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# Phenol transformation and dimerisation, photosensitised by the triplet state of 1-nitronaphthalene: A possible pathway to humic-like substances (HULIS) in atmospheric waters.

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## Abstract

The nitroderivatives of polycyclic aromatic hydrocarbons are potentially important photosensitisers in the atmospheric condensed phase. Here we show that the triplet state of 1-nitronaphthalene ( $^3\text{1NN}^*$ ) is able to directly react with phenol, causing its transformation upon irradiation of 1NN in aqueous solution. Additional but less important processes of phenol degradation are reactions with  $\bullet\text{OH}$  and  $^1\text{O}_2$ , both photogenerated by irradiated 1NN. Dihydroxybiphenyls and phenoxyphenols were detected as main phenol transformation intermediates, likely formed by dimerisation of phenoxy radicals that would be produced upon phenol oxidation by  $^3\text{1NN}^*$ . Very interestingly, irradiation with 1NN shifted the fluorescence peaks of phenol (Ex/Em = 220-230/280-320 nm and 250-275/280-320 nm, with Ex/Em = excitation and emission wavelengths) to a region that overlaps with “M-like” fulvic substances (Ex/Em = 250-300/330-400 nm). Moreover, at longer irradiation times a further peak appeared (Ex/Em = 300-450/400-450 nm), which is in the region of HULIS fluorescence. Irradiated material was also able to photoproduce  $^1\text{O}_2$ , thus showing photosensitisation properties. Therefore, compounds with fluorescence properties that closely resemble those of HULIS (they would be identified as HULIS by fluorescence if present in environmental samples) can be formed upon triplet-sensitised transformation of phenol by 1NN.

**Keywords:** photoinduced transformation; singlet oxygen; phenoxy radical; kinetic modelling.

## 1. Introduction

Photosensitised reactions involving phenolic compounds in the atmospheric condensed phase have recently gained considerable attention, as a potential source of humic-like substances (HULIS) (Net et al., 2010 and 2011). HULIS are secondary atmospheric constituents (El Haddad et al., 2011) and account for an important fraction of the organic aerosol mass (Decesari et al., 2001). They significantly absorb sunlight, especially at shorter wavelengths (Hoffer et al., 2006), and would play an important atmospheric role as cloud condensation nuclei (Dinar et al., 2007; Wex et al., 2008).

Phenolic compounds may occur in the atmosphere as primary pollutants because of biomass burning (Kuo et al., 2011; Shakya et al., 2011). As secondary pollutants, they derive from  $\bullet\text{OH}$ -induced oxidation of aromatic hydrocarbons emitted by combustion processes (Volkamer et al., 2002). Among the hypothesised pathways of HULIS formation, one involves oxidation of phenols to phenoxy radicals (Net et al., 2011), which yield phenol dimers upon dimerisation and H-atom rearrangement (Neta and Grodkowski, 2005; Sun et al., 2010). Dimers can follow similar reaction pathways until higher oligomers are formed (Net et al., 2010).

Atmospheric photosensitisers include a wide range of compounds with different structures. Common functionalities are aromatic carbonyls including benzophenone-like compounds, anthraquinones and aromatic nitroderivatives (Jammoul et al., 2009; Maurino et al., 2011). Brigante et al. (2010) have shown that 1-nitronaphthalene (1NN) is a potentially important photosensitiser, which forms a very reactive triplet state ( $^3\text{1NN}^*$ ) with high yield.

Interestingly, photosensitised processes in gas-solid systems usually need  $\text{O}_3$  for the reaction to proceed (Net et al., 2009 and 2010; Nieto-Gligorovski et al., 2010; Styler et al., 2009 and 2011), while  $\text{O}_3$  is not required in aqueous systems (Maddigapu et al., 2011; Maurino et al., 2011). The difference might be due to far easier contact of reactants in aqueous solution.

We here investigate the transformation of phenol in solution (proxy of the atmospheric aqueous phase), photosensitised by 1NN. The reaction was studied by laser flash photolysis (LFP) to assess the reactivity of  $^3\text{1NN}^*$  toward dissolved species. Moreover, steady irradiation experiments were carried out and a kinetic model developed to account for results. Key input data for the kinetic model were obtained by LFP. The potential for phenol to act as HULIS precursor under the studied conditions was assessed by identification of its main phototransformation intermediates. Moreover, fluorescence spectroscopy (excitation-emission matrix, EEM; Muller et al., 2008) was used to monitor a possible shift of fluorescence bands into the HULIS region as irradiation progressed. Photogeneration of singlet oxygen ( $^1\text{O}_2$ ), by material formed upon prolonged irradiation of phenol + 1NN, was assessed with furfuryl alcohol (FFA) as  $^1\text{O}_2$  probe.

## 2. Experimental section

**2.1. Laser flash photolysis experiments.** A Nd:YAG laser system instrument (Quanta Ray GCR 130-01) operated at 355 nm (third harmonic) with typical energies of 60 mJ (the single pulse was

~9 ns in duration) was used to study the reactions involving the excited state of 1NN ( $^3\text{1NN}^*$ ). Individual cuvette samples (3 mL volume) were used for a maximum of two consecutive laser shots. The transient absorbance at the pre-selected wavelength was monitored by a detection system consisting of a pulsed xenon lamp (150 W), monochromator and photomultiplier (1P28). A spectrometer control unit was used for synchronising the pulsed light source and programmable shutters with the laser output. The signal from the photomultiplier was digitised by a programmable digital oscilloscope (HP54522A) and analysed by a 32 bits RISC-processor workstation.

Solutions were prepared in Milli-Q water and stability was regularly checked by UV spectroscopy. The decay of  $^3\text{1NN}^*$  and the formation of the radical anion ( $1\text{NN}^{\bullet-}$ ) were monitored at 620 and 380 nm. The pseudo-first order decay and growth constants were obtained by fitting the absorbance vs. time data with single or double exponential equations. The error was calculated from the scattering of experimental data around the fit function. Experiments were performed at ambient temperature ( $295 \pm 2$  °K) on air-equilibrated solutions.

**2.2. Irradiation experiments.** The samples (5 mL total volume) were placed into cylindrical Pyrex glass cells (4.0 cm diameter, 2.3 cm height) closed with a lateral screw cap, and were magnetically stirred during irradiation. Irradiation was carried out under a Philips TLK 05 UVA lamp, with emission maximum at 365 nm. The incident radiation reached the cells mainly from the top, and the optical path length of the solution was  $b = 0.4$  cm. The lamp irradiance between 300 and 400 nm was  $20 \pm 2$  W m<sup>-2</sup>, measured with a power meter by CO.FO.ME.GRA. (Milan, Italy). The incident photon flux ( $(1.6 \pm 0.2) \cdot 10^{-5}$  Einstein L<sup>-1</sup> s<sup>-1</sup>) was actinometrically determined using the ferrioxalate method (Kuhn et al., 2004), taking into account the absorption spectrum of  $\text{Fe}(\text{C}_2\text{O}_4)_3^{3-}$  and the variation with wavelength of the quantum yield of  $\text{Fe}^{2+}$  generation (Albinet et al., 2010a).

Figure 1 reports the spectral photon flux density over irradiated solutions ( $p^\circ(\lambda)$ ), measured with an Ocean Optics SD 2000 CCD spectrophotometer and normalised to the actinometry results, also taking Pyrex transmittance into account. The Figure also reports the absorption spectrum of 1NN ( $\epsilon_{1\text{NN}}(\lambda)$ ), measured with a Varian Cary 100 Scan UV-Vis spectrophotometer.

**2.3. Liquid chromatography determinations.** After irradiation the solutions were allowed to cool for 10-15 min under refrigeration, to minimise volatilisation of 1NN. Analysis was then carried out by High Performance Liquid Chromatography coupled with UV-Vis detection (HPLC-UV). The adopted Merck-Hitachi instrument was equipped with AS2000A autosampler (100  $\mu\text{L}$  sample volume), L-6200 and L-6000 pumps for high-pressure gradients, Merck LiChroCART RP-C18 column packed with LiChrospher 100 RP-18 (125 mm  $\times$  4.6 mm  $\times$  5  $\mu\text{m}$  particle diameter), and L-4200 UV-Vis detector (detection wavelength 220 nm). Isocratic elution was carried out with 60:40  $\text{CH}_3\text{OH}$ : aqueous  $\text{H}_3\text{PO}_4$  (pH 2.8), at 1.0 mL min<sup>-1</sup> flow rate. The retention time of phenol was 2.5 min, that of 1NN 11.2 min, column dead time 0.90 min. FFA analysis was performed using an HPLC system (Waters Alliance) equipped with a diode array detector and a C18 Zorbax column

(Agilent, 15 mm × 4.6 mm × 5 μm). The isocratic eluent was 15:85 CH<sub>3</sub>OH:H<sub>2</sub>O (1:1000 formic acid) at 1.0 mL min<sup>-1</sup> flow rate. The retention time of FFA (detected at 220 nm) was 6.5 min.

**2.4. Fluorescence measurements.** A Varian Cary Eclipse fluorescence spectrofluorimeter was used, adopting a 10 nm bandpass on both excitation and emission. Fluorescence excitation-emission matrix (EMM) was obtained at 5 nm intervals for excitation wavelengths from 200 to 500 nm and emission from 220 to 600 nm. Identification of fluorescent constituents in water samples was performed on the basis of literature data, using the main contours to identify fluorescence peaks.

**2.5. Kinetic data treatment.** The time evolution data of 1NN and phenol were fitted with pseudo-first order equations of the form  $(C_t C_0^{-1}) = e^{-kt}$ , where  $C_t$  is the concentration of the substrate (1NN or phenol) at time  $t$ ,  $C_0$  its initial concentration, and  $k$  (fit variable) the pseudo-first order degradation rate constant. The initial transformation rate is  $R = k C_0$ . Errors on  $k$  were derived from the scattering of experimental data around each fit curve. The reported errors on  $R$  ( $\mu \pm \sigma$ ) largely depended on  $k$  errors, because errors on  $C_0$  (derived from calibration curves) were much lower. The reproducibility of repeated runs was around 10-15%. In some cases the experimental rate data were fitted with rather complex equations having one fit variable. The error associated to such variable depended both on data scattering around the fit function and on uncertainty of constant parameters, which was taken into account with error propagation rules.

**2.6. Identification of transformation intermediates.** Intermediate identification was carried out with gas chromatography interfaced with mass spectrometry (GC-MS). Aqueous solutions after irradiation (salted with NaCl and acidified with HCl) were extracted with 3 mL CH<sub>2</sub>Cl<sub>2</sub>, dehumidified with anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness. Each sample was reconstructed with 100 μL CH<sub>2</sub>Cl<sub>2</sub>. The solution was transferred into a vial and injected into a capillary gas chromatograph (Agilent 6890) coupled with a mass spectrometer (Agilent 5973 inert). The injection system used was a Gerstel CIS4 PTV. Initial injection temperature was 40 °C, programmed at 5 °C/s; final temperature was 320 °C, held for 9 min. The injection volume was 2 μL in the splitless mode. The capillary column used was a HP-5MS, 30 m × 0.25 mm × 0.25 μm film thickness. Initial column temperature was 40 °C and was increased by 15 °C/min to 300 °C. The carrier gas was ultrapure He (flow 1.0 mL min<sup>-1</sup>; SIAD, Bergamo, Italy). The ionisation source worked in the electronic impact (EI) mode and the mass spectrometer worked in the Scan mode from 44 to 450 Th. Identification of spectra was performed with the Wiley 7n library (Agilent Part No. G1035B).

### 3. Results and Discussion

#### 3.1. Laser flash photolysis (LFP) experiments

LFP runs were carried out to study the reaction between  $^3\text{1NN}^*$  and phenol. Figure 2 reports the absorption spectra obtained, 65 ns after the laser pulse, upon 355-nm flash photolysis of 0.1 mM 1NN (1), of 0.1 mM 1NN + 1 mM phenol (2), as well as the difference spectrum ((2)-(1)). The difference spectrum shows a maximum at 380 nm, where both phenoxy radical (phenoxy) and  $1\text{NN}^\bullet$  have absorption peaks (Gadosy et al., 1999; Brigante et al., 2010). Such a finding suggests formation of at least one of the two species. However, the formation of *e.g.* phenoxy from phenol would imply a one-electron reduction of  $^3\text{1NN}^*$ , which would yield  $1\text{NN}^\bullet$  as well (Görner, 2008).

The transient  $^3\text{1NN}^*$  could react with both phenol and ground-state 1NN, in addition to reacting with  $\text{O}_2$  to yield  $^1\text{O}_2$  (Brigante et al., 2010). Figure 3a shows the trend of the pseudo-first order rate constant of  $^3\text{1NN}^*$  disappearance ( $k_{^3\text{1NN}^*}$ ), as a function of phenol concentration. The Stern-Volmer approach (Laws and Contino, 1992) yields  $k_{^3\text{1NN}^*} = k_{^3\text{1NN}^*,\text{Phenol}}[\text{Phenol}] + k_{^3\text{1NN}^*}^o$ , where  $k_{^3\text{1NN}^*,\text{Phenol}}$  is the second-order reaction rate constant between  $^3\text{1NN}^*$  and phenol, and  $k_{^3\text{1NN}^*}^o$  is the pseudo-first order rate constant of  $^3\text{1NN}^*$  disappearance in the absence of phenol. Therefore, the value of  $k_{^3\text{1NN}^*,\text{Phenol}}$  can be obtained as the slope of the line  $k_{^3\text{1NN}^*}$  vs. [Phenol] (Figure 3a). Linear data fit yielded  $k_{^3\text{1NN}^*,\text{Phenol}} = (4.5 \pm 0.1) \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1}$ .

Figure 3b shows the trend of  $k_{^3\text{1NN}^*}$  vs. [1NN], which suggests that some reaction would take place between  $^3\text{1NN}^*$  and 1NN ground state. It is  $k_{^3\text{1NN}^*} = k_{^3\text{1NN}^*,1\text{NN}}[1\text{NN}] + k_{^3\text{1NN}^*}^o$ , from which the second-order rate constant between  $^3\text{1NN}^*$  and 1NN ( $k_{^3\text{1NN}^*,1\text{NN}}$ ) can be determined as the line slope. One gets  $k_{^3\text{1NN}^*,1\text{NN}} = (7.1 \pm 0.9) \cdot 10^8 \text{ M}^{-1} \text{ s}^{-1}$ . From the line intercept ( $k_{^3\text{1NN}^*}^o = (4.97 \pm 0.50) \cdot 10^5 \text{ s}^{-1}$ ) one can estimate  $\text{O}_2$  concentration (Brigante et al., 2010). The resulting  $[\text{O}_2] = 0.25 \pm 0.03 \text{ mM}$  is quite reasonable for an aqueous solution in contact with the atmosphere.

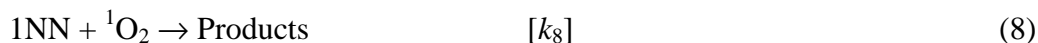
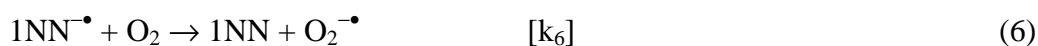
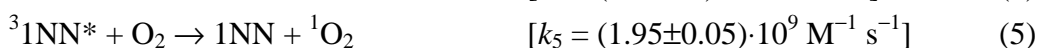
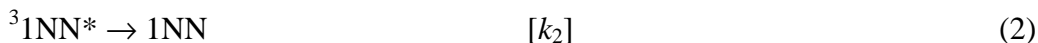
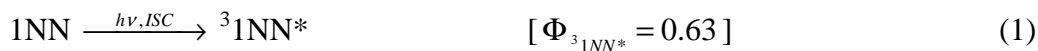
#### 3.2. Steady irradiation experiments

##### 3.2.1. Irradiation of 1NN alone

Solutions containing 0.1 mM 1NN were irradiated at the natural pH  $\sim 6$ , in presence of air and under nitrogen atmosphere. The relevant transformation rates of 1NN were  $R_{1\text{NN}}^{\text{air}} = (6.31 \pm 0.95) \cdot 10^{-9} \text{ M s}^{-1}$  and  $R_{1\text{NN}}^{\text{N}_2} = (9.83 \pm 1.18) \cdot 10^{-9} \text{ M s}^{-1}$ . There is evidence that around 10% of the direct phototransformation of 0.1 mM 1NN in the presence of air is accounted for by photogenerated  $^\bullet\text{OH}$ , probably produced upon water oxidation (Sur et al., 2011). The same process would most likely take place under nitrogen, and the photochemical formation rate of  $^\bullet\text{OH}$  by 0.1 mM 1NN under the adopted irradiation conditions would be  $R_{\text{OH}}^{1\text{NN}} \approx (6.3 \pm 0.9) \cdot 10^{-10} \text{ M s}^{-1}$ .

Considering that 1NN would scavenge practically all photogenerated  $^\bullet\text{OH}$ , the rate of 1NN transformation by  $^\bullet\text{OH}$  would be equal to  $R_{\text{OH}}^{1\text{NN}}$ . Therefore, the rates of 1NN transformation not

involving  $\bullet\text{OH}$  would be  $(R'_{1\text{NN}})_{\text{air}} = (5.68 \pm 1.04) \cdot 10^{-9} \text{ M s}^{-1}$  and  $(R'_{1\text{NN}})_{\text{N}_2} = (9.20 \pm 1.27) \cdot 10^{-9} \text{ M s}^{-1}$  under air and  $\text{N}_2$ , respectively. When the  $\bullet\text{OH}$  pathway is not considered, the following processes would account for the transformation of irradiated 1NN (ISC: inter-system crossing;  $\Phi_{^3\text{1NN}^*}$ : Fournier et al., 1997;  $k_4$ : this work;  $k_5$ : Brigante et al., 2010;  $k_7$ : Rodgers and Snowden, 1982):



Brigante et al. (2010) determined  $k_2 + k_3 + k_4 \cdot [1\text{NN}] = (8.0 \pm 0.1) \cdot 10^4 \text{ s}^{-1}$ . Reaction (3) would contribute to the transformation of 1NN. In presence of air, reaction (4) is expected to take part in 1NN disappearance, because of the likely evolution of  $1\text{NN}^{\bullet+}$  into transformation intermediates, while  $1\text{NN}^{\bullet-}$  would be recycled back to 1NN by oxygen. Under nitrogen atmosphere, the most likely pathway after reaction (4) (in analogy with the behaviour of anthraquinone-2-sulphonate; Maurino et al., 2008) is recombination between  $1\text{NN}^{\bullet+}$  and  $1\text{NN}^{\bullet-}$  to give back 1NN, producing a null cycle.

Although  $\bullet\text{OH}$  photoproduction plays some role in the transformation of 1NN, the fraction of  $^3\text{1NN}^*$  that would be involved in  $\bullet\text{OH}$  generation is negligible (Sur et al., 2011). This also means that only a minor fraction of  $^3\text{1NN}^*$  would be involved into 1NN degradation, the remainder giving back 1NN *via* some process. For this reason,  $\bullet\text{OH}$  production was not considered among the significant  $^3\text{1NN}^*$  sinks in steady-state calculations of  $[^3\text{1NN}^*]$ . Under  $\text{N}_2$  atmosphere, 1NN transformation would be most likely accounted for by reactions (1-4) followed by recombination between  $1\text{NN}^{\bullet+}$  and  $1\text{NN}^{\bullet-}$ . In air-equilibrated solutions, reactions (1-8) would apply.

The fact that 1NN transformation is faster under  $\text{N}_2$  atmosphere than in air-equilibrated solutions, despite the  $1\text{NN}^{\bullet+} + 1\text{NN}^{\bullet-}$  recombination without  $\text{O}_2$ , would be accounted for by the effective scavenging of  $^3\text{1NN}^*$  by  $\text{O}_2$  in reaction (5) (Brigante et al., 2010). The process yields  $^1\text{O}_2$  that contributes to 1NN transformation (reaction (8)), but most  $^1\text{O}_2$  would actually undergo inactivation in reaction (7). Because  $[\text{O}_2] \sim 0.3 \text{ mM}$ , as per its solubility in air-equilibrated solutions and  $[1\text{NN}] = 0.1 \text{ mM}$ , the rate of reaction (5) would be  $\sim 8$  times higher than that of reaction (4). Reaction (5) would act as a very effective quenching process for  $^3\text{1NN}^*$ , inhibiting reactions (3) and (4) and decreasing 1NN transformation rate in the presence of oxygen.

The formation rate of  $^3\text{1NN}^*$  can be calculated as  $R_{^3\text{1NN}^*} = \Phi_{^3\text{1NN}^*} \cdot P_a^{1\text{NN}}$ , where  $P_a^{1\text{NN}} = \int_{\lambda} p^\circ(\lambda) (1 - 10^{-\varepsilon_{1\text{NN}}(\lambda) \cdot b \cdot [1\text{NN}]}) d\lambda = (2.5 \pm 0.3) \cdot 10^{-6} \text{ Einstein L}^{-1} \text{ s}^{-1}$  with  $b = 0.4 \text{ cm}$  and  $[1\text{NN}] = 0.1 \text{ mM}$ . Values of  $p^\circ(\lambda)$  and  $\varepsilon_{1\text{NN}}(\lambda)$  are reported in Figure 1. By applying the steady-state



approximation to  $^3\text{1NN}^*$  in the reaction system relevant to nitrogen atmosphere, one gets  $(R'_{1\text{NN}})_{N_2} = R_{^3\text{1NN}^*} \frac{k_3}{k_2 + k_3 + k_4[1\text{NN}]}$ . Considering that  $R_{^3\text{1NN}^*} = \Phi_{^3\text{1NN}^*} P_a^{1\text{NN}} = (1.6 \pm 0.2) \cdot 10^{-6} \text{ M s}^{-1}$  and  $(R'_{1\text{NN}})_{N_2} = (9.20 \pm 1.27) \cdot 10^{-9} \text{ M s}^{-1}$  (from irradiation experiments), one obtains  $k_3 = (4.6 \pm 1.3) \cdot 10^2 \text{ s}^{-1}$ .

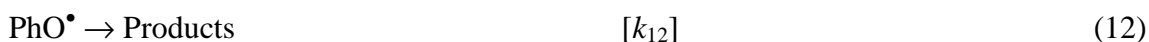
In the presence of air, by applying the steady-state approximation to  $^3\text{1NN}^*$  and  $^1\text{O}_2$  one gets:

$$(R'_{1\text{NN}})_{\text{air}} = R_{^3\text{1NN}^*} \frac{k_3 k_7 + k_3 k_8 [1\text{NN}] + k_5 k_8 [\text{O}_2] [1\text{NN}]}{(k_2 + k_3 + k_4 [1\text{NN}] + k_5 [\text{O}_2]) (k_7 + k_8 [1\text{NN}])} \quad (9)$$

where  $(R'_{1\text{NN}})_{\text{air}} = (5.68 \pm 1.04) \cdot 10^{-9} \text{ M s}^{-1}$ . With the value  $k_3 = (4.6 \pm 1.3) \cdot 10^2 \text{ s}^{-1}$  obtained above, one gets  $k_8 = (8.1 \pm 3.5) \cdot 10^6 \text{ M}^{-1} \text{ s}^{-1}$  as the reaction rate constant between 1NN and  $^1\text{O}_2$ . Under such circumstances, only  $\sim 0.3\%$   $^1\text{O}_2$  would be involved into 1NN transformation in reaction (8), the remainder undergoing thermal inactivation in reaction (7).

### 3.2.2. Irradiation of 1NN with phenol (air-equilibrated solutions)

Figure 4 reports the trends of the initial transformation rates of 1NN ( $R_{1\text{NN}}$ ) and phenol ( $R_{\text{phenol}}$ ), as a function of phenol concentration. The direct photolysis of phenol was negligible when it was irradiated alone. The Figure shows that phenol inhibits the transformation of 1NN, which suggests that the two compounds compete for one or more reactive species. In the presence of air, irradiation of 1NN + phenol (PhOH) would involve reactions (1-8) plus the following, where  $\text{PhO}^\bullet$  is the phenoxy radical (Machado and Boule, 1995; Mark et al., 1996). The constant  $k_{10}$  was obtained by LFP (Figure 3a) and  $k_{13}$  is reported by Mártire et al. (1993):



Reactions (11, 12) are pseudo-first order approximations of more complex processes. Rationale for the approximation is to obtain manageable solutions of kinetic equations. For instance, reaction (11) would most likely involve  $\text{PhO}^\bullet + \text{O}_2^\bullet$  (the latter produced in reaction (6)) to give back phenol (Mack and Bolton, 1999; Vione et al., 2011). Formal inclusion of  $\text{O}_2^\bullet$  in the reaction system would introduce quadratic terms, which would prevent solution of the already complex kinetic equations.

Phenol and 1NN would compete for  $^\bullet\text{OH}$ , which is photogenerated by 1NN with  $R_{\text{OH}}^{1\text{NN}} \approx (6.3 \pm 0.9) \cdot 10^{-10} \text{ M s}^{-1}$  as estimated before. The reaction rate constants with  $^\bullet\text{OH}$  of phenol and 1NN are  $k_{\text{OH}, \text{Phenol}} = (1.4 \pm 0.1) \cdot 10^{10} \text{ M}^{-1} \text{ s}^{-1}$  (Buxton et al., 1988) and  $k_{\text{OH}, 1\text{NN}} = (8.2 \pm 0.4) \cdot 10^7 \text{ M}^{-1} \text{ s}^{-1}$  (Sur et al., 2011). The kinetic system made up of equations (1-8) and (10-13), plus  $^\bullet\text{OH}$ -related processes qualitatively predicts inhibition by phenol of 1NN phototransformation, as

actually observed (Figure 4). However, the relevant calculations overestimate the effect of phenol concentration on 1NN degradation rate: experimental data show that inhibition of 1NN degradation by phenol is significantly lower than predicted. The key point is that reaction (10) transforms  $^3\text{1NN}^*$  into  $1\text{NN}^\bullet$ , which is recycled back to 1NN by  $\text{O}_2$  in reaction (6). In this way, the  $^3\text{1NN}^*$ -assisted transformation of 1NN upon reactions (3,4,5,8) is inhibited. To account for experimental data, one should assume that a fraction  $\alpha$  of  $1\text{NN}^\bullet$  reacts with phenol-derived species to produce transformation intermediates, instead of giving back 1NN. By applying the steady-state approximation to  $^{\bullet}\text{OH}$ ,  $^3\text{1NN}^*$  and  $^1\text{O}_2$ , one gets the following expressions for  $R_{1\text{NN}}$  and  $R_{\text{Phenol}}$ :

$$R_{1\text{NN}} = R_{^3\text{1NN}^*} \frac{(k_3 + \alpha k_{10} [\text{Phenol}])(k_7 + k_8 [1\text{NN}] + k_{13} [\text{Phenol}] + k_5 k_8 [\text{O}_2] [1\text{NN}])}{(k_2 + k_3 + k_4 [1\text{NN}] + k_5 [\text{O}_2] + k_{10} [\text{Phenol}])(k_7 + k_8 [1\text{NN}])} + R_{^{\bullet}\text{OH}}^{1\text{NN}} \frac{k_{\bullet\text{OH},1\text{NN}} [1\text{NN}]}{k_{\bullet\text{OH},\text{Phenol}} [\text{Phenol}] + k_{\bullet\text{OH},1\text{NN}} [1\text{NN}]} \quad (14)$$

$$R_{\text{Phenol}} = R_{^3\text{1NN}^*} \frac{[k_{12} (k_{11} + k_{12})^{-1} k_{10} [\text{Phenol}](k_7 + k_8 [1\text{NN}] + k_{13} [\text{Phenol}])] + k_5 k_{13} [\text{O}_2] [\text{Phenol}]}{(k_2 + k_3 + k_4 [1\text{NN}] + k_5 [\text{O}_2] + k_{10} [\text{Phenol}])(k_7 + k_8 [1\text{NN}] + k_{13} [\text{Phenol}])} + R_{^{\bullet}\text{OH}}^{1\text{NN}} \frac{k_{\bullet\text{OH},\text{Phenol}} [\text{Phenol}]}{k_{\bullet\text{OH},\text{Phenol}} [\text{Phenol}] + k_{\bullet\text{OH},1\text{NN}} [1\text{NN}]} \quad (15)$$

The fit with equation (14) of the  $R_{1\text{NN}}$  data reported in Figure 4, with  $\alpha$  as the only floating variable, yielded  $\alpha = (1.5 \pm 1.3) \cdot 10^{-3}$ . This means that  $<0.3\%$  of  $1\text{NN}^\bullet$  would evolve into transformation intermediates, the remainder being recycled back to 1NN by oxygen. The fit with equation (15) of the  $R_{\text{Phenol}}$  data of Figure 4, with  $\beta = k_{12} (k_{11} + k_{12})^{-1}$  as the only floating variable yielded  $\beta = (6.8 \pm 3.4) \cdot 10^{-2}$ . This means that  $\leq 10\%$   $\text{PhO}^\bullet$  would evolve into transformation intermediates, the remainder yielding back phenol (reaction 11).

The good agreement between experimental data and model predictions (Figure 4) suggests that both 1NN and phenol would undergo transformation *via*  $^3\text{1NN}^*$ ,  $^1\text{O}_2$  and  $^{\bullet}\text{OH}$ . The scavenging of  $^3\text{1NN}^*$  by phenol (reaction 10) would account for most of phenol sensitised transformation, and it would inhibit the degradation of 1NN. The latter issue applies because over 99% of  $1\text{NN}^\bullet$  produced in reaction (10) would be recycled back to 1NN by  $\text{O}_2$ . In the absence of reaction (10),  $^3\text{1NN}^*$  would be involved in 1NN transformation *via* reactions (3,4,5,8).

### 3.3. Transformation intermediates

Several important transformation intermediates of phenol could be detected by GC-MS. Figure 5 shows their mass spectra and reports comparison with library spectra (match  $>90\%$ ). The relevant intermediates are 2- and 4-phenoxyphenol, 1,1'-dihydroxybiphenyl, and 4,4'-dihydroxybiphenyl. These compounds would arise upon dimerisation of  $\text{PhO}^\bullet$  (Gopalan and Savage 1994; Platz et al., 1998; Neta and Grodkowski, 2005), produced in reaction (10). The dimerisation rate is expected to increase with increasing phenol concentration (Maurino et al., 2008) and, coherently, GC-MS peaks of intermediates were considerably higher at 1 mM than at 0.1 mM phenol. Note that such concentration values are somewhat higher than the levels of phenolic compounds in fog water (up

to 30  $\mu\text{M}$ , Harrison et al., 2005), but they are in the range of concentrations that phenols can reach in dew water (Polkowska et al., 2008) or in particles from biomass burning (Vicente et al., 2012).

Considering that most of the electron density of  $\text{PhO}^\bullet$  would be localised at the oxygen atom and at the *ortho* and *para* carbon atoms of the ring (Fehir and McCusker, 2009), the structure of the identified intermediates can be easily accounted for. The phenoxyphenols would be formed by addition of the O atom of  $\text{PhO}^\bullet$  to the electron-rich (*i.e.*, 2- and 4-) ring positions of another  $\text{PhO}^\bullet$ . The dihydroxybiphenyls would result from an addition process, involving the #2 and #4 ring C atoms of  $\text{PhO}^\bullet$ . Further H-atom rearrangement would then yield the detected intermediates.

No transformation intermediates of 1NN were detected. Possible explanations are instability preventing accumulation, poor extraction by  $\text{CH}_2\text{Cl}_2$ , or non-amenability by GC.

### 3.4. Fluorescence measurements

The variation of fluorescence intensity of solutions upon irradiation was studied by means of the Excitation-Emission Matrix (EEM) technique. Preliminary experiments (data not shown) indicated that 1NN has no fluorescence. Figure 6 reports the EEM spectra of: (a) 0.1 mM 1NN + 0.1 mM phenol before irradiation (fluorescence is accounted for by phenol); (b) 0.1 mM 1NN + 0.1 mM phenol after 10 min irradiation; (c) 0.1 mM 1NN + 0.1 mM phenol after 4 h irradiation, and (d) 0.1 mM 4-phenoxyphenol without irradiation. Irradiation of phenol and 1NN shifted the fluorescence emission maximum from  $\sim 300$  nm to 330-400 nm (Figure 6b). Upon prolonged irradiation (4 h) a further, small fluorescence peak appeared at  $\text{Ex/Em} = 300\text{-}450/400\text{-}450$  nm (Figure 6c).

The fluorescence peak of Figure 6b partially overlaps with fluorescence emission by 4-phenoxyphenol (Figure 6d), a transformation intermediate detected in the studied system (see section 3.3), but the fluorescence maximum at  $\text{Ex/Em} = 275/350$  nm does not. Substances with  $\text{Ex/Em} = 250\text{-}300/330\text{-}400$  nm would be classified as “M-like” fulvic acids (Parlanti et al., 2000; Mostofa et al., 2011). Moreover, the interval  $\text{Ex/Em} = 300\text{-}450/400\text{-}450$  nm (Figure 6c) is right into the range of atmospheric humic-like substances (HULIS) (Muller et al., 2008). Fluorescence data suggest that irradiation of phenol, in the presence of 1NN as photosensitiser, induces formation of compounds that would be classified as “M-like” fulvic acids and HULIS if found in environmental samples (Muller et al., 2008; Mostofa et al., 2011). The fluorescence of this material only partially overlaps with that of phenol dimer, suggesting that additional and more important fluorescent compounds are being formed. Interestingly, the sensitised phototransformation of 4-phenoxyphenol is known to produce higher oligomers, such as phenol tetramers (Net et al., 2010). Moreover, HULIS formation from phenols might be an important process in the atmosphere, where phenolic compounds arising from wood burning are quite common (Kuo et al., 2011; Shakya et al., 2011).

### 3.5. Formation of singlet oxygen by the irradiated system 1NN + phenol

Figure 7 reports the time evolution of 1NN, phenol and two fluorescence contours, monitored at Ex/Em = 275/375 nm (fulvic-like) and Ex/Em = 330/415 nm (HULIS), upon irradiation of 0.1 mM 1NN and 60  $\mu$ M phenol. After 8 h irradiation, phenol and 1NN practically disappeared and “fulvic-like” fluorescence reached a plateau (intensity  $\sim$ 150 A.U.). Fluorescence intensity was remarkably stable in the plateau region, with almost no change at least up to 24 h irradiation. The second fluorescence signal, attributed to HULIS, peaked at around 3 h irradiation (intensity  $\sim$  95 A.U.). The insert of Figure 7 reports the trends up to 2 h irradiation, showing very fast increase of the “fulvic-like” signal.

The material formed after 8 h irradiation was additionally studied for photochemical properties, in particular concerning the ability to produce  $^1\text{O}_2$ . Therefore, 10  $\mu$ M FFA was added as  $^1\text{O}_2$  probe to the irradiated system, and FFA degradation was monitored. Figure 8 reports the time trend of 10  $\mu$ M FFA when irradiated alone (direct photolysis, filled circles), and when it was added to solutions that initially contained 0.1 mM 1NN (empty circles) and 0.1 mM 1NN + 60  $\mu$ M phenol (triangles). The latter two solutions were both irradiated for 8 h before the FFA spike. The rationale for irradiating FFA alone, or spiked to pre-irradiated 1NN (blank experiments), was to assess direct FFA photolysis and photochemical activity of possible 1NN photointermediates. Indeed, irradiated 1NN is an effective  $^1\text{O}_2$  source (Brigante et al., 2010), and its (unidentified) intermediates might produce singlet oxygen as well. Note that irradiation of 1NN or FFA alone yielded no fluorescent compounds. Moreover, phenol alone does not absorb above 300 nm, and it did not undergo direct photolysis nor showed fluorescence variation upon irradiation. It was also unable to sensitise FFA transformation.

When FFA was spiked to pre-irradiated 1NN + phenol, its degradation rate was considerably higher compared to blank experiments. Therefore, Figure 8 suggests that the material formed upon irradiation of phenol + 1NN induced photochemical production of  $^1\text{O}_2$ , in addition to showing fluorescence properties analogous to “M-like” fulvic acids and HULIS.

## 4. Conclusions

The triplet state of 1NN reacts with phenol, with rate constant  $k_{^3\text{1NN}^*,\text{Phenol}} = (4.5 \pm 0.1) \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1}$ . The reaction gives phenoxyphenols and dihydroxybiphenyls, coherently with a primary process that would likely produce  $^1\text{NN}^\bullet$  and the phenoxy radical,  $\text{PhO}^\bullet$ . Note that  $\leq 10\%$   $\text{PhO}^\bullet$  is expected to undergo dimerisation, the remainder giving back phenol upon reaction with *e.g.*  $\text{O}_2^\bullet$ . Moreover,  $^1\text{NN}^\bullet$  would be mostly ( $>99\%$ ) recycled back to 1NN by oxygen. Experimental data are accounted for by a kinetic model made up of reactions (1-8) and (10-13), plus  $^\bullet\text{OH}$  formation by  $^3\text{1NN}^*$  and  $<0.3\%$  transformation of  $^1\text{NN}^\bullet$  into intermediates. The majority of phenol would be transformed by  $^3\text{1NN}^*$ , while  $^\bullet\text{OH}$  and  $^1\text{O}_2$  would play a less important role. Phenol inhibits the transformation of 1NN, because the two substrates compete for reaction with  $^3\text{1NN}^*$ ,  $^1\text{O}_2$  and  $^\bullet\text{OH}$ .

A non-irradiated solution containing phenol and INN shows the typical fluorescence emission of phenol. Irradiation causes a red shift of fluorescence excitation and emission wavelengths, from Ex/Em = 220-230/280-320 nm and 250-275/280-320 nm (phenol), to Ex/Em = 250-300/330-400 nm and Ex/Em = 300-450/400-450 nm (contours detected upon irradiation). Compounds with Ex/Em = 250-300/330-400 nm would be identified as “M-like” fulvic substances in environmental samples, while Ex/Em = 300-450/400-450 nm is in the fluorescence range of HULIS. This means that the relevant compounds would be identified as HULIS if found in environmental samples, based on their fluorescence properties. Another interesting finding is that the irradiated material is able to photochemically produce  $^1\text{O}_2$ , thereby showing photosensitisation properties. The potential atmospheric importance of the studied reactions is supported by the fact that both nitroaromatics and phenols are primary and/or secondary atmospheric pollutants, derived from combustion processes. The present work also suggests that EEM fluorescence would be an extremely useful tool to assess the photochemical formation of humic-like and fulvic-like substances.

An issue that deserves further investigation deals with the comparison between humic (or humic-like) substances in the atmosphere *vs.* surface waters. As far as the latter compartment is concerned, photosensitisers such as quinones and aromatic carbonyls can be found in humic and fulvic acids and account for a considerable fraction of their photoactivity (Cory and McKnight, 2005; Canonica et al., 2005; Jammoul et al., 2009; Maurino et al., 2011). There is evidence that atmospheric photosensitisers are less active than those found in surface waters (Albinet et al., 2010b; De Laurentiis et al., 2012). The finding here reported that material with HULIS properties can produce  $^1\text{O}_2$  when irradiated might give some hints about HULIS photoactivity, and could be scope for further research.

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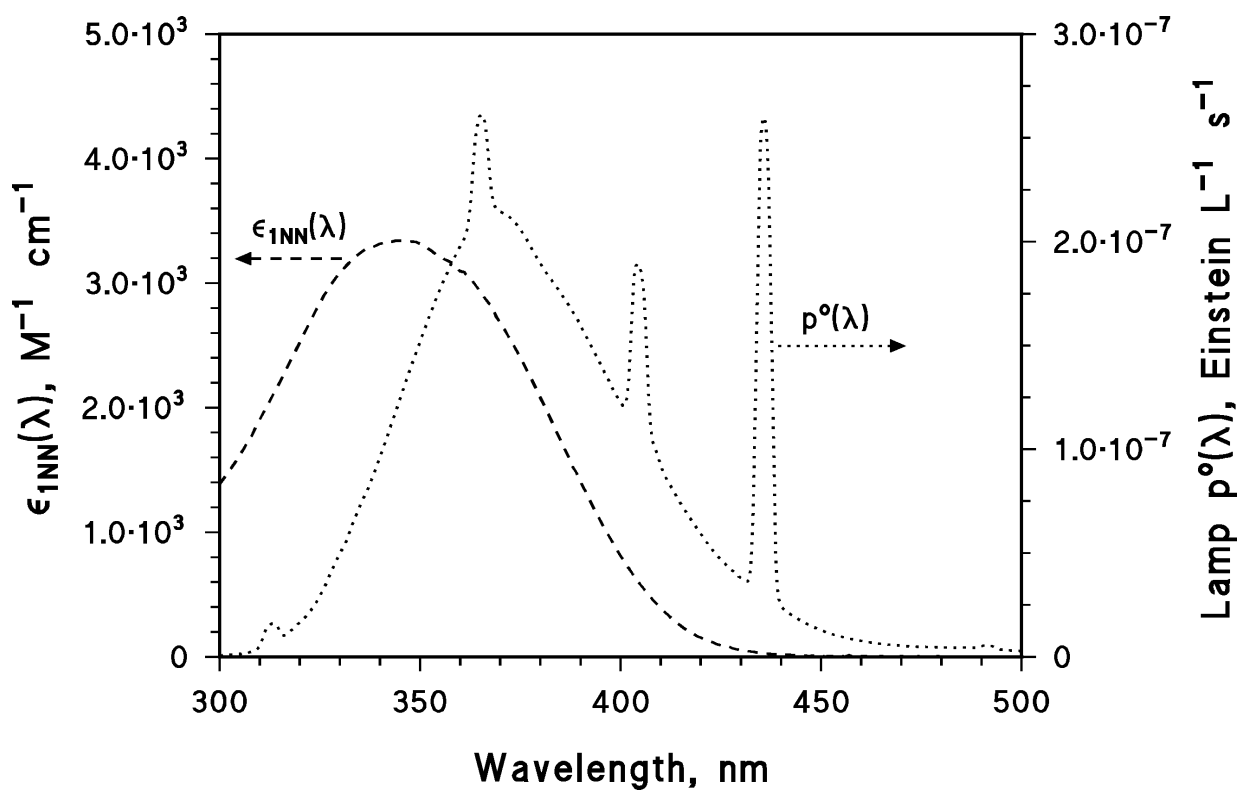
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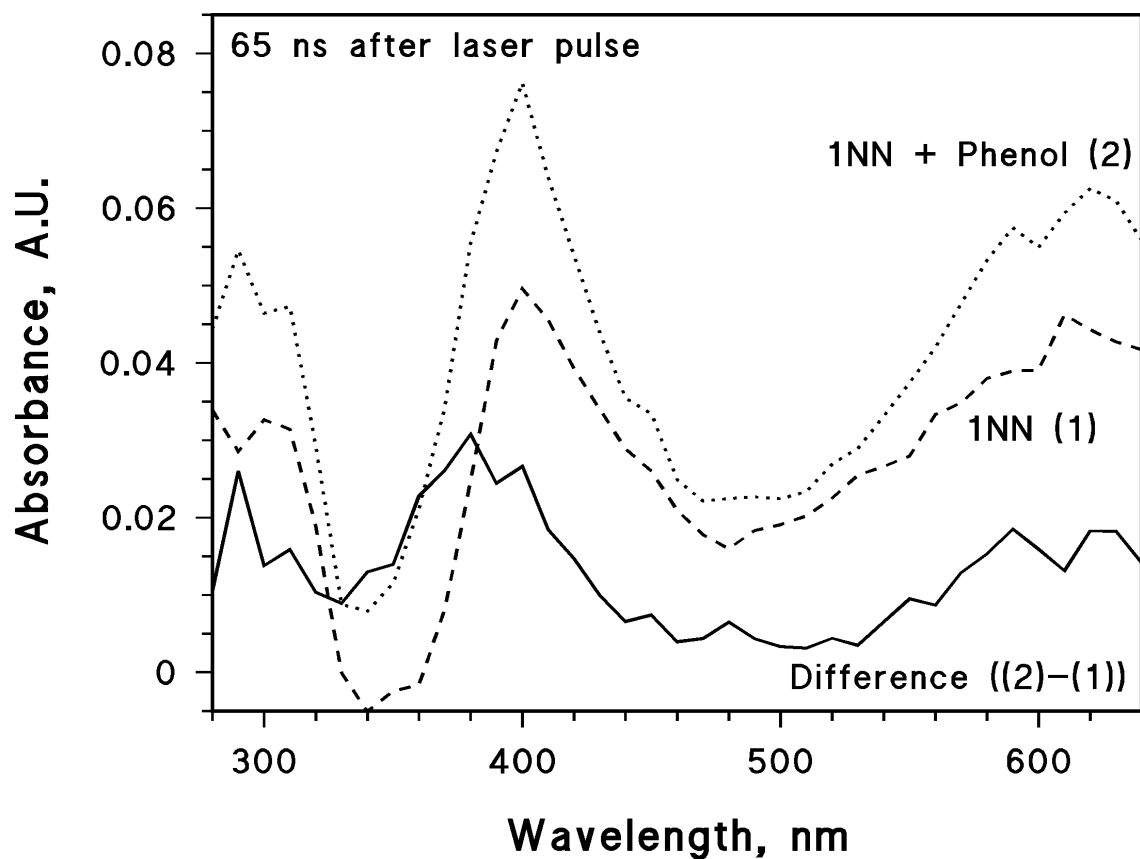
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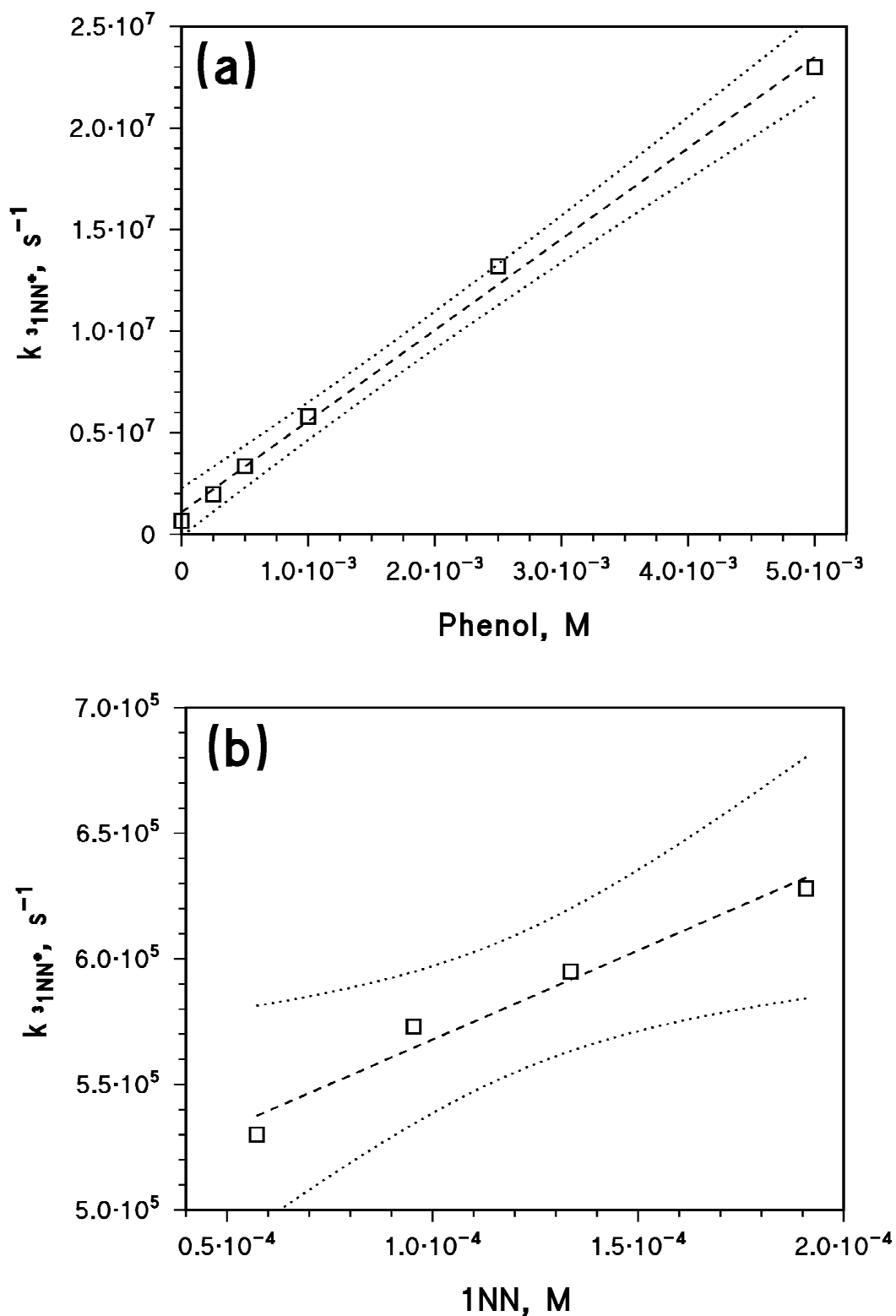
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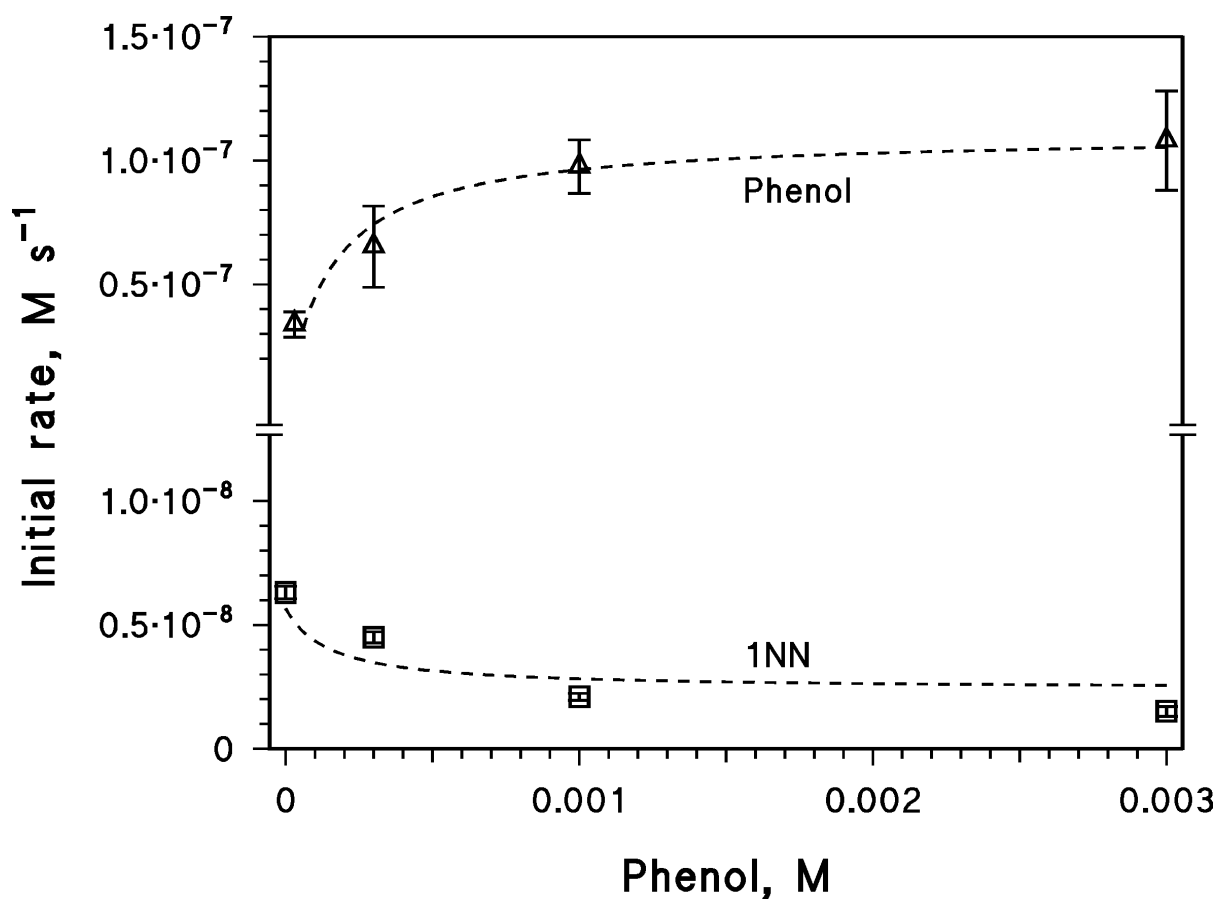
**Figure 1.** Absorption spectrum (molar absorption coefficient  $\epsilon_{1NN}(\lambda)$ ) of 1NN. Incident spectral photon flux density ( $p^\circ(\lambda)$ ) of the adopted UVA lamp (Philips TLK 05).



**Figure 2.** Absorption spectra of transient species formed 65 ns after the laser pulse (355 nm, 60 mJ), upon flash photolysis of air-equilibrated solutions containing: (1) 0.1 mM 1NN; (2) 0.1 mM 1NN + 1 mM phenol. The difference spectrum ((2)-(1)) is also reported.

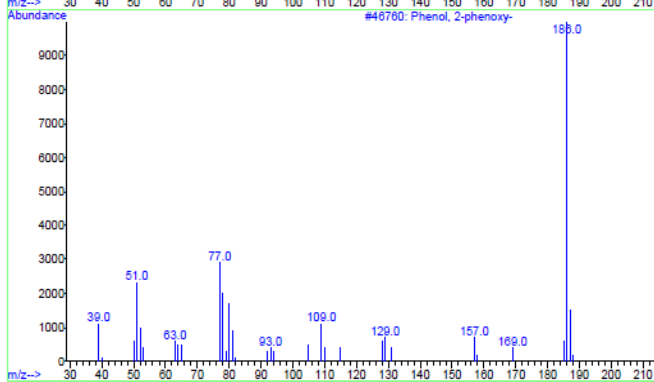
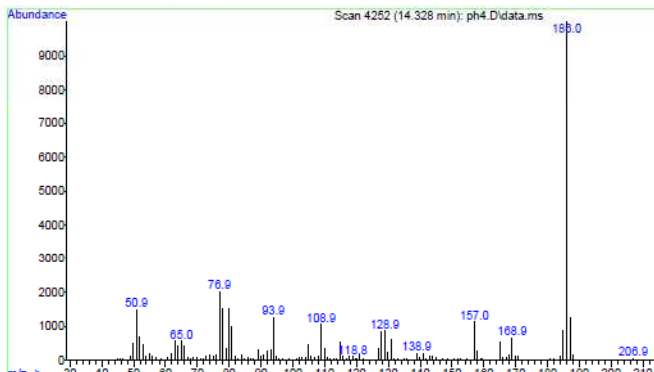


**Figure 3.** Trend of the first-order rate constant of  $^3\text{1NN}^*$  decay ( $k_{^3\text{1NN}^*}$ ), as a function of (a) phenol concentration, and (b) concentration of 1NN. Laser pulse: 355 nm, 60 mJ, air-equilibrated solutions.

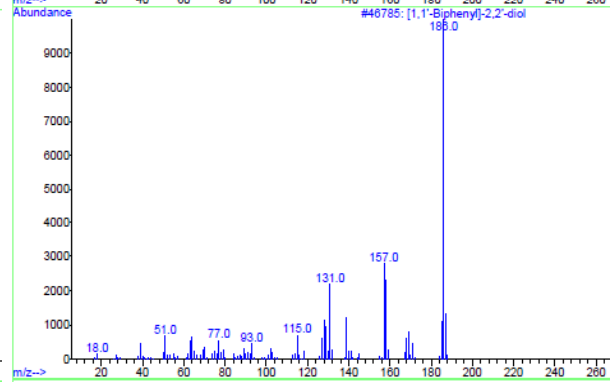
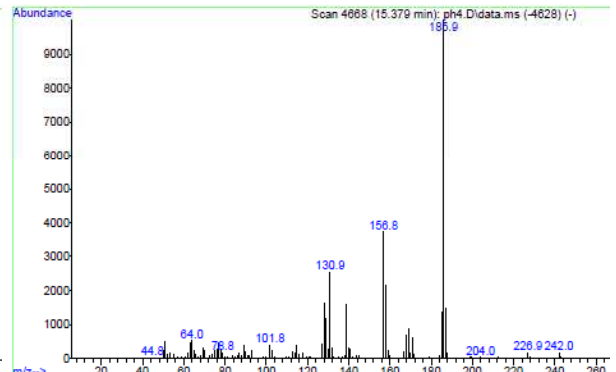


**Figure 4.** Trend with phenol concentration of the initial transformation rates of phenol and 1NN. The dashed lines are the fit of experimental data with equation (15) (phenol) and equation (14) (1NN). Air-equilibrated solutions, UVA irradiation, pH 6.

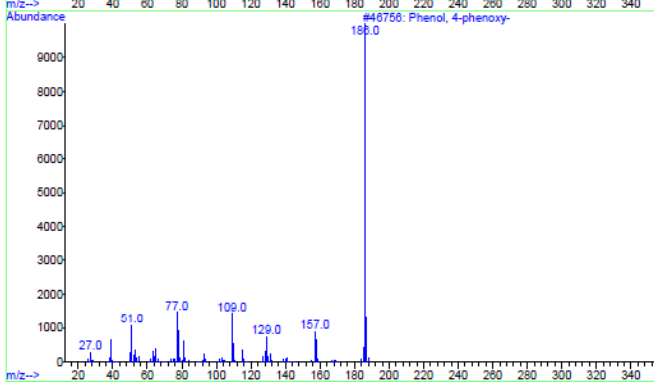
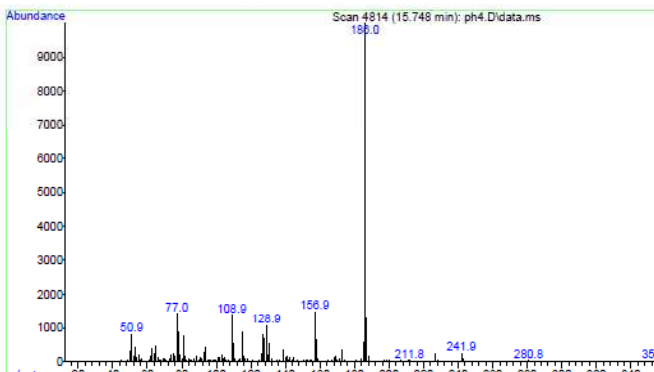
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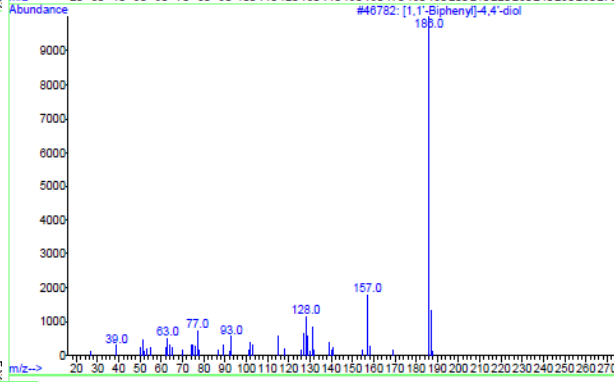
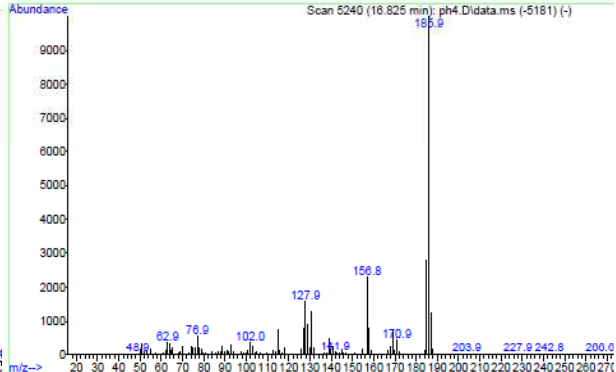
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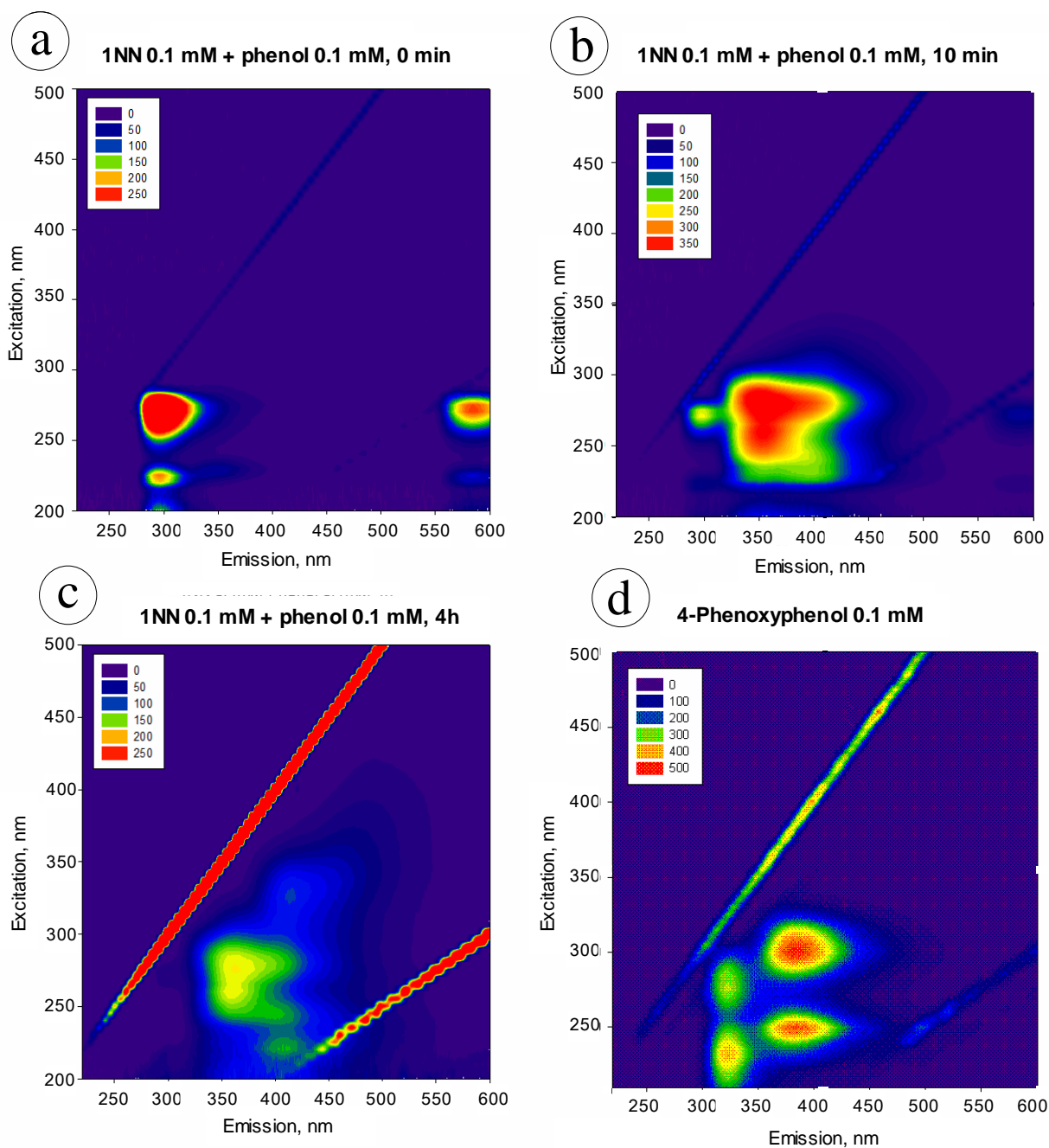
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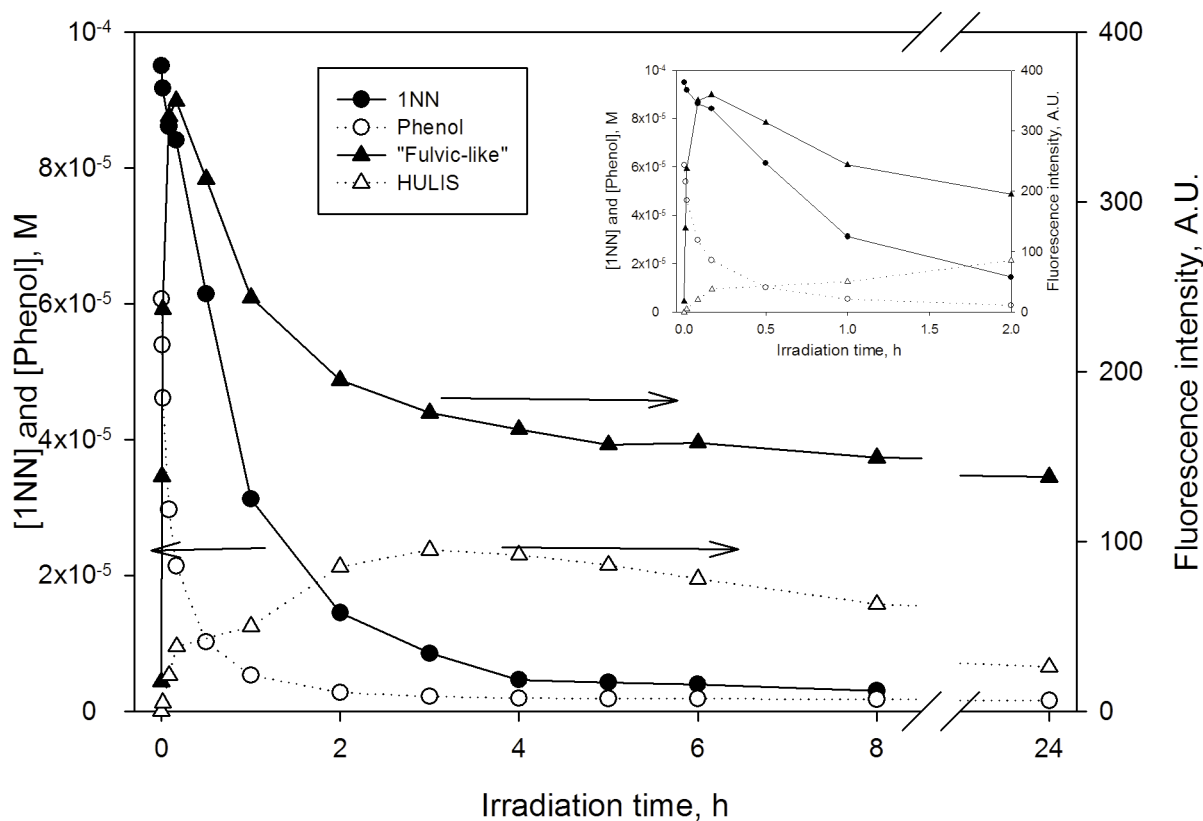
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**Figure 5.** Mass spectra of the detected transformation intermediates (GC-MS analysis). Intermediate structure and library mass spectra are also reported. Peak retention times (minutes) are in brackets over each spectrum.

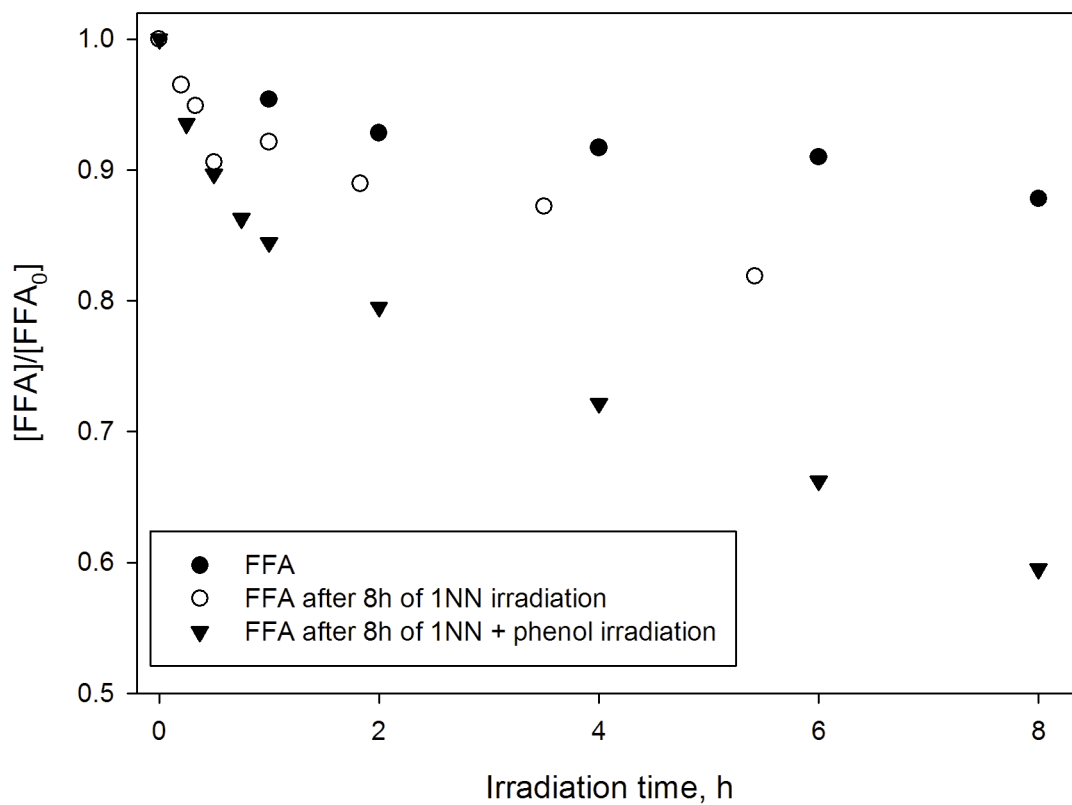


**Figure 6.** Excitation-Emission Matrix (EEM) fluorescence spectra of: (a) 0.1 mM 1NN + 0.1 mM phenol, before irradiation; (b) 0.1 mM 1NN + 0.1 mM phenol, after 10 min irradiation; (c) 0.1 mM 1NN + 0.1 mM phenol, after 4 h irradiation; (d) 0.1 mM 4-phenoxyphenol.



**Figure 7.** Time trend of 0.1 mM 1NN and 60  $\mu$ M phenol upon irradiation. Trend of the related intensities of fluorescence, monitored at  $\lambda_{\text{Ex,max}} = 275 \text{ nm}$  /  $\lambda_{\text{Em,max}} = 375 \text{ nm}$  (Fulvic-like, filled triangles) and at  $\lambda_{\text{Ex,max}} = 330 \text{ nm}$  /  $\lambda_{\text{Em,max}} = 415 \text{ nm}$  (HULIS, empty triangles). The insert shows the trends from 0 up to 2 hours of irradiation.





**Figure 8.** Degradation of 10  $\mu\text{M}$  FFA in Milli-Q water (filled circles), when added to 0.1 mM 1NN solution previously irradiated for 8 h (empty circles), and when added to 0.1 mM 1NN + 60  $\mu\text{M}$  phenol previously irradiated for 8 h (filled triangles).