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# UNIVERSITÀ DEGLI STUDI DI TORINO

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***This is an author version of the contribution published on:***

*Questa è la versione dell'autore dell'opera:*

*[Spadaro D., Garibaldi A., Gullino M.L. (2008) – Occurrence of patulin and its dietary intake through pear, peach and apricot juices in Italy. Food Additives and Contaminants: Part B, 1 (2), 134-139, DOI: 10.1080/02652030802363790]*

***The definitive version is available at:***

*La versione definitiva è disponibile alla URL:*

*[<http://www.tandfonline.com/doi/full/10.1080/02652030802363790#.U4sjqCjm6eA>]*

15 **Occurrence of patulin and its dietary intake through pear, peach and apricot juices in Italy**

16

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25

26 **Abstract**

27 One hundred and twenty five pear, peach, apricot and mixed juices were bought in Italian  
28 supermarkets and organic food shops and analysed for the patulin content. An overall incidence of  
29 34.4% was observed in the fruit juices, with a mean contamination level of 3.6  $\mu\text{g kg}^{-1}$ . No one  
30 sample trespassed 50  $\mu\text{g kg}^{-1}$ , the maximum permitted threshold according to the European  
31 legislation. Anyway, 19 positive samples contained more than 10  $\mu\text{g kg}^{-1}$  of patulin, the maximum  
32 level admitted in baby foods. The incidence of patulin contamination was significantly higher in  
33 pear juices (64.1%) than in apricot, peach or mixed juices. Statistical analysis ( $p = 0.002$ , Kruskal-  
34 Wallis test) showed a significantly higher level of patulin in pear (5.1  $\mu\text{g kg}^{-1}$ ) and mixed juices  
35 (4.9  $\mu\text{g kg}^{-1}$ ) than in the other ones. A slightly higher incidence of positive samples was found in  
36 conventional juices (35.7%) compared to the organic ones (29.6%). The magnitude between the  
37 mean contamination levels was similar in conventional (3.6  $\mu\text{g kg}^{-1}$ ) and organic (3.3  $\mu\text{g kg}^{-1}$ ) juices  
38 ( $p = 0.474$ , Mann-Whitney test). Samples were further divided based on the fruit content in fruit  
39 juices, fruit nectars and fruit drinks. In fruit nectars the incidence of positive samples (37.8%) was  
40 slightly higher than in fruit drinks (27.2%), but the mean patulin contamination was higher in fruit  
41 drinks (5.2  $\mu\text{g kg}^{-1}$ ) than in fruit nectars (3.4  $\mu\text{g kg}^{-1}$ ). Anyway, the magnitude between the two  
42 means was not statistically different ( $p = 0.734$ ; Mann-Whitney test). Italian juices had a higher  
43 incidence of patulin (35.3%), with a lower mean content (3.5  $\mu\text{g kg}^{-1}$ ), compared to the incidence  
44 (22.2%) and level (4.1  $\mu\text{g kg}^{-1}$ ) of foreign juices commercialized in Italy. The magnitude of the  
45 means between Italian and foreign juices was not statistically different, according to Mann-Whitney  
46 test ( $p = 0.616$ ). The estimated intake of patulin ranged from 0.1 to 1.5  $\text{ng kg}^{-1}$  bw for the whole

47 population and from 0.3 to 5.1 ng kg<sup>-1</sup> bw for the consumers only. The highest patulin intake was  
48 estimated for consumers of pear juices, followed by consumers of mixed juices. The daily intake of  
49 patulin deriving by the consumption of pear juices suggests that also pear juices, though a minor  
50 patulin source, could be monitored for their patulin content in order to control the mycotoxin  
51 contamination, especially in countries, such as Italy, where pear nectars are preferred as fruit drinks.

52  
53 **Key words:** fruit juices, HPLC, mycotoxin, *Penicillium expansum*, risk assessment

## 54 55 **Introduction**

56  
57 Patulin (4-hydroxy-4H-furo [3,2c] pyran-2[6H]-one) is a mycotoxin produced by a large group of  
58 fungal species, belonging to over 30 genera, including *Penicillium* spp., *Aspergillus* spp. and  
59 *Byssochlamys* spp. (Weidenböner 2001). The most important producer of patulin is *Penicillium*  
60 *expansum* that is able to attack most fruit both in orchard and in warehouse, causing a disease  
61 known as blue mould. The pathogen is usually associated with other microorganisms that colonize  
62 the fruit first causing lesions used by *P.expansum* to begin the growth (Snowdon 2007).  
63 *P.expansum* is an ubiquitous organism, that can survive to many adverse conditions and  
64 contaminate different foodstuffs (Moake et al. 2005). Almost all *P.expansum* strains are patulin  
65 producers (Andersen et al. 2004).

66 In animal studies, patulin has been found to damage the gastrointestinal and respiratory systems  
67 (Hayes et al. 1979; Mahfoud et al. 2002), DNA and many enzymes (Cooray et al. 1982; Wichmann  
68 et al. 2002), so that a provisional maximum tolerable daily intake (PMTDI) of 0.4 µg kg<sup>-1</sup> body  
69 weight (bw) has been set (JEFCA 1995). Based on this PMTDI, patulin is regulated in the European  
70 Union (EU) at levels of 50 µg kg<sup>-1</sup> in fruit juices and fruit nectars, 25 µg kg<sup>-1</sup> in solid apple products  
71 and 10 µg kg<sup>-1</sup> in apple-based products for infants and young children (European Commission  
72 2006).

73 A study carried out by the Directorate General – Health and Consumer Protection (2002) of the EU  
74 on the assessment of the dietary intake of patulin by the population of EU member states showed  
75 that apple juice and apple nectar are the main sources of patulin intake in most countries,  
76 particularly for young children. However, typical Italian fruits such as peaches, pears and apricots  
77 lead the nectar sector in Italy (Zenith International 2004) and fruit nectars are mainly used as  
78 children and kids snacks (Muraca 2000). Although most of the researches about the contamination  
79 level of patulin focused on apple and apple products (Moake et al. 2005), other fruit can be heavily  
80 damaged by blue mould. Some studies, carried out by using LC/UV or LC/MS, reported the

81 presence of patulin in commercial fruit juices coming from blueberry (Piemontese et al. 2005, 1  
82 sample with 13.5  $\mu\text{g kg}^{-1}$ ), citrus (Majerus and Kapp 2002; 2 samples, mean 25  $\mu\text{g kg}^{-1}$ ), grape  
83 (Rychlik and Schieberle 1999; 2 samples with 4.9 and 5.2  $\mu\text{g kg}^{-1}$ ; Majerus and Kapp 2002; 324  
84 samples, 39% positive, range 3-750  $\mu\text{g kg}^{-1}$ ), and pear (Majerus and Kapp 2002, 100 samples, 17%  
85 positive, mean 5.8  $\mu\text{g kg}^{-1}$ , range 0.03-91  $\mu\text{g kg}^{-1}$ ; Piemontese et al. 2005, 15 samples, 33%  
86 positive, mean 6.2  $\mu\text{g kg}^{-1}$ , max 61.0  $\mu\text{g kg}^{-1}$ ). Few data about the contamination of peach and  
87 apricots juices are available, although Buchanan et al. (1974) and Frank et al. (1977) showed the  
88 contamination by patulin produced by *P. expansum* on pears, peaches and apricots.

89 The aim of this work was to evaluate the occurrence and level of patulin in peach, apricot and pear  
90 juices. Moreover, different fruit juices containing pear, peach and/or apricot juices in different  
91 concentrations were considered (mixed juices). A second aim of the work, was the investigation of  
92 the possible influence of the agricultural production process employed (conventional or organic), of  
93 the fruit product typology (fruit juices, fruit nectars, or fruit drinks) on the occurrence and level of  
94 patulin contamination. Finally, the exposure to patulin in the Italian population and, particularly, in  
95 the consumers of fruit juices was assessed.

96

## 97 **Materials and methods**

98

### 99 *Sampling*

100 One hundred and twenty five fruit juices were bought in supermarkets and organic food shops of  
101 north-western Italy during the period January-September 2007. The products analyzed included  
102 most of the national and imported pear, peach, apricot and mixed fruit drink products sold in Italy.  
103 They were classified based on the agricultural production practices used (conventional/organic), the  
104 origin (Italian/foreign), and the fruit content (fruit juices/fruit nectars/fruit drinks). According to the  
105 European legislation (European Council 2001), fruit juices should have 100% fruit content, while  
106 pear, peach, and apricot nectars should have respectively at least 50%, 45%, and 40% fruit content.  
107 The other fruit based products are called fruit drinks.

108

### 109 *Sample preparation and analysis*

110 The samples were analyzed according to the method described by Spadaro et al. (2007) with slight  
111 variations. Cloudy products were left overnight in contact with pectinase enzyme solution (Sigma  
112 Chemical Co.<sup>®</sup>, St Louis, MO, USA; 5U/g of juice), and then centrifuged at 4,500 rpm for 5 min.  
113 Thirty grams of clarified juice were collected and liquid-liquid extracted with ethyl acetate (Sigma  
114 Chemical Co.<sup>®</sup>). The procedure was carried out three times. The organic phase was dehydrated with

115 15 g of sodium sulphate anhydrous (Merck<sup>®</sup>, Darmstadt, Germany) and then evaporated to dryness  
116 (Rotavapor Laborota 4000, Heidolph<sup>®</sup>, Schwalback, Germany). The clean up was performed using  
117 C<sub>18</sub> SPE column (200 mg, 6 ml, J.T. Baker<sup>®</sup>, Phillipsburg, NJ, USA), previously activated with 10  
118 ml of toluene (Sigma Chemical Co.<sup>®</sup>). The columns were cleaned up with 4 ml of toluene and, for  
119 elution, 4 ml of toluene and ethyl acetate solution (1:1) were used. The samples were collected in 4  
120 ml vials and completely dried (Reack-Therm III, Pierce<sup>®</sup>, Rockford, IL, USA). Samples were  
121 resuspended with 1.5 ml of acetic acid solution (4.35 mM, pH 4.0) and transferred into HPLC vials.  
122 The HPLC apparatus was an Agilent<sup>®</sup> 1100 series equipped with G 1379 degasser, G 1313A  
123 autosampler, G1316A column thermostat set at 30 °C, G 1315B UV diode array detector set at 276  
124 nm, G1311 quaternary pump and Agilent Chemstation G2170AA Windows XP operating system  
125 (Agilent<sup>®</sup>, Waldbronn, Germany). A stainless steel analytical column (250 x 4.6mm i.d., 4 µm,  
126 Synergy Hydro-RP C18; Phenomenex<sup>®</sup>, Torrance, CA, USA) preceded by a guard column (4 x 3  
127 mm i.d.) with the same stationary phase was used. The mobile phase, eluting at a flow rate of  
128 1ml/min, consisted of an isocratic mixture of water – acetonitrile – perchloric acid (96:4:0.1) for 16  
129 min, followed by a washing step with an isocratic mixture of water–acetonitrile (35:65). Hundred  
130 microliters of sample were injected onto the HPLC column and the retention time of patulin was  
131 11.82 min. The amount of patulin in the final solution was determined by using a calibration graph  
132 of concentration versus peak area and expressed as ng/ml, achieved by injection onto the HPLC  
133 column of 100 µl of standard solutions of patulin (Sigma Chemical Co. <sup>®</sup>) prepared according to the  
134 method described by Arranz et al. (2004). The standard solutions had concentrations of 500 ng ml<sup>-1</sup>,  
135 400 ng ml<sup>-1</sup>, 250 ng ml<sup>-1</sup>, 100 ng ml<sup>-1</sup> and 50 ng ml<sup>-1</sup> of patulin and were prepared the same day of  
136 HPLC analysis, in order to avoid patulin degradation.

137 The recovery was determined on a blank fruit juice spiked at three concentrations of patulin (8, 30  
138 and 50 µg kg<sup>-1</sup>). Each test was performed three times and the mean recovery values were  
139 respectively 85.2, 89.5 and 88.9%. The repeatability ranged from 4.5 to 8.3% for triplicate analyses.  
140 The limit of detection (LOD) and the limit of quantification (LOQ) based on the IUPAC definition  
141 (Thompson et al., 2002) were respectively 1.2 and 1.7 µg kg<sup>-1</sup>. The high value of the regression  
142 coefficient ( $R^2 \geq 0.99$ ) obtained indicated a good linearity of the analytical response.

143

#### 144 *Statistical analysis*

145 Samples with a concentration of patulin higher than the LOD were considered positive, whereas  
146 samples with concentrations lower than LOD were considered negative. Mean patulin  
147 concentrations were calculated by using 0 for negative samples. Experimental results are reported as  
148 mean ± standard deviation and maximum. The Kruskal-Wallis test was used to compare the mean

149 patulin levels in apricot / peach / pear / mixed juices, and in fruit juices / fruit nectars / fruit drinks,  
150 while the Mann-Whitney test was used to compare the mean patulin levels in conventional / organic  
151 juices, and in Italian / foreign juices, using the null hypothesis that the levels were not different. The  
152  $\chi^2$  test was used to compare the patulin contamination frequencies of the different categories of fruit  
153 juiciness. Statistical analyses were performed by using the programme SPSS Release 12.01 (2003).

154

## 155 **Results and discussion**

156 Out of 125 samples analyzed, 43 having a patulin concentration higher than the LOD were  
157 considered positive (Table 1), ranging from 1.2 to 33.4  $\mu\text{g kg}^{-1}$ . An overall incidence of 32.0% was  
158 observed in the fruit juices, with 24 samples having less than 10  $\mu\text{g kg}^{-1}$  and 19 samples having  
159 more than 10  $\mu\text{g kg}^{-1}$ . Positive samples were further divided as having more or less than 10  $\mu\text{g kg}^{-1}$ ,  
160 the threshold limit established for baby foods for infants and young children (European  
161 Commission 2006). Anyway no one sample trespassed 50  $\mu\text{g kg}^{-1}$ , the maximum permitted  
162 threshold according to the European legislation. A mean contamination level of 3.6  $\mu\text{g kg}^{-1}$  was  
163 found in the samples analyzed.

164 The levels and incidence of patulin contamination have been largely assessed in apples and apple-  
165 derived products (Moake et al. 2005). This study aimed at assessing the contamination level of  
166 patulin in fruit juices containing apricot, peach, pear, or one or more of the previously named fruits  
167 mixed with other fruits. To our knowledge, this study represents the most extensive investigation  
168 performed on such fruit juice typology. Looking at the fruit species content, the incidence of patulin  
169 contamination was significantly higher in pear juices (64.1%) than in apricot, peach or mixed  
170 juices. Statistical analysis ( $p = 0.002$ , Kruskal-Wallis test) showed a significantly higher level of  
171 patulin in pear (5.1  $\mu\text{g kg}^{-1}$ ) and mixed juices (4.9  $\mu\text{g kg}^{-1}$ ) than in the other ones.

172 *P. expansum* can cause significant losses in pear fruits (Snowdon 2007). By considering that most  
173 of this pathogen isolates are able to produce patulin (Andersen et al. 2004), patulin can be easily  
174 found in pears and pear derived products. Moreover, patulin is stable in pH ranges of 3.3-6.3  
175 (Jefferys 1952) and in substrates with low levels of sulfhydryl compounds (Scott and Somers 1968),  
176 both features are typical of pears. Based on studies carried out by Laidou et al. (2001), removal of  
177 blue mould rot of pears cannot protect consumers from the presence of patulin, because up to 25%  
178 of the mycotoxin can be detected in the sound tissues around the diseased ones.

179 The mixed juices analyzed in this study represent the second source of contamination of patulin,  
180 based on the incidence and level of the mycotoxin. This could be explained either with the lower  
181 care deserved to the production of mixed juices, often containing higher quantities of sugars and

182 other additives, or with the general presence of a certain percentage of apple juice, the major source  
183 of patulin, together with other fruit juices.

184 In Table 2, the fruit juices analyzed were divided based on the agricultural production process  
185 employed (conventional or organic), on the fruit content (fruit juices, fruit nectars, or fruit drinks),  
186 and on the origin (Italian or foreign juices). As already found by analyzing pure and mixed apple  
187 juices (Spadaro et al. 2007), a slightly higher incidence of positive samples was found in  
188 conventional juices (35.7%) compared to the organic ones (29.6%). The  $\chi^2$ -test showed that the  
189 frequencies of patulin occurrence in conventional and organic juices were comparable ( $p = 0.483$ ).  
190 Also the magnitude between the mean contamination levels was similar in conventional ( $3.6 \mu\text{g kg}^{-1}$ )  
191 and organic juices ( $3.3 \mu\text{g kg}^{-1}$ ). The hypothesis that the mean patulin contamination levels in  
192 conventional and organic juices were not different was accepted ( $p = 0.692$ , Mann-Whitney test). It  
193 should be noticed anyway that organic juices contain, on an average, higher fruit content (54%)  
194 than conventional ones (45%), being more exposed to patulin contamination. Generally sound fruit,  
195 which is properly picked and stored, is less subjected to blue mould rot and, consequently, to  
196 contain patulin (Sydenham et al. 1995). The absence of significant differences, based on the  
197 agricultural production methodologies used, can be explained with the fact that the presence of  
198 patulin is a strong indicator of poor agricultural practice, either conventional or organic (Codex  
199 Alimentarius 2003), although *P. expansum* mainly develops after harvesting, where the main  
200 discriminating factor becomes the removal of decayed and damaged fruits during juice processing.  
201 Samples were further divided based on the fruit content: fruit juices should have 100% fruit content,  
202 while pear, peach, and apricot nectars should have respectively at least 50%, 45%, and 40% fruit  
203 content. The fruit based products with lower fruit content are called fruit drinks. Statistical data  
204 (Russo et al. 2002) indicate that national per capita consumption of nectars equals approximately to  
205 6 liters, while 100% fruit juice consumption is lower (4 liters per capita). Per capita consumption of  
206 fruit drinks – with a lower fruit content and supplemented with vitamins and other nutritious  
207 ingredients – is approximately 4 liters per person. In the 5 samples containing 100% fruit, patulin  
208 was not found. In fruit nectars the incidence of positive samples (37.8%) was slightly higher than in  
209 fruit drinks (27.2%), but the mean patulin contamination was higher in fruit drinks ( $5.2 \mu\text{g kg}^{-1}$ )  
210 than in fruit nectars ( $3.4 \mu\text{g kg}^{-1}$ ). Anyway, the magnitude between the two means was not  
211 statistically different ( $p = 0.734$ ; Mann-Whitney test). As already hypothesized for the mixed juices,  
212 and most of the fruit drinks are mixed fruit drinks, the higher patulin content could be explained by  
213 the lower attention used in the production process of such products, where the lower quality of the  
214 fruits can be easily corrected by adding sugars and additives. Since fruit drinks are often produced



215 by using raw materials poor in quality, the probability to retrieve patulin and others similar  
216 contaminants in fruit drinks is increased (Moake et al. 2005).

217 Finally the samples were divided by considering their origin. Italian juices had a higher incidence of  
218 patulin (35.3%), with a lower mean content ( $3.5 \mu\text{g kg}^{-1}$ ), compared to the incidence (22.2%) and  
219 level ( $4.1 \mu\text{g kg}^{-1}$ ) of foreign juices commercialized in Italy. The magnitude of the means between  
220 Italian and foreign juices was not statistically different, according to Mann-Whitney test ( $p =$   
221  $0.616$ ). Anyway it should be noticed that it was difficult to find pear, peach and apricot foreign  
222 juices commercialized in Italy (only 9 samples out of 125), because pears, peaches and apricots are  
223 typical products of the Italian horticulture.

224 The estimated intake of patulin in the total population and, particularly, in the consumers was  
225 calculated based on the patulin contamination in the pear, apricot, peach and mixed fruit juices  
226 analyzed. Apple juices are the most consumed fruit juices around the world. However, in Italy apple  
227 juice is not so popular as in other Countries. Apricot, peach and pear juices are highly consumed by  
228 a large share of the Italian population, and they are used as baby-foods (Zenith International 2004).  
229 By considering (Turrini et al. 2001) that the mean daily consumption of fruit juices in Italy is 21.0  
230 g, and that only 29.8% of the population consume fruit juices ( $70.5 \text{ g day}^{-1}$ ), the intake of patulin  
231 was calculated in four different scenarios, where the fruit juice consumed was only apricot, or pear,  
232 or peach, or mixed juice (Table 1). The daily intake of patulin ranged from 0.1 to  $1.5 \text{ ng kg}^{-1} \text{ bw}$  for  
233 the whole population and from 0.3 to  $5.1 \text{ ng kg}^{-1} \text{ bw}$  for the consumers only. The highest patulin  
234 intake was estimated for consumers of pear juices, followed by consumers of mixed juices. All of  
235 the values are from two to three magnitude orders lower than the PMTDI recommended by JEFCA  
236 (1995) which is  $400 \text{ ng kg}^{-1} \text{ bw day}^{-1}$ . The intake of patulin from apricot, pear, peach and mixed  
237 juices could be considered similar to the values found by Piemontese et al. (2005) that anyway  
238 showed a higher intake of patulin by consuming organic than conventional fruit juices.

239 In conclusion, the average level of contamination of apricot, peach, pear and mixed juices analyzed  
240 in this work appears to be generally low, by considering that none of the samples exceeded the  
241 threshold of  $50 \mu\text{g kg}^{-1}$  imposed by the European regulations. Anyway, the incidence of patulin  
242 found in pear juices showed that also pears – besides apples – can be substrates for patulin  
243 contamination. The daily intake of patulin deriving by the consumption of pear juices suggests that  
244 also pear juices, though a minor patulin source, could be monitored for their patulin content in order  
245 to control the mycotoxin contamination, especially in countries, such as Italy, where pear nectars  
246 are preferred as fruit drinks.

247

248 **Acknowledgements**

249 This research was carried out with a grant from the Italian Ministry for the Environment, Land and  
250 Sea within the Framework Agreement “Sustainable Agriculture” and with a grant from Piedmont  
251 Region “STONE SAFE - Production of stone fruits in Piedmont: monitoring, prevention and  
252 control of pathogenic and mycotoxigenic fungi to guarantee food safety”. The authors acknowledge  
253 Dr. Sandro Frati and Dr. Fabio Fanella for helping in the experimental trials.

254

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329 Table 1 - Patulin contamination in apricot, pear, peach and mixed juices marketed in Italy.  
 330

	Mean fruit concentration (%)	Positive (%)	Number of samples			Mean <sup>a</sup> (µg kg <sup>-1</sup> )
			Positive/total	<10 µg kg <sup>-1</sup>	>10 µg kg <sup>-1</sup>	
Apricot juices	41	25.9	7/27	2	5	3.6 ±
Pear juices	54	64.1	25/39	17	8	5.1 ±
Peach juices	48	6.7	2/30	2	0	0.3 ±
Mixed juices	43	31.0	9/29	3	6	4.9 ±
<b>TOTAL JUICES</b>	<b>47</b>	<b>34.4</b>	<b>43/125</b>	<b>24</b>	<b>19</b>	<b>3.6 ±</b>

331  
 332 <sup>a</sup>Mean level was calculated using 0 for samples lower than LOD.

333

334 Table 2 – Patulin contamination in different categories of apricot, peach, pear and mixed juices  
 335 marketed in Italy.  
 336

	Mean fruit concentration (%)	Positive (%)	Number of samples		
			Positive/total	<10 µg kg <sup>-1</sup>	>10 µg kg <sup>-1</sup>
Conventional juices	45	35.7	35/98	21	14
Organic juices	54	29.6	8/27	3	5
Fruit juices	100	0.0	0/5	0	0
Fruit nectars	49	37.8	37/98	23	14
Fruit drinks	28	27.2	6/22	1	5
Italian juices	48	35.3	41/116	24	17
Foreign juices	39	22.2	2/9	0	2

337 <sup>a</sup>Mean level was calculated using 0 for samples lower than LOD.  
 338

339 Table 3 – Estimated intake of patulin in the total population and in the consumers only calculated  
340 for pear, apricot, peach and mixed fruit juices.  
341

	Mean Consumption (g day <sup>-1</sup> ) <sup>a</sup>	Intake of patulin (ng kg <sup>-1</sup> bw <sup>b</sup> day <sup>-1</sup> )			
		From apricot products	From pear products	From peach products	From mixed juices
Population	21.0	1.1	1.5	0.1	1.5
Consumers only	70.5	3.6	5.1	0.3	4.9

342 <sup>a</sup> from Turrini et al. 2001: consumers represent 29.8% of the population; <sup>b</sup> bw=body weight; <sup>c</sup> mean  
343 patulin concentration used in the calculations were 3.6 µg kg<sup>-1</sup> for apricot juices, 5.1 µg kg<sup>-1</sup> for  
344 pear juices, 0.3 µg kg<sup>-1</sup> for peach juices and 4.9 µg kg<sup>-1</sup> for mixed juices.  
345