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Updated genetic testing of Italian patients referred with a clinical diagnosis of primary hyperoxaluria.

Alessandra Pelle^{1, 2}, Alessandra Cuccurullo^{1, 2}, Cecilia Mancini³, Regina Sebastiano², Giovanni Stallone⁴, Susanna Negrisola⁵, Elisa Benetti⁵, Licia Peruzzi⁶, Michele Petrarulo⁷, Mario De Marchi^{1, 2}, Martino Marangella⁸, Antonio Amoroso³, Daniela Giachino^{1, 2}, Giorgia Mandrile¹

¹ University of Torino, Department of Clinical & Biological Sciences, Orbassano, Italy

² Medical Genetics Unit, San Luigi University Hospital, Orbassano, Italy

³ University of Torino, Department of Medical Sciences, Città della Salute e della Scienza, Torino, Italy

⁴ University of Foggia, Department of Medical and Surgical Sciences, Foggia, Italy

⁵ Pediatric Nephrology, Dialysis and Transplant Unit, Department of Pediatrics, University of Padova, Italy

⁶ Nephrology Dialysis Transplantation Unit, Regina Margherita Children's Hospital, Città della Salute e della Scienza, Torino, Italy

⁷ S.C. Laboratorio Centrale, A.O. Ordine Mauriziano, Torino, Italy

⁸ Fondazione Scientifica, A.O. Ordine Mauriziano, Torino, Italy

Corresponding author:

Alessandra Pelle, University of Torino, Department of Clinical & Biological Sciences, San Luigi University Hospital, Regione Gonzole 10 - 10040 Orbassano (TO) - Italy. phone: +39-0119026205, fax: +39-0116705428, genmed-dscb@unito.it, alessandraelisapelle@gmail.com

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ABSTRACT

Background: Primary hyperoxaluria (PH) is a rare autosomal recessive disease commonly arising in childhood and presenting with nephrolithiasis, nephrocalcinosis and/or chronic renal failure. Three responsible genes are known: alanine-glyoxylate aminotransferase (*AGXT*, PH type 1), glyoxylate reductase/hydroxypyruvate reductase (*GRHPR*, PH type 2), and 4-hydroxy-2-oxoglutarate aldolase (*HOGAI*, PH type 3). In our centre at the end of 2014 molecular diagnosis of PH1 had been performed in 80 patients, while one patient received a PH2 diagnosis.

Material and Methods: Fifteen patients referred to our Centre and suspected to have PH on clinical grounds were negative for pathogenic variants in the entire coding sequence and exon-intron boundaries of the *AGXT* gene. Therefore, we extended the analysis to the *AGXT* promoter region and the *GRHPR* and *HOGAI* genes.

Results: Two patients were heterozygous for two novel *AGXT*-promoter variants (c.-647C>T, c.-424C>T) that were probably non pathogenic. One patient was homozygous for a novel *HOGAI* variant of intron 2 (c.341-81delT), whose pathogenicity predicted by *in silico* splicing tools was not confirmed by a minigene splicing assay in COS-7 and HEK293T cells.

Conclusion: New genetic subtypes of PH can be hypothesized in our patients, that may be caused by mutations in (an)other gene/genes encoding proteins of glyoxylate metabolism.

Alternatively, some kind of mutations (e.g. deletions/duplications, deep intronic splicing regulatory variants) could be missed in a few cases, similarly to other genetic diseases.

Keywords: *AGXT*, glyoxylate metabolism, *GRHPR*, *HOGAI*, primary hyperoxaluria.

Abbreviation footnote:

AGT: alanine-glyoxylate aminotransferase

ESRD: end stage renal disease

GFR: glomerular filtration rate

GRHPR: glyoxylate reductase/hydroxypyruvate reductase

GO: glycolate oxidase

HOGA: 4-hydroxy-2-oxoglutarate aldolase

LDH: lactate dehydrogenase

nv: normal value

PH: primary hyperoxaluria

PH1: primary hyperoxaluria type 1

PH2: primary hyperoxaluria type 2

PH3: primary hyperoxaluria type 3

Pox: plasma oxalate

Uox: urinary oxalate

WT: wild type

Introduction

Primary hyperoxaluria (PH) is a rare autosomal recessive disease (prevalence $1-3 \times 10^{-6}$) with onset mainly in childhood [1]. It is characterized by increased levels of urinary and plasma oxalate, nephrolithiasis and/or nephrocalcinosis leading to end stage renal disease (ESRD) and systemic oxalosis [2]. Three types of PH are currently known, but further genetic heterogeneity has been hypothesized, since in some patients the clinical suspicion of PH is not confirmed by molecular analysis [1].

PH1 (MIM #259900), the most frequent and severe form, is caused by an impaired function of the alanine-glyoxylate aminotransferase enzyme (AGT), encoded by the *AGXT* gene (MIM *604285) [3,4]. Deficiency of glyoxylate reductase/hydroxypyruvate reductase enzyme, encoded by the *GRHPR* gene (MIM *604296), is responsible of PH2 (MIM #260000) [5]. PH2 patients rarely develop systemic oxalosis and only 20% of them progress to ESRD [5-7]. A peculiar biochemical PH2 characteristic is the presence of increased urinary L-glyceric acid [8]. PH3 (MIM #613616) is caused by mutations in the *HOGA1* gene (MIM*613597), that encodes the mitochondrial enzyme 4-hydroxy-2-oxoglutarate aldolase (HOGA), catalysing the final step of 4-hydroxy-2-oxoglutarate catabolism to glyoxylate and pyruvate [9]. The age at onset of signs and symptoms is similar to those of PH1 and PH2, but in most PH3 patients a clinical remission is observed in the adult, despite the persistence of high oxalemia and oxaluria levels [10,11].

High 24-hour urinary oxalate (Uox) levels are suggestive of PH in the absence of secondary causes of hyperoxaluria, such as inflammatory bowel diseases or intestinal malabsorption [1,4]. Another important parameter is the molar urinary Oxalate/Creatinine ratio, that should always be related to the age of the subject, since higher ratios are observed in children due to lower urinary creatinine concentrations [12]. In patients with ESRD, Uox can be within the normal range or even below because of the decreased glomerular filtration; in such cases Plasma Oxalate (Pox) measurement it is preferable [1].

The assay of *AGXT* activity on hepatic tissue, once considered as the gold standard in PH1 diagnosis, is at present rarely performed, since the less invasive genetic testing has become available. In addition, the clinical presentation of the three PH types is largely overlapping, thus genetic analysis is an important tool to differentiate the three subtypes. According to the current recommendations, a different treatment is indicated for each PH type: combined kidney-liver transplantation is recommended only for PH1 [3], while isolated kidney transplantation is mostly performed in PH2, and no specific need of transplantation has been so far reported for PH3.

Moreover, *AGXT* genotype can help in predicting the effectiveness of vitamin B6 supplementation [1].

Here we report the 80 Italian PH1 and the only PH2 patients currently known in our Country, collected until the end of 2014 and we describe a group of 15 patients with a clinical suspicion of PH but without a mutation in the three known genes. Moreover, we characterize the functional assay of a *HOGAI* variant of unknown pathogenicity.

Material and methods

Major clinical criteria to suspect PH included nephrolithiasis, nephrocalcinosis and precocious age at onset of symptoms, while macro/micro hematuria and recurrent urinary infections were considered as indirect signs of renal stone disease. Glomerular filtration rate (GFR) in children was evaluated using the appropriate Schwartz's formula according to creatinine dosage assay while for adults the MDRD or CKD-EPI was employed. Urinary tract anomalies and secondary causes of hyperoxaluria, such as intestinal malabsorption and inflammatory bowel disease, were excluded. Biochemical parameters included oxalemia, oxaluria, urinary glycolate (see ref [12] for normal values) and renal function (plasma creatinine and GFR). Data about parents' consanguinity and the familiar history were collected.

Sanger sequencing of AGXT, GRHPR and HOGAI genes

Genomic DNA was extracted from peripheral blood using Maxwell System (Promega, WI, USA). Amplification and sequencing primers and protocols are available on request. Analysis of *AGXT* was performed at first; in case of negative result *GRHPR*, *HOGAI* and the *AGXT* promoter-region have been analyzed.

Minigene assay

In vitro analysis of the potential splice-site *HOGAI* c.341-81delT variant was performed using the pSPL3 splicing assay previously described [10,13,14].

A 449 bp genomic fragment of DNA centered on the change c.341-81delT from the patient and one from a wild type (WT) control sample were amplified and the PCR products were cloned into a TA-vector (pGEM-T Easy Vector - Promega, Madison, WI, USA) and transformed into DH5a bacterial cells (RBC Bioscience, Chung Ho City, Taiwan). Plasmids containing the WT or the variant

sequence were extracted and digested with EcoRI and NotI restriction endonuclease. Inserts were sub-cloned into a pSPL3 exon-trapping vector (Life Technologies, Paisley, UK). PSPL3 minigene constructs containing the genomic change c.341-81delT and the WT sequence were extracted from DH5a bacterial cells and transfected into both COS-7 and HEK293T cells. After 24 h, total RNA was extracted and retro-transcribed with the Cells-To-Ct kit (Life Technologies). The cDNA was amplified by standard and nested PCR and the different amplicons were separated on a TBE-agarose 1.8% gel (Online Resource, protocols available upon request).

Results

Clinical and biochemical data of the 105 patients referred to our Centre were evaluated according to recent guidelines [3] and the clinical suspicion of PH was supported by clinical findings in 96 of them. A different diagnosis was established in nine patients: four were affected by secondary hyperoxaluria, three by kidney malformations; one was diagnosed as Dent and one as Pseudo-Bartter syndrome.

Clinical, biochemical and genetic characteristics of PH1 and PH2 patients

At the end of 2014 the Italian database included 80 PH1 patients (28 females and 52 males) and one PH2 patient. Thirteen patients (16%) had consanguineous parents (information not available for 16 patients), a familiar history was reported in 30 cases (24.6%).

Symptoms at onset were mainly nephrolithiasis (52.5%) and nephrocalcinosis (35.2%). Three patients were diagnosed at family screening before symptoms developed. Median age at onset was 4 years (range 0.5-11.5), median age at diagnosis was 11 years (range 4-30).

Biochemical measurements at diagnosis were not available for 11 patients. Median Pox at diagnosis was 130 $\mu\text{mol/L}$ (24-187; normal values (nv): $<2.5 \mu\text{mol/L}$; data not available in 23 patients) and median Uox 230 $\mu\text{M/mM creat per day}$ (151-435.5; data not available in 41 patients).

Liver biopsy for AGT activity measurement was available for 35 patients: mean residual AGT activity was negligible in 27, only in 8/35 (22.9%) samples a partial AGT activity was detected (mean: $39 \pm 13\%$ of the normal).

At the end of 2014, 55 patients had ESRD and the median age was 14 years (range 4-31).

Ten patients had a kidney-only transplant and two of them had a second kidney-only transplant. In another six cases a second kidney transplant was combined with liver transplant. Median age at first kidney transplant was 18 years old (range 10-35). In 17 patients a combined kidney-liver transplant was performed and in one a sequential kidney-liver transplant. Median age at kidney-liver transplant was 13 years old (range 4-27). In two cases a second kidney transplant was performed after the combined kidney-liver. One patient underwent two combined kidney-liver transplants. In one case a preemptive liver transplant was performed.

As reported in literature, also in the Italian cohort the most common *AGXT* mutation is p.Gly170Arg (28% of the alleles) detected in 14 patients in homozygosis, in 11 in heterozygosis with a missense mutation and in four with a null mutation.

The patient affected by PH2 suffered from recurrent nephrolithiasis since 9 years old. Neither familiar history of nephrolithiasis nor parents consanguinity was referred. Uox was 77 $\mu\text{M}/\text{mM}$ creat per day at 31 years of age. Urinary L-glycerate was 4600 $\mu\text{M}/24\text{h}$ (nv <30). His renal functional was normal until the end of 2014. Molecular analysis revealed the c.295C>T, p.Arg99* mutation in *GRHPR* gene in homozygosis.

Clinical, biochemical and genetic characteristics of non-PH1 non-PH2 patients

Among the fifteen mutation-negative patients (six males), eleven (73%) had nephrolithiasis (two bilateral), five (33%) had nephrocalcinosis and one was affected with both. The median age at onset of symptoms was 5.3 years (range: 2 months-32 years). Renal function was normal in nine; three had already progressed to ESRD; data were not available for three patients. Three patients had recurrent urinary infections, four hematuria (three of them microhematuria).

Uox was available for eight patients; in children < 12 years, mean oxaluria was 190.3 mmol/mmol creatinine (range: 106-366) while in adults 78 mmol/mmol creatinine (nv < 40 mmol/mmol creatinine [12]).

Mean Pox was available in four children (7.3 $\mu\text{mol}/\text{L}$) and three adults (33.9 $\mu\text{mol}/\text{L}$) in ESRD state. Urinary calcium in patients with preserved renal function was in the normal range.

Four patients had family history of nephrolithiasis (Online Resource table 1).

Among these patients no pathogenic mutations in the three PH genes were found.

Sequencing of the *AGXT* promoter revealed two novel variants: one patient was heterozygous for the c.-647C>T variant and another patient resulted heterozygous for the c.-424C>T variant.

Neither variants were reported in dbSNP141, 1000Genomes (<http://www.1000genomes.org>, 29/07/2015), EVS (Exome variant server, 29/07/2015), EXAC (29/07/2015) and, because they do not lie in any known regulatory site, were classified as probably non pathogenic.

A third patient carried a homozygous variant in the intron 2 of *HOGAI* (c.341-81delT). This variant is not reported in EXAC Browser or EVS, but it is reported in dbSNP and 1000Genomes (<http://www.1000genomes.org> 29/07/2015) as a variant of unknown pathogenicity (rs570638580, MAF/minor allele: 0,0030/15).

In silico tools for splicing prediction [15-19] reported c.341-81delT as pathogenic, because it generates a novel acceptor splicing site (AG in c.341-79 position). Four out of five tools (SSF, MaxEntScan, Gene Splicer, HSF) predicted a possible damaging role of the variant, with an alternative transcript 79bp longer r.(340_341ins341-79_341-1). This transcript would cause a reading frameshift and a truncated protein product (premature stop codon in exon 3: p.(Thr115Glufs*43)).

Clinical characteristics of the patient homozygous for the c.341-81delT HOGAI variant

The patient homozygous for the *HOGAI* variant suffered from nephrocalcinosis since two months of life and died at four months of respiratory failure. Pox levels were normal (<1 mmol/L), while values of oxaluria/urinary creatinine were at the upper limit of the normal range (366 µmol/mmol, nv for 0-6 months 325-360 µmol/mmol creatinine). Urinary calcium level was at the upper limit (uCa/uCr 0,56 mmol/mmol, nv for children < 1 years old: < 0.4). The renal function at four month of life was decreased (GFR 53 ml/min/1,73m²). His parents were not consanguineous.

HOGAI minigene assay

In order to substantiate the predicted effect of the *HOGAI* c.341-81delT variant on splicing we set up a minigene assay by cloning the intron 2 and exon 3 WT and variant sequences into a pSPL3 vector (Fig.1). We expected a 391bp product resulting from the canonical splicing of *HOGAI* exon 3, a 471bp product from the alternative splicing generated by the deletion of T in c.341-81, and a 263bp product resulting from the complete skipping of the inserted exon. Analysis of the cDNA showed only the presence of the two 263bp and 391bp bands, without any difference from WT (Fig. 2).

Discussion

The genetic basis of PH has been widened in recent years thanks to second line studies of *AGXT* negative patients, leading to identify PH2 and PH3[1,9].

Nevertheless, a grey zone of cases in which suspicion based on clinical and biochemical findings is not confirmed by genetic testing still exists. The exact prevalence of this subset is currently unknown: several Authors reported the existence of undefined PH, but did not specify its prevalence or its clinical and genetic characteristics [1,3,20].

In our cohort of patients, 80 resulted homozygous or compound heterozygous for *AGXT* mutations (83.3%) and one homozygous for a recurrent *GRHPR* mutation (1.1%). The 15.6% of the subjects with clinical PH suspicion resulted negative for any PH mutation. A recent UK study performed in 200 patients detected mutations in *AGXT*, *GRHPR* and *HOGA1* in 50%, 8% and 8% of the patients respectively, while in 25% of the subjects no mutations were found [21]. In the USA, the analysis of 301 PH families revealed PH1 in 68.4% of them, PH2 in 9.3% and PH3 in 11.0% [20]. Compared to these results, PH1 in Italy seems to be far more frequent than the other PH types and, surprisingly, no Italian PH3 cases were identified up to now.

In two patients two novel heterozygous *AGXT*-promoter variants, c.-647C>T and c.-424C>T were identified. We considered these variants not to be pathogenic since they do not lie in any known regulatory site. The validity of this conclusion is limited by the scanty literature on the *AGXT* promoter [22] that precludes comparison with previous studies.

In one patient we detected a homozygous *HOGA1* variant in intron 2 (c.341-81delT); we could not perform genetic analysis of the *HOGA1* variant in the patient's parents, since their DNA samples were not accessible.

The c.341-81delT variant was predicted to affect splicing by *in silico* analysis and to introduce a frameshift change (p.Thr115Glufs*43) in isoform 1, disruptive of HOGA enzymatic activity. Indeed, the two minor HOGA isoforms described [23] include exons 1, 5, 6, 7 (isoform 2), and exons 1, 6, 7 (isoform 3) but not exons 2 and 3, and are probably not functional.

Even if the clinical characteristics of the c.341-81delT homozygous child were suggestive for PH3, a definite diagnosis cannot be established on a clinical basis alone. High urinary or plasma levels of dihydroxyglutarate and hydroxyoxoglutarate are indicative for PH3 [24], however the measurement of these metabolites is not yet performed in Italy.

Neither RNA nor HOGA enzymological studies were possible because further biological samples of the patient were not available. To evaluate the effect of the variant, we set up a minigene

assay that in contrast to *in silico* predictions did not reveal any difference between the product of WT and c.341-81delT variant constructs. A possible explanation is that the power of the splicing site may be weaker than the canonical splicing site, so it could not be revealed by the simple minigene protocol in COS-7 and HEK293T cells.

In the Primary Hyperoxaluria Database [25] it has been reported a *HOGAI* c.341-82delA variant of undetermined significance: this description and our finding of the delT at c.341-81 highlight the potential pathogenic effect of variations in this region, as pointed out also by *in silico* tools predictions. No functional studies are available for the c.341-82delA and we believe that this report could be a useful base for further functional studies, despite the limitations of the minigene approach.

Negative findings in our cohort of patients with a clinical diagnosis of hyperoxaluria may be ascribed to the presence of gross rearrangements (e.g. deletions/duplications), not identifiable by Sanger sequencing. However, such mutations implicate homozygosity of the SNPs haplotype and this is in contrast with our finding of heterozygous patients for several known *AGXT*, *GRHPR* and *HOGAI* SNPs (Online Resource table 2).

In our patients the presence of rare deep intronic splicing mutations in the three PH genes could not be excluded, as we did not sequence the whole intronic regions. However, such mutations are rarely searched in genetic testing and their prevalence is unknown; in literature few data regarding deep intronic mutations are reported: *AGXT* and *GRHPR* promoter analysis did not identify any causative mutations, while no Author has analyzed the whole intronic regions of the three PH genes [22,26].

In four out of 15 cases we carried out genetic analysis on the basis of suggestive clinical history alone, although neither plasma nor urinary oxalate determinations were available since the collection was difficult; in these patients, secondary causes were excluded and a genetic etiology could be hypothesized because of their early onset of nephrolithiasis/nephrocalcinosis.

These large number of undefined patients without a definitive diagnosis (11 patients) could be explained by other nephropathies or by the hypothesis of (an)other PH type(s), as postulated by several Authors [10,1,20], whose responsible gene/s is/are still unknown.

Conversely, a putatively causative homozygous or compound heterozygous genotype may not always lead to phenotypic expression, and a recent extensive population genetic screen suggested an unexpectedly high frequency of such asymptomatic/underdiagnosed cases [20].

Oxalate metabolism is complex and not fully described, it is possible that more causative or modifier genes exist [1]. As candidates, the three human lactate dehydrogenases (*LDHA*, *LDHB*,

LDHC) have been proposed, because of their important role in excessive oxalate production: LDH is able to convert glyoxylate to oxalate, glyoxylate to glycolate and hydroxypyruvate to L-glycerate, using NAD(H) as cofactor [27]. Genetic variants in these three genes could influence the total LDH activity and the oxalate accumulation in PH patients.

Another candidate involved in the pathogenesis of PH is the hepatocytes peroxisomal enzyme Glycolate Oxidase, (GO, encoded by *HAOI* gene, OMIM *605023), also known as “hydroxyacid oxidase 1”. This enzyme catalyses the oxidation of glycolate to glyoxylate, the AGT substrate. The absence of GO activity causes isolated glycolic aciduria [28], but it could be hypothesized that hypermorphic genetic *HAOI* variants could modulate the enzyme activity leading to a different production of glyoxylate, responsible of the variable phenotype in PH patients.

Finally, even a multigenic PH inheritance has been suggested [10,29], on the basis of the wide clinical variability among patients with the same *AGXT* genotype [30]. Therefore, further studies are needed to evaluate the relative contribution of genotype and environment in the development of the clinical phenotype.

To explore this hypothesis and discover novel genes involved in oxalate metabolism, whole exome sequencing and SNPs array techniques and also further phenotype-genotype studies are needed.

In conclusion, this study is the first report of *HOGAI* genetic analysis in an Italian population. Despite PH3 being frequently reported both in Europe and in USA [10,20,21], no PH3 Italian cases were so far identified. It could be possible that some PH3 cases are missed because of the clinical remission of PH3 in adult age.

In this study the analysis of *GRHPR* and *HOGAI* genes after *AGXT* analysis did not increase the sensitivity of PH genetic testing. This negative report should prompt a closer interaction between clinicians and geneticists in evaluating the clinical and biochemical data and we propose a multiple step diagnostic algorithm (Fig 3) based on clinical and biochemical data to achieve this goal.

Compliance with ethical standard

Ethical approval:

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

This article does not contain any studies with animals performed by any of the authors.

Informed consent:

Informed consent was obtained from all individual participants included in the study.

Figures and legend

Fig.1 Minigene assay: the WT or mutant fragments of the human *HOGA1* gene containing exon 3 were cloned into the pSPL3 splicing vector to construct plasmids pSPL3-*HOGA1*-Exon3-WT and pSPL3-*HOGA1*-delT-Exon3. White boxes mark the pSPL3 exons A and B (derived from the B-globin gene) and *HOGA1* exon 3, and the adjacent black lines depict the flanking intronic sequence of pSPL3. The grey box represents the 79bp insertion expected from the generation of a novel splice-site by the c.341-81delT variant. The interrupted black lines mark the intronic sequences expected to be spliced out

Fig. 2 1,8% agarose gel electrophoresis of the WT and c.341delT (delT) PCR products

Fig. 3 Proposed algorithm for the diagnosis of primary hyperoxaluria. PH: Primary hyperoxaluria, Uox: Urinary Oxalate; Pox: Plasma Oxalate

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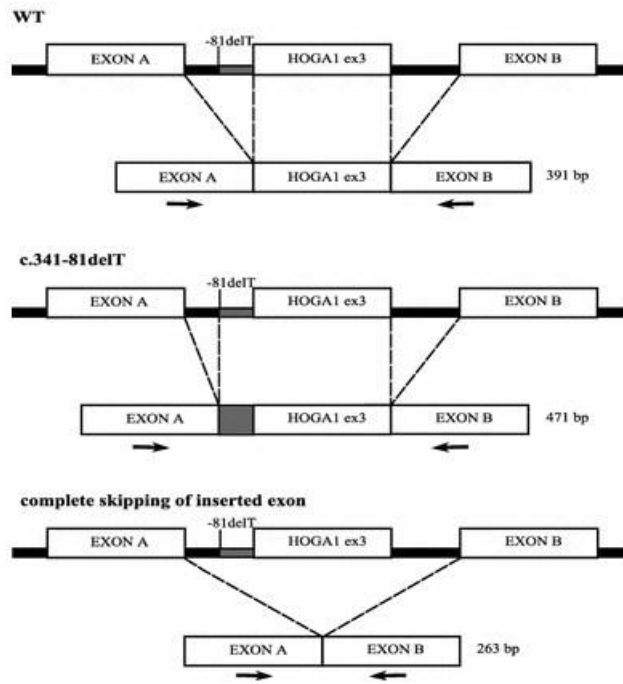


Fig. 1

Minigene assay: the WT or mutant fragments of the human *HOGA1* gene containing exon 3 were cloned into the pSPL3 splicing vector to construct plasmids pSPL3-*HOGA1*-Exon3-WT and pSPL3-*HOGA1*-delT-Exon3. *White boxes* mark the pSPL3 exons A and B (derived from the B-globin gene) and *HOGA1* exon 3, and the *adjacent black lines* depict the flanking intronic sequence of pSPL3. The *grey box* represents the 79 bp insertion expected from the generation of a novel splice-site by the c.341-81delT variant. The *interrupted black lines* mark the intronic sequences expected to be spliced out

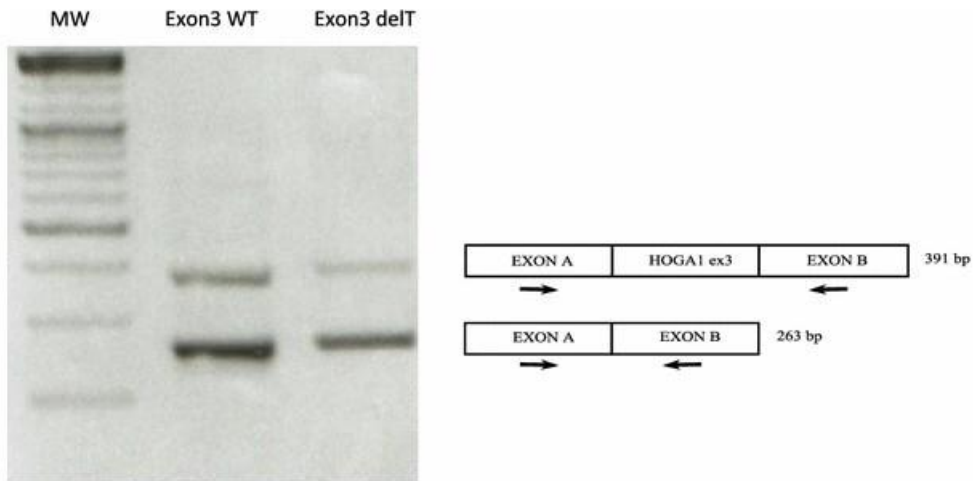


Fig. 2

1.8 % agarose gel electrophoresis of the WT and c.341delT (delT) PCR products

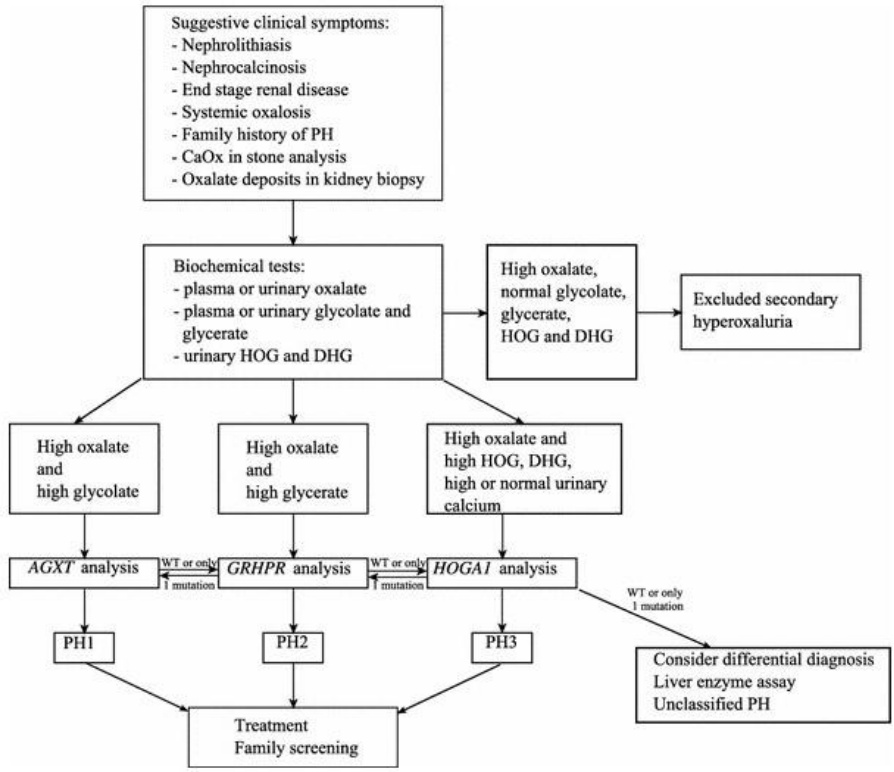


Fig. 3

Proposed algorithm for the diagnosis of primary hyperoxaluria. PH Primary hyperoxaluria, Uox urinary oxalate, Pox plasma oxalate

SUPPLEMENTARY

Minigene assay

A 449 bp genomic fragment of DNA centered on the genomic change c.341-81delT and from a WT control sample was amplified, using the following primers:

Forward 5'-ACTGGAATTCGTGGGTGGATGTGCAGGAT-3'

Reverse 5'-GGTCAGGTAAGCCTCCCAAT-3'.

Nested PCR

A nested PCR of the c-DNA WT and c-DNA variant products was performed using the following primers: Fw 5'-AGTCGACCCAGCACTGAG; Rev 3'-CCAGGTCTTGGTGTAGTGG.

Amplicons were separated on a TBE-agarose 1,8% gel and visualized after GelRed staining, the sequences were checked.

Table 1 Clinical characteristics of patients. Y: years, mo: months, nd: not detected

ID patient	Sex	Age	Age at onset	ESRD	Uox uMol/mMol creatinine	Age at Uox dosage	Pox uMol/L	Clinical characteristics
1	M	44 y	32 y	yes	nd	-	80	bilateral nephrolithiasis, vascular calcifications, familiarity for nephrolithiasis
2	F	13 y	9 y	no	70	9 y	5,6	nephrolithiasis
3	M	46 y	3 y	yes	nd	-	16	nephrolithiasis since 3 years old, mono kidney
4	M	4 y	nd	nd	nd	-	nd	nephrocalcinosis
5	F	6 y	6 mo	no	106	2 y	4	nephrolithiasis, hematuria recurrent urinary infections
6	M	deceased at 4 mo	2 mo	no	366	2 mo	nd	nephrocalcinosis
7	F	16 y	9 y	no	86	13 y	nd	nephrolithiasis, pyelonephritis
8	F	3 y	4 mo	no	115	8 mo	10	nephrolithiasis, nephrocalcinosis, microhematuria, crystalluria
9	F	5 y	2 y	no	180	3 y	nd	nephrolithiasis familiarity for nephrolithiasis
10	M	2 y	6 mo	no	205	1 y	nd	bilateral nephrolithiasis, hematuria, familiarity for nephrolithiasis
11	M	37 y	7 y	yes	nd	-	nd	nephrolithiasis
12	F	3 y	1 y	no	190	2 y	6	nephrolithiasis, hematuria
13	M	2 y	14 mo	no	170	14 mo	9	nephrocalcinosis, familiarity for nephrolithiasis
14	F	13 y	9 y	no	nd	-	nd	nephrolithiasis, left nephrectomy for pyonephrosis
15	M	1 y	2 mo	nd	nd	-	nd	nephrocalcinosis at 3 mo, recurrent urinary infections since 2 mo

Table 2 Results of genetic analysis for each patient; common SNPs founded in heterozygosis are reported in order to demonstrate the presence of both alleles for the heterozygous exons. *AGXT* haplotype: M major, m minor. Ex exon. WT: wild type. SNP: single nucleotide polymorphism. UV: unknown [pathogenicity] variant. SNPs code reference in dbSNP (<http://www.ncbi.nlm.nih.gov/SNP/>).

Patient ID	<i>AGXT</i>		<i>GRHPR</i>			<i>HOGAI</i>		
	Haplotype	Exons in homozygosis	Result	SNP in heterozygosis	Exons in homozygosis	Result	SNP in heterozygosis	Exons in homozygosis
1	Mm	ex 3, 4, 5	WT	rs10973332CA	ex 2-9	WT	rs11817730AG rs7078003CT rs2297644CT	ex 1, 3, 5, 7
2	MM	all	WT	-	all	WT	-	all
3	Mm	ex 3-5; 7-9	WT	rs35891798delG rs72275009insCT	ex 2-8	WT	-	all
4	Mm	ex 3-5; 7-9	WT	rs10973332CA rs72275009insCT	ex 2-8	WT	rs11527932GT	ex 1, 2, 3, 4, 5, 7
5	MM	ex 1, 2, 6, 10	WT	rs72275009insCT	ex 2-8	WT	-	all
6	MM	ex 1-5, 7-10	WT	rs72275009insCT	ex 1-8	UV	-	all
7	MM	ex 1-5, 7-10	WT	-	all	WT	rs11817730AG rs7078003CT rs 297644CT rs12261752AC	ex 1, 3, 5, 6
8	MM	ex 1-5, 7-10	WT	-	all	WT	rs75929214CT rs12261752CA	ex 1, 2, 3, 5, 6
9	MM	ex 2, 3, 5-10	WT	rs35891798etdelG rs76299266GA	ex 2-8	WT	rs12261752CA	ex 1, 2, 3, 5, 6
10	Mm	ex 3, 5, 7, 8, 9	WT	rs72275009insCT	ex 1-8	WT	rs75929214CT rs2297643AG rs12261752CA	ex 1, 2, 3, 6

Patient ID	<i>AGXT</i>		<i>GRHPR</i>			<i>HOGAI</i>		
	Haplotype	Exons in homozygosis	Result	SNP in heterozygosis	Exons in homozygosis	Result	SNP in heterozygosis	Exons in homozygosis
11	MM	all	WT	-	all	WT	rs2297643AG rs115279832GT rs12261752CA	ex 1, 2, 3, 4
12	MM	ex 2-3; 6-10	WT	rs35891798delG rs72275009insCT	ex 2-8	WT	-	all
13	MM	ex 1, 6, 11	WT	-	all	WT	rs75929214CT	ex 1, 2, 3, 5, 6, 7
14	MM	ex 1-4; 7-10	WT	-	all	WT	rs75929214CT rs74155508GC	ex 1, 2, 5, 6, 7
15	MM	ex 1-5; 7-10	WT	-	all	WT	rs11817730AG rs75929214CT rs7078003CT rs2297644CT	ex 1, 3, 5, 6, 7