REVIEW ARTICLE



Molecular genetics in aquaculture

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ABSTRACT

Great advances in molecular genetics have deeply changed the way of doing research in aquaculture, as it has already done in other fields. The molecular revolution started in the 1980's, thanks to the widespread use of restriction enzymes and Polymerase Chain Reaction technology, which makes it possible to easily detect the genetic variability directly at the DNA level.

In aquaculture, the molecular data are used for several purposes, which can be clustered into two main groups. The first one, focused on individuals, includes the sex identification and parentage assignment, while the second one, focused on populations, includes the wide area of the genetic characterization, aimed at solving taxonomic uncertainties, preserving genetic biodiversity and detecting genetic tags. For the future, the increase in the number of molecular markers and the construction of high density genetic maps, as well as the implementation of genomic resources (including genome sequencing), are expected to provide tools for the genetic improvement of aquaculture species through Marked Assisted Selection. In this review the characteristics of different types of molecular markers, along with their applications to a variety of aquaculture issues are presented.

Key words: Aquaculture, Molecular markers, Practical applications.

RIASSUNTO

LA GENETICA MOLECOLARE IN ACQUACOLTURA

La genetica molecolare sta profondamente modificando il modo di fare ricerca nell'ambito dell'acquacoltura, così come è già avvenuto in altri settori. La rivoluzione molecolare è iniziata durante gli anni '80, grazie all'utilizzo degli enzimi di restrizione e della Reazione a Catena della Polimerasi, che permettono di studiare, in modo rapido e relativamente poco costoso, la variabilità genetica direttamente a livello del DNA. Nell'ambito dell'acquacoltura, i dati molecolari sono stati utilizzati per numerose applicazioni, che possono essere raggruppate in due categorie. Nella prima, riferita agli individui, rientrano il riconoscimento del sesso e la verifica di parentela, mentre nella seconda, riferita alle popolazioni, rientra il vasto settore della caratterizzazione genetica, finalizzata alla risoluzione di incertezze tassonomiche, alla tutela della biodiversità e all'individuazione di etichette genetiche. I recenti progressi nell'identificazione di un numero sempre più elevato di marcatori, unitamente alla costruzione di mappe genetiche a maggiore densità, forniranno informazioni utili anche nell'ambito del miglioramento genetico. Questa rassegna illustra brevemente le caratteristiche dei diversi marcatori molecolari, soffermandosi più in dettaglio sulle loro applicazioni nel settore dell'acquacoltura.

Parole chiave: Acquacoltura, Marcatori molecolari, Applicazioni pratiche.

Introduction

Mendelian genetics deduced the principles of the heredity by observing the individual phenotypes resulting from appropriate breeding experiments. Its integration with information and methods made available by the progressive advances of molecular biology led to a new branch of genetics, called molecular genetics, whose aim is to investigate all aspects of the gene, such as its structure and functions. The molecular approach provided geneticists with very powerful tools, the molecular markers, which opened exciting perspectives of application in many research fields. The objective of this work is to review the characteristics and potential power of different types of genetic markers, with a major focus on their applications to a variety of aquaculture issues.

Molecular markers

A genetic marker can be defined as any trait which allows the identification of the genotype of an individual. Until the 1980's, the most used markers were proteins (allozymes), because amino acid differences in the polypeptide chain detected by electrophoresis reflect mutations in the coding gene. However, the protein markers have a reduced power in detecting the DNA variability both within and between populations (Valenta *et al.*, 1977; Valenta *et al.*, 1978; Šlechtová *et al.*, 1995; Kohlmann and Kersten, 1998; Antunes *et al.*, 1999), because some DNA mutations do not lead to amino acid substitutions, or the amino acid substitution does not always change the protein total electric charge.

The advances in molecular genetics have made it possible to detect the genetic polymorphism directly at the DNA level, thanks to the widespread use of restriction endonucleases and Polymerase Chain Reaction (PCR) technology. Used alone or in association, they allow the identification of different types of DNA variability: base substitutions, commonly referred to as Single Nucleotide Polymorphism (SNP), insertion or deletion of nucleotides (indel) and Variable Number of Tandem Repeat (VNTR). The last group includes microsatellites (or Simple Sequence Repeats, SSRs), which consist of short sequences (mostly 2-4 base pairs) tandemly repeated up to tens or hundreds of times along the DNA strand (Levinson and Gutman, 1987; Tautz, 1989) and mostly located in non coding regions. Thousands of microsatellites have been found in farm animals; in fish, in particular, the presence of a microsatellite has been estimated every 10 kb (O'Connell and Wright, 1997), with a mutation rate at 10⁻²-10⁻⁶ per locus per generation, which is much greater than that of non repetitive DNA (10⁻⁹) (Weber and Wong, 1993), resulting in a very high level of polymorphism. Moreover, microsatellites are co-dominant and abundantly distributed throughout the genome. For their peculiar features, microsatellites are extensively used in a wide range of research fields and applications.

In aquaculture genetics, mitochondrial

DNA (mtDNA) is also quite popular, for its intrinsic and technical features: a relatively high mutation rate, haploid and maternal inheritance, which reduces the effective population size and thus increases the sensitivity to genetic drift, ease of isolation and manipulation (Avise, 1994; Moritz, 1994). The analysis of mtDNA has been generally carried out by PCR-RFLP and more recently by sequencing. Mitochondrial DNA complete sequences for some aquaculture species, such as common carp (Cyprinus carpio) (Chang and Huang, 1994), tench (Tinca tinca) (Saitoh et al., 2006) and red grouper (Plectropomus leopardus) (Zhu and Yue, 2008), are now available.

The last decade has seen a renewed interest in coding genes for studying the association of their variability with economically important traits. In this respect, comparative genomics can greatly accelerate the identification of effective markers, because many genes are very conservative, allowing the transmission of information between species, with reduction of time and costs.

Applications

Molecular markers can be used in a variety of aquaculture studies, at the individual or population level, and the choice of the markers to be used depends on both the aim of the research and on the marker characteristics. Some examples are briefly presented.

Gene mapping

Gene mapping provides fundamental information for genetic studies, including QTL identification, marker assisted selection and comparative genomics (Danzmann and Gharbi, 2001). In brief, a physical map defines, by *in situ* hybridization, the physical location of DNA segments on a chromosome, while a genetic map depicts the relative distance and the order of the loci along a chromosome on the basis of segregation analyses in reference populations or families: if two markers segregate together, they will locate very close on the chromosome, so they will define a *linkage group*, where the proportion of recombinants between the linked markers is used as a measure of the distance between them. The microsatellite markers represent the tool of choice for the construction of a primary framework map, which can be further enriched with SNP markers in coding genes (Gregory *et al.*, 2004).

Although less developed than in other animal species, microsatellite and SNP-based linkage maps are now available for several aquaculture species (Table 1). As some studies have revealed extensive homology among vertebrates (Morizot, 1983; Woods *et al.*, 2005), comparative evolutionary studies will greatly benefit from the increasing knowledge on the linkage groups arrangements in different species (Matsuoka *et al.*, 2004; Gharbi *et al.*, 2006).

Medium-term genome research will focus on the integration of genetic linkage and physical maps, which would significantly enhance the possibility to apply genomebased technologies to the genetic improvement (Somridhivej *et al.*, 2008).

Sex identification

Sex identification is important in several biological sciences, such as genetics and conservation biology, but in fish it is often difficult because of the reduced sexual dimorphism and the frequent absence of heteromorphic sex chromosomes. Moreover, in fish species where one sex has better performances, monosex stocks have been developed, usually by sex reversal and family selection, with the need to discriminate between genetic and phenotypic sex (Devlin and Nagahama, 2002). Molecular genetics has proven to be very effective in solving the

Species	Reference Knapik <i>et al.</i> , 1998; Shimoda <i>et al.</i> , 1999; Woods <i>et al.</i> , 2005	
Zebrafish (Danio rerio)		
Nile tilapia (Oreochromis niloticus)	Kocher <i>et al.</i> , 1998; Agresti <i>et al.</i> , 2000	
Medaka (Oryzias latipes)	Naruse <i>et al.</i> , 2000	
Channel catfish (Ictalurus punctatus)	Waldbieser et al., 2001; Liu et al., 2003	
Rainbow trout (Onchorhynchus mykiss)	Young <i>et al.</i> , 1998; Nichols <i>et al.</i> , 2003; Rexroad <i>et al.</i> , 2008	
Arctic charr (Salvelinus alpinus)	Woram <i>et al.</i> , 2004	
Atlantic salmon <i>(Salmo salar)</i>	Moen <i>et al.</i> , 2004, 2008; Gilbey <i>et al.</i> , 2004	
Pacific oyster (Crassostrea gigas)	Li and Guo, 2004	
European sea bass (Dicentrarchus labrax)	Christiakov <i>et al.</i> , 2005	
Brown trout (Salmo trutta)	Gharbi <i>et al.</i> , 2006	
Gilthead sea bream (Sparus aurata)	Franch <i>et al.</i> , 2006	
Atlantic halibut (Hippoglossus hippoglossus)	Reid <i>et al.</i> , 2007	
European flat oyster (Ostrea edulis)	Lallias <i>et al.</i> , 2007	
Bighead carp (Aristichthys nobilis)	Liao <i>et al.</i> , 2007	
Silver carp (Hypophthamichthys molitrix)	Liao <i>et al.</i> , 2007	
Coho salmon (Oncorhynchus kisutch)	McClelland and Naish, 2008	

Table 1. Genetic linkage maps in aquaculture species.

problem through the possibility of detecting sex-linked markers in different species, including Atlantic salmon (Devlin *et al.*, 1991; Du *et al.*, 1993; Clifton and Rodriguez, 1997), African catfish (Kovacs *et al.*, 2000), rainbow trout (Iturra *et al.*, 2001; Felip *et al.*, 2005), tongue sole (Chen *et al.*, 2008).

$\label{eq:individual} Individual \ identification \ and \ parentage \\ assignment$

In many fields, such as conservation genetics or selection, the unambiguous identification of the individuals and reliable genealogical records are required for the management programmes. Unfortunately, these data are difficult to obtain in fish populations, because physical tags are impossible to apply in juveniles and, when possible, such as in farmed mussels, they are lost in 40-90% of the cases (MacAvoy *et al.*, 2008). The alternative to keep different families in separate ponds is expensive and limits the number of animals available for selection.

Several studies have demonstrated the ability to identify a unique genetic profile for each individual and to establish parentage in fish using highly polymorphic markers, especially microsatellites. Investigations on rainbow trout (*Onchorhynchus mykiss*) (Herbinger *et al.*, 1995), Atlantic salmon (*Salmo salar*) (Norris *et al.*, 2000) and the New Zealand mussel *Perna canaliculus* (MacAvoy *et al.*, 2008) showed that appropriate sets of microsatellites allow a success rate of 95-99.9% in parentage assignment. The disadvantage of using molecular markers is the relatively high cost, which can be limited by choosing the minimum number of markers compatible with the maximum level of accuracy and developing multiplex systems able to co-amplify the markers used, as already done for many aquaculture species (O'Reilly *et al.*, 1996; Fishback *et al.*, 1999; Porta *et al.*, 2006; Johnson *et al.*, 2007).

Population genetics

The genetic characterization of the individuals leads to the possibility of describing a population by means of allele frequencies. In recent decades, the molecular markers have been extensively used to define the genetic structure of many aquaculture populations, which represents the fundamental step for the definition of the taxonomic status, that is for species, subspecies, breeds and strains identification. From a practical point of view, once a status has been assigned to each population, the information can be used for either understanding their role in determining the whole variability of the species, or detecting cases of crossbreeding and/or hybridization.

For the identification of species, separated by large genetic distances, almost all markers can be used. On the contrary, markers with a very high resolution power are needed for breeds and strains identification because the genetic distances between the phylogenetic units are often quite small. The microsatellite variability made it possible to distinguish European and American populations of Atlantic salmon (McConnell et al., 1995), as well as seven populations of masu salmon living in the Atsuta river (Kitanishi et al., 2009), while significant differences between two adjacent Canadian populations were revealed by mtDNA (Tessier et al., 1995). David et al. (2001) successfully applied AFLP markers to distinguish nine common carp populations, showing that the Amur carp was the most different, so that

one marker was sufficient to recognize it, while two or more markers were necessary to distinguish the other populations.

Molecular markers are widely used in conservation genetics as well, to indirectly estimate the inbreeding level by measuring the heterozygosity degree, whose changes during time reveal population size fluctuations and possible bottlenecks occurred in the past (Gross et al., 2007). The main purpose of these studies is to provide tools for preserving the existing genetic variability, which is fundamental for the survival of the species, because it allows individuals to face changes in the environmental conditions. Focusing on farmed animals, the genetic diversity represents the possibility of both adapting to new rearing conditions/food sources/diseases, and providing improved products in answer to new requirements. Nevertheless, any selection programme leads to a reduction of the genetic diversity, which should be limited as much as possible in order to avoid the negative effects, known as inbreeding depression (Pante et al., 2001). From this point of view, molecular information can be used to maximise the genetic diversity when assembling a founder population in order to ensure maximum long-term genetic response from the breeding programmes (Hayes et al., 2006).

Fortunately, most of aquaculture species have a great advantage compared to other species: as the domestication process started only recently, wild populations still exist and can represent the source of genetic diversity in the future. In this context, molecular genetics plays a basic role because molecular markers are able to detect differences between wild and cultured populations and to reveal processes which determined the observed differences. Mitochondrial DNA is especially employed for this purpose in several aquaculture species. For example, Hansen *et al.* (1997) used mtDNA polymorphism to detect differences between brown trout cultured strains and wild populations from three river basins, revealing a greater reduction of genetic variability in hatchery strains, mainly due to the small numbers of founder females. Also data on microsatellite markers showed that farmed strains were genetically quite similar, while clearly separated from wild populations in rainbow trout (Gross *et al.*, 2007) and tench (Kohlmann *et al.*, 2007). The ability to discriminate wild and cultured populations can be also exploited to identify escaped domesticated animals, as demonstrated for Chinook salmon reared in marine netpens (Withler *et al.*, 2007).

The genetic characterization offers the possibility to trace back to the origin of a processed product (traceability). Species-specific, strain-specific or population-specific markers can be used as genetic tags, which make it possible to detect the original taxon and secure the consumer's rights to be informed about the purchased product. For example, the polymorphism of the mitochondrial Cyt b gene made it possible to discriminate between 23 species, including European eel (Anguilla anguilla), Atlantic salmon (Salmo salar), Atlantic cod (Gadus morhua), bass (Dicentrarchus labrax), sea bream (Sparus aurata), tuna (Thunnus thynnus) and plaice (Pleuronectes platessa) (Wolf et al., 2000). Recently a species database of fish, molluscs and crustaceans has been created with the aim to identify species of origin of seafood products by previously defined AFLP patterns (Maldini et al., 2006).

A strictly related application is in forensic science to detect cases of fraud, for example in caviar trade (Wuertz *et al.*, 2007), or illegal poaching of threatened species. Primmer *et al.* (2000) reported a funny case of fraud that occurred during a fishing competition in Finland: the microsatellite genotyping, together with software able to assign an individual to its population, provided a highly significant power for excluding the possibility of a suspected fish originating from the competition lake. At the end, the offender confessed to purchasing it in a local fish shop!

Selection

Selection is aimed at modifying the genetic structure of a breed in order to obtain animals with superior performances for the traits of interest. The classical approach is to estimate the breeding value of the individuals on the basis of phenotypic values, to select the ones with the best genetic performances and to mate them within appropriate breeding schemes.

Although the basic concepts are the same, the selection strategies used in most farm animals do not directly apply to fish species for their biological and breeding characteristics. On one hand, the extremely high reproductive capacity and the external fertilization of fish offer a great flexibility in the implementation of selection programmes with a high precision in the estimates and permits the use of higher intensities of selection. On the other hand, the peculiar breeding management has some practical limitations, such as the difficulty in obtaining accurate genealogical and phenotypic data, or the influence of the competition between individuals in the same pond, which induces a distortion in the estimates of the genetic parameters, with negative effects on the selection response (Moav and Wohlfart, 1974).

Apart from these differences, the traits objectives of selection in aquaculture are similar to those of other species (including growth, carcass composition and quality) and the genetic improvement realized so far mainly depends on the application of traditional methods, involving selection, crossing and hybridation (Wohlfarth, 1993; Bakos and Gorda, 1995; Hulata, 1995; Hulata, 2001). However, the progress in molecular genetics provides perspectives for implementing the marker assisted selection (MAS), aimed at choosing the genetically superior individuals using molecular information.

To perform MAS, markers tightly linked to the loci responsible for quantitative traits (QTL) or major genes directly involved in the phenotypic expression should be found. In the first case, a QTL can be identified and localized due to the co-segregation with a molecular marker; in this respect the microsatellites are very helpful, being highly polymorphic and widely distributed throughout the genome. In the second case, candidate genes possibly responsible for quantitative traits are chosen, based on previous knowledge of their position and/or function, and the statistical associations between their SNPs and the phenotypic expression of the trait of interest are investigated. The identification of genetic markers related to traits objective of selection would have a great impact on the selection response, mainly for traits with low heritability, such as reproduction or disease resistance. In fact, if a given allele were associated to a given trait, it would be sufficient to select the individuals carrying that allele in order to improve the associated trait. If so, the selection process would be greatly simplified, because simple Mendelian traits would be concerned, instead of complex quantitative traits.

Up to now, the QTL analysis in fish is in its infancy because the genetic mapping is not so advanced as in other species, even if medium-density linkage maps are available at least for the principal aquaculture species, as mentioned above. The first data on QTLs date back to the late 1990's, when Jackson *et al.* (1998) identified two markers associated with the upper temperature tolerance in rainbow trout. Later on, other QTLs were reported in different aquaculture species (Table 2).

In aquaculture many efforts are devoted to the search of markers for disease resist-

ance, due to both the enormous economical implications and the difficulties of the classical selection, which requires the genetic evaluation of the individuals through exposure to the virus. Ozaki et al. (2001) first identified, in rainbow trout, two markers associated with the infectious pancreatic necrosis (IPN), a highly contagious disease against which the presently available vaccines offer only a partial protection. More recently, a marker for resistance to lymphocystis disease in Japanese flounder has been identified (Fuji et al., 2007); the potential of MAS for improving the disease resistance is demonstrated by the fact that no affected fish were observed in an experimental population selected for the allele associated to the lymphocystis resistence, compared to about 5% of the control population.

Concerning major genes, the available data are still limited (Table 3). However, aquaculture species have the great advantage that the economically important traits are similar to those included in the selection programmes of many farm animals, so that the huge amount of information already available for other vertebrates can be exploited to find homologous genes in fish, where the research in this field is less advanced. For example, growth rate, which is one of the main objectives of selection in most fish species, has been thoroughly investigated for many years in other animals, where the genes associated to the somatotropic axis have been identified, mapped and sequenced. These results have accelerated the research in fish, leading to the identification in different species of GH (Growth Hormone), GHR (Growth Hormone Receptor), GHRH (Growth Hormone Releasing Hormone), IGF-I (Insulin-like Growth Factor I) genes (De-Santis and Jerry, 2007). The identification of polymorphic sites in these genes is the further step towards investigations on the possible associations with production

Table 2. Examples of QTL in aquaculture species.				
Species	Trait	Reference		
Rainbow trout (Oncorhyncus mykiss)	Upper temperature tolerance	Jackson <i>et al.</i> , 1998		
	Spawning time	Sakamoto <i>et al.</i> , 1999		
	IPNV resistance	Ozaki <i>et al.</i> , 2001		
	Embryonic development rate	Robison et al., 2001		
	Sex	Iturra <i>et al.</i> , 2001		
	Early maturation	Haidle <i>et al.</i> , 2007		
	Cortisol level	Drew <i>et al.</i> , 2007		
Tilapia (Oreochromis hybrid)	Stress/immune response	Cnaani <i>et al.</i> , 2004		
	Body colour	Lee <i>et al.</i> , 2005		
Coho salmon (Oncorhynchus kisutch)	Flesh colour	Araneda <i>et al.</i> , 2005		
	Spawning date	Araneda <i>et al.</i> , 2007		
Atlantic salmon (Salmo salar)	IPNV resistance	Houston <i>et al.</i> , 2007		
	Body lipid percentage	Derayat <i>et al.</i> , 2007		
Sea bass (Dicentrarchus labrax)	Morphometric traits	Chatziplis et al., 2007		
Japanese flounder (Paralichthys olivaceus)	Lymphocystis disease	Fuji <i>et al.</i> , 2007		
Eastern oyster (Crassostrea virginica)	Disease resistance	Yu and Guo, 2006		

Table 2.	Examples of	QTL in	aquaculture	species.
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Table 3. Examples of candidate genes in fish species.						
Species	Gene	Trait	Reference			
Zebrafish (Danio rerio)	МҮО	Growth	Acosta <i>et al.</i> , 2005			
Rainbow trout (Oncorhyncus mykiss)	Clock	Spawning time	Leder <i>et al.</i> , 2006			
Asian seabass (Lates calcarifer)	PVALB1	Body weight/ length	Xu <i>et al.</i> , 2006			
Grouper (Epinephelus coioides)	Epinecidin-1	Antimicrobial activity	Yin <i>et al.</i> , 2006			
Tilapia <i>(Nile tilapia)</i>	ACTB, ATP2B1, HBB, POMC	Water salt tolerance	Rengmark et al., 2007			
Masu salmon (Oncorhyncus masou)	MELO	HUFA biosynthesis	Alimuddin <i>et al.</i> , 2008			

traits. Another promising candidate gene is Myostatin, whose variability is responsible for the double muscled phenotype in cattle. Recently, Acosta et al (2005) reported that

the inactivation of the Myostatin gene in zebrafish resulted in an increased weight gain (+45% compared to the control), as observed in mice (McPherron et al., 1997). Therefore, the polymorphism of the gene could be associated to growth differences also in fish.

Transgenesis

The knowledge of molecular genetics gives a basic contribution to the genetic engineering, which in fish is more advanced compared to other animals, for the simpler manipulation, due to the external fertilization and embryogenesis (Maclean, 2003).

Following the milestone experiment of Palmiter et al. (1982) in mice, the main application in aquaculture concerned the growth rate, with the successful transfer of *GH* gene from mammals and more recently from fish. The results were impressive, with growth rate four times higher in salmon (Devlin et al., 1994) and 2.5 - 4 times in tilapia (Rahman et al., 2001). However, some results showed that the growth enhancement is relatively low in species selected for growth over centuries, as they had less 'capacity' for extra growth (Devlin et al., 2001). Therefore, the existence of biological limitations could reduce the usefulness of the transgenesis. The consumer acceptance of the transgenic fish, probably related to the perceived risk, and possible adverse environmental impacts are also to be taken into account (Maclean, 2003).

Considerable work in the field of transgenesis concerns the pathogen resistance and freeze resistance, but, even if transgenic fish have been produced, the research remains at a preliminary stage (Maclean, 2003). The development of ornamental fish

REFERENCES

- Acosta, J., Carpio, Y., Borrato, I., Gonzales, O., Estrada, M.P., 2005. Myostatin gene silenced by RNAi shows a zebrafish giant phenotype. J. Biotechnol. 119:342-331.
- Agresti, J.J., Seki, S., Cnaani, A., Poompuang, S., Hallerman, E.M., Umiel, N., Hulata, G., Gall,

expressing naturally fluorescent proteins in the skeletal muscle (Gong *et al.*, 2003) and the construction of models for human diseases (Kari *et al.*, 2007) demonstrate the wide range of application of the transgenic technology.

Conclusions

For the last twenty years the genetic research in aquaculture has been exponentially increasing thanks to the widespread use of molecular technologies together with the possibility to exploit the impressive amount of data available for other species.

Up to now the knowledge on molecular genetics has been mainly applied to the genetic characterization of the populations, covering a variety of aspects, with special emphasis on diversity analysis and conservation. For the future considerable progress can be expected from gene mapping thanks to the efforts presently devoted to both the enrichment of the genetic maps and to the integration of genetic linkage and physical maps, which is essential for the understanding of genes responsible for performance traits, including growth and disease resistance. With the increasing global demand for aquaculture products and the early stage of selection for most aquatic species, molecular genetics is expected to play a major role in the management of breeding programmes aimed at developing improved strains for the most economically important species.

G.A.E., May, B., 2000. Breeding new strains of tilapia: development of an artificial center of origin and linkage map based on AFLP and microsatellite loci. Aquaculture 185:43-56.

Alimuddin, Kiron, V., Satoh, S., Takeuchi, T., Yoshizaki, G., 2008. Cloning and over-expression of a masu salmon (*Oncorhynchus masou*) fatty acid elongaselike gene in zebrafish. Aquaculture 282:13-18.

- Antunes, A., Alexandrino, P., Ferrand, N., 1999. Genetic characterization of Portuguese brown trout (*Salmo trutta* L.) and comparison with other European populations. Ecol. Freshw. Fish 8:194-200.
- Araneda, C., Díaz, N.F., Gómez, G., Iturra, P., 2007. Identification and characterization of three new molecular markers associated with spawning date in coho salmon (*Oncorhynchus kisutch*). Aquaculture 272(Suppl.1):S240 (abstr.).
- Araneda, C., Neira, R., Iturra, P., 2005. Identification of a dominant SCAR marker associated with colour traits in Coho salmon (*Oncorhynchus kisutch*). Aquaculture 247:67-73.
- Avise, J.C., 1994. Molecular Markers, Natural History and Evolution. Chapman and Hall, New York, NY, USA.
- Bakos, J., Gorda, S., 1995. Genetic improvement of common carp strains using intraspecific hybridization. Aquaculture 129:183-186.
- Chang, Y., Huang, H.D., 1994. The complete nucleotide sequence and gene organisation of carp (*Cyprinus carpio*) mitochondrial genome. Molec. Evol. 38:138-155.
- Chatziplis, D., Batargias, C., Tsigenopoulos, C.S., Magoulas, A., Kollias, S., Kotoulas, G., Volckaert, F.A.M., Haley, C.S., 2007. Mapping quantitative trait loci in European sea bass (*Dicentrarchus labrax*): the BASSMAP pilot study. Aquaculture 272(Suppl. 1):S171-S182.
- Chen, S.L., Deng, S.P., Ma, H.Y., Tian, Y.S., Xu, J.Y., Yang, J.F., Wang, Q.Y., Ji, X.S., Shao, C.W., Wang, X.L., Wu, P.F., Deng, H., Zhai, J.M., 2008. Molecular marker-assisted sex control in half-smooth tongue sole (*Cynoglossus semilaevis*). Aquaculture 283:7-12.
- Christiakov, D.A., Hellemans, B., Haley, C.S., Law, A.S., Tsigenopoulos, B., Kotoulas, G., Bertotto, D., Libertini, A., Volckaert, F.A.M., 2005. A microsatellite linkage map of the European sea bass *Dicentrarchus labrax* L. Genetics 170:1821-1826.
- Clifton, D.R., Rodriguez, R.J., 1997. Characterization and application of a quantitative DNA marker that discriminates sex in chinook salmon (*Oncorhynchus tshawytscha*). Can. J. Fish Aquat. Sci. 54:2647-2652.

- Cnaani, A., Zilberman, N., Tinman, S., Hulata, G., Ron, M., 2004. Genome-scan analysis for quantitative trait loci in an F2 tilapia hybrid. Mol. Genet. Genomics 272:162-172.
- Danzmann, R.G., Gharbi, K., 2001. Gene mapping in fishes: a means to an end. Genetica 111:3-23.
- David, L., Rajasekaran, P., Fang, J., Hollet, J., Lavi, U., 2001. Polymorphism in ornamental and common carp strains (*Cyprinus carpio* L.) as revealed by AFLP analysis and a new set of microsatellite markers. Mol. Genet. Genomics 266:353-362.
- Derayat, A., Houston, R.D., Guy, D.R., Hamilton, A., Ralph, J., Spreckley, N., Taggart, J.B., McAndrew, B.J., Haley, C.S., 2007. Mapping QTL affecting body lipid percentage in Atlantic salmon (*Salmo salar*). Aquaculture 272(Suppl. 1):S250-S251.
- De-Santis, C., Jerry, D.R., 2007. Candidate growth genes in finfish – Where should we be looking?. Aquaculture 272:22-38.
- Devlin, R.H., Biagi. C.A., Yesaki, T.Y., Smailus, D.E., Byatt, J.C., 2001. Growth of domesticated transgenic fish. Nature 409:781-782.
- Devlin, R.H., McNeil, B.K., Groves, T.D.D., Donaldson, E.M., 1991. Isolation of a Y-chromosomal DNA probe capable of determining genetic sex in chinook salmon (*Oncorhynchus tshawytscha*). Can. J. Fish Aquat. Sci. 48:1606-1612.
- Devlin, R.H., Nagahama, Y., 2002. Sex determination and sex differentiation in fish: an overview of genetic, physiological, and environmental influences. Aquaculture 208:191-364.
- Devlin, R.H., Yesaki, T.Y., Biagi, C.A., Donaldson, E.M., Swanson, P., Chan, W.K., 1994. Extraordinary salmon growth. Nature 371:209-210.
- Drew, R.E., Schwabl, H., Wheeler, P.A., Thorgaard, G.H., 2007. Detection of QTL influencing cortisol levels in rainbow trout (*Oncorhynchus mykiss*). Aquaculture 272(Suppl.1):S183-S194.
- Du, S.J., Devlin, R.H., Hew, C.L., 1993. Genomic structure of growth hormone genes in chinook salmon (*Oncorhynchus tshawytscha*): presence of two functional genes, GH-I and GH-II, and a male specific pseudogene, GH-ψ. DNA Cell Biol. 12:739-751.
- Felip, A., Young, W.P., Wheeler, P.A., Thorgaard, G.H.,

2005. An AFLP-based approach for the identification of sex-linked markers in rainbow trout (*Oncorhynchus mykiss*). Aquaculture 247:35-43.

- Fishback, A.G., Danzmann, R.G., Sakamoto, T., Ferguson, M.M., 1999. Optimization of semi-automated microsatellite multiplex polymerase chain reaction systems for rainbow trout (Oncorhynchus mykiss). Aquaculture 172:247-254.
- Franch, R., Louro, B., Tsalavouta, M., Chatziplis, D., Tsigenopoulos, C.S., Sarropoulou, E., Antonello, J., Magoulas, A., Mylonas, C.C., Babbucci, M., Patarnello, T., Power, D.M., Kotoulas, G., Bargelloni, L., 2006. A genetic linkage map of the hermaphrodite teleost fish *Sparus aurata* L. Genetics 174:851-861.
- Fuji, K., Hasegawa, O., Honda, K., Kumasaka, K., Sakamoto, T., Okamoto, N., 2007. Marker-assisted breeding of a lymphocystis disease-resistant Japanese flounder (*Paralichthys olivaceus*). Aquaculture 272:291-295.
- Gharbi, K., Gautier, A., Danzmann, R.G., Gharbi, S., Sakamoto, T., Høyheim, B., Taggart, J.B., Cairney, M., Powell, R., Krieg, F., Okamoto, N., Ferguson, M.M., Holm, L.E., Guyomard, R., 2006. A linkage map for brown trout (*Salmo trutta*): chromosome homeologies and comparative genome organization with other salmonid fish. Genetics 172:2405-2419.
- Gilbey, J., Verspoor, E., McLay, A., Houlihan, D., 2004. A microsatellite linkage map for Atlantic salmon (*Salmo salar*). Anim. Genet. 35:98-105.
- Gong, Z., Wan, H., Tay, T.L., Wang, H., Chen, M., Yan, T., 2003. Development of transgenic fish for ornamental and bioreactor by strong expression of fluorescent proteins in the skeletal muscle. Biochem. Bioph. Res. Co. 308:58-63.
- Gregory, D.J., Waldbieser, G.C., Bosworth, B.G., 2004. Cloning and characterization of myogenic regulatory genes in three Ictalurid species. Anim. Genet. 35:425-430.
- Gross, R., Lulla, P., Paaver, T., 2007. Genetic variability and differentiation of rainbow trout (*Oncorhynchus mykiss*) strains in northern and Eastern Europe. Aquaculture 272(Suppl. 1):S139-S146.
- Haidle, L., Janssen, J., Gharbi, K., Danzmann, R.G., Ferguson, M.M., 2007. Identifying regions of the

genome responsible for early maturation and associated traits in rainbow trout (*Oncorhynchus mykiss*). Aquaculture 272(Suppl.1):S265-S266.

- Hansen, M.M., Menserberg, K.D., Rasmussen, G., Simonsen, V., 1997. Genetic variation within and among Danish brown trout (*Salmo trutta* L.) hatchery strains, assessed by PCR-RFLP analysis of mitochondrial DNA segments. Aquaculture 153:15-29.
- Hayes, B., He, J., Moen, T., Bennewitz, J., 2006. Use of molecular markers to maximise diversity of founder populations for aquaculture breeding programs. Aquaculture 255:573-578.
- Herbinger, C.M., Doyle, R.W., Oitman, E.R., Paquet, D., Mesa, K.A., Morris, D.B., Wright, J.M., Cook, D., 1995. DNA fingerprint-based analysis of paternal and maternal effects on offspring growth and survival in communally reared rainbow trout. Aquaculture 137:245-256.
- Houston, R.D., Guy, D.R., Hamilton, A., Ralph, J., Spreckley, N., Taggart, J.B., McAndrew, B.J., Haley, C.S., Bishop, S.C., 2007. Mapping QTL affecting resistance to infectious pancreatic necrosis (IPN) in Atlantic salmon (*Salmo salar*). Aquaculture 272(Suppl. 1):S269.
- Hulata, G., 1995. A review of genetic improvement of the common carp (*Cyprinus carpio* L.) and other hybrids by crossbreeding, hybridization and selection. Aquaculture 129:143-155.
- Hulata, G., 2001. Genetic manipulations in aquaculture: a review of stock improvement by classical and modern technologies. Genetica 11:155-173.
- Iturra, P., Bagley, M., Vergara, N., Imbert, P., Medrano, J.F., 2001. Development and characterization of DNA sequence *OMYP9* associated with the sex chromosomes in rainbow trout. Heredity 86:412-419.
- Jackson, T.R., Ferguson, M.M., Danzmann, R.G., Fishback, A.G., Ihssen, P.E., O'Connell, M., Crease, T.J., 1998. Identification of two QTL influencing upper temperature tolerance in three rainbow trout (Oncorhynchus mykiss) half-sib families. Heredity 80:143-151.
- Johnson, N.A., Rexroad, C.E., Hallerman, E.M., Vallejo, R.L., Palti, Y., 2007. Development and evalua-

tion of a new microsatellite multiplex system for parental allocation and management of rainbow trout (*Oncorhynchus mykiss*) broodstocks. Aquaculture 266:53-62.

- Kari, G., Rodeck, U., Dicker, A.P., 2007. Zebrafish: an emerging model system for human disease and drug discovery. Clin. Pharmacol. Ther. 82:70-80.
- Kitanishi, S., Yamamoto, T., Higashi, S., 2009. Microsatellite variation reveals fine-scale genetic structure of masu salmon, *Oncorhynchus masou*, within the Atsuta River. Ecol. Freshw. Fish 18:65-71.
- Knapik, E.W., Goodman, A., Ekker, M., Chevrette, M., Delgado, J., Neuhauss, S., Shimoda, N., Driever, W., Fishman, M.C., Jacob, H.J., 1998. A microsatellite genetic linkage map for zebrafish (*Danio rerio*). Nat. Genet. 18:338-343.
- Kocher, T.D., Lee, W.J., Sobolewska, H., Penman, D., McAndrew, B., 1998. A genetic linkage map of a cichlid fish, the tilapia (*Oreochromis niloticus*). Genetics 148:1225-1232.
- Kohlmann, K., Kersten, P., 1998. Enzyme variability in a wild population of tench (*Tinca tinca* L.). Pol. Arch. Hydrobiol. 45:303-310.
- Kohlmann, K., Kersten, P., Flajšhans, M., 2007. Comparison of microsatellite variability in wild and cultured tench (*Tinca tinca*). Aquaculture 272(Suppl. 1):S147-S151.
- Kovacs, B., Egedi, S., Bartfai, R., Orban, L., 2000. Male-specific DNA markers from African catfish (*Clarias gariepinus*). Genetica 110:267-276.
- Lallias, D., Beaumont, A.R., Haley, C.S., Boudry, P., Heurtebise, S., Lapègue, S., 2007. A first generation genetic linkage map of the European flat oyster Ostrea edulis (L.) based on AFLP and microsatellite markers. Anim. Genet. 38:560-568.
- Leder, E.H., Danzmann, R.G., Ferguson, M.M., 2006. The candidate gene, Clock, localizes to a strong spawning time quantitative trait locus region in rainbow trout. J. Hered. 97:74-80.
- Lee, B.Y., Lee, W.J., Streelman, J.T., Carleton, K.L., Howe, A.E., Hulata, G., Slettan, A., Stern, J.E., Terai, Y., Kocher, T.D., 2005. A second-generation genetic linkage map of Tilapia (*Oreochromis* ssp.). Genetics 170:237-244.

- Levinson, G., Gutman, G.A., 1987. High frequency of short frameshift in poly-CA/GT tandem repeats borne by bacteriophage M13 in Escherichia coli K-12. Nucleic Acids Res. 15:5323-5338.
- Li, L., Guo, P., 2004. AFLP-based genetic linkage maps of the pacific oyster *Crassostrea gigas* Thunberg. Mar. Biotechnol. 6:26-36.
- Liao, M., Zhang, L., Yang, G., Zhu, M., Wang, D., Wie, Q., Zou, G., Chen, D., 2007. Development of silver carp (*Hypophthamichthys molitrix*) and bighead carp (*Aristichthys nobilis*) genetic maps using microsatellite and AFLP markers and a pseudotestcross strategy. Anim. Genet. 38:364-370.
- Liu, Z.J., Karsi, A., Li, P., Cao, D., Dunham, R., 2003. An AFLP-based genetic linkage map of channel catfish (*Ictalurus punctatus*) constructed by using an interpecific hybrid resource family. Genetics 165:687-694.
- MacAvoy, E.S., Wood, A.R., Gardner, J.P.A., 2008. Development and evaluation of microsatellite markers for identification of individual Greenshell[™] mussels (*Perna canaliculus*) in a selective breeding programme. Aquaculture 274:41-48.
- Maclean, N., 2003. Genetically modified fish and their effects on food quality and human health and nutrition. Trends Food Sci. Tech. 14:242-252.
- Maldini, M., Nonnis Marzano, F., Gonzales Fortes, G., Papa, R., Gandolfi, G., 2006. Fish and seafood traceability based on AFLP markers: elaboration of a species database. Aquaculture 261:487-494.
- Matsuoka, M.P., Gharret, A.J., Wilmot, R.L., Smoker, W.W., 2004. Genetic linkage mapping of allozyme loci in even- and odd-year pink salmon (Oncorhynchus gorbuscha). J. Hered. 95:421-429.
- McClelland, E.K., Naish, K.A., 2008. A genetic linkage map for coho salmon (Oncorhynchus kisutch). Anim. Genet. 39:169-179.
- McConnell, S.K., O'Reilly, P., Hamilton, L., Wright, J.M., Bentzen, P., 1995. Polymorphic microsatellite loci from Atlantic salmon (*Salmo salar*): genetic differentiation of North American and European populations. Can. J. Fish Aquat. Sci. 52:1863-1872.
- McPherron, A.C., Lawler, A.M., Lee, S.J., 1997. Regulation of skeletal muscle mass in mice by a new

TGF- β superfamily member. Nature 387:83-90.

- Moav, R., Wohlfarth, D.W., 1974. Magnification through competition of genetic differences in yield capacity in carp. Heredity 33:181-202.
- Moen, T., Hayes, B., Baranski, M., Berg, P.R., Kjøglum, S., Koop, B.F., Davidson, W.S., Omholt, S.W., Lien, S., 2008. A linkage map of the Atlantic salmon (*Salmo salar*) based on EST-derived SNP markers. BMB Genomics 9:223-237.
- Moen, T., Hoyheim, B., Munck, H., Gomez-Raya, L., 2004. A linkage map of Atlantic salmo (Salmo salar) reveals an uncommonly large difference in recombination rate between sexes. Anim. Genet. 35:81-92.
- Moritz, C., 1994. Applications of mitochondrial DNA analysis in conservation: A critical review. Mol. Ecol. 3:401-411.
- Morizot, D.C., 1983. Tracking linkage groups from fishes to mammals. J. Hered. 74:413-416.
- Naruse, K., Fukamachi, S., Mitani, H., Kondo, M., Matsuoka, T., Kondo, S., Hanamura, N., Morita, Y., Hasegawa, K., Nishigaki, R., Simada, A., Wada, H., Kusakabe, T., Suzuki, N., Kinoshita, M., Kanamori, A., Terado, T., Kimura, H., Nonaka, M., Shima, A., 2000. A detailed linkage map of medaka, *Oryzias latipes*: comparative genomics and genome evolution. Genetics 154:1773-1784.
- Nichols, K.M., Young, W.P., Danzmann, R.G., Robinson, B.D., Rexroad, C., Noakes, M., Phillips, R.B., Bentzen, P., Spies, I., Knudsen, K., Allendorf, F.W., Cunningham, B.M., Brunelli, J., Zhang, H., Ristow, S., Drew, R., Brown, K.H., Wheeler, P.A., Thorgaard, G.H., 2003. A consolidated linkage map for rainbow trout (*Oncorhynchus mykiss*). Anim. Genet. 34:102-115.
- Norris, A.T., Bradley, D.G., Cunningham, E.P., 2000. Parentage and relatedness determination in farmed Atlantic salmon (*Salmo salar*) using microsatellite markers. Aquaculture 182:73-83.
- O'Connell, M., Wright, J.M., 1997. Microsatellite DNA in fishes. Rev. Fish Biol. Fisher. 7:331-363.
- O'Reilly, P.T., Hamilton, L.C., McConnell, S.K., Wright, J.M., 1996. Rapid analysis of genetic variation in Atlantic salmon (*Salmo salar*) by PCR multiplexing of dinucleotide and tetranucleotide

microsatellites. Can. J. Fish Aquat. Sci. 53:2292-2298.

- Ozaki, A., Sakamoto, T., Khoo, S., Nakamura, K., Coimbra, M., Akutsu, T., Okamoto, N., 2001. Quantitative trait loci (QTLs) associated with resistance/susceptibility to infectious pancreatic necrosis virus (IPNV) in rainbow trout (*Oncorhynchus mykiss*). Mol. Genet. Genomics 265:23-31.
- Palmiter, R.D., Brinster, R.L., Hammer, R.E., Trumbauer, M.E., Rosenfeld, M.G., Birnberg, N.C., Evans, R.M., 1982. Dramatic growth of mice that develop from eggs microinjected with metallothionein-growth hormone fusion genes. Nature 300:611-615.
- Pante, M.J.R., Gjerde, B., McMillan, I., 2001. Inbreeding levels in elected populations of rainbow trout, *Oncorhynchus mykiss*. Aquaculture 192:201-211.
- Porta, J., Porta, J.M., Martinez-Rodriguez, G., Alvarez, M.C., 2006. Development of a microsatellite multiplex PCR for Senegalese sole (*Solea senegalensis*) and its application to broodstock management. Aquaculture 256:159-166.
- Primmer, C.R., Koskinen, M.T., Piironen, J., 2000. The one that did not get away: individual assignment using microsatellite data detects a case of fishing competition fraud. P. Roy. Soc. Lond. B Bio. 267:1699-1704.
- Rahman, M.A., Ronyai, A., Engidaw, B.Z., Jauncey, K., Hwang, G.L., Smith, A., Roderick, E., Penman, D., Varadi, L., Maclean, N., 2001. Growth and nutritional trials on transgenic Nile tilapia containing an exogenous fish growth hormone gene. J. Fish Biol. 59:62-78.
- Reid, D.P., Smith, C.A., Rommens, M., Blanchard, B., Martin-Robichaud, D., Reith, M., 2007. A genetic linkage map of Atlantic halibut (*Hippoglossus hippoglossus* L.). Genetics 177:1193-1205.
- Rengmark, A.H., Slettan, A., Lee W.J., Lie, Ø., Lingaas, F., 2007. Identification and mapping of genes associated with salt tolerance in tilapia. J. Fish Biol. 71(Suppl. C):409-422.
- Rexroad, C.E., Palti, Y., Gahr, S.A., Vallejo, R.L., 2008. A second generation genetic map for rainbow trout (*Oncorhynchus mykiss*). BMC Genet. 9:74-87.

- Robison, B.D., Wheeler, P.A., Sundin, K., Sikka, P., Thorgaard, G.H., 2001. Composite interval mapping reveals a major locus influencing embryonic development rate in rainbow trout (*Oncorhynchus mykiss*). J. Hered. 92:16-22.
- Saitoh, K., Sado, T., Mayden, R.L., Hanzawa, N., Nakamura, K., Nishida, M., Miya, M., 2006. Mitogenomic evolution and interrelationships of the Cypriniformes (*Actinopterygii: Ostariophysi*): The first evidence toward resolution of higher-level relationships of the world's largest freshwater fish clade based on 59 whole mitogenome sequences. J. Mol. Evol. 63:826-841.
- Sakamoto, T., Danzmann, R.G., Okamoto, N., Ferguson, M.M., Ihssen, P.E., 1999. Linkage analysis of quantitative trait loci associated with spawning time in rainbow trout (*Oncorhynchus mykiss*). Aquaculture 173:33-43.
- Shimoda, N., Knapik, E.W., Ziniti, J., Sim, C., Yamada, E., Kaplan, S., Jackson, D., DeSauvage, F., Jacob, H., Fishman, M.C., 1999. Zebrafish genetic map with 2000 microsatellite markers. Genomics 58:219-232.
- Šlechtová, V., Šlechtá, V., Valenta, M., 1995. Genetic protein variability in tench (*Tinca tinca* L.) stocks in Czech Republic. Pol. Arch. Hydrobiol. 42:133-140.
- Somridhivej, B., Wang, S., Sha, Z., Liu, H., Quilang, J., Xu, P., Li, P., Hu, Z., Liu, Z., 2008. Characterization, polymorphism assessment, and database construction for microsatellites from BAC end sequences of channel catfish (*Ictalurus punctatus*): A resource for integration of linkage and physical maps. Aquaculture 275:76-80.
- Tautz, D., 1989. Hypervariability of simple sequences as a general source for polymorphic DNA markers. Nucleic Acids Res. 17:6463-6471.
- Tessier, N., Bernatchez, L., Presa, P., Angers, B., 1995. Gene diversity analysis of mitochondrial DNA, microsatellites and allozymes in landlocked Atlantic salmon. J. Fish Biol. 47(Suppl. A):156-163.
- Valenta, M., Šlechtová, V., Kàlal, L., Stratil, A., Janatkova, J., Šlechtá, V., Rab, P., Pokorny, J., 1978. Polymorphic proteins of the common carp (*Cyprinus carpio*) and tench (*Tinca tinca*) and the possibility of using them in breeding. Zivocisna

vyroba 23:797-809.

- Valenta, M., Stratil, A., Kàlal, L., 1977. Polymorphism and heterogeneity of transferrins in some species of the fish family Cyprinidae. Anim. Blood Grps. biochem. Genet. 8:93-109.
- Waldebieser, G.C., Bosworth, B.G., Nonneman, D.J., Wolters, W.R., 2001. A microsatellite-based genetic linkage map for channel catfish, *Ictalurus punctatus*. Genetics 158:727-734.
- Weber, J.L., Wong, C., 1993. Mutation of human short tandem repeats. Hum. Mol. Genet. 2:1123-1128.
- Withler, R.E., Rundle, T., Beacham, T.D., 2007. Genetic identification of wild and domesticated strains of chinook salmon (*Oncorhynchus tshawytscha*) in southern British Columbia, Canada. Aquaculture 272(Suppl.1):S161-S171.
- Wohlfarth, G.W., 1993. Heterosis for growth rate in common carp. Aquaculture 113:31-46.
- Wolf, C., Burgener, M., Hubner, P., Luthy, J., 2000. PCR-RFLP analysis of mitochondrial DNA: differentiation of fish species. Lebensm.-Wiss. U.-Technol. 33:144-150.
- Woods, I.G., Wilson, C., Friedlander, B., Chang, P., Reyes, D.K., Nix, R., Kelly, P.D., Chu, F., Postlethwait, J.H., Talbot, W.S., 2005. The zebrafish gene map defines ancestral vertebrate chromosomes. Genome Res. 15:1307-1314.
- Woram, R.A., McGowan, C., Stout, J.A., Gharbi, K., Ferguson, M.M., Hoyheim, B., Davidson, E.A., Davidson, W.S., Rexroad, C., Danzmann, R.G., 2004. A genetic linkage map for Artic char (*Salvelinus alpinus*): evidence for higher recombination rates and segregation distortion in hybrid versus pure strain mapping parents. Genome 47:304-315.
- Wuertz, S., Belay, M., Kirschbaum, F., 2007. On the risk of criminal manipulation in caviar trade by intended contamination of caviar with PCR products. Aquaculture 269:130-134.
- Xu, Y.X., Zhu, Z.Y., Lo, L.C., Wang, C.M., Lin, G., Feng, F., Yue, G.H., 2006. Characterization of two parvalbumin genes and their association with growth traits in Asian seabass (*Lates calcarifer*). Anim. Genet. 37:266-268.
- Yin, Z.X., He, W., Chen, W.J., Yan, J.H., Yang, J.N., Chan, S.M., He, J.G., 2006. Cloning, expression

and antimicrobial activity of an antimicrobial peptide, epinecidin-1, from the orange-spotted grouper, *Epinephelus coioides*. Aquaculture 253:204-211.

- Young, W.P., Wheeler, P.A., Coryell, V.H., Keim, P., Thorgaard, G.H., 1998. A detailed linkage map of rainbow trout produced using doubled haploids. Genetics 148:839-850.
- Yu, Z., Guo, X., 2006. Identification and mapping of disease-resistance QTLs in the eastern oyster, *Crassostrea virginica* Gmelin. Aquaculture 254:160-170.
- Zhu, Z.Y, Yue, G.H., 2008. The complete mitochondrial genome of red grouper *Plectropomus leopardus* and its applications in identification of grouper species. Aquaculture 276:44-49.