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(Article begins on next page)

A REVIEW IN USING AGAR GELS FOR CLEANING ART SURFACES

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Abstract

Cleaning an artwork is an extremely delicate and irreversible procedure in a conservation work. Art conservators constantly work to identify an appropriate and correct cleaning method according to the requirements of different substrates and in order to solve different removal problems. In recent years, gels positively responded to the conservation requests becoming a valid help and, in some cases, the best alternative compared to traditional cleaning methods. This paper proposes a summary of the most recent results regarding agar gels and their applications for cleaning art surfaces; moreover, the application of agar gels specifically formulated to address different conservation problems is also discussed.

Introduction

Cleaning is often a challenge when conservators are called to work on a delicate art surface. In the so-called “chemical cleaning”, several different systems can be chosen depending on the desired results to be obtained: pure solvents, solvent mixtures and solutions. The addition of surfactants or chelating agents or other chemicals also contributes to the required conservation achievements. The obtained fluid could be used by swab, brush or by **gel pad** directly on the art surface. As an alternative, the fluid system can be supported by an inert material, such as cellulose microfibers, clay or micronized silica, preparing a paste which can be applied to the area of interest [1-5]. In doing this, the operator improves the fine control of the cleaning procedure.

It is worth noting that many scientific publications [6-28] and conferences [29] have focused on this specific topic. Since the **1990s**, when thickeners based on cellulose ethers and solvent gels were introduced, the development of **these types** of cleaning systems and the study of their effectiveness

37 have been gradual and have pointed to different **conservative** problems (e.g. residues on the treated
38 surface) [6-8].

39 Recently, gelling agents able to control the fluid release to the substrate avoiding evident residues
40 have raised the interest of the conservation community. In fact, in recent decades, many efforts have
41 been made to prepare new gelling systems capable of retaining liquid compounds and being easily
42 removable. Responsive gels were considered the new frontier of thickeners because of their
43 promising properties [11,12]. Unfortunately, the specialized literature shows that peelable gels have
44 had the greatest success so far in the **conservation** practice [16,17, 29]. Agar gel and gellan gum
45 can be included in this category even if they are natural gelling systems [9,10,13-15,22]. Both gels
46 are usually applied in the cleaning of artworks thanks to their great effectiveness and ease of
47 application on a huge variety of substrates, from the firmest, like stone surfaces or plaster with
48 consistent soiling [20, 23, 27], to the most fragile, like paper [18,21,30].

49 In general, gels can be considered as soft materials consisting of interconnected long polymer chains
50 dispersed into a fluid (water and/or organic solvent), forming a three-dimensional network [31]. They
51 allow to use less solvent than in “free solvent” cleaning, thus improving the sustainability of the
52 treatment [32,33]. Moreover, reproducible cleaning processes can be easily and effectively obtained
53 by controlling the contact time of gels with the surface underneath. However, the risk of leaving
54 organic residues on the treated surfaces must be taken into account for these systems, because of
55 the difficulty of completely removing the gel residues from the substrates. Such a risk can be
56 prevented with several washings by swabs soaked in suitable solvents, or it can be avoided by using
57 semi-rigid gels [17] and, in particular, agar gels [10, 34].

58 Agar is a natural polysaccharide extracted from several orders of red seaweeds (Gelidiales and
59 Gracilariales), able to form semi-rigid, thermo-reversible and hydrophilic gels by simply dispersing
60 the agar raw powder in water, subsequent heating and final cooling. During the last step, agar chains
61 arrange into an ordered structure, where aggregates of co-axial helices form the junctions of the
62 three-dimensional gel network [9,35]. This peculiar supramolecular structure provides interesting
63 properties for its application as salt bridge in electrochemistry, gelatine substitute in the food industry,
64 growth media for microbiological cultures, and many others [36]. Hence, agar gels allow the confined
65 release of the solvent at the interface between gel and substrate, and display other interesting
66 features for the conservation field: high effectiveness and versatility in removing different types of
67 soiling from different substrates, low impact on the artwork, low cost, “green” approach and
68 applicability in different forms and environmental conditions [10]. These properties are particularly
69 useful when water sensitive substrates are involved. In fact, water is one of the most important
70 solvents used in cleaning painting materials, murals, metals and stones. Despite its fundamental role
71 in cleaning processes, several materials (such as gypsum, stuccoworks, etc.) cannot tolerate water
72 effects (e.g. solvent action and decrease of the substrate mechanical features). Thus, especially in

73 these situations, the use of agar gel widens the potential of water cleaning by controlling the contact
74 time and, consequently, the release of water.

75 The conservation of sensitive substrates, i.e. plaster and gypsum works, introduces some issues,
76 which steered conservators toward the use of agar gels already during the first decade of the new
77 century. These substrates exhibit specific physical, mechanical and chemical properties which
78 demand care procedures: light colour, porosity and consequent water absorption, weak mechanical
79 resistance, especially when the materials are wet, just to name a few. Moreover, the artworks,
80 especially gypsum works, quite often display a composite structure, with insertion of materials as
81 iron bars, wood elements or vegetable fibres; in those cases, cleaning problems, such as the
82 application of a liquid phase and its confinement, can dramatically increase.

83 The practical effects provided by the use of agar gels in the cleaning of artworks have been studied
84 for more than a decade. Best practices suggest preparing agar gels by dissolving raw powders in
85 water in the range 0.5-5% w/v [10,37]. Moreover, new application methods were developed drawing
86 inspiration by the polysaccharide characteristics, and new trends of use were opened in different
87 types of conservation interventions [37, 38]. In particular, for the removal of metal stains from building
88 materials, an improvement in agar gel performances was obtained by adding functional chemicals,
89 such as ethylenediaminetetraacetic acid (EDTA), triammonium citrate (TAC) or specific amino acids
90 [23,39,40].

91 However, the effectiveness of gel cleaning is strictly related to the soiling removal mechanism, which
92 is not known in detail yet [23]. On the other hand, it is known that the environmental conditions of
93 growth of red seaweed affect the chemical composition of the raw material, thus influencing the gel
94 structure and its cleaning performances [24,41,42].

95 Given these premises, this review provides an overview of recent works published on agar gels as
96 cleaning materials, focusing especially on the following aspects:

- 97 1. summarize the chemical differences among the raw materials currently present on the
98 market, in order to understand how the supplier influences the final cleaning performances;
- 99 2. elucidate the water bonds in the agar network (in the following refereed as water
100 state), that could be linked to the “release issue”;
- 101 3. highlight the chelating action of agar gels towards metals;
- 102 4. exemplify the practical versatility of agar gels in several conservation cases

103

104 **The applied multi-analytical approach**

105 Compositional, structural and functional features of different agar raw materials and gels have been
106 elucidated by a multi-analytical approach [24, 26]. Each analytical technique provided specific
107 information, and, in several cases, one technique completed the information recorded by the others.
108 In the attempt to identify and clarify all the factors that might influence the effectiveness of agar gels
109 in cleaning procedures, different formulations of agar gels were used, both pristine and added with

110 disodium EDTA and triammonium citrate [23,24,26]. Four raw materials used in several application
111 fields were compared and studied. Table 1 reports the name and the supplier of the selected raw
112 materials. AgarArt and Agar Purissimo are the most used material by conservators in Italy, whereas
113 the other materials are usually applied in biological cultures and in the food industry (in the following
114 named Agar Food). In particular, agar purchased by Sigma Aldrich was used as a reference. Agar
115 solutions of 1, 3 and 5 % w/v were chosen since they are the most used concentrations applied by
116 **conservators**. For gel preparation, agar powder was dissolved in the proper amount of Milli-Q water,
117 then the solution was heated for 2 min in a microwave oven at 700W and then it was cooled in the
118 air. In the case of EDTA and TAC-added gels, the proper amount of additives was introduced in the
119 solution just before cooling.

120 The general compositional screening and the chemical characterization of the different agar
121 formulations were performed by infrared spectroscopy (FTIR), Fourier Transform Raman
122 Spectroscopy (FT-Raman) and pyrolysis - gas chromatography/mass spectrometry (Py-GC/MS). In
123 particular, FT-Raman analyses were carried out to avoid the fluorescence effects observed in
124 conventional Raman spectroscopy. For Py-GC/MS analyses, the thermally assisted hydrolysis and
125 methylation method (THM) [43-45] was applied to obtain detailed information on the
126 monosaccharides present in agarose and agaropectin chains and on the way the monosaccharide
127 units are linked to each other.

128 The characterization of the agar gel structure was focussed on the evaluation of several issues, such
129 as the water state and its release, investigating the chelating properties and performances as well
130 [23, 26]. Different **stone substrates** were used for different purposes: Noto calcarenite for the
131 determination of water absorption and veined Carrara marble for the reproduction of the copper
132 green stains observed on real cases. Water release was determined by capillarity test according to
133 Italian Standard UNI 10859 [46] substituting the multifilter paper with agar gels at different
134 concentrations. ¹H NMR analyses were also performed to evaluate the water state in the gel and in
135 a highly porous (36.2%) stone [47].

136 Copper stains were reproduced on specimens according to the recipe suggested by Bakhtiani et al.
137 [48], synthesizing brochantite [Cu₄(SO₄)(OH)₆] directly on the marble surface. Before staining (at
138 time t₀), laboratory marble surfaces display at the optical microscope a compact surface without
139 morphological flaws (Figure 1a), whereas after staining copper compounds do not remain as external
140 formations, but penetrate into subsurface regions (Figure 1b), probably in correspondence with more
141 porous areas (at time t₁). After application of the agar gel (Figure 1c) on the stained specimens for
142 the required time (t₂, 30 and 60 min), the microscope images allowed to verify that in general copper
143 compounds were removed from the intergranular spaces, leaving them brighter in colour [23].

144 Moreover, thermal analyses, such as thermogravimetric analyses (TGA) and differential scanning
145 calorimetry (DSC), were fundamental to give an insight into the water state in agar gel at different
146 concentrations [26].

147 The agar gel chelating properties and its performances for metal stain removal were investigated by
148 electron paramagnetic resonance (EPR) and induced coupled plasma optical emission spectrometry
149 (ICP-OES) [23]. In particular, EPR spectroscopy characterized Cu(II) centres symmetry in gels,
150 highlighting that also agar gels have the intrinsic capacity to coordinate metals. Furthermore, ICP-
151 OES spectroscopy allowed to quantify the total copper amount extracted by agar gels from
152 substrates, suggesting the best formulation applicable to a specific conservative problem.

153

154 **The chemical differences among raw materials**

155 As already reported in the introduction, literature strongly correlates the specificity of the raw
156 materials to their properties and performances. In particular, the environmental conditions of algae
157 growth can affect agar gel properties in terms of chemical composition, gel strength and, as a
158 consequence, in cleaning performances. The research published by the authors demonstrates the
159 compositional differences among the four different commercial raw materials reported in Table 1
160 [24]. Table 2 lists the assignments of all specific FTIR absorption and FT-Raman scattering bands
161 for agar samples. In general, the FTIR patterns of different raw agar powders are too similar to
162 discriminate differences in composition [24, 49-53]. On the other hand, spectral subtraction of FT-
163 Raman spectra was found to be a valuable tool to highlight the presence of specific spectral pattern,
164 e.g. in Agar Purissimo and in Agar Food, which can be well correlated to the presence of crystalline
165 glucose [54].

166 These data were confirmed also by Py-GC/MS analyses, which show the presence of galactose
167 linked on position C3, both in agarose and agaropectine, as well as glucose signals, thus supporting
168 the spectroscopic results. Moreover, three additional peaks of 4-O-substituted glucose, typical
169 compounds of cellulose, were detected in AgarArt and Agar Purissimo, pointing to a different
170 purification level of the seaweed from which the samples were extracted (Table 3).

171 Importantly, Py-GC/MS analyses carried out without derivatization allowed to perform a semi-
172 quantitative evaluation of the anhydrous fraction responsible of the gel strength [55]. The analyses
173 demonstrate a variable content of anhydrous and non-anhydrous units. In particular, under pyrolysis
174 Agar Sigma, which is the purest raw agar, produced comparable amounts of galactopyranose and
175 anhydro-galactopyranose, whereas Agar Purissimo recorded the lowest content of the anhydrous
176 compound. This compositional information was in good agreement with rheological measurements.
177 In fact, Agar Food gels contains the highest percentage of anhydrous units and the greatest
178 viscoelastic moduli has been detected [55].

179 Finally, thermogravimetric analyses also showed slight differences in the weight loss profiles of
180 different types of agar. AgarArt and Agar Sigma were found to be more thermally stable than Agar
181 Purissimo and Agar Food. The first two samples exhibited a fast and steep weight loss at around
182 200 °C, which was shifted at approximately 140 °C in the other two agar gels. This difference was
183 possibly correlated to the free glucose volatilization.

184

185 **The water state and its release**

186 The cleaning selectivity of agar gel is strongly correlated to several specific properties of gels, such
187 as the spontaneous release of the liquid phase (syneresis), its diffusion capacity into porous
188 substrates thanks to an ion concentration gradient, and finally osmosis phenomena [56]. The main
189 advantages in using gel cleaning systems, **compared to the traditional cleaning methods (free**
190 **solvent, mechanical methods or their combination)**, are the fine control of the liquid release into the
191 substrate and the reduced evaporation of solvents, thus limiting the aggressiveness of the cleaning
192 process.

193 Water in gels is commonly classified into three main types: non-freezable bound water with strong
194 water-polymer connection, freezable bound water with weak water-polymer connection and free
195 (freezable) water without water-polymer connection [57,58]. The quantification of the different types
196 of water allows to understand at what extent the agar structure and composition influence the
197 transport phenomena.

198 Starting from the evaluation of the water release, modified capillary test were performed on Noto
199 calcarenite, substituting the multilayer filter paper pad with AgarArt gel at increasing concentrations
200 [23]. The amount of water (Q_i) absorbed by capillarity into substrates was studied by capillarity
201 absorption measurements on five Noto calcarenite specimens kept in contact with agar gel.
202 Absorption capillary coefficients (CA), proportional to the absorption rate, were calculated as
203 suggested in UNI 10859 [46] and are reported in Table 4. Results show that both the amount of
204 water absorbed by capillarity and the water absorption rate decrease when the agar gel
205 concentration increases. In particular, the asymptotic value of the gel 1 % curve is very similar to the
206 one obtained for a multilayer filter paper pad [59], used as reference. Thus, agar gel 1% almost
207 performs as a “free” water reservoir.

208 Unilateral $^1\text{H-NMR}$ analyses were also carried out to support the water transport capillarity results
209 with further information on mobility of the various water components as a function of gel
210 concentration. The longitudinal and transverse magnification decays of the different types of water
211 exhibit a multi-exponential trend, different from the mono-exponential decay trend of bulk water [26].
212 In fact, agar samples display a mono-exponential trend in longitudinal relaxation time (T_1), whereas
213 a bi-exponential trend is recorded in transverse relaxation time (T_2). Longitudinal relaxation time
214 exhibits a linear correlation with agar concentration, indicating that water mobility decreases with the
215 density of the polymeric network. For what concern the “bound” and “free” water, transverse
216 relaxation time can be described by two proton species characterized by a short (T_{2A}) and long (T_{2B})
217 transverse relaxation time, respectively. Both relaxation times decrease when the agar concentration
218 increases, which affects more the mobility of free water than bound water, further confirming that
219 water mobility is strongly hampered by the polymer network and its inner interconnections.

220 The amount of bound water is very low, around 4-7% of the total water, while the free water content
221 is in the range of 93-96% in all samples. Moreover, the self-diffusion coefficient of water molecules
222 was investigated by diffusion ^1H NMR measurements on Noto calcarenite mock-ups with agar gels
223 at different concentration. In all commercial agar gels, also the diffusion decays exhibit a linear
224 correlation with agar concentration: the self-diffusion coefficients of water in agar gels were slower
225 than those observed for bulk water ($2.5 \times 10^{-9} \text{ m}^2/\text{s}$ at $25 \text{ }^\circ\text{C}$) and decreased with increasing agar
226 concentration. At higher concentrations of agar, water mobility is hindered both by interactions with
227 the agar hydroxyls and by the greater density of the polysaccharide matrix [26]. AgarArt exhibits a
228 slower translational motion of water molecules and therefore more water release if compared to Agar
229 Sigma, in which a tendency to collapse has been observed after a certain release of water (between
230 60 and 240 min).

231 The amount of freezable bound water and freezable free water in different agar gels was also
232 estimated by thermal analyses [26]. In general, freezable bound water melts at lower temperature
233 than free water. By monitoring the gel drying by DSC, the melting signal of bound water increased
234 at the expense of the free water melting signal. Moreover, the heat of melting of freezable free water
235 is always lower than in bulk water, confirming the different behaviour of water in bulk or in a network.
236 Concerning the correlation between the different types of water and agar concentration, DSC
237 analyses highlighted that freezable water is not affected by agar concentration, but the amount of
238 bound water changes reaching approximately 20 wt% of the total water content. This is probably
239 correlated to the gel microstructure: the smaller is the porosity, the greater the interfacial area and
240 the greater the number of bound water sites. This happens until a concentration limit, in this case
241 5% w/v, beyond which the binding sites for water become less accessible.

242 The correlation of the water release with the gel microstructure is a difficult task because porosity
243 and especially the pore connectivity are difficult to observe on hydrogels, even with an Environmental
244 SEM (E-SEM) equipment. For such an investigation, hydrogels can be lyophilised (i.e. freeze-dried)
245 immediately after contact with stained specimens, in order to obtain a polymeric network without the
246 liquid phase, called xerogels [60]. In fact, ad hoc prepared xerogels may be considered
247 representative of the morphology of the related hydrogels, since fast freezing in liquid nitrogen leads
248 to the formation of amorphous ice, which does not expand the hydrogel network structure unlike ice
249 crystals. The authors' research is moving exactly in this direction, trying to correlate water release
250 and cleaning effectiveness with the gel microstructure.

251

252 **Chelating properties of agar gels**

253 In order to better understand the chemical mechanisms involved and to evaluate the cleaning
254 effectiveness, the removal of artificial copper stains from marble specimens was investigated by ICP-
255 OES and the chelating properties of agar gels were investigated by EPR spectroscopy [23].

256 As prepared agar gels contain negligible amounts of copper. After 1 h contact with artificially stained
257 marble specimens, ICP-OES data showed that the amount of copper detected in pristine gels (1, 3,
258 5 % w/v) was rather small, below 15 $\mu\text{g}/\text{cm}^2$ (Figure 2a, b, c). By using agar gel 3 % containing
259 additives (EDTA and TAC), the removal of copper was about twice more effective, with no significant
260 differences among the used additives (Figure 2d, e). The higher effectiveness displayed by agar gels
261 containing additives with respect to pristine gels is in agreement with the results obtained in other
262 researches about copper ion removal from wastewaters [61].

263 For what concerns the study of copper coordination, all pristine agar gels displayed weak axial EPR
264 signals (Figure 3a), having values of g and A tensor components (Table 5) consistent with those of
265 Cu (II) centers in a tetragonal symmetry field of oxygen atoms [62,63]. Thus, these signals were
266 attributed to Cu (II) centres coordinated by agar gels, from now on termed as Cu-agar.

267 For gel 3 % w/v added with TAC (Fig. 3b), resonances attributable to Cu-agar species were
268 observed. In addition, an isotropic intense broad band was observed ($g \sim 2$), attributable to interacting
269 Cu (II) centres in frozen water solution [63,64].

270 Gel 3 % added with EDTA showed two different axial signals, attributable to two different Cu (II)
271 centres in a tetragonal symmetry field of oxygen atoms (Figure 3c). The weaker one has the same
272 EPR parameters as those observed for pristine agar gels (Cu-agar), while the other signal has higher
273 intensity and tensor components ($g_{\perp} = 2.07$, $g_{\parallel} = 2.29$, $A_{\parallel} = 149$ G), which can be attributed to Cu
274 (II) centres coordinated by the additive, from now on termed Cu-EDTA.

275 Thus, EPR spectra showed that in all the studied samples copper centres are coordinated both by
276 agar gels and by additives, when present. Moreover, in 3% w/v gels added with TAC some copper
277 dispersed in water was also found. Thus, all agar gels with chelators were found to be equally
278 effective in removing copper stains, even if they host metals in different ways [23].

279

280 **Understanding some agar features through the practical experience of conservators**

281 Usually conservators identify the proper cleaning procedure with agar gels taking into consideration
282 the soiling, the substrate characteristics and the following variables:

- 283 • specific type of raw material
- 284 • agar concentration
- 285 • gel thickness
- 286 • application mode
- 287 • contact time

288 Some of the above parameters have been already discussed. As for thickness, it usually ranges in
289 the millimetre scale, depending on the application mode and substrate morphology. Rigid gels are
290 usually the thickest, reaching 1 cm. Considering the empirical experiences collected so far, the
291 thickness of the gel significantly influences the cleaning effectiveness, and consequently, the release
292 of liquid phase. Although this issue is not scientifically confirmed by a systematic study aimed at

293 clarifying the correlation between thickness and a quantitative value of the released liquid, if one
294 wants to apply gels for a prolonged contact time, their thickness must be carefully controlled to
295 prevent the gel from drying out on the artwork surface. Moreover, it is worth noting that several
296 practical experiences of conservation interventions suggest the combined action of mechanical and
297 chemical processes to remove particles detached from the surface of artworks, especially when the
298 gel is used in the viscous sol form, without compromise the integrity of the treated surfaces [18, 20,
299 23, 65].

300 Aqueous solutions of agar show a sol–gel transition temperature in the range from 32°C to 47°C
301 [66]. Hence, the use of agar gel at room temperature allows its application in the rigid form, cutting
302 it in pads of the desired shape to be placed on flat surfaces. Otherwise, warm agar solutions can be
303 applied as viscous fluids, with viscosity depending on the temperature of the solution and on the
304 agar concentration.

305 Agar gels, both in the rigid and fluid form, have successfully cleaned large three-dimensional figures
306 as well as detailed decorative architectural surfaces. Gullotta reported one of the first researches
307 where empirical observations have been completed and interpreted through scientific laboratory
308 studies [20].

309 So far a lot of experience in the use of agar gels on several different materials such as marble (both
310 conserved in indoor environment or exposed to outdoor conditions), wall paintings, panel or canvas
311 paintings or wooden sculptures, has been collected [8-10, 18, 20, 22, 27- 29, 32, 37-40, 66-71]. Such
312 a variety of case studies led to addressing a wide range of different cleaning problems, related to
313 both the substrate features (morphology, dimensions, hydrophilicity, absorption features, etc.), and
314 the soiling nature (chemical composition, thickness, adhesiveness, cohesion). The versatility of agar
315 gels makes the **conservation** procedure flexible, according to operational needs. This happens
316 because agar ensures a good control of water release and limited colour changes in the substrates.
317 A sequence of repeated applications allows a satisfactory selectivity, even when the desired result
318 does not entail the total removal of soiling. This is possible because rigid agar gels can be easily
319 removed by a simple peeling, avoiding more aggressive mechanical actions and limiting subsequent
320 rinsing.

321 A double process of heating/cooling is recommended as standardized procedure for the preparation
322 of rigid agar gels in order to obtain a transparent gel with a better consistency, which leads to an
323 improved cleaning effectiveness.

324 Rigid gels prepared at increasing concentrations exhibit different final mechanical properties [55].
325 Moreover, the increase in concentration implies a decrease in water release. These empirical
326 features were confirmed by Bertasa et al. [26], who found a linear dependence of the polysaccharide
327 concentration versus the total amount of *free* water in the gel. *Free* water is considered to be the
328 main responsible of soiling extraction.

329 It is also possible to use agar gels in a paste form, purchasable on the market under the registered

330 trademark name of Nevek®; the product is ready-to use and sold in the form of small rigid particles
331 stabilized by ethanol and a very low amount of isopropanol (~ 0.1 %). Nevek® can be added to
332 apolar solvents such as ligroin, thus increasing the variety of possible removable soiling. In general,
333 the possibility of adding different chemicals, such as organic solvents (both polar and apolar),
334 chelating agents or surfactants, enormously widens the potential of agar gels as cleaning materials.
335 Finally, it is worth mentioning a recent study in which good results for sensitive materials were
336 obtained combining the cleaning with agar gels with Nd:YAG laser sources [67] and paving the way
337 for new strategies of intervention. The combined cleaning of laser-agar demonstrated an interesting
338 sensitivity in removing different kind of soiling: colorimetric measurements recorded minimal
339 chromatic variation under the threshold ($\Delta E \sim 3$) of eye sensibility. Moreover, optical microscope
340 observations showed no changes in the morphology and integrity of the treated surfaces.

341 In Table 6, the possible application modes are summarized, each one with related benefits and
342 drawbacks.

343 When the gel is prepared with a standard procedure its pH is close to neutrality (in the range 6.5/7.5
344 pH units); as mentioned above, several additives and solvents can be added to the gel modifying its
345 chemical and physical properties, including pH. It is worth noting that the gelling properties are
346 strongly dependent on pH values, hence it is possible that moving from neutrality, the gel does not
347 form correctly.

348 The gel cleaning potential changes by working on a specific pH: for instance, agar gels can work on
349 hydrophobic surfaces by adding a surfactant, or as an alternative, they can be used to extract
350 insoluble salts by adding chelating agents. More specifically, above pH 8 insoluble calcium salts
351 (sulphate, carbonates and oxalate) can be extracted by gels added with EDTA and citrate [72]: EDTA
352 has a higher capacity of chelating Ca^{2+} compared to citrate.

353 According to the practical experience, the cleaning action of the system gel-chelating agent is
354 improved at minor chelator concentration (lower than 1%) [40]. As already mentioned, Nevek®
355 achieved good results when added with solvents (both polar and apolar), up to about 40% of the
356 weight, depending on the solvent. As a matter of fact, on the same artwork cleaning methods can be
357 varied depending on the specific state of conservation and decay phenomena. In the following, some
358 examples of conservative work with agar gel are reported, showing the wide range of potential that
359 it can offer.

360 Stoldo Lorenzi was an Italian mannerist sculptor active both in Tuscany and in Milan: the “Adamo”
361 marble figure, sculpted in 1575 and currently in the Milan Museum of Ancient Art collection (Sforza
362 Castle), exhibited an extremely critical conservation state. Fluid agar gel at 3% w/v was used in
363 several repeated applications, focusing on the removal of thick and adherent surface deposits,
364 patination and retouching, while the stains on the back side of the sculpture were treated by Nevek
365 ® *ready-to-use*, with the addition of acetone (Figure 4a). Moreover, fluid agar gel added with TAC at
366 0.2%, was applied on specific zones, characterized by an intense coloration due to decay, with

367 different contact time, until a complete drying of the gel itself. In particular, the halo of the stain was
368 progressively contained by applying the agar gel at slightly greater size than the stain itself,
369 increasing the chelating agent concentration or prolonging the contact time [68].

370 A combination of gel and laser cleaning were used on two extremely delicate areas: i) at the head of
371 the sculpture (Figure 4b), characterized by very weak conditions, since even a minimal mechanical
372 treatment could have created a weakening of the surface material, and ii) on the black areas of the
373 lower part of the sculpture as well, in order to reduce the coloured retouching still present on the
374 sculpture. After the laser ablation and several applications of fluid agar, a veil of yellowish deposits
375 remained on the head surface, thus matching the correct chromatic tone, which is suggested by the
376 museum curators and the supervisors of the Italian Ministry of Cultural Heritage [40,67,68].

377 Agar gel in different application modes was used with good results also on the XIV century mural
378 painting in San Miguel Chapel (Pedralbes Monastery, Barcelona) [71]. The variety of the observed
379 painting techniques was respected by intervening with a targeted and diversified way according to
380 the conservation state of the painting and the chemical composition of pigments and binders. In this
381 case, agar in three different application forms, such as rigid, fluid form and Nevek® with and without
382 additives, was tested in order to evaluate the best formulation in accordance to the specific
383 conservative problem (Figure 5a). All formulations were adapted to obtain different removal levels of
384 vegetal glue (probably gum arabic). After evaluating advantages and drawbacks (see Table n. 6),
385 Nevek® agar was applied both pristine and with chelating agents, such as triammonium citrate with
386 tetraethylammonium and trisodic EDTA. Chelating agents were used in the concentration range of
387 0.3 -1% w/v, with different contact times (from 15 min to 4 hour) continuously controlling pH level
388 (safety range according to the different painting layer: 6.53 for oil paint and 7.25 for fresco).
389 Furthermore, two levels of Japanese paper were applied on the surface to precisely control the
390 cleaning level and detect the eventual paint detachment [71].

391 An extremely innovative application of the agar gel system, **not yet published**, undoubtedly regards
392 the wax sculpture “Portinaia” by Medardo Rosso, part of a private collection. The alkyd paint on the
393 wax sculpture was removed after a softening step obtained by a layer of Nevek® directly applied on
394 the surfaces. Nevek® agar gel has been also used to remove adherent deposits on the surface of
395 another wax sculpture entitled “Ecce Puer” by the same author: in this particular case, Nevek® was
396 applied on Japanese tissue in order to protect the rippled and uneven surface (Figure 5b).

397

398 **Summary and open questions**

399 The research reviewed in this article has the ambition to provide a deep and innovative insight into
400 agar gel properties and cleaning mechanisms. Several issues were investigated with a multi-
401 analytical approach in order to characterize compositional, structural and functional features which
402 affect agar-based cleaning methods. Agar raw materials purchased by different supplier are
403 characterized by intrinsic chemical differences depending on the application field and the different

404 purification level. Such a result was neither confirmed nor denied by the companies which sell the
405 product.

406 The evaluation of water release and state in different gels shows significant differences in the overall
407 content of water according to the polymer concentration. The research underlined that agar at 1%
408 w/v performs as a free water source. This result suggests the application of a more concentrated
409 gels in order to better control the water release at the surface, as confirmed by ¹H-NMR analyses.
410 In fact, water mobility is strongly hindered by the polymer network due to the increase of double
411 helices population and their interconnection. These data are in agreement with thermal analyses,
412 which detected a lower amount of freezable water compared to bulk water in the different gels,
413 suggesting that water in gels is in a more dispersed state than bulk water.

414 Copper centres can be coordinated both by agar gels and by additives (when present). All copper
415 centres are coordinated in gels added with EDTA, while they are also dispersed in water within gels
416 added with TAC. Thus, all agar gels with additives are equally effective for copper stain removal,
417 even if they host copper in different ways. This suggests that the cleaning of stains on stones starts
418 with the diffusion of water at the gel-stone interface, but copper coordination, both by agar sites and
419 additives, is the driving force.

420 The conservators' experience highlights the extreme versatility of the agar gel that can be applied
421 on different types of material and according to the **conservation** needs.

422 Overall, agar gel has renewed the concept of the conservation intervention, starting from the
423 sustainability of the material and the versatility of the application mode, to the possibility of solving
424 open **conservative** problems. The presence of gel residues on the surface of the treated artworks or,
425 in a pessimistic view, inside the artwork has not yet been thoroughly investigated, although a few
426 works have been reported in the literature [10, 18, 65, 73].

427 In a sense agar gel is comparable to other gelling agents, but it becomes superior in the ability to
428 adapt to different situations, with the possibility to tailor its physical and chemical properties.

429

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437

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665 **Tables**

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Table 1 - Agar raw materials

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Agar name	Purchased by
AgarArt	CTS, Italy
Agar Purissimo	Bresciani, Italy
Agar Sigma	Sigma Aldrich (A7002_CAS:9002-18-0)
Agar Food	UK supplier

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Table 2 - Assignments of specific absorption and scattering bands of agar samples

Wavenumber (cm ⁻¹)	Assignment		Intensity		References
			FT-Raman	IR	
3400-3300	ν O–H		w	m	[24]
2900-2800	ν C–H	R ₃ -CH; R-CH ₂	vs	vs	[24]
1470-1460	δ_s CH ₂		m-s	m-s	[24]
1430-1200	δ O–H		m-w	w	[24]
1390-1370	ν_{as} S=O		m	vs-s	[53], [24]
1200-1188	ν_a S=O		vs-s	vs-s	[49], [50], [51], [52], [53], [24]
1150-1075	$\nu_{out-of-phase}$ C–C–O	Secondary alcohols	m-s	s	[52], [24]
1070	$\nu_{out-of-phase}$ C–C–O	3,6-anhydrogalactose	s	s	[50], [24]
1075-1000	$\nu_{out-of-phase}$ C–C–O	Primary alcohols	m-s	s	[51], [53], [24]

930	ν C–O	3,6-anhydrogalactose	vw	w	[49], [51], [52], [53], [24]
900-800	$\nu_{\text{in-phase}}$ C–C–O	Primary /secondary alcohols	vs-m	m	[50], [51], [52], [53], [24]
867	C–O–SO ₃	On C6 of galactose	vw	vw	[50], [51], [24]
845	C–O–SO ₃	On C4 of galactose	m	-	[49], [50], [52], [24]
815–820	C–O–SO ₃	On C6 of galactose	sh	-	[49], [50], [52], [24]
770-740	Skeletal vibr	Galactose ring	vw-m	m-s	[51], [53], [24]
500-400	δ C–C–O	Primary /secondary alcohols	-	m-w	[24]

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Table 3 - Characteristic pyrolysis products of agar samples derivatized with TMAH
(table modified from Bertasa M. et al., 2017)

Assignment	Tentative sugar assignments	m/z	MM*
2-hydroxypropanoic acid, methyl ester	-	45	104
Methoxyacetic acid, methyl ester	-	45	104
2-methoxypropionic acid, methyl ester	-	59	118
Unknown (m/z 144, 131, 113, 99, 71)	-	71	-
1,2,4-trimethoxybenzene	-	168	168
tri-O-methyl-3-deoxy-2-methoxymethyl-d-erythro- pentonic acid, methyl ester	GLU 4-O substituted	173	236
tri-O-methyl-3-deoxy-2-methoxymethyl-d-threo- pentonic acid, methyl ester	GLU 4-O substituted	173	236
tri-O-methyl-3-deoxy-d-arabino-hexonic acid, methyl ester	GLU	129	206
tetra-O-methyl-3-deoxy-d-arabino-hexonic acid, methyl ester	GLU	129	220

tri-O-methyl-3-deoxy-d-xylo-hexonic acid, methyl ester	GAL	129	236
tetra-O-methyl-3-deoxy-2-methoxymethyl-d-erythro/threo-pentonic acid, methyl ester	GLU 4-O substituted	129	264
tetra-O-methyl-3-deoxy-d-xylo-hexonic acid, methyl ester	GAL	129	250
tri-O-methyl-3-deoxy-d-ribo-hexonic acid, methyl ester	GLU	129	206
tetra-O-methyl-3-deoxy-d-ribo-hexonic acid, methyl ester	GLU	129	220
tri-O-methyl-3-deoxy-d-lyxo-hexonic acid, methyl ester	GAL	129	236
tetra-O-methyl-3-deoxy-d-lyxo-hexonic acid, methyl ester	GAL	129	250

674 * MM = Molecular Mass

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676 **Table 4** - Adsorption capillarity coefficients (CA) on Noto calcarenite in contact with AgarArt gel at
677 different concentration.

Gels	CA (mg/cm ² s ^{-1/2})
UNI 10859	11.3 ± 0.9
AgarArt 1%	6.1 ± 0.9
AgarArt 3%	0.6 ± 0.2
AgarArt 5%	0.13 ± 0.01

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679 **Table 5** - Parameters of EPR signals observed on agar gels after 1 h contact with laboratory-stained
680 specimens.



Sample		g// ± 0.01	A// ± 5 G	g⊥ ± 0.01	Attribution
AgarArt 1 %	Pristine	2.39	135	2.08	Cu-agar
AgarArt 3 %	Pristine	~ 2.4	"	"	Cu-agar
	with TAC	2.39	"	"	Cu-agar
		~ 2.1	n.d.	"	Interacting Cu(II) centres
	with EDTA	2.39	135	"	Cu-agar
		2.29	149	2.07"	Cu-EDTA (predominant)
AgarArt 5 %	Pristine	"	135	"	Cu-agar



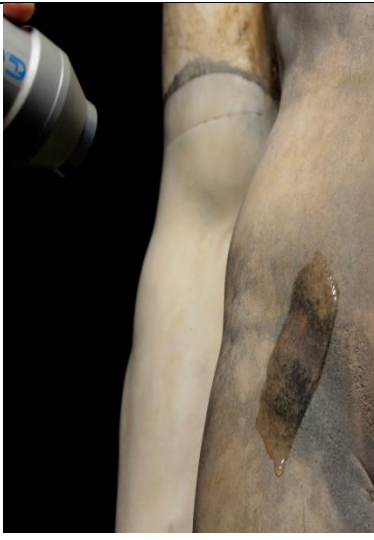
681 n.d. = not detected

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Table 6 - Agar gel properties according to the application methods

[Images - courtesy of M. Anzani]

Rigid agar gel		<ul style="list-style-type: none">• Minimal adhesion• Limited water release• Moderate and controllable removal of the soiling• Substrate should present perfect planarity• The gel can be cut and shaped, according to the substrate surface, desired regular thickness and the gel concentration• Good transparency• Possibility to be simply peeled off
Fluid agar gel		<ul style="list-style-type: none">• Very good adhesion• Light increase in water release• Irregularity in thickness• Combination of chemical and mechanical cleaning• Good Transparency• Possibility to be simply peeled off

<p>Nevek ®</p>		<ul style="list-style-type: none"> · Adhesion minimised · Slight increase in water release · Moderate soiling removal · Limited transparency · Easy removal · Possibility to add apolar solvents
<p>Foam agar gel</p>		<ul style="list-style-type: none"> · Very good adhesion · Minimal water release · Moderate soiling removal · Higher effectiveness increasing the volume due to the increase of specific surface · No transparency · Easy removal
<p>Agar gel in combination with laser methodology</p>		<ul style="list-style-type: none"> · Preliminary step in cleaning · Wetting agent on the surface during the laser irradiation · Control of the thermic peak and limitation of the mechanical stress · No fall down mechanisms and fixing of soiling · Limitation of colour variations

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687 **Figure Captions**

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689 **Figure 1** - Optical microscope images of Carrara marble laboratory specimens before (a), after the
690 staining process (b) and after cleaning for 30 and 60 min with AgarArt 3% w/v added with EDTA 1%
691 w/v (c) (scale bar 1 cm).

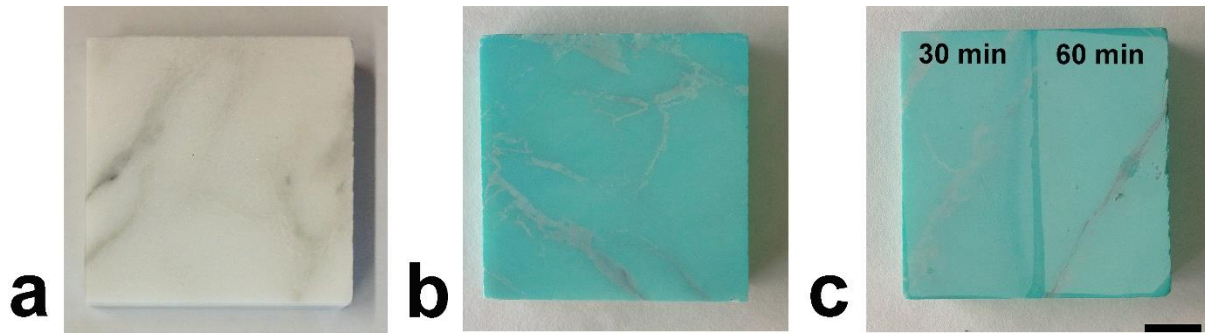
692 **Figure 2** - Copper content per agar surface area after 1 h cleaning of laboratory specimens with
693 agar gels: 1% (a), 3% (b), 5 % (c), 3% + TAC (d), 3% + EDTA (e).

694 **Figure 3** - EPR spectra of agar gels after 1 h cleaning of laboratory specimens with agar gels: 5 %
695 (a), 3% +TAC (b), 3% + EDTA (c).

696 **Figure 4** - The peelable agar gel with yellowish deposit (a); thinning of the veil of yellowish deposits
697 by laser ablation (b). *[Images - courtesy of M. Anzani]*

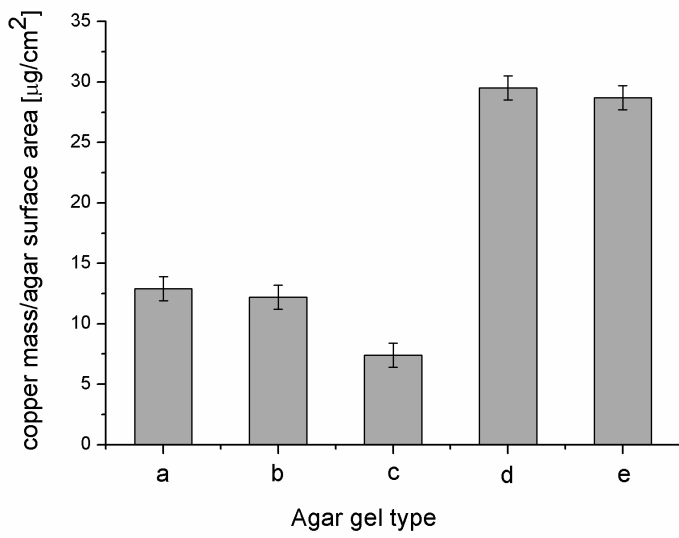
698 **Figure 5** - Application of Nevek ® formulation and Japanese paper on XII century mural painting (a)
699 and on Medardo Rosso sculpture (b). *[Images - courtesy of M. Anzani]*

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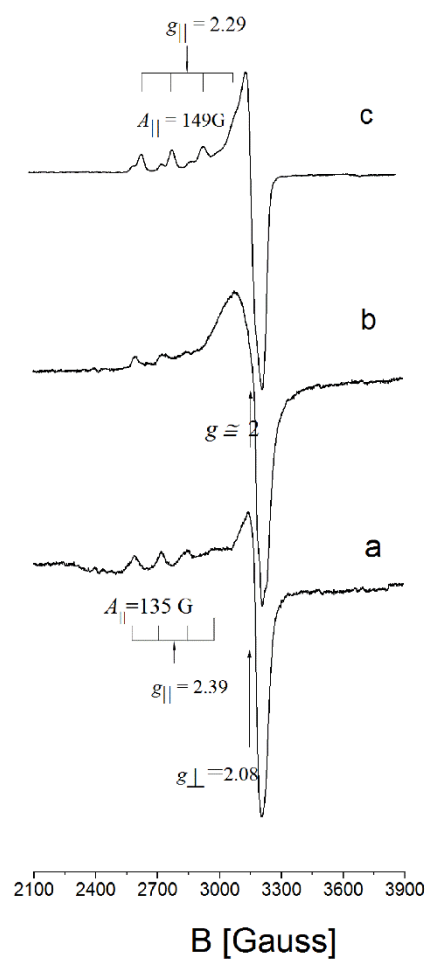
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Figure 1



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Figure 2



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711 **Figure 3**

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716 **Figure 4**

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a



b

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Figure 5