

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Methods for and Implementation of Pregnancy Diagnosis in Dairy Cows

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1596973> since 2016-09-26T13:15:45Z

Published version:

DOI:10.1016/j.cvfa.2015.09.006

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

Methods for and Implementation of Pregnancy Diagnosis in Dairy Cows

Paul M. Fricke, PhD^{a,*}, Alessandro Ricci, DVM^b,
Julio O. Giordano, PhD^c, Paulo D. Carvalho, MS^a

KEYWORDS

Transrectal palpation Transrectal ultrasonography Pregnancy loss Progesterone Pregnancy-associated glycoproteins

KEY POINTS

Although coupling a nonpregnancy diagnosis with a management strategy to quickly re-initiate artificial insemination (AI) may improve reproductive efficiency by decreasing the interval between AI services, early pregnancy loss limits the accuracy of many direct and indirect methods for early pregnancy diagnosis currently under development. These limitations make the benefits of many currently available methods for early pregnancy diagnosis questionable and require that all cows diagnosed pregnant early after insemination be scheduled for pregnancy reconfirmations at later times during gestation to identify cows experiencing pregnancy loss. Although research and development efforts are being made toward development of an in-direct pregnancy test for dairy cows, it remains to be seen whether these indirect tests will replace transrectal palpation or transrectal ultrasonography as the primary methods used for pregnancy diagnosis in dairy cows or whether veterinarians will combine these methods in a reproductive management program. Future technologies for pregnancy diagnosis in dairy cows may someday overcome current limitations of direct and indirect methods for pregnancy diagnosis, thereby improving reproductive performance.

ATTRIBUTES OF THE IDEAL PREGNANCY TEST

An ideal early pregnancy test for dairy cows would fulfill the following criteria:

1. High sensitivity (ie, correctly identify pregnant animals)
2. High specificity (ie, correctly identify nonpregnant animals)
3. Inexpensive to conduct
4. Simple to conduct under field conditions
5. Ability to determine pregnancy status at the time the test is performed

A final attribute of an ideal early pregnancy test would be the ability to determine pregnancy status without the need to physically handle the cow to conduct the test. Such a test may overcome the inherent limitations of current tests caused by pregnancy loss and may make pregnancy diagnosis before 28 to 35 days postpartum in dairy cows an economically viable reproductive management strategy. Although all of the methods described in this article require physical handling of individual cows to administer the test, future technologies for early pregnancy diagnosis may someday realize all of these criteria.

From an economic perspective, the sensitivity of an early nonpregnancy test (ie, correct identification of pregnant cows) is more important than the specificity (ie, correct identification of nonpregnant cows) based on an economic simulation.¹ Inaccurate diagnosis of nonpregnancy (ie, false negatives), however, increases the rate of iatrogenic pregnancy loss when prostaglandin F_{2a} (PGF_{2a}) or one of its analogues is administered to synchronize estrus or ovulation to reduce the interval to the next artificial insemination (AI) service. The economic loss incurred because of

pregnancy loss depends on many factors and has been estimated to range from \$462 to \$300.³ Because a management intervention can only be implemented for nonpregnant cows, it is critical that a pregnancy test accurately identify nonpregnant cows to avoid iatrogenic pregnancy loss. Nonetheless, a high rate of false-positive results diminishes the usefulness and cost-effectiveness of an early pregnancy test by failing to present a management opportunity to return nonpregnant cows to AI service early after AI and potentially increasing the interval to the subsequent AI.

RETURN TO ESTRUS AS A DIAGNOSTIC INDICATOR OF PREGNANCY STATUS

Accurate identification of cows returning to estrus from 18 to 32 days after AI is the easiest and least costly method for determining nonpregnancy early after insemination. This assumption, however, is being challenged by new research and long-recognized reproductive problems. First, estrous detection efficiency is estimated to be less than 50% on most dairy farms in the United States.⁴ Only 51.5% of the eligible cows were detected in estrus and inseminated in a recent study in which detection of estrus was performed through continuous monitoring with activity tags after a previous insemination until pregnancy diagnosis 32 days after AI.⁵ Second, estrous cycle duration varies widely with a high degree of variability among individual cows.⁶ Finally, the high rate of pregnancy loss in dairy cows can increase the interval from insemination to return to estrus for cows that establish pregnancy early then undergo pregnancy loss later during gestation.⁷

PREGNANCY LOSS IN LACTATING DAIRY COWS

Pregnancy loss contributes to reproductive inefficiency because fertility assessed at any point during pregnancy is a function of both conception rate and pregnancy loss.⁸ Pregnancy loss can be monitored using a variety of methods, including measurement of milk progesterone concentration or

pregnancy-specific proteins, transrectal ultrasonography, and transrectal palpation. Since the widespread application of transrectal ultrasonography for reproductive research in cattle,⁹ many studies have reported rates of pregnancy loss during early gestation under field conditions. In a summary of 14 studies,¹⁰ pregnancy loss from 27 to 31 and 38 to 50 days of gestation averaged 13% based on transrectal ultrasonography. Vasconcelos and colleagues¹¹ characterized pregnancy loss at various stages of gestation using transrectal ultrasonography and reported pregnancy losses of 11% from 28 to 42 days, 6% from 42 to 56 days, and 2% from 56 to 98 days after AI (Fig. 1), supporting that the rate of loss is greater early during gestation and then decreases as gestation proceeds. It has long been accepted that pregnancy status should be determined in dairy cows as soon as possible after insemination but without having the diagnosis confounded by subsequent pregnancy loss.^{12,13} Pregnancy loss diminishes the benefit of early pregnancy diagnosis. Because of the high rate of pregnancy loss that occurs around the gestational period that most direct and indirect pregnancy tests are performed, the magnitude of pregnancy loss observed is greater the earlier after breeding that a positive diagnosis is made. Thus, the earlier that pregnancy is diagnosed after insemination, the fewer nonpregnant cows are identified to which a management strategy can be implemented to reinseminate them. If left unidentified, cows diagnosed pregnant early after insemination that subsequently undergo pregnancy loss decreases reproductive efficiency by extending the interval from calving to the insemination that results in a full-term pregnancy. To compensate for pregnancy loss, cows diagnosed pregnant early after insemination must undergo one or more subsequent pregnancy examinations to identify and reinseminate cows that experience pregnancy loss. Most dairy farms

conduct an early nonpregnancy diagnosis around 28 to 35 days after AI and then reconfirm pregnancies.

for cows diagnosed pregnant around 4 to 6 weeks later and around dry off to identify cows that have lost pregnancies. For many herds, particularly those with low estrus detection efficiency, pregnancy reconfirmation is critical to reinseminate cows that undergo pregnancy loss. Problems caused by pregnancy loss apply to all currently available methods for assessing pregnancy status early after breeding and may make pregnancy testing before 25 days after insemination impractical unless pregnancy diagnosis can be made continually and cost-effective on a daily basis or at each milking until the rate of pregnancy loss decreases or until the underlying causes of pregnancy loss are understood and mitigated.

DIRECT METHODS FOR PREGNANCY DIAGNOSIS

By definition, direct methods for early pregnancy diagnosis involve direct detection of the tissues and/or associated fluids of the conceptus either manually or via electronic instrumentation. Currently used direct methods for diagnosis of pregnancy include transrectal palpation and B-mode ultrasonography. Technical expertise, operator proficiency, and the stage after insemination that the technique is performed can affect the specificity and sensitivity of the test; however, experienced bovine practitioners can achieve high sensitivity and specificity with either method.

Transrectal palpation

Transrectal palpation of the uterus for pregnancy diagnosis in cattle was first described in the 1800s¹⁴ and is the oldest and most widely used direct method for early pregnancy diagnosis in dairy cows. Transrectal palpation of the amniotic vesicles as an aid in determining pregnancy status in dairy cows was described by Wisnicky and Cassida,¹⁵ whereas slipping of the chorioallantoic membranes between the thumb and forefinger

beginning on about 30 days in gestation was described by Zemjanis.¹⁶ Because pregnancy in cows can be intentionally terminated by manual rupture of the amniotic vesicle,^{17,18} several studies have investigated the extent of iatrogenic pregnancy loss induced by transrectal palpation. Examining pregnant cows early in gestation by transrectal palpation has been reported to increase the risk of iatrogenic pregnancy loss in some studies,^{19–23} whereas other studies have reported that cows submitted for transrectal palpation had a decreased risk for pregnancy loss or that palpation had no effect on subsequent pregnancy loss.^{12,24} Although controversy still exists regarding the extent of iatrogenic pregnancy loss induced by transrectal palpation, other factors have a greater influence on calving rates than pregnancy examination using transrectal palpation.²⁵ Because of its widespread use and the number of bovine practitioners trained to perform the procedure, transrectal palpation will likely remain a popular method for pregnancy diagnosis in dairy cows until newer direct or indirect methods for pregnancy diagnosis are developed and adopted. Furthermore, because of its widespread use, high accuracy, and low cost per cow, transrectal palpation is the standard that newer direct and indirect methods for pregnancy diagnosis in dairy cows must displace as the method of choice for pregnancy diagnosis.

B-Mode Ultrasonography

Applications of and detailed methods for performing transrectal ultrasonography for reproductive research have been extensively reviewed and described elsewhere.^{8,9,26} Although early pregnancy diagnosis is among the most practical application for reproductive management using transrectal ultrasonography, additional information gathered using the technology that may be useful for reproductive management include evaluation of ovarian structures, identification of cows carrying twin fetuses, and determination of fetal sex.⁸ Recently, changes in endometrial

thickness using transrectal ultrasonography near the time of AI were reported to be a good indicator of ovulation failure and pregnancy success.²⁷ Transrectal ultrasonography has not been implicated as a direct cause of pregnancy loss in cows,^{28,29} and ultrasound is a less invasive technique for early pregnancy diagnosis than is transrectal palpation.^{21,22} As a pregnancy diagnosis method, transrectal ultrasonography is accurate and rapid; the outcome of the test is known immediately at the time the test is conducted. Transrectal ultrasonography has begun to displace transrectal palpation as the direct method of choice by veterinarians for pregnancy diagnosis.³⁰ Because many experienced bovine practitioners can accurately diagnose pregnancy as early as 35 days after insemination using transrectal palpation, pregnancy examination using transrectal ultrasonography 28 to 34 days after insemination only reduces the interval from insemination to pregnancy diagnosis by a few days. Although ultrasound conducted at 45 or more days after breeding did not increase the accuracy of pregnancy diagnosis for an experienced palpator, it may improve diagnostic accuracy of a less experienced one.³¹ The rate of pregnancy loss and the efficacy of strategies to reinseminate cows at various stages after breeding also play a role in determining the advantages and disadvantages on the timing of pregnancy diagnosis and resynchronization.³² Another potential benefit of transrectal ultrasonography over transrectal palpation is the opportunity to more accurately determine the ovarian status of cows at a nonpregnancy diagnosis facilitating the assignment of cows to different treatment alternatives. For example, use of an Ovsynch protocol for resynchronization of cows identified not pregnant 32 days after AI resulted in greater conception rates when cows were identified with a corpus luteum (CL) compared with cows without a CL at the first gonadotropin-releasing hormone (GnRH) treatment of the protocol.^{33,34}

Treatment of cows without a CL at the first GnRH treatment of an Ovsynch protocol with exogenous progesterone (ie, a intravaginal progesterone insert) increased fertility at first as well as resynch timed AI in lactating dairy cows.^{35,36} Treatment of cows with a CL of 20 mm or greater at nonpregnancy diagnosis with a PGF_{2a} injection increased the overall proportion of cows inseminated after a detected estrus for second and subsequent AI services.⁵ Based on these data, many veterinarians now use the presence or absence of a CL at a nonpregnancy diagnosis to improve outcomes to timed AI protocols used to resynchronize nonpregnant cows or to increase the proportion of cows inseminated in estrus after a previous insemination.

Problems with Early Pregnancy Diagnosis Using Transrectal Ultrasonography

Early studies in which transrectal ultrasonography was used to assess embryonic development in vivo reported that a fetal heartbeat could be visualized at around 21 days in gestation under controlled experimental conditions and using a high-quality scanner and transducer.³⁷ Several studies reported that pregnancy diagnosis can be rapidly and accurately diagnosed using ultrasound as early as 26 days after AI.^{38,39} A recent report evaluated using transrectal ultrasonography as early as 18 to 21 days after insemination in Irish Holstein Friesian dairy cows.⁴⁰ Because of these reports, many bovine practitioners focused on pushing the lower limit of early pregnancy diagnosis to conduct pregnancy diagnosis using transrectal ultrasonography. Use of transrectal ultrasonography before about 30 days after insemination under field conditions on a commercial dairy, however, can negatively affect the accuracy of pregnancy diagnosis outcomes.⁴¹

To determine the accuracy of early pregnancy diagnosis using transrectal ultraso-nography, we conducted a field trial on a commercial dairy farm milking approximately 2000 cows.⁴¹ Pregnancy status was determined 29 days after timed AI using transrec-tal ultrasonography (Easi-scan, BCF Technology Ltd, Rochester, MN) based on the following criteria: presence or absence of a CL; presence, absence, volume, and appearance of uterine fluid typical for a 29-day conceptus; presence or absence of an embryo with a heartbeat. Cows were classified as (1) not pregnant: presence or absence of a CL, absence of uterine fluid or insufficient uterine fluid, and absence of an embryo; (2) pregnant: CL present, normal uterine fluid, and no embryo; (3) preg-nant embryo: CL present, normal uterine fluid, and at least one embryo visualized; and (4) questionable pregnant: CL present and one or more of the following: uterine fluid, insufficient uterine fluid, and either no embryo or a nonviable embryo. At 39 and 74 days after timed AI, pregnancy status was determined using transrectal palpation and pregnancy loss occurring between each pregnancy examination was calculated. Results from this experiment are shown in [Table 1](#). Overall, 802 cows were classi-fied as not pregnant 29 days after timed AI, whereas 799 cows were classified as not pregnant 39 days after timed AI resulting in a not-pregnant misdiagnosis rate of 0.5% (4 of 802) for transrectal ultrasonography 29 days after timed AI. At 29 days after timed AI, 1116 cows were classified as either pregnant with an embryo visualized (68%), pregnant based on uterine fluid alone (29%), or questionable pregnant (3%). Among questionable pregnant cows, 69% were classified as not pregnant 39 days after timed AI and an additional 46% were classified as not pregnant 74 days after timed

AI. For cows classified pregnant 29 days after timed AI, more ($P<0.01$) cows diagnosed based on uterine fluid only than fluid and the presence of an embryo were classified as not pregnant using transrectal palpation 39 days after timed AI. Similarly, more ($P<0.01$) cows diagnosed pregnant based on uterine fluid alone than cows diagnosed pregnant based on visualization of an embryo with a heartbeat were classified as not pregnant

Table 1
 Pregnancy loss by pregnancy classification for lactating Holstein cows diagnosed pregnant using ultrasonography 29 days after timed AI

Item	Pregnancy Classification ^b		Question able
	Pregnant	Uterine Fluid	
	% (n/n)		
29 d after timed AI	68 (758 of 1116)	29 (322 of 1116)	3 (36 of 1116)
Pregnancy loss			
29–39 d	4a (30 of 758)	18a (57 of 322)	69a (25 of 36)
39–74 d	5a (39 of 728)	12a (32 of 265)	46a (5 of 11)

	9a (69 of		83a (30
Total loss	758)	28a (89 of 322)	of 36)

a Within a row, proportions with different superscripts differ ($P < .001$).

b Lactating Holstein cows diagnosed pregnant were classified based on the following criteria using transrectal ultrasonography: pregnant: visualization of a CL ipsilateral to the gravid uterine horn, visualization of an amount of nonechogenic uterine fluid in accordance to stage of pregnancy, and visualization of an embryo with a heartbeat; uterine fluid: visualization of a CL ipsilateral to the gravid uterine horn, visualization of an amount of nonechogenic uterine fluid in accordance to stage of pregnancy but without visualization of the embryo; questionable: visualization of a CL ipsilateral to the gravid uterine horn with insufficient uterine fluid for the stage of pregnancy.

Adapted from Giordano JO, Fricke PM. Accuracy of pregnancy diagnosis outcomes using trans-rectal ultrasonography 29 days after artificial insemination in lactating dairy cows. *J Dairy Sci* 2012;95(Suppl 2):75; with permission.

using transrectal palpation 74 days after timed AI. From the initial pregnancy examination at 29 days to the last examination 74 days after timed AI, more ($P < 0.01$) cows diagnosed pregnant based on uterine fluid alone than cows diagnosed pregnant based on visualization of an embryo with a heartbeat were classified as not pregnant using transrectal palpation 74 days after timed AI. Cows classified pregnant based on uterine fluid alone 29 days after timed AI were 3.8 (95% confidence interval 2.7–5.4) times more

likely to be classified as not pregnant 74 days after timed AI than cows diagnosed pregnant based on visualization of an embryo with a heartbeat. Based on these data, the authors concluded that the accuracy of pregnancy outcomes using transrectal ultrasonography increase dramatically when an embryo with a heartbeat is visualized compared with outcomes based only on the presence of a CL and the volume of uterine fluid in the absence of a visualized embryo with a heartbeat. The presence of a large proportion of cows with a CL and fluid was visualized in the absence of an embryo with a heartbeat is likely due to a high degree of early pregnancy loss in dairy cows. In 2 experiments, 35% to 44% of dairy cows diagnosed not pregnant 32 days after timed AI had extended luteal phases.^{7,42} Based on the authors' results, early pregnancy diagnosis should not be conducted earlier than an embryo with a heartbeat can be rapidly and reliably detected in pregnant cows under on-farm conditions using transrectal ultrasonography (w30 days after AI) to reduce the negative impact of false-positive results.

INDIRECT METHODS FOR PREGNANCY DIAGNOSIS IN DAIRY COWS

Indirect methods for early pregnancy diagnosis use qualitative or quantitative measures of hormones or conceptus-specific substances in maternal body fluids as indirect indicators of the presence of a viable pregnancy. Commercially available indirect methods for pregnancy diagnosis in dairy cows include milk progesterone tests and tests for pregnancy-associated glycoproteins (PAGs) in blood or milk.

Progesterone

Progesterone is the most biologically active progestagen in cattle and is primarily produced and secreted by the corpus luteum during the estrous cycle and the placenta during pregnancy. Quantification of progesterone in blood or milk can be achieved in a laboratory using radioimmunoassay (RIA) or enzyme-linked immunosorbent assay (ELISA) methods. The biology of early pregnancy and maintenance of the CL results in distinct progesterone profiles for pregnant compared with nonpregnant cows. Lactating dairy cows were synchronized for first timed AI, and resulting progesterone profiles based on thrice weekly (Monday, Wednesday, Friday) blood sampling are shown in Fig. 2. The upper panel of Fig. 2 indicates a cow that failed to become pregnant and had a normal luteal phase followed by a subsequent estrous cycle. By contrast, the middle panel of Fig. 2 indicates a cow that maintained pregnancy. The lower panel of Fig. 2 is representative of cows that fail to maintain a pregnancy and had an extended luteal phase. Extended luteal phases are common in dairy cows after AI. In one experiment, 35% of dairy cows diagnosed not pregnant 32 days after timed AI had extended luteal phases⁴²; in another experiment, the proportion of cows with extended luteal phases was 44%.⁷ Unfortunately, sequential sampling of milk or blood for determination of progesterone using RIA or ELISA methods is not practical or cost-effective for use on commercial dairy farms. Future technologies to monitor milk progesterone profiles of individual cows on a daily or even a weekly basis could revolutionize reproductive management strategies for dairy cows. 172

Rapid on-farm qualitative tests for assessing progesterone levels in milk were commercialized for pregnancy diagnosis in dairy cows in the 1980s,⁴³ and a few remain commercially available today. Manufacturers recommended these tests be conducted 18 to 24 days after insemination to determine pregnancy status. Based

on the progesterone profiles in Fig. 2, cows with low progesterone 18 to 24 days after AI would be classified as not pregnant, whereas cows with high progesterone 18 to 24 days after AI would be classified as pregnant. Although not-pregnant outcomes are highly accurate for identifying cows that truly are not pregnant, the accuracy of high progesterone 18 to 24 days after AI for accurately diagnosing pregnant cows is poor. This poor accuracy is due to the biology associated with pregnancy loss that confounds early pregnancy diagnosis using transrectal ultrasonography discussed previously. Most of these extended luteal phases may be explained by cows that establish a pregnancy early by signaling maternal recognition of pregnancy and maintenance of the CL past the normal time of luteal regression but then subsequently undergo pregnancy loss.⁷ Thus, although future technologies may allow for on-farm sampling of milk progesterone, the use of cow-side milk progesterone tests conducted 18 to 24 days after insemination should focus on identifying not-pregnant cows rather than pregnant cows.

Pregnancy-Associated Proteins

Proteins produced and secreted by the placenta early during pregnancy are obvious candidates for development of an early pregnancy test; however, proteins produced by the placenta vary widely among eutherotic mammals. For example, only the higher primates produce a chorionic gonadotropin homologous to the human protein (human chorionic gonadotropin) required for luteal support early during pregnancy, whereas only ruminant ungulates are known to produce type I interferon as an antiluteolytic hormone.⁴⁴ Because cattle do not produce a chorionic gonadotropin, research has focused on discovery and characterization of

pregnancy-specific proteins suitable for determining pregnancy status in cattle early after insemination. Some pregnancy-associated factors, such as the early conception factor, have not proven to be accurate in dairy cows.⁴⁵ It is now possible to detect a viable conceptus between 15 to 22 days after AI by measuring the expression of interferon-stimulated genes in circulating white blood cells^{46–48}; however, this method has not yet been commercialized. The most recent advance in this area has been made in the commercialization of tests for PAGs.

Pregnancy-Associated Glycoproteins

Bovine PAGs were discovered through attempts to develop indirect early pregnancy tests in dairy cows.⁴⁴ In 1982, two proteins, pregnancy-specific protein (PSP) A and B, were isolated from bovine fetal membrane extracts.⁴⁹ Development of a specific RIA for PSPB⁵⁰ allowed for quantification of PSPB in maternal serum as an indirect method for pregnancy diagnosis and pregnancy loss in dairy cows.^{51,52} Molecular cloning and sequencing studies revealed that PAGs belong to a large family of inactive aspartic proteinases expressed by the placenta of domestic ruminants, including cows, ewes, and goats.⁵³

In cattle, the PAG gene family comprises at least 22 transcribed genes as well as some variants.⁵⁴ Bovine PAGs have been immunologically localized to trophoblast binucleate cells present in fetal cotyledonary villi and to a lesser extent to caruncular epithelium.⁵⁵ Migration of binucleate cells from the trophoctoderm to the uterine epithelium allows for exocytosis of granules containing PAG into the maternal circulation.⁵⁶ Because cellular products of binucleate cells are released into maternal circulation, the ideal antigen for an indirect early pregnancy test in dairy cows would be a PAG expressed in

binucleate cells around the time of implantation.⁵⁷ Mean PAG concentrations in cattle increase from 15 to 35 days in gestation⁴²; however, variation in plasma PAG levels among cows precludes PAG testing as a reliable indicator of pregnancy until about 26 to 30 days after AI.^{58,59} Several experiments have evaluated the use of commercial PAG tests to determine pregnancy status in dairy cows and heifers (Table 2). The sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and accuracy obtained using PAG tests are summarized based on several experiments^{58,60–66} in Table 2. Few studies have compared factors associated with PAG levels in blood and milk of dairy cows early in gestation and the impact these factors may have on the accuracy of pregnancy diagnosis. The authors recently conducted an experiment to determine the factors affecting PAG levels in blood and milk of dairy cows during early gestation.⁶⁶ Lactating Holstein cows (n = 141) were synchronized to receive their first timed artificial AI. Blood and milk samples were collected 25 and 32 days after timed AI (TAI), and pregnancy status was determined 32 days after TAI using transrectal ultrasonography. Cows diagnosed pregnant with singletons (n = 48) continued the experiment in which blood and milk samples were collected, and pregnancy status was assessed weekly using transrectal ultrasonography from 39 to 102 days after TAI. Plasma and milk samples were assayed for PAG levels using commercial ELISA kits.

To evaluate pregnancy outcomes from the plasma and milk PAG tests in cows of unknown pregnancy status, 2 by 2 contingency tables were constructed to calculate sensitivity, specificity, PPV, NPV, and accuracy of the pregnancy outcomes for the plasma and

milk PAG tests 32 days after timed AI; these outcomes were compared with those based on transrectal ultrasonography 32 days after timed AI (Table 3). Sensitivity for both the plasma and milk PAG tests in the present experiment was high (100% and 98%, respectively) compared with specificity (87% and 83%, respectively). As a result, the NPV for the plasma and milk PAG tests in the present experiment was high (100% and 99%, respectively) compared with the PPV of both tests (84% and 79%, respectively). The overall accuracy of the plasma and milk PAG tests 32 days after timed AI was 92% and 89%, respectively. Results from this sensitivity analysis support that the accuracy of using plasma or milk PAG levels as an indicator of pregnancy status in dairy cows 32 days after AI is high, and the authors' results agree with others who have conducted similar analyses from 27 to 39 days in gestation when PAG levels in both plasma and milk are at early peak levels.^{61,64,65}

The incidence of pregnancy loss in the present study for cows diagnosed with singleton pregnancies 32 days after TAI during the experiment was 13% (7 of 55), which agrees with the 13% loss reported to occur from 27 to 31 and 38 to 50 days of gestation based on transrectal ultrasonography in a summary of 14 studies.¹⁰ For the plasma PAG ELISA, all but one cow that underwent pregnancy loss tested positive, whereas all cows undergoing pregnancy loss tested positive at one or more time points for the milk PAG test. Similarly, 5 of 7 cows tested recheck based on the plasma PAG test before the loss occurred compared with 3 of 7 cows based on the milk PAG test. Thus, PAG levels detected by these ELISA tests in the present study have a half-life in maternal circulation resulting in a 7- to 14-day delay in

identification of cows undergoing pregnancy loss based on plasma or milk PAG levels compared with transrectal ultrasonography.

Profiles of PAG in plasma and milk of cows that maintained pregnancy from 25 to 102 days in gestation are shown in [Fig. 3](#). Compared with transrectal ultrasonography, accuracy was 92% for the plasma PAG test and 89% for the milk PAG test 32 days after timed AI. Factors associated with PAG levels in dairy cows included stage of gestation, parity, pregnancy loss, and milk production. Based on plasma and milk PAG profiles, the optimal time to conduct a first pregnancy diagnosis is around 32 days after AI coinciding with an early peak in PAG levels. The authors concluded that because of the

Adapted from Ricci A, Carvalho PD, Amundson MC, et al. Factors associated with pregnancy-associated glycoprotein (PAG) levels in plasma and milk of Holstein cows during early pregnancy and their effect on the accuracy of pregnancy diagnosis. *J Dairy Sci* 2015;98:2502–14; with permission

Factors associated with pregnancy-associated glycoprotein (PAG) levels in plasma and milk of Holstein cows during early pregnancy and their effect on the accuracy of pregnancy diagnosis. *J Dairy Sci* 2015;98:2502–14; with permission.) occurrence of pregnancy loss, all pregnant cows should be retested 74 days after AI or later when plasma and milk PAG levels in pregnant cows have rebounded from their nadir.

FUTURE TECHNOLOGIES FOR PREGNANCY DIAGNOSIS

A novel approach to the problem of early pregnancy diagnosis in dairy cows would be to monitor a pregnancy-specific substance or hormone secreted in milk in sufficient quantities to be detected by an in-line milk-sensing device during normal milking periods on

a dairy. Obviously, this pregnancy-specific substance must first be discovered or a known marker must be used and the in-line milk sampling technology developed to accurately detect and monitor this substance. If sensitive and specific, such a system would have a minimal marginal cost per test once the initial capital outlay was made to install the equipment on the dairy. By using such a system, a pregnancy diagnosis would be conducted during each milking period for all lactating cows on a dairy so that nonpregnancy, pregnancy, and pregnancy loss could be continually monitored and tracked on a daily basis. Integration of this information into a computerized dairy management software system would allow dairy managers to review the pregnancy status of individual cows in the herd on a daily or weekly basis so that reproductive management strategies could be implemented to establish, maintain, or attempt to reinitiate a pregnancy. Finally, such a system would achieve the heretofore-unrealized characteristic of conducting the pregnancy test without having to handle the cow to administer the test. Limitations imposed by pregnancy loss during early gestation will not be overcome until such a system is developed.

SUMMARY

Although coupling a nonpregnancy diagnosis with a management strategy to quickly reinitiate AI may improve reproductive efficiency by decreasing the interval between AI services, early pregnancy loss limits the accuracy of many direct and indirect methods for early pregnancy diagnosis currently under development. These limitations make the benefits of many currently available methods for early pregnancy diagnosis questionable and require that all cows diagnosed pregnant early after

insemination be scheduled for pregnancy reconfirmations at later times during gestation to identify cows experiencing pregnancy loss. Although much research and development efforts are being made toward development of an indirect pregnancy test for dairy cows, it remains to be seen whether these indirect tests will replace transrectal palpation or transrectal ultrasonography as the primary method used for pregnancy diagnosis in dairy cows or whether veterinarians will combine these methods in a reproductive management program. Future technologies for pregnancy diagnosis in dairy cows may someday overcome current limitations of direct and indirect methods for pregnancy diagnosis, thereby improving reproductive performance.

REFERENCES

1. Giordano JO, Fricke PM, Cabrera VE. Economics of resynchronization strategies including chemical tests to identify nonpregnant cows. *J Dairy Sci* 2013;96: 949–61.
2. Ferguson JD, Galligan DT. The value of pregnancy diagnosis—a revisit to an old art. Annual Conference Symposium. Society of Theriogenology. Montgomery, AL; Milwaukee, WI. August 9–13, 2011.
3. Galligan DT, Ferguson J, Munson R, et al. Economic concepts regarding early pregnancy testing. In: Proceedings of the American Association of Bovine Practitioners. Omaha, NE; Auburn, AL. 2009. p. 48–53.
4. Senger PL. The estrus detection problem: new concepts, technologies, and possibilities. *J Dairy Sci* 1994;77:2745–53.
5. Giordano JO, Stangaferro ML, Wijma R, et al. Reproductive performance of dairy cows managed with a program aimed at increasing insemination of cows in estrus based on increased

- physical activity and fertility of timed artificial inseminations. *J Dairy Sci* 2015;98:2488–501.
6. Remnant JG, Green MJ, Huxley JN, et al. Variation in the interservice intervals of dairy cows in the United Kingdom. *J Dairy Sci* 2015;98:889–97.
 7. Ricci A, Carvalho PD, Amundson MC, et al. Characterization of luteal dynamics in lactating dairy cows for 32 days after synchronization of ovulation and timed artificial insemination. *J Dairy Sci* 2014;97(Suppl 1):693.
 8. Fricke PM. Scanning the future – ultrasonography as a reproductive management tool for dairy cattle. *J Dairy Sci* 2002;85:1918–26.
 9. Griffin PG, Ginther OJ. Research applications of ultrasonic imaging in reproductive biology. *J Anim Sci* 1992;70:953–72.
 10. Santos JEP, Thatcher WW, Chebel RC, et al. The effect of embryonic death rates in cattle on the efficacy of estrus synchronization programs. *Anim Reprod Sci* 2004;82–83:513–35.
 11. Vasconcelos JLM, Silcox RW, Lacerda JA, et al. Pregnancy rate, pregnancy loss, and response to heat stress after AI at two different times from ovulation in dairy cows [abstract]. *Biol Reprod* 1997;56(Suppl 1):140.
 12. Studer E. Early pregnancy diagnosis and fetal death. *Vet Med Small Anim Clin* 1969;64:613–7.
 13. Melrose DR. The need for, and possible methods of application of, hormone assay techniques for improving reproductive efficiency. *Br Vet J* 1979;135:453–9.
 14. Cowie TA. Pregnancy diagnosis tests: a review. Commonwealth agricultural bureaux joint publication No. 13, Oxford, UK; 1948. p. 11–7.

15. Wisnicky W, Cassida LE. A manual method for diagnosis of pregnancy in cattle. *J Am Vet Med Assoc* 1948;113:451.
16. Zemjanis R. Diagnostic and therapeutic techniques in animal reproduction. 2nd edition. Baltimore (MD): Williams and Wilkins; 1970. p. 29–45.
17. Ball L, Carroll EJ. Induction of fetal death in cattle by manual rupture of the am-niotic vesicle. *J Am Vet Med Assoc* 1963;142:373–4.
18. Lo´pez-Gatius F. The effect on pregnancy rate of progesterone administration after manual reduction of twin embryos in dairy cattle. *J Vet Med A Physiol Pathol Clin Med* 2005;52:199–201.
19. Abbitt BL, Ball G, Kitto P, et al. Effect of three methods of palpation for pregnancy diagnosis per rectum on embryonic and fetal attrition in cows. *J Am Vet Med As-soc* 1978;173:973–7.
20. Franco OJ, Drost M, Thatcher MJ, et al. Fetal survival in the cow after pregnancy diagnosis by palpation per rectum. *Theriogenology* 1987;27:631–44.
21. Paisley LG, Mickelsen WD, Frost OL. A survey of the incidence of prenatal mor-tality in cattle following pregnancy diagnosis by rectal palpation. *Theriogenology* 1978;9:481–9.
22. Vaillancourt D, Vierschwal CJ, Ogwu D, et al. Correlation between pregnancy diagnosis by membrane slip and embryonic mortality. *J Am Vet Med Assoc* 1979;175:466–8.
23. White ME, LaFaunce N, Mohammed HO. Calving outcomes for cows diagnosed pregnant or nonpregnant by per rectum examination at various intervals after insemination. *Can Vet J* 1989;30:867–70.
24. Thurmond MC, Picanso JP. Fetal loss associated with palpation per rectum to di-agnose pregnancy in cows. *J Am Vet Med Assoc* 1993;203:432–5.

25. Thompson JA, Marsh WE, Calvin JA, et al. Pregnancy attrition associated with pregnancy testing by rectal palpation. *J Dairy Sci* 1994;77:3382–7.
26. Ginther OJ. Ultrasonic imaging and animal reproduction: cattle. Book 3. Cross Plains (WI): Equiservices Publishing; 1998.
27. Souza AH, Silva EPB, Cunha AP, et al. Ultrasonographic evaluation of endometrial thickness near timed AI as a predictor of fertility in high-producing dairy cows. *Theriogenology* 2011;75:722–33.
28. Ball PJH, Logue DDN. Ultrasound diagnosis of pregnancy in cattle. *Vet Rec* 1994; 134:532.
29. Baxter SJ, Ward WR. Incidence of fetal loss in dairy cattle after pregnancy diagnosis using an ultrasound scanner. *Vet Rec* 1997;140:287–8.
30. Caraviello DZ, Weigel KA, Fricke PM, et al. Survey of management practices on reproductive performance of dairy cattle on large US commercial farms. *J Dairy Sci* 2006;89(12):4723–35.
31. Galland JC, Offenbach LA, Spire MF. Measuring the time needed to confirm fetal age in beef heifers using ultrasonographic examination. *Vet Med* 1994;89:795–804.
32. Fricke PM, Caraviello DZ, Weigel KA, et al. Fertility of dairy cows after resynchronization of ovulation at three intervals after first timed insemination. *J Dairy Sci* 2003;86:3941–50.
33. Giordano JO, Wiltbank MC, Guenther JN, et al. Increased fertility in lactating dairy cows resynchronized with Double-Ovsynch compared with Ovsynch initiated 32 d after timed artificial insemination. *J Dairy Sci* 2012;95:639–53.

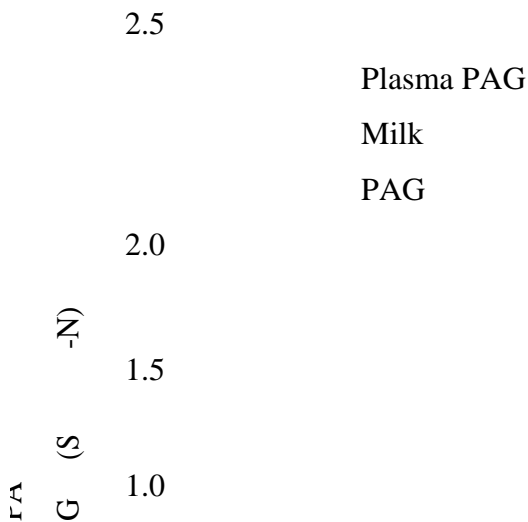
34. Lopes G Jr, Giordano JO, Valenza A, et al. Effect of timing of initiation of resynchro-nization and presynchronization with gonadotropin-releasing hormone on fertility of resynchronized inseminations in lactating dairy cows. *J Dairy Sci* 2013;96:3788–98.
35. Chebel RC, Al-Hassan MJ, Fricke PM, et al. Supplementation of progesterone via controlled internal drug release inserts during ovulation synchronization protocols in lactating dairy cows. *J Dairy Sci* 2010;93:922–31.
36. Bilby TR, Brune RGS, Lager KJ, et al. Supplemental progesterone and timing of resynchronization on pregnancy outcomes in lactating dairy cows. *J Dairy Sci* 2013;96:7032–42.
37. Curran S, Pierson RA, Ginther OJ. Ultrasonographic appearance of the bovine conceptus from days 20 through 60. *J Am Vet Med Assoc* 1986;189:1295–302.
38. Pieterse MC, Szenci O, Willemsse AH, et al. Early pregnancy diagnosis in cattle by means of linear-array real-time ultrasound scanning of the uterus and a qualitative and quantitative milk progesterone test. *Theriogenology* 1990;33:697–707.
39. Nation DP, Malmo J, Davis GM, et al. Accuracy of bovine pregnancy detection using transrectal ultrasonography at 28 to 35 days after insemination. *Aust Vet J* 2003;81:63–5.
40. Scully S, Butler ST, Kelly AK, et al. Early pregnancy diagnosis on days 18 to 21 postinsemination using high-resolution imaging in lactating dairy cows. *J Dairy Sci* 2014;97:3542–57.
41. Giordano JO, Fricke PM. Accuracy of pregnancy diagnosis outcomes using transrectal ultrasonography 29 days after artificial insemination in lactating dairy cows. *J Dairy Sci* 2012;95(Suppl 2):75.

42. Giordano JO, Guenther JN, Lopes G Jr, et al. Changes in serum pregnancy-associated glycoprotein, pregnancy-specific protein B, and progesterone concentrations before and after induction of pregnancy loss in lactating dairy cows. *J Dairy Sci* 2012;95:683–97.
43. Nebel RL. On-farm milk progesterone tests. *J Dairy Sci* 1988;71:1682–90.
44. Xie S, Green J, Bixby JB, et al. The diversity and evolutionary relationships of the pregnancy-associated glycoproteins, an aspartic proteinase subfamily consisting of many trophoblast-expressed genes. *Proc Natl Acad Sci U S A* 1997;94:12809–16.
45. Cordoba MC, Sartori R, Fricke PM. Assessment of a commercially available early conception factor (ECF) test for determining pregnancy status of dairy cattle. *J Dairy Sci* 2001;84:1884–9.
46. Stevenson JL, Dalton JC, Ott TL, et al. Correlation between reproductive status and steady-state messenger ribonucleic acid levels of the Myxovirus resistance gene, MX2, in peripheral blood leukocytes of dairy heifers. *J Anim Sci* 2007;85:2163–72.
47. Gifford CA, Racicot K, Clark DS, et al. Regulation of interferon-stimulated genes in peripheral blood leukocytes in pregnant and bred, nonpregnant dairy cows. *J Dairy Sci* 2008;90:274–80.
48. Green JC, Okamura CS, Poock SE, et al. Measurement of interferon-tau (IFN-t) stimulated gene expression in blood leukocytes for pregnancy diagnosis within 18–20 d after insemination in dairy cattle. *Anim Reprod Sci* 2010;121:24–33.

49. Butler JE, Hamilton WC, Sasser RG, et al. Detection and partial characterization of two bovine pregnancy-specific proteins. *Biol Reprod* 1982;26:925–33.
50. Sasser RG, Ruder CA, Ivani KA, et al. Detection of pregnancy by radioimmuno-assay of a novel pregnancy-specific protein in serum of cows and a profile of serum concentration during gestation. *Biol Reprod* 1986;35:936–42.
51. Humblot P, Camous S, Martal J, et al. Diagnosis of pregnancy by radioimmuno-assay of a pregnancy-specific protein in the plasma of dairy cows. *Theriogenology* 1988;30:257–68. Fricke et al.
52. Humblot P, Camous S, Martal J, et al. Pregnancy-specific protein B, progesterone concentrations and embryonic mortality during early pregnancy in dairy cows. *J Reprod Fertil* 1988;83:215–23.
53. Haugejorden G, Waage S, Dahl E, et al. Pregnancy associated glycoproteins (PAG) in postpartum cows, ewes, goats and their offspring. *Theriogenology* 2006;66:1976–84.
54. Prakash B, Telugu VL, Walker AM, et al. Characterization of the bovine pregnancy-associated glycoprotein gene family – analysis of gene sequences, regulatory regions within the promoter and expression of selected genes. *BMC Genomics* 2009;10:185–202.
55. Zoli AP, Demez P, Beckers JF, et al. Light and electron microscopic immunolocalization of bovine pregnancy-associated glycoprotein in the bovine placentome. *Biol Reprod* 1992;46:623–9.
56. Wooding FBP. Current topic: the synepitheliochorial placenta of ruminants: binucleate cell fusions and hormone production. *Placenta* 1992;13:101–13.

57. Green JA, Xie S, Quan X, et al. Pregnancy-associated bovine and ovine glycoproteins exhibit spatially and temporally distinct expression patterns during pregnancy. *Biol Reprod* 2000;62:1624–1631.
58. Zoli AP, Guilbault LA, Delahaut P, et al. Radioimmunoassay of a bovine pregnancy-associated glycoprotein in serum: its application for pregnancy diagnosis. *Biol Reprod* 1992;46:83–92.
59. Humblot P. Use of pregnancy specific proteins and progesterone assays to monitor pregnancy and determine the timing, frequencies and sources of embryonic mortality in ruminants. *Theriogenology* 2001;56:1417–33.
60. Szenci O, Beckers JF, Humblot P, et al. Comparison of ultrasonography, bovine pregnancy-specific protein B, and bovine pregnancy-associated glycoprotein 1 tests for pregnancy detection in dairy cows. *Theriogenology* 1998;50:77–88.
61. Silva E, Sterry RA, Kolb D, et al. Accuracy of a pregnancy-associated glycoprotein ELISA to determine pregnancy status of lactating dairy cows twenty-seven days after timed artificial insemination. *J Dairy Sci* 2007;90:4612–22.
62. Romano JE, Larson JE. Accuracy of pregnancy specific protein-B test for early pregnancy diagnosis in dairy cattle. *Theriogenology* 2010;74:932–9.
63. Piechotta M, Bollwein J, Friedrich M, et al. Comparison of commercial ELISA blood tests for early pregnancy detection in dairy cows. *J Reprod Dev* 2011; 57:72–5.
64. Sinedino LDP, Lima FS, Bisinotto RS, et al. Effect of early or late resynchronization based on different methods of pregnancy diagnosis on reproductive performance of dairy cows. *J Dairy Sci* 2014;97:4932–41.

65. Lawson BC, Shahzad AH, Dolecheck KA, et al. A pregnancy detection assay using milk samples: evaluation and considerations. *J Dairy Sci* 2014;97:6316–25.
66. Ricci A, Carvalho PD, Amundson MC, et al. Factors associated with pregnancy-associated glycoprotein (PAG) levels in plasma and milk of Holstein cows during early pregnancy and their effect on the accuracy of pregnancy diagnosis. *J Dairy Sci* 2015;98:2502–14.



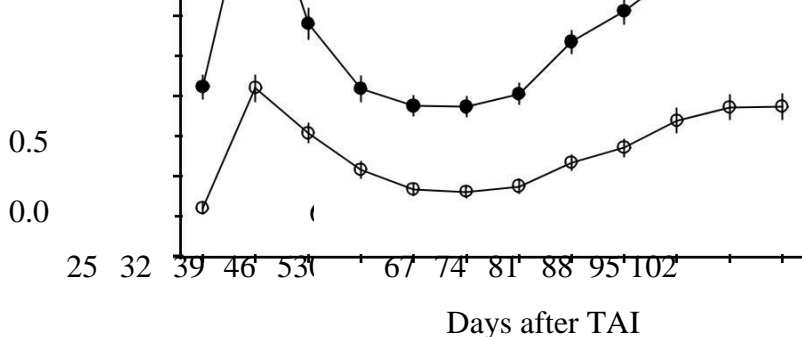


Fig. 3. Plasma and milk PAG profiles for Holstein dairy cows (n 5 48) that maintained pregnancy from 25 to 102 days after AI. ELISA outcomes were calculated from the optical density (OD) of the sample (corrected by subtraction of the reference wavelength OD of the sample [S] minus the OD of the negative control [N] at 450 nm with both values corrected by subtraction of the reference wavelength OD of the negative control), which resulted in an S-N value. Plasma and milk PAG levels were affected by week after AI (P<.01). (Adapted from Ricci A, Car-valho PD, Amundson MC, et al.)

Table 3

Sensitivity, specificity, PPV, NPV, and accuracy of plasma and milk PAG ELISA tests for determination of pregnancy status 32 days after AI

	PPV ^a %	NPV ^b %	Sensitivity c %	Specificity d %	Accuracy %
PAG					
ELISA	(No./No.)	(No./No.)	(No./No.)	(No./No.)	(No./No.)
Plasma	84 (57 of 68)	100 (73 of 73)	100 (57 of 57)	87 (73 of 84)	92 (130 of 141)
Milk	79 (52 of 66)	99 (68 of 69)	98 (52 of 53)	83 (68 of 82)	89 (120 of 135)

- a Proportion of cows diagnosed pregnant using the PAG ELISA that truly were pregnant.
- b Proportion of cows diagnosed as not pregnant using the PAG ELISA that truly were not pregnant.
- c Proportion of pregnant cows with a positive PAG ELISA outcome.
- d Proportion of not-pregnant cows with a negative PAG ELISA outcome.
- e Proportion of pregnancy status outcomes, pregnant and not pregnant, that were correctly classified by the PAG ELISA.

Table 2

Sensitivity, specificity, PPV, NPV, and accuracy for RIA and ELISA PAG test results

Reference	Days After AI	Test	Sensitivity ^a	Specificity ^b	PPV ^c	NPV ^d	Accuracy ^e
Zoli et al, ⁵⁸ 1992	22–30	RIA (blood PAG)	98.8	87.5	93.0	97.9	94.5
Szenci et al, ⁶⁰ 1998	26–58	RIA (PSPB)	75–100	85–92	81–91	80–100	80–96
		RIA (blood PAG)	81–100	57–71	62–74	78–100	69–84
Silva et	27	ELISA	94–96	92–97	90–	97–98	94–96

al, 61		(blood					98
2007		PAG)					
		ELISA					92–
Romano	28–35 (PSPB)	94–97	94–96	95	95–98	95–96	
&							
Larson, 6							
2							
2010							
		ELISA					
Piechotta	26–58 (PSPB)	98.0	97.1	99.3	91.9	97.8	
et al, 63		ELISA					
2011	(blood	97.8	91.2	97.8	91.2	96.4	
	PAG)						
		ELISA					
Sinedino	28	(blood	95	89	90	95	92
et al, 64							
2014		PAG)					
		ELISA					99– 83– 98–
Lawson	30–95 (milk	98–100	98–100	100	100	100	
et al, 65							
2014		PAG)					
		ELISA					
Ricci et		(blood	100	87	84	100	92
al, 66	32						
2015		PAG)					
		ELISA					
		(milk	98	83	79	99	89
		PAG)					

Abbreviations: NPV, negative predictive value; PAG, pregnancy-associated glycoprotein; PPV, positive predictive value; PSPB, pregnancy specific protein B.

^aProportion of serum samples from pregnant cows with a positive PAG test result.

^bProportion of serum samples from nonpregnant cows with a negative PAG test result.

^cProbability that a positive PAG test result is from a pregnant cow.

^dProbability that a negative PAG test result is from a nonpregnant cow.

^eProbability of correctly identifying pregnancy status.

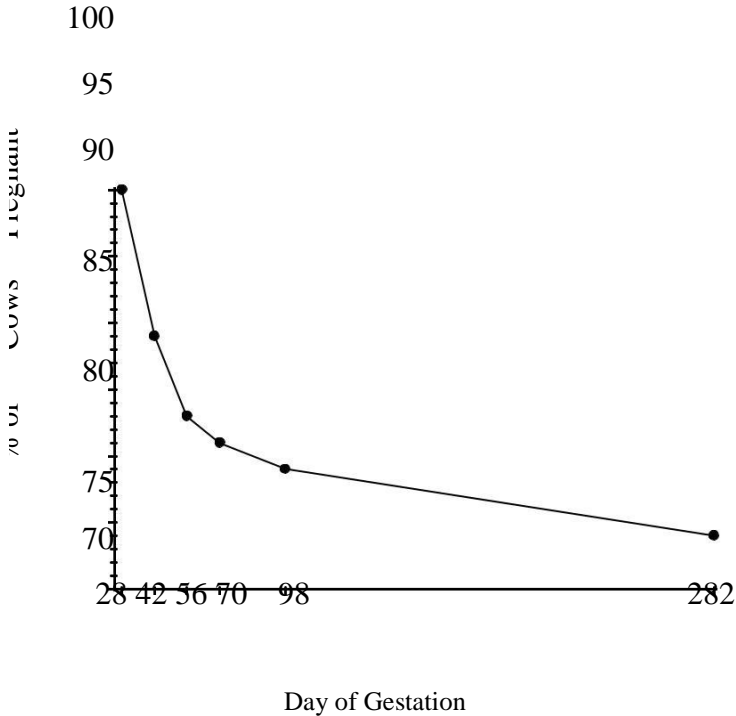


Fig. 1. Pregnancy loss in lactating Holstein cows assessed using transrectal ultrasonography from 28 days after AI to calving. (Adapted from Vasconcelos JLM, Silcox RW, Lacerda JA, et al. Pregnancy rate, pregnancy loss, and response to heat stress after AI at two different times from ovulation in dairy cows [abstract]. Biol Reprod 1997;56(Suppl 1):140; with permission.

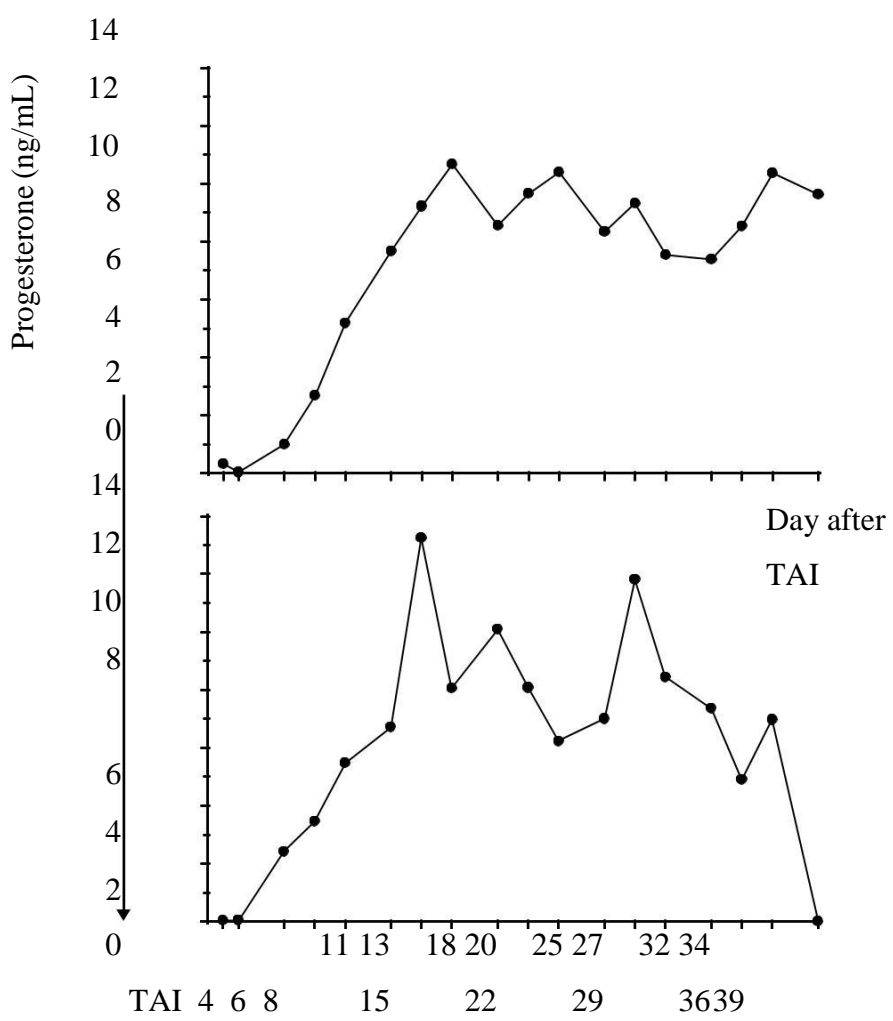
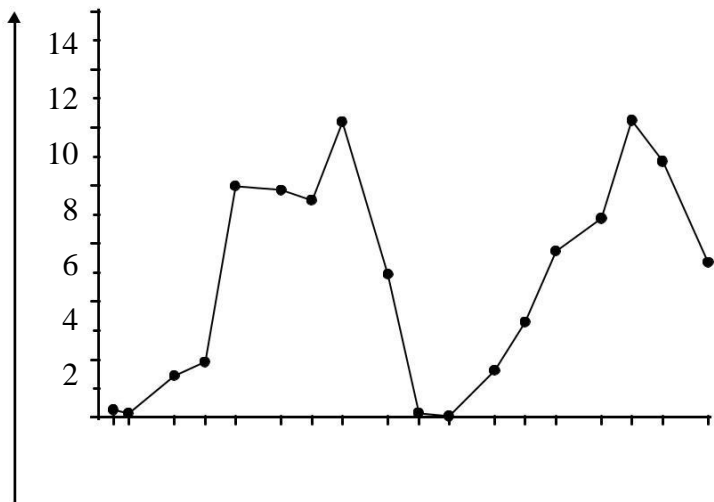


Fig. 2. Representative progesterone profiles from blood samples collected thrice weekly (Monday, Wednesday, Friday) from Holstein dairy cows after synchronization of ovulation and timed AI. Upper panel: a cow that failed to become pregnant after timed AI and had a normal luteal phase; middle panel: a cow that became pregnant after timed AI; lower panel: a cow that failed to become pregnant after timed AI and had an extended luteal phase.

