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Molecular and cellular mechanisms of skeletal muscle atrophy: an update

Alessandro Fanzani, Viviane M. Conraads, Fabio Penna, Wim Martinet.

Addresses:

Alessandro Fanzani, PhD:

Department of Biomedical Sciences and Biotechnologies and Interuniversity Institute of Myology (IIM), University of Brescia, viale Europa 11, 25123 Brescia, Italy. * Corresponding author. Tel +39-030-3717567. Fax +39-030-3701157.

E-mail address: fanzani@med.unibs.it (Alessandro Fanzani)

Viviane Conraads, MD, PhD:

Antwerp University Hospital, Wilrijkstraat 1, 2650 Edegem, Belgium; University of Antwerp, Universiteitsplein 1, B-2610 Antwerp, Belgium.

Fabio Penna, PhD:

Department of Biochemistry and Molecular Biology, University of Barcelona, Diagonal 643, 08028 Barcelona, Spain.

Wim Martinet, PhD

Laboratory of Physiopharmacology, University of Antwerp, Universiteitsplein 1, B-2610 Antwerp, Belgium.

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Abstract

Skeletal muscle atrophy is defined as a decrease in muscle mass and occurs when protein degradation exceeds protein synthesis. Potential triggers of muscle wasting are long-term immobilization, malnutrition, severe burns, aging as well as various serious and often chronic diseases, such as chronic

heart failure, obstructive lung disease, renal failure, AIDS, sepsis, immune disorders, cancer and dystrophies. Interestingly, a cooperation between several pathophysiological factors, including inappropriately adapted anabolic (e.g. growth hormone, insulin-like growth factor 1) and catabolic proteins (e.g. tumor necrosis factor alpha, myostatin), may tip the balance towards muscle-specific protein degradation through activation of the proteasomal and autophagic systems or the apoptotic pathway. Based on the current literature, we present an overview of the molecular and cellular mechanisms that contribute to muscle wasting. We also focus on the multi-faceted therapeutic approach that is currently employed to prevent the development of muscle wasting and to counteract its progression. This approach includes adequate nutritional support, implementation of exercise training and possible pharmacological compounds.

1. Introduction

Skeletal muscle is the most abundant tissue in the body of vertebrates and is involved in many different important functions. For instance, skeletal muscle is the primary target of glucose uptake, can regenerate after injury, and is subjected to size remodelling. Overall, skeletal muscle mass represents a determinant of strength, endurance and physical performance. Reduced muscle bulk or muscle atrophy takes place when protein degradation exceeds protein synthesis, leading to a reduction of the cross-sectional area of myofibers and decreased muscle strength. Though muscle atrophy is a consequence of certain physiological processes, such as aging, it may also be a prominent feature present in several disparate diseases. In this latter case, a general loss of body mass, not exclusively limited to muscle, inherently implies an underlying, usually chronic and severe syndrome with impaired prognosis, known as cachexia. In both physiological and pathological conditions, the clinical implications of muscle wasting translate into poor quality of life, implying exercise intolerance and inability to manage daily activity. Since our knowledge on the mechanisms contributing to muscle wasting has remarkably increased in the past 20 years, the aim of the present review is to provide a comprehensive overview of the different cues, mechanisms and degradative machineries involved in muscle atrophy. Furthermore, an update is given on current therapeutic approaches.

2. Muscle atrophy

Muscle atrophy is a physiological consequence of aging (i.e. age-related sarcopenia), defined as the presence of both low muscle mass and low muscle function (strength or performance) [1], but it may also result from prolonged periods of rest or a sedentary lifestyle. Moreover, muscle atrophy represents a clinical feature of cachexia, a multifactorial syndrome implying reduced life expectancy and

accompanying many illnesses like chronic heart failure (CHF), chronic obstructive pulmonary disease (COPD), chronic kidney disease (CKD), cancer, HIV, sepsis, immune disorders and dystrophies [2].

A reduced cross-sectional area of myofibers with subsequent impaired strength is the main characteristic of muscle atrophy [3], mirroring a consistent depletion of contractile proteins. Muscle atrophy is also frequently characterised by a switch of the fiber-type composition. For instance, while the loss of contractile proteins predominantly affects the type II fast fibers during aging or cancer cachexia [3], giving rise to a higher proportion of slow versus fast fibers [4], an increase in type IIX fiber proportions at the expense of either type I or type IIA muscle fibers has been observed in CHF [5-7], suggesting that different pathophysiological cues may differently affect the fiber-type muscular pattern. Besides the loss of muscle mass, inherent deficits in the “quality” of skeletal muscle have been demonstrated. For instance, in symptomatic CHF patients, quadriceps muscle strength is reduced compared to controls, even after correction for reduced circumferential area [8]. Equally important, however, are the prognostic implications of cachexia and muscle loss. Anker et al. [9] provided strong evidence that the identification of a heart failure patient as being cachectic significantly increased the risk of dying, even after correction for age, gender, disease severity and treatment allocation.

Until recently, the lack of a clear definition for cachexia has hampered structured research as well as the development of treatment targets. Therefore, in order to allow timely clinical recognition, objective criteria to define cachexia have become mandatory. According to the last consensus criteria, the condition sine qua non to define cachexia is weight loss greater than 5%, or weight loss greater than 2% in individuals already showing depletion according to current body weight and height (body-mass index [BMI] <20) or skeletal muscle mass [10]. However, the assessment for classification and clinical management of cachectic patients should include additional domains such as anorexia or reduced food intake, catabolic drive, muscle mass and strength as well as functional and psychosocial impairment.

Altogether, these observations clearly indicate that different, somewhat apparently unrelated mechanisms may synergistically cooperate to trigger the impairment of body’s performance through loss of bulk muscle, thereby inducing critical and often fatal health complications.

3. Triggers of muscle atrophy

A dysbalance of anabolic versus catabolic factors may alter nitrogen balance leading to consistent protein depletion and muscle atrophy (Fig. 1). During embryonic development as well as in post-natal life, different anabolic factors are required to ensure proper muscle growth. In this context, growth hormone (GH), androgens (testosterone), insulin and insulin-like growth factor-1 (IGF-1) play a prominent role, with the latter being central due to its unrivalled pleiotropic ability to regulate different muscular mechanisms such as cell proliferation [11] and differentiation [12], myofiber growth [13,14] and regeneration [13]. Accordingly, low levels of circulating IGF-1 have been associated with sarcopenia [15], CHF [16], cancer [17] and other clinical syndromes [18]. Keeping in mind that IGF-1 exerts pleiotropic effects by stimulating proliferation of resident satellite cells during regeneration as well as growth of pre-existing myofibers, it is conceivable that its decline in circulating levels during aging or in consequence of disease may impinge on the muscle architecture. Unlikely, liver-specific IGF-1 knock-out mice exhibit

normal growth rates [19,20], suggesting that muscle-produced IGF-1 may compensate for the lack of systemic (hepatic) IGF-1. In support of the pivotal role of IGF-1, it has been reported that muscle-specific transgenic overexpression of IGF-1 promotes muscle hypertrophy and increases physiological muscle strength [13,21], improves muscle regeneration [13,22], ameliorates the dystrophic phenotype of mdx mice [23,24], protects motor neurons in a mouse model of amyotrophic lateral sclerosis (ALS) [25] and also attenuates disease in a mouse model of spinal and bulbar muscular atrophy [26]. In this regard, the beneficial IGF-1 effects mostly rely on the activation of the PI3K/AKT pathway, which promotes protein synthesis and blunts protein degradation [27-29]. Since insulin activates the PI3K/AKT pathway, analogously to IGF-1, insulin resistance plays a consistent role in muscle atrophy of diabetic patients [30]. Indeed, transgenic mice with a dominant-negative IGF-1 receptor specifically targeted to the skeletal muscle developed insulin resistance, due to formation of hybrid receptors between the mutant and the endogenous IGF-1 and insulin receptors, thus providing an excellent model to study the molecular mechanisms underlying the development of human type 2 diabetes [31]. While sarcopenia occurring during aging mainly relates to impaired anabolism, because of decreased anabolic factors or reduced anabolic response [32,33], different conditions leading to muscle wasting show an increase of certain catabolic factors, including tumor necrosis factor- α (TNF α) [34,35], transforming growth-factor beta (TGF β) ligands such as myostatin [36-39] and glucocorticoids [40-43]. Several tumor bearing animal models developing cachexia are characterized by an excess of circulating TNF β [44] as well as by the increased expression of other cachectic cytokines, such as interleukin-1 (IL-1) [45] and IL-6 [46,47]. Another master regulator of muscle size [48], the TGF β family member termed myostatin, elicits atrophy when administered to an adult animal [49], as, analogously, glucocorticoids do [50]. Finally, it is worth mentioning that protein degradation in skeletal muscle is stimulated by excessive oxidative stress [51], as discussed in more detail below.

4. Molecular pathways underlying muscle atrophy

Unraveling the signaling network that contributes to muscle depletion is pivotal to develop therapeutic interventions (Fig. 1). Among the different pathways controlling muscle size [28,29,52,53], the PI3K/Akt pathway is central and is mainly recruited upon IGF-1/insulin stimuli to promote protein synthesis and to block degradation [54-56]. In the cytoplasm, Akt1 activates S6 kinase 1 (S6K1) via mammalian target of rapamycin (mTOR), which leads to increased protein synthesis [57]. Akt1 phosphorylates also FoxO transcription factors, specifically making FoxO3 unable to enter the nucleus and to drive transcription of certain ubiquitin ligases (e.g. muscle atrophy F-Box (MAFbx/atrogin-1) and muscle RING finger-1 (MuRF1)) [56] or autophagy-related genes (e.g. LC3 and Bnip3) [58], which account for the protein breakdown mediated by the ubiquitin-proteasomal and autophagic/lysosomal pathways [59], respectively (vide infra). Thus, constitutive Akt1 activation produces rapid muscle hypertrophy in transgenic mice [60,61], whereas Akt1/Akt2 double-knockout mice exhibit severe skeletal muscle atrophy [62]. Moreover, S6K1(-/-) myotubes are atrophic and their hypertrophic response to IGF-1, nutrients and membrane-targeted Akt1 is blunted [63], definitely corroborating the importance of the IGF-1/Akt axis pathway on the muscle maintenance.

In support of this evidence, muscle atrophy due to reduced Akt signaling is observed in different diseases or pathophysiological conditions, including ALS [64-66], CKD [67,68], diabetes [30], chronic hypoxia [69], statin-induced myopathy [70], sepsis [71,72], burn injury [73] and aging [74]. In particular, different molecules behave as pro-atrophic factors by reducing Akt signaling, including TNF α [75], TNF-related weak inducer of apoptosis [76], glucocorticoids [77,78], angiotensin [79] and cisplatin [80]. It is noteworthy that one of the most powerful inducers of muscle atrophy, namely myostatin, can elicit activation of Smad2/3 pathway and increased expression of MAFbx/MURF1 ubiquitin ligases followed by increased proteasomal activity [81,82], thus contributing to the breakdown of sarcomeric proteins. Moreover, it has been recently shown that most of the myostatin effects rely on its ability to reduce the Akt signaling [83,84], as myostatin can inhibit the IGF-1-induced myotube hypertrophy by lowering Akt signaling [85].

Overall, these findings suggest that Akt is a valuable target for improvement of muscle performance in different pro-atrophic conditions. Of note, sustaining the Akt pathway confers also several advantages to Duchenne's Muscular Dystrophy (DMD) muscles, as first observed upon IGF-1 administration to mdx mice [24]. Other studies showed that the activation of Akt in DMD muscles promotes hypertrophy [86,87], prevents the force drop induced by eccentric contractions [88], upregulates the utrophin-glycoprotein complex, promotes sarcolemma stability [89], attenuates muscular degeneration, promotes myofiber regeneration, improves muscle function [90] and mediates the hypertrophic effects promoted by matrix metalloproteinase-9 [91,92].

Another pathway playing a consistent role in muscle atrophy relies on the activity of transcription factor NF- κ B [93,94], which is frequently activated by several cytokines through a cascade involving the phosphorylation and consequent degradation of NF- κ B inhibitor I κ -B α [95]. NF- κ B pathway was first implicated in a model of skeletal muscle disuse atrophy [96]. Interestingly, NF- κ B is able to directly bind the MURF1 promoter, thus triggering a proteasome-dependent muscle wasting [97]. In addition, NF- κ B activation is tightly linked to increased oxidative stress, which in turn controls the rate of protein degradation in skeletal muscle [51]. Indeed, NF- κ B induces muscle wasting through an increase in the expression of iNOS, leading to increased nitric oxide (NO) [98-101], as confirmed by the increased levels of protein nitration in aged muscle [102-105]. Thus, elevated NO levels are implicated in age-induced skeletal muscle apoptosis [106], in cytokine-induced cachexia [98,99,107] and iNOS expression is elevated in COPD [108] and human cancer and AIDS patients suffering from cachexia [109]. In addition, nNOS is commonly dislocated in neuromuscular disorders [110], such as ALS [111], and mediates muscle atrophy via regulation of FoxO transcription factors in a model of unloading-induced muscle atrophy [112]. Thus, the NO pathway may represent an effective target for the therapy of both muscle sarcopenia and cachexia [113].

In physiological conditions, a variety of reactive oxygen species (ROS) are produced in the skeletal muscle to allow the adaptation of the muscle to several stimuli [114]. In contrast, prolonged muscle disuse causes increased radical production and subsequent injury in inactive muscle fibers, as seen upon experimental immobilization of skeletal muscles. However, this effect can be delayed by exogenous antioxidants [115-117]. When ROS levels exceed the healthy threshold, at least three different signaling systems are supposed to play a central role in mediating the pro-atrophic effects on inactive muscles,

namely the cytosolic calcium-dependent calpains, the mitogen activated protein kinase (MAPK) system that includes Jnk, p38 and Erk1/2 kinases, and the NF- κ B pathway indeed [51,118].

5. Cellular mechanisms of protein degradation during muscle atrophy

The two major protein degradation pathways in eukaryotic cells are the UPS (ubiquitin-proteasome system) and the autophagy-lysosome pathway (Fig. 1). In normal physiological conditions, the UPS is responsible for the degradation of short-lived proteins, whereas autophagy regulates levels of long-lived proteins and organelles. However, a large body of evidence suggests that both pathways also play an important role in skeletal muscle wasting.

5.1. The ubiquitin-proteasome system

Muscle-specific ubiquitin ligases

Ubiquitin is an abundant 8 kDa peptide found in all cell types. To date, free ubiquitin has no biological function, but exerts its effects by covalent attachment to other proteins [119]. The substrate protein can be mono-ubiquitinated or additional ubiquitin molecules can be added to produce a poly-ubiquitinated protein that is targeted for degradation by the proteasome [120], while the mono-ubiquitinated substrates are likely degraded in the lysosomes [121]. The ubiquitination of proteins is a multi-step reaction [122,123]. The first step comprises activation of ubiquitin by ubiquitin activating enzyme (E1), an ATP hydrolyzing protein that adenylates ubiquitin on its C-terminal glycine and then links the ubiquitin molecule on a cysteine residue at its catalytic site. Subsequently, E1 transfers the activated ubiquitin to the active site of a ubiquitin conjugating enzyme (E2). Finally, E2 binds to a ligase (E3) that recognizes the substrate and transfers ubiquitin to the target protein. In mammalian cells, there is only one ubiquitin activating enzyme. The ubiquitin conjugating enzyme family is composed of several members that specifically interact with a subset of ubiquitin ligases which in turn recognize specific substrates [124,125].

In 2001, two muscle-specific ubiquitin ligases involved in the ubiquitination process of skeletal muscle proteins have been characterized, namely MAFbx/atrogen-1 and MuRF1 [126,127]. MAFbx is member of a multi-protein SCF (Skp1, Cullin1 and F-box) complex, which needs to be assembled first in order to establish E3 activity [127]. MuRF1 contains an N-terminal RING domain, a MuRF family conserved region, a zinc-finger domain (B-box), leucine-rich coiled-coil domains and an acidic C-terminal tail [128]. Knock-out mice for either MAFbx or MuRF1 are phenotypically normal. However, under atrophy conditions, loss of a muscle-specific ubiquitin ligase confers partial protection against muscle wasting [126]. The molecular characterization of these ubiquitin ligases has led to the discovery that MuRF1 binds to the myofibrillar protein titin [128,129] and more importantly to the myosin heavy chain (MyHC), the most abundant protein in the skeletal muscle [130]. Indeed, MyHC was shown to be degraded by the proteasome [131] and loss of MyHC was blocked by siRNA-mediated inhibition of MuRF1 [130]. Moreover, MuRF1 is able to ubiquitinate MyHC in vitro [130]. More recent work showed that other myofibrillar proteins including myosin light chain and myosin-binding protein C are specifically

ubiquitinated by MuRF1 [132]. The relevance of MAFbx in the stimulation of myofibrillar protein degradation is less clear. Hitherto, the only confirmed substrates for MAFbx are MyoD [133] and eIF3f [134]. MyoD is a transcription factor mainly involved in the myogenic program and partially in protein synthesis induction [135], while eIF3f is a translation initiation factor [136]. The role of MAFbx in protein synthesis rather than degradation was then suggested by reports showing no correlation between the rates of protein breakdown and the MAFbx mRNA levels both in vivo [137,138] and in vitro [139]. Moreover, expression of MAFbx/Atrogin-1 and MuRF-1 undergoes different regulation [140,141]. Because muscle atrophy results from both increased protein degradation and/or reduced synthesis, the two E3 ligases contribute most likely in a distinct manner to the wasting process. These observations should be taken into account when the measurement of mRNA levels of such E3s is taken as an indicator of proteasome-dependent protein degradation.

After characterization of MAFbx and MuRF1, another muscle-specific ubiquitin ligase termed TRIM32 was discovered in 2005 [142]. This protein was a tripartite motif (TRIM) family member containing in its structure a RING-finger, a B-box, and a coiled-coil motif. TRIM32 was shown to associate with the thick myofibrillar filament, and is able to ubiquitinate actin and dysbindin [142], a protein involved in lysosomal-endosomal trafficking [143]. Mutations in the E3 ubiquitin ligase TRIM32 are involved in skeletal muscle disorders such as Limb-girdle muscular dystrophy type 2H (LGMD2H) and the multisystemic disorder, Bardet-Biedl syndrome type 11 [144]. Skeletal muscles from murine TRIM32 knock-out mice (T32KO) reveal mild myopathic changes such as Z-line streaming and a dilated sarcotubular system with vacuoles, thereby replicating phenotypes of LGMD2H and sarcotubular myopathy [145]. Analysis of T32KO neural tissue shows a decreased concentration of neurofilaments and a reduction in myelinated motoraxon diameters, suggesting a shift toward a slower motor unit type, as observed in T32KO soleus muscle. These data suggest that muscular dystrophy due to TRIM32 mutations involves both neurogenic and myogenic characteristics [145]. Finally, in 2007 a novel muscle-specific F-box protein (FBXO40) was found to be upregulated after denervation, though not after starvation [146]. FBXO40 was previously identified in LGMD patients that showed a dramatic decrease in comparison to healthy individuals [147]. Very recently, FBXO40 was found to induce IRS1 ubiquitination in skeletal muscle, thereby providing a negative feedback on the pro-hypertrophic IGF1R/IRS1/PI3K/Akt pathway [148], as demonstrated by the enhanced body and muscle size of mice null for FBXO40 during the growth phase associated with elevated IGF1 levels. Future studies will reveal new substrates and specific new roles for E3 ubiquitin ligases, while more investigations in humans are needed to validate their potential role as therapeutic targets for the prevention of muscle atrophy.

Proteasome induction/activation in muscle atrophy

Several physiological factors such as classical hormones, insulin, IGF-1 and TNF α can modulate the expression of enzymes controlling ubiquitination and proteasome-dependent protein degradation. For example, insulin reduction as a consequence of fasting or experimental diabetes leads to increased expression of ubiquitin [30,41], MAFbx [149], as well as UBR1, UBR2, UBR3 [150,151]. Similar effects can be found after glucocorticoid administration [40-43], even though most of the glucocorticoid-induced effects are reverted by IGF-1 [42,43,152]. Also pro-inflammatory cytokines such as TNF α are responsible

for increased ubiquitin expression and accumulation of ubiquitinated proteins [35,153]. However, the first observations indicating a role for the ubiquitin system during atrophy came from cancer cachexia models, showing increased ubiquitin and proteasome subunits expression or protein ubiquitination in tumor bearing animals [154-158]. Later, as MAFbx and MURF1 were discovered, their levels were reported to be increased in AH-130 bearing rats [17], as well as in MAC16 [159] and Colon-26 bearing mice [74,160]. After the identification of proteasome activation in cancer cachexia, similar observations were reported in atrophying muscles following denervation or starvation [161]. These experimental procedures led to the discovery of the ubiquitin ligases MAFbx and MURF1 [126,127]. Worth of note, the atrophy of skeletal muscle following denervation is dependent on the upregulation of specific histone deacetylases (HDAC), namely HDAC4 and HDAC5, which repress Dach2, a negative regulator of myogenin, resulting in the activation of myogenin which in turn drives the increase of MAFbx/Atrogin-1 and MuRF-1 expression, leading to proteolytic muscle wasting [162]. Also sepsis-induced muscle atrophy associates with increased expression and enzymatic activity of the proteasome [163]. Subsequently, the raise in the mRNA levels of the ubiquitin ligases was reported [164]. Hitherto, a report showing increased MAFbx or MURF1 increase exists for almost every experimental condition characterized by muscle wasting, even if evidence for increased proteasome degradation is lacking. These conditions include disuse [126], diabetes [127,149], CKD [149], glucocorticoid administration [55], rheumatoid arthritis [165], COPD [166], alcohol-myopathy [167], AIDS [168], aging [132] and spinal cord injury [169].

5.2 Autophagy

Autophagy is an evolutionarily conserved subcellular process for bulk destruction of proteins and entire organelles [170]. The process is classically considered to be a pathway contributing to cellular homeostasis and adaptation to stress, and thus may act as a pro-survival mechanism. However, excessive stimulation of the autophagic machinery is deleterious and may lead to caspase-independent cell death [171]. Different autophagic pathways have been described, all sharing the import of cytoplasmic components into the lysosome. For the most prevalent form of autophagy (macroautophagy, further referred to as autophagy), double-membrane vacuoles known as autophagosomes will form, originating from a largely undefined structure known as the phagophore or isolation membrane [170]. The process proceeds in a step-wise fashion, with the elongation of the isolation membrane, maturation of the autophagosome followed by its fusion with a lysosome, thereby generating an autolysosome. During this final step, the incorporation of the outer autophagosomal membrane with the lysosomal membrane eventually allows the degradation of the remaining inner single-membrane and the cytoplasmic content of the autophagosome by lysosomal hydrolases. So far, more than 30 autophagy-related (Atg) genes have been identified that regulate autophagy induction and autophagosome formation [172].

Autophagy is constitutively active at basal levels in skeletal muscle, as shown by the accumulation of autophagosomes in human myopathies caused by genetic deficiency of lysosomal proteins (e.g. Pompe's and Danon's disease) [173,174], or by pharmacological inhibition of lysosomal function, as in chloroquine myopathy [175]. Muscle-specific deletion of the essential autophagy gene Atg7 results in profound muscle atrophy and an age-dependent decrease in muscle force [176,177], suggesting that basal autophagy plays a beneficial role in controlling muscle mass. Moreover, Atg7 null muscles show

accumulation of abnormal mitochondria, sarcoplasmic reticulum distension, disorganization of sarcomere, and formation of aberrant concentric membranous structures [176,177], indicating that basal autophagy in muscle is responsible for the removal of dysfunctional organelles and required for maintaining myofiber integrity. Because of an abnormal sarcoplasmic reticulum, Atg7 null muscles also reveal activation of ER chaperones, such as BiP, and phosphorylation of eIF2a [176,177], suggesting induction of an unfolded protein response. This condition leads to inhibition of protein synthesis, which further contributes to muscle atrophy.

Even though basal autophagy is constitutively active and sufficient to keep myofibers in a healthy state during normal physiological conditions, several studies demonstrate that autophagy is strongly induced in skeletal muscle following oxidative stress [65,178] or in response to severe catabolic conditions such as denervation [179] or fasting [177]. The classical pathway that regulates autophagy in mammalian cells acts through mammalian target of rapamycin (mTOR), a protein kinase that plays a key role as an intracellular nutrient sensor and regulator of protein synthesis, cell growth and metabolism [172]. Surprisingly, mTOR signaling is not the major factor that controls autophagy in adult muscles. Detailed biochemical studies have shown that rapamycin-mediated mTOR inhibition only barely (~10%) increases protein breakdown in differentiated myotubes [180]. Instead, the transcription factor FoxO3 was found to be necessary (and sufficient) to activate autophagy in myotubes. Indeed, FoxO3 controls the expression of many autophagy-related genes, including microtubule-associated protein-1 light chain 3 (LC3) involved in cargo recruitment and biogenesis of autophagosomes as well as Bnip3 [58,59]. Because autophagosome formation is not induced in (fed) mice overexpressing GFP-LC3 [177], LC3 induction per se is not sufficient to trigger autophagy. Most likely, LC3 is continuously consumed when autophagy is activated. In this way, upregulation of LC3 is required to replenish the LC3 protein pool and allows progression of autophagy. In contrast, upregulation of Bnip3 has a major role in mediating the effect of FoxO3 on the autophagic process, as shown by the fact that induction of autophagy is markedly decreased by Bnip3 knockdown [58]. Other autophagy-related genes that are induced by FoxO3 during muscle atrophy are the LC3 homologue Gabarapl1, Atg12l and the lysosomal proteinase cathepsin L [58,59]. The role of cathepsin L upregulation is still unclear. Possibly, cathepsin L is involved in the degradation of muscle-specific proteins, as shown recently for the sialidase Neu2 [181], a protein involved in myoblast differentiation. Unfortunately, and quite similar to autophagy inhibition, excessive activation of autophagy aggravates muscle wasting by removing portions of cytoplasm, proteins and organelles [182]. Therefore, tight control of FoxO3-induced gene expression is needed to avoid unbalanced autophagy induction and muscle loss.

6. Apoptosis and muscle atrophy

Cells undergoing apoptosis are characterized by cytoplasmic and nuclear condensation, oligonucleosomal cleavage of the chromosomal DNA, violent blebbing of the plasma membrane, packaging of the cellular contents into membrane-enclosed vesicles (apoptotic bodies) and elimination of these vesicles by heterophagocytosis [183,184]. Changes in membrane fluidity and ionic charge, altered behaviour of sugar chains or lipids, and conformational changes in membrane proteins are also likely to occur [183,184]. The morphological features of apoptosis are mainly the result of a subset of proteolytic enzymes, called

caspases. These proteases are activated by two principal pathways: an intrinsic and an extrinsic one. The intrinsic pathway begins within the cell and is triggered by stress stimuli such as DNA damage. Hallmark events include mitochondrial outer membrane permeabilization (MOMP) and the release of cytochrome C through the permeability transition pore complex. Once released, cytochrome C binds apoptotic protease-activating factor 1 (Apaf-1) to form a complex known as the apoptosome. This complex also recruits procaspase-9, promoting its self-activation. Active caspase-9 in turn cleaves downstream effector caspases such as caspase-3 and caspase-7, which rapidly degrade intracellular substrates. The extrinsic pathway overlaps with the intrinsic pathway, and is activated by ligand binding to specific death receptors on the cell surface. Examples of ligands to death receptors include TNF α , Fas ligand (FasL) and TNF α -related apoptosis-inducing ligand (TRAIL) that bind to the TNF-receptor, Fas or TRAIL receptor, respectively.

There is overwhelming evidence that apoptosis increases in skeletal muscle during normal aging [185,186], and thus may contribute to sarcopenia [187] (Fig. 1). First, the mitochondrion-mediated (intrinsic) signalling pathway is activated in aged skeletal muscle. Indeed, the content of several Bcl-2 family members (Bax and Bcl-2) is altered with age, expression of Apaf-1 is enhanced, and the presence of cleaved caspases increases. There is also evidence that caspase-independent mechanisms may contribute to apoptosis through elevated nuclear levels of endonuclease G in aged muscle as well as raised expression levels of apoptosis inducing factor (AIF). Secondly, receptor-mediated apoptotic signalling may be activated in aging muscle, as shown by increased activation of caspase-8, increased expression of FADD, and increased mitochondrial content of tBid. These events most likely occur in response to elevated circulating levels of TNF α . Thirdly, p53 appears to be elevated, although it is unclear whether this may lead to apoptotic cell death as some studies have shown elevated cytosolic levels while others have seen increases in nuclear p53.

Apart from aging, apoptosis might contribute to muscle atrophy in various other conditions including cancer cachexia [188], CHF [189], muscle denervation [190], dystrophies [191], sepsis [192], hindlimb unweighting [193], muscle disuse [194] and burn injury [195]. However, with regard to interpretation of apoptosis data in muscle, several critical issues should be mentioned. Since skeletal myocytes are multinucleated, apoptosis of a particular nucleus may not equate to apoptosis of the entire muscle fiber. It is not yet known how many nuclei must be lost before the entire fiber undergoes apoptosis or how this process is regulated in skeletal muscle. Therefore, it has been suggested that nuclear apoptosis in skeletal muscle does not necessarily translate into death of the myofiber [186]. On the other hand, the classical terminal deoxynucleotidyl transferase end-labeling (TUNEL) technique is frequently used to detect DNA fragmentation during apoptosis, even though this method lacks specificity as it may also detect single-stranded DNA or RNA breaks. Accordingly, false positive TUNEL reactions can be observed in living muscle cells during active gene transcription [196]. Other difficulties in interpreting cell death data in atrophying muscle stem from the observation that caspase-3 is not only involved in apoptosis, but also in myofibrillar protein degradation [197]. As occurs for calpains, caspase-3 might be involved in the release of myofibrillar proteins targeted to proteasome degradation. Also of note, caspase-3 enzyme activity has been reported to be elevated in aging muscle [198], while others have found no change in activity [185]. This discrepancy cannot simply be explained by differences in muscle type, age or animal strain, although differential expression of some anti-apoptotic proteins such as XIAP may help to understand contradictory study results [186].

7. Therapeutic inhibition of muscle atrophy

It is beyond the scope of this paper to extensively discuss the multitude of therapeutic approaches that have been explored until now to counteract or prevent muscle atrophy and excellent reviews on this subject have been published [199,200]. Given that muscle wasting is a complex systemic disorder, it is likely that multiple actions are required to prevent its development and progression. Ideally, a multi-faceted approach involves adequate nutritional support, the implementation of exercise training to stimulate muscle hypertrophy and, hopefully one or more pharmacological compounds that specifically, or because of pleiotropic actions, antagonize “pro-atrophic” pathways (Table 1).

7.1. Malnutrition and weight loss

Patients who suffer from life-threatening diseases are characterized by anorexia. Nevertheless, even bypassing the obstacle of reduced appetite by introducing enteral nutrition, or – if inevitable - total parenteral nutrition, does not reverse the wasting process. Therefore, nutritional optimisation should be considered as a supportive measure in malnourished patients. Deficiencies in micronutrients, vitamins, amino acids and electrolyte abnormalities need correction. For instance, previous observations in yeast suggested that branched-chain amino acids (BCAAs), leucine, isoleucine, and valine, might be potential candidates in promoting survival [201]. BCAAs increase the activity of mTORC1 (mTORC1; mTOR in complex with raptor), a key regulator of protein synthesis and cell growth [202], thus favouring cell oxidative capacity [203] and PGC-1 α -mediated mitochondrial gene expression [204]. In addition, long-term dietary supplementation with a specific BCAA-enriched amino acid mixture reduced oxidative damage, both in cardiac and skeletal muscles of middle aged mice, increasing average lifespan [205]. As such, BCAAs have been reported to have anti-cachectic effects, presumably resulting from interference with the UPS [206]. Moreover, in vitro experiments indeed demonstrated suppressed expression of MaFbx/atrogen-1 mRNA in a C2C12 cell line [207]. Fish oil derived N-3 polyunsaturated fatty acids (PUFAs) may have anti-catabolic effects through anti-inflammatory actions [208,209]. Despite their long term application on empirical grounds, appetite stimulants such as anti-histaminic drugs, as well as orally active progestational agents (megestrol acetate and medroxyprogesterone acetate) are still evaluated in clinical trials and have been approved for certain indications. The associated tendency for fluid retention limits their use in patients with CHF.

The orexigenic hormone ghrelin as a potential means to target cachexia has recently received considerable attention [210]. The effects of synthetic human ghrelin, as well as ghrelin receptor agonists, are being evaluated in several trials. Ghrelin, predominantly produced by the stomach, increases food intake through its actions on the hypothalamus [211]. In addition, serving as a growth-hormone secretagogue and mainly mediating its effects via IGF-1, ghrelin also has anabolic effects. Both GH and insulin resistance have indeed been demonstrated in cachexia-related conditions.

Angiotensin II directly induces muscle protein catabolism through the ubiquitin-proteasome proteolytic pathway and may play a role in cancer cachexia, as well as in CHF patients [212]. Chronic heart failure

patients treated with enalapril in the SOLVD study were less likely to develop weight loss [213]. Reduced energy expenditure and weight increase are seen as an undesirable side-effect of beta-blockers in general. In CHF patients, similar post-hoc observations in large beta-blocker trials, have confirmed that antagonising high catecholamine levels increases body weight.

7.2. Compounds with anabolic activity

In patients with CHF, an overall depletion of circulating anabolic hormones (i.e.; circulating total testosterone and free testosterone, dehydroepiandrosterone sulfate, and IGF-1) is associated with poor outcome [214]. Male hypogonadism is also often found in patients with cancer cachexia. In CHF patients, replacement therapy with testosterone did not seem to impact muscle bulk, but elicited a significant improvement in functional status [215]. The positive effects of anabolic steroids, as well as testosterone administration, on protein synthesis, have to be weighted against possible hazards, such as hepatic toxicity, virilising effects, fluid retention and - in the case of testosterone - the development of prostate complications and malignancy. Further development and testing of non-steroidal androgen receptor modulators (SARMs) might lead to useful alternatives [216], as indeed observed in the case of ostarine [217] and enobosarm [218].

Several cachectic-prone diseases represent a state of GH resistance: high levels of GH and inappropriately low circulating and locally expressed IGF-1, which is considered the main GH mediator [219]. Whereas studies with GH have yielded disappointing results, a more comprehensive approach might be the direct administration of IGF-1. By inhibiting FoxO-1, leading to downregulation of the ubiquitin E3 ligases Murf-1 and MAFbx, skeletal proteosomal protein degradation is reduced, whereas, mainly through the PI3K/Akt pathway, synthesis is stimulated [54,56]. Recent experimental evidence suggests that IGF-1 supplementation might be a viable option to treat cancer cachexia [220].

Beta-adrenergic agonists inhibit both Ca²⁺ dependent proteolysis, and the FoxO mediated transcription of E3 ubiquitin ligases. Despite their effective promotion of muscle hypertrophy, their side-effects, especially short (tachycardia) as well as long-term (cardiac hypertrophy) cardiovascular risks, preclude current application [221]. In animal models, the newer generation beta-agonist, formoterol, elicited an anabolic response in skeletal muscle at very low doses, and might therefore be pursued in future investigations [222].

7.3. Anti-inflammatory drugs

Increased levels of pro-inflammatory cytokines, both in the circulation, as well as locally at the level of the skeletal muscle, have been attributed a key role in the pathophysiology of cachexia and muscle wasting. Hence, the release of potent monoclonal TNF α -antibodies opened a new avenue of possible applications. Although not specifically studied as anti-cachectic drugs, CHF trials with etanercept [223] and infliximab [224] were prematurely stopped because of futility and toxicity, respectively. Nevertheless, these drugs are now successfully used to achieve disease control in patients with rheumatoid arthritis and Crohn's disease. In addition, their potential anti-cachectic effects are still

actively explored in the setting of COPD and cancer. The use of interleukin-6 monoclonal antibodies is still in its early stages. Since a whole cascade of pro- and anti-inflammatory adaptations characterizes cachexia-associated wasting, the “single target approach” of specific cytokine antibodies has been scrutinized. A more general anti-inflammatory approach, as well as the use of so-called “immunomodulators” could be of interest. Cyclooxygenase-2 inhibitors may be beneficial in counteracting muscle wasting. In patients with cancer cachexia, celecoxib indeed led to a significant increase of lean body mass and muscle strength [225]. Thalidomide, pentoxifylline and statins all exert anti-inflammatory effects, but have, until now, not been specifically investigated in cachectic patients. Despite their potent anti-inflammatory effects, the successful application of 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors in cardiovascular disease was not confirmed in clinical CHF trials [226]. Interestingly, low cholesterol levels, often already found in cachectic patients, may offset the pleiotropic characteristics of statins. Indeed, several in vitro experiments [227] support the neutralising effects of lipopolysaccharide by cholesterol micelles. The latter may at least partially explain for the unexpectedly improved outcome in CHF patients with elevated plasmalipids [228].

7.4. Modulating the energetic crisis of skeletal muscle

Together with GH resistance and downregulation of IGF-1, insulin resistance contributes to the anabolic/catabolic imbalance [229]. The exact mechanisms underlying the initiation of insulin resistance and its relation to muscle wasting remain to be further explored. In CHF patients, increased plasma fatty acids, pro-inflammatory cytokines, as well as local muscle adiponectin resistance, may culminate into an insulin resistant state. The adipocyte-derived cytokine adiponectin exerts beneficial insulin-sensitising and anti-inflammatory effects in situations of insulin resistance, such as obesity and diabetes [230]. Paradoxically, increased adiponectin levels infer poor prognosis in several chronic conditions, such as in the setting of CHF [231] and renal failure [232]. In skeletal biopsies taken from CHF patients, adiponectin expression was significantly upregulated, whereas the reverse was seen for the skeletal muscle adiponectin receptor AdipoR1 [233]. Skeletal muscle metabolic deficiency in CHF was further corroborated by the finding of a deactivated PPAR α /AMPK pathway and a concordant down-regulation of several target genes, involved in the metabolism of free fatty acids and glucose. Interestingly, these changes were reversible with the introduction of a 4 months exercise training protocol [234]. Targeting insulin resistance by using insulin sensitizer such as thiazolidinediones [235], as well as the application of PPAR α agonists have been suggested to beneficially affect protein energy metabolism [236].

7.5. Specific proteasome inhibitors

Several compounds designed to act as proteasome inhibitors such as the peptidyl aldehydes carbobenzoxyl-Ile-Glu(O-t-butyl)-Ala-leucinal (PSI), N-acetyl-leucyl-leucyl-norleucinal (LLN) or CBZ-leucyl-leucyl-leucinal (MG132) have helped immensely in understanding the biological role and importance of the ubiquitin-proteasome system during muscle wasting. These compounds reduce the accelerated proteolysis in atrophying muscle and preserve muscle fiber diameter [237-241]. Moreover, inhibition of the proteasome machinery prevents the degradation of I κ B, which maintains NF- κ B in its inactive state,

thus preventing the upregulation of MuRF1 and Mafbx/atrogen-1 as well as the synthesis of TNF α , IL-1 and IL-6 [239,240]. Other beneficial effects are restoration of Akt phosphorylation, decreased apoptosis, enhanced locomotive activity and shortening of the rehabilitation period [239,240]. Besides peptide aldehyde inhibitors, the N-protected dipeptide Pyz-Phe-boroLeu (also known as PS-341, bortezomib or Velcade) is frequently used to inhibit nonlysosomal protein breakdown. It is a potent proteasome inhibitor and approved by the American Food and Drug Administration for treatment of multiple myeloma. Even though bortezomib can prevent excessive protein degradation and muscle wasting in animal models [242,243], its use is not yet approved in patients for treatment of muscle atrophy. Indeed, clinical trials with bortezomib for the treatment of cancer cachexia have not shown univocal results [244]. In this light, it is noteworthy that only a partial reduction of denervation-induced atrophy was observed in rat soleus muscle after treatment with bortezomib [242]. This partial effect could be due to other proteolytic mechanisms that are not inhibited by bortezomib, such as lysosomal pathways, which are also involved in muscle wasting (vide supra). Peptide aldehyde inhibitors such as MG132 can repress certain lysosomal cysteine proteases and calpains, which might explain the total rescue of muscle mass in some studies [240].

7.6. Exercise training – an all-in-one approach?

The multiple effects of exercise training in reversing atrophy and – especially – the hypertrophic stimulus of resistance exercises are well known. Regular physical activity, for instance, in patients with CHF, has local and general anti-inflammatory [245,246] and anti-oxidative effects [247], decreases insulin and adiponectin resistance [234], has a quiescent influence on the over-activated UPS [248] and reduces skeletal muscle apoptosis [247]. The resulting improvement in terms of physical capacity and quality of life in CHF and COPD patients has stimulated investigators to explore this treatment modality in other conditions, such as cancer, renal failure and rheumatoid arthritis. Very recent experimental data suggest that in pressure overload-induced heart failure [249], myostatin, produced in the heart itself, translates into remote effects of peripheral muscle wasting. These data are somewhat contradictory to findings from an earlier study, using a different heart failure model (LAD ligation), in which local skeletal muscle myostatin upregulation was shown [250]. Whereas targeted inhibition of myostatin, by using myostatin antibodies or activin receptor IIB blockers might be a future therapeutic option, it is interesting to mention that exercise training of animals following LAD ligation significantly downregulated skeletal muscle and myocardial myostatin protein overexpression [251]. In that same study, using a C2C12 cell line, TNF α induced the expression of myostatin through a p38MAPK-dependent pathway involving NF- κ B.

Very recently, a combined approach of eicosapentaenoic acid (EPA) and exercise training has been tested on Lewis lung carcinoma-bearing mice [252]. While EPA alone did not prevent the muscle loss induced by tumor growth, the combination with exercise induced a partial rescue of muscle strength and mass in cachectic animals, as corroborated by reduced PAX-7 accumulation and increased PCG-1 protein [252].

8. Conclusions

Muscle atrophy may arise as a consequence of many different physiological and pathological processes. Regardless of these underlying conditions, counteracting loss of muscle mass is a major issue that should be addressed in order to increase the quality of life in severely ill patients. Based on research conducted in different areas, it is strongly suggested that a plethora of combinations, comprising unbalanced triggers and activated or deactivated pathways, eventually culminate into a hypercatabolic status. These observations clearly lead up to the necessity of elaborating a multi-faceted therapeutic approach, that involves adequate nutritional support, the implementation of exercise training to stimulate muscle hypertrophy and, one or more pharmacological compounds that specifically antagonize pro-atrophic pathways. Because of the complexity of each disease and the variation of skeletal muscle wasting among patients, the clinical approach needs to be personalized in order to obtain a targeted therapy and to avoid dangerous side-effects.

Authors' contributions

AF, VC, FP and WM contributed equally to this paper. All authors read and approved the final manuscript.

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Conflicts of interest

The authors confirm that there are no conflicts of interest.

Fig. 1: Pathophysiological events associated with muscle atrophy.

Table 1: Overview of different therapeutic approaches employed to counteract muscle wasting.

Therapeutic approach	Class	Molecule	References
Dietary supplementation			
	BCCAs	Leu, Ile, Val, Hmb	[206,207]
	PUFAs	EPA, DHA	[208,209]
	Appetite stimulants, ghrelin	Anti-histaminic drugs,	progestational agents, [210]
	ACE inhibitors	Enalapril	[213]
Compounds with anabolic activity		Steroidal androgens	Testosterone [215]
	SARMs	Ostarine	
and enobosarm			[216-218]
	Growth factors	IGF-1, insulin	[220]
	Beta2-agonist	Formoterol	[222]

Anti-inflammatory drugs

TNF α antibodies Etancercept, infliximab [223,224]

Cyclooxygenase-2 inhibitors Celecoxib [225]

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adipocyte-derived cytokine Adiponectin (diabetes and obesity) [230]

Insulin sensitizer Thiazolidinediones [235]

PPAR α agonists Fibrates [236]

Muscle wasting inhibitors

Proteasome inhibitors PSI, LLN, MG132 [237-241]

PS-341, bortezomib or Velcade [242-244]

Exercise training PRT PRT, endurance [234,245-247]

PRT combined with therapeutic strategies PRT and Eicosapentaenoic acid [252]

Table abbreviations: ACE, angiotensin converting enzyme; BCCAs, branched chain aminoacids; Hmb, β -hydroxy β -methylbutyrate; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; IGF-1, insulin-like growth factor 1; PPAR α , peroxisome proliferator-activated receptors alpha; PRT, regular physical training; PUFAs, polyunsaturated fatty acids; SARMs, non-steroidal androgen receptor modulators; TNF α tumor necrosis alpha.

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Figure 1

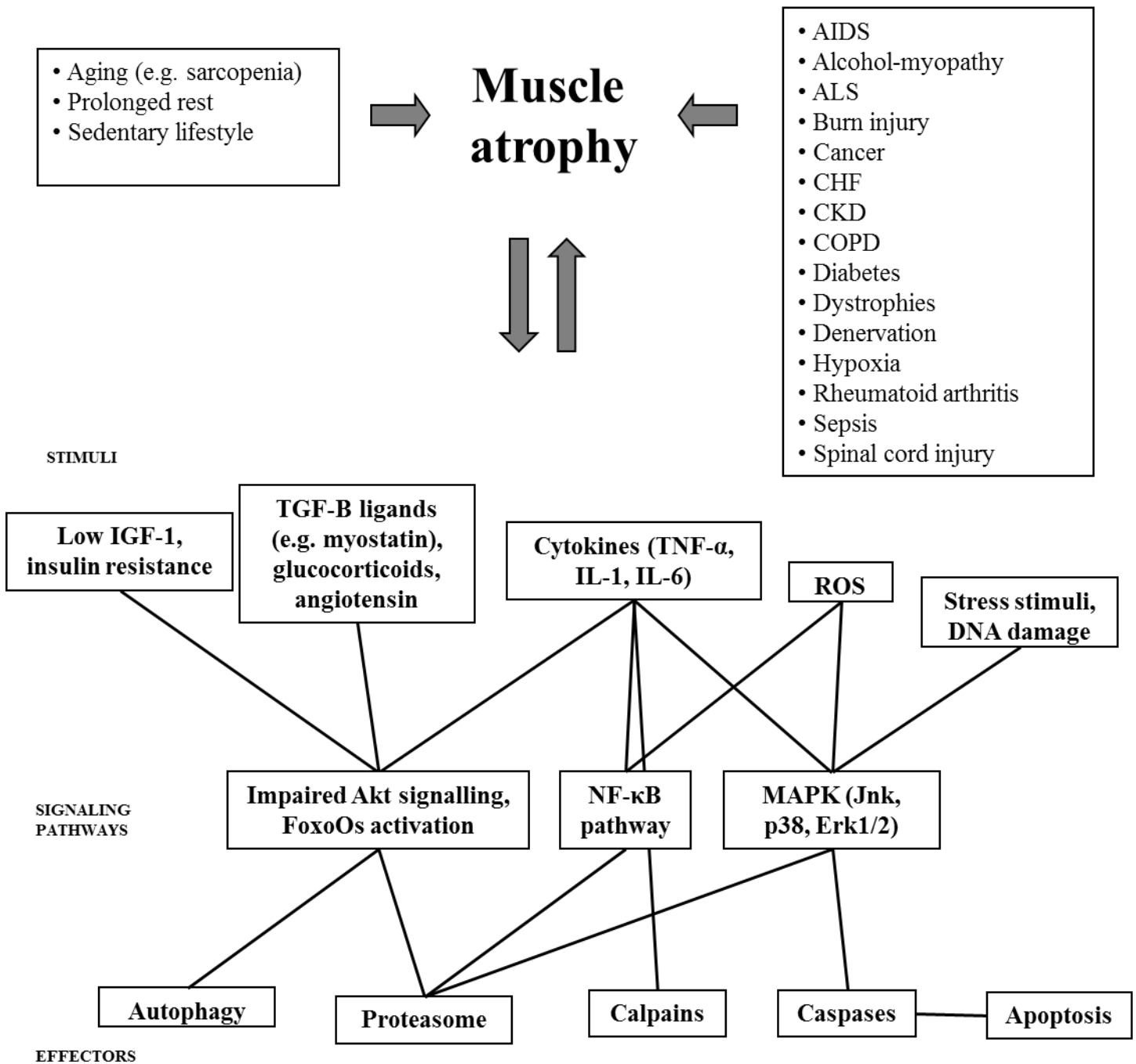


Fig. 1: Pathophysiological events associated to muscle atrophy.