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1 **DIFFERENT COATINGS FOR THE HS-SBSE GRAPE VOLATILE ANALYSIS IN**
2 **MODEL SOLUTION: PRELIMINARY RESULTS**

3
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32 **ABSTRACT**

33 Head space stir bar sorptive extraction was used to evaluate the efficacy of two
34 stir bar coatings, poly(ethylene glycol) (PEG)-modified silicone (EG-Silicone) and
35 polyacrylate (PA), in comparison with polydimethyl siloxane (PDMS) coating. Two
36 temperatures (0 °C and 25 °C) of the CIS injection system were compared with the
37 aim to propose an analytical procedure faster respect to traditional extractive
38 methods and environmental friendly, avoiding the use of solvents and high-energy
39 consumption for cooling. Fifteen volatiles with different polarities were tested. The
40 PDMS coating showed higher relative areas for terpenes and norisoprenoids. C6
41 volatiles, benzaldehyde and eugenol, obtained greater area response using EG-
42 Silicone and PA coatings, regardless the temperatures. EG Silicone and PA coatings
43 showed a higher memory effect for a certain compounds, probably due to the
44 lower desorption temperature that EG Silicone and PA coatings can be submitted
45 to, respect to PDMS (220 °C respect to 295 °C).

46

47 **Keywords:** volatiles, PDMS, EG-Silicone, Polyacrylate, grape.

48

49 **1. Introduction**

50 Aroma is one of the most important characteristics of wine, being related to
51 product quality and consumer acceptance. Among the constituents of wine aroma,
52 the ones from grapes play an important role in determining quality and
53 authenticity. Isolation, identification and quantification of grape volatiles are
54 pivotal to understand key events of vine response to external stimuli. Thus, it is
55 necessary to find rapid and efficient analytical methods that can help technicians

56 and grapegrowers to make the most correct choice at harvest and during
57 winemaking, when time is often the critic point.

58 Traditional analytical methods require time-consuming extraction and enrichment
59 steps prior to chromatography. Modern techniques in sample preparation have
60 evolved towards simplification by reducing solvents and sample handling. Stir bar
61 sorptive extraction (SBSE; commercially named 'Twister®') (Baltussen, Sandra,
62 David & Cramers, 1999; Baltussen, Cramers & Sandra, 2002) is an extraction
63 technique that is less time consuming respect to traditional extractive techniques
64 and does not employ solvents. Moreover, it has been largely automated. The stir
65 bar is coated with a polymeric extracting phase that traps the volatile analytes in
66 liquid samples (immersion mode) or in headspace. SBSE technique was used to
67 analyze grape and wine volatiles by the immersion mode (Ferrandino, Carlomagno,
68 Baldassarre & Schubert, 2012; Pardo-García, Serrano de la Hoz, Zalacain, Alonso &
69 Salinas, 2014; Zalacain, Marín, Alonso & Salinas, 2007), by the headspace mode
70 (Maggi, Zalacain, Mazzoleni, Alonso & Salinas, 2008) or by both of them (Arbulu,
71 Sampedro, Sánchez-Ortega, Gómez-Caballero, Unceta, Goicolea & Barrio, 2013). In
72 comparison with the immersion mode, the headspace SBSE mode extends the life
73 of the stir bar, especially in complex matrix. The SBSE technique showed analytical
74 simplicity, high reproducibility and very low detection limits. Polydimethyl
75 siloxane (PDMS) was the universal commercially available stationary phase
76 coating material. PDMS is a sorptive material that at temperature above its glass
77 transition temperature acquires a gum-like state with chemical properties such as
78 diffusion and distribution constants similar to those of organic solvents. SBSE-
79 PDMS has a stationary phase volume much more higher respect to SPME-PDMS:
80 the maximum volume of PDMS in a SPME fiber is 0.5 microliter (film thickness of

81 100 micrometer) whereas PDMS in SBSE can provide from 24 to 126 microliters of
82 volume. Thus, the sensitivity of PDMS-SBSE is higher, resulting in a higher volatile
83 extraction respect to SPME and in the detection and identification of compounds
84 present at very low concentrations, such as sesquiterpenes (May & Wüst, 2012;
85 Ferrandino, Carlomagno, Baldassarre & Schubert, 2012) or malodors in wastewaters
86 (Loughrin, 2006). PDMS stir bars were also used for the detection of stilbene molecules
87 in wines (Cacho, Campillo, Viñas & Hernández-Córdoba, 2013). However, doubts
88 about its efficaciousness arose due to the PDMS non-polar nature that was thought
89 to limit the applicability of SBSE-PDMS to hydrophobic compounds. Slightly polar
90 or polar coatings have recently been introduced but their use is limited. Cacho,
91 Campillo, Viñas & Hernández-Córdoba (2015) compared the use of these three
92 different coatings for the determination of volatile terpenes in honey, of chlorophenols
93 and chloroanisoles in wines (Cacho, Campillo, Viñas & Hernández-Córdoba, 2014).
94 EG-Silicone stir-bars were recently used to measure ochratoxin A in beers (Nguyen &
95 Dojin, 2014).

96 For the thermo-desorption of the retained compounds on the stir-bar, a specific
97 injector, constituted by a Thermal Desorption Unit (TDU) and of a Cooled Injection
98 System (CIS), is generally used. Different CIS temperatures, such as: -50 °C, (Caven-
99 Quantrill & Bouglass, 2011); -30 °C (Martínez-Gil, Garde-Cerdán, Martínez, Alonso &
100 Salinas, 2011; Martínez-Gil, Pardo-García, Zalacain, Alonso & Salinas, 2013); 0 °C
101 (Ferrandino, Carlomagno, Baldassarre & Schubert, 2012); 15 °C (Cacho, Campillo,
102 Viñas, & Hernández-Córdoba, 2015); 30 °C (Ochiai, Sasamoto, Ieda, David & Sandra,
103 2014), were used to measure volatiles with the PDMS-SBSE/GC-MS technique, but at
104 present no studies deal with the comparison of specific CIS temperatures, and with
105 different coatings.

106 The CIS is an injection system with programmed temperature vaporization
107 (PTV) that allows the controlled vaporization of the sample after an enrichment
108 step. The CIS liner can be rapidly heated or cooled. Moderate cooling can be
109 accomplished using a Peltier Cooling Unit but, to achieve more important cooling,
110 liquid nitrogen (N₂) or liquid carbon dioxide (CO₂) are used, increasing the costs
111 and the complexity of the analysis. The analytes on the stir bar can be cooled very
112 quickly, but when the temperature ranges are very wide, more time is required to
113 get the desired temperatures; besides, extreme ranges of temperature imply a
114 more rapid deterioration of the analytical equipment. Thus, analysis are more
115 expensive and a higher energy consumption is required. In most articles, the CIS
116 temperature is a parameter systematically described but not further commented.
117 No studies comparing the effect of different CIS temperatures on the analytical
118 response of the different volatiles have been conducted. We deem necessary to
119 make an effort in the direction of 'energetic' resource reduction also in the
120 laboratory, adapting, when possible, the analytical technique for 'energy saving'.

121 In the present study, two stir bar coatings: poly(ethylene glycol) (PEG)-modified
122 silicone (EG Silicone) and polyacrylate (PA) were compared to the traditional
123 PDMS stir bar to analyse some grape volatile compounds. The stir bars were used
124 in the HS mode. Two different CIS temperatures (0 °C and 25 °C) were assayed to
125 identify running temperatures as high as possible without limiting the analytical
126 results, to propose a more environmental friendly analytical procedure. However,
127 results are still preliminary as validation on musts and wines and on a larger
128 number of molecules is required.

129

130 **2. Materials and methods**

131

132 *2.1. Chemicals and reagents*

133 Benzaldehyde, β -citronellol, eugenol, geraniol, 1-hexanol, (*E*)-2-hexenal, (*E*)-2-
134 hexen-1-ol, (*Z*)-3-hexen-1-ol, α -ionone, β -ionone, D-limonene, linalool, nerolidol
135 and α -terpineol were purchased from Sigma-Aldrich (Steinheim, Germany). A
136 mixture of commercial standards of high purity grade (>97%) in methanol was
137 prepared, using concentration ranges for each compound commonly found in
138 wines. A model solution was prepared: an aqueous ethanol solution at 12% with 5
139 g L⁻¹ of tartaric acid was and pH adjusted to 3.6 with 1M sodium hydroxide
140 (Zalacain, Marin, Alonso & Salinas, 2007) (Table 1).

141 The internal standard was γ -hexalactone (Sigma-Aldrich, Steinheim, Germany)
142 solution at 1 μ L mL⁻¹ in absolute ethanol (Merck, Darmstad, Germany).

143 were employed to prepare model solutions.

144

145 *2.2. Stir bar coatings*

146 To perform the sorptive extraction process, commercial stir bars coated with
147 polydimethylsiloxane (PDMS Twister®), poly (ethylene glycol) (PEG)-modified
148 silicone (EG Silicone Twister®) and polyacrylate (Acrylate Twister® PA) were used,
149 the last two freely provided by Gerstel (Mülheim and der Ruhr, Germany towards
150 SRA Instruments, Cernusco sul Naviglio, MI, Italy). They consisted of a 10 mm long
151 glass-encapsulated magnetic stir bar with three different external coatings: 24 μ L
152 of PDMS, 32 μ L of EG Silicone and 25 μ L of PA.

153

154 *2.3. Volatile extraction*

155 The model solution was extracted by headspace (HS-SBSE; Martínez-Gil, Garde-
156 Cerdán, Martínez, Alonso & Salinas, (2011). Twenty-two mL of model solution were
157 placed into a 50 mL headspace vial. For each milliliter of sample, 0.1 g of NaCl and
158 10 μ L of internal standard (γ -hexalactone) were added. HS-SBSE extraction was
159 carried out by exposing the coated stir bar to the vial headspace hermetically
160 closed, stirring the sample was stirred at 500 rpm and heating at 60 °C for 60 min.
161 After sorption, the stir bar was rinsed with pure water, dried with a lint-free tissue
162 and desorbed.

163

164 *2.4. Analytical procedure*

165 The sample introduction system was composed of an automated thermal
166 desorption unit (TDU), combined with a MultiPurpose Sampler (MPS) and a
167 Programmed Temperature Vaporization (PTV) Cooled Injection System (CIS-4),
168 from Gerstel . The GC was an Agilent 7890A gas chromatograph coupled to an
169 Agilent 5975C quadrupole mass spectrometer detector (MS, Agilent Technologies,
170 Palo Alto, CA, USA) equipped with a DB-WAX column (J&W 122-7032; 30 m length,
171 0.25 mm I.D. and 0.25 μ m film thickness). The carrier gas was helium with a
172 constant column pressure of 20.75 psi.

173 Stir bars were thermally desorbed using helium as carrier gas at a flow rate of
174 75 mL/min; TDU was programmed from 40 °C to 295 °C (5 min) at a rate of 60
175 °C/min for PDMS and from 40 °C to 220 °C (5 min) at a rate of 60 °C/min for EG
176 Silicone and PA coatings, following the supplier's recommendations. The analytes
177 were focused on the CIS-4, containing a packed liner (20 mg of Tenax®), at two
178 different temperatures: 25 °C and 0 °C with liquid CO₂ cooling prior to injection.

179 After desorption and focusing, the CIS-4 temperature was programmed to reach
180 260 °C (5 min) at 12 °C/s to transfer volatiles onto the analytical column. The stir
181 aliquot of volatiles still sorbed on the the stir bar after desorption and rinsing
182 ('memory effect') was evaluated both at 0 and at 25 °C.

183 The TDU operated in the splitless desorption mode, the CIS-4 operated in PTV
184 solvent vent mode (purge flow to split vent of 80 mL/min, vent 75 mL/min and
185 pressure 20.85 psi). The GC oven temperature was programmed at 40 °C (2 min),
186 raised to 150 °C (10 °C/min, held for 5 min) then to 230 °C (10 °C/min, held 2
187 min). The MS operated in scan mode (27–300 amu) with ionization energy set at
188 70 eV. The temperature of the MS transfer line was 230 °C. Each molecule was
189 identified by comparing mass-spectrum with those of the NIST library and
190 chromatographic retention index of each standard. To avoid matrix interferences,
191 the MS quantification was carried out in the single ion monitoring mode using each
192 molecule characteristic m/z value (Table 1).

193 The HS- SBSE-GC/MS analysis were performed in sevenfold.

194

195 *2.5. Statistical Analysis*

196 Means and standard errors of the seven analysed replicates were calculated.
197 Data were statistically treated with SPSS (Version 19.0 statistical package for
198 Windows; SPSS, Chicago, IL, USA) and processed using the analysis of variance
199 (ANOVA). Statistical differences among means were evaluated using the Duncan's
200 test ($P \leq 0.05$). Normalized data underwent a discriminant analysis, using the
201 coating type as differentiating variable.

202

203 **3. Results and Discussion**

204 Each compound relative area was calculated as the ratio between its response area
205 and the response area of the internal standard after HS-SBSE-GC/MS analysis,
206 setting the CIS at 0 °C and 25 °C (Tables 2 and 3). Standard errors were very low,
207 regardless the type of Twister® coating used, demonstrating high precision and
208 reproducibility of the methodology.

209 Compounds whose Log K_{ow} was near 3 are the most apolar (Table 1), so it would
210 be expected that they were more retained by the PDMS due to its nonpolar phase;
211 the other two coatings (more polar) were expected to exhibit higher affinity for
212 compounds with Log K_{ow} around 1.

213 *CIS temperature at 0 °C.*

214 When the CIS temperature was set at 0 °C (Table 2), EG Silicone coating showed a
215 behaviour similar to PDMS for (*E*)-2-hexenal and (*E*)-2-hexen-1-ol. On the
216 contrary, more apolar compounds showed lower relative areas, whereas more
217 polar compounds such as benzaldehyde, 1-hexanol and (*Z*)-3-hexen-1-ol showed
218 higher values with a higher percentage of recovery in EG Silicone respect to PDMS
219 coatings.

220 PA coating showed lower relative areas for most compounds respect to PDMS;
221 only for 1-hexanol similar relative areas in PA and PDMS coatings were obtained.
222 Benzaldehyde, (*E*)-2-hexen-1-ol and (*Z*)-3-hexen-1-ol were the only compounds
223 showing an opposite trend, as they displayed higher relative areas in PA respect to
224 PDMS; so, the highest percentages of recovery were detected. Similarly to EG
225 Silicone coating, also in PA coating a relation between the percentages of recovery
226 and the compound polarities was detected, as higher relative areas respect to
227 PDMS were detected for compounds with lower LogK_{ow}.

228 *CIS temperature at 25 °C.*

229 Comparing EG Silicone and PDMS coatings with the CIS temperature set at 25 °C
230 (Table 3) resulted in significant differences for most compounds except for (*E*)-2-
231 hexenal, D-limonene and linalool. Higher relative areas and percentage recoveries
232 were detected for benzaldehyde, 1-hexanol, (*E*)-2-hexen-1-ol and (*Z*)-3-hexen-1-ol;
233 lower values were detected for all remaining compounds. Again, it emerged that
234 compounds with lower LogK_{ow} were more efficaciously sorbed onto more polar
235 coatings.

236 *Comparison of the CIS temperatures.*

237 With PDMS coating, individual compound relative areas were always
238 significantly higher when the CIS was set at 25 °C respect to 0 °C, except for D-
239 limonene (Table 3). For EG Silicone stir bars, setting the CIS at 25 °C increased the
240 relative area values for all compounds except for α -terpineol. For PA, setting the
241 CIS at 25 °C increased the relative area values of 1-hexanol, (*E*)-2-hexenal, (*E*)-2-
242 hexen-1-ol, (*Z*)-3-hexen-1-ol and of α -ionone. However, areas were lower respect
243 to those obtained with PDMS.

244 Generally, for all three types of coatings, standard errors were always lower when
245 the CIS temperature was set at 25 °C.

246 Compounds with Log K_{ow} between 1.58 and 1.82 were more efficaciously sorbed
247 by EG Silicone and PA coatings even though the standard errors were always
248 higher, regardless the CIS temperature. For the other studied compounds, PDMS
249 showed higher relative areas.

250 *Area detection and quantification limits.*

251 We calculated the limits of the relative detection area and of the relative
252 quantification area for each compound (ADL and AQL, respectively) detected by
253 using the three coatings operating with the two CIS temperatures (Table 4); to do

254 that we used a procedure similar to the one used to calculate detection and
255 quantification limits when working with concentrations. ADL and AQL were
256 calculated based on a signal to noise ratio of 3 and 10. Using areas is less
257 appropriate respect to using concentrations; however, we deem preliminary
258 information reported in this study useful and informative. In fact, a number of
259 recent studies reported analytical areas from SBSE-GC/MS analysis for specific
260 compounds (Ochiai, Sasamoto, Ieda, David & Sandra, 2013; Gilart, Miralles, Mercé,
261 Borrull & Fontanals, 2013; Gilart, Mercé, Borrull & Fontanals, 2014). The necessity
262 to report information expressed in areas rather than in concentrations is tied to
263 the lack of standards of many molecules and/or to their high costs.

264 ADL values calculated when the CIS was set at 0 °C ranged between 0.53×10^{-6}
265 (nerolidol, EG Silicone) and 12.83×10^{-6} (D-limonene, PDMS); AQL values for the
266 same temperature between 2.05×10^{-6} (nerolidol, PA) and 35.27×10^{-6} (α -terpineol,
267 PDMS).

268 When the CIS temperature was set at 25 °C ADL values ranged between
269 0.97×10^{-6} (nerolidol, PA) and 8.29×10^{-6} (1-hexanol, PA), whereas AQL values from
270 3.77×10^{-6} (nerolidol, EG Silicone) to 27.62×10^{-6} (1-hexanol, PA). The area limit
271 values for the identification and quantification of these compounds show an
272 adequate sensitivity for the two CIS temperatures and the three coatings. However,
273 we consider that the two tested coatings (Table 2 and 3) do not offer important
274 improvements of analytical results respect to PDMS, except for benzaldehyde, 1-
275 hexanol, (E)-2-hexenal, (E)-2-hexen-1-ol, (Z)-3-hexen-1-ol, particularly when the
276 CIS was set at 25 °C. The discriminant analysis (93.9 % of explained variance on
277 function 1 and 6.1 on function 2, Figure 1) supported these conclusions: the
278 centroid distances for CIS 25 °C and CIS 0 °C are the shortest, meaning that results

279 obtained with the PDMS are very similar, regardless the CIS temperature. On the
280 contrary, PA centroids are the most distant and EG Silicone presented an
281 intermediate situation. We can conclude that using the CIS at 0 °C or at 25 °C
282 brought to quite different analytical results when EG Silicone and PA coatings were
283 used.

284 *Stir bar memory effect*

285 Another important aspect in HS-SBSE-GC/MS analysis is to show that
286 compounds are as much as possible desorbed from the stir bar after analysis to
287 avoid the memory effect. EG Silicone and PA coatings showed a higher memory
288 effect for 1-hexanol, (*Z*)-3-hexen-1-ol, (*E*)-2-hexen-1-ol, linalool, α -terpineol and
289 nerolidol respect to PDMS coatings at both CIS temperatures (Figure 2). This
290 higher memory effect could be due to the lower desorption temperature that EG
291 Silicone and PA coatings require respect to PDMS (220 °C the former two, 295 °C
292 the latter).

293

294 **4. Conclusions**

295 SBSE used in the HS mode allowed a highly reliable and reduced time-consuming
296 detection of some wine volatiles of grape origin. EG-Silicone and Polyacrylate,
297 being more polar respect to PDMS, resulted to be more efficient in the
298 measurement of polar volatiles, with Log K_{ow} minor than 1.82, particularly when
299 the CIS temperature was set at 25 °C. As CIS temperature set at 25 °C always
300 allowed the detection of larger volatile relative areas, regardless the coating type,
301 we deem not necessary to use a forced cryo-focalization at very low temperatures
302 for the assessment of the volatiles tested in this work. This is an important point

303 to focus on for SBSE-GC/MS analysis as cryo-focusing, particularly when
304 performed by liquid CO₂ is expensive for the laboratory.

305

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310

311 **References**

312

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373

374 **Figure 1.** Discriminant analysis between the three type of coatings (PDMS, EG Silicone
375 and PA) and the two CIS temperatures (0 °C and 25 °C).

376

377 **Figure 2.** Overlay of HS-SBSE-GC-MS chromatograms obtained after a second
378 injection of each Twister® in the Total Ion Chromatogram (TIC) mode. A = CIS
379 temperature at 0 °C; B = CIS temperature at 25 °C. Some molecules showed higher
380 memory effects in EG Silicone and PA Twister® respect to PDMS. 1, 1-hexanol; 2, (Z)-
381 3-hexen-1-ol; 3, (E)-2-hexen-1-ol; 4, linalool; 5, α -terpineol; 6, nerolidol.