

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

**International Society for Extracellular Vesicles and International Society for Cell and Gene Therapy statement on extracellular vesicles from mesenchymal stromal cells and other cells: considerations for potential therapeutic agents to suppress coronavirus disease-19**

**This is a pre print version of the following article:**

*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/1758797> since 2020-10-20T09:56:57Z

*Published version:*

DOI:10.1016/j.jcyt.2020.05.002

*Terms of use:*

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

## EVs from MSCs and other cells: potential therapeutic agents to suppress COVID-19?

Verena Börger<sup>1,\*,</sup>, Daniel J. Weiss<sup>2,\*,</sup>, Johnathon D. Anderson<sup>3,¶</sup>, Francesc E. Borràs<sup>4</sup>, Benedetta Bussolati<sup>5,¶</sup>, David R.F. Carter<sup>6,¶</sup>, Juan M. Falcón-Pérez<sup>7,8,9,¶</sup>, Mario Gimona<sup>10,¶</sup>, Andrew F. Hill,<sup>11,¶</sup> Andrew M. Hoffman<sup>12,¶</sup>, Dominique de Kleijn<sup>13,¶</sup>, Bruce L. Levine<sup>14, #</sup>, Rebecca Lim<sup>15,¶</sup>, Sai Kiang Lim<sup>16, #, ¶</sup>, Jan Lötvall<sup>17,¶</sup>, S. Alex Mitsialis<sup>18,¶</sup>, Marta Monguió-Tortajada<sup>19,¶</sup>, Maurizio Muraca<sup>20,¶</sup>, Rienk Nieuwland<sup>21,¶</sup>, Anna Nowocin<sup>22,¶</sup>, Lorraine O'Driscoll<sup>23,¶</sup>, Luis A. Ortiz<sup>24, #, ¶</sup>, Donald G Phinney<sup>25, #</sup>, Ilona Reischl<sup>26,5</sup>, Eva Rohde<sup>27,28, #, ¶</sup>, Ralf Sanzenbacher<sup>29</sup>, Clotilde Théry<sup>30,¶</sup>, Wei Seong Toh<sup>31,¶</sup>, Kenneth W. Witwer<sup>32,33,¶, ‡</sup>, Bernd Giebel<sup>1, #, ¶, ‡</sup>

<sup>1</sup> Institute for Transfusion Medicine, University Hospital Essen, University of Duisburg-Essen, Essen, Germany

<sup>2</sup> Department of Medicine, University of Vermont, Burlington, VT, USA

<sup>3</sup> Department of Otolaryngology, Stem Cell Program, University of California, Davis, Davis, USA

<sup>4</sup> REMAR-IVECAT Group, Health Science Research Institute Germans Trias i Pujol (IGTP), Can Ruti Campus, and Nephrology Service, Germans Trias i Pujol University Hospital, Badalona, Spain

<sup>5</sup> Department of Molecular Biotechnology and Health Sciences, University of Torino, Torino, Italy

<sup>6</sup> Department of Biological and Medical Sciences, Oxford Brookes University, Oxford, UK

<sup>7</sup> Exosomes Laboratory, Center for Cooperative Research in Biosciences (CIC bioGUNE), Basque Research and Technology Alliance (BRTA). Derio, Bizkaia, Spain;

<sup>8</sup> Centro de Investigación Biomédica en Red de enfermedades hepáticas y digestivas (CIBERehd), Instituto de Salud Carlos III, Madrid, Spain;

<sup>9</sup> IKERBASQUE, Basque Foundation for Science, Bilbao, Bizkaia, Spain

<sup>10</sup> Spinal Cord Injury & Tissue Regeneration Center Salzburg (SCI-TReCS) GMP Unit and EV-TT Transfer Center, Paracelsus Medizinische Privatuniversität, Salzburg, Austria

<sup>11</sup> La Trobe Institute for Molecular Science, La Trobe University, Bundoora, Australia

<sup>12</sup> School of Veterinary Medicine, University of Pennsylvania, Philadelphia, USA

<sup>13</sup> Department of Vascular Surgery, University Medical Center Utrecht and Netherlands Heart Institute, Utrecht, The Netherlands

<sup>14</sup> Center for Cellular Immunotherapies at the Perelman School of Medicine, University of Pennsylvania Philadelphia, USA

<sup>15</sup> Department of Obstetrics and Gynaecology, Monash University and The Ritchie Centre, Hudson Institute of Medical Research, Melbourne, Australia

<sup>16</sup> Institute of Molecular and Cellular Biology, Agency for Science, Technology and Research, Singapore

<sup>17</sup> Krefting Research Centre, Institute of Medicine, Sahlgrenska Academy at University of Gothenburg, Gothenburg, Sweden

<sup>18</sup> Departments of Pediatrics, Harvard Medical School & Boston Children's Hospital, Boston, USA

<sup>19</sup> ICREC Research Program, Health Science Research Institute Germans Trias i Pujol (IGTP), Can Ruti Campus, and Cardiology Service, Germans Trias i Pujol University Hospital, Badalona, Spain

<sup>20</sup> Department of Women's and Children's Health, University of Padova, Italy

21 Laboratory of Experimental Clinical Chemistry, Department of Clinical Chemistry, and Vesicle Observation Center, Amsterdam UMC, Location AMC, University of Amsterdam, Amsterdam, The Netherlands

22 Biotherapeutics, National Institute for Biological Standards and Control (NIBSC), Medicines and Healthcare products Regulatory Agency, Hertfordshire, UK

23 School of Pharmacy and Pharmaceutical Sciences & Trinity Biomedical Sciences Institute, Trinity College Dublin, Dublin, Ireland

24 Department of Environmental and Occupational Health, University of Pittsburgh, Pittsburgh, Pennsylvania, USA

25 Department of Molecular Therapeutics, The Scripps Research Institute, Jupiter, FL, USA

26 Federal Office for Safety in Health Care (BASG) and Austrian Agency for Health and Food Safety (AGES), Institute Surveillance, Vienna, Austria

27 Department of Transfusion Medicine, University Hospital, Salzburger Landeskliniken GesmbH (SALK), Austria

28 GMP Unit, Spinal Cord Injury & Tissue Regeneration Centre Salzburg (SCI-TReCS), Paracelsus Medical University (PMU), Salzburg, Austria

29 Section Tissue Engineering and Cell Therapeutics, Paul-Ehrlich-Institut, Federal Institute for Vaccines and Biomedicines, Langen, Germany

30 Institut Curie/INSERM U932/PSL Research University, Paris, France

31 Faculty of Dentistry, National University of Singapore, Singapore

32 Department of Molecular and Comparative Pathobiology, 32 Department of Neurology, The Johns Hopkins University School of Medicine, Baltimore, USA

#Authors who are members of the International Society for Cell and Gene Therapy (ISCT)

¶Authors who are members of the International Society for Extracellular Vesicles (ISEV)

All authors but the first (\*) and communicating authors (‡) are listed in an alphabetic order

#### Correspondence:

Kenneth W. Witwer, Department of Molecular and Comparative Pathobiology, Department of Neurology, The Johns Hopkins University School of Medicine, 733 N. Broadway/MRB 829, Baltimore, MD 21205, USA Phone: +1 410-955-9770. e-mail: [kwitwer1@jhmi.edu](mailto:kwitwer1@jhmi.edu)

Bernd Giebel, Institute for Transfusion Medicine, University Hospital Essen, University of Duisburg-Essen, Virchowstr. 179, 45147 Essen, Germany. Phone: +49-201-7234204, e-mail: [bernd.giebel@uk-essen.de](mailto:bernd.giebel@uk-essen.de)

*STATEMENT: The International Society for Cellular and Gene Therapies (ISCT) and the International Society for Extracellular Vesicles (ISEV) recognize the potential of extracellular vesicles (EVs, including exosomes) from mesenchymal stromal cells (MSCs) and possibly other cell sources as treatments for COVID-19. Research and trials in this area are encouraged. However, ISEV and ISCT do not currently endorse the use of EVs or exosomes for any purpose in COVID-19, including but not limited to reducing cytokine storm, exerting regenerative effects, or delivering drugs, pending the generation of appropriate manufacturing and quality control provisions, pre-clinical safety and efficacy data, rational clinical trial design, and proper regulatory oversight.*

First described in December 2019, the severe acute respiratory syndrome associated with coronavirus disease 19 (COVID-19) quickly evolved into a pandemic with severe and increasing worldwide morbidity and mortality. Although most infected patients have mild to moderate symptoms or are even asymptomatic, older patients or those with existing chronic diseases are at greater risk of developing serious complications such as pneumonia or multiple organ failures. COVID-19 respiratory infection is marked by dysregulated immune responses leading to significant respiratory pathology as well as increased probabilities for multi-organ pathologies. While the inflammatory pathways are still being elucidated, notable components include increased circulating levels of pro-inflammatory cytokines and other mediators including interleukin-6 (IL-6), interleukin-1 $\beta$  (IL-1 $\beta$ ), induced protein 10 (IP10) and monocyte chemoattractant protein-1 (MCP-1) (Chen et al., 2020; Diao et al., 2020; Yang et al., 2020). There are also significant alterations in circulating inflammatory cell populations with initial lymphocytosis followed by severe lymphopenia, with increased ratios of helper to regulatory T cells (Chen et al., 2020; Diao et al., 2020; Qin et al., 2020). Since dysregulated immune responses and the cytokine storm are triggers for development of acute respiratory distress syndrome (ARDS), an increasing effort and current clinical trials are focused on immune therapeutic approaches such as IL-1 blockade (anakinra), IL-6 receptor blockade (tocilizumab) or Janus kinase (JAK) inhibition (Mehta et al., 2020). In parallel, there are a rapidly increasing number of cell-based therapy investigations, mostly utilizing mesenchymal stromal cells (MSCs) (Khoury et al., 2020a). These are based on supporting pre-clinical data for use of MSCs delivered either systemically or intratracheally in pre-clinical models of acute lung injuries and on demonstration of safety of systemic MSC administration in recent trials for ARDS resulting from other etiologies (Laffey and Matthay, 2017; Matthay et al., 2019).

Among the cell-based therapy investigations for COVID-19, some registered clinical trials aim to utilize extracellular vesicles (EVs) prepared from MSC conditioned media rather than the cells themselves. They MSC-EVs will be administered intravenously (ChiCTR2000030484), or by inhalation (NCT04276987, ChiCTR2000030261). The rationale for these approaches is

also based on a relatively small but growing number of investigations in pre-clinical lung injury and sepsis models in which MSC-EV preparations were described as safe and effective, if not more effective than their parent cells (Mahida et al., 2020; Worthington and Hagood, 2020). This approach is further supported by a growing body of literature on the therapeutic potential and mechanisms of EVs in a wide range of diseases. This includes recent positive results in a steroid-refractory Graft-versus-Host Disease (GvHD) patient treated with MSC-EVs, and in a single centre, randomized, placebo-controlled phase II/III clinical pilot study on MSC-EV treated patients suffering from chronic kidney disease (CDK) (Kordelas et al., 2014; Nassar et al., 2016).

The mechanisms, by which EVs exert their beneficial effects, as well as their site(s) of action, remain incompletely understood. Nonetheless, effects observed in a range of pre-clinical non-COVID-19 model systems suggest that they may also have efficacy against COVID-19. For example, systemic administration of MSC-EV preparations modulated the immune responses including elevated cytokine storms in relevant lung disease models, including acute lung injury and sepsis (Liu et al., 2019; Mansouri et al., 2019; Monsel et al., 2015; Morrison et al., 2017; Park et al., 2019; Varkouhi et al., 2019; Willis et al., 2018; Zhu et al., 2014). Noteworthy, in *E. coli* induced pneumonia mouse models, MSC-EV administration was found to enhance phagocytosis of bacteria (Hao et al., 2019; Monsel et al., 2015). In a pig model MSC-EVs were shown to attenuate influenza virus-induced acute lung injury, amongst others by inhibiting influenza virus replication (Khatri et al., 2018). Disease attenuating effects on inflammatory immune responses following MSC-EV administration have also been observed in other disease models (Börger et al., 2017). In an ischemic stroke model, for example, systemic MSC-EV administration reduced stroke-induced lymphopenia and pro-inflammatory immune responses in the brain and periphery, resulting in overall improvement of disease symptoms (Doepfner et al., 2015; Wang et al., 2020). These preliminary observations support MSC-EV administration as a potential treatment option for COVID-19.

However, the specific scientific rationale for MSC-EV and other EV administration to COVID-19 patients needs to be better understood and justified. For example, MSC-EVs do not necessarily suppress immune responses, but rather modulate them. Specifically, they seem to moderate acute immune responses towards regulatory responses, with the latter inducing tolerance and restoring homeostasis (Giebel and Hermann, 2019; Zhang et al., 2018; Zhang et al., 2014). While tolerance induction in GvHD and other non-infectious diseases may be beneficial, it might have severe adverse effects in the presence of replicating pathogens. Although influenza and *E. coli* infections were attenuated in selected models (Hao et al.,

2019; Khatri et al., 2018; Monsel et al., 2015), other viruses and bacteria might conceivably expand in uncontrolled manners in induced tolerogenic environments.

There are a number of additional issues that need to be considered before administering MSC-EVs to COVID-19 patients. These include the source of MSC-EVs. MSCs are a heterogeneous cell entity that can be obtained from different tissues. Even if derived from the same tissues, they may display interindividual and eventually clone specific functional differences (Phinney, 2012; Phinney et al., 1999; Radtke et al., 2016; Vogel et al., 2003). Indeed, side by side comparison of four MSC-EV preparations harvested from the conditioned media of different donor derived bone marrow MSCs demonstrated significant variations in cytokine content (Kordelas et al., 2014). Whether this correlates with therapeutic potency is not yet clear; however, in the example of the ischemic stroke model, it was demonstrated that MSC-EV preparations with comparable particle and protein contents can significantly differ in potency. While some preparations effectively suppressed stroke symptoms, others failed to exert therapeutic activities (Wang et al., 2020). Furthermore, in an acute lung injury model, EVs from young but not from aged MSCs alleviated LPS-induced acute lung injury (Huang et al., 2019).

Potentially, heterogeneity of EV potency due to different sources, preparations, aging, and other factors could be resolved by generating immortalized clonal MSC lines that could be rigorously tested for EV production and potency (Chen et al., 2011). Still, apart from their immunomodulatory capabilities, MSC-EVs apparently also control additional biological processes, some with approved therapeutic functions (Arslan et al., 2013), and others that might trigger unforeseen side effects. Just recently, it was found that adipose-derived MSC-EVs had higher thrombogenic activities than bone-marrow-derived MSC-EVs (Chance et al., 2019; Silachev et al., 2019). Thus, the source of parental cells might increase thrombosis risks. Coupled to the finding that activation of complement pathways and an associated procoagulant state seem to result in catastrophic microvascular injury syndrome in a proportion of severe COVID-19 cases (Magro et al., 2020), MSC-EV administration could even be counterproductive in COVID-19.

To this end, it is imperative that stringent “identity” and “potency” parameters are defined and potential side effects addressed before MSC-EV or other EV preparations are released for therapeutic applications (Lener et al., 2015; Reiner et al., 2017; Witwer et al., 2019). To date, many groups use in-house MSC-EV manufacturing and characterization strategies, mainly for preclinical studies (Börger et al., 2017). Protocols fulfilling good manufacturing practice (GMP) criteria are sparse, and just a few have been published (Gimona et al., 2017; Pachler et al., 2017; Rohde et al., 2019). With those product candidates subsequent studies focusing

on safety and clinical pharmacology need to be performed. Results of such studies are mandatory to provide guidance for adjustment of manufacturing, storage, dosing, and administration of EV-based therapeutics in specific target diseases.

We would like to refer to a recent statement by ISCT on the use of MSCs in COVID-19 (Khoury et al., 2020b), as many of the same considerations apply to MSC-EVs or other EVs. Governmental organizations, healthcare providers, and clinical investigators must take the lead by insisting that clinical use of EVs follow appropriate scientific, regulatory and ethical guidelines and are approved only after a rigorous review by duly empowered agencies. The ethical guidelines produced by the World Health Organization (WHO) are a useful baseline.<sup>1</sup> The urgency of the current outbreak does not justify administration of EVs in rather uncontrolled compassionate use settings and does not obviate the need to register clinical trials, obtain informed consent from patients or proxies, and otherwise comply with good clinical practice (GCP). In particular, even limited compassionate use should employ well-characterized MSC-EV preparations produced through strict GMP conditions under the oversight of the relevant national regulatory entity. Additional outbreak-specific measures may be needed, including establishment of simplified clinical protocols for hospitalized patients, such as the WHO COVID-19 core protocol, minimizing risks to trial integrity,<sup>2</sup> changing logistics of trial participant visits (e.g. implementation of remote assessments), and protocol changes for the sake of hazard minimization that may need to be implemented and reported, in Europe, to the Institute for Research in Biomedicine (IRB) Barcelona after the fact. Certainly, regulatory flexibility and support is helpful to foster developments, such as the US Food and Drug Administration (FDA) special emergency program for possible therapies, the Coronavirus Treatment Acceleration Program (CTAP),<sup>3</sup> the European Medicines Agency (EMA) COVID-19 pandemic Task Force (COVID-ETF) activities,<sup>4</sup> the EMA guidance for medicine developers and companies on COVID-19<sup>5</sup> and the guidelines for clinical trials published by an EMA coordinated group<sup>6</sup> or the Medicines and Healthcare products

---

<sup>1</sup> Organisation WH. Guidance for managing ethical issues in infectious disease outbreaks. World Heal Organ 2016:62.

<sup>2</sup> FDA Guidance on Conduct of Clinical Trials of Medical Products during COVID-19 Pandemic Guidance for Industry, Investigators, and Institutional Review Boards. 2020.

<sup>3</sup> Coronavirus Treatment Acceleration Program (CTAP) | FDA n.d. <https://www.fda.gov/drugs/coronavirus-covid-19-drugs/coronavirus-treatment-acceleration-program-ctap> (accessed April 1, 2020).

<sup>4</sup> COVID-19 EMA pandemic Task Force (COVID-ETF): "to help EU Member States and the European Commission to take quick and coordinated regulatory action on the development, authorisation and safety monitoring of treatments and vaccines intended for the treatment and prevention of COVID-19." <https://www.ema.europa.eu/en/human-regulatory/overview/public-health-threats/coronavirus-disease-covid-19/emas-governance-during-covid-19-pandemic# covid-19-ema-pandemic-task-force-section>

<sup>5</sup> <https://www.ema.europa.eu/en/human-regulatory/overview/public-health-threats/coronavirus-disease-covid-19/guidance-medicine-developers-companies-covid-19>

<sup>6</sup> [https://ec.europa.eu/health/sites/health/files/files/eudralex/vol-10/guidanceclinicaltrials\\_covid19\\_en.pdf](https://ec.europa.eu/health/sites/health/files/files/eudralex/vol-10/guidanceclinicaltrials_covid19_en.pdf)

Regulatory Agency (MHRA),<sup>7</sup> respectively. Most or all of the considerations covered for cell-based therapies are also applicable to EV investigations.

In conclusion, to mitigate the risk of potential of life-threatening side effects, ISCT and ISEV strongly urge that the potential benefits and risks in the use of MSC-EVs for COVID-19 be weighed carefully against available pre-clinical data in relevant animal models and clinical data from relevant MSC clinical trials, and that any use of EVs be carefully evaluated through rational clinical trial design, employing well-characterized EV preparations produced under strict GMP conditions and under the proper regulatory oversight.

### **Disclosures:**

JDA: Co-founder of an exosome therapeutics company called Somos Therapeutics, Inc.

BG: Scientific advisory board member of Evox Therapeutics and Innovex Therapeutics SL.

MG: Consulting and Advisory Role: MDimune

BLL: Stock and Other Ownership Interests: Tmunity Therapeutics. Honoraria: Novartis, Terumo, AstraZeneca. Consulting or Advisory Role: Brammer Bio/ThermoFisher Viral Vector Services, Avectas, Immuneel, Ori Biotech, Vycellix.

SKL: Founder, Paracrine Therapeutics, Scientific advisory role: Ilias Biologics and ExoCo

S.A.M. is the inventor of Intellectual Property licensed by BCH to United Therapeutics Corp.

### **Literature:**

Arslan, F., Lai, R.C., Smeets, M.B., Akeroyd, L., Choo, A., Agnor, E.N., Timmers, L., van Rijen, H.V., Doevendans, P.A., Pasterkamp, G., *et al.* (2013). Mesenchymal stem cell-derived exosomes increase ATP levels, decrease oxidative stress and activate PI3K/Akt pathway to enhance myocardial viability and prevent adverse remodeling after myocardial ischemia/reperfusion injury. *Stem Cell Res* 10, 301-312.

Börger, V., Bremer, M., Ferrer-Tur, R., Gockeln, L., Stambouli, O., Becic, A., and Giebel, B. (2017). Mesenchymal Stem/Stromal Cell-Derived Extracellular Vesicles and Their Potential as Novel Immunomodulatory Therapeutic Agents. *Int J Mol Sci* 18.

Chance, T.C., Rathbone, C.R., Kamucheka, R.M., Peltier, G.C., Cap, A.P., and Bynum, J.A. (2019). The effects of cell type and culture condition on the procoagulant activity of human mesenchymal stromal cell-derived extracellular vesicles. *J Trauma Acute Care Surg* 87, S74-S82.

Chen, G., Wu, D., Guo, W., Cao, Y., Huang, D., Wang, H., Wang, T., Zhang, X., Chen, H., Yu, H., *et al.* (2020). Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest* 130.

Chen, T.S., Arslan, F., Yin, Y., Tan, S.S., Lai, R.C., Choo, A.B., Padmanabhan, J., Lee, C.N., de Kleijn, D.P., and Lim, S.K. (2011). Enabling a robust scalable manufacturing process for therapeutic exosomes through oncogenic immortalization of human ESC-derived MSCs. *J Transl Med* 9, 47.

Diao, B., Wang, C., Tan, Y., Chen, X., Liu, Y., Ning, L., Chen, L., Li, M., Liu, Y., Wang, G., *et al.* (2020). Reduction and Functional Exhaustion of T Cells in Patients with Coronavirus Disease 2019 (COVID-19). *medRxiv*, 2020.2002.2018.20024364.

---

<sup>7</sup> <https://www.gov.uk/guidance/mhra-regulatory-flexibilities-resulting-from-coronavirus-covid-19#clinical-trials>



- Doepfner, T.R., Herz, J., Gorgens, A., Schlechter, J., Ludwig, A.K., Radtke, S., de Miroshedji, K., Horn, P.A., Giebel, B., and Hermann, D.M. (2015). Extracellular Vesicles Improve Post-Stroke Neuroregeneration and Prevent Postischemic Immunosuppression. *Stem Cells Transl Med* 4, 1131-1143.
- Gimona, M., Pachler, K., Laner-Plamberger, S., Schallmoser, K., and Rohde, E. (2017). Manufacturing of Human Extracellular Vesicle-Based Therapeutics for Clinical Use. *Int J Mol Sci* 18.
- Hao, Q., Gudapati, V., Monsel, A., Park, J.H., Hu, S., Kato, H., Lee, J.H., Zhou, L., He, H., and Lee, J.W. (2019). Mesenchymal Stem Cell-Derived Extracellular Vesicles Decrease Lung Injury in Mice. *J Immunol* 203, 1961-1972.
- Huang, R., Qin, C., Wang, J., Hu, Y., Zheng, G., Qiu, G., Ge, M., Tao, H., Shu, Q., and Xu, J. (2019). Differential effects of extracellular vesicles from aging and young mesenchymal stem cells in acute lung injury. *Aging (Albany NY)* 11, 7996-8014.
- Khatri, M., Richardson, L.A., and Meulia, T. (2018). Mesenchymal stem cell-derived extracellular vesicles attenuate influenza virus-induced acute lung injury in a pig model. *Stem Cell Res Ther* 9, 17.
- Khoury, M., Cuenca, J., Cruz, F.F., Figueroa, F.E., Rocco, P.R.M., and Weiss, D.J. (2020a). Current Status of Cell-Based Therapies for Respiratory Virus Infections: Applicability to COVID-19. *Eur Respir J*, 2000858.
- Khoury, M., Rocco, P.R.M., Phinney, D.G., Krampera, M., Martin, I., Viswanathan, S., Nolte, J.A., LeBlanc, K., Galipeau, J., and Weiss, D.J. (2020b). Cell-Based Therapies for COVID-19: Proper Clinical Investigations are Essential. *Cytotherapy*.
- Kordelas, L., Rebmann, V., Ludwig, A.K., Radtke, S., Ruesing, J., Doepfner, T.R., Epple, M., Horn, P.A., Beelen, D.W., and Giebel, B. (2014). MSC-derived exosomes: a novel tool to treat therapy-refractory graft-versus-host disease. *Leukemia* 28, 970-973.
- Laffey, J.G., and Matthay, M.A. (2017). Fifty Years of Research in ARDS. Cell-based Therapy for Acute Respiratory Distress Syndrome. *Biology and Potential Therapeutic Value. Am J Respir Crit Care Med* 196, 266-273.
- Lener, T., Gimona, M., Aigner, L., Borger, V., Buzas, E., Camussi, G., Chaput, N., Chatterjee, D., Court, F.A., Del Portillo, H.A., *et al.* (2015). Applying extracellular vesicles based therapeutics in clinical trials - an ISEV position paper. *J Extracell Vesicles* 4, 30087.
- Liu, J., Chen, T., Lei, P., Tang, X., and Huang, P. (2019). Exosomes Released by Bone Marrow Mesenchymal Stem Cells Attenuate Lung Injury Induced by Intestinal Ischemia Reperfusion via the TLR4/NF-kappaB Pathway. *Int J Med Sci* 16, 1238-1244.
- Magro, C., Mulvey, J.J., Berlin, D., Nuovo, G., Salvatore, S., Harp, J., Baxter-Stoltzfus, A., and Laurence, J. (2020). Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: A report of five cases. *Transl Res*.
- Mahida, R.Y., Matsumoto, S., and Matthay, M.A. (2020). Extracellular Vesicles: A New Frontier for Research in Acute Respiratory Distress Syndrome. *Am J Respir Cell Mol Biol*.
- Mansouri, N., Willis, G.R., Fernandez-Gonzalez, A., Reis, M., Nassiri, S., Mitsialis, S.A., and Kourembanas, S. (2019). Mesenchymal stromal cell exosomes prevent and revert experimental pulmonary fibrosis through modulation of monocyte phenotypes. *JCI Insight* 4.
- Matthay, M.A., Calfee, C.S., Zhuo, H., Thompson, B.T., Wilson, J.G., Levitt, J.E., Rogers, A.J., Gotts, J.E., Wiener-Kronish, J.P., Bajwa, E.K., *et al.* (2019). Treatment with allogeneic mesenchymal stromal cells for moderate to severe acute respiratory distress syndrome (START study): a randomised phase 2a safety trial. *Lancet Respir Med* 7, 154-162.
- Mehta, P., McAuley, D.F., Brown, M., Sanchez, E., Tattersall, R.S., Manson, J.J., and Hlth Across Speciality Collaboration, U.K. (2020). COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet* 395, 1033-1034.
- Monsel, A., Zhu, Y.G., Gennai, S., Hao, Q., Hu, S., Rouby, J.J., Rosenzweig, M., Matthay, M.A., and Lee, J.W. (2015). Therapeutic Effects of Human Mesenchymal Stem Cell-derived Microvesicles in Severe Pneumonia in Mice. *Am J Respir Crit Care Med* 192, 324-336.
- Morrison, T.J., Jackson, M.V., Cunningham, E.K., Kissenpfennig, A., McAuley, D.F., O'Kane, C.M., and Krasnodembskaya, A.D. (2017). Mesenchymal Stromal Cells Modulate Macrophages in Clinically Relevant Lung Injury Models by Extracellular Vesicle Mitochondrial Transfer. *Am J Respir Crit Care Med* 196, 1275-1286.
- Nassar, W., El-Ansary, M., Sabry, D., Mostafa, M.A., Fayad, T., Kotb, E., Temraz, M., Saad, A.N., Essa, W., and Adel, H. (2016). Umbilical cord mesenchymal stem cells derived extracellular vesicles can safely ameliorate the progression of chronic kidney diseases. *Biomater Res* 20, 21.

- Pachler, K., Lener, T., Streif, D., Dunai, Z.A., Desgeorges, A., Feichtner, M., Oller, M., Schallmoser, K., Rohde, E., and Gimona, M. (2017). A Good Manufacturing Practice-grade standard protocol for exclusively human mesenchymal stromal cell-derived extracellular vesicles. *Cytotherapy* 19, 458-472.
- Park, K.S., Svennerholm, K., Shelke, G.V., Bandeira, E., Lasser, C., Jang, S.C., Chandode, R., Gribonika, I., and Lotvall, J. (2019). Mesenchymal stromal cell-derived nanovesicles ameliorate bacterial outer membrane vesicle-induced sepsis via IL-10. *Stem Cell Res Ther* 10, 231.
- Phinney, D.G. (2012). Functional heterogeneity of mesenchymal stem cells: implications for cell therapy. *J Cell Biochem* 113, 2806-2812.
- Phinney, D.G., Kopen, G., Righter, W., Webster, S., Tremain, N., and Prockop, D.J. (1999). Donor variation in the growth properties and osteogenic potential of human marrow stromal cells. *J Cell Biochem* 75, 424-436.
- Qin, C., Zhou, L., Hu, Z., Zhang, S., Yang, S., Tao, Y., Xie, C., Ma, K., Shang, K., Wang, W., *et al.* (2020). Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clin Infect Dis*.
- Radtke, S., Gorgens, A., Liu, B., Horn, P.A., and Giebel, B. (2016). Human mesenchymal and murine stromal cells support human lympho-myeloid progenitor expansion but not maintenance of multipotent haematopoietic stem and progenitor cells. *Cell Cycle* 15, 540-545.
- Reiner, A.T., Witwer, K.W., van Balkom, B.W.M., de Beer, J., Brodie, C., Corteling, R.L., Gabriëlsson, S., Gimona, M., Ibrahim, A.G., de Kleijn, D., *et al.* (2017). Concise Review: Developing Best-Practice Models for the Therapeutic Use of Extracellular Vesicles. *Stem Cells Transl Med* 6, 1730-1739.
- Rohde, E., Pachler, K., and Gimona, M. (2019). Manufacturing and characterization of extracellular vesicles from umbilical cord-derived mesenchymal stromal cells for clinical testing. *Cytotherapy* 21, 581-592.
- Silachev, D.N., Goryunov, K.V., Shpilyuk, M.A., Beznoschenko, O.S., Morozova, N.Y., Kraevaya, E.E., Popkov, V.A., Pevzner, I.B., Zorova, L.D., Evtushenko, E.A., *et al.* (2019). Effect of MSCs and MSC-Derived Extracellular Vesicles on Human Blood Coagulation. *Cells* 8.
- Varkouhi, A.K., Jerkic, M., Ormesher, L., Gagnon, S., Goyal, S., Rabani, R., Masterson, C., Spring, C., Chen, P.Z., Gu, F.X., *et al.* (2019). Extracellular Vesicles from Interferon-gamma-primed Human Umbilical Cord Mesenchymal Stromal Cells Reduce Escherichia coli-induced Acute Lung Injury in Rats. *Anesthesiology* 130, 778-790.
- Vogel, W., Grunebach, F., Messam, C.A., Kanz, L., Brugger, W., and Buhning, H.J. (2003). Heterogeneity among human bone marrow-derived mesenchymal stem cells and neural progenitor cells. *Haematologica* 88, 126-133.
- Wang, C., Börger, V., Sardari, M., Murke, F., Skuljec, J., Pul, R., Hagemann, N., Dzyubenko, E., Dittrich, R., Gregorius, J., *et al.* (2020). Mesenchymal Stromal Cell-Derived Small Extracellular Vesicles Induce Ischemic Neuroprotection by Modulating Leukocytes and Specifically Neutrophils. *Stroke*, STROKEAHA119028012.
- Willis, G.R., Fernandez-Gonzalez, A., Anastas, J., Vitali, S.H., Liu, X., Ericsson, M., Kwong, A., Mitsialis, S.A., and Kourembanas, S. (2018). Mesenchymal Stromal Cell Exosomes Ameliorate Experimental Bronchopulmonary Dysplasia and Restore Lung Function through Macrophage Immunomodulation. *Am J Respir Crit Care Med* 197, 104-116.
- Witwer, K.W., Van Balkom, B.W.M., Bruno, S., Choo, A., Dominici, M., Gimona, M., Hill, A.F., De Kleijn, D., Koh, M., Lai, R.C., *et al.* (2019). Defining mesenchymal stromal cell (MSC)-derived small extracellular vesicles for therapeutic applications. *J Extracell Vesicles* 8, 1609206.
- Worthington, E.N., and Hagood, J.S. (2020). Therapeutic Use of Extracellular Vesicles for Acute and Chronic Lung Disease. *Int J Mol Sci* 21, 2318.
- Yang, Y., Shen, C., Li, J., Yuan, J., Yang, M., Wang, F., Li, G., Li, Y., Xing, L., Peng, L., *et al.* (2020). Exuberant elevation of IP-10, MCP-3 and IL-1ra during SARS-CoV-2 infection is associated with disease severity and fatal outcome. medRxiv, 2020.2003.2002.20029975.
- Zhu, Y.G., Feng, X.M., Abbott, J., Fang, X.H., Hao, Q., Monsel, A., Qu, J.M., Matthay, M.A., and Lee, J.W. (2014). Human mesenchymal stem cell microvesicles for treatment of Escherichia coli endotoxin-induced acute lung injury in mice. *Stem Cells* 32, 116-125.