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1 Efficacy of biocontrol agents and natural compounds against powdery mildew of zucchini

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12 13 Abstract

14
15 The activity of different types of natural compounds and of two biofungicides based on *Bacillus subtilis* and
16 *Ampelomyces quisqualis* alone and in combination with fungicides against powdery mildew of zucchini was
17 tested. The efficacy was compared to the activity of fungicides used alone in four experimental trials carried out
18 in open field and under greenhouse conditions. The *P. xanthii* population used throughout the work was partially
19 resistant to azoxystrobin, while was susceptible to mychlobutanil. Sulphur plus terpenes and mustard oil
20 consistently controlled powdery mildew, followed by mychlobutanil alone or combined with *A. quisqualis*. *B.*
21 *subtilis* and *A. quisqualis* when tested alone were partially effective. The combination of azoxystrobin and *B.*
22 *subtilis* was only delaying the spread of the pathogen.

23
24 **Key words:** *Podosphaera xanthii*; natural compounds; biological control; integrated disease management

25 26 INTRODUCTION

27 Powdery mildew, incited by *Podosphaera xanthii*, previously known as *Sphaerotheca fuliginea* and *S. fusca*
28 (Braun and Takamatsu 2000) is a severe disease of cucurbits and one of two species of powdery mildew of
29 cucurbits worldwide (Sitterly 1978; Zitter et al. 1996). The disease is particularly important in the
30 Mediterranean countries, where it causes severe losses on crops grown in open field as well as under
31 greenhouse. Powdery mildew in Italy is particularly serious on crops such as melon and zucchini.

32 The most common strategy to control powdery mildew of zucchini includes the use of resistant cultivar and the
33 application of fungicides. Actually, chemical control has a key role and it is the principal tool to manage
34 cucurbit powdery mildew (McGrath 2001). However, in spite of this, powdery mildew continues to cause
35 serious losses worldwide (Zitter et al. 1996). The intensive use of chemicals against *P. xanthii* often resulted in
36 the development of resistance: this has happened in the case of most of the groups of chemicals applied
37 (McGrath 2001 and 2007). During the past few years, resistance became widespread also in the case of
38 Quinone outside Inhibitors (QoIs) fungicides (McGrath, 2007; Ishii, 2010).

39 Biological control agents as well as natural compounds are possible alternatives to the use of chemicals, that
40 have been proposed and evaluated in numerous pathosystems, with different degrees of success. Among

41 biocontrol agents, *Ampelomyces quisqualis* and *Bacillus subtilis* have been widely tested and are registered for
42 use in several countries (Copping 2004). In many cases, their application within integrated disease management
43 strategies offered interesting results (Paulitz and Bélanger 2001; Gilardi et al., 2008). Moreover, a synergistic
44 effect between *B. subtilis* and QoI fungicides was observed in the control of powdery mildew of zucchini
45 (Gilardi et al., 2008).

46 Different types of so called natural compounds, ranging from salts such as sodium bicarbonate to plant extracts
47 and oils have been largely exploited against several agents of powdery mildews on a number of crops (Horst et
48 al., 1992; Pasini et al., 1997; Hagiladi and Ziv, 1986; Martin et al., 2005; Stephan et al., 2005; Rongai et al.,
49 2009), providing in many cases very interesting results. Moreover, in some cases a positive effect of mineral
50 fertilisers has been shown (Reuveni and Reuveni, 1998).

51 The main objective of this study was to evaluate the activity of different types of natural compounds, mineral
52 fertilisers, and of two biofungicides based on *B. subtilis* and *A. quisqualis* alone and in combination with
53 fungicides, in comparison with fungicides (included sulphur) used alone against *P. xanthii* on zucchini
54 (*Cucurbita pepo* L.) under open field and greenhouse conditions.

55

56 MATERIALS AND METHODS

57

58 **Field trials.** Two trials were carried out in open field at Boves, in the Cuneo province (Northern-Italy). Zucchini
59 plants (cv. Xsara) 18 day-old, were transplanted into soil covered with black plastic mulch by following a
60 randomized block design, with three replicates and 8 plants/replicate.

61

62 **Greenhouse trials.** Two trials were carried out under greenhouse at Grugliasco, in the Turin province
63 (Northern-Italy). Zucchini plants (cv. Genovese) were grown in pots (14x14 cm, 2 L volume of soil) in a peat:
64 clay: perlite substrate (65:30:5 v/v). Two plants/pot were planted. Plants were maintained at temperatures
65 ranging between 24 and 27 °C, at 60-70% RH. Fifteen-day old plants with their second true expanded leaf were
66 used. A randomised block design with four replicates was used.

67

68 **Sensitivity of the pathogen to the fungicides used during the trials.** The strain AG 1 of *P. xanthii* was
69 collected in Piedmont (Northern Italy) from infected zucchini. The sensitivity of *P. xanthii* AG1 strain towards
70 azoxystrobin and mychlobutanil was evaluated by treating zucchini seedlings at the cotyledon stage with
71 increasing rates of the two fungicides up to twice their field dosages, corresponding respectively to 0.186 ml L⁻¹
72 for azoxystrobin and 0.056 ml L⁻¹ for mychlobutanil. The seedlings treated were placed in a greenhouse at a
73 temperature of 22-25°C. The artificial inoculation was carried out 24 h after the fungicide treatment by using a
74 paint-brush, with 1x10⁵ conidia cm⁻². Inoculated and not treated plants were used as control. After 7-14 days
75 from the last treatment, the percentage of zucchini leaves affected by *P. xanthii* (disease incidence) was
76 evaluated by using a scale from 0 to 5 (0: No infection, 1= 0 to 0.99 % of infected leaf area; 2 = 1- 4.99 %
77 infected leaf area; 3 = 5-19.9 % infected leaf area; 4 = 20-40% infected leaf area; 5 = > 40%). The minimal
78 inhibitory concentration (MIC) and the concentrations able to inhibit 50% (ED₅₀) of the development of *P.*
79 *xanthii* in comparison with the inoculated and non-treated control were evaluated.

80

81 **Treatments.** *Bacillus subtilis* QST 713 (Serenade WP, AgraQuest Inc, USA, 10% a.i.) and *Ampelomyces*
82 *quisqualis* (AQ 10, Intrachem Bio Italia S.p.A., Bergamo, Italy, 58% a.i.) were used as commercial
83 formulations and applied, as foliar sprays, at the suggested dosages, as reported under Tables 2-8. AQ 10 was
84 applied in combination with Nu-Film P, as recommended by the company.

85 Azoxystrobin (Ortiva, Syngenta Crop Protection S.p.A., Milano, Italy, 23.2% a.i.), mychlobutanil (Thiocur
86 forte, DowAgrosciences, 4.5 % a.i.), sulphur plus terpenes (Heliosoufre S, Intrachem Bio Italia S.p.A.,
87 Bergamo, Italy, 51,1% a.i.), mustard oil (Duolif, Cerealtoscana S.p.A., Livorno, Italy, soluble organic nitrogen
88 3%, soluble sulphur 15%, organic matter 80%), organic-mineral fertiliser N:K (Kendal, soluble organic nitrogen
89 3.5%, soluble potassium oxide 15.5%, organic carbon 3-4% Valagro, Atessa, Chieti, Italy), mineral fertiliser
90 N:K+ B, and Mo (Silvest, soluble organic nitrogen 8%, soluble potassium oxide 8%, soluble boron 0.1%,
91 soluble molybdenum 0.01%, Green Has Italia S.p.A., Canale d'Alba, Cuneo, Italy) were applied at the dosages
92 reported under Tables 2 - 8.

93 When applied together, chemicals and biofungicides were mixed before spraying. Treatments were carried out,
94 at 6-8 day intervals, by using 800 l ha⁻¹ with a EFCO atomizer. Treatments were carried out 24 h before the
95 artificial inoculation with the pathogen. Two to three sprays were carried out in the different trials (Table 1).

96
97 **Data collection.** Typical symptoms of powdery mildew started to be visible 7-20 days after artificial
98 inoculation. Plants were checked every 7 days after the last treatment for disease development and the
99 percentage of zucchini leaves affected by *P. xanthii* (disease incidence) was evaluated. The evaluations were
100 carried out by assessing the upper surfaces of 50 (first and second evaluation, Trial 1) and 100 leaves. Disease
101 severity was evaluated by using a disease index ranging from 0 to 5 (EPPO 2004). The disease index used
102 throughout the experiments ranged from 0 to 100 (0 = healthy plant; 1 = 0-0.99 % of infected leaf area; 2 = 1-
103 4.99 % infected leaf area; 3 = 5-19.99 % infected leaf area; 4 = 20-40% infected leaf area; 5 = > 40%). The
104 final disease rating took place 30-37 days after inoculation. Biomass, expressed as fresh weight of zucchini
105 plants at beginning of flowering, was also evaluated at the end of trials 3 and 4.

106
107 **Statistical analysis.** The data from all the experiments were analysed using ANOVA (SPSS software 18) and
108 means were spread according to Tukey's test ($P = 0.05$; WINER 1962). Disease index data were transformed to
109 the respective arcsin values prior to statistical analysis.

110
111 **RESULTS**
112 **Sensitivity of *P. xanthii* AG1 strain towards azoxystrobin and mychlobutanil.** The population of *P. xanthii*
113 AG1 used throughout the work for artificial inoculation was able to cause slight infections on zucchini plants
114 treated with the field dosages of 186 mg L⁻¹ of azoxystrobin. In the case of azoxystrobin, ED₅₀ of *P. xanthii*
115 population after 7 days from the last treatment ranged between 23.2 and 46.4 mg L⁻¹, while MIC was higher
116 than 372 mg L⁻¹. In the case of mychlobutanil, its ED₅₀ was 14-28 mg L⁻¹, while the MIC was 56 mg L⁻¹. MIC.
117 The decreased sensitivity of the population of *P. xanthii* to QoI was confirmed by the low to poor efficacy
118 shown by azoxystrobin in all trials (Tables 2-8).

119

120 **Efficacy of biocontrol agents and natural compounds against powdery.** The artificial inoculation with *P.*
121 *xanthii* resulted in high infection levels in all trials (Tables 2-7), with disease incidence ranging, at the end of the
122 trials in the inoculated untreated controls, from 61 to 96% and disease severity ranging from 20 to 57 %.

123 In trial 1, carried out in open field, the best results, in terms of reduction of disease incidence and disease
124 severity were provided, at the end of the trial, by mustard oil and sulphur, followed by the organic-mineral
125 fertiliser N:K 3.5-15.5 (Kendal), *A. quisqualis* alone and in mixture with mychlobutanil and by the mixture of
126 *B. subtilis* with azoxystrobin. The two biocontrol agents, *B. subtilis* and *A. quisqualis*, when applied alone, only
127 partially controlled the disease. Azoxystrobin and the mineral fertilizer Silvest did not satisfactorily control
128 powdery mildew (Table 2). In particular, at the last reading, in the presence of 70.7% disease incidence in the
129 control plots, mustard oil reduced disease incidence to 27.3%, sulphur to 32.7%, Kendal to 44%, *A. quisqualis*
130 to 45.3%, when applied alone and to 48% when applied in mixture with mychlobutanil (Table 2). Disease
131 severity was reduced from 22.5 % in the untreated control to 5.4 and 5.8% respectively by mustard oil and
132 terpenic sulphur. The mixture of *B. subtilis* + azoxystrobin reduced disease severity to 10.3% and mychlobutanil
133 + *A. quisqualis* to 14%. *A. quisqualis* and *B. subtilis* alone reduced disease severity respectively to 15 and
134 15.4% (Table 2).

135 In trial 2, in the open field, in the presence of 85.3 % disease incidence and 36.0% disease severity in the
136 untreated control at the end of the trial, mychlobutanil provided the best control of powdery mildew (reducing
137 disease incidence to 40.6 and disease severity to 9.8%), followed by sulphur plus terpenes, which reduced
138 disease incidence to 58.0 and disease severity to 12.8%. Mustard oil provided a partial control of the disease.
139 The other tested compounds were only partially effective. In particular, azoxystrobin alone and in mixture with
140 *B. subtilis* provided a limited disease control. The same poor disease control was observed by applying the
141 mineral fertilizer N:K+Mo and B (Silvest) (Table 3).

142 In trial 3, under greenhouse conditions, the best disease control was offered by sulphur plus terpenes, followed
143 by mustard oil and mychlobutanil (Tables 4 and 5). Disease incidence, which was 95.5% in the untreated plots,
144 was reduced to 46.5% by terpenic sulphur, 57.0% by mustard oil and 59.5% by mychlobutanil (Table 4).
145 Disease severity, which was 57.0 in the untreated control, was reduced to 11.3 % by sulphur, to 17.1 % by
146 mustard oil and to 18.3% by mychlobutanil (Table 5). Azoxystrobin, alone and in mixture with *B. subtilis*
147 provided a only partial control of powdery mildew as well as the mineral fertilizer N:K+Mo and B (Silvest),
148 while *B. subtilis* alone was not effective (Tables 4 and 5).

149 In trial 4, under greenhouse conditions, sulphur plus terpenes and mustard oil confirmed their good activity,
150 followed by mychlobutanil alone and in mixture with *A. quisqualis* (Tables 6 and 7). Disease incidence was
151 reduced from 77.6% in the control plots to 41.5% by sulphur, 44.0 % by mustard oil, 49.8 % by mychlobutanil
152 and 50.5% by the mixture mychlobutanil + *A. quisqualis* (Table 6). Disease severity was 39.9 % in the control
153 plots and was reduced to 9.9 % by sulphur plus terpenes and mustard oil, 13,1 % by mychlobutanil and 17.2%
154 by the mixture mychlobutanil + *A. quisqualis* (Table 7). Azoxystrobin and the mineral fertilizer Silvest were
155 less effective.

156 In trials 3 and 4, where also biomass at the end of the trials was considered, sulphur plus terpenes provided the
157 best results, followed by mustard oil (Table 8).

158
159 **DISCUSSION**

160

161 The cucurbit powdery mildew fungus *P. xanthii* has a high potential for developing fungicide resistance, thus
162 complicating disease management. Actually, resistance developed to benzimidazoles, DMIs, organophosphates,
163 hydroxypyrimidines, QoIs, and quinozalines (McGrath 2001). Resistance did develop quickly in some cases,
164 such as DMIs and QoIs. Following resistance development towards DMIs, it was shown that control with this
165 class of fungicides could be improved by decreasing spray intervals, increasing water volumes, and increasing
166 fungicide dosages (Huggenberger et al. 1984). In 1999, after only two years of commercial use, strains of *P.*
167 *xanthii* resistant to QoIs were found in field and greenhouse crops of melon and cucumber in Japan, Taiwan,
168 Spain and France (Heaney et al. 2000)

169 In Italy, resistance to demethylation inhibitors and QoI fungicides has been reported (Gilardi et al., 2008). The
170 widespread presence of populations of the pathogen resistant to several of the most commonly used fungicides
171 makes very interesting the exploitation of control strategies, also based on non-chemical measures (McGrath,
172 2007).

173 In this study, sulphur consistently provided a good disease control both in the open field and under greenhouse
174 conditions. The same good results were provided by mustard oil, Vegetable oil-based fungicides could
175 represent a good alternative to chemical fungicides. They are effective in controlling a number of plant
176 pathogens at low dosages and induce little or no resistance in target fungi (Martin et al., 2005). They have very
177 good spreading and leaf surface adhesion characteristics, and, due to their quick biodegradation rate, they have a
178 low toxicity for human beings and cause a limited environmental impact.

179 Serenade biofungicide is based on a naturally occurring strain of *B. subtilis* QST-713 and is registered and used
180 in several countries (Paulitz and Bélanger 2001; Copping 2004). It works through complex modes of action that
181 entail biological action of the bacteria and also lipopeptide compounds (iturins, agrastatin/plipastatins and
182 surfactins) produced by it, well known for their antimicrobial properties (Marrone 2002; Manker, 2005). The
183 complex mode of action of *B. subtilis* (Jacobsen et al., 2004; Romero et al, 2007) is well suited for its use under
184 integrated control strategies.

185 AQ 10, based on strain AQ 10 of *A. quisqualis* and commercialized in several countries, parasitizes powdery
186 mildew colonies and is active against several powdery mildews on different hosts (Hofstein et al. 1996; Paulitz
187 and Bélanger 2001; Copping 2004). Also AQ 10 is intended for use as part of an integrated disease
188 management programme and is compatible with a wide range of chemicals (McGrath and Shishkoff 1999;
189 Shishkoff and McGrath, 2002). Previous works carried out on cucurbits showed that the same formulation of *B.*
190 *subtilis* showed inconsistent results (from ineffective to very effective) against powdery mildews when applied
191 alone. In alternation with QoIs, *B. subtilis* was significantly more effective (Keinath and DuBose 2004). *B.*
192 *subtilis* QST 713 alternated with sulphur, mychlobutanil and trifloxystrobin provided good control of powdery
193 mildew of lettuce (Matheron and Porchas 2000). A synergistic effect among *B. subtilis* and QoI fungicides when
194 applied against *P. xanthii* on zucchini was reported by Gilardi et al. (2008).

195 In this work, in the presence of high disease pressure, it was possible to manage effectively powdery mildew of
196 zucchini with both sulphur plus terpenes and mustard oil. Mychlobutanil alone and in combination with *A.*
197 *quisqualis* provided interesting results.

198 The good activity shown by the formulation containing sulphur and terpenes as well as mychlobutanil, and the
199 possibility of introduction of natural product such as mustard oil, and biocontrol agents in integrated disease
200 management strategies provides choices for extension services and growers.

201 Azoxystrobin, due to the presence of resistance, did not provide a satisfactory control of the pathogen.

202 This study offers further development to the previous ones, showing the possibility of introducing natural
203 compounds such as mustard oil within management strategies. In the mean time, it shows that an old fungicide
204 such as sulphur plus terpenes can perform well, if applied properly.

205

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207

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210

211 **REFERENCES**

212

213 Braun, U., & Takamatsu S. (2000). Phylogeny of *Erysiphe*, *Microsphaera*, *Uncinula* (Erysipheae) and
214 *Cystotheca*, *Podosphaera*, *Sphaerotheca* (Cystothecaceae) inferred from rDNA ITS sequences – some taxonomic
215 consequences. *Schlechtendalia* 4, 1-33.

216 Copping, L.G. (ed.) (2004). The Manual of Biocontrol Agents. British Crop Protection Council, Alton,
217 Hampshire, UK.

218 EPPO, (2004). EPPO Standards PP1, 2nd Edition, Vol. 2. European and Mediterranean Plant Protection
219 Organization, Paris.

220 Gilardi, G., Manker, D.C., Garibaldi, A., & Gullino, M.L. (2008). Efficacy of the biocontrol agents *Bacillus*
221 *subtilis* and *Ampelomyces quisqualis* applied in combination with fungicides against powdery mildew of
222 zucchini. *Journal of Plant Diseases and Protection*, 115, 208-213.

223 Hagiladi, A., & Ziv, O. (1986). The use of antitranspirants for the control of powdery mildew of roses in the
224 field. *Journal Environmental Horticulture* 4, 69-71.

225 Heaney, S.P., Hall, A.A., Davies, S.A., & Olaya, G. (2000). Resistance to fungicides in the QoI-STAR cross-
226 resistance group: current perspectives. *Proc. BCPC Conf. Pests Dis.* 2, 755-762.

227 Hofstein, R., Daoust, R.A., & Aeschlimann, J.P. (1996). Constraints to the development of biofungicides: the
228 example of AQ 10, a new product for controlling powdery mildews. *Entomophaga* 41, 455-460.

229 Horst, R.K., Kawamoto, S.O. & Porter, L.L. (1992). Effect of sodium bicarbonate and oils on the control of
230 powdery mildew and black spot of roses. *Plant Disease* 76, 247-251.

231 Huggenberger, F., Collins, M.A., & Skylakakis, G. (1984). Decreased sensitivity of *Sphaerotheca fuliginea* to
232 fenarimol and other ergosterol-biosynthesis inhibitors. *Crop Prot.* 3, 137-149.

233 Ishii, H. (2010). QoI fungicide resistance: current status and the problems associated with DNA-based
234 monitoring. In: Recent developments in management of plant diseases (Gisi U., Chet I., Gullino M.L. ds.),
235 Springer, Dordrecht, The Netherlands, 37-45.

236 Jacobsen, B.J., Zidack, N.K., & Larson, B.J. (2004). The role of *Bacillus*-based biological control agents in
237 integrated pest management systems: plant diseases. *Phytopathology* 94, 1272-1275.

238 Keinath, A.P., & DuBose, V.B. (2004). Evaluation of fungicides for prevention and management of powdery
239 mildew on watermelon. *Crop Prot.* 23, 35-42.

240 Manker, D.C. (2005). Natural products as green pesticides. In: J.M. Clark, H. Ohkawa (eds.): *New Discoveries*
241 *in Agrochemicals*, pp. 283-294. American Chemical Society, Washington, DC and Columbus, OH, USA.

242 Marrone, P.G. (2002). An effective biofungicide with novel modes of action. *Pestic. Outl.* 13, 193-194.

243 Martin, B., Hernandez, S., Silvarrey, C., Jacas, J.A., & Cabaleion, C. (2005). Vegetable , fish and mineral oils
244 control grapevine powdery mildew. *Phytopathologia Mediterranea*, 44, 169-179.

245 Matheron, M.E., & Porchas, M. (2000). Evaluation of fungicide performance for control of powdery mildew on
246 lettuce in 2000. Online publication no. AZ1177 in: *Vegetable: College of Agriculture Report 2000*, College of
247 Agriculture, University of Arizona, Tucson, AZ, USA.

248 McGrath, M.T. (2001). Fungicide resistance in cucurbit powdery mildew, experiences and challenges. *Plant*
249 *Dis.* 85, 236-245.

250 McGrath M.T. (2007). Managing cucurbit powdery mildew and fungicide resistance. *Acta Horticulturae*, 731:
251 211-216.

252 McGrath, M.T., & Shishkoff, N. (1999). Evaluation of biocompatible fungicides for managing cucurbit powdery
253 mildew. *Crop Prot.* 18, 471-478.

254 Pasini, C., D'Aquila, F., Curir, P., & Gullino, M.L. (1997). Effectiveness of antifungal compounds against rose
255 powdery mildew (*Sphaerotheca pannosa* var. *rosae*) in glasshouses. *Crop Protection* 16, 251-256.

256 Paulitz, T.C., & Bélanger, R.B. (2001). Biological control in greenhouses systems. *Annu. Rev. Phytopathol.* 39,
257 103-133.

258 Reuveni, R., & Reuveni, M. (1998). Foliar-fertilizer therapy – a concept in integrated pest management. *Crop*
259 *Prot.* 17, 111-118.

260 Romero, D., Devicente, A., Rakotoaly, R.H., Dufour, S.E., Veening, J.W., Arrebola, E., Cazorta, F.M., Kuipers,
261 O.P., Paquot, M., & Perez-Garcia, A. (2007). The iturin and fengycin families of lipopeptides are key factors in
262 antagonism of *Bacillus subtilis* toward *Podosphaera fusca*. *Mol. Plant-Microbe Interact.* 20, 430-440.

263 Rongai, D., Cerato, C., & Lazzeri, L. (2009). A natural fungicide for the control of *Erysiphe betae* and *Erysiphe*
264 *cichoracearum*. *European Journal of Plant Pathology*, 124, 613-619.

265 Shishkoff, N. & McGrath, M.T. (2002). AQ10 biofungicide combined with chemical fungicides or AddQ spray
266 adjuvant for control of cucurbit powdery mildew in detached leaf culture. *Plant Dis.* 86, 915-918.

267 Sitterly, W.R. (1978). Powdery mildew of cucurbits. In: D.M. Spencer (ed.): *The Powdery Mildews*, pp. 359-
268 379. Academic Press, London.

269 Stephan, D., Schmitt, A., Martins Carvalho, S., Seddon, B., & Koch, E. (2005). Evaluation of biocontrol
270 preparations and plant extracts for the control of *Phytophthora infestans* on potato leaves. *Eur. J. Plant Pathol.*
271 112, 235-246.

272 Winer, B.J. (1962). *Statistical Principles in Experimental Design*, 2nd Edition. McGraw-Hill, New York.

273 Zitter, T.A., Hopkins, D.L., & Thomas, C.E. (1996). *Compendium of Cucurbit Diseases*. APS Press, St. Paul,
274 MN, USA.

275

276 **Table 1** Time table for the four powdery mildew experiments

Operation	Field trials ^x		Greenhouse trials	
	1	2	3	4
First treatment	6 ^y	7	4	5
Artificial inoculation with <i>Podosphaera xanthii</i>	7	24	6	6
Second treatment	15	15	11	12
Third treatment	-	31	19	20
First evaluation	35	37	11	19
Second evaluation	49	44	19	25
Third evaluation	-	-	26	32
Fourth evaluation	-	-	33	-
Biomass evaluation	-	-	33	32

277 ^xData of transplant for the four trials: August 7 (Trial 1); July 13 (Trial 2); February 10 (Trial
 278 3); February 25 (Trial 4). The first trial was conducted on 2008, the second in 2010 and the
 279 third and fourth on 2011

280 ^yNumbers indicate days after transplanting

281

282

283

284 **Table 2** Effect of different treatments, expressed as disease incidence and disease severity,
 285 against *Podosphaera xanthii* on zucchini (cv. Xsara) (Trial 1, Boves)

Treatment	Dosage	Disease incidence ^x at		Disease severity ^y at	
	a.i. g or ml L ⁻¹	DAT 35 ^k	DAT 49	DAT 35	DAT 49
<i>Bacillus subtilis</i>	0.4	40.8 bc ^w	52.0 bcd	8.8 a	15.4 ab
<i>Ampelomyces quisqualis</i>	0.029	51.8 cd	45.3 abc	12.3 ab	15.0 ab
Azoxystrobin	0.186	54.7 cd	63.3 cd	11.5 ab	17.8 ab
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	45.0 bcd	48.0 abc	8.9 a	10.3 ab
Mychlobutanil + <i>A. quisqualis</i>	0.056+0.029	34.9 ab	48.0 abc	6.6 a	14.0 ab
Sulphur	1.53	21.3 a	32.7 ab	2.5 a	5.8 a
Kendal (N:K, organic C)	3.0 ^z	46.7 bcd	44.0 abc	11.5 ab	10.4 ab
Duolif (mustard oil)	10.0 ^z	44.7 bcd	27.3 a	8.9 a	5.4 a
Inoculated control	-	57.5 d	70.7 d	26.0 b	22.5 b

286 ^xExpressing the percent of infected leaves

287 ^y Expressing the percent of infected leaf area

288 ^k Numbers indicate days after transplanting

289 ^wMeans within a column, followed by the same letter do not significantly differ following
 290 Tukey's Test $P < 0.05$

291 ^z Dosage (ml L⁻¹) of the commercial formulation

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296 **Table 3** Effect of different treatments, expressed as disease incidence and severity, against
 297 *Podosphaera xanthii* on zucchini (cv. Xsara) (Trial 2, Boves)

Treatment	Dosage	Disease incidence ^x at		Disease severity ^y at	
	a.i. g or ml L ⁻¹	DAT 37 ^k	DAT 44	DAT 37	DAT 44
<i>Bacillus subtilis</i>	0.4	62.8 cd ^w	80.7 cd	22.4 bcd	28.4 bc
Azoxystrobin	0.186	59.4 cd	66.7 bc	22.1 bcd	23.6 abc
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	65.0 cd	63.3 bc	17.6 bcd	15.1 ab
Mychlobutanil	0.056	11.0 a	40.6 a	2.3 a	9.8 a
Sulphur	1.53	34.0 ab	58.0 ab	8.2 ab	12.8 a
Silvest (N:K+B, Mo)	3.5 ^z	64.5 cd	74.0 bcd	23.1 cd	20.4 ab
Duolif (mustard oil)	10.0 ^z	44.2 bc	60.7 b	12.1 abc	19.4 ab
Inoculated control	-	79.9 d	85.3 d	32.3 d	36.0 c

298 ^x Expressing the percent of infected leaves

299 ^y Expressing the percent of infected leaf area

300 ^k Numbers indicate days after transplanting

301 ^w Means within a column, followed by the same letter do not significantly differ following
 302 Tukey's Test $P < 0.05$

303 ^z Dosage (ml L⁻¹) of the commercial formulation

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306 **Table 4** Effect of different treatments, expressed as disease severity, against *Podosphaera*
 307 *xanthii* on zucchini (cv. Genovese) (Trial 3, Grugliasco)

Treatment	Dosage a.i. g or ml L ⁻¹	Disease incidence ^x at			
		DAT 11 ^k	DAT 19	DAT 26	DAT 33
<i>Bacillus subtilis</i>	0.4	5.0 a ^w	40.0 b	48.5 abc	87.0 c
Azoxystrobin	0.186	30.5 b	44.3 b	51.0 bc	71.0 abc
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	5.5 a	41.5 b	56.7 c	83.0 bc
Mychlobutanil	0.056	1,5 a	10.9 a	31.8 ab	59.5 ab
Sulphur	1.53	0.5 a	9.5 a	29.3 a	46.5 a
Duolif (mustard oil)	10.0 ^z	0.5 a	9.5 a	33.3 ab	57.0 ab
Silvest (N:K+B, Mo)	3.5 ^z	41.5 c	47.3 b	54.5 c	70.0 abc
Inoculated and not treated control	-	43.8 c	63.0 c	79.0 d	95.5 c

308 ^x Expressing the percent of infected leaves

309 ^k Numbers indicate days after transplanting

310 ^w Means within a column, followed by the same letter do not significantly differ following
 311 Tukey's Test $P < 0.05$

312 ^z Dosage (ml L⁻¹) of the commercial formulation

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314 **Table 5** Effect of different treatments, expressed as disease severity, against *Podosphaera*
 315 *xanthii* on zucchini (cv. Genovese) (Trial 3, Grugliasco)

Treatment	Dosage a.i. g or ml L ⁻¹	Disease severity ^y at			
		DAT 11 ^k	DAT 19	DAT 26	DAT 33
<i>Bacillus subtilis</i>	0.4	0.3 a ^w	5.6 b	13.8 bc	44.8 de
Azoxystrobin	0.186	5.1 c	13.6 d	18.5 c	37.0 cd
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	0.6 a	6.7 bc	20.7 c	41.6 de
Mychlobutanil	0.056	0.1 a	0.8 a	3.7 ab	18.3 abc
Sulphur	1.53	0.1 a	1.0 a	3.0 a	11.3 a
Duolif (mustard oil)	10.0 ^z	0.0 a	1.0 a	3.2 a	17.1 ab
Silvest (N:K+B, Mo)	3.5 ^z	3.6 b	11.2 cd	14.3 c	31.5 bcd
Inoculated and not treated control	-	5.6 c	27.6 e	44.5 d	57.0 e

316 ^y Expressing the percent of infected leaf area

317 ^k Numbers indicate days after transplanting

318 ^w Means within a column, followed by the same letter do not significantly differ following
 319 Tukey's Test $P < 0.05$

320 ^z Dosage (ml L⁻¹) of the commercial formulation

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322 **Table 6** Effect of different treatments, expressed as disease severity, against *Podosphaera*
 323 *xanthii* on zucchini (cv. Genovese) (Trial 4, Grugliasco)

Treatment	Dosage a.i. g or ml L ⁻¹	Disease incidence ^x at		
		DAT 19 ^k	DAT 25	DAT 32
<i>Bacillus subtilis</i>	0.4	44.7 c ^w	48.0 de	71.5 de
Azoxystrobin	0.186	17.9 b	41.0 cde	56.7 abcd
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	19.4 b	33.7 cd	56.0 abcd
<i>Ampelomyces quisqualis</i>	0.029	39.1 c	56.0 e	56.7 abcd
Mychlobutamil + <i>A. quisqualis</i>	0.056+0.029	13.4 ab	27.0 bc	50.5 abc
Mychlobutanil	0.056	13.3 ab	26.5 bc	49.8 ab
Sulphur	1.53	4.5 a	10.5 ab	41.5 a
Duolif (mustard oil)	10.0 ^z	4.0 a	9.0 a	44.0 ab
Kendal (N:K, organic C)	3.0 ^z	48.8 c	53.5 e	62.5 bcde
Silvest (N:K+B, Mo)	3.5 ^z	39.5 c	46.4 de	69.5 cde
Inoculated and not treated control	-	63.5 d	73.5 f	77.6 e

324 ^x Expressing the percent of infected leaves

325 ^k Numbers indicate days after transplanting

326 ^w Means within a column, followed by the same letter do not significantly differ following
 327 Tukey's Test $P < 0.05$

328 ^z Dosage (ml L⁻¹) of the commercial formulation.

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331 **Table 7** Effect of different treatments, expressed as disease severity, against *Podosphaera*
 332 *xanthii* on zucchini (cv. Genovese) (Trial 4, Grugliasco)

Treatment	Dosage a.i. g or ml L ⁻¹	Disease severity ^y at		
		DAT 19 ^k	DAT 25	DAT 32
<i>Bacillus subtilis</i>	0.4	7.8 b ^w	13.8 de	30.1 cd
Azoxystrobin	0.186	1.8 a	7.7 bc	22.1 abc
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	1.3 a	8.3 cd	19.8 abc
<i>Ampelomyces quisqualis</i>	0.029	9.6 b	20.7 f	22.8 abc
Mychlobutamyl + <i>A. quisqualis</i>	0.056+0.029	0.8 a	2.0 ab	17.3 abc
Mychlobutanil	0.056	0.8 a	3.9 abc	13.1 ab
Sulphur	1.53	0.3 a	1.3 a	9.6 a
Duolif (mustard oil)	10.0 ^z	0.2 a	1.1 a	9.9 a
Kendal (N:K, organic C)	3.0 ^z	10.8 b	18.2 ef	24.9 bc
Silvest (N:K+B, Mo)	3.5 ^z	8.3 b	14.6 e	28.9 cd
Inoculated and not treated control	-	21.9 c	35.9 g	40.0 d

333 ^y Expressing the percent of infected leaf area

334 ^k Numbers indicate days after transplanting

335 ^w Means within a column, followed by the same letter do not significantly differ following
 336 Tukey's Test $P < 0.05$

337 ^z Dosage (ml L⁻¹) of the commercial formulation

338

339 **Table 8** Effect of different treatments, against *Podosphaera xanthii* on zucchini
 340 (cv. Genovese) on biomass (Trials 3 and 4, Grugliasco)

Treatment	Dosage a.i. g or ml L ⁻¹	Biomass (g)	
		Trial 3	Trial 4
<i>Bacillus subtilis</i>	0.4	118.1 abcd ^w	120.0 cde
Azoxystrobin	0.186	82.1 cd	141.7 abc
Azoxystrobin + <i>B. subtilis</i>	0.186+0.4	106.3 bcd	150.4 ab
<i>Ampelomyces quisqualis</i>	0.029	n.t.	110.5 de
Mychlobutamil + <i>A. quisqualis</i>	0.056+0.029	n.t.	93.5 e
Mychlobutanil	0.056	138.3 abc	146.4 abc
Sulphur	1.53	169.8 a	203.5 a
Duolif (mustard oil)	10.0 ^z	158.1 ab	165.1 b
Kendal (N:K, organic C)	3.0 ^z	n.t.	126.9 cde
Silvest (N:K+B, Mo)	3.5 ^z	102.3 bcd	152.6 ab
Inoculated and not treated control	-	71.9 d	126.3 cde

341 ^w Means within a column, followed by the same letter do not significantly differ following

342 Tukey's Test $P < 0.05$

343 ^z Dosage (ml L⁻¹) of the commercial formulation

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