

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Treating pathological pain: is KCC2 the key to the gate?

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/132380> since

Published version:

DOI:10.1586/ERN.13.40

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)



UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

Questa è la versione dell'autore dell'opera:

Expert Review of Neurotherapeutics vol. 13(5), 2013, pp. 469-71, doi:
10.1586/ern.13.40.

The definitive version is available at:

La versione definitiva è disponibile alla URL:

<http://informahealthcare.com/doi/abs/10.1586/ern.13.40>

Treating pathological pain: is KCC2 the key to the gate?

Nicolas Doyon^{1,2}, Francesco Ferrini³, Martin Gagnon^{1,2} and Yves De Koninck^{1,2}

¹Institut Universitaire de santé mentale de Québec, Québec, G1J 2G3, Canada

²Department of Psychiatry and Neuroscience, Université Laval, Québec, Québec, G13 7P4, Canada

³Department of Veterinary Sciences, University of Turin, 10095, Grugliasco, Turin, Italy

Despite intensive efforts, development of novel treatments of neuropathic pain has encountered mitigated success. While common and incapacitating, neuropathic pain remains poorly treated, with few patients experiencing greater than 50% symptom relief⁽¹⁾. As an example, Pregabalin, which targets the $\alpha_2\delta$ subunit of the voltage-dependent calcium channel and which is the current gold standard for neuropathic pain, reduces the pain score significantly in only one out of four patients⁽²⁾. Targeting TrpV1 channel, which has attracted considerable resources, has only yielded the weak-selling capsaicin patch and has recently been denied indication expansion into HIV-related neuropathic pain by the FDA. In addition to weak efficacy issues, most approved treatments cause common and significant central side effects such as sedation and dizziness, diminishing the quality of life of affected patients. Opiate-based treatments, for example, are plagued with several side effects, beyond sedation, including psychomotor issues, abuse liability, tolerance and even paradoxical hyperalgesia⁽³⁾. Recent promising alternatives, such as anti-nerve growth factor (NGF) antibodies have demonstrated very strong analgesic efficacy, yet, these strategies were put on hold due to severe side effects; many patients developing joint damage⁽⁴⁾.

The unmet needs of improved efficacy and better adverse event profile will require a different approach. One potential way of treating neuropathic pain is to restore proper inhibitory pathways in the central nervous system. Indeed, central disinhibition has long been known to play a major role in the pathophysiology of neuropathic pain states⁽⁵⁾. After nerve injury, there is a marked decrease in the inhibitory efficacy of GABA_A and glycine receptor-mediated transmission which results in aberrant transmission of sensory and nociceptive information to the brain⁽⁶⁾. A first step to develop therapeutics that would restore proper inhibition is to understand the underlying mechanism of disinhibition. While several causes have been proposed⁽⁶⁾, a key mechanism that has emerged involves the disruption of Cl⁻ homeostasis resulting from loss of activity of the K⁺-Cl⁻ co-transporter KCC2, a molecule responsible for maintaining efficient Cl⁻ mediated inhibition in central neurons⁽⁷⁾. This mechanism appears implicated across several pathological pain syndromes with diverse aetiologies, including spinal cord

injury⁽⁸⁾, inflammation⁽⁹⁾, painful diabetic neuropathy⁽¹⁰⁾, trigeminal pain⁽¹¹⁾ and more recently even morphine-induced paradoxical hyperalgesia⁽³⁾.

Several components of the cellular signalling cascade underlying impaired Cl⁻ homeostasis in spinal nociceptive pathways in pathological conditions have been identified, uncovering several potential therapeutic targets^(12,3). Enhanced BDNF release either from microglia^(12,3,13) or from sensory nerves⁽¹⁴⁾, acting on TrkB receptors to cause downregulation of KCC2, appears as a common culprit in several conditions. In processes involving microglia, a key event is the *de novo* expression of the purinergic P2X4 receptors (P2X4R) in these cells at the spinal level^(15,12,3,13). Each step of the P2X4R-BDNF-TrkB-KCC2 pathway thus emerges as a potential druggable target for combating pathological pain symptoms. Indeed, the ablation of P2X4R prevents the development of neuropathic pain⁽¹³⁾.

Importantly, ablating microglia, chelating BDNF or blocking P2X4R or TrkB receptors reverses established pain hypersensitivity, indicating that the identified pathway plays an active role in maintaining pathological pain^(12,3). Targeting signalling events upstream of KCC2 may thus serve to reverse the pathology, not only prevent its development. Yet, altering microglial function may be detrimental as they are involved in essential support and immune activities. Similarly, interfering with BDNF-TrkB signalling is likely to impact on a multitude of trophic and survival mechanisms in neurons. An obvious alternative approach to counter disinhibition is to increase GABA_A receptor-mediated function. However, a fundamental limitation of drugs acting on GABA_A receptors such as benzodiazepines or barbiturates may be found in the ionic mechanisms underlying their action.

Indeed, since KCC2 is responsible for Cl⁻ extrusion, disruption of its function causes a collapse of the transmembrane Cl⁻ gradient and a depolarizing shift in GABA_A reversal potential (E_{GABA}). This in turn leads to a decrease in inhibitory efficacy⁽¹⁶⁾. The strategy of compensating the decrease in Cl⁻ driving force by an increase in Cl⁻ conductance to restore inhibitory current may be at first met by success. Yet, it inherently has its limitations because it may exacerbate the collapse of the Cl⁻ gradient. This appears to be the case whereby low dosage of midazolam is effectively anti-hyperalgesic in neuropathic pain models, while the efficiency of the drug is reduced at high doses⁽¹⁷⁾. The mechanistic explanation for this paradoxical effect may have an ionic basis. For hyperpolarization to result from Cl⁻ currents, it is mandatory that the Cl⁻ reversal potential (E_{Cl}) be more hyperpolarized than the membrane potential which is only possible at low intracellular Cl⁻ concentration ($[Cl^-]_i$). The value of $[Cl^-]_i$ results from a dynamical equilibrium between Cl⁻ influx through channels and its efflux through KCC2. It follows

that KCC2 hypofunction does not only lead to depolarization of the GABA_A reversal potential (E_{GABA}), it also renders E_{GABA} more labile⁽¹⁸⁾. For example, large Cl⁻ loads exacerbate E_{GABA} depolarization⁽¹⁹⁾ and repeated or sustained activity can cause dynamic collapse of inhibition, especially in dendrites^(20,18). Thus, increased GABA_A receptor-mediated activity will have two opposing effects in cells with depleted KCC2 activity: first, an increase in Cl⁻ mediated inhibition, followed by an activity dependant collapse of Cl⁻ transmembrane gradient. This negative feedback implies that increasing GABA_A activity will rapidly reach a limit.

This phenomenon in itself is not sufficient however to explain the decrease in drug efficacy occurring at high dose. The latter paradoxical effect can be explained by the depolarizing flow of HCO₃⁻ to which GABA_A and glycine channels are also permeable⁽²¹⁾. In contrast to E_{Cl} , E_{HCO_3} is maintained relatively stable by pH buffering mechanisms and rapid diffusion of CO₂ across the membrane allowing efficient [HCO₃⁻]_i replenishment by conversion of H₂O and CO₂ via a carbonic anhydrase (CA) mediated reaction⁽²¹⁾. Accordingly, upon collapse of Cl⁻ currents due to a drug-induced heightened GABA_A function, the opposing HCO₃⁻ current becomes dominant, effectively counteracting the pro-inhibitory action of the drug.

One strategy to mitigate the negative effects of HCO₃⁻ currents through GABA_A channels is to block the CA activity, for example using acetazolamide, enabling activity-dependant depletion of [HCO₃⁻]_i. The decrease in HCO₃⁻ current will parallel the collapse of the Cl⁻ current⁽³⁾ extending the efficacy of pro-GABA_A drugs⁽¹⁷⁾. However, this strategy also has its limits because it does not prevent the dynamic collapse in Cl⁻ gradient, meaning that a certain amount of hyperexcitability will remain uncompensatable. Furthermore, impaired Cl⁻ extrusion can cause a positive feedback loop between excitation and Cl⁻ accumulation in which case sustained inputs can lead to catastrophic failure of inhibition⁽¹⁸⁾. In addition, potential problems associated with impaired intracellular pH regulation in absence of normal CA activity may limit the usefulness of targeting HCO₃⁻ currents.

For the reasons exposed above, a promising analgesic strategy is to enhance KCC2 activity not only to restore E_{GABA} to its normal value, but also to prevent activity dependant Cl⁻ accumulation and its associated side effects. Furthermore, since KCC2 already operates near its equilibrium point⁽²²⁾, potential excess activity caused by treatment aiming to restore its normal function is unlikely to have adverse effects on neuron dynamics⁽¹⁸⁾. How can KCC2 function be restored? The most

straightforward strategy is to directly target KCC2 function, expression and/or turnover. Thus, enhancing KCC2 function can be achieved by promoting its synthesis and/or reducing its degradation, but also potentially by modulating transporter function. For example, it has been suggested that the quaternary structure of KCC2 is an important determinant of its function. Indeed, KCC2 oligomerization has been suggested as a rapid mechanism by which KCC2 activity is modulated during development⁽²³⁾, the oligomeric form being the active one. Consistent with these findings, BDNF-dependent pain hypersensitivity appears to be associated with an increase in the monomer/oligomer ratio⁽³⁾. Compounds stabilizing the oligomeric state may thus be effective KCC2 enhancers. Proof of principle that positive modulation of KCC2 function can have therapeutic benefits has recently been provided for analgesia⁽²⁴⁾ and treatment of motor spasticity⁽²⁵⁾.

Because KCC2 expression is restricted to central neurons⁽²⁶⁾, targeting this transporter may reduce the risk of unwanted effects in peripheral nervous system or in other tissues. And because disrupted KCC2 activity appears to be involved in several neurological and psychiatric disorders including epilepsy, motor spasticity, stress and schizophrenia^(27,28,20,29,30), KCC2 enhancing drugs may have broad therapeutic potential.

Financial & competing interest disclosure

The authors acknowledge the financial support from the Canadian Institutes of Health Research, the Natural Sciences and Engineering Research Council of Canada, the Fonds de recherche Québec – Santé, the Krembil foundation and the Regione Piemonte. The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript.

References

1. Finnerup NB, Otto M, Mcquay HJ, Jensen TS, Sindrup SH. Algorithm for neuropathic pain treatment: An evidence based proposal. *Pain* 118(3), 289-305 (2005).
2. Freynhagen R, Strojek K, Griesing T, Whalen E, Balkenohl M. Efficacy of pregabalin in neuropathic pain evaluated in a 12-week, randomised, double-blind, multicentre, placebo-controlled trial of flexible- and fixed-dose regimens. *Pain* 115(3), 254-263 (2005).
3. Ferrini F, Trang T, Mattioli TA *et al.* Morphine hyperalgesia gated through microglia-mediated disruption of neuronal Cl(-) homeostasis. *Nature Neuroscience* 16(2), 183-192 (2013).

4. Garber K. Fate of novel painkiller mAbs hangs in balance (vol 29, pg 173, 2011). *Nature Biotechnology* 30(5), 466-466 (2012).
5. Berger JV, Knaepen L, Janssen SP *et al.* Cellular and molecular insights into neuropathy-induced pain hypersensitivity for mechanism-based treatment approaches. *Brain Research Reviews* 67(1-2), 282-310 (2011).
6. Labrakakis C, Ferrini F, De Koninck Y. Mechanisms of plasticity of inhibition in chronic pain conditions. In: *Inhibitory Synaptic Plasticity*. Springer, 91-105 (2011)
7. Coull JAM, Boudreau D, Bachand K *et al.* Trans-synaptic shift in anion gradient in spinal lamina I neurons as a mechanism of neuropathic pain. *Nature* 424(6951), 938-942 (2003).
8. Lu Y, Zheng J, Xiong L, Zimmermann M, Yang J. Spinal cord injury-induced attenuation of GABAergic inhibition in spinal dorsal horn circuits is associated with down-regulation of the chloride transporter KCC2 in rat. *Journal of Physiology-London* 586(23), 5701-5715 (2008).
9. Zhang W, Liu L, Xu T. Reduced potassium-chloride co-transporter expression in spinal cord dorsal horn neurons contributes to inflammatory pain hypersensitivity in rats. *Neuroscience* 152(2), 502-510 (2008).
10. Jolivald CG, Lee CA, Ramos KM, Calcutt NA. Allodynia and hyperalgesia in diabetic rats are mediated by GABA and depletion of spinal potassium-chloride co-transporters. *Pain* 140(1), 48-57 (2008).
11. Wei B, Kumada T, Furukawa T *et al.* Pre- and Post-Synaptic Switches of Gaba Actions Associated with Cl(-) Homeostatic Changes Are Induced in the Spinal Nucleus of the Trigeminal Nerve in A Rat Model of Trigeminal Neuropathic Pain. *Neuroscience* 228, 334-348 (2013).
12. Coull JAM, Beggs S, Boudreau D *et al.* BDNF from microglia causes the shift in neuronal anion gradient underlying neuropathic pain. *Nature* 438(7070), 1017-1021 (2005).
13. Ulmann L, Hatcher JP, Hughes JP *et al.* Up-Regulation of P2X(4) Receptors in Spinal Microglia after Peripheral Nerve Injury Mediates BDNF Release and Neuropathic Pain. *Journal of Neuroscience* 28(44), 11263-11268 (2008).
14. Zhao J, Seereeram A, Nassar MA *et al.* Nociceptor-derived brain-derived neurotrophic factor regulates acute and inflammatory but not neuropathic pain. *Molecular and Cellular Neuroscience* 31(3), 539-548 (2006).
15. Tsuda M, Shigemoto-Mogami Y, Koizumi S *et al.* P2X(4) receptors induced in spinal microglia gate tactile allodynia after nerve injury. *Nature* 424(6950), 778-783 (2003).
16. Prescott SA, Sejnowski TJ, De Koninck Y. Reduction of anion reversal potential subverts the inhibitory control of firing rate in spinal lamina I neurons: towards a biophysical basis for neuropathic pain. *Molecular Pain* 2 (2006).

17. Asiedu M, Ossipov MH, Kaila K, Price TJ. Acetazolamide and midazolam act synergistically to inhibit neuropathic pain. *Pain* 148(2), 302-308 (2010).
18. Doyon N, Prescott SA, Castonguay A, Godin AG, Kroeger H, De Koninck Y. Efficacy of Synaptic Inhibition Depends on Multiple, Dynamically Interacting Mechanisms Implicated in Chloride Homeostasis. *Plos Computational Biology* 7(9) (2011).
19. Cordero-Erausquin M, Coull JAM, Boudreau D, Rolland M, De Koninck Y. Differential maturation of GABA action and anion reversal potential in spinal lamina I neurons: Impact of chloride extrusion capacity (vol 19, 9613, 2005). *Journal of Neuroscience* 25(45), 10574-10575 (2005).
20. Hewitt SA, Wamsteeker JI, Kurz EU, Bains JS. Altered chloride homeostasis removes synaptic inhibitory constraint of the stress axis. *Nature Neuroscience* 12(4), 438-443 (2009).
21. Kaila K. Ionic Basis of Gaba(A) Receptor-Channel Function in the Nervous-System. *Progress in Neurobiology* 42(4), 489-537 (1994).
22. Buzsaki G, Kaila K, Raichle M. Inhibition and brain work. *Neuron* 56(5), 771-783 (2007).
23. Blaesse P, Guillemain I, Schindler J *et al.* Oligomerization of KCC2 correlates with development of inhibitory neurotransmission. *Journal of Neuroscience* 26(41), 10407-10419 (2006).
24. Coull JA, Gagnon M. The manipulation of cation-chloride co-transporters as a novel means to treat persistent pain, epilepsy and other neurological disorders. *Curr.Opin.Investig.Drugs* 10(1), 56-61 (2009).
25. Bos R, Sadlaoud K, Boulenguez P *et al.* Activation of 5-HT_{2A} receptors upregulates the function of the neuronal K-Cl cotransporter KCC2. *Proc.Natl.Acad.Sci.U.S.A* 110(1), 348-353 (2013).
26. Rivera C, Voipio J, Payne JA *et al.* The K⁺/Cl⁻ co-transporter KCC2 renders GABA hyperpolarizing during neuronal maturation. *Nature* 397(6716), 251-255 (1999).
27. Arion D, Lewis DA. Altered Expression of Regulators of the Cortical Chloride Transporters NKCC1 and KCC2 in Schizophrenia. *Archives of General Psychiatry* 68(1), 21-31 (2011).
28. Boulenguez P, Liabeuf S, Bos R *et al.* Down-regulation of the potassium-chloride cotransporter KCC2 contributes to spasticity after spinal cord injury. *Nature Medicine* 16(3), 302-U97 (2010).
29. Huberfeld G, Wittner L, Clemenceau S *et al.* Perturbed chloride homeostasis and GABAergic signaling in human temporal lobe epilepsy. *Journal of Neuroscience* 27(37), 9866-9873 (2007).
30. Hyde TM, Lipska BK, Ali T *et al.* Expression of GABA Signaling Molecules KCC2, NKCC1, and GAD1 in Cortical Development and Schizophrenia. *Journal of Neuroscience* 31(30), 11088-11095 (2011).