

ADOPTED: 21 November 2019 doi: 10.2903/j.efsa.2020.5935

## Pest categorisation of potato virus A (non-EU isolates)

EFSA Panel on Plant Health (PLH),

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## Abstract

Following a request from the EU Commission, the Panel on Plant Health has addressed the pest categorisation of non-EU isolates of potato virus A (PVA). The information currently available on geographical distribution, biology, epidemiology, potential entry pathways, potential additional impact over the current situation and availability of control measures of non-EU isolates of PVA has been evaluated with regard to the criteria to qualify as potential Union quarantine pest. Because non-EU isolates of PVA are absent from the EU, they do not meet one of the requirements to be regulated as a regulated non-quarantine pest (RNQP) (presence in the EU); as a consequence, the Panel decided not to evaluate the other RNQP criteria for these isolates. This categorisation was performed considering two groups of isolates: those reported in *Solanum betaceum* (PVA-TamMV, not reported from the EU) and all other isolates (hereafter referred to as PVA, worldwide distribution). Non-EU isolates of PVA and of PVA-TamMV do not meet one of the criteria evaluated by EFSA to be regarded as a potential Union quarantine pest, since they are not expected to have an additional impact in the EU.

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Keywords: European Union, pest risk, plant health, plant pest, PVA, PVA-TamMV, quarantine

Requestor: European Commission

Question number: EFSA-Q-2019-00506

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**Acknowledgements:** This opinion was prepared in cooperation with the National Plant Protection Organization, Netherlands Food and Consumer Product Safety Authority under the tasking grant (GP/ EFSA/ALPHA/2017/04).

**Suggested citation:** EFSA PLH Panel (EFSA Panel on Plant Health), Bragard C, Dehnen-Schmutz K, Gonthier P, Jacques M-A, Jaques Miret JA, Justesen AF, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, van der Werf W, Vicent Civera A, Yuen J, Zappalà L, Candresse T, Lacomme C, Bottex B, Oplaat C, Roenhorst A, Schenk M and Di Serio F, 2020. Scientific Opinion on the pest categorisation of potato virus A (non-EU isolates). EFSA Journal 2020;18 (1):5935, 33 pp. https://doi.org/10.2903/j.efsa.2020.5935

#### **ISSN:** 1831-4732

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## 1. Introduction

## **1.1.** Background and Terms of Reference as provided by the requestor

## 1.1.1. Background

Council Directive 2000/29/EC<sup>1</sup> on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community establishes the present European Union plant health regime. The Directive lays down the phytosanitary provisions and the control checks to be carried out at the place of origin on plants and plant products destined for the Union or to be moved within the Union. In the Directive's 2000/29/EC annexes, the list of harmful organisms (pests) whose introduction into or spread within the Union is prohibited, is detailed together with specific requirements for import or internal movement.

Following the evaluation of the plant health regime, the new basic plant health law, Regulation (EU) 2016/2031<sup>2</sup> on protective measures against pests of plants, was adopted on 26 October 2016 and will apply from 14 December 2019 onwards, repealing Directive 2000/29/EC. In line with the principles of the above mentioned legislation and the follow-up work of the secondary legislation for the listing of EU regulated pests, EFSA is requested to provide pest categorisations of the harmful organisms included in the annexes of Directive 2000/29/EC, in the cases where recent pest risk assessment/pest categorisation is not available.

#### **1.1.2.** Terms of reference

EFSA is requested, pursuant to Article 22(5.b) and Article 29(1) of Regulation (EC) No 178/2002<sup>3</sup>, to provide scientific opinion in the field of plant health.

EFSA is requested to prepare and deliver a pest categorisation (step 1 analysis) for each of the regulated pests included in the appendices of the annex to this mandate. The methodology and template of pest categorisation have already been developed in past mandates for the organisms listed in Annex II Part A Section II of Directive 2000/29/EC. The same methodology and outcome is expected for this work as well.

The list of the harmful organisms included in the annex to this mandate comprises 133 harmful organisms or groups. A pest categorisation is expected for these 133 pests or groups and the delivery of the work would be stepwise at regular intervals through the year as detailed below. First priority covers the harmful organisms included in Appendix 1, comprising pests from Annex II Part A Section I and Annex II Part B of Directive 2000/29/EC. The delivery of all pest categorisations for the pests included in Appendix 1 is June 2018. The second priority is the pests included in Appendix 2, comprising the group of *Cicadellidae* (non-EU) known to be vector of Pierce's disease (caused by *Xylella fastidiosa*), the group of *Tephritidae* (non-EU), the group of potato viruses and virus-like organisms, the group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L., and the group of *Margarodes* (non-EU species). The delivery of all pest categorisations for the pests included in Appendix 3 cover pests of Annex I part A section I and all pest categorisations should be delivered by end 2020.

For the above mentioned groups, each covering a large number of pests, the pest categorisation will be performed for the group and not the individual harmful organisms listed under "such as" notation in the Annexes of the Directive 2000/29/EC. The criteria to be taken particularly under consideration for these cases, is the analysis of host pest combination, investigation of pathways, the damages occurring and the relevant impact.

Finally, as indicated in the text above, all references to 'non-European' should be avoided and replaced by 'non-EU' and refer to all territories with exception of the Union territories as defined in Article 1 point 3 of Regulation (EU) 2016/2031.

<sup>&</sup>lt;sup>1</sup> Council Directive 2000/29/EC of 8 May 2000 on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community. OJ L 169/1, 10.7.2000, p. 1–112.

<sup>&</sup>lt;sup>2</sup> Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants. OJ L 317, 23.11.2016, p. 4–104.

<sup>&</sup>lt;sup>3</sup> Regulation (EC) No 178/2002 of the European Parliament and of the Council of 28 January 2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. OJ L 31/1, 1.2.2002, p. 1–24.



## 1.1.2.1. Terms of Reference: Appendix 1

List of harmful organisms for which pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

## Annex IIAI

## (a) Insects, mites and nematodes, at all stages of their development

Aleurocanthus spp. Anthonomus bisignifer (Schenkling) Anthonomus signatus (Say) Aschistonyx eppoi Inouye Carposina niponensis Walsingham Enarmonia packardi (Zeller) Enarmonia prunivora Walsh Grapholita inopinata Heinrich Hishomonus phycitis Leucaspis japonica Ckll. Listronotus bonariensis (Kuschel)

## (b) Bacteria

Citrus variegated chlorosis *Erwinia stewartii* (Smith) Dye

## (c) Fungi

*Alternaria alternata* (Fr.) Keissler (non-EU pathogenic isolates) *Anisogramma anomala* (Peck) E. Müller *Apiosporina morbosa* (Schwein.) v. Arx *Ceratocystis virescens* (Davidson) Moreau *Cercoseptoria pini-densiflorae* (Hori and Nambu) Deighton *Cercospora angolensis Carv. and Mendes* 

## (d) Virus and virus-like organisms

Beet curly top virus (non-EU isolates) Black raspberry latent virus Blight and blight-like Cadang-Cadang viroid Citrus tristeza virus (non-EU isolates) Leprosis

## Annex IIB

## (a) Insect mites and nematodes, at all stages of their development

Anthonomus grandis (Boh.) Cephalcia lariciphila (Klug) Dendroctonus micans Kugelan Gilphinia hercyniae (Hartig) Gonipterus scutellatus Gyll. Ips amitinus Eichhof Numonia pyrivorella (Matsumura) Oligonychus perditus Pritchard and Baker Pissodes spp. (non-EU) Scirtothrips aurantii Faure Scirtothrips citri (Moultex) Scolytidae spp. (non-EU) Scrobipalpopsis solanivora Povolny Tachypterellus quadrigibbus Say Toxoptera citricida Kirk. Unaspis citri Comstock

*Xanthomonas campestris* pv. *oryzae* (Ishiyama) Dye and pv. *oryzicola* (Fang. et al.) Dye

*Elsinoe* spp. Bitanc. and Jenk. Mendes *Fusarium oxysporum* f. sp. *albedinis* (Kilian and Maire) Gordon *Guignardia piricola* (Nosa) Yamamoto *Puccinia pittieriana* Hennings *Stegophora ulmea* (Schweinitz: Fries) Sydow & Sydow *Venturia nashicola* Tanaka and Yamamoto

Little cherry pathogen (non- EU isolates) Naturally spreading psorosis Palm lethal yellowing mycoplasm Satsuma dwarf virus Tatter leaf virus Witches' broom (MLO)

*Ips cembrae* Heer *Ips duplicatus* Sahlberg *Ips sexdentatus* Börner *Ips typographus* Heer *Sternochetus mangiferae* Fabricius



## (b) Bacteria

*Curtobacterium flaccumfaciens pv. flaccumfaciens* (Hedges) Collins and Jones

## (c) Fungi

*Glomerella gossypii* Edgerton *Gremmeniella abietina* (Lag.) Morelet

1.1.2.2. Terms of Reference: Appendix 2

List of harmful organisms for which pest categorisation is requested per group. The list below follows the categorisation included in the annexes of Directive 2000/29/EC.

## Annex IAI

## (a) Insects, mites and nematodes, at all stages of their development

Group of Cicadellidae (non-EU) known to be vector of Pierce's disease (caused by Xylella fastidiosa), such as:

- 1) Carneocephala fulgida Nottingham
- 2) Draeculacephala minerva Ball

Group of Tephritidae (non-EU) such as:

- 1) Anastrepha fraterculus (Wiedemann)
- 2) Anastrepha ludens (Loew)
- 3) Anastrepha obliqua Macquart
- 4) Anastrepha suspensa (Loew)
- 5) Dacus ciliatus Loew
- 6) Dacus curcurbitae Coquillet
- 7) Dacus dorsalis Hendel
- 8) Dacus tryoni (Froggatt)
- 9) Dacus tsuneonis Miyake
- 10) Dacus zonatus Saund.
- 11) Epochra canadensis (Loew)

## (c) Viruses and virus-like organisms

Group of potato viruses and virus-like organisms such as:

- 1) Andean potato latent virus
- 2) Andean potato mottle virus
- 3) Arracacha virus B, oca strain
- 4) Potato black ringspot virus

- 5) Potato virus T
- non-EU isolates of potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc) and Potato leafroll virus

Group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L., such as:

- 1) Blueberry leaf mottle virus
- 2) Cherry rasp leaf virus (American)
- 3) Peach mosaic virus (American)
- 4) Peach phony rickettsia
- 5) Peach rosette mosaic virus
- 6) Peach rosette mycoplasm
- 7) Peach X-disease mycoplasm

- 8) Peach yellows mycoplasm
- 9) Plum line pattern virus (American)
- 10) Raspberry leaf curl virus (American)
- 11) Strawberry witches' broom mycoplasma
- 12) Non-EU viruses and virus-like organisms of *Cydonia Mill., Fragaria L., Malus Mill., Prunus L., Pyrus L., Ribes L., Rubus L.* and *Vitis L.*

3) *Graphocephala atropunctata* (Signoret)

Hypoxylon mammatum (Wahl.) J. Miller

- 12) Pardalaspis cyanescens Bezzi
- 13) Pardalaspis quinaria Bezzi
- 14) Pterandrus rosa (Karsch)
- 15) Rhacochlaena japonica Ito
- 16) Rhagoletis completa Cresson
- 17) Rhagoletis fausta (Osten-Sacken)
- 18) Rhagoletis indifferens Curran
- 19) Rhagoletis mendax Curran
- 20) Rhagoletis pomonella Walsh

## 21) Rhagoletis suavis (Loew)



## Annex IIAI

## (a) Insects, mites and nematodes, at all stages of their development

Group of Margarodes (non-EU species) such as:

- 1) *Margarodes vitis* (Phillipi)
- 2) Margarodes vredendalensis de Klerk

## 1.1.2.3. Terms of Reference: Appendix 3

List of harmful organisms for which pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

#### Annex IAI

#### (a) Insects, mites and nematodes, at all stages of their development

Longidorus diadecturus Eveleigh and Allen Acleris spp. (non-EU) Amauromyza maculosa (Malloch) *Monochamus* spp. (non-EU) Anomala orientalis Waterhouse Myndus crudus Van Duzee Nacobbus aberrans (Thorne) Thorne and Allen Arrhenodes minutus Drury Choristoneura spp. (non-EU) Naupactus leucoloma Boheman Conotrachelus nenuphar (Herbst) *Premnotrypes* spp. (non-EU) Dendrolimus sibiricus Tschetverikov Pseudopityophthorus minutissimus (Zimmermann) Diabrotica barberi Smith and Lawrence Pseudopityophthorus pruinosus (Eichhoff) Diabrotica undecimpunctata howardi Barber Scaphoideus luteolus (Van Duzee) Diabrotica undecimpunctata undecimpunctata Spodoptera eridania (Cramer) Mannerheim Spodoptera frugiperda (Smith) Spodoptera litura (Fabricus) Diabrotica virgifera zeae Krysan & Smith Diaphorina citri Kuway Thrips palmi Karny Xiphinema americanum Cobb sensu lato (non-EU Heliothis zea (Boddie) populations) Hirschmanniella spp., other than Hirschmanniella gracilis (de Man) Luc and Goodey Xiphinema californicum Lamberti and Bleve-Zacheo Liriomyza sativae Blanchard

#### (b) Fungi

Ceratocystis fagacearum (Bretz) Hunt Chrysomyxa arctostaphyli Dietel Cronartium spp. (non-EU) Endocronartium spp. (non-EU) Guignardia laricina (Saw.) Yamamoto and Ito Gymnosporangium spp. (non-EU) Inonotus weirii (Murril) Kotlaba and Pouzar Melampsora farlowii (Arthur) Davis

#### (c) Viruses and virus-like organisms

Tobacco ringspot virus Tomato ringspot virus Bean golden mosaic virus Cowpea mild mottle virus Lettuce infectious yellows virus Mycosphaerella larici-leptolepis Ito et al. Mycosphaerella populorum G. E. Thompson Phoma andina Turkensteen Phyllosticta solitaria Ell. and Ev. Septoria lycopersici Speg. var. malagutii Ciccarone and Boerema Thecaphora solani Barrus Trechispora brinkmannii (Bresad.) Rogers

3) Margarodes prieskaensis Jakubski

Pepper mild tigré virus Squash leaf curl virus Euphorbia mosaic virus Florida tomato virus



## (d) Parasitic plants

Arceuthobium spp. (non-EU)

#### <u>Annex IAII</u>

## (a) Insects, mites and nematodes, at all stages of their development

*Meloidogyne fallax* Karssen *Popillia japonica* Newman Rhizoecus hibisci Kawai and Takagi

## (b) Bacteria

*Clavibacter michiganensis* (Smith) Davis et al. ssp. *Ralstonia solanacearum* (Smith) Yabuuchi et al. *sepedonicus* (Spieckermann and Kotthoff) Davis et al.

## (c) Fungi

Melampsora medusae Thümen

Synchytrium endobioticum (Schilbersky) Percival

## Annex I B

#### (a) Insects, mites and nematodes, at all stages of their development

Leptinotarsa decemlineata Say

Liriomyza bryoniae (Kaltenbach)

#### (b) Viruses and virus-like organisms

Beet necrotic yellow vein virus

## **1.2.** Interpretation of the Terms of Reference

European Food Safety Authority (EFSA) is asked to develop pest categorisations for non-European Union (EU) isolates of seven potato viruses, i.e. potato leafroll virus and potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc), which are defined by their geographical origin outside the EU. As such, isolates of these viruses occurring outside the EU territory are considered as non-EU isolates. Accordingly, a plant infected with one of these viruses originating in a non-EU country is considered to be infected with a non-EU isolate. All seven viruses are important pathogens of potato and, therefore, there is no uncertainty about the fact that non-EU isolates have an impact on potato crops in absolute terms. However, EU isolates of these viruses already have an impact in the EU; consequently, the Panel decided to evaluate whether the non-EU isolates would have an additional impact compared to the current situation, upon introduction and spread in the EU. This interpretation was agreed with the European Commission.

This scientific opinion presents the pest categorisation of non-EU isolates of potato virus A (PVA). Non-EU isolates of PVA are listed in the Appendices of the Terms of Reference (ToR) to be subject to pest categorisation to determine whether they fulfil the criteria of a quarantine pest for the area of the EU excluding Ceuta, Melilla and the outermost regions of Member States referred to in Article 355(1) of the Treaty on the Functioning of the European Union (TFEU), other than Madeira and the Azores.

Because non-EU isolates of PVA are absent from the EU, they do not meet one of the requirements to be regulated as an regulated non-quarantine pest (RNQP) (presence in the EU); as a consequence, the Panel decided not to evaluate the other RNQP criteria for these isolates.

The new Plant Health Regulation (EU) 2016/2031<sup>4</sup>, on the protective measures against pests of plants, will be applying from December 2019. The regulatory status sections (Section 3.3) of the present opinion are still based on Council Directive 2000/29/EC, as the document was adopted in November 2019.

<sup>&</sup>lt;sup>4</sup> Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants, amending Regulations (EU) 228/2013, (EU) 652/2014 and (EU) 1143/2014 of the European Parliament and of the Council and repealing Council Directives 69/464/EEC, 74/647/EEC, 93/85/EEC, 98/57/EC, 2000/29/EC, 2006/91/EC and 2007/33/EC. OJ L 317, 23.11.2016, pp. 4–104.

## 2. Data and methodologies

## 2.1. Data

#### 2.1.1. Literature search

A literature search on potato virus A (PVA) was conducted in the ISI Web of Science bibliographic database. The scientific name of the pest was used as search term. Relevant papers were reviewed with a focus on potential differences between isolates and strains. Further references and information were obtained from experts, as well as from citations in the reviewed papers and grey literature. The search was continued until no further information could be found or until the collected information was considered sufficient to perform the pest categorisation; consequently, the presented data are not necessarily exhaustive.

#### 2.1.2. Database search

Information on hosts, vectors and distribution at species level, was retrieved from CABI Crop Protection Compendium (CABI cpc) and relevant publications. Additional data on isolates distribution were obtained from the literature.

Data about the import of commodity types that could potentially provide a pathway for the pest to enter the EU and about the area of hosts grown in the EU were obtained from EUROSTAT (Statistical Office of the European Communities).

The Europhyt database was consulted to identify interceptions of non-EU isolates of PVAF. Europhyt is a web-based network run by the Directorate General for Health and Food Safety (DG SANTÉ) of the European Commission and is a subproject of PHYSAN (Phyto-Sanitary Controls) specifically concerned with plant health information. The Europhyt database manages notifications of interceptions of plants or plant products that do not comply with EU legislation, as well as notifications of plant pests detected in the territory of the Member States (MSs) and the phytosanitary measures taken to eradicate or avoid their spread.

#### 2.2. Methodologies

The Panel performed the pest categorisation for non-EU isolates of PVA, following the guiding principles and steps presented in the EFSA guidance on quantitative pest risk assessment (EFSA PLH Panel, 2018) and in the International Standard for Phytosanitary Measures No 11 (FAO, 2013) and No 21 (FAO, 2004).

General information on PVA will be provided at species level. Further information will be added at the level of strains, lineages and/or non-EU isolates when available and/or applicable.

This work was initiated following an evaluation of the EU plant health regime. Therefore, to facilitate the decision-making process, in the conclusions of the pest categorisation, the Panel addresses explicitly each criterion for a Union quarantine pest in accordance with Regulation (EU) 2016/2031 on protective measures against pests of plants, and includes additional information required in accordance with the specific ToR received by the European Commission. As explained in the interpretation of the ToR, the criterion on impact focuses on additional impact of non-EU isolates of PVA. For each conclusion, the Panel provides a short description of its associated uncertainty.

Table 1 presents the Regulation (EU) 2016/2031 pest categorisation criteria on which the Panel bases its conclusions. All relevant criteria have to be met for the pest to potentially qualify as a quarantine pest. If one of the criteria is not met, the pest will not qualify.

It should be noted that the Panel's conclusions are formulated respecting its remit and particularly with regard to the principle of separation between risk assessment and risk management (EFSA founding regulation (EU) No 178/2002); therefore, the Panel will present a summary of the reported impacts. Impacts are expressed in terms of yield and quality losses and not in monetary terms, whereas addressing social impacts is outside the remit of the Panel.



Table 1:	Pest categorisation criteria under evaluation, as defined in Regulation (EU) 2016/2031 on
	protective measures against pests of plants (the number of the relevant sections of the
	pest categorisation is shown in brackets in the first column)

Criterion of pest categorisation	Criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Criterion in Regulation (EU) 2016/2031 regarding protected zone quarantine pest (articles 32–35)	Criterion in Regulation (EU) 2016/2031 regarding Union regulated non-quarantine pest
Identity of the pest (Section 3.1)	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?	Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?
Absence/ presence of the pest in the EU territory (Section 3.2)	Is the pest present in the EU territory? If present, is the pest widely distributed within the EU? Describe the pest distribution briefly!	Is the pest present in the EU territory? If not, it cannot be a protected zone quarantine organism	Is the pest present in the EU territory? If not, it cannot be a RNQP. (A regulated non- quarantine pest must be present in the risk assessment area)
Regulatory status (Section 3.3)	If the pest is present in the EU but not widely distributed in the risk assessment area, it should be under official control or expected to be under official control in the near future	The protected zone system aligns with the pest-free area system under the International Plant Protection Convention (IPPC) The pest satisfies the IPPC definition of a quarantine pest that is not present in the risk assessment area (i.e. protected zone)	Is the pest regulated as a quarantine pest? If currently regulated as a quarantine pest, are there grounds to consider its status could be revoked?
Pest potential for entry, establishment and spread in the EU territory (Section 3.4)	Is the pest able to enter into, become established in, and spread within, the EU territory? If yes, briefly list the pathways!	Is the pest able to enter into, become established in, and spread within, the protected zone areas? Is entry by natural spread from EU areas where the pest is present possible?	Is spread mainly via specific plants for planting, rather than via natural spread or via movement of plant products or other objects? Clearly state if plants for planting is the main pathway!
Potential for consequences in the EU territory (Section 3.5)	Would the pests' introduction have an economic or environmental impact on the EU territory?	Would the pests' introduction have an economic or environmental impact on the protected zone areas?	Does the presence of the pest on plants for planting have an economic impact as regards the intended use of those plants for planting?
Available measures (Section 3.6)	Are there measures available to prevent the entry into, establishment within or spread of the pest within the EU such that the risk becomes mitigated?	Are there measures available to prevent the entry into, establishment within or spread of the pest within the protected zone areas such that the risk becomes mitigated? Is it possible to eradicate the pest in a restricted area within 24 months (or a period longer than 24 months where the biology of the organism so justifies) after the presence of the pest was confirmed in the protected zone?	Are there measures available to prevent pest presence on plants for planting such that the risk becomes mitigated?



Criterion of pest categorisation	Criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Criterion in Regulation (EU) 2016/2031 regarding protected zone quarantine pest (articles 32–35)	Criterion in Regulation (EU) 2016/2031 regarding Union regulated non-quarantine pest
Conclusion of pest categorisation (Section 4)	A statement as to whether (1) all criteria assessed by EFSA above for consideration as a potential quarantine pest were met and (2) if not, which one(s) were not met	A statement as to whether (1) all criteria assessed by EFSA above for consideration as potential protected zone quarantine pest were met, and (2) if not, which one(s) were not met	A statement as to whether (1) all criteria assessed by EFSA above for consideration as a potential RNQP were met, and (2) if not, which one(s) were not met

The Panel will not indicate in its conclusions of the pest categorisation whether to continue the risk assessment process, but following the agreed two-step approach, will continue only if requested by the risk managers. However, during the categorisation process, experts may identify key elements and knowledge gaps that could contribute significant uncertainty to a future assessment of risk. It would be useful to identify and highlight such gaps so that potential future requests can specifically target the major elements of uncertainty, perhaps suggesting specific scenarios to examine.

#### 2.3. Nomenclature

Virus nomenclature is reported using the latest release of the official classification by the International Committee on Taxonomy of Viruses (ICTV, Release 2018b.v1, https://talk.ictvonline.org/taxonomy/). Virus names are not italicised throughout this opinion, corresponding to ICTV instructions.

## 3. Pest categorisation

#### **3.1.** Identity and biology of the pest

#### 3.1.1. Identity and taxonomy

Is the identity of the pest established, or has it been shown to produce consistent symptoms and to be transmissible?

**Yes.** PVA is a well-known virus and the definition of `non-EU isolates', as used in the present opinion has been clarified (see Section 1.2).

Potato virus A (PVA) is a well-characterised virus in the genus *Potyvirus*, family *Potyviridae* (Adams et al., 2011). PVA has a single-stranded positive-sense RNA genome and complete and/or partial genomic sequences are available for a number of isolates.

#### **3.1.2.** Biology of the pest

PVA isolates, including PVA-TamMV, are not reported to be transmitted by pollen or true seeds (Bartels, 1971). They are transmitted by vegetative propagation (via tubers) and are expected to be transmitted mechanically since they have been shown to be readily transmitted under experimental conditions (Bartels, 1971; de Bokx, 1972).

In addition, PVA isolates, including PVA-TamMV, are reported to be transmitted by several aphid species (Hemiptera: Aphididae) including *Aphis fabae* (Scopoli), *Macrosiphum euphorbiae* (Thomas), *Metopolophium dirhodum* (Walker), *Myzus persicae* (Sulzer), *Sitobion avenae* (Fabricius) and *Rhopalosiphum padi* (Linnaeus) (Bartels, 1971; Mossop, 1977; Andrejeva et al., 1996; Fox et al., 2017; CABI, 2019). However, for some isolates, aphid transmission failed or is reported as inefficient (Rajamaki et al., 1998). Currently, a relationship between aphid transmission and phylogenetic grouping or geographic distribution has not been identified (Rajamaki et al., 1998).

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## 3.1.3. Intraspecific diversity

Viruses generally exist as quasispecies, which means that they accumulate as a cluster of closely related sequence variants in a single host (Andino and Domingo, 2015). This is likely due to competition among the genomic variants that are generated as a consequence of the error-prone viral replication (higher in RNA than in DNA viruses) and the ensuing selection of the most fit variants in a given environment (Domingo et al., 2012). This genetic variability may have consequences on the virus' biological properties (e.g. host range, transmissibility and pathogenicity) as well as on the reliability of detection methods, especially when they target variable genomic regions.

This pest categorisation focuses on taxonomic levels below the species level, i.e. on isolates, lineages, and strains, which are defined in this opinion as follows:

- **Isolate**: virus population as present in a plant;
- Lineage: group of isolates belonging to a distinct phylogenetic cluster;
- **Strain**: group of isolates sharing biological, molecular and/or serological properties (Garcia-Arenal et al., 2001).

ICTV does not address taxonomic levels below the species level and, therefore, the names of lineages and/or strains are based on reports in literature. In the past, the term 'strain' has also often been used as a synonym for 'isolate'. As a consequence of this inconsistent use of terminology, the literature is often unclear.

Studies showing an unambiguous relationship between specific virus genotypes (isolates/lineage/ strains) and biological properties are limited. Moreover, the interpretation of such data may be hampered because discrimination between strains based on biological data is not always supported by genomic data. Historically, strains have been distinguished for many viruses, including PVA, based on differences in reactions on a set of indicator plants. This differentiation became further established by serology, by using monoclonal antibodies specifically selected to discriminate between the earlier distinguished strains. However, with the advent of molecular techniques, it became apparent that the initial biological and/or serological strain differentiation was not always supported by phylogenetic analyses of isolates based on genomic data. Moreover, the discrimination between strains might be further complicated by the existence of recombinant isolates, hampering an unambiguous assignment of isolates to recognised strains. This implies that there is frequent uncertainty about the interpretation of (older) data on strain differentiation and on geographical distribution.

There have been several approaches to distinguish PVA strains; i.e. on the basis of reactions on indicator plants, serological properties and genomic data. PVA strains have been distinguished based on differential responses of potato cultivars and of *Nicandra physalodes* (Bartels, 1971; Valkonen et al., 1995). Phylogenetic sequence analysis, using the coding region of the CP gene separated two major lineages, PVA-I and PVA-II (Mortensen et al., 2010; He et al., 2014). PVA-I isolates are reported to occur both in and outside the EU, PVA-II isolates are only reported from the EU. Currently, there is no evidence for the existence of PVA-II isolates outside the EU. However, since this grouping is based only on sequence comparison, without a link to biological properties, PVA will not be categorised along the PVA-I/PVA-II lineages.

Furthermore, tamarillo mosaic virus (TamMV) has been reported from *Solanum betaceum* (Eagles et al., 1990; Andrejeva et al., 1996) and, based on sequence analysis, was later considered an isolate of PVA, hereafter referred to as PVA-TamMV (Rajamaki et al., 1998; Kekarainen et al., 1999; Mortensen et al., 2010). PVA-TamMV does not group within lineages PVA-I or PVA-II and, based on whole genome sequences, has been reported to be the most distant isolate of PVA (Kekarainen et al., 2002; He et al., 2014). Based on sequence comparisons, He et al. (2014) provided evidence for recombination between PVA isolates and PVA-TamMV within their CP gene. The directionality of this recombination event is not known, so it is not possible to know whether PVA-TamMV or PVA isolates is (are) the actual recombinant(s). Because of its distant phylogenetic position and the difference in natural host range, PVA-TamMV will be categorised separately (see Table 2).

Group of isolates	Acronym	Other information	Key references
Potato virus A	PVA	All PVA isolates, except	Rajamaki et al. (1998), Mortensen et al. (2010),
isolates		PVA-TamMV	He et al. (2014)
Tamarillo mosaic	PVA-TamMV	NCBI GenBank	Eagles et al. (1990), Andrejeva et al. (1996),
virus isolates		accession AJ131403	Rajamaki et al. (1998), Mortensen et al. (2010)

Table 2:	Categorised	virus and	isolate in	the i	present opinion
	Categoriseu	vii us ariu	isolate in		

## **3.1.4.** Detection and identification of the pest

Are detection and identification methods available for the pest?

**Yes.** Methods are available for detection and identification of PVA at species level, and therefore for the identification of non-EU isolates. Identification of PVA-TamMV would require partial genomic sequencing.

As mentioned in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), virus detection and identification is complicated by several recurrent uncertainties. ICTV lists species demarcation criteria, but it is not always clear whether these are met in diagnostic tests. Furthermore, in the absence or near absence of information on genetic variability, it is not possible to guarantee that a given test will detect all variants of a species. On the contrary, generic tests may detect closely related viruses in addition to the target species. This implies that the reliability of a test depends on its validation for the intended use. For initial screening, it is important to prevent false-negative results, which means that the following performance characteristics are most relevant: analytical sensitivity, inclusivity of analytical specificity (coverage of the intra-species variability) and selectivity (matrix effects). For identification, it is important to prevent false positives and, therefore, the possible occurrence of cross-reactions should be determined, i.e. the exclusivity of the analytical specificity (the resolution should be sufficient to discriminate between related species).

PVA is a well-known virus for which detection methods are available. Bioassays associated with ELISA and/or (real-time) RT-PCR are available for the detection and identification of PVA at the species level (Rajamaki et al., 1998; Spetz et al., 2003; Agindotan et al., 2007; Loebenstein and Gaba, 2012).

Isolates of PVA-TamMV can be detected using bioassays associated with ELISA (Rajamaki et al., 1998). They can be identified by partial genomic sequencing. Based on the two coat protein sequences available (NCBI GenBank accession AY995213 and X54804) (Eagles et al., 1990), additional molecular methods could be developed to identify PVA-TamMV, with uncertainty on their specificity (inclusivity and exclusivity).

#### **3.2.** Pest distribution

#### **3.2.1.** Pest distribution outside the EU

PVA occurs worldwide wherever potato is grown (CABI cpc, 2019). Isolates of PVA are reported in Asia, Africa, Europe, North America, South America and Oceania (Rajamaki et al., 1998; Spetz et al., 2003; Maoka et al., 2010; Robertson et al., 2011; Larbi et al., 2012; He et al., 2014; de Neergaard et al., 2014; Were et al., 2014; Priegnitz et al., 2018).

In the absence of specific surveys, there is only limited information on the geographical distribution of the PVA lineages. PVA-I isolates are reported worldwide, including the EU (He et al., 2014), whereas PVA-II isolates are only reported in the EU (Mortensen et al., 2010; He et al., 2014). PVA-TamMV isolates are only reported from New Zealand (Eagles et al., 1990, 1994). In addition, PVA has been reported from *Solanum betaceum* in Rwanda (Anastase et al., 2019). However, in the absence of sequence information, the identity of the involved isolate(s) remains uncertain.

## **3.2.2.** Pest distribution in the EU

Is the pest present in the EU territory? If present, is the pest widely distributed within the EU?

**Yes.** PVA is present in the EU.

**No.** PVA-TamMV is not known to be present in the EU.



As indicated in the previous section, PVA-I isolates are reported worldwide including several EU Member States (Finland, Germany, Hungary, the Netherlands and the United Kingdom) (Mortensen et al., 2010; He et al., 2014). PVA-II isolates are only reported in the EU (Mortensen et al., 2010).

PVA-TamMV isolates are not reported in the EU. However, this assessment is associated with uncertainties in the absence of specific surveys.

#### 3.3. Regulatory status

#### 3.3.1. Council Directive 2000/29/EC

Non-EU isolates of PVA are specifically listed in Council Directive 2000/29/EC and are regulated in Annex IAI (See Table 3).

Annex I, Part A	Harmful organisms whose introduction into, and spread within, all member states shall be banned	
Section I	ection I Harmful organisms not known to occur in any part of the community and releva the entire community	
(d)	Viruses and virus-like organisms	
2.	Potato viruses and virus-like organisms such as: (g) non-European isolates of potato viruses A, M, S, V, X and Y (including Y <sup>o</sup> , Y <sup>n</sup> and Y <sup>c</sup> ) and Potato leafroll virus	

Table 3: Non-EU isolates of PVA in Council Directive 2000/29/EC

#### 3.3.2. Legislation addressing potato

Table 4 reports on the articles in Council Directive 2000/29/EC which address potato or tuberforming species of *Solanum* L. PVA may also infect other hosts; references to the corresponding legislation are reported in Section 3.4.1.



## **Table 4:** Overview of the regulation in Annexes III, IV and V of Council Directive 2000/29/EC that applies to potato or tuber-forming Solanum species

Annex III, Part A	Plants, plant products and other object	s the introduction of which shall be prohibited in all Member States			
	Description	Country of origin			
10.	Tubers of <i>Solanum tuberosum</i> L., seed potatoes	Third countries other than Switzerland			
11.	Plants of stolon- or tuber-forming species of <i>Solanum</i> L. or their hybrids, intended for planting, other than those tubers of <i>Solanum tuberosum</i> L. as specified under Annex III A (10)	Third countries			
12.	Tubers of species of <i>Solanum</i> L., and their hybrids, other than those specified in points 10 and 11	Without prejudice to the special requirements applicable to the potato tubers listed in Annex IV, Part A Section I, third countries other than Algeria, Egypt, Israel, Libya, Morocco, Syria, Switzerland, Tunisia and Turkey, and other than European third countries which are either recognised as being free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., in accordance with the procedure referred to in Article 18(2), or in which provisions recognised as equivalent to the Community provisions on combating <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. in accordance with the procedure with the procedure referred to in Article 18(2), have been complied with			
Annex IV, Part A	Special requirements which shall be lai other objects into and within all Memb	d down by all member states for the introduction and movement of plants, plant products and er States			
Section I	Plants, plant products and other objects originating outside the Community				
	Plants, plant products and other objects	Special requirements			
25.1	Tubers of <i>Solanum tuberosum</i> L., originating in countries where <i>Synchytrium</i> <i>endobioticum</i> (Schilbersky) Percival is known to occur	<ul> <li>Without prejudice to the prohibitions applicable to the tubers listed in Annex III(A) (10), (11) and (12), official statement that:</li> <li>(a) the tubers originate in areas known to be free from <i>Synchytrium endobioticum</i> (Schilbersky) Percival (all races other than Race 1, the common European race), and no symptoms of <i>Synchytrium endobioticum</i> (Schilbersky) Percival have been observed either at the place of production or in its immediate vicinity since the beginning of an adequate period;</li> <li>or</li> <li>(b) provisions recognised as equivalent to the Community provisions on combating <i>Synchytrium endobioticum</i> (Schilbersky) Percival in accordance with the procedure referred to in Article 18(2) have been complied with, in the country of origin</li> </ul>			



Tubers of <i>Solanum tuberosum</i> L.	Without prejudice to the provisions listed in Annex (A) (10), (11) and (12) and Annex IV(A)(I) (25.1), official statement that:
	(a) the tubers originate in countries known to be free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al.; or
	(b) provisions recognised as equivalent to the Community provisions on combating <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al. in accordance with the procedure referred to in Article 18(2), have been complied with, in the country of origin
Tubers of <i>Solanum tuberosum</i> L., other than early potatoes, originating in countries where Potato spindle tuber viroid is known to occur	Without prejudice to the provisions applicable to the tubers listed in Annex III(A) (10), (11) and (12) and Annex $IV(A)(I)$ (25.1) and (25.2), suppression of the faculty of germination
Tubers of <i>Solanum tuberosum</i> L., intended for planting	Without prejudice to the provisions applicable to the tubers listed in Annex III(A)(10), (11) and (12) and Annex IV(A)(I) (25.1), (25.2) and (25.3), official statement that the tubers originate from a field known to be free from <i>Globodera rostochiensis</i> (Wollenweber) Behrens and <i>Globodera pallida</i> (Stone) Behrens and
	(aa) either, the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known not to occur; or
	(bb) in areas where <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known to occur, the tubers originate from a place of production found free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al., or considered to be free thereof, as a consequence of the implementation of an appropriate procedure aiming at eradicating <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. which shall be determined in accordance with the procedure referred to in Article 18(2)
	and (cc) either the tubers originate in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known not to occur; or
	<ul> <li>(dd) in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known to occur,</li> <li>— either the tubers originate from a place of production which has been found free from <i>Meloidogyne</i></li> </ul>
	<i>chitwoodi</i> Golden et al. (all populations), and <i>Meloidogyne fallax</i> Karssen based on an annual survey of host crops by visual inspection of host plants at appropriate times and by visual inspection both externally and by cutting of tubers after harvest from potato crops grown at the place of production, or
	— the tubers after harvest have been randomly sampled and, either checked for the presence of symptoms after an appropriate method to induce symptoms, or laboratory tested, as well as inspected visually both externally and by cutting the tubers, at appropriate times and in all cases at the time of closing of the packages or containers before marketing according to the provisions on closing in Council Directive 66/403/EEC of 14 June 1996 on the marketing of seed potatoes (1) and no symptoms of <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen have been found
	Tubers of Solanum tuberosum L., other than early potatoes, originating in countries where Potato spindle tuber viroid is known to occurTubers of Solanum tuberosum L., intended



25.4.1.	Tubers of <i>Solanum tuberosum</i> L., other than those intended for planting	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (12) and Annex IV(A)(I) (25.1), (25.2) and (25.3), official statement that the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is not known to occur
25.4.2.	Tubers of Solanum tuberosum L.	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (10), (11) and (12) and Annex IV(A)(I) (25.1), (25.2), (25.3), (25.4) and (25.4.1), official statement that: (a) the tubers originate in a country where <i>Scrobipalpopsis solanivora</i> Povolny is not known to occur; or (b) the tubers originate in an area free from <i>Scrobipalpopsis solanivora</i> Povolny, established by the national plant protection organisation in accordance with relevant International Standards for Phytosanitary Measures.
25.5.	Plants of Solanaceae, intended for planting, other than seeds, originating in countries where Potato stolbur mycoplasm is known to occur	Without prejudice to the provisions applicable to tubers listed in Annex III(A) (10), (11), (12) and (13), and Annex IV(A)(I) (25.1), (25.2), (25.3) and (25.4), official statement that no symptoms of Potato stolbur mycoplasm have been observed on the plants at the place of production since the beginning of the last complete cycle of vegetation
Section II	Plants, plant products and other object	ts originating in the Community
	Plants, plant products and other objects	Special requirements
18.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting	<ul> <li>Official statement that:</li> <li>(a) the Union provisions to combat <i>Synchytrium endobioticum</i> (Schilbersky) Percival have been complied with; and</li> <li>(b) either the tubers originate in an area known to be free from <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i></li> <li>(Spieckermann and Kotthoff) Davis et al. or the Union provisions to combat <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i></li> <li>(Spieckermann and Kotthoff) Davis et al. or the Union provisions to combat <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i></li> <li>(Spieckermann and Kotthoff) Davis et al. have been complied with; and</li> <li>(d) (aa) either, the tubers originate in areas in which <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al.is known not to occur; or</li> <li>(bb) in areas where <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. is known to occur, the tubers originate from a place of production found free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al., or considered to be free thereof, as a consequence of the implementation of an appropriate procedure aiming at eradicating <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al.;</li> </ul>
		and (e) either, the tubers originate in areas in which <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known not to occur, or in areas where <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen are known to occur: — either, the tubers originate from a place of production which has been found free from <i>Meloidogyne chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen based on an annual survey of host crops by visual inspection of host plants at appropriate times and by visual inspection both externally and by cutting of tubers after harvest from potato crops grown at the place of production, or — the tubers after harvest have been randomly sampled and, either checked for the presence of symptoms after an appropriate method to induce symptoms or laboratory tested, as well as inspected visually both externally and by cutting the tubers, at appropriate times and in all cases at the time of closing of the packages or containers before marketing according to the provisions on closing in Council Directive 66/403/EEC, and no symptoms of <i>Meloidogyne</i> <i>chitwoodi</i> Golden et al. (all populations) and <i>Meloidogyne fallax</i> Karssen have been found



18.1.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting, other than those to be planted in accordance with Article 4.4(b) of Council Directive 2007/33/EC	Without prejudice to the requirements applicable to the tubers of <i>Solanum tuberosum</i> L., intended for planting in Annex IV, Part A, Section II (18.1), official statement that the Union provisions to combat <i>Globodera pallida</i> (Stone) Behrens and <i>Globodera rostochiensis</i> (Wollenweber) Behrens are complied with
18.2	Tubers of <i>Solanum tuberosum</i> L., intended for planting, other than tubers of those varieties officially accepted in one or more Member States pursuant to Council Directive 70/457/EEC of 29 September 1970 on the common catalogue of varieties of agricultural plant species (1)	<ul> <li>Without prejudice to the special requirements applicable to the tubers listed in Annex IV(A)(II) (18.1), official statement that the tubers:</li> <li>belong to advanced selections such a statement being indicated in an appropriate way on the document accompanying the relevant tubers,</li> <li>have been produced within the Community, and</li> <li>have been derived in direct line from material which has been maintained under appropriate conditions and has been subjected within the Community to official quarantine testing in accordance with appropriate methods and has been found, in these tests, free from harmful organisms.</li> </ul>
18.3	Plants of stolon or tuber-forming species of <i>Solanum</i> L., or their hybrids, intended for planting, other than those tubers of <i>Solanum tuberosum</i> L. specified in Annex IV(A)(II) (18.1) or (18.2), and other than culture maintenance material being stored in gene banks or genetic stock collections	<ul> <li>(a) The plants shall have been held under quarantine conditions and shall have been found free of any harmful organisms in quarantine testing;</li> <li>(b) the quarantine testing referred to in (a) shall:</li> <li>(aa) be supervised by the official plant protection organisation of the Member State concerned and executed by scientifically trained staff of that organisation or of any officially approved body;</li> <li>(b) be executed at a site provided with appropriate facilities sufficient to contain harmful organisms and maintain the material including indicator plants in such a way as to eliminate any risk of spreading harmful organisms;</li> <li>(cc) be executed on each unit of the material;</li> <li>by visual examination at regular intervals during the full length of at least one vegetative cycle, having regard to the type of material and its stage of development during the testing programme, for symptoms caused by any harmful organisms,</li> <li>by testing, in accordance with appropriate methods to be submitted to the Committee referred to in Article 18:</li> <li>in the case of all potato material at least for:</li> <li>Andean potato latent virus,</li> <li>Arracacha virus B. oca strain,</li> <li>Potato black ringspot virus,</li> <li>Potato spindle tuber viroid,</li> <li>Potato virus T,</li> <li>Andean potato nottle virus,</li> <li><i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al.,</li> <li><i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al.,</li> <li>in the case of true seed potato of least for the viruses and viroid listed above;</li> <li>(d) by appropriate testing on any other symptom observed in the visual examination in order to identify the harmful organisms such a way esting in accordance with symptom observed in the visual examination in order to identify the harmful organisms.</li> </ul>



		<ul> <li>(c) any material, which has not been found free, under the testing specified under (b) from harmful organisms as specified under (b) shall be immediately destroyed or subjected to procedures which eliminate the harmful organism(s);</li> <li>(d) each organisation or research body holding this material shall inform their official Member State plant protection service of the material held</li> </ul>
18.3.1.	Seeds of <i>Solanum tuberosum</i> L., other than those specified in point 18.4.	Official statement that: The seeds derive from plants complying, as applicable, with the requirements set out in points 18.1., 18.1.1, 18.2 and 18.3; and (a) the seeds originate in areas known to be free from <i>Synchytrium endobioticum</i> (Schilbersky) Percival, <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., <i>Ralstonia solanacearum</i>
		(Smith) Yabuuchi et al. and Potato spindle tuber viroid; or
		<ul> <li>(b) the seeds comply with all of the following requirements:</li> <li>(i) they have been produced in a site where, since the beginning of the last cycle of vegetation, no symptoms of disease caused by the harmful organisms referred to in point (a) have been observed;</li> <li>(ii) they have been produced at a site where all of the following actions have been taken: separation of the site from other solanaceous plants and other host plants of Potato spindle tuber viroid; prevention of contact with staff and items, such as tools, machinery, vehicles, vessels and packaging material, from other sites producing solanaceous plants and other host plants of Potato spindle tuber viroid, or appropriate hygiene measures concerning staff or items from other sites producing solanaceous plants and other host plants of Potato spindle tuber viroid, on appropriate hygiene measures concerning staff or items from other sites producing solanaceous plants and other host plants of Potato spindle tuber viroid to prevent infection; only water free from all harmful organisms referred to in this point is used</li> </ul>
18.4	Plants of stolon, or tuber-forming species of <i>Solanum</i> L., or their hybrids, intended for planting, being stored in gene banks or genetic stock collections	Each organisation or research body holding such material shall inform their official Member State plant protection service of the material held
18.5.	Tubers of <i>Solanum tuberosum</i> L., other than those mentioned in Annex IV(A)(II) (18.1), (18.1.1), (18.2), (18.3) or (18.4)	There shall be evidence by a registration number put on the packaging, or in the case of loose-loaded potatoes transported in bulk, on the vehicle transporting the potatoes, that the potatoes have been grown by an officially registered producer, or originate from officially registered collective storage or dispatching centres located in the area of production, indicating that the tubers are free from <i>Ralstonia solanacearum</i> (Smith) Yabuuchi et al. and that (a) the Union provisions to combat <i>Synchytrium endobioticum</i> (Schilbersky) Percival, and
		(b) where appropriate, the Union provisions to combat <i>Clavibacter michiganensis</i> ssp. <i>sepedonicus</i> (Spieckermann and Kotthoff) Davis et al., and
		(c) the Union provisions to combat <i>Globodera pallida</i> (Stone) Behrens and <i>Globodera rostochiensis</i> (Wollenweber) Behrens are complied with



Annex IV, Part B	Special requirements which shall be laid down by all member states for the introduction and movement of plants, plant products and other objects into and within certain protected zones				
	Plants, plant products and other objects	Special requirements	Protected zone(s)		
20.1.	Tubers of <i>Solanum tuberosum</i> L., intended for planting	Without prejudice to the provisions applicable to the plants listed in Annex III(A) (10), (11), Annex IV(A)(I) (25.1), (25.2), (25.3), (25.4), (25.5), (25.6), Annex IV(A)(II) (18.1), (18.2), (18.3), (18.4), (18.6), official statement that the tubers: (a) were grown in an area where Beet necrotic yellow vein virus (BNYVV) is known not to occur; or (b) were grown on land, or in growing media consisting of soil that is known to be free from BNYVV, or officially tested by appropriate methods and found free from BNYVV; or	F (Britanny), FI, IRL, P (Azores), UK (Northern Ireland)		
		(c) have been washed free from soil			
20.2.	Tubers of <i>Solanum tuberosum</i> L., other than those mentioned in Annex IV(B) (20.1)	<ul> <li>(a) The consignment or lot shall not contain more than 1% by weight of soil, or</li> <li>(b) the tubers are intended for processing at premises with officially approved waste disposal facilities which ensures that there is no risk of spreading BNYVV</li> </ul>	F (Britanny), FI, IRL, P (Azores), UK (Northern Ireland)		
Annex V		er objects which must be subject to a plant health inspection (at the place of pro ed within the Community—in the country of origin or the consignor country, if or nitted to enter the Community			
Part A	Plants, plant products and othe	er objects originating in the Community			
Section I	Plants, plant products and othe which must be accompanied by	er objects which are potential carriers of harmful organisms of relevance for the y a plant passport	entire Community and		
1.3.	Plants of stolon- or tuber-forming s	species of Solanum L. or their hybrids, intended for planting.			
			tain protected zenes and		
		er objects which are potential carriers of harmful organisms of relevance for cert y a plant passport valid for the appropriate zone when introduced into or moved			
Section II	which must be accompanied by				
Section II 1.5.	which must be accompanied by	y a plant passport valid for the appropriate zone when introduced into or moved ant products and other objects listed in Part I			
	<ul><li>which must be accompanied by</li><li>Without prejudice to the plants, pla</li><li>Tubers of <i>Solanum tuberosum</i> L., ir</li></ul>	y a plant passport valid for the appropriate zone when introduced into or moved ant products and other objects listed in Part I	within that zone		
1.5.	<ul> <li>which must be accompanied by</li> <li>Without prejudice to the plants, pla</li> <li>Tubers of <i>Solanum tuberosum</i> L., in</li> <li>Plants, plant products and other</li> </ul>	y a plant passport valid for the appropriate zone when introduced into or moved ant products and other objects listed in Part I ntended for planting.	within that zone		

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## **3.3.3. Legislation addressing the organisms that vector PVA** (Directive/2000/29/EC)

PVA and PVA-TamMV are reported to be transmitted by aphid vectors (see Section 3.1.2) which are not subject to specific regulation.

## 3.4. Entry, establishment and spread in the EU

## 3.4.1. Host range

Table 5 provides information on reports of natural hosts of PVA and PVA-TamMV isolates including the associated uncertainties and regulation.

Due to the absence of specific surveys for PVA-TamMV, any isolate of the PVA-TamMV lineage in a host other than *S. betaceum* would likely have been identified as a PVA isolate. Therefore, it cannot be excluded that the natural host range of PVA-TamMV lineage is similar to that of PVA.

Table 5:	Natural hosts of PVA. Data regarding natural hosts was retrieved from the CABI cpc and
	literature up to 2 October 2019

Group of isolates	Hosts	Rationale and/or uncertainty	Regulation
PVA	<b>CABI cpc:</b> <i>Capsicum</i> spp., <i>Nicotiana</i> <i>tabacum, Solanum tuberosum</i> <b>Literature:</b> <i>Begonia, Celosia, Impatiens</i> (unknown species) (Pásztor et al., 2017), <i>S. nigrum</i> (Takács et al., 2018)	Additional natural hosts may exist	<b>Begonia L.:</b> IVB 24.3, VAII 2.1 <b>Capsicum sp.:</b> IVAI 16.6, 25.7, 36.3, IVAII 18.6.1, 18.7; VBI 1,3. <b>Nicotiana sp.:</b> IVAI 25.7; IVAII 18.7. <b>Solanum sp.:</b> IIIA 10,11,12; IVAI 25.1, 25.2, 25.3, 25.4, 25.4.1, 25.4.2, 25.5, 25.6, 25.7, 25.7.1,
PVA- TamMV	<i>Solanum betaceum</i> (Eagles et al., 1990)	Limited information. Additional natural hosts may exist Narrow experimental host range reported, including two <i>S.</i> <i>tuberosum</i> cultivars (Mossop, 1977; Rajamaki et al., 1998)	25.7.2, 28.1, 36.2, 45.3, 48; IVAI 18.1, 18.1.1, 18.2, 18.3, 18.3.1,

#### 3.4.2. Entry

Is the pest able to enter into the EU territory? If yes, identify and list the pathways.

**Yes.** Non-EU isolates of PVA, including PVA-TamMV, may enter the EU territory via plants for planting, i.e. seed potatoes (tubers) and/or microplants. Additional pathways include: ware potatoes (i.e. tubers intended for consumption or processing), plants for planting and fruits of other hosts, and viruliferous aphid vectors.

The following pathways can be considered for entry of non-EU isolates of PVA into the EU: potato plants for planting (seed potatoes, microplants), ware potatoes (i.e. tubers intended for consumption or processing), plants for planting and fruits of other natural hosts and viruliferous aphid vectors (see Table 6 for the major pathways).

PVA is transmitted by vegetative propagation and therefore seed potatoes and more generally, potato plants for planting, are considered the most important pathway for entry. The potential pathways for entry of non-EU isolates via seed potatoes of *Solanum tuberosum* and plants for planting of other tuber-forming *Solanum* species and their hybrids is addressed by the current EU legislation (Table 4; (EU) 2000/29 Annex IIIA, 10 and 11), which sets that import is not allowed from third countries except Switzerland. However, import of seed potatoes from Canada into Greece, Spain, Italy, Cyprus, Malta and Portugal is allowed by a derogation (2011/778/EU, 2014/368/EU, document C (2014) 3878). PVA is reported from Canada (Rajamaki et al., 1998) and, by definition, the PVA isolates present in this country are considered to be non-EU isolates. Therefore, the pathway of plants for planting is considered partially regulated for non-EU isolates of PVA. Potato is not reported as a natural host for PVA-TamMV and plants for planting of potato are therefore not considered a pathway. However, potato is known to be an experimental host of PVA-TamMV. PVA-TamMV isolates are not



reported in Canada and Switzerland. Should these isolates infect potato naturally, the pathway would be closed by legislation given their geographical distribution.

Entry of ware potatoes is addressed by the current EU legislation (Table 4, Annex IIIA, 12). Import of ware potatoes is prohibited from third countries other than Algeria, Egypt, Israel, Libya, Morocco, Syria, Switzerland, Tunisia and Turkey, and from European non-EU countries which do not meet a series of requirements addressing several other pathogens (see Table 4). As reported in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), the majority of the imported ware potatoes comes from Equpt and Israel (47 and 47.2%, respectively). Note that as long as ware potatoes are used for the intended use (consumption or processing), the ability of the non-EU isolates of PVA to establish is low. In addition, there are specific measures in place (Annex IV 25.3) for countries where potato spindle tuber viroid is known to occur (according to EPPO: Egypt, Israel and Turkey) aimed at mitigating the risk of establishment by suppression of the faculty of germination of ware potatoes, other than early potatoes, from these countries. PVA is, or is considered to be, present in these specified countries and, by definition, the PVA isolates present in these countries are considered non-EU isolates. They can in principle enter the EU via the ware potato pathway as there are no specific measures in place that mitigate the risk of entry. Therefore, the pathway of ware potatoes is considered partially regulated for non-EU isolates of PVA. Potato is not reported as a natural host for PVA-TamMV and ware potatoes are therefore not considered a pathway. However, potato is known to be an experimental host of PVA-TamMV. PVA-TamMV is not reported from countries subject to import derogations. Should PVA-TamMV isolates infect potato naturally, the pathway would be closed by legislation given their geographical distribution.

PVA has a limited number of natural hosts in addition to potato (see Section 3.4.1). Plants for planting of solanaceous hosts can be imported from European and Mediterranean countries; plants for planting of non-solanaceous hosts (*Celosia* and *Impatiens*) can be imported irrespective of the country of origin. In both cases, plants for planting provide an additional pathway of entry, although the magnitude of the trade is unclear. Overall, the pathway of plants for planting of other hosts than potato is considered partially regulated for non-EU isolates of PVA. The only known natural host of PVA-TamMV is *S. betaceum*, but it cannot be excluded that the natural host range of PVA-TamMV is comparable to that of PVA (see Section 3.4.1). Since PVA-TamMV is not reported from European and Mediterranean countries, the pathway of plants for planting of solanaceous hosts other than potato is considered possibly open for non-EU isolates of PVA-TamMV. This assessment is affected by uncertainties on trade and host range.

Viruliferous aphid vectors are a possible pathway of entry for non-EU isolates of PVA (see Section 3.1.3). Since the relevant aphid species are not subject to specific regulation, this pathway is open for non-EU isolates of PVA and PVA-TamMV. However, PVA is transmitted by aphids in a non-persistent manner, which implies that viruliferous aphids will lose the ability to transmit the virus within a short period. Therefore, this pathway is considered to be of minor importance and is not listed in Table 6.

Import of fruits can be an additional pathway for entry of non-EU isolates of PVA, including PVA-TamMV. However, the lack of seed transmission (see Section 3.1.3) reduces the relevance of this potential pathway. Aphid vectors can probe the infected fruits and acquire the virus for later transmission, as shown for other potyviruses such as papaya ringspot virus and zucchini yellow mosaic virus from melons, and plum pox virus from peaches (Lecoq et al., 2003; Gildow et al., 2004). Fruits of *Capsicum annuum* can be imported from a range of countries where PVA isolates have been reported. Overall, this pathway is considered to be open for non-EU isolates of PVA. The pathway is considered similarly open for non-EU isolates of PVA-TamMV. In both cases, there are uncertainties about the volume of trade. However, given the relatively unlikely series of events involved (aphids feeding on imported fruits followed by moving to susceptible plants) and the absence of seed transmission, this pathway is considered as minor and therefore not listed in Table 6.



Group of isolates	Potato plants for planting <sup>(1)</sup>	Ware potatoes <sup>(1)</sup>	Plants for planting of other hosts <sup>(1),(2)</sup>	Uncertainties
PVA	Pathway partially regulated: plants for planting of potato can be imported from Canada	Pathway partially regulated: import of ware potatoes is allowed from third countries subject to derogations	Pathway partially regulated: no import ban for some hosts	Geographic distribution Existence of other natural hosts
PVA- TamMV	Not a pathway: potato is not reported as a natural host. Should potato be a host, the pathway would be closed by legislation given the geographical distribution of these isolates.	Not a pathway: potato is not reported as a natural host. Should potato be a host, the pathway would be closed by legislation given the geographical distribution of these isolates	Pathway closed for solanaceous plants for planting: import is banned from New Zealand where PVA-TamMV is reported Pathway possibly open for non- solanaceous hosts: the existence of additional natural hosts not subject to import restrictions cannot be excluded	Geographic distribution Potato as natural host Existence of other natural hosts

**Table 6:** Identified major pathways for potential entry of non-EU isolates of PVA and the extent to which these pathways are addressed by current legislation

(1): 'Pathway open': no regulation or ban that prevents this pathway, 'Pathway closed' (as opposed to 'pathway open'): ban that prevents entry. 'Pathway possibly open': no direct evidence of the existence of the pathway (not closed by current legislation), but existence cannot be excluded based on comparisons with the biology of closely related viruses (in the same genus or family). 'Pathway regulated': regulations exist that limit the probability of entry along the pathway, but there is not a complete ban on imports. 'Pathway partially regulated': pathway consists of several sub-pathways, some are open, while others are closed (e.g. regulation for some hosts, but not for others; a ban exists for some non-EU MSs but not for all). 'Not a pathway': no evidence supporting the existence of the pathway

(2): Plants for planting, including seeds and pollen, of other hosts which are listed in Table 5.

Table 7 reports the only interception of PVA by EU member states during the period between 1995 and 8 August 2019. Only interceptions involving consignments imported from outside the EU were considered.

Table 7:	Interceptions of PVA by EU MSs on imported material from outside the EU. Data retrieved
	from the Europhyt database on 8 August 2019

Virus	Europhyt interception ID	Year of interception	Origin	Plant species on which it has been intercepted
PVA	109175	2017	Peru	Solanum tuberosum <sup>(1)</sup>

(1): Illegal import.

#### **3.4.3. Establishment**

Is the pest able to become established in the EU territory?

**Yes.** Non-EU isolates of PVA are likely to become established in the EU territory, as EU isolates and the main hosts are already present in the EU. This statement is associated with uncertainty for non-EU isolates of PVA-TamMV because their ability to naturally infect potato is not fully established and because of uncertainties about the presence of the only known natural host of PVA-TamMV (*S. betaceum*, tamarillo) in the EU.

#### 3.4.3.1. EU distribution of main host plants

Potato is widely grown in the EU, as reported in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020).

There is no evidence in Eurostat that *S. betaceum* (tamarillo) is commercially grown in the EU. Still, some cultivation might exist on a limited scale (commercial or non-commercial) in the frost-free southernmost part of the EU (e.g. citrus-growing areas). There is thus uncertainty about the presence of the only known natural host of PVA-TamMV in the EU.



#### 3.4.3.2. Climatic conditions affecting establishment

Except for those conditions affecting survival of the host plants, no eco-climatic constrains exist for the PVA isolates categorised here. Therefore, it is expected that these isolates are able to establish wherever their hosts may live. Potato is widely cultivated in the EU, and therefore, the Panel considers that climatic conditions will not impair the ability of the viruses addressed here to establish in the EU. However, it must be taken into consideration that virus impact, accumulation and distribution within natural hosts are dependent on environmental conditions. The same applies to expression of symptoms, vector populations and virus transmission being affected by climatic conditions.

#### 3.4.4. Spread

#### Is the pest able to spread within the EU territory following establishment?

**Yes.** Non-EU isolates of PVA, including PVA-TamMV, can spread via plants for planting, by mechanical transmission, and in addition, most of them can be spread by aphid vectors.

Non-EU isolates of PVA, including PVA-TamMV, can be transmitted by aphids (see Section 3.1.3), including *Myzus persicae* (Sulzer), which is widespread in and outside the EU (see Figure 1).

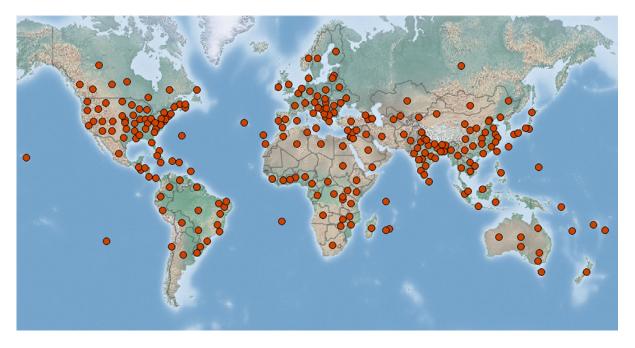


Figure 1: Global distribution map of *Myzus persicae* (Sulzer). Extracted from CABI cpc on 8 August 2019

#### 3.5. Impacts

Would the pests' introduction have an economic or environmental impact on the EU territory?

**No.** Non-EU isolates of PVA are not known to differ from PVA isolates already present in the EU and no additional impact is therefore expected on the EU territory.

Non-EU isolates of PVA-TamMV are not expected to have an additional impact in the EU territory, because the only known natural host (*S. betaceum*, tamarillo) is not grown on a significant scale in the EU. Should these isolates be able to infect potato, no additional impact is expected because under experimental conditions induced symptoms in *S. tuberosum* were identical to those of the potato-infecting PVA isolates already present in the EU.

As mentioned in the pest categorisation of non-EU viruses and viroids of potato (EFSA PLH Panel, 2020), symptoms caused by viruses are influenced by different factors, such as the isolate of the virus, the host and variety, and environmental conditions. A causal relation between a virus and reported



symptoms is not always clear, for example, in the case of mixed infections. Mixed infections are especially common in vegetative-propagated crops such as potato and the presence of additional viruses might increase or attenuate the observed symptoms. Therefore, reports on the symptomatology of individual viruses might not be conclusive, leading to uncertainties on the causal relation between a virus and the symptoms reported.

Infection of PVA in potatoes are usually mild, but yield losses up to 40% have been observed in extreme cases (Bartels, 1971). Limited biological data are available for most isolates of PVA. There is no evidence that PVA isolates present outside of the EU might have different biological properties than those already present in the EU. Therefore, non-EU isolates of PVA are not expected to have an additional impact over the current situation, with uncertainties.

There are no reports of PVA-TamMV naturally infecting potato. PVA-TamMV could infect some potato cultivars experimentally and induce symptoms identical to those of the other PVA isolates studied, although fewer potato cultivars could be infected by PVA-TamMV (Rajamaki et al., 1998). Therefore, additional impact of PVA-TamMV is not expected on potato.

PVA-TamMV is reported to infect *Solanum betaceum* (tamarillo), affecting both plant appearance and fruit quality (MacDiarmid, 1994). There is no information in Eurostat regarding the commercial production in the EU of this plant. However, the Panel takes note (of the fact) that in part of the EU the climate might be suitable to grow *S. betaceum*. It is nevertheless unclear whether *S. betaceum* is grown on any significant scale in the EU, and additional impact is therefore not expected, with uncertainty. Further uncertainty stems from the lack of information on the ability of isolates of PVA present in the EU to infect *S. betaceum*.

## **3.6.** Availability and limits of mitigation measures

Are there measures available to prevent the entry into, establishment within or spread of the pest within the EU such that the risk becomes mitigated?

**Yes.** See section 3.3 for measures already implemented in the current legislation. Additional measures could be implemented to further regulate the identified pathways or to limit entry, establishment or spread of non-EU isolates of PVA.

#### **3.6.1.** Identification of additional measures

Phytosanitary measures are currently applied to potato and other hosts (see Sections 3.3 and 3.4.1). Potential additional measures to mitigate the risk of entry of the isolates categorised in this opinion may include:

- Repel import derogations for potato plants for planting;
- Set specific phytosanitary requirements addressing the isolates categorised in this opinion for imported seed potatoes and/or ware potatoes;
- Extension of phytosanitary measures to specifically include hosts other than potato;
- Banning import of non-potato hosts plants for planting from countries where PVA is present;
- Extension of certification schemes and testing requirements to non-solanaceous natural hosts;
- Extension of plant passport requirements to specifically include hosts other than stolon- and tuber-forming *Solanum* species.

In addition, non-EU isolates of PVA, including PVA-TamMV, may enter in the EU through viruliferous aphids. Measures against aphids may include chemical treatment of consignments identified as potential entry pathways.

## **3.6.1.1. Additional control measures**

Table 8 reports on the potential additional control measures to reduce the likelihood of entry, establishment and/or spread of the categorised non-EU isolates of PVA. The additional control measures are selected form a longer list reported in EFSA PLH Panel (2018). Control measures are measures that have a direct effect on pest abundance.



Information sheet <u>(with</u> <u>hyperlink to</u> <u>information</u> <u>sheet if</u> <u>available)</u>	Control measure summary	Risk component	Rationale	
Growing plants in isolation	Description of possible exclusion conditions that could be implemented to isolate the crop from pests and if applicable relevant vectors. E.g. a dedicated structure such as glass or plastic greenhouses	Spread	Growing plants in insect proof greenhouses may prevent infestation by viruliferous aphid vectors. This measure would not be applicable for potato, with the exception of early stages of seed potato production Production of seed potatoes in areas with low aphid pressure (e.g. high altitude) would minimise the risk of infestation	
Chemical treatments on consignments or during processing	Use of chemical compounds that may be applied to plants or to plant products after harvest, during process or packaging operations and storage. The treatments addressed in this information sheet are: a) fumigation; b) spraying/dipping pesticides; c) surface disinfectants; d) process additives; e) protective compounds	Entry	a), b) and c) could remove viruliferous aphid vectors PVA is transmitted by aphids in a non-persistent way, which implies that viruliferous aphids will lose the ability to transmit the virus within a short period Therefore, the additional effect on preventing entry is minimal	
Cleaning and disinfection of facilities, tools and machinery	The physical and chemical cleaning and disinfection of facilities, tools, machinery, transport means, facilities and other accessories (e.g. boxes, pots, pallets, palox, supports, hand tools). The measures addressed in this information sheet are: washing, sweeping and fumigation	Spread	Cleaning tools may limit the spread via mechanical transmission	
Roguing and pruning	Roguing is defined as the removal of infested plants and/or uninfested host plants in a delimited area, whereas pruning is defined as the removal of infested plant parts only, without affecting the viability of the plant	Establishment and spread	Roguing of infested plants is efficient, in particular to prevent spread of PVA via contact. Pruning is not effective to remove a virus from infected plants	
Crop rotation, associations and density, weed/ volunteer control	Crop rotation, associations and density, weed/ volunteer control are used to prevent problems related to pests and are usually applied in various combinations to make the habitat less favourable for pests The measures deal with (1) allocation of crops to field (over time and space) (multi-crop, diversity cropping) and (2) to control weeds and volunteers as hosts of pests/vectors	Spread and impact	Viruses are maintained by vegetative propagation and, therefore, control of volunteers is important. Control of weed hosts may be of relevance	
Use of resistant and tolerant plant species/varieties	Resistant plants are used to restrict the growth and development of a specified pest and/or the damage they cause when compared to susceptible plant varieties under similar environmental conditions and pest pressure It is important to distinguish resistant from tolerant species/varieties	Spread and impact	Resistant and tolerant cultivars are available and could be used	

Table 8:	Selected additional control measures to consider to reduce the likelihood of pest entry,
	establishment and/or spread of non-EU isolates of PVA



Information sheet <u>(with</u> hyperlink to information sheet if available)	Control measure summary	Risk component	Rationale
Timing of planting and harvesting	The objective is to produce phenological asynchrony in pest/crop interactions by acting on or benefiting from specific cropping factors such as: cultivars, climatic conditions, timing of the sowing or planting and level of maturity/ age of the plant seasonal timing of planting and harvesting	Spread and impact	Relevant to prevent transmission by aphid vectors
Chemical treatments on crops including reproductive material	Chemical treatments on crops may prevent infestations by vectors and seed transmission	Spread and impact	Desiccation/removal of the foliage reduces the risk of transmission via aphid vectors and may prevent transport to the tubers of infected plants
Post-entry quarantine and other restrictions of movement in the importing country	This information sheet covers post-entry quarantine of relevant commodities; temporal, spatial and end-use restrictions in the importing country for import of relevant commodities; prohibition of import of relevant commodities into the domestic country Relevant commodities are plants, plant parts and other materials that may carry pests, either as infection, infestation or contamination	Entry and spread	Identifying virus-infected plants and banning their movement limit the risks of entry and spread in the EU

#### 3.6.1.2. Additional supporting measures

Table 9 reports on the possible additional supporting measures which are selected from the list reported in EFSA PLH Panel (2018). Supporting measures are organisational measures or procedures supporting the choice of appropriate risk reduction options that do not directly affect pest abundance.

**Table 9:**Selected supporting measures in relation to currently unregulated hosts and pathways.Supporting measures are organisational measures or procedures supporting the choice of<br/>appropriate risk reduction options that do not directly affect pest abundance

Information sheet title <u>(with</u> <u>hyperlink to</u> <u>information</u> <u>sheet if</u> <u>available</u> )	Supporting measure summary	Risk component	Comments
Inspection and trapping	Inspection is defined as the official visual examination of plants, plant products or other regulated articles to determine if pests are present or to determine compliance with phytosanitary regulations (ISPM 5) The effectiveness of sampling and subsequent inspection to detect pests may be enhanced by including trapping and luring techniques	Entry and spread	Visual inspection may detect potentially infected material Only applicable when visible symptoms on leaves and/or propagating tissues occur, which is dependent on the isolate, host/cultivar and environmental conditions
Laboratory testing	Examination, other than visual, to determine if pests are present using official diagnostic protocols. Diagnostic protocols describe the minimum requirements for reliable diagnosis of regulated pests	Entry and spread	Laboratory testing may detect/identify non-EU isolates of PVA on sampled material



Information sheet title <u>(with</u> <u>hyperlink to</u> <u>information</u> <u>sheet if</u> <u>available)</u>	Supporting measure summary	Risk component	Comments
Certified and approved premises	Mandatory/voluntary certification/approval of premises is a process including a set of procedures and of actions implemented by producers, conditioners and traders contributing to ensure the phytosanitary compliance of consignments. It can be a part of a larger system maintained by a National Plant Protection Organization in order to guarantee the fulfilment of plant health requirements of plants and plant products intended for trade. Key property of certified or approved premises is the traceability of activities and tasks (and their components) inherent the pursued phytosanitary objective. Traceability aims to provide access to all trustful pieces of information that may help to prove the compliance of consignments with phytosanitary requirements of importing countries	Entry and spread	Certified and approved premises may guarantee the absence of the harmful viruses imported for research and/or breeding purposes
Delimitation of Buffer zones	ISPM 5 defines a buffer zone as 'an area surrounding or adjacent to an area officially delimited for phytosanitary purposes in order to minimize the probability of spread of the target pest into or out of the delimited area, and subject to phytosanitary or other control measures, if appropriate' (ISPM 5). The objectives for delimiting a buffer zone can be to prevent spread from the outbreak area and to maintain a pest-free production place, site or area	Spread	Buffer zones may contribute to reduce the spread of non- EU isolates of PVA after entry in the EU
Sampling	According to ISPM 31, it is usually not feasible to inspect entire consignments, so phytosanitary inspection is performed mainly on samples obtained from a consignment. It is noted that the sampling concepts presented in this standard may also apply to other phytosanitary procedures, notably selection of units for testing For inspection, testing and/or surveillance purposes the sample may be taken according to a statistically based or a non-statistical sampling methodology	Spread	
Phytosanitary certificate and plant passport	An official paper document or its official electronic equivalent, consistent with the model certificates of the IPPC, attesting that a consignment meets phytosanitary import requirements (ISPM 5) a) export certificate (import) b) plant passport (EU internal trade)	Entry and spread	
Certification of reproductive material (voluntary/official)	Certification of reproductive material when not already implemented would contribute to reduce the risk associated with spread	Spread	



Information sheet title <u>(with</u> hyperlink to information sheet if available)	Supporting measure summary	Risk component	Comments
Surveillance	Official surveillance may contribute to early detection of non-EU isolates of PVA, favouring immediate adoption of control measures if they come to establish	Spread	

# **3.6.1.3.** Biological or technical factors limiting the effectiveness of measures to prevent the entry, establishment and spread of the pest

- Symptomless infections for some of the non-EU isolates of PVA in some hosts
- Uneven virus distribution or low concentrations limiting the reliability of the detection
- Absence of a validated diagnostic protocol allowing the identification of PVA-TamMV isolates

## 3.7. Uncertainty

The Panel identified the following knowledge gaps and uncertainties:

#### **Identity and biology**

- Lack of biological data, i.e. on host range, transmission and pathogenicity.
- Uncertainty on the existence of other non-EU isolates of PVA that have not yet been identified and might have additional impact on the EU territory.

#### Pest distribution

• Uncertainty on the geographical distribution and prevalence of PVA and PVA-TamMV because of the absence of systematic surveys.

## **Regulatory status**

• The concept of 'non-EU isolates' leaves some room for interpretation, which may create confusion or difficulties when enforcing the legislation (see Section 1.2).

#### Entry, establishment and spread in the EU (host range, entry, establishment, spread)

- Uncertainty on the host range of the categorised groups of isolates of PVA, particularly in the case of PVA-TamMV.
- Uncertainty on the cultivation of *Solanum betaceum* in the EU.
- Uncertainty on the trade volumes of some commodities that constitute pathways of entry.

#### Impact

• Uncertainty on the impact of non-EU isolates and whether this impact would exceed that of the isolates already present in the EU.

## 4. Conclusions

The information currently available on geographical distribution, biology, epidemiology, potential additional impact over the present situation and potential entry pathways of non-EU isolates of potato virus A (PVA) has been evaluated with regard to the criteria to qualify as potential Union quarantine pest. The conclusions of the Panel are summarised in Table 10.

Non-EU isolates of PVA and PVA-TamMV do not meet one of the criteria evaluated by EFSA to be regarded as a potential Union quarantine pest, since they are not expected to have an additional impact in the EU.

The Panel wishes to stress that these conclusions are associated with uncertainties because of limited information on distribution, biology and impact of PVA and PVA-TamMV isolates. Furthermore, other potentially harmful non-EU isolates of PVA might exist that are currently unknown.



**Table 10:**The Panel's conclusions on the pest categorisation criteria defined in Regulation (EU)<br/>2016/2031 on protective measures against pests of plants (the number of the relevant<br/>sections of the pest categorisation is shown in brackets in the first column) for non-EU<br/>isolates of PVA

Criterion of pest categorisation	Panel's conclusions against criterion in Regulation (EU) 2016/2031 regarding Union quarantine pest	Key uncertainties
Identity of the pest (Section 3.1)	The identity of PVA is well established Methods are available for detection and identification of non-EU isolates of PVA and PVA-TamMV	Uncharacterised PVA isolates may exist
Absence/presence of the pest in the EU territory (Section 3.2)	PVA is present in several EU MSs PVA-TamMV is not reported in the EU	Unreported presence of PVA- TamMV isolates in the EU
Regulatory status (Section 3.3)	Non-EU isolates of PVA are currently regulated in Annex IAI	Interpretation of the concept of 'non-EU isolate'
Pest potential for entry, establishment and spread in the EU territory (Section 3.4)	Non-EU isolates of PVA, including PVA-TamMV are able to enter into the EU The pathways of plants for planting of potato and ware potatoes are partially regulated for non-EU isolates of PVA and are not pathways for non-EU isolates of PVA-TamMV, since potato has not been reported as a natural host. Should potato be a host of PVA-TamMV, the pathways would be closed by legislation given the geographical distribution of these isolates For plants for planting of other hosts, the pathway is partially regulated for non-EU isolates of PVA. For non-EU isolates of PVA-TamMV, this pathway is closed for solanaceous hosts and possibly open for non-solanaceous	<ul> <li>Geographical distributions</li> <li>Existence of other natural hosts, in particular for PVA-TamMV</li> <li>Cultivation of <i>Solanum betaceum</i> in the EU</li> <li>Existence and relevance of trade of plants for planting of non-solanaceous hosts</li> </ul>
	hosts For non-EU isolates of PVA and PVA-TamMV, the minor pathways of viruliferous aphids and fruits of host species are open If non-EU isolates of PVA and PVA-TamMV were to enter the EU territory, they could become established and spread	
Potential for consequences in the EU territory (Section 3.5)	There is no indication that non-EU isolates of PVA differ biologically from PVA isolates already present in the EU and, therefore, they are not expected to have an additional impact Non-EU isolates of PVA-TamMV are also not expected to have an additional impact because the only known natural host ( <i>S. betaceum</i> , tamarillo) is not significantly grown in	Uncertainty on the impact of non-EU isolates of PVA and PVA-TamMV
	the EU and, should these isolates be able to infect potato, no additional impact is expected	
Available measures (Section 3.6)	Phytosanitary measures are available to reduce the likelihood of entry and spread of non-EU isolates of PVA and PVA-TamMV in the EU	No uncertainty
Conclusion on pest categorisation (Section 4)	Non-EU isolates of PVA and PVA-TamMV do not meet one of the criteria evaluated by EFSA to be regarded as a potential Union quarantine pest, since they are not expected to have an additional impact in the EU	
Aspects of assessment to focus on/scenarios to address in future if appropriate	<ul> <li>The main knowledge gaps or uncertainties identified concer</li> <li>Lack of information on the biology of the categorised grange, distribution and pathogenicity)</li> <li>Uncertainty on the cultivation of <i>Solanum betaceum</i> in t</li> <li>Volume of trade and countries of origin of plants for plant</li> <li>Uncertainty on the impact of non-EU isolates of PVA and</li> </ul>	oups of isolates (e.g. host he EU nting of non-potato hosts



## References

- Adams MJ, Zerbini FM, French R, Rabenstein F, Stenger DC and Valkonen JPT, 2011. Potyviridae. In: King AMQ, Adams MJ, Carstens EB and Lefkowitz EJ (eds.). Virus Taxonomy, 9th Report of the International Committee on Taxonomy of Viruses, 1069–1089.
- Agindotan BO, Shiel PJ and Berger PH, 2007. Simultaneous detection of potato viruses, PLRV, PVA, PVX and PVY from dormant potato tubers by TaqMan real-time RT-PCR. Journal of Virological Methods, 142, 1–9.
- Anastase H, Anatole N, Judith U and Jeanne U, 2019. Potyvirus infection analyses in Tamarillo trees in the northern province of Rwanda. African Journal of Food Science and Technology, 10, 38–41.
- Andino R and Domingo E, 2015. Viral quasispecies. Virology, 479–480, 46–51.
- Andrejeva J, Merits A, Rabenstein F, Puurand U and Jarvekulg L, 1996. Comparison of the nucleotide sequences of the 3'-terminal regions of one aphid and two non-aphid transmissible isolates of potato A potyvirus. Archives of Virology, 141, 1207–1219.
- Bartels R, 1971. DPV 54. Potato virus A. Viruses of Plants. Descriptions and Lists From the VIDE Database. CAB International, Wallingford, UK. http://www.dpvweb.net/dpv/showdpv.php?dpvno=054
- Bokx de JA, 1972. Viruses of potato and seed-potato production.
- CABI cpc, 2019. Datasheet of potato virus A (PVA). Available online: https://www-cabi-org/cpc/datasheet/42524 [Accessed September 17, 2019].
- Domingo E, Sheldon J and Perales C, 2012. Viral quasispecies evolution. Microbiology and Molecular Biology Reviews, 76, 159–216.
- Eagles RM, Gardner RC and Forster RL, 1990. Nucleotide sequence of the tamarillo mosaic virus coat protein gene. Nucleic Acids Research, 18, 7166.
- Eagles RM, Gardner RC and Forster RLS, 1994. Incidence and distribution of six viruses infecting tamarillo (Cyphomandra betacea) in New Zealand. New Zealand Journal of Crop and Horticultural Science, 22, 453–458.
- EFSA PLH Panel (EFSA Panel on Plant Health), 2018. Guidance on quantitative pest risk assessment. EFSA Journal 2018;16(8):5350, 86 pp. https://doi.org/10.2903/j.efsa.2018.5350
- EFSA PLH Panel (EFSA Panel on Plant Health), 2020. Pest categorisation of non-EU viruses and viroids of potato. EFSA Journal 2020;18(1):5853, 134 pp. https://doi.org/10.2903/j.efsa.2020.5853
- FAO (Food and Agriculture Organization of the United Nations), 1995. ISPM (International standards for phytosanitary measures) No 4. Requirements for the establishment of pest free areas. Available online: https://www.ippc.int/en/publications/614/
- FAO (Food and Agriculture Organization of the United Nations), 2004. ISPM (International Standards for Phytosanitary Measures) 21—Pest risk analysis of regulated non-quarantine pests. FAO, Rome, 30 pp. Available online: https://www.ippc.int/sites/default/files/documents//1323945746\_ISPM\_21\_2004\_En\_2011-11-29\_Refor. pdf
- FAO (Food and Agriculture Organization of the United Nations), 2013.ISPM (International Standards for Phytosanitary Measures) 11—Pest risk analysis for quarantine pests. FAO, Rome, 36 pp. Available online: https://www.ippc.int/sites/default/files/documents/20140512/ispm\_11\_2013\_en\_2014-04-30\_201405121523-494.65%20KB.pdf
- FAO (Food and Agriculture Organization of the United Nations), 2017. ISPM (International standards for phytosanitary measures) No 5. Glossary of phytosanitary terms. Available online: https://www.ippc.int/en/ publications/622/
- Fox A, Collins LE, Macarthur R, Blackburn LF and Northing P, 2017. New aphid vectors and efficiency of transmission of potato virus A and strains of potato virus Y in the UK. Plant Pathology, 66, 325–335.
- Garcia-Arenal F, Fraile A and Malpica JM, 2001. Variability and genetic structure of plant virus populations. Annual Review of Phytopathology, 39, 157–186.
- Gildow F, Damsteegt V, Stone A, Schneider W, Luster D and Levy L, 2004. Plum pox in North America: identification of aphid vectors and a potential role for fruit in virus spread. Phytopathology, 94, 868–874.
- He C, Zhang W, Hu X, Singh M, Xiong X and Nie X, 2014. Molecular characterization of a Chinese isolate of potato virus A (PVA) and evidence of a genome recombination event between PVA variants at the 3'-proximal end of the genome. Archives of Virology, 159, 2457–2462.
- Kekarainen T, Merits A, Oruetxebarria I, Rajamäki M-L and Valkonen JPT, 1999. Comparison of the complete sequences of five different isolates of potato virus A (PVA), genus *Potyvirus*. Archives of Virology, 144, 2355–2366.
- Kekarainen T, Savilahti H and Valkonen JP, 2002. Functional genomics on potato virus A: virus genome-wide map of sites essential for virus propagation. Genome Research, 12, 584–594.
- Larbi I, Djilani-Khouadja F, Khamassay N and Fakhfakh H, 2012. Potato virus surveys and wide spread of recombinant PVY-NTN variant in central Tunisia. African Journal of Microbiology Research, 6, 2109–2115.
- Lecoq H, Desbiez C, Wipf-Scheibel C and Girard M, 2003. Potential involvement of melon fruit in the long distance dissemination of cucurbit Potyviruses. Plant Disease, 87, 955–959.

Loebenstein G and Gaba V, 2012. Viruses of Potato. Advance Virus Research, 84, 209–246.

MacDiarmid R, 1994. Tamarillo mosaic potyvirus: characterization and resistance. PhD thesis. Plant Science school of biological sciences. University of Auckland.



- Maoka T, Sugiyama S, Maruta Y and Hataya T, 2010. Application of cDNA macroarray for simultaneous detection of 12 potato viruses. Plant Disease, 94, 1248–1254.
- Mortensen RJ, Shen X, Reid A and Mulholland V, 2010. Characterization of viruses infecting potato plants from a single location in Shetland, an isolated Scottish archipelago. Journal of Phytopathology, 158, 633–640.
- Mossop DW, 1977. Isolation, purification, and properties of tamarillo mosaic virus, a member of the potato virus Y group. New Zealand Journal of Agricultural Research, 20, 535–541.
- de Neergaard E, Munk L and Nielsen SL, 2014. First report of potato leafroll virus, potato virus A, potato virus X and potato virus Y in potato in Greenland. New Disease Reports, 30, 20.
- Pásztor G, Ihárosi EN and Takács A, 2017. Virological examination of public ornamental plants. VIII International Scientific Agriculture Symposium, "Agrosym 2017", Jahorina, Bosnia and Herzegovina, October 2017. Book of Proceedings, pp. 1203–1207.
- Priegnitz U, Lommen WJM, van der Vlugt RAA and Struik PC, 2018. Impact of positive selection on incidence of different viruses during multiple generations of potato seed tubers in Uganda. Potato Research, 62, 1–30.
- Rajamaki M, Merits A, Rabenstein F, Andrejeva J, Paulin L, Kekarainen T, Kreuze JF, Forster RL and Valkonen JP, 1998. Biological, serological, and molecular differences among isolates of potato a potyvirus. Phytopathology, 88, 311–321.
- Robertson NL, Smeenk J and Anderson JM, 2011. Molecular characterization of potato leafroll virus, potato virus A, and potato virus X isolates from potatoes in Alaskan cities and villages. Plant Health Progress, 12, 39.
- Spetz C, Taboada AM, Darwich S, Ramsell J, Salazar LF and Valkonen JP, 2003. Molecular resolution of a complex of potyviruses infecting solanaceous crops at the centre of origin in Peru. Journal of General Virology, 84, 2565–2578.
- Takács A, Cserpes M and Pásztor G, 2018. The role of Solanum nigrum in the spread of potato viruses. IX International Scientific Agriculture Symposium "AGROSYM 2018", Jahorina, Bosnia and Herzegovina. 4–7 October 2018. Book of Proceedings 2018, pp. 895–900.
- Valkonen JPT, Puurand U, Slack SA, Makinen K and M S, 1995. Three strain groups of potato A potyvirus based on hypersensitive Responses in potato, serological properties, and coat protein sequences. Plant Disease, 79, 748–753.
- Were HK, Kabira JN, Kinyua ZM, Olubayo FM, Karinga JK, Aura J, Lees AK, Cowan GH and Torrance L, 2014. Occurrence and distribution of potato pests and diseases in Kenya. Potato Research, 56, 325–342.

## Abbreviations

- EPPO European and Mediterranean Plant Protection Organization
- FAO Food and Agriculture Organization
- IPPC International Plant Protection Convention
- ISPM International Standards for Phytosanitary Measures
- MS Member State
- PLH EFSA Panel on Plant Health
- PVA potato virus A
- RNQP Regulated non-quarantine pests
- PZ Protected Zone
- TFEU Treaty on the Functioning of the European Union
- ToR Terms of Reference

## Glossary

Containment (of a pest)	Application of phytosanitary measures in and around an infested area to prevent spread of a pest (FAO, 1995, 2017)
Control (of a pest)	Suppression, containment or eradication of a pest population (FAO, 1995, 2017)
Entry (of a pest)	Movement of a pest into an area where it is not yet present, or present but not widely distributed and being officially controlled (FAO, 2017)
Eradication (of a pest)	Application of phytosanitary measures to eliminate a pest from an area (FAO, 2017)
Establishment (of a pest)	Perpetuation, for the foreseeable future, of a pest within an area after entry (FAO, 2017)
Impact (of a pest)	The impact of the pest on the crop output and quality and on the environment in the occupied spatial units
Introduction (of a pest) Isolate	The entry of a pest resulting in its establishment (FAO, 2017) Virus population as present in a plant



Lineage Measures	Group of isolates belonging to a distinct phylogenetic cluster Control (of a pest) is defined in ISPM 5 (FAO 2017) as 'Suppression, containment or eradication of a pest population' (FAO, 1995). Control measures are measures that have a direct effect on pest abundance. Supporting measures are organisational measures or procedures supporting the choice of appropriate Risk Reduction Options that do not directly affect pest abundance.
Pathway	Any means that allows the entry or spread of a pest (FAO, 2017)
Phytosanitary measures	Any legislation, regulation or official procedure having the purpose to prevent the introduction or spread of quarantine pests, or to limit the economic impact of regulated non-quarantine pests (FAO, 2017)
Protected zones (PZ)	A Protected zone is an area recognised at EU level to be free from a harmful organism, which is established in one or more other parts of the Union
Quarantine pest	A pest of potential economic importance to the area endangered thereby and not yet present there, or present but not widely distributed and being officially controlled (FAO, 2017)
Regulated non-quarantine pest	A non-quarantine pest whose presence in plants for planting affects the intended use of those plants with an economically unacceptable impact and which is therefore regulated within the territory of the importing contracting party (FAO, 2017)
Risk reduction option (RRO)	A measure acting on pest introduction and/or pest spread and/or the magnitude of the biological impact of the pest should the pest be present. A RRO may become a phytosanitary measure, action or procedure according to the decision of the risk manager
Spread (of a pest)	Expansion of the geographical distribution of a pest within an area (FAO, 2017)
Strain	Group of isolates sharing biological, molecular and/or serological properties