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# 1 A first report of rare earth elements in Northwestern Mediterranean seaweeds

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# 11 Abstract

12 The concentrations of rare earth elements (REE) were determined by ICP-MS in dominant seaweed 13 species, collected from three locations of the northwestern Mediterranean Sea. This is the first study 14 to define levels and patterns of REE in macro algae from these coastal areas.

- Rare elements are becoming emerging inorganic contaminants in marine ecosystems, due to their worldwide increasing applications in industry, technology, medicine and agriculture.
- 17 Significant inter-site and interspecies differences were registered, with higher levels of REE in 18 brown and green macro algae than in red seaweeds. Levels of light REE were also observed to be 19 greater compared to heavy REE in all samples.

20 One of the investigated locations (Bergeggi, SV) had higher REE and  $\Sigma$ REE concentrations, 21 probably due to its proximity to an important commercial and touristic harbor, while the other two 22 sites were less affected by anthropogenic contaminations, and showed comparable REE patterns 23 and lower concentrations.

- 23 24
- 25 **Capsule**: rare earth elements in seaweeds
- 26
- 27 Keywords: REE, seaweeds, Mediterranean Sea, pollution tracers.
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### 1. Introduction

Rare earth elements (REE) are a group of chemical elements including vttrium (Y), scandium (Sc) 32 and lanthanides (from lanthanum to lutetium). Despite their name, REE are not that rare in the 33 natural environment, being the fifteenth most abundant component of the earth's crust (USEPA, 34 2012). REE are further subdivided into light REE (LREE), including lanthanum (La), cerium (Ce), 35 praseodymium (Pr), neodymium (Nd) and samarium (Sm); and heavy REE (HREE), including 36 37 gadolinium (Gd), europium (Eu), terbium (Tb), dysprosium (Dy), thulium (Tm), ytterbium (Yb), holmium (Ho), erbium (Er), lutetium (Lu) and yttrium (Y) (Anastopoulos et al., 2016). 38 REE mainly enter into oceans through atmospheric fallout (De Baar et al., 1983) and fluvial inputs 39 (Frost et al., 1986), and have been frequently investigated as natural tracers of biogeochemical 40 processes (Oliveri et al., 2010). As the distribution patterns of REE in the water column are 41 already known, it is possible to utilize these patterns for tracing water masses or to identify 42

43 pollution sources in seawater (Censi et al., 2004).

In fact, in the last decade, the worldwide use of REE in industrial applications (electronics, nuclear energy, metallurgy, medicine, computer manufacturing) and in some countries (such as China) for use in fertilizer and feed additives, has increased levels of REE in water environments (Mashitah et al., 2012; Hermann et al., 2016). Thus, REE can be considered as emerging contaminants and pose a potential risk for marine and freshwater ecosystems.

The Mediterranean Sea is a semi-enclosed sea; concentrations of trace elements and REE in this basin are higher than those registered in other nutrient-depleted surficial waters (Greaves et al., 1994; Strady et al., 2015). Numerous investigations regarding patterns of dissolved and particulate REE have been performed in this basin (e.g. Censi et al., 2004; Martinez-Boti et al., 2009; Tranchida et al., 2011; Roussiez et al., 2013; Ayache et al., 2016); conversely, occurrence and distribution of REE in marine biota have scarcely been investigated. To our knowledge, there are only two studies that have analyzed the distribution of REE in plankton from the Mediterranean Sea (Strady et al., 2015; Battuello et al., 2017); examining REE in marine organisms is of great
importance because of their increasing levels in seawater environments and, consequently, in the
marine food chain.

Of the marine organisms that can be utilized as bioindicators of trace elements and REE in marine environments, seaweeds have several advantages as they are widespread, easy to collect and have a considerable ability to take-up trace elements in solution and concentrate them. Moreover, as they are at the base of the marine food chain, macro algae are essential in the transfer of trace elements to higher trophic levels.

We determined REE concentrations and distributions in seaweeds from three different sites located in Northwestern Mediterranean coastal areas. These sites have different environmental protection in the Ligurian and Northern Tyrrhenian Sea. The macro algae species collected for this study were the most abundant and widespread in all three sampling sites and were represented by the three phylum Chlorophyta (green algae), Ochrophyta (brown algae) and Rodophyta.

Macro algae species from these three locations were the subject of a previous investigation that focused on essential and nonessential trace elements, in the perspective of identifying the species potentially suitable for human and animal nutrition, as well to identify any potential risks for consumers due to the presence of toxic metals such as lead, cadmium and mercury in seaweeds of Mediterranean origin (Squadrone et al., under review).

In this study, we aimed to measure, for the first time, the concentrations of REE in marine Mediterranean seaweeds, identifying patterns and fractionations of REE, and verifying the potential use of REE as pollution tracers in the studied area.

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### 78 2. Materials and methods

79 2.1. Sampling area

80 All three sampling locations were situated in the northwestern Mediterranean Sea (Figure 1).

The first sampling site was located in Bergeggi (SV), a Marine Protected Area of the Ligurian Sea (44°14'26. 94"N, 8°26'50. 98"E, General Reserve named B zone.) Here, human activities are restricted and regulated by the Italian law. Close to this site is located the industrial and commercial harbor of Vado Ligure (SV), characterized by high shipping traffic.

The second site was in the Island of Elba (Tyrrhenian Sea, 42°42'35. 17"N, 10°24'44. 97"E), five nautical miles off the Tuscan coast. Elba is the most populated island of the Tuscan archipelago, especially in summer.

The third sampling site was located in the little Capraia Island, (43°4'26. 90"N, 9°49'39.63"E, in the National Park of the Tuscan Archipelago, PNAT), another Marine Protected Area of the Ligurian Sea, about 30 nautical miles off Tuscan coast. The island has few inhabitants and no industrial activities.

Seaweed samples were collected in summer 2016. After collection, the macro algae were washed on board with seawater and then stored in refrigerated conditions. The specimens were transported to the laboratory and examined under the stereomicroscope, after being cut into thin sections, in order to identify the macro algae species. Before analyzing the seaweeds for REE content in the chemical laboratory, samples were rinsed with tap water, followed by a rinse with distilled water, then freezedried and homogenized to obtain a fine powder. Approximately 1-1.5 g of each sample were utilized for quantitation of REEs.

99 2.2 Determination of REE

Samples mineralization was performed using a microwave digestion lab station (Ethos 1, Milestone,
 Shelton, CT, USA), equipped with a 10 positions rotor for high pressures polytetrafluoroethylene
 (PTFE) digestion tubes.

All digestion tubes were cleaned with concentrated acid, rinsed with ultrapure water and dried at room temperature under a chemical hood. Disposable polypropylene tubes were used to storage mineralized samples. Freeze-dried samples (1.0-1.5 g) were directly weighed into PTFE digestion tubes. 7 mL of HNO<sub>3</sub> (70% v/v) and 1.5 mL of  $H_2O_2$  (30% v/v) were then added before the microwave digestion process, programmed as follows: heating to 130°C in 8 min, hold for 2 min, heating to 200°C in 8 min, hold for 5 min; cooling for 30 min. Digested samples were then quantitatively transferred to 50 mL polypropylene tubes and gravimetrically diluted to a final weight of 50 g with ultrapure water.

REE determination was performed by Inductively Coupled Plasma-Mass Spectrometer (ICP-MS Xseries II, Thermo Scientific, Bremen, Germany) equipped with a multi-vial auto sampler (ASX 520, CETAC Technologies, Omaha, NE, USA). Instrument was tuned daily before each analytical trial. Certified Reference Materials (REE-1 from the National Institute of Standard and Technology), blank reagents and standard solutions were processed during each analytical session to verify performances of the methods. The limit of quantification (LOQ) was 0.010 mg Kg<sup>-1</sup>.

117 Results were expressed in mg  $Kg^{-1}$  dry weight as the mean for each site with standard deviation; the

sum of REE ( $\Sigma$ REE), of light REE (LREE) and of heavy REE (LREE) are also shown (Table 1).

119 *2.3 Statistical analysis* 

The one-way analysis of variance (ANOVA) was employed to compare the average contents of the sum of REE ( $\Sigma$ REE) in macro algae between the three sampling sites and between the macro algae species in the same site (Table 2). The unpaired t-test was used to compare the average contents of  $\Sigma$ REE in macro algae between sites 2 and 3. Results were considered statistically significant at p values of < 0.05. Graph Pad Statistics Software Version 6.0 (GraphPad Software, Inc., USA) was used for statistical evaluations.

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### 3. Results and Discussion

REE are typical lithophile elements, with scarce presence in biological tissues. Seaweeds mostly develop in marine environments in contact with sediments and suspended particulate from geologic origin is present in their environment. Therefore, fine geological particulate, containing REE, are incorporated in plants tissues in different amounts. The concentrations of REE in marine macro algae from the three collection sites of the Northwestern Mediterranean Sea are shown in Table 1 (mg Kg<sup>-1</sup> dry weight). REE and  $\Sigma$ REE are also graphically presented for the three sites (Figure 2) and for the analyzed seaweeds (Figures 3 and 4), to facilitate comparison.

A high variability in REE concentrations between the three sampling sites and between species was recorded; the REE concentrations, however, consistently followed the same trend, and concentrations of LREE were always higher than HREE.

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### 3.1 Inter-site variability

In Figures 2a and 2b, mean REE levels in the three sampling sites are graphically represented. As 139 shown, the specific area of collection greatly affected REE concentrations. In fact, macro algae 140 from site 1 (Bergeggi, SV) had the highest concentrations for all the analyzed REE (Table 1). In site 141 1, the highest REE values were found in the Ocrophyta Halopteris filicina (Ce 8.8 mg Kg<sup>-1</sup>, La 4.3 142 mg Kg<sup>-1</sup>, Nd 4.1 mg Kg<sup>-1</sup>, Y 3.4 mg Kg<sup>-1</sup>, Sc 1.5 mg Kg<sup>-1</sup>, Pr 1.1 mg Kg<sup>-1</sup>, Sm and Gd 0.87 mg Kg<sup>-1</sup> 143 <sup>1</sup>, Dy 0.64 mg Kg<sup>-1</sup>, Er 0.32 mg Kg<sup>-1</sup>, Yb 0.27 mg Kg<sup>-1</sup>, Tb and Ho 0.22 mg Kg<sup>-1</sup>, Eu 0.17 mg Kg<sup>-1</sup>, 144 Tm 0.80 mg Kg<sup>-1</sup>, Lu 0.78 mg Kg<sup>-1</sup>) and the lowest concentrations were found in the Rodhopyta 145 Ganonema farinosum (Ce 4.8 mg Kg<sup>-1</sup>, La 2.5 mg Kg<sup>-1</sup>, Nd 2.2 mg Kg<sup>-1</sup>, Y 2.0 mg Kg<sup>-1</sup>, Sc 0.84 mg 146 Kg<sup>-1</sup>, Pr 0.55 mg Kg<sup>-1</sup>, Sm and Gd 0.46 mg Kg<sup>-1</sup>, Dy 0.35 mg Kg<sup>-1</sup>, Er 0.19 mg Kg<sup>-1</sup>, Yb 0.15 mg 147 Kg<sup>-1</sup>, Eu 0.10 mg Kg<sup>-1</sup>, Tb and Ho 0.080 mg Kg<sup>-1</sup>, Tm and Lu 0.030 mg Kg<sup>-1</sup>). 148

Despite being located in a marine protected area, site 1 seems to be greatly affected by beingsituated close to an important industrial and touristic harbor.

- 151 In site 2 (Elba Island, LI), we detected the highest levels of all elements in the Clorophyta Flabellia
- 152 *petiolata* (Ce 3.5 mg Kg<sup>-1</sup>, Y 2.3 mg Kg<sup>-1</sup>, La 2.2 mg Kg<sup>-1</sup>, Nd 1.9 mg Kg<sup>-1</sup>, Sc 0.66 mg Kg<sup>-1</sup>, Pr
- 153  $0.47 \text{ mg Kg}^{-1}$ , Gd  $0.45 \text{ mg Kg}^{-1}$ , Sm  $0.40 \text{ mg Kg}^{-1}$ , Dy  $0.34 \text{ mg Kg}^{-1}$ , Er  $0.18 \text{ mg Kg}^{-1}$ , Yb 0.15 mg
- 154  $Kg^{-1}$ , Eu and Ho 0.10 mg Kg<sup>-1</sup>, Tb 0.091 mg Kg<sup>-1</sup>, Tm 0.040 and Lu 0.033 mg Kg<sup>-1</sup>).
- 155 In site 3 (Capraia Island, LI), the highest REE concentrations were found in the Ocrophyta
- 156 *Halopteris scoparia* (Ce 6.2 mg Kg<sup>-1</sup>, La and Nd 2.7 mg Kg<sup>-1</sup>, Y 1.9 mg Kg<sup>-1</sup>, Sc 0.87 mg Kg<sup>-1</sup>, Pr

157  $0.65 \text{ mg Kg}^{-1}$ , Sm and Gd  $0.51 \text{ mg Kg}^{-1}$ , Dy  $0.34 \text{ mg Kg}^{-1}$ , Er  $0.17 \text{ mg Kg}^{-1}$ , Yb  $0.13 \text{ mg Kg}^{-1}$ , Eu 158  $0.10 \text{ mg Kg}^{-1}$ , Ho  $0.073 \text{ mg Kg}^{-1}$ , Tb  $0.070 \text{ mg Kg}^{-1}$ , Tm  $0.025 \text{ mg Kg}^{-1}$ , Lu  $0.022 \text{ mg Kg}^{-1}$ ) and the 159 lowest levels were found in the Rodhopyta *Dudresnaya verticillata*.

Sites 2 and 3 seem to be less affected by anthropogenic contamination, and REE mean values werehalf of those registered in site 1.

In Table 2, the comparison between the  $\Sigma$ REE by one-way ANOVA showed a highly significant difference in concentrations between the three locations (p < 0.0001). However, the comparison between only sites 2 and 3 using the unpaired t-test resulted in a non-significant difference (p > 0.05), highlighting that site 1 showed very different levels of REE in seaweeds, while between sites 2 and 3,  $\Sigma$ REE values did not differ significantly.

167 *3.2 Inter* 

# Interspecies variability

168 The total levels of REE ( $\Sigma$ REE) are shown for each analyzed species in the three examined 169 locations (Figure 3).

In site 1, Bergeggi (SV), the highest values were found overall, especially in green and brown 170 macro algae, while in sites 2 and 3, REE values were lower; however, there was a high interspecies 171 variability in the same sampling site. In fact, in site 1 (Figure 3), the total REE content was in the 172 following decreasing order Halopteris filicina ( $\Sigma REE 27 \text{ mg Kg}^{-1} \text{ d.w.}$ ) > Flabellia petiolata > 173 Padina pavonica > Codium bursa > Ganonema farinosum ( $\Sigma REE 15 \text{ mg Kg}^{-1} \text{ d.w.}$ ); in site 2, the 174 order was *Flabellia petiolata* ( $\Sigma REE 15 \text{ mg Kg}^{-1}$  dry weight (d.w.) > *Dictyota dichotoma* > *Codium* 175 bursa = Padina pavonica > Peyssonnelia squamaria > Laurencia obtusa > Caulerpa racemosa > 176 Halopteris filicina ( $\Sigma REE 2.1 \text{ mg Kg}^{-1} \text{ d.w.}$ ); and finally, in site 3, the order was Halopteris 177 scoparia ( $\Sigma REE 17 \text{ mg Kg}^{-1} \text{ d.w.}$ ) > Padina pavonica > Halimeda tuna > Peyssonnelia squamaria 178 > Cystoseira spp> Flabellia petiolata> Codium bursa> Dudresnava verticillata ( $\Sigma$ REE 2.3 mg 179  $Kg^{-1}$  d.w.). A comparison between the three species that were collected in all three locations (*F*. 180 petiolata, C. bursa, P. pavonica) is shown in Figure 4. The REE pattern was similar, but 181

182 concentrations differed in the same species in the two green macro algae *F. petiolata* and *C. bursa*183 from the three sites; the brown macro alga *P. pavonica* showed almost the same levels of REE in
184 sites 2 and 3, but a higher value in site 1.

As shown in Table 2, the one-way interspecies comparison revealed highly significant differences between the different macro algae species for each site. This finding is in line with the scarce literature regarding REE in macro algae. In fact, other authors have underlined the interspecies variability in REE levels. Fu and coauthors (2000) suggested that REE patterns are divisiondependent; while Sakamoto and coauthors (2008) indicated that the mechanism of uptake could be different and related to seaweed morphology, even if the accumulation of REE in seaweed is still not elucidated.

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### 3.3 Chondrite-normalized REE pattern (Leedey Oklahoma chondrite)

To define a normalized REE pattern (Figure 5), we utilized the normalized values reported by Masuda (1975) for the Leedey chondrite, which is considered the most primitive chondrite (Sakamoto et al., 2008). Chondrite meteorites are, in fact, used as a reference for the normalization of REE, as they are thought to be similar to the original composition of the Earth's crust (Masuda et al., 1973). Moreover, during this meteorite formation, lanthanide fractionation did not occur (Song et al., 2006; Antonina et al., 2013), therefore, if fractionation between REE in seaweeds occurred, the comparison with the chondrite pattern could disclose this phenomenon.

In our study, the REE patterns were comparable in the three sites (Figure 5), even if, as previously 200 indicated, site 1 (Bergeggi, SV) showed higher overall REE concentrations than the other two 201 locations. The REE Ce and Eu have additional valences compared to the other lanthanides, and 202 when Ce and/or Eu concentrations are enriched or depleted compared to the levels recorded in 203 chondrite, this phenomenon is defined as a Ce or Eu (positive or negative) anomaly. In seaweeds 204 205 from the three sites, a Eu negative anomaly was observed (Figure 5), while the REE ratios (Ce/La, Gd/Yb, La/Yb) were almost the same in the three different stations (mean values 1.8, 3.3 and 17, 206 respectively). 207

The Eu anomaly is thought to be strictly dependent on lithology (Moller et al., 2004) and enrichment or depletion was explained by the Eu capacity to be mostly incorporated into plagioclase minerals.

Despite the very different sites of origin, the chondrite-normalized pattern profile of Mediterranean seaweeds appeared to be very similar to the pattern found by Mashitah and co-authors (2012) in brown seaweeds from Malaysian coasts.

The REE patterns normalized by chondrite (Figure 5) are typical of geological materials such as sediments, confirming that REE measurements are compatible with a geological material incorporated in macro algae tissues in different amounts. Moreover, the patterns are similar among sampling sites suggesting that correspond to sediments of similar origin.

218 *3.4 Comparison with REE in biota* 

In a previous study, we analyzed Ce and La concentrations in marine zooplankton from the Northwestern Mediterranean Sea (Battuello et al., 2017). We observed that concentrations for both these elements decreased from herbivorous to carnivorous copepods, and were in the average range of  $0.50 - 1.86 \text{ mg Kg}^{-1}$  for Ce and  $0.28 - 0.88 \text{ mg Kg}^{-1}$  d.w. for La (lowest values in carnivores).

In this investigation, Ce and La in seaweeds were in the medium range of 7.2 - 2.5 mg Kg<sup>-1</sup> and 3.7 223 - 1.3 mg Kg<sup>-1</sup> d.w., respectively, showing the higher ability of REE to accumulate in seaweeds 224 compared to zooplankton. As for as we know, we cannot compare these results with other seaweeds 225 226 from the Mediterranean Sea, but a few studies have been performed in other parts of the world. For example, Hou and Yan (1998) analyzed La levels in Chinese coast seaweeds, finding the highest 227 values reported in seaweeds to date, 10.14 mg Kg<sup>-1</sup> d.w. in green macro algae and 6.73 mg Kg<sup>-1</sup> d.w. 228 in red macro algae, while the highest value we found for lanthanum was 4.3 mg Kg<sup>-1</sup> in *H. filicina* 229 and P. pavonica from site 1. 230

231 Masitah (2012) analyzed REE concentrations in *P. pavonica* (Malaysian coast), and found  $\Sigma$ REE 232 values ranging from 62 to 8.4 mg Kg<sup>-1</sup>, higher concentrations than in the Mediterranean area, where the range we registered was 22 - 7.9 mg Kg<sup>-1</sup> (Table 1, Figure 2). REE in *Padina sp.* from the
Malaysia areas decreased in the following order:
Ce>Nd>La>Pr>Gd>Sm>Dy>Er>Yb>Eu=Tb>Ho>Tm>Lu.

In our 236 study, in site 1 (Bergeggi, SV), the order of REE was: Ce>La>Nd>Y>Pr>Gd=Sm>Sc>Dy>Er>Yb>Eu>Tb>Ho>Tm>Lu, while in sites 2 and 3, the order 237 Ce>Y>La>Nd>Sc>Pr>Gd>Sm>Dy>Er>Yb>Eu>Ho>Tb>Tm>Lu, namely 238 was the same. demonstrating that *P. pavonica* samples in these two sites have the same geological "fingerprint". 239 240 Moreover, it was evident that these findings reflected a different pattern of REE in seaweeds, not only between the two different marine areas (Malaysia and Mediterranean), but also between the 241 Mediterranean stations that we investigated. Sakamoto and co-authors (2008) investigated REE 242 patterns in seaweed species collected in the Pacific Ocean (Japan). They found that seaweeds 243 accumulated REE at levels  $10^3$  times higher than concentrations detected in seawater, and that the 244 245 accumulation factor was higher for the heavy REE.

In order to estimate the bio concentration factor (BCF), which is defined as the accumulation of a chemical from water in an organism (Landis et al., 2011), we used the REE concentrations measured in surface waters of the Mediterranean Sea (Censi et al., 2004), ranging from 0.00013  $\mu$ g L<sup>-1</sup> (Yb) to 0.0029  $\mu$ g L<sup>-1</sup> (La). In fact, as REE partitioning is controlled by complexation and binding constants on an ocean-wide basis (Sholkovitz et al., 1994; Strady et al., 2015), we decided to apply them on a regional basin-wide basis, with the approach already utilized in the study by Strady and co-authors (2015).

BCF was usually expressed as the ratio of the concentration of the chemical in the organism and inwater; our findings are shown in Figure 6.

The BCFs were expressed in the following decreasing order:
Ce>Yb>Pr>La>Nd>Sm>Eu>Tb>Gd>Dt>Ho>Tm>Lu>Er, and with the exception of Yb, the bio
concentration factors were higher in LREEs than in HREEs.

In a previous investigation in the studied area (Squadrone et al., under review), we found that 258 different macro algae accumulated trace elements from seawater to different extents, and that brown 259 and green macro algae had higher values than red macro algae. In fact, in seaweeds from site 1, 260 important concentrations of iron, aluminum, manganese, copper, nickel and chromium were 261 registered. 262

In this study, the same site (Bergeggi, SV) showed the highest levels of REE. 263

We suggest that REE concentrations in macro algae from sites 2 and 3, being non-contaminated 264 areas, could constitute a baseline record for these elements in Mediterranean seaweeds, due to their 265 characteristic elemental profile. In the presence of anthropogenic sources, such as in site 1, this 266 unique profile was altered, and we can, therefore, suggest the use of REEs as pollution tracers. 267

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269 4.

### Conclusions

270 REE, due to their unique chemical properties have become crucially important in many industrial applications, and the global demand is still increasing. Following the consequent release of REE in 271 terrestrial and aquatic environments, due to the extraction process and production of several 272 273 industrial components, REE can be considered new emerging inorganic contaminants, for which the potential risks for human health and ecosystems have still not been investigated. Seaweeds have 274 been shown to constitute a useful tool for biomonitoring REE, as they can concentrate REE at 275 higher levels than in seawater. This study constitutes the first investigation of REE in seaweeds 276 from the Northwestern Mediterranean Sea, providing the first record, which can be utilized for 277 future comparisons. 278

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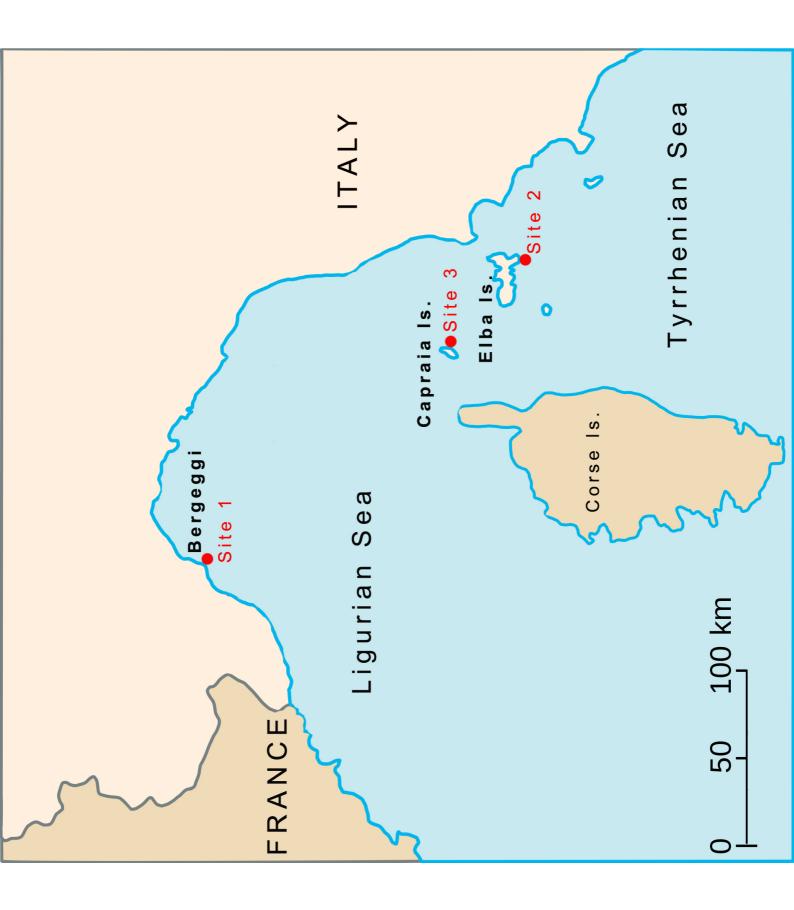
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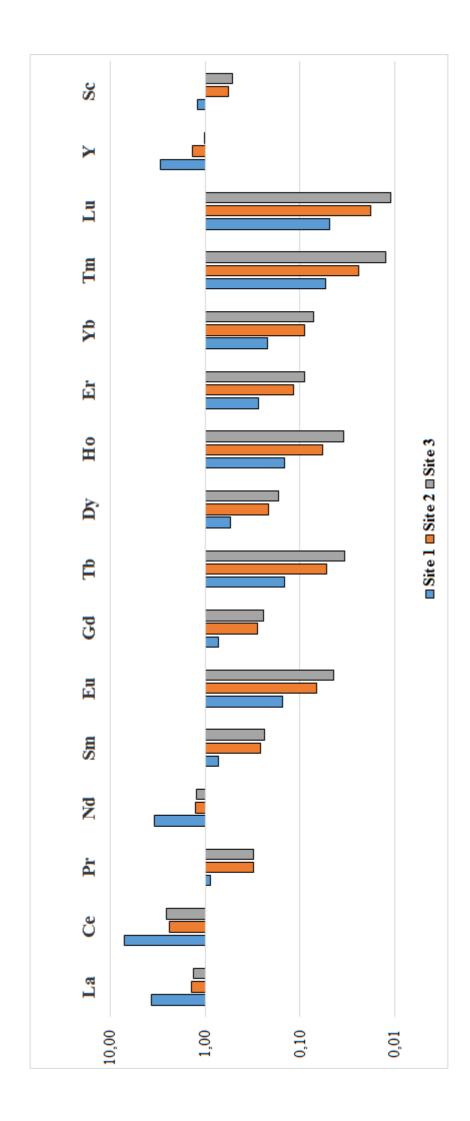
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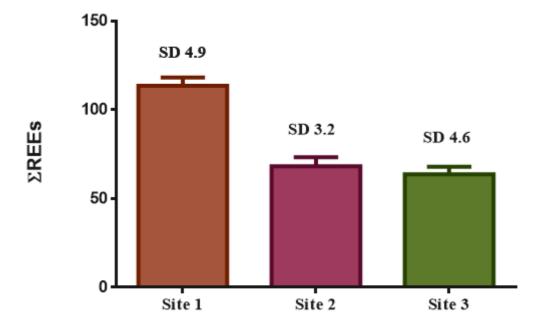
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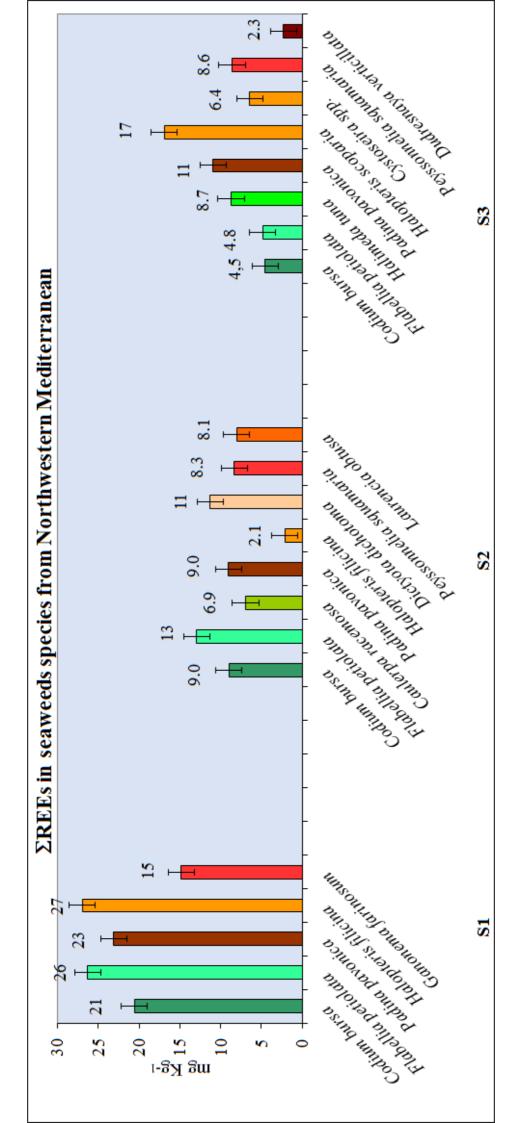
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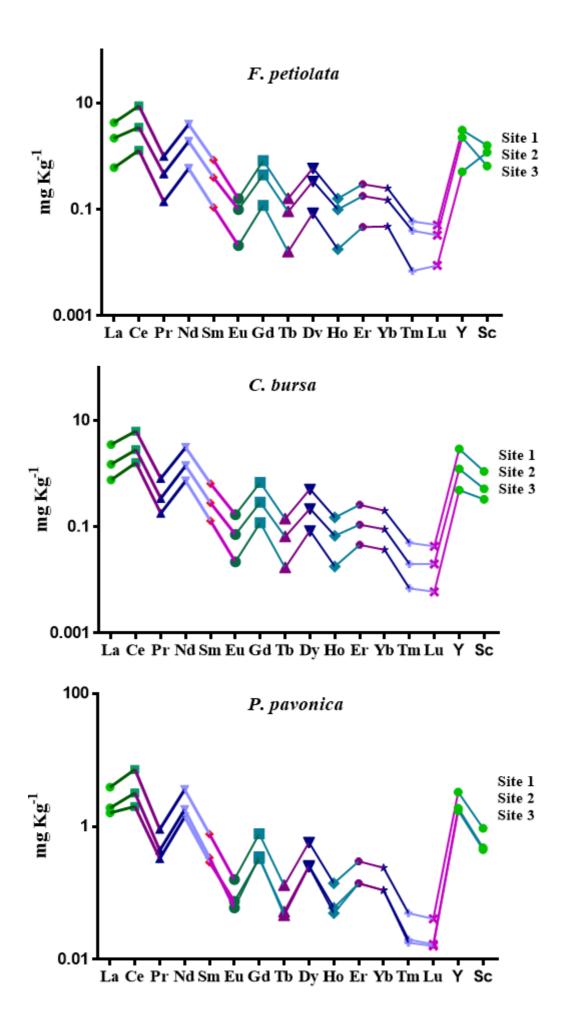
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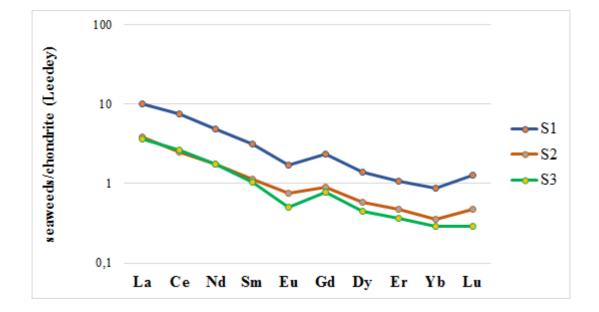


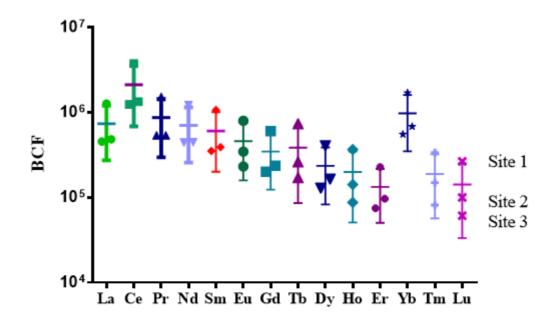












Flabell. Padina Halopto Ganone farinosu mean SD 32 Codium Flabell. Caulerp Padina Halopto Squama Lauren mean SD 33	lium bursa bellia petiolata lina pavonica	3.5				Sm	Eu	Gd	Tb	Dy	Ho	Er	Yb	Tm	Lu	Y	Sc	ΣREE	LREE	HREE
Padina Halopte Ganone farinost mean SD 32 Codium Flabell Caulerp Padina Halopte Dictyot Peysson squama Laurem Mean SD 33 Codium Flabell Halime Padina Halopte	1		6.3	0.81	3.1	0.65	0.17	0.68	0.14	0.51	0.15	0.26	0.20	0.050	0.043	2.9	1.1	21	14	5.1
Halopte Ganone farinost mean SD 32 Codium Flabell Cauler Padina Halopte Dictyot Peysson squama Lauren mean SD 33	ina navonica	4.3	8.8	1.0	4.0	0.85	0.16	0.83	0.16	0.60	0.16	0.30	0.25	0.060	0.051	3.1	1.6	26	19	5.6
Ganone farinosu mean SD 32 Codium Flabell. Caulerp Padina Halopte Dictyot Peysson squama Lauren. Mean SD 33 Codium Flabell. Halime Padina Halopte	ina pavonica	3.9	7.2	0.91	3.6	0.77	0.16	0.78	0.13	0.58	0.14	0.30	0.24	0.050	0.041	3.3	0.94	23	16	5.8
mean SD S2 Codium Flabell. Caulery Padina Halopte Dictyot Peysson squama Lauren. Mean SD S3 Codium Flabell. Halime Padina Halopte Cystose		4.3	8.8	1.1	4.1	0.87	0.17	0.87	0.22	0.64	0.22	0.32	0.27	0.080	0.078	3.4	1.53	27	19	6.3
SD Codium Flabell Caulerp Padina Halopte Dictyot Peysson squama Lauren mean SD 33		2.5	4.8	0.55	2.2	0.46	0.10	0.47	0.08	0.35	0.08	0.19	0.15	0.030	0.030	2.0	0.84	15	10	35
Codium Flabell Caulery Padina Halopte Dictyot Peysson squama Laurent <b>mean</b> SD 33 Codium Flabell Halime Padina Halopte Cystose	an	3.7	7.2	0.87	3.4	0.72	0.15	0.73	0.15	0.53	0.15	0.27	0.22	0.054	0.048	2.9	1.2	22	16	6.5
Codium Flabell Caulerp Padina Halopte Dictyot Peyssor squama Lauren <b>mean</b> <b>SD</b> 33 Codium Flabell Halime Padina Halopte Cystose		0.77	1.72	0.21	0.77	0.17	0.03	0.16	0.051	0.11	0.049	0.051	0.048	0.018	0.018	0.55	0.36	4.9	3.6	1.1
Flabell. Caulerp Padina Halopte Dictyot Peyssor squama Lauren. <b>mean</b> <b>SD</b> 33 Codium Flabell. Halime Padina Halopte Cystose		La	Ce	Pr	Nd	Sm	Eu	Gd	Tb	Dy	Ho	Er	Yb	Tm	Lu	Y	Sc	ΣREE	LREE	HREE
Caulery Padina Halopto Dictyot Peyssor squama Laurent <b>mean</b> <b>SD</b> 33 Codium Flabell Halime Padina Halopto Cystose	lium bursa	1.5	2.8	0.34	1.4	0.28	0.072	0.29	0.065	0.22	0.068	0.11	0.090	0.020	0.020	1.22	0.52	9.0	6.3	2.2
Padina Padina Halopte Dictyot Peyssor squama Lauren <b>mean</b> SD S3 Codium Flabell Halime Padina Halopte Cystose	bellia petiolata	2.2	3.5	0.47	1.9	0.40	0.10	0.45	0.091	0.34	0.10	0.18	0.15	0.040	0.033	2.26	0.66	13	8.5	3.8
Halopte Dictyot Peyssor squama Lauren <b>mean</b> SD S3 Codium Flabell Halime Padina Halopte Cystose	lerpa racemosa	1.1	1.8	0.24	1.0	0.22	0.050	0.24	0.040	0.20	0.050	0.11	0.080	0.020	0.020	1.30	0.50	6.9	4.3	2.1
Dictyot Peyssor squama Lauren mean SD 33 Codium Flabell Halime Padina Halopte Cystose	ina pavonica	1.6	2.0	0.33	1.4	0.29	0.075	0.33	0.052	0.26	0.060	0.14	0.11	0.020	0.017	1.88	0.48	9.0	5.6	2.9
Peysson squama Lauren mean SD 33 Codium Flabell Halime Padina Halopto Cystose	opteris filicina	0.29	0.64	0.07	0.31	0.070	0.010	0.070	0.010	0.050	0.010	0.030	0.020	0.031	0.003	0.23	0.25	2.1	1.4	0.5
squama Lauren mean SD 33 Codium Flabell Halime Padina Halopto Cystose	tyota dichotoma	1.8	3.3	0.41	1.6	0.34	0.090	0.36	0.060	0.27	0.070	0.15	0.11	0.020	0.020	1.61	1.13	11	7.4	2.8
mean SD S3 Codium Flabell Halime Padina Halopto Cystose	ssonnelia amaria	1.6	2.6	0.32	1.3	0.25	0.070	0.26	0.050	0.18	0.050	0.090	0.070	0.020	0.010	1.02	0.45	8.3	6.0	1.8
SD Codium Flabell Halime Padina Halopte Cystose	rencia obtusa	1.3	2.3	0.28	1.2	0.25	0.060	0.27	0.050	0.21	0.050	0.11	0.080	0.020	0.020	1.33	0.61	8.1	5.2	2.2
5 <b>3</b> Codium Flabell Halime Padina Halopto Cystose	an	1.4	2.4	0.31	1.3	0.26	0.066	0.28	0.052	0.22	0.057	0.12	0.089	0.024	0.018	1.4	0.58	8.5	5.6	2.9
Codium Flabell Halime Padina Halopto Cystose		0.56	0.91	0.12	0.47	0.10	0.028	0.11	0.023	0.084	0.025	0.046	0.038	0.008	0.009	0.60	0.26	3.2	2.1	1.0
Flabell Halime Padina Halopte Cystose		La	Ce	Pr	Nd	Sm	Eu	Gd	Tb	Dy	Ho	Er	Yb	Tm	Lu	Y	Sc	ΣREE	LREE	HREE
Halime Padina Halopte Cystose	lium bursa	0.76	1.6	0.18	0.73	0.13	0.022	0.12	0.017	0.085	0.018	0.046	0.037	0.0070	0.006	0.49	0.33	4.5	3.4	0.9
Padina Halopte Cystose	bellia petiolata	0.61	1.3	0.14	0.59	0.11	0.021	0.12	0.016	0.086	0.018	0.047	0.048	0.0070	0.0089	0.51	1.19	4.8	2.7	0.9
Halopte Cystose	imeda tuna	1.51	3.0	0.37	1.4	0.26	0.040	0.25	0.030	0.17	0.030	0.090	0.071	0.012	0.0090	1.04	0.42	8.7	6.5	1.7
Cystose	ina pavonica	1.9	3.2	0.44	1.8	0.34	0.060	0.35	0.046	0.25	0.050	0.14	0.11	0.018	0.016	1.74	0.45	11	7.7	2.8
	opteris scoparia	2.73	6.2	0.65	2.7	0.51	0.10	0.51	0.070	0.34	0.073	0.17	0.13	0.025	0.022	1.87	0.871	17	13	3.3
Peysson		1.00	1.5	0.23	0.99	0.22	0.050	0.25	0.040	0.20	0.040	0.12	0.096	0.016	0.015	1.38	0.32	6.4	3.9	2.2
squama Dudres	toseira spp. ssonnelia		2.9	0.35	1.4	0.26	0.050	0.26	0.040	0.18	0.040	0.090	0.070	0.013	0.011	1.01	0.40	8.6	6.4	1.8
	ssonnelia amaria Iresnaya	1.52			0.25	0.070	0.010	0.070	0.010	0.040	0.010	0.000	0.012	0.0000	0.00000	0.00	0 1 0		1.0	<u> </u>
mean SD	ssonnelia amaria Iresnaya icillata	1.52 0.52 <b>1.32</b>		0.090 <b>0.31</b>	0.35	0.060 <b>0.24</b>	0.010 <b>0.044</b>	0.070 <b>0.24</b>	0.010 <b>0.034</b>	0.040 <b>0.17</b>	0.010 <b>0.035</b>	0.020 0.090	0.013 0.072	0.0020 <b>0.013</b>	0.0020 <b>0.011</b>	0.20	0.10 <b>0.51</b>	2.3 <b>7.9</b>	1.8 <b>5.7</b>	0.4 <b>2.3</b>

# Table 2 Statistical evaluation

P value	P value
P < 0.0001	****
P = 0.0811	
(P > 0.05)	NS
P < 0.0001	****
P < 0.0001	****
P < 0.0001	****
	P < 0.0001 P = 0.0811 (P > 0.05) P < 0.0001 P < 0.0001

# Figure 1

Sampling sites, study area.

### Figure 2a

Rare earth elements in macro algae from three Northwestern Mediterranean locations (mg Kg<sup>-1</sup> dry weight, log scale).

# Figure 2b

Box-plot diagrams of  $\Sigma REE$  (mean  $\pm$  SD) in the three sampling locations (mg Kg<sup>-1</sup> dry weight).

# Figure 3

Box-plot diagrams of  $\Sigma REE$  (mean ± SEM) in the macro algae species (mg Kg<sup>-1</sup> dry weight) from the three examined locations.

# Figure 4

REE distribution (mg Kg<sup>-1</sup> dry weight) in *Flabellia petiolata*, *Codium bursa* and *Padina pavonica* in the three sampling sites.

# Figure 5

Chondrite (Leedey, Oklahoma) normalized patterns of REE in Northwestern Mediterranean macro algae (log scale).

# Figure 6

Bioconcentration factors in macro algae from the Northwestern Mediterranean Sea.