

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Conditional inactivation of Neuropeptide Y-Y1 receptors unravels the role of Y1 and Y5 receptors co-expressing neurons in anxiety.

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/143746> since 2016-01-07T16:45:54Z

Published version:

DOI:10.1016/j.biopsych.2014.01.009

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)



UNIVERSITÀ DEGLI STUDI DI TORINO

This Accepted Author Manuscript (AAM) is copyrighted and published by Elsevier. It is posted here by agreement between Elsevier and the University of Turin. Changes resulting from the publishing process - such as editing, corrections, structural formatting, and other quality control mechanisms - may not be reflected in this version of the text. The definitive version of the text was subsequently published in *BIOLOGICAL PSYCHIATRY*, ahead of print, 2014, 10.1016/j.biopsych.2014.01.009.

You may download, copy and otherwise use the AAM for non-commercial purposes provided that your license is limited by the following restrictions:

- (1) You may use this AAM for non-commercial purposes only under the terms of the CC-BY-NC-ND license.
- (2) The integrity of the work and identification of the author, copyright owner, and publisher must be preserved in any copy.
- (3) You must attribute this AAM in the following format: Creative Commons BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/deed.en>), 10.1016/j.biopsych.2014.01.009

The definitive version is available at:

<http://linkinghub.elsevier.com/retrieve/pii/S0006322314000481>

Conditional Inactivation of Neuropeptide Y Y1 Receptors Unravels the Role of Y1 and Y5 Receptors Coexpressing Neurons in Anxiety

Angela Longo, Paolo Mele, Ilaria Bertocchi, Alessandra Oberto, Alessia Bachmann, Alessandro Bartolomucci, Paola Palanzaf, Rolf Sprengelg, Carola Eva

From the Neuroscience Institute of the Cavalieri-Ottolenghi Foundation (AL, PM, IB, AO, CE), Orbassano (Turin), and Neuroscience Institute of Turin (AO, CE), Department of Neuroscience (AL, PM, IB, AO, CE), and Departments of Anatomy, Pharmacology, and Forensic Medicine (ABac), University of Turin, Turin, Italy; Department of Integrative Biology and Physiology (ABar), University of Minnesota, Minneapolis, Minnesota; Department of Evolutionary and Functional Biology (PP), University of Parma, Parma, Italy; and Department of Molecular Neurobiology (RS), Max Planck Institute for Medical Research, Heidelberg, Germany.

Address correspondence to Carola Eva, Ph.D., Neuroscience Institute of the Cavalieri-Ottolenghi Foundation (NICO), Regione Gonzole 10, University of Turin, 10043 Orbassano (Turin), Italy; E-mail: carola.eva@unito.it.

Authors AL and PM contributed equally to this work.

Abstract

Background The Y1 receptor (Y1R) and Y5 receptor (Y5R) for neuropeptide Y share similar actions in the regulation of anxiety. Previously demonstrated that conditional removal of the Y1R during postnatal development in the forebrain excitatory neurons leads to higher anxiety, increased hypothalamus-pituitary-adrenocortical axis activity, and decreased body growth rate in male mice raised by foster mothers that exhibit high levels of maternal care. In the present study, we used the same conditional system to analyze the specific contribution to emotional behavior and stress response of the Y1R coexpressed with the Y5R.

Methods Using the Cre-loxP recombination system, we investigated anxious behavior, spatial memory, and metabolic functions of conditional knockout mice in which the inactivation of the *Npy1r* gene was induced in the Y5Rs expressing neurons of juvenile mice (*Npy1rY5R^{-/-}*).

Results *Npy1rY5R^{-/-}* mice show increased anxiety-related behavior but no changes in hypothalamus-pituitary-adrenocortical axis activity or in body weight growth, independently of gender and mouse strain used as foster mothers. Also, *Npy1rY5R^{-/-}* mice of both genders display increased spatial reference memory in the Morris water maze test.

Conclusions The results suggest that neuropeptide Y Y1R differentially expressed in the limbic system regulates anxiety and stress responses via distinct neurochemical circuits. In addition, we provide the first experimental genetic evidence that the Y1Rs coexpressed with the Y5R are involved in retention of spatial memory in male and female mice.

Key words: Behavioral flexibility; conditional knockout mice; Cre-loxP system; GABA; hypothalamus-pituitary-adrenocortical axis; spatial memory.

Introduction

Neuropeptide Y (NPY) is widely distributed in the central nervous system, where it is involved in the regulation of several biological functions, including emotionality and stress reactions, energy balance, and cognition 1, 2 and 3. In humans, NPY haploinsufficiency is correlated with brain responses to emotional and stress challenges and to anxiety (4). In rodents, injection of NPY into the third ventricle or into the limbic system reduces both anxiety and stress responses 5, 6 and 7. In the brain, NPY interacts with a family of G-protein-coupled receptors that includes the Y1 (Y1R), Y5 (Y5R), and Y2 receptors (the last one considered to function mainly as a presynaptic receptor on NPYergic terminals) 8 and 9.

Pharmacologic and genetic studies suggest that NPY induces anxiolytic effects via activation of the Y1Rs in amygdala, hippocampus, and locus coeruleus 10, 11, 12 and 13. We showed more recently that conditional inactivation of *Npy1r* gene in limbic excitatory neurons of *Npy1rrfb* (*rfb* = reduced forebrain expression) mice increases anxiety level and hypothalamus-pituitary-adrenocortical (HPA) axis activity and decreases body weight growth. Differences in phenotype between *Npy1rrfb* conditional mutants and their control littermates became apparent only when both genotypes were raised by dams exhibiting high levels of maternal care, suggesting that the Y1Rs expressed in limbic principal neurons are key targets of maternal care-induced programming of anxiety and energy homeostasis (10).

The Y5Rs and Y1Rs have overlapping function in regulating anxiety (14). The genes *Npy1r* and *Npy5r* are located on the same chromosome in humans and rodents, displaying an opposite transcriptional orientation and a partly overlapping gene structure (15). In rodents, the Y1R and Y5R are colocalized in several forebrain regions, including the basolateral amygdala (BLA) and hippocampal neurons 16 and 17. We hypothesized that the coordinated expression of the Y1R and Y5R might be required for the regulation of anxiety, spatial learning, and memory. In agreement, pharmacologic studies indicated that NPY induces anxiolytic effects via activation of the Y5R in the BLA. More recently, Domschke et al. (18) suggested that *Npy5r* gene variants may be associated with panic disorder, which is consistent with a role for the Y5R in anxiety disorders 14 and 19.

In the present study, we generated conditional knockout mice in which the inactivation of *Npy1r* was induced in Y5R-expressing neurons of adolescent mice (*Npy1rY5R^{-/-}*). We achieved this by combining the gene targeted floxed *Npy1r* alleles (10) and the inducible Cre recombinase transgene (Cre) that is transcriptionally controlled by a bacterial artificial chromosome-encoded *Npy5r* promoter-driven tetracycline suppressible transactivator (tTA) (16). Our data reveal that conditional ablation of the Y1R in Y5R-expressing neurons results in increased anxiety-related behavior and improved spatial reference memory but no changes in basal or stress-activated HPA axis or body weight growth.

Methods and Materials

Animals

Mice were housed in groups of two to six in a temperature-controlled ($22 \pm 1^\circ\text{C}$) and humidity-controlled ($50 \pm 10\%$) room on a 12-hour light/dark cycle (8:00 am–8:00 pm) and had ad libitum access to food and water. All experiments were conducted in accordance with the European Community Council Directive of 24 November 1986 86/609/EEC and 6106/10/EU and approved by the University of Turin Ethical Committee for animal research and by the Italian Ministry of Health (License No. 180/2006-B).

Generation of *Npy1r*^{Y5R-/-} Conditional Mutants

The region-specific inactivation of *Npy1r* gene in Y5R-expressing neurons in adolescent mice was achieved by using three different mouse lines: 1) *Npy1r*^{2lox}, carrying a modified *Npy1r* allele in which two loxP sites flank the coding region of the *Npy1r* gene (10); 2) *Tg*^{Y5R-htTA/Y1RVenus}, expressing a doxycycline (Dox)-sensitive, synthetic transcription factor (tTA) under the control of the *Npy5r* promoter (16); and 3) *Tg*^{LC1}, encoding a tTA inducible Cre transgene (20) (Figure 1A). Both loxP sites are DNA targets for Cre, which catalyzes site-specific recombination between the loxP sites and the removal of the *Npy1r* coding region (21). First, we generated two mouse lines, *Npy1r*^{2lox}/*Tg*^{Y5R-htTA/Y1RVenus} mice and *Npy1r*^{2lox}/*Tg*^{LC1} mice, which were inter-crossed providing offspring that were used as control animals (summarized as *Npy1r*^{2lox} mice) as well as offspring *Npy1r*^{2lox}/*Tg*^{Y5R-htTA/Y1RVenus}/*LC1* that contained all genetic elements necessary for the inactivation of the *Npy1r* gene specifically in Y5R-containing neurons (summarized as *Npy1r*^{Y5R-/-} mice) (Figure 1B).

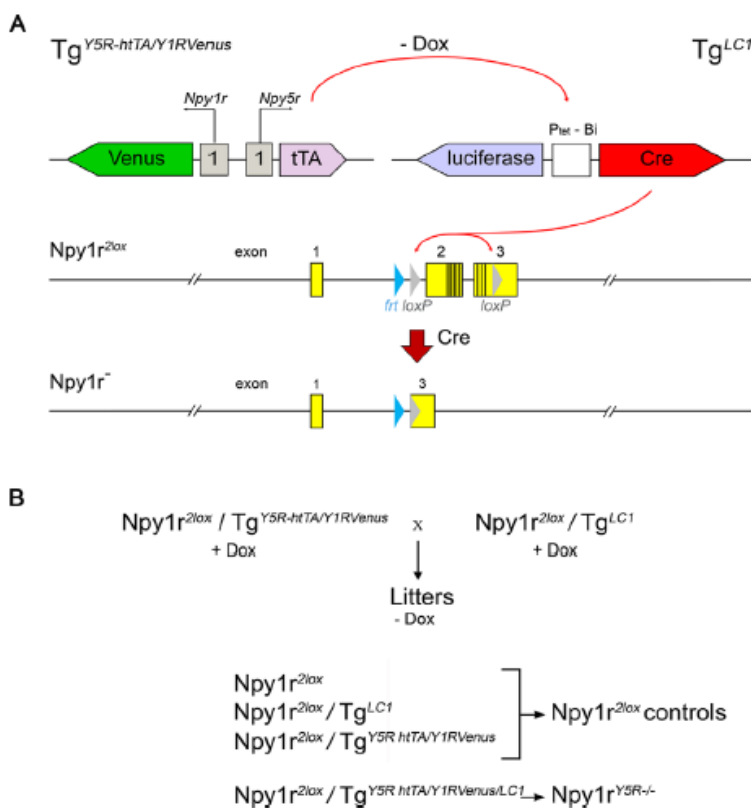


Figure 1. Generation of *Npy1r*^{Y5R-/-} mutants. **(A)** Diagram depicting the interaction of the different genetic components. After doxycycline (Dox) removal at postnatal day (P) 0, the *Npy5r* promoter-driven Dox-sensitive, synthetic transcription activator (tTA) activates transcription of the transgene *Tg*^{LC1}, inducing Cre recombinase (Cre) expression in neurons containing the Y5 receptor (Y5R) subtype. Cre interacts with loxP sites in the gene-targeted *Npy1r*^{2lox} alleles (10) and removes the *Npy1r*^{2lox} coding region leading to the inactivation of the *Npy1r* gene (*Npy1r*⁻). *Frt* and loxP sites are in blue and gray triangles, respectively; exons are in open boxes, coding regions are in gray boxes; and transmembrane spanning codons are in black boxes. **(B)** By mating the compound transgenic mice *Npy1r*^{2lox}/*Tg*^{Y5R-htTA/Y1RVenus} and *Npy1r*^{2lox}/*Tg*^{LC1} under Dox treatment, pups with four different genotypes were generated and found in a Mendelian ratio. At the day of birth (P0), the litters were transferred to Dox-naïve foster mothers to induce the Cre-mediated *Npy1r* gene inactivation in Y5R-containing neurons of *Npy1r*^{2lox}/*Tg*^{Y5R-htTA/Y1RVenus}/*LC1* (*Npy1r*^{Y5R-/-}). Littermates comprising *Npy1r*^{2lox}/*Tg*^{Y5R-htTA/Y1RVenus}, *Npy1r*^{2lox}/*Tg*^{LC1}, and *Npy1r*^{2lox} genotypes were used as controls (*Npy1r*^{2lox} controls).

Long-term treatment of mothers with Dox (50 mg/L in drinking water, 1% sucrose; Sigma-Aldrich, Milano, Italy) prevents early Npy1r gene inactivation by an efficient suppression of tTA-dependent Cre expression 10 and 22. The Dox withdrawal at birth, by fostering litters to Dox-naïve dams at postnatal day (P) 0, slowly activates tTA, which is specifically expressed in Y5R-containing neurons. Activated tTA now induces Cre expression and, subsequently, Npy1r gene inactivation in Npy1rY5R^{-/-} mice that is fully achieved between P45 and P50 (16) (Figure 1A,B and Figure S1 in Supplement 1). Mice (4–11 litters) were fostered at P0 to Dox-free FVB/J dams and analyzed between P60 and P90. To investigate whether early maternal environment may unmask the phenotype of Npy1rY5R^{-/-} conditional mutants, mice from four to nine litters were fostered at P0 to Dox-free C57BL/6J dams exhibiting poor levels of maternal care compared with FVB/J dams (10).

Restraint Stress

The method used for restraint stress can be found in Supplemental Methods and Materials in Supplement 1.

Behavior

Open field (OF) and elevated plus maze (EPM) (to test anxiety and locomotor activity) were performed between P65 and P70 from 8:00–10:00 am (OF) and 5:00–7:00 pm (EPM). The Morris water maze (MWM) to test spatial memory was performed around P90 from 2:00–4:00 pm in an independent cohort of mice. Data were recorded automatically from the digitized image by using a computerized video tracking software. Detailed information can be found in Supplemental Methods and Materials in Supplement 1.

Body Weight and Food Intake

Body weight of mice housed in groups was measured twice a week from P30 to P90. For food intake and poststarvation refeeding test, mice were housed individually in cages starting at P30. At P70, food intake was measured every 24 hours for 5 days, and then mice were starved for 30 hours. After this period, food consumption was again measured 3 hours after refeeding and once a day for the subsequent 5 days.

Histologic Examination

Methods used for immunostaining, in situ hybridization, and quantification analysis can be found in Supplemental Methods and Materials in Supplement 1.

Real-Time Polymerase Chain Reaction

Methods used for real-time polymerase chain reaction and primer sequences can be found in Supplemental Methods and Materials in Supplement 1.

Serology

Methods used for serum collection and corticosterone analysis (radioimmunoassay) can be found in Supplemental Methods and Materials in Supplement 1.

Data Analysis

Three-way analysis of variance for repeated measures was used to compare mean body weight over time and food intake. All the other quantitative results were analyzed by two-way or three-way analysis of

variance, followed by Newman-Keuls Test, Bonferroni test for multiple comparisons, or Student t test when indicated. All data are expressed as mean \pm SEM, and the level of statistical significance was set at $p < .05$.

Results

Verification of the Region-Specific and Temporal-Specific *Npy1r* Gene Inactivation in *Npy1r^{Y5R-/-}* Mice

Using the TgY5RitTA/Y1R^{Venus}/LC1 mice, we previously demonstrated that *Npy1r* gene promoter-directed Venus and *Npy5r* promoter-directed tTA colocalize in several telencephalic and diencephalic structures, including cerebral cortex, hippocampus, amygdala, and hypothalamus (16). We expected the strongest decrease of Y1R expression in the hypothalamic and forebrain regions, similar to the reduction induced in the limbic structures of *Npy1r^{rfb}* mice using the α -calcium/calmodulin-dependent protein kinase II (α -CamKII) promoter-controlled, Dox-attenuated Cre expression for postnatal inactivation of the *Npy1r* gene in excitatory neurons (10).

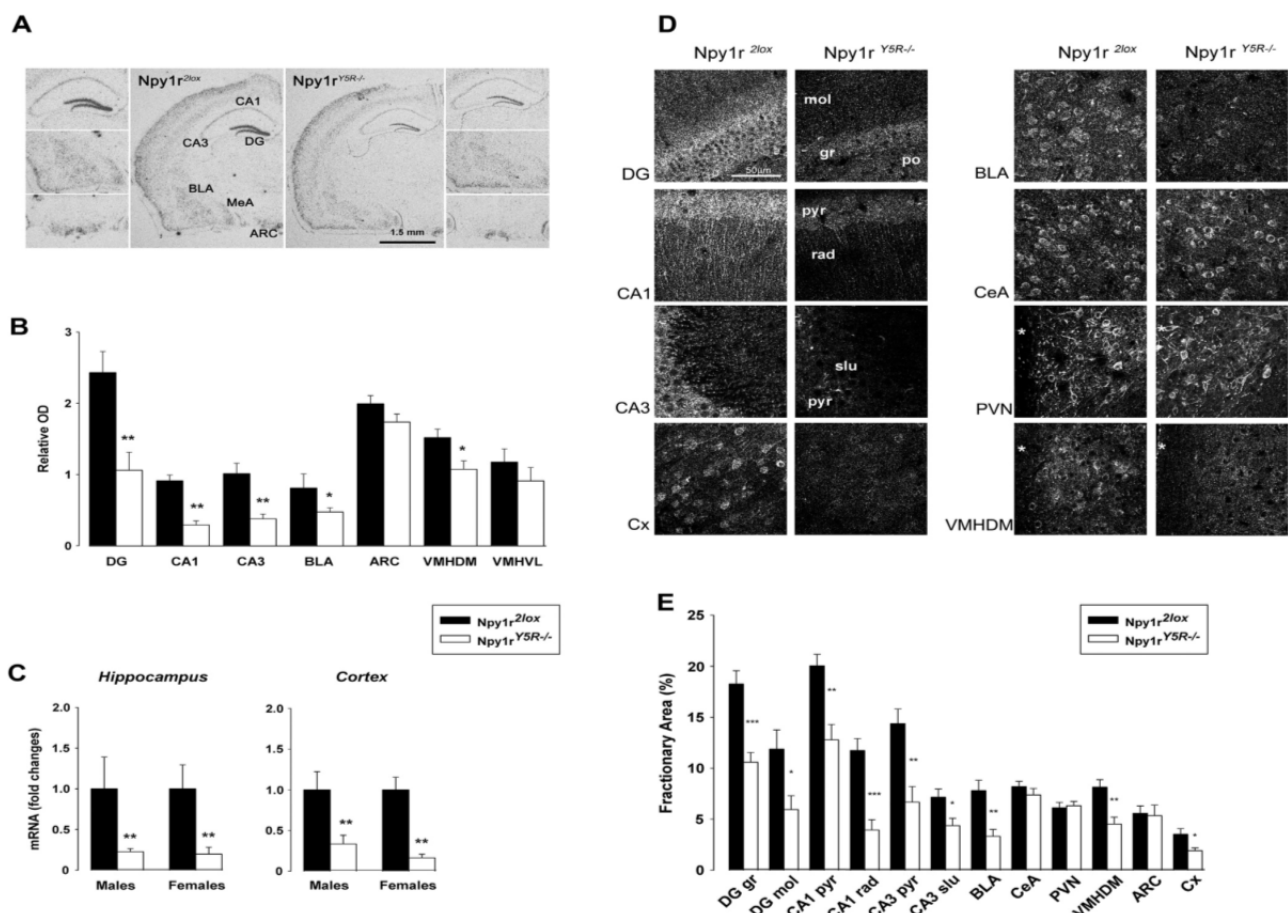


Figure 2. Expression of *Npy1r* messenger RNA (mRNA) and Y1 receptor (Y1R) peptide in the brain of *Npy1r^{2lox}* control and *Npy1r^{Y5R-/-}* conditional mutants. (A) Representative autoradiograms of in situ hybridization of *Npy1r* mRNA on brain coronal sections from *Npy1r^{2lox}* and *Npy1r^{Y5R-/-}* male mice. (B) Quantitative signal intensity (OD) analysis of in situ hybridization showed a significant decrease of *Npy1r* mRNA expression in the hippocampus (DG granule cell layer and CA1 and CA3 pyramidal cell layer), in the BLA, and in the VMHDM nucleus of *Npy1r^{Y5R-/-}* mice compared with control *Npy1r^{2lox}* littermates. Data are expressed as relative optical density and are the mean \pm SEM from two independent experiments; $n = 5$. $**p < .01$ and $*p < .05$ vs. *Npy1r^{2lox}* mice (Student *t* test). (Scale bar = 1.5 mm.) (C) *Npy1r^{Y5R-/-}* male and female mice showed lower *Npy1r* mRNA expression in the hippocampus and cerebral cortex compared with *Npy1r^{2lox}* siblings. Data are expressed as fold change variations after real-time polymerase chain reaction analysis and are mean \pm SEM; $n = 3$ –5 from three litters. Two-way analysis of variance revealed a significant effect of genotype for *Npy1r* in the hippocampus [$F_{1,15} = 9.03$, $p = .0089$] and in the cerebral cortex [$F_{1,8} = 25.94$, $p = .0009$]. $**p < .01$ vs. male and female *Npy1r^{2lox}* mice, respectively (Bonferroni posttest). (D) The Y1R immunofluorescence confirmed the reduced Y1R expression in CA1, CA3, and DG; in the Cx; in the BLA; and in the VMHDM of *Npy1r^{Y5R-/-}* mice compared with *Npy1r^{2lox}* littermates. The Y1R immunosignal was similar in the PVN and in the CeA. (Scale bar = 50 μ m.) (E) Semiquantitative analysis of Y1R immunostaining showed a significant decrease of Y1R in the hippocampus (DG molecular and granule cell layers, CA1 and CA3 pyramidal cell layer, CA1 stratum radiatum, and CA3 stratum lucidum), in the BLA, in the VMHDM nucleus, and in the Cx of *Npy1r^{Y5R-/-}* mice compared with control *Npy1r^{2lox}* littermates. Data are expressed as fractional area (%) and are mean \pm SEM from three independent experiments; $n = 5$ –11. $***p < .001$, $**p < .01$, and $*p < .05$ vs. *Npy1r^{2lox}* mice (Student *t* test). ARC, arcuate nucleus; BLA, basolateral amygdala; CA1, CA1 stratum pyramidal; CA3, CA3 stratum pyramidal; CeA, central amygdala; Cx, cerebral cortex; DG, dentate gyrus; gr, granular cell layer; Mea, medial amygdala; mol, molecular layer; po, polymorphic layer; PVN, paraventricular nucleus; pyr, pyramidal cell layer; rad, stratum radiatum; slu, stratum lucidum; VMHDM, ventromedial hypothalamic nucleus, dorsomedial part; VMHVL, ventromedial hypothalamic nucleus, ventrolateral part; *third ventricle.

Conditional Cre-mediated inactivation of the *Npy1r* gene was first verified in *Npy1rY5R^{-/-}* mice by semiquantitative in situ hybridization. A significant reduction of *Npy1r* messenger RNA (mRNA) expression was observed in the cerebral cortex, in the hippocampal CA1 and CA3 pyramidal cell layers, in the dentate gyrus (DG) granule cell layer, and in the BLA of *Npy1rY5R^{-/-}* mice compared with their control littermates (Figure 2A,B). In the hypothalamus, *Npy1rY5R^{-/-}* conditional mutants also displayed a significantly lower *Npy1r* mRNA expression in the dorsomedial part of the ventromedial nucleus compared with *Npy1r2lox* siblings (Figure 2A,B). As expected, no significant differences were observed in the hypothalamic arcuate nucleus where no colocalization of Venus and Cre reporter gene was previously observed (16). *Npy1r* mRNA in the paraventricular nucleus (PVN) was below the detection limit of the method and could not be measured.

In situ analysis was confirmed by real-time polymerase chain reaction. *Npy1r* mRNA was decreased in the hippocampus and in the cerebral cortex of *Npy1rY5R^{-/-}* mice compared with control littermates of both genders (Figure 2C). The *Npy1r* gene expression profile was analyzed by immunofluorescence staining of coronal brain sections using an anti-Y1R antibody (Figure 2D). Semiquantitative analysis of Y1R immunoreactivity (IR) revealed a significant decrease of the receptor protein in the cerebral cortex, the DG (molecular and granule cell layers), the CA1 (stratum pyramidale and radiatum), the CA3 (stratum pyramidale and lucidum), the BLA, and the dorsomedial part of the ventromedial nucleus of *Npy1rY5R^{-/-}* male mice compared with their control littermates (Figure 2E). No significant differences were observed in the central amygdala (CeA), the PVN, and the arcuate nucleus hypothalamic nuclei (Figure 2E).

Y1R and Y5R Coexist in Both GABAergic and Glutamatergic Neurons of the BLA

After postnatal Cre activation, the *Npy1r* gene was selectively removed in some brain areas of *Npy1rY5R^{-/-}* mice, but residual Y1R still could be detected (Figure 2D,E and Figure S1 in Supplement 1). The residual Y1R is due to incomplete removal of the *Npy1r* gene in Y5R-expressing cells and to some nonoverlapping activities of the *Npy1r* and the *Npy5r* promoters (16), assuming that the Venus-induced and tTA-induced Cre expression reflects the endogenous *Npy1r* and the *Npy5r* promoter activity, respectively.

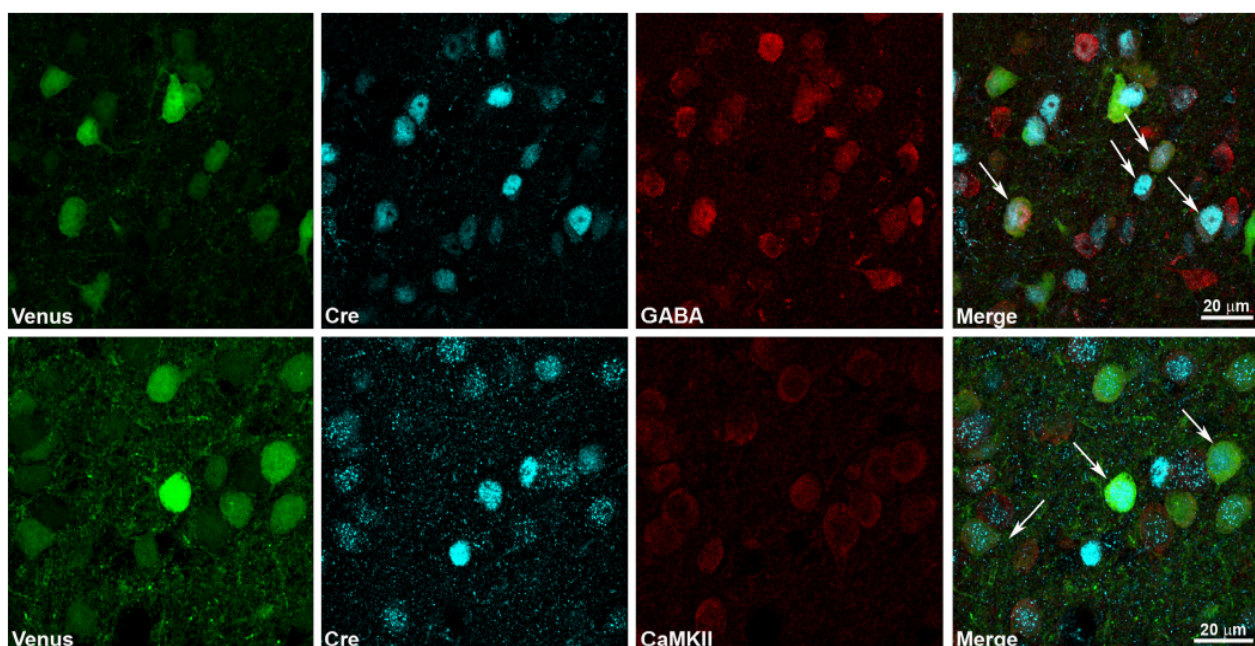


Figure 3. Colocalization of Y1 receptor (Y1R), Y5 receptor (Y5R), and gamma-aminobutyric acid (GABA) or α -calcium/calmodulin-dependent protein kinase II (α -CamKII) in the basolateral amygdala. Confocal images showed the distribution of Venus fluorescence, Cre recombinase (Cre) immunostaining, and GABA immunostaining or α -CamKII immunostaining in neurons of the basolateral amygdala. Merged images show the degree of colocalization of Venus, Cre, and GABA or α -CamKII as indicated by arrows. (Scale bar = 20 μ m.)

To analyze in detail the predicted colocalization of Y1R and Y5R in interneurons and principal cells of the BLA, we monitored Venus fluorescence (Npy1r promoter) and Cre recombinase (Npy5r promoter) and gamma-aminobutyric acid (GABA) and α -CamKII (glutamate) immunoreactivities in coronal brain sections from Npy1rY5R^{-/-} mice using confocal microscopy. The number of Y5Rs (Cre-positive cells) was higher and only partly overlapping with Y1Rs (Venus-positive cells). The analysis of sections from different animals (four per mouse) suggests that \approx 50% of GABA immunofluorescent cells coexpress Venus and Cre, whereas only a subpopulation (\approx 25%) of glutamatergic α -CamKII-IR cells are positive for Venus-IR and Cre-IR (Figure 3).

Analysis of the Phenotype of Npy1rY5R^{-/-} Mice

The removal of Y1Rs from α -CaMKII-expressing neurons in the limbic system of juvenile mice was correlated with decreased body weight growth, increased activity of the HPA axis, and increased anxiety (10). We analyzed whether the conditional inactivation of the Npy1r gene in Y5R-expressing neurons of the forebrain of adolescent Npy1rY5R^{-/-} mice was also affecting these physiologic and behavioral parameters. Additionally, because it was described that transgenic rats overexpressing hippocampal NPY show signs of spatial memory impairments (23), we analyzed the Npy1rY5R^{-/-} mice in the MWM test.

Conditional Inactivation of Y1R in Y5R-Expressing Neurons Does Not Affect Physiologic Functions

Conditional deletion of the Y1R in Y5R-expressing cells had no effect on the HPA axis activity of Npy1rY5R^{-/-} mice under basal conditions, independent of gender. This lack of effect was demonstrated by measuring 1) the density of corticotropin-releasing hormone immunoreactive fibers and cell bodies in the CeA and in the medial parvocellular division of the PVN, 2) the density of glucocorticoid receptor (GR) immunoreactive neurons in the CA1, 3) the fold changes of GR mRNA in the whole hippocampus, and 4) the serum levels of corticosterone (Figure S2 and Table S1 in Supplement 1). The exposure to 30 minutes of restraint stress significantly increases corticosterone serum levels and GR-IR nuclear expression in the CA1 in both male and female control mice and conditional mutants, but no differences were observed between the two phenotypes (Figure S2 and Table S1 in Supplement 1). Similarly, conditional Npy1r gene inactivation failed to affect body weight growth (Figure 4A) and food intake (Figure 4B) of Npy1rY5R^{-/-}, independent of gender.

Conditional Inactivation of Y1R in Y5R-Expressing Neurons Increases Anxiety

Npy1rY5R^{-/-} mice showed increased anxietylike behavior as determined by a significantly lower frequency of entries and time spent in the open arms of the EPM (Figure 5A) and less time and shorter distance traveled in the center of the OF (Figure 5B) compared with Npy1r2lox siblings. The effect was similar in males and females and independent of locomotor activity (OF, total distance traveled [m], males, Npy1r2lox = $4.8 \pm .34$ and Npy1rY5R^{-/-} = $4.5 \pm .31$, and females, Npy1r2lox = $5.2 \pm .40$ and Npy1rY5R^{-/-} = $4.4 \pm .22$; EPM, total entries [no.], males, Npy1r2lox = 32 ± 3.5 and Npy1rY5R^{-/-} = 28 ± 2.1 , and females, Npy1r2lox = 31 ± 2.8 and Npy1rY5R^{-/-} = 27 ± 2.5). The anxiety-like behavior of Npy1rY5R^{-/-} mice is significantly correlated with the Npy1r mRNA expression in hippocampal CA1 pyramidal cell layers and, to a lower extent, in CA3 and in the BLA, suggesting that the degree of reduced expression of the Y1R in the limbic system can predict the level of anxiety in the individual animals (Figure S3 in Supplement 1).

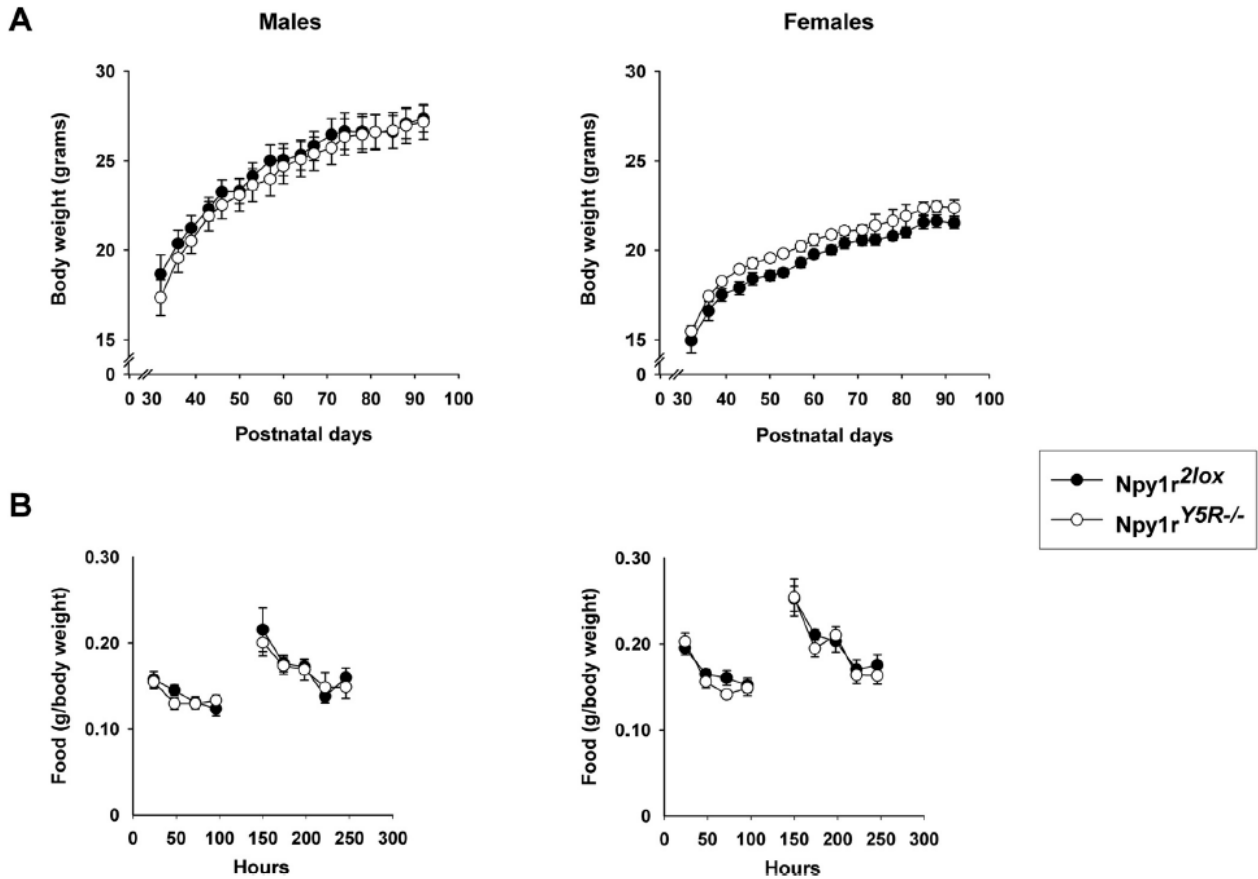


Figure 4. Body weight growth and food intake. (A) *Npy1r^{2lox}* and *Npy1r^{Y5R-/-}* male and female mice showed a similar body weight growth curve throughout the 3-month period of monitoring. (B) Food intake was similar between *Npy1r^{2lox}* controls and *Npy1r^{Y5R-/-}* mice during the 4 days before 30-hour fasting, for 3 hours after refeeding, and for the subsequent 4 days. Data are mean \pm SEM; $n = 5-13$ from six to eight litters.

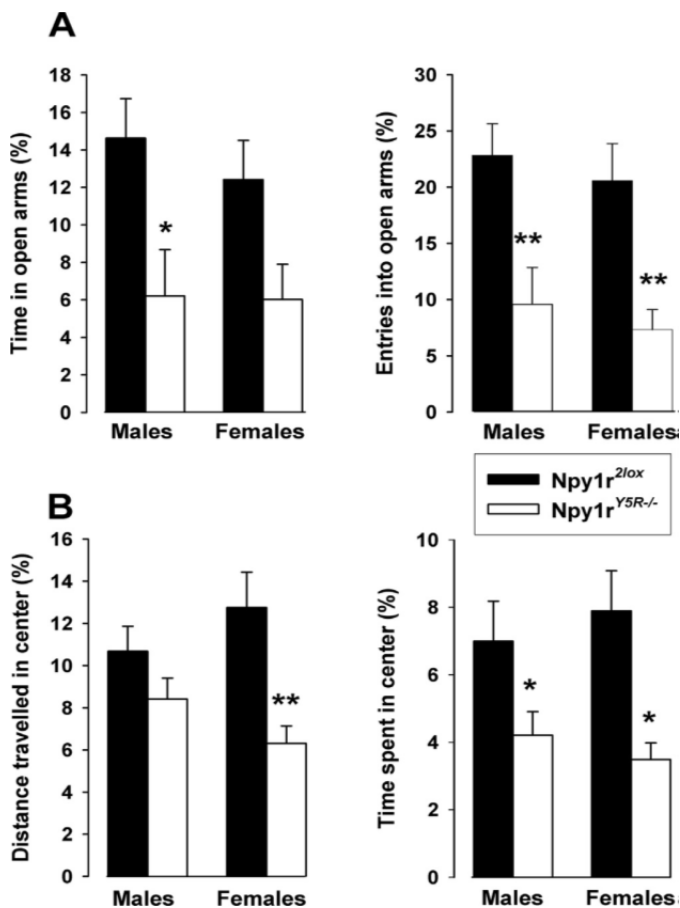


Figure 5. Anxiety-like behavior in the elevated plus maze and open field. *Npy1r^{Y5R-/-}* mice showed higher anxiety levels in the elevated plus maze (A) and open field (B) tests compared with *Npy1r^{2lox}* mice, independent of gender. Data are mean \pm SEM; $n = 12-17$ from 9-11 litters. For elevated plus maze, two-way analysis of variance revealed a significant effect of genotype for percent of time in open arms [$F_{1,55} = 10.13, p < .005$] and for percent of entries in open arms [$F_{1,55} = 23.78, p = .00001$], $**p < .01$ and $*p < .05$ *Npy1r^{Y5R-/-}* versus male and female *Npy1r^{2lox}* mice, respectively (Newman-Keuls test). For open field, two-way analysis of variance revealed a significant effect of genotype for percent of time spent in the center [$F_{1,47} = 15.32, p < .001$] and for percent of distance traveled in the center [$F_{1,47} = 14.98, p < .001$], $**p < .01$ and $*p < .05$ *Npy1r^{Y5R-/-}* versus male and female *Npy1r^{2lox}* mice, respectively (Newman-Keuls test).

Conditional Inactivation of Y1R in Y5R-Expressing Neurons Does Not Affect Learning but Increases Spatial Reference Memory

In the acquisition phase, Npy1r2lox and Npy1rY5R-/- mice showed similar decline of latency to find the hidden platform over 4 days of training indicating normal acquisition of spatial reference memory (Figure 6A). In the probe trial (conducted 24 hours after the last training trial), no significant differences in latency to reach the platform area were observed between Npy1rY5R-/- and Npy1r2lox mice independent of gender (latency [sec], males, Npy1r2lox = 7.9 ± 1.3 and Npy1rY5R-/- = 9.0 ± 2.5, and females, Npy1r2lox = 13 ± 2.3 and Npy1rY5R-/- = 13 ± 5.0; n = 9–16). Npy1rY5R-/- male and female mice showed a significantly higher percentage of time spent in the target zone (Figure 6B). Npy1rY5R-/- mice demonstrated a stronger preference for the probe quadrant because they spent more time in the target zone (north east) compared with the cumulative time spent in the no target zones (south east, south west, north west) (Figure 6C).

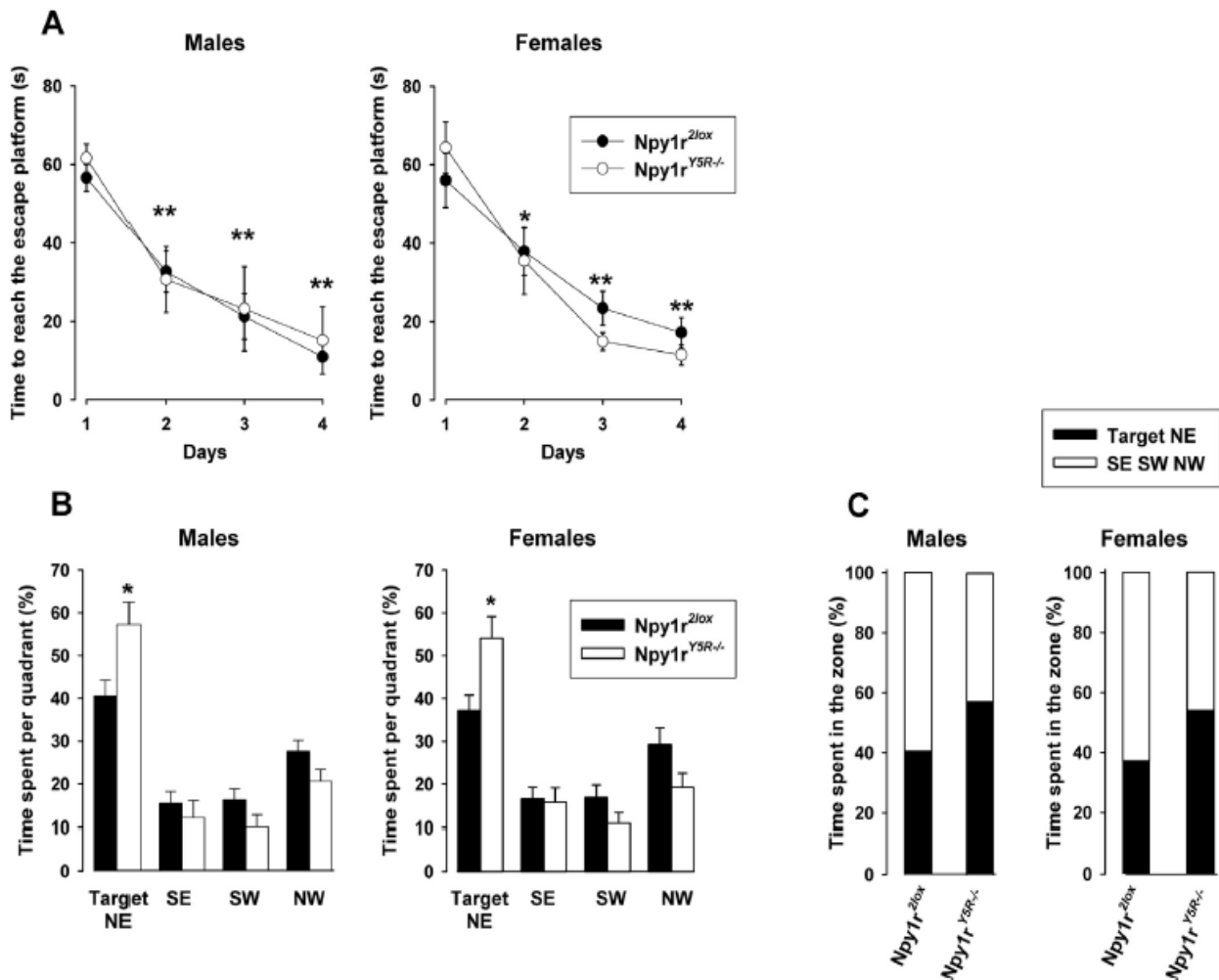


Figure 6. Spatial learning and memory retention in the Morris water maze. Npy1r^{Y5R-/-} mice showed higher percentage of time spent in the target zone compared with Npy1r^{2lox} mice, independent of gender. (A) Acquisition phase. No significant difference in escape time was observed between Npy1r^{2lox} and Npy1r^{Y5R-/-} mice each day. Data are mean ± SEM; n = 9–16 from four to nine litters. After 4 days of training, the time to reach the escape platform for all groups decreased, indicating normal spatial learning memory. Two-way analysis of variance for repeated measures revealed a significant effect of days [$F_{3,138} = 63.15, p = .000$]. * $p < .05$ and ** $p < .01$ compared with the time to reach the escape platform at day 1. (B) Probe trial. After training, the platform was removed from the north east quadrant (NE, target zone) where it was originally placed. The y axis indicates the percentage of total time spent in one specific quadrant. Data are mean ± SEM; n = 9–16 from four to nine litters. Two-way analysis of variance revealed a significant effect of genotype for percentage of total time spent in NE target quadrant [$F_{1,46} = 16.376, p = .0002$] and for percentage of time spent in south west (SW) [$F_{1,46} = 5.206, p = .0272$] and north west (NW) [$F_{1,46} = 7.369, p = .00093$] quadrants. * $p < .05$ Npy1r^{Y5R-/-} vs. Npy1r^{2lox} mice (Newman-Keuls Test). (C) Percentage of time spent in NE target and no target (south east [SE], SW, NW) zones. Data are mean ± SEM; n = 9–16 from four to nine litters. Two-way analysis of variance revealed a significant effect of genotype for percentage of total time spent in target and no target quadrants [$F_{1,46} = 16.39, p = .0002$].

We also measured the path length after mice reached the target zone in the probe trial (24). An increased path length outside the target zone, after reaching the original location of the platform, would suggest increased flexibility in an attempt to look for a new location of the platform. On the contrary, longer path length in the target zone would suggest persistency possibly related to increased anxiety. Both female and male *Npy1r^{Y5R-/-}* mice displayed a significantly shorter path length outside the target zone after reaching the original position occupied by the platform compared with their control siblings, whereas *Npy12lox* mice showed an increased path length outside the target zone, independent of gender (Figure 7A,B).

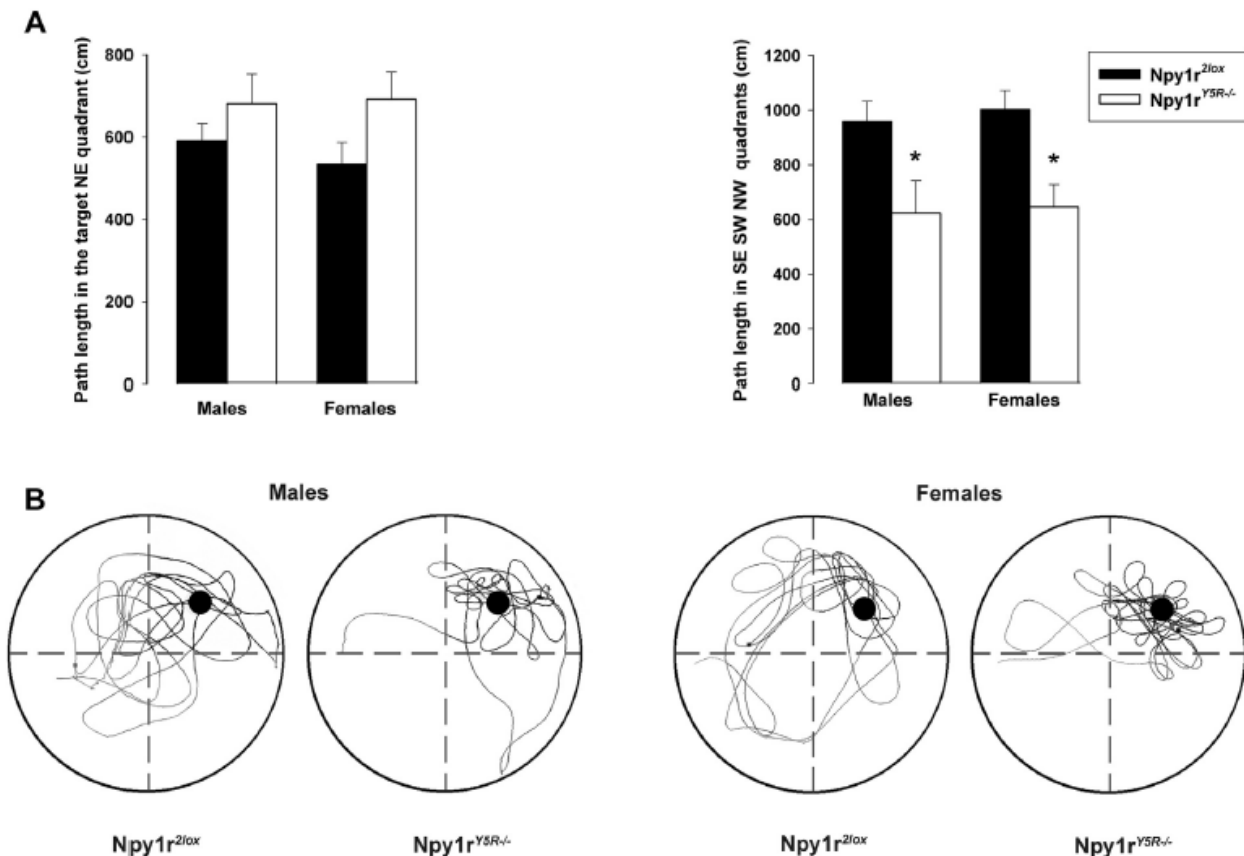


Figure 7. Path length in Morris water maze. Measures of the total distance covered (path length) by *Npy1r^{2lox}* and *Npy1r^{Y5R-/-}* mice after they reached the target zone in the probe trial. (A) *Npy1r^{Y5R-/-}* mice showed longer path length (cm) in the target north east (NE) quadrant and shorter path length in south east (SE), south west (SW), and north west (NW) quadrants compared with *Npy1r^{2lox}* mice, independent of the gender. Data are mean \pm SEM; $n = 9-16$ from four to nine litters. Two-way analysis of variance revealed a significant effect of genotype for path length in NE target quadrant [$F_{1,46} = 5.058$, $p = .0293$] and no target (SE, SW, NW) zones [$F_{1,46} = 17.97$, $p = .00011$]. * $p < .05$ vs. *Npy1r^{2lox}* mice (Newman-Keuls Test). (B) Representative swim paths of *Npy1r^{2lox}* and *Npy1r^{Y5R-/-}* male and female mice in the Morris water maze. The examples show that *Npy1r^{Y5R-/-}* male and female mice cover the longest path length in the target NE quadrant, and *Npy1r^{2lox}* male and female mice cover the longest path length in the SE, SW, and NW quadrants.

Differences in Phenotypes Between *Npy1r^{Y5R-/-}* and *Npy1r^{2lox}* Mice Are Independent of Maternal Care Received

We previously demonstrated that conditional inactivation of the *Npy1r* gene in the limbic excitatory neurons of adult *Npy1rfb* mice leads to different behavioral and physiologic consequences depending on early maternal environment. We compared the differences in phenotype between *Npy1r^{2lox}* and *Npy1r^{Y5R-/-}* mice within animals from the same litter raised either by FVB/J dams, displaying high levels of maternal care, or by C57BL/6J dams, showing low maternal behavior versus the adopted pups (10).

As previously reported (10), low levels of maternal care has long-lasting impact on C57BL/6J-fostered *Npy1r^{2lox}* control mice that show lower body weight, higher anxiety level, and corticotropin-releasing hormone IR in the PVN compared with FVB/J-fostered *Npy1r^{2lox}* mice from the same litter (Table 1). *Npy1r^{Y5R-/-}* mice showed increased anxiety-related behavior in the EPM and improved spatial reference

memory in the MWM compared with Npy1r2lox siblings, demonstrating that the physiologic and behavioral phenotype of conditional mutants is unaffected by the maternal care received.

Table 1. Comparison of Phenotypes of Npy1r^{2lox} and Npy1r^{Y5R-/-} Male Mice Fostered to FVB/J and C57BL/6J Dams

Foster Mother Strain Phenotype	FVB/J		C57BL/6J	
	Npy1r ^{2lox}	Npy1r ^{Y5R-/-}	Npy1r ^{2lox}	Npy1r ^{Y5R-/-}
BW (Grams)	25.6 ± .57	27.2 ± 1.0	23.2 ± .63 ^a	23.5 ± .67 ^{a,b}
EPM (% Time in Open Arms)	14.6 ± 2.1	6.2 ± 2.5 ^c	6.5 ± 2.0 ^d	1.2 ± .71 ^{d,e}
PVN CRH-IR (Fractional Area × 10 ⁻³)	7.8 ± .8	7.0 ± 1.1	11 ± 1.1 ^f	13 ± 1.9 ^{f,g}
MWM (% of Time in NE Target Zone)	41.01 ± 2.8	59.57 ± 4.8 ^h	42.52 ± 3.4	55.5 ± 5.2 ⁱ

Body weight, anxiety-like behavior (elevated plus maze), corticotropin-releasing hormone immunoreactivity in the paraventricular nucleus of the hypothalamus, and spatial memory retention (Morris water maze) of Npy1r^{Y5R-/-} conditional mutant male mice and their control littermates fostered to FVB/J and C57BL/6J dams at postnatal day (P) 0. Mice were analyzed at P60 (body weight), P65 (elevated plus maze), and P90 (Morris water maze and corticotropin-releasing hormone immunoreactivity). FVB/J-fostered Npy1r^{2lox} control mice showed higher body weight, lower anxiety levels, and lower density of corticotropin-releasing hormone immunoreactive cell bodies in the paraventricular nucleus than the C57BL/6J-fostered Npy1r^{2lox} controls, as previously reported (10). Data are expressed as mean ± SEM (body weight, *n* = 7–13 from five to six litters; elevated plus maze, *n* = 8–14 from eight to nine litters; paraventricular nucleus corticotropin-releasing hormone immunoreactivity, *n* = 3–5 from two litters; Morris water maze, *n* = 8–18 from four to six litters).

BW, body weight; CRH-IR, corticotropin-releasing hormone immunoreactivity; EPM, elevated plus maze; MWM, Morris water maze; NE, north east; PVN, paraventricular nucleus.

BW: Two-way analysis of variance (ANOVA) revealed a significant effect of strain of foster mother [$F_{1,35} = 19.3, p < .0001$]. ^a*p* < .05 vs. FVB fostered Npy1r^{2lox} mice; ^b*p* < .01 versus FVB fostered Npy1r^{Y5R-/-} mice (Newman-Keuls Test).

EPM: Two-way ANOVA revealed a significant effect of strain of foster mother and genotype [$F_{1,46} = 9.71, p = .0032$; $F_{1,46} = 8.81, p = .0047$, respectively]. ^c*p* < .05 and ^d*p* < .01 vs. FVB fostered Npy1r^{2lox} mice (Newman-Keuls Test); ^e*p* < .05 vs. C57BL/6J fostered Npy1r^{2lox} mice (Student *t* test).

PVN CRH-IR: Two-way ANOVA revealed a significant effect of strain of foster mother [$F_{1,13} = 16.9, p = .0012$]; ^f*p* < .05 vs. FVB fostered Npy1r^{2lox} mice; ^g*p* < .05 vs. FVB fostered Npy1r^{Y5R-/-} mice (Newman-Keuls Test).

MWM: Two-way ANOVA revealed a significant effect of genotype [$F_{1,49} = 15.4, p = .00027$]; ^h*p* < .01 vs. FVB fostered Npy1r^{2lox} mice and ⁱ*p* < .05 vs. C57BL/6J fostered Npy1r^{2lox} mice.

Discussion

To analyze the specific contribution of Y1Rs coexpressed with Y5Rs on NPY mediated physiologic functions, we generated conditional Npy1r knockout mice (Npy1rY5R^{-/-}) exhibiting inactivation of Y1Rs selectively in Y5R-containing neurons of adult animals. In situ hybridization, quantitative real-time polymerase chain reaction, and immunofluorescence revealed reduced Y1R mRNA and protein levels in the cerebral cortex; in the CA1, CA3, and DG of the hippocampus; in the BLA; and in the hypothalamic area dorsomedial part of the ventromedial nucleus. Npy1rY5R^{-/-} mice showed increased anxiety and higher behavioral persistency possibly related to improved spatial reference memory compared with control littermates.

The anxiogenic effect of specific inactivation of the Y1R in Y5R-containing neurons became apparent in the EPM and OF tests. In the EPM, Npy1rY5R^{-/-} mice showed a lower frequency of entries and spent significantly less time in the open arms. In the OF, Npy1rY5R^{-/-} conditional mutants of both genders spent less time and traveled a shorter distance in the center of the arena compared with their control littermates. These results suggest that neurons containing both Y1Rs and Y5Rs play a major role in the control of anxious behavior, without affecting HPA axis activity, as shown by substantial identical levels of serum corticosterone and corticotropin-releasing hormone expression in CeA and PVN and hippocampal GR-IR and mRNA in control and Npy1rY5R^{-/-} mice under basal conditions and after 30 minutes of restraint stress.

By using the same conditional system, we generated knockout mice (Npy1rrfb) exhibiting reduced levels of Y1Rs in adult limbic excitatory neurons (10). Npy1rrfb and Npy1rY5R^{-/-} targeted mice display a few similarities and differences. Both Npy1rrfb and Npy1rY5R^{-/-} male mice displayed anxiety-like behavior in EPM and OF; however, Npy1rrfb male mice, but not Npy1rY5R^{-/-} mice, showed increased HPA activity. Npy1rrfb male mice showed a significant decrease of body weight growth compared with their control littermates, whereas no changes were observed in Npy1rY5R^{-/-} male mice. When we used the α -CamKII promoter-controlled Cre to remove Y1Rs from principal cells of the forebrain of juvenile mice (Npy1rrfb),

the development of a behavioral and physiologic impairment compared with wild-type littermates was gender-specific and inversely correlated with maternal care (10). On the contrary, the phenotype of *Npy1rY5R^{-/-}* mice was largely independent of gender and maternal care. These differences between phenotypes of *Npy1rY5R^{-/-}* and *Npy1rrfb* mice may be explained by the different regional and cellular inactivation pattern of the *Npy1r* gene.

Limbic stress effector pathways relay through basal forebrain, hypothalamic, and brainstem neurons that innervate the medial parvocellular division of the PVN to modulate the HPA axis. This neuronal circuitry more recently has been proposed to be involved in long-lasting impact of maternal care on stress resilience (25). The hippocampus is an important component of this circuit controlling anxiety-related behaviors and stress responses and seems to inhibit the HPA axis through glutamate-GABA connections (26). We previously postulated that in *Npy1rrfb* mice, the selective inactivation of Y1Rs in principal excitatory neurons of hippocampus lead to increased HPA axis activity via the glutamatergic output, which possibly depends on early maternal conditions (10).

Both *Npy1rY5R^{-/-}* and *Npy1rrfb* mice display a significant reduction of *Npy1r* mRNA and protein in the hippocampal CA1 and CA3 pyramidal and DG granule cell layers compared with their control littermates. However, comparison of Cre recombinase expression and *Npy1r* inactivation pattern in limbic system of these conditional mutants demonstrates that *Npy1rY5R^{-/-}* mice, but not *Npy1rrfb* mice, show a significant decrease of *Npy1r* mRNA and protein in the BLA. Our data also indicate that in the BLA \approx 50% of GABA-IR cells coexpress the Y1R and Y5R, whereas a small population of α -CamKII-positive neurons (\approx 25%) colocalize with both NPY receptor subtypes, suggesting that the conditional inactivation of the *Npy1r* gene in *Npy1rY5R^{-/-}* mice might occur mainly in GABAergic neurons of the BLA. This finding indicates that the loss of Y1Rs also on the cellular level is different between *Npy1rY5R^{-/-}* and *Npy1rrfb* mice, which may account for the diverse *Npy1rY5R^{-/-}* and *Npy1rrfb* mice expressed phenotypes.

The BLA is an important component of neuronal circuitry controlling anxiety-related behaviors where both the Y1R and the Y5R appear to be required for the anxiolytic response to NPY. In the BLA, GABAergic interneurons have strong synaptic control over its principal neurons, the pyramidal cells (27). The presence of the Y1Rs on this population of interneurons could have profound effects on the synchronous activity of the BLA (28).

These results suggest that the circuitry by which NPY exerts its anxiolytic action is likely to involve more neuronal pathways. Molosh et al. (29) suggested more recently that two distinct, cyclic adenosine monophosphate-mediated intracellular mechanisms are operative in the Y1R-mediated control of N-methyl-D-aspartate and GABAA receptor functions in the BLA. Previous studies showed a striking dichotomy between CeA and BLA in the NPY regulation of behavioral, autonomic, and endocrine responses during stress, suggesting that different components of stress response not only are elicited through a wide range of pathways but also are differentially regulated (7). Consistent with these studies, we now suggest that a similar dichotomy may exist between the BLA GABAergic interneurons and hippocampal excitatory neurons in Y1R-mediated regulation of anxiety and the HPA axis that may explain the differences between phenotypes of *Npy1rrfb* and *Npy1rY5R^{-/-}* mice.

In the present study we also demonstrate that conditional inactivation of *Npy1r* increased spatial reference memory. In the MWM, *Npy1rY5R^{-/-}* mice showed similar latencies to reach the platform as control *Npy1r2lox* mice but a significant increase of time spent in the target zone, 1 day after 4 days of training. This finding suggests that the deficit in memory retention, previously observed in transgenic rats

overexpressing NPY in the hippocampus (23), might be mediated by the Y1R coexpressed in Y5R-containing neurons, providing novel evidence for a role of the Y1R in retention of spatial memory. The higher persistence of *Npy1rY5R*^{-/-} mice in the target zone might also be related to lower behavioral flexibility, which would be consistent with the high anxiety profile of these animals. The behavioral flexibility requires that the animal does not persist in choosing a wrong solution (i.e., perseverating in the area where the platform was before) and rapidly modifies its behavior to search for a new solution 22 and 30. The underlying mechanisms allowing such adaptive behavior belong to a behavioral inhibition system, whose functions are controlled, in particular, by hippocampal formation 31, 32 and 33. However, further experiments are necessary to test this issue specifically by using a target-reversal design.

Previous studies have shown that GRs participate in the negative feedback inhibition of the HPA axis and modulate learning and memory (34) and that forebrain GR overexpression leads to cognitive deficits (35). Our findings demonstrate that hippocampal GR mRNA and protein expression and corticosterone levels were not affected by the conditional inactivation of *Npy1r* in Y5R-expressing neurons. It is unlikely that dysregulated hippocampal GRs accounts for the improvement of spatial reference memory observed in *Npy1rY5R*^{-/-} conditional knockout mice.

It was postulated more recently that Y1R and Y5R colocalization results in receptor heterodimerization and in enhanced or altered function of several physiologic systems (36). The anxious phenotype of *Npy1rY5R*^{-/-} mice might be due to both the conditional inactivation *Npy1r* and the altered Y5R functions. Domschke et al. (18) suggested that a coding variant and an intronic single nucleotide polymorphism in *Npy5r*, found to be associated with panic disorders, might directly or indirectly downregulate Y5R expression, contributing to an increased vulnerability to anxiety states. In line with these observations, it is reasonable to expect that conditional inactivation of *Npy5r* in Y1R-coexpressing neurons might similarly result in increased anxiety-related behavior and improved spatial reference memory. Although the interaction mechanisms between Y1Rs and Y5Rs require further evaluation, their synergistic action in the regulation of emotional behavior and behavioral flexibility may have new therapeutic potentials for human anxiety.

In conclusion, we established a genetic tool to inactivate spatially and temporally *Npy1r* gene in neurons coexpressing both the Y1R and the Y5R. Our analysis demonstrates that *Npy1rY5R*^{-/-} mutants show a strong anxious phenotype that is not associated with increased HPA axis activity; this phenotype is independent of gender and early maternal care, and we suggest it might be mechanistically linked to inactivation of the Y1R in GABAergic interneurons of the BLA. In addition, we provide experimental genetic evidence that Y1Rs coexpressed with Y5Rs are involved in spatial reference memory and possibly in behavioral flexibility in male and female mice. Together, these data strongly support the concept that NPY Y1R, acting on distinct limbic neurotransmitter systems, may play different physiologic roles in the regulation of anxiety and stress response. If we consider our findings in the context of human studies, showing a direct relationship between NPY and interindividual variation in emotion and stress resilience (4), our data suggest a potential strategy of novel therapeutics for various disorders related to anxiety and stress.

Acknowledgements

We thank Dr J. Urban for polyclonal antibody and protocols for Y1 receptor immunohistochemistry. The following grants were awarded: Grant Nos. PRIN 2005057519-002 and 2008PLKP3E-003 from MIUR (Ministero dell'Istruzione, dell'Università e della Ricerca), Fondazione CRT (Cassa di Risparmio di Torino), Torino, to CE; "Neuroscience program," Compagnia di San Paolo, Torino, to CE and PP; fellowships from the

Health Ministry of Regione Piemonte and Fondazione CRT to AL; and German Research Foundation Grant No. SFB636/A4 to RS. IB is currently affiliated with the Department of Molecular Neurobiology, Max Planck Institute for Medical Research, Heidelberg, Germany.

The authors report no biomedical financial interests or potential conflicts of interest.

References

1. C. Eva, M. Serra, P. Mele, G. Panzica, A. Oberto. Physiology and gene regulation of the brain NPY Y1 receptor. *Front Neuroendocrinol*, 27 (2006), pp. 308–339
2. T. Hokfelt, C. Broberger, X. Zhang, M. Diez, J. Kopp, Z. Xu, et al. Neuropeptide Y: Some viewpoints on a multifaceted peptide in the normal and diseased nervous system. *Brain Res Brain Res Rev*, 26 (1998), pp. 154–166
3. Z. Zukowska, G.Z. Feuersteing. *NPY Family of Peptides in Neurobiology, Cardiovascular and Metabolic Disorders: From Genes to Therapeutics*. Springer, New York (2006)
4. Z. Zhou, G. Zhu, A.R. Hariri, M.A. Enoch, D. Scott, R. Sinha, et al. Genetic variation in human NPY expression affects stress response and emotion. *Nature*, 452 (2008), pp. 997–1001
5. P. Broqua, J.G. Wettstein, M.N. Rocher, B. Gauthier-Martin, J.L. Junien. Behavioral effects of neuropeptide Y receptor agonists in the elevated plus-maze and fear-potentiated startle procedures. *Behav Pharmacol*, 6 (1995), pp. 215–222
6. M. Heilig, S. McLeod, M. Brot, S.C. Heinrichs, F. Menzaghi, G.F. Koob, et al. Anxiolytic-like action of neuropeptide Y: Mediation by Y1 receptors in amygdala, and dissociation from food intake effects. *Neuropsychopharmacology*, 8 (1993), pp. 357–363
7. T.J. Sajdyk, P.L. Johnson, R.J. Leitermann, S.D. Fitz, A. Dietrich, M. Morin, et al. Neuropeptide Y in the amygdala induces long-term resilience to stress-induced reductions in social responses but not hypothalamic-adrenal-pituitary axis activity or hyperthermia. *J Neurosci*, 28 (2008), pp. 893–903
8. W.F. Colmers, D. Bleakman. Effects of neuropeptide-Y on the electrical-properties of neurons. *Trends Neurosci*, 17 (1994), pp. 373–379
9. E.P. Finta, J.T. Regenold, P. Illes. Depression by neuropeptide-Y of noradrenergic inhibitory postsynaptic potentials of locus coeruleus neurons. *Naunyn Schmiedebergs Arch Pharmacol*, 346 (1992), pp. 472–474
10. Bertocchi, A. Oberto, A. Longo, P. Mele, M. Sabetta, A. Bartolomucci, et al. Regulatory functions of limbic Y1 receptors in body weight and anxiety uncovered by conditional knockout and maternal care. *Proc Natl Acad Sci U S A*, 108 (2011), pp. 19395–19400
11. E.J. Lin, S. Lin, A. Aljanova, M.J. During, H. Herzog. Adult-onset hippocampal-specific neuropeptide Y overexpression confers mild anxiolytic effect in mice. *Eur Neuropsychopharmacol*, 20 (2010), pp. 164–175
12. T.J. Sajdyk, M.G. Vandergriff, D.R. Gehlert. Amygdalar neuropeptide Y Y1 receptors mediate the anxiolytic-like actions of neuropeptide Y in the social interaction test. *Eur J Pharmacol*, 368 (1999), pp. 143–147
- A. Kask, M. Eller, L. Orelund, J. Harro. Neuropeptide Y attenuates the effect of locus coeruleus denervation by DSP-4 treatment on social behaviour in the rat. *Neuropeptides*, 34 (2000), pp. 58–61
12. G. Sorensen, C. Lindberg, G. Wortwein, T.G. Bolwig, D.P. Woldbye. Differential roles for neuropeptide Y Y1 and Y5 receptors in anxiety and sedation. *J Neurosci Res*, 77 (2004), pp. 723–729

13. H. Herzog, K. Darby, H. Ball, Y. Hort, A. Beck-Sickinger, J. Shine. Overlapping gene structure of the human neuropeptide Y receptor subtypes Y1 and Y5 suggests coordinate transcriptional regulation. *Genomics*, 41 (1997), pp. 315–319
- A. Oberto, E. Acquadro, T. Bus, R. Sprengel, C. Eva. Expression patterns of promoters for NPY Y(1) and Y(5) receptors in Y(5)RitTA and Y(1)RVenus BAC-transgenic mice. *Eur J Neurosci*, 26 (2007), pp. 155–170
14. M.L. Wolak, M.R. DeJoseph, A.D. Cator, A.S. Mokashi, M.S. Brownfield, J.H. Urban. Comparative distribution of neuropeptide Y Y1 and Y5 receptors in the rat brain by using immunohistochemistry. *J Comp Neurol*, 464 (2003), pp. 285–311
15. K. Domschke, C. Hohoff, C. Jacob, W. Maier, J. Fritze, B. Bandelow, et al. Chromosome 4q31-34 panic disorder risk locus: association of neuropeptide Y Y5 receptor variants. *Am J Med Genet B Neuropsychiatr Genet*, 147B (2008), pp. 510–516
16. T.J. Sajdyk, D.A. Schober, D.R. Gehlert. Neuropeptide Y receptor subtypes in the basolateral nucleus of the amygdala modulate anxiogenic responses in rats. *Neuropharmacology*, 43 (2002), pp. 1165–1172
17. K. Schonig, F. Schwenk, K. Rajewsky, H. Bujard. Stringent doxycycline dependent control of CRE recombinase in vivo. *Nucleic Acids Res*, 30 (2002), p. e134
18. H. Bouabe, K. Okkenhaug. Gene targeting in mice: A review. *Methods Mol Biol*, 1064 (2013), pp. 315–336
19. D.M. Bannerman, T. Bus, A. Taylor, D.J. Sanderson, I. Schwarz, V. Jensen, et al. Dissecting spatial knowledge from spatial choice by hippocampal NMDA receptor deletion. *Nat Neurosci*, 15 (2012), pp. 1153–1159
- A. Thorsell, M. Michalkiewicz, Y. Dumont, R. Quirion, L. Caberlotto, R. Rimondini, et al. Behavioral insensitivity to restraint stress, absent fear suppression of behavior and impaired spatial learning in transgenic rats with hippocampal neuropeptide Y overexpression. *Proc Natl Acad Sci U S A*, 97 (2000), pp. 12852–12857
20. C.V. Vorhees, M.T. Williams. Morris water maze: Procedures for assessing spatial and related forms of learning and memory. *Nat Protoc*, 1 (2006), pp. 848–858
21. C.A. Karsten, T.Z. Baram. How does a neuron “know” to modulate its epigenetic machinery in response to early-life environment/experience? *Front Psychiatry*, 4 (2013), p. 89
22. J.P. Herman, N.K. Mueller, H. Figueiredo. Role of GABA and glutamate circuitry in hypothalamo-pituitary-adrenocortical stress integration. *Ann N Y Acad Sci*, 1018 (2004), pp. 35–45
23. J.F. Muller, F. Mascagni, A.J. McDonald. Pyramidal cells of the rat basolateral amygdala: Synaptology and innervation by parvalbumin-immunoreactive interneurons. *J Comp Neurol*, 494 (2006), pp. 635–650
24. D.G. Rainnie, I. Mania, F. Mascagni, A.J. McDonald. Physiological and morphological characterization of parvalbumin-containing interneurons of the rat basolateral amygdala. *J Comp Neurol*, 498 (2006), pp. 142–161
25. A.I. Molosh, T.J. Sajdyk, W.A. Truitt, W. Zhu, G.S. Oxford, A. Shekhar. NPY Y(1) receptors differentially modulate GABA(A) and NMDA receptors via divergent signal-transduction pathways to reduce excitability of amygdala neurons. *Neuropsychopharmacology*, 38 (2013), pp. 1352–1364
26. G. Malleret, R. Hen, J.L. Guillou, L. Segu, M.C. Buhot. 5-HT1B receptor knock-out mice exhibit increased exploratory activity and enhanced spatial memory performance in the Morris water maze. *J Neurosci*, 19 (1999), pp. 6157–6168
27. F. Cirulli, A. Berry, F. Chiarotti, E. Alleva. Intrahippocampal administration of BDNF in adult rats affects short-term behavioral plasticity in the Morris water maze and performance in the elevated plus-maze. *Hippocampus*, 14 (2004), pp. 802–807

28. J.A. Gray. The hippocampus as an interface between cognition and emotion. H.L. Roitblat, T.G. Bever, H.S. Terrace (Eds.), *Animal Cognition*, Erlbaum, Hillsdale, NJ (1984), pp. 607–625
29. K.R. Kleinknecht, B.T. Bedenk, S.F. Kaltwasser, B. Grunecker, Y.C. Yen, M. Czisch, et al. Hippocampus-dependent place learning enables spatial flexibility in C57BL6/N mice. *Front Behav Neurosci*, 6 (2012), p. 87
30. E.R. de Kloet, M. Joels, F. Holsboer. Stress and the brain: From adaptation to disease. *Nat Rev Neurosci*, 6 (2005), pp. 463–475
31. Q. Wei, E.K. Hebda-Bauer, A. Pletsch, J. Luo, M.T. Hoversten, A.J. Osetek, et al. Overexpressing the glucocorticoid receptor in forebrain causes an aging-like neuroendocrine phenotype and mild cognitive dysfunction. *J Neurosci*, 27 (2007), pp. 8836–8844
32. S. Mashiko, R. Moriya, A. Ishihara, A. Gomori, H. Matsushita, S. Egashira, et al.. Synergistic interaction between neuropeptide Y1 and Y5 receptor pathways in regulation of energy homeostasis. *Eur J Pharmacol*, 615 (2009), pp. 113–117