

# BIOLOGICAL & MEDICINAL CHEMISTRY

BMCS Postgraduate Symposium **XVII**

**Tuesday 9th January, 2024**



**The Medical Sciences Teaching  
Centre, Oxford University, Oxford**

# Contents

Click on a heading in this Contents page (bookmarked) to be directed to the relevant page(s) within the handbook.

[Sponsors](#)

[Information for Delegates](#)

[Programme](#)

[Speaker Abstracts](#)

[Poster Listing](#)

[Poster Abstracts](#)

[Organising Committee](#)

[Forthcoming Events](#)

[Initiatives and Announcements](#)

[Exhibitor floor plan](#)

[Catalogue of Exhibitors](#)

[Attendee List](#)

## Sponsors

The organising committee would like to thank the organisations below for their generous support and sponsorship of the symposium.



# Information for Delegates

## Fire Assembly Points

There is no fire alarm test planned on the day of the meeting. The fire assembly point is outside of the Medical Sciences Teaching Centre building on Darlington Link, please [click here](#) for map with the fire evacuation point.

## Security

Please note that conference badges must be worn at all times (access may be denied if not worn).

## Mobile phones and Photography

Please ensure that your phone is switched off or in silent mode in the lecture theatre.

No photography or filming is permitted during presentations – please contact the speaker directly if you would like a copy of their presentation.

## Twitter

Please use this hashtag when tweeting about the conference: #BMCSPostGrad24

## Smoking

Smoking is not permitted in any rooms or buildings.

## Internet Access

Wi-Fi connection is available free-of-charge throughout the venue, please collect Wi-fi ticket from the registration desk.

## Poster Presenters

Please refer to the author letter you would have received confirming your successful abstract, or the poster log, in this handbook for your poster number. Poster presenters are asked to stand by their boards during their allocated poster session which is detailed in the programme. Posters should be removed from boards by 15:30 pm on the day of the meeting, as the poster boards will be dismantled thereafter.

## Poster Voting and Prizes

There will be a People's poster prize. You are asked to vote for the People's Prize, using the voting slip included in your delegate badge. Please ensure this is posted in the ballot box on the Registration desk for your vote to be included. You should vote on the quality of the science being presented, and voting will be closed at 15:05 pm.

## Post-event Feedback

Promptly after the event, you will be sent an evaluation request to your registered email. Your feedback is very important to us so please take a few minutes to complete this.

## Luggage and Cloakroom

There is no cloakroom at the Medical Sciences Teaching Centre, please keep all your belongings with you at all times.



## Information for Delegates

### Parking

There is no parking available at the Medical Sciences Teaching Centre. If you are driving to Oxford, the best option is to use park and ride, please see a few suggestions below:

- Redbridge Park and Ride, please [click here](#) for the website;
- Pear Tree Park and Ride, please [click here](#) for the website;
- Thornhill Park and Ride, please [click here](#) for the website;

More park and ride locations near Oxford can be found [here](#).

### Taxis

Here are a few taxi suggestions:

001 Taxis                      01865240000

Royal cars                     01865778866

Radio taxis                    01865242424

The pickup address is: The Medical Sciences Teaching Centre, South parks road, Oxford, OX1 3PL.

The MSTC is 20 minutes' walk from the railway station. There are also express coach services that run between Oxford and London (1-1 1/2 hours) and Oxford is conveniently reached from Heathrow Airport (1 1/4 hours) and Gatwick Airport (2 hours) by frequent coach services. Further information how to get around Oxford can be found [here](#).

[Click here](#) for the online map of Medical Sciences Teaching Centre.

# Programme

Tuesday 9<sup>th</sup> January

## 9.30 Welcome address

*Douglas Williamson, Transition Bio, UK*

## 9.35 Opening remarks

*Angela Russell, University of Oxford, UK*

First session

Chair: *Tom Lanyon-Hogg, University of Oxford, UK*

## 9.40 Keynote lecture: Chemical Biology in Drug Discovery

*David Hewings, Vertex, UK*

## 10.10 Novel inhibitors of efflux pump NorA to target antimicrobial resistance

*Janine Gray, Imperial College London, UK*

## 10.30 Design and synthesis of novel allosteric modulators for the prostaglandin EP2 G protein-coupled receptor

*Constance Dalton, University of Nottingham, UK*

## 10.50 – 11.20 Refreshment break and exhibition

Second session

Chair: *Katherine Jones, Charles River, UK*

## 11.20 Discovery of Hedgehog Acyltransferase Inhibitors to Target Hedgehog Signalling in Cancer

*Efthymios S. Gavriil, Imperial College London, UK*

## 11.40 Flash Oral poster presentations

11.40 **FO01** Assessment of the Bioorthogonality of the Nitrile Imine 1,3-Dipole

*Mhairi Gibson, University of Strathclyde, UK*

11.42 **FO02** Investigation of the central core of MEDS433: A Bioisosteric voyage into the Amide's Role

*Elena Martino, University of Turin, Italy*

11.44 **FO03** Design, Synthesis, and In Vitro Characterisation of Activin Receptor-like Kinase 2 Degradable as a Novel Therapeutic Strategy Towards the Treatment of Diffuse Intrinsic Pontine Glioma

*Daniel Webb, University of Strathclyde, UK*

11.46 **FO04** Design, Synthesis and Pharmacological Testing of Novel Therapeutic Agents Designed to Enhance Insulin Secretion at the Trace Amine-associated Receptor 1

*Rhianna Lenham, University of Nottingham, UK*

11.48 **FO05** Vectorial Functionalisation of Pyrazolo[3,4-c]pyridines for Fragment-based Drug Discovery

*Elizabeth Bedwell, University of Durham, UK*

11.50 **FO06** Discovery of PINK1 Activators as Treatments for Parkinson's Disease

*Arwa AlGhamdi, University of Cardiff, UK*

11.52 **FO08** The Development of an on-DNA Micelle-Promoted Reductive Amination of DNA-Conjugated Amines to Access Previously Underexplored Peptidomimetics

*Matt Anderson, University of Newcastle, UK*

## Programme

11.54 **FO10** Development of Cleavable Linkers for Polymer-Drug Conjugates

*Sarah Phillips, University of Cambridge, UK*

11.56 **FO12** Targeting the cell-adhesion molecule PSGL-1 with a small molecule inhibitor

*Ciyana James, Queen's University Belfast, UK*

11.58 **FO16** The Development of 1,2,4-Triazine G-Protein Coupled Receptor 84 (GPR84) Antagonists

*Michael Malone, University of Glasgow, UK*

12.00 **FO17** Discovery of Novel Small Molecules for the Treatment of Human Coronaviruses

*Elliott Smyth, LifeArc, UK*

12.02 **FO18** Synthesis of Macrocyclic Ligands for the Bromodomain of CREBBP/p300

*Alistair Boyd, University of Oxford, UK*

12.04 **FO19** Bivalent chemical tools for investigating the tandem plant homeodomain finger-bromodomain cassette in TRIM24

*Michael Platt, University of Oxford, UK*

12.06 **FO20** Design, Synthesis, Biological Evaluation, and Molecular Modeling of Novel Benzofuran Derivatives as Targeted Cancer Chemotherapy

*Alaa Awad Taha Gad Osman, University of Cairo, Germany*

**12.08** Lunch, poster session and exhibition

Third session

Chair: *John Skidmore, University of Cambridge, UK*

**13.35 Keynote lecture: Design and Synthesis of Metathesis-Stapled Peptides**

*Alison Hulme, University of Edinburgh, UK*

**14.05 Synthesis of Novel Small Molecule Inhibitors for Snakebite Therapy**

*Nada Mosallam, University of Liverpool, UK*

**14.25 Identification Of Covalent Tools For Essential Leishmania Bromodomain LdBDF5**

*Claudine Greenwood, University of Strathclyde and GSK, UK*

**14.45 – 15.05** Refreshment break and exhibition

Fourth session

Chair: *Mary Wheldon, University of Dundee, UK*

**15.05 Structure-guided optimisation of N-hydroxythiazole-derived inhibitors of Factor Inhibiting Hypoxia-Inducible Factor- $\alpha$**

*Thomas Corner, University of Oxford, UK*

**15.25 3-D Building Blocks: A Modular Synthetic Platform for Elaborating Fragments to 3-D Lead Compounds**

*Andres Gomez Angel, University of York, UK*

**15.45 Keynote lecture: Development of Selective, Brain Penetrant PI5P4Kg Inhibitors with Contrasting Binding Modes**

*Helen Boffey, University of Cambridge, UK*

16.15 Closing remarks

*John Skidmore, University of Cambridge, UK*

16.25 Wine mixer & prizes

## **Programme**

17.00 Meeting close

**Chemical Biology in Drug Discovery***David Hewings*

Vertex Pharmaceuticals (Europe)  
86-88 Jubilee Avenue, Milton Park, Abingdon, Oxfordshire OX14 4RW  
david\_hewings@vrtx.com

David completed his MChem in Chemistry, MSc in Medicinal Chemistry for Cancer, and DPhil in Organic Chemistry (under the supervision of Prof. Stuart Conway) at the University of Oxford. He moved to the USA for postdoctoral research in Chemical Biology, first at Stanford University with Prof. Eric Kool, developing new techniques for the detection of nucleic acids, and then at Genentech. There, he worked on activity-based protein profiling (ABPP) of deubiquitinase enzymes. In 2018 he moved to Roche in Basel, Switzerland, as a Scientist in Medicinal Chemistry, and in 2020 returned to the UK, where he is currently a Principal Scientist at Vertex Pharmaceuticals. His work focuses on the application of Chemical Biology to drug discovery projects.

Chemical Biology, in its broadest sense, is the application of chemistry to improve our understanding of biology, but this gives little indication of what Chemical Biology can mean in practice in the context of drug discovery. I suggest that in a pharmaceutical setting, Chemical Biology encompasses both a set of challenges, such as assessing target engagement or developing hit finding strategies for previously-undruggable targets, and a set of techniques, such as chemical proteomics or induced proximity. New techniques, which contain chemistry at their core, are constantly expanding the range of challenges that Chemical Biology can help to tackle. I will present two case studies, one based on my postdoctoral research at Genentech, and another around the E3 ligase cereblon, which illustrate how chemical approaches can uncover new biology and how these new biological discoveries can in turn drive pharmaceutical innovation.



### **Novel inhibitors of efflux pump NorA to target antimicrobial resistance**

**J Gray**<sup>1</sup>, E Ledger<sup>2</sup>, K Arvaniti<sup>2</sup>, T Burden<sup>1</sup>, T Lanyon-Hogg<sup>3</sup>, J Riley<sup>4</sup>, K Reed<sup>4</sup>, I Gilbert<sup>4</sup>, A Edwards<sup>2</sup>, E Tate<sup>1</sup>

<sup>1</sup>Chemistry, Imperial College London, London, UK, <sup>2</sup>Medicine, Imperial College London, London, UK,

<sup>3</sup>Pharmacology, University of Oxford, Oxford, UK, <sup>4</sup>Life Sciences, DDU, University of Dundee, Dundee, UK

The emergence of antimicrobial resistance (AMR) is a global health challenge expected to cause 10 million deaths per year by 2050, and the incidence of multidrug resistance has been accelerated by excessive prescription and misuse of existing antibiotics. This pressing problem is further exacerbated by a lack of financial incentives, and by regulatory barriers to novel antibiotic development. There is an urgent need for the development of novel antibiotics with a new mechanism of action to combat bacterial infections.

A key facet of AMR is driven by expression of efflux pumps which extrude drugs from the bacterium, preventing them from reaching their targets at a sufficient concentration to inhibit growth. This presentation details the identification of the first potent and druglike inhibitors of the *Staphylococcus aureus* NorA multidrug efflux pump in a novel high-throughput phenotypic screen. NorA was established as the cellular target through transposon mutagenesis and assays demonstrating inhibition of substrate efflux. Initial SAR exploration improved potency to low nanomolar concentrations in NorA EtBr efflux and synergistic antibiotic susceptibility assays. Pharmacokinetic studies in vitro and in vivo demonstrated that the novel lead series was suitable for testing in animal models. Subsequent in vivo experiments showed that the compounds were active in combination therapy with existing antibiotics in murine infection models.

## Design and synthesis of novel allosteric modulators for the prostaglandin EP2 G protein-coupled receptor

C Dalton<sup>1</sup>, C Laughton<sup>1</sup>, N Holliday<sup>2</sup>, S Mistry<sup>1</sup>

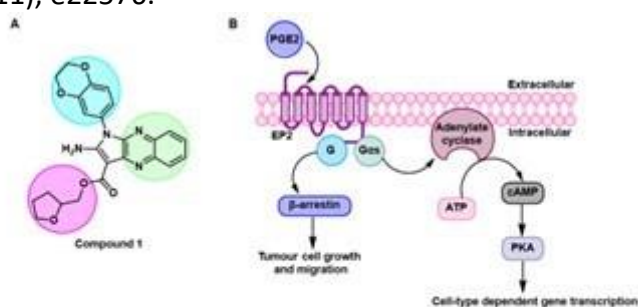
<sup>1</sup>School of Pharmacy, University of Nottingham, Nottingham, UK, <sup>2</sup>School of Life Sciences, University of Nottingham, Nottingham, UK

The prostaglandin EP2 receptor (EP2) is a widely expressed G protein-coupled receptor activated endogenously by prostaglandin E2 (PGE2), which contributes to the development of chronic inflammation in cancer and has roles in diseases such as Parkinson's, endometriosis, arthritis and intercranial aneurysms etc. (Figure 1B).<sup>1, 2</sup> EP2 antagonism is considered a possible therapeutic approach to treat these diseases. Previously, numerous orthosteric antagonists (i.e. those that bind to the PGE2 binding site) have been synthesised.<sup>1, 3</sup> In 2020, "**Compound 1**" (Figure 1A) was reported as the first allosteric EP2 antagonist that demonstrates a reversible, agonist dependent mode of action.

As part of this communication we will report the synthetic route to "**Compound 1**" and our exploration of an expanded structure-activity-relationship dataset with modifications at the tetrahydrofuran moiety. Initial work identified two novel analogues of "**Compound 1**" displaying improved affinity for EP<sub>2</sub> compared to the literature compound whilst demonstrating an insurmountable mode of action indicative of a negative allosteric modulator. Pharmacological characterisation was using conducted using both a NanoBRET competition binding study employing the G protein mimetic peptide TMR-Gαs19cha18 and a NanoBiT complementation assay with comparison against a known orthosteric antagonist.<sup>3</sup>

### References

1. C. Jiang, R. A., T. Ganesh and R. Dingleline, An Agonist Dependent Allosteric Antagonist of Prostaglandin EP2 receptors. *ACS. Chem. Neurosci.* **2020**, *11* (10), 1436-1446.
2. X. Sun, L. Q., Prostaglandin EP2 receptor: Novel therapeutic target for human cancers (Review). *Int. J. Mol. Med.* **2018**, *42* (3), 1203-1214.
3. J. P. Farmer, S. N. Mistry, C. A. Laughton, N. D. Holliday, Development of fluorescent peptide G protein-coupled receptor activation biosensors for NanoBRET characterization of intracellular allosteric modulators. *FASEB J* **2022**, *36* (11), e22576.



**Figure 1.** A. Structure of "**Compound 1**" highlighting three regions of interest for structural modification; dioxane (blue- where most structural changes occurred in the literature), tetrahydrofuran (pink) and quinaxaline (green). B. PGE<sub>2</sub> binds and activates EP<sub>2</sub>, G<sub>αs</sub>-mediated induction of adenylate cyclase to increase cytoplasmic cAMP levels. Downstream events are then mediated through protein kinase A. EP<sub>2</sub> activation also induces β-arrestin2 which is known to promote tumor cell growth and migration.<sup>1, 2</sup>

## Discovery of Hedgehog Acyltransferase Inhibitors to Target Hedgehog Signalling in Cancer

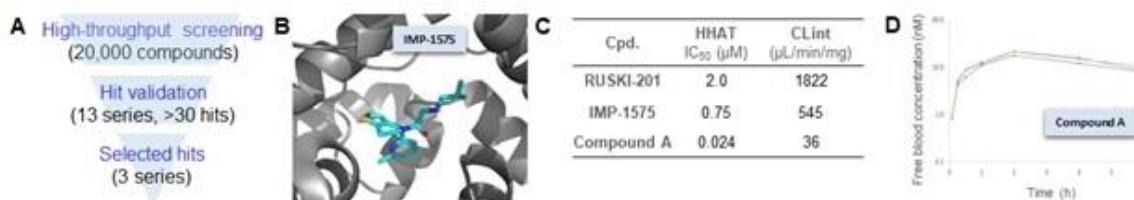
E Gavrili<sup>1</sup>, Z Xiao<sup>1</sup>, S Andrei<sup>1</sup>, GL Senatla<sup>1</sup>, A Chatwin<sup>1</sup>, CE Coupland<sup>2</sup>, P Kumar<sup>2</sup>, L Carrique<sup>2</sup>, S Afolaranmi<sup>3</sup>, A Hayes<sup>4</sup>

<sup>1</sup>Department of Chemistry, Imperial College London, London, UK, <sup>2</sup>Division of Structural Biology, Wellcome Centre for Human Genetics, University of Oxford, Oxford, UK, <sup>3</sup>Cancer Research UK Cambridge Institute, University of Cambridge, Cambridge, UK, <sup>4</sup>Division of Cancer Therapeutics, Centre for Cancer Drug Discovery, The Institute of Cancer Research, London, UK, <sup>5</sup>Department of Pharmacology, University of Oxford, Oxford, UK, <sup>6</sup>National Heart and Lung Institute, Imperial College London, London, UK

The Hedgehog (HH) pathway is aberrantly activated in cancer and has an important role in cancer stem cell maintenance and in tumour-stroma crosstalk [1]. Hedgehog Acyltransferase (HHAT), a transmembrane protein at the endoplasmic reticulum, catalyses the palmitoylation of HH peptides (SHH, DHH, IHH), which is a critical step for their signalling activity [2]. HHAT has been identified as an interesting target in breast, lung, colon and pancreatic cancer [3-7]. Previously reported HHAT inhibitors are characterised by non-HH specific cytotoxicity [8] and high clearance [7]. We aim to discover potent, selective and *in vivo* active HHAT inhibitors, as a novel approach to target hedgehog signalling in cancer.

We have elucidated key structural details in the HHAT active site using photochemical probes [9] and Cryo-EM [10]. We have also developed a high-throughput biochemical assay, termed 'Acyl-cLIP' [11], and screened a library of 20,000 compounds against purified HHAT. Subsequent hit validation and selection provided three series for optimisation (Fig. 1A). Cryo-EM structure-based drug design (Fig. 1B) has guided hit optimisation, generating lead Compound A, a low nM HHAT inhibitor both in enzymatic and cellular assays. Compound A is also characterised by acceptable metabolic stability and excellent permeability (Fig. 1C). Furthermore, *in vivo* pharmacokinetic studies have shown that Compound A is orally bioavailable and achieves sufficient levels of free drug concentration (Fig. 1D). Therefore, Compound A is a suitable and unique chemical probe to validate the role of HHAT *in vivo* and target HH signalling in cancer models. Further optimisation of lead Compound A along with progression of additional hit series is ongoing.

**References:** [1] Cell Chem Biol 2017, 24, 252. [2] PLoS ONE 2015, 9, e89899. [3] Mol Cancer 2015, 14, 72. [4] Cancer Cell 2014, 25, 139. [5] Oncogene 2013, 32, 2335. [6] Cell Rep 2017, 21, 2813. [7] Oncogene 2015, 34, 263. [8] ACS Chem Biol 2016, 11, 3256. [9] Angew. Chem. Int. Ed. 2021, 60, 13542. [10] Mol Cell 2021, 81, 5025. [11] Chem Sci 2019, 10, 8995.



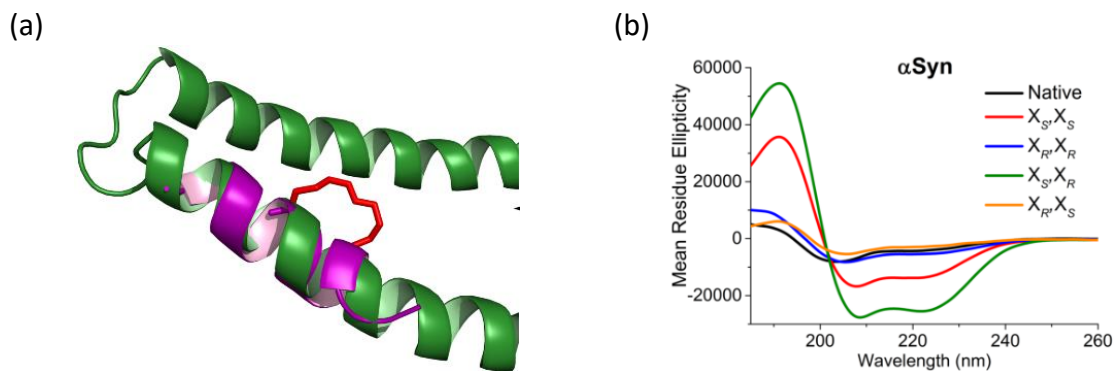
## Design and Synthesis of Metathesis-Stapled Peptides

Alison N. Hulme

School of Chemistry, The University of Edinburgh  
David Brewster Road, Edinburgh, EH9 3FJ, UK  
Alison.Hulme@ed.ac.uk

Stapled peptides, whereby cross-linking of two or more amino acid sidechains on an  $\alpha$ -helical peptide is carried out via chemical synthesis, generally have a more compact structure, enhanced cell penetration, and are more resistant to proteolysis than their non-stapled counterparts.<sup>1</sup> Moreover, their metabolites are relatively safe, and recent studies suggest that stapled peptides offer an advantage over traditional drugs by averting the development of drug resistance. Hence stapled peptides have predictably sparked a growing interest from the scientific community since the early 2000s; with the first “all-hydrocarbon” metathesis-stapled peptides currently in late-stage clinical trials.

Stapled peptide sequences may be designed from natural peptide interaction sequences; by the excision of a helical fragment from a protein-protein interface (Figure 1a); through biological screening; and computationally.<sup>1,2</sup> Staples can be varied by sequence position, cross-linking chemistry, length of the tether and stereochemistry at the  $\alpha$ -position (Figure 1b). Successful stapling is often indicated in the first instance by CD analysis, with a trend to higher peptide helicity upon stapling. We have shown that whilst  $\alpha,\alpha$ -disubstituted alanine derivatives are often used to promote helicity in metathesis-stapled peptides, their simpler (and more readily synthesized) mono-substituted counterparts can be equally as efficient.<sup>2</sup> Our recent work has focused on the development and application of add-on force fields for AMBER for the non-natural amino acids in stapled peptides.<sup>3</sup> These allow us to model stapled peptides with high efficiency and herald a new era in metathesis-stapled peptide design.



**Figure 1.** (a) Backbone alignment of an NMR refined solution structure of a metathesis-stapled  $\alpha$ -helical segment (magenta) of  $\alpha$ -Synuclein with the parent protein (green; PDB: 1XQ8). (b) Variation in helicity with  $\alpha$ -position stereochemistry ( $X_S/X_R$ ) in the metathesis-stapled peptide.<sup>2</sup>

- (1) Bluntzer M. T. J.; O'Connell J.; Baker T. S.; Michel J.; Hulme A. N. Designing stapled peptides to inhibit protein-protein interactions: An analysis of successes in a rapidly changing field. *Peptide Science* **2021**, *113*, e24191.
- (2) McWhinnie F. S.; Sepp K.; Wilson C.; Kunath T.; Hupp T. R.; Baker T. S.; Houston D. R.; Hulme A. N. Mono-Substituted Hydrocarbon Diastereomer Combinations Reveal Stapled Peptides with High Structural Fidelity. *Chem. Eur. J.* **2018**, *24*, 2094-2097.
- (3) Bluntzer M. T. J.; Notari, E.; O'Connell J.; Michel J.; Hulme A. N. *unpublished results*.

**Synthesis of Novel Small Molecule Inhibitors for Snakebite Therapy**

**N Mosallam**\*<sup>1</sup>, D Chong<sup>1</sup>, R Gunasekar<sup>1</sup>, N James<sup>1</sup>, A Westhorpe<sup>2</sup>, R Clare<sup>2</sup>, S Hall<sup>2</sup>, N Casewell<sup>2</sup>, N Berry<sup>1</sup>, P O'Neill<sup>1</sup>

<sup>1</sup>Department of Chemistry, University of Liverpool, Liverpool, UK, <sup>2</sup>Centre for Snakebite Research & Interventions, Liverpool School of Tropical Medicine, Liverpool, UK

Snakebite envenoming is responsible for 81000-138000 deaths annually and 400000 cases of long-term morbidity, making snakebite the most lethal neglected tropical disease in the world. The only current treatment for snakebite is antivenom, which has various side effects; in addition to its administration in healthcare facilities only, which limits rapid treatment for rural communities. These factors highlight the need for novel snakebite treatments. An oral snakebite treatment would be ideal as a standalone therapy or to be given as a rapid pre-hospital treatment to neutralise the effects of snakebite envenoming. Although there are various toxins found in any specific snake venom, those encoded by the phospholipase A2 (PLA2), snake venom metalloproteinase (SVMP) are the most abundant ones across different snake species, which makes them attractive targets for snake venom treatment. This presentation will focus on SVMP small molecule inhibitors for snakebite treatment. Since SVMP shares structural similarities with human matrix metalloproteinase (MMP), we aim to repurpose existing MMP inhibitors for SVMP inhibition.

In our programme, we have successfully developed several scaffolds as SVMP inhibitors with optimum potency against five main snake venoms. Three series of SVMP inhibitors were developed. Thirty analogues were prepared in total with the majority of them displaying potent activity against SVMP and good *in vitro* drug metabolism and pharmacokinetic (DMPK) profiles. Molecular modelling has been used to define key binding interactions of homology and crystal structures of bound inhibitors enabling us to target molecules with selectivity against human MMP isoforms.

Five of the most potent derivatives with optimum DMPK profiles were tested for their *in vivo* pharmacokinetic properties and preliminary proof of concept *in vivo* experiments. Interestingly, three analogues displayed good oral bioavailability in mice, snake venom protection in a mouse model and modest half-lives which encouraged us to focus on these scaffolds for lead optimisation.

In conclusion, novel SVMP inhibitors have been developed with several analogues displaying good potency and pharmacokinetic properties. Further investigation is underway to improve molecule potency to develop a novel oral drug for SVMP inhibition to be used in combination with a PLA2 inhibitor as a new approach for snakebite therapy.



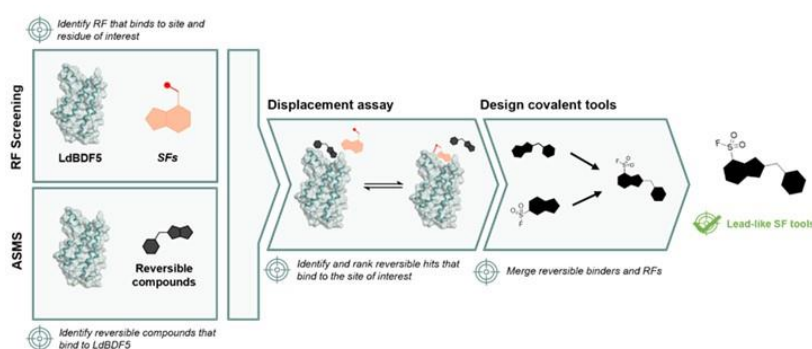
**Identification Of Covalent Tools For Essential Leishmania Bromodomain LdBDF5****C Greenwood<sup>1,2</sup>, A Aatkar<sup>1,2</sup>, N Tomkinson<sup>1</sup>, J Bush<sup>2</sup>, E Grant<sup>2</sup>**

<sup>1</sup>Pure and Applied Chemistry, University of Strathclyde, Glasgow, UK, <sup>2</sup>Chemical Biology, GSK, Stevenage, UK

Parasitic infections cause a tremendous burden of disease in tropic and sub-tropic regions of the world. Visceral leishmaniasis (VL), the second largest parasitic killer in the world after malaria, is caused by *Leishmania donovani*, a human blood parasite transmitted by sandflies. Whilst treatments are available for VL, recent years have seen an emergence of parasite resistance to current therapies. Our work aims to identify novel inhibitors of essential *Leishmania donovani* bromodomain factor 5 (LdBDF5). The essential nature of this protein results in a high resistance to mutations, key to prevent drug resistance. Identifying LdBDF5 inhibitors would therefore provide a novel method of treating VL.

Due to the key role bromodomains (BRDs) play in gene expression, LdBDF5 tools must be specific over human orthologs. To identify the most appropriate approach to target LdBDF5 with small molecules, crystal structures and protein sequences were examined. This study highlighted nucleophilic tyrosine residues unique to parasites that could be leveraged to gain selectivity over human BRDs. As such, it was believed that covalent inhibition *via* tyrosine targeting would be a valuable method to identify selective LdBDF5 inhibitors. A sulfonyl fluoride (SF) reactive fragment (RF) library was screened against human and parasite BRDs. Indeed, LdBDF5-selective SFs were identified, having modified the aforementioned tyrosine residues.

To rapidly identify chemical matter beyond a fragment space, libraries of drug-like reversible compounds were screened in an affinity selection-mass spectrometry (AS-MS) assay against LdBDF5 (Figure 1). The AS-MS assay identified a number of hits that bound reversibly to the protein, which were further assessed in a displacement assay to confirm site of binding (Figure 1). Finally, the most potent reversible binders and our hit SF fragments were merged to design two series of lead-like SFs. These demonstrated significant covalent modification and high binding efficiencies against LdBDF5. The results thus far have shown that a covalent strategy is suitable for selectively engaging parasite BRDs via the covalent modification of unique nucleophilic residues. Furthermore, an invaluable workflow was established, which through the rapid screening of covalent and reversible libraries and merging of the most potent hits, leads to the identification of potent lead-like SFs.



## Structure-guided optimisation of *N*-hydroxythiazole-derived inhibitors of Factor Inhibiting Hypoxia-Inducible Factor- $\alpha$

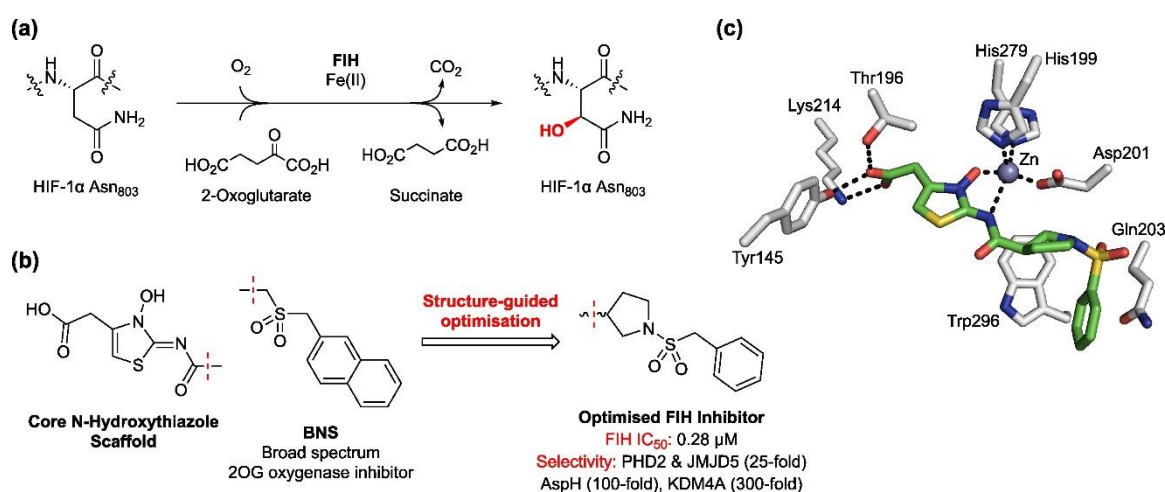
T Corner<sup>1</sup>, R Teo<sup>1</sup>, Y Wu<sup>2</sup>, E Salah<sup>1</sup>, Y Nakashima<sup>3</sup>, G Fiorini<sup>1</sup>, A Tumber<sup>1</sup>, X Zhang<sup>2</sup>, L Brewitz<sup>1</sup>, CJ Schofield<sup>1</sup>

<sup>1</sup>Department of Chemistry and the Ineos Oxford Institute, University of Oxford, Oxford, UK, <sup>2</sup>Jiangsu Key Laboratory of Drug Design and Optimization, China Pharmaceutical University, Nanjing, China, <sup>3</sup>Institute of Natural Medicine, University of Toyama, Toyama, Japan

The human 2-oxoglutarate (2OG)- and Fe(II)-dependent oxygenases factor inhibiting hypoxia-inducible factor- $\alpha$  (FIH) and HIF- $\alpha$  prolyl residue hydroxylases 1-3 (PHD1-3) regulate the response to hypoxia in humans via catalysing hydroxylation of the  $\alpha$ -subunits of the hypoxia-inducible factors (HIFs). Small-molecule PHD inhibitors are used for anaemia treatment; by contrast, few selective inhibitors of FIH have been reported, despite their potential to regulate the hypoxic response, either alone or in combination with PHD inhibition.

We report molecular, biophysical, and cellular evidence that BNS, a reported *N*-hydroxythiazole-based PHD2 inhibitor, is, in fact, a broad spectrum 2OG oxygenase inhibitor, the inhibition potential of which can be tuned to achieve selective FIH inhibition. Structure-guided optimisation resulted in the discovery of *N*-hydroxythiazole derivatives that manifest efficient FIH inhibition ( $IC_{50} < 0.3 \mu M$ ) and a substantially improved selectivity for FIH inhibition over PHD2 and other structurally-related 2OG oxygenases, including Jumonji-C domain-containing protein 5, aspartate/asparagine- $\beta$ -hydroxylase and histone *N*<sup>E</sup>-lysine demethylase 4A.

The optimised *N*-hydroxythiazole-based FIH inhibitors modulate the expression of FIH-dependent HIF target genes and, consistent with reports that FIH regulates cellular metabolism, suppressed lipid accumulation in adipocyte cells. Crystallographic studies reveal that the *N*-hydroxythiazole derivatives compete with both 2OG and the HIF- $\alpha$  substrate for binding to the FIH active site. Given the modular structure and broad-spectrum 2OG oxygenase inhibitory activity of BNS, derivatisation of the *N*-hydroxythiazole scaffold has the potential to afford selective inhibitors for 2OG oxygenases other than FIH, which play important roles in fundamental pathophysiological process including epigenetic gene regulation, DNA/RNA damage repair and lipid metabolism.



**Figure 1.** (A) FIH catalyses the C3 hydroxylation of Asn803 in HIF-1 $\alpha$ . (B) Structure-guided optimisation afforded *N*-hydroxythiazole analogues that manifest efficient FIH inhibition and high levels of selectivity over structurally-related 2OG oxygenases. (C) Crystallographic studies reveal a 2OG- and substrate-competitive FIH binding mode.

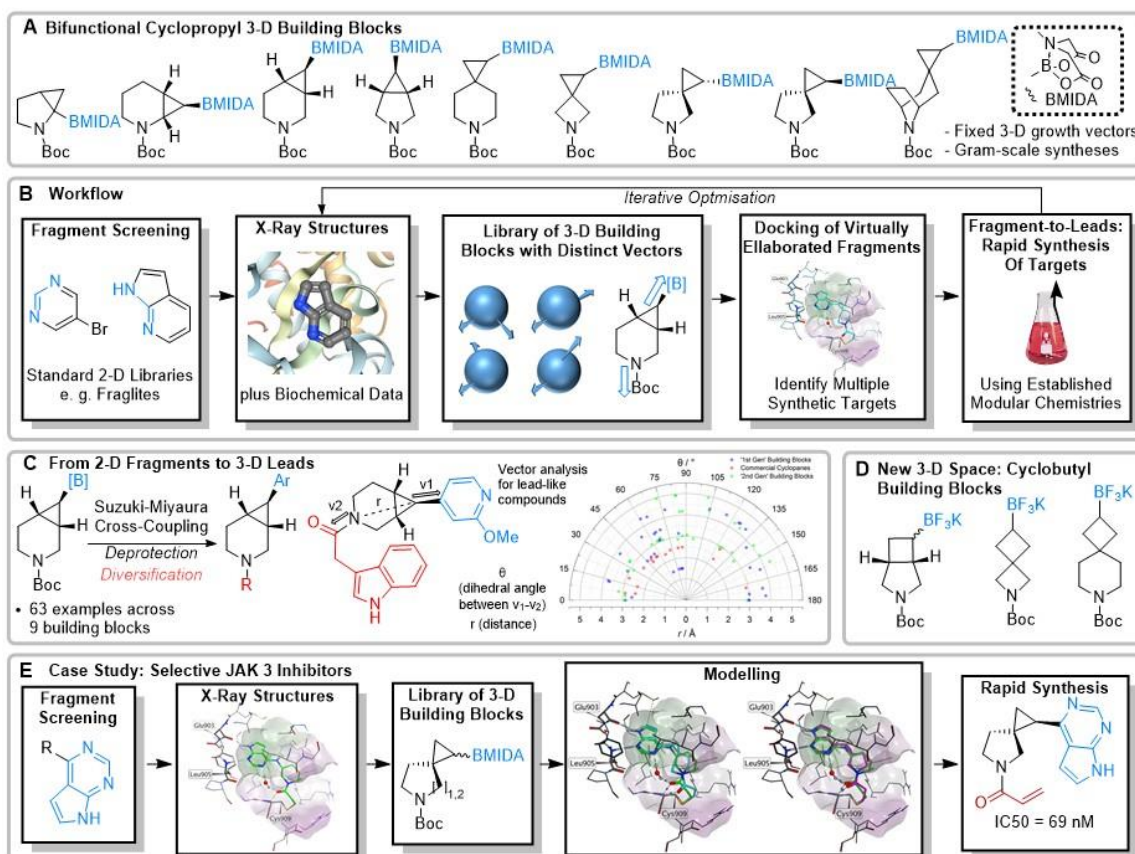
### 3-D Building Blocks: A Modular Synthetic Platform for Elaborating Fragments to 3-D Lead Compounds

Andres R Gomez-Angel<sup>1</sup>, William T Butler<sup>1</sup>, James R Donald<sup>1</sup>, Hanna F Klein<sup>1</sup>, Stephen Y Yao<sup>1</sup>, Rebecca Appiani<sup>1</sup>, James D Firth<sup>1</sup>, Lucia Fusani<sup>2</sup>, Simon C.C. Lucas<sup>2</sup>, Peter O'Brien<sup>1</sup>

<sup>1</sup>Department of Chemistry, University of York, York, UK, <sup>2</sup>Hit Discovery, Discovery Sciences, Cambridge, UK

With the advent of Fragment Based Drug Discovery (FBDD) for the efficient sampling of chemical space, the overall rate of discovery of potential drug candidates starting from fragments has increased.<sup>1</sup> However, this increase has highlighted the need to further develop synthetic chemistry to support FBDD.<sup>2</sup> One of these needs is increasing the 3-D shape of potential drug candidates<sup>3</sup> and interest in 3-D shaped fragments has emerged.<sup>4</sup> Nonetheless, as current libraries possess many compounds with low 3-D shapes<sup>5</sup> and elaborating such compounds is challenging, we now present a new, modular approach for the conversion of 2-D fragments into 3-D lead-like compounds with potential for automation.

Our technology platform enables the rapid elaboration of 2-D fragments in three-dimensions. A series of bifunctional 3-D building blocks with defined elaboration vectors has been designed and synthesised (**A**, available from Key Organics). Making use of structural information acquired during standard FBDD campaigns, our technology platform allows for a computationally guided elaboration of fragments (**B**). Utilising the cyclopropyl MIDA boronate handle, elaboration with medicinally relevant aryl bromides via Suzuki-Miyaura cross-coupling can be achieved. Additionally, a variety of *N*-functionalisation reactions are demonstrated to give access to a series of lead-like compounds by the use of precedented pharmacophores (**C**)<sup>6,7</sup> – this provides access to a wide range of 3-D vector space. In order to expand this access a new series of cyclobutyl containing building blocks is also being investigated (**D**). Finally, the utility of our modular synthetic platform is further highlighted by the design and synthesis of selective JAK3 inhibitors utilising two of the designed 3-D building blocks (**E**). Full details will be presented.



## Development of Selective, Brain Penetrant Phosphatidylinositol 5-phosphate 4-kinase Gamma Inhibitors with Contrasting Binding Modes

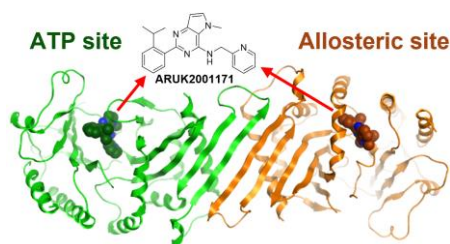
*Helen K. Boffey<sup>a</sup>, Timothy P. C. Rooney<sup>a</sup>, Henriette M. G. Willems<sup>a</sup>, Simon Edwards<sup>a</sup>, Greg G. Aldred, Christopher Green<sup>a</sup>, Tina Howard<sup>b</sup>, Derek Ogg<sup>b</sup>, Tamara Romero<sup>a</sup>, Duncan E. Scott<sup>a</sup>, David Winpenny<sup>a</sup>, James Duce<sup>a</sup>, John Skidmore<sup>a</sup>, Jonathan H. Clarke<sup>a</sup>, and Stephen P. Andrews<sup>a</sup>*

a. The ALBORADA Drug Discovery Institute, University of Cambridge, CB2 0AH, U.K.

b. Peak Proteins, Macclesfield SK10 4TG, U.K.

hkb32@cam.ac.uk

Phosphatidylinositol 5-phosphate 4-kinases (PI5P4Ks) are emerging as attractive therapeutic targets in diseases, such as cancer, immunological disorders, and neurodegeneration, owing to their central role in regulating cell signalling pathways that are either dysfunctional or can be modulated to promote cell survival. Different modes of binding may enhance inhibitor selectivity and reduce off-target effects in cells. By improving the physicochemical properties of the selective PI5P4K $\gamma$  inhibitor, NIH-12848<sup>1</sup>, we have demonstrated that this chemotype engages PI5P4K $\gamma$  in intact cells and is selective over PI5P4K $\alpha$  and PI5P4K $\beta$ . Furthermore, the first X-ray structure of PI5P4K $\gamma$  bound to an inhibitor has been determined with this chemotype, confirming an allosteric binding mode (pdb: 7QIE). An exemplar from this chemical series (ARUK2001171) adopted two distinct modes of inhibition, including through binding to a putative lipid interaction site which is 18 Å from the ATP pocket<sup>2</sup>.



In addition to this work, a virtual screening campaign using the PI5P4K $\alpha$  isoform afforded tractable biochemical hit compounds for both PI5P4K $\alpha$ <sup>3</sup> and PI5P4K $\gamma$ <sup>4</sup>. One series optimised towards PI5P4K $\gamma$  provided compounds with enhanced potency compared to ARUK2001171, isoform selectivity and favourable physicochemical properties. In contrast to ARUK2001171, an X-ray structure of a compound from this novel series (ARUK2001607) showed binding occurs exclusively in the ATP pocket of PI5P4K $\gamma$ . Significantly, these compounds demonstrate brain permeability in mice, and enabled the development of in vivo tool compounds for further evaluation of PI5P4Ks as a therapeutic target for neurodegenerative diseases.

1. Clarke, J. H., Irvine, R. F., The Activity, Evolution and Association of Phosphatidylinositol 5-Phosphate 4-Kinases. *Advances in Biological Regulation*, 2012, **52**, 40–45.
2. Boffey, H.K., Rooney, T. P. C., Willems, H. M. G., Edwards, S., Green, C., Howard, T., Ogg, D., Romero, T., Scott, D. E., Winpenny, D., Duce, J., Skidmore, J., Clarke, J. H., Andrews, S. P., Development of Selective Phosphatidylinositol 5-Phosphate 4-Kinase  $\gamma$  Inhibitors with a Non-ATP-competitive, Allosteric Binding Mode, *Journal of Medicinal Chemistry*, 2022, **65**, 3359-3370.
3. Willems, H. M. G., Edwards, S., Boffey, H.K., Chawner, S. J., Green, C., Romero, T., Winpenny, D., Duce, J., Skidmore, J., Clarke, J. H., Andrews, S. P., Identification of ARUK2002821 as an isoform-selective PI5P4K $\alpha$  inhibitor, *RSC Medicinal Chemistry*, 2023, **14**, 934.
4. Rooney, T. P. C., Aldred, G. A., Boffey, H.K., Willems, H. M. G., Edwards, S., Chawner, S. J., Scott, D. E., Green, C., Winpenny, D., Skidmore, J., Clarke, J. H., Andrews, S. P., The Identification of Potent, Selective, and Brain Penetrant PI5P4K $\gamma$  Inhibitors as In Vivo-Ready Tool Molecules, *Journal of Medicinal Chemistry*, 2023, **66**, 804-821.

## Poster Listing

No.	Title	Authors	Affiliations	Country
<b>FO01 &amp; P01</b>	Assessment of the Bioorthogonality of the Nitrile Imine 1,3-Dipole	<b>M Gibson</b> <sup>1</sup> , C Jamieson <sup>1</sup> , J Pettinger <sup>2</sup>	<sup>1</sup> Pure and Applied Chemistry, University of Strathclyde, Glasgow, UK, <sup>2</sup> Chemical Biology, GlaxoSmithKline, Stevenage, UK	UK
<b>FO02 &amp; P02</b>	Investigation of the central core of MEDS433: A Bioisosteric voyage into the Amide's Role	<b>E Martino</b> <sup>1</sup> , S Sainas <sup>1</sup> , M Alberti <sup>2</sup> , I Mannella <sup>1</sup> , M Giorgis <sup>1</sup> , P Circosta <sup>3</sup> , D Boschi <sup>1</sup> , ML Lolli <sup>1</sup>	<sup>1</sup> Department of Science and Drug Technology, University of Turin, Turin, Italy, <sup>2</sup> Department of Science and Drug Technology, University of Oriental Piedmont, Novara, Italy, <sup>3</sup> Molecular Biotechnology Center, University of Turin, Turin, Italy	Italy
<b>FO03 &amp; P03</b>	Design, Synthesis, and In Vitro Characterisation of Activin Receptor-like Kinase 2 Degradable as a Novel Therapeutic Strategy Towards the Treatment of Diffuse Intrinsic Pontine Glioma	<b>Daniel Webb</b> <sup>1,2</sup> , William Esmieu <sup>1</sup> , Katherine Jones <sup>1</sup> , Ryan Tinson <sup>1</sup> , Natsuko Macabuag <sup>1</sup> , Ruzica Bago <sup>1</sup> , Steve Clifton <sup>1</sup> , Lampros Milanos <sup>1</sup> , David Lindsay <sup>2</sup> , William Kerr <sup>2</sup>	<sup>1</sup> Early Discovery, Charles River Laboratories, Saffron Walden, UK, <sup>2</sup> Pure and Applied Chemistry, University of Strathclyde, Glasgow, UK	UK
<b>FO04 &amp; P04</b>	Design, Synthesis and Pharmacological Testing of Novel Therapeutic Agents Designed to Enhance Insulin Secretion at the Trace Amine-associated Receptor 1	<b>R Lenham</b> <sup>1,2</sup> , S Mistry <sup>1</sup> , C Loughton <sup>1</sup> , M Turner <sup>2</sup>	<sup>1</sup> School of Pharmacy, University of Nottingham, Nottingham, UK, <sup>2</sup> School of Science and Technology, Nottingham Trent University, Nottingham, UK	UK
<b>FO05 &amp; P05</b>	Vectorial Functionalisation of Pyrazolo[3,4-c]pyridines for Fragment-based Drug Discovery	<b>EV Bedwell</b> <sup>1</sup> , F Emery <sup>2</sup> , GC Clososki <sup>2</sup> , I Baxendale <sup>1</sup> , PG	<sup>1</sup> Department of Chemistry, Durham University, Durham, UK, <sup>2</sup> Faculdade de Ciências Farmacéuticas, Universidade de São Paulo, Ribeirão Preto, SP, Brazil	



## Poster Listing

<b>FO06 &amp; P06</b>	Discovery of PINK1 Activators as Treatments for Parkinson's Disease	<b>A Alghamdi</b> <sup>1</sup> , O Lambourne <sup>1</sup> , S Bell <sup>2</sup> , L Wilhelm <sup>3</sup> , E Yarbrough <sup>4</sup> , G Holly <sup>4</sup> , O Russell <sup>2</sup> , I Ganley <sup>3</sup> , M Goldbery <sup>4</sup> , Y Mehellou	<sup>1</sup> School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK, <sup>2</sup> Wellcome Centre for Mitochondrial Research, Newcastle University, Tyne, UK, <sup>3</sup> MRC Protein Phosphorylation and Ubiquitylation Unit, University of Dundee, Dundee, UK, <sup>4</sup> Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>5</sup> Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>6</sup> Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>7</sup> Wellcome Centre for Mitochondrial Research, Newcastle University, Tyne, UK, <sup>8</sup> MRC Protein Phosphorylation and Ubiquitylation Unit, University of Dundee, Dundee, UK, <sup>9</sup> Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, United States, <sup>10</sup> School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK	UK
<b>FO08 &amp; P08</b>	The Development of an on-DNA Micelle-Promoted Reductive Amination of DNA-Conjugated Amines to Access Previously Underexplored Peptidomimetics	<b>M Anderson</b> , T Carton, M Waring	Department of Chemistry, Newcastle University, Newcastle upon Tyne, UK	UK
<b>FO10 &amp; P10</b>	Development of Cleavable Linkers for Polymer-Drug Conjugates	<b>S Phillips</b> <sup>1</sup> , R Brewster <sup>2</sup> , R England <sup>2</sup> , M Park <sup>1</sup> , J Mandrup Kandemir <sup>1</sup> , D Spring <sup>1</sup>	<sup>1</sup> Yusuf Hamied Department of Chemistry, University of Cambridge, Cambridge, UK, <sup>2</sup> AstraZeneca, Macclesfield, UK	UK
<b>FO12 &amp; P12</b>	Targeting the cell-adhesion molecule PSGL-1 with a small molecule inhibitor	<b>C James</b> <sup>1</sup> , V Kanabar-Raivadera <sup>2</sup> , CP Page <sup>2</sup> , L Martin <sup>1</sup> , GK Wagner <sup>1</sup>	<sup>1</sup> School of Pharmacy, Queen's University, Belfast, UK, <sup>2</sup> School of Cancer and Pharmaceutical Sciences, King's College, London, UK	UK
<b>FO16 &amp; P16</b>	The Development of 1,2,4-Triazine G-Protein Coupled Receptor 84 (GPR84) Antagonists	<b>Michael Malone</b> <sup>1</sup> , Graeme Milligan <sup>2</sup> , Andrew Jamieson <sup>1</sup>	<sup>1</sup> Chemistry, University of Glasgow, Glasgow,Scotland, <sup>2</sup> Molecular Bioscience, University of Glasgow, Glasgow,Scotland	UK

## Poster Listing

<b>FO17 &amp; P17</b>	Discovery of Novel Small Molecules for the Treatment of Human Coronaviruses	<b>Elliott Smyth</b> <sup>1,2</sup> , Joao Pisco <sup>1</sup> , Nathalie Bouloc <sup>1</sup> , Jonathan Large <sup>1</sup> , Richard Foster <sup>2</sup>	<sup>1</sup> Chemical Biology, LifeArc, Stevenage, UK, <sup>2</sup> School of Chemistry, University of Leeds, Leeds, UK	UK
<b>FO18 &amp; P18</b>	Synthesis of Macrocyclic Ligands for the Bromodomain of CREBBP/p300	<b>Alistair Boyd</b> <sup>1</sup> , Mustafa Moroglu <sup>2</sup> , Stuart Conway <sup>3</sup>	<sup>1</sup> Organic Chemistry, University of Oxford, Oxford, UK, <sup>2</sup> Medicinal Chemistry, GSK, Stevenage, UK, <sup>3</sup> Chemistry, UCLA, Los Angeles, United States of America	UK
<b>FO19 &amp; P19</b>	Bivalent chemical tools for investigating the tandem plant homeodomain finger-bromodomain cassette in TRIM24	<b>Michael Platt</b> , Stuart Conway	Department of Chemistry, University of Oxford, Oxford, UK	UK
<b>FO20 &amp; P20</b>	Design, Synthesis, Biological Evaluation, and Molecular Modeling of Novel Benzofuran Derivatives as Targeted Cancer Chemotherapy	<b>A A. Osman</b> <sup>1,2</sup> , E R. Mohamed <sup>2</sup> , H A. Abdel-Aziz <sup>3</sup> , A M. El Kerdawy <sup>2,4</sup> , H Abdelrasheed Allam <sup>2</sup>	<sup>1</sup> Department of Pharmaceutical Chemistry, Faculty of Pharmacy, NewGiza University, Newgiza, Cairo 12256, Egypt, <sup>2</sup> Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Cairo University, Cairo, P.O. Box 11562, Egypt, <sup>3</sup> Applied Organic Chemistry Department, National Research Center, Dokki, Cairo 12622, Egypt, <sup>4</sup> School of Pharmacy, College of Health and Science, University of Lincoln, Green Lane, Lincoln, Lincolnshire, UK	UK

# Poster Abstracts

## FO01 & P01 Assessment of the Bioorthogonality of the Nitrile Imine 1,3-Dipole

M Gibson<sup>1</sup>, C Jamieson<sup>1</sup>, J Pettinger<sup>2</sup>

<sup>1</sup>Pure and Applied Chemistry, University of Strathclyde, Glasgow, UK, <sup>2</sup>Chemical Biology, GlaxoSmithKline, Stevenage, UK

The nitrile imine (NI) 1,3-dipole is a highly reactive and readily accessible synthetic intermediate generated via the photolysis of 2,5-disubstituted tetrazoles. Its ability to participate in 1,3-dipolar cycloadditions has enabled its application in a variety of synthetic methods. Of note, NIs have found application in medicinal chemistry, materials chemistry, and more recently, bioorthogonal chemistry. NI-mediated click reactions have recently found traction in bioorthogonal labelling techniques due to the light-activated, traceless nature of the system and the formation of stable, fluorescent adducts with biomolecules modified with an appropriate dipolarophile. While NIs are renowned for their proclivity towards cycloadditions, this species exhibits broad reactivity with a range of nucleophilic functionalities. Such functionalities are ubiquitous in biomolecules and therefore their promiscuous reactivity with the NI dipole may hinder its application as a true bioorthogonal labelling tool.

Previous work in our group has sought to explore the reactivity profile of the NI species through a series of competition experiments utilising a library of nucleophilic model substrates and dipolarophiles. Interestingly, the quantification of NI dipole reactivity with a range of carboxylic acid moieties revealed an enhancement in reactivity with decreasing  $pK_a$  of the acidic coupling partner. These findings have been expanded to assess the bioorthogonality of the dipole through the competitive reaction of an electronically activated dipolarophile versus a highly acidic fluorinated carboxylic acid. A range of NI species were generated through photolysis of a 2,5-disubstituted tetrazole and their reactivity with a model substrate was quantified.

The selectivity observed demonstrated that NI reactivity can be tuned *via* modulation of the  $pK_a$  of an acidic coupling partner. The next phase of this work sought to exploit this observation by exemplifying the application of highly acidic carboxylic acid moieties as novel bioorthogonal handles for NI-mediated photoclick reactions. A suitable NI precursor has been identified which enhances chemoselectivity for the bioorthogonal handle, suppressing the reactivity of endogenous competing nucleophiles. Current work is ongoing to incorporate a selection of highly acidic bioorthogonal handles into model peptide sequences containing multiple nucleophilic amino acid residues, allowing the chemoselectivity of the NI with these novel bioorthogonal handles to be assessed.



## Poster Abstracts

### FO02 & P02 Investigation of the central core of MEDS433: A Bioisosteric voyage into the Amide's Role

E Martino<sup>1</sup>, S Sainas<sup>1</sup>, M Alberti<sup>2</sup>, I Mannella<sup>1</sup>, M Giorgis<sup>1</sup>, P Circosta<sup>3</sup>, D Boschi<sup>1</sup>, ML Lolli<sup>1</sup>

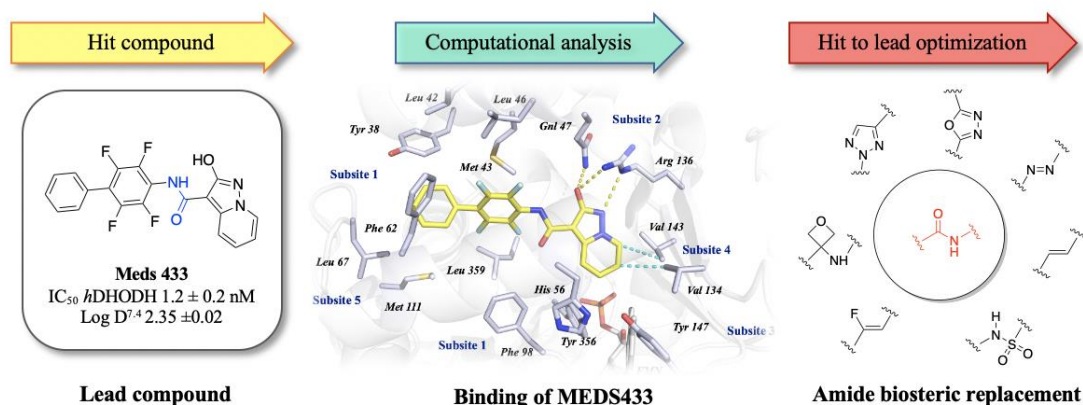
<sup>1</sup>Department of Science and Drug Technology, University of Turin, Turin, Italy, <sup>2</sup>Department of Science and Drug Technology, University of Oriental Piedmont, Novara, Italy, <sup>3</sup>Molecular Biotechnology Center, University of Turin, Turin, Italy

*Human Dihydroorotate Dehydrogenase (hDHODH)* it's a key enzyme involved in the stereoselective oxidation of L-dihydroorotate (DHO) to orotate (ORO) in the *de novo* pyrimidine biosynthesis. Recent studies proved that inhibiting *hDHODH* constitutes a promising pharmacological strategy for treatment of hematological malignancies, such as acute myeloid leukemia (AML)<sup>1,2</sup>, and viral pathologies<sup>3,4</sup>.

In recent years, the MEDSynth research group has studied and designed a first generation of inhibitors based on hydroxypyrazolo[1,5-a]pyridine core scaffold, which acts as an unusual carboxylic acid bioisosters. The heterocycle is therefore linked to a biphenyl or diaryl ether via an amide linker<sup>4</sup>.

Within the series, **MEDS433** emerged as the most promising compound, showing *in vitro* nanomolar activity against *hDHODH* and *in vivo* ability to achieve differentiation in AML cells. Currently, its development as a drug candidate is progressing through the pre-clinical phase for the treatment of AML. Despite its high potency, this lead compound suffers from poor solubility and optimization of its drug-like properties is necessary in order to perform further pre-clinical studies.

In this work, aiming to improve the solubility without losing in binding affinity, new analogues series was developed with the aim to study the role of the amide moiety through a bioisosteric approach. Theoretical design, synthesis and enzymatic assay are here presented and discussed.



**Figure 1:** Hit to lead optimization of MEDS433 by bioisosteric approach.

1. Lolli, M. L. *et al. Recent Pat. Anticancer. Drug Discov.* **2018**, *13*, 86–105.
2. Sykes, D. B. *Expert Opin. Ther. Targets* **2018**, *22*, 893–898.
3. Dunn, M. C. C., Knight, D. A. & Waldman, W. J. *Antivir. Ther.* **2011**, *16*, 309–317.
4. Sainas, S. *et al., J. Med. Chem.* **2018**, *61*, 6034–6055.

# Poster Abstracts

## FO03 & P03 Design, Synthesis, and *In Vitro* Characterisation of Activin Receptor-like Kinase 2 Degraders as a Novel Therapeutic Strategy Towards the Treatment of Diffuse Intrinsic Pontine Glioma.

Daniel Webb<sup>1,2</sup>, William Esmieu<sup>1</sup>, Katherine Jones<sup>1</sup>, Ryan Tinson<sup>1</sup>, Natsuko Macabuag<sup>1</sup>, Ruzica Bago<sup>1</sup>, Steve Clifton<sup>1</sup>, Lampros Milanos<sup>1</sup>, David Lindsay<sup>2</sup>, William Kerr<sup>2</sup>

<sup>1</sup>Early Discovery, Charles River Laboratories, Saffron Walden, UK, <sup>2</sup>Pure and Applied Chemistry, University of Strathclyde, Glasgow, UK

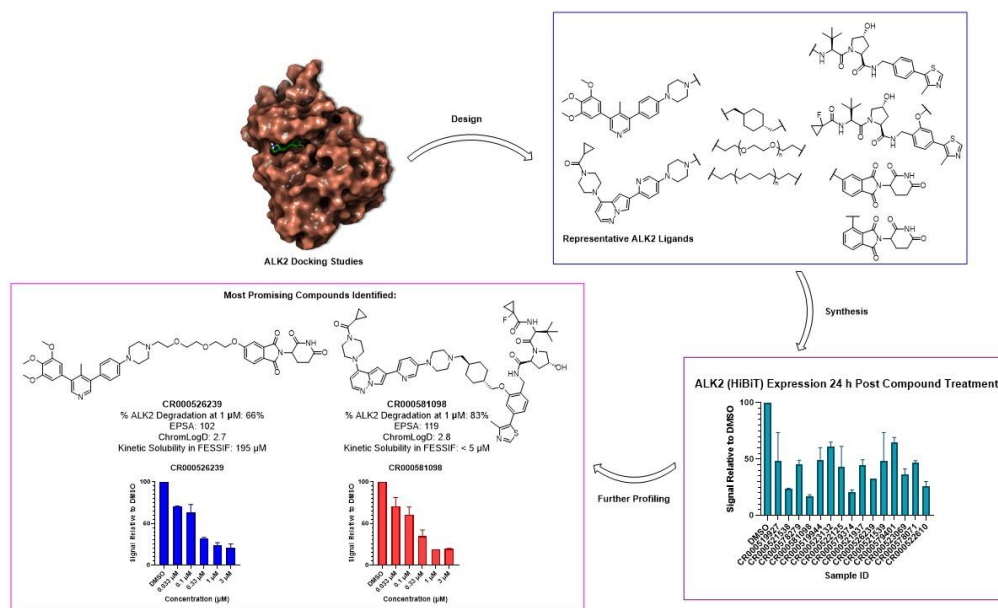
Diffuse Intrinsic Pontine Gliomas (DIPG) are grade IV tumours originating in the pons region of the brainstem. They account for 10–20% of all paediatric brain tumours and the median overall survival rate is 8–11 months after diagnosis.<sup>1</sup> Focal radiation therapy remains the only treatment option for DIPG patients and, to date, over 200 clinical trials using chemotherapeutic agents have failed to extend survival. Activin receptor-like kinase 2 (ALK2) has been identified as a potential target for therapeutic intervention in this disease, and several pre-clinical ALK2 inhibitors have been reported recently.<sup>2,3</sup> Herein, the design, synthesis, and biological evaluation of ALK2 proteolysis targeting chimeras (PROTACs) is explored as an alternative ALK2 targeting strategy.

The X-ray crystal structure of ALK2 bound to a small molecule inhibitor, LDN-212854, was used to dock ALK2 ligands across two distinct chemotypes, to identify suitable PROTAC-linker attachment points. A set of 15 ALK2 degraders was then designed, incorporating ligands for the E3 ligases, von Hippel-Lindau (VHL) and Cereblon (CRBN). Towards the synthesis of these targets, methods including high-throughput experimentation (HTE) were employed to optimise the efficiency of key chemical transformations. This initial set of compounds spanned a broad range of physicochemical property space, including a CLogD range of 1-6 and a TPSA range of 140-230. Western blot screening experiments were conducted to identify promising compounds that induced ALK2 degradation at a concentration of 1  $\mu$ M, before profiling selected compounds further and demonstrating that our PROTACs induced ALK2 degradation in a dose-dependent manner. Chromatographic measures were used to experimentally measure the physicochemical properties of our degraders, EPSA and ChromLogD, and investigate how these values differed from the calculated properties, TPSA and CLogD. *In vitro* ADME studies have been used to assess compound permeability, solubility, and metabolic stability, and to determine how these parameters are influenced by compound structure. Using the *in vitro* data obtained, a further iteration of compounds has been designed with the goal of improving potency and ADME properties. Synthesis and evaluation of these compounds is currently underway.

1. *J Korean Neurosurg Soc* **2018**, 61 (3), 343-351.

2. *Commun Biol* **2019**, 2, 156.

3. *ACS Omega* **2021**, 6 (32), 20729-20734.





## Poster Abstracts

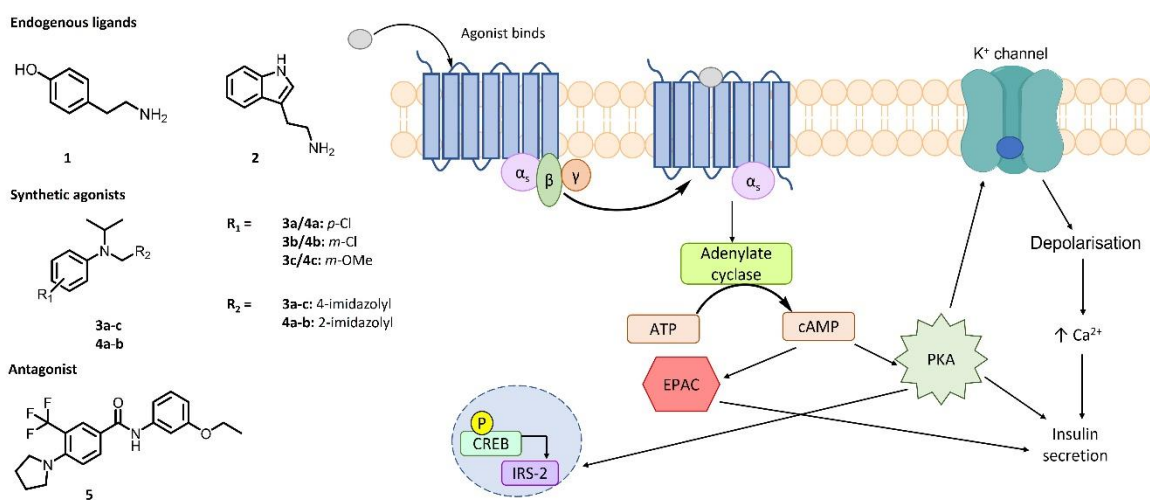
### FO04 & P04 Design, Synthesis and Pharmacological Testing of Novel Therapeutic Agents Designed to Enhance Insulin Secretion at the Trace Amine-associated Receptor 1.

R Lenham<sup>1,2</sup>, S Mistry<sup>1</sup>, C Laughton<sup>1</sup>, M Turner<sup>2</sup>

<sup>1</sup>School of Pharmacy, University of Nottingham, Nottingham, UK, <sup>2</sup>School of Science and Technology, Nottingham Trent University, Nottingham, UK

Insulin is widely considered the most important hormone involved in metabolic homeostasis with defects in its action or secretion causing diabetes mellitus, a condition afflicting over 400 million people worldwide. Although several oral hyperglycaemic agents have been developed to manage type 2 diabetes, their effectiveness often declines over time. Trace amine-associated receptor 1 (TAAR1) is a G protein-coupled receptor expressed throughout the body, including pancreatic  $\beta$ -cells. Pancreatic TAAR1 (below figure) can amplify insulin secretion via the activation of protein kinase A and EPAC signalling cascades, thus it is a potential therapeutic target for novel, oral hyperglycaemic drugs.

Five known TAAR1 agonists (**3a-c** and **4a-b**,  $K_i = 4\text{--}138$  nM) were resynthesized to validate the proposed pharmacology assays by determining their insulin secretion ability and capability to enhance cyclic AMP levels. Furthermore, as no crystal structure of TAAR1 has been determined, machine learning techniques were implemented for structure determination. Over 1000 potential models were generated and ranked based on their ability to dock the four known TAAR1 ligands (**1-3a** and **5**) with 2 models showing promising results. Novel analogues were designed, then docked into the homology models and their pharmacological properties evaluated *in vitro*. As part of this communication, we will describe the synthesis and pharmacological activity of the novel and repurposed literature compounds as well as the molecular docking studies.



## Poster Abstracts

### FO05 & P05 Vectorial Functionalisation of Pyrazolo[3,4-c]pyridines for Fragment-based Drug Discovery

EV Bedwell<sup>1</sup>, F Emery<sup>2</sup>, GC Clososki<sup>2</sup>, I Baxendale<sup>1</sup>, PG Steel<sup>1</sup>

<sup>1</sup>Department of Chemistry, Durham University, Durham, UK, <sup>2</sup>Faculdade de Ciências Farmacéuticas, Universidade de São Paulo, Riberião Preto, SP, Brazil

Fragment-based drug discovery (FBDD) is now a well-established method of identifying novel drug candidates that is being applied to diverse targets from DNA to protein-protein interactions. Heterocycles have a privileged place in FBDD due to an ability to engage with the target protein through a wide variety of intermolecular interactions, coupled with the potential to optimise drug-like properties (lipophilicity, hydrogen bonding capacity, and polarity) through modification of substituents. However, the range of heterocycles in common usage remains limited which hinders development of new bioactive agents. Novel heterocyclic ring systems are therefore needed as valuable inputs to unlock elusive therapeutic targets.

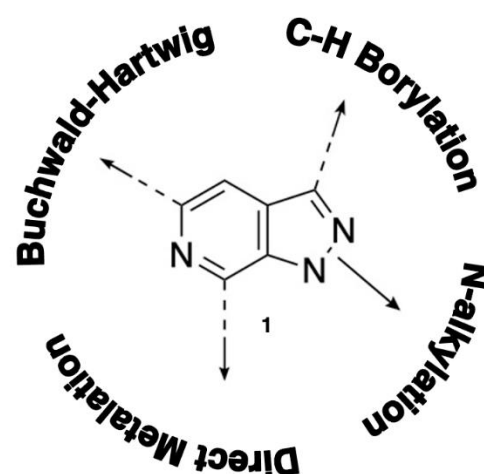
A challenge of working with novel heterocycles arises when introducing new substitution patterns requires complex *de novo* synthesis and strategies for late-stage functionalization are lacking. This limits the structural diversity that can be rapidly generated for testing in drug-discovery regimes and progress may stall while new synthetic routes are established.

To address these issues, we developed strategies for late-stage functionalisation of novel heterocyclic scaffolds to generate a diverse library of fragments. The pyrazolo[3,4-c]pyridine scaffold **1** was identified through a comprehensive enumeration of plausible yet unreported heteroaromatic rings in a virtual library designed to spark exploration of new chemical space.<sup>[1]</sup>

This poster presents synthetic routes to the pyrazolo[3,4-c]pyridine scaffold **1** optimised for large-scale batch or flow chemistries, and details regioselective methods to access the major growth-vectors. Specifically, using (1) metal-catalysed C-H borylation and Suzuki-Miyaura cross-coupling, (2) N-alkylation, (3) Buchwald-Hartwig amination, and (4) direct metalation chemistry with magnesium bases. These methods can be used individually or in sequence to generate an elaborated library of compounds based on the pyrazolo[3,4-c]pyridine scaffold **1**.

In addition to developing this synthetic methodology, this project aims to demonstrate the medicinal chemistry potential of the generated heterocycles. Computational docking and enzymatic assays with the sirtuin proteins of *Leishmania* parasites have been chosen to establish antileishmanial potential.

The future of this project will focus on synthesis and functionalization of other heterocycles of interest in the development of antileishmanial agents.



[1] WR. Pitt, DM. Parry, BG. Perry, CR. Groom, *J. Med. Chem.* **2009**, *52*, 2952–2963

## Poster Abstracts

### FO06 & P06 Discovery of PINK1 Activators as Treatments for Parkinson's Disease

A AlGhamdi<sup>1</sup>, O Lambourne<sup>1</sup>, S Bell<sup>2</sup>, L Wilhelm<sup>3</sup>, E Yarbrough<sup>4</sup>, G Holly<sup>4</sup>, O Russell<sup>2</sup>, I Ganley<sup>3</sup>, M Goldbery<sup>4</sup>,  
Y Mehellou<sup>1</sup>

<sup>1</sup>School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK, <sup>2</sup>Wellcome Centre for Mitochondrial Research, Newcastle University, Tyne, UK, <sup>3</sup>MRC Protein Phosphorylation and Ubiquitylation Unit, University of Dundee, Dundee, UK, <sup>4</sup>Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>5</sup>Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>6</sup>Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, USA, <sup>7</sup>Wellcome Centre for Mitochondrial Research, Newcastle University, Tyne, UK, <sup>8</sup>MRC Protein Phosphorylation and Ubiquitylation Unit, University of Dundee, Dundee, UK, <sup>9</sup>Center for Neurodegeneration and Experimental Therapeutics, University of Alabama, Birmingham, United States, <sup>10</sup>School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK

Parkinson's disease (PD) is a slowly progressive neurodegenerative disease that affects almost 10 million people worldwide. Among the causes of PD are loss of function mutations in the serine/threonine protein kinase PINK1.<sup>(1)</sup> These mutations lead to impairment in mitophagy, a cellular process that removes damaged or depolarized mitochondria, which protects cells from oxidative stress and eventually neuronal cell death. The accumulation of damaged mitochondria can result in increased oxidative stress and inflammation, which are believed to contribute to the death of dopaminergic neurons in PD.<sup>(2)</sup> Upon mitochondrial depolarization, PINK1 gets stabilized on the outer mitochondrial membrane (OMM) and phosphorylates ubiquitin and the E3 Ubiquitin ligase, parkin, consequently triggering mitophagy. PINK1 activators have gained great attention as a potential therapeutic approach for PD by promoting neuroprotection through mitophagy. Several attempts have been made to develop small molecule PINK1 activators, including nucleobases, nucleosides, and their nucleotide prodrugs.<sup>(3)</sup> Encouraged by these efforts, the aim of this work is to design and synthesize small molecule activators of the PINK1 activators as a potential treatment of PD. We designed and synthesized a small series of PINK1 activators. These compounds were obtained in high purity (>95%) and with yields ranging from 10-35%. These compounds were found to be able to activate PINK1 in cells as judged by the phosphorylation of its physiological substrate parkin. Using *mito*-QC reporter assay in MEFs, PINK1 activation by nucleoside analogues resulted in triggering PINK1-dependent mitophagy, which was not observed in MEF PINK1 KO. Interestingly, the PINK1-dependent mitophagy caused by our nucleoside analogues was able to suppress the accumulation of phospho-ubiquitin caused by mitochondrial damaging agents such as niclosamide and CCCP, which is a hallmark of PD. In conclusion, we have discovered the first series of PINK1 activators that have the capability of suppressing the accumulation of phosphoubiquitin, which is toxic to neurons. Such compounds hold promise for future development as PD treatments.

## Poster Abstracts

### FO08 & P08 The Development of an on-DNA Micelle-Promoted Reductive Amination of DNA-Conjugated Amines to Access Previously Underexplored Peptidomimetics

M Anderson, T Carton, M Waring

Department of Chemistry, Newcastle University, Newcastle upon Tyne, UK

Hit generation is a crucial component of any drug discovery process. A plethora of methods exist, allowing for the screening of potential candidate molecules against a variety of targets, including high-throughput screening (HTS), fragment-based drug discovery (FBDD), and several *in silico* methods. DNA-encoded libraries (DELs) have emerged as a promising technology within the field of screening, due to their comparative benefits: large library sizes ( $>10^6$  members) covering a diverse range of chemical space, time- and cost-effective synthesis and storage, and the relatively small amount of protein required for the screening.

Despite the numerous advantages of DELs, there remain limitations in the chemistries applicable to on-DNA synthesis as reactions occur in the presence of unprotected DNA, therefore, any reactions employed must be DNA-compatible. Specifically, reactions must be conducted in water and efficient at high dilutions while preserving the integrity of the DNA, i.e. cause no damage to the DNA tag. Ideally, transformations should be high-yielding for a broad substrate scope. One approach to facilitating reactions under aqueous conditions is through the application of micelle-forming surfactants, such as TPGS-750-M.

Reductive aminations play a crucial role in medicinal chemistry towards the synthesis of secondary and tertiary amines. In spite of this, reported on-DNA approaches are few and far between, typically utilising large excesses of amine to force the reaction. In fact, there are no published procedures applicable to a wide range of aldehydes, using amine-tagged DNA. **Herein is detailed the development of an on-DNA reductive amination, with amine on DNA, applicable to an incredibly diverse range of substrates ( $> 50$ ), including aliphatic, aromatic, and heterocyclic aldehydes. The optimised micellar conditions yield high conversions ( $> 75\%$ ) to the desired amine for various on-DNA amines, both primary and secondary in nature.**

Peptoids are a class of peptidomimetics bearing resistance to proteolytic degradation and improved cell permeability, relative to many peptides, while maintaining the ability to mimic peptidic structures. Peptoids also boast an expansive side-chain diversity; hence, peptoid synthesis from a broad selection of aldehydes on DNA allows for the synthesis of a peptoidic DEL.

# Poster Abstracts

## FO10 & P10 Development of Cleavable Linkers for Polymer-Drug Conjugates

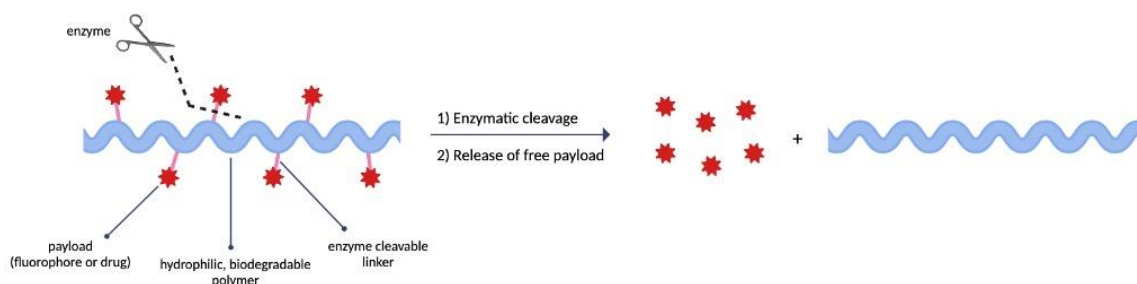
S Phillips<sup>1</sup>, R Brewster<sup>2</sup>, R England<sup>2</sup>, M Park<sup>1</sup>, J Mandrup Kandemir<sup>1</sup>, D Spring<sup>1</sup>

<sup>1</sup>Yusuf Hamied Department of Chemistry, University of Cambridge, Cambridge, UK, <sup>2</sup>AstraZeneca, Macclesfield, UK

Polymer-drug conjugates (PDCs) are an important class of polymer therapeutic that consist of a polymer backbone conjugated to a small-molecule drug via a chemical linker. PDCs provide many benefits to the delivery of small molecular weight drugs for the treatment of cancer, such as increased circulation half life, high drug loading and increased hydrophilicity. Despite the benefits of PDCs as anti-cancer treatments, currently no polymer-small molecule drug conjugates have reached the drug market for oncology, due to off-target effects or reduced efficacy in comparison to the free drug.

For PDCs, the linker chemistry between the drug and the polymer is key to their efficacy: the linkers must enable efficient drug release from the polymer and the release may be selective within a particular disease environment, which reduces off-target toxicity. Enzyme cleavable linkers have exhibited effective and selective drug release in other cancer therapeutics, such as antibody-drug conjugates (ADCs); these linkers are susceptible to cleavage by enzymes that are upregulated in tumours. However, the linker chemistry for PDCs has typically focused on potentially sub-optimal and non-selective hydrolytic cleavage mechanisms. In addition, the use of enzyme cleavable linkers within PDCs has been limited to the peptidic cathepsin B cleavable linkers, which can be problematic due to their hydrophobicity and instability in rodent blood. Therefore, the toolbox of linkers used in PDCs needs to be expanded to include other types of cleavable linkers.

In this work, a series of enzyme cleavable linkers, with a fluorophore payload, were designed and synthesised to investigate their compatibility within PDCs. These enzyme cleavable linkers were conjugated to amino acid-based biodegradable polymers via amide couplings. The model PDCs were then incubated with their relevant enzymes and the fluorescence intensity was measured; they exhibited effective release of the fluorophore payload, with some PDCs showing selective release at lysosomal pH compared to the pH of blood plasma. The enzyme cleavable linkers that exhibited the most effective payload release were consequently conjugated to the anti-cancer drug doxorubicin and the biological activity of these PDCs will be tested.



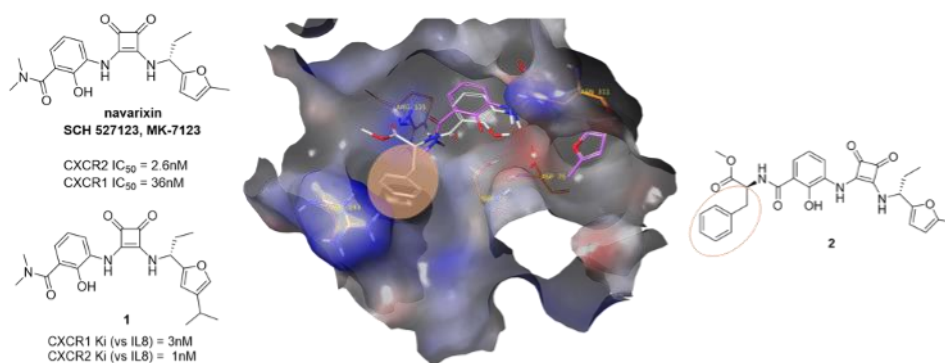
## Poster Abstracts

### P11 Exploration of Intracellular Binding Pockets of CXC Chemokine Receptors for Allosteric Modulation

R Francis<sup>1</sup>, S Mistry<sup>1</sup>, C Laughton<sup>1</sup>, N Holliday<sup>2</sup>

<sup>1</sup>School of Pharmacy, University of Nottingham, Nottingham, UK, <sup>2</sup>School of Life Sciences, University of Nottingham, Nottingham, UK

Chronic inflammation is a major contributor to tumour growth. The CXC chemokine receptor (CXCR)/CXC ligand (CXCL) axis plays a vital role in the cancer microenvironment and is attributed to proliferation, angiogenesis, invasion, and metastasis. Drug discovery programmes have attempted to target CXCR1/2 using small molecule therapeutic agents (navarixin and danirixin), however toxic side effects resulted in phase II clinical trials being terminated. Mutation studies of CXCR2 have shown that the downstream signalling can be influenced by allosteric modulators binding to an intracellular site close to the G-protein binding region present in both CXCR1 and CXCR25. The most advanced class of compounds acting at this binding site are 3,4-diaminocyclo-3-ene-1,2-dione based antagonists (navarixin and **1**). Development of novel antagonists can be aided by in-silico techniques however one of the barriers to rational drug design for CXCR1 antagonists is the limited structural data available. The aim of this work is to develop novel CXCR1 antagonists using a combination of in-silico and in-vitro methods. Molecular dynamic techniques and docking studies are being explored to construct and validate a homology model structure for the inactive, intracellular allosteric ligand bound state of CXCR1. Docking of known ligands for CXCR1 correlate with literature, validating the design and set a basis for further CXCR1 inhibitor design. Pocket and docking analysis have revealed a potential extension to the binding region between TM3 and 5. Extension of the previously described druggable binding pocket may play a key role in the design of a new class of CXCR1/2 antagonists. Ten 3,4-diaminocyclo-3-ene-1,2-dione based analogues were designed to probe this region and showed promising in-silico docking results. The series of analogues was subsequently synthesised and are under evaluation for activity at CXCR1. The in-vitro results obtained support the continued development of an accurate CXCR1 homology model along with structure-activity relationship studies for future drug design.



## Poster Abstracts

### FO12 & P12 Targeting the cell-adhesion molecule PSGL-1 with a small molecule inhibitor

C James<sup>1</sup>, V Kanabar-Raivadera<sup>2</sup>, CP Page<sup>2</sup>, L Martin<sup>1</sup>, GK Wagner<sup>1</sup>

<sup>1</sup>School of Pharmacy, Queen's University, Belfast, UK, <sup>2</sup>School of Cancer and Pharmaceutical Sciences, King's College, London, UK

P-selectin glycoprotein ligand-1 (PSGL-1), a major selectin ligand expressed on leukocytes, plays a major part in tethering blood cells to endothelial selectins. It facilitates transmigration of leukocytes and platelets into inflamed tissue and drives chronic inflammatory processes [1]. PSGL-1 is upregulated on leukocytes from patients with chronic obstructive pulmonary disease (COPD) [2]. Pharmacological reduction of elevated PSGL-1 levels may therefore be a promising strategy for regulating PSGL-1-mediated cell adhesion in chronic inflammatory airway diseases.

We have previously shown that treatment of primary human peripheral blood mononuclear cells with the pro-inflammatory cytokine IL-1 $\beta$  leads to elevated PSGL-1 levels *in vitro* [3]. Herein, we report that these elevated PSGL-1 levels can be reduced to basal levels with a drug-like small molecule (Fig.1).

The molecular target(s) and mode of action of this inhibitor are currently unknown. To enable target identification studies, we have developed an optimised synthetic scheme for this inhibitor. The target molecule was obtained in 12% total yield over 6 synthetic steps.

To identify potential molecular targets involved in the inhibitor-mediated PSGL-1 downregulation, we used 5 computational target and activity prediction servers (RF QSAR, Swiss target prediction, SEA Search Server, SuperPRED and LigTMap) [4-8]. These algorithms compare the structure and binding profiles of a query inhibitor with that of the natural ligand. Ranking of potential targets from all servers identified common candidate targets, including several kinases.

This inhibitor is hypothesized to elicit a pro-drug effect, i.e., cleaved to the subsequent acid on entry into cell. The inhibitor and its acid analogue were tested against a panel of 140 kinases using a radioactive filter binding assay. As expected, the inhibitor was less active compared to the acid analogue against these enzymes. It reduced the residual activity of six kinases to less than 60%, and of one kinase to less than 30%.

Some preliminary docking studies were conducted on the hits obtained from the screening to identify potential strategies for further optimization. Docking results suggest that both the terminal carboxylate and the formylthienyl group may engage in critical interactions with the target enzyme. Evaluation of the inhibitor and analogues in cell models are currently underway and preliminary results will also be reported.

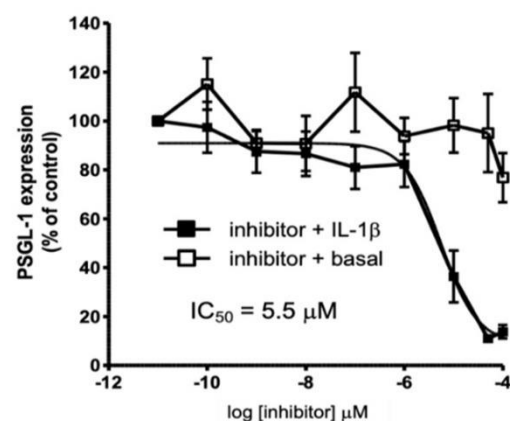


Fig. 1: Incubation with inhibitor leads to reduction of induced cell-surface PSGL-1 levels on hPBMCs treated with IL-1 $\beta$ . The inhibitor has no significant effect on constitutive cell surface PSGL-1 levels



## Poster Abstracts

### FO16 & P16 The Development of 1,2,4-Triazine G-Protein Coupled Receptor 84 (GPR84) Antagonists

Michael Malone<sup>1</sup>, Graeme Milligan<sup>2</sup>, Andrew Jamieson<sup>1</sup>

<sup>1</sup>Chemistry, University of Glasgow, Glasgow, Scotland, <sup>2</sup>Molecular Bioscience, University of Glasgow, Glasgow, Scotland

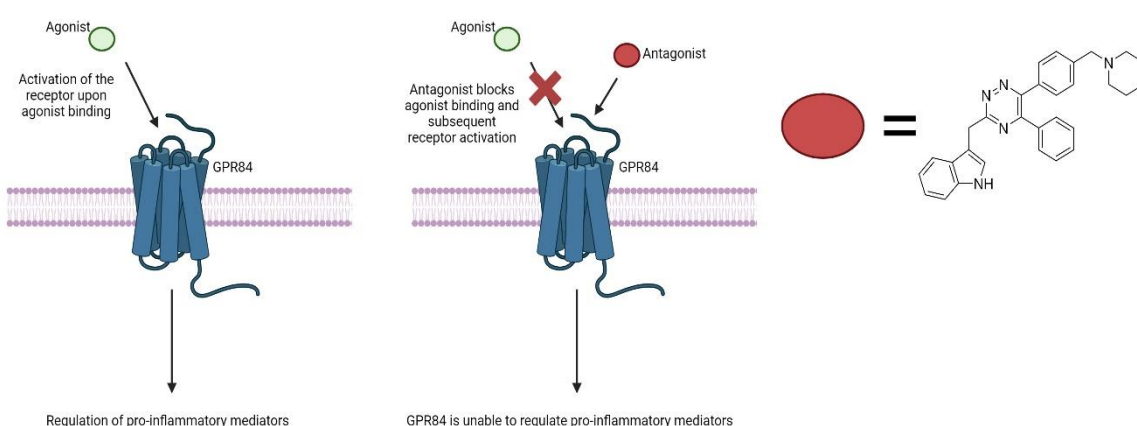
Over 800 G protein-coupled receptors (GPCRs) are encoded by the human genome with more than 30% of prescribed drugs in clinic targeting these proteins. Stimulation of GPCRs is a consequence of ligand binding, whereby the specific ligand-GPCR interaction elicits a range of cellular responses. Although extensively researched, a significant number of GPCRs require further pharmacological evaluation.

GPR84 is activated by medium-chain free fatty acids. However, due to the low potency of this class of ligand GPR84 remains categorised as an orphan receptor.<sup>1</sup> Due to the documented upregulation of GPR84 in various pro-inflammatory conditions, it is proposed that antagonising the receptor may be a potential therapeutic route for pro-inflammatory mediated diseases, such as ulcerative colitis.<sup>2</sup> Milligan, in collaboration with Jamieson, have reported the structure activity relationship (SAR) of a 1,2,4-triazine GPR84 antagonist series.<sup>1, 2</sup> However, the SAR was limited due to no current facile method known for the regioselective synthesis of unsymmetrical trisubstituted 1,2,4-triazines. Such a method is required for late-stage diversification and to further explore this series.<sup>2</sup>

This poster will outline some of the key findings surrounding the SAR of the 1,2,4-triazine GPR84 antagonist series. Additionally, the development of a regioselective synthesis of trisubstituted 1,2,4-triazines will be presented.<sup>1, 2</sup>

1 - Jenkins L, Marsango S, Mancini S, Al Mahmud Z, Morrison A, McElroy, Kirstie A. Bennett, Matt Barnes, Andrew B. Tobin, Irina G. Tikhonova, and Graeme Milligan, *ACS Pharmacol Transl Sci.* **2021**, 4(5):1598-1613.

2 - Amit Mahindra, Laura Jenkins, Sara Marsango, Mark Huggett, Margaret Huggett, Lindsay Robinson, Jonathan Gillespie, Muralikrishnan Rajamanickam, Angus Morrison, Stuart McElroy, Irina G. Tikhonova, Graeme Milligan, and Andrew G. Jamieson, *J. Med. Chem.* **2022**, 65(16): 11270-11290.



## Poster Abstracts

### FO17 & P17 Discovery of Novel Small Molecules for the Treatment of Human Coronaviruses

Elliott Smyth<sup>1,2</sup>, Joao Pisco<sup>1</sup>, Nathalie Bouloc<sup>1</sup>, Jonathan Large<sup>1</sup>, Richard Foster<sup>2</sup>

<sup>1</sup>Chemical Biology, LifeArc, Stevenage, UK, <sup>2</sup>School of Chemistry, University of Leeds, Leeds, UK

Three deadly coronavirus (CoV) infections have emerged in the last 20-years: SARS-CoV, MERS-CoV and the recent SARS-CoV-2 pandemic. SARS-CoV-2 is the latest coronavirus to infect humans, and the resulting pandemic created unprecedented challenges for healthcare systems around the world. Many of those infected with the virus experience mild symptoms or are asymptomatic, however some individuals encounter a more severe disease with long-lasting respiratory symptoms, and side-effects causing debilitating impairment and multiple organ damage. While vaccines are now widely available, patients need regular booster doses, so novel small molecule therapeutics may offer a complimentary method to addressing this disease.

One possible route to treat COVID-19, the disease caused by SARS-CoV-2, is to target the CoV-specific protein, non-structural protein 14 (NSP14). NSP14 is an essential CoV-specific protein responsible for maintaining replication fidelity, resistance to immune response and nucleoside analogue inhibitors, and increased virulence. SARS-CoV-2 NSP14 is well characterised structurally and biochemically but has not been subjected to significant inhibitor development. No marketed drug exists that targets NSP14, and there is evidence to suggest that inhibition could be a promising therapeutic strategy. The objective of this project is to synthesise and develop small molecules to selectively target NSP14, in order to create novel therapeutics for the treatment of past, present and future human coronaviruses.

A hit-to-lead medicinal chemistry campaign has been carried out for a non-nucleoside chemotype against NSP14, utilising a literature starting point with poor physicochemical properties. Biochemical assay and ADME data, as well as structural information, have been used to inform a design-make-test-analyse cycle. This cascade has ultimately led to the development of sub micromolar inhibitors of NSP14 with desirable physicochemical property profiles, which are positioned for further *in vitro* characterisation and lead development.

# Poster Abstracts

## FO18 & P18 Synthesis of Macrocyclic Ligands for the Bromodomain of CREBBP/p300

Alistair Boyd<sup>1</sup>, Mustafa Moroglu<sup>2</sup>, Stuart Conway<sup>3</sup>

<sup>1</sup>Organic Chemistry, University of Oxford, Oxford, UK, <sup>2</sup>Medicinal Chemistry, GSK, Stevenage, UK,

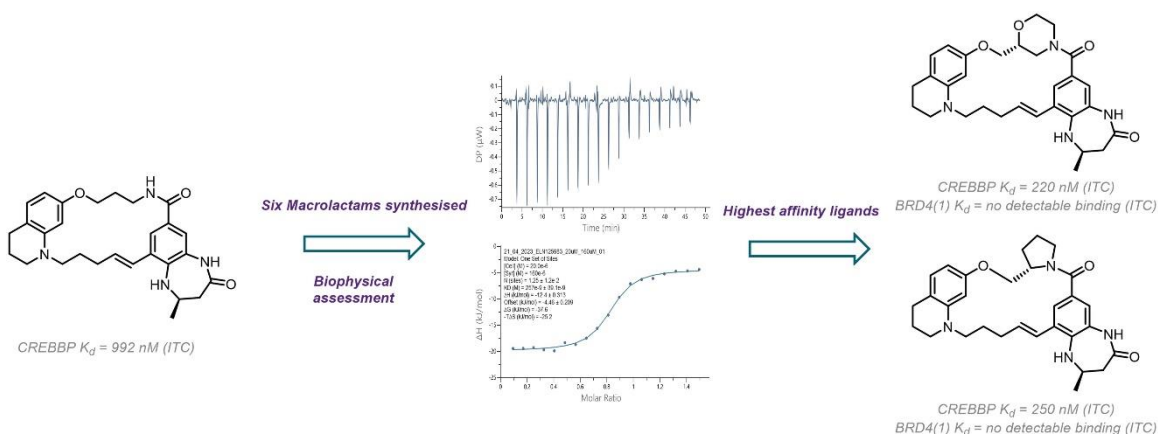
<sup>3</sup>Chemistry, UCLA, Los Angeles, United States of America

CREBBP/p300 are important proteins, comprised of ten distinct domains. One domain of particular interest within these paralogues' is the bromodomain which recognises the acetylated lysine tails of histones around which DNA is supercoiled.

Dysregulation of these proteins is implicated in several cancers including lymphatic and prostate, with therapies currently undergoing clinical trials that target these proteins.

In this work a novel approach has been undertaken to selectively target the bromodomains of CREBBP/p300, through the design and synthesis of macrocyclic ligands. These compounds are based upon an existing scaffold demonstrating an alternative conformation upon CREBBP when compared to that of the main off-target protein of BRD4(1).

This work builds upon previous macrolactam based ligands which bound the bromodomain of CREBBP with an affinity of around 1  $\mu$ M. The work presented herein improves upon the previous design via synthesis of a small series of macrocyclic ligands, derived from a common intermediate. The resulting compounds' biophysical properties were assessed, and the data produced gave a range of binding affinities, with the two highest affinity compounds producing  $K_d$  values below 300 nM by ITC. Utilising the same assay, no binding was observed to BRD4(1) with all but one of the compounds. These data mean that four of the compounds have improved binding affinities compared to the macrocycle from which they were derived. With this data in-hand we hope to further assess the selectivity and potency of these compounds. This work demonstrates the synthesis of the first series of high affinity macrocyclic ligands for the CREBBP/p300 bromodomain.



## Poster Abstracts

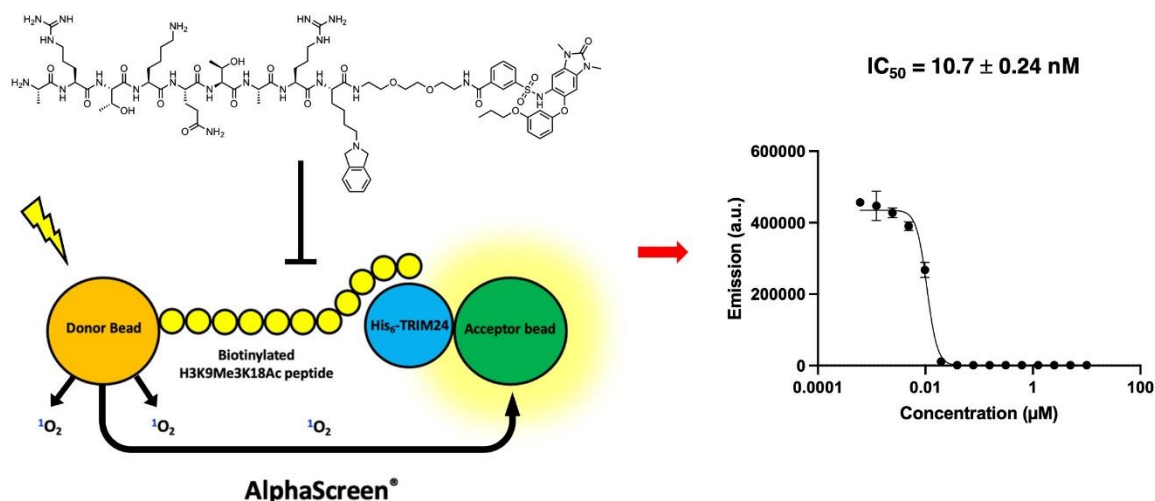
### FO 19 & P19 Bivalent chemical tools for investigating the tandem plant homeodomain finger-bromodomain cassette in TRIM24

Michael Platt, Stuart Conway

Department of Chemistry, University of Oxford, Oxford, UK

Tripartite motif-containing protein 24 (TRIM24) is an epigenetic reader and E3 ligase protein involved in multiple biological processes including innate immunity and cell proliferation. Aberrant expression of TRIM24 has been observed in breast and prostate cancers, making the protein an attractive therapeutic target. Among its various functions, TRIM24 acts as an epigenetic reader of post translational modifications (PTMs) on histone tails. Its tandem plant homeodomain (PHD) finger-bromodomain (BRD) binds to histone 3 proteins carrying the K9 trimethylation (H3K9Me<sub>3</sub>) or K18/K23 acetylation (H3K18/K23Ac) marks, respectively. In addition, studies have shown binding of a histone peptide *in cis* to the TRIM24 PHD and BRD, which raises the possibility of combinatorial readout.<sup>1</sup> However, the biological consequences of TRIM24 reading multiple PTMs on a single histone remain unknown; and there is, therefore, a need for development of chemical probes binding to *both* the PHD and BRD.

Currently, only two high affinity TRIM24 BRD ligands exist in the literature; while no TRIM24 PHD inhibitors have been reported.<sup>2,3</sup> BRD inhibition alone only partially displaces histones as it does not prevent binding to the PHD. Whereas bivalent chemical tools, which can simultaneously bind to both domains, could completely abolish histone binding to TRIM24. We have taken this approach and synthesised the first peptidomimetic bivalent compounds targeting the TRIM24 PHD and BRD. In AlphaScreen<sup>®</sup> competition assays, these compounds demonstrated low nanomolar IC<sub>50</sub> values (10–30 nM) for the displacement of a dual-modified PHD-BRD binding histone peptide (H3K9Me<sub>3</sub> K18Ac); and an approximate 80-fold decrease in IC<sub>50</sub> compared to the parent BRD ligand. Furthermore, these compounds showed superior inhibition over the current highest affinity TRIM24 BRD ligand, IACS-9571.<sup>3</sup> Future work with these compounds will enable elucidation of the biological consequences of chromatin association to TRIM24 and help us understand the biological functions of TRIM24 in more detail.



1 Tsai, W. W. *et al. Nat.* **2010**, 468 (7326), 927–932. [doi:10.1038/NATURE09542]

2 Bennett, J. *et al. J. Med. Chem.* **2015**, 59 (4), 1642–1647. [doi:10.1021/ACS.JMEDCHEM.5B00458]

3 Palmer, W. S. *et al. J. Med. Chem.* **2015**, 59 (4), 1440–1454. [doi:10.1021/ACS.JMEDCHEM.5B00405]

## Poster Abstracts

### FO20 & P20 Design, Synthesis, Biological Evaluation, and Molecular Modeling of Novel Benzofuran Derivatives as Targeted Cancer Chemotherapy

A A. Osman<sup>1,2</sup>, E R. Mohamed<sup>2</sup>, H A. Abdel-Aziz<sup>3</sup>, A M. El Kerdawy<sup>2,4</sup>, H Abdelrasheed Allam<sup>2</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmacy, NewGiza University, Newgiza, Cairo 12256, Egypt,

<sup>2</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Cairo University, Cairo, P.O. Box 11562, Egypt,

<sup>3</sup>Applied Organic Chemistry Department, National Research Center, Dokki, Cairo 12622, Egypt, <sup>4</sup>School of Pharmacy, College of Health and Science, University of Lincoln, Green Lane, Lincoln, Lincolnshire, UK

Cancer remains one of the major causes of morbidity and mortality worldwide, and for decades researchers have been trying to identify the origin of human cancer 1,2. Cancer is characterized by the dysregulation of several enzymes that play essential roles in cell division and differentiation such as protein kinases. For instance, epidermal growth factor receptor (EGFR) plays a substantial role in regulating tissue development and homeostasis 3. Thus, inappropriate activation of EGFR can lead to cancer initiation and progression 4. Benzofuran is a privileged scaffold that has been used to design several new anticancer agents which could inhibit the growth of multiple human cancer cell lines 5,6. Moreover, thiazol-2-yl-hydrazone moiety is a key fragment in several EGFR inhibitors 7,8. Thus, in the current work, a set of hybrid compounds containing both privileged fragments were designed and synthesized as novel EGFR inhibitors. Additionally, molecular docking simulations were performed for the designed compounds in the EGFR active site to confirm their binding capability to the EGFR kinase domain. All the newly synthesized compounds were capable of binding to the key amino acid Met768 in the hinge region of the kinase domain. Furthermore, SwissADME tool was used to predict the physicochemical and pharmacokinetic properties of the target compounds and to check their medicinal chemistry friendliness. Our future prospective is to evaluate the EGFR inhibitory activity as well as the cytotoxic activity of the designed compounds.

## Organising Committee



# RSC INTEREST GROUP BIOLOGICAL AND MEDICINAL CHEMISTRY SECTOR

*David Alker (Treasurer)*

David Alker Associates

*Katherine Jones*

Charles River

*Tom Lanyon-Hogg*

University of Oxford

*Angela Russell*

University of Oxford

*John Skidmore*

University of Cambridge

*Mary Wheldon*

University of Dundee

*Douglas Williamson (Chair)*

Transition Bio

Hg3 Conferences Ltd

Secretariat



## RSC BMCS Forthcoming Events



# RSC INTEREST GROUP BIOLOGICAL AND MEDICINAL CHEMISTRY SECTOR

### *Forthcoming Events in 2024*

2024

3 <sup>rd</sup> – 5 <sup>th</sup> March	9 <sup>th</sup> RSC-BMCS Fragment-based Drug Discovery Meeting Cambridge, UK
7 <sup>th</sup> March	BMCS Conformational Design in Drug Discovery Stevenage, UK
15 <sup>th</sup> March	The BMCS Mastering MedChem VIII: 8 <sup>th</sup> RSC-BMCS Symposium on Mastering Medicinal Chemistry London, UK
25 <sup>th</sup> April	35 <sup>th</sup> Medicinal Chemistry in Eastern England Stevenage, UK
16 <sup>th</sup> May	Hot Topics: Covalent Drug Discovery 2024 virtual
4 <sup>th</sup> – 5 <sup>th</sup> June	4 <sup>th</sup> Synthesis in Drug Discovery and Development Virtual
9 <sup>th</sup> – 11 <sup>th</sup> September	3 <sup>rd</sup> RSC / SCI New Therapeutics for Global Health Milton Keynes, UK
16 <sup>th</sup> - 18 <sup>th</sup> September	7 <sup>th</sup> RSC-CICAG/RSC-BMCS Artificial Intelligence in Chemistry Cambridge, UK
10 <sup>th</sup> – 13 <sup>th</sup> October	9 <sup>th</sup> GPCR's in Medicinal Chemistry Verona, Italy



## RSC BMCS Initiatives and Announcements



# RSC INTEREST GROUP BIOLOGICAL AND MEDICINAL CHEMISTRY SECTOR

Royal Society of Chemistry – Biological and Medicinal Chemistry Sector

### Social Media

#### Social Media Links

Please join as many of the groups below as you wish to receive the latest news from the RSC BMCS:

Website	<a href="https://www.rscbmcs.org">https://www.rscbmcs.org</a>
Twitter	@RSC_BMCS
LinkedIn	linkedin.com/company/rsc-bmcs

### Initiatives and Announcements

#### RSC-BMCS Membership Subscription Renewal

The BMCS organises a range of high quality conferences and runs initiatives that support the education and training of the next generation of medicinal chemists. We also have an outreach programme which supports chemistry clubs and provides enhanced equipment to schools. When renewing your RSC membership subscription, please remember to tick the BMCS membership box as this will allow us to keep you informed of future BMCS events and initiatives. Please pass this notice on to any non-member colleagues who may be interested in joining the BMCS or attending any of our events.

#### PhD Student and Post-Doc Conference Bursaries

Did you know that most BMCS sponsored meetings have a number of bursaries available for PhD and post-doctoral students? Normally these awards are up to a value of £350 for one-day events and up to £700 for multi-day events, and help to cover costs, including registration and travel. Preference will be given to members of the RSC (and meeting co-sponsors if applicable), especially those who are selected to give posters.

# RSC BMCS Initiatives and Announcements

## The BMCS International Travel Prize

Each year, the BMCS offers a number of International Travel Prizes for use in the following year, up to a value of £2,000, to postgraduate students enrolled in research programmes (PhD or Masters) in UK institutions. The call for nominations will open on 1<sup>st</sup> March 2024.

## The RSC BMCS Lectureship

Every two years, the BMCS seeks nominations of BMCS lectures to deliver lectures. These include an initial lecture as part of the RSC series of Thursday evening public lectures at Burlington House. The next call for nominations will be September 2024.

BMCS Lecturer for 2019/2020 was Dr David Witty formerly of Convergence.

BMCS Lecturers for 2020/2021 was Elisabetta Chiarparin of AstraZeneca and James Duffy of Medicines for Malaria Venture.

BMCS Lecturer for 2021/2022 was David Rees of Astex Pharmaceuticals.

BMCS Lecturer for 2022/2023 is James Crawford of Genetech.

BMCS Lecturer for 2023/2024 is Dafydd Owen of Pfizer

## The RSC BMCS Researcher Mobility Fellowship

The BMCS is pleased to announce a new call for the Researcher Mobility Fellowships, to drive excellence in the UK-based science. These fellowships of up to £5,000 aim to support short-term placements (up to 3 months) between industry and academia in areas of interest to the BMCS, namely bioorganic chemistry, medicinal chemistry, chemical biology and agriscience. Applications should be jointly submitted by academic and industrial partners. Call for nominations open in January 2024.

## Hall of Fame and Medal

The BMCS is pleased to announce a new call for nominations for The Hall of Fame and associated medal, to recognize prominent chemists for outstanding, sustained, significant contributions to any area of interest to the BMCS, e.g. medicinal chemistry, agriscience, bioorganic chemistry, chemical biology. Call for nominations for the Hall of Fame 2024 close on 31<sup>st</sup> March 2024.

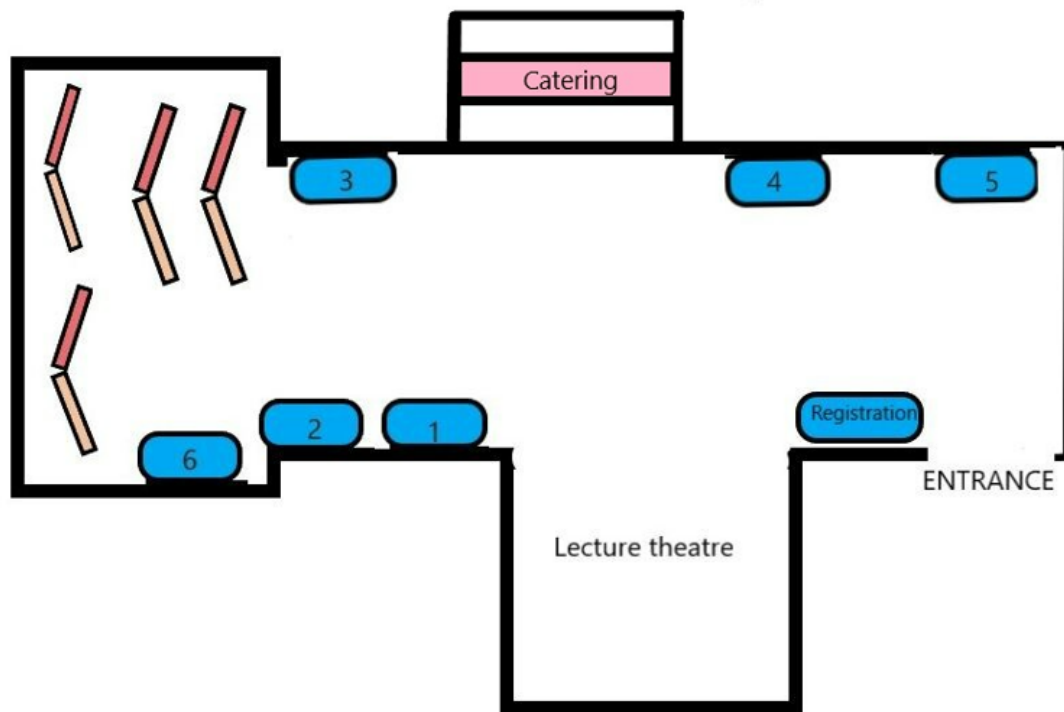
## Websites

<https://www.rscbmcs.org>

<https://www.rscbmcs.org/awards/> and <https://www.rscbmcs.org/grants/>

## Exhibitor floor plan

### BMCS Postgraduate Symposium XVII Exhibitor Floor plan



#### Exhibition stands

- 1 – Asynt
- 2 – Biosynth
- 3 – Sygnature Discovery
- 4 – Collaborative Drug Discovery
- 5 – Medchemexpress
- 6 - Stratech Scientific Ltd

## Catalogue of Exhibitors

We are grateful to our exhibitors for their support of this meeting

Asynt 

BIOSYNTH® 

 **CDD.VAULT**<sup>®</sup>  
Complexity Simplified

 **MCE**<sup>®</sup>  
MedChemExpress

 **Stratech**

**SYGNATURE**  
**DISCOVERY** 



# Asynt

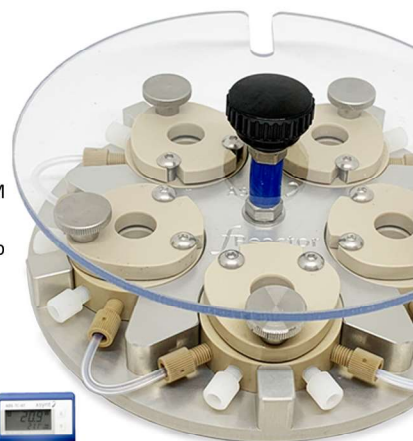
## Cutting edge technologies for every laboratory

Get up and  
with **GLOW**  
LightSyn Illumin8



Revel in the pressure  
with **PressureSyn & Quadracell**

Go with the *flow*  
with **fReactor™**  
...one for every lab



**DrySyn:**  
**CondensSyn:**  
**Spiral Evaporator:**  
**DrySyn OCTO:**

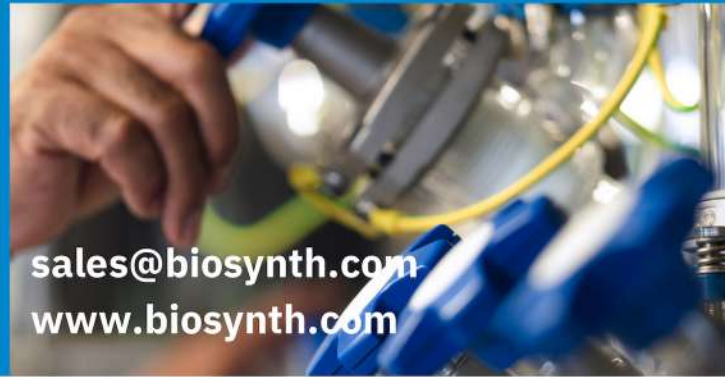
single or parallel safe, clean, oil-free heating  
outstanding performance waterless air condensers  
brilliant, even with high-boiling solvents  
powerful magnetic stirring and heating, inert  
atmosphere and reflux all in a tiny footprint

**Asynt.com**

**Talk to us today... [enquiries@asynt.com](mailto:enquiries@asynt.com)**



Welcome to  
the Edge of  
Innovation



[sales@biosynth.com](mailto:sales@biosynth.com)  
[www.biosynth.com](http://www.biosynth.com)

Products



Services

Where Products meets Services  
Where Chemistry meets Biology  
Where Innovation meets Quality

On the Edge of Innovation:  
We are **Biosynth**

Our new branding highlights our key strength - providing everything for our customers in one place.

Science led and customer focused from the first idea to the finished product.

Our new brand pattern evokes The Edge and our previous DNA Helix pattern



## Securing Life Sciences Supply Chains

### Complex Chemicals

Route Scouting & Optimization  
Small-Scale Manufacture  
GMP Manufacture  
Large-Scale Manufacture  
Analytical Development and QC  
Quality Assurance and Regulatory Support

Carbohydrates, Nucleosides'  
Enzyme Substrates  
Chemical APIs and intermediates  
Formulation ingredients and Excipients

### Quality Peptides

Lead Discovery & Optimization  
Small Scale Synthesis & Arrays  
GMP Manufacture  
Large-Scale Manufacture  
Analytical Development and QC  
Quality Assurance and Regulatory Support

Neoantigen Peptides  
Peptide APIs  
Proteomics Tools

[sales@biosynth.com](mailto:sales@biosynth.com)

### Key Biologics

Antibodies  
Antigens  
Proteins  
Enzymes  
Epitope Mapping

Custom raw materials for in vitro diagnostics  
Custom Antibody Production  
Bioprocessing Enzymes

[www.biosynth.com](http://www.biosynth.com)

### Research Products

0.5M+ products

Carbohydrates  
Nucleosides  
Enzyme Substrates  
Natural Products  
Ligands  
Biochemicals  
Peptides  
Biologics





# CDD VAULT® Complexity Simplified

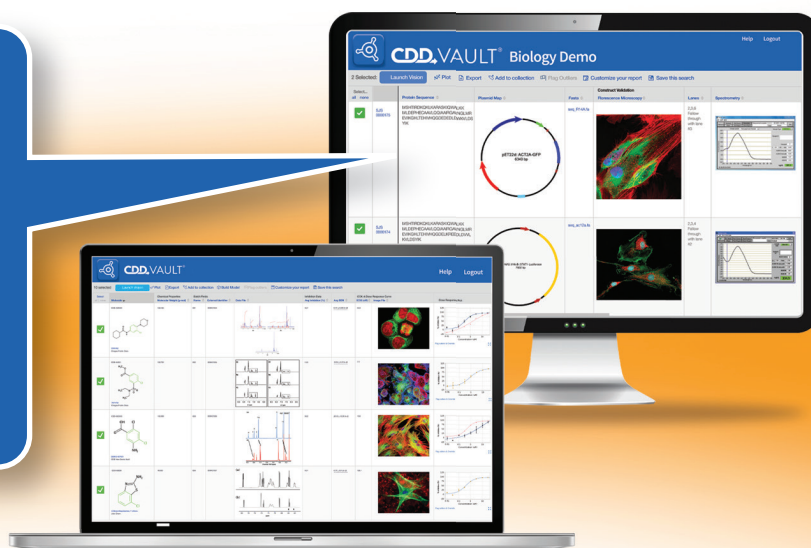
## futureproof Your CRO Laboratory

### Plan, capture, and share experimental data in one secure informatics platform

In today's digital age, customers are expecting CROs to document high-quality experiment results electronically. Give yourself a competitive edge by using CDD Vault, a flexible

and intuitive hosted software that allows you to organize chemical and biological data in one platform and securely collaborate with external partners.

- ✓ Digitalize your lab the easy way
- ✓ Onboard your team in 90 minutes
- ✓ Document experiments with CDD Vault Registration and ELN
- ✓ Securely share data in real time with customers





# CDD VAULT® Complexity Simplified

## flexible licensing

Choose the model that meets your needs



### Basic Model

The CRO sets up one Vault with multiple Projects to hold data for each customer. Data can be exported and sent to the customer as needed.

**Best for: Low cost and maintenance**



### Collaborative Model

Customers are granted access to their Project within the CRO's Vault. This allows for real-time data visibility.

**Best for: Quicker data access without needing to send emails**



### Individual Vault Model

An individual Vault is created for each customer. The CRO works and generates data within the customer's Vault.

**Best for: The customer has a fully functional Vault and the CRO never needs to transfer data**

Contact us to learn more! [info@collaborativedrug.com](mailto:info@collaborativedrug.com)

Collaborative Drug Discovery, Inc. 1633 Bayshore Hwy., Suite 342, Burlingame, CA 94010. (650) 242-5259. [www.collaborativedrug.com](http://www.collaborativedrug.com)



## Inhibitors & Agonists

**50,000+** Specific Inhibitors & Agonists

Targeting **1,000+** Key Proteins in 20 Signaling Pathways

Widely Used in Hot Spots Areas of Biology, Medicine and Pharmacology



## Drug Discovery

**200+** Bioactive Compound Libraries

Diversity Compound Libraries (**50,000+** Compounds)

Virtual Screening



## Recombinant Proteins

**10,000+** Recombinant Proteins

High Purity

Superior Biological Activity

Excellent Lot-to-lot Consistency

GMP-Grade Proteins



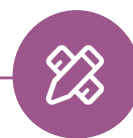
## Life Science Products

**4,000+** Bioactive Peptides

**1,000+** Ion Indicators, ROS/NO Probes, etc.

Inhibitory Antibodies

CCK-8, Magnetic Beads, Inhibitor Cocktails, etc.



## Customized Services

R&D platform for Chemistry small molecules, Oligonucleotides, ADC, PROTAC and Stable Isotope-Labeled Compounds



## Peer Scientists Recognition –

Cited in **30,000+** Publications Including CNS Series

## MCE Global Partners



### MedChemExpress EU

+46 86500910

eu.sales@MedChemExpress.com

Bergkällavägen 37C, 192 79 Sollentuna, Sweden

### MedChemExpress USA

+1609-228-6898

+1609-228-5909

1 Deer Park Dr, Suite Q, Monmouth Junction, NJ 08852, USA

sales@MedChemExpress.com

tech@MedChemExpress.com

## Who are Stratech?

Stratech Scientific Ltd supplies an extensive range of innovative and specialist life science research tools for researchers who need consistent reproducible results. We have built an excellent reputation over 40 years for supplying high quality, competitively priced, reliable products to our end users.

### our catalogue includes:



Primary  
Antibody



Secondary  
Antibody



Assay And  
Kits



DNA &  
RNA



Protein  
Science



Cell Culture  
And Media



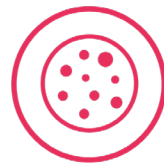
Reagents



Lipids



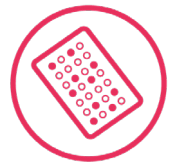
Inhibitors



Cells and  
Tissue



Equipment



ELISA kits

## contact us

[info@stratech.co.uk](mailto:info@stratech.co.uk)  
[sales@stratech.co.uk](mailto:sales@stratech.co.uk)  
[orders@stratech.co.uk](mailto:orders@stratech.co.uk)  
[technical@stratech.co.uk](mailto:technical@stratech.co.uk)



@stratech\_uk



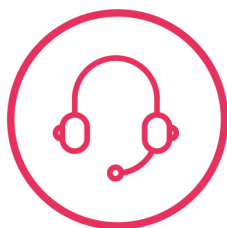
@stratechscientificltd



@stratech-scientific-ltd



plant a tree  
for free with  
every order



outstanding  
technical  
support



free delivery  
for UK  
universities



we offer a  
full product  
guarantee



See us at booth  
**#3**

# SYGNATURE DISCOVERY

## Discover your next breakthrough

Providing experienced, innovative and multi-disciplinary drug discovery teams to support customer projects across a wide range of therapeutic areas and target classes

WITH 1,000 TEAM MEMBERS  INCLUDING >900 SCIENTISTS

WHO HAVE SUCCESSFULLY DELIVERED:

**>40**

PRECLINICAL  
COMPOUNDS

**24**

CLINICAL  
COMPOUNDS

**>170**

PATENT  
APPLICATIONS



TARGET  
VALIDATION



HIT  
IDENTIFICATION



HIT TO  
LEAD



LEAD  
OPTIMIZATION



LATE LEAD  
OPTIMIZATION

[sygnaturediscovery.com](http://sygnaturediscovery.com)

Upcoming Events

**EFMC-ACSMEDI Medicinal Chemistry Frontiers 2024**  
**Joint Symposium on Medicinal Chemistry**  
Utrecht, The Netherlands | April 8-11, 2024



**18th EFMC Short Course on Medicinal Chemistry**  
**Transformative Approaches in Medicinal Chemistry,**  
**with Applications in Cancer Research**  
Oegstgeest, The Netherlands | April 21-24, 2024



**EFMC-ISM 2024**  
**XXVIII EFMC International Symposium on Medicinal Chemistry**  
Rome, Italy | September 1-5, 2024



**EFMC-YMCS 2024**  
**11th EFMC Young Medicinal Chemists' Symposium**  
Rome, Italy | September 5-6, 2024



Awards

— The Nauta Pharmacochimistry Award for Medicinal Chemistry and Chemical Biology  
— The “UCB-Ehrlich Award for Excellence in Medicinal Chemistry”  
— Prous Institute - Overton and Meyer Award for New Technologies in Drug Discovery  
— The “EFMC-WuXi AppTec Award for Excellence in Chemical Biology”  
Visit [www.efmc.info/awards](http://www.efmc.info/awards) for more information

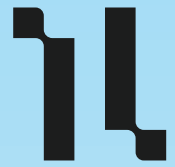
Prizes

— EFMC Prizes for Young Medicinal Chemist or Chemical Biologist in Industry & Academia  
— EFMC-YSN PhD Prize  
— Symeres PhD Prize for Excellence in Chemistry in Life Sciences Research  
Visit [www.efmc.info/prizes](http://www.efmc.info/prizes) for more information

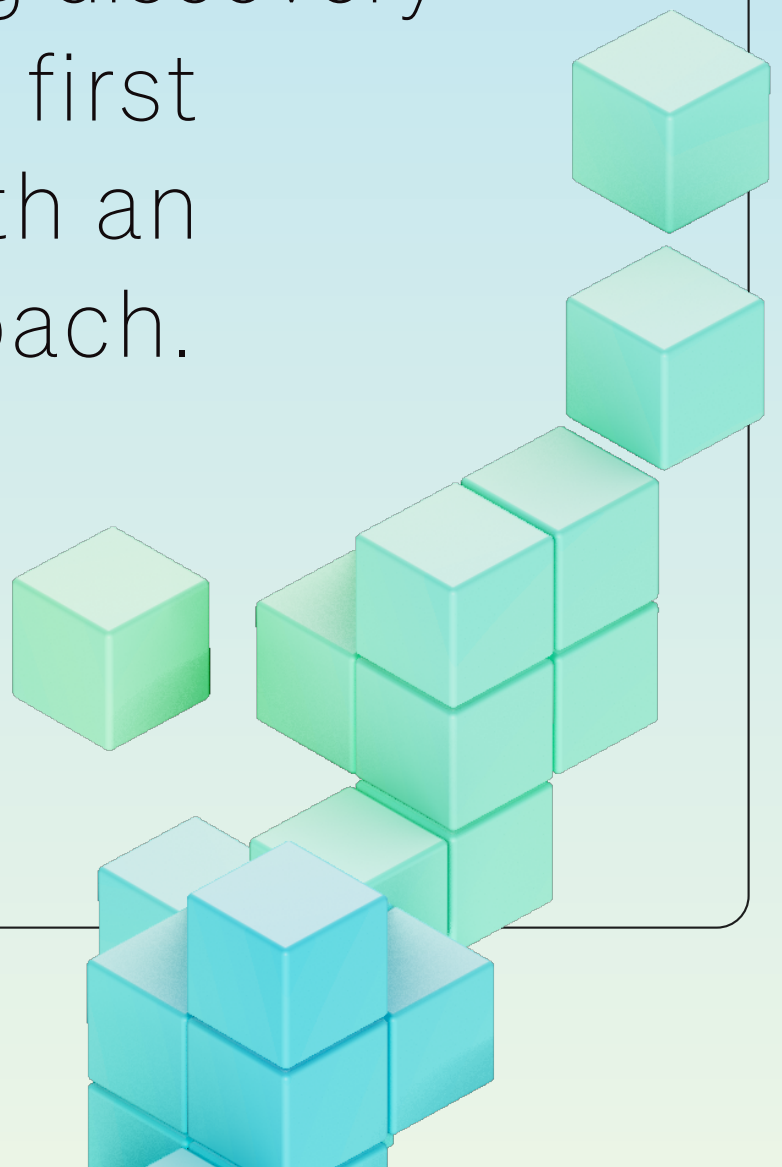
EFMC-YSN

**The Young Scientists Network**  
Building a strong network at an early stage in your career is crucial!  
The aim of the EFMC-YSN is to **inspire, connect** and **provide opportunities** to medicinal chemists and chemical biologists in their Early Career.  
Visit [www.efmc.info/ysn](http://www.efmc.info/ysn) for more information

Join us  
[isomorphiclabs.com/join](https://isomorphiclabs.com/join)  
[talent-acquisition@isomorphiclabs.com](mailto:talent-acquisition@isomorphiclabs.com)



Be a part of reimaging  
the entire drug discovery  
process from first  
principles with an  
AI-first approach.



## Attendee List

Firstname	Lastname	Company	Job Title	Registration contact email
Ruqaiya	A	King's College London	Student	ruqaiya_786@hotmail.co.uk
Gulijiannaiti	Abulaiti	King's College London	PhD student	k21073545@kcl.ac.uk
Md Sabir	Alam	SGT University	Associate Professor	mdsabir_pharmacy@sgtuniversity.org
David	Alker	David Alker Associates	Consultant	dave.alker@btinternet.com
Danah	Alshaer	Evotec	Scientist I	danah.alshaer@evotec.com
Cedric	Amengor	University of Health and Allied Sciences	Lecturer	camengor@uhas.edu.gh
Matt	Anderson	University Newcastle	PhD student	m.anderson9@newcastle.ac.uk
Charlotte	Avery	UCB	Medicinal Chemist IP Student	charlotte.avery@ucb.com
Christos	Avgerinos	University of Copenhagen	PhD	christos.avgerinos@sund.ku.dk
Alaa	Awad Taha Gad Osman	Cairo University	Master student	alaaawad.osman@icloud.com
Hannah	Baillie	University of Oxford	DPhil Student	Hannah.baillie@linacre.ox.ac.uk
Benoit	Baillif	University of Cambridge	PhD student	bb596@cam.ac.uk
Gabrielle	Bangay	Universidade Lusófona, Lisbon	PhD student	p6978@ulusofona.pt
Patricia	Barbosa	Charnwood Discovery	Research Scientist	patriciasbarb@gmail.com
Charlotte	Bayley	The Institute of Cancer Research	PhD student	charlotte.bayley@icr.ac.uk
Daniil	Bazanov	Lomonosov Moscow State University	Senior lecturer	daniil_bazanov@mail.ru
Elizabeth	Bedwell	University of Durham	PhD Candidate	Elizabeth.bedwell@durham.ac.uk
Alex	Bentley	GSK/University of Strathclyde	PhD student	alex.x.bentley@gsk.com
Dmitriy	Berillo	Satbayev University	Associate Professor	berillo.d@kaznmu.kz
Siddheshwar	Bhujbal	Gwangju institute of science and technology	PhD student	sidbhujbal77@gm.gist.ac.kr
Helen	Boffey	The Alborada Drug Discovery Institute	Team Leader	hkb32@medschl.cam.ac.uk
Jacob	Bradbury	University of Oxford	DPhil Student	jacob.bradbury@sjc.ox.ac.uk
Skye	Brettell	University of Glasgow	Post Graduate Researcher	2196732b@student.gla.ac.uk
Jemima	Brimacombe	University of Oxford	DPhil student	jemima.brimacombe@new.ox.ac.uk
Annabel	Brunt	University of Oxford	DPhil Student	annabel.brunt@chem.ox.ac.uk
Martin	Buttenschoen	University of Oxford	DPhil student	martin.buttenschoen@stats.ox.ac.uk
Bekir Caglar	Celikkaya	University of Strathclyde	PhD student	bekir.celikkaya@strath.ac.uk
Lal	Cheema	N/A	Chemist	lcheema10@talktalk.net
Julian	Chesti	University of Leeds	PhD Researcher	cmjch@leeds.ac.uk
Thomas	Corner	University of Oxford	PhD student	thomas.corner@chem.ox.ac.uk
Constance	Dalton	University of Nottingham	PhD student	pcycd4@nottingham.ac.uk
Josef	Dann	Evotec	Discovery chemist	josef.dann@evotec.com



## Attendee List

Yao	Ding	King's College London	PhD candidate	yaoyu.ding@kcl.ac.uk
Kristina	Elersic	N/A	Researcher	kristina.elersic@guest.arnes.si
Holly	Evans	Science Solutions Recruitment	Scientific Recruitment Specialist	holly.evans@sciencesr.com
Azeza	Fdel	Cardiff University	Research Student	fdela@cardiff.ac.uk
Viktor	Filatov	SkyLab AG	Researcher, Lead Innovation Manager	fivitya@yandex.ru
Barbara	Forte	University of Dundee	Team Leader/Project manager	b.forte@dundee.ac.uk
Ceri	Foster	University of Oxford	DPhil student	ceri.foster@chem.ox.ac.uk
Rhys	Francis	University of Nottingham	PhD student	rhys.francis@nottingham.ac.uk
Miguel	Garcia-Ortegon	University of Cambridge	PhD student	mg770@cam.ac.uk
Efthymios S.	Gavriil	Imperial College London	Research Associate	e.gavriil@imperial.ac.uk
Mhairi	Gibson	University of Strathclyde	PhD student	mhairi.gibson.2015@uni.strath.ac.uk
Nick	Gocher	Evotec	Scientist	nick.gocher@evotec.com
Andres	Gomez Angel	University of York	PhD student	arga500@york.ac.uk
Janine	Gray	Imperial College London	Research Associate	janine.gray@imperial.ac.uk
Abdulbasit	Haliru Yakubu	University of Southampton	PhD Reseacher	a.h.yakubu@soton.ac.uk
Charlotte	Hampton	Domainex Ltd	Senior Scientist I	charlotte.hampton@domainex.co.uk
Renate	Hans	University of Namibia	SEnior Lecturer/Researcher	rhans@unam.na
Irene	Herrera González	Universidad de Salamanca	Postdoctoral Researcher	irehego@gmail.com
David	Hewings	Vertex Pharmaceuticals	Principal Scientist	david_hewings@vrtx.com
Matthew	Holland	Centre for Medicines Discovery, University of Oxford	Doctoral Student	matthew.holland@cmd.ox.ac.uk
Romilly	Hryczanek	Evotec	Scientist	romilly.hryczanek@evotec.com
Ali	Hussain	University of Tameer-e-Millat		faizanmahmood042000@gmail.com
Nazir	Hussain	University of Ancient Sciences	Associate Professor	abrahamad071994@gmail.com
Ciyana	James	Queen's University Belfast	PhD student	cjames04@qub.ac.uk
Risy Namratha	Jamullamudi	Koneru Lakshmaiah College of Pharmacy	Assistant Professor	r.namratha747@gmail.com
Mohan	Karwal	MedchemExpress	Sales Manager	mohan.karwal@medchemexpress.com
Jean-Pierre	Kayembe Kayembe	North-West	PDF fellow	jeanpkayembe@gmail.com
Md. Zaved Hossain	Khan	Jashore University of Science and Technology	Chairman and Associate Professor	zaved.khan@just.edu.bd
Evangelia	Konstantinou	University of Peloponnese	Msc Chemist, PhDc	chrisaeva0@gmail.com
Kristina	Kostadinova	University of Cambridge	PhD student	kak48@cam.ac.uk
Jedrzej	Kukuloiwcz	Jagiellonian University Medical College	PhD candidate	jedrzej.kukulowicz@doctoral.uj.edu.pl

## Attendee List

Thomas	Lanyon-Hogg	University of Oxford	Group Leader	thomas.lanyon-hogg@pharm.ox.ac.uk
Gennady B	Lapa	Pirogov Medical University	Associate professor	lapa_g@mail.ru
Aditya	Lavekar	Pi industries	Research scientist	aaditya.lavekar@gmail.com
Christine	Law	Monash University	PhD student	christine.law@monash.edu
Rhianna	Lenham	University of Nottingham	PhD student	Rhianna.Lenham@nottingham.ac.uk
Roba	Maghrebi	Postgraduate	Medical product specialist	roba.mo@hotmail.com
Michael	Malone	University of Glasgow	Post-graduate Researcher	m.malone.1@research.gla.ac.uk
Francesca	Mangiavacchi	University of Florence	Post-Doc Fellow	francesca.mangiavacchi@unifi.it
Ezekiel	Mariam	Cambridge Crystallographic Data Centre	Snr Business Development Executive	emariam@ccdc.cam.ac.uk
Jared	Marklew	Vernalis Research	Senior Scientist II	j.marklew@yahoo.co.uk
Elena	Martino	University of Turin	PhD	e.martino@unito.it
David	Matthews	Queen's University Belfast	PhD Candidate	dmatthews12@qub.ac.uk
Reuben	McKay	University of Bradford	Postgraduate Researcher	rcmckay@yahoo.com
Eilidh	McKay	GSK/University of Strathclyde	PhD student	eilidh.x.mckay@gsk.com
Sam	McKee	Evotec	Senior Scientist	sam.mckee@evotec.com
Anuradha	Mehra	Lovely Professional University	Assistant Professor	anu.2004m@gmail.com
Ana	Mesa	University of Antioquia	Professor	amaria.mesa@udea.edu.co
Rangan	Mitra	Banaras Hindu University	PhD Research Scholar	ranganmitra.rs.phe19@itbhu.ac.in
Jahasultana	Mohammed	Koneru Lakshmaiah College of Pharmacy	Assistant Professor	sohnivya786@gmail.com
Rebecca	Morley	Evotec	Scientist I	rebecca.morley@evotec.com
Nada	Mosallam	University of Liverpool	Post-Doctoral Research Associate	Nada.Mosallam@liverpool.ac.uk
VENUGOPAL	MURALIDHARAN	Vishnu Institute of Pharmaceutical Education and Research	Associate Professor	vmd1213@gmail.com
Gauri alias Pooja Mathurdas	Naik	Lovely Professional University	Research Scholar	pooja90gauri@gmail.com
Venkata Naga Jyothi	Nakka	Koneru Lakshmaiah College of Pharmacy	Research Scholar	nvn.jyothi567@gmail.com
Subhendu	Nayak	CSIR-National Chemical Laboratory,Pune	Project Associate-I	snjio1998@gmail.com
Elena	Nomerotskaia	Immunocore	Process Development Scientist	elena.nomerotskaia@immunocore.com
Ramya	Nuti	Queen's University Belfast	PhD student	rnuti01@qub.ac.uk
Oluwatobi	Otun	University of the Witwatersrand (WITS)	Postdoctoral Research Fellow	oluwatobi.otun@wits.ac.za
Harry	Palmer	University of Strathclyde	PhD student	harry.x.palmer@gsk.com
Anjoomaara Hidayatullah	Patel	University of Central Lancashire	Student	AHPatel4@uclan.ac.uk

## Attendee List

Rachel	Pearce	Domainex Ltd	Director of Business Development	rachel.pearce@domainex.co.uk
Julian	Perfect	New York University London	Senior Demonstrator in Organic Chemistry	julianperfect@hotmail.com
George	Phillips	Domainex Ltd	Senior Scientist I	george.phillips@domainex.co.uk
Sarah	Phillips	University of Cambridge	PhD student	sp2103@cam.ac.uk
Michael	Platt	University of Oxford	DPhil student	map1197@yahoo.co.uk
Erika	Plazas	Acadia University	Postdoctoral fellow	eaplazasg@unal.edu.co
Maria	Popescu	UCL	PhD student	maria.popescu.23@ucl.ac.uk
Jonathon	Puleston	University of Strathclyde	PhD student	jonathonpuleston@gmail.com
Alina	Qaisar	Trinity College Dublin	PhD student	qaisara@tcd.ie
Reg	Richardson	ChemBridge Corp	Business Manager	reg@chembridge.com
Subrata	Roy	CSIR-CDRI	RESEARCHER	SUBRATAROY65@GMAIL.COM
Massimiliano	Runfola	University of Oxford	Marie Curie Postdoctoral Fellow	massimiliano.runfola@phar.ox.ac.uk
Wadhah	Salem	Yemen Supreme Board of Drugs and Medical Appliances	Manager of QMS	wadhahatef@gmail.com
Emily	Sampey	University of Manchester	PhD student	Emily.sampey@postgrad.manchester.ac.uk
Lame	Senatla	Imperial College London	PhD student	gaseitsiwe.senatla16@imperial.ac.uk
Mobeen	Shaik	Koneru Lakshmaiah College of Pharmacy	Assistant Professor	mobeenshaik@kluniversity.in
Ganesh	Shelke	Biosynth Ltd	Senior synthetic chemist	Ganesh.shelke@biosynth.com
Monika	Singh	Shoolini University	Assistant Professor	monikasagar999@gmail.com
Shivam Kumar	Singh	Lovely Professional University	Student	shivamkumarsingh435@gmail.com
Christos	Siokatas	Aristotle University of Thessaloniki	PhD Candidate	siokatas@chem.auth.gr
John	Skidmore	University of Cambridge	CSO, ALBORADA Drug Discovery Institute	js930@cam.ac.uk
Sofia	Srdanovic	Imperial College London	PhD researcher	ss8422@ic.ac.uk
Andrew	Stachulski	University of Liverpool	Research fellow	stachuls@liv.ac.uk
Dorota	Stary	Jagiellonian University Medical College	PhD student	dorota.stary@doctoral.uj.edu.pl
Sadiya	Tanga	Ashoka University	Phd Student	sadiya.tanga_phd21@ashoka.edu.in
Emilia	Taylor	University of Oxford	PhD Candidate	Emilia.taylor@stx.ox.ac.uk
Shivani	Thakur	Indian Institute of technology Bhilai	PhD student	shivanihakur@iitbhilai.ac.in
Biagio	Todaro	Scuola Normale Superiore	Postdoctoral Researcher	biagio.todaro@sns.it
Ibrahim	Tolaymat	Anglia Ruskin University	Senior lecturer	ibrahim.tolaymat@aru.ac.uk
Dritan	Topi	University of Tirana	Lecturer	dritan.topi@unitir.edu.al

## Attendee List

Harashkumar	V T	Vellore Institute of Technology	PhD Scholar	harashkumar.vt2020@vitstudent.ac.in
Muhammad	Wahajuddin	University of Bradford	Lecturer	wahajuddin@gmail.com
Rebekah	West	University of Cambridge	PhD Student	rw603@cam.ac.uk
Declan	Wolverson	Activate Scientific	Head of Sales	dwolverson@activate-scientific.com
Dan	Wu	RCSI	Postdoc	danwu@rcsi.ie
Muhammad	Zeeshan	Zhengzhou University	PhD student	m.zee1882@gmail.com
Shengxin	Zhang	TU Dublin	PhD student	D22127061@mytudublin.ie

Correct as of 03.01.2024