



AperTO - Archivio Istituzionale Open Access dell'Università di Torino

## Dupilumab to target interleukin 4 for inflammatory bowel disease? Hypothesis based on a translational message

# This is the author's manuscript Original Citation: Availability: This version is available http://hdl.handle.net/2318/1707539 since 2019-08-31T13:54:47Z Published version: DOI:10.23736/S1120-4826.19.02556-4 Terms of use: Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

# Dupilumab to target interleukin 4 for inflammatory bowel disease? Hypothesis based on a translational message

Angelo Armandi<sup>1</sup>, Silvia Bonetto<sup>1</sup>, Rinaldo Pellicano<sup>2</sup>, Gian Paolo Caviglia<sup>1</sup>, Marco Astegiano<sup>2</sup>, Giorgio Maria Saracco<sup>1</sup>, Davide Giuseppe Ribaldone<sup>3</sup>

<sup>1</sup>Department of Medical Sciences, Division of Gastroenterology, University of Torino, Turin, Italy <sup>2</sup>Molinette Hospital, Unit of Gastroenterology, Turin, Italy

<sup>3</sup>Department of Surgical Sciences, University of Torino, Turin, Italy

Rinaldo Pellicano ORCID identifiers: 0000-0003-3438-0649 Gian Paolo Caviglia ORCID identifiers: 0000-0002-0529-9481 Marco Astegiano ORCID identifiers: 0000-0003-0916-1188 Giorgio Maria Saracco ORCID identifiers: 0000-0001-5310-4143 Davide Giuseppe Ribaldone ORCID identifiers: 0000-0002-9421-3087

### ABSTRACT

Dupilumab is a human monoclonal antibody that has recently been approved for the treatment of mild-tosevere atopic dermatitis. Dupilumab inhibits the interleukin (IL)-4/IL-13 signalling by binding specific subunits of the receptors. IL-4 is involved in the regulation of both innate and adaptive intestinal immune system, thus its possible role in the pathogenesis of inflammatory bowel disease (IBD) has been investigated. Several polymorphisms of IL-4 have been found in patients with IBD. Furthermore, the crucial role of IL-4 has been demonstrated in Th2-driven models of colitis as well as in Th1-related models of this disease. Finally, many studies have shown the increased production of IL-4 in two models of experimental colitis: the dextran sulphate sodium – related colitis and the oxazolone-related colitis. In the latter, a dual antagonist of IL-4/IL-13 has proved to ameliorate the course of the disease. In conclusion, IL-4 antagonists might play a role in the treatment of IBD, especially in ulcerative colitis, which shows a classic pattern of Th2-related disease.

*Keywords:* Crohn's disease; Dupilumab; inflammatory bowel disease; interleukin-4 antagonists; interleukin-13; ulcerative colitis

### Introduction

The burden of autoimmune disorders is progressively increasing worldwide. The first data about the epidemiology of autoimmune diseases were published in 1997 by Jacobson et al., and showed a prevalence of 3.2% between 1967 and 1997 in the United States.<sup>1</sup> Subsequently, one Danish study performed by Eaton et al. estimated a prevalence of 5.3%, between 1977 and 2001, based on national hospitalization registry data.<sup>2</sup> The most recent data, which cover 20 years, from 1989 to 2008, are extrapolated from a review conducted by Cooper et al., which showed a worldwide prevalence of 7.6-9.4%.<sup>3</sup>

As a consequence of the increased prevalence of autoimmune disorders, new drugs, specifically monoclonal antibodies, which target critical cytokines of the signalling pathways involved in the mechanisms of disease, are progressively studied. Interestingly, clinical trials contribute to translational message from basic research to the ultimate understanding of the functioning of targeted molecules with the hope to optimize and perhaps personalize treatments.<sup>4</sup> Thanks to monoclonal antibodies, it has become clearer which are the sites of human body (i.e. intestinal mucosa and skin) where the targeted cytokines mainly work, and which is their role in the pathogenesis of each specific disease.

On the basis of already known pathogenetic models of inflammatory bowel diseases (IBD), this narrative review describes the recent development of the new drug dupilumab, targeting interleukin-4 (IL-4), explaining the reasons of its potential application in the treatment of IBD.

### The role of dupilumab in atopic dermatitis

Atopic dermatitis is a widespread chronic inflammatory skin disease. It is characterized by a damaged skin barrier function, with an increased transepidermal water loss and inflammation, a reduction of long chain fatty acid in the lipid bilayer and a lympho-histiocytic infiltration.<sup>5-7</sup> Although its causal agents remain unknown, in the last years also gastrointestinal microbiota alterations, with consequences in the development of the host's immune system, have been involved as potential mechanisms of damage.<sup>8,9</sup> These processes are mainly Th2-cell mediated and therefore Th2-related cytokines, such as IL-4 and IL-13, are increased in the

involved skin, as well as dendritic epidermal cells. Conversely, Th1-related cytokines, like interferon (INF)- $\gamma$  and IL-2, are decreased.<sup>10-12</sup> IL-4 and IL-13 induce spongiosis of the skin and reduce filaggrin gene expression in keratinocytes, leading to a damage of the barrier function.<sup>13,14</sup> Furthermore, IL-4 stimulates B-cell differentiation and an immunoglobulin (Ig) subclass switch, resulting in IgE production.<sup>15</sup>

Dupilumab is a human monoclonal antibody approved by the Federal and Drugs Adiministration on March 28, 2017 for the treatment of adult patients with moderate-to severe atopic dermatitis which is not adequately controlled with topical therapies.<sup>16</sup> It inhibits the IL-4 and IL-13 signalling by binding the IL-4R- $\alpha$  subunit<sup>17</sup>, which is part of both IL-4R and IL-13R. It thereby downregulates the JAK-STAT downstream signalling pathway.<sup>18</sup>

In patients with atopic dermatitis, many genes are regulated through the JAK-STAT pathway<sup>19</sup>: its activation in fact, leads to the downregulation of skin barrier proteins, such as loricrin, involucrin and filaggrin<sup>14,20</sup> and it promotes the production of chemokines such as CCL3L1, CCL8, CCL24, CCL25, CCL26, CXCL6 and CXCL16 by keratinocytes.<sup>21</sup> Moreover, it stimulates the Th2-cell differentiation, resulting in an increased production of IL-3, IL-4, IL-5, IL-6, IL-10 and IL-13<sup>22</sup>, and causes the B-cell activation which is involved in the IgE production.<sup>23,24</sup> Hence, by blocking the IL-4 and IL-13 signalling pathways, dupilumab acts on the critical disease mechanisms of atopic dermatitis.

Dupilumab has been approved for subcutaneous administration, starting from an initial dose of 600 mg, followed by a weekly dose of 300 mg.<sup>17</sup> Steady-state concentrations are obtained after 16 weeks of treatment.<sup>17</sup> This drug was firstly tested in patients with persistent allergic asthma and eosinophilia, in whom it improved lung function and reduced exacerbations.<sup>25</sup> Since allergic asthma and atopic dermatitis are genetically linked and share the same immunopathological mechanisms<sup>26</sup>, randomized clinical trials with dupilumab for the indication of atopic dermatitis were subsequently conducted.<sup>27</sup> The safety profile of dupilumab was superior to conventional immunosuppressive drugs, like cyclosporine or methotrexate.<sup>27</sup> The most common side effects were erythema or edema at the injection-site and conjunctivitis.<sup>28</sup>

### The role of IL-4 in the intestinal immune system

IBD are distinguished in two main phenotypes, Crohn's disease (CD) and ulcerative colitis (UC), both characterized by chronic inflammation of the intestinal mucosa and variable course <sup>29</sup>

While the etiology of IBD remains unclear, the hypothesis of a multifactorial genesis supposes that an antigenic stimulus may act as a "trigger" for an abnormal activation of the immune reaction towards colorectal mucosa. The intraluminal antigenic stimulation would facilitate the loss of the immunotolerance towards indigenous bacterial flora and consequently activates the gut-associated immune system. <sup>30,31</sup> Pharmacological management of IBD, partly reflects our limited understanding of the pathogenesis of the disease, and its complex phenotypic presentation.<sup>32-35</sup>

IL-4 has also been investigated as a possible pathogenetic factor in the onset of IBD. Changes in cytokines' expression have been frequently found in patients with IBD,<sup>36</sup> and several speculations about the role of the intestinal lymph cells have been made. Th17-cells, which produce IL-17, IL-21 and IL-22, are thought to cause inflammatory and autoimmune reactions.<sup>37</sup> Conversely, T-reg cells, which produce IL-10, and transforming growth factor (TGF)-beta), are known for their regulatory effects on immune responses.<sup>38</sup> IL-4 and IL-5, produced by Th2-cells, have been classically considered as anti-autoimmune cytokine by

inhibiting Th17-cells.<sup>39</sup> Thus, IL-4 and IL-10 have always been thought to be protective cytokines in the pathogenesis of IBD.

Nonetheless, IL-4 directly regulates intestinal epithelial cell functions: it modulates the growth of epithelial cells, can alter the intestinal mucosal barrier function (a key element in the IBD damage<sup>40</sup>, and stimulates the adherence and migration of neutrophils to the epithelium.<sup>41</sup> Therefore, overproduction of IL-4 may subvert the immunological homeostasis between the intestinal immune system and the environmental antigens<sup>42</sup>, including the gut microbiota<sup>43</sup>, by promoting enhanced translocation of luminal pathogens and recruiting inflammatory cells at the very beginning of the disease.

### The possible role of IL-4 polymorphisms in the genesis of IBD

To support of the potential involvement of IL-4 alterations in the pathogenesis of IBD , a recent study conducted by Ebrahimi Daryani et al. investigated the presence of polymorphisms that may predispose to the onset of this disease. They found that all genotypes studied were correlated to IBD: C-allele of IL-4-590 polymorphism and T-allele of IL-4-1098 polymorphism were found in the whole group of IBD patients.<sup>44</sup>

Another study pointed out the importance of the above-mentioned polymorphisms.<sup>45</sup> The gene of IL-4 maps on the long arm (q23-31) of chromosome 5.<sup>46</sup> A C $\rightarrow$ T exchange has been identified in position -590, which correlates with an increased transcriptional activity and secretion of IgE.<sup>47</sup> It has been suggested a correlation between this promoter region polymorphism and allergic asthma, atopic dermatitis and rheumatoid arthritis in US and Japanese families, despite this evidence has not been confirmed in UK and Australian families.<sup>48-51</sup> Another C $\rightarrow$ T substitution has been subsequently discovered at position -34 in the untranslated region which is in linkage disequilibrium with the position -590 in the promoter region.<sup>52</sup>

The IL-4R, instead, maps on the short arm (16p12) of chromosome 16. A G $\rightarrow$ A substitution at position 1902 (change from glutamine to arginine at amino acid 576) leads to an enhanced signalling activity and is commonly found in patients with atopy.<sup>53,54</sup>

Since the pericentromeric region of chromosome 16, which contains the IL-4R gene<sup>55</sup> and the long arm of chromosome 5, which contains the IL-4 gene<sup>56</sup>, have been previously identified as susceptibility loci for CD, this study examined the possible role of the above-mentioned polymorphisms in the onset of the disease. The authors found that both variant IL-4 and the association between variant IL-4 and variant IL-4R are related to CD, suggesting a role of IL-4 in the pathogenesis of CD.<sup>45</sup>

### T-cell receptor (TCR) a chain-knockout mice as IL-4-driven model of Th2-related colitis.

The murine models have been surprisingly useful for understanding the genesis of IBD, in which the cytokines' cascades are tightly connected and the role of each signal pathway receptor and the cytokines involved are barely understood. Table 1 shows murine models regarding the role of IL-4 and IL-13 in experimental colitis.

It has been shown that TCR- $\alpha$  chain-deficient mice developed IBD, and the colitis was associated with increased number of Th2-cells producing eminently IL-4.<sup>57-59</sup> Iijima et al. showed that approximately 60% of these mice, including those treated with mock (inactive) anti-IL-4 antibodies and those left untreated, spontaneously developed IBD. Conversely, those mice treated with anti-IL-4 antibodies did not express any clinical or histological signs of IBD, with a decrease of Th2 related cytokines and an increase in INF- $\gamma$  expression.<sup>60</sup> These findings suggest that aberrant mucosal Th2-cells are involved in the development of IBD

and the treatment with anti-IL-4 antibodies altered the cytokines profile from Th2 to Th1 types, as a result of the decrease of B-cells.<sup>60</sup>

A study, conducted by another group, gave direct evidence of the involvement of IL-4 in the disease processes, showing that double knockout murine models (TCR- $\alpha$  chain and IL-4 deficient) had a reduced incidence of colitis.<sup>61</sup>

Using the same model of colitis, Mizoguchi et al. have studied the possible role of IFN- $\gamma$ , rather than IL-4, for the onset of the disease. They created double-mutant mice which were negative for TCR- $\alpha$ , and negative for either IL-4 or IFN- $\gamma$ . The lack of IL-4 was confirmed to suppress the onset of colitis, whilst the lack of IFN- $\gamma$  did not interfere with the onset of colitis, which resulted as similar as the colitis developed in the single TCR- $\alpha$  mutant mice.<sup>61</sup>

### The role of IL-4 in the T cell transfer model of colitis.

Although IL-4 is involved in Th2 immune responses<sup>62</sup>, it can also interfere with the Th1 responses. The latter are evocated when antigen presenting cells (APC) release IL-12, inducing the differentiation of CD4+ T cells, which produce IFN- $\gamma$  and IL-2.<sup>62</sup> CD shows a typical Th1 pattern of disease, with a large amount of IFN- $\gamma$  and TNF- $\alpha$ .<sup>63</sup> IL-4 is classically known to suppress Th1-type inflammatory response.<sup>64</sup>

However, there is increasing evidence that IL-4 can worsen Th1 related immune responses.<sup>65</sup> In a study conducted by Fort et al.<sup>66</sup>, the effect of IL-4 treatment was tested in the CD4(+)CD45RB T cell transfer model of colitis, an established model of Th1-related disease.<sup>67</sup> IL-4 treatment was shown to exacerbate Th1-colonic inflammation in this model and did not switch the colitis to a Th2-type immune response.<sup>66</sup> As the receptor of IL-4 is expressed in many types of cells other than T cells (i.e. B cells, monocytes, intestinal epithelial cells)<sup>68-71</sup>, in this study the effect of IL-4 treatment was also tested using CD4(+)CD45RB T cells from mice deficient for  $\alpha$  chain receptor of IL-4 (IL-4R- $\alpha$ ). The treatment was able to exacerbate colitis in this model, suggesting that IL-4 has a proinflammatory effect in both T cells and non-T cells.<sup>66</sup> To support this evidence, it has been shown that IL-4 therapy for human cancers leads to diarrhea and abdominal pain, related to mucosal ulcerations which resemble the early lesions of CD.<sup>72</sup> On biopsies obtained from ileal resection it has been found an increase of IL-4 m RNA.<sup>73</sup>

### The role of IL-4 in the dextran sulphate sodium-induced colitis and in the oxazolone-induced colitis.

The effect of IL-4 in IBD has also been studied by inducing experimental colitis. A study, conducted by Stevceva et al.<sup>74</sup>, used the dextran sulphate sodium to induce an experimental colitis, that resembles UC<sup>74-79</sup>, in IL-4 deficient mice. The experimental colitis was significantly milder in IL-4 deficient mice, with lower histological activity score. In parallel, they found and increased amount of IFN-y.<sup>74</sup> This result suggests a role of IL-4 in influencing the severity of the colitis. Another study investigated the role of IL-33 in the same model of colitis.<sup>80</sup> Through its receptor ST2, IL-33 acts in different ways according to the context.<sup>81,82</sup> It is expressed in the epithelium and endothelium, and increases in response to inflammatory stimuli or necrosis.<sup>81,82</sup> It is expressed in almost all innate immune cells<sup>81-87</sup> and can promote Th1/Th17 responses<sup>88,89</sup>, as well as Th2 responses and increase of IL-5.<sup>81-88</sup> The study conducted by Pushparaj et al.<sup>80</sup>, demonstrated that IL-33 and ST2 are the earliest genes induced in dextran sulphate sodium-related colitis and act as major pathogenic factor in the exacerbation of UC. Furthermore, they showed, for the first time, that the immune response led by IL-33 mandatorily requires the activation of IL-4 to be fulfilled.<sup>80</sup> Furthermore, one study conducted by Boirivant et al. used a haptenating agent, oxazolone, to induce a Th2-related colitis.<sup>90</sup> This colitis only involves the distal half of the colon and its pattern of disease resembles UC.<sup>90</sup> Due to its haptenating power, oxazolone has been shown to stimulate T cells, resulting in increased amount of IL-4, counterbalanced by an increase of TGF-B.90 Oxazolone colitis is prevented by administering anti-IL-4 agents.<sup>90</sup> This colitis is immunologically similar to that colitis occurring in the TCR-α chain knockout mice described above.<sup>57-59</sup> The two kinds of colitis differ from the extension of disease: oxazolone-induced colitis only involves the distal colon and it is an acute, self-limited inflammation<sup>94</sup>, whereas colitis occurring in TCR- $\alpha$  chain knockout mice involves the appendix and then involves the entire colon, persisting as chronic inflammation.<sup>91,92</sup> The difference may be related to the different expression of TGF-β among the two kinds of colitis.<sup>90</sup> Finally, Kasaian et al. have evaluated the possible therapeutic role of IL-4/IL-13 dual antagonist in oxazolone colitis.<sup>93</sup> Due to the pleiotropic activity of most cytokines<sup>94-100</sup>, the authors developed a dual antagonist to strengthen the therapeutic activity.<sup>93</sup> IL-13, like IL-4, contribute to intestinal inflammation, as it causes a permeabilization of the epithelial barrier mucosa by inducing the apoptosis of epithelial cells and disruption of tight junctions.<sup>97</sup> By elevating the expression of claudin-2, a tight junction pore protein, IL-13 promotes ion flux across the barrier.<sup>101,102</sup>

This dual antagonist was proved to block both cytokines <sup>93,103</sup> and was shown to ameliorate the course of the disease.<sup>93</sup> Serum concentration of the IL-4/IL-13 antagonist was inversely proportional to disease severity, colon tissue expression of pro-inflammatory genes, and serum amyloid P concentration.<sup>93</sup> This result emphasizes the role of IL-4 and IL-13 in the onset of UC and suggests a possible role for IL-4/IL-13 antagonists in the treatment of UC.

### **Conclusions.**

Many studies have provided evidence for a critical role of IL-4 in the immunopathogenesis of IBD . Specific polymorphisms of IL-4 have been found in crucial loci<sup>44,45</sup>; furthermore, the study of different models of colitis, both in knockout mice<sup>60,61,66</sup> and in experimentally-induced colitis<sup>74,80,90,93</sup>, has permitted to understand the eminent proinflammatory activity of IL-4, acting either in the genesis of disease, or during the exacerbations. IL-4 is involved both in innate<sup>41,43</sup> and adaptive immune system<sup>62,64,65</sup>; it can influence the intestinal barrier function<sup>41</sup> and has been proved to stimulate Th2-related responses<sup>62</sup>, therefore it seems to play a role in Th2-related disease, like bronchial asthma, atopic dermatitis and UC. Nonetheless IL-4 may interfere with Th1 responses, intensifying the inflammatory process, and may play a role in CD, which is eminently a Th1-related disease [61-62]. For these reasons we hypothesize that antagonists of IL-4, like dupilumab, that has been recently approved for the treatment of mild-to-severe atopic dermatitis<sup>16,17</sup>, might play a role in the treatment of IBD and specific clinical trial, at least in UC, should be encouraged.

### **Conflict of interest**

The authors declare that they have no conflict of interest.

### References

1. Jacobson DL, Gange SJ, Rose NR, Graham NM. Epidemiology and estimated population burden of selected autoimmune diseases in the United States. Clin Immunol Immunopathol 1997;84:223-43.

2. Eaton WW, Rose NR, Kalaydjian A, Pedersen MG, Mortensen PB. Epidemiology of autoimmune diseases in Denmark. J Autoimmun 2007;29:1-9.

3. Cooper GS, Bynum MLK, Somers EC. Recent insights in the epidemiology of autoimmune diseases: improved prevalence estimates and understanding of clustering of diseases. J Autoimmun 2009;33:197-207.

4. Actis GC, Pellicano R. Chronic (inflammatory) diseases: precision medicine versus comprehensive understanding? Minerva Med 2018; 109(2):150-151.

5. Wollenberg A, Ehmann LM. Long term treatment concepts and proactive therapy for atopic eczema. Ann Dermatol 2012;24:253-60

6. Proksch E, Folster-Holst R, Jensen JM. Skin barrier function, epidermal proliferation and differentiation in eczema. J Dermatol Sci 2006;43:159-69.

7. Macheleidt O, Kaiser HW, Sandhoff K. Deficiency of epidermal protein-bound omegahydroxyceramides in atopic dermatitis. J Invest Dermatol 2002;119:166-73.

8. Gibiino G, Ianito G, Cammarota G, Gasbarrini A. The gut microbiota: its anatomy and physiology over a lifetime. Minerva Gastroenterol Dietol 2017;63:329-36.

9. Pellicano R, Cisarò F, Durazzo M, Ribaldone DG. When is it useful to act on microbiota in atopic children? A recent experience. Minerva Pediatr. 2019;71:1-3.

10. Esnault S, Benbernou N, Lavaud F, Shin HC, Potron G, Guenounou M. Differential spontaneous expression of mRNA for IL-4, IL-10, IL-13, IL-2 and interferon-gamma (IFN-gamma) in peripheral blood mononuclear cells (PBMC) from atopic patients. Clin Exp Immunol 1996;103:111-8.

11. Jujo K, Renz H, Abe J, Gelfand EW, Leung DY. Decreased interferon gamma and increased interleukin-4 production in atopic dermatitis promotes IgE synthesis. J Allergy Clin Immunol 1992; 90:323-31.

12. Kaminishi K, Soma Y, Kawa Y, Mizoguchi M. Flow cytometric analysis of IL-4, IL-13 and IFN-gamma expression in peripheral blood mononuclear cells and detection of circulating IL-13 in patients with atopic

dermatitis provide evidence for the involvement of type 2 cytokines in the disease. J Dermatol Sci 2002;29:19-25.

13. Danso MO, van Drongelen V, Mulder A, van Esch J, Scott H, van Smeden J et al. TNF- $\alpha$  and Th2 cytokines induce atopic dermatitis-like features on epidermal differentiation proteins and stratum corneum lipids in human skin equivalents. J Invest Dermatol 2014;134:1941-50

14. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A et al. Cytokine modulation of atopic dermatitis filaggrin skin expression. J Allergy Clin Immunol 2009;124:R7-R12.

15. Bieber T. Atopic dermatitis. N Eng J Med 2008; 358:1483-94.

16. Regeneron Pharmaceuticals Inc. Regeneron and Sanofi announce FDA approval of DUPIXENT® (dupilumab), the first targeted biologic therapy for adults with moderate-to-severe atopic dermatitis. PR Newswire; 2017.

17. Regeneron. Dupixent® (dupilumab) prescribing information; 2017

 Andrews R, Rosa L, Daines M, Khurana Hershey G. Reconstitution of a functional human type II IL-4/IL-13 receptor in mouse B cells: demonstration of species specificity. J Immunol 2001;166:1716-22.

19. Bao L, Zhang H, Chan LS. The involvement of the JAK-STAT signaling pathway in chronic inflammatory skin disease atopic dermatitis. Jakstat 2013; 2:e24137.

20. Kim BE, Leung DY, Boguniewicz M, Howell MD. Loricrin and involucrin expression is down-regulated by Th2 cytokines through STAT-6. Clin Immunol 2008;126:332–7.

21. Bao L1, Shi VY, Chan LS. IL-4 up-regulates epidermal chemotactic, angiogenic, and pro-inflammatory genes and down-regulates antimicrobial genes in vivo and in vitro: relevant in the pathogenesis of atopic dermatitis. Cytokine. 2013;61:419-25.

22. Chen L, Martinez O, Overbergh L, Mathieu C, Prabhakar BS, Chan LS. Early up-regulation of Th2 cytokines and late surge of Th1 cytokines in an atopic dermatitis model. Clin Exp Immunol 2004;138:375-87.

23. McKenzie AN, Culpepper JA, de Waal Malefyt R, Brière F, Punnonen J, Aversa G et al. Interleukin 13, a T-cell derived cytokine that regulates human monocyte and B-cell function. Proc Natl Acad Sci USA 1993; 90:3735-9.

24. Pernis AB, Rothman PB. JAK-STAT signaling in asthma. J Clin Invest 2002;109:1279-83.

25. Wenzel S, Ford L, Pearlman D, Spector S, Sher L, Skobieranda F et al. Dupilumab in persistent asthma with elevated eosinophil levels. N Engl J Med 2013;68:2455-66.

26. Wollenberg A, Oranje A, Deleuran M, Simon D, Szalai Z, Kunz B et al. ETFAD/EADV Eczema task force 2015 position paper on diagnosis and treatment of atopic dermatitis in adult and paediatric patients. J Eur Acad Dermatol Venereol 2016; 30:729-47.

27. Seegraber M, Srour J, Walter A, Knop M, Wollenberg A. Dupilumab for treatment of atopic dermatitis. Expert Rev Clin Pharmacol 2018;11:467-74.

28. Wollenberg A, Ariens L, Thurau S, Van Luijk C, Seegräber M, De Bruin-Weller M. Conjunctivitis occurring in atopic dermatitis patients treated with dupilumab – clinical characteristics and treatment. J Allergy Clin Immunol Pract 2018;6:1778-80.

29. Ribaldone DG, Simondi D, Manca A, Demarchi B, Pulitanò R, Astegiano M et al. Features of inflammatory bowel disease followed in a second level center in Northern Italy. Minerva Med. 2017; 108(5):481-482.

30. Ribaldone DG, Pellicano R, Actis GC. Inflammation: a highly conserved, Janus-like phenomenon-a gastroenterologist' perspective. J Mol Med (Berl). 2018;96:861–71.

31. Fagoonee S, Pellicano R, Actis GC. ADAM17 and gastrointestinal tract diseases: clinical aspects with translational messages. Minerva Biotecnol. 2018;30(1):22–8.

32. Gargallo CJ, Lué A, Gomollón F. Biosimilars in inflammatory bowel disease. Minerva Med. 2017;108:239-54.

33. Actis GC, Pellicano R, Ribaldone DG. A concise history of thiopurines for inflammatory bowel disease: From anecdotal reporting to treat-to-target algorithms. Rev Recent Clin Trials. 2019;14:4-9.

34. Tursi A, Allegretta L, Chiri S, Della Valle N, Elisei W, Forti G, et al. Effectiveness and safety of infliximab biosimilar CT-P13 in treating ulcerative colitis: a real life experience in IBD primary centers. Minerva Gastroenterol Dietol. 2017;63:313-8.

35. Ribaldone DG, Dileo I, Pellicano R, Resegotti A, Fagoonee S, Vernero M, et al. Severe ulcerative colitis: predictors of response and algorithm proposal for rescue therapy. Ir J Med Sci. 2018;187:385-92.

36. Niessner M, Volk BA. Altered Th1/Th2 cytokine profiles in the intestinal mucosa of patients with inflammatory bowel disease as assessed by quantitative reversed transcribed polymerase chain reaction (RT-PCR). Clin Exp Immunol 1995;101:428-35.

37. Noack M, Miossec P. Th17 and regulatory T cell balance in autoimmune and inflammatory diseases. Autoimmun Rev 2014;13:668-77.

38. Sakaguchi S. Naturally arising CD4+ regulatory T cells for immunologic self-tolerance and negative control of immune responses. Ann Rev Immunol 2004;22:531-62.

39. Harrington LE, Hatton RD, Mangan PR. Interleukin 17-producing CD+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. Nat Immunol 2005;6:1123-32.

40. Caviglia GP, Dughera F, Ribaldone DG, Rosso C, Abate ML, Pellicano R, et al. Serum zonulin in patients with inflammatory bowel disease: a pilot study. Minerva Med. 2019 Apr;110(2):95-100.

41. Colgan SP, Resnick MB, Parcos CA, Delp-Archer C, McGuirk D, Bacarra AE et al. IL-4 directly modulates function of a model human intestinal epithelium. J Immunol 1994;153:2122-9.

42. Vajdy M, Kosco-Vilbois MH, Kopf M, Kohler G, Lycke N. Impaired mucosal immune responses in interleukin 4-targeted mice. J Exp Med 1995;181:41-53.

43. Hermon-Taylor J. Causation of Crohn's disease: the impact of clusters. Gastroenterology 1995;104:643-6.

44. Ebrahimi Daryani N, Saghazadeh A, Moossavi S, Sadr M, Shahkarami S, Soltani S et al. Interleukin-4 and Interleukin-10 Gene Polymorphisms in patients with Inflammatory Bowel Disease. Immunol Invest 2017;46:714-29.

45. Aithal GP, Day CP, Leathart J, Daly AK, Hudson M. Association of single nucleotide polymorphisms in the interleukin-4 gene and interleukin-4 receptor gene with Crohn's disease in a British population. Genes Immun 2001;2:44-7.

46. Le Beau MM, Lemons RS, Espinosa R, Larson RA, Arai N, Rowley JD. Interleukin-4 and high levels of transforming growth factors and receptors are deleted in myeloid leukemia with del (5q). Blood 1989;73:647-50.

47. Rosenwasser LJ1, Klemm DJ, Dresback JK, Inamura H, Mascali JJ, Klinnert M et al. Promoter polymorphisms in the chromosome 5 gene cluster in asthma and atopy. Clin Exp Allergy 1995;25:74-8.

48. Kawashima T, Noguchi E, Arinami T, Yamakawa-Kobayashi K, Nakagawa H, Otsuka F et al. Linkage and association of an interleukin 4 gene polymorphism with atopic dermatitis in Japanese families. J Med Genet 1998;35:229-39.

49. Noguchi E, Shibasaki M, Arinami T, Takeda K, Yokouchi Y, Kawashima T et al. Association of asthma and the interleukin-4 promoter gene in Japanese. Clin Exp Allergy 1998;28:449-53.

50. Cantagrel A, Navaux F, Loubet-Lescoulié P, Nourhashemi F, Enault G, Abbal M et al. Interleukin-1β, Interleukin-1 receptor antagonist, Interleukin-4, and Interleukin-10 gene polymorphisms: relationship to occurrence and severity of rheumatoid arthritis. Arthritis Rheum 1999;42:1093-100.

51. Walley AJ, Cookson WOCM. Investigation of an interleukin-4 promoter polymorphisms for associations with asthma and atopy. J Med Genet 1996;33:689-92.

52. Takabayashi A, Ihara K, Sasaki Y, Kusuhara K, Nishima S, Hara T. Novel polymorphisms in the 5'untranslated region of the interleukin-4 gene. J Hum Genet 1999;44:352-3.

53. Shirakawa T, Deichmann KA, Izuhara K, Mao X-Q, Adra CN, Hopkin JM. Atopy and asthma: genetic variants of IL-4 and IL-13 signaling. Immunology Today 2000;21:60-4.

54. Ober C1, Leavitt SA, Tsalenko A, Howard TD, Hoki DM, Daniel R et al. Variation in the interleukin 4receptor  $\alpha$  gene confers susceptibility to asthma and atopy in ethnically diverse populations. Am J Hum Genet 2000;66:517-26.

55. Hugot JP, Laurent-Puig P, Gower-Rousseau C, Olson JM, Lee JC, Beaugerie L et al. Mapping of a susceptibility locus for Crohn's disease on chromosome 16. Nature 1996;379:821-3.

56. Rioux JD, Silverberg MS, Daly MJ, Steinhart AH, McLeod RS, Griffiths AM et al. Genomewide search in Canadian families with inflammatory bowel disease reveals two novel susceptibility loci. Am J Hum Genet 2000;66:1863-70.

57. Mombaerts P, Mizoguchi E, Grusby MJ, Glimcher LH, Bhan AK, Tonegawa S. Spontaneous development of inflammatory bowel disease in T cell receptor mutant mice. Cell 1993;75:274-82.

58. Mizoguchi A, Mizoguchi E, Chiba C, Splekermann GM, Tonegawa S, Nagler-Anderson C et al. Cytokine imbalance and autoantibody production in T cell receptor-α mutant mice with inflammatory bowel disease. J Exp Med 1996; 183:847-56. 59. Takahashi I, Kiyono H, Hamada S. CD4+ T-cell population mediates development of inflammatory bowel disease in T-cell receptor  $\alpha$  chain-deficient mice. Gastroenterology 1997;112:1876-86

60. Iijima H, Takahashi I, Kishi D, Kim JK, Kawano S, Hori M et al. Alteration of Interleukin 4 production results in the inhibition of T Helper Type 2 Cell- dominated Inflammatory Bowel Disease in T Cell Receptor  $\alpha$  Chain-deficient mice. J Exp Med 1999;190:607-15.

61. Mizoguchi A, Mizoguchi E, Bhan AK. The critical role of Interleukin 4 but not Interferon Gamma in the pathogenesis of Colitis in T-Cell receptor  $\alpha$  mutant mice. Gastroenterology 1999;116:320-6.

62. O'Garra A, Murphy K. Role of cytokines in determining T-lymphocyte function. Curr Opin Immunol 1994; 6(3):458-66

63. Fuss LJ, Neurath M, Boirivant M, Klein JS, De La Motte C, Strong SA et al. Disparate CD4+ lamina propria (LP) lymphokine secretion profiles in inflammatory bowel disease. J Immunol 1996; 157(3):1261-70.

64. Finnegan A, Mikecz K, Tao P, Glant TT. Proteoglycan (aggrecan)-induced arthritis in BALB/c mice is a Th1-type disease regulated by Th2 cytokines. J Immunol 1999; 163(10):5383-90.

65. Ramanathan S, de Kozak Y, Saoudi A, Goureau O, Van der Meide PH, Druet P et al. Recombinant IL-4 aggravates experimental autoimmune uveoretinitis in rats. J Immunol 1996; 157(5):2209-15.

66. Fort MM, Lesley R, Davidson NJ, Menon S, Brombacher F, Leach MW et al. IL-4 Exacerbates diasese in a Th1 Cell Transfer Model of Colitis. J Immunol 2001;166:2793-800.

67. Blumberg R, Saubermann LJ, Strober W. Animal models of mucosal inflammation and their relation to human inflammatory bowel disease. Curr Opin Immunol 1999; 11(6):648-56.

68. De Vries JE, Punnonen J. Interleukin-4 and interleukin-13. In: C. Snapper, editor. Cytokine regulation of humoral immunity. London: John Wiley & Sons, 1996.

69. De Waal Malefyth R . Role of interleukin-10, interleukin-4 and interleukin-13 in resolving inflammatory responses. In: Snyderman GA, editor. Inflammation: basic principles and clinical correlates. Lippincott-Williams & Wilkins, Philadelphia, 1999; pp 837-849.

70. Chomarat P, Banchereau J. Interleukin-4 and interleukin-13: their similarities and discrepancies. Int Rev Immunol 1998; 17(1-4):1-52.

71. Reincker H, Podolsky D. Human intestinal epithelial cells express functional cytokine receptors sharing the common γc chain of the interleukin 2 receptor. Proc Natl Acad Sci USA 1995; 92(18):8353-7.

72. Margolin K, Aronson FR, Sznol M, Atkins MB, Gucalp R, Fisher RI, et al. Phase II studies of recombinant interleukin-4 in advanced renal cancer and malignant melanoma. J Immunother 1994;15:147-53.

73. Desreumaux P, Brandt E, Gambiez L, Emilie D, Geboes K, Klein O et al. Distinct cytokine patterns in early and chronic ileal lesions of Crohn's disease. Gastroenterology 1997;113:118-26.

74. Stevceva L, Pavli P, Husband A, Ramsay A, Doe WF. Dextran sulphate sodium-induced colitis is ameliorated in interleukin 4 decificent mice. Genes Immun 2001;2:309-16.

75. Stevceva L, Pavli P, Buffinton G, Wozniak A, Doe WF. Dextran sodium sulphate-induced colitis activity varies with mouse strain but develops in lipopolysaccharide-unresponsive mice. J Gastroenterol Hepatol 1999;14:54-60.

76. Okayasu I, Hatakeyama S, Yamada M, Ohkusa T, Inagaki Y, Nakaya R. A novel method in the induction of reliable experimental acute and chronic ulcerative colitis in mice. Gastroenterology 1990; 98:694-702.

77. Yamada M, Ohkusa T, Okayasu I. Occurrence of dysplasia and adenocarcinoma after experimental chronic ulcerative colitis in hamsters induced by dextran sulphate sodium. Gut 1992;33:1521-7.

78. Tamaru T, Kobayashi H, Kishimoto S, Kajiyama G, Shimamoto F, Brown WR. Histochemical study of colonic cancer in experimental colitis of rats. Dig Dis Sci 1993;38:529-37.

79. Stevceva L, Buffinton GD, Wozniak A, Doe WF. Disease activity and histopathological changes in the chronic dextran sulphate sodium (DSS)-induced colitis resemble those in chronic human inflammatory bowel disease. J Gastroenterol Hepatol 1994;9:5-A:105.

80. Pushparaj PN, Li D, Komai-Koma M, Guabiraba R, Alexander J, McSharry C et al. Interleukin-33 exacerbates acute colitis via interleukin-4 in mice. Immunology 2013;140:70-7.

81. Schmitz J, Owyang A, Oldham E, Song Y, Murphy E, McClanahan TK et al. IL-33, an interleukin-1like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. Immunity 2005; 23:479-90. 82. Liew FY, Pitman NI, McInnes IB. Disease-associated functions of IL-33: the new kid in the IL-1 family. Nat Rev Immunol 2010;10:103-10.

83. Kurowska-Stolarska M1, Kewin P, Murphy G, Russo RC, Stolarski B, Garcia CC et al. IL-33 induces antigen-specific IL-5+ cells and promotes allergic-induced airway inflammation independent of IL-4. J Immunol 2008;181:4780-90.

84. Xu D, Chan WL, Leung BP, Wheeler R, Piedrafita D, Robinson IH et al. Selective expression of a stable cell surface molecule on type 2 but not type 1 helper T cells. J Exp Med 1998;187:787-94.

85. Stolarski B, Kurowska-Stolarska M, Kewin P, Xu D, Liew FY. IL-33 exacerbates eosinophil-mediated airway inflammation. J Immunol 2010;185:3472-80.

86. Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TK et al. Nuocyes represent a new innate effector leukocyte that mediates type-2 immunity. Nature 2010; 464:1367-70.

87. Komai-Koma M, Brombacher F, Pushparaj PN, Arendse B, McSharry C, Alexander J et al. Interleukin-33 amplifies IgE synthesis and triggers mast cell degranulation via interleukin-4 in naïve mice. Allergy 2012;67:1118-26.

88. Xu D, Jiang HR, Kewin P, Li Y, Mu R, Fraser AR et al. IL-33 exacerbates antigen-induced arthritis by activating mast cells. Proc Natl Acad Sci USA 2008;105:10913-8.

89. Verri WA Jr, Guerrero AT, Fukada SY, Valerio DA, Cunha TM, Xu D et al. IL-33 mediates antigeninduced cutaneous and articular hypernociception in mice. Proc Natl Acad Sci USA 2008;105:2723-8.

90. Boirivant M, Fuss IJ, Chu A, Strober W. Oxazolone colitis: a murine model of T helper cell type 2 colitis treatable with antibodies to interleukin 4. J Exp Med 1998;188:1929-39.

91. Mizoguchi A, Mizoguchi E, Chiba C, Bhan AK. Role of appendix in the development of inflammatory bowel disease in TCR-α mutant mice. J Exp Med 1996;184:707-15.

92. Mizoguchi E, Mizoguichi A, Bhan AK. Role of cytokines in the early stages of chronic colitis in TCR α-mutant mice. Lab Investig 1997;76:385-397.

93. Kasaian MT, Page KM, Fish S, Brennan A, Cook TA, Moreira K et al. Therapeutic activity of an interleukin-4/interleukin-13 dual antagonist on oxazolone-induced colitis in mice. Immunology 2014;143:416-27.

94. Kasaian MT, Miller DK. IL-13 as a therapeutic target for respiratory disease. Biochem Farmacol 2008;76:147-55

95. Inoue S, Matsumoto T, Iida M, Mizuno M, Kuroki F, Hoshika K et al. Characterization of cytokine expression in the rectal mucosa of ulcerative colitis: correlation with disease activity. Am J Gastroenterol 1999;94:2441-6.

96. Heller F, Fuss IJ, Nieuwenhuis EE, Blumberg RS, Strober W. Oxazolone colitis, a Th2 colitis model resembling ulcerative colitis, is mediated by IL-13 producing NK-T cells. Immunity 2002;17:629-38.

97. Heller F, Florian P, Bojarski C, Richter J, Christ M, Hillenbrand B et al. Interleukin-13 is the key effector Th2 cytokine in ulcerative colitis that affects epithelial tight junctions, apoptosis, and cell restitution. Gastroenterology 2005;129:550-64.

98. Olsen T, Cui G, Goll R, Husebekk A, Florholmen J. Infliximab therapy decreases the levels of TNF- $\alpha$  and IFN- $\gamma$  m RNA in colonic mucosa of ulcerative colitis. Scand J Gastroenterol 2009;44:727-35.

99. Berin MC, Yang PC, Ciok L, Waserman s, Perdue MH. Role for IL-4 in macromolecular transport across human intestinal epithelium. Am J Physiol 1999;276:1046-52.

100. Mannon PJ, Hornung RL, Yang Z, Yi C, Groden C, Friend J et al. Suppression of inflammation in ulcerative colitis by interferon- $\beta$ -1a is accompanied by inhibition of IL-13 production. Gut 2010;60:449-55.

101. Weber CR, Raleigh DR, Su L, Shen L, Sullivan EA, Wang Y et al. Epithelial myosin light chain kinase activation induces mucosal interleukin-13 expression to alter tight junction ion selectivity. J Biol Chem 2010;285:12037-46.

102. Prasad S, Mingrino R, Kaukinen K, Hayes KL, Powell RM, MacDonald TT et al. Inflammatory processes have differential effects on claudins 2,3 and 4 in colonic epithelial cells. Lab Invest 2005;85:1139-62.

103. Kasaian MT, Marquette K, Fish S, DeClercq C, Agostinelli R, Cook TA et al. An IL-4/IL-13 dual antagonist reduces lung inflammation, airway hyperresponsiveness, and IgE production in mice. Am J Respir Cell Mol Biol 2013;49:37-46.

Iijima H et al <sup>60</sup>	Anti-IL4 mAb treatment in TCR-alpha deficient mice
Mizoguchi A et al <sup>61</sup>	Double-mutant TCR- $\alpha$ -/- IL-4- mice and double mutant TCR- $\alpha$ -/- IFN- $\gamma$ - mice.
Fort MM et al <sup>66</sup>	IL-4 treatment in CD4(+)CD45RB(high) T cell transfer colitis
Stevceva L et al <sup>74</sup>	Dextran sulphate sodium induced colitis in IL4- and IL4+ mice
Pushparaj PN et al <sup>80</sup>	IL33 treatment in DSS induced colitis and DSS-induced colitis in ST2(-/-) BALB/c mice
Boirivant M et al <sup>90</sup>	Anti IL4, anti IL12 and anti TGF- $\beta$ treatment in oxazolone induced colitis
Kasaian MT <sup>93</sup>	IL-4/IL-13 antagonist treatment oxazolone induced colitis

Table 1