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**Thymus atrophy is an efficient marker of illicit treatment with dexamethasone in veal calves:  
Results from a triennial experimental study**

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1 **Thymus atrophy is an efficient marker of illicit treatment with**  
2 **dexamethasone in veal calves: results from a triennial experimental study**

3

4 **Thymus and dexamethasone illicit treatments**

5

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15 **Abstract**

16 Glucocorticoids, used in a wide range of pathologies thank to their therapeutical properties,  
17 are also illegally used as growth-promoters in animal breeding even if the European Union  
18 regulates their use to protect consumers' health from the adverse effects of residues in food.  
19 The first aim of the study was to establish the applicability of two histological parameters –  
20 atrophy and cortex-medulla ratio - to detect glucocorticoids misuse in calves. The second aim  
21 was to test the potentiality of both parameters to discriminate between treated and untreated  
22 animals.

23 One hundred and seventytwo male Friesian veal calves were raised for six months and  
24 divided into two groups: Group A (106 calves) was given dexamethasone *per os* for twenty  
25 days (0.4 mg/day), Group B (66 calves) used as control. Thymic samples were  
26 microscopically examined. Fat infiltration was evaluated and a degree of atrophy from 1 to 3  
27 (mild, moderate, severe) was attributed; thymic cortex-medulla ratio was calculated too.  
28 Fisher's exact test and a Wilcoxon–Mann–Whitney test were performed to investigate the  
29 differences in thymic atrophy and cortex-medulla ratio between the groups.

30 Results demonstrate that the thymic atrophy grading was significantly increased in group A  
31 ( $p= 0,006$ ), whereas the cortex-medulla ratio was decreased ( $p<0,004$ ) when compared to  
32 group B; moreover, the parallel testing with fixed degree of atrophy and cortex-medulla ratio  
33 cut-off thresholds optimize the sensitivity (90%) in the detection of glucocorticoids anabolic  
34 treatments.

35 These data suggest that microscopic thymus analysis represent a valid tool for the screening  
36 and monitoring of glucocorticoid illicit treatments.

37

38

39 **Keywords:** glucocorticoids; illegal treatment; dexamethasone; calves; thymus; histological  
40 screening method

41

## 42 **Introduction**

43 Glucocorticoids (GCs) are widely used in bovine internal medicine for their anti-  
44 inflammatory properties, but are also illegally used, despite the European Union ban stated in  
45 the Council Directive 2003/74, in food-producing animals as growth-promoters constituting a  
46 health risk for the consumers (Botsoglou NA et al., 2001).

47 Natural corticosteroids are hormones secreted by the adrenal cortex that are involved in a  
48 wide range of physiological processes, such as stress response, inflammation, immune  
49 function, hydro electrolyte balance, reproduction, behaviour (Osamu N 2001; Schuerholz T  
50 et al., 2007). The discovery of their anti-inflammatory properties has led to the chemical  
51 synthesis of more active synthetic glucocorticoids, e.g. dexamethasone and prednisolone, that  
52 are used as therapeutic drugs. In veterinary medicine, the legal utilization of these compounds  
53 is strictly regulated, with withdrawal periods between treatment and slaughtering and  
54 maximal residue levels (MRLs) established for some compounds (Commission Regulation  
55 EU N. 37/2010).

56 In particular dexamethasone seems to be often involved in bovine illegal treatment protocols,  
57 as confirmed by the technical report published by the European Food Safety Authority  
58 (EFSA) on the “Results from the monitoring of veterinary medicinal product residues and  
59 other substances in live animals and animal products” that summarises the monitoring data  
60 collected in 2014 on the presence of residues of veterinary medicinal products in live animals  
61 and animal products all over the European Union (EFSA Report 2014) .

62 Dexamethasone, in fact, can be illegally used at very low dosage for a prolonged period of  
63 time before slaughtering to obtain an enhancement of bovine carcass yield at slaughterhouse.

64 It can be illegally used alone or in association with other growth-promoting substances to  
65 constitute unauthorized cocktails; the simultaneous administration of dexamethasone and  
66 oestrogens or  $\beta$ -agonists to synergize their growth promoting effects is widely reported  
67 (Abraham G. et al., 2004; Odore R. et al., 2007; Lopparelli RM et al., 2011).

68 Moreover this molecule, if fraudulently used, has the advantage of having a high and rapid  
69 rate of urine excretion that makes it not detectable by the official analytical chemical methods  
70 already a few days after the last administration (Vincenti M. et al., 2009; Ferranti C. et al.,  
71 2011).

72 Given the powerful pharmacological action carried out by dexamethasone, the illegal use of  
73 this molecule poses a serious issue for consumers' health: the possible accumulation of  
74 residues of dexamethasone in edible organs of cattle is, in fact, potentially dangerous. Among  
75 all the adverse effects, the most dangerous for human health include: the immunosuppressive  
76 action and the alterations of glucose metabolism, with increased glycemia and resistance to  
77 insulin (Lupu F. et al., 2001). Furthermore, the transfer of GCs and their metabolites through  
78 the placental barrier to the foetus is associated to abortions, premature births, adrenal  
79 insufficiency of the newborn and delays in skeletal growth and brain development (Allen DB  
80 1996). Even the exposure through breast milk may hesitate in a series of negative effects,  
81 such as decreased growth, metabolic disorders and bone mineralization impairment (Yeh TF.  
82 et al., 2004).

83 For all the reasons stated above, many research groups are focusing their efforts in  
84 overcoming the limits of official control methods through shifting the target from the  
85 detection of the illegally-administrated molecules, to the study of their biological effects. New  
86 approaches have been recently investigated: tissue and serum biomarkers, biosensors, -omic  
87 techniques (Biancotto G. et al., 2013; Bovee TF. et al., 2013; Cacciatore G et al., 2009;

88 Courant F. et al., 2009 ; Divari S. et al., 2010; McGrath TF. et al., 2013; Nebbia C. et al.,  
89 2011; Pezzolato M. et al., 2013; Pirro V. et al. 2015)  
90 In the attempt to develop an accurate biological method to detect illicit glucocorticoids  
91 treatments in food-producing animals, microscopic modifications of the thymus induced by  
92 the administration of low-dose dexamethasone were preliminary investigated in veal calves.  
93 The results of this previous works were obtained on a limited number of calves and data  
94 published needed to be confirmed in order to be applicable in a routine-based workflow  
95 (Biolatti B. et al., 2005; Bozzetta E. et al., 2011).  
96 The main purpose of this experimental trial was to study thoroughly and concurrently  
97 histological alterations, i.e. fat infiltration and cortex-medulla ratio, induced by low-dose  
98 dexamethasone treatment as robust and reliable parameters to accurately evaluate the  
99 performance of a relatively simple and not time consuming analytical approach, in order to  
100 correctly identify treated calves. The objective was to implement the Italian Histological  
101 Residues Control Plan that has been successfully applied since 2008 in the context of the  
102 Italian Residues Control Plan. Therefore the final aim of this work is to protect the consumers  
103 against the possible adverse effects caused by the ingestion of glucocorticoids' residues.

104

## 105 **Materials and Methods**

### 106 **Animals and experimental design**

107 The study was set up as a randomised controlled blind clinical trial. The whole experimental  
108 trial was carried out in accordance with the European Council Directive 86/609, recognised  
109 and adopted by the Italian Government (DLgs 27/01/1992 no. 116). The experiment was  
110 authorized by the Italian Ministry of Health and the Ethics Committee of the University of  
111 Turin. At the end of the sampling procedure, the carcasses of the treated animals were  
112 destroyed according to the law in force (Directive 2003/74/EC)

113 Due to the wide number of animals included in the experiment, the study was developed  
114 during three years (from may to October) with three cycles of farming.  
115 The sample size was calculated to detect a statistically significant difference in  
116 Cortical/Medulla ratio between group A (treated) and group B (control) with a power of 95%  
117 and level of significance of 5%.

118 Overall 180 male veal calves were recruited, randomly divided into two groups (group A and  
119 B) and raised in multiple pens for 5 months under the same conditions. Each pen had its own  
120 crib, multiple drinking troughs, and a dedicated automated milk feeder system. The calves  
121 were vaccinated against Bovine Infectious Rhinotracheitis (IBR), Parainfluenza3 (PI3),  
122 Bovine Syncytial (BRS) and Bovine Viral Diarrhoea Viruses (CATTLEMASTER® 4 Pfizer  
123 Animal Health; New York, USA). Clinical controls were carried out daily by a veterinarian  
124 and treatments for occurring infections were performed without using hormonally active  
125 substances.

126 The calves were fed through an automatic milk feeder; corn silage was increasingly added up  
127 to 1 Kg/day during the fourth month according to the indications suggested by European  
128 Commission Decision 97/182. Before administration, all feeds, milk replacer and corn were  
129 analyzed with an Enzyme-Linked Immuno Assay (ELISA) to exclude the presence of  
130 hormonally active substances.

131 During the sixth month, the animals without insurgence of clinical signs, hence did not  
132 require medical treatments, entered the experiment ( $n=172$ ). Calves belonging to Group A ( $n$   
133 = 106) were given a daily dose of 0,4 mg of dexamethasone-21-disodium-phosphate *per os*  
134 *per capita* (dexadreson) for 20 consecutive days orally, according to a presumed anabolic  
135 protocol of treatment. Animals belonging to Group B ( $n = 66$ ) were used as control.

136 The animals were all slaughtered ten a day in an EC certified slaughterhouse about 10 days  
137 after the last drug administration, control animals were slaughtered after the treated ones.

138

## 139 **Histopathology**

### 140 **Sample preparation**

141 At the slaughterhouse the central portion of the thoracic thymus of each animal was sampled,  
142 fixed in 4% buffered formaldehyde at room temperature for about three days, routinely  
143 processed, embedded in a paraffin wax, sectioned in 3-5  $\mu\text{m}$  slices and stained with  
144 haematoxylin and eosin (HE).

### 145 **Histopathological characterization of thymus atrophy**

146 The morphology of the thymus parenchyma was evaluated by two expert pathologists using  
147 light microscopy in two different session works and in blind.

148 The presence of adipose tissue, as indirect marker of thymus atrophy, was evaluated by light  
149 microscopy at low magnification (1x and 4x) and a grading was attributed to the amount of fat  
150 infiltration: grade 1 was attributed to minimal or mild invasion of adipose tissue localized  
151 within the thymus septa; grade 2 was attributed to moderate invasion of adipose tissue in  
152 septa with minimal invasion of cortex part of the thymus; grade 3 was attributed to severe  
153 invasion of adipose tissue in the cortex of the thymus with invasion of the medullar part (Fig.  
154 1).

### 155 **Morphometry**

156 The thymus sections were also examined at low magnification (4x) using a digital  
157 microimaging device (Leica DMD108 Digital micro imaging device for clinical diagnostics  
158 labs) to evaluate cortex and medulla thickness. Assuming that the lobule is the morpho-  
159 functional unit of the thymus and that every lobule is composed of an outer cortex and inner  
160 medulla and surrounded by connective tissue, for each slide, five lobules were randomly  
161 selected and measured against a graduated line, starting and ending at level of the interlobular  
162 connective tissue; a second line was drawn just in correspondence of the first to measure



163 medullar diameter (Fig 2). Cortex thickness was obtained by subtracting the medullar  
164 diameter from the corresponding diameter of the entire lobule, then the cortex-medulla ratio  
165 was calculated.

166 All measures were recorded on a spreadsheet.

### 167 **Statistical analysis**

168 A descriptive analysis on the outcome variables was carried out.

169 The association between thymus fat score and group (Group A vs Group B ) was assessed by  
170 Fisher's exact test.

171 In order to fit the Gaussian distribution of the data, a logarithmic transformation was applied.

172 Then, the distribution of cortex-medulla ratio (C/M ratio), of the cortex and of the medulla  
173 thickness among groups was evaluated using a linear mixed model. A mixed-effects model  
174 consists of two parts, fixed effects and random effects. Fixed-effects terms are usually the  
175 conventional linear regression part and are not modelled, and the random effects represent the  
176 varying coefficients and they are associated with individual experimental units drawn at  
177 random from a population. It is a term that refers to the randomness in the probability model  
178 for the group-level coefficients. The fixed effect was the group of animals while the random  
179 effect was the animal nested within group; lobules, collected for each animal, represented the  
180 residual. The analysis of residuals was adopted for checking the models.

181 To verify the presence of a significant difference in the degree of thymic atrophy between the  
182 two groups, a test for linear trend was performed and the difference between three increasing  
183 degrees of atrophy was expressed as Odds Ratios (OR).

184 Fixed cutoff thresholds optimizing sensitivity and specificity values for thymic degree of  
185 atrophy and cortex-medulla ratio were calculated using the Receiver Operating Characteristic  
186 (ROC) curves which plot sensitivity against (1 – specificity) across all possible cut-off  
187 thresholds. The combination of the two histological tests was evaluated using parallel test

188 interpretation and assuming conditionally dependence of the two binary tests (Gardner IA. Et  
189 al., 2000). The extent of the dependence has been calculated estimating the covariance  
190 between the two test results. The covariance in the two groups has been calculated as  
191 suggested by Dohoo et al. (2009).

192 All data analyses (thymus atrophy, cortex thickness, medulla thickness and cortex-medulla  
193 ratio) were performed using Stata 11.2 (Copyright 1985-2013 StataCorp LP Statistics/Data  
194 Analysis StataCorp). The P value of 0.05 was considered as the level of statistical  
195 significance.

196

## 197 **Results**

198 One hundred and seventy-two animals concluded the trial without insurgence of clinical signs  
199 or requiring medical treatment and entered the data analysis process.

### 200 **Histopathological characterization of thymus atrophy**

201 The results obtained by the microscopic observation showed that the scores of 1, mild, and 2,  
202 moderate infiltration of fat, were detected in both treated and control animals, whereas the  
203 score of 3, severe infiltration of fat, even if detected in both group was mainly attributed to  
204 the group of the treated animals and recorded in only four controls.

205 The amount of fat infiltrating the thymic tissue resulted significantly associated with the  
206 treated group by Fisher's exact test ( $p=0.001$ ), even if the trend was not linear as shown in  
207 Table 1 in terms of Odds Ratio.

208 The Cohen's Kappa of 0.78 (95% CI = 0.69 - 0.88) established a good repeatability among  
209 pathologists .

### 210 **Comparative morphometry**

211 Microscopic examination at a high magnification showed a decrease in the cortical area of the  
212 treated animals verified by measured recorded results.

213 Descriptive statistics of cortex thickness, medulla thickness and cortex-medulla ratio (C/M  
214 ratio) stratified by treatment group are provided in Table 2.

215 The distribution of the C/M ratio between the two groups is provided in Graph 1: a  
216 significantly different distribution between the two groups ( $p < 0.0001$ ) was evident.

217 Table 3 shows the results of the comparison performed by the linear regression model. The  
218 three parameters taken into account (Cortex thickness, medulla thickness and C/M ratio)  
219 resulted significantly different between the two groups; In particular the model results put in  
220 evidence that the C/M ratio is one-third lower in the treated animals compared to the control  
221 ones.

222 To assess the combined performance of thymic degree of atrophy and cortex-medulla ratio to  
223 detect glucocorticoids illicit treatments the ROC curves were examined to select optimal fixed  
224 cutoff thresholds for each histological test. In table 4 test sensitivity, specificity and the cutoff  
225 values are reported for the two histological methods. Parallel testing, with fixed cutoff  
226 thresholds of  $\geq 2$  thymic degree of atrophy and  $\leq 1.24$  cortex-medulla ratio, increases  
227 sensitivity to 88% (95% CI 80%-93%) at the expense of specificity (63% - 95%CI:54% -  
228 78%).

229 Considering the conditionally dependence of the two tests, the sensitivity and specificity are,  
230 respectively, 90% and 61%.

231

## 232 **Discussion**

233 Our findings confirm that the administration of low-dose dexamethasone as a growth promoter  
234 in veal calves, according to a protocol often illegally adopted in farm practice (Biolatti B. et al.,  
235 2005), can induce morphologic changes in the thymus, resulting in a significant reduction in the  
236 cortex-medulla ratio (Bozzetta E. et al., 2011). In 1998 Schilt (Schilt R. et al., 1998) first  
237 reported this finding as a valuable feature to distinguish between beclomethasone treated and

238 untreated veal calves. Later Elmore in 2006 included this parameter among criteria to  
239 distinguish between chemical-induced and age or stress-related thymic atrophy. Here we show  
240 the performance of this parameter calculated on a large number of animals raised under  
241 controlled conditions. Cortical atrophy and fat infiltration has been previously reported by  
242 several research groups (Biolatti B, et al., 2005; Cannizzo FT. et al., 2008; Groot MJ. Et al.,  
243 1998) as co-existing finding in dexamethasone-fed veal calves; our experiment confirms that  
244 fat infiltration and cortex-medulla ratio could be associated with low-dose dexamethasone  
245 treatment, with high statistical support.

246 Prolonged treatment with glucocorticoids at low doses causes perceivable changes in thymus  
247 architecture.

248 In a previous work, two histological parameters i.e. fat infiltration and cortex-medulla ratio,  
249 were investigated and results showed that the cortex-medulla ratio performed well in  
250 discriminating treated versus untreated calves (Bozzetta E, et al., 2011). Nevertheless, in  
251 contrast with data reported in literature, no statistically significant differences were found  
252 regarding fat infiltration between the two groups (Biolatti B. et al., 2005; Cannizzo FT. et al.,  
253 2010; Vascellari M. et al., 2012; Groot MJ, et al., 1998)

254 This work was set up in order to investigate the performance of the histological method in order  
255 to evaluate its efficacy as screening method in the field of the histological part of the Italian  
256 Residues Control Plan. Final results from the analyses of all animals' thymus strengthened the  
257 preliminary results obtained on the reliability of cortex-medulla ratio, that was decreased in  
258 treated animals according to the results obtained by Bozzetta et al., 2011 even if in the latter  
259 thymus atrophy of treated calves was not supported by the statistical analysis. At the end of our  
260 triennial experimental study results are different and the statistical analysis confirms that  
261 treated calves can be successfully separated by untreated animals considering the atrophy  
262 parameter.

263 The parallel testing analysis allowed to evaluate the detection performance for the combined  
264 evaluation of the thymus degree of atrophy and cortex-medulla ratio.

265 Parallel testing with fixed cutoff thresholds of  $\geq 2$  for thymus degree of atrophy and  $\leq 1.24$  for  
266 cortex-medulla ratio maximizes the ability to detect anabolic treatments increasing the  
267 sensitivity of the method. The results of the combined employment of the two parameters  
268 shows the possibility to use the parallel analysis in the screening phase of the glucocorticoids  
269 illicit treatments monitoring.

270

## 271 **Conclusion**

272 This scientific and statistically robust finding can finally confirm that this simple parameter is  
273 able to distinguish the presence of illicit treatment achieved with synthetic glucocorticoids and  
274 can be successfully applied in the Italian Residue Control Plan. The European Food Safety  
275 Agency recommends that all the efforts to improve the control of illegal use of growth  
276 promoters in cattle should include novel screening methods, able to highlight the biological  
277 effects of growth promoters in livestock rather than measuring their residues concentration in  
278 food or feed (European Food Safety Authority 2013).

279 For that reason and based on these results we can conclude that histological investigations i.e.  
280 specifically, the assessment of thymus morphometry represent a standardized and powerful  
281 method for screening purposes.

282

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285

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288

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388

389

390 **Table 1. The linear trend in the degree of thymic atrophy. The trend is not statistically**

391 **significant. Score test for trend of Odds  $p=0.26$**

392

| FAT SCORE | OR   | CI95% |   |
|-----------|------|-------|---|
| 1         | -    | -     | 1 |
| 2         | 1.43 | 0.09  | 2 |
| 3         | 5.63 | 0.32  | 3 |

393

394

395

396 **Table 2. Descriptive statistics for the parameters of thymus samples**

397

| PARAMETER            | GROUP   | MEAN  | MEDIAN | SD    | Min   | Max    |
|----------------------|---------|-------|--------|-------|-------|--------|
| Cortex thickness     | Treated | 666.9 | 634.8  | 261.1 | 153.8 | 1795   |
|                      | Control | 909   | 858.4  | 356.1 | 169.6 | 2175.4 |
| Medulla thickness    | Treated | 685   | 610.3  | 297.8 | 81.4  | 2100   |
|                      | Control | 581.1 | 550    | 208.6 | 117.5 | 1300   |
| Cortex-Medulla ratio | Treated | 1.1   | 1      | 0.6   | 0.2   | 5.3    |
|                      | Control | 1.7   | 1.5    | 0.9   | 0.3   | 7.5    |

398

399 **Table 3. Results of the linear mixed models. The dependent variables are the features of**  
400 **the thymus and are expressed as log. All the variables resulted significantly different**  
401 **between the two groups.**

402

| DEPENDENT VARIABLE   | ESTIMATE | STANDARD ERROR | ADJUSTED <i>p</i> |
|----------------------|----------|----------------|-------------------|
| Cortex thickness     | -0.31    | 0.34           | 0.000             |
| Medulla thickness    | 0.15     | 0.32           | 0.000             |
| Cortex-Medulla ratio | -0.47    | 0.44           | 0.000             |

403 **Table 4. Sensitivity and specificity of the thymic degree of atrophy test and cortex-**  
 404 **medulla ratio test, including 95% CIs and cutoff thresholds values**  
 405

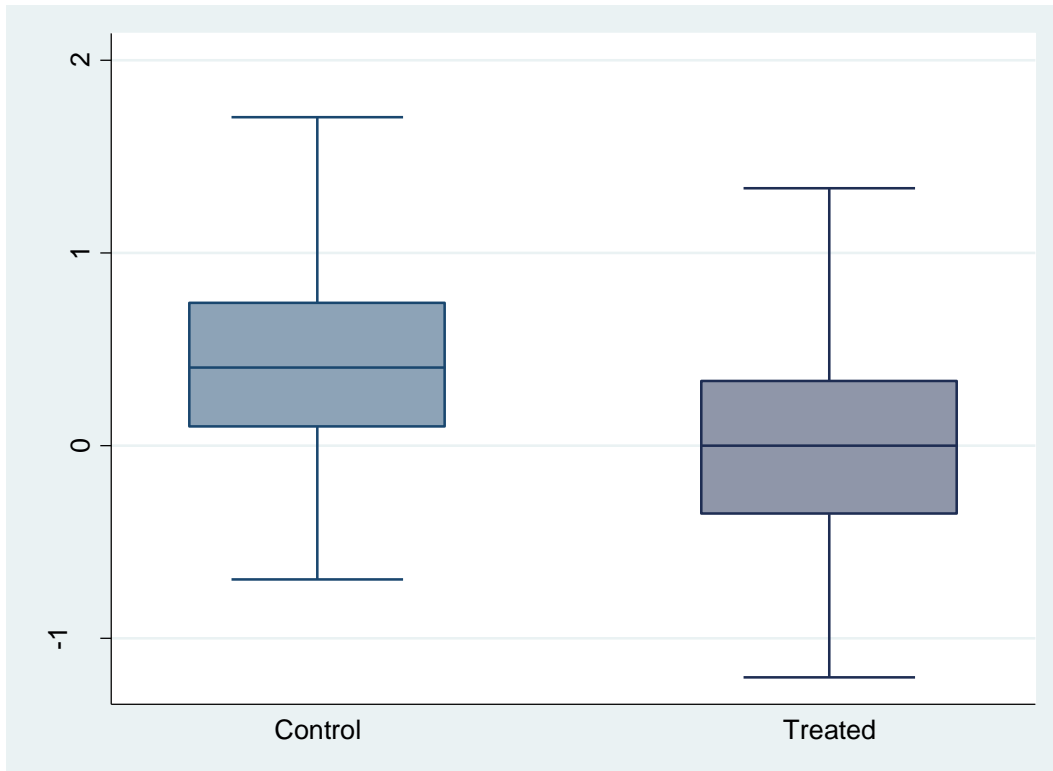
| DIAGNOSTIC METHOD        | Sensitivity | Sensitivity CI 95% | Specificity | Specificity CI 95% | CUTOFF      |
|--------------------------|-------------|--------------------|-------------|--------------------|-------------|
| Thymic degree of atrophy | 76%         | 66% - 83%          | 82%         | 70% - 90%          | $\geq 2$    |
| Cortex-medulla ratio     | 52%         | 42% - 62%          | 77%         | 65%-87%            | $\leq 1,24$ |

406

407 **Graph 1. distribution of the C/M ratio values between the two groups. Outliers have**

408 **been excluded. The difference is resulted statistically significant ( $p < 0.0001$ ).**

409



410

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412

413 **Figures captures**

414 Fig. 1. Thymus atrophy grading; mild (A), moderate (B), severe (C).

415 Fig. 2. Thymic morphometry: Extension of the cortex was measured against a graduated line  
416 (red) starting and ending at the interlobular connective or adipose tissue; a second parallel line  
417 was drawn to measure medulla thickness (green) (HE 4X).