

BIODEGRADABLE AND COMPOSTABLE FILM AND MODIFIED ATMOSPHERE PACKAGING IN POSTHARVEST SUPPLY CHAIN OF RASPBERRY FRUITS (CV. GRANDEUR)

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ABSTRACT

In this study, noncommercial biodegradable and compostable nonperforated films (F2, F3, F4) were evaluated for modified atmosphere packaging storage for 5 days at 1 ± 1 C and 7 days at 20 ± 1 C of raspberries cv. Grandeur. After measuring the CO₂ (PeCO₂) and the O₂ (PeO₂) permeability of new films at different temperatures, the most important qualitative traits and nutraceutical components of fruits were analyzed and compared with those of unwrapped raspberries and macroperforated film (F1). As the temperature increased, the F2 and F3 were the only films to allow storage of the fruits up to 12 days, but the F3 film (5.7 kPa O₂ and values of 31.6 kPa CO₂) was the best for maintaining the color parameter (*L* 29.0, chroma 36.4) close to the value observed at harvest because of a similar ratio between the PeCO₂ and the PeO₂ (4.2 and 4.1, respectively) at both low and high temperatures.

PRACTICAL APPLICATIONS

This study of the permeability of new films from renewable sources for packaging raspberries in a temperature range that can simulate the postharvest supply chain could indicate that storing these perishable fruits under modified atmospheric packaging could increase environmental sustainability.

INTRODUCTION

The raspberry industry has recently undergone significant changes due to increasing customer quality requirements, health and lifestyle (sustainable consumption) concerns, and the need to stock fresh raspberries in supermarkets year-round (Paraušić *et al.* 2010; Girgenti *et al.* 2013). Such changes require producers and traders to develop new strategies to reduce soft fruit losses and improve shelf life, and access to new varieties and innovation in packaging technology could play a key role in improving the postharvest quality during storage. Genetically improved varieties have led to the cv. Grandeur, a new everbearing variety of red raspberry (*Rubus idaeus* L.) that is of particular interest to the European market because of its good resistance to climate stresses, long picking period, large size, high pulp consistency and the bright color of the epidermis (Ackerman and Adams 2009).

Because raspberries provide an important source of chemical compounds that are essential for human health, their marketing must integrate the themes of sustainable production and distribution processes (Girgenti *et al.* 2013). Supply chain sustainability must consider packaging management, particularly in sectors where packaging is integral to handling and transportation and where cool temperatures are not guaranteed. The use of materials derived from renewable sources could provide a good opportunity for the development of the postharvest supply chain.

Raspberry fruits stored at 0–0.5C and 90–95% relative humidity (RH) can be maintained in a normal atmosphere (NA) for 5–7 days (Salunkhe and Desai 1984), although high CO₂ treatments and controlled atmospheres (15–20% CO₂ and 5–10% O₂) have also been studied (Callesen and Møller Holm 1989; Agar and Streif 1996; Kader 2001) for improving the shelf life of berries. Modified atmosphere packaging (MAP) can supplement proper temperature

management and translate into reduced qualitative and quantitative losses of soft fruits. The modified atmosphere extends the shelf life of berries, and the sealed container protects them from exposure to disease and other environmental contaminants. MAP techniques involve the use of plastic films that limit gas diffusion, leading to CO₂ enrichment and O₂ reduction (Waghmare and Annapure 2013). When raspberries are stored in gaseous mixtures containing 10% O₂ and 15% CO₂, decay is significantly reduced, and the berries have more attractive color compared with berries stored under NA conditions (Haffner *et al.* 2002). The final gaseous composition depends on a series of factors such as the weight of the product packed, the storage temperature, the commodity respiration rate, the cultivar and the ripening stage. In addition, the exchange of gases between the atmosphere in the container and the exterior is affected by concentration differences both inside and outside the package, the exposed surface and the permeability of the selected film. Several films with various permeability values for water vapor, CO₂ and O₂ for packaging fruits and vegetables under modified atmosphere conditions are commercially available (Linke and Geyer 2013), but the gas permeability values of most plastic films are too low to allow gas exchange and permit slow respiration (Guillaume *et al.* 2010). When temperatures change during shipping, handling or retail display, a MAP storage system could cause O₂ depletion and CO₂ accumulation due to an increase in respiration that exceeds the increase in the permeability of the film (Exama *et al.* 1993; Jeong *et al.* 2013). Thus, the permeability of the packaging film or perforation must change at the same rate as the respiration over the temperature range of interest (Talasila *et al.* 1995). The fluctuating temperatures encountered in the postharvest supply chain can negatively influence MAP storage because of the development of high humidity inside the package, which promotes the development of decay and blocks O₂ diffusion into fruit tissues and through the film (Cameron *et al.* 1995; Brecht *et al.* 2003). Further, CO₂ levels greater than 20% can cause discoloration, softening and an off-flavor in raspberries (Agar and Streif 1996).

Packaging in the soft fruit sector is changing, and the reduction of packaging weights and the use of sustainable materials are essential to respond to the desires of retailers and customers. Alternative packaging materials that are “eco-friendly,” biodegradable, and made from natural resources and can be used in place of petroleum-based polymers such as polyethylene terephthalate and high- and low-density polyethylene are being developed to package fresh and cut produce (Peelman *et al.* 2013). Several studies have shown interest in the use of these alternative materials for postharvest storage (Marsh and Bugusu 2007; Joo *et al.* 2011; Peelman *et al.* 2013). For example, the shelf life of strawberries cv. Camarosa was improved by including an

oxygen absorber in bio-based packages (Aday and Caner 2013). A biodegradable laminate was found to be suitable as a MAP material in the inert temperature range for fresh products, such as shredded lettuce and cabbage, head lettuce, cut broccoli, whole broccoli, tomatoes, sweet corn and blueberries (Makino and Hirata 1997). Seglina *et al.* (2009) examined the possibility of extending the shelf life of the raspberry cultivar Polana by packaging the fruits in different materials. The authors observed that the samples stored in MAP with a polylactic acid (PLA) film maintained the best headspace gas composition. Research on starch-based films has shown that such films could be suitable alternatives to conventional plastics for different food products (Peelman *et al.* 2013), but data on the use of these materials to store highly perishable raspberry fruits under passive MAP conditions are limited (Peano *et al.* 2013).

The objectives of this study were as follows: (1) to evaluate the performance in terms of gas transfer of the noncommercial biodegradable and compostable films under different temperature conditions; (2) to evaluate the capacity of the new films to manage a passive MAP to store a new everbearing variety of red raspberry, the cv. Grandeur, for up to 12 days at a cool storage temperature (1 ± 1°C) and at the most common temperature at European retail points (20 ± 1°C); and (3) to evaluate the impact of the resulting gas conditions on the quality and nutraceutical compounds of raspberry fruits at various storage times.

MATERIALS AND METHODS

Plant Material

Red raspberry (*R. idaeus* L.) cv. Grandeur fruits were obtained from a commercial orchard of the Agrifrutta Soc. Coop. SRL (Piedmont, Italy). This cultivar is a new everbearing variety that is characterized by a large fruit size, high fruit firmness, a light red color, conical shape, even color and good flavor (Ackerman and Adams 2009). The fruits were hand-picked in the middle of September at the red-ripe stage of maturity. The fruits were graded for the uniformity of color and size, and damaged berries were removed. The fruits were individually packed in PLA trays and transported to the packing house (Peveragno, Cuneo, Italy) in less than an hour. The different storage treatments were started approximately 3 h after harvest. The raspberries were packed in rigid PLA trays, each containing 0.125 kg of fruit. Each tray (size 9.5 × 14 × 5 cm; consumer unit) was hermetically sealed with a flowpack machine using one commercial polypropylene macroperforated film (6-mm holes) that is actually used in the retailer distribution (Trepack, Siena, Italy) (F1) and three noncommercial biodegradable, nonperforated and compostable films

TABLE 1. MAIN MECHANICAL AND CHEMICAL CHARACTERISTICS OF THE NONCOMMERCIAL BIODEGRADABLE AND COMPOSTABLE FILMS

Film*	Thickness (μm)	Tensile strength at break (S_b) (MPa)	Elongation at break (e_b) (%)	Young's modulus (E)† (MPa)	Haze‡ (%)
F2	25	50	325	1,900	15
F3	15	55	250	2,100	5
F4	25	45	350	1,750	20

* Prototypes films by Novamont.

† American Society for Testing for Materials ASTM-D882-02 (ASTM 2000a).

‡ American Society for Testing for Materials ASTM-D1003-00 (ASTM 2000b).

(prototypes, Novamont, Novara, Italy) (F2, F3 and F4). Table 1 contains the main mechanical and chemical properties for the noncommercial films obtained from the provider.

Permeation Tests on the Biodegradable and Compostable Films

The Multiperm Oxygen and Carbon Dioxide Analyzer (Extra Solution s.r.l., Pisa, Italy) was used to measure the oxygen transmission rate (ASTM 2008) and the carbon dioxide transmission rate (ASTM 2005) values ($\text{cm}^3/\text{m}^2/\text{day}/\text{bar}$) of the three noncommercial biodegradable and compostable films. Permeability tests were conducted at 38, 30, 23, 20, 15 and 10°C and at 90% RH instead of 1°C, the temperature at which raspberry fruits were stored for up to 5 days because it was not possible to reach temperatures as low as 1°C with the apparatus used to measure the permeability of the films. The system consisted of two chambers, with the test film ($S = 50 \text{ cm}^2$) hermetically separating the two chambers. Gases at atmospheric pressure flowed continuously through the upper chamber. Nitrogen was used as the sweep gas in the lower chamber. The flow rate of the permeation gas was 73 mL/min. At steady state, the permeated gases in the sweep gas stream were analyzed to obtain the gas transmission rates. The pressure from the instrument was given in bar units. To obtain the data value in kPa, the primary SI unit, it is necessary to use the following conversion factor: 1 bar = 100 kPa, according to NIST Special Publication 811 (National Institute of Standards and Technology (NIST) 2008).

Gas permeability ($\text{mmol}\cdot\text{cm}/\text{cm}^2/\text{h}/\text{kPa}$) was calculated according to Eq. (1) based on Fick's first law of diffusion for thin and infinite films (Crank and Park 1968):

$$J = \frac{Pe \times S}{e} \times \Delta P \quad (1)$$

To determine the gas barrier properties of the films under real working conditions, the equation from an Arrhenius plot of the data was used (Beaudry *et al.* 1992; Exama *et al.* 1993).

Packaging Procedure and Postharvest Storage Conditions

A set of three trays of 0.375 kg for each time point was left unpacked and used as the control. For the flowpack equipment, a Taurus 700 (Delphin, Italy) electronic horizontal wrapping machine including a take-up reel with translational movement of the clamping jaws was used. All of the packages were sealed under ordinary atmospheric conditions (0.2 kPa CO_2 and 21.2 kPa O_2).

The fruits were stored at $1 \pm 1^\circ\text{C}$ at 90–95% RH in a cold room for 5 days. After cool storage, the fruits were removed and held in the laboratory for 7 additional days at $20 \pm 1^\circ\text{C}$ to simulate retailer conditions.

Sampling Procedures

All analyses, with the exception of the headspace gas composition, were performed for each sample at five time points: at harvest (0); after 3 and 5 days at the constant temperature of $1 \pm 1^\circ\text{C}$; at 24 h from the change of the storage temperature at 6 days ($20 \pm 1^\circ\text{C}$); and at the end of the storage period (12 days: 5 days at low temperature + 7 days at high temperature). Three randomly selected trays (0.375 kg of raspberry fruits) were used for each package.

Headspace Gas Composition

The headspace gas composition inside each package changed during storage due to the combined effect of the respiration of the cv. Grandeur, the films acting as a barrier to gases, and the temperature. Therefore, to measure the relative changes of the carbon dioxide and oxygen concentrations, we used a CO_2 and O_2 analyzer (CheckPoint II, PBI Dansensor, Milan, Italy). The changes in gas composition values were measured daily over the trial period and are expressed as v/v kPa. To avoid modifications in the headspace gas composition due to gas sampling, the same air volume (free volume 330 mL) was maintained in the packages during the trial period (due to a modification to the made by the supplier) because the analyzer introduced

the same quantity of air that it removed for the analyses. To prevent gas leakage during the measurement, an adhesive single septum system (Septum white 15-mm diameter, PBI Dansensor) was placed on the surface of the package. The results are expressed as an average of three replicates.

Fruit Quality Assessment

Weight Loss. The weight (water) loss of each raspberry tray was measured using an electronic balance (SE622, WVR, Radnor, PA) with an accuracy of 0.01 g. The weight of each tray was recorded at harvest and at the end of each storage period. The weight losses are reported as a percentage of the initial fruit weight of each package. The results are expressed as an average of three replicates.

Total Soluble Solids (TSS) and Titratable Acidity (TA). TSS analysis was conducted using squeezed raspberries at 20°C. The TSS concentration was determined through the homogenization of five individual fruits from each lot with an Atago Pal-1 pocket refractometer (Atago Co. Ltd., Tokyo, Japan) and is expressed in units of °Brix at 20°C (Aday and Caner 2011). The TA was measured using an automatic titrator (Titritino 702, Metrohm, Herisau, Switzerland) and was determined potentiometrically using 0.1 N NaOH to an end point of pH 8.1 in 5 mL of juice diluted in 50 mL of distilled water.

Color. Color was measured on the first 15 sound, nonmoldy fruits from each basket (three baskets were randomly chosen for each package). The mean of the 30 fruit measurements was used for data analysis. Color was measured on the side of a slightly flattened whole fruit using a tristimulus color analyzer (Chroma Meter, model CR-400, Minolta, Langenhagen, Germany) equipped with a measuring head with an 8-mm-diameter measuring area. The analyzer was calibrated to a standard white reflective plate and used Commission Internationale d'Eclairage (CIE) Illuminant C.

CIELAB or $L^*a^*b^*$ space was used to describe the color. This color space is device-independent and able to create consistent colors regardless of the device used to acquire the image. L^* is the luminance or lightness component, which ranges from 0 to 100, whereas a^* (green to red) and b^* (blue to yellow) are two chromatic components, with values varying from -120 to +120 (Yam and Papakadis 2004). These values were used to calculate chroma, which indicates the intensity or color saturation, using the following equation:

$$C^* = [a^{*2} + b^{*2}]^{1/2} \quad (2)$$

Hue angle was calculated as follows:

$$h^\circ = \arctangent[b^*/a^*] \quad (3)$$

where 0° = red-purple, 90° = yellow, 180° = bluish-green and 270° = blue (McGuire 1992).

Total Anthocyanin, Phenolic Content and Antioxidant Activity. To determine the total anthocyanin content, the total phenolic content and the total antioxidant capacity, an extract of berries was obtained using 10 g of fruit added to 25 mL of extraction buffer (500 mL of methanol, 23.8 mL of de-ionized water and 1.4 mL of 37% hydrochloric acid). After 1 h in the dark at room temperature, the samples were thoroughly homogenized for a few minutes with an ULTRA TURRAX (IKA, Staufen, Germany) and centrifuged for 15 min at 3,019 g.

The supernatant obtained by centrifugation was collected and transferred into glass test tubes and stored at -20°C until analysis.

The total anthocyanin content was quantified according to the pH differential method of Cheng and Breen (1991). Anthocyanins were estimated by the difference in absorbance at 510 and 700 nm in a buffer at pH 1.0 and pH 4.5, where $A_{\text{tot}} = (A_{515} - A_{700})_{\text{pH 1.0}} - (A_{515} - A_{700})_{\text{pH 4.5}}$. The results are expressed as milligrams of cyanidin-3-glucoside (C3G) equivalents per 100 g of fresh weight (FW). The total phenolic content was measured using Folin-Ciocalteu reagent with gallic acid as a standard at 765 nm following the method of Slinkard and Singleton (1977). The results are expressed as milligrams of gallic acid equivalents (GAE) per 100 g of FW. Antioxidant activity was determined using the ferric reducing antioxidant power (FRAP) assay following the methods of Pellegrini *et al.* (2003), with some modifications.

The antioxidant capacity of the dilute berry extract was determined by its ability to reduce ferric iron to ferrous iron in a solution of 2,4,6-tripyridyl-s-triazine (TPTZ) prepared in sodium acetate at pH 3.6. The reduction of iron in the TPTZ-ferric chloride solution (FRAP reagent) results in the formation of a blue-colored product (ferrous tripyridyl-triazine complex), the absorbance of which was read spectrophotometrically at 595 nm 4 min after the addition of appropriately diluted berry extracts or antioxidant standards to the FRAP reagent. The results are expressed as mmol Fe^{2+} per 1 kg of fresh berries. All of these analyses were performed using the UV-vis spectrophotometer 1600 PC VWR International.

Statistical Analysis

For the qualitative analysis and the nutraceutical compounds, a bifactorial model (time of storage \times film) analysis of variance (ANOVA) was applied, and significant

TABLE 2. VALUES OF THE OXYGEN TRANSMISSION RATE (O₂TR) AND CARBON DIOXIDE TRANSMISSION RATE (CO₂TR) OF NONCOMMERCIAL BIODEGRADABLE AND COMPOSTABLE FILMS

Film	Temperature (C)	O ₂ TR (ASTMF2622-08) (cm ³ /m ² /day/bar)	CO ₂ TR (ASTMF2476-05) (cm ³ /m ² /day/bar)
F2	38	1,312	4,164
	30	1,018	3,265
	23	798	2,581
	20	729	2,544
	15	622	2,226
	10	518	1,913
F3	38	2,192	7,874
	30	1,637	6,221
	23	1,316	5,135
	20	1,197	4,748
	15	1,031	4,086
	10	897	3,545
F4	38	1,559	5,355
	30	1,185	4,255
	23	956	3,576
	20	865	3,272
	15	732	2,875
	10	615	2,492

differences were calculated using Tukey's test. When the interactions were significant, the mean values were compared by a least significant difference multiple range test, with $P < 0.05$ considered significant. SPSS Statistics 20 statistical package software (SPSS Statistics 20, 2013, IBM, Milan, Italy) for Windows was used.

RESULTS AND DISCUSSION

Gas Barrier Properties of the Biodegradable and Compostable Films

The measured oxygen transmission rate (O₂TR) and the carbon dioxide transmission rate (CO₂TR) for the tested films are listed in Table 2. The changes in the barrier properties of the biodegradable and compostable films for both

gases are expressed as a function of temperature, as reported by Beaudry *et al.* (1992). The Arrhenius plot of the data measured indicated that the natural log of the permeability coefficient for both gases depended linearly on the reciprocal temperature (K):

$$\ln Pe = \ln A + (E_a/RT) \quad (4)$$

where Pe is the O₂ or CO₂ permeability (mmol·cm/cm²/h/kPa), A is the Arrhenius constant (mmol·cm/cm²/h/kPa), E_a is the activation energy of O₂ or CO₂ permeation (kJ/mol), R is the universal gas constant (0.00831448 kJ/mol/K) and T is the temperature (K).

In Figs. 1 and 2, the oxygen (O₂) and carbon dioxide (CO₂) permeability values for all of the noncommercial films used for packages (F2, F3 and F4) correlated well with the Arrhenius equation. All of the innovative films showed a

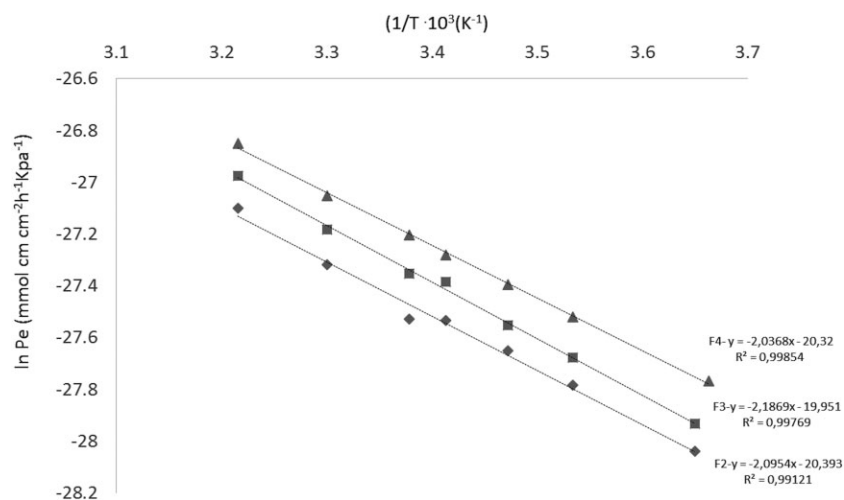


FIG. 1. ARRHENIUS PLOT OF CARBON DIOXIDE PERMEABILITY FOR THE NONCOMMERCIAL BIODEGRADABLE AND COMPOSTABLE FILMS

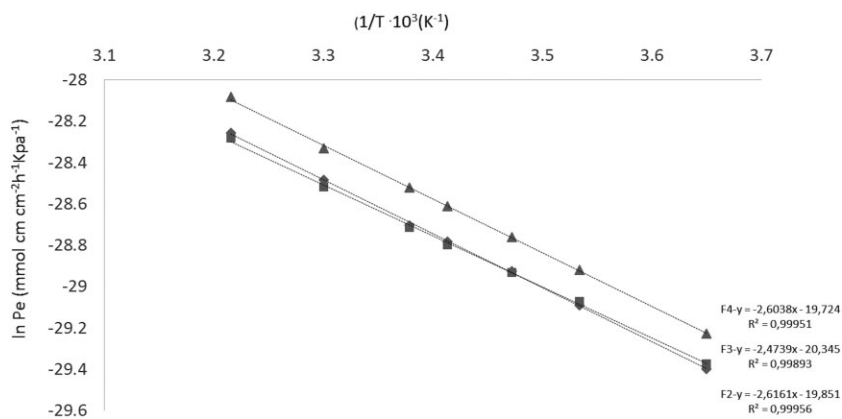


FIG. 2. ARRHENIUS PLOT OF OXYGEN PERMEABILITY FOR THE NONCOMMERCIAL BIODEGRADABLE AND COMPOSTABLE FILMS

high determination coefficients ($R^2 = 0.99$), and the slope of the fitted line allowed the estimation of the E_a/R .

For both O_2 or CO_2 and for all of the biodegradable and compostable films, the permeability values increased with an increase in temperature, but the range within which the O_2 and CO_2 values changed was different for the two storage temperatures (Table 3).

As for other materials, the selectivity (Pe_{CO_2}/Pe_{O_2}) of the biodegradable and compostable films suggested that the CO_2 flow was greater than the O_2 flow, but the ratio varied as a function of temperature (Cameron *et al.* 1995).

For all of the biodegradable and compostable films at 1C, the Pe_{CO_2} and Pe_{O_2} values showed small differences. Therefore, the selectivity values (Pe_{CO_2}/Pe_{O_2}) were similar and ranged between 3.9 (F2) and 4.3 (F4).

Conversely, for both gases, the values of gas permeability for the films at 20C were much more variable. The Pe_{CO_2} values were one order of magnitude higher than those for Pe_{O_2} .

At 20C, the selectivity values (Pe_{CO_2}/Pe_{O_2}) of each biodegradable and compostable film were lower, suggesting increased O_2 mobility at high temperatures. The values ranged between 3.5 (F2) and 4.1 (F3).

These differences could suggest higher solubility and greater hydrophilicity of CO_2 than O_2 in the matrix of the noncommercial films used and inside each package, which could explain the highest difference in the headspace CO_2 to O_2 ratio with the change of temperature. Based on the

values of most permeable plastic films (Cameron *et al.* 1995), the new tested films showed intermediate selectivity, which is important for controlling the respiratory exchange of fresh fruits, particularly at high temperature (Guilbert *et al.* 1996). The F3 film was the only one able to maintain a similar value of selectivity (Pe_{CO_2}/Pe_{O_2}) at both temperatures (4.2 and 4.1 at low and high temperatures, respectively), which could suggest better control of the atmosphere in the packed raspberries throughout the storage time.

Headspace Gas Composition

The O_2 and CO_2 levels detected in the sample package headspace during storage are reported in Figs. 3 and 4, respectively. The atmosphere inside the packages with the F1 film did not change at any storage time because of the macro hole (6-mm diameter).

The initial atmosphere gas composition changed in the packages with all of the innovative films (0.2 kPa CO_2 and 21.2 kPa O_2); the exchange area (550 cm^2) through the film packages was constant, so the evolution of the internal atmosphere inside the trays was passively created by the respiration rate of the cv. Grandeur and the permeability of the films to O_2 and CO_2 (Beaudry *et al.* 1992), both of which were affected by temperature (Kader *et al.* 1989). Differences had already been observed between the gas levels

Film	Temperature (C)	Pe_{CO_2} (mmol·cm/cm ² /h/kPa)	Pe_{O_2} (mmol·cm cm ² /h/kPa)	Pe_{CO_2}/Pe_{O_2}
F2	20	1.10E ⁻¹²	3.20E ⁻¹³	3.5
	1	6.64E ⁻¹³	1.70E ⁻¹³	3.9
F3	20	1.28E ⁻¹²	3.11E ⁻¹³	4.1
	1	7.39E ⁻¹³	1.75E ⁻¹³	4.2
F4	20	1.42E ⁻¹²	3.75E ⁻¹³	3.8
	1	8.71E ⁻¹³	2.03E ⁻¹³	4.3

TABLE 3. O_2 AND CO_2 PERMEABILITIES OF NONCOMMERCIAL BIODEGRADABLE AND COMPOSTABLE FILMS

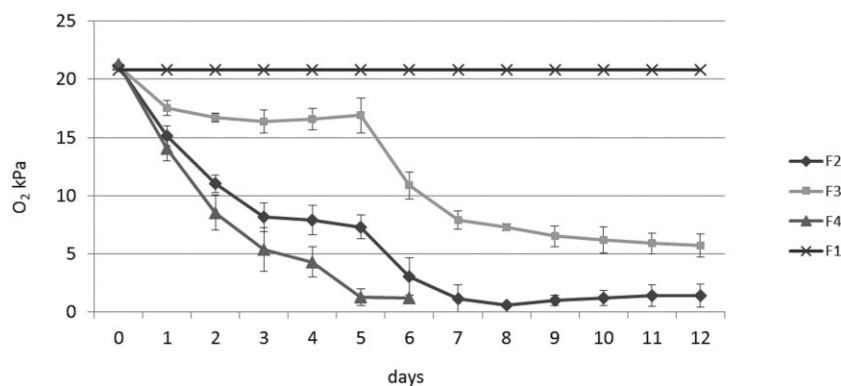


FIG. 3. HEADSPACE O₂ CONCENTRATION (KPA) OF RASPBERRY CV. GRANDEUR FRUITS STORED

established in the packages wrapped with the three different biodegradable and compostable films a few hours after packaging.

As expected, a decrease in the headspace O₂ and an increase in the headspace CO₂ during the entire storage time were observed.

A characterization of the F2, F3 and F4 films indicates that the ratio between the O₂TR and the CO₂TR is approximately 1:4 (Table 2). Thus, assuming that the respiratory quotient RQ = 1 and that the O₂ changes by 5% on the first day, the CO₂ should increase by less than 8%. This did not occur, most likely due to the different velocities of the respiration of the fruits and because the mass transfer rate of the gas must account for the concentration gradient between the inside and the outside of the package, which differs for O₂ and CO₂.

The O₂ decreased by up to 1.3, 7.3 and 16.9 kPa, whereas CO₂ increased by up to 18.6, 13.2 and 7.8 kPa with F4, F2 and F3 films, respectively, during fruit storage at 1 ± 1°C for up to 5 days. The lowest O₂ observed for the F4 film was due to the highest PeO₂ (Table 3), which affected the respiration rate of the cv. Grandeur. As a result, there was an increase in the concentration of CO₂, although the PeCO₂ for this film was greater than that of the other biodegradable and compostable films. The equilibrium of partial pressures

achieved with the F3 film could be considered the same as that recommended for raspberry fruits at low temperature (Callesen and Møller Holm 1989; Agar and Streif 1996; Kader 2001).

At 6 days, with an increase in storage temperature (20 ± 1°C), the changes in the atmospheric composition for all packages were greater than those at the previous temperature, most likely due to the relative increase in the respiration rate of fruits and the changes in film permeability. In particular, CO₂ permeability responded more than O₂ permeability (Table 3), and the rate of growth of CO₂ was higher than that of O₂. In fact, the O₂ values observed were 1.2, 3.1 and 10.9 kPa for the F4, F2 and F3 films, whereas CO₂ increased to 37.1, 17.0 and 14.8 kPa. At this point, the O₂ and CO₂ gas composition achieved for the F4 film became potentially harmful to fruit quality; according to Joles *et al.* (1994), the induction of anaerobic respiration in a high CO₂ atmosphere can cause multiple undesirable changes in the fruits, including the development of off-flavors (Argenta *et al.* 2002). In fact, the raspberry fruits packed with the F4 film could not be held to the end of the storage period, instead lasting only up to 6 days. However, as early as the fourth day, the gas values were critical, and the fruits were not marketable by visual analysis. At the end of the storage period (12 days), the O₂ values were 1.4 and

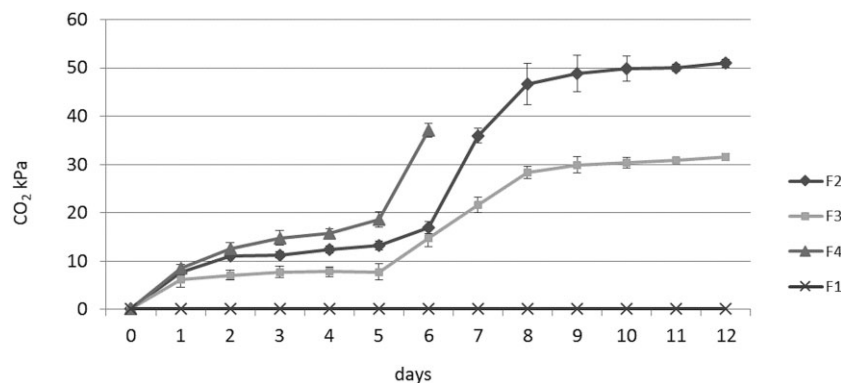


FIG. 4. HEADSPACE CO₂ CONCENTRATION (KPA) OF RASPBERRY CV. GRANDEUR FRUITS STORED

5.7 kPa, and the CO₂ values were 51.0 and 31.6 kPa for the F2 and F3 films, respectively. Although the fruits packed with the F2 film were kept until the end of the storage period, the gas values had reached the peak. In fact, the CO₂ value was already high from the eighth day.

The F3 film was the only one able to establish and maintain an equilibrated headspace gas composition, with O₂ values of 5.7 kPa and CO₂ values of 31.6 kPa as a result of the higher CO₂ permeability in these packages compared with those with the F2 film (Table 3).

Fruit Quality Assessment

Weight Loss. Generally, the weight loss (water loss) of the cv. Grandeur due to transpiration and respiration of the fruits increased with time, and its rate was dependent on the film used (Table 4). In fact, statistically significant differences were observed among the films used and different time points.

All of the wrapped fruits (F1, F2, F3 and F4 films) showed minimal weight loss, with values below 1% for 5 days in cold storage (1 ± 1°C). As reported in the literature, these values cannot be considered critical aspects of

raspberry marketability (Callesen and Møller Holm 1989; Haffner *et al.* 2002).

With the change and increase in temperature (20 ± 1°C) already evident after 6 days, the weight loss was significant for all of the packed fruits and unpacked fruits (control), showing a significant difference in weight of 2.96% compared with the other films. The lower weight loss in fruits packed with the biodegradable and compostable films compared with unpacked fruits (control) stemmed from the low rates of water loss in the MAP fruits, which occurred because the atmosphere of the packed fruits was limited; however, for the macroperforated film (F1), the reduction of weight loss was due to protection from mechanical damage. At 6 days of storage, neither the fruits stored with the F1 film nor the unwrapped fruits (control) were marketable, unlike the fruits packed with the other films.

The F2 and F3 films were the only films suitable for storing fruits for up to 12 days because they limited weight loss at high temperature (20 ± 1°C) due a good barrier to humidity. In fact, water condensation did not develop inside the packages, and no fungi were observed on the fruits (data not shown). It is well documented in the literature that bioplastics made from starch have a natural high

TABLE 4. WEIGHT LOSS, TOTAL SOLUBLE SOLIDS (TSS), TITRATABLE ACIDITY (TA) AND COLOR OF CV. GRANDEUR RASPBERRY FRUITS

Storage times (days)	Film	% weight loss	TSS (°Brix)	TA (meq/L)	<i>L</i>	Chroma	Hue
						(C)	(°h)
0	Harvest	–	9.2 ± 0.2	15.9 ± 0.2	31.6 ± 2.7	44 ± 4.4	29.8 ± 1.9
3	F1	0.44 ± 0.1	9.6 ± 0.1	13.1 ± 0.2	27.1 ± 1.3	39.5 ± 5.9	25.9 ± 2.8
	F2	0.30 ± 0.1	9.2 ± 0.1	13.5 ± 0.4	28.7 ± 2.0	40.6 ± 2.4	28.7 ± 3.1
	F3	0.51 ± 0.1	9.2 ± 0.0	14.9 ± 0.3	29.3 ± 2.3	37.6 ± 4.9	26.6 ± 2.6
	F4	0.38 ± 0.1	9.3 ± 0.1	12.9 ± 0.1	29.8 ± 2.5	38.9 ± 5.8	27.5 ± 2.9
	Control	1.18 ± 0.3	9.9 ± 0.1	13.7 ± 0.2	26.9 ± 2.6	32.8 ± 6.1	24.1 ± 2.4
5	F1	0.66 ± 0.66	9.4 ± 0.1	11.4 ± 0.6	26.5 ± 2.0	31.7 ± 4.7	24.4 ± 1.7
	F2	0.45 ± 0.08	9.2 ± 0.2	12.6 ± 0.3	28.7 ± 3.2	36.2 ± 5.2	25.9 ± 3.0
	F3	0.75 ± 0.08	9.2 ± 0.2	13.3 ± 0.2	29.4 ± 3.7	38.6 ± 6.1	26.1 ± 2.3
	F4	0.62 ± 0.10	9.3 ± 0.1	13.9 ± 0.1	30.6 ± 3.3	40.0 ± 6.1	27.5 ± 2.4
	Control	1.76 ± 0.35	9.5 ± 0.1	11.4 ± 0.9	26.3 ± 2.4	31.9 ± 4.8	23.6 ± 1.7
6	F1	1.02 ± 0.15	9.6 ± 0.1	12.6 ± 0.1	25.8 ± 2.1	29.9 ± 4.8	23.1 ± 1.6
	F2	0.81 ± 0.18	9.3 ± 0.2	15.4 ± 0.2	29.7 ± 2.5	38.7 ± 7.2	27.9 ± 3.1
	F3	1.15 ± 0.14	9.3 ± 0.2	15.1 ± 0.3	28.9 ± 2.1	33.6 ± 4.7	25.2 ± 2.0
	F4	1.19 ± 0.21	9.4 ± 0.1	14.8 ± 0.5	29.6 ± 3.7	39.4 ± 6.6	27.1 ± 2.9
	Control	2.96 ± 0.83	9.9 ± 0.1	13.4 ± 0.3	26.0 ± 2.2	29.3 ± 4.1	22.7 ± 1.2
12	F1	–	–	–	–	–	–
	F2	2.2 ± 0.23	9.1 ± 0.1	15.3 ± 0.2	26.6 ± 2.4	34.1 ± 1.7	28.7 ± 1.9
	F3	2.9 ± 0.30	9.2 ± 0.1	13.9 ± 0.1	29.0 ± 2.3	36.4 ± 2.1	26.5 ± 1.6
	F4	–	–	–	–	–	–
	Control	–	–	–	–	–	–
LSD (5%)							
Storage time		0.000	0.000	0.14	0.476	0.000	0.000
Film		0.001	0.000	0.122	0.000	0.000	0.000
Storage time × Film		0.155	0.000	0.022	0.310	0.000	0.001

Least significant difference (LSD) values for $P < 0.05$.

permeability to water vapor (Muller *et al.* 1991); at the end of the storage period, weight loss values of 2.2 and 2.9% were observed for the F2 and F3 films, respectively. The data showed that weight loss was not a limiting factor for the quality of the cv. Grandeur fruits during storage, even when the berries were subjected to metabolic stress due to the change of temperature.

TSS and TA. The TSS and TA contents contribute to fruit flavor, and high values are required for good berry flavor (Kader 1991). After 3 days of storage at $1 \pm 1\text{C}$, the macroperforated film (F1) and unwrapped fruits (control) already showed the highest TSS values (9.6 and 9.9°Brix, respectively), and higher values than those of the biodegradable and compostable films were maintained throughout the storage time as a result of the highest water loss. The storage times, the film used and their interaction were the factors that significantly affected the TSS values. The F2 and F3 films maintained TSS values close to those of the fresh fruits (9.2°Brix) throughout the storage time and particularly at 6 days of storage, whereas the F4 film showed intermediate TSS values with the increase of temperature. This was most likely due to the good barrier properties with respect to water vapor for the films used (Peelman *et al.* 2013), even at high temperatures.

The TA of red raspberries cv. Grandeur decreased with storage time compared with the TA of fresh fruits (15.9 meq/L), thus corroborating the findings of Robbins *et al.* (1989) and Haffner *et al.* (2002). The lowest value was achieved after 12 days of storage with the F3 treatment (13.9 meq/L), but the values were generally maintained within a range that is not critical for consumer acceptability. The interaction between storage time and the film was statistically significant. The fruits maintained at low temperature ($1 \pm 1\text{C}$) (F1 and control) showed the lowest acidity (11.4 meq/L) after 5 days of storage because of the highest weight loss. An increase in the TA values was observed for all of the packages as the temperature increased. In particular, the highest value was observed for the fruits stored in MAP. At the end of the storage period, the F2 film showed the highest TA value (15.3 meq/L) because it had the highest CO₂ headspace concentration, thus corroborating the results of Malhotra and Prasad (1999) and of Remon *et al.* (2003) for packed cherries.

Color. The chromatic characteristics of stored cv. Grandeur are shown in Table 4. The external color derived from anthocyanins is related to consumers' perception of quality and is an important parameter for fruit ripeness and freshness (Krüger *et al.* 2011). According to the literature, fruit color changes after storage (Varseveld and Richardson 1980; Sjulín and Robbins 1983).

All of the color parameters (L^* , C^* and h°) were significantly affected by the film used (Table 4). At both storage temperatures (1 ± 1 and $20 \pm 1\text{C}$), the biodegradable and compostable films (F2, F3 and F4) showed a positive effect on the retention of external color of the cv. Grandeur compared with the macroperforated film (F1) and unwrapped fruits (control) due to the modification of the atmosphere inside each package, with a relative accumulation of CO₂. A decrease in the L value reflected the darkening of fruits by anthocyanin accumulation and indicated that the ripening process had occurred in the fruits. At the end of refrigerated storage (5 days), berries packed with the macroperforated film (F1) and unwrapped fruits (control) lost more of their luminosity (i.e., showed greater decreases in L^*) than berries packed with the other films (F2, F3 and F4), with L^* values of 26.5 and 26.3, respectively. The F4 film showed the highest L^* value (30.6), as a consequence of the high CO₂ in the headspace gas composition (Fig. 4). At 6 days of storage, L^* values decreased, and the same trend was found for each film; however, at the end of the storage period (12 days), differences in the L^* values were observed between the F2 and F3 films due to the different gas compositions inside the packages.

During storage, the color of the fruits became less vivid than at the time of harvest (44.0; lower chroma), and this trend was more evident for fruits packed with the macroperforated film (F1) and unwrapped fruits (control) than the others. The C value was significantly affected by the storage time, the film used and their interaction.

A reduction in the fruit hue angle (h° ; less vivid fruits) from harvest (29.8°) was observed during storage for all of the packages, and this parameter was significantly affected by the storage time, the film used and their interaction. This decrease reflected the change in fruit colour from light green to deep purple due to the accumulation of anthocyanin (Fig. 6). The h° values were directly related to the humidity during storage (Goncalves *et al.* 2007); in fact, the highest h° value for each quality control was achieved by the biodegradable and compostable films (F2, F3 and F4), corresponding to high water vapor barrier properties at both storage temperatures. The h° values of the cv. Grandeur stored at $20 \pm 1\text{C}$ decreased faster than did those stored at $1 \pm 1\text{C}$; in particular, the lowest values were observed after 6 days for the macroperforated film (F1) and unwrapped fruits (control), with values of 23.1° and 22.7°, respectively.

Total Anthocyanin and Phenolic Contents and Antioxidant Activity. Although anthocyanin, phenolic content and antioxidant activity of raspberry fruits have been previously reported, this is the first report on changes in cv. Grandeur after storage using different biodegradable and compostable films for packages.

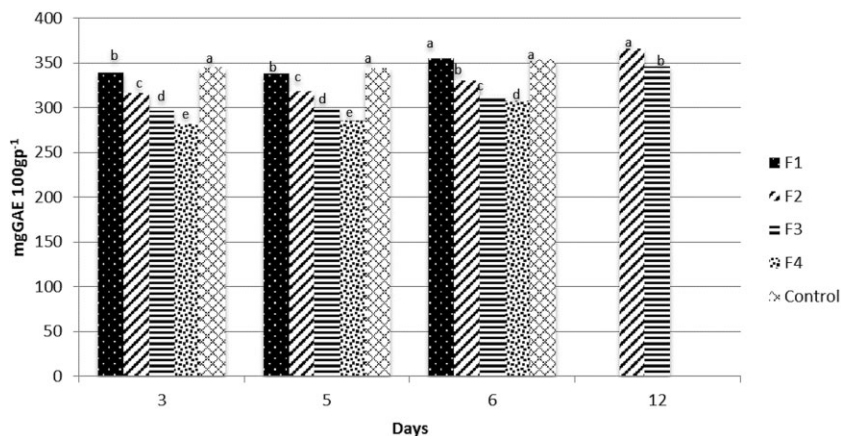


FIG. 5. TOTAL ANTHOCYANIN CONTENT OF RASPBERRY CV. GRANDEUR FRUITS STORED IN BIODEGRADABLE AND COMPOSTABLE FILMS

Anthocyanins are the main compounds that contribute to the bright, red color of fruits; their synthesis continues after fruits are harvested, and factors that favor the stability of the pigments include the absence of oxygen, low pH and low processing/storage temperatures (Kalt *et al.* 1999). The cv. Grandeur was confirmed as a valuable source of potentially healthy compounds. In fact, the total content of anthocyanins in fresh raspberry fruits of cv. Grandeur (37.60 mg/100 g FW; Fig. 5) was in the range of values reported in the literature for red raspberries (Deighton *et al.* 2000; Weber *et al.* 2008; Sariburun *et al.* 2010). All of the packages showed an increase in total anthocyanins compared with the values at harvest because of the decreases in TA and increases in weight loss during storage, as reported by Mazza and Minati (1993). The interconversion of organic acids and carbohydrates may provide carbon skeletons for the synthesis of phenolics, including both anthocyanin and non-anthocyanin phenolics. The anthocyanin content was significantly different between packages. Raspberries stored with the macroperforated film (F1) and unwrapped fruits (control) showed darker colors (Table 4), with the pigment levels increasing after 5 days, whereas

storage in packages with a modified atmosphere (F2, F3 and F4) protected the integrity of the fruits and kept the pigment content relatively unchanged, thereby preventing color change during storage at 1 ± 1°C. According to Kalt *et al.* (1999), smaller changes in anthocyanin content were reported after storage at low temperature. In fact, after 5 days of storage, the total anthocyanin content values were 49.38, 47.18 and 44.33 mg C3G/100 g FW for the F2, F3 and F4 films, respectively. An increase was observed for each package as the temperature changed (6 days) due to an increase in sugar synthesis (Table 4), as described in other soft fruit species (Mori and Sakurai 1994; Perkins-Veazie *et al.* 2000) and in agreement with Seglina *et al.* (2009). The F2 and F3 films showed a similar increase (12–13.8%), whereas the F4 film showed an increase of 20.5%. At the end of the storage period (12 days), the total anthocyanin content of the cv. Grandeur ranged from 79.47 (F2) to 64.74 mg C3G/100 g FW (F3; Fig. 6).

The behaviour of the polyphenol content in the cv. Grandeur was similar to that observed for the anthocyanin content. A strong positive correlation ($r = 0.81$) was found between the total anthocyanin and the phenolic content,

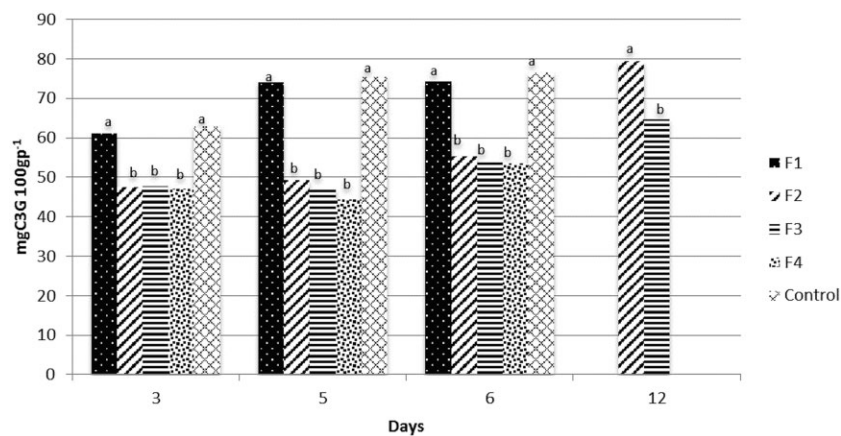
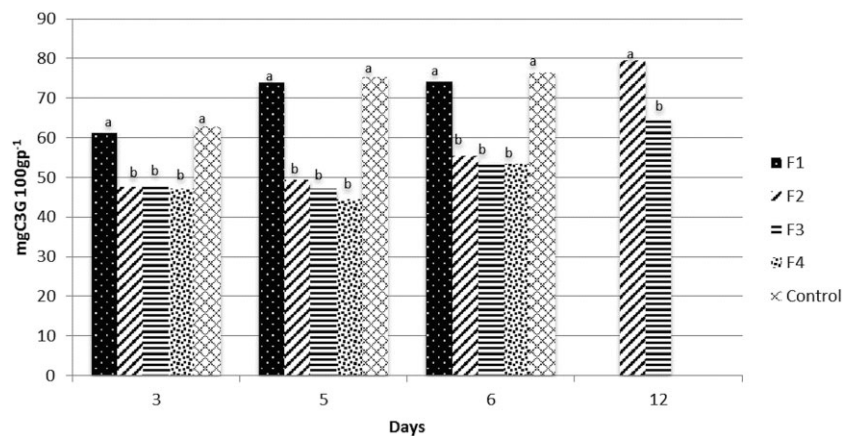


FIG. 6. TOTAL POLYPHENOL CONTENT OF RASPBERRY CV. GRANDEUR FRUITS STORED IN BIODEGRADABLE AND COMPOSTABLE FILMS

For each time point, the means followed by different letters are significantly different at $p \leq 0.05$ according to Tukey's test.

FIG. 7. TOTAL ANTIOXIDANT CAPACITY OF RASPBERRY CV. GRANDEUR FRUITS STORED IN BIODEGRADABLE AND COMPOSTABLE FILMS

For each time point, the means followed by different letters are significantly different at $p \leq 0.05$ according to Tukey's test.



showing that fruits with a higher anthocyanin content had a higher phenolic content. The evolution of the total polyphenol content of the cv. Grandeur (Fig. 6) was in agreement with that reported by Wang and Lin (2000), and all stored raspberries showed values higher than those of fresh fruits (210.15 mg GAE/100 g FW).

In our study, the change in the total polyphenol content occurred more slowly at the lower temperature (1 ± 1 C), and the macroperforated film (F1) and unwrapped fruits (control) generally maintained higher values compared with the biodegradable and compostable films. After 5 days of storage, the total polyphenol contents were 318.78, 299.16 and 286.26 mg GAE/100 g FW for the F2, F3 and F4 films, respectively. As reported by Kalt *et al.* (1999), raspberries stored at temperatures greater than 1C showed an increase in the total phenolic content, and the magnitude of the increase was related to the storage temperature. The highest value was observed at the end of the storage period (12 days) for the F2 and F3 films (366.02 and 345.92 mg GAE/100 g FW, respectively).

Due to its high anthocyanin and polyphenol content, cv. Grandeur has good antioxidant potential. Wang and Lin (2000) observed an increase in the antioxidant activity of red raspberries from the pink to the red ripe stage, corresponding to increased levels of anthocyanin and total phenolics. Similarly, storage had an important influence on the evolution of antioxidant capacity, and all of the packages showed values greater than those of fresh fruits (0.81 mmol Fe⁺²; Fig. 7) throughout the supply chain. The increase in antioxidant capacity corresponded to the evolution of the total phenolic and anthocyanin content, and at the end of the storage period (12 days), the highest value was found for the F2 film (2.20 mmol Fe⁺²).

CONCLUSION

This study indicated that the new films used to wrap the packages are good substitutes for traditional plastic film

(polypropylene macroperforated film) and could be used throughout the supply chain of the cv. Grandeur. The new tested films minimized changes in weight loss and color of the raspberries, showing the best performance at a lower temperature (1 ± 1 C) and changed the initial atmospheric composition inside the packages due to the good permeability properties of films to the gas and the respiration of cv. Grandeur. As expected, all of the qualitative traits of the fruits were affected by the temperature increase, but the F2 and F3 films were able to maintain the qualitative traits of the cv. Grandeur for up to 12 days. However, the F2 film showed critical gas values during storage, particularly after the temperature increase. Generally, the thinner biodegradable and compostable film (F3) was more likely to maintain the most important qualitative and nutraceutical traits closest to those at harvest because it showed more equilibrated film selectivity throughout the storage time, preserving the metabolic stress of the fruit that was inevitable with a change of temperature. The potential application of these noncommercial films for packages combined with the introduction of the new cv. Grandeur could increase the accessibility on the market affected by potential temperature fluctuations during commercial postharvest, and the extension of the marketability up to 12 days could justify the likely high costs of using these materials.

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REFERENCES

- ACKERMAN, S.M. and ADAMS, S.W. 2009. Inventors. Raspberry plant named “Grandeur”. U.S. Patent PP20459P3.

- ADAY, M.S. and CANER, C. 2011. The applications of “active packaging and chlorine dioxide” for extended shelf life of fresh strawberries. *Packag. Technol. Sci.* 24, 123–136.
- ADAY, M.S. and CANER, C. 2013. The shelf life extension of fresh strawberries using an oxygen absorber in the biobased package. *LWT – Food Sci. Technol.* 52, 102–109.
- AGAR, I.T. and STREIF, J. 1996. Effect of high CO₂ and controlled atmosphere (CA) storage on the fruit quality of raspberry. *Gartenbauwissenschaft* 61, 261–267.
- ARGENTA, L.C., FAN, X. and MATTHEIS, J.P. 2002. Responses of “Fuji” apples to short and long duration exposure to elevated CO₂ concentration. *Postharvest Biol. Technol.* 24, 13–24.
- ASTM. 2000a. *ASTM-D882-02. Standard Test Method for Tensile Properties of Thin Plastic*, American Society for Testing and Materials, Philadelphia, PA.
- ASTM. 2000b. *ASTM-D1003-00. Standard Test Method for Haze and Luminous Transmittance of Transparent Plastics*, American Society for Testing and Materials, Philadelphia, PA.
- ASTM. 2005. *ASTM F2476-05. Test Method for the Determination of Carbon Dioxide Gas Transmission Rate (CO₂TR) Through Barrier Materials Using an Infrared Detector*, American Society for Testing and Materials, Philadelphia, PA.
- ASTM. 2008. *ASTM F2622-08. Standard Test Method for Oxygen Gas Transmission Rate Through Plastic Film and Sheeting Using Various Sensors*, American Society for Testing and Materials, Philadelphia, PA.
- BEAUDRY, R.M., CAMERON, A.C., SHIRAZI, A. and DOSTALLANGE, D.L. 1992. Modified atmosphere packaging of blueberry fruit – effect of temperature on package O₂ and CO₂. *J. Am. Soc. Hortic. Sci.* 117, 436–441.
- BRECHT, J.K., CHAU, K.V., FONSECA, S.C., OLIVEIRA, F.A.R., SILVA, F.M., NUNES, M.C.N. and BENDER, R.J. 2003. Maintaining optimal atmosphere conditions for fruits and vegetables throughout the postharvest handling chain. *Postharvest Biol. Technol.* 27, 87–101.
- CALLESEN, O. and MØLLER HOLM, B. 1989. Storage results with red raspberries. *Acta Hortic.* 262, 247–254.
- CAMERON, A.C., TALASILA, P.C. and JOLLES, D.W. 1995. Predicting film permeability needs for modified-atmosphere packaging of lightly processed fruits and vegetables. *HortScience* 30, 25–34.
- CHENG, G.W. and BREEN, P.J. 1991. Activity of phenylalanine ammonia-lyase (PAL) and concentrations of anthocyanins and phenolics in developing strawberry fruit. *J. Am. Soc. Hortic. Sci.* 116, 865–869.
- CRANK, J. and PARK, G.S. 1968. *Diffusion in Polymers*, p. 360, Academic Press, London.
- DEIGHTON, N., BRENNAN, R., FINN, C. and DAVIES, H.V. 2000. Antioxidant properties of domesticated and wild *Rubus* species. *J. Sci. Food Agric.* 80, 1307–1313.
- EXAMA, A., ARUL, J., LENCKI, R.W., LEE, L.Z. and TOUPIN, C. 1993. Suitability of plastic films for modified atmosphere packaging of fruits and vegetables. *J. Food Sci.* 58, 1365–1370.
- GIRGENTI, V., PEANO, C., BOUNOUS, M. and BAUDINO, C. 2013. A life cycle assessment of non-renewable energy use and greenhouse gas emission associated with blueberry and raspberry production in northern Italy. *Sci. Total Environ.* 458–460, 415–418.
- GONCALVES, B., SILVA, A.P., MOUTINHO-PEREIRA, J., BACELAR, E., ROSA, E. and MEYER, A.S. 2007. Effect of ripeness and postharvest storage on the evolution of color and anthocyanins in cherries (*Prunus avium* L.). *Food Chem.* 103, 976–984.
- GUILBERT, S., GONTARD, N. and GORRIS, L.G.M. 1996. Prolongation of the shelf life of perishable food products using biodegradable films and coatings. *LWT – Food Sci. Technol.* 29, 10–17.
- GUILLAUME, C., SCHWAB, I., GASTALDI, E. and GONTARD, N. 2010. Biobased packaging for improving preservation of fresh common mushrooms (*Agaricus bisporus* L.). *Innov. Food Sci. Emerg. Technol.* 11, 690–696.
- HAFFNER, K., ROSENFELD, H.J., SKREDE, G. and WANG, L. 2002. Quality of red raspberry *Rubus idaeus* L. cultivars after storage in controlled and normal atmospheres. *Postharvest Biol. Technol.* 24, 279–289.
- JEONG, M., AN, D.S., AHN, G.H. and LEE, D.S. 2013. Master packaging system for sweet persimmon applicable to produce supply chains. *Postharvest Biol. Technol.* 86, 141–146.
- JOLLES, D.W., CAMERON, A.C., SHIRAZI, A., PETRACEK, P.D. and BEAUDRY, R.M. 1994. Modified-atmosphere packaging of “Heritage” red raspberry fruit: Respiratory response to reduced oxygen, enhanced carbon dioxide, and temperature. *J. Am. Soc. Hortic. Sci.* 119, 540–545.
- JOO, M.J., LEWANDOWSKI, N., AURAS, R., HARTE, J. and ALMENAR, E. 2011. Comparative shelf life study of blackberry fruit in bio-based and petroleum-based containers under retail storage conditions. *Food Chem.* 126, 1734–1740.
- KADER, A.A. 1991. Quality and its maintenance in relation to the postharvest physiology of strawberry. In *The Strawberry into the Twenty-First Century* (J.J. Luby and A. Dale, eds.) pp. 145–152, Timber Press, Portland, OR.
- KADER, A.A. 2001. A summary of CA requirements and recommendations for fruits other than apples and pears. In *Postharvest Horticultural Series* (A.A. Kader, ed.) pp. 29–70, University of California, Davis, CA.
- KADER, A.A., ZAGORY, D. and KERBEL, E.L. 1989. Modified atmosphere packaging of fruits and vegetables. *Crit. Rev. Food Sci. Nutr.* 28, 1–30.
- KALT, W., FORNEY, C.F., MARTIN, A. and PRIOR, R.L. 1999. Antioxidant capacity, vitamin C, phenolics, and anthocyanins after fresh storage of small fruits. *J. Agric. Food Chem.* 47, 4638–4644.
- KRÜGER, E., DIETRICH, H., SCHÖPPLEIN, E., RASIM, S. and KÜRBEL, P. 2011. Cultivar, storage conditions and ripening effects on physical and chemical qualities of red raspberry fruit. *Postharvest Biol. Technol.* 60, 31–37.

- LINKE, M. and GEYER, M. 2013. Condensation dynamics in plastic film packaging of fruit and vegetables. *J. Food Eng.* 116, 144–154.
- MAKINO, Y. and HIRATA, T. 1997. Modified atmosphere packaging of fresh produce with a biodegradable laminate of chitosan-cellulose and polycaprolactone. *Postharvest Biol. Technol.* 10, 247–254.
- MALHOTRA, J. and PRASAD, D.N. 1999. Role of carbon dioxide in enhancing the microbiological quality of perishable foods: A review. *Microbiol. Alims Nutr.* 17, 155–168.
- MARSH, K. and BUGUSU, B. 2007. Food packaging-roles, materials, and environmental issues. *J. Food Sci.* 72, R39–R54.
- MAZZA, G. and MINATI, E. 1993. *Anthocyanins in Fruits, Vegetables, and Grains*, CRC, Boca Raton, FL.
- MCGUIRE, R.G. 1992. Reporting of objective color measurements. *HortScience* 27, 1254–1255.
- MORI, T. and SAKURAI, M. 1994. Production of anthocyanin from strawberry cell suspension cultures; effects of sugar and nitrogen. *J. Food Sci.* 59, 588–593.
- MULLER, H.P., TILLMAN, H. and GUNTER, W. 1991. Inventors. U.S. Patent EP449041 A2.
- NATIONAL INSTITUTE OF STANDARDS AND TECHNOLOGY (NIST). 2008. *Guide for the Use of the International System of Units (SI), Special Publication 811* (A. Thompson and B.N. Taylor, eds.) U.S. Dept. of Commerce, Gaithersburg, MD.
- PARAŠIĆ, V., MIHAILOVIĆ, B. and HAMOVIĆ, V. 2010. Imperfect competition in the primary agricultural commodity market in Serbia. *Econ. Ann.* 184, 113–150.
- PEANO, C., GIRGENTI, V., PALMA, A., FONTANELLA, E. and GIUGGIOLI, N.R. 2013. Film type and MAP on cv. Himbo Top raspberry fruit quality, composition and volatiles. *Ital. J. Food Sci.* 4, 1–12.
- PEELMAN, N., RAGAERT, P., DE MEULENAER, B., ADONS, D., PEETERS, R., CARDON, L., VAN IMPE, F. and DEVLIEGHERE, F. 2013. Application of bioplastics for food packaging. *Trends Food Sci. Technol.* 32, 128–141.
- PELLEGRINI, N., SERAFINI, M., COLOMBI, B., DEL RIO, D., SALVATORA, S. and BIANCHI, M. 2003. Total antioxidant capacity of plant foods, beverages and oils consumed in Italy by three different *in vitro* assays. *J. Nutr.* 133, 2812–2819.
- PERKINS-VEAZIE, P., CLARK, J.R., HUBER, D.J. and BALDWIN, E.A. 2000. Ripening physiology in “Navaho” thornless blackberries: Color, respiration, ethylene production, softening, and compositional changes. *J. Am. Soc. Hortic. Sci.* 125, 357–363.
- REMON, S., VENTURINI, M.E., LOPEZ-BUESA, P. and ORIA, R. 2003. Burlat cherry quality after long range transport: Optimisation of packaging conditions. *Innov. Food Sci. Emerg. Technol.* 4, 425–434.
- ROBBINS, J., SJULIN, T.M. and PATTERSON, M. 1989. Postharvest storage characteristics and respiration rates in five cultivars of red raspberries. *HortScience* 24, 980–982.
- SALUNKHE, D.K. and DESAI, B.B. 1984. *Small Fruits – Berries. Postharvest Biotechnology of Fruits*, p. 111, CRC Press, Boca Raton, FL.
- SARIBURUN, E., SALIHA, S., CEVDET, D., CIHAT, T. and VILDAN, S. 2010. Phenolic content and antioxidant activity of raspberry and blackberry cultivars. *J. Food Sci.* 75, C328–C335.
- SEGLINA, D., KRASNOVA, I., HEIDEMANE, G., KAMPUSE, S., DUKALSKA, L. and KAMPUSS, K. 2009. Packaging technology influence on the shelf life extension of fresh raspberries. *Acta Hortic.* 877, 433–440.
- SJULIN, T.M. and ROBBINS, J.A. 1983. Shelf life studies of red raspberry varieties. Proceedings of the 73rd Annual Meeting Western Washington Horticultural Association, Mount Vernon, WA, pp. 116–122.
- SLINKARD, K. and SINGLETON, V.L. 1977. Total phenol analysis: Automation and comparison with manual methods. *Am. J. Enol. Vitic.* 28, 49–55.
- TALASILA, P.C., CHAU, K.V. and BRECHT, J.K. 1995. Modified atmosphere packaging under varying surrounding temperature. *Trans. ASAE* 38, 869–876.
- VARSEVELD, G.W. and RICHARDSON, D.G. 1980. Evaluation of storage and processing quality of mechanically and hand-harvested *Rubus* spp. fruit. *Acta Hortic.* 112, 265–272.
- WAGHMARE, R.B. and ANNAPURE, U.S. 2013. Combined effect of chemical treatment and or modified atmosphere packaging (MAP) on quality of fresh-cut papaya. *Postharvest Biol. Technol.* 85, 147–153.
- WANG, S.Y. and LIN, H.S. 2000. Antioxidant activity in fruits and leaves of blackberry, raspberry, and strawberry varies with cultivar and developmental stage. *J. Agric. Food Chem.* 48, 140–146.
- WEBER, C.A., PERKINS-VEAZIE, P., MOORE, P.P. and HOWARD, L. 2008. Variability of antioxidant content in raspberry germplasm. *Acta Hortic.* 77, 493–498.
- YAM, K.L. and PAKADIS, S.E. 2004. A simple digital imaging method for measuring and analysing color of food surfaces. *J. Food Eng.* 61, 137–142.