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Thymus atrophy is an efficient marker of illicit treatment with dexamethasone in veal calves: Results from a triennial experimental study

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(Article begins on next page)

1	Thymus atrophy is an efficient marker of illicit treatment with
2	dexamethasone in veal calves: results from a triennial experimental study
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4	Thymus and dexamethasone illicit treatments
5	
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15 Abstract

Glucocorticoids, used in a wide range of pathologies thank to their therapeutical properties, are also illegally used as growth-promoters in animal breeding even if the European Union regulates their use to protect consumers' health from the adverse effects of residues in food. The first aim of the study was to establish the applicability of two histological parameters – atrophy and cortex-medulla ratio - to detect glucocorticoids misuse in calves. The second aim was to test the potentiality of both parameters to discriminate between treated and untreated animals.

23 One hundred and seventy two male Friesian veal calves were raised for six months and

24 divided into two groups: Group A (106 calves) was given dexamethasone per os for twenty

25 days (0.4 mg/day), Group B (66 calves) used as control. Thymic samples were

26 microscopically examined. Fat infiltration was evaluated and a degree of atrophy from 1 to 3

27 (mild, moderate, severe) was attributed; thymic cortex-medulla ratio was calculated too.

28 Fisher's exact test and a Wilcoxon–Mann–Whitney test were performed to investigate the

29 differences in thymic atrophy and cortex-medulla ratio between the groups.

30 Results demonstrate that the thymic atrophy grading was significantly increased in group A

(p=0,006), whereas the cortex-medulla ratio was decreased (p<0,004) when compared to

32 group B; moreover, the parallel testing with fixed degree of atrophy and cortex-medulla ratio

cut-off thresholds optimize the sensitivity (90%) in the detection of glucocorticoids anabolictreatments.

35 These data suggest that microscopic thymus analysis represent a valid tool for the screening36 and monitoring of glucocorticoid illicit treatments.

37

39 <u>Keywords</u>: glucocorticoids; illegal treatment; dexamethasone; calves; thymus; histological
 40 screening method

41

42 Introduction

43 Glucocorticoids (GCs) are widely used in bovine internal medicine for their anti-

44 inflammatory properties, but are also illegally used, despite the European Union ban stated in

45 the Council Directive 2003/74, in food-producing animals as growth-promoters constituting a

46 health risk for the consumers (Botsoglou NA et al., 2001).

47 Natural corticosteroids are hormones secreted by the adrenal cortex that are involved in a

48 wide range of physiological processes, such as stress response, inflammation, immune

49 function, hydro electrolyte balance, reproduction, behaviour (Osamu N 2001; Schuerholz T

50 et al., 2007). The discovery of their anti-inflammatory properties has led to the chemical

51 synthesis of more active synthetic glucocorticoids, e.g. dexamethasone and prednisolone, that

52 are used as therapeutic drugs. In veterinary medicine, the legal utilization of these compounds

is strictly regulated, with withdrawal periods between treatment and slaughtering and

54 maximal residue levels (MRLs) established for some compounds (Commission Regulation

55 EU N. 37/2010).

56 In particular dexamethasone seems to be often involved in bovine illegal treatment protocols,

as confirmed by the technical report published by the European Food Safety Authority

58 (EFSA) on the "Results from the monitoring of veterinary medicinal product residues and

59 other substances in live animals and animal products" that summarises the monitoring data

60 collected in 2014 on the presence of residues of veterinary medicinal products in live animals

61 and animal products all over the European Union (EFSA Report 2014).

62 Dexamethasone, in fact, can be illegally used at very low dosage for a prolonged period of

time before slaughtering to obtain an enhancement of bovine carcass yield at slaughterhouse.

It can be illegally used alone or in association with other growth-promoting substances to
constitute unauthorized cocktails; the simultaneous administration of dexamethasone and
oestrogens or β-agonists to synergize their growth promoting effects is widely reported
(Abraham G. et al., 2004; Odore R. et al., 2007; Lopparelli RM et al., 2011).
Moreover this molecule, if fraudulently used, has the advantage of having a high and rapid
rate of urine excretion that makes it not detectable by the official analytical chemical methods
already a few days after the last administration (Vincenti M. et al., 2009; Ferranti C. et al.,

71 2011).

72 Given the powerful pharmacological action carried out by dexamethasone, the illegal use of 73 this molecule poses a serious issue for consumers' health: the possible accumulation of 74 residues of dexamethasone in edible organs of cattle is, in fact, potentially dangerous. Among 75 all the adverse effects, the most dangerous for human health include: the immunosuppressive 76 action and the alterations of glucose metabolism, with increased glycemia and resistance to 77 insulin (Lupu F. et al., 2001). Furthermore, the transfer of GCs and their metabolites through 78 the placental barrier to the foetus is associated to abortions, premature births, adrenal 79 insufficiency of the newborn and delays in skeletal growth and brain development (Allen DB 80 1996). Even the exposure through breast milk may hesitate in a series of negative effects, 81 such as decreased growth, metabolic disorders and bone mineralization impairment (Yeh TF. 82 et al., 2004).

For all the reasons stated above, many research groups are focusing their efforts in
overcoming the limits of official control methods through shifting the target from the
detection of the illegally-administrated molecules, to the study of their biological effects. New
approaches have been recently investigated: tissue and serum biomarkers, biosensors, -omic
techniques (Biancotto G. et al., 2013; Bovee TF. et al., 2013; Cacciatore G et al., 2009;

88 Courant F. et al., 2009; Divari S. et al., 2010; McGrath TF. et al., 2013; Nebbia C. et al.,

89 2011; Pezzolato M. et al., 2013; Pirro V. et al. 2015)

In the attempt to develop an accurate biological method to detect illicit glucocorticoids
treatments in food-producing animals, microscopic modifications of the thymus induced by
the administration of low-dose dexamethasone were preliminary investigated in veal calves.
The results of this previous works were obtained on a limited number of calves and data
published needed to be confirmed in order to be applicable in a routine-based workflow
(Biolatti B. et al., 2005; Bozzetta E. et al., 2011).

The main purpose of this experimental trial was to study thoroughly and concurrently 96 97 histological alterations, i.e. fat infiltration and cortex-medulla ratio, induced by low-dose 98 dexamethasone treatment as robust and reliable parameters to accurately evaluate the 99 performance of a relatively simple and not time consuming analytical approach, in order to 100 correctly identify treated calves. The objective was to implement the Italian Histological 101 Residues Control Plan that has been successfully applied since 2008 in the context of the 102 Italian Residues Control Plan. Therefore the final aim of this work is to protect the consumers 103 against the possible adverse effects caused by the ingestion of glucocorticoids' residues.

104

105 Materials and Methods

106 Animals and experimental design

The study was set up as a randomised controlled blind clinical trial. The whole experimental trial was carried out in accordance with the European Council Directive 86/609, recognised and adopted by the Italian Government (DLgs 27/01/1992 no. 116). The experiment was authorized by the Italian Ministry of Health and the Ethics Committee of the University of Turin. At the end of the sampling procedure, the carcasses of the treated animals were destroyed according to the law in force (Directive 2003/74/EC)

113 Due to the wide number of animals included in the experiment, the study was developed

114 during three years (from may to October) with three cycles of farming.

115 The sample size was calculated to detect a statistically significant difference in

116 Cortical/Medulla ratio between group A (treated) and group B (control) with a power of 95%

117 and level of significance of 5%.

118 Overall 180 male veal calves were recruited, randomly divided into two groups (group A and

B) and raised in multiple pens for 5 months under the same conditions. Each pen had its own

120 crib, multiple drinking troughs, and a dedicated automated milk feeder system. The calves

121 were vaccinated against Bovine Infectious Rhinotracheitis (IBR), Parainfluenza3 (PI3),

122 Bovine Syncytial (BRS) and Bovine Viral Diarrhoea Viruses (CATTLEMASTER® 4 Pfizer

123 Animal Health; New York, USA). Clinical controls were carried out daily by a veterinarian

and treatments for occurring infections were performed without using hormonally activesubstances.

126 The calves were fed through an automatic milk feeder; corn silage was increasingly added up

127 to 1 Kg/day during the fourth month according to the indications suggested by European

128 Commission Decision 97/182. Before administration, all feeds, milk replacer and corn were

129 analyzed with an Enzyme-Linked Immuno Assay (ELISA) to exclude the presence of

130 hormonally active substances.

131 During the sixth month, the animals without insurgence of clinical signs, hence did not

132 require medical treatments, entered the experiment (*n*=172). Calves belonging to Group A (*n*

133 = 106) were given a daily dose of 0,4 mg of dexamethasone-21-disodium-phosphate *per os*

134 *per capita* (dexadreson) for 20 consecutive days orally, according to a presumed anabolic

135 protocol of treatment. Animals belonging to Group B (n = 66) were used as control.

136 The animals were all slaughtered ten a day in an EC certified slaughterhouse about 10 days

137 after the last drug administration, control animals were slaughtered after the treated ones.

139 Histopathology

140 Sample preparation

141 At the slaughthouse the central portion of the thoracic thymus of each animal was sampled,

- 142 fixed in 4% buffered formaldehyde at room temperature for about three days, routinely
- 143 processed, embedded in a paraffin wax, sectioned in 3-5 µm slices and stained with

144 haematoxylin and eosin (HE).

145 Histopathological characterization of thymus atrophy

146 The morphology of the thymus parenchyma was evaluated by two expert pathologists using

147 light microscopy in two different session works and in blind.

The presence of adipose tissue, as indirect marker of thymus atrophy, was evaluated by light microscopy at low magnification (1x and 4x) and a grading was attributed to the amount of fat infiltration: grade 1 was attributed to minimal or mild invasion of adipose tissue localized within the thymus septa; grade 2 was attributed to moderate invasion of adipose tissue in septa with minimal invasion of cortex part of the thymus; grade 3 was attributed to severe invasion of adipose tissue in the cortex of the thymus with invasion of the medullar part (Fig.

154 1).

155 Morphometry

156 The thymus sections were also examined at low magnification (4x) using a digital

157 microimaging device (Leica DMD108 Digital micro imaging device for clinical diagnostics

158 labs) to evaluate cortex and medulla thickness. Assuming that the lobule is the morpho-

159 functional unit of the thymus and that every lobule is composed of an outer cortex and inner

- 160 medulla and surrounded by connective tissue, for each slide, five lobules were randomly
- 161 selected and measured against a graduated line, starting and ending at level of the interlobular
- 162 connective tissue; a second line was drawn just in correspondence of the first to measure

163 medullar diameter (Fig 2). Cortex thickness was obtained by subtracting the medullar

diameter from the corresponding diameter of the entire lobule, then the cortex-medulla ratiowas calculated.

166 All measures were recorded on a spreadsheet.

167 Statistical analysis

168 A descriptive analysis on the outcome variables was carried out.

169 The association between thymus fat score and group (Group A vs Group B) was assessed by170 Fisher's exact test.

171 In order to fit the Gaussian distribution of the data, a logarithmic transformation was applied.

172 Then, the distribution of cortex-medulla ratio (C/M ratio), of the cortex and of the medulla

thickness among groups was evaluated using a linear mixed model. A mixed-effects model

174 consists of two parts, fixed effects and random effects. Fixed-effects terms are usually the

175 conventional linear regression part and are not modelled, and the random effects represent the

176 varying coefficients and they are associated with individual experimental units drawn at

177 random from a population. It is a term that refers to the randomness in the probability model

178 for the group-level coefficients. The fixed effect was the group of animals while the random

179 effect was the animal nested within group; lobules, collected for each animal, represented the

180 residual. The analysis of residuals was adopted for checking the models.

181 To verify the presence of a significant difference in the degree of thymic atrophy between the 182 two groups, a test for linear trend was performed and the difference between three increasing 183 degrees of atrophy was expressed as Odds Ratios (OR).

184 Fixed cutoff thresholds optimizing sensitivity and specificity values for thymic degree of

185 atrophy and cortex-medulla ratio were calculated using the Receiver Operating Characteristic

186 (ROC) curves which plot sensitivity against (1 – specificity) across all possible cut-off

187 thresholds. The combination of the two histological tests was evaluated using parallel test

188	interpretation and assuming conditionally dependence of the two binary tests (Gardner IA. Et
189	al., 2000). The extent of the dependence has been calculated estimating the covariance
190	between the two test results. The covariance in the two groups has been calculated as
191	suggested by Dohoo et al. (2009).
192	All data analyses (thymus atrophy, cortex thickness, medulla thickness and cortex-medulla
193	ratio) were performed using Stata 11.2 (Copyright 1985-2013 StataCorp LP Statistics/Data
194	Analysis StataCorp). The P value of 0.05 was considered as the level of statistical
195	significance.
196	

197 **Results**

One hundred and seventy-two animals concluded the trial without insurgence of clinical signsor requiring medical treatment and entered the data analysis process.

200 Histopathological characterization of thymus atrophy

201 The results obtained by the microscopic observation showed that the scores of 1, mild, and 2,

202 moderate infiltration of fat, were detected in both treated and control animals, whereas the

score of 3, severe infiltration of fat, even if detected in both group was mainly attributed to

the group of the treated animals and recorded in only four controls.

205 The amount of fat infiltrating the thymic tissue resulted significantly associated with the

treated group by Fisher's exact test (p=0.001), even if the trend was not linear as shown in

207 Table 1 in terms of Odds Ratio.

The Cohen's Kappa of 0.78 (95% CI = 0.69 - 0.88) established a good repeatability among
pathologists .

210 **Comparative morphometry**

211 Microscopic examination at a high magnification showed a decrease in the cortical area of the

treated animals verified by measured recorded results.

213 Descriptive statistics of cortex thickness, medulla thickness and cortex-medulla ratio (C/M
214 ratio) stratified by treatment group are provided in Table 2.

The distribution of the C/M ratio between the two groups is provided in Graph 1: a

significantly different distribution between the two groups (p<0.0001) was evident.

Table 3 shows the results of the comparison performed by the linear regression model. The

218 three parameters taken into account (Cortex thickness, medulla thickness and C/M ratio)

219 resulted significantly different between the two groups; In particular the model results put in

220 evidence that the C/M ratio is one-third lower in the treated animals compared to the control

221 ones.

222 To assess the combined performance of thymic degree of atrophy and cortex-medulla ratio to

detect glucocorticoids illicit treatments the ROC curves were examined to select optimal fixed

224 cutoff thresholds for each histological test. In table 4 test sensitivity, specificity and the cutoff

values are reported for the two histological methods. Parallel testing, with fixed cutoff

thresholds of ≥ 2 thymic degree of atrophy and ≤ 1.24 cortex-medulla ratio, increases

227 sensitivity to 88% (95% CI 80%-93%) at the expense of specificity (63% - 95% CI:54% -

228 78%).

Considering the conditionally dependence of the two tests, the sensitivity and specificity are,respectively, 90% and 61%.

231

232 **Discussion**

Our findings confirm that the administration of low-dose dexamethasone as a growth promoter in veal calves, according to a protocol often illegally adopted in farm practice (Biolatti B. et al., 2005), can induce morphologic changes in the thymus, resulting in a significant reduction in the cortex-medulla ratio (Bozzetta E. et al., 2011). In 1998 Schilt (Schilt R . et al., 1998) first reported this finding as a valuable feature to distinguish between beclomethasone treated and

238 untreated veal calves. Later Elmore in 2006 included this parameter among criteria to 239 distinguish between chemical-induced and age or stress-related thymic atrophy. Here we show 240 the performance of this parameter calculated on a large number of animals raised under 241 controlled conditions. Cortical atrophy and fat infiltration has been previously reported by 242 several research groups (Biolatti B, et al., 2005; Cannizzo FT. et al., 2008; Groot MJ. Et al., 243 1998) as co-existing finding in dexamethasone-fed veal calves; our experiment confirms that 244 fat infiltration and cortex-medulla ratio could be associated with low-dose dexamethasone 245 treatment, with high statistical support.

246 Prolonged treatment with glucocorticoids at low doses causes perceivable changes in thymus247 architecture.

In a previous work, two histological parameters i.e. fat infiltration and cortex-medulla ratio,

249 were investigated and results showed that the cortex-medulla ratio performed well in

250 discriminating treated versus untreated calves (Bozzetta E, et al., 2011). Nevertheless, in

251 contrast with data reported in literature, no statistically significant differences were found

regarding fat infiltration between the two groups (Biolatti B. et al., 2005; Cannizzo FT. et al.,

253 2010; Vascellari M. et al., 2012; Groot MJ, et al., 1998)

254 This work was set up in order to investigate the performance of the histological method in order 255 to evaluate its efficacy as screening method in the field of the histological part of the Italian 256 Residues Control Plan. Final results from the analyses of all animals' thymus strengthened the 257 preliminary results obtained on the reliability of cortex-medulla ratio, that was decreased in 258 treated animals according to the results obtained by Bozzetta et al., 2011 even if in the latter 259 thymus atrophy of treated calves was not supported by the statistical analysis. At the end of our 260 triennial experimental study results are different and the statistical analysis confirms that 261 treated calves can be successfully separated by untreated animals considering the atrophy 262 parameter.

The parallel testing analysis allowed to evaluate the detection performance for the combinedevaluation of the thymus degree of atrophy and cortex-medulla ratio.

Parallel testing with fixed cutoff thresholds of ≥ 2 for thymus degree of atrophy and ≤ 1.24 for cortex-medulla ratio maximizes the ability to detect anabolic treatments increasing the sensitivity of the method. The results of the combined employment of the two parameters shows the possibility to use the parallel analysis in the screening phase of the glucocorticoids illicit treatments monitoring.

270

271 Conclusion

This scientific and statistically robust finding can finally confirm that this simple parameter is able to distinguish the presence of illicit treatment achieved with synthetic glucocorticoids and can be successfully applied in the Italian Residue Control Plan. The European Food Safety Agency recommends that all the efforts to improve the control of illegal use of growth promoters in cattle should include novel screening methods, able to highlight the biological effects of growth promoters in livestock rather than measuring their residues concentration in food or feed (European Food Safety Authority 2013).

For that reason and based on these results we can conclude that histological investigations i.e.

280 specifically, the assessment of thymus morphometry represent a standardized and powerful

- 281 method for screening purposes.
- 282

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285

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- **Table 1. The linear trend in the degree of thymic atrophy. The trend is not statistically**
- 391 significant. Score test for trend of Odds p=0.26

FAT SCORE	OR	CI95%	
1	-	-	1
2	1.43	0.09	2
3	5.63	0.32	3

Table 2. Descriptive statistics for the parameters of thymus samples

PARAMETER	GROUP	MEAN	MEDIAN	SD	Min	Max
Cortex thickness	Treated	666.9	634.8	261.1	153.8	1795
	Control	909	858.4	356.1	169.6	2175.4
Medulla thickness	Treated	685	610.3	297.8	81.4	2100
	Control	581.1	550	208.6	117.5	1300
Cortex-Medulla ratio	Treated	1.1	1	0.6	0.2	5.3
	Control	1.7	1.5	0.9	0.3	7.5

- **Table 3. Results of the linear mixed models. The dependent variables are the features of**
- 400 the thymus and are expressed as log. All the variables resulted significantly different
- **between the two groups.**

DEPENDENT VARIABLE	ESTIMATE	STANDARD ERROR	ADJUSTED <i>p</i>
Cortex thickness	-0.31	0.34	0.000
Medulla thickness	0.15	0.32	0.000
Cortex-Medulla ratio	-0.47	0.44	0.000

403 Table 4. Sensitivity and specificity of the thymic degree of atrophy test and cortex-

404 medulla ratio test, including 95% CIs and cutoff thresholds values

DIAGNOSTIC		Sensitivity		Specificity	CUTOFF
METHOD	Sensitivity	CI 95%	Specificity	CI 95%	
Thymic degree of atrophy	76%	66% - 83%	82%	70% - 90%	≥2
Cortex-medulla ratio	52%	42% - 62%	77%	65%-87%	≤1,.24

- 407 Graph 1. distribution of the C/M ratio values between the two groups. Outliers have
- 408 been excluded. The difference is resulted statistically significant (p < 0.0001).



413 Figures captures

- 414 Fig. 1. Thymus atrophy grading; mild (A), moderate (B), severe (C).
- 415 Fig. 2. Thymic morphometry: Extension of the cortex was measured against a graduated line
- 416 (red) starting and ending at the interlobular connective or adipose tissue; a second parallel line
- 417 was drawn to measure medulla thickness (green) (HE 4X).