

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Silica nanoparticles actively engage with mesenchymal stem cells in improving acute functional cardiac integration

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1676901> since 2019-05-09T22:35:49Z

Published version:

DOI:10.2217/nnm-2017-0309

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

Silica nanoparticles actively engage with mesenchymal stem cells in improving acute functional cardiac integration

Jasmin Popara†, Lisa Accomasso†, Emanuela Vitale, Clara Gallina, Dorotea Roggio, Ambra Iannuzzi, Stefania Raimondo, Raffaella Rastaldo, Gabriele Alberto, Federico Catalano, Gianmario Martra, Valentina Turinetti, Pasquale Pagliaro & Claudia Giachino

Published Online: 8 Jun 2018 <https://doi.org/10.2217/nnm-2017-0309>

Aim: To assess functional effects of silica nanoparticles (SiO₂-NPs) on human mesenchymal stem cell (hMSC) cardiac integration potential.

Methods: SiO₂-NPs were synthesized and their internalization effects on hMSCs analyzed with particular emphasis on interaction of hMSCs with the cardiac environment

Results: SiO₂-NP internalization affected the area and maturation level of hMSC focal adhesions, accounting for increased *in vitro* adhesion capacity and augmented engraftment in the myocardial tissue upon cell injection in infarcted isolated rat hearts. SiO₂-NP treatment also enhanced hMSC expression of Connexin-43, favoring hMSC interaction with cocultured cardiac myoblasts in an ischemia-like environment.

Conclusion: These findings provide strong evidence that SiO₂-NPs actively engage in mediating biological effects, ultimately resulting in augmented hMSC acute cardiac integration potential.

Keywords:

cell adhesion, gap junction, heart, mesenchymal stem cell, regeneration, silica nanoparticle

Human mesenchymal stem cells (hMSCs) represent an attractive therapeutic source as they appear to have beneficial effects in regenerating injured tissues [1], including the cardiac tissue [2,3].

Although both preclinical and clinical studies suggest the therapeutic potential of hMSCs, several questions remain open. An important limitation of hMSCs is the low survival rate within the first 1–2 weeks after transplantation, due to anoikis in damaged tissues [4]. Anoikis is a programmed cell death that occurs in anchorage-dependent cells when they detach from the extracellular matrix (ECM) [5,6]. Indeed, a loss of matrix anchorage and a subsequent low propensity to adhere may cause the anoikis of the transplanted MSCs. Therefore, to enhance cell survival in the site of transplantation, one should enhance the adhesion of the transplanted MSCs to limit anoikis. This might improve the success of the MSC-based therapeutic approach. Although a number of studies aim to improve MSC survival at the site of transplantation, yet few effective solutions have been proposed to solve the problem of weak cell-ECM adhesion [7,8].

In the context of stem cell transplantation, another crucial aspect is the capability of hMSCs to correctly interact with the surrounding cells in the injured tissue. In the context of myocardium

regeneration, cross-talk between cardiomyocyte and hMSCs represents the key point for paracrine factors exchange: protective factors released by hMSCs and factors released by cardiomyocyte may affect the fate of cell transplantation [9]. The possibility to form functional gap junctions allows the connection of the cytoplasm of adjacent cells. This will permit various small molecules, including ions and secondary messengers, to directly pass through a regulated gate between cells and this may be a modality for MSCs to exert cytoprotective effects [10]. It has been demonstrated that hMSCs express cardiac connexins, the building block proteins of gap junctions, and can form functional gap junctions with other cells expressing cardiac connexins including adult cardiac myocytes [11]. Therefore, the ability to increase connexin expression in MSCs would be critical to promoting cardiac regeneration.

Nanoparticle (NP)-based stem cell imaging has emerged as an essential tool for regenerative medicine, in order to track the MSCs and document both their persistence and promotion of regenerative effects [12–14]. Up to now, however, there are only a few reports on the effects exerted on MSC functions by cellular uptake of different NP types [14,15], even though this would be of great interest for the design of high-performance NP-based imaging systems for regenerative medicine processes. We have developed a method of fluorescent staining with silica NPs (SiO₂-NPs) suited to track hMSCs and validated this approach both *in vitro* [16,17] and *ex vivo* [18]. We already demonstrated that SiO₂-NP uptake by hMSCs was well tolerated in the long term, did induce neither cell death nor genotoxic stress and did not alter proliferative activity and differentiation potential of hMSCs [16–18].

In order to extend the use of SiO₂-NPs in cardiac regenerative medicine *in vivo*, it is of great interest to point out any possible stem cell responses to this nanomaterial internalization. Thus, the aim of the present work was to evaluate whether any phenotypic effects could be exerted by internalization of SiO₂-NPs that might ultimately impact on hMSC regenerative potential, with particular emphasis on the interaction of hMSCs with the cardiac environment.

Materials & methods

SiO₂-NPs: production, size & photoemission properties

Pure SiO₂-NPs and red fluorescent cyanine dye-doped SiO₂-NPs were prepared exploiting reverse micelles formed in a water-in-oil microemulsion, as detailed by Alberto *et al.* [19]. For the preparation of dye-doped nanoparticles, the aminopropyltriethoxysilane derivative of IRIS3 cyanine (by Pianeta Srl, formerly Cyanine Technologies Srl, Torino, Italy) was added to the reaction system. The absorption and photoluminescence spectra of IRIS 3 range from ca. 450 to 575 nm and from 500 to 700 nm, respectively. At the end of the process cyanine molecules resulted stably entrapped in the bulk of NPs based on solvatochromism test, thus the surface chemistry of NPs was not affected by the presence of IRIS3 molecules. Obtained SiO₂-NPs (both pure and dye-doped) exhibited a diameter of 50 ± 2 nm and possessed elevated morphologic homogeneity. When containing cyanine molecules, they displayed bright fluorescence emission and high photostability [19,20].

SiO₂-NP characterization

Dynamic light scattering

Dynamic light scattering measurements were performed in a 90Plus Particle Size Analyzer (Brookhaven Instruments, NY, USA) at a laser wavelength of 660 nm and a detection angle of 90 degrees at 293 K. Samples were prepared by suspending for up to 1 h SiO₂-NPs in distilled water (pH 5.5) and DMEM 1% fetal bovine serum (FBS). In all cases the SiO₂-NPs

concentration just before measurements was 20 µg/ml. Dynamic light scattering plots are reported as mass distribution. Measurements were performed in triplicates.

ζ-potential

The surface potential of both bare SiO₂-NPs (suspended for 1 h in distilled water and in DMEM) and SiO₂-NPs with hard corona proteins (suspended for 1 h in DMEM) was evaluated by electrophoretic light scattering using a Zetasizer Nano-ZS, Malvern Instruments (Worcestershire, UK). SiO₂-NPs with the protein hard corona were obtained by the FBS adsorption procedure reported in the [Supplementary Materials](#) document, with final washing in DMEM.

Cell culture & drug treatments

hMSCs isolated from the bone marrow of healthy donors were commercially obtained from Lonza (Lonza Group Ltd, Switzerland). For all the experiments cells from passage 4 to 8 were used. Briefly, hMSCs were cultured in DMEM supplemented with 1% sodium pyruvate, 1% nonessential amino acids, 1% kanamycin, 1% L-glutamine, 0.1% β-mercaptoethanol (complete DMEM) and 10% FBS (standard medium) (all from Sigma-Aldrich, MO, USA) and kept in an atmosphere of 5% CO₂, 95% air at 37°C in a humidified incubator. Cells were expanded at a seeding density of 3500 cell/cm² and subcultured twice a week. Exponentially growing hMSCs were seeded at 6,500 cell/cm² 24 h before the two following protocols: SiO₂-NPs: cells were exposed for 16 h to a suspension of water dissolved SiO₂-NPs 50 µg/ml in complete DMEM 1% FBS; SiO₂-NPs used for some *in vitro* experiments contained no fluorophore to avoid interference of fluorescence spectra. Control (CTR): cells were incubated for 16 h with complete DMEM 1% FBS supplemented with the same volume of sterile H₂O in which SiO₂-NPs were dispersed. Subsequently, samples of both conditions were washed twice with warm phosphate-buffered saline (PBS) and analyzed after the treatment (Day 0) or after 1, 4, 8 and 14 days of *in vitro* recovery in standard medium. H9C2 cell lines were purchased from the American Type Culture Collection and cultured in complete DMEM standard medium.

Transmission electron microscopy

High-resolution transmission electron microscopy (HR-TEM) images of SiO₂-NPs in their bare form and after suspension in DMEM 1% FBS followed by washing with distilled water were obtained with a Jeol 3010 microscope operated at 300 kV. Specimens were prepared by suspending the samples in distilled water and depositing a drop of the suspensions on Cu grids coated with a lacey carbon film, waiting until dryness.

For HR-TEM analysis, cells were prepared following procedures described by Raimondo *et al.* [21] See [Supplementary](#) for detailed explanation.

Confocal microscopy

hMSCs were seeded on 24-well ibiTreat microplate (Ibidi GmbH, Planegg, Germany) and, at the end of experiments, were processed for confocal immunofluorescence. Briefly, cells were fixed with 4% paraformaldehyde (PFA), permeabilized with 1% Triton-X-100 and blocked with 6% (w/v) BSA and 2.5% (v/v) normal goat serum.

For focal adhesion complex (FAC) analysis cells were stained for 2 h at room temperature (RT) with the primary antibody mouse anti-vinculin (Sigma-Aldrich) (1:600) and then incubated for 30 min at 37°C with the secondary antibody anti-mouse Alexa Fluor 488 (Thermo Fisher,

MA, USA) (1:500). Quantification was performed evaluating at least 40 cells per sample using ImageJ® [22].

For Connexin-43 analysis, both permeabilized and nonpermeabilized cells were incubated for 16 h at 4°C with the primary antibody mouse anti-Connexin-43 (1:400) and then the secondary antibody anti-mouse Alexa Fluor 488 (1:500) was added. Nuclear staining was performed with Hoechst-33342 (5 µg/ml). Quantification was assessed evaluating at least 120 cells per sample using ImageJ®.

For LC3 analysis, permeabilized cells were incubated for 16 h at 4°C with the primary antibody rabbit anti-LC3 (1:200) and stained for 60 min at RT with the secondary antibody anti-rabbit Alexa Fluor 488 1:500 in PBS.

To mark late endosomes and lysosomes, cells were incubated for 15 min at 37°C with LysoTracker Green (2 µmol/l) (Life Technologies, Italy) in complete DMEM and analyzed with inverted confocal laser scanning microscope LSM 800 (Carl Zeiss, Germany).

Flow cytometry

For Connexin-43 surface expression, approximately 400,000 hMSCs were trypsinized and maintained in 10% FBS culture medium for 2 h at RT, on a rocker platform to enable regeneration of the receptors. Cells were then washed with PBS, fixed 15 min at RT with 4% PFA, washed with PBS supplemented with 1% FBS and stained for 30 min at 4°C with rabbit polyclonal anti Connexin-43 primary antibody (Sigma-Aldrich) 1:60 in PBS, followed by incubation with secondary antibody goat anti-rabbit conjugated with phycoerythrin (PE, Southern Biotech, USA) 1:500 in PBS for 30 min at 4°C. Then cells were washed once with PBS, suspended in cold PBS, acquired with CyAN ADP flow cytometer (Beckman Coulter, CA, USA) and analyzed by Summit 4.3 software. At least 20,000 events per sample were collected.

Cell adhesion & cell detachment assays

For cell adhesion assay, cells were detached with 0.25% Trypsin-0.2% EDTA and resuspended in DMEM with 10% FBS in a 15 ml sterile tube to a final concentration of 1×10^5 /ml. Cells were maintained in rotation at RT for 2 h to allow surface receptor recovery. Then cells were spin down and resuspended at a final concentration of 2×10^4 /ml. A 100 µl of cells' suspension (2×10^4 cells) was then seeded in fibronectin (10 µg/ml) coated 96-well plates and cells were allowed to adhere for 5 or 15 min at 37°C in the CO₂ chamber. At least three wells were analyzed for each condition. The assay was stopped discarding the medium carefully without scratching the bottom of wells and rinsing cells once with PBS and washing three-times with 0.5% BSA in PBS.

For cell detachment assay, cells were seeded in fibronectin (10 µg/ml) coated 96-well plates at a density of 10,000/cm² and cells were allowed to adhere overnight at 37°C in the CO₂ chamber. Cells were treated with either control medium or SiO₂-NPs and then incubated with 0.025% Trypsin-0.02% EDTA for 4 or 8 min. The assay was stopped discarding the detached cells carefully without scratching the bottom of wells and rinsing cells once with DMEM 10% FBS and washing for three-times with PBS -0.5% BSA.

For both assays, cells were then fixed with 4% PFA for 10 min at RT and washed with PBS. To quantify the amount of adherent cells 0.1% crystal violet was added to each well and incubated for 10 min at RT; after three washes with ddH₂O (200 µl/well), the plates were let to dry up completely. Finally, 100 µl of 2% SDS were added to each well and incubated for

10 min with gentle shaking at RT. Fluorescence emission (excitation filter: 485 ± 20 nm; emission filter: 535 ± 25 nm) was read with an Infinite F200 microplate reader (Tecan Group Ltd., Switzerland).

Cell injection in *ex vivo* perfused infarcted rat hearts

Perfusion of isolated rat hearts & injection of SiO₂-NPs-hMSCs

Experiments were performed on adult male Wistar rats (450–550 g body-weight) ($n = 4$). Animals were heparinized (2500 U im., Roche, Italy) and anaesthetized with urethane (1 g/kg ip.) 10 min later. The hearts were rapidly excised, cannulated via the aorta and retrogradely perfused with oxygenated Krebs-Henseleit buffer containing (in mmol/l) 127 NaCl, 17.7 NaHCO₃, 5.1 KCl, 1.5 CaCl₂, 1.26 MgCl₂ and 11 D-glucose, supplemented with 5 µg/ml lidocaine. A constant flow was adjusted with a proper pump (Watson-Marlow 313, UK) to obtain a typical coronary perfusion pressure of 80–85 mmHg during initial stabilization. Thereafter, the same flow level (9 ± 1 ml/min/g) was maintained throughout the experiment. The temperature of perfusate and hearts were kept constant at 37°C throughout the experiments.

To produce an infarction, after 30-min stabilization, the left descending coronary artery was occluded for 30 min ([Supplementary Figure 2](#), 'Ischemia', recognized by pale-colored tissue after coronary occlusion and by a fall in left ventricular developed pressure) and then re-opened to full reperfusion of the left ventricle ([Supplementary Figure 2](#), 'Reperfusion'). Hearts were divided into two groups, one group received 5-chloromethylfluorescein diacetate (CMFDA)-treated and the other received red fluorescent cyanine dye-doped SiO₂-NPs-treated hMSCs. A total of 1×10^6 cells were injected in the apex after 30 min of reperfusion. All the experiments were then stopped after total 6 h, as this time was considered the end point for proper *ex vivo* experiments, according to Penna *et al.* [23].

Heart processing & immunofluorescence analysis of tissue slices

At the end of perfusion, atria and vasa were discarded and ventricles were fixed 3 h RT with 4% PFA with gentle stirring. After four washes with PBS, tissues were submerged in PBS supplemented with 30% sucrose, allowed to precipitate overnight at 4°C and then incubated 30 min RT in a solution 1:1 of 30% sucrose and TissueTek® Optimal Cutting Temperature (OCT™, Sakura FineTek, CA, USA). Finally, tissues were embedded in OCT and stored at -80°C. Transverse slices 10 µm thick were obtained starting from the apex with a CM 1900 cryostat (Leica Microsystems GmbH, Wetzlar, Germany) and processed. Sections for ultrastructural analysis were processed for immunofluorescence. Briefly, they were rinsed in PBS, permeabilized 20 min at RT with 0.5% Triton-X-100 and blocked 1 h at RT with 6% wt/vol BSA and 2.5% NGS in PBS. The primary antibody mouse anti-sarcomeric α -actinin 1:600 in PBS was incubated overnight 4°C and the secondary antibody anti-mouse Alexa Fluor 488 1:1000 in PBS was incubated 1 h at RT. After several washes with PBS, samples were finally mounted with Mowiol and conserved at 4°C.

Dye transfer assay

To monitor diffusion of fluorescent molecules through gap junctions (dye coupling) we used a method similar to the 'parachute' assay [24]. The method we used considered donors (hMSC cells – untreated or treated with SiO₂-NPs) loaded with a gap junction permeant dye, in other words, Calcein-AM and unlabeled acceptors (cardiomyoblasts H9C2 cells).

For confocal microscopy analysis, hMSCs were labeled with Calcein-AM (Sigma-Aldrich) membrane permeant dye (1 μ M) for 30 min at 37°C. After labeling, the cells were cocultivated (hMSC:H9C2 ratio 1:2) for 6 h at 37°C and then analyzed for their fluorescence. The two cell types were distinguished morphologically. Carbenoxolone-disodium (Sigma-Aldrich) was used to inhibit Calcein-AM transfer through gap junctions. Cells were preincubated with the inhibitor (100 μ M) overnight at 37°C. After labeling, the cells were cocultivated (hMSC:H9C2 ratio 1:2) for 6 h at 37°C and then analyzed for their fluorescence. The two cell types were distinguished morphologically.

For flow cytometry analysis hMSCs were labeled with Calcein-AM in the same way as for confocal microscopy analysis. After labeling, hMSCs were cocultivated with H9C2 for 24 h at 37°C and incubated with 21% O₂ and 5% CO₂, in other words, normoxia or in a hypoxic chamber with 1% O₂ and 5% CO₂, in other words, hypoxia. Before starting a coculture, H9C2 intended for hypoxia were maintained for 150 min in 0% O₂, 5% CO₂, glucose-free medium to mimic ischemia as reported by Cselenyák *et al.* [25]. CD90 staining was performed using specific MSC marker anti CD90-PE antibody (BD Pharmingen, CA, USA) 1:10 in PBS for 30 min at 4°C to distinguish hMSCs and H9C2. Cells were acquired with CyAN ADP flow cytometer (Beckman Coulter) and analyzed by Summit 4.3 software. At least 100,000 events per sample were collected.

As a negative control, Vybrant®-5-(and-6)-carboxyfluorescein diacetate, succinimidyl ester (CFDA SE; Molecular Probes) labeled hMSCs were used. CFDA-SE is a fluorescein-based tracer. Once inside the cell, nonspecific esterases cut its lipophilic groups and the dye remains in the cell. The concentration of 2.5 μ M in PBS was used for 15 min at 37°C. At least 20,000 events per sample were collected.

Cell metabolism

To evaluate cell metabolism, CellTiter-Blue® (Promega, Leiden, the Netherlands) assay was used. See [Supplementary Materials](#) for more information.

Western blot analyzes

hMSCs were lysed with RIPA buffer and 12 μ g of protein samples was loaded. SDS-PAGE (12% Bis-Tris gel, Invitrogen, CA, USA), Polyvinylidene difluoride transfer membrane and quantification BCA assay (Thermo Scientific, MA, USA) were used. Membranes were blocked in 5% milk for 1 h at room temperature and probed with the primary antibodies (anti-LC3B, anti-p62/SQSTM1 purchased from Sigma-Aldrich) (1:1000) overnight at 4°C, followed by secondary antibodies (1:5000) for 1 h at RT. Bands were visualized using an enhanced chemiluminescence kit (SuperSignal™ West Pico PLUS, Thermo Scientific).

Statistical analysis

Data are expressed as a mean \pm standard error of the mean (SEM) of at least three different experiments or as the mean \pm standard deviation (SD) of representative experiments. Statistical comparisons were performed with either paired or unpaired Student's *t*-test. Differences with $p < 0.05$ were regarded as statistically significant.

Results

The state of SiO₂-NPs when administered to cells and their efficient internalization by hMSCs

For the administration of NPs to cells, an aliquot of the water suspension of NPs, where they were almost monodispersed ([Figure 1A](#)), was added to complete DMEM – 1% FBS, and this resulted in the formation of agglomerates of ca. 1 μm in size, stable over at least 1 h ([Figure 1B](#)), in other words, a time corresponding to the incubation stage with cells inspected by HR-TEM. It is worth mentioning that a protein corona was formed on NPs agglomerates when suspended in DMEM-1% FBS [[17](#)], that was possible to image for the very few small clusters of NPs found on the specimen grid when depositing a drop of NPs-DMEM 1% FBS suspension ([Figure 1A & B](#)). According to the formation of the protein corona, the ζ -potential of NPs changed in a large extent, passing from -25 to -9 mV for SiO_2 -NPs in the bare form and in DMEM 1% FBS, respectively ([Figure 1A & B](#)). Some agglomeration of NPs was retained during incubation with cells, as indicated by HR-TEM analysis, which allowed observing invaginations of the hMSC plasma membrane that surrounded ensembles of NPs, explainable as the beginning of an endocytic process ([Figure 1C](#)).

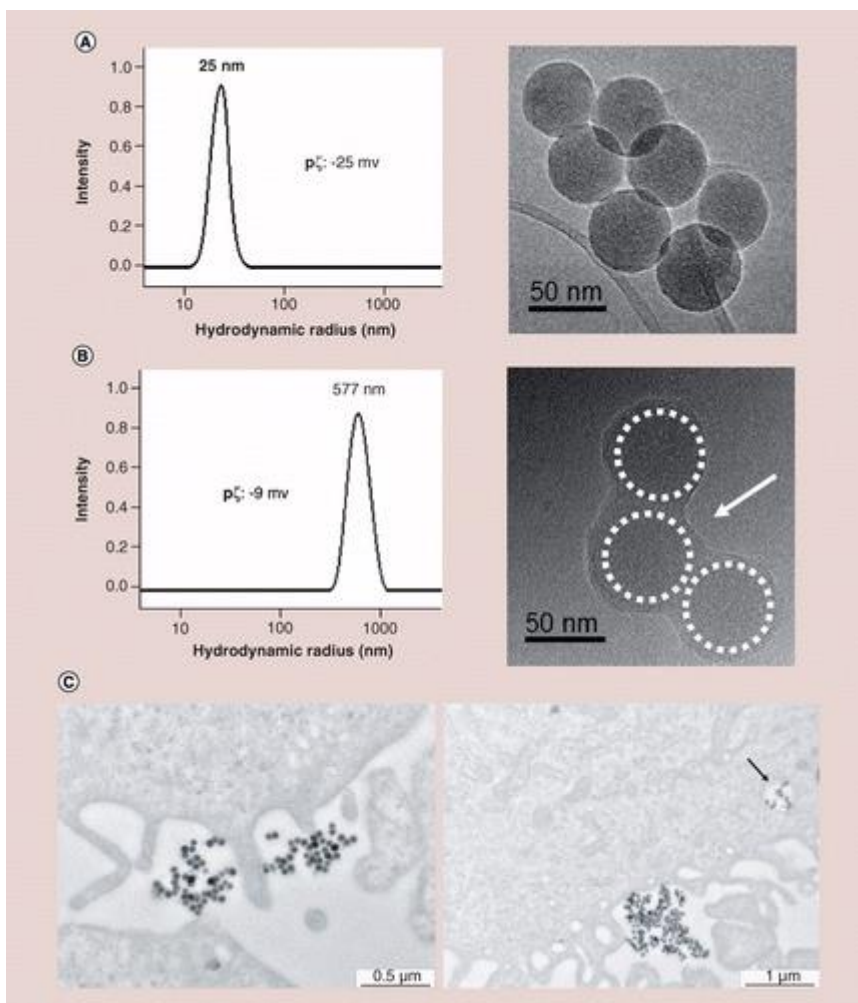


Figure 1. Silica nanoparticle characterization and interaction with human mesenchymal stem cells.

(A) Hydrodynamic radius (by dynamic light scattering) and ζ -potential (left panel) and TEM image (right panel) of silica-nanoparticle (SiO_2 -NP) dispersed in water. **(B)** Hydrodynamic radius (by dynamic light scattering) and ζ -potential (left panel) and TEM image (right panel) of SiO_2 -NP suspended for 1 h in complete medium supplemented with 1% fetal bovine serum. Arrow shows the corona formed by serum protein adsorbed on the NPs. Dotted circle represents the shape of NPs. **(C)** TEM images of human mesenchymal stem cells after 1 h of incubation with SiO_2 -NP in complete medium supplemented with 1% fetal bovine serum. Magnification 50,000 \times .

TEM: Transmission electron microscopy; NP: Nanoparticle.

The cytofluorimetric analysis confirmed that virtually all hMSCs possessed internalized NPs by the end of the treatment (16 h) ([Supplementary Figure 1](#)).

SiO₂-NP internalization modifies focal adhesion structure in hMSCs

Adhesion is an essential process of anchorage-dependent cells involved in regulation of cell functions and indeed, in regenerative medicine, one crucial aspect concerns the capability of stem cells to interact with the surrounding environment. One of the earliest and best-characterized adhesion structures are focal adhesion complexes, in other words, large macromolecular complexes that link the actin cytoskeleton to the ECM to provide traction, and properly regulated FAC dynamics are critical for cell attachment and migration [[26–28](#)].

To identify possible effects of SiO₂-NP internalization on cellular FACs, we evaluated the universal focal adhesion marker vinculin [[29,30](#)] by immunofluorescence and confocal microscopy analysis. Qualitative evaluation of images strongly suggested that, following SiO₂-NP internalization, FACs acquired a more elongated shape and an increased size ([Figure 2A](#)). FAC analyzes showed that after SiO₂-NP internalization the total number of FA remained unchanged, yet their mean size was significantly increased ([Figure 2B & C](#)). Considering the growth model of force-induced focal adhesion driven by actomyosin-mediated tension [[26,31](#)], large focal adhesions tend to be elongated, while small focal adhesions tend to be round. FAC length and width measurements showed that in control hMSCs a low length/width ratio occurred more frequently, indicative of a prevalently round structure, while in SiO₂-NP-hMSCs a high length/width ratio was prevalent, indicating an elongated structure ([Figure 2D & E](#)).

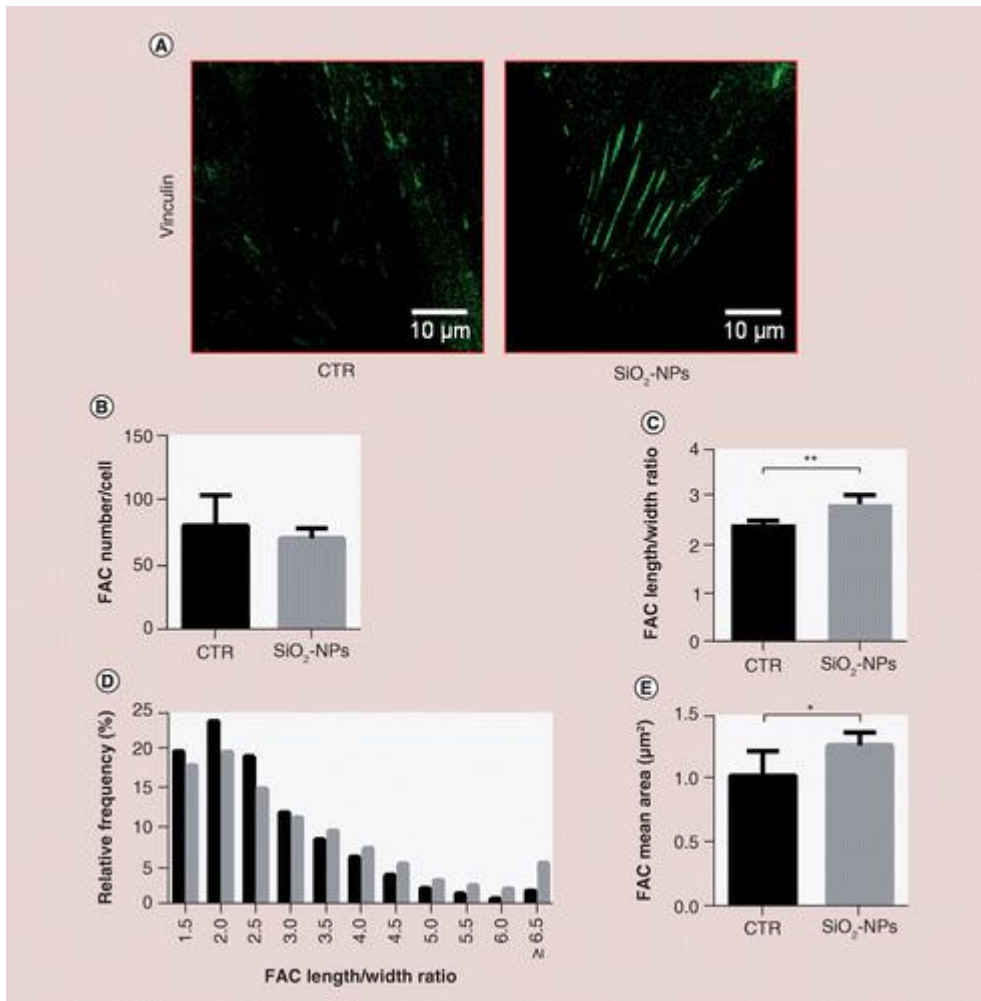


Figure 2. Silica

nanoparticle effects on human mesenchymal stem cell focal adhesion complexes.

Cells were treated for 16 h with either control medium (CTR) or silica nanoparticles (SiO₂-NPs) and then stained with an anti-vinculin antibody conjugated with a green fluorophore before confocal microscopy acquisition. **(A)** Representative confocal images of stained cells shown the qualitative differences of the FACs. Magnification 63x. **(B & C)** Quantification of the number of FACs/cell **(B)** and FAC mean area **(C)** in CTR and SiO₂-NP treated human mesenchymal stem cells (hMSCs). Data are shown as mean ± standard deviation (SD). **(D)** Quantification of the FAC elongation distribution (expressed as %) in CTR and SiO₂-NP treated hMSCs. FA elongation index was obtained calculating the length/width ratio of every FA. **(E)** Quantification of FAC elongation in CTR and SiO₂-NPs treated hMSCs. Data are shown as mean ± SD.

*p < 0.05; **p < 0.01.

FAC: Focal adhesion complex.

These data, taken together, demonstrate that SiO₂-NP treatment increases hMSC FAC area and elongation, suggesting the possibility that SiO₂-NP internalization induces FAC stabilization contributing to confer an enhanced capability of SiO₂-NP treated hMSCs to adhere to a substrate [32].

SiO₂-NP internalization augments hMSC adhesion behavior *in vitro* & *ex vivo*

The adhesive strength of SiO₂-NP-hMSCs was assessed using two different *in vitro* assays. One was the trypsin/EDTA-induced detachment assay, which measures the strength of cell attachment to fibronectin (FN)-coated plates. FN was used as it is an important component of cardiac ECM and it is strongly up-regulated *in vivo* after myocardial infarction [33]. After treatment with trypsin/EDTA, SiO₂-NP-hMSCs remained attached to the plate at a significantly higher level than control hMSCs after 4 and 8 min (Figure 3A).

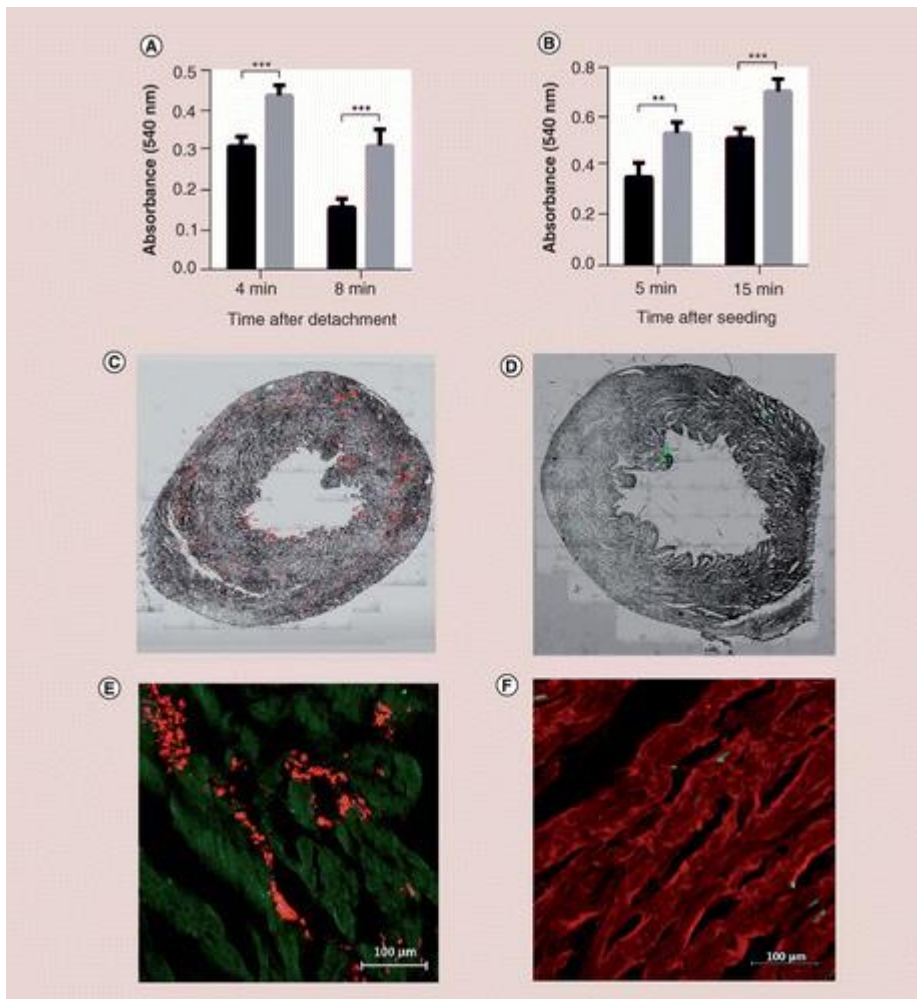


Figure 3. Silica

nanoparticles effect on human mesenchymal stem cell adhesion.

(A) Trypsin-based cell detachment assay. Control (CTR) and silica nanoparticle-human mesenchymal stem cells (SiO₂-NP-hMSCs) were treated with 0.025% trypsin-0.02% EDTA for the indicated time points. The ability of hMSCs to remain attached to the substrate was evaluated. (B) Adhesion assay. CTR and SiO₂-NP-hMSCs were allowed to adhere on fibronectin coated plates for the indicated time points. For both assays (A & B), adherent cells were stained with 0.1% crystal-violet and their relative absorbance was determined spectrophotometrically at 540 nm. Data are shown as mean \pm standard deviation. (C) Imaging of CMFDA-labeled hMSCs inside infarcted hearts. (D) Imaging of red fluorescent cyanine dye-doped SiO₂-NPs-labeled hMSCs inside infarcted hearts. For both images (C & D), reconstruction of a 10 μ m transverse slice to show the distribution of labeled hMSCs in infarcted hearts. Green/Red points put to highlight the clusters of hMSCs. Magnification 10 \times , scale bar 200 μ m. (E & F) Superposition of sarcomeric α -actinin staining (red in E and green in F) and labeled hMSCs (green with CMFDA in E and red with SiO₂-NPs in F). Magnification 20 \times . **p < 0.01; ***p < 0.005.

CMFDA: 5-chloromethylfluorescein diacetate.

Another technique used was the adhesion assay, which measures the adhesion of hMSCs that have been allowed to attach on FN-coated plates. Results showed an increased adhesion of SiO₂-NP-hMSCs in comparison to control cells. After 5 and 15 min of adhesion on FN-coated plates, the presence of SiO₂-NPs conferred a significantly higher ability to adhere to the substrate ([Figure 3B](#)).

We next assessed the possibility to highlight an increased adhesion behavior following SiO₂-NP internalization in the challenging condition of a beating heart, subjected to ischemia/reperfusion (I/R). To this aim, an *ex vivo* model of perfused rat heart was used and cells were injected into the apex 30 min after the ligation of left descending coronary artery was removed ([Supplementary Figure 2](#)). Cells employed for these investigations were exposed for 16 h to SiO₂-NPs and then immediately injected; as a control, we used hMSCs labeled with the CMFDA green fluorescent probe, a well-known labeling procedure commonly used to highlight cell integration *in vivo* and that does not interfere with hMSC adhesion properties (our data not shown). Reconstruction of a representative transverse section highlighted that SiO₂-NP labeled cells were clearly visible and widespread distributed inside the walls of both right and left ventricles ([Figure 3C](#)). Differently, few CMFDA labeled cells can be detected ([Figure 3D](#)). These findings are confirmed with a higher amplification and α -actinin immunostaining ([Figure 3E & F](#)).

These results demonstrate that SiO₂-NP internalization confers increased adhesion ability to hMSCs both *in vitro* and *ex vivo*.

SiO₂-NP internalization enhances Connexin-43 surface expression & favors the communication between hMSCs & cardiac myoblasts

In the context of cardiac regenerative medicine, it is necessary that hMSCs form gap junctions with the surrounding cells, to become an effective member of the myocardium [[11](#)]. As Cx43 is the most important gap junction complex in the myocardium, we investigated whether SiO₂-NPs internalization by hMSCs could affect Cx43 expression. Surface expression of Cx43 was first evaluated by flow cytometry analysis. Control cells were positive for Cx43, presenting a homogenous, low level of fluorescence. After SiO₂-NPs treatment, fluorescence relative to Cx43 increased significantly, indicating an increase in Cx43 surface membrane expression ($p < 0.001$) ([Figure 4A](#)). These data were confirmed by confocal microscopy analysis. Qualitative observation of confocal images suggested that the dot-like expression of Cx43 was increased in SiO₂-NPs treated hMSCs ([Supplementary Figure 3](#)). Quantification of the total area of the Cx43 confirmed that following SiO₂-NPs internalization hMSCs significantly increased Cx43 expression ([Figure 4B](#)).

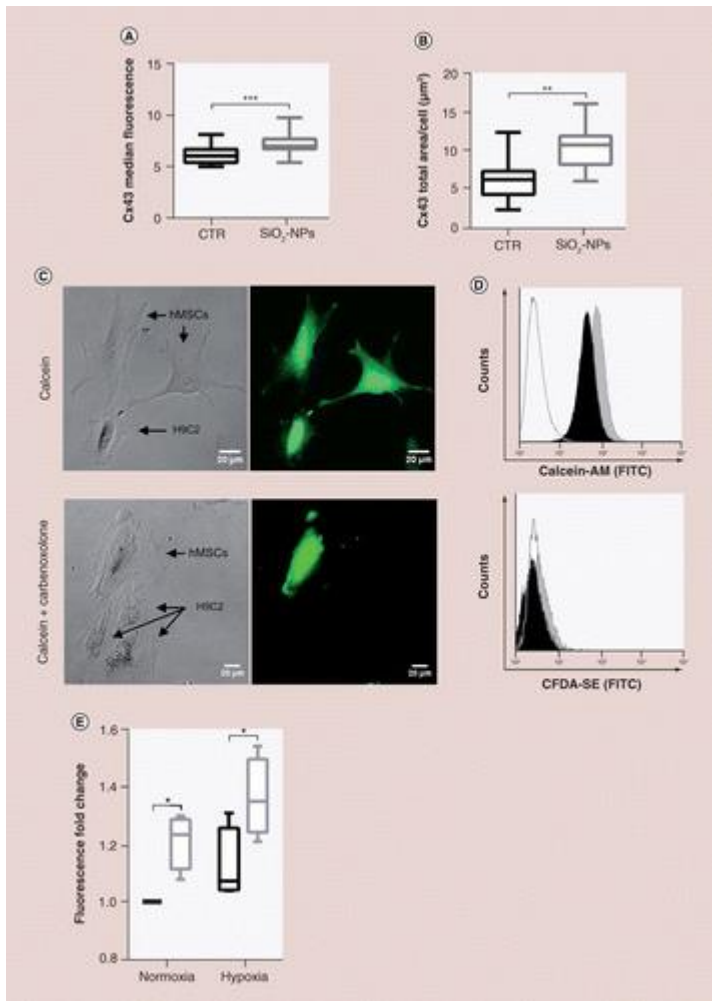


Figure 4. Silica nanoparticles effect on human mesenchymal stem cell intercellular communication.

(A) Box plot showing the distribution of the median fluorescence relative to Cx43 in control (CTR) and silica nanoparticles (SiO₂-NPs) treated human mesenchymal stem cells (hMSCs) (eight independent experiments) obtained by flow cytometry. Paired *t*-test were performed ****p* < 0.001. **(B)** Box plot showing the quantification of Cx43 area in CTR and SiO₂-NPs treated hMSCs obtained by confocal microscopy. At least 120 cells were analyzed. Unpaired *t*-test was performed. *****p* < 0.0001. One representative experiment out of three. **(C)** Representative dye transfer image in confocal microscopy. hMSCs were labeled with Calcein-AM (green cells) without (top panel) or in presence (lower panel) of the gap junction inhibitor Carbenoxolone and H9C2 were discriminated morphologically by phase contrast images. Cells were cocultured for 24 h and the presence of H9C2 cells that received Calcein-AM through intercellular communication with hMSCs were detected as green color in H9C2 cells. **(D)** Representative dye transfer experiment in flow cytometry. Upper panel: H9C2 cells were cultured for 24 h alone (white histogram) or in the presence of either CTR (light gray histogram) or SiO₂-NP treated hMSCs (black histogram) labeled for 24 h with Calcein-AM (gap junction permeant dye). The green fluorescence detected in H9C2 cells was due to the Calcein-AM they received through intercellular communication with hMSCs. Lower panel: H9C2 cells were cultured for 24 h alone (white histogram) or in the presence of either CTR (light gray histogram) or SiO₂-NPs treated hMSCs (black histogram) labeled for 24 h with CFDA-SE (gap junction impermeant dye). The absence of H9C2 cells that received CFDA-SE was verified. H9C2 and hMSCs were distinguished through CD90 hMSC specific surface marker. **(E)** Box plot showing the distribution of the mean fluorescence relative to Calcein-AM that H9C2 received from CTR (black borders) and SiO₂-NPs treated hMSCs (gray borders)

obtained by flow cytometry. At least 80,000 cells were analyzed for each experiment. Data were normalized on control cells in normoxia. T-test was performed. * $p < 0.05$.

These data show that SiO₂-NP treatment induces the surface expression of gap junction Cx43 protein, suggesting the interesting possibility that SiO₂-NP-hMSCs might have an increased capability to communicate with the surrounding cells. To demonstrate this hypothesis, gap junction-mediated dye transfer assays between SiO₂-NP-hMSCs and the cardiomyoblast H9C2 cell line were performed. hMSCs were labeled with the gap junction permeant dye Calcein-AM or the gap junction impermeant dye CFDA-SE and cocultured with unlabeled H9C2 cells. Transfer of Calcein-AM between hMSCs and H9C2 cells was observed in confocal microscopy (Figure 4C, upper panel) and it was blocked when a gap junction inhibitor was added, as expected (Figure 4C, lower panel). Analysis of cocultures by flow cytometry demonstrated that H9C2 cells gained Calcein-AM from hMSCs while no CFDA-SE was transferred, demonstrating that gap junction intercellular communication occurs between these cells. (Figure 4D). After SiO₂-NP treatment, hMSCs increased the amount of Calcein-AM transferred to H9C2 cells, in both, normoxia and hypoxia, demonstrating the correct functionality of the enhanced surface Cx43 expression. (Figure 4E). The increment of dye exchange between CTR and SiO₂-NP-hMSCs was statistically significant in both, normoxic and hypoxic conditions. It was also evident that more dye was exchanged in hypoxia than in normoxia, although not in a statistically significant way. (Figure 4E).

Transient lysosomal degradation impairment induced by SiO₂-NP uptake could explain the observed functional effects on hMSCs

In order to find a possible explanation for the observed functional effects, we focused our attention on the mechanism of intracellular accumulation of SiO₂-NPs. Internalized SiO₂-NPs displayed a punctuate distribution with some enrichment in the perinuclear area, suggestive of their accumulation within intracellular organelles. We previously demonstrated that while no colocalization of internalized SiO₂-NPs with the Golgi complex, mitochondria or the endoplasmic reticulum was detected, the accumulation of SiO₂-NPs in late endosomes/lysosomes was clearly observed in living cells incubated with LysoTracker Green, an organic heterotricyclic 4-bora-3a,4a-diaza-s-indacene compound that accumulates within the acidic lumen of late endosomes/lysosomes [16,18]. In accordance with our previously published results, LysoTracker staining of living cells highlighted that untreated hMSCs prevalently displayed few and small lysosomes around nuclei, while hMSCs exposed to SiO₂-NPs presented a greater number of larger organelles, most of which were perinuclear (Figure 5A – day 0). A time course analysis of LysoTracker experiments on SiO₂-NP-treated hMSCs highlighted that at 1 and, to a lesser extent, 4 days after the treatment larger organelles were still observed (Figure 5A – days 1 and 4). However, after 8 days of culture hMSCs displayed a normalized lysosome compartment, demonstrating that lysosomal compartment perturbation is a transient event induced by NP accumulation that can be spontaneously recovered by hMSCs (Figure 5A – day 8). Accordingly, no metabolic impairment was observed in SiO₂-NP treated hMSCs up to 8 days, as evaluated through a fluorometric assay (Figure 5B). We next investigated whether the phenotypic changes observed in SiO₂-NP treated hMSCs are maintained over time or are transient, analogously to lysosomal accumulation. Interestingly, we found that both increased cell adhesion and Cx43 expression returned to baseline levels in parallel with lysosomal compartment normalization (Figure 5C & D). Further, we found that inhibition of lysosomal degradation activity, obtained using 100 nM bafilomycin A1 for 2 h, reproduced in CTR hMSCs both, increased adhesion and Cx43 expression observed in SiO₂-NP-treated cells (Figure 6A & B). This suggested that SiO₂-NP-filled lysosomes might be transiently impaired in their degradation activity.

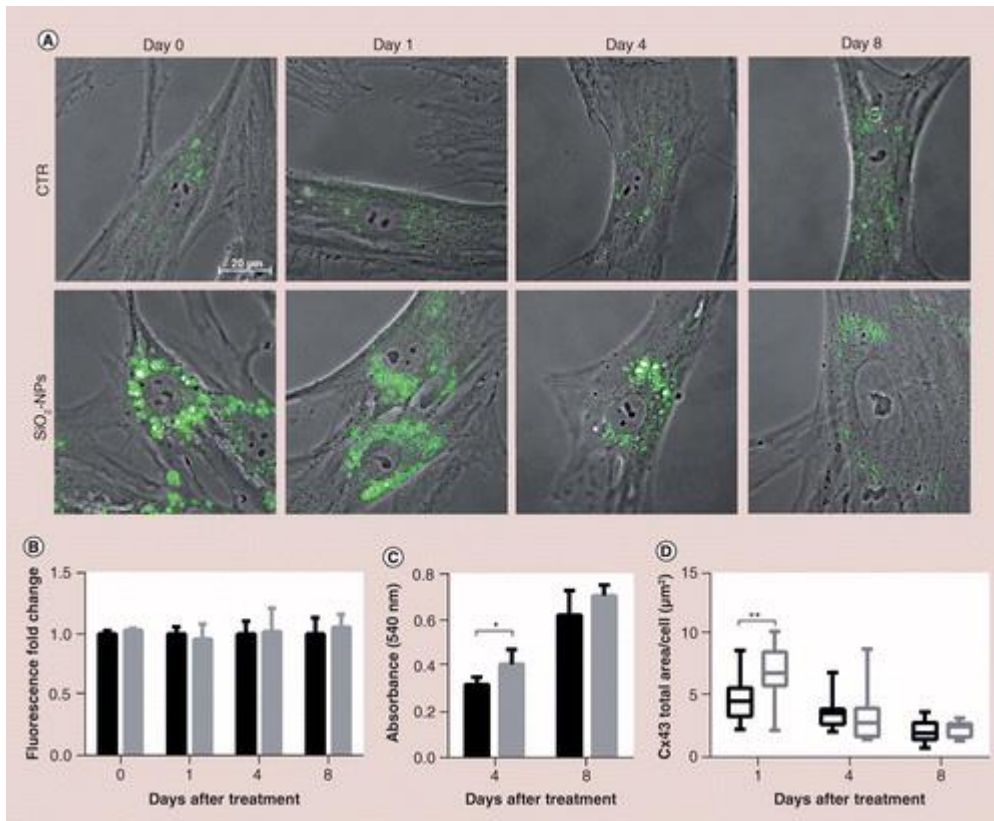


Figure 5. Time course, metabolism and phenotype analysis of silica nanoparticles effects on human mesenchymal stem cells.

(A) Control (CTR) and silica nanoparticles (SiO₂-NP) treated human mesenchymal stem cells (hMSCs) were labeled with LysoTracker (green) at day 0 – day 1 – day 4 – day 8 time points and representative images were acquired by confocal microscopy. Magnification 63x. **(B)** CellTiter-Blue fluorescence were analyzed at day 0 – day 1 – day 4 – day 8 and the fluorescence values were normalized on control cells and represented as mean ± standard deviation (SD). **(C)** CTR and SiO₂-NP treated hMSCs were allowed to recover in complete medium for 4, 8 and 14 days and then adhesion assay was performed after 5 min of adhesion. Adherent cells were stained with 0.1% crystal-violet and their relative absorbance was determined spectrophotometrically at 540 nm. Data are shown as mean ± SD; *p < 0.05. **(D)** CTR and SiO₂-NP treated hMSCs were allowed to recover in complete medium for 1, 4 and 8 days and then Cx43 expression was evaluated by confocal microscopy. Box plot showing the quantification of Cx43 area in CTR and SiO₂-NPs treated hMSCs. At least 120 cells were analyzed. Unpaired *t*-test was performed. **p < 0.01.

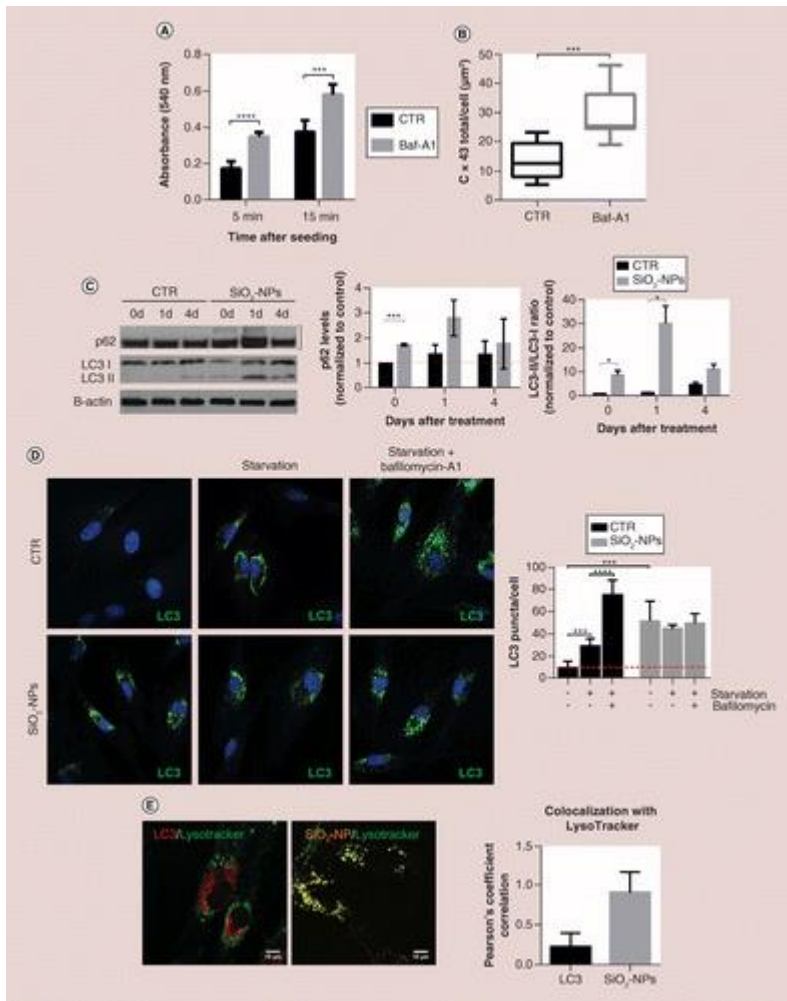


Figure 6. Inhibition of lysosomal degradation activity.

(A) Adhesion assay. Human mesenchymal stem cells (hMSCs) were treated for 2 h with either control medium or Bafilomycin A1 100 nM. Cells were allowed to adhere on fibronectin coated plates for the indicated time points. Adherent cells were stained with 0.1% crystal-violet and their relative absorbance was determined spectrophotometrically at 540 nm. Data are shown as mean \pm standard deviation (SD); *** $p < 0.001$, **** $p < 0.0001$. **(B)** hMSCs were treated for 2 h with either control medium or Bafilomycin A1 100 nM and then Cx43 expression was evaluated by confocal microscopy. Unpaired t -test was performed. Data are shown as mean \pm SD; *** $p < 0.001$. **(C)** Control (CTR) and silica nanoparticle (SiO_2 -NP) treated hMSCs were analyzed by western blotting for p62 and LC3. The level of p62 and the LC3-II/LC3-I ratio were quantified and normalized to untreated control cells. Data are shown as mean \pm SD; * $p < 0.05$, *** $p < 0.001$. **(D)** Left, representative confocal images of CTR or SiO_2 -NP treated hMSCs incubated in conditions as above. After fixation, cells were stained for LC3. Right, quantification of LC3 puncta per cell. Data are shown as mean \pm SD; *** $p < 0.001$, **** $p < 0.0001$. **(E)** Left, representative confocal images of SiO_2 -NP treated hMSCs stained with LC3 and/or LysoTracker green. Right, Pearson's correlation analyzes showing near complete lack of colocalization between LC3 and LysoTracker (0.223 ± 0.172), and strong colocalization between SiO_2 -NPs and LysoTracker (0.909 ± 0.255). Data are the mean \pm SEM, $n = 3$ independent experiments.

Considering that bafilomycin A1, a widely used inhibitor of autophagosome–lysosome fusion *in vitro* to determine the activity of autophagic flux, reproduced the phenotype of SiO_2 -NP treated cells, we addressed the hypothesis that altered lysosomal degradation might result from impaired cargo delivery to SiO_2 -NP-filled lysosomes from upstream donor compartments, such

as autophagosomes. If this were the case, one would expect that autophagosomal components, like LC3-II and p62, being substrates for lysosomal proteolysis, accumulate in SiO₂-NP treated cells and they do so in compartments (autophagosomes) lacking SiO₂-NPs (accumulated in lysosomes), due to impaired autophagosome–lysosome fusion. In accordance with this hypothesis, WB analysis indicated a transient cellular accumulation of both LC3-II and p62 proteins in SiO₂-NP treated cells that peaked at 1 day and began to attenuate at 4 days (Figure 6C). Further, confocal imaging revealed that starvation of control cells resulted in increased levels of LC3 due to the accumulation of LC3-positive autophagosomes, and this phenotype was exacerbated by concomitant application of bafilomycin to prevent lysosomal turnover of autophagosomes (Figure 6D, upper panel); cells treated with SiO₂-NPs, instead, displayed high levels of LC3 and accumulated LC3-positive autophagosomes largely irrespective of whether cells had been starved and/or treated with bafilomycin (Figure 6D, lower panel). Finally, a near complete lack of colocalization between LC3 and SiO₂-NP-filled lysosomes (e.g., positive for LysoTracker) was revealed (Figure 6E). Altogether, these data strongly indicated an NP-induced transient impairment of autophagic flux as a likely mechanistic explanation.

Discussion

While NP-based stem cell imaging is an essential tool for regenerative medicine, cellular uptake of NPs might trigger numerous intracellular signaling events through which cells control their function and state. The central aim of this work was to evaluate any possible functional effects of SiO₂-NPs on hMSC therapeutic potential, with particular emphasis on cardiac regeneration and interaction with the cardiac environment. The data collected here suggest that SiO₂-NP internalization does have an influence on hMSC cell behavior, providing strong evidence that nanostructures not only passively interact with cells but also actively engage and mediate the molecular processes that are essential for regulating cell functions.

First, we found that SiO₂-NP treatment is associated with significant changes in focal adhesion structures. In particular, the FACs of SiO₂-NP treated hMSCs are increased in mean area and elongation index as evidenced by vinculin staining. This biological effect seems not to be peculiar of SiO₂-NPs, however, as interestingly, Tay and collaborators obtained similar results by examining vinculin surface expression in TR146 oral mucosa cells treated with SiO₂, TiO₂ and hydroxyapatite NPs [34]. The higher occurrence of long mature FACs near cell–cell boundaries was detected through vinculin immunofluorescence analysis, suggesting the promotion of an increased adhesive phenotype [34]. In this work, we describe this phenotype for the first time in hMSCs.

These findings made it plausible to hypothesize an increased adhesion capability of hMSCs due to SiO₂-NP treatment, which was confirmed using *in vitro* and *ex vivo* models. While an increased *in vitro* cell adhesion was previously described in MSCs treated with other NP types (metal oxide NPs), we here document for the first time an augmented engraftment of SiO₂-NP treated hMSCs in the myocardial tissue upon injection in infarcted isolated rat hearts. Considering that adhesion of the transplanted hMSCs is necessary not only for cell engraftment and generation of new tissue but also for a cross-talk between transplanted cells and receiving an organ, our data suggest the possibility that the internalization of a SiO₂-NP stimulates a cell function (engraftment) that is required for an effective regenerative therapy. Using non-NP-based, *ex vivo* genetic modification approaches, several studies demonstrated that an increased adhesion phenotype of MSCs represents a crucial aspect to enhance cell survival and therapeutic potency upon transplantation to sites of injury [35,36]. MSCs derived from transgenic rats overexpressing tissue transglutaminase, a coreceptor for FN implicated in cell adhesion associated with integrins [37], displayed enhanced cell attachment and spreading features. In these cells, the formation of FACs and the increased phosphorylation of

the focal adhesion-related kinases PI3K and Src were observed. Moreover, the implantation of tissue transglutaminase-transfected MSCs in rat infarcted myocardium improved cardiac functions [38]. Other studies demonstrated that an increased adhesion phenotype enhances cell survival and prevents anoikis [38–40]. In particular, MSC transfection with an integrin-linked kinase, a Ser/Thr kinase that interacts with the cytoplasmic domain of β 1 integrin, plays a crucial role in ECM assembly, cell spreading, integrin-mediated cell adhesion, signaling and survival [41].

Further, we showed that SiO₂-NP treatment increases Cx43 surface expression in hMSCs and dye transfer experiments demonstrated the functionality of these gap junctions and the possibility to increase intercellular communication with cardiomyoblasts. Increased Cx43 expression suggests that hMSCs might readily integrate into electrical syncytia of cardiac tissue, promoting repair or serving as the substrate for a therapeutic delivery system. After acute myocardial infarction, severe loss of healthy myocardium impairs intercellular communications in the peri-infarct area and induces cardiac conduction disturbances [42,43]. Administration of naive MSCs at this phase may pose arrhythmic risks [44,45]. Previous studies addressed that abnormal cardiac conduction and arrhythmogenic remodeling after MI could be worsened by low Cx43 expression of naive MSCs [45,46]. Besides the use of NPs, growth factor (FGF-2, IGF-1, BMP-2) treatment of MSCs has been reported to augment intercellular connections with host cardiomyocytes via Cx40- and Cx43-mediated junctions [47]. Another study demonstrated that Ang II pre-treatment enhanced MSC gap junction protein expression *in vitro* [10]. Interestingly, injection of genetically modified MSCs that overexpressed Cx43 conferred a better reparative effect of the postinfarcted myocardium [46–48]. Previous studies also showed that overexpression of Cx43 in MSCs can greatly improve the therapeutic efficacy of the cell therapy in MI regardless of the cellular arrangement of Cx43 [46–49].

A possible effect of NPs on cellular gap junction crosstalk has been recently reported by Han and colleagues [50], although not in MSCs. Those authors demonstrated that iron oxide NPs significantly augmented the expression of Cx43 in H9C2, which would be critical for gap junctional communication with MSCs in coculture for the generation of therapeutic potential-improved MSCs [50]. MSCs cocultured with iron oxide NP-harboring H9C2 cells showed active cellular crosstalk and displayed significantly higher levels of electrophysiological cardiac biomarkers and a cardiac repair-favorable paracrine profile, both of which are responsible for MI repair [50]. Our results, together with the above study, highlight an effect of NPs in developing gap junctional crosstalk among cells, extending the potential application of NPs in cell biology and stem cell-based therapies. Furthermore, this is a first study that shows an increase of Cx43 in hMSCs induced by SiO₂-NPs internalization.

What might be the reason for the two observed biological effects in hMSCs? After internalization, SiO₂-NPs readily accumulate in lysosomes, and this might transiently impact on the lysosomal function. Based on the results we obtained in this work, including the spontaneous normalization of the biological phenotypes by 8 days from SiO₂-NP treatment, a time when normalization of lysosomal compartment could also be described, and the capability of reproducing the same biological effects by the use of a lysosomal activity inhibitor that impedes autophagosome–lysosome fusion, our conclusion is that accumulation of SiO₂-NPs in lysosomes cause a transient block of autophagic flux in hMSCs. In accordance, we found an accumulation of two autophagosome marker proteins, LC3-II and p62, in SiO₂-NP treated cells and a near complete lack of colocalization between LC3 and SiO₂-NP-containing lysosomes. Accumulation of SiO₂-NPs in lysosomes is thus predicted to result in transient inhibition of autophagy-mediated protein turnover in hMSCs, leading to perturbed degradation of cytosolic proteins, in perfect accordance with two recently published papers concerning lysosomal dysfunction caused by silica NPs [51,52]. Indeed, cell adhesion is one of the cellular

processes described to be regulated by autophagy [53,54] and, early on in connexin research, it became clear that lysosomal pathways also contributed to the connexin degradation [55,56]; more recently, autophagy has emerged as a crucial connexin-degradation pathway for different connexin isoforms, including Cx-43 [57]. Notably, this transient dysfunction in lysosomal activity occurs in complete absence of any metabolic impairment or cytotoxicity in our cells, at variance with what has been reported for human cervix carcinoma cells [51] and human hepatocytes [52]. Lysosomal activity inhibition is subsequently overcome in hMSCs presumably due to SiO₂-NP distribution to daughter cells and their consequent intracellular dilution [18], and the phenotypic effects released in parallel.

Conclusion

The data presented in this work suggest that SiO₂-NPs should no longer be viewed as simple imaging agents for biomedical applications, but can also play an active role in mediating biological effects, some of which might ultimately result in an augmented stem cell engraftment potential. Considering, in particular, the modified microenvironment due to a myocardial infarction, which includes ischemia, inflammation and anoikis, where substrate adhesion and early intercellular communication abilities of the transplanted hMSCs are potentially the major contributors to cell engraftment and tissue regeneration, the observed positive SiO₂-NP functional effects may represent a desirable biological improvement.

Future Perspective

Increased adhesion and engraftment of stem cells in the damaged myocardium as well as enhancement of functional gap junctions in stem cell plasma membrane due to interaction with lysosomal activity are interesting effects of NP internalization in human mesenchymal stem cells. NPs could represent a feasible tool to overcome some important limitations of stem cell-based therapies such as the low percentage of surviving and engrafted cells as well as the enhanced occurrence of arrhythmias in the injured cardiac tissue. Further study is needed to better understand whether NP internalization could influence also other aspects of stem cell behavior.

Summary points

- The therapeutic potential of human mesenchymal stem cells (hMSCs) in regenerating injured cardiac tissue is limited due to a low survival rate within the first week after transplantation.
- To enhance transplanted cell survival, adhesion capacity and interaction with the surrounding cells in the injured tissue should be improved.
- The aim of the present work was to evaluate whether any phenotypic effects could be exerted by internalization of silica nanoparticles (SiO₂-NPs) that might ultimately impact on the interaction of hMSCs with the cardiac environment.

Results

- SiO₂-NP internalization impacted hMSC focal adhesion complexes, thus increasing *in vitro* adhesion capacity and engraftment in the myocardial tissue upon cell injection in infarcted isolated rat hearts.
- Furthermore, SiO₂-NP treatment enhanced hMSC expression of Connexin-43, favoring hMSC interaction with cocultured cardiac myoblasts in an ischemia-like environment.

- SiO₂-NPs affected lysosomal function by causing transient inhibition of autophagosome–lysosome fusion.

Conclusion

- Besides the use of NPs as simple imaging agents for biomedical applications, they should be considered for playing an active role in mediating biological effects, some of which might ultimately result in an augmented stem cell engraftment potential.

References

- 1Rohban R, Pieber TR. Mesenchymal stem and progenitor cells in regeneration: tissue specificity and regenerative potential. **Stem Cells Int.** 2017, 5173732 (2017).[Crossref](#), [Medline](#), [Google Scholar](#)
- 2Gallina C, Turinetti V, Giachino C. A new paradigm in cardiac regeneration: the mesenchymal stem cell secretome. **Stem Cells Int.** 2015, 765846 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)
- 3Singh A, Singh A, Sen D. Mesenchymal stem cells in cardiac regeneration: a detailed progress report of the last 6 years (2010–2015). **Stem Cell Res. Ther.** 7(1), 82 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
- 4Lee S, Choi E, Cha M-J, Hwang K-C. Cell adhesion and long-term survival of transplanted mesenchymal stem cells: a prerequisite for cell therapy. **Oxid. Med. Cell. Longev.** 2015, 632902 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)
- 5Taddei ML, Giannoni E, Fiaschi T, Chiarugi P. Anoikis: an emerging hallmark in health and diseases. **J. Pathol.** 226(2), 380–393 (2012).[Crossref](#), [Medline](#), [Google Scholar](#)
- 6Michel J-B. Anoikis in the cardiovascular system: known and unknown extracellular mediators. **Arterioscler. Thromb. Vasc. Biol.** 23(12), 2146–2154 (2003).[Crossref](#), [Medline](#), [Google Scholar](#)
- 7Kavanagh DPJ, Robinson J, Kalia N. Mesenchymal stem cell priming: fine-tuning adhesion and function. **Stem Cell Rev.** 10(4), 587–599 (2014).[Crossref](#), [Medline](#), [Google Scholar](#)
- 8den Haan MC, van Zuylen V-L, Pluijmer NJ *et al.* Discrepant results of experimental human mesenchymal stromal cell therapy after myocardial infarction: are animal models robust enough? **PLoS ONE** 11(4), e0152938 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
- 9Gnecchi M, Zhang Z, Ni A, Dzau VJ. Paracrine mechanisms in adult stem cell signaling and therapy. **Circ. Res.** 103(11), 1204–1219 (2008).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
- 10Liu C, Fan Y, Zhou L *et al.* Pretreatment of mesenchymal stem cells with angiotensin II enhances paracrine effects, angiogenesis, gap junction formation and therapeutic efficacy for myocardial infarction. **Int. J. Cardiol.** 188, 22–32 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)
- 11Valiunas V, Doronin S, Valiuniene L *et al.* Human mesenchymal stem cells make cardiac connexins and form functional gap junctions. **J. Physiol.** 555(Pt 3), 617–626 (2004).[Crossref](#), [Medline](#), [Google Scholar](#)
- 12Patel S, Lee K-B. Probing stem cell behavior using nanoparticle-based approaches. **Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.** 7(6), 759–778 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)

-
- 13Accomasso L, Gallina C, Turinetti V, Giachino C. Stem cell tracking with nanoparticles for regenerative medicine purposes: an overview. **Stem Cells Int.** 2016, 7920358 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 14Liao L. Nanoparticle-dependent labeling of mesenchymal stem cell. **J. Nanosci. Nanotechnol.**14(1), 958–968 (2014).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 15Bokara KK, Ogbu GS, Vidyasagar AJ, Asthana A, Lee JE, Rao CM. Modulation of stem cell differentiation by the influence of nanobiomaterials/carriers. **Curr. Stem Cell Res. Ther.** 9(6), 458–468 (2014).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 16Accomasso L, Cibrario Rocchietti E, Raimondo S *et al.* Fluorescent silica nanoparticles improve optical imaging of stem cells allowing direct discrimination between live and early-stage apoptotic cells. **Small** 8(20), 3192–200 (2012).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 17Catalano F, Accomasso L, Alberto G *et al.* Factors ruling the uptake of silica nanoparticles by mesenchymal stem cells: agglomeration versus dispersions, absence versus presence of serum proteins. **Small** 11(24), 2919–2928 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 18Gallina C, Capeloa T, Saviozzi S *et al.* Human mesenchymal stem cells labelled with dye-loaded amorphous silica nanoparticles: long-term biosafety, stemness preservation and traceability in the beating heart. **J. Nanobiotechnol.** 13, 77 (2015). •• Demonstrates biosafety of silica-nanoparticles we use and optimizes protocol for labeling human mesenchymal stem cells.[Crossref](#), [Medline](#), [Google Scholar](#)
 - 19Alberto G, Miletto I, Viscardi G *et al.* Hybrid cyanine–silica nanoparticles: homogeneous photoemission behavior of entrapped fluorophores and consequent high brightness enhancement. **J. Phys. Chem. C.** 113(50), 21048–21053 (2009).[Crossref](#), [Google Scholar](#)
 - 20Miletto I, Gilardino A, Zamburlin P *et al.* Highly bright and photostable cyanine dye-doped silica nanoparticles for optical imaging: photophysical characterization and cell tests. **Dye Pigment**84(1), 121–127 (2010).[Crossref](#), [Google Scholar](#)
 - 21Raimondo S, Penna C, Pagliaro P, Geuna S. Morphological characterization of GFP stably transfected adult mesenchymal bone marrow stem cells. **J. Anat.** 208(1), 3–12 (2006).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 22Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. **Nat. Methods** 9(7), 671–675 (2012).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - 23Penna C, Raimondo S, Ronchi G *et al.* Early homing of adult mesenchymal stem cells in normal and infarcted isolated beating hearts. **J. Cell. Mol. Med.** 12(2), 507–521 (2008).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 24Ziambaras K, Lecanda F, Steinberg TH, Civitelli R. Cyclic stretch enhances gap junctional communication between osteoblastic cells. **J. Bone Miner. Res.** 13(2), 218–228 (1998).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 25Cselenyák A, Pankotai E, Horváth EM *et al.* Mesenchymal stem cells rescue cardiomyoblasts from cell death in an *in vitro* ischemia model via direct cell-to-cell connections. **BMC Cell Biol.**11(1), 29 (2010).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 26Parsons JT, Horwitz AR, Schwartz MA. Cell adhesion: integrating cytoskeletal dynamics and cellular tension. **Nat. Rev. Mol. Cell Biol.** 11(9), 633–643 (2010).[Crossref](#), [Medline](#), [Google Scholar](#)
 - 27Ridley AJ. Life at the leading edge. **Cell** 145(7), 1012–1022 (2011).[Crossref](#), [Medline](#), [Google Scholar](#)

-
- **28**Webb DJ, Parsons JT, Horwitz AF. Adhesion assembly, disassembly and turnover in migrating cells - over and over and over again. **Nat. Cell Biol.** 4(4), E97–E100 (2002).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **29**Humphries JD, Wang P, Streuli C, Geiger B, Humphries MJ, Ballestrem C. Vinculin controls focal adhesion formation by direct interactions with talin and actin. **J. Cell Biol.** 179(5), 1043–1057 (2007).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **30**Wang C, Gong Y, Zhong Y, Yao Y, Su K, Wang D-A. The control of anchorage-dependent cell behavior within a hydrogel/microcarrier system in an osteogenic model. **Biomaterials** 30(12), 2259–2269 (2009).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **31**Besser A, Safran SA. Force-induced adsorption and anisotropic growth of focal adhesions. **Biophys. J.** 90(10), 3469–3484 (2006).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **32**Wolfenson H, Henis YI, Geiger B *et al.* The heel and toe of the cell's foot: a multifaceted approach for understanding the structure and dynamics of focal adhesions. **Cell Motil. Cytoskeleton** 66(11), 1017–1029 (2009).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **33**Valiente-Alandi I, Schafer AE, Blaxall BC. Extracellular matrix-mediated cellular communication in the heart. **J. Mol. Cell. Cardiol.** 91, 228–237 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **34**Tay CY, Cai P, Setyawati MI *et al.* Nanoparticles strengthen intracellular tension and retard cellular migration. **Nano Lett.** 14(1), 83–88 (2014).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **35**Hou Y, Cai K, Li J *et al.* Effects of titanium nanoparticles on adhesion, migration, proliferation, and differentiation of mesenchymal stem cells. **Int. J. Nanomedicine** 8, 3619–3630 (2013).[Medline](#), [Google Scholar](#)
 - **36**Hackenberg S, Scherzed A, Technau A, Froelich K, Hagen R, Kleinsasser N. Functional responses of human adipose tissue-derived mesenchymal stem cells to metal oxide nanoparticles *in vitro*. **J. Biomed. Nanotechnol.** 9(1), 86–95 (2013).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **37**Akimov SS, Krylov D, Fleischman LF, Belkin AM. Tissue transglutaminase is an integrin-binding adhesion coreceptor for fibronectin. **J. Cell Biol.** 148(4), 825–838 (2000).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **38**Song H, Chang W, Lim S *et al.* Tissue transglutaminase is essential for integrin-mediated survival of bone marrow-derived mesenchymal stem cells. **Stem Cells** 25(6), 1431–1438 (2007).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **39**Benoit DSW, Tripodi MC, Blanchette JO, Langer SJ, Leinwand LA, Anseth KS. Integrin-linked kinase production prevents anoikis in human mesenchymal stem cells. **J. Biomed. Mater. Res. A** 81(2), 259–268 (2007).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **40**Song S-W, Chang W, Song B-W *et al.* Integrin-linked kinase is required in hypoxic mesenchymal stem cells for strengthening cell adhesion to ischemic myocardium. **Stem Cells** 27(6), 1358–1365 (2009).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **41**Chang W, Song BW, Hwang KC. Mesenchymal stem cell survival in infarcted myocardium: adhesion and anti-death signals. In: **Stem Cells and Cancer Stem Cells, Volume 10: Therapeutic Applications in Disease and Injury**. Hayat MA (Ed.). Springer, Dordrecht, the Netherlands, 35–43 (2013).[Google Scholar](#)
 - **42**Sato D, Xie L-H, Sovari AA *et al.* Synchronization of chaotic early afterdepolarizations in the genesis of cardiac arrhythmias. **Proc. Natl Acad. Sci. USA** 106(9), 2983–2988 (2009).[Crossref](#), [Medline](#), [Google Scholar](#)

-
- **43**de Bakker JM, van Capelle FJ, Janse MJ *et al.* Slow conduction in the infarcted human heart. “Zigzag” course of activation. **Circulation** 88(3), 915–926 (1993).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **44**Beeres SLMA, Zeppenfeld K, Bax JJ *et al.* Electrophysiological and arrhythmogenic effects of intramyocardial bone marrow cell injection in patients with chronic ischemic heart disease. **Heart Rhythm** 4(3), 257–265 (2007).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **45**Chen H-SV, Kim C, Mercola M. Electrophysiological challenges of cell-based myocardial repair. **Circulation** 120(24), 2496–2508 (2009).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **46**Roell W, Lewalter T, Sasse P *et al.* Engraftment of connexin 43-expressing cells prevents post-infarct arrhythmia. **Nature** 450(7171), 819–824 (2007).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **47**Hahn J-Y, Cho H-J, Kang H-J *et al.* Pre-treatment of mesenchymal stem cells with a combination of growth factors enhances gap junction formation, cytoprotective effect on cardiomyocytes, and therapeutic efficacy for myocardial infarction. **J. Am. Coll. Cardiol.** 51(9), 933–943 (2008).[Crossref](#), [Medline](#), [CAS](#), [Google Scholar](#)
 - **48**Song H, Hwang HJ, Chang W *et al.* Cardiomyocytes from phorbol myristate acetate-activated mesenchymal stem cells restore electromechanical function in infarcted rat hearts. **Proc. Natl Acad. Sci. USA** 108(1), 296–301 (2011).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **49**Carvalho JL, Braga VBA, Melo MB *et al.* Priming mesenchymal stem cells boosts stem cell therapy to treat myocardial infarction. **J. Cell. Mol. Med.** 17(5), 617–625 (2013).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **50**Han J, Kim B, Shin J-Y *et al.* Iron oxide nanoparticle-mediated development of cellular gap junction crosstalk to improve mesenchymal stem cells' therapeutic efficacy for myocardial infarction. **ACS Nano** 9(3), 2805–2819 (2015).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **51**Schütz I, Lopez-Hernandez T, Gao Q *et al.* Lysosomal dysfunction caused by cellular accumulation of silica nanoparticles. **J. Biol. Chem.** 291(27), 14170–14184 (2016). • Highlights a role of silica-nanoparticles in cell proteins turnover.[Crossref](#), [Medline](#), [Google Scholar](#)
 - **52**Wang J, Yu Y, Lu K *et al.* Silica nanoparticles induce autophagy dysfunction via lysosomal impairment and inhibition of autophagosome degradation in hepatocytes. **Int. J. Nanomed.** 12, 809–825 (2017).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **53**Ballabio A. The awesome lysosome. **EMBO Mol. Med.** 8(2), 73–76 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **54**Pu J, Guardia CM, Keren-Kaplan T, Bonifacino JS. Mechanisms and functions of lysosome positioning. **J. Cell Sci.** 129(23), 4329–4339 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **55**Laing JG, Beyer EC. The gap junction protein connexin43 is degraded via the ubiquitin proteasome pathway. **J. Biol. Chem.** 270(44), 26399–26403 (1995).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **56**Laing JG, Tadros PN, Westphale EM, Beyer EC. Degradation of connexin43 gap junctions involves both the proteasome and the lysosome. **Exp. Cell Res.** 236(2), 482–492 (1997).[Crossref](#), [Medline](#), [Google Scholar](#)
 - **57**Iyyathurai J, Decuyper J-P, Leybaert L, D'hondt C, Bultynck G. Connexins: substrates and regulators of autophagy. **BMC Cell Biol.** 17(Suppl. 1), 20 (2016).[Crossref](#), [Medline](#), [Google Scholar](#)

Supplementary data

To view the supplementary data that accompany this paper please visit the journal website at: <http://www.futuremedicine.com/doi/suppl/10.2217/nnm-2017-0309>

Financial & competing interests disclosure

This study was supported by GIAC_RILO_17_02 and PAGP_RILO_16_01. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

Ethical conduct of research

This study conforms to the *Guide for the Care and Use of Laboratory Animals* published by the US National Institutes of Health (NIH Publication No. 85–23, revised 1996) and in compliance with the European Directive 2010/63/EU on the protection of animals used for scientific purposes. The local ethical committee approved the research project.

Open access

This work is licensed under the Attribution-NonCommercial-NoDerivatives 4.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>