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UNIVERSITÀ DEGLI STUDI DI TORINO

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1 **Host taxon-derived Sarcoptes mite in European wild animals**

2
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15
16 **Abstract** Microsatellite genotyping was applied on individual *Sarcoptes* mites from 15
17 wild mammalian populations belonging to 10 host species in three European countries,
18 using 10 *Sarcoptes* mite specific markers. The results showed that the geographical
19 separations had real biological significance for the definition of mite sub-populations,
20 and that the degree of genetic exchange occurring between mites from different
21 localities was related to the geographical distance between locations. Wild host-derived
22 mite populations were clustered into three main groups: herbivore-, carnivore- and
23 omnivore-derived *Sarcoptes* mite populations. Omnivore-derived was halfway between
24 herbivore- and carnivore-derived *Sarcoptes* mite populations. The separation between
25 the three mite groups (herbivore-, carnivore- and omnivore-derived mite) was more

26 supported than that by the geographical separations; nevertheless a kind of sub-
27 clustering was detected within each group (carnivore-, omnivore- and herbivore-),
28 scattering mite populations up to their geographical localities (countries). The lack of
29 gene flow between *Sarcoptes* populations may have improved parasitic adaptations and
30 led to, what we called host taxon-derived (carnivore host-, herbivore host- and omnivore
31 host-derived) *Sarcoptes* mite populations in European wild animals. Our results
32 demonstrated that *Sarcoptes* is not a single panmictic population even within locations,
33 which will have important ramifications on the study of population genetic structure,
34 life cycle, diagnosis and monitoring protocols, and could contribute to the better
35 understanding of the epidemiology of the ubiquitous *Sarcoptes* mite.

36

37 **Keywords:** *Sarcoptes scabiei*; Genetic epidemiology; Genetic structure; Microsatellite
38 markers; Omnivore-derived; Carnivore-derived; Herbivore-derived; Host taxon-derived.

39 **Introduction**

40

41 Predicting the spread of a disease to wildlife is critical in order to identify populations at
42 risk, target surveillance, and design proactive management programmes (Blanchong et
43 al. 2008). Recently, there has been an increased interest in diseases of free-living
44 animals and the ecological role of diseases in populations, particularly their ability to
45 regulate animal abundance (Scott 1988; Lyles & Dobson 1993; Robinson 1996; Daszak
46 et al. 2000). There is also an awareness that free-living species may act as reservoirs of
47 diseases of man and domestic animals (Robinson 1996; Daszak et al. 2000).

48 Although neglected as a pathogen, the ectoparasite *Sarcoptes scabiei* continues
49 to affect humans and a wide range of mammalian hosts on a worldwide scale (Bornstein
50 et al. 2001; Pence & Ueckermann 2002; Walton et al. 2004a). The introduction of
51 infected domestic animals and the succeeding adaptation of *Sarcoptes* mite to a new
52 highly susceptible and receptive wild host have been proposed to give rise to sarcoptic
53 mange epizootics in previously mange-free wildlife populations (Arlian 1989).

54 In several European wild mammal populations, *Sarcoptes* mite infections are
55 endemic and cause devastating mortality as reported especially for Alpine and Pyrenean
56 chamois, Iberian ibex, aoudad, and red fox (Fandos 1991; Mörner 1992; Pérez et al.
57 1997; León-Vizcaino et al. 1999; González-Candela et al. 2004; Rossi et al. 2007).
58 Notwithstanding, only few cases have been reported in other sympatric host species,
59 like stone marten, badger, lynx, and roe deer (Ryser-Degiorgis et al. 2002; Oleaga et al.
60 2008).

61 Morphological studies have failed to identify any significant differences among
62 mite populations (Fain 1978), and experimental cross contamination of *Sarcoptes* mite
63 between hosts of different species is commonly unsuccessful (Arlian et al. 1984; Arlian

64 1989). Apparently no epidemiological relationship exists, in Europe, between mange
65 foci affecting wild ruminants, wild boars, and carnivores (Berrilli et al. 2002).

66 The question as to whether *Sarcoptes* mites might be divided into different
67 species or whether they are, in fact, monospecific has been the subject of an ongoing
68 debate (Zahler et al. 1999; Burgess 1999; Berrilli et al. 2002; Gu & Yang 2008; Alasaad
69 et al. 2009c). Zahler et al. (1999) and Berrilli et al. (2002), using the ITS-2 sequences as
70 genetic markers, did not detect clear-cut evidence of genetic separation related to host
71 species or geographic location. As well as, in our previous study, we have shown that
72 ITS-2 rDNA does not appear to be suitable marker for examining genetic diversity
73 among *Sarcoptes* mite populations from different wild host species and/or geographical
74 localities (Alasaad et al. 2009c). In phylogenetic analyses bootstrapping support for the
75 closest relationships may be relatively poor due to reduced time to accumulate
76 informative changes in the sequences examined. Further resolution is therefore provided
77 in faster evolving hypervariable sequences such as nuclear polymorphic microsatellite
78 loci (Walton et al. 2004b).

79 Walton et al. (1999; 2004b), using multi-locus genotyping applied to
80 microsatellite markers, substantiated previous data that gene flow between scabies mite
81 populations on human and dog hosts is extremely rare in northern Australia. As well as,
82 genetic differences were detected between geographically distinct populations, even
83 between householders. Microsatellite markers were used by Alasaad et al. (2008b) to
84 describe a new phenomenon of genetic structuring among *S. scabiei* at individual host
85 skin-scale.

86 Taking into account all the above-mentioned information, the aim of the present
87 study was to test the extent of genetic relationship between sympatric wild host-derived
88 *Sarcoptes* mite populations, and to study the influence of the geographical isolation on

89 the genetic structuring of *Sarcoptes*. This is pivotal for wildlife health management in
90 order to understand the geographic variation among bordered mite populations, and to
91 measure the patterns of host specific differences, especially in sympatric hosts.

92

93 **Materials and methods**

94

95 Collection of *S. scabiei*

96

97 Using Postponed Isolation (Post-frozen Isolation) and Direct Isolation (Live Isolation)
98 methods, as described by Alasaad et al. (2009b), 251 *Sarcoptes* mites were collected
99 from the crusted skin of 100 animals belonged to 15 populations of 10 European wild
100 mammalian species, as listed in Table 1, which were sampled in Italy, France and Spain
101 (Fig. 1). *Rupicapra rupicapra rupicapra*, *Cervus elaphus*, *Martes martes*, *Ovis*
102 *musimon*, *Capra ibex*, and *Vulpes vulpes* were sympatric in Northeast Italian Alps. *V.*
103 *vulpes*, *Martes foina* and *Sus scrofa* were sympatric in Northwest Italian Alps. Taking
104 into account the topography of Sierra Nevada Mountain and that the first case of
105 *Sarcoptes* mite infection was reported in Dílar Valle (East Sierra Nevada) (Pérez et al.
106 1997), mites from Sierra Nevada were divided into two different groups, East and West
107 populations. All mites were identified as *S. scabiei* based on known morphological
108 criteria (Fain 1968).

109 In Table 1, term ‘Code’ refers to all mites belonging to the same geographical
110 and/or host species-derived population, from now onwards called ‘component
111 population’ or, simply, population (Bush et al. 1997).

112

113 Preparation of *Sarcoptes* gDNA

114

115 The DNA of individual Sarcoptes mites was extracted with the NucleoSpin Tissue kit
116 procedure (Macherey-Nagel, Düren, Germany) with some modifications proposed by
117 Soglia et al. (2009), and recently with HotSHOT Plus ThermalSHOCK technique
118 (Alasaad et al. 2008a).

119

120 Fluorescent-based polymerase chain reaction analysis of microsatellite DNA

121

122 From the panel described by Walton et al. (1997), ten microsatellites (Sarms 33-38, 40,
123 41, 44, and 45) were selected and analysed with one 10× multiplex PCR. Each 15 µl
124 PCR reaction mixture consisted of 3 µl of the single mite DNA, together with the PCR
125 mixture containing all primer pairs (ranged from 0.04 to 0.1 µM per primer), 200 µM of
126 each dATP, dCTP, dGTP, and dTTP, 1.5 µl of 10× PCR buffer (200 mM KCl and 100
127 mM Tris-HCl, pH 8.0), 1.5 mM MgCl₂, and 0.15 µl (0.5 U/reaction) HotStartar Taq
128 (QIAGEN, Milano, Italy). Samples were subjected to the following thermal profile for
129 amplification in a 2720 thermal cycler (Applied Biosystems, Foster City, CA, USA): 15
130 min at 95°C (initial denaturing), followed by 37 cycles of three steps of 30 s at 94°C
131 (denaturation), 45 s at 55°C (annealing) and 1.5 min at 72°C (extension), and a final
132 elongation of 7 min at 72°C.

133

134 Microsatellite analysis

135

136 Using 96-well plates, aliquots of 12 µL of formamide with Size Standard 500 Liz
137 (Applied Biosystems, Foster City, CA, USA) and 2 µl PCR product were prepared.
138 Then, the plates were heated for 2 min at 95°C and chilled to 4°C. Fluorescent PCR

139 amplification products were analyzed by ABI PRISM 310 Genetic Analyzer with pop4.
140 Allele calling was performed using the GeneMapper v. 4.0 software (Applied
141 Biosystems, Foster City, CA, USA). To track and minimize the amount of error
142 associated with genotyping, the genetic data were collected twice by SA and DS.

143

144 Descriptive statistics and cluster analysis

145

146 CONVERT 1.31 software (Glaubitz 2004) was used to reformat files for the statistical
147 softwares. Descriptive statistics and diversity analyses were carried out with GenAlEx
148 v. 6.2 (Peakall & Smouse 2006), Genepop v. 4.0 (Raymond & Rousset 1995), Fstat v.
149 2.9.3 (Goudet 1995), and Arlequin v. 3.1 (Excoffier et al. 2005) softwares, i.e. allelic
150 richness (R), number of private alleles, allele frequencies, unbiased expected (H_e) and
151 observed (H_o) heterozygosity, test for Hardy-Weinberg equilibrium (HWE), test for
152 linkage equilibrium (LE), and F statistics. All pairs of the component populations were
153 compared for homogeneity of genetic variation using the Wilcoxon's matched-pairs
154 signed-rank test (GraphPad InStat software).

155 Analysis of structure and relationships among host-specific mite populations
156 were studied using two different approaches:

157 (i) Multilocus proportion of shared alleles (Dps) was computed between all possible
158 pairs of individual mites using the Microsat software (Minch 1997) ignoring any
159 preliminary information on origins of parasites. One thousand datasets were generated
160 by resampling the input data (bootstrapping), the Neighbor-Joining algorithm was used
161 as implemented by the Phylip v. 3.6 package (Felsenstein 1989), and a consensus
162 dendrogram was obtained. The dendrogram was visualized using the Dendroscope v.
163 2.2.2 software (Huson et al. 2007).

164 (ii) The analysis of relationships among mites was then improved by a Bayesian
165 assignment test using the method implemented by the Structure v. 2.2 software
166 (Pritchard et al. 2000). We performed 50000 MCMC (Markov chain Monte Carlo)
167 replicates following a burn-in period of 10000 steps. This parameter set was run 20
168 times for each of different numbers, K, of genetic clusters of multilocus genotypes; all
169 values of K from 1 to 20 were tested. The probability of the multilocus genotype of any
170 individual mite to occur in each of the K clusters was computed. We used the admixture
171 model (each mite drew some fraction of its multilocus genotype from each of the K
172 clusters) allowing the allele frequencies to be correlated among clusters. This
173 configuration has been considered the best in the case of subtle population structure
174 (Falush et al. 2003). We used the height of the modal value of the distribution of ΔK in
175 order to estimate the uppermost number of clusters capturing the overall mite sample
176 structure, as suggested by Evanno et al. (2005). We then associated any individual mite
177 with the cluster that corresponded to its greatest membership, q , that is fraction of its
178 multilocus genotype; a threshold value $q \geq 0.9$ was used. Finally, each of the inferred
179 clusters was associated with the component populations of its mites. If a cluster was
180 labelled with multiple mite populations, an additional substructure analysis for K values
181 from 1 to 5 was performed testing only the mites assigned to that cluster.

182

183 **Results**

184

185 Descriptive statistics

186 Ten marker loci were analysed on 251 mites belonging to 15 populations of 10
187 European wild mammalian species from Italy, France and Spain; 101 alleles were
188 detected. Allele count for each of the 10 loci ranged from six (Sarms 37) to 15 (Sarms

189 34). Proportion of missing genotypes was as low as 0.04 and it did not affect single
190 locus or population. Forty two private alleles were detected in 11 wild host-derived mite
191 populations, ranging from 1 (ItNWMf, SpNEVv and SpNWRp) to 10 (FrNESs).
192 Whereas in ItNECe, SpEMf, ItNEOam, and ItNEMm no private alleles were identified
193 (Table 2). The highest within-population genetic variability was observed for the two *S.*
194 *scrofa* mite populations, in spite of their small size, whereas little variation was found in
195 ItNWVv and SpNWRp (Table 3).

196 Allelic richness (R) and heterozygosity (H_e) were used as most informative
197 parameters of diversity. In particular, allelic richness provided a measure of the number
198 of alleles independent of sample size, hence allowing comparison among different
199 populations. Level of genetic diversity varied both across loci and among populations.
200 Wilcoxon's test stated ($P < 0.001$) that *C. ibex* mites showed more variability ($R = 2.6$,
201 $H_e = 0.339$) than mites from the other ruminants whereas *S. scrofa* mites were the most
202 variable at all ($R = 3.4$, $H_e = 0.667$).

203 LE test (Lewontin 1964; Slatkin 1994; Slatkin & Excoffier 1996) was
204 performed for all loci and significant linkage disequilibrium ($P < 0.05$) was observed for
205 18 pairs when all the mite populations were pooled. In no cases disequilibrium was
206 observed at the same loci in more than two populations individually analysed. HWE
207 estimates were assessed of 70 locus-by-population comparisons, 40 (57%) showed
208 significant heterozygosity deficiencies. Deviations from HWE did not point at any locus
209 in particular. All populations deviated from HWE across loci after sequential Bonferroni
210 correction ($P < 0.001$).

211 Population differentiation based on allele frequencies for all 15 populations gave
212 an overall $F_{st} = 0.721$. Each locus significantly ($P < 0.001$) contributed to distribution of
213 variability among populations with per-locus values ranging from 0.290 to 0.821. This

214 very high estimate means that most of the global *Sarcoptes* genetic variability resided
215 among rather than within component populations different in geographical and host-
216 derived distribution of mites.

217

218 Structure and relationships among mite populations

219

220 (i) Multilocus proportion of shared alleles (Dps) as a measure of genetic similarity
221 between all pairs of mites.

222

223 Genetic variability among populations of *Sarcoptes* mites collected from the same host
224 species from different localities.

225 The proportion of shared alleles between pairs of individual mites from the two *C.*
226 *pyrenaica* mite populations in Spain (SpSWCp and SpSECp) were scattered randomly
227 with no evidence of distribution based on geographical location of hosts. Individual
228 *Sarcoptes* mites belonging to the three *V. vulpes* mite populations from the Northeast
229 and Northwest Italian Alps as well as from Northeast Spain showed clear clustering up
230 to their original populations. *V. vulpes* mite population from Spain was the most
231 different one, supported by 980/1000 bootstraps. The *V. vulpes* mites from Northeast
232 and Northwest Italian Alps were relatively more similar between each other, their
233 distribution across two distinct clusters being very poorly supported (169/1000
234 bootstraps) (Fig. 2). Regarding mites from *S. scrofa* populations from Northwest Italian
235 Alps and from Northeast France, strong separation was detected between them
236 (1000/1000 bootstraps, data not shown).

237

238 Genetic variability among populations of *Sarcoptes* mites collected from sympatric host

239 species.

240 The dendrogram of individual mites from six sympatric host-derived populations from
241 Northeast Italian Alps, and three sympatric host-derived populations from Northwest
242 Italian Alps allowed the clustering of the mites into three groups (Fig. 3). The first
243 group was formed by all carnivore-derived mites from East and West Italian Alps,
244 mainly mites from *V. vulpes* and, in addition, *M. foina* (Figure 3, a) and *M. martes*
245 (Figure 3, b), nevertheless it included also a mite from *C. ibex* (Figure 3, c). The second
246 group included the herbivore-derived mite populations from Northeast Italian Alps,
247 mainly mites from *C. ibex* and *R. Rupicapra*, all scattered across the cluster. In addition,
248 the cluster included *O. aries musimon* (Figure 3, d) and *C. elaphus* (Figure 3, e) mites.
249 The separation of herbivore- and carnivore-derived mites was then quite clear-cut
250 (470/1000 bootstraps). The *S. scrofa* mites from Northwest Italian Alps were near the
251 carnivore-derived mites but distinct from them (572/1000 bootstraps).

252

253 Genetic variability among *Sarcoptes* populations distributed according to both host
254 species and geographical localities.

255 Five clusters resulted from the analysis of the overall 15 wild host-derived mite
256 populations (Fig. 4). Cluster I included almost all the herbivore-derived mites from
257 Spain (SpNWRp, SpSWCp and SpSECp) and the *C. ibex* mite which clustered with
258 carnivore-derived parasites in Figure 3 (c). Although some mites scattered around,
259 evidence of separation between SpNWRp and SpSWCp-SpSECp was observed.
260 Clusters IIa and IIb contained all the omnivore-derived mites, ItNWSs and FrNESs
261 respectively, and one ItNECi (included in cluster IIb, Figure 4, arrow). Cluster III
262 included all the carnivore-derived mites, i.e. ItNEMm, ItNWMf, ItNWWv, ItNEVv as
263 well as SpNEVv (Figure 4, a) and SpEMf (Figure 4, b). Cluster IV contained almost all

264 the herbivore-derived mites from Italy (ItNERr, ItNECi, ItNECe, and ItNEOam,) and
265 one SpNWRp.

266

267 (ii) Analysis of mite population structure by the Bayesian method.

268

269 The modal value of the statistic ΔK for the overall dataset (251 mites) stated that the
270 uppermost cluster value was $K = 4$ (Evanno et al. 2005). Each of the four inferred
271 clusters was then associated with the information of its mites. For each cluster the
272 average membership and number of mites assigned with the greatest membership were
273 computed (Fig. 1). Cluster I shared all mites with the cluster I in Fig. 4, i.e. nearly all
274 mites of Spanish ruminants (SpNWRp, SpSECp and SpSWCp) and one ItNECi. High
275 proportions of membership were always obtained ($q > 0.97$). Cluster II grouped all the
276 *S. scrofa* mites (ItNWSs and FrNESs) (see Fig. 4, clusters IIa and IIb) and their
277 membership fraction was $q > 0.98$. Two ItNECi (for one of them see Fig. 4, cluster IIb)
278 and three SpNEVv mites were also added but they lacked to show high membership in
279 this cluster ($q < 0.70$). Cluster III shared most mites with the cluster III in Fig. 4
280 grouping all mites of Italian *V. vulpes* (ItNEVv and ItNWVv), *M. martes* (ItNEMm), and
281 *M. foina* (ItNWMf and SpEMf) with very robust membership ($q > 0.95$ for all mites but
282 SpEMf parasite with $q = 0.86$). One SpNEVv mite showed some similarity to this cluster
283 although with poor membership ($q = 0.56$). Cluster IV grouped the majority of the mites
284 of Italian ruminants (ItNERr, ItNECi, ItNECe, and ItNEOam) and one SpNWRp with
285 very high membership ($q > 0.97$). Its members corresponded to those of the cluster IV
286 in Fig. 4.

287 The computation of the statistic ΔK was repeated separately for four subsets of
288 samples made up of the main geographical and host-specific mite groups, i.e. mites

289 belonging to *S. scrofa* (ItNWSs and FrNESs), Italian ruminants (ItNERr, ItNECi,
290 ItNECe, and ItNEOam), *V. vulpes* (ItNEVv, ItNwVv, and SpNEVv), and Spanish
291 ruminants (SpNWRp, SpNECp and SpNWCp). No evidence of substructure was
292 detected in any case.

293 Spanish *V. vulpes* mites received ambiguous assignment as in case of mixed
294 ancestry. However, they were collected from a single host animal and lacked a
295 substantial component population as a reference. Two ItNECi were misplaced in *S.*
296 *scrofa* mite cluster with low membership. One of them was also assigned to the
297 sympatric *V. vulpes* mite cluster (Fig. 3, c) or to the allopatric Spanish ruminant mite
298 cluster (Fig. 4, Cluster I) depending on the populations used for the comparison. Such
299 individual parasites seemed to be randomly assigned since they carried multilocus
300 genotypes infrequent in their population, so the algorithm could not recognize their
301 ancestry.

302 In synthesis, the proportion of shared alleles as a similarity measure among
303 mites and the assessment of structure using the Bayesian method provided patterns in
304 agreement with each other. When the full data set was used, four distinct genetic
305 clusters of mites were inferred, i.e. omnivore-, Italian herbivore-, carnivore-, and Spain
306 herbivore-derived parasites.

307

308 **Discussion**

309

310 Differentiation of host-specific mites using morphological traits, apart from being very
311 difficult and time consuming, proved to be impossible to implement when mites of the
312 same host-specific variant, but belonging to different geographical component
313 populations, have to be compared (Arlian et al. 1984; Arlian 1989). Short fragments of

314 mitochondrial or ribosomal DNA spacer regions have been shown not to be suitable
315 markers for examining genetic diversity among Sarcoptes mite populations (e.g.
316 Alasaad et al. 2009c). Further resolution is therefore provided in faster evolving
317 hypervariable sequences such as nuclear polymorphic microsatellite loci. Microsatellites
318 have previously demonstrated to provide strong support for geographically discrete
319 populations, they showed congruence with evolutionary patterns at the population level,
320 and reported genetic differentiation at the skin-scale of individual mangy hosts
321 (Bowcock et al. 1994; Walton et al. 2004b; Alasaad et al. 2008b).

322 The number of the mites utilized in this study (251 samples) has to be
323 considered high as compared with previous studies in this field (see Alasaad et al.
324 2009a for review).

325 All the component populations showed a strong deficiency of heterozygosity
326 over all loci and mites belonging to the same component population showed to be
327 scattered through the same cluster more than subdivided across individual host animals.

328 Sarcoptes mites lack free-living stages. Individual hosts, depending on their
329 susceptibility and behaviour, are more or less ephemeral habitats and may provide
330 patchy environments which hamper random mating (Price 1980; Criscione et al. 2005).
331 All mites on an individual host could form an ‘infrapopulation’ (Bush et al. 1997) with
332 some recurrent generations on that host. Number of generations is affected by short
333 generation interval of the parasite, about three weeks, as well as by life expectancy of
334 the infested host, depending on its susceptibility.

335 In our data set, the reduced gene pools made mites alike each other and hid
336 possible equilibrium between dispersive process and gene flow among infrapopulations.
337 This may be due to rapid diffusion of few genotypes as for an epidemic population
338 structure (Oura et al. 2005).

339 Another evident feature of our results is the lack of homogeneity of genetic
340 diversity across populations. *S. scrofa* mites were the most variable at all. Wild boar
341 populations are widespread and growing, and generally show higher resistance to
342 parasites than other mammalian species (Rodrigues & Hiraoka 1996; Nejsum et al.
343 2009). Consequently, a single host can be affected by repeated infestation events
344 through mites from other infrapopulations or, even, from other component populations.

345 The other major determinants of gene flow among mites are the degree of host
346 specificity and geographical structure of host populations. Previous investigations
347 showed that approaches to individual clustering provide appropriate characterisation of
348 population structure at high F_{st} values (Rosenberg et al. 2001; Manel et al. 2002; Latch
349 et al. 2006). In the presence of very diverging taxa, few loci are needed to achieve high
350 performance, regardless of the sample sizes (Manel et al. 2002; Tadano et al. 2008). In
351 fact, the ideal marker locus for our purposes should be monomorphic within any taxon
352 and polymorphic across taxa (Reed 1973).

353 In our data set, the unusually high value of F_{st} and the high number of private
354 alleles, in most populations, indicated that the mite component populations were very
355 unlike each other. All the 10 loci provided a significant component of among-
356 population diversity. As a consequence, our marker panel provided good accuracy for
357 analysis of the genetic characteristics of *Sarcoptes* populations.

358 *Sarcoptes scabiei* (1) from different host species belonging to different
359 geographical localities, (2) from the same host species belonging to different
360 geographical localities, and (3) from closely related host species belonging to different
361 geographical localities clustered up to their original populations. Clear genetic diversity
362 among mite populations from different geographical localities exists. The differences
363 show to be as stronger as the geographical separation between host populations is

364 larger. In the case of short geographical separations (East and West Sierra Nevada)
365 mites from *C. pyrenaica* scattered randomly in the dendrogram and no clear separation
366 was detected. The differentiation between *V. vulpes* mites from Spain and Italy was
367 highly supported whereas the genetic separation between *V. vulpes* mites from East and
368 West Italian Alps was poorly supported. This finding suggests that gene flow occurring
369 among mites from different localities is related to the geographical distances.

370 The individual mites belonged to the six sympatric host-derived mite
371 populations from East Italian Alps, and the three sympatric host-derived mite
372 populations from West Italian Alps clustered into three main groups (Fig. 1, 3, and 4):
373 herbivore-derived mites (ItNECi, ItNERr, ItNEOm and ItNECe), carnivore-derived
374 mites (ItNEVv, ItNEMm, ItNWMf and ItNWVv), and omnivore-derived mites (ItNWSs).
375 In particular, mites from *S. scrofa* were distinct from both herbivore- and carnivore-
376 derived mites and they did not cluster with the sympatric Northwest Italian populations.
377 In other words, the host-specific separation among the three clusters was stronger than
378 that by the geographical separation between East and West Italian Alps.

379 Similar results were obtained when the overall mite samples of our investigation
380 were analysed (Fig. 1 and 4). For example, Cluster III contained all the carnivore-
381 derived mites regardless of their geographical origins from different European countries
382 under study.

383 Our results from the sympatric wild animals in Italy and from the general
384 analysis of all mite populations show unambiguously lack of gene flow or recent
385 admixture among *Sarcoptes* populations carnivore-, herbivore-, and omnivore-derived.
386 Mite transmission may occur within each mite cluster herbivore-, carnivore-, and
387 omnivore-derived but it seems to be extremely rare or absent among them. This might

388 improve parasite adaptations and led to, what we called, host taxon-derived (carnivore,
389 herbivore, and omnivore host-derived) *Sarcoptes* mite populations.

390 Population structure of *Sarcoptes* would be that of a species subdivided into
391 genetically small populations with restricted gene flow among local demes (Price 1980;
392 Martínez et al. 1999; Nadler & Hafner 1990). Strong specialisation could be the result
393 of a host taxon-derived shift and, even if two host taxon-derived species are sympatric
394 as for their host species, they should be considered as allopatric if the parasites have no
395 possibility of host choice. In other words, the host sympatry is not the same as the
396 parasite sympatry.

397 The probability of disease transfer between sympatric host taxon-derived
398 species could be reduced by evolved intrinsic mechanisms, this means that the
399 behaviour has been selected to impede crosses between individuals from two different
400 host taxon-derived, and this could represent the first step of sympatric speciation
401 (McCoy 2003). Host taxon-derived effect is stronger than the geographical separation in
402 the definition of speciation events.

403 The existence of host taxon-derived *Sarcoptes* mites could be the explanation of
404 the mange-free wildlife populations in sympatry with other mangy wild animals, like
405 the mange-free *C. ibex* and *R. rupicapra* of West Italian Alps which are in sympatry
406 with the endemically mangy population of *V. vulpes*. This effect could be the reason
407 behind the successful of the cross-transmission/infection in some *Sarcoptes* varieties
408 e.g. *S. scabiei* var *vulpes/canis* readily infect dogs and other canids as well as felids
409 including domestic European cats, as all of them belong to the same host taxon-derived
410 (Bornstein 1995).

411 We have no clear explanation of this taxonomic affiliation. Further studies on
412 dispersal capability of host animals and their disposition to interact with each other, host

413 behaviour and parasite adaptation are needed to explain the host taxon-derived
414 Sarcoptes. Characterization of host genetic structure in addition to mite population
415 genetic structure would contribute additional valuable information.

416

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431 **References**

432

433 Alasaad S, Rossi L, Maione S, Sartore S, Soriguer RC, Pérez JM, Rasero R, Zhu XQ,
434 Soglia D (2008a) HotSHOT Plus ThermalSHOCK, a new and efficient
435 technique for preparation of PCR-quality *Sarcoptes* mite genomic DNA.
436 *Parasitology Research*, **103**, 1455–1457.

437 Alasaad S, Soglia D, Sarasa M, Soriguer RC, Pérez JM, Granados JE, Rasero R, Zhu
438 XQ, Rossi L (2008b) Skin-scale genetic structure of *Sarcoptes scabiei*
439 populations from individual hosts: empirical evidence from Iberian ibex-derived
440 mites. *Parasitology Research*, **104**, 101–105.

441 Alasaad S, Rossi L, Soriguer RC, Rambozzi L, Soglia D, Pérez JM, Zhu XQ (2009a)
442 *Sarcoptes* mite from collection to DNA extraction: the lost realm of the
443 neglected parasite. *Parasitology Research*, **104**, 723–732.

444 Alasaad S, Soglia D, Maione S, Sartore S, Soriguer RC, Pérez JM, Rasero R, Rossi L
445 (2009b) Effectiveness of the postponed isolation (post-frozen isolation) method
446 for PCR-quality *Sarcoptes* mite gDNA. *Experimental and Applied Acarology*,
447 **47**, 173–178.

448 Alasaad S, Soglia D, Spalenza V, Maione S, Soriguer RC, Pérez JM, Rasero R, Ryser
449 Degiorgis MP, Nimmervoll H, Zhu XQ, Rossi L (2009c) Is ITS-2 rDNA suitable
450 marker for genetic characterization of *Sarcoptes* mites from different wild
451 animals in different geographic areas?. *Veterinary Parasitology*, **159**, 181–185.

452 Arlian LG (1989) Biology, host relations and epidemiology of *Sarcoptes scabiei*.
453 *Annual Review Entomology*, **34**, 139–161.

454 Arlian LG, Runyan RA, Estes SA (1984) Cross infestivity of *Sarcoptes scabiei*. *Journal*
455 *of American Academy of Dermatology*, **10**, 979–986.

456 Berrilli F, D'Amelio S, Rossi L (2002) Ribosomal and mitochondrial DNA sequence
457 variation in *Sarcoptes* mites from different hosts and geographical regions.
458 *Parasitology Research*, **88**, 772–777.

459 Blanchong JA, Samuel MD, Scribner KT, Weckworth BV, Langenberg JA, Filcek KB
460 (2008) Landscape genetics and the spatial distribution of chronic wasting
461 disease. *Biology Letters*, **4**, 130–133.

462 Bornstein, S (1995) *Sarcoptes scabiei* infections of the domestic dog, red fox and pig:
463 clinical and serodiagnostic studies. Doctoral thesis, Swedish University of

464 Agricultural Sciences, Department of Veterinary Microbiology, Section of
 465 Parasitology, Uppsala, Sweden

466 Bornstein S, Mörner T, Samuel WM (2001) *Sarcoptes scabiei* and sarcoptic mange. In:
 467 Samuel WM, Pybus MJ, Kocan AA (Eds) *Parasitic diseases of wild mammals*,
 468 2nd edn. Iowa State University Press, Ames, pp 107–119.

469 Bowcock AM, Ruiz-Linares A, Tomfohrde J, Minch E, Kidd J, Cavalli-Sforza LL
 470 (1994) High resolution of human evolutionary trees with polymorphic
 471 microsatellites. *Nature*, **368**, 455–457.

472 Burgess I (1999) Biology and epidemiology of scabies. *Current Opinion in Infectious*
 473 *Diseases*, **12**, 177–180.

474 Bush AO, Lafferty KD, Lotz JM, Shostak AW (1997) Parasitology meets ecology on its
 475 own terms: Margolis et al. revisited. *Journal of Parasitology*, **83**, 575–583.

476 Criscione CD, Poulin R, Blouin S (2005) Molecular ecology of parasites: elucidating
 477 ecological and microevolutionary processes. *Molecular Ecology*, **14**, 2247–2257.

478 Daszak P, Cunningham AA, Hyatt AD (2000) Emerging infectious diseases of wildlife-
 479 threats to biodiversity and human health. *Science*, **287**, 443–449.

480 Excoffier L, Laval G, Schneider S (2005) ARLEQUIN ver 3.0: An integrated software
 481 package for population genetics data analysis. *Evolutionary Bioinformatics*
 482 *Online*, **1**, 47–50.

483 Evanno G, Regnaut S, Goudet J (2005) Detecting the number of clusters of individuals
 484 using the software STRUCTURE: a simulation study. *Molecular Ecology*, **14**,
 485 2611–2620.

486 Fain A (1978) Epidemiological problems of scabies. *International Journal of*
 487 *Dermatology*, **17**, 20–30.

488 Fain A (1968) Étude de la variabilité de *Sarcoptes scabiei* avec une revision des
 489 *Sarcoptidae*. *Acta zoologica et pathologica Antverpiensia*, **47**, 1–196.

490 Falush D, Stephens M, Pritchard JK (2003) Inference of population structure using
 491 multilocus genotype data: linked loci and correlated allele frequencies. *Genetics*,
 492 **164**, 1567–1587.

493 Fandos P (1991) *La cabra montés (Capra pyrenaica) en el Parque Natural de Las*
 494 *Sierras de Cazorla, Segura y Las Villas*. ICONA-CSIC, Madrid.

495 Felsenstein J (1989) PHYLIP - Phylogeny inference package (Version 3.2). *Cladistics*,
 496 **5**, 164–166.

- 497 Glaubitz J (2004) CONVERT: A user-friendly program to reformat diploid genotypic
498 data for commonly used population genetic software packages. *Molecular*
499 *Ecology Notes*, **4**, 309–310.
- 500 González-Candela M, León-Vizcaino L, Cubero-Pablo MJ (2004) Population effects of
501 sarcoptic mange in Barbary sheep (*Ammotragus lervia*) from Sierra Espuña
502 Regional Park, Spain. *Journal of Wildlife Diseases*, **40**, 456–465.
- 503 Goudet J (1995) FSTAT (vers. 1.2): a computer program to calculate F-statistics.
504 *Journal of Heredity*, **86**, 485–486.
- 505 Gu XB, Yang GY (2008) A study on the genetic relationship of mites in the genus
506 *Sarcoptes* (Acari: Sarcoptidae) in China. *International Journal of Acarology*, **32**,
507 183–190.
- 508 Huson DH, DeZulian T, Franz M, Rausch C, Richter DC, Rupp R (2007) Dendroscope -
509 an interactive tree drawer. *BMCB*, **8**, 460.
- 510 Johnson KP, Williams BL, Drown DM, Adams RJ, Clayton DH (2002) The population
511 genetics of host specificity: genetic differentiation in dove lice (Insecta:
512 Phthiraptera). *Molecular Ecology*, **11**, 25–38.
- 513 Latch EK, Dharmarajan G, Glaubitz JC, Rhodes OL Jr (2006) Relative performance of
514 Bayesian clustering software for inferring population substructure and individual
515 assignment at low levels of population differentiation. *Conservation Biology*, **7**,
516 295–302.
- 517 León-Vizcaíno L, Ruíz de Ybáñez MR, Cubero MJ (1999) Sarcoptic mange in Spanish
518 ibex from Spain. *Journal of Wildlife Diseases*, **35**, 647–659.
- 519 Lewontin R C (1964) The interaction of selection and linkage. I. General
520 considerations; heterotic models. *Genetics*, **49**, 49–67.
- 521 Lyles AM, Dobson AP (1993) Infectious disease and intensive management: population
522 dynamics, threatened hosts, and their parasites. *Journal of Zoo and Wildlife*
523 *Medicine*, **24**, 315–326.
- 524 McCoy KD (2003) Sympatric speciation in parasites—what is sympatry?. *Trends in*
525 *Parasitology*, **19**, 400–404.
- 526 Manel S, Berthier P, Luikart G (2002) Detecting wildlife poaching: identifying the
527 origin of individuals with Bayesian assignment tests and multilocus genotypes.
528 *Conservation Biology*, **16**, 650–659.
- 529 Martínez JG, Soler JJ, Soler M, Møller AP, Burke T (1999) Comparative population
530 structure and gene flow of a brood parasite, the great spotted cuckoo (*Clamator*

531 glandarius) and its primary host, the magpie (*Pica pica*). *Evolution*, **53**, 269–
532 278.

533 Minch E (1997) <http://hpgl.stanford.edu/projects/microsat/>

534 Mörner T (1992) Sarcoptic mange in Swedish wildlife. *Revue Scientifique et Technique*
535 *de l'Office International des Epizooties*, **11**, 115–121.

536 Nadler SA, Hafner MS (1990). Genetic differentiation among chewing louse
537 populations (Mallophaga: Trichodectidae) in a pocket gopher contact zone
538 (Rodentia: Geomyidae). *Evolution*, **44**, 942–951.

539 Nejsum P, Roepstorff A, Jørgensen CB, Fredholm M, Göring HH, Anderson TJ,
540 Thamsborg SM (2009) High heritability for *Ascaris* and *Trichuris* infection
541 levels in pigs. *Heredity*, **102**, 357–64.

542 Oleaga A, Balseiro A, Gortázar C (2008) Sarcoptic mange in two roe deer (*Capreolus*
543 *capreolus*) from northern Spain. *European Journal of Wildlife Research*, **54**,
544 134–137.

545 Oura CAL, Asiimwe BB, Weir W, Lubega GW, Tait A (2005) Population genetic
546 analysis and sub-structuring of *Theileria parva* in Uganda. *Molecular and*
547 *Biochemical Parasitology*, **140**, 229–239.

548 Peakall R, Smouse PE (2006) GENALEX 6: genetic analysis in Excel. Population
549 genetic software for teaching and research. *Molecular Ecology Notes*, **6**, 288–
550 295.

551 Pence DB, Ueckermann E (2002) Sarcoptic mange in wildlife. *Revue Scientifique Et*
552 *Technique*, **21**, 385–398.

553 Pérez JM, Ruíz-Martínez I, Granados JE, Soriguer RC, Paulino F (1997) The dynamics
554 of sarcoptic mange in the ibex population of Sierra Nevada in Spain - influence
555 of climatic factors. *Journal of Wildlife Research*, **2**, 86–89.

556 Price PW (1980) *Evolutionary biology of parasites*. Princeton University Press,
557 Princeton NJ.

558 Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using
559 multilocus genotype data. *Genetics*, **155**, 945–959.

560 Raymond M, Rousset F (1995) GENEPOP (version 1.2): Population genetics software
561 for exact tests and ecumenism. *Journal of Heredity*, **86**, 248–249.

562 Reed TE (1973) Number of gene loci required for accurate estimation of ancestral
563 population proportions in individual human hybrids. *Nature*, **244**, 575–576.

- 564 Robinson T (1996) Wildlife-a reservoir for disease. *Microbiology Australia*, **7**, 30–32.
- 565 Rosenberg NA, Burke T, Elo K, Feldman MW, Freidlin PJ, Groenen MAM, Hillel J,
566 Mäki-Tanila A, Tixier-Boichard M, Vignal A, Wimmers K, Weigend S (2001)
567 Empirical evaluation of genetic clustering methods using multilocus genotypes
568 from 20 chicken breeds. *Genetics*, **159**, 699–713.
- 569 Rodrigues DL, Hiraoka M (1996) *Sus scrofa domestica* endoparasitic resistance in the
570 Amazonas. *Annals of the New York Academy of Sciences*, **791**, 473–477.
- 571 Rossi L, Fraquelli C, Vesco U, Permunion R, Somnavilla GM, Carmignola G, Da
572 Pozzo M, Meneguz PG (2007) Descriptive epidemiology of a scabies epidemic
573 in chamois in the Dolomite Alps, Italy. *European Journal of Wildlife Research*,
574 **53**, 131–141.
- 575 Ryser-Degiorgis MP, Ryser A, Bacciarini LN, Angst C, Gottstein B, Janovsky M,
576 Breitenmoser U (2002) Notoedric and sarcoptic mange in free-ranging lynx from
577 Switzerland. *Journal of Wildlife Diseases*, **38**, 228–232.
- 578 Scott ME (1988) The impact of infection and disease on animal populations:
579 implications for conservation biology. *Conservation Biology*, **2**, 40–56.
- 580 Slatkin M (1994) Linkage disequilibrium in growing and stable populations. *Genetics*,
581 **137**, 331–336.
- 582 Slatkin M, Excoffier L (1996) Testing for linkage disequilibrium in genotypic data
583 using the EM algorithm. *Heredity*, **76**, 377–383.
- 584 Soglia D, Rambozzi L, Maione S, Spalenza V, Sartore S, Alasaad S, Sacchi P, Rossi L
585 (2009) Two simple techniques for the safe *Sarcoptes* collection and individual
586 mite DNA extraction. *Parasitology Research*, **105**, 1465–1468.
- 587
- 588 Tadano R, Nishibori M, Tsudzuki M (2008) High accuracy of genetic discrimination
589 among chicken lines obtained through an individual assignment test. *Animal*
590 *Genetics*, **39**, 567–571.
- 591 Walton SF, Choy JL, Bonson A, Valle A, McBroom J, Taplin D, Arlian L, Mathews JD,
592 Currie B, Kemp DJ (1999) Genetically distinct dog-derived and human-derived
593 *Sarcoptes scabiei* in scabies-endemic communities in northern Australia.
594 *American Journal of Tropical Medicine and Hygiene*, **61**, 542–547.
- 595 Walton SF, Holt DC, Currie BJ, Kemp DJ (2004a) Scabies: New future for a neglected
596 disease. *Advances in Parasitology*, **57**, 309–376.
- 597 Walton SF, Currie BJ, Kemp DJ (1997) A DNA fingerprinting system for the

598 ectoparasite *Sarcoptes scabiei*. *Molecular and Biochemical Parasitology*, **85**,
599 187–196.

600 Walton SF, Dougall A, Pizzutto S, Holt D, Taplin D, Arlian LG, Morgan M, Currie BJ,
601 Kemp DJ (2004b) Genetic epidemiology of *Sarcoptes scabiei* (Acari:
602 Sarcoptidae) in northern Australia. *International Journal for Parasitology*, **34**,
603 839–849.

604 Zahler M, Essig A, Gothe R, Rinder H (1999) Molecular analyses suggest
605 monospecificity of the genus *Sarcoptes* (Acari: Sarcoptidae). *International*
606 *Journal for Parasitology*, **29**, 759–766.

Table 1 Countries, geographical locations and host species used in this study, together with the number of host animals and Sarcoptes mite samples.

Code	Countries	Geographical	Host taxon	Host species	N°. of animals	N°. of mites
ItNERr	Italy	Northeast	Herbivore	<i>Rupicapra rupicapra</i>	20	63
ItNECi	Italy	Northeast	Herbivore	<i>Capra ibex</i>	10	25
ItNECe	Italy	Northeast	Herbivore	<i>Cervus elaphus</i>	1	1
ItNEOam	Italy	Northeast	Herbivore	<i>Ovis aries musimon</i>	1	2
ItNEVv	Italy	Northeast	Carnivore	<i>Vulpes vulpes</i>	7	23
ItNEMm	Italy	Northeast	Carnivore	<i>Martes martes</i>	1	3
ItNwVv	Italy	Northwest	Carnivore	<i>Vulpes vulpes</i>	11	30
ItNwMf	Italy	Northwest	Carnivore	<i>Martes foina</i>	1	2
ItNwSs	Italy	Northwest	Omnivore	<i>Sus scrofa</i>	1	3
FrNESs	France	Northeast	Omnivore	<i>Sus scrofa</i>	4	5
SpNEVv	Spain	Northeast	Carnivore	<i>Vulpes vulpes</i>	1	4
SpNWRp	Spain	Northwest	Herbivore	<i>Rupicapra pyrenaica</i>	9	26
SpSECp	Spain	Southeast	Herbivore	<i>Capra pyrenaica</i>	21	33
SpSWCp	Spain	Southwest	Herbivore	<i>Capra pyrenaica</i>	11	30
SpEMf	Spain	West	Carnivore	<i>Martes foina</i>	1	1

Table 2 Private alleles detected at the 10 microsatellite loci for the host-associated mite populations, together with their frequencies.

Pop (N°mites)	Locus	Allele	Freq	Pop (N° mites)	Locus	Allele	Freq
ItNERr (63)	ms33	224	0,008	ItNEVv (23)	ms35	150	0,065
	ms33	244	0,025		ms41	232	0,022
	ms34	170	0,016	ItNwVv (30)	ms35	146	0,200
		192	0,190		ms41	264	0,033
	ms41	214	0,008	SpNEVv(23)	ms38	205	1,000
	ms41	250	0,083	FrNESs(5)	ms33	266	0,800
	ms38	290	0,008		ms33	268	0,100
ItNECi (25)	ms34	188	0,104		ms33	270	0,100
	ms34	208	0,042		ms34	182	0,200
	ms35	138	0,022		ms35	126	0,200
	ms37	176	0,045		ms35	128	0,300
	ms41	244	0,026		ms36	287	0,400
	ms38	223	0,043		ms37	178	0,900
SpSWCp(30)	ms35	160	0,333		ms41	228	0,600
	ms36	263	0,017		ms44	274	0,700
	ms36	273	0,017	ItNWSs(3)	ms33	274	1,000
	ms40	217	0,100		ms34	200	0,250
	ms40	225	0,067		ms40	235	1,000
ms35		158	0,015		ms45	176	0,500
SpSECp(33)	ms36	277	0,015	ItNWMf(2)	ms38	219	0,250
	ms45	164	0,030	SpNWRp (26)	ms45	198	0,679
		ItNECe(1); SpEMf(1); ItNEOam(2) and ItNEMm(3): no private allele detected					

Table 3 Descriptive statistics for the main mite populations (R=allelic richness; He=expected heterozygosity; Ho=observed heterozygosity)

	ItNERr	ItNECi	ItNEVv	ITNwVv	SpNWRp	SpSWCp	SpSECp	FrNESs- ItNWSs
R	1.9	2.6	2.0	1.5	1.6	1.8	1.8	3.4
He	0.215	0.339	0.232	0.119	0.119	0.217	0.216	0.667
Ho	0.048	0.097	0.126	0.020	0.051	0.077	0.103	0.340

Fig. 1 Europe map showing approximate sites for sample collection, together with structure clusters. The colours within bars show the proportion of membership of each individual to the genetic clusters for each *Sarcoptes* population separately. The pie charts give the genetic membership per *Sarcoptes* population. 1=SpNWRp, 2=SpSWCp, 3=SpSECp, 4=ItNWSs, 5=FrNESs, 6=SpNEVv, 7=ItNEVv, 8=ItNWVv, 9=ItNEMm, 10=ItNWMf, 11=SpEMf, 12=ItNERr, 13=ItNECi, 14=ItNECe, and 15=ItNEOam. For site abbreviations see Table 1.

Fig. 2 Unrooted Dps consensus dendrogram for individual *Sarcoptes* mites from three *V. vulpes*-derived mite populations from Northwest and Northeast Italian Alps, and from Northeast Spain. Numbers at the nodes are percentage values of 1000 bootstraps supporting the same branching structure. Codes in this figure (bold for Northwest Italian Alps, italic for Northeast Italian Alps, grey for Northeast Spain) represent the sample codes in Table 1 and Fig.1.

Fig. 3 Unrooted Dps consensus dendrogram for individual *Sarcoptes* mites from six sympatric host-derived mite populations in Northeast Italian Alps, and three sympatric host-derived mite populations in Northwest Italian Alps (Table 1). Numbers at the nodes are percentage values of 1000 bootstraps supporting the same branching structure. Thick branches for all carnivore-derived mites, thin branches for all herbivore-derived mites, grey branches for *S. scrofa*-derived mites. Carnivore-derived mites: *V. vulpes* and Herbivore-derived mite: *C. ibex* and *R. Rupicapra*. a: *M. Foina*, b: *M. martes*, c: *C. ibex*, d: *O. aries musimon* and e: *C. elaphus*.

Fig. 4 Unrooted Dps consensus dendrogram for individual *Sarcoptes* mites from the 15 wild host derived populations (Table 1) using a similarity matrix based on the proportion of shared alleles. Thick branches for all carnivore-derived mites, thin branches for all herbivore-derived mites, grey branches for *S. scrofa*-derived mites. Cluster I (SpNWRp, SpSWCp and SpSECp), Clusters IIa and IIb (ItNWSs and FrNESSs), Cluster III (ItNEMm, ItNWMf, ItNWWv and ItNEVv) and Cluster IV (ItNERr, ItNECi, ItNECe, and ItNEOam). a: SpNEVv, b: SpEMf and arrow: ItNECi.

Fig. 1

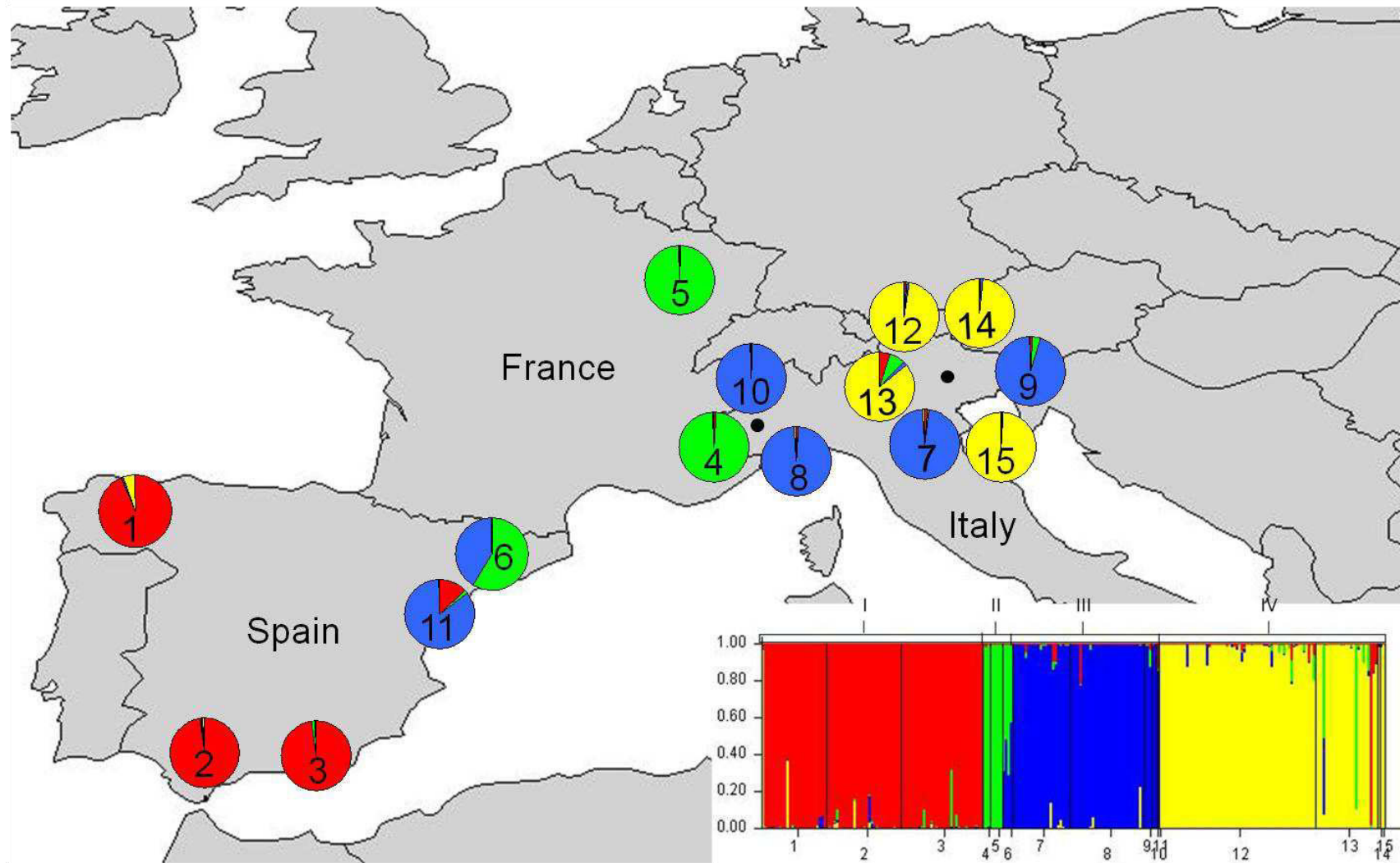


Fig. 3

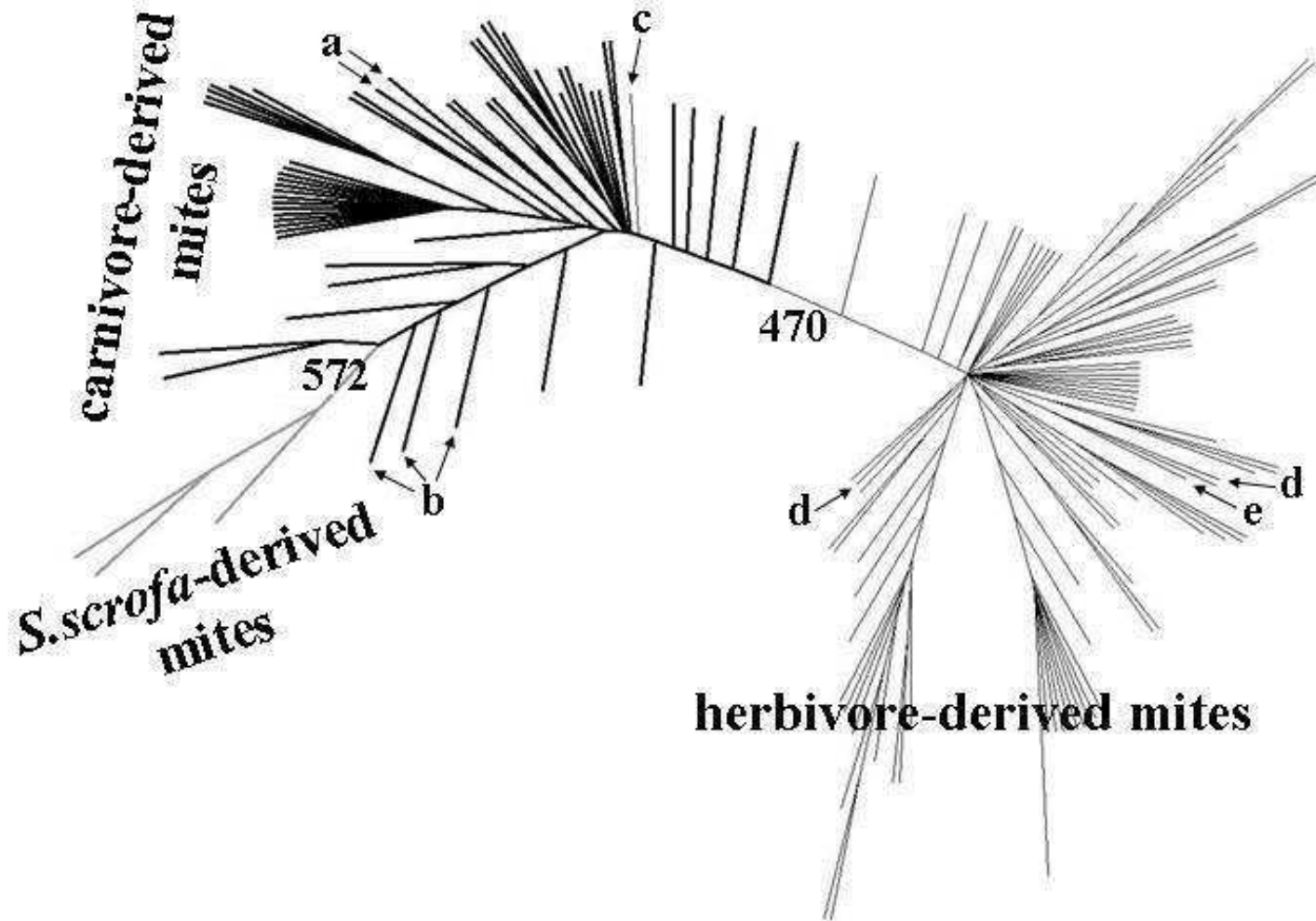


Fig. 4

