

This is the author's manuscript



AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Organic versus conventional systems in viticulture: Comparative effects on spiders and carabids in vineyards and adjacent forests

since 2016-10-11T15:19:55Z
Open Access". Works made available rms and conditions of said license. Use blisher) if not exempted from copyright

(Article begins on next page)



UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

Questa è la versione dell'autore dell'opera:

Organic versus conventional systems in viticulture: Comparative effects on spiders and carabids in vineyards and adjacent forests

Agricultural Systems Volume 136, June 2015, Pages 61–69 - ELSEVIER doi:10.1016/j.agsy.2015.02.009

Enrico Caprio, Beatrice Nervo, Marco Isaia, Gianni Allegro, Antonio Rolando

The definitive version is available at:

La versione definitiva è disponibile alla URL: http://www.sciencedirect.com/science/article/pii/S0308521X15000311

Organic versus conventional systems in viticulture: comparative effects on

spiders and carabids in vineyards and adjacent forests

Enrico Caprio^a, Beatrice Nervo^a, Marco Isaia^a, Gianni Allegro^b, Antonio Rolando^a

^a Department of Life Sciences and Systems Biology, University of Torino, Via Accademia

Albertina 13, Torino, Italy.

^b CRA- Unità di Ricerca per le Produzioni Legnose Fuori Foresta, Strada per Frassineto 35, Casale

Monferrato (AL), Italy

*Correspondence author. E-mail: beatrice.nervo@unito.it

Tel. +39 011 6704535

Abstract

Farming systems and management regimes of vineyards may affect local biodiversity of plants and invertebrates. While most studies have focused on the overall biodiversity of vineyards, there has been little consideration of the response of different ecological guilds to vineyard management, nor to how vineyard management affects communities of adjacent semi-natural habitats.

We study here two functional guilds of carabids and five of spiders in Langa Astigiana (NW-Italy) with the following aims: *i*) to assess the comparative effects of organic and conventional farming systems, along with associated habitat and landscape variables, on species richness and abundance in vineyards; and *ii*) to compare the same within forest patches *surrounding* organic and conventional vineyards.

The different guilds exhibited distinct preferences for habitat characteristics (i.e. grass cover), landscape context and farming systems. Generalized Linear Mixed Models showed that spider preferences mostly depended upon habitat variables, while carabid preferences depended on small-scale landscape variables. In general, organic farming increased biodiversity and abundance of arthropod predators, even though different guilds of carabids and spiders responded differently. Brachypterous carabids, ambush spiders, ground-hunter spiders and other hunters preferred organic vineyards, whereas macropterous carabids, specialist spiders (mostly ant-eating spiders) and sheet web weavers selected conventional vineyards. The research we report here shows that preferences for vineyards with different farming systems has been driven by farming systems *per se* (i.e. omission of synthetic pesticides), but also by habitat characteristics and small-scale landscape structure. Arthropod diversity was greater in the forest patches adjacent to organic vineyards than to conventional ones. This suggests that organic systems may sustain a higher diversity of carabids and spiders both in vineyards and in the adjacent forest patches as well. We conclude that although conventional systems may promote the diversity of some guilds, organic systems should take priority.

Keywords

Biodiversity, carabids, spiders, organic farming, vineyards, forest patches.

1. Introduction

Agroecosystems are characterized by diverse inputs, such as human labour and petrochemical energy and products, which replace and supplement the functioning of many ecosystems. While such substitutions may buffer some of these functions, they also run the risk of damaging others. For instance, the use of pesticides may control diseases that have negative impact on crops, but these may also kill non-target organisms with other positive functions such as pollination or soil fertility enhancement (Swift and van Noordwijk, 2004; Power, 2010).

The current intensification of agriculture is leading to growing concern about the sustainability of farming systems, since farmland biodiversity has severely declined (Vickery et al., 2004; Kleijn et al., 2011). Biodiversity is certainly important to the functioning of ecosystems: insights from Biodiversity and Ecosystem Function (BEF) experiments are likely to underestimate, rather than overestimate, the importance of biodiversity to ecosystem functioning and the provision of ecosystem services (Duffy, 2009). One of the major threats to farmland biodiversity is the simplification of landscape structure, with diminution of non-crop habitat deriving from the expansion of intensive arable crops (Stoate et al., 2001; Benton et al., 2003). Organisms at higher trophic levels seem to be more vulnerable to disturbance than those at the lower trophic levels (Kruess and Tscharntke, 1994), suffering decreases both in their diversity and abundance. Disturbance affects predatory arthropods both directly and indirectly through reduced densities of their prey and hosts. This process in turn decreases the *natural* control of important crop pests (Riechert and Lawrence, 1997; Schmidt et al., 2003). Considering that many ecosystem services of particular importance for agriculture such as pollination and natural pest control often depend on the number of species in an ecosystem (Tilman et al., 2002; Cardinale et al., 2012), the impoverishment of natural communities by agriculture should be minimized to avoid negative feedbacks on production (Diaz et al., 2007).

Organic systems have been shown to support higher biodiversity than conventional ones across many different taxa (Fuller et al., 2005; Bengtsson et al., 2005). These systems aim to promote beneficial organisms by prohibiting the use of synthetic pesticides, herbicides and mineral fertilizers. Moreover, they minimize tillage in order to reduce soil erosion. Studies on organic farming in vineyards are particularly prominent because these agroecosystems are important not just for agriculture, but for conservation as well. In temperate Europe, vineyards (which typically occupy sites with particularly warm and dry climates) may host rare and endangered species of plants and invertebrates. General biodiversity is also typically high (Costello and Daane, 1998; Gliessman, 2000; Isaia et al., 2006).

Vineyards are an ancient crop of Mediterranean mountain environments, cultivated on steep slopes or terraces probably since the early middle ages (Wicherek, 1991; Aldighieri et al., 2006; Cots-Folch et al., 2006). Predicted northward shifts in the climate of European viticultural regions over the coming decades (Kenny and Shao, 1992; Maracchi et al., 2005) may alter both the spectrum and the distribution of grape varieties currently used (Schultz, 2000; Metzger et al., 2008). Several studies have shown that farming systems and regimes of vineyards are important factors determining biodiversity of plants and invertebrates (Di Giulio et al., 2001; Costello and Daane, 2003; Thomson and Hoffman, 2007; Bruggisser et al., 2010; Trivellone at al., 2012). Carabids and spiders are important components of the vineyards. They are potentially important natural agents of pest-control because of their predatory polyphagous habits, and they may be helpful to maintain ecosystem functions and services and promote sustainable agriculture (Kromp, 1999).

Vineyard landscapes of north-western Italy represent peculiar agroecosystems which deserve high conservation priority because of ecological, historical and economic importance (high quality wine production). The research we report here investigated how species richness and abundance of spiders and carabids respond to organic and conventional farming systems in the context of habitat and landscape variables. We also studied the effects of these systems on spider and carabid diversity in the forest patches surrounding the vineyards because, to our knowledge, little attention has been

addressed to study the effect of management on surrounding habitats while more consideration has been addressed to analyze how landscape context influences arthropod communities in organic and conventional farms.

Furthermore, while most studies have focused on the overall biodiversity of vineyards, less attention has addressed the effect of organic versus conventional systems on the different ecological guilds (Krauss et al., 2011). Accordingly, we considered functional guild identity of carabids and spiders instead of the overall community, since species with varying ecological requirements may respond differently to different farming systems.

2. Material and methods

2.1. Study area and sampling design

The study was carried out in the Langa Astigiana (NW Italy which ranges for about 28.000 ha), a rural region where vineyards cover 19% of the territory (5343 ha). The present landscape is the result of centuries of historically documented activities. Other main land uses include oak (*Quercus robur*), chestnut (*Castanea sativa*) and black locust (*Robinia pseudoacacia*) groves/forests (28%, 7873 ha), hazelnut orchard areas and other fruit crops (21%, 5905 ha), arable lands (16%, 4499 ha), grasslands and pastures (9.5%, 2671 ha), shrub lands (3%, 843 ha), urban areas (3%, 843 ha), and uncultivated lands (0.11%, 31 ha). The climate belongs to type Cfa (temperate, without dry season and with hot summer), in terms of Köppen-Geiger's classification (Peel et al., 2007). During the last five years, annual precipitation ranged from 567 to 894 mm with minimum values in July, January and February and with a maximum peak in April and November. Total annual rainfall averaged 757.4 mm, while the mean annual temperature was 11.9°C (Loazzolo climatic station, 600 m a.s.l.). We investigated 12 vineyards, of which 6 were certified for organic production whereby no chemical treatments except sulphur and copper sulfate spraying were used. In some cases pyrethrum was sprayed against the principal vector (*Scaphoideus titanus*) of flavescence dorèe (*Candidatus*)

Phytoplasma vitis IRPCM 2004) which is a bacterial disease of the vine. The other 6 vineyards were cultivated according to conventional production methods. These involved chemical treatments with pre- and post-emergence herbicides, insecticides (mostly against flavescence dorèe), anti-rot compounds, sulphur, copper and zinc spraying, products with esaconazol and copper oxiclorur sulphate against oidium and rots, carbamate pesticides and fungicide, and the use of mineral fertilizers with average concentration of P, K and N at 6.5 q/ha. In particular, during the study period, conventional vineyards were treated with 1.5 l/ha of chlorpyrifos-ethyl and 1.5 l/ha of chlorpyrifos-methyl against bacterial infection (flavescence dorèe) in the months of June and July respectively. Treatment against downy mildew consisted of three treatments of copper oxychloride (40%) and Dimetomorf 6% (3.5 kg/ha) in June and three treatments of Bordeaux mixture (6 kg/ha). Treatment against Oidium consisted of powdered sulphur (50 kg/ha), one treatment of Trifloxystrobin (125 g/ha), and two treatments of wettable sulphur powder (3 kg/ha) in June and two in July.

We placed five pitfall traps in the core of each vineyard and five in the last row of the vines at the edge of the vineyards. For each vineyard, we selected the closest, possibly adjacent, broad leaved forest patch (mixed black locust-oak forest in each site), where we placed five traps as well. Traps were arranged 10 m apart along line transects. Pitfall traps were 7.5 cm in diameter and 9 cm deep, filled with 150 ml of a standard mixture of wine vinegar and saturated sodium chloride solution, designed to preserve individuals. They were placed at the beginning of July 2009 and emptied three times at two-week intervals. Trapped arthropods were sorted and identified, whenever possible, to the species level using updated standard keys or specialist works. For spiders, only adults were considered. Nomenclature follows Platnick, 2014 for spiders and Vigna Taglianti, 2005 for carabids.

Three habitat variables were recorded in vineyards around each pitfall in a circular area of 5 meter radius: the percentage of grass cover, leaf litter cover (estimated by eye), and the mean grass height (ten random measurements, in centimeters). Five habitat variables were recorded in the forests close

to the vineyards around each pitfall in a circular area of 5 meter radius: the percentage of grass cover, leaf litter cover, bare ground cover and dead wood cover (estimated by eye), and the mean grass height (ten random measurements, in centimeters).

2.2. Data analysis

We used land cover data digitized from 1:10000 aerial photographs to describe the landscape composition and structure. We considered a small scale (focused on the vineyard and forest patches) and a large scale (focused on the landscape, i.e. vineyard and adjacent land uses). At the small scale, we created a buffer of 200 m of radius with the center coincident with the third trap (i.e. in the middle of the transect) of each transect. At the large scale, we created a buffer of 1500 m of radius with the center coincident with the centroid of the triangle whose vertices coincided with the third trap of each of the three transects (two in the vineyard and one in the forest patch).

Thirteen local landscape variables were measured using Geographical Information System (ESRI, 2006): the area of forests, grasslands, shrubs, vineyards, croplands, hazelnut orchards, urban and uncultivated patches, total number of patches, Shannon diversity index of patches, total mean area of patches, the distance from the closest patch of forest (in meters) and the largest patch index (LPI). LPI corresponds to the area of the largest patch (m²) of the corresponding patch type divided by total landscape area (m²), and multiplied by 100. In other words, LPI equals the percentage of the landscape comprised within the largest patch. The number of collinear variables was reduced by applying a Principal Component Analysis (PCA) with a Varimax rotation (Kaiser 1958). At large scale we considered the areas of forests, grasslands, shrubs, vineyards, croplands, hazelnut orchards, urban and uncultivated patches.

Differences in landscape and habitat between conventional and organic systems were tested using a Kruskal-Wallis test due to evidence of a non-Normal distribution.

The diversity of carabid and spider assemblages was described in terms of species richness and total abundance. Two functional guilds were considered for carabids: the macropterous and the brachypterous. We identified seven functional guilds for spiders according to the recent classification provided by Cardoso et al., 2011. Specifically, we considered: ambush hunters (namely Thomisids), ground hunters (dominated by Gnaphosids and Lycosids), sheet web weavers (mostly Agelenids), space web weavers (Theridiids), specialists (mostly Zodariids - ant-eating spiders), sensing web weavers (Atypids) and the mixed group of other hunters either runners and stalkers (Philodromids and Salticids) or small ballooners (Erigonids).

The relative contribution of vineyard systems (conventional or organic), transect location (core or edge of the vineyard), habitat variables (grass cover, grass height, leaf litter cover) and landscape variables on species richness and abundance in the vineyards were tested using generalized linear mixed models, GLMMs (Zuur et al., 2009). Vineyards (N=12) and pitfalls inside each transect (N=5) were considered as random factors. The fixed factors were represented by: farming systems (organic or conventional), transect location (core or edge of the vineyard), sampling period, habitat variables and landscape variables. Conditioning scatter plots were used to evaluate possible interactions among these variables. The significance of factor levels in the models was tested through maximum likelihood methods, and model simplification was undertaken. Akaike's information criteria (AIC) was used to test the goodness of fit of the estimated statistical models, and a model with a lower AIC was preferred to one with a higher AIC. Likelihood ratios were used for testing the explanatory power of the models and, using the drop1 function, we selected the minimum adequate model best explaining the data (Crawley, 2002). A Poisson distribution of errors was specified since variables were based on count data. All models were checked for overdispersion via the ratio between Pearson residuals of the model and the degrees of freedom. Observation level was treated as a random factor when models showed overdispersion (Elston et al., 2001).

The effects of farming systems, habitat and landscape structure *on the adjacent forest patches* were also tested on the abundance and species richness of carabids and spiders using univariate GLMMs.

The farming system, habitat and landscape variables were set as fixed factors, while the vineyards (N=12) and the pitfalls inside each transect (N=5) as random effects.

In all GLMM analyses, the pitfall was the basic sampling unit, and the number of species and the abundance of arthropods per trap was measured.

All statistical analyses were run using R package (R Core Team, 2013; Roberts, 2012).

3. Results

3.1. Assemblage composition

A total of 1541 carabids and 1204 adult spiders were collected, corresponding to 49 and 95 species respectively (Table. 1). Juveniles of spiders (261) were also collected; however, they were excluded from the analyses because they could not be identified at the species level.

In organic systems, the average number of individuals per pitfall was 3.73 ± 6.09 in vineyards and 5.62 ± 7.5 in forest patches. In conventional systems, the average number of individuals was 5.59 ± 14.69 in vineyards versus 1.33 ± 2.54 in forest patches.

Most of the arthropods were collected inside the vineyards (85% of individuals and 74% of species), because the sampling effort was twice as high in vineyards (two transects, ten pitfall traps) than in adjacent forest patches (one transect, five traps). Macropterous carabids were the most abundant guild in vineyards with 64% of sampled individuals. *Calathus fuscipes graecus* and *Brachinus crepitans* were the predominant brachypterous species, while *Harpalus dimidiatus* was the most abundant macropterous species.

Spiders were dominated by the ground hunters guild with 58% of sampled individuals, followed by specialists (14.7%), space web weavers (8.8%), ambush hunters (8.3%), other hunters (5.8%) and sheet web weavers (4.6%). Sensing web weavers were very poorly represented (only one individual found in a conventional vineyard) and were therefore discarded from analyses. *Zodarion rubidum*, an ant-eating specialist, and the ground hunter, *Haplodrassus dalmatensis*, were the predominant

spider species. The lists of carabid and spider species are given in supplementary material Appendix A and B, respectively.

3.2. Landscape and habitat characterization of vineyards

On a large scale within the 1.5 km radius buffer, landscape variables did not differ significantly between organic and conventional systems. On the contrary, on a small scale within a 200 m radius buffer, the area of vineyards was smaller (Kruskal-Wallis chi-squared = 4.20, df = 1, residual df=26, p-value = 0.04), while the area of adjacent forests (Kruskal-Wallis chi-squared = 10.17, df = 1, residual df=26, p-value = 0.001), and Shannon- Wiener diversity index (Kruskal-Wallis chi-squared = 3.84, df = 1, residual df=26, p-value = 0.05) were greater in organic than in conventional landscapes.

Since organic and conventional vineyards were located in the same contexts, as shown by landscape analysis on a large scale, only small scale variables were used to identify landscape factors affecting species richness and total abundance/trap in vineyards. The first four principal components (PC1, PC2, PC3, PC4) accounted for 81.8% of the total variation in the landscape structure matrix, with eigenvalues > 1 (Table. 2). The Shannon diversity index along with grassland, crop and shrubland areas were positively correlated with PC1 while vineyards areas and largest patch index (LPI) were negatively correlated with PC1. This shows a gradient from landscapes dominated by vineyards to more diverse and rich landscapes. PC2 was correlated negatively with woodland areas and positively with the distance from woodland. PC3 was positively correlated with patch richness, urban and uncultivated areas, and PC4 was positively correlated with hazelnut orchards.

Habitat analyses showed that grass height (Kruskal-Wallis chi-squared = 12.27, df = 1, residual df=26, p-value = 0.0005) and the percentage of leaf cover (Kruskal-Wallis chi-squared = 13.98, df = 1, residual df=26, p-value = 0.0002) were significantly higher in organic than in conventional vineyards.

3.3. Factors affecting diversity in vineyards

GLMM models regarding the richness and abundance of carabid and spider species are shown in Table 3a-3b. Sampling period was included in most of the models (with the exception of those relative to spider specialists and sheet web weavers), with total abundance and species richness higher in the first than in the second and third periods. Carabid species richness and abundance were higher in the core transect (fig. 1) and were negatively correlated with PC2, increasing therefore when forests were larger and closer to the vineyards. Spider species richness was lower in conventional vineyards, and increased according to grass cover and PC3. That is, it increased with urban and uncultivated areas and patch richness. Spider abundance responded in the same way as the species richness (fig. 2), in addition to being greater in the core transect.

Carabid functional guilds

Species richness and abundance of brachypterous species were negatively correlated with PC2, increasing therefore when forests were larger and closer to the vineyards. Also, the abundance was significantly lower in the core than in the edge transect.

The number of macropterous species was higher in the core than in the edge transects. Abundance of macropterous species was higher in conventional than organic vineyards and in core than in edge transects; it also increased with taller grass and a lower percentage of grass cover. Finally, abundance was positively correlated with PC1 and negatively correlated with PC4, meaning that it increased with larger grassland, shrubland and crop areas and smaller hazelnut areas (Table. 3a).

Spider functional guilds

Species richness of ground hunters, ambush hunters and other hunters was greater in organic than in conventional vineyards as well as the abundance of ground and other hunters.

The abundance of ambush and other hunters increased with larger grass cover. Ambush hunters showed also a significant interaction 'grass cover * farming system', suggesting a negative effect of

grass cover in conventional vineyards. Species richness of ground hunters also increased with taller grass.

Species richness and abundance of specialists (namely ant-eating spiders) were higher in conventional than in organic vineyards, while species richness and abundance of sheet web weavers were associated with grass height only, decreasing significantly with taller grasses (Table. 3b).

3.4. Differences between organic and conventional forest patches

Univariate GLMMs showed that diversity parameters of the overall carabid community (species richness and abundance of individuals), macropterous (species richness) and brachypterous carabids (species richness and abundance) were lower in the forest patches adjacent to conventional than in the patches close to organic vineyards, and their values increased along with the size of the forest patch (supplementary material Appendix C). Carabid species richness was also positively correlated with leaf litter and dead wood cover and negatively correlated with grass cover and mean grass height. Macropterous carabids were also positively correlated with bare ground and dead wood cover, shrub areas and heterogeneous landscape. Brachypterous species richness was also positively correlated with the size of the forest patch, litter and dead wood cover, and negatively correlated with grass cover and grass height.

Concerning spiders, the overall community (abundance and species richness), ambush hunters (abundance) and specialists (abundance and species richness) increased significantly in forest patches adjacent to organic vineyards compared to those adjacent to conventional vineyards (supplementary material Appendix D). The diversity parameters of the overall community (abundance and species richness) were also positively correlated with grassland area, forest patch area, heterogeneous landscape, and negatively correlated with LPI and vineyard area. Also, the abundance of spiders significantly increased with shrubland area.

Species richness of ground hunters responded positively to bare ground cover, forest patch and grassland area, while their abundance was positively correlated with bare ground, grassland and shrubland area, grass cover and Shannon patch diversity index. Abundance was also negatively correlated with the area of the vineyards, LPI and grass height. Ambush hunters (species richness and abundance) were positively correlated with bare ground, grass height, the area of shrubs and heterogeneous landscape. Sheetweb weavers (species richness and abundance) were positively correlated with grassland and shrub area and heterogeneous landscape. The diversity parameters of the specialist guild showed a positive correlation with litter and dead wood cover and a negative correlation with grassland cover, grass height and homogenous landscapes (i.e. LPI).

4. Discussion

In our study, we considered carabid and spider functional guilds to monitor the effects of two farming systems in addition to habitat characteristics and landscape context. Our approach allowed us to take into account the heterogeneity of the ecological requirements of distinct functional groups within carabid and spider assemblages (Cole et al., 2002, Clough et al., 2007, Negro et al., 2009, Batáry et al., 2012). Our results confirmed the robustness of this approach, because different guilds of carabids and spiders responded in different ways to habitat, landscape and farming systems. Considering all the species of carabids or spiders together may be misleading in two ways: the ecological preference of the dominant guild may become representative of the overall assemblage; or the ecological preferences of different groups may mask a potential trend in the community response to a possible disturbance. As a caveat, we acknowledge that, by using pitfall traps, sampling was not exhaustive for spiders, as we mainly detected ground dwelling spiders.

4.1. Habitat variables

Habitat variables appeared to have minimal influence on carabids. Only the abundance of macropterous species were linked to grass cover and grass height. On the contrary, spiders seemed

to be more dependent on habitat structure. In particular, species richness and abundance of ambush, ground and other hunters were positively linked to grass cover and/or grass height, while species richness and abundance of sheet web weavers were negatively correlated with grass height. Higher grass height and grass cover may provide protection and favorable thermal conditions for prey, which may attract a large number of spider species in turn. In particular, the preference of ambush hunters for higher grass cover accords with their hunting strategy, since they typically lie motionless in ambush for prey. Ambush hunters were mainly represented by *Xysticus kochi* (Thomisidae) whose abundance has also been shown to increase with higher litter and grass cover in other studies (Clark et al. 1994, Zrubecz et al. 2008). Ground hunters are dominated by species such as *Haplodrassus dalmatensis* and *Pardosa hortensis* belonging to the Gnaphosidae and Lycosidae families, respectively, while other hunters are mainly represented by *Thanatus arenarius* (Philodromidae). This species is known to select typically open and dry habitats. The negative correlation of sheet web weavers with grass height seems to be related to their preference to construct webs at low heights (Janetos, 1982).

4.2. Landscape structure

On a large scale, conventional and organic vineyards did not differ with respect to landscape structure variables, suggesting that they were located in the same general landscape context. Nonetheless, small scale analysis showed that *landscape structure* in organic farms differed significantly from the conventional ones: the former were characterized by smaller vineyards, larger forest areas and greater landscape heterogeneity. Moreover, organic systems favored the maintenance of bushes, trees and small forest patches. In this framework, carabids appeared to depend on landscape structure, while no guild of spiders seemed to be affected by the small scale landscape. This result seems to contrast with Isaia et al., 2006, in which landscape heterogeneity and distance from forest patches affected significantly the composition of the spider assemblage, both on the ground (pitfall trapped) and on the vines (visual standardized search).

Species richness and abundance of brachypterous carabids increased with large forest patches close to the vineyards; while abundance of macropterous carabids was linked to large grassland, shrubland and crop areas and to small hazelnut areas. Brachypterous species are mainly predators. They are medium-large body size species, either wingless or with reduced wings, and hence incapable of long movements or dispersal by flight (den Boer, 1970; Negro et al. 2009). It is sensible that they are mainly associated with less managed sites (Ribera et al., 2001). Large forests represented a potential source habitat for this functional guild. A greater proximity of the forests to vineyards allowed them to disperse with short movements and reach areas with high availability of prey. On the contrary, macropterous species are small body sized, flying, pioneer species which prefer open and disturbed areas and are able to colonize new habitats (Negro et al., 2009, Ribera et al., 2001)

4.3. Farming systems

In general, organic farming exhibited greater biodiversity and abundance of arthropod predators, allowing us to assume a better top-down control of insect pests. However, it need to be considered that generalist predators like several species of carabids and spiders may strongly reduce pest insects, but they may also act as an intraguild predator, reducing the control by other specialist predators or parasitoids (Snyder et al., 2001). The different guilds of carabids and spiders showed different preferences according to farming system.

For example, macropterous carabids were more abundant in conventional vineyards than the organic ones. On the contrary, brachypterous species richness and abundance were explained mainly by landscape context in the models instead of farming system (Table. 3a), suggesting that the main driver influencing brachypterous carabids was the small scale landscape structure surrounding the vineyards. Conventional vineyards which cover larger areas and have less ground cover were selected by macropterous species. These commonly prefer disturbed habitat (Ribera et al., 2001). Apart from differences in farming system, macropterous and brachytpterous species

showed different patterns of abundance according to its location: the former were more abundant in the core transect, while the latter in the edge transect. The vineyard cores are probably the most disturbed habitat in terms of natural vegetation development. For this reason they might be more attractive to macropterous species. Conversely, field edges may have benefitted from lower farming intensities and from edge effects from the forest patches close to the vineyards (Rand et al., 2006). Our results showed that field edges and field cores may often contain communities that vary in diversity and abundance according to functional group, with consequent provisioning of ecosystem service varying in the edge compared to the core. Brachypterous species are indeed predators, while most of phytophagous carabids belong to macropterous species.(Brandmayr et al., 2005). Moreover, the surrounding landscape matrix, and specifically the distance of forests to the vineyard edges, may act as a source for farmland brachypterous carabids in that they provide refuges and corridors for beetles dispersing between and across fields.

The effect of farming system in addition to habitat variables was particularly evident in spiders since variations in the community indices were explained in most of the models by organic versus conventional systems (Table. 3b). The influence of farming system on spider communities implies that some unmeasured factor such as pesticides may affect spiders. Omitting pesticides would both directly reduce spider mortality, and increase food availability through a reduction in the mortality of spider prey (Schmidt et al., 2005). However, the different guilds of spiders exhibited opposite preferences in relation to farming system. In particular, organic farming enhanced predators like ground, ambush and other hunters, relevant for ecosystem services. In contrast to our expectations, specialists (mostly ant eating spiders) appeared to prefer conventional vineyards. However such a trend appears unclear: considering the negative effect of conventional management on ants (Lobry de Bryuyn 1999, Dauber 2001), a positive effect on ant spiders would have been expected. On the other hand, conventional farming may favor ant nesting for two reasons: (1) the use of herbicides in conventional vineyards may determine more open soil surface that is favorable for ants, strongly

depending on high soil temperatures; (2) mechanical treatments for the weed control in organic vineyards may increase soil disturbance. Less soil disturbance in conventional vineyards because of the use of herbicides could favor the ground-nests of ants.

The different farming systems, chemical treatments and habitats did not affect ambush hunter abundance, but only species richness. This was probably due to the higher diversification of microhabitats found in organic vineyards and to the high sensitivity of spiders to pesticides (Ripper 1956, Mansour 1987, Mansour & Nentwig 1988, Pekar 1998, Fountain et al. 2007). A similar explanation can be given concerning ground hunters (both diurnal and nocturnal spiders) and for the mixed guild of other hunters (foliage dwellers and stalkers).

Ecosystem services provided by the increasing abundance and number of functional guilds in organic fields may benefit farmers due to better top-down control of pest species (Krauss et al., 2011).

The preference patterns of spiders for farming systems is strongly linked to the habitat features characterizing organic and conventional vineyards. Organic vineyards, for instance, were characterized by higher grass height and leaf cover which provide higher structural complexity and hence refuges at the soil surface, and may potentially increase the availability of herbivore prey (Zrubecz et al., 2008; Purtauf et al., 2005).

The functional guild of the specialists showed a preference for conventional vineyards. Since most of the specialists are ant-eating spiders (Zodariidae) (Pekar, 2004), we hypothesize that the conventional vineyards might have higher availability of specialist prey.

A rather surprising result of this study was that species richness and abundance of carabids and spiders were higher in forest patches adjacent to organic than in patches close to conventional vineyards, irrespective of functional guilds. It should be noted that forest patches were usually located below the vineyards. This result could be determined by a possible leaching of chemicals and fertilizers coming from conventional systems and/or smaller forest patch areas surrounding

conventional vineyards. The possible leaching of chemicals may have caused arthropod mortality and/or a decrease of food availability for predators such as spiders and carabids in forest patches adjacent to conventional vineyards. Other drivers influencing the arthropod community in the forest patches were characterized by habitat and landscape variables. In carabids, the flying macropterous are strongly influenced by landscape features such as presence of bushes and patch richness, showing the importance of the hedges for the maintenance of good disperses in the agricultural landscape (Fischer et al., 2013), which may enhance the biological pest control for adjacent agricultural crops via carabids' colonisation potential (Niemelä, 2001). Conversely, brachypterous which have a limited dispersal abilities are mainly influenced by habitat variables and by the size of forest patches (Pearce et al., 2005). However, the models ranked based on the AIC value showed that in most cases species richness in carabids was mainly influenced by the farming system while abundance of individuals responded to habitat/landscape variables. Moreover, our results showed that spiders are strongly influenced by landscape heterogeneity and in particular by the presence of grasslands (Lacasella et al., 2014).

Many studies have considered how landscape context in organic and conventional farms influences arthropod communities (Schimdt et al., 2005; Purtauf et al., 2005), but much less consideration has been devoted to evaluating the effects of farming systems on the communities of the surrounding habitats and the spillover in the managed to natural direction (Blitzer et al., 2011).

Here, we evaluated both the effect of landscape context on arthropods sampled inside the vineyards, and the effect of vineyard systems on the arthropod communities sampled outside the vineyards. The preservation of forest patches surrounding the farmland is likely to be useful for biodiversity conservation in all types of agro-ecosystems. In crop ecosystems, for instance, forest patches, field margins and grasslands are important refuges for shelter, breeding and dispersal, as well as for hibernation, especially for spring breeding carabids (Holland & Luff, 2000; Wamser et al., 2011; Jonason et al., 2013).

5. Conclusions

Vineyard landscapes of north-western Italy (Langhe, Roero and Monferrato, in Piedmont region) are included among World Heritage Sites listed by UNESCO. These areas form a spectacular expanse of rolling hills where the various combinations of climate, cultivation techniques, type of graft and grape variety determine the development of a wide range of agro-ecosystems. Our results showed that organic farming systems enhance arthropod predators belonging to several functional guilds, and influence the diversity of carabids and spiders in adjacent forest patches as well. Therefore, although conventional systems may promote the diversity of macropterous carabids and specialist spiders, we suggest organic systems should take priority. Our conclusions are also supported by several general considerations. The presence of predator carabids and spiders in crops is particularly important because the control of herbivores depends on high predator densities (Landis et al., 2000; Symondson et al., 2002; Schmidt et al., 2003). The increase, or even the mere preservation of species richness and abundance of spider and carabid predator guilds through organic farming may improve natural pest control, contributing thereby to enhanced agricultural productivity (Östman et al., 2003). Furthermore, conventional farming systems can severely reduce the economic value of some ecosystem services in agriculture (supporting and regulating services, explained in Millennium Ecosystem Assessment, 2005), whereas organic practices may enhance their value (Sandhu et al., 2010). Finally, several studies have shown that organic agriculture enhances the nutritional value of plant foods themselves, the dry matter, the minerals and antioxidant micronutrients such as phenols and salicylic acid (Brandt and Mølgaard, 2001; Lairon, 2010).

5. Acknowledgements

We wish to thank Gabriele Demichelis and Deborah Solarolo for their help in the field, and the owners of the vineyards for their collaboration, in particular Mariuccia Borio of Cascina Castlèt and

Gianni Scaglione of Forteto della Luja. We are very grateful to Frank Lad and Dan Chamberlain who checked the English and gave us useful advice and to two anonymous referees for their comments.

6. References

Aldighieri, B., Bonardi, L., Comolli, R., Conforto, A., Mariani, L., Mazzoleni, G., Rizzotti, T., 2006. La viticoltura in Valchiavenna (SO): il progetto Pianazzola. Boll. Della Soc. Geol. Ital. Vol. Spec. 17–27.

Batáry, P., Holzschuh, A., Orci, K.M., Samu, F., Tscharntke, T., 2012. Responses of plant, insect and spider biodiversity to local and landscape scale management intensity in cereal crops and grasslands. Agric. Ecosyst. Environ. 146, 130–136. doi:10.1016/j.agee.2011.10.018

Bengtsson, J., Ahnström, J., Weibull, A.C., 2005. The effects of organic agriculture on biodiversity and abundance: a meta-analysis. J. Appl. Ecol. 42, 261–269. doi:10.1111/j.1365-2664.2005.01005.x

Benton, T.G., Vickery, J.A., Wilson, J.D., 2003. Farmland biodiversity: is habitat heterogeneity the key? Trends Ecol. Evol. 18, 182–188. doi:10.1016/S0169-5347(03)00011-9

Boer, P.J.D., 1970. On the significance of dispersal power for populations of carabid-beetles (Coleoptera, Carabidae). Oecologia 4, 1–28. doi:10.1007/BF00390612

Blitzer, E.J., Dormann, C.F., Holzschuh, A., Klein, A.M., Rand, T.A., Tscharntke, T., 2012. Spillover of functionally important organisms between managed and natural habitats. Agriculture, Ecosystems & Environment, 146, 34-43

Brandmayr, P., Zetto, T., Pizzolotto, R., Casale, A., Vigna Taglianti, A., 2005. I Coleotteri Carabidi per la valutazione ambientale e la conservazione della biodiversità — Italiano, Manuale operativo. APAT, Roma.

Brandt, K., Mølgaard, J.P., 2001. Organic agriculture: does it enhance or reduce the nutritional value of plant foods? J. Sci. Food Agric. 81, 924–931. doi:10.1002/jsfa.903

Bruggisser, O.T., Schmidt-Entling, M.H., Bacher, S., 2010. Effects of vineyard management on biodiversity at three trophic levels. Biol. Conserv. 143, 1521–1528. doi:10.1016/j.biocon.2010.03.034

Cardinale, B.J., Duffy, J.E., Gonzalez, A., Hooper, D.U., Perrings, C., Venail, P., Narwani, A., Mace, G.M., Tilman, D., Wardle, D.A., Kinzig, A.P., Daily, G.C., Loreau, M., Grace, J.B., Larigauderie, A., Srivastava, D.S., Naeem, S., 2012. Biodiversity loss and its impact on humanity. Nature 486, 59–67. doi:10.1038/nature11148

Cardoso, P., Erwin, T.L., Borges, P.A.V., New, T.R., 2011. The seven impediments in invertebrate conservation and how to overcome them. Biol. Conserv. 144, 2647–2655. doi:10.1016/j.biocon.2011.07.024

Clark, M.S., Luna, J.M., Stone, N.D., Youngman, R.R., 1994. Generalist Predator Consumption of Armyworm (Lepidoptera: Noctuidae) and Effect of Predator Removal on Damage in No-Till Corn. Environ. Entomol. 23, 617–622.

Clough, Y., Kruess, A., Tscharntke, T., 2007. Organic versus conventional arable farming systems: Functional grouping helps understand staphylinid response. Agric. Ecosyst. Environ. 118, 285–290. doi:10.1016/j.agee.2006.05.028

Cole, L.J., McCracken, D.I., Dennis, P., Downie, I.S., Griffin, A.L., Foster, G.N., Murphy, K.J., Waterhouse, T., 2002. Relationships between agricultural management and ecological groups of ground beetles (Coleoptera: Carabidae) on Scottish farmland. Agric. Ecosyst. Environ. 93, 323–336. doi:10.1016/S0167-8809(01)00333-4

Costello, M., Daane, K.M., 2003. Spider and Leafhopper (Erythroneura spp.) Response to Vineyard Ground Cover. Environ. Entomol. 32, 1085–1098.

Costello, M.J., Daane, K.M., 1998. Influence of ground cover on spider populations in a table grape vineyard. Ecol. Entomol. 23, 33–40. doi:10.1046/j.1365-2311.1998.00108.x

Cots-Folch, R., Martínez-Casasnovas, J.A., Ramos, M.C., 2006. Land terracing for new vineyard plantations in the north-eastern Spanish Mediterranean region: Landscape effects of the EU Council Regulation policy for vineyards' restructuring. Agric. Ecosyst. Environ. 115, 88–96. doi:10.1016/j.agee.2005.11.030

Crawley, M.J., 2002. Statistical Computing: An Introduction to Data Analysis using S-Plus. Wiley.

Di Giulio, M., Edwards, P.J., Meister, E., 2001. Enhancing insect diversity in agricultural grasslands: the roles of management and landscape structure. J. Appl. Ecol. 38, 310–319. doi:10.1046/j.1365-2664.2001.00605.x

Dauber, J., 2001. Ant communities of an agricultural landscape: Relationships to landscape structure and land-use management, Ph.D. thesis, Justus Liebig- University of Giessen, Germany, 120 p

Díaz, S., Lavorel, S., Bello, F. de, Quétier, F., Grigulis, K., Robson, T.M., 2007. Incorporating plant functional diversity effects in ecosystem service assessments. Proc. Natl. Acad. Sci. 104, 20684–20689. doi:10.1073/pnas.0704716104

Duffy, J.E., 2008. Why biodiversity is important to the functioning of real-world ecosystems. Front. Ecol. Environ. 7, 437–444. doi:10.1890/070195

Elston, D.A., Moss, R., Boulinier, T., Arrowsmith, C., Lambin, X., 2001. Analysis of aggregation, a worked example: numbers of ticks on red grouse chicks. Parasitology 122, 563–569.

ESRI (Environmental Systems Research Institute), 2006. ArcGIS 9.2.

Fischer, C., Schlinkert, H., Ludwig, M., Holzschuh, A., Gallé, R., Tscharntke, T., Batáry, P., 2013. The impact of hedge-forest connectivity and microhabitat conditions on spider and carabid beetle assemblages in agricultural landscapes. J Insect Conserv 17, 1027–1038. doi:10.1007/s10841-013-9586-4

Fountain, M.T., Brown, V.K., Gange, A.C., Symondson, W.O.C., Murray, P.J., 2007. The effects of the insecticide chlorpyrifos on spider and Collembola communities. Pedobiologia 51, 147–158. doi:10.1016/j.pedobi.2007.03.001

Fuller, R. J., Norton, L. R., Feber, R. E., Johnson, P. J., Chamberlain, D. E., Joys, A. C., Mathews, F., Stuart, R.C., Townsend, M.C., Manley, W.J., Wolfe, M.S., Macdonald, D.W., Firbank, L. G., 2005. Benefits of organic farming to biodiversity vary among taxa. Biology Lett, 1(4), 431-434. doi: 10.1098/rsbl.2005.0357

Gliessman, S.R., 2000. Agroecosystem Sustainability: Developing Practical Strategies, 1 edition. ed. CRC Press, Boca Raton, Fla.

Holland, J.M., Luff, M.L., 2000. The Effects of Agricultural Practices on Carabidae in Temperate Agroecosystems. Integr. Pest Manag. Rev. 5, 109–129. doi:10.1023/A:1009619309424

IRPCM 2004 Phytoplasma/Spiroplasma Working Team – Phytoplasma Taxonomy Group (IRPCM). 2004.

Isaia, M., Bona, F., Badino, G., 2006. Influence of Landscape Diversity and Agricultural Practices on Spider Assemblage in Italian Vineyards of Langa Astigiana (Northwest Italy). Environ. Entomol. 35, 297–307. doi:10.1603/0046-225X-35.2.297

Janetos, A.C., 1982. Foraging tactics of two guilds of web-spinning spiders. Behav. Ecol. Sociobiol. 10, 19–27. doi:10.1007/BF00296392

Jonason, D., Smith, H.G., Bengtsson, J., Birkhofer, K., 2013. Landscape simplification promotes weed seed predation by carabid beetles (Coleoptera: Carabidae). Landsc. Ecol. 28, 487–494. doi:10.1007/s10980-013-9848-2

Kaiser, H.F., 1958. The varimax criterion for analytic rotation in factor analysis. Psychometrika 23, 187–200. doi:10.1007/BF02289233

Kenny, G.J., Shao, J., 1992. An assessment of a latitude-temperature index for predicting climate suitability for grapes in Europe. J. Hortic. Sci. 67, 239–246.

Kleijn, D., Rundlöf, M., Scheper, J., Smith, H.G., Tscharntke, T., 2011. Does conservation on farmland contribute to halting the biodiversity decline? Trends Ecol. Evol. 26, 474–481. doi:10.1016/j.tree.2011.05.009

Krauss, J., Gallenberger, I., Steffan-Dewenter, I., 2011. Decreased Functional Diversity and Biological Pest Control in Conventional Compared to Organic Crop Fields. Plos One 6, e19502. doi:10.1371/journal.pone.0019502

Kromp, B., 1999. Carabid beetles in sustainable agriculture: a review on pest control efficacy, cultivation impacts and enhancement. Agric. Ecosyst. Environ. 74, 187–228. doi:10.1016/S0167-8809(99)00037-7

Kruess, A., Tscharntke, T., 1994. Habitat Fragmentation, Species Loss, and Biological Control. Science 264, 1581–1584. doi:10.1126/science.264.5165.1581

Lacasella, F., Gratton, C., Felici, S.D., Isaia, M., Zapparoli, M., Marta, S., Sbordoni, V., 2014. Asymmetrical responses of forest and "beyond edge" arthropod communities across a forest–grassland ecotone. Biodivers Conserv 1–19. doi:10.1007/s10531-014-0825-0

Lairon, D., 2010. Nutritional quality and safety of organic food. A review. Agron. Sustain. Dev. 30, 33–41. doi:10.1051/agro/2009019

Landis, D.A., Wratten, S.D., Gurr, G.M., 2000. Habitat Management to Conserve Natural Enemies of Arthropod Pests in Agriculture. Annu. Rev. Entomol. 45, 175–201. doi:10.1146/annurev.ento.45.1.175

Lobry de Bruyn, L.A., 1999. Ants as bioindicators of soil function in rural environments. Agriculture, Ecosystems & Environment 74, 425–441. doi:10.1016/S0167-8809(99)00047-X

Mansour, F., 1987. Effect of pesticides on spiders occurring on apple and citrus in Israel. Phytoparasitica 15, 43–50. doi:10.1007/BF02980518

Mansour, F., Nentwig, W., 1988. Effects of agrochemical residues on four spider taxa: Laboratory methods for pesticide tests with web-building spiders. Phytoparasitica 16, 317–325. doi:10.1007/BF02979507

Maracchi, G., Sirotenko, O., Bindi, M., 2005. Impacts of Present and Future Climate Variability on Agriculture and Forestry in the Temperate Regions: Europe, in: Salinger, J., Sivakumar, M.V.K., Motha, R.P. (Eds.), Increasing Climate Variability and Change. Springer Netherlands, pp. 117–135.

Metzger, M.J., Schröter, D., Leemans, R., Cramer, W., 2008. A spatially explicit and quantitative vulnerability assessment of ecosystem service change in Europe. Reg. Environ. Change 8, 91–107. doi:10.1007/s10113-008-0044-x

Millennium Ecosystem Assessment, 2005. Millennium ecosystem assessment synthesis report.

Millennium Ecosystem Assessment.

Negro, M., Isaia, M., Palestrini, C., Rolando, A., 2009. The impact of forest ski-pistes on diversity of ground-dwelling arthropods and small mammals in the Alps. Biodivers. Conserv. 18, 2799–2821. doi:10.1007/s10531-009-9608-4

Niemelä, J., 2001. Carabid beetles (Coleoptera: Carabidae) and habitat fragmentation: a review. Eur J Entomol 98:127–132.

Östman, Ö., Ekbom, B., Bengtsson, J., 2003. Yield increase attributable to aphid predation by ground-living polyphagous natural enemies in spring barley in Sweden. Ecol. Econ. 45, 149–158. doi:10.1016/S0921-8009(03)00007-7

Pearce, J.L., Venier, L.A., Eccles, G., Pedlar, J., McKenney, D., 2005. Habitat islands, forest edge and spring-active invertebrate assemblages. Biodivers. Conserv. 14, 2949–2969.

Peel, M.C., Finlayson, B.L., McMahon, T.A., 2007. Updated world map of the Köppen-Geiger climate classification. Hydrol Earth Syst Sci 11, 1633–1644. doi:10.5194/hess-11-1633-2007

Pekár, S., 1998. Effect of selective insecticides on the beneficial spider community of a pear orchard in the Czech Republic. In Selden, P.A. (ed.) Proceedings of the 17th European Colloquium of Arachnology, Edinburgh.

Pekár, S., 2004. Predatory behavior of two European ant-eating spiders (Araneae, Zodariidae). J. Arachnol. 32 (1): 31-41.

Platnick, N., 2014. The World Spider Catalog, Version 14.5, American Museum of Natural History.

Power, A.G., 2010. Ecosystem services and agriculture: tradeoffs and synergies. Philos. Trans. R. Soc. B Biol. Sci. 365, 2959–2971. doi:10.1098/rstb.2010.0143

Purtauf, T., Roschewitz, I., Dauber, J., Thies, C., Tscharntke, T., Wolters, V., 2005. Landscape context of organic and conventional farms: Influences on carabid beetle diversity. Agric. Ecosyst. Environ. 108, 165–174. doi:10.1016/j.agee.2005.01.005

R Development Core Team, 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Rand, T.A., Tylianakis, J.M., Tscharntke, T., 2006. Spillover edge effects: the dispersal of agriculturally subsidized insect natural enemies into adjacent natural habitats. Ecol. Lett. 9, 603–614. doi:10.1111/j.1461-0248.2006.00911.x

Ribera, I., Dolédec, S., Downie, I.S., Foster, G.N., 2001. Effect of land disturbance and stress on species traits of ground beetle assemblages. Ecology 82, 1112–1129. doi:10.1890/0012-9658(2001)082[1112:EOLDAS]2.0.CO;2

Riechert, S.E., Lawrence, K., 1997. Test for predation effects of single versus multiple species of generalist predators: spiders and their insect prey. Entomol. Exp. Appl. 84, 147–155. doi:10.1046/j.1570-7458.1997.00209.x

Ripper, W.E., 1956. Effect of Pesticides on Balance of Arthropod Populations. Annual Review of Entomology 1, 403–438. doi:10.1146/annurev.en.01.010156.002155

Roberts, D.W., 2012. labdsv: Ordination and Multivariate Analysis for Ecology. R package version 1.5-0. http://CRAN.R-project.org/package=labdsv

Sandhu, H.S., Wratten, S.D., Cullen, R., 2010. Organic agriculture and ecosystem services. Environ. Sci. Policy 13, 1–7. doi:10.1016/j.envsci.2009.11.002

Schmidt, M.H., Lauer, A., Purtauf, T., Thies, C., Schaefer, M., Tscharntke, T., 2003. Relative importance of predators and parasitoids for cereal aphid control. Proc. R. Soc. Lond. B Biol. Sci. 270, 1905–1909. doi:10.1098/rspb.2003.2469

Schmidt, M.H., Roschewitz, I., Thies, C., Tscharntke, T., 2005. Differential effects of landscape and management on diversity and density of ground-dwelling farmland spiders. J. Appl. Ecol. 42, 281–287. doi:10.1111/j.1365-2664.2005.01014.x

Schultz, H., 2000. Climate change and viticulture: A European perspective on climatology, carbon dioxide and UV-B effects. Aust. J. Grape Wine Res. 6, 2–12. doi:10.1111/j.1755-0238.2000.tb00156.x

Snyder, W.E., Ives, A.R. Generalist Predators Disrupt Biological Control by a Specialist Parasitoid. Ecology, 82 (3), 705-716

Stoate, C., Boatman, N.D., Borralho, R.J., Carvalho, C.R., Snoo, G.R. de, Eden, P., 2001. Ecological impacts of arable intensification in Europe. J. Environ. Manage. 63, 337–365. doi:10.1006/jema.2001.0473

Swift, M.J., Izac, A.-M.N., van Noordwijk, M., 2004. Biodiversity and ecosystem services in agricultural landscapes—are we asking the right questions? Agric. Ecosyst. Environ. 104, 113–134. doi:10.1016/j.agee.2004.01.013

Symondson, W.O.C., Sunderland, K.D., Greenstone, M.H., 2002. Can Generalist Predators Be Effective Biocontrol Agents?. Annu. Rev. Entomol. 47, 561–594. doi:10.1146/annurev.ento.47.091201.145240

Thomson, L.J., Hoffmann, A.A., 2007. Effects of ground cover (straw and compost) on the abundance of natural enemies and soil macro invertebrates in vineyards. Agric. For. Entomol. 9, 173–179. doi:10.1111/j.1461-9563.2007.00322.x

Tilman, D., Cassman, K.G., Matson, P.A., Naylor, R., Polasky, S., 2002. Agricultural sustainability and intensive production practices. Nature 418, 671–677. doi:10.1038/nature01014

Trivellone, V., Paltrinieri, L.P., Jermini, M., Moretti, M., 2012. Management pressure drives leafhopper communities in vineyards in Southern Switzerland. Insect Conserv. Divers. 5, 75–85. doi:10.1111/j.1752-4598.2011.00151.x

Vickery, J.A., Bradbury, R.B., Henderson, I.G., Eaton, M.A., Grice, P.V., 2004. The role of agrienvironment schemes and farm management practices in reversing the decline of farmland birds in England. Biol. Conserv. 119, 19–39. doi:10.1016/j.biocon.2003.06.004

Vigna Taglianti, A., 2005. Checklist e corotipi delle specie di Carabidae della fauna italiana. Appendice B. In: Brandmayr P., Zetto T. & Pizzolotto R. (a cura di): I Coleotteri Carabidi per la valutazione ambientale e la conservazione della biodiversità. Manuale operativo. APAT, Manuali e linee guida, 34, 186-225.

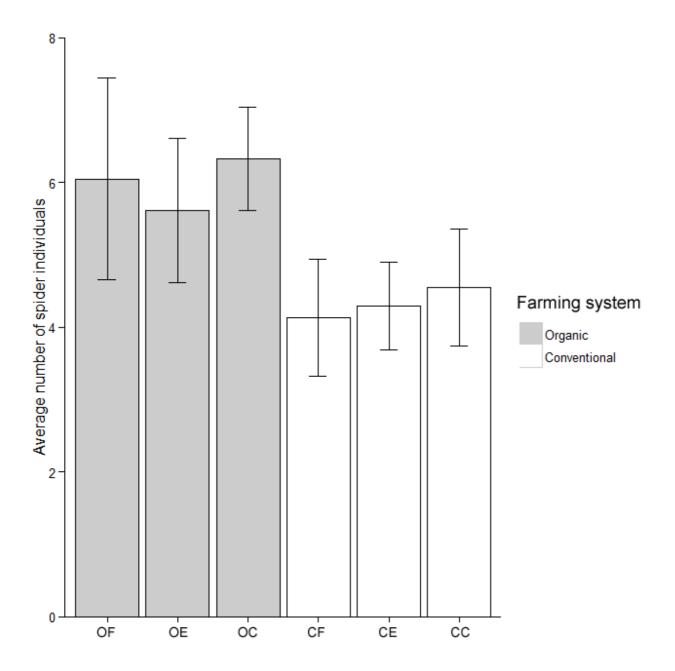
Wamser, S., Dauber, J., Birkhofer, K., Wolters, V., 2011. Delayed colonisation of arable fields by spring breeding ground beetles (Coleoptera: Carabidae) in landscapes with a high availability of hibernation sites. Agric. Ecosyst. Environ. 144, 235–240. doi:10.1016/j.agee.2011.08.019

Wicherek, S., 1991. Viticulture and soil erosion in the north of Parisian basin. Example: The mid Aisne region. Z Geomorph Nf Suppl-Bd 83, 115–126.

Zrubecz, P., Toth, F., Nagy, A., 2008. Is Xysticus kochi (Araneae: Thomisidae) an efficient indigenous biocontrol agent of Frankliniella occidentalis (Thysanoptera: Thripidae)? BioControl 53, 615–624. doi:10.1007/s10526-007-9100-6

Zuur, A., Ieno, E.N., Walker, N., Saveliev, A.A., Smith, G.M., 2009. Mixed effects models and extensions in ecology with R. Springer.

Fig. 1. Average number of individuals of carabids sampled per pitfall, in each transect. Bars stand for standard errors. OF: Forest patch transect close to organic vineyards; OE: Edges transect in organic vineyards; OC: Core transect in organic vineyards; CF: Forest patch transect close to conventional vineyards; CE: Edges transect in conventional vineyards; CC: Core transect in conventional vineyards.



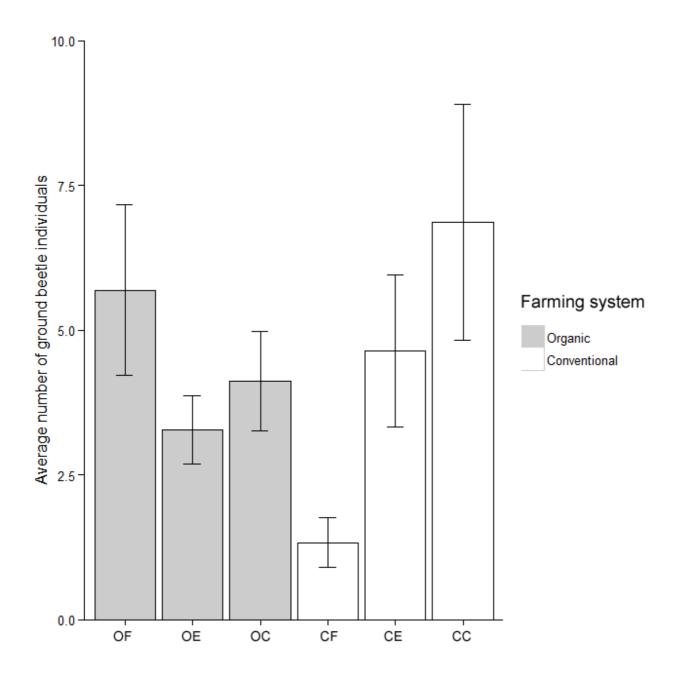


Fig. 2. Average number of individuals of spiders sampled per pitfall, in each transect. Bars stand for standard errors. OF: Forest patch transect close to organic vineyards; OE: Edges transect in organic vineyards; OC: Core transect in organic vineyards; CF: Forest patch transect close to conventional vineyards; CE: Edges transect in conventional vineyards; CC: Core transect in conventional vineyards.

Table 1. Number of individuals and number of species (in brackets) of carabid and spider functional guilds in organic and conventional vineyards, and in forest patches close to organic and conventional vineyards.

Vineyards	ORGANIC VINEYARDS	CONVENTIONAL VINEYARDS	Forest patches	CLOSE TO ORGANIC	CLOSE TO CONVENTIONAL
Carabids			Ground beetles		
Brachypterous	292 (11)	194 (11)	Brachypterous	129 (7)	34 (5)
Macropterous	194 (36)	675 (27)	Macropterous	14 (6)	9 (6)
Spiders			Spiders		
Ambush hunters	57 (4)	25 (4)	Ambush hunters	6 (4)	3 (3)
Ground hunters	363 (36)	207 (36)	Ground hunters	60 (16)	54 (21)
Other hunters	49 (12)	28 (11)	Other hunters	13 (3)	11 (7)
Space web weavers	40 (4)	47 (6)	Space web weavers	2 (2)	13 (3)
Sheet web weavers	9 (2)	17 (2)	Sheet web weavers	15 (3)	10(1)
Sensing web weavers	0	1 (1)	Sensing web weavers	0	0
Specialists	21 (2)	124 (3)	Specialists	25 (4)	4 (2)

Table 2. Results of Principal Component Analysis carried out on small scale landscape variables. The highest loadings are given in bold type.

LAND USE TYPE	PC1	PC2	PC3	PC4
Urban areas	0.351	0.502	0.644	0.014
Woodland areas	0.365	-0.849	-0.23	0.048
Uncultivated areas	0.028	-0.111	0.894	0.007
Hazelnut orchard areas	0.027	-0.06	0.028	0.969
Grassland areas	0.757	0.05	0.007	-0.05
Crops areas	0.83	0.292	-0.015	-0.213
Shrubland areas	0.735	-0.231	0.138	-0.291
Vineyard areas	-0.908	0.373	-0.01	-0.134
LPI	-0.918	0.256	-0.059	-0.169
Mean areas of patches	-0.74	-0.188	-0.203	-0.107
Patch Richness	0.699	0.013	0.564	0.051
Shannon Diversity Index	0.92	-0.167	0.279	0.118
Distance from woodland	0.053	0.848	-0.172	-0.031
Eigenvalues	5.612	2.116	1.757	1.15
Total variance %	43.166	16.281	13.514	8.847

Table 3a. GLMM results of carabid species richness and abundance, in organic and conventional vineyards. PC: principal component; SP: sampling period.

CARABIDS				
Overall community species richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	0.689	0.171	4.018	***
Transect location-Core	0.246	0.098	2.491	*
PC2	-0.201	0.082	-2.503	*
SP 2	-1.257	0.131	-9.565	***
SP 3	-0.991	0.118	-8.356	***
Overall community abundance of individuals				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	1.183	0.268	4.42	***
Transect location-Core	0.355	0.175	2.025	*
PC2	-0.316	0.129	-2.45	*
SP 2	-1.857	0.211	-8.792	***
SP 3	-1.736	0.206	-8.422	***
Brachypterous				
Species richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-0.375	0.182	-2.065	*
PC2	-0.209	0.091	-2.281	*
SP 2	-0.803	0.204	-3.944	***
SP 3	-0.772	0.202	-3.823	***
Abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-0.00142	0.324	0	0.997
PC2	-0.336	0.159	-2.11	*
Gradient-Core	-0.981	0.1743	5.627	***

SP 2	-1.359	0.266	-5.12	***
SP 3	-1.631	0.274	-3.823	***
Macropterous				
Species richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	0.115	0.215	0.54	0.591
Transect Location-Core	0.376	0.187	2.9	**
SP 2	-1.557	0.187	-8.33	***
SP 3	-1.101	0.155	-7.12	***
Abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-0.899	0.647	-1.39	0.165
System-Conventional	1.273	0.747	1.7	*
Transect Location-Core	0.549	0.229	2.39	*
Grass cover	-0.01	0.007	-1.55	**
Grass height	0.063	0.0089	7.12	***
PC1	0.421	0.148	2.84	**
PC4	-0.655	0.111	-5.88	***
SP 2	-2.461	0.29	-8.48	***
SP 3	-1.782	0.233	-7.65	***

Table 3b. GLMM results of spider species richness and abundance, in organic and conventional vineyards. PC: principal component; SP: sampling period.

SPIDERS				
Overall community specie richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z
Intercept	0.899	0.172	5.22	***
System-Conventional	-0.203	0.098	-2.07	*
Grass cover	0.008	0.002	3.52	***
PC 3	0.114	0.054	2.11	*
SP 2	-0.43	0.117	-3.66	***
SP 3	-0.187	0.094	-1.98	*
Overall community abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z
Intercept	1.5731	0.18	8.73	***
System-Conventional	-0.203	0.098	-2.07	*
Transect location-Core	0.138974	0.069813	3.52	*
Grass cover	0.007301	0.001812	2.11	**
PC3	0.067	0.012	2.373	*
SP 2	-0.68	0.166	-4.11	***
SP 3	-0.19	0.13	5.22 -2.07 3.52 2.11 -3.66 -1.98 z value 8.73 -2.07 3.52 2.11 2.373 -4.11 -1.42 z value -2.734 -3.042 -2.909 0.57 z value	NS
Ambush hunters				
Species Richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z
Intercept	-0.694	0.254	-2.734	**
System-Conventional	-0.894	0.294	-3.042	**
SP 2	-1.7675	0.607	-2.909	**
SP 3	0.15	0.279	0.57	NS
Abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z
Intercept	-1.579	0.552	-2.859	**

System-Conventional	0.451	0.673	0.67	NS
Grass cover	0.018	0.008	2.422	*
Grass cover: Systems (Conventional)	-0.025	0.01	-2.511	*
SP 2	-2.036	0.606	-3.359	***
SP 3	-0.025	0.01	-2.365	
Ground hunters				
Species Richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	0.796	0.242	3.285	**
System-Conventional	-0.449	0.113	-3.972	***
Grass height	0.027147	0.007	3.626	***
SP 2	-1.019	0.146	-6.989	***
SP 3	-1.056	0.141	-7.474	***
Abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	0.952	0.294	3.238	**
System-Conventional	-0.615	0.149	-4.126	***
Grass height	0.032	0.009	3.351	***
SP 2	-1.226	0.185	-6.624	***
SP 3	-1.206	0.174	-6.895	***
Other hunters	-1.200	0.174	-0.075	
Species Richness				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-0.537	0.26	-2.066	*
System-Conventional	-0.512	0.25	-2.000	*
SP 2	-1.02	0.3697	-2.76	**
SP 3	-0.561	0.292	-1.922	NS
Abundance	-0.501	0.272	-1.722	140
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-1.646	0.53	-3.105	**
System-Conventional	-0.626	0.317	-1.979	*
Grass cover	0.014	0.0064	2.28	*
SP 2	-1.031	0.445	-2.319	*
SP 3	-0.669	0.443	-2.319	NS
Sheet Web Weavers	-0.009	0.57	-1.01	110
Species Richness	Estimata	Ctd Emon	z voluo	D _m (> _m)
Fixed Factors	Estimate 2 20412	Std. Error	z value	Pr(> z)
Intercept	-2.29413	1.2525	-1.832 -2.284	0.067 *
Grass height	-0.06292	0.02755	-2.284	-4
Abundance	Dating	Ctd D	g1 ·	D (s. 1. 1)
Fixed Factors	Estimate -2.20571	Std. Error	z value	Pr(> z)
Intercept		1.25809	-1.753	0.0796 *
Grass height	-0.06319	0.02705	-2.336	<u>т</u>
Specialists				
Species Richness			_	
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-1.8954	0.4193	-4.521	***
System-Conventional	0.7933	0.2749	2.885	***
Abundance				
Fixed Factors	Estimate	Std. Error	z value	Pr(> z)
Intercept	-2.718	0.583	-4.663	***
System-Conventional	1.258	0.37	3.399	***

Appendix A List of carabid species collected in each transect (core, edge, forest) of organic and conventional vineyards. OF: Forest patch transect close to organic vineyards; OE: Edges transect in organic vineyards; OC: Core transect in organic vineyards; CF: Forest patch transect close to conventional vineyards; CE: Edges transect in conventional vineyards; CC: Core transect in conventional vineyards. The functional guild of each species is specified (B: Brachypterous, M: Macropterous).

Carabid species	Functional guild	OF	OE	ос	CF	CE	сс	Total N
Abax continuus Ganglbauer, 1891	В	3	0	0	0	0	0	3
Abax parallelepipedus contractus Heer, 1841	В	12	0	0	3	0	0	15
Calathus fuscipes graecus Dejean, 1831	В	52	97	176	4	36	65	430
Carabus convexus Fabricius, 1775	В	14	1	0	10	0	1	26
Carabus germarii fiorii Born, 1901	В	0	1	0	4	2	7	14
Carabus glabratus latior Born, 1895	В	14	1	0	0	0	0	15
Carabus problematicus inflatus Kraatz, 1878	В	33	1	0	0	6	0	40
Carabus solieri liguranus Breuning, 1933	В	1	0	0	13	1	0	15
Licinus cassideus (Fabricius, 1792)	В	0	1	0	0	0	0	1
Poecilus koyi viaticus (Dejean, 1828)	В	0	3	3	0	18	36	60
Pterostichus melas italicus (Dejean, 1828)	В	0	0	6	0	16	6	28

Pterostichus micans Heer, 1841	В	0	0	2	0	0	0	2
Amara aenea(De Geer, 1774)	M	1	1	1	0	0	3	6
Amara anthobia A. Villa & G.B. Villa, 1833	M	0	3	0	0	0	0	3
Amara eurynota (Panzer, 1796)	М	0	0	0	0	0	1	1
Amara lucida (Duftschmid, 1812)	M	2	3	2	0	0	0	7
Anchomenus dorsalis (Pontoppidan, 1763)	М	0	0	0	0	0	0	0
Brachinus crepitans (Linné, 1758)	M	0	1	6	1	194	272	474
Brachinus explodens Duftschmid, 1812	М	0	0	17	0	1	5	23
Brachinus sclopeta (Fabricius, 1792)	М	0	1	16	0	0	0	17
Callistus lunatus (Fabricius, 1775)	M	0	0	0	0	1	3	4
Calosoma maderae (Fabricius, 1775)	M	0	1	3	0	0	0	4
Dinodes decipiens (L. Dufour, 1820)	М	0	0	2	0	4	19	25
Harpalus affinis (Schrank, 1781)	M	1	0	0	0	0	0	1
Harpalus dimidiatus (P. Rossi, 1790)	М	0	12	31	1	14	51	109
Harpalus distinguendus (Duftschmid, 1812)	M	0	2	3	0	24	16	45
Harpalus flavicornis Dejean, 1829	M	0	3	0	0	0	5	8
Harpalus pumilus Sturm, 1818	M	0	1	0	0	0	0	1
Harpalus rubripes (Duftschmid, 1812)	М	2	12	15	0	0	0	29
Harpalus serripes (Quensel in Schönherr, 1806)	M	0	8	6	1	0	0	15
Harpalus subcylindricus Dejean, 1829	M	0	3	1	0	0	0	4
Harpalus sulphuripes Germar, 1824	M	2	1	2	0	0	0	5
Harpalus tardus (Panzer, 1797)	M	0	0	9	0	0	5	14
Leistus spinibarbis (Fabricius, 1775)	М	0	0	0	1	0	0	1
Nebria brevicollis (Fabricius, 1792)	М	0	0	0	0	3	0	3
Ophonus cribricollis (Dejean, 1829)	M	0	0	2	0	0	4	6
Ophonus melletii (Heer, 1837)	М	0	0	0	0	0	0	0
Ophonus sabulicola (Panzer, 1796)	М	0	0	0	0	1	1	2
Parophonus maculicornis (Duftschmid, 1812)	M	0	4	9	0	1	2	16
Parophonus planicollis (Dejean, 1829)	M	0	0	3	0	0	0	3
Pseudoophonus calceatus (Duftschmid, 1812)	M	0	0	0	0	0	0	0
Pseudoophonus griseus (Panzer, 1796)	М	0	1	3	4	16	14	38
Pseudoophonus rufipes (De Geer, 1774)	M	1	3	3	6	8	6	27
Scybalicus oblongiusculus (Dejean, 1829)	M	0	0	0	0	0	1	1

Appendix B: List of spider species collected in each transect (core, edge, forest) of organic and conventional vineyards. OF: Forest patch transect close to organic vineyards; OE: Edges transect in organic vineyards; OC: Core transect in organic vineyards; CF: Forest patch transect close to conventional vineyards; CE: Edges transect in conventional vineyards; CC: Core transect in conventional vineyards. The functional guild of each species is specified (AH: Ambush hunters, GH: Ground hunters, OH: Other hunters, SEW: Sensing web weavers, SHW: Sheet web weavers, SP: Specialists, SPW: Space web weavers).

Family	Spider species	Functional guild	OF	OE	ос	CF	CE	СС	Total N
AGELENIDAE	Eratigena fuesslini Pavesi, 1873	SHW	13	5	3	10	6	10	47
AGELENIDAE	Textrix denticulata (Olivier, 1789)	SHW	1	0	0	0	0	0	1
AMAUROBIIDAE	Amaurobius ferox (Walckenaer, 1830)	SHW	1	0	0	0	0	0	1
ATYPIDAE	Atypus muralis Bertkau, 1890	SEW	0	0	0	0	0	1	1
CLUBIONIDAE	Clubiona terrestris Westring, 1851	ОН	0	0	0	1	0	0	1
DICTYNIDAE	Argenna patula (Simon, 1874)	GH	0	11	9	0	2	1	23
DYSDERIDAE	Dasumia taeniifera Thorell, 1875	SP	5	3	1	3	0	5	17
DYSDERIDAE	Dysdera crocata C. L. Koch, 1838	SP	6	0	0	0	0	0	6
DYSDERIDAE	Harpactocrates apennicola Simon, 1914	SP	2	0	0	0	0	0	2
EUTICHURIDAE	Cheiracanthium mildei L. Koch, 1864	ОН	0	0	0	1	0	0	1
GNAPHOSIDAE	Callilepis schuszteri (Herman, 1879)	GH	0	0	0	0	1	0	1
GNAPHOSIDAE	Drassodes lapidosus (Walckenaer, 1802)	GH	3	6	7	2	6	4	28
GNAPHOSIDAE	Drassodes pubescens (Thorell, 1856)	GH	0	4	8	0	0	1	13
GNAPHOSIDAE	Drassyllus praeficus (L. Koch, 1866)	GH	4	3	12	0	0	0	19
GNAPHOSIDAE	Drassyllus pumilus (C. L. Koch, 1839)	GH	0	0	2	2	3	7	14
GNAPHOSIDAE	Drassyllus pusillus (C. L. Koch, 1833)	GH	1	4	1	1	2	0	9
GNAPHOSIDAE	Drassyllus villicus (Thorell, 1875)	GH	10	0	1	0	4	0	15
GNAPHOSIDAE	Gnaphosa lucifuga (Walckenaer, 1802)	GH	0	0	3	0	0	0	3
GNAPHOSIDAE	Haplodrassus dalmatensis (L. Koch, 1866)	GH	2	11	18	2	16	7	56
GNAPHOSIDAE	Haplodrassus signifer (C. L. Koch, 1839)	GH	0	6	2	1	4	5	18
GNAPHOSIDAE	Haplodrassus silvestris (Blackwall, 1833)	GH	0	0	0	0	1	0	1
GNAPHOSIDAE	Micaria albovittata (Lucas, 1846)	GH	0	8	8	0	0	0	16
GNAPHOSIDAE	Micaria coarctata (Lucas, 1846)	GH	0	2	7	1	2	0	12
GNAPHOSIDAE	Micaria formicaria (Sundevall, 1831)	GH	0	0	4	0	1	0	5
GNAPHOSIDAE	Nomisia exornata (C. L. Koch, 1839)	GH	0	0	0	1	2	0	3
GNAPHOSIDAE	Phaeocedus braccatus (L. Koch, 1866)	GH	0	0	2	0	1	1	4
GNAPHOSIDAE	Trachyzelotes fuscipes (L. Koch, 1866)	GH		0	0	1	0	0	1
GNAPHOSIDAE		GH	0	0	1	2	0	0	6
GNAPHOSIDAE	Zelotes apricorum (L. Koch, 1876 Zelotes atrocoeruleus (Simon, 1878)	GH	0	_		2	6	3	14
GNAPHOSIDAE	Zelotes longipes (L. Koch, 1866)	GH		1 0	0	0		0	
GNAPHOSIDAE	Zelotes oblongus (C. L. Koch, 1883)	GH	0	_	0	0	1 0	0	1 2
GNAPHOSIDAE	Zelotes paroculus Simon, 1914		0	0	_	_		_	
GNAPHOSIDAE		GH	1	_	0	0	0	1	2
GNAPHOSIDAE	Zelotes pygmaeus (Miller, 1943)	GH	0	0	_	0	0	1	1
GNAPHOSIDAE	Zelotes subterraneus (C. L. Koch, 1833)	GH GH	1	0	10	0	0	0	1 10
HAHNIIDAE	Zelotes tenuis (L. Koch, 1866)	GH SHW/	0	_	10		0	_	
LINYPHIIDAE	Hahnia pusilla C. L. Koch, 1841	SHW	0	0	1	0	1	0	2
	Diplocephalus alpinus (O. P Cambridge, 1872)	ОН	0	0	0	0	1	1	2
LINYPHIIDAE	Erigone dentipalpis (Wider, 1834)	ОН	0	0	1	0	0	2	3
LINYPHIIDAE	Metopobactrus nadigi Thaler, 1976	ОН	1	0	0	0	0	0	1
LINYPHIIDAE	Porrohomma microphtalmum (O. PCambridge, 1871)	ОН	0	0	1	0	0	0	1
LINYPHIIDAE	Saaristoa abnormis (Blackwall, 1841)	ОН	0	0	0	0	0	1	1
LINYPHIIDAE	Diplostyla concolor (Wider, 1834)	SHW	0	0	3	3	3	8	17

Agyneta rurestris (C. L. Koch, 1836)	SHW	0	0	1	0	0	1	2
Tenuiphantes flavipes (Blackwall, 1854)	SHW	0	0	0	0	0	2	2
Tenuiphantes tenuis (Blackwall, 1852)	SHW	0	0	0	1	1	1	3
Robertus arundineti (O. PCambridge, 1871)	SPW	0	0	0	0	1	0	1
Liocranum rupicola (Walckenaer, 1830)	GH	0	0	1	0	0	0	1
Agroeca cuprea Menge, 1873	GH	0	1	4	4	0	1	10
Scotina celans (Blackwall, 1841)	GH	2	0	0	0	0	0	2
Alopecosa aculeata (Clerck, 1757)	GH	0	2	1	0	0	0	3
Alopecosa albofasciata (Brullé, 1832)	GH	19	3	24	1	0	0	47
Alopecosa cuneata (Clerck, 1757)	GH	0	5	5	0	0	1	11
Alopecosa etrusca Lugetti & Tongiorgi, 1969	GH	0	0	0	0	0	1	1
Alopecosa mariae (Dahl, 1908)	GH	2	7	4	0	1	2	16
• • • • • • • • • • • • • • • • • • • •	GH	0	14	21	3	0	4	42
	GH	1	8	25	0	2	3	39
Aulonia albimana (Walckenaer, 1805)	GH	0	2	2	6	1	3	14
· · · · ·	GH	1	0	5	2	3	1	12
Pardosa agrestis (Westring, 1861)	GH	0	2	3	2	18	13	38
Pardosa bifasciata (C. L. Koch, 1834)	GH	0	0	0	0	0	1	1
Pardosa gr. lugubris	GH	0	1	0	0	0	0	1
Pardosa hortensis (Thorell, 1872)	GH	0	9	20	1	4	12	46
Pardosa palustris (Linnaeus, 1758)	GH	0	0	0	0	1	0	1
<i>Pardosa vlijmi</i> den Hollander & Dijkstra, 1974	GH	0	0	6	0	0	1	7
Trochosa hispanica Simon, 1870	GH	2	1	9	12	8	7	39
-	GH	0	0	0	0	1	0	1
	SP	0	0	0	0	1	0	1
Zora manicata Simon, 1878	GH	0	0	1	3	0	0	4
, i	ОН	1	1	0	0	0	0	2
Philodromus cespitum (Walckenaer, 1802)	ОН	0	0	2	0	0	0	2
·	ОН	0	16	11	2	2	1	32
Phrurolithus festivus (C. L. Koch, 1835)	GH	3	8	6	3	10	11	41
	GH	5	5	3	2	3	10	28
Aelurillus v-insignitus (Clerck, 1757)	ОН	0	1	0	0	2	0	3
Euophrys frontalis (Walckenaer, 1802)	ОН	0	2	3	2	0	0	7
Heliophanus flavipes Simon, 1900	ОН	0	2	0	0	0	0	2
Phlegra fasciata (Hahn, 1826)	ОН	0	0	0	0	1	0	1
Phylaeus chrysops (Poda, 1761)	ОН	0	0	0	0	0	1	1
Saitis barbipes (Simon, 1868)	ОН	11	1	3	1	0	0	16
Salticus scenicus (Clerck, 1757)	ОН	0	0	1	0	0	0	1
Asagena italica (Knoflach, 1996)	SPW	0	0	0	0	1	0	1
Enoplognatha thoracica (Hahn, 1833)	SPW	0	1	1	0	0	2	4
Episinus angulatus (Hahn, 1833)	SPW	1	1	0	5	3	5	15
Steatoda albomaculata (De Geer, 1778)	SPW	0	0	0	0	1	0	1
	АН	1	0	0	1	0	0	2
Cozyptila blackwalli (Simon, 1875)	All	_						
Cozyptila blackwalli (Simon, 1875) Ozyptila atomaria (Panzer, 1801)	AH	0	0	0	1	0	1	2
			0	0	1 0	0	1 1	2 1
	Tenuiphantes flavipes (Blackwall, 1854) Tenuiphantes tenuis (Blackwall, 1852) Robertus arundineti (O. PCambridge, 1871) Liocranum rupicola (Walckenaer, 1830) Agroeca cuprea Menge, 1873 Scotina celans (Blackwall, 1841) Alopecosa aculeata (Clerck, 1757) Alopecosa albofasciata (Brullé, 1832) Alopecosa cuneata (Clerck, 1757) Alopecosa etrusca Lugetti & Tongiorgi, 1969 Alopecosa mariae (Dahl, 1908) Alopecosa pulverulenta (Clerck, 1757) Arctosa personata (L. Koch, 1872) Aulonia albimana (Walckenaer, 1805) Hogna radiata (Latreille, 1817) Pardosa agrestis (Westring, 1861) Pardosa bifasciata (C. L. Koch, 1834) Pardosa fr. lugubris Pardosa hortensis (Thorell, 1872) Pardosa palustris (Linnaeus, 1758) Pardosa vlijmi den Hollander & Dijkstra, 1974 Trochosa robusta (Simon, 1870 Trochosa robusta (Simon, 1876) Ero furcata (Villers, 1789) Zora manicata Simon, 1878 Philodromus aureolus (Clerck, 1757) Philodromus cespitum (Walckenaer, 1802) Thanatus arenarius L. Koch, 1872 Phrurolithus festivus (C. L. Koch, 1835) Phrurolithus minimus (C. L. Koch, 1839) Aelurillus v-insignitus (Clerck, 1757) Euophrys frontalis (Walckenaer, 1802) Heliophanus flavipes Simon, 1900 Phlegra fasciata (Hahn, 1826) Phylaeus chrysops (Poda, 1761) Saitis barbipes (Simon, 1868) Salticus scenicus (Clerck, 1757) Asagena italica (Knoflach, 1996) Enoplognatha thoracica (Hahn, 1833) Episinus angulatus (Hahn, 1833)	Tenuiphantes flavipes (Blackwall, 1854) Tenuiphantes tenuis (Blackwall, 1852) Robertus arundineti (O. PCambridge, 1871) Liocranum rupicola (Walckenaer, 1830) Agroeca cuprea Menge, 1873 Scotina celans (Blackwall, 1841) Alopecosa aculeata (Clerck, 1757) Alopecosa albofasciata (Brullé, 1832) Alopecosa cuneata (Clerck, 1757) Alopecosa etrusca Lugetti & Tongiorgi, 1969 Alopecosa mariae (Dahl, 1908) Alopecosa pulverulenta (Clerck, 1757) Arctosa personata (L. Koch, 1872) Aulonia albimana (Walckenaer, 1805) Hogna radiata (Latreille, 1817) Pardosa agrestis (Westring, 1861) Pardosa bifasciata (C. L. Koch, 1834) Pardosa pulustris (Linnaeus, 1758) GH Pardosa vilimi den Hollander & Dijkstra, 1974 Trochosa hispanica Simon, 1870 GH Trochosa robusta (Simon, 1876) Ero furcata (Villers, 1789) Spara manicata Simon, 1878 Philodromus aureolus (Clerck, 1757) Philodromus cespitum (Walckenaer, 1802) Thanatus arenarius L. Koch, 1835) Phrurolithus festivus (C. L. Koch, 1835) Phrurolithus minimus (C. L. Koch, 1839) Aelurillus v-insignitus (Clerck, 1757) Euophrys frontalis (Walckenaer, 1802) Heliophanus flavipes Simon, 1900 Phlegra fasciata (Hahn, 1826) Phyloga tha fulcia (Knoflach, 1996) Spw Enoplognatha thoracica (Hahn, 1833) Spw Episinus angulatus (Hahn, 1833)	Tenuiphantes flavipes (Blackwall, 1854) Tenuiphantes tenuis (Blackwall, 1852) Robertus arundineti (O. PCambridge, 1871) Liocranum rupicola (Walckenaer, 1830) Agroeca cuprea Menge, 1873 Scotina celans (Blackwall, 1841) Alopecosa aculeata (Clerck, 1757) Alopecosa aubofasciata (Brullé, 1832) Alopecosa autoreata (Clerck, 1757) Alopecosa autoreata (Clerck, 1757) Alopecosa ariuse (Dahl, 1908) Alopecosa mariae (Dahl, 1908) Alopecosa pulverulenta (Clerck, 1757) Arctosa personata (L. Koch, 1872) Aulonia albimana (Walckenaer, 1805) Hogna radiata (Latreille, 1817) Pardosa agrestis (Westring, 1861) Pardosa pulveris (Thorell, 1872) GH Pardosa pulveris (Linnaeus, 1758) Pardosa pulveris (Linnaeus, 1758) Pardosa viljmi den Hollander & Dijkstra, 1974 Trochosa hispanica Simon, 1870 Trochosa robusta (Simon, 1876) Fro furcata (Villers, 1789) Zora manicata Simon, 1878 Philodromus aureolus (Clerck, 1757) Philodromus cespitum (Walckenaer, 1802) Phrurolithus festivus (C. L. Koch, 1835) Phrurolithus rinnimus (C. L. Koch, 1839) Aelurillus v-insignitus (Clerck, 1757) Euophrys frontalis (Walckenaer, 1802) Heliophanus flavipes Simon, 1900 Phlegra fasciata (Hahn, 1826) Phylaeus chrysops (Poda, 1761) Saitis barbipes (Simon, 1868) Salticus scenicus (Clerck, 1757) Asagena italica (Knoflach, 1996) Enoplognatha thoracica (Hahn, 1833) SPW 1	Tenuiphantes flavipes (Blackwall, 1854) SHW 0 0 Robertus arundineti (O. PCambridge, 1871) SPW 0 0 Liocranum rupicola (Walckenaer, 1830) GH 0 0 Agroeca cuprea Menge, 1873 GH 0 1 Scotina celans (Blackwall, 1841) GH 2 0 Alopecosa aculeata (Clerck, 1757) GH 0 2 Alopecosa albofasciata (Brullé, 1832) GH 19 3 Alopecosa detrusca Lugetti & Tongiorgi, 1969 GH 0 5 Alopecosa etrusca Lugetti & Tongiorgi, 1969 GH 0 0 Alopecosa mariae (Dahl, 1908) GH 2 7 Alopecosa pulverulenta (Clerck, 1757) GH 0 14 Arctosa personata (L. Koch, 1872) GH 1 1 Alopecosa pulverulenta (Clerck, 1757) GH 0 1 Alopecosa pulverulenta (Clerck, 1757) GH 0 1 Alopecosa pulverulenta (Clerck, 1757) GH 0 2 Aulonia albimana (Walckenaer, 1805) <	Tenuiphantes flavipes (Blackwall, 1854) SHW O O O O O O O O O	Tenuiphantes flavipes (Blackwall, 1854) SHW O O O O O Tenuiphantes tenuis (Blackwall, 1852) SHW O O O O O O Tenuiphantes tenuis (Blackwall, 1852) SHW O O O O O O D Tenuiphantes tenuis (Blackwall, 1852) SHW O O O O O O D Tenuiphantes tenuis (Blackwall, 1852) SHW O O O O O O D D Tenuiphantes tenuis (Blackwall, 1830) GH O O O O O O O O O	Tenuiphantes flavipes (Blackwall, 1854) SHW 0 0 0 0 0 1 1 1 1 1	Tenuiphantes flavipes (Blackwall, 1854) SHW O O O O O O O O C Tenuiphantes tenuis (Blackwall, 1852) SHW O O O O O O O I I I

THOMISIDAE	<i>Xysticus kochi</i> Thorell, 1872	АН	2	11	13	1	7	7	41
THOMISIDAE	Xysticus ninnii Thorell, 1872	АН	1	9	20	0	2	7	39
THOMISIDAE	Xysticus robustus (Hahn, 1832)	АН	2	0	2	0	0	0	4
TITANOECIDAE	Nurscia albomaculata (Lucas, 1846)	SPW	0	5	12	5	11	12	45
TITANOECIDAE	Titanoeca tristis L. Koch, 1872	SPW	1	2	18	3	4	7	35
TRACHELIDAE	Cetonana laticeps (Canestrini, 1868)	GH	0	0	0	0	1	0	1
ZODARIIDAE	Zodarion rubidum Simon, 1914	SP	12	4	13	1	64	54	148

Appendix C: Univariate GLMM results of carabid species richness and abundance in forest patches close to organic and conventional vineyards. PC: principal component; SP: sampling period.

CARABIDS					
Overall community species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
System-Conventional	1.70 ± 0.60	-0.055	0.019	**	194
Leaf litter cover	-0.848±0.381	0.225	0.063	***	196
Grass cover	0.90 ± 0.23	-1.275	0.359	***	196
Dead wood cover	0.858 ± 0.389	-0.0162	0.006	*	196
Grass height	-0.66 ± 0.23	0.0264	0.011	*	197
Forest patch area	-0.519 ± 0.405	0.0306	0.0133	*	197
Overall community abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Grass height	3.11±0.58	-0.090	0.020	***	374
Forest patch area	-0.71±0.61	0.310	0.110	**	402
System-Conventional	1.71±0.49	-1.720	0.660	**	403
Macropterous					
Species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
LPI	-0.49 ± 0.01	-0.014	0.004	***	92.1
Shannon patch diversity index	-2.13±0.02	0.822	0.005	***	92.1
Shrubland area	-1.54 ± 0.01	0.339	0.004	***	92.6
Dead wood cover	-1.68±0.00	0.011	0.004	*	92.7
Patch Richness	-1.78±0.00	0.087	0.005	***	92.7
System-Conventional	-1.42 ± 0.00	-0.040	0.005	***	92.9

•					
Bare ground cover	-1.47±0.00	0.012	0.005	*	92.9
Forest patch area	-1.52±0.02	0.018	0.004	***	92.9
Brachypterous					
Species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
System-Conventional	-1.08±0.46	0.039	0.018	**	161
Leaf litter cover	1.75±0.68	-0.072	0.022	**	161
Grass cover	0.66 ± 0.24	-1.484	0.422	***	164
Dead wood cover	-1.47±0.53	0.274	0.086	**	164
Grass height	-1.26±0.51	0.033	0.012	**	166
Forest patch area	0.65±0.40	-0.020	0.008	*	167
Abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
System-Conventional	3.42 ± 0.72	-0.119	0.021	***	339
Grass height	-1.49±0.73	0.4017	0.1298	**	378
Forest patch area	1.58±0.55	-2.188	0.772	**	378

Appendix D: Univariate GLMM results of spider species richness and abundance in forest patches close to organic and conventional vineyards. PC: principal component; SP: sampling period.

SPIDERS					
Overall community species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
LPI	-0.01±0.33	-0.013	0.005	**	108
System-Conventional	1.47 ± 0.26	-0.307	0.146	*	108
Grassland area	0.46 ± 0.21	0.536	0.112	***	108
Forest patch area	1.04 ± 0.33	0.074	0.036	*	108
Vineyard area	1.79 ± 0.30	-0.060	0.027	*	108
Shannon patch diversity index	0.79 ± 0.34	0.612	0.277	*	108
Overall community abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Grass height	2.56 ± 0.41	-0.028	0.007	***	392
System-Conventional	2.07 ± 0.35	-0.656	0.123	***	393
LPI	3.03 ± 0.35	-0.020	0.004	***	393
Vineyard area	2.59 ± 0.35	-0.102	0.022	***	400
Shannon patch diversity index	0.91±0.38	1.016	0.225	***	402
Forest patch area	1.30±0.43	0.123	0.032	***	406
Grassland area	0.77 ± 0.30	0.643	0.146	***	409
Shrubland area	1.74 ± 0.38	0.358	0.109	***	411
Patch Richness	1.48 ± 0.41	0.094	0.039	*	416
Ground hunters					
Species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Forest patch area	-1.47±0.53	0.274	0.086	**	164
Bare ground cover	0.14 ± 0.16	0.096	0.022	***	212

Grassland area	0.19±0.21	0.325	0.129	*	226
Leaf litter cover	0.96±0.17	-0.011	0.005	*	226
Abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Bare ground cover	0.21±0.16	0.135	0.021	***	301
Grass height	2.18±0.44	-0.048	0.014	***	332
Leaf litter cover	1.36±0.24	-0.018	0.006	**	333
Grassland area	0.39 ± 0.23	0.372	0.128	**	336
LPI	1.71±0.34	-0.001	0.000	**	336
Shannon patch diversity index	0.18±0.29	0.744	0.259	**	336
Shrubland area	0.64 ± 0.20	0.477	0.185	*	337
Grass cover	0.45±0.26	0.008	0.003	*	338
Vineyard area	0.99±0.20	-0.008	0.004	*	340
Ambush hunters	_				
Species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Bare ground cover	-2.62±0.74	0.161	0.071	*	75
Shrubland area	-2.25±0.50	1.054	0.439	*	75.8
LPI	0.02 ± 0.62	-0.003	0.001	*	76.5
Grass height	-4.95±1.81	0.095	0.040	*	76.8
Patch richness	-3.02±0.85	0.304	0.141	*	77.1
Shannon patch diversity index	-3.08 ± 0.88	1.050	0.701	*	77.2
Abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
System-Conventional	-2.00± 0.01	-0.293	0.010	***	80
Bare ground cover	-2.75±0.77	0.182	0.069	**	80.4
Shrubland area	-2.25±0.50	1.190	0.423	*	81.6
LPI	0.33±0.58	-0.003	0.001	**	82.4
Patch Richness	-3.15±0.84	0.351	0.137	*	82.7
Shannon patch diversity index	-3.23 ± 0.88	1.730	0.685	*	83.1
Sheetweb weavers					
Species richness					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Grassland area	-2.17±0.64	0.871	0.370	*	99.7
Shannon patch diversity index	-2.03 ± 059	1.093	0.494	*	102
LPI	0.14±0.48	-0.002	0.001	*	103
Shrubland area	-2.25±0.50	1.054	0.439	*	104
Abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Grassland area	-2.16±0.63	0.977	0.361	**	116
Shannon patch diversity index	-2.07± 0.46	1.290	0.460	**	118
LPI	0.49±0.43	-0.002	0.001	**	119
Patch Richness	-1.81 ± 0.52	0.223	0.090	*	120
Shrubland area	-1.15±0.29	0.674	0.277	*	121
Specialists		5.071			
Special sis					
Fixed Factors	Intoncent	Estimata	Ctd Ermor	D	AIC
FIXEU FACIOIS	Intercept	Estimate	Std Error	P	AIC

Grass cover	0.37±0.31	-0.040	0.013	**	104
System-Conventional	-0.39±0.33	-0.293	0.010	***	106
Dead wood cover	-2.70±0.68	0.071	0.020	***	106
Vineyard area	0.15±0.29	-0.127	0.050	*	107
Leaf litter cover	-2.16±0.82	0.054	0.018	**	107
Grass height	2.00 ± 0.94	-0.108	0.037	**	110
Shrubland area	-1.44±0.33	1.003	0.296	***	110
LPI	0.64 ± 0.43	-0.003	0.001	**	113
Patch Richness	-1.99±0.54	0.256	0.092	**	115
Shannon patch diversity index	-2.04±0.56	1.267	0.461	**	115
Abundance					
Fixed Factors	Intercept	Estimate	Std Error	P	AIC
Vineyard area	0.88 ± 0.23	-0.200	0.046	***	146
Shrubland area	-1.44±0.33	1.424	0.276	***	149
LPI	1.62±0.35	-0.003	0.001	***	153
Patch Richness	-2.42±0.54	0.404	0.085	**	156
Shannon patch diversity index	-2.52±0.56	2.011	0.429	**	156
Grass cover	0.43 ± 0.34	-0.030	0.009	**	160
System-Conventional	-0.18±0.54	-1.900	0.572	***	161
Dead wood cover	-1.97±0.52	0.056	0.016	***	161
Leaf litter cover	-2.0±0.60	0.038	0.013	**	164
			•		•
Grass height	1.38 ± 0.84	-0.076	0.029	**	164