IRIS AperTO

UNIVERSITÀ DEGLI STUDI DI TORINO

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Performance Assessment in Fingerprinting and Multi Component Quantitative NMR Analyses

This is a pre print version of the following article:

Original Citation:

Availability:

This version is available http://hdl.handle.net/2318/1528415 since 2015-11-18T18:15:33Z

Published version:

DOI:10.1021/acs.analchem.5b00919

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

05 May 2024

analytical chemistry

Article

pubs.acs.org/ac

Performance Assessment in Fingerprinting and Multi Component Quantitative NMR Analyses

Vito Gallo,*[,1,2,3] Nicola Intini,[3,4] Piero Mastrorilli,[1,3] Mario Latronico,[1,3] Pasquale Scapicchio,[5] Maurizio Triggiani,[6] Vitoantonio Bevilacqua,[6] Paolo Fanizzi,[7] Domenico Acquotti,[8] Cristina Airoldi,[9] Fabio Arnesano,[10] Michael Assfalg,[11] Francesca Benevelli,[12,13] Davide Bertelli,[14] Laura R. Cagliani,[15] Luca Casadei,[16] Flaminia Cesare Marincola,[17] Giuseppe Colafemmina,[10,18] Roberto Consonni,[15] Cesare Cosentino,[19] Silvia Davalli,[20] Sandra A. De Pascali,[21] Virginia D'Aiuto,[22] Andrea Faccini,[8] Roberto Gobetto,[23] Raffaele Lamanna,[24] Francesca Liguori,[25] Francesco Longobardi,[10] Domenico Mallamace,[26] Pierluigi Mazzei,[27] Ileana Menegazzo,[28] Salvatore Milone,[29] Adele Mucci,[30] Claudia Napoli,[12] Thelma Pertinhez,[8] Antonino Rizzuti,[1] Luca Rocchigiani,[31] Elisabetta Schievano,[28] Fabio Sciubba,[16] Anatoly Sobolev,[32] Leonardo Tenori,[33] and Mariacristina Valerio[16]

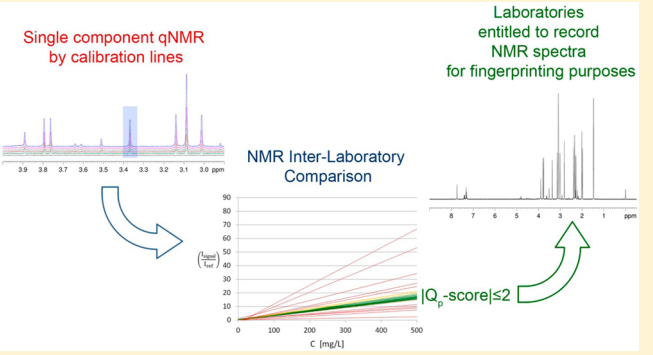
[1]Dipartimento di Ingegneria Civile, Ambientale, del Territorio, Edile e di Chimica, Politecnico di Bari, Via Orabona 4 CAMPUS, I-70125, Bari, Italy

[2]SAMER (Special Agency of the Chamber of Commerce of Bari), Via E. Mola 19, I-70121, Bari, Italy

[3]Innovative Solutions S.r.l., Spin Off del Politecnico di Bari, Zona H 150/B, I-70015, Noci, Bari, Italy

Supporting Information

ABSTRACT: An interlaboratory comparison (ILC) was organized with the aim to set up quality control indicators suitable for multicomponent quantitative analysis by nuclear magnetic resonance (NMR) spectroscopy. A total of 36 NMR data sets (corresponding to 1260 NMR spectra) were produced by 30 participants using 34 NMR spectrometers. The calibration line method was chosen for the quantification of a five-component model mixture. Results show that quantitative NMR is a robust quantification tool and that 26 out of 36 data sets resulted in statistically equivalent calibration lines for all considered NMR signals. The performance of each laboratory was assessed by means of a new performance index (named Q_p-score) which is related to the difference between the experimental and the consensus values of the slope of the calibration lines. Laboratories endowed with a Q_p-score falling within the suitable acceptability range are qualified to produce NMR spectra that can be considered statistically equivalent in terms of relative intensities of the signals. In addition, the specific response of nuclei to the experimental excitation/relaxation conditions was addressed by means of the parameter named NR. NR is related to the difference between the theoretical and the consensus slopes of the calibration lines and is specific for each signal produced by a well-defined set of acquisition parameters.



Since the first successful experiments on the detection of nuclear resonance signals back in 1945−1946,[1−3] nuclear magnetic resonance (NMR) spectroscopy has become a powerful technique for investigating the finer properties of matter showing no sign of slackening even 70 years later. In the field of quantitative analytical chemistry, the use of NMR as a quantification tool has become very common for many applications in both academic and industrial research such as pharmacy, food, and materials science. Recently, the needs and advantages of using NMR spectroscopy as a quantification tool have been exhaustively reviewed by Bharti and Roy.[4]

NMR spectroscopy is considered a primary analytical technique due to the possibility to derive a full uncertainty budget by mathematical equations. As a consequence, NMR spectroscopy is enabled for quantitative determinations at the highest metrological level. The main feature making NMR a powerful technique in quantitative determinations concerns the direct proportionality existing between the intensity of the NMR signal and the number of nuclei generating the signal. Quantitative NMR does not need reference standard molecules showing chemical structure similarity with the analyzed sample as conversely requested, for instance, in chromatographic methods. Quantification is typically obtained by integrating the

Received: March 3, 2015
Accepted: May 28, 2015

ACS Publications © XXXX American Chemical Society

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

signal of interest and scaling it to the peak area of a selected signal generated by an arbitrary reference material, whose concentration is known. Notwithstanding these advantages, official qNMR methods are still rare,[5] when compared to other analytical techniques officially recognized for quantification. The lack of official qNMR methods is a serious limitation for the exploitation of NMR potential in single component quantification analyses and represents a critical problem when NMR potential is considered for multicomponent and fingerprinting purposes. In fact, NMR spectroscopy is gaining ever growing popularity for the development of analytical approaches focusing on multicomponent untargeted analyses.[6−29] Among the many reasons for the gap between the use of NMR and the use of other techniques for official purposes, high costs of NMR spectrometers and high limits of detection (LODs) are commonly invoked. However, the lack of reproducibility data for specific methods also plays certainly an important role in preventing recognition of NMR measurements by institutions and certification bodies. This has to be probably ascribed to the fact that academic researchers are rarely involved in design of formal standardization procedures.

Measurement uncertainty is typically evaluated by three models: one laboratory−one method (1L1M), many laboratories−many methods (mLmM), and many laboratories−one method (mL1M). In the NMR community, the first limit model is the rule and several validation processes[30] are available, demonstrating the suitability of NMR spectroscopy as a quantification tool. For single component quantification, the mLmM limit model is required for a wide acceptance of the quantification method. Such a model was followed in the first German and international interlaboratory comparisons organized by the Federal Institute of Materials Research and Testing (BAM) in 1999.[31] At that time, it was found that results differed enormously (up to 100%) among the participating laboratories. The unacceptable result was attributed to the individual and independent setup of the measurements, the data processing, and the evaluation procedure of each single laboratory. To overcome these drawbacks, approximately 5 years later, another interlaboratory comparison was organized by Melz and Jancke using the mL1M model for uncertainty evaluation.[31] The 33 participants used spectrometers working at ^{1}H frequencies ranging from 200 to 600 MHz and adopted a common protocol for the experimental setup and data processing. The NMR experiment considered for this second comparison consisted of a single 30° excitation pulse followed by a suitable relaxation delay. Data elaboration, valid for determination of mole ratios of each compound, turned out a measurement uncertainty of 1.5% for a confidence level of 95% (k = 2), thus demonstrating the importance of acquisition and processing protocols for accurate and precise quantitative NMR measurements. Moreover, it was demonstrated that precision could be improved when a single operator processed all the NMR spectra.

An interesting advantage of the NMR technique deals with the possibility to suppress selectively one or more intense signals with the consequent opportunity to enhance dramatically the signal-to-noise ratio of weak signals. Typically, this kind of experiment allows one to remove solvent signals thus reducing the manipulation of the samples and avoiding the use of large amounts of deuterated solvents. In routine experiments, signal suppression can be simply obtained by implementing the pulse sequence with a presaturation scheme consisting of a low power radio frequency pulse able to saturate a specific resonance.

In principle, the introduction of the presaturation scheme should not affect the quantitative NMR measurements. The reproducibility of a single pulse experiment preceded by presaturation of the solvent signal has been evaluated by application of principal component analysis (PCA) to ^{1}H NMR data in the framework of two interlaboratory comparisons.[32,33] PCA offers the advantage to estimate measurement reproducibility by easy visual inspection of the *scores* plot but quality control indexes to be used as general reference parameters for quality assessment of NMR spectra are still lacking.

With the aim to set up new quality control parameters suitable for multi component quantitative NMR analysis as well as for NMR fingerprinting methods, we have organized the first Italian interlaboratory comparison according to the internationally agreed procedures ISO/IEC 17043:2010,[34] which specifies general requirements for development and operation of proficiency testing schemes, and ISO/IEC 17025:2005,[35] which specifies the general requirements for the competence to carry out tests and calibrations performed using standard methods, nonstandard methods, and laboratory-developed methods. The conventional statistical elaboration of data was carried out according to ISO 13528:2005[36] and ISO 5725, parts 1−6.[37] The analytical target of the comparison was the quantification of analytes in a five-component model mixture by the calibration curve approach and using the mL1M model for uncertainty evaluation. Two different data elaborations were considered: the first one was carried out by a single operator who processed NMR spectra and developed calibration lines with signal areas as input data, without referencing to any standard molecule;[38] the second one was characterized by the involvement of each participant in NMR spectra processing and signal area calculation. In the second elaboration, signal areas were scaled to a standard molecule and calibration lines were developed by a specifically designed Web application.

In this paper, the comparison between results obtained by the two data elaborations are discussed in terms of coefficient of variation. The performance assessment in the second data elaboration was carried out by means of the parameter (z-score) usually considered as performance index in single component quantifications as well as by means of a new parameter, named Q_p-score, better suited for performance assessment in multicomponent and fingerprinting analyses. Moreover, a third index (NR), specific for each NMR signal, was introduced to gain insights into the possible effects of the acquisition parameters on signal intensities.

EXPERIMENTAL SECTION

Materials. 2-Methyl-2-(methylthio)propanal-*O*-(*N*-methylcarbamoyl)oxime (Aldicarb, CAS No. 116-06-3, neat purity 99.9%, Sigma-Aldrich, Milan, Italy), 2-methoxy-*N*-(2-oxo-1,3-oxazolidin-3-yl)-acet-2′,6′-xylidide (Oxadixyl, CAS No. 77732-09-3, neat purity 99.9%, Sigma-Aldrich, Milan, Italy), *O*,*S*-dimethylphosphoramidothioate (Methamidophos, CAS No. 102658-92-6, neat purity 98.5%, Sigma-Aldrich, Milan, Italy), (2-dimethylamino-5,6-dimethylpyrimidin-4-yl)-*N*,*N*-dimethylcarbamate (Pirimicarb, CAS No. 23103-98-2, neat purity 99.0%, Sigma-Aldrich, Milan, Italy), 3-(trimethylsilyl)-2,2,3,3-tetradeutero-propionic acid sodium salt (TSP, CAS No. 24493-21-8, 99% D, Armar Chemicals, Döttingen, Switzerland), deuterium oxide (D_2O, CAS No. 7789-20-0, 99.86% D, Sigma-Aldrich, Milan, Italy) were used for sample preparation.

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

Chemical structures of compounds are reported in Chart S1 in the Supporting Information.

Sample Preparation. Standard and test mixtures were prepared under thermic and hygrometric control (20 ± 5 °C, 40−60 R.H.%) by gravimetric method using a certified analytical balance KERN ABT 100-5 M (KERN & Sohn GmbH, Balingen, Germany) with weighing range 1−101.000 mg, readability 0.01 mg, and reproducibility 0.05 mg. The balance was periodically calibrated by the certified test weight set KERN DKD-K-11801, 11-06, s/n G0703552. Uncertainty for each analyte mass was calculated taking into account uncertainty parameters of the balance. A factor $k = 2$, corresponding to a confidence level of 95%, was considered to determine extended uncertainties.

A solution made up of TSP in D_2O (20.33 ± 0.29 mg/L) was used to prepare six standard (labeled as A−E and Blank) and one test (labeled as X) mixtures at the levels listed in Table S1 in the Supporting Information (in the range 0−500 mg/L). Standard mixtures were used to obtain the calibration curves considered to determine the concentration values of the test mixture X. Mixtures were prepared by diluting stock solutions to the desired concentration using class A glassware. NMR tubes were filled in with 1.0 mL of the solution.

Experimental Procedures. Nine NMR signals were selected for this study: three for Aldicarb (A1, A2 and A3), one for Methamidophos (M1), two for Oxadixyl (O1 and O2), two for Pirimicarb (P1 and P2), and the singlet of TSP which was taken as reference. A typical ^{1}H NMR spectrum of the mixture is reported in Figure S1 in the Supporting Information and the integration ranges used for calculation of the peak areas are listed in Table S2 in the Supporting Information.

In order to choose the optimal recycle delay, T_1 values were determined taking into proper account all signals listed in Table S2 in the Supporting Information. T_1 determination was carried out by inversion recovery experiments applied to single component solutions (analyte in D_2O) at two different magnetic fields, 9.4 T (400 MHz) and 16.5 T (700 MHz), and two concentration levels, ∼37 mg/L and ∼600 mg/L. The highest T_1 value (5.4 s, measured for M1 signal of a 37.4 mg/L solution of Methamidophos at 9.4 T) was taken into account to set the recycle delay to 30 s. D_2O was not degassed before preparation of the solutions. Single component solutions and test mixtures were prepared in the same laboratory using the same batch of D_2O. NMR tubes were filled with 0.5 mL solution, sealed, and delivered to the participants.

Data Acquisition and Processing. The NMR experiment considered for the interlaboratory comparison consisted of a single 90° excitation pulse preceded by a selective presaturation step. Even though it was organized before the publication of the EUROLAB technical report on NMR method development and validation,[39] this work produced results coherent with guidelines described therein. For each NMR tube, 5 spectra were recorded to comply with conditions for repeatability (measurements performed under the same operating conditions over a short period of time) considering the same NMR tube, same spectrometer, same user, consecutive runs without removing the NMR tube from the magnet and to comply with conditions for intermediate precision (measurements performed under repeatability condition devoid of only one obligation) considering the same NMR tube, same spectrometer, same user, at least 24 h delay between runs, removal of the NMR tube from the magnet from run to run. Summarizing, each participant recorded 35 NMR spectra (5 replicates for each of the 7 NMR tubes) in three different sessions: (i) 3 consecutive runs per NMR tube (run 1, run 2, and run 3); (ii) 1 run per NMR tube delayed at least 24 h from the first session (run 4); (iii) 1 run per NMR tube delayed at least 24 h from the second session (run 5). It has been demonstrated[38] that results obtained in repeatability conditions (considering only data obtained by runs 1−3), in intermediate precision conditions (considering only data obtained by runs 1, 4, and 5) and both conditions (considering data obtained by runs 1−5) can be safely considered as substantially equivalent. In the present paper, calculation on all available replicates (runs 1−5) will be described. More details on NMR data acquisition and processing are reported in the Supporting Information.

Statistical Elaboration. Signal integrals were scaled to the TSP integral and the corresponding (I_{signal}/I_{TSP}) values were uploaded on a Web application specifically designed and validated for data elaboration in agreement with internationally accepted requirements.[35−37] (I_{signal}/I_{TSP}) values were uploaded reporting at least four decimal places. The five (I_{signal}/I_{TSP}) replicates collected for each signal and for each NMR tube were submitted to the Shapiro-Wilk test to ascertain their normal distribution and to Huber, Dixon, and Grubbs tests for identification of possible outliers. Throughout the paper, Grubbs tests refer to application of both the classical Grubbs test identifying one outlier and the double Grubbs test which enables the identification of two outliers. Data identified as outliers by all the four tests were not considered in successive steps. Data derived from standard mixtures A−E and Blank were used to plot (I_{signal}/I_{TSP}) versus analyte concentrations and to develop an equation for the calibration line by least-squares linear regression. The equation of general formula $y = ax + b$ (with $y = (I_{signal}/I_{TSP})$ and x = concentration as mg/L) was used to calculate concentration values of analytes in test mixture X. Then, the 5 concentration values calculated for the test mixture X were submitted to the Shapiro-Wilk test to ascertain their normal distribution and to Huber, Dixon, and Grubbs tests for identification of possible outliers. After removing outliers, calculated concentrations were used to determine the mean concentration values and the corresponding standard deviations which were considered as intralaboratory uncertainties of the method. Results from all participants (36 sets of results from 34 NMR spectrometers) were submitted to data elaboration for proficiency test and for determination of the assigned values for analytes in mixture X. The lack of official qNMR analyses for this case study prompted us to determine assigned values as consensus values from participants.[34] Thus, for each analyte, according to the flowchart suggested by Horwitz,[40] the 36 standard deviation values were submitted to the Cochran test (provided that all of the 5 replicates of mixture X successfully passed the above-mentioned tests for outliers) with the aim to identify and remove outliers for successive calculations. In turn, mean concentration values from data sets which passed successfully the Cochran test were submitted to Grubbs tests with the aim to further refine the quality of the results. The remaining sets of data were submitted to the Shapiro-Wilk test to ascertain the normal distribution of the population (data were always normal distributed after refinement by the Cochran and Grubbs tests) and were used to calculate, for each analyte in test mixture X, the assigned concentration value, the interlaboratory standard deviation, the coefficient of variation (CV%), and the reproducibility limits.

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

RESULTS AND DISCUSSION

Performance Assessment for Single Component Quantitative NMR Measurements. Among the quantification approaches available for NMR spectroscopy,[4] the calibration line method was chosen in this work as it allows for identification of a theoretical line to be taken as reference in performance assessment. Moreover, this method has a general applicability in analytical chemistry and has the advantage to nullify the effects of nuclei relaxation on quantitative accuracy, provided that all the acquisition parameters are kept constant for standard and test solutions.[4] Thus, it is expected that systematic errors deriving from hardware features or from the set of acquisition parameters should be minimized.

A first statistical data elaboration of the ILC was carried out by a single operator who processed NMR spectra (Fourier transformation, phase and baseline correction, signal integration) and obtained calibration lines with no scaled signal areas as input data.[38] In a second data elaboration, NMR data processing was carried out by each participant and signal areas were scaled to the TSP area. Therefore, the main difference between the two elaboration approaches relays on different processing conditions. Results of both elaborations are summarized in Table S3 in the Supporting Information where assigned concentration values along with the corresponding standard deviations, coefficients of variation, and reproducibility limits are reported. It is apparent that changing the processing conditions of the NMR spectra, from "one operator−all NMR data sets" to "one operator−one NMR data set", has a little impact on the final result in terms of mean value. Conversely, standard deviations (and consequently the related coefficients of variation and reproducibility limits) are affected by the different NMR processing conditions. Notwithstanding the deterioration of their quality in terms of coefficient of variation (CV%), these results are quite satisfactory if this test is considered as a confirmatory method for organic residues and contaminants. Indeed, according to the European Commission decision concerning the performance of analytical methods and the interpretation of results,[41] the interlaboratory coefficient of variation (CV%) for repeated analysis of a reference material, under reproducibility conditions, shall not exceed 5.7% for concentration values higher than 1000 ppm, according to the Horwitz equation:

$$\mathrm{CV\%} = 2^{(1-0.5 \log C)}$$

where C is the mass fraction expressed as a power of 10 (e.g., 1 mg/g = 10^{-3}). Being the concentration values considered in this work are lower than 500 mg/L, the highest obtained CV% value of 4.9% indicates that single excitation pulse preceded by selective presaturation of the solvent is a reliable NMR experiment for quantification purposes.

Once the assigned values for all the analytes were determined, performance statistics were carried out with the aim to estimate the deviation of the mean concentration values from the assigned value for each participant, including those producing data sets rejected by the Cochran and Grubbs tests. A commonly used parameter estimating the performance for quantitative results is the z-score, which is defined as

$$z = \frac{C_i - \bar{C}}{\sigma}$$

where C_i is the mean concentration value determined by the ith data set, $\bar{C}$ is the assigned concentration value, and σ is the interlaboratory standard error, all referred to as a single NMR signal. Satisfactory performance is indicated by $|z| \leq 2.0$, questionable performance is obtained when $2.0 < |z| < 3.0$, while $|z| \geq 3.0$ indicates unsatisfactory performance. In the latter case, suitable actions are required to identify and to solve the analytical problems.

Figure 1 shows the z-scores of Aldicarb quantification by the NMR A1 singlet. It is apparent that, even though results of 10

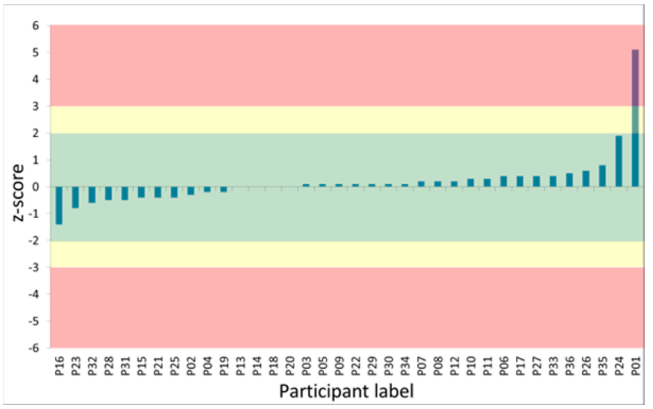


Figure 1. z-score for quantification of Aldicarb by means of A1 signal (green, $|z| \leq 2.0$; yellow, $2.0 < |z| < 3.0$; red, $|z| \geq 3.0$. Assigned concentration value, 94.57 mg/L; interlaboratory standard deviation, 3.64; reproducibility limit, 12.46; CV%, 3.8%).

participants were excluded from calculation of the assigned value, the quality of the result was satisfactory for 35 sets of NMR data and only 1 unsatisfactory performance was registered. Very similar results were obtained using each of all other NMR signals (Supporting Information, Figures S2−S8). High-performance quantifications are obtained also when signals different from singlets were taken into account (as in the case of M1 and O2 where a doublet and a group of signals were considered, respectively). It is worth noting that performance in terms of result quality was not affected by the magnetic field, hardware configuration, manufacturer, and production year of the spectrometer. These findings highlight the robustness of NMR spectroscopy when calibration curve approaches are adopted.

The z-score represents a satisfactory indicator for performance assessment in single component analyses, but it cannot account for performance assessment in multicomponent analyses because a single z-score refers to only a single quantification measurement. Thus, for performance assessment in fingerprinting measurements and quantitative multicomponent analyses, the introduction of indicators more appropriate than the z-score is desirable.

Basics of Quantitative NMR. Before discussing the new quality control parameters proposed by us in performance assessment for fingerprinting measurements and quantitative multicomponent analyses, recall of the basic equation of quantitative NMR is advisible (eq 1).

$$I = kn \quad (1)$$

Equation 1 provides the direct proportionality between the number of moles (n) of nuclei generating a signal and the intensity (I) of the same signal with a proportionality constant k being the spectrometer constant which remains the same for all resonances in a NMR spectrum.[4]

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

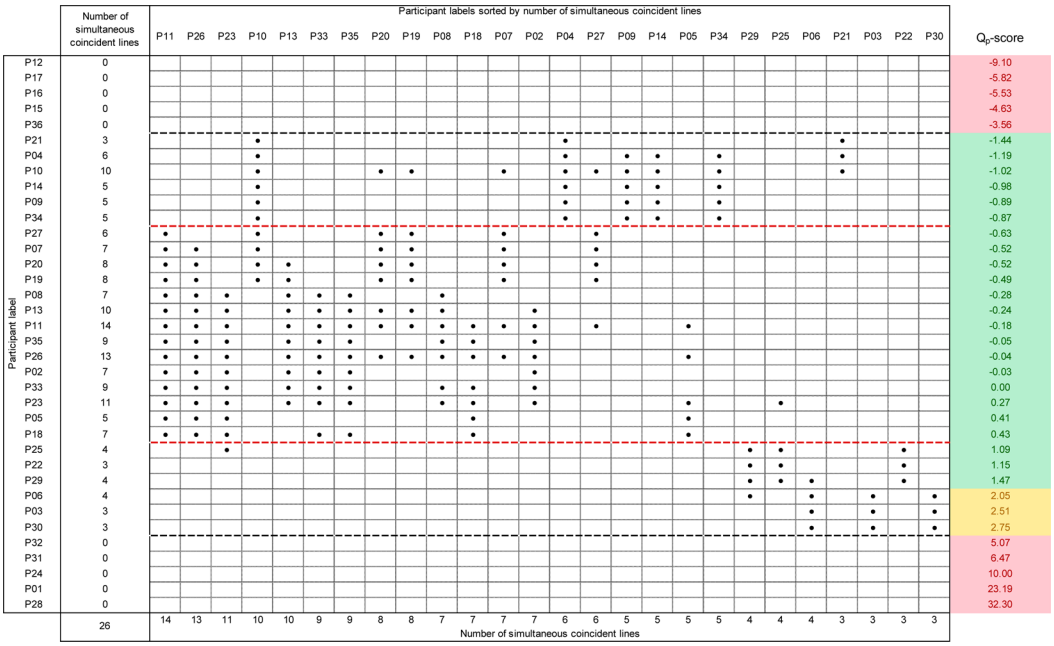


Figure 2. Results of the paired t test for statistical equivalence of pairs of calibration lines and laboratory Q_p-scores (referred to the A1 signal). Equivalent lines are cross-linked by the ●; green, $|Q_p| \leq 2.0$; yellow, $2.0 < |Q_p| < 3.0$; red, $|Q_p| \geq 3.0$.

Let us consider, in a NMR spectrum, the signal (a) having intensity I_a generated by specific protons belonging to the analyte of interest and the signal (r) having intensity I_r generated by specific protons in a reference compound. Applying eq 1 to I_a and I_r gives

$$I_a = kn_a$$

$$I_r = kn_r$$

Hence the ratio $(I_a/I_r) = (n_a/n_r)$ (eq 2) is independent from the proportionality constant k and, as a consequence, it does not depend on the spectrometer. Thus, taking the methyl protons signal of TSP as reference signal, all of the calibration lines obtained plotting (I_a/I_{TSP}) versus analyte concentration (C) should be independent from the spectrometer and statistically equivalent to each other. In other words, all the participants to an ILC should develop equivalent calibration lines

$$\left(\frac{I_a}{I_{TSP}}\right) = aC + b \tag{3}$$

where intercept b should have a null value due to the fact that no signal is generated if no nuclei (C = 0 mg/L) are contained in the mixture. Thus, eq 3 can be rewritten as

$$\left(\frac{I_a}{I_{TSP}}\right) = \left(\frac{n_a}{n_{TSP}}\right) = \left(\frac{\frac{m_{analyte}}{M_{analyte}}N_a}{\frac{m_{TSP}}{M_{TSP}}N_{TSP}}\right) = aC = a\frac{m_{analyte}}{V} \tag{4}$$

where $m_{analyte}$ is the mass of the analyte, m_{TSP} is the mass of TSP, $M_{analyte}$ is the molar mass of the analyte, M_{TSP} is the molar mass of TSP, N_a is the number of protons generating the signal (a), N_{TSP} is the number of methyl protons (nine) generating the reference signal, and V the solution volume.

Equation 4 can be rearranged into

$$\left(\frac{\frac{m_{analyte}}{M_{analyte}}N_a}{\frac{m_{TSP}}{M_{TSP}}N_{TSP}}\right) = a\frac{m_{analyte}}{V} \tag{5}$$

From eq 5 the theoretical value that slope must assume for a given TSP concentration can be extracted:

$$a_{theoretical} = \frac{M_{TSP}}{M_{analyte}}\frac{N_a}{N_{TSP}}\frac{V}{m_{TSP}} = \frac{M_{TSP}}{M_{analyte}}\frac{N_a}{9}\frac{1}{C_{TSP}} \tag{6}$$

The need to harmonize NMR protocols prompted us to propose a new parameter suited for checking the equivalence of the calibration lines. Such a parameter will be shown to represent a quality control index of the NMR spectra to use in fingerprinting applications and multicomponent NMR quantifications.

Quality Control Parameters for Performance Assessment in Fingerprinting Measurements and Quantitative Multicomponent Analyses. In order to assess the laboratory performance in multicomponent analyses without considering as many z-scores as the number of analytes, we propose a new parameter, named Q_p-score, accounting for participant performance as the result of instrumental adequacy and operator skill. Knowing that, for each signal, calibration lines developed by each participant must be equivalent to each other, let us define the indicator of the line equivalence Q_p as

$$Q_p = \frac{a_i - \bar{a}}{\sigma_{slope}} \tag{7}$$

where a_i is the slope of the calibration line determined by the ith participant, $\bar{a}$ is the consensus slope value, and σ_{slope} is the interlaboratory standard deviation on slopes, all referred to a single NMR signal. The values $\bar{a}$ and σ_{slope} are determined using a_i successfully passing the Huber test. By an analogous reasoning followed for the z-score, performance assessment by the Q_p-score is considered satisfactory when $|Q_p| \leq 2.0$,

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

Participant label	Signal A1	A2	A3	M1	O1	O2	P1	P2
P12	-9.10	-8.07	-6.46	-6.62	-8.55	-6.67	-6.59	-6.68
P17	-5.82	-5.29	-4.03	-4.21	-5.50	-4.29	-4.29	-4.19
P16	-5.53	-4.90	-4.08	-4.07	-5.35	-4.18	-4.01	-4.00
P15	-4.63	-4.12	-3.25	-3.34	-4.33	-3.33	-3.31	-3.32
P36	-3.56	-3.20	-2.67	-2.60	-3.39	-2.83	-2.56	-2.58
P21	-1.44	-1.34	-1.11	0.22	-0.68	0.78	-1.21	-1.01
P04	-1.19	-1.11	-0.84	-0.82	-1.20	-0.96	-0.82	-0.82
P10	-1.02	-0.99	-0.71	-0.71	-1.04	-0.78	-0.72	-0.68
P14	-0.98	-1.00	-0.96	-0.86	-0.89	-0.66	-1.05	-1.03
P09	-0.89	-0.96	-0.84	-0.71	-0.86	-0.73	-0.75	-0.74
P34	-0.87	-0.91	-0.77	-0.58	-1.04	-0.75	-0.49	-0.83
P27	-0.63	-0.62	-0.42	-0.36	-0.71	-0.47	-0.29	-0.60
P07	-0.52	-0.65	-0.39	-0.29	-0.48	-0.39	-0.35	-0.31
P20	-0.52	-0.50	-0.37	-0.48	-0.59	-0.46	-0.30	-0.25
P19	-0.49	-0.55	-0.32	-0.29	-0.67	-0.39	-0.29	-0.14
P08	-0.28	-0.39	-0.20	-0.06	-0.34	-0.23	-0.18	-0.01
P13	-0.24	-0.37	-0.13	-0.07	-0.25	-0.19	-0.14	0.01
P11	-0.18	-0.22	1.03	-0.72	-1.13	-0.90	-0.46	-0.82
P35	-0.05	-0.26	0.21	-0.04	-0.03	-0.20	-0.10	0.08
P26	-0.04	-0.23	-0.10	-1.28	-0.06	0.41	-0.38	-0.25
P02	-0.03	-0.15	-0.31	0.13	0.16	-0.45	0.10	0.01
P33	0.00	0.06	-0.22	-0.03	0.15	-0.05	0.14	-0.01
P23	0.27	0.54	0.65	0.11	-0.07	0.86	0.50	0.59
P05	0.41	0.22	0.33	0.44	0.17	0.21	0.32	0.54
P18	0.43	0.40	0.22	0.49	0.50	0.44	0.47	0.62
P25	1.09	0.80	0.77	0.83	0.75	0.71	0.86	1.04
P22	1.15	0.80	0.77	0.99	0.83	0.84	1.02	0.80
P29	1.47	1.20	1.08	1.14	1.16	0.97	1.12	1.01
P06	2.05	1.86	1.63	1.56	1.80	1.53	1.59	1.53
P03	2.51	2.08	1.72	1.85	2.15	1.68	1.90	1.74
P30	2.75	2.32	1.96	2.13	2.38	1.99	2.08	2.11
P32	5.07	4.28	3.56	3.83	4.49	3.68	3.82	3.73
P31	6.47	5.54	4.62	4.91	5.82	4.77	4.88	4.76
P24	10.00	8.62	7.08	7.52	9.27	7.27	7.50	7.43
P01	23.19	20.03	16.33	15.56	20.26	16.45	16.51	16.56
P28	32.30	28.19	22.62	22.61	28.75	23.11	23.21	23.35

Figure 3. Q_p-scores for all NMR signals as labeled in Table S2 in the Supporting Information. Green, $|Q_p| \leq 2.0$; yellow, $2.0 < |Q_p| < 3.0$; red, $|Q_p| \geq 3.0$.

questionable when $2.0 < |Q_p| < 3.0$ and unsatisfactory when $|Q_p| \geq 3.0$.

In the case study of Aldicarb quantification by NMR signal A1, Huber tests applied to the 36 slope values gave 11 outliers, the 5 lowest and the 6 highest values. The 25 remaining values resulted in a normal distribution after the Shapiro-Wilk test and were considered for the calculation of $\bar{a}$ and σ_{slope}. The values of $\bar{a}$ and σ_{slope} were 0.0340 L/mg and 0.0032 L/mg, respectively.

Concerning the experimental intercept values, the population was not normal and was too scattered so that iterated Huber test gave meaningless results (all values were identified as outliers). The mean value of the intercept was −0.048, and the related standard deviation was 0.513 indicating that the null value can be well considered as the experimental intercept.

These results give $y = 0.0340x$ as the consensus equation for the calibration line but do not yet demonstrate the statistical equivalence of the calibration lines. In order to evaluate the statistical parallelism, and then the equivalence of the calibration lines, all possible slope pairs were submitted to the paired t test with 95% confidence level. Computational part of the test consists of calculation of parameter t_{slopes} as the difference between two slopes divided by the standard error of the difference between the same two slopes.[42] Then, t_{slopes} was compared with Student's t at the desired confidence level (95%) to evaluate whether the null hypothesis was supported, that is whether no relationship between two data sets existed. If the slope obtained by one data population is significantly different from that generated with another (and independent) data set, then $t_{slopes} > t$ or else $t_{slopes} \leq t$, and the two slopes can be considered statistically equivalent. Results of the paired t tests applied to all possible slope pairs are summarized in Figure 2 where statistically equivalent lines are cross-linked by black circles. For instance, participant P11 produced a calibration line which is equivalent to those produced by participants P27, P07, P20, P19, P08, P13, P35, P26, P02, P33, P23, P05, and P18.

It is apparent from Figure 2 that the slopes of 26 calibration lines (bordered by black dashed lines) are statistically equivalent. Of these 26 lines, 23 were characterized by $|Q_p|$ lower than 2, i.e., in the range of satisfactory performance assessment. Moreover, with the exception of participant P10, the highest number of simultaneous coincidences (7−14) was recorded for participants endowed with very low Q_p-scores (ranging from −0.63 to 0.43, bordered by red dashed lines in Figure 2).

The statistical equivalence of the calibration lines is in agreement with the theoretical treatment described above. It represents the experimental evidence that, for a given TSP concentration, slopes assume a certain value depending on the signal and not on the spectrometer constants. Once defined, the concentration range of the analytes, slopes associated with satisfactory Q_p-scores indicate that the corresponding NMR spectra were recorded and processed under similar conditions. Deviation from the consensus value of the slope is explained in terms of hardware reliability, acquisition and processing parameters. Therefore, Q_p-score represents a quality control index which accounts for hardware functioning conditions and operator skills. It is important to point out that questionable and unsatisfactory Q_p-scores did not prevent successful single component quantifications as the latter depend only on the quality of the calibration line. Indeed, good fitting in the linear regression allows for a good performance in terms of z-score but it does not account for deviation of the slope from its theoretical value.

Given that the Q_p-score is a quality parameter of the NMR spectrum as a whole, it can be expected that, as far as multicomponent analysis is concerned, Q_p-score based performance assessment of a laboratory should be almost independent from the considered signal. This is indeed the case, as clearly

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

Table 1. NR Values (%) Calculated for All NMR Signals As Labeled in Table S2 in the Supporting Information[a]

signal	A1	A2	A3	M1	O1	O2	P1	P2
$M_{analyte}$ (g/mol)	190	190	190	141	278	278	238	238
N_a	6	3	1	3	3	3	3	3
$a_{theoretical} \times 10^2$ (L/mg)	2.97	1.48	0.49	2.00	1.01	1.01	1.19	1.19
$\bar{a} \times 10^2$ (L/mg)	3.40	1.72	0.54	1.95	0.95	1.20	1.39	1.38
NR (%)	−14.6	−15.7	−9.3	2.4	6.2	−18.2	−17.0	−16.7

[a] M_{TSP} = 172.27 g/mol; N_{TSP} = 9; C_{TSP} = 20.33 mg/L.

demonstrated by inspection of Figure 3, where Q_p-scores obtained by each participant by considering each of the NMR signals selected for this study are reported. Apart from the variations of the Q_p-scores falling in the proximity of the limiting value ±2, the performance category ($|Q_p| \leq 2.0$, $2.0 < |Q_p| < 3.0$ and $|Q_p| \geq 3.0$) is retained for all considered signals. These findings are in good agreement with the high reproducibility of ^{1}H NMR experiments evaluated by PCA in previous studies.[32,33]

The introduction of Q_p-score paves the way to validation of multicomponent quantification methods, of great importance for fingerprinting and profiling applications. In fact, such validation procedures might be carried out in the future by an interlaboratory comparison where laboratory performance could be preliminarily assessed developing calibration lines for any arbitrary compound mixture. Once a Q_p acceptability range is fixed (for instance, $|Q_p| < 1$), all laboratories within such a range will be qualified to produce NMR spectra of a given mixture that are statistically equivalent in terms of relative intensities of the signals. In other words, for a given set of acquisition parameters, laboratories gaining satisfactory Q_p-scores will be accredited to record NMR spectra on every kind of mixture, thus allowing for pooling of NMR data in suitable databanks.

It is worth noting that, in real experiments, a deviation from theoretical slope is expected due to the specific response of the nuclei to the experienced excitation/relaxation conditions during spectrum acquisition. Such a response depends on several factors including (i) hard excitation pulse which must be uniform throughout all the spectral width; (ii) proximity of the signals to the offsets; (iii) recycle delay, which must be long enough to allow for complete magnetization recovery of all nuclei; (iv) energy exchange effects (NOE, spin diffusion, etc.) introduced by soft pulses. Therefore, in any interlaboratory comparison the consensus slope may differ from the theoretical one as an effect of the specific set of acquisition parameters.

In order to gain insights into the effects of the experimental excitation/relaxation conditions on the nuclei response, we introduce a new indicator as the relative deviation of the consensus slope from the theoretical value, according to eq 8.

$$NR = \frac{a_{theoretical} - \bar{a}}{a_{theoretical}} \times 100 \quad (8)$$

NR calculated for all signals considered in this study are reported in Table 1.

NR values for the various signals ranged from −18.2% to 6.2% indicating that signals are not affected to the same extent by the used acquisition parameters. Moreover, NR values were different also for signals generated by inequivalent nuclei in the same molecule.

In the present case, NR represents an index of the response of the nuclei submitted to a NMR experiment characterized by a single 90° excitation pulse preceded by a selective presaturation step with the specific set of acquisition parameters. In our opinion, among the above-mentioned factors affecting the nuclei response, energy exchange effects introduced by soft pulses can be considered the most relevant to interpret the NR values obtained in the present study. Energy exchange effects are certainly operative in the acquisition condition characterized by a selective pulse acting during the long recycle delay (30 s). The other factors are thought to affect NR values only marginally because possible incorrect setting of the pulses (factors i and ii) will give random contributions averaged to almost null deviation of the calibration line and because the adopted recycle delay (longer than 5 times the highest measured T_1) ensures complete recovery of the magnetization (factor iii). Anyway, a deeper study on factors affecting the nuclei response to experimental acquisition conditions to give the NR values reported in Table 1 requires further NMR experiments. This is out of the scope of the present paper.

CONCLUSION

This study introduces a new quality control parameter, Q_p-score, suitable for harmonization of fingerprinting protocols and quantitative multicomponent analysis. Such a parameter, that was designed considering consolidated internationally agreed statistics, represents an unbiased evaluation tools for NMR method validations.

The Q_p-score accounts for laboratory performance in terms of both instrumental adequacy and operator skill and enables laboratories to pooling of NMR data in suitable databanks. Moreover, Q_p can be valuable for the development of multilaboratory metabolomic platforms. In fact, it was shown that participants having a Q_p-score in a suitable acceptability range are able to produce NMR spectra of a given mixture that can be considered statistically equivalent in terms of relative intensities of the signals. Another practical use of Q_p-score consists of the entitlement of laboratories endowed with acceptable $|Q_p|$ values to carry out quantifications by using relative intensity of the signal of interest after fitting with the consensus calibration line deriving from the interlaboratory comparison. For instance, in suitable networking conditions, equivalent calibration lines could be shared to enable different laboratories to carry out quantitative analyses without wasting time in calibration steps, with a consequent increase of productivity.

Since basic equations of quantitative NMR are independent from the type of solvent, it can be expected that performance assessment by Q_p-score applies also to experiments carried out in solvents others than D_2O (for instance, in organic solvents such as $CDCl_3$ widely used in lipidomics and in complex mixtures such as biofluids which are mostly used in their native state).

Another parameter, NR, has been proposed, which is related to differences between the theoretical and the consensus slopes

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

of the calibration lines and which is specific for each signal produced by a well-defined set of acquisition parameters. For a given molecule in a defined solvent, NR represents an index of the specific response of the various nuclei submitted to a definite NMR experiment.

■ ASSOCIATED CONTENT

Supporting Information

Complete list of affiliations and complete list of acknowledged individuals; chart of chemical structures of five components contained in the model mixture; Tables S1 (analyte concentration values assessed by gravimetric method in standard and test mixtures), S2 (signal labels, chemical shifts and integration ranges used for the calculation of peak areas), and S3 (comparison between two data elaboration approaches); data acquisition protocol; NMR data processing; and a typical ^{1}H NMR spectra of the mixture (Figure S1) and additional *z*-score plots (Figures S2−S8). The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.analchem.5b00919.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: vito.gallo@poliba.it. Phone: +39 0805963607. Fax: +39 0805963611.

Author Contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

Notes

The authors declare no competing financial interest.

[4−33]The complete list of the affiliations is reported in the Supporting Information.

■ ACKNOWLEDGMENTS

This work was carried out with voluntary contribution of all participants under the hint of the Chamber of Commerce of Bari. Alessandro Ambrosi (President of the Chamber of Commerce of Bari), Angela Patrizia Partipilo (Secretary General of the Chamber of Commerce of Bari), Piero Pontrelli (Director of SAMER), Umberto Bucci and Giuseppe Margiotta (President and Vice President of SAMER, respectively) are gratefully acknowledged for financial support in organization of promotion events. GIDRM (Gruppo Italiano di Discussione sulle Risonanze Magnetiche) and GIRM (Gruppo Interdivisionale di Risonanze Magnetiche della Società Chimica Italiana) are also acknowledged. The complete list of acknowledged persons is reported in the Supporting Information.

■ REFERENCES

(1) Purcell, E. M.; Torrey, H. C.; Pound, R. V. *Phys. Rev.* **1946**, *69*, 37−38.

(2) Bloch, F.; Hansen, W. W.; Packard, M. *Phys. Rev.* **1946**, *69*, 127−127.

(3) Bloch, F.; Hansen, W. W.; Packard, M. *Phys. Rev.* **1946**, *70*, 474−485.

(4) Bharti, S. K.; Roy, R. *Trends Anal. Chem.* **2012**, *35*, 5−26.

(5) For site-specific natural isotope fractionation (SNIF-NMR) see (a) method OIV-MA-AS311-05; (b) AOAC Official method 995.17; (c) AOAC Official method 2006.05; (d) AOAC Official Method 2000.19; for solid fat content determination see (e) AOCS Official Method Cd 16b-93; (f) AOCS Official Method Cd 16-81; for moisture and fat in meats see (g) AOAC Official Method 2008.06.

(6) Del Coco, L.; Assfalg, M.; D'Onofrio, M.; Sallustio, F.; Pesce, F.; Fanizzi, F. P.; Schena, F. P. *Metabolomics* **2013**, *9*, 740−751.

(7) Napoli, C.; Sperandio, N.; Lawlor, R. T.; Scarpa, A.; Molinari, H.; Assfalg, M. *J. Proteome Res.* **2012**, *11*, 1274−1283.

(8) Assfalg, M.; Bortoletti, E.; D'Onofrio, M.; Pigozzi, R.; Molinari, H.; Boner, A. L.; Peroni, D. G.; Piacentini, G. L. *Br. J. Derm.* **2012**, *166*, 1123−1125.

(9) Mucci, A.; Parenti, F.; Righi, V.; Schenetti, L. *Food Chem.* **2013**, *141*, 3167−3176.

(10) Pertinhez, T. A.; Casali, E.; Lindner, L.; Spisni, A.; Baricchi, R.; Berni, P. *Blood Transfus.* **2014**, *12*, 548−556.

(11) Tomassini, A.; Vitalone, A.; Marini, F.; Praticò, G.; Sciubba, F.; Bevilacqua, M.; Delfini, M.; Di Sotto, A.; Di Giacomo, S.; Mariani, P.; Mammola, C. L.; Gaudio, E.; Miccheli, A.; Mazzanti, G. J. *Proteome Res.* **2014**, *13*, 5848−5859.

(12) Brasili, E.; Praticò, G.; Marini, F.; Valletta, A.; Capuani, G.; Sciubba, F.; Miccheli, A.; Pasqua, G. *Metabolomics* **2014**, *10*, 1186−1196.

(13) Calvani, R.; Brasili, E.; Praticò, G.; Sciubba, F.; Roselli, M.; Finamore, A.; Marini, F.; Marzetti, E.; Miccheli, A. *J. Clin. Gastroenterol.* **2014**, *48*, S5−S7.

(14) Sciubba, F.; Capuani, G.; Di Cocco, M. E.; Avanzato, D.; Delfini, M. *Food Res. Int.* **2014**, *62*, 66−73.

(15) Caligiani, A.; Coisson, J. D.; Travaglia, F.; Acquotti, D.; Palla, G.; Palla, L.; Arlorio, M. *Food Chem.* **2014**, *148*, 77−85.

(16) Caligiani, A.; Palla, L.; Acquotti, D.; Marseglia, A.; Palla, G. *Food Chem.* **2014**, *157*, 94−99.

(17) Airoldi, C.; Tripodi, F.; Guzzi, G.; Nicastro, R.; Coccetti, P. *Mol. BioSyst.* **2015**, *11*, 379−383.

(18) Petrakis, E. A.; Cagliani, L. R.; Polissiou, M. G.; Consonni, R. *Food Chem.* **2015**, *173*, 890−896.

(19) Cagliani, L. R.; Pellegrino, G.; Giugno, G.; Consonni, R. *Talanta* **2013**, *106*, 169−173.

(20) Consonni, R.; Cagliani, L. R.; Stocchero, M.; Porretta, S. *J. Agric. Food Chem.* **2010**, *58*, 7520−7525.

(21) Mallamace, D.; Corsaro, C.; Salvo, A.; Cicero, N.; Macaluso, A.; Giangrosso, G.; Ferrantelli, V.; Dugo, G. *Physica A* **2014**, *401*, 112−1170.

(22) Cicero, N.; Corsaro, C.; Salvo, A.; Vasi, S.; Giofrè, S. V.; Ferrantelli, V.; Di Stefano, V.; Mallamace, D.; Dugo, G. *Nat. Prod. Res.* **2015**, DOI: 10.1080/14786419.2015.1012166.

(23) Rizzuti, A.; Aguilera-Saez, L. M.; Gallo, V.; Cafagna, I.; Mastrorilli, P.; Latronico, M.; Pacifico, A.; Matarrese, A. M.; Ferrara, G. *Food Chem.* **2015**, *171*, 341−350.

(24) Gallo, V.; Mastrorilli, P.; Cafagna, I.; Nitti, G. I.; Latronico, M.; Longobardi, F.; Minoja, A. P.; Napoli, C.; Romito, V. A.; Schäfer, H.; Schütz, B.; Spraul, M. J. *Food Compos. Anal.* **2014**, *35*, 44−52.

(25) Ferrara, G.; Mazzeo, A.; Pacucci, C.; Netti, G.; Cafagna, I.; Mastrorilli, P.; Vezzoso, M.; Gallo, V. *Am. J. Enol. Viticult.* **2014**, *65*, 381−387.

(26) Longobardi, F.; Ventrella, A.; Bianco, A.; Catucci, L.; Cafagna, I.; Gallo, V.; Mastrorilli, P.; Agostiano, A. *Food Chem.* **2013**, *141*, 3028−3033.

(27) Rizzuti, A.; Caliandro, R.; Gallo, V.; Mastrorilli, P.; Chita, G.; Latronico, M. *Food Chem.* **2013**, *141*, 1908−1915.

(28) Aghemo, C.; Albertino, A.; Gobetto, R.; Lussiana, C.; De Maria, A.; Isocrono, D. *Eur. J. Lipid. Sci. Technol.* **2012**, *114*, 1409−1416.

(29) Aghemo, C.; Albertino, A.; Gobetto, R.; Spanna, F. *J. Sci. Food Agric.* **2011**, *91*, 2088−2094.

(30) A nonexhaustive list of some recent examples is reported: (a) Weber, M.; Hellriegel, C.; Rück, A.; Sauermoser, R.; Wüthrich, J. *Accredit. Qual. Assur.* **2013**, *18*, 91−98. (b) Ohtsuki, T.; Sato, K.; Sugimoto, N.; Akiyama, H.; Kawamura, Y. *Talanta* **2012**, *99*, 342−348. (c) Schoenberger, T. *Anal. Bioanal. Chem.* **2012**, *403*, 247−254.

(31) Malz, F.; Jancke, H. *J. Pharm. Biomed. Anal.* **2005**, *38*, 813−823.

(32) Ward, J. L.; Baker, J. M.; Miller, S. J.; Deborde, C.; Maucourt, M.; Biais, B.; Rolin, D.; Moing, A.; Moco, S.; Vervoort, J.; Lommen, A.; Schäfer, H.; Humpfer, E.; Beale, M. H. *Metabolomics* **2010**, *6*, 263−273.

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX

(33) Viant, M. R.; Bearden, D. W.; Bundy, J. B.; Burton, I. W.; Collette, T. W.; Ekman, D. R.; Ezernieks, V.; Karakach, T. K.; Yu Lin, C.; Rochfort, S.; De Ropp, J. S.; Teng, Q.; Tjeerdema, R. S.; Walter, J. A.; Wu, H. *Environ. Sci. Technol.* **2009**, *43*, 219−225.
(34) *ISO/IEC 17043:2010* and reference norms cited therein.
(35) *ISO/IEC 17025:2005* and reference norms cited therein.
(36) *ISO 13528:2005* and reference norms cited therein.
(37) (a) *ISO 5725-1:1994*; (b) *ISO 5725-2:1994*; (c) *ISO 5725-3:1994*; (d) *ISO 5725-4:1994*; (e) *ISO 5725-5:1998*; (f) *ISO 5725-6:1994* and reference norms cited therein.
(38) Gallo, V.; Scapicchio, P.; Vezzoso, M.; Fanizzi, P. *Risonanza Magnetica e Controllo Qualità−Presentazione dei Risultati del Primo Confront Interlaboratorio Nazionale*; Camera di Commercio di Bari, 2013 (available free of charge at http://www.samer.it/articoli/allegato/Samer_rev1_def.pdf and at http://www.innovative-solutions.it/wp-content/uploads/2013/09/Pubblicazione-con-ISBN.pdf).
(39) EUROLAB. *Technical Report 1/2014, Guide to NMR Method Development and Validation−Part 1: Identification and Quantification*, 2014.
(40) Horwitz, W. *Pure Appl. Chem.* **1995**, *67*, 331−343.
(41) COMMISSION DECISION of 12 August 2002 implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results (2002/657/EC), *Official Journal of the European Communities*, August 17, 2002.
(42) Snedecor, G. W.; Cochran, W. G. *Statistical Methods*, 8th ed.; Iowa State University Press: Ames, IA, 1989.

DOI: 10.1021/acs.analchem.5b00919
Anal. Chem. XXXX, XXX, XXX−XXX