/114	43.86%	6	1/9	11.11%	2	16/142	11.27%	6
ASFLV	1/12	8.33%	1	-	-	-	-	-	-
AstV	48/132	36.36%	3	-	-	-	7/145	4.83%	3
CosV	-	-	-	-	-	-	9/18	50.00%	1
EV	106/544	19.49%	18	13/1667	0.78%	11	266/3439	7.73%	24
HAV	63/268	23.51%	7	0/994	0.00%	3	17/2335	0.73%	10
HCV	3/30	10.00%	1	-	-	-	-	-	-
HEV	26/176	14.77%	5	0/15	0.00%	2	24/80	30.00%	5
KV	2/12	16.67%	1	-	-	-	-	-	-
NoV GI	311/950	32.74%	21	4/1634	0.24%	10	97/4454	2.18%	20
NoV GII	463/1310	35.34%	26	13/1768	0.74%	11	71/4794	1.48%	22
NoV GIII	56/173	32.37%	2	-	-	-	-	-	-
NoV GIV	0/64	0.00%	1	-	-	-	-	-	-
PMMoV	273/320	85.31%	8	64/1078	5.94%	4	34/120	28.33%	7
BK/JC/MC/KI/WU	273/617	44.25%	11	15/1541	0.97%	6	22/170	12.94%	6
PyV									
ReV	0/16	0.00%	1	-	-	-	-	-	-
RoV	347/830	41.81%	14	57/1168	4.88%	4	201/3671	5.48%	18
SaliV	-	-	-	-	-	-	13/18	72.22%	1
SaV	5/144	3.47%	3	-	-	-	1/20	5.00%	1
TMV	8/12	66.67%	1	-	-	-	17/30	56.67%	2
TTV	16/79	20.25%	2	-	-	-	19/69	27.54%	3

Table 2. Total number of articles that investigated each viral agent and cumulative percentages of positive samples divided by type of water and type of virus.

- = not assessed; AdV = adenovirus; AiV = aichivirus; ASFLV = asfarvirus-like virus; AstV = astrovirus; CosV = cosavirus; DW = drinking water (water used for human consumption, not bottled); EV = enterovirus; GW-D = groundwater used as a source for DW production; HAV = hepatitis A virus; HCV = hepatitis C virus; HEV = hepatitis E virus; KV = klassevirus; NoV = norovirus; PMMoV = pepper mild mottle virus; PyV = polyomavirus; ReV = reovirus; RoV = rotavirus; SaliV = salivirus; SaV = sapovirus; SW-D = surface water used as a source for DW production; TMV = tobacco mosaic virus; TTV = torque teno virus.

Primary concentration	Secondary concentration	Number of articles	Reference
F-	None	17	Ahmad et al., 2018; Bortagaray et al., 2020; Canh et al., 2021; de Souza et al., 2018; Dos Santos et al., 2015; Gad et al., 2019; Kato et al., 2018; Kishida et al., 2012; Kluge et al., 2014; Miagostovich et al., 2020; Miura et al., 2019; Rashid et al., 2021; Rizk and Allayeh, 2018; Spilki et al., 2013; Staggemeier et al., 2015; Tandukar et al., 2020b; Vecchia et al., 2013
	UF	11	Asami et al., 2016; Assis et al., 2015; Da Silva Luz et al., 2020; Diston et al., 2015; Fongaro et al., 2015, 2013; Garcia et al., 2012; Haramoto et al., 2013, 2012; Kuroda et al., 2015; Tandukar et al., 2018
	PEG	1	Mackowiak et al., 2018
	None	10	Bonanno Ferraro et al., 2021; Ferrer et al., 2015; Iaconelli et al., 2017; Joung et al., 2013; Jung et al., 2011; Salvador et al., 2020; Silva et al., 2015; Steyer et al., 2011; Sylvestre et al., 2021; Varughese et al., 2018
F+	PEG	7	Grøndahl-Rosado et al., 2014; Kiulia et al., 2014; Opere et al., 2021; Pérez-Sautu et al., 2012; Potgieter et al., 2020; Shi et al., 2021; Ye et al., 2012
	F	1	Lee et al., 2018
	UF	1	Teixeira et al., 2020
	UF	6	Chigor and Okoh, 2012a, 2012b; Dienus et al., 2016; Kuroda et al., 2015; Malla et al., 2019; Sangsanont et al., 2016
F	None	5	Ferguson et al., 2012; Guerrero-Latorre et al., 2011; Hssaine et al., 2011; Kittigul and Pombubpa, 2021; Mattioli et al., 2013
	PEG	3	Borchardt et al., 2012; Gotkowitz et al., 2016; Lambertini et al., 2011
TEE	UF	1	Aw and Gin, 2011
IFF	PEG	1	Marie and Lin, 2017
	PEG	5	Charest et al., 2015; Cuevas-Ferrando et al., 2020; Murphy et al., 2020; Stokdyk et al., 2020; Williamson et al., 2011
UF	UF	4	Gibson et al., 2011; Gibson and Schwab, 2011a, 2011b; Hata et al., 2021
	None	3	Jacob et al., 2015; Knappett et al., 2011; Shoeib et al., 2011
PEG	F-	1	Ahmad et al., 2015
SMF	None	3	Calgua et al., 2013; Gamazo et al., 2018; Vieira et al., 2016

Table 3. Primary and secondary concentration methods reported in the analysed articles (total articles= 79, two concentration methods were applied by the study of Kuroda et al., 2015).

TFF = tangential flow filtration; UF = ultrafiltration; F = filtration; F- = filtration with electronegative charged membrane; F+ = filtration with electropositive charged membrane; PEG = precipitation with polyethylene glycol; SMF = skimmed milk flocculation.

Declaration of interests

 \boxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

□The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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