

## UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

Avalle L, Regis G, Poli V. Universal and Specific Functions of STAT3in Solid Tumours Editor: Springer-Verlag 2012 ISBN: 9783709108901

in

Decker T, Muller M Jak-Stat Signaling: From Basics to Disease 305 - 333

The definitive version is available at: http://www.springerlink.com/index/pdf/10.1007/978-3-7091-0891-8\_17

# Universal and Specific Functions of STAT3 in Solid Tumours

Lidia Avalle\*, Gabriella Regis\*, and Valeria Poli

#### Abstract

STAT3 is constitutively activated in a high percentage of tumours and tumourderived cells of both liquid and solid origin, often correlating with aggressive disease and bad prognosis. Persistent STAT3 activity, to which tumours often become addicted, is mostly due to the aberrant activation of pro-oncogenic/proinflammatory signals that can trigger its phosphorylation, such as oncogenes, growth factor receptors and cytokines. Among STAT3-mediated functions are increased survival and proliferation, enhanced angiogenesis, motility and invasion, and down-modulation of anti-tumour immune responses. Moreover, STAT3 was recently shown to play unexpected roles in regulating cell metabolism and mitochondrial activity via both transcriptional and non-transcriptional mechanisms. Here, we review the main knowledge about the role of STAT3 in solid tumours, with a particular focus on breast cancer and our recent work with mouse models.

#### Introduction

Signal Transducers and Activators of Transcription (STAT) factors mediate the signalling downstream of cytokine and growth factor receptors, and often their activity is deregulated in cancer (Turkson and Jove 2000; Siddiquee et al. 2007). Once activated by tyrosine-phosphorylation via receptor-associated JAK kinases,

<sup>\*</sup>These two authors equally contributed to this work.

L. Avalle • G. Regis • V. Poli (🖂)

Molecular Biotechnology Center (MBC) and Department of Genetics, Biology and Biochemistry, University of Turin, Via Nizza 52, Turin 10126, Italy e-mail: valeria.poli@unito.it

STATs form parallel dimers that concentrate into the nucleus regulating the expression of target genes (Schindler et al. 2007). The family member STAT3 can be activated by a wide variety of cytokines and growth factors (e.g. IL-6 family, leptin, IL-12, IL-17, IL-10, Interferons, G-CSF, EGF, PDGF) and by a number of oncogenes such as Src, Abl, Sis, Fps, Ros, Met and ErbB2 (Turkson and Jove 2000). Accordingly, STAT3 is found to be constitutively tyrosinephosphorylated in a high percentage of tumours and tumour-derived cell lines of both liquid and solid origin, which often become addicted to its activity for continuous survival and growth (Yu et al. 1995; Turkson and Jove 2000; Kortylewski et al. 2005; Siddiquee et al. 2007), and is considered a good target for anti-cancer therapy. Indeed, STAT3 tyrosine phosphorylation and consequent transcriptional activation was shown to be required for cell transformation downstream of several oncogenes, the prototype being v-Src (Yu et al. 1995; Bromberg et al. 1998; Silva 2004). Although a unique core activity determining addiction to STAT3 by a wide spectrum of biologically distinct tumors has still not been identified, STAT3-mediated gene expression signature is mostly consistent with tumour cell survival and proliferation (Pensa 2008; Yu et al. 2009). In addition, STAT3 constitutive activity in tumour cells can down-modulate antitumour immune responses (Yu et al. 2009) as well as promote tumour angiogenesis (Niu et al. 2002a). STAT3 can also regulate cell movement, contributing to cytoskeleton reorganization and controlling cell adhesion properties, and is thought to play a role in tumour invasion and metastasis by inducing the expression of matrix metalloproteinases (MMP) and promoting the epithelial to mesenchymal transition (EMT) (Pensa 2008; Yu et al. 2009). Finally, recent work has shown that STAT3 acts as an important regulator of cell metabolism, promoting aerobic glycolysis and downregulating mitochondrial activity via its canonical, nuclear functions (Demaria et al. 2010) while preserving mitochondrial respiratory activity via mitochondrial localization of its serinephosphorylated form (Gough et al. 2009; Wegrzyn et al. 2009). Both activities contribute to tumor transformation downstream of distinct signals, which promote STAT3 phosphorylation on either tyrosine or serine. The pro-oncogenic role of STAT3 was first directly demonstrated in vitro by the finding that overexpression of the constitutively active mutant form STAT3C can transform fibroblasts and epithelial cells (Bromberg et al. 1999; Dechow et al. 2004) followed by in vivo experiments in transgenic or knock-in mice demonstrating oncogenic potential in the lung, skin and breast (Li et al. 2007; Chan et al. 2008; Barbieri et al. 2010a).

Many aspects of STAT3 biology in tumours have been extensively reviewed. Here, we chose to summarize the main knowledge about the role of STAT3 in solid tumours, with a particular focus on breast cancer and our recent work with mouse models (Barbieri et al. 2010a, b; Demaria et al. 2010).

The solid tumours where STAT3 has been found to be constitutively active, as well as its main functions and target genes are sketched in Table 1 and detailed below.

Table 1 How ac	stivated STAT3 affects different aspect	cts of tumorigene	sis. The table highlights the exp	erimental findings concerning the role of activated
STAT3 in progn indicating in which	osis, proliferation/survival, metastasi ch model system the data were obtain	is/angiogenesis, c	lrug resistance, immune respon 3 target genes identified in each	se, cancer stem cells in different solid tumours, system
Tumor	Role in tumorigenesis <sup>a</sup>	Model <sup>b</sup>	Target genes	References
Epidermal non-	Proliferation/survival	Mouse	Ciclin D1, Bcl-xL, c-Myc	Chan et al. (2004a, b, 2008)
melanoma tumours	Metastasis/angiogenesis	Cell, clinic.	bFGF	Jee et al. (2004), Suiqing et al. (2005)
Melanoma	Correlation with prognosis not evaluated (90%)	Clinic.		Niu et al. (2002a)
	Proliferation/survival	Cell	Bcl-xL, Mcl1	Niu et al. (2002a)
	Metastasis/angiogenesis	Cell	MMP2, VEGF, bFGF	Niu et al. (2002a), Xie et al. (2004, 2006)
	Immune response	Cell		Wang et al. (2004)
Head and neck cancer	Poor prognosis/highest expression in early stages (40–80%)	Clinic.		Masuda et al. (2002b), Nagpal et al. (2002)
	Proliferation/survival	Cell	Cyclin D1, Bcl-xL, Bcl2	Masuda et al. (2002a, b, 2007)
	Metastasis/angiogenesis	Cell	VEGF	Masuda et al. (2002a, 2007), Lui et al. (2009)
	Drug resistance	Cell		Masuda et al. (2002c), Gu et al. (2010)
	Immune response	Cell		Albesiano et al. (2010)
	Cancer stem cells	Clinic.		Chen et al. (2008b)
Colorectal	Poor prognosis, metastasis (50%)	Clinic.		Kusaba et al. (2005, 2006)
carcinoma	Proliferation/survival	Cell, clinic.	Bcl-xL, survivin, miR-21, miR-181b1	Turkson et al. (2004), Lassmann et al. (2007), Iliopoulos et al. (2010)
	Metastasis	Cell	MMP1, MMP3	Tsareva et al. (2007)
	Immune response	Cell		Nefedova et al. (2004), Wang et al. (2004)
Hepatocellular	Poor prognosis (60–75%)	Clinic.		Feng et al. (2001), Zhang et al. (2010)
carcinoma	Proliferation/survival	Mouse	Bcl-xL, cyclin D1	Yoshida et al. (2002)
	Metastasis/angiogenesis	Cell	VEGF, MMP2	Li et al. (2006)
				(continued)

iversal and Specific Functions of STAT3 in So

Table 1 (continu	ued)			
Tumor	Role in tumorigenesis <sup>a</sup>	Model <sup>b</sup>	Target genes	References
Pancreatic	Proliferation/survival	Cell		Wei et al. (2003)
cancer	Metastasis/angiogenesis	Cell	VEGF	Wei et al. (2003)
	Immune response	Cell		Bharadwaj et al. (2007)
Gastric	Poor prognosis (30%)	Clinic.		Lee et al. (2009)
carcinoma	Proliferation/survival	Cell, clinic.	Survivin, Bcl2	Kanda et al. (2004), Choi et al. (2006)
	Angiogenesis	Clinic.	VEGF	Choi et al. (2006)
Ovarian cancer	Poor prognosis $(90\%)$	Clinic.		Rosen et al. (2006)
	Proliferation/survival	Cell	Cyclin D1, Bcl-xL	Huang et al. (2000)
	Metastasis/angiogenesis	Cell		Silver et al. (2004), Nilsson et al. (2005)
	Drug resistance	Cell	Bcl-xL	Burke et al. (2001)
Prostate cancer	Highest malignancy (80%)	Clinic.		Mora et al. (2002)
	Proliferation/survival	Cell	Bcl2, cyclin D1, c-Myc, Bcl-xL, Mcl1	Lou et al. (2000), Gao et al. (2001, 2005), Flowers et al. (2005), Hellsten et al. (2008)
	Metastasis/angiogenesis	Cell	HIF1, VEGF, E-cadherin	Xu et al. (2005), Azare et al. (2007), Abdulghani et al. (2008)
	Drug resistance	Cell	Bcl-xL (partially)	Pu et al. (2004)
	Cancer stem cells	Cell		Mathews et al. (2010)
Lung carcinoma	Poor prognosis but not significant (70%)	Clinic.		van Cruijsen et al. (2009)
	Proliferation/survival	Cell	Bcl2, Bcl-xL, Mcl1, survivin, cyclin D1, c-Myc	Weerasinghe et al. (2007)
	Metastasis/angiogenesis	Cell	VEGF	Weerasinghe et al. (2007), Pfeiffer et al. (2009)
	Immune response	Mouse	Ccl5, IL-6, VEGF	Li et al. (2007)
Glioblastoma	Correlation with prognosis not evaluated (95%)	Clinic.		Rahaman et al. (2002)
	Proliferation/survival	Cell, clinic.	Bcl-xL, Mcl1, Bcl2	Rahaman et al. (2002), Sherry et al. (2009)
	Metastasis/angiogenesis	Cell	MMP9; VEGF	Loeffler et al. (2005), Liu et al. (2010)
	Immune response	Cell		Hussain et al. (2007)
	Cancer stem cells	Cell		Sherry et al. (2009), Villalva et al. (2010)

308

Rabdo-	Proliferation/survival	Mouse, cell	Bcl-xL	Lee et al. (2006), Chen et al. (2007b)
myosarcoma	Immune response	Cell		Nabarro et al. (2005)
Renal carcinoma	Poor prognosis, metastasis (100%)	Clinic.		Horiguchi et al. (2002b)
	Proliferation/survival	Cell	Survivin, Bcl-xL, Bcl2, cyclin E; VEGF	Horiguchi et al. (2002a), Xin et al. (2009)
	Angiogenesis	Cell	VEGF	Jung et al. (2007), Xin et al. (2009)
	Immune response	Cell		Xin et al. (2009)
Cervical cancer	Poor prognosis (60%)	Clinic.		Takemoto et al. (2009)
	Proliferation/survival	Clinic.	Bcl-xL, survivin, Mcl1	Chen et al. (2007a)
	Angiogenesis	Clinic.	VEGF	Wei et al. (2003)
Bladder cancer	Proliferation/survival	Cell	Bcl2, Bcl-xL, survivin, cyclin D1	Chen et al. (2008a)
	Metastasis	Cell	MMP1	Itoh et al. (2006)
Breast cancer	Better prognosis of node- negative breast cancer; metastasis in regional lymph node; poor response to therapy (30–60%)	Clinic.		Dolled-Filhart et al. (2003), Dechow et al. (2004), Hsieh et al. (2005), Diaz et al. (2006)
	Proliferation/survival	Cell	TIMP3	Garcia et al. (2001), Selander et al. (2004)
	Metastasis/angiogenesis	Cell	VEGF	Niu et al. (2002a), Selander et al. (2004)
	Drug resistance	Cell	Bcl2	Real et al. (2002)
	Immune response	Cell	SOCS3	Sun et al. (2006)
	Cancer stem cells	Cell		Zhou et al. (2007)
<sup>a</sup> The percentage r <sup>b</sup> Model: <i>mouse</i> m	efers to the clinical samples where ac ouse model, <i>cell</i> cell lines, <i>clinic</i> . clin	ctivated STAT3 we nical samples	as detected in the cited reference,	/s

Universal and Specific Functions of STAT3 in Solid Tumours

#### **Epidermal Non-Melanoma Tumours**

STAT3 constitutive activation was observed in several types of human epidermal non melanoma cutaneous tumours, correlating with poor differentiation, tumour invasion and metastasis in clinical samples of cutaneous squamous cell carcinoma (SCC) (Suiqing et al. 2005). STAT3 involvement with metastatic potential was also confirmed in xenograft experiments of basal cell carcinoma cells (BCC) over-expressing IL-6, where IL-6-mediated angiogenesis supported tumour development in part via STAT3 activation (Jee et al. 2004).

The two-stage chemical carcinogenesis model is considered a good model of epithelial carcinogenesis, recapitulating the different phases from tumor initiation to progression (Chan et al. 2004a). The first experimental evidence that STAT3 activation is required for epithelial tumorigenesis in vivo was obtained using this model, where STAT3 ablation in keratinocytes completely abrogated skin tumour development (Chan et al. 2004b). STAT3-deficient keratinocytes were more sensitive to DMBA-induced apoptosis and STAT3 inhibition with an oligonucleotide decoy injected into primary skin papillomas led to significant reduction of tumour volume. STAT3 was also implicated in ultraviolet B (UVB)-induced skin carcinogenesis by the observation that UVB irradiation promoted proliferation and survival of keratinocytes via STAT3 activation (Sano et al. 2005). Conversely, transgenic mice overexpressing the constitutively active form STAT3C in keratinocytes developed skin tumours with a shorter latency and in greater number compared to non-transgenic mice (Kim et al. 2007; Chan et al. 2008). STAT3C acted in both tumour initiation and promotion by upregulating genes involved in survival, proliferation, angiogenesis and metastasis.

One key step in STAT3-mediated epithelial tumorigenesis may be the induction of Bcl-xL, which plays a fundamental role in early skin carcinogenesis by enhancing survival of keratinocyte stem cells in the bulge region of the hair follicle, where mutations are believed to arise during the initiation stage and whose clonal expansion occurs during tumour promotion, as shown by studies in Bcl-xL deficient mice (Kim et al. 2009).

#### Melanoma

STAT3 is constitutively active in the vast majority of melanoma tumours and cell lines (Niu et al. 2002a), most often downstream of activated c-Src (Kortylewski et al. 2005), where it favours proliferation, escape from apoptosis and angiogenesis via induction of Bcl-xL, Mcl1 and VEGF (Niu et al. 2002a; Xie et al. 2006). STAT3 activation is apparently crucial to promote the metastatic process in melanoma, through direct upregulation of MMP2, VEGF and bFGF expression (Xie et al. 2004, 2006). STAT3 activation was shown to promote an immunosuppressive environment leading to impaired dendritic cell (DC) maturation and tumour-specific T cell response in melanoma B16 cells (Wang et al. 2004). Accordingly WP1066, a STAT3 inhibitor that blocks melanoma cells growth, was found to interfere with melanoma brain metastasis by inhibiting the production of immunosuppressive cytokines such as TGF-b, MCP1, RANTES and VEGF by tumour cells, thus enhancing cytotoxic T lymphocyte responses and inhibiting T regulatory (Treg) cells differentiation (Kong et al. 2008).

#### **Head and Neck Cancer**

STAT3 constitutive activation has been observed in 40–80% of human head and neck squamous cell cancer (HNSCC), correlating with poor prognosis and with proliferation and apoptosis resistance via induction of cyclin D1, Bcl2 and Bcl-xL expression (Masuda et al. 2002b; Nagpal et al. 2002). Apoptosis could be reinstated and tumour growth blocked by inhibiting STAT3 activity (Leong et al. 2003; Jing et al. 2006). Therapeutic blockade of STAT3 activity also resulted in impaired angiogenesis due to direct STAT3-mediated regulation of VEGF expression (Masuda et al. 2007). STAT3 activation occurring upon EBV infection or deregulated EGFR signalling in nasopharingeal cancer promotes anchorage-independent growth and invasion (Lui et al. 2009; Wheeler et al. 2010). High STAT3 phosphorylation levels in cells from HNSCC patients are associated with the expression of CD44 and aldehyde dehydrogenase 1 (ALDH1) stem cell markers and with typical features of Cancer Stem Cells (CSCs) such as high tumorigenicity, radioresistance, expression of the stemness markers Bmi, Oct4 and Nanog and of the EMT genes Snail and Twist (Chen et al. 2008b). STAT3 inhibition in these cells reinstated responsiveness to chemotherapy, favoured differentiation and impaired tumorigenesis and metastasis formation (Chen et al. 2008b). Moreover, anti-tumour immune responses were affected, with enhanced production of proinflammatory cytokines and chemokines which, in turn, triggered DC activation and lymphocytes migration and prompted anti-tumour immune response (Albesiano et al. 2010).

#### **Colorectal Carcinoma**

STAT3 constitutive activity was observed in colorectal carcinoma cells in about 50% of clinical samples (Kusaba et al. 2005, 2006), correlating with proliferation and tumour growth rate (Becker et al. 2005; Corvinus et al. 2005), and with tumour invasion, lymph node metastasis and poor prognosis (Kusaba et al. 2005, 2006). STAT3-related enhanced invasiveness correlates with strong expression of the MMP1, -3, -7, and -9. MMP1 and MMP3 are direct STAT3 targets, and activated STAT3 colocalizes with MMP1 in tumour specimens (Tsareva et al. 2007). Importantly, both NF-kB and STAT3 activation have been shown to be crucial for inflammation-driven colon carcinogenesis (Greten et al. 2004; Bollrath et al. 2009; Grivennikov et al. 2009). NF-kB activation in myeloid cells drives IL-6 expression, whose levels are indeed increased in patients serum, and in turn IL-6 is responsible for STAT3 constitutive activation in colon tumour cells, a paradigm that is thought to hold true for several other inflammation-related tumours (Grivennikov and Karin 2010).

Mice lacking STAT3 in intestinal epithelial cells (IECs) showed an almost complete protection from the development of the AOM/DSS model of colitis-associated cancer (CAC), correlating with decreased epithelial proliferation and enhanced sensitivity to treatment-induced apoptosis (Bollrath et al. 2009; Grivennikov et al. 2009). However, the role of STAT3 in colon tumorigenesis appears to be context-dependent, as we have recently shown in Apc<sup>Min</sup> mice that this factor promotes early tumorigenesis steps but impairs tumour progression at later stages, via regulation of the adhesion molecule CEACAM1 (Musteanu et al. 2010).

STAT3-mediated immune suppression was shown to play an important role in colon cancer cells (Nefedova et al. 2004; Wang et al. 2004), where tumour-derived factors, inducing STAT3 activation in infiltrating immature myeloid cells, prevented their differentiation into mature dendritic cells (Nefedova et al. 2004).

#### **Other Tumours of the Gastrointestinal Tract**

STAT3 constitutive activation plays a role in several other tumours that develop in the gastrointestinal tract. In hepatocellular carcinoma (HCC) its activity is often induced by HCV infection or IL-6 and other inflammatory cytokines and drives tumorigenesis by promoting proliferation, survival (via Bcl-xL and cyclin D1 induction) and anchorage-independent growth (Yoshida et al. 2002; He et al. 2010). STAT3 inhibition in HCC cells impaired growth, angiogenesis and metastasis while enhancing apoptosis and sensitivity to chemotherapy (Li et al. 2006; Choudhari et al. 2007; Sun et al. 2008).

Constitutively activated STAT3 is widely observed also in pancreatic cancer, often downstream of IL-6 or G-CSF, promoting tumour cell growth, metastasis and angiogenesis (Wei et al. 2003) and impairing dendritic cells differentiation and activation (Bharadwaj et al. 2007).

STAT3 phosphorylation is relatively infrequent in gastric carcinoma, but when present it correlates with tumour cell proliferation, survival and angiogenesis via induction of Bcl2, VEGF and survivin expression (Kanda et al. 2004; Choi et al. 2006). The analysis of human gastric cancer specimens identified correlations between STAT3 activation and lymph node metastasis (Deng et al. 2010). Moreover, STAT3 uncontrolled activity can lead to gastric cancer, as shown by the spontaneous development of gastric carcinomas following disruption of the integrity of mucosal epithelium in gp130 knock-in mutant mice (gp130<sup>757F</sup>) that are unable to respond to SOCS3-mediated negative feedback (Tebbutt et al. 2002).

#### **Ovarian Cancer**

Constitutive activation of STAT3 was detected in a high percentage of ovarian cancer cell lines and human tumour specimens (94%) with respect to normal ovary epithelium, correlating with aggressive clinical behavior and tumour progression, and it was shown to enhance proliferation and inhibition of apoptosis through induction of cyclin D1 and Bcl-xL expression, respectively (Huang et al. 2000; Rosen et al. 2006). Moreover, studies in cell lines have shown that enhanced

STAT3 activity and expression contribute to resistance to apoptosis in response to chemotherapeutic drugs (Burke et al. 2001; Duan et al. 2006). STAT3 inhibition leads to decreased Bcl-xL expression, sensitizing tumour cells to chemotherapy-induced apoptosis (Burke et al. 2001). One of the main signals responsible for STAT3 constitutive activation in ovarian cancer is its canonical activator IL-6, which promotes angiogenesis, leading to tumour proliferation and dissemination of malignant cells (Nilsson et al. 2005).

#### **Prostate Cancer**

Phosphorylated STAT3 was detected in the majority of human prostate cancers, correlating with the degree of malignancy (Mora et al. 2002) and with JAK2/IL-6 signalling, which enhances proliferation and survival (Lou et al. 2000; Flowers et al. 2005). STAT3 inhibition lead to apoptosis of tumour cell lines, both in vitro and in vivo, through downregulation of Bcl2, cyclin D1, c-Myc, Bcl-xL and Mcl1 expression (Lou et al. 2000; Jing et al. 2004; Turkson et al. 2004; Gao et al. 2005; Hellsten et al. 2008), and inhibited angiogenesis and tumour growth via downregulation of both HIF1a and VEGF expression (Xu et al. 2005). The expression of the constitutively active form STAT3C in immortalized prostate epithelial cells caused tumour transformation and enhanced cell motility by decreasing E-cadherin level and increasing the number of lamellipodia and stress fibers (Azare et al. 2007), suggesting a role in EMT and metastasis that was subsequently confirmed by the observation of constitutive STAT3 activation in clinical samples of prostate cancer metastasis, where it promoted cell motility by reorganizing the actin and microtubule network (Abdulghani et al. 2008). IL-6 dependent STAT3 activation was shown to contribute to resistance of human prostate cancer cells to chemotherapy (Pu et al. 2004). There appear to be correlations between STAT3 activation, tumour invasion and CSCs. In particular, invasive prostate cancer cells were shown to display promoter methylation patterns reminiscent of those observed in CSCs, with many differentially methylated genes belonging to the IL-6/STAT3 pathway (Mathews et al. 2010). Additionally, STAT3 was shown to interact with the CSC marker SOX1, whose silencing decreased STAT3 activation and in vitro invasiveness (Mathews et al. 2010). Accordingly, IL-6 was recently shown to induce the conversion of prostate non stem cancer cells (NSCCs) into sphereforming CSCs, similar to what observed in breast cancer cells (Iliopoulos et al. 2011a).

#### Lung Carcinoma

About 50–70% of human non-small cell lung carcinomas (NSCLC) and cell lines were shown to display constitutive STAT3 activation, correlating with enhanced proliferation and survival (Song et al. 2003; Haura et al. 2005; van Cruijsen et al. 2009). STAT3 inhibition in these cells lead to decreased expression of a number of

known STAT3 targets (e.g. Bcl2, Bcl-xL, Mcl1, survivin, VEGF, cyclin D1 and c-Myc), thereby promoting apoptosis, impairing proliferation and reducing angiogenesis (Weerasinghe et al. 2007). Interestingly, mutant EGFR forms in primary human lung adenocarcinomas lead to STAT3 activation via IL-6 upregulation (Gao et al. 2007). In contrast with these data, Pfeiffer and colleagues demonstrated that STAT3 constitutive activation is characteristic of primary tumour samples from patients with small cell lung cancer (SCLC) but not from NSCLC, and that blocking STAT3 activation impaired anchorage-independent tumour cell growth, suggesting the implication of STAT3 in the rapid metastasizing phenotype of SCLC (Pfeiffer et al. 2009).

The lung was the first tissue where over-expressed constitutively active STAT3 was shown to play an autonomous pro-oncogenic role, since transgenic expression of the STAT3C mutant form in alveolar type II epithelial cells induced lung bronchoalveolar adenocarcinomas preceded by remarkable infiltration of inflammatory cells (Li et al. 2007). Tumour development correlated with enhanced secretion of pro-inflammatory molecules and with reactivation of genes critical for epithelial cell growth during embryonic lung development, similar to what observed in human bronchoalveolar adenocarcinomas (Li et al. 2007). Accordingly, STAT3 downstream genes were proposed to serve as biomarkers in human lung adenocarcinoma and chronic obstructive pulmonary disease, which are both induced by chronic inflammation of the lung (Qu et al. 2009).

#### Glioblastoma

High levels of STAT3 activation are also detected in about 95% of glioblastoma cell lines and tumour samples, inducing proliferation and apoptosis resistance through upregulation of Bcl-xL, Mcl1 and Bcl2 expression (Rahaman et al. 2002), and promoting angiogenesis, invasion and metastasis via upregulation of VEGF and MMP9 expression (Loeffler et al. 2005; Liu et al. 2010). Hypoxia resistance is a common feature of both stem cells and CSCs, which are thought to act as tumour initiating cells (TICs) in different types of tumours, including glioblastoma (Hemmati et al. 2003; Zhou and Zhang 2008). Resistance of these cells to chemotherapy is often responsible for relapses and/or metastasis (Villalva et al. 2010). The highly hypoxic glioblastoma microenvironment triggers STAT3mediated induction of VEGF, HIF1, MMP2 and Twist1, which in turn promote angiogenesis and tumour invasion (Kang et al. 2010). Interestingly, STAT3 activation was shown to be essential for glioblastoma stem cells proliferation and ability to form neurospheres, and inhibition of its activity triggered the downregulation of genes associated with the stem cell phenotype (Sherry et al. 2009) and sensitization to chemotherapeutic treatment, suggesting that combined chemotherapy and STAT3 inhibition may allow more efficient killing of CSCs. STAT3 activation in glioblastoma is often supported by the constitutive expression of IL-6 in tumour cells, and indeed IL6<sup>-/-</sup> mice were protected from glioblastoma development (Brantley and Benveniste 2008). Abnormal activation of the FGFR and EGFR

pathways also correlated with STAT3 phosphorylation (Brantley and Benveniste 2008). Interestingly, however, while STAT3 could cooperate with the oncogenic mutant form EGFRvIII to mediate cell transformation, it accelerated disease progression in glioblastomas induced by PTEN-loss. Thus, depending on the genetic background, STAT3 activity in glioblastoma can be either tumour-suppressive or tumour-promoting (de la Iglesia et al. 2008).

#### **Other Solid Tumours**

A low percentage of rhabdomyosarcomas showed STAT3 activation that is linked to enhanced proliferation and resistance to apoptosis (Chen et al. 2007b) and correlating with the overexpression of the stem cell marker Piwil2, recently found associated to different tumours. Piwil2 can activate STAT3, which in turn enhances tumour cell survival through Bcl-xL induction (Lee et al. 2006). Moreover, STAT3 can interact with PAX3-FKHR, an oncogenic fusion protein specifically associated with an aggressive rhabdomyosarcoma metastatic subtype. This association leads to a reduction in tumour MHC expression and to an altered cytokine microenvironment that inhibits inflammatory cells action and hampers immune detection of tumour (Nabarro et al. 2005).

STAT3 activation was observed in 100% of renal carcinomas, correlating with poor prognosis and metastatic disease and promoting proliferation and survival (Horiguchi et al. 2002a, b). Pharmacological inhibition of STAT3 not only favoured the apoptotic action of chemotherapeutic agents on tumour cells, but also downmodulated their angiogenic and metastatic potential while improving antitumour immune response by reducing myeloid suppressor and Tregs cells (Xin et al. 2009).

About 60% of cervical cancers display STAT3 phosphorylation, correlating with poor prognosis (Takemoto et al. 2009) and linked to increased proliferation and apoptosis resistance via induced expression of Bcl2, survivin, Mcl1 (Chen et al. 2007a) and enhanced angiogenesis mediated by VEGF (Wei et al. 2003).

Finally STAT3 activation in bladder cancer cells, although limited, was implicated in tumour cells proliferation and invasion (Itoh et al. 2006; Chen et al. 2008a).

#### **Breast Cancer**

Persistently phosphorylated STAT3 is detected in 30–60% of primary breast carcinomas (Garcia et al. 2001) correlating with poor response to therapy (Diaz et al. 2006) and with regional lymph node metastasis (Hsieh et al. 2005), although a correlation with a good prognosis of node-negative cancers was suggested (Dolled-Filhart et al. 2003). High STAT3 phosphorylation levels are detected in several human breast cancer cell lines, where its inactivation leads to growth arrest and cell death (Garcia et al. 1997, 2001). Similar to most other solid tumours, STAT3

activity in breast cancer has been linked to enhanced proliferation and survival, to resistance to apoptosis and to cell movement, invasion and metastasis.

#### Pathways Leading to Persistent STAT3 Activation in Breast Cancer

Despite the wide range of tumours where STAT3 is constitutively active, so far no activating genetic mutations have been described, suggesting that abnormal STAT3 activity in neoplastic cells must be triggered by deregulated upstream signalling. In breast cancer, STAT3 activation shows positive correlation with EGF and ErbB2 receptors overexpression and with Src activation (Berclaz et al. 2001; Diaz et al. 2006; Leslie et al. 2006), all of which have been shown to lead to STAT3 phosphorylation, albeit not directly (Berishaj et al. 2007). V-Src was the first oncogene whose transforming activities were shown to require STAT3 (Bromberg et al. 1998). Additionally, STAT3 was reported to be a substrate of the breast tumour kinase (Brk), distantly related to the Src family (Liu et al. 2006).

An impressive body of data points towards IL-6 as the main trigger for STAT3 aberrant activation in solid tumours, which at hindsight is perhaps not surprising since IL-6 and its family of related cytokines are among the most prominent inducers of STAT3 activity. In breast cancer patients, serum IL-6 levels are elevated (Jiang et al. 2000; Kozłowski et al. 2003), and correlate with advanced tumour stage (Kozłowski et al. 2003), increased number of metastatic sites (Salgado et al. 2003) and overall poor prognosis (Zhang and Adachi 1999) (Bachelot et al. 2003; Salgado et al. 2003). High local IL-6 production is also detected, correlating with tumour grade (Chavey et al. 2007). Indeed, inflammationinduced IL-6 produced either systemically or locally by tumour infiltrating inflammatory cells is believed to start a positive loop by activating STAT3 in cancer cells (Grivennikov and Karin 2010). This in turn induces the secretion of soluble factors promoting STAT3 activation and anergy in the antigen presenting cells, finally leading to enhanced tumour cell survival and growth both via cell autonomous and immune-mediated mechanisms (Yu et al. 2009). An oncogene-driven inflammatory loop was also implicated in the initial stages of tumour transformation. Indeed, transient Src activation generates an inflammatory signal which triggers an epigenetic switch to cancer cells via a positive feedback loop involving NF-kB, Lin28, let-7 and IL-6 (Iliopoulos et al. 2009). IL-6-activated STAT3 is essential for this switch via direct induction of miR-21 and miR-181b-1, which target the PTEN and CYLD tumour suppressor genes, respectively. Their downregulation in turn leads to NF-kB activation, required to maintain the transformed state (Iliopoulos et al. 2010). The importance of this circuit was first demonstrated in transformation of mammary epithelial cells and subsequently confirmed in prostate, colon, lung and hepatocellular carcinoma cells.

IL-6-induced STAT3 activation is normally transient, due to tight negative feedback control such as that mediated by SOCS3 (Yoshimura 2005). What are the mechanisms helping to maintain persistent STAT3 phosphorylation in tumours?

Loss of negative feedback via silencing of SOCS factors has been shown to occur in several systems (Baltayiannis et al. 2008). Recently, it was shown that the low but constitutive activation of STAT3 in different tumours, including the breast, is at least partly mediated by the elevated expression of S1PR1, the receptor for the lysophospholipid sphingosine-1-phospate. S1PR1 is a STAT3 transcriptional target which in turn upregulates IL-6 expression and enhances STAT3 activation, establishing a positive feedback loop resulting in STAT3 persistent activation in both the tumour cells and the tumour microenvironment, accelerated tumour growth and malignant progression (Lee et al. 2010).

Other cytokines belonging to the IL-6 family, such as LIF (Quaglino et al. 2007) and leptin (Park et al. 2010), are also elevated in breast tumours, driving STAT3 activation. In particular, adipocyte-derived leptin is present at high concentrations within the mammary gland of obese individuals, is considered as a risk factor in several types of cancers and is proposed to correlate with breast cancer progression (Garofalo et al. 2004). Estrogen receptor alpha was shown to enhance leptin-mediated STAT3 activation (Binai et al. 2010), and inactivation of the peripheral leptin receptor attenuates tumour progression and metastasis in an MMTV-PyMT model of breast cancer, via inactivation of the ERK1/2 and Jak2/STAT3 pathways (Park et al. 2010).

#### STAT3-Mediated Features: Proliferation and Survival

Most cell lines displaying persistent STAT3 phosphorylation are addicted to its activity for proliferation and survival, both in vitro and in vivo (Garcia et al. 2001; Hsieh et al. 2005; Diaz et al. 2006), at least partly correlating with the induction of the anti-apoptotic genes survivin/BIRC5 and Bcl-xL and of cyclin D1 (Siddiquee et al. 2007). Indeed, high levels of activated STAT3 correlate positively with elevated cyclin D1 mRNA and protein expression in breast tumours and cell lines (Leslie et al. 2006) and STAT3 can directly bind to the promoter of the human cyclin D1 gene (Leslie et al. 2006; Saxena et al. 2007). Moreover, cyclin D1 appears to be required for mouse fibroblasts anchorage-independent growth downstream of constitutively active STAT3C or v-Src (Leslie et al. 2006). Interestingly, the progesteron receptor was shown to act as STAT3 coactivator by inducing ErbB2 nuclear translocation and the assembly of a transcriptional complex on the cyclin D1 promoter (Béguelin et al. 2010).

Immunohistochemical analyses of invasive breast carcinomas also showed a positive correlation between activated Src, phosphorylated STAT3 and the expression of survivin, a member of the inhibitor of apoptosis protein family (Diaz et al. 2006). Like cyclin D1, also survivin is a direct STAT3 transcriptional target, and STAT3 silencing leads to survivin downregulation and apoptotic death in a human breast cancer cell line (Gritsko et al. 2006). In addition to downregulating survivin and Bcl-xL expression, STAT3 silencing in human breast cancer cells was recently shown to lead to Fas-mediated intrinsic apoptotic pathway via the activation of caspases-8, -9, -3 and PARP1 cleavage (Kunigal et al. 2009).

The pro-survival role of STAT3 might be exploited for therapeutic purposes in combined treatments. For example, the inhibition of STAT3 in metastatic breast cancer cells enhanced the proapoptotic effects of doxorubicin, at least in part interfering with survivin and Bcl-xL expression (Gariboldi et al. 2007). Recently, ErbB2-activated STAT3 was shown to directly upregulate the p21(Cip1) gene in breast cancer cells, resulting in increased Taxol resistance and suggesting that Src and STAT3 inhibitors may be used in Taxol sensitization of ErbB2-overexpressing breast cancers (Hawthorne et al. 2009).

#### STAT3-Mediated Features: Migration, Invasion and Metastasis

Activated STAT3 levels have been reported to correlate with invasiveness and metastasis in breast cancer (Hsieh et al. 2005), and indeed a leading role for STAT3 in driving migration, invasion and metastatic disease of breast cancer cells has emerged in the past years, and thoroughly explored in mouse models of ErbB2-driven tumorigenesis genetically modified for STAT3 (see next section). Both transcriptional and non-transcriptional mechanisms have been proposed to drive STAT3-induced migration.

Intriguingly, activated STAT3 was shown for the first time in ovarian cancer cells to localize not only to the nucleus but also to the focal adhesions, interacting with activated paxillin and focal adhesion kinase, implying local regulation of focal adhesions and integrin-mediated cell movement (Silver et al. 2004). We also have observed STAT3 localization to focal adhesions in mouse breast cancer cell lines derived from MMTV-Her2 transgenic tumours, which was enhanced in cells derived from mice expressing constitutively active STAT3C and displaying more aggressive and invasive tumour phenotype (Barbieri et al. 2010a). Cytoplasmic, non-phosphorylated STAT3 was reported to induce cell migration by interacting with, and inhibiting, the microtubules destabilizer stathmin, thus enhancing microtubules polymerization in murine embryonic fibroblasts (Ng et al. 2006). Conversely, several microtubule-based drugs were shown to modulate STAT3 activity by reducing its phosphorylation in breast tumour cell lines, possibly explaining part of their therapeutic mechanism (Walker et al. 2010).

STAT3-mediated invasion has been linked to the ability to directly upregulate the transcription of MMP9, whose expression levels correlated with those of phosphorylated STAT3 in primary breast cancers. MMP9 was required for mammary epithelial cells transformation mediated by constitutively active STAT3 (Dechow et al. 2004), and its downregulation by the trimeric resveratrol derivative LYR71 correlates with suppression of STAT3 activation, tumour migration and invasion in mouse breast cancer cells (Kim et al. 2008). Additionally, upregulation of the Fra-1 oncogene in response to tumour associated macrophages lead to a malignant switch in breast tumour cells, via activation of the IL-6/JAK/STAT3 loop and increased release of MMP9, VEGF and TGF-b (Luo et al. 2010). Interestingly, STAT3 apparently regulates different subsets of MMPs in different kinds of cancer

including MMP2 in melanoma (Xie et al. 2004), MMP1 in the bladder (Itoh et al. 2006) and MMP1, -3, -7, and -9 in the colon (Tsareva et al. 2007).

STAT3-driven metastasis formation was also linked to its ability to induce anchorage-independent growth, EMT and angiogenesis. Impaired in vivo metastasis due to reduced angiogenesis was reported to occur as a consequence of inhibiting STAT3 activation by expressing a dominant negative form of gp130 in a human breast cancer cell line (Selander et al. 2004), correlating with increased expression of the tissue inhibitor of metallo-proteinase 3 (TIMP-3). On the other hand, VEGF is a direct STAT3 transcriptional target, and its upregulated production by STAT3 is believed to induce angiogenesis in different cancer types including the breast (Niu et al. 2002b).

Metastatic breast cancer cells display increased expression of the EMT transcription factor Twist1, which is required for EMT and breast cancer metastasis. A strong positive correlation between active STAT3 and Twist1 levels was detected in late stage breast cancer tissues and in subpopulations of human breast cancer cell lines displaying enhanced invasiveness (Lo et al. 2007; Cheng et al. 2008c). In these cells, STAT3 inhibition lead to Twist1 downregulation correlating with impaired migration, invasion and colony formation, all of which could be rescued by Twist1 re-expression. Interestingly, it was recently shown that IL-6, the canonical STAT3 activator, induces EMT in the ERa-positive human breast cancer cell line MCF-7, including impaired E-cadherin expression and induction of vimentin, N-cadherin, Snail and Twist1. Conversely, constitutive expression of Twist1 triggered aberrant IL-6 production and STAT3 activation, suggesting a positive loop promoting autocrine IL-6 production (Sullivan et al. 2009). Finally, STAT3 was required for EGF-induced Twist1 upregulation in human breast carcinoma cells by directly binding to its promoter (Lo et al. 2007). Correlations between STAT3 and Twist1 were observed also in mouse cells. The silencing of STAT3 in the metastatic mouse breast cancer 4T1 cell line is sufficient to impair tumour formation in vivo and invasion ability in vitro, correlating with reduction of c-Myc, activated Src and Twist1 (Ling and Arlinghaus 2005). However, no putative STAT3 binding site was detected in the murine Twist1 promoter, suggesting different modes of STAT3dependent activation in the human and the mouse (Lo et al. 2007).

Finally, IL-6 paracrine/autocrine production and STAT3 activation were recently shown to take part in the cross-talk between cancer cells and tumour microenvironment to regulate motility, aggressiveness, angiogenesis and metastasis. Mesenchymal stem cells (MSCs), which reside in the bone marrow, are likely to come in contact with extravasated, metastasis-initiating breast cancer cells. These cells were shown to enhance tumour aggressivity and growth rates in ER-alphanegative breast cancer cell lines via IL-6 secretion and STAT3 activation (Sasser et al. 2007). MSCs have also been shown to selectively migrate to hypoxic breast tumours, where they are thought to play a tumour-promoting role. Tumour-produced IL-6 acts as an attractant for MSCs, leading to their cytoskeletal reorganization via STAT3 activation (Rattigan et al. 2010). Once within strict contact, a positive loop is likely to get started, whereby infiltrating MSCs in turn produce IL-6 and enhance STAT3 activation in the cancer cells. Due to their specific ability to

migrate to and engraft into primary breast tumours, genetically modified MSCs over-expressing Interferon-b (IFN-b) have been explored as potential therapeutic means. IFN-b producing MSCs suppressed breast cancer cells growth and pulmonary and hepatic metastases mainly via inhibition of STAT3 signalling (Ling et al. 2010).

Growth and spread of cancer is thought to be mainly driven by a small subpopulation of CSCs, the only cells capable of long-term self renewal and of generating phenotypically diverse tumour cell populations. These slowly-replicating, selfmaintaining cells are resistant to most chemotherapeutics, thus driving relapse. STAT3 is prominently involved in maintaining the undifferentiated status of mouse embryonic stem cells (Burdon et al. 2002), and was shown to be critical for the viability and maintenance of the stem-like side population in the MCF-7 breast cancer cell line (Zhou et al. 2007). Additionally, experimental evidence implied IL-6 signalling in driving formation and malignancy of breast cancer stem cells. Sansone and co-authors reported that mammospheres from node invasive basal-like breast carcinoma tissues, an aggressive breast carcinoma variant showing stem cell features, produce high levels of IL-6, and that autocrine IL-6 signalling sustain the aggressive features of hypoxia-selected MCF-7 cells (Sansone et al. 2007). Recently, IL-6 was shown to drive the conversion of nonstem cancer cells in CSCs in human breast tumours and cell lines (Iliopoulos et al. 2011a). The intimate relationship of STAT3 with the IL-6 pathway leads to postulate its involvement in these systems, even though its activation was not specifically explored.

### Role of STAT3 in ErbB2-Driven Mammary Tumorigenesis: Lessons from Mouse Models

Overexpression of the rat oncogenic form of the human EGF receptor ErbB2 (Neu) in the mammary gland under the MMTV promoter triggers the onset of invasive multifocal breast carcinomas at high multiplicity and is widely used as a model for human breast cancer (Guy et al. 1996). The role of STAT3 in Neu-mediated tumorigenesis has been studied by several groups including ours, suggesting a pivotal role of STAT3 in driving tumour progression and metastasis that is in agreement with the clinical and experimental observations reported above. All studies suggest that, although not required for Neu-driven breast tumours onset and growth, STAT3 is heavily implicated in the formation of lung metastasis by a variety of mechanisms. Analyzing the role of b4 integrin in ErbB2 signalling by deleting the b4 signalling domain in the context of MMTV-Neu transgenic mice, Guo and colleagues have shown that b4 integrin forms a complex with ErbB2, enhancing the activation of the transcription factors STAT3 and c-Jun. While c-Jun is required for hyperproliferation, STAT3 contributes to disruption of epithelial adhesion and polarity, and is required for cell invasion and experimental metastasis (Guo et al. 2006). In agreement with this finding, Cre-mediated STAT3 loss of function in MMTV-Neu transgenic mice has shown that STAT3 is not required for the onset and growth of breast tumours, but its deletion results in a dramatic reduction of lung metastasis by both primary and xenografted tumours (Ranger et al. 2009; Barbieri et al. 2010b). The reduced malignancy of STAT3-deficient tumours was partly due to an inhibition of both inflammatory and angiogenic responses, normally activated in a STAT3-dependent transcriptional cascade involving C/EBP $\delta$  (Ranger et al. 2009). Additionally, STAT3 is required in a cell autonomous fashion to warrant anchorage-independent growth and the ability to produce lung metastasis in immuno-depressed mice (Barbieri et al. 2010b).

In an effort to reproduce the relatively low but persistent activation of STAT3 observed in most tumours, we have generated knock-in mice expressing at physiological levels the constitutively active mutant form STAT3C. In agreement with the results obtained with the loss of function mutants, MMTV-Neu transgenic mice carrying the STAT3C allele developed earlier onset, more aggressive tumours with lower levels of spontaneous apoptosis but similar proliferation rates (Barbieri et al. 2010a). Tumour-derived STAT3C/Neu cell lines displayed enhanced migration and invasion in vitro and increased tumorigenic and metastatic potential in vivo, correlating with a profoundly modified organization of cell-cell contacts showing altered, irregular distribution of both adherent and tight junctions components such as E-cadherin, b-catenin and Zo-1. Cytoskeletal organization was also perturbed, with actin redistributing from a cortical localization typical of well differentiated epithelial cells to form abundant actin stress fibres, typical of highly migratory cells (Pellegrin and Mellor 2007). Several genes consistently expressed at higher levels in all three STAT3C/Neu cell lines are known players in regulating cell migration and/or tumour metastasis, including the STAT3 transcriptional target Twist1, involved in tumour invasiveness and EMT (Lo et al. 2007; Cheng et al. 2008c).

In addition, we identified the atypical tensin family member Cten as a novel STAT3 target. Cten was recently shown to mediate EGF-induced migration (Katz et al. 2007), to promote colon cancer tumorigenicity and cell motility (Albasri et al. 2009; Liao et al. 2009), and to correlate positively with tumour stage in thymomas, lung tumours and gastric tumours (Sasaki et al. 2003a, b; Sakashita et al. 2008), all displaying constitutive STAT3 activity. It is the most consistently upregulated gene in both STAT3C-expressing cell lines and tumours, and is involved in both their increased migration and disruption of cell junctions organization (Fig. 1, adapted from Barbieri et al. [2010a]). Moreover, we could show that Cten is induced by IL-6 in MCF10 mammary epithelial cells. IL-6-mediated induction is STAT3-dependent, suggesting that indeed Cten may represent an important functional mediator in the inflammation-STAT3-migration-metastasis loop. Indeed, CTEN expression is particularly elevated in the extremely aggressive and invasive inflammatory breast cancers, correlating with high EGFR and HER2 levels, loss of oestrogen receptor, high tumour grade and node metastasis (Katz et al. 2007). Thus, CTEN may represent an important point of functional convergence between inflammationdriven STAT3 activity, altered EGFR/ErbB2-mediated signalling and invasion of the surrounding tissues.



**Fig. 1** Both Stat3 inhibition and Cten silencing partially revert the aggressive phenotype of Stat3C expressing cells. Adapted from REF. STAT3C/Neu cells were either treated with the S3I inhibitor for the indicated lengths of time (**a**, **b**) or transfected with an siRNA against Cten (**c**–**e**). S3I treatment downregulates STAT3 phosphorylation (p-Stat3) and Cten expression, as shown by Western blot (**a**). Both treatments significantly impaired FCS-stimulated Transwell migration (**b**, **d**). Values are shown as mean numbers  $\pm$  SEM of migrated cells per microscopic field (20×) of triplicates in one representative experiment out of two independently performed (p < 0.05). (**e**) phase contrast and immunofluorescence images of Cten-silenced cells. *Arrows* indicate evident discontinuous (si-ctrl) versus tight (si-Cten) cell-cell contacts. *Blue*, nuclei; *green*,  $\beta$ -catenin; *red*, Zo-1. The insects (4× magnification) correspond to the areas indicated by an *asterisk*. Scale bar, 20 µm

#### **Concluding Remarks**

STAT3 has come a long way since its discovery in the 90s as STAT1's little brother. Initially thought to be almost an IL-6-family-dedicated factor, it has soon emerged as one of the most pleiotropic STATs from many points of view, all contributing to its widespread role in tumours. First, its ever-growing number of upstream activating pathways including many that are aberrantly active in tumours, as initially hinted by the lethal phenotype of STAT3 null embryos. Second, the tissue-dependent variety of target genes, reflected in its variegated functions. Third, its novel non-canonical roles, which apparently do not involve its transcriptional activities. Importantly, its improperly prolonged activity is pro-oncogenic both in tumour and stromal cells, and indeed STAT3 is emerging as a key factor in mediating the cross talk between microenvironment and tumour cells and a main player in inflammation-driven tumorigenesis. Despite the intense research for STAT3 inhibitors, transcription factors are certainly not easily druggable targets. The understanding of STAT3 biology therefore, including which upstream events drive its activation and which are its main effectors in specific tumours, is still highly relevant on the agenda.

#### References

- Abdulghani J, Gu L, Dagvadorj A, Lutz J, Leiby B, Bonuccelli G, Lisanti MP, Zellweger T, Alanen K, Mirtti T, Visakorpi T, Bubendorf L, Nevalainen MT (2008) STAT3 promotes metastatic progression of prostate cancer. Am J Pathol 172:1717–1728
- Albasri A, Seth R, Jackson D, Benhasouna A, Crook S, Nateri AS, Chapman R, Ilyas M (2009) Cterminal tensin-like (CTEN) is an oncogene which alters cell motility possibly through repression of E-cadherin in colorectal cancer. J Pathol 218:57–65
- Albesiano E, Davis M, See AP, Han JE, Lim M, Pardoll DM, Kim Y (2010) Immunologic consequences of signal transducers and activators of transcription 3 activation in human squamous cell carcinoma. Cancer Res 70:6467–6476
- Azare J, Leslie K, Al-Ahmadie H, Gerald W, Weinreb PH, Violette SM, Bromberg J (2007) Constitutively activated STAT3 induces tumorigenesis and enhances cell motility of prostate epithelial cells through integrin beta 6. Mol Cell Biol 27:444–4453
- Bachelot T, Ray-Coquard I, Menetrier-Caux C, Rastkha M, Duc A, Blay J-Y (2003) Prognostic value of serum levels of interleukin 6 and of serum and plasma levels of vascular endothelial growth factor in hormone-refractory metastatic breast cancer patients. Br J Cancer 88:1721–1726
- Baltayiannis G, Baltayiannis N, Tsianos EV (2008) Suppressors of cytokine signaling as tumor repressors. Silencing of SOCS3 facilitates tumor formation and growth in lung and liver. J BUON 13:263–265
- Barbieri I, Pensa S, Pannellini T, Quaglino E, Maritano D, Demaria M, Voster A, Turkson J, Cavallo F, Watson CJ, Provero P, Musiani P, Poli V (2010a) Constitutively active STAT3 enhances neu-mediated migration and metastasis in mammary tumors via upregulation of Cten. Cancer Res 70:2558–2567
- Barbieri I, Quaglino E, Maritano D, Pannellini T, Riera L, Cavallo F, Forni G, Musiani P, Chiarle R, Poli V (2010b) STAT3 is required for anchorage-independent growth and metastasis but not for mammary tumor development downstream of the ErbB-2 oncogene. Mol Carcinog 49:114–120
- Becker C, Fantini MC, Wirtz S, Nikolaev A, Lehr HA, Galle PR, Rose-John S, Neurath MF (2005) IL-6 signaling promotes tumor growth in colorectal cancer. Cell Cycle 4:217–220
- Béguelin W, Díaz Flaqué MC, Proietti CJ, Cayrol F, Rivas MA, Tkach M, Rosemblit C, Tocci JM, Charreau EH, Schillaci R, Elizalde PV (2010) Progesterone receptor induces ErbB-2 nuclear translocation to promote breast cancer growth via a novel transcriptional effect: ErbB-2 function as a coactivator of STAT3. Mol Cell Biol 30:5456–5472
- Berclaz G, Altermatt HJ, Rohrbach V, Siragusa A, Dreher E, Smith PD (2001) EGFR dependent expression of STAT3 (but not STAT1) in breast cancer. Int J Oncol 19:1155–1160
- Berishaj M, Gao SP, Ahmed S, Leslie K, Al-Ahmadie H, Gerald WL, Bornmann W, Bromberg JF (2007) STAT3 is tyrosine-phosphorylated through the interleukin-6/glycoprotein 130/Janus kinase pathway in breast cancer. Breast Cancer Res 9:R32

- Bharadwaj U, Li M, Zhang R, Chen C, Yao Q (2007) Elevated interleukin-6 and G-CSF in human pancreatic cancer cell conditioned medium suppress dendritic cell differentiation and activation. Cancer Res 67:5479–5488
- Binai NA, Damert A, Carra G, Steckelbroeck S, Löwer J, Löwer R, Wessler S (2010) Expression of estrogen receptor alpha increases leptin-induced STAT3 activity in breast cancer cells. Int J Cancer 127:55–66
- Bollrath J, Phesse TJ, von Burstin VA, Putoczki T, Bennecke M, Bateman T, Nebelsiek T, Lundgren-May T, Canli O, Schwitalla S, Matthews V, Schmid RM, Kirchner T, Arkan MC, Ernst M, Greten FR (2009) gp130-mediated STAT3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. Cancer Cell 15:91–102
- Brantley EC, Benveniste EN (2008) Signal transducer and activator of transcription-3: a molecular hub for signaling pathways in gliomas. Mol Cancer Res 6:675–684
- Bromberg JF, Horvath CM, Besser D, Lathem WW, Darnell JE (1998) STAT3 activation is required for cellular transformation by v-src. Mol Cell Biol 18:2553–2558
- Bromberg JF, Wrzeszczynska MH, Devgan G, Zhao Y, Pestell RG, Albanese C, Darnell JE (1999) STAT3 as an oncogene. Cell 98:295–303
- Burdon T, Smith A, Savatier P (2002) Signalling, cell cycle and pluripotency in embryonic stem cells. Trends Cell Biol 12:432–438
- Burke WM, Jin X, Lin HJ, Huang M, Liu R, Reynolds RK, Lin J (2001) Inhibition of constitutively active STAT3 suppresses growth of human ovarian and breast cancer cells. Oncogene 20:7925–7934
- Chan KS, Carbajal S, Kiguchi K, Clifford J, Sano S, DiGiovanni J (2004a) Epidermal growth factor receptor-mediated activation of STAT3 during multistage skin carcinogenesis. Cancer Res 64:2382–2389
- Chan KS, Sano S, Kiguchi K, Anders J, Komazawa N, Takeda J, DiGiovanni J (2004b) Disruption of STAT3 reveals a critical role in both the initiation and the promotion stages of epithelial carcinogenesis. J Clin Invest 114:720–728
- Chan KS, Sano S, Kataoka K, Abel E, Carbajal S, Beltran L, Clifford J, Peavey M, Shen J, Digiovanni J (2008) Forced expression of a constitutively active form of STAT3 in mouse epidermis enhances malignant progression of skin tumors induced by two-stage carcinogenesis. Oncogene 27:1087–1094
- Chavey C, Bibeau F, Gourgou-Bourgade S, Burlinchon S, Boissière F, Laune D, Roques S, Lazennec G (2007) Oestrogen receptor negative breast cancers exhibit high cytokine content. Breast Cancer Res 9:R15
- Chen CL, Hsieh FC, Lieblein JC, Brown J, Chan C, Wallace JA, Cheng G, Hall BM, Lin J (2007a) STAT3 activation in human endometrial and cervical cancers. Br J Cancer 96:591–599
- Chen CL, Loy A, Cen L, Chan C, Hsieh FC, Cheng G, Wu B, Qualman SJ, Kunisada K, Yamauchi-Takihara K, Lin J (2007b) Signal transducer and activator of transcription 3 is involved in cell growth and survival of human rhabdomyosarcoma and osteosarcoma cells. BMC Cancer 7:111
- Chen CL, Cen L, Kohout J, Hutzen B, Chan C, Hsieh FC, Loy A, Huang V, Cheng G, Lin J (2008a) Signal transducer and activator of transcription 3 activation is associated with bladder cancer cell growth and survival. Mol Cancer 7:78
- Chen YW, Chen KH, Huang PI, Chen YC, Chiou GY, Lo WL, Tseng LM, Hsu HS, Chang KW, Chiou SH (2008b) Cucurbitacin I suppressed stem-like property and enhanced radiationinduced apoptosis in head and neck squamous carcinoma-derived CD44(+)ALDH1(+) cells. Mol Cancer Ther 9:2879–2892
- Cheng GZ, Zhang WZ, Sun M, Wang Q, Coppola D, Mansour M, Xu LM, Costanzo C, Cheng JQ, Wang L-H (2008) Twist is transcriptionally induced by activation of STAT3 and mediates STAT3 oncogenic function. J Biol Chem 283:14665–14673
- Choi JH, Ahn MJ, Park CK, Han HX, Kwon SJ, Lee YY, Kim IS (2006) Phospho-STAT3 expression and correlation with VEGF, p53, and Bcl-2 in gastric carcinoma using tissue microarray. APMIS 114:619–625

- Choudhari SR, Khan MA, Harris G, Picker D, Jacob GS, Block T, Shailubhai K (2007) Deactivation of Akt and STAT3 signaling promotes apoptosis, inhibits proliferation, and enhances the sensitivity of hepatocellular carcinoma cells to an anticancer agent. Atiprimod Mol Cancer Ther 6:112–121
- Corvinus FM, Orth C, Moriggl R, Tsareva SA, Wagner S, Pfitzner EB, Baus D, Kaufmann R, Huber LA, Zatloukal K, Beug H, Ohlschlager P, Schutz A, Halbhuber KJ, Friedrich K (2005) Persistent STAT3 activation in colon cancer is associated with enhanced cell proliferation and tumor growth. Neoplasia 7:545–555
- de la Iglesia N, Konopka G, Puram SV, Chan JA, Bachoo RM, You MJ, Levy DE, Depinho RA, Bonni A (2008) Identification of a PTEN-regulated STAT3 brain tumor suppressor pathway. Genes Dev 22:449–462
- Dechow TN, Pedranzini L, Leitch A, Leslie K, Gerald WL, Linkov I, Bromberg JF (2004) Requirement of matrix metalloproteinase-9 for the transformation of human mammary epithelial cells by STAT3-C. Proc Natl Acad Sci USA 101:10602–10607
- Demaria M, Giorgi C, Lebiedzinska M, Esposito G, D'Angeli L, Bartoli A, Gough DJ, Turkson J, Levy DE, Watson CJ, Wieckowski MR, Provero P, Pinton P, Poli V (2010) A STAT3mediated metabolic switch is involved in tumour transformation and STAT3 addiction. Aging (Albany NY) 2:823–842
- Deng JY, Sun D, Liu XY, Pan Y, Liang H (2010) STAT-3 correlates with lymph node metastasis and cell survival in gastric cancer. World J Gastroenterol 16:5380–5387
- Diaz N, Minton S, Cox C, Bowman T, Gritsko T, Garcia R, Eweis I, Wloch M, Livingston S, Seijo E, Cantor AB, Lee J-H, Beam CA, Sullivan D, Jove R, Muro-Cacho CA (2006) Activation of STAT3 in primary tumors from high-risk breast cancer patients is associated with elevated levels of activated SRC and survivin expression. Clin Cancer Res 12:20–28
- Dolled-Filhart M, Camp RL, Kowalski DP, Smith BL, Rimm DL (2003) Tissue microarray analysis of signal transducers and activators of transcription 3 (STAT3) and phospho-STAT3 (Tyr705) in node-negative breast cancer shows nuclear localization is associated with a better prognosis. Clin Cancer Res 9:594–600
- Duan Z, Foster R, Bell DA, Mahoney J, Wolak K, Vaidya A, Hampel C, Lee H, Seiden MV (2006) Signal transducers and activators of transcription 3 pathway activation in drug-resistant ovarian cancer. Clin Cancer Res 12:5055–5063
- Feng DY, Zheng H, Tan Y, Cheng RX (2001) Effect of phosphorylation of MAPK and STAT3 and expression of c-fos and c-jun proteins on hepatocarcinogenesis and their clinical significance. World J Gastroenterol 7:33–36
- Flowers LO, Subramaniam PS, Johnson HM (2005) A SOCS-1 peptide mimetic inhibits both constitutive and IL-6 induced activation of STAT3 in prostate cancer cells. Oncogene 24:2114–2120
- Gao B, Shen X, Kunos G, Meng Q, Goldberg ID, Rosen EM, Fan S (2001) Constitutive activation of JAK-STAT3 signaling by BRCA1 in human prostate cancer cells. FEBS Lett 488:179–184
- Gao L, Zhang L, Hu J, Li F, Shao Y, Zhao D, Kalvakolanu DV, Kopecko DJ, Zhao X, Xu DQ (2005) Down-regulation of signal transducer and activator of transcription 3 expression using vector-based small interfering RNAs suppresses growth of human prostate tumor in vivo. Clin Cancer Res 11:6333–6341
- Gao SP, Mark KG, Leslie K, Pao W, Motoi N, Gerald WL, Travis WD, Bornmann W, Veach D, Clarkson B, Bromberg JF (2007) Mutations in the EGFR kinase domain mediate STAT3 activation via IL-6 production in human lung adenocarcinomas. J Clin Invest 117:3846–3856
- Garcia R, Yu CL, Hudnall A, Catlett R, Nelson KL, Smithgall T, Fujita DJ, Ethier SP, Jove R (1997) Constitutive activation of STAT3 in fibroblasts transformed by diverse oncoproteins and in breast carcinoma cells. Cell Growth Differ 8:1267–1276
- Garcia R, Bowman TL, Niu G, Yu H, Minton S, Muro-Cacho CA, Cox CE, Falcone R, Fairclough R, Parsons S, Laudano A, Gazit A, Levitzki A, Kraker A, Jove R (2001) Constitutive activation of STAT3 by the Src and JAK tyrosine kinases participates in growth regulation of human breast carcinoma cells. Oncogene 20:2499–2513

- Gariboldi MB, Ravizza R, Molteni R, Osella D, Gabano E, Monti E (2007) Inhibition of STAT3 increases doxorubicin sensitivity in a human metastatic breast cancer cell line. Cancer Lett 258:181–188
- Garofalo C, Sisci D, Surmacz E (2004) Leptin interferes with the effects of the antiestrogen ICI 182,780 in MCF-7 breast cancer cells. Clin Cancer Res 10:6466–6475
- Gough DJ, Corlett A, Schlessinger K, Wegrzyn J, Larner AC, Levy DE (2009) Mitochondrial STAT3 supports Ras-dependent oncogenic transformation. Science 324:1713–1716
- Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M (2004) IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. Cell 118:285–296
- Gritsko T, Williams A, Turkson J, Kaneko S, Bowman T, Huang M, Nam S, Eweis I, Diaz N, Sullivan D, Yoder S, Enkemann S, Eschrich S, Lee J-H, Beam CA, Cheng J, Minton S, Muro-Cacho CA, Jove R (2006) Persistent activation of STAT3 signaling induces survivin gene expression and confers resistance to apoptosis in human breast cancer cells. Clin Cancer Res 12:11–19
- Grivennikov SI, Karin M (2010) Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. Cytokine Growth Factor Rev 21:11–19
- Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, Scheller J, Rose-John S, Cheroutre H, Eckmann L, Karin M (2009) IL-6 and STAT3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. Cancer Cell 15:103–113
- Gu F, Ma Y, Zhang Z, Zhao J, Kobayashi H, Zhang L, Fu L (2010) Expression of STAT3 and Notch1 is associated with cisplatin resistance in head and neck squamous cell carcinoma. Oncol Rep 23:671–676
- Guo W, Pylayeva Y, Pepe A, Yoshioka T, Muller WJ, Inghirami G, Giancotti FG (2006) Beta 4 integrin amplifies ErbB2 signaling to promote mammary tumorigenesis. Cell 126:489–502
- Guy CT, Cardiff RD, Muller WJ (1996) Activated neu induces rapid tumor progression. J Biol Chem 271:7673–7678
- Haura EB, Zheng Z, Song L, Cantor A, Bepler G (2005) Activated epidermal growth factor receptor-STAT-3 signaling promotes tumor survival in vivo in non-small cell lung cancer. Clin Cancer Res 11:8288–8294
- Hawthorne VS, Huang W-C, Neal CL, Tseng L-M, Hung M-C, Yu D (2009) ErbB2-mediated Src and signal transducer and activator of transcription 3 activation leads to transcriptional upregulation of p21Cip1 and chemoresistance in breast cancer cells. Mol Cancer Res 7:592–600
- He G, Yu GY, Temkin V, Ogata H, Kuntzen C, Sakurai T, Sieghart W, Peck-Radosavljevic M, Leffert HL, Karin M (2010) Hepatocyte IKKbeta/NF-kappaB inhibits tumor promotion and progression by preventing oxidative stress-driven STAT3 activation. Cancer Cell 17:286–297
- Hellsten R, Johansson M, Dahlman A, Dizeyi N, Sterner O, Bjartell A (2008) Galiellalactone is a novel therapeutic candidate against hormone-refractory prostate cancer expressing activated Stat3. Prostate 68:269–280
- Hemmati HD, Nakano I, Lazareff JA, Masterman-Smith M, Geschwind DH, Bronner-Fraser M, Kornblum HI (2003) Cancerous stem cells can arise from pediatric brain tumors. Proc Natl Acad Sci USA 100:15178–15183
- Horiguchi A, Oya M, Marumo K, Murai M (2002a) STAT3, but not ERKs, mediates the IL-6induced proliferation of renal cancer cells, ACHN and 769P. Kidney Int 61:926–938
- Horiguchi A, Oya M, Shimada T, Uchida A, Marumo K, Murai M (2002b) Activation of signal transducer and activator of transcription 3 in renal cell carcinoma: a study of incidence and its association with pathological features and clinical outcome. J Urol 168:762–765
- Hsieh F-C, Cheng G, Lin J (2005) Evaluation of potential STAT3-regulated genes in human breast cancer. Biochem Biophys Res Commun 335:292–299
- Huang M, Page C, Reynolds RK, Lin J (2000) Constitutive activation of STAT 3 oncogene product in human ovarian carcinoma cells. Gynecol Oncol 79:67–73
- Hussain SF, Kong LY, Jordan J, Conrad C, Madden T, Fokt I, Priebe W, Heimberger AB (2007) A novel small molecule inhibitor of signal transducers and activators of transcription 3 reverses immune tolerance in malignant glioma patients. Cancer Res 67:9630–9636

- Iliopoulos D, Hirsch HA, Struhl K (2009) An epigenetic switch involving NF-kappaB, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. Cell 139:693–706
- Iliopoulos D, Jaeger SA, Hirsch HA, Bulyk ML, Struhl K (2010) STAT3 activation of miR-21 and miR-181b-1 via PTEN and CYLD are part of the epigenetic switch linking inflammation to cancer. Mol Cell 39:493–506
- Iliopoulos D, Hirsch HA, Wang G, Struhl K (2011) Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. Proc Natl Acad Sci USA 108:1397–1402
- Itoh M, Murata T, Suzuki T, Shindoh M, Nakajima K, Imai K, Yoshida K (2006) Requirement of STAT3 activation for maximal collagenase-1 (MMP-1) induction by epidermal growth factor and malignant characteristics in T24 bladder cancer cells. Oncogene 25:1195–1204
- Jee SH, Chu CY, Chiu HC, Huang YL, Tsai WL, Liao YH, Kuo ML (2004) Interleukin-6 induced basic fibroblast growth factor-dependent angiogenesis in basal cell carcinoma cell line via JAK/STAT3 and PI3-kinase/Akt pathways. J Invest Dermatol 123:1169–1175
- Jiang XP, Yang DC, Elliott RL, Head JF (2000) Reduction in serum IL-6 after vacination of breast cancer patients with tumour-associated antigens is related to estrogen receptor status. Cytokine 12:458–465
- Jing N, Li Y, Xiong W, Sha W, Jing L, Tweardy DJ (2004) G-quartet oligonucleotides: a new class of signal transducer and activator of transcription 3 inhibitors that suppresses growth of prostate and breast tumors through induction of apoptosis. Cancer Res 64:6603–6609
- Jing N, Zhu Q, Yuan P, Li Y, Mao L, Tweardy DJ (2006) Targeting signal transducer and activator of transcription 3 with G-quartet oligonucleotides: a potential novel therapy for head and neck cancer. Mol Cancer Ther 5:279–286
- Jung JE, Kim HS, Lee CS, Park DH, Kim YN, Lee MJ, Lee JW, Park JW, Kim MS, Ye SK, Chung MH (2007) Caffeic acid and its synthetic derivative CADPE suppress tumor angiogenesis by blocking STAT3-mediated VEGF expression in human renal carcinoma cells. Carcinogenesis 28:1780–1787
- Kanda N, Seno H, Konda Y, Marusawa H, Kanai M, Nakajima T, Kawashima T, Nanakin A, Sawabu T, Uenoyama Y, Sekikawa A, Kawada M, Suzuki K, Kayahara T, Fukui H, Sawada M, Chiba T (2004) STAT3 is constitutively activated and supports cell survival in association with survivin expression in gastric cancer cells. Oncogene 23:4921–4929
- Kang SH, Yu MO, Park KJ, Chi SG, Park DH, Chung YG (2010) Activated STAT3 regulates hypoxia-induced angiogenesis and cell migration in human glioblastoma. Neurosurgery 67:1386–1395, discussion 1395
- Katz M, Amit I, Citri A, Shay T, Carvalho S, Lavi S, Milanezi F, Lyass L, Amariglio N, Jacob-Hirsch J, Ben-Chetrit N, Tarcic G, Lindzen M, Avraham R, Liao Y-C, Trusk P, Lyass A, Rechavi G, Spector NL, Lo SH, Schmitt F, Bacus SS, Yarden Y (2007) A reciprocal tensin-3-Cten switch mediates EGF-driven mammary cell migration. Nat Cell Biol 9:961–969
- Kim DJ, Chan KS, Sano S, Digiovanni J (2007) Signal transducer and activator of transcription 3 (STAT3) in epithelial carcinogenesis. Mol Carcinog 46:725–731
- Kim JE, Kim HS, Shin YJ, Lee CS, Won C, Lee SA, Lee JW, Kim Y, Kang JS, Ye SK, Chung MH (2008) LYR71, a derivative of trimeric resveratrol, inhibits tumorigenesis by blocking STAT3mediated matrix metalloproteinase 9 expression. Exp Mol Med 40:514–522
- Kim DJ, Kataoka K, Sano S, Connolly K, Kiguchi K, DiGiovanni J (2009) Targeted disruption of Bcl-xL in mouse keratinocytes inhibits both UVB- and chemically induced skin carcinogenesis. Mol Carcinog 48:873–885
- Kong LY, Abou-Ghazal MK, Wei J, Chakraborty A, Sun W, Qiao W, Fuller GN, Fokt I, Grimm EA, Schmittling RJ, Archer GE Jr, Sampson JH, Priebe W, Heimberger AB (2008) A novel inhibitor of signal transducers and activators of transcription 3 activation is efficacious against established central nervous system melanoma and inhibits regulatory T cells. Clin Cancer Res 14:5759–5768
- Kortylewski M, Jove R, Yu H (2005) Targeting STAT3 affects melanoma on multiple fronts. Cancer Metastasis Rev 24:315–327

- Kozłowski L, Zakrzewska I, Tokajuk P, Wojtukiewicz MZ (2003) Concentration of interleukin-6 (IL-6), interleukin-8 (IL-8) and interleukin-10 (IL-10) in blood serum of breast cancer patients. Rocz Akad Med Białymst 48:82–84
- Kunigal S, Lakka SS, Sodadasu PK, Estes N, Rao JS (2009) STAT3-siRNA induces Fas-mediated apoptosis in vitro and in vivo in breast cancer. Int J Oncol 34:1209–1220
- Kusaba T, Nakayama T, Yamazumi K, Yakata Y, Yoshizaki A, Nagayasu T, Sekine I (2005) Expression of p-STAT3 in human colorectal adenocarcinoma and adenoma; correlation with clinicopathological factors. J Clin Pathol 58:833–838
- Kusaba T, Nakayama T, Yamazumi K, Yakata Y, Yoshizaki A, Inoue K, Nagayasu T, Sekine I (2006) Activation of STAT3 is a marker of poor prognosis in human colorectal cancer. Oncol Rep 15:1445–1451
- Lassmann S, Schuster I, Walch A, Gobel H, Jutting U, Makowiec F, Hopt U, Werner M (2007) STAT3 mRNA and protein expression in colorectal cancer: effects on STAT3-inducible targets linked to cell survival and proliferation. J Clin Pathol 60:173–179
- Lee JH, Schütte D, Wulf G, Füzesi L, Radzun H-J, Schweyer S, Engel W, Nayernia K (2006) Stem-cell protein Piwil2 is widely expressed in tumors and inhibits apoptosis through activation of STAT3/Bcl-XL pathway. Hum Mol Genet 15:201–211
- Lee J, Kang WK, Park JO, Park SH, Park YS, Lim HY, Kim J, Kong J, Choi MG, Sohn TS, Noh JH, Bae JM, Kim S, Lim do H, Kim KM, Park CK (2009) Expression of activated signal transducer and activator of transcription 3 predicts poor clinical outcome in gastric adenocarcinoma. APMIS 117:598–606
- Lee H, Deng J, Kujawski M, Yang C, Liu Y, Herrmann A, Kortylewski M, Horne D, Somlo G, Forman S, Jove R, Yu H (2010) STAT3-induced S1PR1 expression is crucial for persistent STAT3 activation in tumors. Nat Med 16:1421–1428
- Leong PL, Andrews GA, Johnson DE, Dyer KF, Xi S, Mai JC, Robbins PD, Gadiparthi S, Burke NA, Watkins SF, Grandis JR (2003) Targeted inhibition of STAT3 with a decoy oligonucleotide abrogates head and neck cancer cell growth. Proc Natl Acad Sci USA 100:4138–4143
- Leslie K, Lang C, Devgan G, Azare J, Berishaj M, Gerald W, Kim YB, Paz K, Darnell JE, Albanese C, Sakamaki T, Pestell R, Bromberg JF (2006) Cyclin D1 is transcriptionally regulated by and required for transformation by activated signal transducer and activator of transcription 3. Cancer Res 66:2544–2552
- Li WC, Ye SL, Sun RX, Liu YK, Tang ZY, Kim Y, Karras JG, Zhang H (2006) Inhibition of growth and metastasis of human hepatocellular carcinoma by antisense oligonucleotide targeting signal transducer and activator of transcription 3. Clin Cancer Res 12:7140–7148
- Li Y, Du H, Qin Y, Roberts J, Cummings OW, Yan C (2007) Activation of the signal transducers and activators of the transcription 3 pathway in alveolar epithelial cells induces inflammation and adenocarcinomas in mouse lung. Cancer Res 67:8494–8503
- Liao Y-C, Chen N-T, Shih Y-P, Dong Y, Lo SH (2009) Up-regulation of C-terminal tensin-like molecule promotes the tumorigenicity of colon cancer through beta-catenin. Cancer Res 69:4563–4566
- Ling X, Arlinghaus RB (2005) Knockdown of STAT3 expression by RNA interference inhibits the induction of breast tumors in immunocompetent mice. Cancer Res 65:2532–2536
- Ling X, Marini F, Konopleva M, Schober W, Shi Y, Burks J, Clise-Dwyer K, Wang R-Y, Zhang W, Yuan X, Lu H, Caldwell L, Andreeff M (2010) Mesenchymal stem cells overexpressing IFN-β inhibit breast cancer growth and metastases through STAT3 signaling in a syngeneic tumor model. Cancer Microenviron 3:83–95
- Liu L, Gao Y, Qiu H, Miller WT, Poli V, Reich NC (2006) Identification of STAT3 as a specific substrate of breast tumor kinase. Oncogene 25:4904–4912
- Liu Q, Li G, Li R, Shen J, He Q, Deng L, Zhang C, Zhang J (2010) IL-6 promotion of glioblastoma cell invasion and angiogenesis in U251 and T98G cell lines. J Neurooncol 100:165–176

- Lo H-W, Hsu S-C, Xia W, Cao X, Shih J-Y, Wei Y, Abbruzzese JL, Hortobagyi GN, Hung M-C (2007) Epidermal growth factor receptor cooperates with signal transducer and activator of transcription 3 to induce epithelial-mesenchymal transition in cancer cells via up-regulation of TWIST gene expression. Cancer Res 67:9066–9076
- Loeffler S, Fayard B, Weis J, Weissenberger J (2005) Interleukin-6 induces transcriptional activation of vascular endothelial growth factor (VEGF) in astrocytes in vivo and regulates VEGF promoter activity in glioblastoma cells via direct interaction between STAT3 and Sp1. Int J Cancer 115:202–213
- Lou W, Ni Z, Dyer K, Tweardy DJ, Gao AC (2000) Interleukin-6 induces prostate cancer cell growth accompanied by activation of STAT3 signaling pathway. Prostate 42:239–242
- Lui VW, Wong EY, Ho Y, Hong B, Wong SC, Tao Q, Choi GC, Au TC, Ho K, Yau DM, Ma BB, Hui EP, Chan AS, Tsang CM, Tsao SW, Grandis JR, Chan AT (2009) STAT3 activation contributes directly to Epstein-Barr virus-mediated invasiveness of nasopharyngeal cancer cells in vitro. Int J Cancer 125:1884–1893
- Luo YP, Zhou H, Krueger J, Kaplan C, Liao D, Markowitz D, Liu C, Chen T, Chuang T-H, Xiang R, Reisfeld RA (2010) The role of proto-oncogene Fra-1 in remodeling the tumor microenvironment in support of breast tumor cell invasion and progression. Oncogene 29:662–673
- Masuda M, Suzui M, Lim JT, Deguchi A, Soh JW, Weinstein IB (2002a) Epigallocatechin-3gallate decreases VEGF production in head and neck and breast carcinoma cells by inhibiting EGFR-related pathways of signal transduction. J Exp Ther Oncol 2:350–359
- Masuda M, Suzui M, Yasumatu R, Nakashima T, Kuratomi Y, Azuma K, Tomita K, Komiyama S, Weinstein IB (2002b) Constitutive activation of signal transducers and activators of transcription 3 correlates with cyclin D1 overexpression and may provide a novel prognostic marker in head and neck squamous cell carcinoma. Cancer Res 62:3351–3355
- Masuda M, Toh S, Koike K, Kuratomi Y, Suzui M, Deguchi A, Komiyama S, Weinstein IB (2002c) The roles of JNK1 and STAT3 in the response of head and neck cancer cell lines to combined treatment with all-trans-retinoic acid and 5-fluorouracil. Jpn J Cancer Res 93:329–339
- Masuda M, Ruan HY, Ito A, Nakashima T, Toh S, Wakasaki T, Yasumatsu R, Kutratomi Y, Komune S, Weinstein IB (2007) Signal transducers and activators of transcription 3 upregulates vascular endothelial growth factor production and tumor angiogenesis in head and neck squamous cell carcinoma. Oral Oncol 43:785–790
- Mathews LA, Hurt EM, Zhang X, Farrar WL (2010) Epigenetic regulation of CpG promoter methylation in invasive prostate cancer cells. Mol Cancer 9:267
- Mora LB, Buettner R, Seigne J, Diaz J, Ahmad N, Garcia R, Bowman T, Falcone R, Fairclough R, Cantor A, Muro-Cacho C, Livingston S, Karras J, Pow-Sang J, Jove R (2002) Constitutive activation of STAT3 in human prostate tumors and cell lines: direct inhibition of STAT3 signaling induces apoptosis of prostate cancer cells. Cancer Res 62:6659–6666
- Musteanu M, Blaas L, Mair M, Schlederer M, Bilban M, Tauber S, Esterbauer H, Mueller M, Casanova E, Kenner L, Poli V, Eferl R (2010) STAT3 is a negative regulator of intestinal tumor progression in Apc(Min) mice. Gastroenterology 138(1003–1011):e1001–1005
- Nabarro S, Himoudi N, Papanastasiou A, Gilmour K, Gibson S, Sebire N, Thrasher A, Blundell MP, Hubank M, Canderan G, Anderson J (2005) Coordinated oncogenic transformation and inhibition of host immune responses by the PAX3-FKHR fusion oncoprotein. J Exp Med 202:1399–1410
- Nagpal JK, Mishra R, Das BR (2002) Activation of STAT-3 as one of the early events in tobacco chewing-mediated oral carcinogenesis. Cancer 94:2393–2400
- Nefedova Y, Huang M, Kusmartsev S, Bhattacharya R, Cheng P, Salup R, Jove R, Gabrilovich D (2004) Hyperactivation of STAT3 is involved in abnormal differentiation of dendritic cells in cancer. J Immunol 172:464–474
- Ng DCH, Lin BH, Lim CP, Huang G, Zhang T, Poli V, Cao X (2006) STAT3 regulates microtubules by antagonizing the depolymerization activity of stathmin. J Cell Biol 172:245–257

- Nilsson MB, Langley RR, Fidler IJ (2005) Interleukin-6, secreted by human ovarian carcinoma cells, is a potent proangiogenic cytokine. Cancer Res 65:10794–10800
- Niu G, Bowman T, Huang M, Shivers S, Reintgen D, Daud A, Chang A, Kraker A, Jove R, Yu H (2002a) Roles of activated Src and STAT3 signaling in melanoma tumor cell growth. Oncogene 21:7001–7010
- Niu G, Wright KL, Huang M, Song L, Haura E, Turkson J, Zhang S, Wang T, Sinibaldi D, Coppola D, Heller R, Ellis LM, Karras J, Bromberg JF, Pardoll DM, Jove R, Yu H (2002b) Constitutive STAT3 activity up-regulates VEGF expression and tumor angiogenesis. Oncogene 21:2000–2008
- Park J, Kusminski CM, Chua SC, Scherer PE (2010) Leptin receptor signaling supports cancer cell metabolism through suppression of mitochondrial respiration in vivo. Am J Pathol 177:3133–3144
- Pellegrin S, Mellor H (2007) Actin stress fibres. J Cell Sci 120:3491-3499
- Pensa S, Regis G, Boselli D, Novelli F, Poli V (2008) STAT1 and STAT3 in tumorigenesis: two sides of the same coin? JAK-STAT Pathway in Disease A. Stephanou, Landes Bioscience, Austin, 100–121
- Pfeiffer M, Hartmann TN, Leick M, Catusse J, Schmitt-Graeff A, Burger M (2009) Alternative implication of CXCR4 in JAK2/STAT3 activation in small cell lung cancer. Br J Cancer 100:1949–1956
- Pu YS, Hour TC, Chuang SE, Cheng AL, Lai MK, Kuo ML (2004) Interleukin-6 is responsible for drug resistance and anti-apoptotic effects in prostatic cancer cells. Prostate 60:120–129
- Qu P, Roberts J, Li Y, Albrecht M, Cummings OW, Eble JN, Du H, Yan C (2009) STAT3 downstream genes serve as biomarkers in human lung carcinomas and chronic obstructive pulmonary disease. Lung Cancer 63:341–347
- Quaglino A, Schere-Levy C, Romorini L, Meiss RP, Kordon EC (2007) Mouse mammary tumors display Stat3 activation dependent on leukemia inhibitory factor signaling. Breast Cancer Res 9:R69
- Rahaman SO, Harbor PC, Chernova O, Barnett GH, Vogelbaum MA, Haque SJ (2002) Inhibition of constitutively active STAT3 suppresses proliferation and induces apoptosis in glioblastoma multiforme cells. Oncogene 21:8404–8413
- Ranger JJ, Levy DE, Shahalizadeh S, Hallett M, Muller WJ (2009) Identification of a STAT3dependent transcription regulatory network involved in metastatic progression. Cancer Res 69:6823–6830
- Rattigan Y, Hsu J-M, Mishra PJ, Glod J, Banerjee D (2010) Interleukin 6 mediated recruitment of mesenchymal stem cells to the hypoxic tumor milieu. Exp Cell Res 316:3417–3424
- Real PJ, Sierra A, De Juan A, Segovia JC, Lopez-Vega JM, Fernandez-Luna JL (2002) Resistance to chemotherapy via STAT3-dependent overexpression of Bcl-2 in metastatic breast cancer cells. Oncogene 21:7611–7618
- Rosen DG, Mercado-Uribe I, Yang G, Bast RC Jr, Amin HM, Lai R, Liu J (2006) The role of constitutively active signal transducer and activator of transcription 3 in ovarian tumorigenesis and prognosis. Cancer 107:2730–2740
- Sakashita K, Mimori K, Tanaka F, Kamohara Y, Inoue H, Sawada T, Hirakawa K, Mori M (2008) Prognostic relevance of tensin4 expression in human gastric cancer. Ann Surg Oncol 15:2606–2613
- Salgado R, Junius S, Benoy I, Van Dam P, Vermeulen P, Van Marck E, Huget P, Dirix LY (2003) Circulating interleukin-6 predicts survival in patients with metastatic breast cancer. Int J Cancer J103:642–646
- Sano S, Chan KS, Kira M, Kataoka K, Takagi S, Tarutani M, Itami S, Kiguchi K, Yokoi M, Sugasawa K, Mori T, Hanaoka F, Takeda J, DiGiovanni J (2005) Signal transducer and activator of transcription 3 is a key regulator of keratinocyte survival and proliferation following UV irradiation. Cancer Res 65:5720–5729
- Sansone P, Storci G, Tavolari S, Guarnieri T, Giovannini C, Taffurelli M, Ceccarelli C, Santini D, Paterini P, Marcu KB, Chieco P, Bonafè M (2007) IL-6 triggers malignant features in mammospheres from human ductal breast carcinoma and normal mammary gland. J Clin Invest 117:3988–4002

- Sasaki H, Moriyama S, Mizuno K, Yukiue H, Konishi A, Yano M, Kaji M, Fukai I, Kiriyama M, Yamakawa Y, Fujii Y (2003a) Cten mRNA expression was correlated with tumor progression in lung cancers. Lung Cancer 40:151–155
- Sasaki H, Yukiue H, Kobayashi Y, Fukai I, Fujii Y (2003b) Cten mRNA expression is correlated with tumor progression in thymoma. Tumour Biol 24:271–274
- Sasser AK, Sullivan NJ, Studebaker AW, Hendey LF, Axel AE, Hall BM (2007) Interleukin-6 is a potent growth factor for ER-alpha-positive human breast cancer. FASEB J 21:3763–3770
- Saxena NK, Vertino PM, Anania FA, Sharma D (2007) Leptin-induced growth stimulation of breast cancer cells involves recruitment of histone acetyltransferases and mediator complex to CYCLIN D1 promoter via activation of STAT3. J Biol Chem 282:13316–13325
- Schindler C, Levy DE, Decker T (2007) JAK-STAT signaling: from interferons to cytokines. J Biol Chem 282:20059–20063
- Selander KS, Li L, Watson L, Merrell M, Dahmen H, Heinrich PC, Muller-Newen G, Harris KW (2004) Inhibition of gp130 signaling in breast cancer blocks constitutive activation of STAT3 and inhibits in vivo malignancy. Cancer Res 64:6924–6933
- Sherry MM, Reeves A, Wu JK, Cochran BH (2009) STAT3 is required for proliferation and maintenance of multipotency in glioblastoma stem cells. Stem Cells 27:2383–2392
- Siddiquee K, Zhang S, Guida WC, Blaskovich MA, Greedy B, Lawrence HR, Yip MLR, Jove R, McLaughlin MM, Lawrence NJ, Sebti SM, Turkson J (2007) Selective chemical probe inhibitor of STAT3, identified through structure-based virtual screening, induces antitumor activity. Proc Natl Acad Sci USA 104:7391–7396
- Silva CM (2004) Role of STATs as downstream signal transducers in Src family kinase-mediated tumorigenesis. Oncogene 23:8017–8023
- Silver DL, Naora H, Liu J, Cheng W, Montell DJ (2004) Activated signal transducer and activator of transcription (STAT) 3: localization in focal adhesions and function in ovarian cancer cell motility. Cancer Res 64:3550–3558
- Song L, Turkson J, Karras JG, Jove R, Haura EB (2003) Activation of STAT3 by receptor tyrosine kinases and cytokines regulates survival in human non-small cell carcinoma cells. Oncogene 22:4150–4165
- Suiqing C, Min Z, Lirong C (2005) Overexpression of phosphorylated-STAT3 correlated with the invasion and metastasis of cutaneous squamous cell carcinoma. J Dermatol 32:354–360
- Sullivan NJ, Sasser AK, Axel AE, Vesuna F, Raman V, Ramirez N, Oberyszyn TM, Hall BM (2009) Interleukin-6 induces an epithelial-mesenchymal transition phenotype in human breast cancer cells. Oncogene 28:2940–2947
- Sun Z, Yao Z, Liu S, Tang H, Yan X (2006) An oligonucleotide decoy for STAT3 activates the immune response of macrophages to breast cancer. Immunobiology 211:199–209
- Sun X, Zhang J, Wang L, Tian Z (2008) Growth inhibition of human hepatocellular carcinoma cells by blocking STAT3 activation with decoy-ODN. Cancer Lett 262:201–213
- Takemoto S, Ushijima K, Kawano K, Yamaguchi T, Terada A, Fujiyoshi N, Nishio S, Tsuda N, Ijichi M, Kakuma T, Kage M, Hori D, Kamura T (2009) Expression of activated signal transducer and activator of transcription-3 predicts poor prognosis in cervical squamous-cell carcinoma. Br J Cancer 101:967–972
- Tebbutt NC, Giraud AS, Inglese M, Jenkins B, Waring P, Clay FJ, Malki S, Alderman BM, Grail D, Hollande F, Heath JK, Ernst M (2002) Reciprocal regulation of gastrointestinal homeostasis by SHP2 and STAT-mediated trefoil gene activation in gp130 mutant mice. Nat Med 8:1089–1097
- Tsareva SA, Moriggl R, Corvinus FM, Wiederanders B, Schütz A, Kovacic B, Friedrich K (2007) Signal transducer and activator of transcription 3 activation promotes invasive growth of colon carcinomas through matrix metalloproteinase induction. Neoplasia 9:279–291
- Turkson J, Jove R (2000) STAT proteins: novel molecular targets for cancer drug discovery. Oncogene 19:6613–6626

- Turkson J, Zhang S, Palmer J, Kay H, Stanko J, Mora LB, Sebti S, Yu H, Jove R (2004) Inhibition of constitutive signal transducer and activator of transcription 3 activation by novel platinum complexes with potent antitumor activity. Mol Cancer Ther 3:1533–1542
- van Cruijsen H, Ruiz MG, van der Valk P, de Gruijl TD, Giaccone G (2009) Tissue micro array analysis of ganglioside N-glycolyl GM3 expression and signal transducer and activator of transcription (STAT)-3 activation in relation to dendritic cell infiltration and microvessel density in non-small cell lung cancer. BMC Cancer 9:180
- Villalva C, Martin-Lanneree S, Cortes U, Dkhissi F, Wager M, Le Corf A, Tourani JM, Dusanter-Fourt I, Turhan AG, Karayan-Tapon L (2010) STAT3 is essential for the maintenance of neurosphere-initiating tumor cells in patients with glioblastomas: a potential for targeted therapy? Int J Cancer 128:826–838
- Walker SR, Chaudhury M, Nelson EA, Frank DA (2010) Microtubule-targeted chemotherapeutic agents inhibit signal transducer and activator of transcription 3 (STAT3) signaling. Mol Pharmacol 78:903–908
- Wang T, Niu G, Kortylewski M, Burdelya L, Shain K, Zhang S, Bhattacharya R, Gabrilovich D, Heller R, Coppola D, Dalton W, Jove R, Pardoll DM, Yu H (2004) Regulation of the innate and adaptive immune responses by STAT-3 signaling in tumor cells. Nat Med 10:48–54
- Weerasinghe P, Garcia GE, Zhu Q, Yuan P, Feng L, Mao L, Jing N (2007) Inhibition of STAT3 activation and tumor growth suppression of non-small cell lung cancer by G-quartet oligonucleotides. Int J Oncol 31:129–136
- Wegrzyn J, Potla R, Chwae YJ, Sepuri NB, Zhang Q, Koeck T, Derecka M, Szczepanek K, Szelag M, Gornicka A, Moh A, Moghaddas S, Chen Q, Bobbili S, Cichy J, Dulak J, Baker DP, Wolfman A, Stuehr D, Hassan MO, Fu XY, Avadhani N, Drake JI, Fawcett P, Lesnefsky EJ, Larner AC (2009) Function of mitochondrial STAT3 in cellular respiration. Science 323:793–797
- Wei D, Le X, Zheng L, Wang L, Frey JA, Gao AC, Peng Z, Huang S, Xiong HQ, Abbruzzese JL, Xie K (2003) STAT3 activation regulates the expression of vascular endothelial growth factor and human pancreatic cancer angiogenesis and metastasis. Oncogene 22:319–329
- Wheeler SE, Suzuki S, Thomas SM, Sen M, Leeman-Neill RJ, Chiosea SI, Kuan CT, Bigner DD, Gooding WE, Lai SY, Grandis JR (2010) Epidermal growth factor receptor variant III mediates head and neck cancer cell invasion via STAT3 activation. Oncogene 29:5135–5145
- Xie T-X, Wei D, Liu M, Gao AC, Ali-Osman F, Sawaya R, Huang S (2004) STAT3 activation regulates the expression of matrix metalloproteinase-2 and tumor invasion and metastasis. Oncogene 23:3550–3560
- Xie TX, Huang FJ, Aldape KD, Kang SH, Liu M, Gershenwald JE, Xie K, Sawaya R, Huang S (2006) Activation of STAT3 in human melanoma promotes brain metastasis. Cancer Res 66:3188–3196
- Xin H, Zhang C, Herrmann A, Du Y, Figlin R, Yu H (2009) Sunitinib inhibition of STAT3 induces renal cell carcinoma tumor cell apoptosis and reduces immunosuppressive cells. Cancer Res 69:2506–2513
- Xu Q, Briggs J, Park S, Niu G, Kortylewski M, Zhang S, Gritsko T, Turkson J, Kay H, Semenza GL, Cheng JQ, Jove R, Yu H (2005) Targeting STAT3 blocks both HIF-1 and VEGF expression induced by multiple oncogenic growth signaling pathways. Oncogene 24:5552–5560
- Yoshida T, Hanada T, Tokuhisa T, Kosai K, Sata M, Kohara M, Yoshimura A (2002) Activation of STAT3 by the hepatitis C virus core protein leads to cellular transformation. J Exp Med 196:641–653
- Yoshimura A (2005) Negative regulation of cytokine signaling. Clin Rev Allergy Immunol 28:205–220
- Yu CL, Meyer DJ, Campbell GS, Larner AC, Carter-Su C, Schwartz J, Jove R (1995) Enhanced DNA-binding activity of a STAT3-related protein in cells transformed by the Src oncoprotein. Science 269:81–83

- Yu H, Pardoll D, Jove R (2009) STATs in cancer inflammation and immunity: a leading role for STAT3. Nat Rev Cancer 9:798–809
- Zhang GJ, Adachi I (1999) Serum interleukin-6 levels correlate to tumor progression and prognosis in metastatic breast carcinoma. Anticancer Res 19:1427–1432
- Zhang CH, Xu GL, Jia WD, Li JS, Ma JL, Ren WH, Ge YS, Yu JH, Liu WB, Wang W (2010) Activation of STAT3 signal pathway correlates with twist and E-cadherin expression in hepatocellular carcinoma and their clinical significance. J Surg Res (in press)
- Zhou J, Zhang Y (2008) Cancer stem cells: models, mechanisms and implications for improved treatment. Cell Cycle 7:1360–1370
- Zhou J, Wulfkuhle J, Zhang H, Gu P, Yang Y, Deng J, Margolick JB, Liotta LA, Petricoin E, Zhang Y (2007) Activation of the PTEN/mTOR/STAT3 pathway in breast cancer stem-like cells is required for viability and maintenance. Proc Natl Acad Sci USA 104:16158–16163