

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

**Degradation of nanoplastics in the environment: Reactivity and impact on atmospheric and surface waters**

**This is the author's manuscript**

*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/1759930> since 2021-02-11T09:19:38Z

*Published version:*

DOI:10.1016/j.scitotenv.2020.140413

*Terms of use:*

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

1 Degradation of nanoplastics in the environment:  
2 reactivity and impact on atmospheric and surface  
3 waters

4 Angelica Bianco<sup>1</sup>, Fabrizio Sordello<sup>2</sup>, Mikael Ehn<sup>1</sup>, Davide Vione<sup>2</sup>, Monica Passananti<sup>1,2,\*</sup>

5

6 <sup>1</sup> Institute for Atmospheric and Earth System Research/Physics, Faculty of Science, University of  
7 Helsinki, FI-00014, Finland.

8 <sup>2</sup> Dipartimento di Chimica, Università di Torino, Via Pietro Giuria 5, 10125, Torino, Italy.

9

10 \* Corresponding author

11 E-mail address: [monica.passananti@unito.it](mailto:monica.passananti@unito.it) (M. Passananti)

12

13

14

15

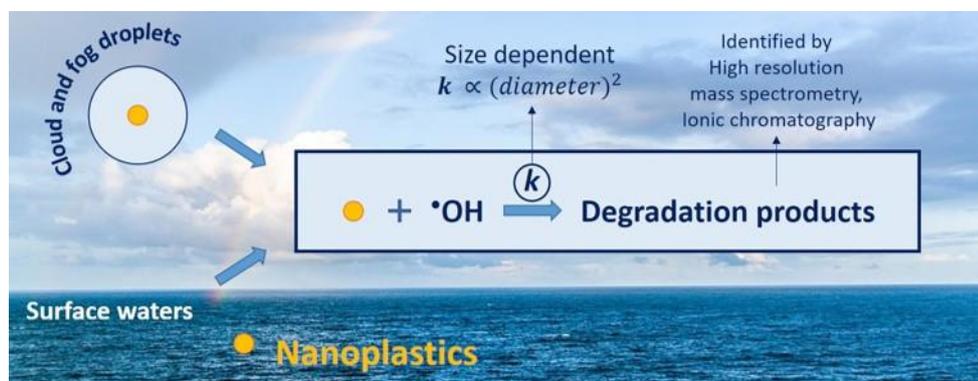
16 **HIGHLIGHTS**

- 17 • Smaller nanoplastics particles show higher reactivity with  $\cdot\text{OH}$ .
- 18 • Reactivity constants of polystyrene nanoparticles with  $\cdot\text{OH}$  are measured.
- 19 • There is potential for nanoplastics to significantly scavenge  $\cdot\text{OH}$  in the environment.
- 20 • Degradation of nanoplastics releases organic compounds in the aqueous phase.

21

## 22 GRAPHICAL ABSTRACT

23



24

25

26

## 27 Abstract

28 Microplastics (MPs) and nanoplastics (NPs) are ubiquitous and contaminate soil, surface waters,  
29 atmospheric aerosol, precipitations, indoor and outdoor environments. However, the occurrence,  
30 transformation and fate of NPs in the environment are still unclear. In this work, polystyrene  
31 nanoparticles (PS-NPs) are used as a proxy of NPs to study their reactivity and potential impact on  
32 atmospheric and surface waters. In particular, the reactivity with hydroxyl radicals ( $\cdot\text{OH}$ ) in the  
33 aqueous phase is investigated. For the first time, a reactivity constant for the reaction of NPs with  
34  $\cdot\text{OH}$  is measured, strongly dependent on the exposed particle surface area of NPs. Degradation  
35 products (short chain carboxylic acids and aromatic compounds), obtained by direct and  $\cdot\text{OH}$ -  
36 mediated photolysis of PS-NPs suspensions, are identified by mass spectrometry. Irradiation of a PS-  
37 NPs suspension under natural sunlight for 1 year has shown the formation of formic acid and organic  
38 compounds similar to those found in riverine and cloud dissolved organic matter, which could  
39 contribute significantly to the dissolved organic matter in the aqueous phase.

40

41 **Keywords:** Nanoplastics; polystyrene; hydroxyl radical; dissolved organic matter; kinetic constant.

42

## 43 **1. Introduction**

44 The large production of plastic material (PlasticsEurope, 2019), together with the mishandling of  
45 plastic waste, has resulted in ubiquitous plastic pollution, which now reaches even the most remote  
46 areas of the Earth (Allen et al., 2019; Bergmann et al., 2019) and has dramatic consequences on flora  
47 and fauna (Acampora et al., 2017; Todd et al., 2010). Plastics undergo a slow process of erosion in  
48 the environment, which is defined as the mass loss of the polymer matrix and is due to the loss of  
49 monomers, oligomers or even pieces of non-degraded polymer (Göpferich, 1996). This process  
50 decreases the size of plastic debris down to microplastics (MPs) and nanoplastics (NPs), defined as  
51 plastic debris with diameters between 1  $\mu\text{m}$  and 5 mm and lower than 1  $\mu\text{m}$ , respectively (Frias and  
52 Nash, 2019). Other possible sources of MPs and NPs are landfills or waste incineration, as well as  
53 synthetic fibers from clothes and houses (Dris et al., 2016; Liu et al., 2019).

54 Microplastics (MPs) contaminate soils, sediments and surface waters (both seawater and fresh  
55 waters) (Besseling et al., 2014; Scheurer and Bigalke, 2018; Yonkos et al., 2014); for example they  
56 represent 94% (in number) of collected plastic debris in the Great Pacific Garbage Patch (Lebreton  
57 et al., 2018). Recent studies have shown that MPs occur also in atmospheric aerosol and precipitations  
58 (Allen et al., 2019; Bergmann et al., 2019; Klein and Fischer, 2019), and their presence has also been  
59 documented in indoor and outdoor environments in Paris and Dongguan (Cai et al., 2017; Dris et al.,  
60 2017; Gasperi et al., 2018). In the last decades there has been a growing interest on MPs occurrence  
61 in the environment and on their potential toxicity (Anbumani and Kakkar, 2018; Dris et al., 2018;  
62 Koelmans et al., 2019), even though their environmental impact is still under debate (Backhaus and  
63 Wagner, 2019). While MPs have been measured in several environmental matrices (Chen et al., 2020;  
64 Han et al., 2019; Li et al., 2019; Xu et al., 2020), the occurrence of NPs in the environment is almost  
65 unknown (Lehner et al., 2019), mainly due to sampling and instrumental limitations (Mintenić et al.,  
66 2018; Schwaferts et al., 2019; Ter Halle et al., 2017). Up to now the occurrence, transformation,  
67 transport and fate of NPs in the environment have still been unclear. Moreover, MPs and NPs are not

68 inert and they can react and interact with chemical species that occur in the surrounding environment  
69 (Jeong et al., 2018; Lee et al., 2019).

70 Erosion of MPs and NPs is the result of mechanical abrasion, oxidation, UV radiation or  
71 microbiological activity (da Costa et al., 2016; Dawson et al., 2018; Lambert et al., 2013; Mattsson  
72 et al., 2018). When a plastic piece degrades down to parts with sizes in the nanometre scale, the  
73 exposed area increases drastically and so does the surface reactivity (Mattsson et al., 2018). However,  
74 the dependence of plastics degradation kinetics upon their particle size is almost unknown, because  
75 most of the degradation studies are carried out on large plastics pieces or on polymeric films (Gewert  
76 et al., 2015; O’Brine and Thompson, 2010).

77 The main abiotic degradation process of plastics is photo-initiated oxidation (Gomiero, 2019),  
78 which leads to polymer chain scission, branching and formation of polymer fragments as well as of  
79 volatile compounds (Gewert et al., 2015). Photochemistry plays a key role in these transformation  
80 processes, by direct photodegradation or by indirect photolysis, through the photogeneration of  
81 reactive transient species such as the hydroxyl radical,  $\cdot\text{OH}$  (Calza and Vione, 2016; Faust, 1994).

82 MPs and NPs in the atmosphere and in surface waters are exposed to sunlight and oxidants (e.g.,  
83  $\cdot\text{OH}$ ). Bubble bursting and wind action can suspend MPs and NPs in air, where they could be present  
84 as dry or deliquescent (in-droplet) particles. In atmospheric waters, like cloud and fog droplets, MPs  
85 and NPs are even more exposed to gas and aqueous phase oxidants: for instance, the  $\cdot\text{OH}$  steady-state  
86 concentration in cloud water is 2 orders of magnitude higher than in sea water (Lallement et al., 2018;  
87 Olasehinde, 2012). Therefore, the degradation pathway(s) of MPs and NPs in atmospheric and surface  
88 waters can be similar, considering that in both cases they are exposed to sunlight, gas-phase and  
89 liquid-phase oxidants.

90 The objective of this work is to better understand the reactivity of an aqueous suspension of  
91 polystyrene NPs (PS-NPs) in the presence of sunlight (direct photolysis) and hydroxyl radical  
92 (indirect photolysis) that is a relevant oxidant in surface waters and atmospheric liquid particles. To

93 develop this new approach to study the reactivity of NPs in the environment, we decided to use well-  
94 defined polymeric NPs. PS standards with different monodisperse particle sizes are commercially  
95 available, and are very suitable for studying the correlation between the reactivity and the exposed  
96 surface. For the first time, a reactivity constant with  $\cdot\text{OH}$  is determined, which can be implemented  
97 into atmospheric and surface waters chemistry models (Bodrato and Vione, 2014) to understand the  
98 environmental impact of NPs degradation. Moreover, the identification of degradation products  
99 allows for the estimation of the contribution of NPs degradation to the natural dissolved organic  
100 matter occurring in the aqueous phase. This work can significantly contribute to assess the impact of  
101 NPs abiotic degradation in the atmospheric aqueous phase, like cloud and fog droplets, and surface  
102 waters.

103

## 104 **2. Material and Methods**

### 105 *2.1. Materials and procedures*

106 Suspensions (1% solids content w/w) of PS-NPs in water were purchased from Thermo Fischer  
107 Scientific (diameter of  $102 \pm 1$  nm,  $203 \pm 5$  nm,  $400 \pm 9$  nm, and  $600 \pm 9$  nm). The suspensions for  
108 irradiation were prepared daily by mixing the stock suspension of PS-NPs, previously sonicated, with  
109 MilliQ water. After irradiation and before analysis, all the suspensions were filtered on  $0.22 \mu\text{m}$  nylon  
110 filters from Sigma-Aldrich. Blank experiments were carried out to verify that no organic material was  
111 released from the filter. Additional information on chemicals used are available in the Supporting  
112 Information (SI), paragraph SI-S1.

### 113 *2.2. Analysis of product formation - UVA Irradiation*

114 Aliquots of 15 mL of PS-NPs suspensions in Milli-Q water (0.05% solid) were irradiated in a  
115 photoreactor placed horizontally in a custom-built wood container covered with aluminium foil. The  
116 photoreactor consisted of a closed quartz tube (diameter 2.5 cm). Two UVA fluorescent lamps (Narva  
117 LT 18W/009), with emission spectrum ranging mainly from 300 to 440 nm, were placed on top of

118 the photoreactor (distance 6 cm), as reported in Figures S1-S2. UVA radiation was chosen because it  
119 represents the main UV fraction of solar light. PS-NPs with diameter of  $400 \pm 9$  nm were chosen for  
120 this series of experiments, because they can be easily filtered and analysed by means of SMPS  
121 (scanning mobility particle sizer, SI-S2), in order to determine the size and concentration of the  
122 suspension and to investigate eventual formation of aggregates. All the experiments were carried out  
123 at  $293 \pm 2$  K using a continuous flow of compressed air to cool the system. An aliquot of the  
124 suspension (3 mL) was withdrawn from the reactor and used for the analysis of degradation products  
125 at fixed times. Filtered solutions were analysed using an ElectroSpray Ionization-Differential  
126 Mobility Analyser (ESI-DMA) coupled with Atmospheric Pressure interface Time Of Flight Mass  
127 Spectrometer (Tofwerk) (ESI-DMA-API-TOF) (Kangasluoma et al., 2016; Passananti et al., 2019).  
128 Acetate and formate anions were determined by ion chromatography. More details on the analytical  
129 methods and instrumentation are available in SI-S2. The size and the concentration of PS-NPs were  
130 measured by SMPS before and after irradiation. No difference was found: PS-NPs keep the same  
131 diameter (no aggregation) and the same concentration.

### 132 2.3. Competition method for the measurement of the reactivity constant between PS and $\cdot\text{OH}$ .

133 The reaction rate constants between  $\cdot\text{OH}$  and PS-NPs were measured using the competition method  
134 described by Zhou and Mopper (1990), employing benzoic acid (hereafter BZ) as a probe.  
135 Suspensions containing 0.05% PS-NPs, 50 mM  $\text{H}_2\text{O}_2$  and BZ with different concentrations, varying  
136 from 0.18 mM to 1.2 mM, were irradiated under UVA light for 60 minutes. In these experiments the  
137 loading of PS-NPs was kept constant, to avoid interference of varying NPs loadings on the light field  
138 and irradiance in the suspensions. In such a way, the light scattering of the suspension would not  
139 change among different experiments and would thus not affect the measured rate constants. To carry  
140 out kinetic competition, BZ concentration was thus varied. The time evolution of BZ was monitored  
141 by High-Performance Liquid Chromatography coupled with Diode Array Detection (HPLC–DAD),

142 using conditions detailed in SI-S3. Blank experiments excluded the degradation of BZ under UVA  
143 irradiation alone (direct photolysis).

#### 144 *2.4. Experiment under real sunlight irradiation*

145 A suspension of PS 100 nm in MilliQ water (0.2% v/v) was exposed for 14 months (August 1st, 2018-  
146 October, 1st, 2019) on the rooftop of the Physicum building in Kumpula Campus in Helsinki  
147 (Finland). The irradiation set-up is shown in Figure S3. The average temperature during this period  
148 was 8.5°C, with a minimum on January 22<sup>nd</sup> (2019) of -18.9°C and a maximum on July 28<sup>th</sup> (2019)  
149 of 32.1°C. The suspension froze during wintertime, but the large amount of air inside the reactor  
150 prevented breaking of the glass. Filters with mesh size lower than 0.2 µm were not available, thus we  
151 filtered the 100 nm PS-NPs suspension on 0.2 µm filters by gravity, without application of pressure.  
152 The resulting solution was limpid and the particles that might have passed through the filter did not  
153 affect the following spectroscopic and spectrometric analysis. Solar radiation was measured by  
154 SMEARIII station with a time resolution of one minute, and the relevant data are available online at  
155 SMEAR website (<https://avaa.tdata.fi/web/smart/smear>).

156

### 157 **3. Results and Discussion**

#### 158 *3.1 Interaction with hydroxyl radicals*

159 Surface modification due to polymers aging by light and gas-phase oxidants, such as O<sub>3</sub>, is well  
160 known (Singh and Sharma, 2008; Yousif and Haddad, 2013). However, experiments are usually  
161 performed on films and with high concentration of O<sub>3</sub>, which are non-representative of environmental  
162 conditions. Concerning PS, attention has been focused on the functionalisation of the polymer chain  
163 and on surface changes, using ATR/FTIR spectroscopy and microscopy techniques (SEM, AFM)  
164 (Yang et al., 2018; Zhang et al., 2000).

165 PS particles occurring in surface waters and in cloud/fog waters can also react with aqueous-phase  
166 oxidants, such as most notably the hydroxyl radicals (·OH) (Arakaki et al., 2013; Faust and Allen,

167 1993; Mopper and Zhou, 1990; Vione et al., 2010). A previous study of oxidation in dichloromethane  
168 reveals that  $\cdot\text{OH}$  addition to PS occurs at the phenyl groups, and that mucondialdehydes are produced  
169 by chain scission (Weir, 1978). Surface modifications and functionalisation of MPs and NPs lead to  
170 increased hydrophilicity, which could have an effect on ice nucleating properties (Ganguly and Ariya,  
171 2019). However, to our knowledge, no study on PS reactivity with  $\cdot\text{OH}$  has been carried out in the  
172 aqueous phase. As first approach, we investigated the potential reactivity between PS-NPs and  
173 photogenerated  $\cdot\text{OH}$  in aqueous solution. Figure 1 reports a preliminary test of UVA-irradiation of  
174 PS-NPs in the presence of  $\text{H}_2\text{O}_2$  and benzoic acid (BZ) in water, using the experimental conditions  
175 detailed in SI-S4. The plot reports the BZ concentration as a function of time. The degradation rate  
176 of BZ decreases in the presence of PS-NPs, suggesting that PS-NPs may be able to significantly  
177 scavenge  $\cdot\text{OH}$ .

### 178 3.2 Reactivity of PS-NPs: measurement of the reactivity constant with hydroxyl radical

179 PS-NPs react with  $\cdot\text{OH}$ , but no information on the reactivity rate constants is reported in previous  
180 works. Reactivity constants are essential for modelling studies, with the aim of assessing PS-NPs  
181 degradation in different environmental scenarios as well as their ability to scavenge  $\cdot\text{OH}$  and to affect  
182 photo-oxidation cycles in natural waters. For the first time, the rate constant of PS-NPs with  $\cdot\text{OH}$  is  
183 determined with the competition method described in Section 2.3, using BZ as reference compound.  
184 Three different sizes of PS-NPs are used, namely 200 nm, 400 nm and 600 nm. As discussed in detail  
185 in SI-S6, the second-order kinetic constants between PS and  $\cdot\text{OH}$  are calculated with the following  
186 equation:

$$187 \quad k_{\text{OH,PS}} = \frac{a}{b} \times k_{\text{OH,BZ}} - [\text{H}_2\text{O}_2] \times k_{\text{OH,H}_2\text{O}_2} \quad (1)$$

188 where  $a/b$  is the ratio between the slope and the intercept of the linear correlation reported in the plot  
189 ( $x =$  inverse of BZ concentration,  $y =$  inverse of BZ degradation rate),  $k_{\text{OH,BZ}}$  is the second-order  
190 reaction rate constant between BZ and  $\cdot\text{OH}$  ( $5.9 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$ ) (Buxton et al., 1988),  $[\text{H}_2\text{O}_2]$  is the  
191 hydrogen peroxide concentration, and  $k_{\text{OH,H}_2\text{O}_2}$  is the second-order reaction rate constant between

192  $\cdot\text{OH}$  and  $\text{H}_2\text{O}_2$  ( $2.7 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ ) (Buxton et al., 1988). Figure S4 shows the plot used to determine  
193 the kinetic constant. BZ is also a degradation product of PS-NPs (*see below*); however, due to the  
194 short time of the kinetic experiments and the large concentration difference between initially added  
195 and photogenerated BZ, there is no interference due to the formation of degradation products. This  
196 applies both to the monitoring of the BZ time trend, and to the ability of the products (including BZ)  
197 to scavenge  $\cdot\text{OH}$  on their own. Indeed, less than 0.1% of  $\cdot\text{OH}$  would react with BZ produced during  
198 PS degradation in one hour.

199 The kinetic constants between  $\cdot\text{OH}$  and PS-NPs ( $k_{\text{OH,PS}}$ ) are reported in Table 1. Because the  
200 concentration of PS-NPs used for the rate-constant calculation was expressed in #particles  $\text{mL}^{-1}$  (see  
201 SI-S6), the measure unit of  $k_{\text{OH,PS}}$  is  $\text{mL s}^{-1} \text{ #particles}^{-1}$ . We observed that the size of the particles  
202 (*i.e.*, their surface area) plays a role in particles reactivity, and that the values of  $k_{\text{OH,PS}}$  are different  
203 for each PS particle size. The kinetic constant is higher for PS 600 nm, which exposes a larger surface  
204 area, and smaller for PS 200 nm, which exposes a surface area of  $0.126 \mu\text{m}^2$  per particle (see Table  
205 1). This result can be explained by the fact that  $\cdot\text{OH}$  would react with the PS surface sites (most likely,  
206 the aromatic rings) and it has no access to the polymer bulk. The measured kinetic constants are  
207 proportional to the square of the diameter  $d$  of the particles ( $k_{\text{OH,PS}} \propto d^2$ , see Figure S5); therefore,  
208 the reactivity constant of PS-NPs can be expressed as a function of their diameter, allowing for the  
209 extrapolation of kinetic constants at different diameters (see Table 1). We decided to express the  
210 kinetic constants as a function of the diameter, in order to employ them in kinetic models in  
211 combination with environmental data. Indeed, environmental data about NPs and MPs provide  
212 information on their concentration (#particles/volume) and size (generally a range), without  
213 specifying the surface area or the specific shape of the collected debris. Therefore, a kinetic constant  
214 expressed as a function of the diameter would be more useful compared to a surface-area-normalized  
215 rate constant. Further studies are needed to obtain data on the surface area of NPs in the environment,  
216 in order to overcome this spherical-approximation. It is important to notice that the reactivity is higher

217 for single particles with larger diameters and higher surface area; however, at equal mass  
218 concentration, PS-NPs with lower particle size (200 nm) expose a higher surface area than PS-NPs  
219 with larger particle size (600 nm) and thus show higher reactivity, as reported in Figure S6 and Table  
220 S3.

### 221 3.3 Impact of environmental stressing agents on PS-NPs: degradation products

222 We have demonstrated that  $\cdot\text{OH}$  reacts with PS-NPs and determined the reaction rate constants, but  
223 little is known about the impact of degraded plastics on the environment, in particular concerning the  
224 release of organic compounds. Vicente et al (2009) studied in detail the effect of  $\text{O}_3$  and UVA  
225 radiation on aerosolised PS-NPs. Davidson et al. (2005) investigated the surface modification after  
226  $\text{O}_3$  and light exposure, and analysed the photodegradation products with GC-MS after washing with  
227 methanol or water. They found that benzaldehyde, acetophenone, BZ, carboxylic acids, aromatic  
228 ketones and esters are produced during PS exposure to  $\text{O}_3$  and light. However, limited information is  
229 available concerning the photodegradation products formed by  $\cdot\text{OH}$  reactions with PS-NPs.  
230 Therefore, we investigated how plastic degradation by  $\cdot\text{OH}$  affects dissolved organic matter chemical  
231 composition. Suspensions of PS-NPs were exposed to 4 experimental conditions described in Table  
232 S1 under the irradiation set-up presented in Figure S2. This set-up allowed for estimating the impact  
233 of: (1) direct photodegradation + photogenerated  $\cdot\text{OH}$  (indirect photolysis); (2) direct  
234 photodegradation alone; (3) reactivity with  $\text{H}_2\text{O}_2$  in dark conditions; and (4) possible  
235 hydrolysis/degradation in dark conditions. Suspensions were sampled at fixed times, then filtered on  
236 nylon filters and analysed by ESI-DMA-APiToF. High-resolution mass spectrometry allows for the  
237 attribution of molecular formulas to degradation products. Trends of the main products formed by  
238 indirect and direct photodegradation are reported in Figure S7a and S7b, respectively, while  
239 degradation products formed by reactivity with and without  $\text{H}_2\text{O}_2$  in dark conditions are reported in  
240 Figure S8. The  $\cdot\text{OH}$  reactivity induces the formation of carboxylic compounds such as formic (also

241 detected as formic acid dimer), acetic, lactic acids, and aromatic acids like BZ, as well as aliphatic  
242 and aromatic aldehydes like muconaldehyde and benzaldehyde.

243 Among the products of PS-NPs photodegradation, benzoquinone, dibenzoyl methane and benzoic  
244 anhydride are also observed. The benzene signal increases by more than 350% of the initial value,  
245 while the phenol signal increases by 40% during the first 24h and later on decreases, as shown in  
246 Figure 2. The formation of degradation products could be explained through the  $\cdot\text{OH}$  addition to  $\pi$ -  
247 systems or hydrogen abstraction. A possible reaction mechanism, detailing the reactivity of  
248 polystyrene and the formation of degradation products, is reported in Figure 3. We considered the  
249 reactions induced by photogenerated  $\cdot\text{OH}$  (light +  $\text{H}_2\text{O}_2$ ), but the mechanism is probably the same  
250 with light only. The tertiary polystyryl radicals, generated by hydrogen abstraction or  $\cdot\text{OH}$  addition,  
251 can efficiently react with molecular oxygen to form peroxy radicals,  $\text{ROO}\cdot$ . The latter can recombine  
252 or give sequential reactions, leading to the formation of hydroxyl, carbonyl or carboxylic functions.  
253 The carboxylic group  $\text{RCOOH}$  can react with  $\cdot\text{OH}$  to give  $\text{RCOO}\cdot$ , which decomposes to form  $\text{R}\cdot$  and  
254  $\text{CO}_2$  (Gardette et al., 1995). Tertiary polystyryl radicals can be generated by absorption of light with  
255 wavelength longer than 300 nm (Gardette et al., 1995), explaining why formation of BZ,  
256 benzaldehyde, benzoic anhydride, dibenzoyl methane, benzene, acetic and formic acids are also  
257 observed in the case of direct photodegradation (Figure 2). However, the formation of the tertiary  
258 polystyryl radicals seems to be enhanced in the presence of  $\cdot\text{OH}$ , as suggested by the much higher  
259 concentration values of photodegradation products in the  $\text{H}_2\text{O}_2$  + light experiments compared to light  
260 alone. A previous study on the irradiation of PS films in both humid and dry conditions revealed the  
261 formation of acetophenone (Gardette et al., 1995). This degradation product is not detected under our  
262 experimental conditions in the presence of  $\cdot\text{OH}$ , while its signal increases during the direct photolysis  
263 process (200% of the initial value). The rationale for the lack of acetophenone in the  $\cdot\text{OH}$  experiments  
264 could be that this compound reacts rapidly with  $\cdot\text{OH}$ , with a second-order reaction rate constant of  
265  $6.5 \times 10^9 \text{ M}^{-1}\text{s}^{-1}$  (Neta and Dorfman, 1968), thereby disappearing immediately after formation. Dark

266 experiments yield degradation products in limited amount, as reported in Figure S8 that shows the  
267 trends observed with PS-NPs and (when relevant) H<sub>2</sub>O<sub>2</sub>.

268 ESI-DMA-APiToF allows for clearly identifying the degradation products, but does not provide  
269 reliable quantification: the complexity of the matrix makes it difficult to perform signal calibration,  
270 even with standard additions. For this reason, PS-NPs suspensions subjected to experimental  
271 condition (1) (H<sub>2</sub>O<sub>2</sub> + light) were analysed with ion chromatography to quantify acetic and formic  
272 acids, while benzoic acid (BZ) concentration was measured with the same HPLC method described  
273 previously. Trends and comparison with mass signals are reported in Figure S9. During the first 24h  
274 of irradiation, the concentration of acetic and formic acids increases rapidly. It then reaches 0.38 mM  
275 and 0.25 mM respectively, after 4 days of irradiation. BZ concentration increases to around 63 μM  
276 after 4 days of irradiation. The formation of these degradation products contributes to the dissolved  
277 organic carbon (DOC): acetic acid is responsible for an increase of 9.1 mgC L<sup>-1</sup>, formic acid of 3.0  
278 mgC L<sup>-1</sup>, and BZ of 5.3 mgC L<sup>-1</sup>. These values are of the same order of magnitude as the DOC in  
279 atmospheric hydrometeors (Bianco et al., 2015; Herckes et al., 2013, 2002; van Pinxteren et al., 2016)  
280 and in surface waters, except for the high-DOC aquatic systems (Bittar et al., 2016; Chen et al., 2004;  
281 Del Castillo et al., 1999; Galgani and Engel, 2016; Massicotte et al., 2017; Stedmon and Cory, 2014).  
282 Therefore, PS-NPs degradation could potentially impact the composition of the aqueous phase in both  
283 atmospheric and surface waters.

284 3.4 *Environmental conditions: PS-NPs exposed for 14 months to sunlight and temperature changes*  
285 *(direct photolysis)*

286 Laboratory irradiations give a first rapid estimate of the degradation of PS-NPs in the aqueous phase.  
287 However, longer irradiations in real environmental conditions are more suitable to investigate the  
288 release of organic compounds during PS-NPs degradation. A suspension of PS 100 nm in MilliQ  
289 water was exposed to sunlight from August 1<sup>st</sup>, 2018 to October, 1<sup>st</sup>, 2019. Figure S10 reports  
290 temperature and radiation averaged on hourly measurements for the considered period. PS-NPs were

291 degraded, they agglomerated in clusters and the aqueous system became yellowish (visual control).  
292 An UV-vis spectrum after filtration was recorded showing the formation of chromophoric organic  
293 compounds that absorb in the UV region up to 400 nm (Figure S11). The suspension was analysed  
294 before and after solar exposure, using the same methods described for the laboratory irradiation  
295 (Section 2.2). Figure S12a reports the variation in MS signals of selected compounds, while Figure  
296 S12b reports the full mass spectrum obtained for the irradiated suspension. Mass spectrometric  
297 analysis shows the formation of formic, benzoic and lactic acids and of benzaldehyde, while the  
298 formation of other compounds is negligible. Probably, during 1-year irradiation, the degradation  
299 products of PS that absorb in the near UV (see Figure S11) are transformed into small organic  
300 compounds by photochemical reactions. However, Figure S12b reveals the presence of many  
301 unidentified compounds, with a spectrum shape similar to those obtained for riverine and cloud  
302 dissolved organic matter.(Bianco et al., 2018; Dubinenkov et al., 2015) This similarity is probably  
303 due to the presence of a large variety of organic compounds, with hydroxyl, carboxylic and carbonyl  
304 functions, which might be similar to the composition of natural dissolved organic matter. This  
305 hypothesis is supported by the Excitation Emission Matrix (EEM) fluorescence spectrum of the liquid  
306 phase (Figure S13). The presence of an emission signal around 410 nm induced by excitation around  
307 300 nm recalls one of the characteristic peaks of humic substances, namely peak C (Coble, 1996) that  
308 is routinely observed in surface waters (Holbrook et al., 2006). A similar peak was also observed for  
309 HULIS in the atmosphere (Muller et al., 2008), and it could be due to highly oxygenated small  
310 compounds (Ghigo et al., 2019) that could derive from several oxidation steps during many months  
311 of sunlight exposure. Formic, acetic and benzoic acid quantification reveals concentration values of  
312 7.83 mM, 4.98 mM and 0.13 mM, respectively (see Table S2), and the detected concentration of  
313 formic acid corresponds to 94.0 mgC L<sup>-1</sup>. Formate represents the last step of the degradation of  
314 organic matter in the aqueous phase, as described in many environmental aqueous phase chemistry  
315 models (Ervens, 2015; Herrmann et al., 2000; Li et al., 2017; Mouchel-Vallon et al., 2017).

#### 316 4. Conclusions

317 This work aims at evaluating the abiotic degradation of PS-NPs in the aqueous phase, as a proxy of  
318 nanoplastics to study the reactivity with hydroxyl radicals. The results obtained can be useful for both  
319 surface and atmospheric waters, like cloud and fog droplets.

320 To get insight into the environmental significance of the present findings we have compared the  
321 potential effect of PS-NPs and DOC as  $\cdot\text{OH}$  scavengers. The DOC is, indeed, the main scavenger of  
322  $\cdot\text{OH}$  radicals occurring in freshwater and cloud-water (Arakaki et al., 2013; Gligorovski et al., 2015).  
323 Figure 4 reports an estimate of  $k_{\text{scavenger}}$  ( $\text{s}^{-1}$ ) of  $\cdot\text{OH}$ , which is the sum of the contributions of  $\cdot\text{OH}$   
324 reaction with DOC and NPs, as a function of the DOC ( $k_{\text{DOC}} = 2 \times 10^4 \text{ (L s}^{-1} \text{ mgC}^{-1}) \times \text{DOC}$ ) and NPs  
325 (Gligorovski et al., 2015). The typical DOC interval in cloud water (0-10  $\text{mgC L}^{-1}$ ) (Deguillaume et  
326 al., 2014; van Pinxteren et al., 2016) is highlighted in light blue in the figure, and that in surface  
327 freshwaters ( $> 1 \text{ mgC L}^{-1}$ ) in violet (Wetzel, 2001). Concentrations of DOC higher than 30  $\text{mgC L}^{-1}$   
328 are measured in surface water in specific cases only (Wetzel, 2001) but they are not considered in  
329 this work, because in such cases DOC is obviously the main scavenger of  $\cdot\text{OH}$  radicals.

330 The concentration of PS-NPs in the environment is difficult to predict considering the lack of data.  
331 For this reason, we varied the PS-NPs concentration from  $10^{10}$  #particles  $\text{mL}^{-1}$ , one order of  
332 magnitude higher than the concentration of NPs in polluted environments ( $10^9$  #particles  $\text{mL}^{-1}$ , as per  
333 Gallego-Urrea et al., (2010)) down to  $10^6$  #particles  $\text{mL}^{-1}$ , a reasonable concentration for lesser  
334 polluted aqueous media (Gallego-Urrea et al., 2010). We considered a reactivity constant between  
335  $\cdot\text{OH}$  and NPs of  $1.93 \times 10^{-4} \text{ mL s}^{-1} \text{ #particles}^{-1}$ , the one determined for PS-NPs 600 nm in this work.  
336 Figure 4 shows that when the DOC concentration is low, like in DOC-poor surface freshwater and  
337 atmospheric cloud-water, there is potential for NPs to significantly scavenge  $\cdot\text{OH}$ . In particular, at  
338  $10^9$  particles  $\text{mL}^{-1}$  NPs could even become the main  $\cdot\text{OH}$  scavenger if  $\text{DOC} < 9 \text{ mgC L}^{-1}$ . Moreover,  
339 at very low DOC values (1  $\text{mgC L}^{-1}$  or below) the organic compounds released by NPs

340 photodegradation, especially if the latter takes place over a long time period, could give a non-  
341 negligible contribution to both the water DOC and the  $\cdot\text{OH}$  scavenging ability.

342 The comparison in the reactivity of NPs and DOC suggests three main issues:

343 (i) The ability of NPs to scavenge  $\cdot\text{OH}$  should be taken into account in models aimed at describing  
344 the role of  $\cdot\text{OH}$  in environmental waters. To do so, however, additional data are needed on the  $\cdot\text{OH}$   
345 reactivity of other plastic types (e.g., polyethylene and polypropylene), on the correlation between  
346 size of particles and reactivity, and on the concentration of NPs in a wide variety of aquatic  
347 environments.

348 (ii) Due to their scavenging ability, it is very important to assess the fate of NPs upon  $\cdot\text{OH}$  reaction,  
349 including the potential to release dissolved organic compounds.

350 (iii) There is also the need to compare the  $\cdot\text{OH}$  scavenging ability of NPs themselves with that of the  
351 compounds that are released by their photodegradation, especially over long irradiation times.

352 In aquatic environments with higher concentration of DOC (or other  $\cdot\text{OH}$  scavengers, such as bromide  
353 in seawater) (Buxton et al., 1988; Gligorovski et al., 2015), PS-NPs can react with sunlight as shown  
354 through direct photolysis experiments, leading to the formation of very similar degradation products  
355 as those obtained under  $\cdot\text{OH}$ -mediated degradation. The identification of degradation products of PS-  
356 NPs suspensions subjected to direct photolysis and  $\cdot\text{OH}$ -induced degradation highlights the formation  
357 of several compounds, mainly small carboxylic acids and some toxic compounds like benzene and  
358 phenol. These compounds could have an effect on the ecosystems by increasing the dissolved organic  
359 matter amount. Moreover, the production of light-absorbing species (Figure S11) could interfere with  
360 the natural photochemical equilibrium in environmental waters.

361 Our results can be implemented in models, allowing for the estimation of the amount of organic matter  
362 produced by NPs abiotic degradation and, therefore, of the impact of NPs on environmental equilibria.

363

364 **Declaration of Competing Interest**

365 The authors declare that they have no known competing financial interests or personal relationships  
366 that could have appeared to influence the work reported in this paper.

367

### 368 **Acknowledgments**

369 We acknowledge funding from the University of Helsinki (Three-Year Research Grants), the ERC  
370 Projects 692891-DAMOCLES and 638703-COALA, ATMATH Project.

371

### 372 **Author contribution**

373 A.B. formal analysis and writing-original draft, F.S. formal analysis and writing-review and editing,  
374 M.E. supervision and writing-review and editing, D.V. software, supervision and writing-review and  
375 editing, M.P. conceptualization, funding, formal analysis, writing-original draft and review.

376

### 377 **Appendix A. Supplementary data**

378 Supplementary data to this article can be found online.

379

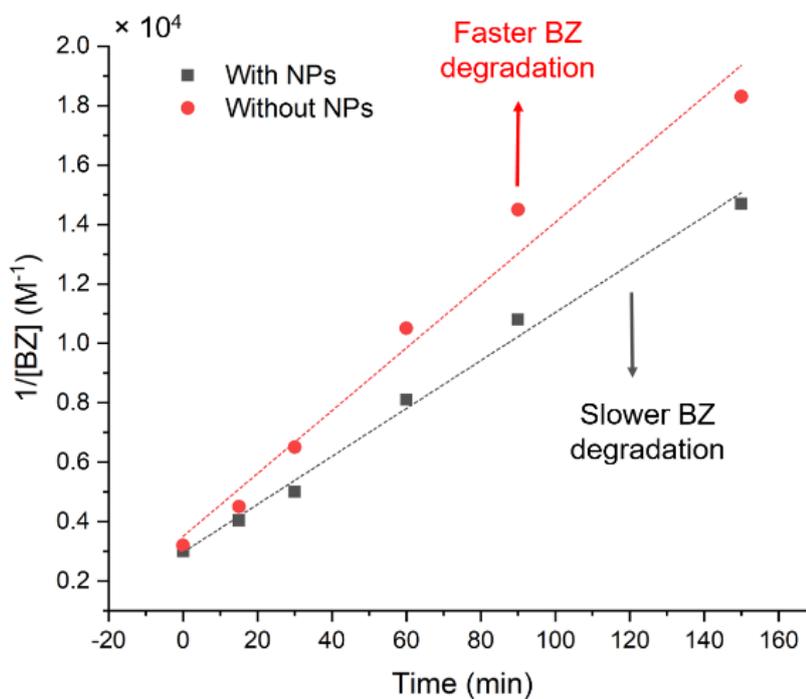
## 380 TABLES

Diameter of PS-NPs ( $\mu\text{m}$ )	Particle surface area ( $\mu\text{m}^2$ )	$k_{OH,PS}$ ( $\text{mL s}^{-1} \text{\#particles}^{-1}$ )
0.20	0.126	$(6.50 \pm 0.38) \times 10^{-5}$
0.40	0.502	$(8.64 \pm 1.30) \times 10^{-5}$
0.60	1.130	$(1.93 \pm 0.22) \times 10^{-4}$
Correlation between $k_{OH,PS}$ ( $\text{mL s}^{-1} \text{\#particles}^{-1}$ ) and diameter of PS-NPs		
$k_{OH,PS} = (5.47 \pm 0.78) \times 10^{-4} d^2$		
d = diameter ( $\mu\text{m}$ )		

381 **Table 1:** Kinetic constants between  $\cdot\text{OH}$  and PS-NPs ( $k_{OH,PS}$ ) as a function of the particle surface area for each PS-NPs  
382 size (200 nm, 400 nm and 600 nm). Correlation between  $k_{OH,PS}$  and the diameter of PS-NPs. The error bounds represent  
383 the sigma-level variability derived from data fit, which was used to obtain the relevant rate constant values.

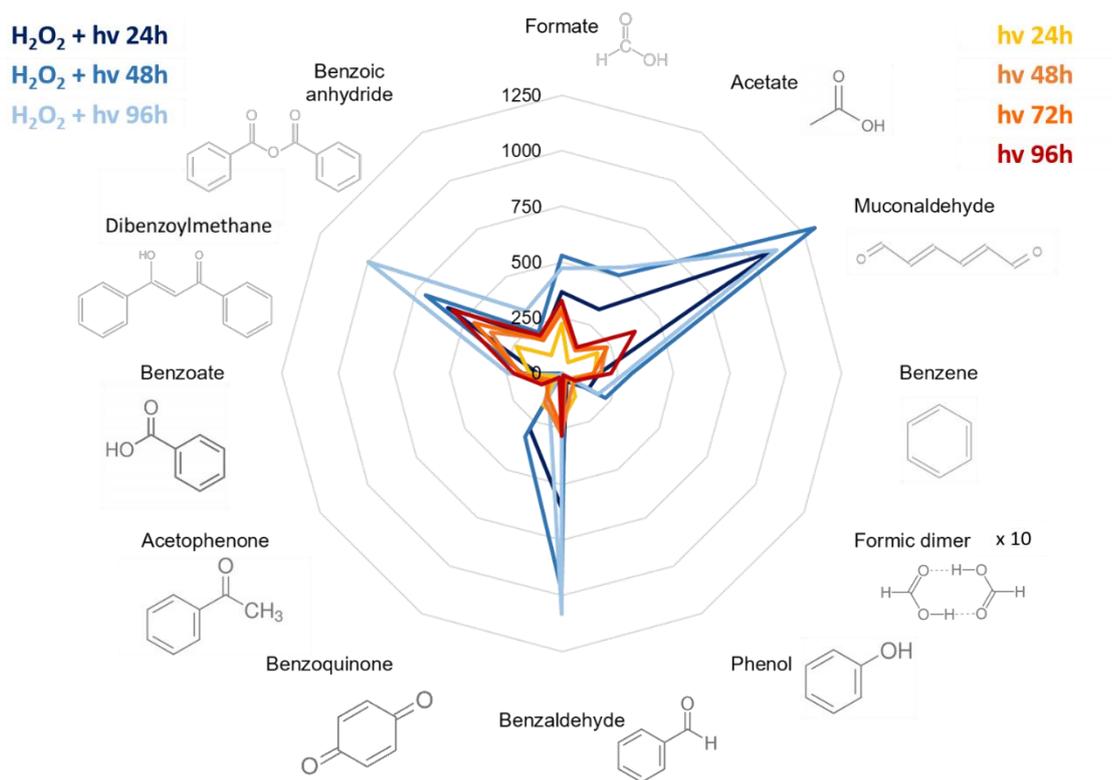
384

## 385 FIGURES



386

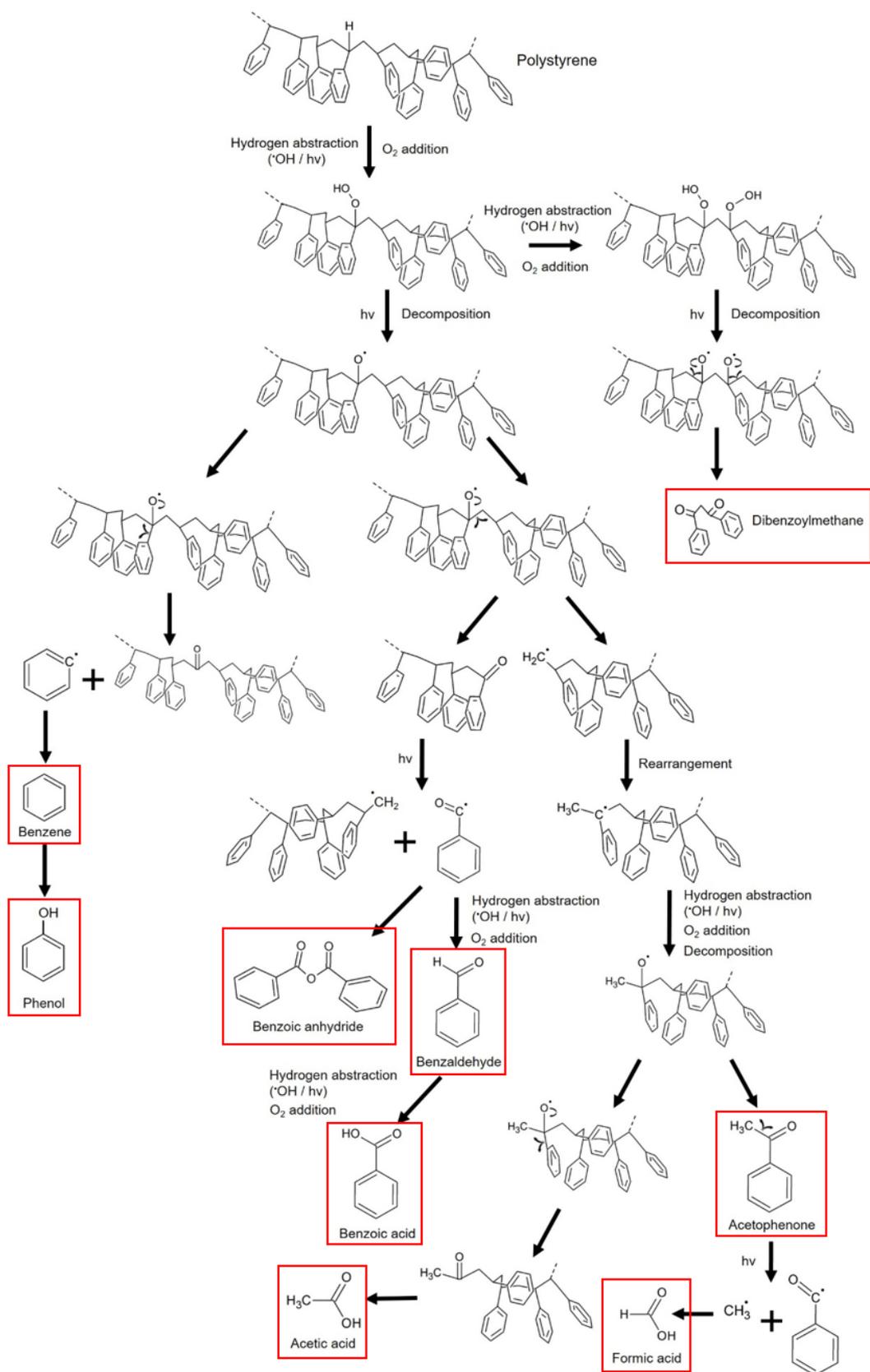
387 **Figure 1:** Degradation of BZ (expressed as  $1/[\text{BZ}]$ ) as function of time with and without NPs in the presence of 50 mM  
388 of  $\text{H}_2\text{O}_2$  under UVA irradiation in water.



389

390 **Figure 2:** Signal increase (%) of degradation products over time under: direct photolysis + hydroxyl radical mediated  
 391 oxidation (H<sub>2</sub>O<sub>2</sub> + hv) (in shade of blue) and b) direct photolysis (hv) (in shade of red).

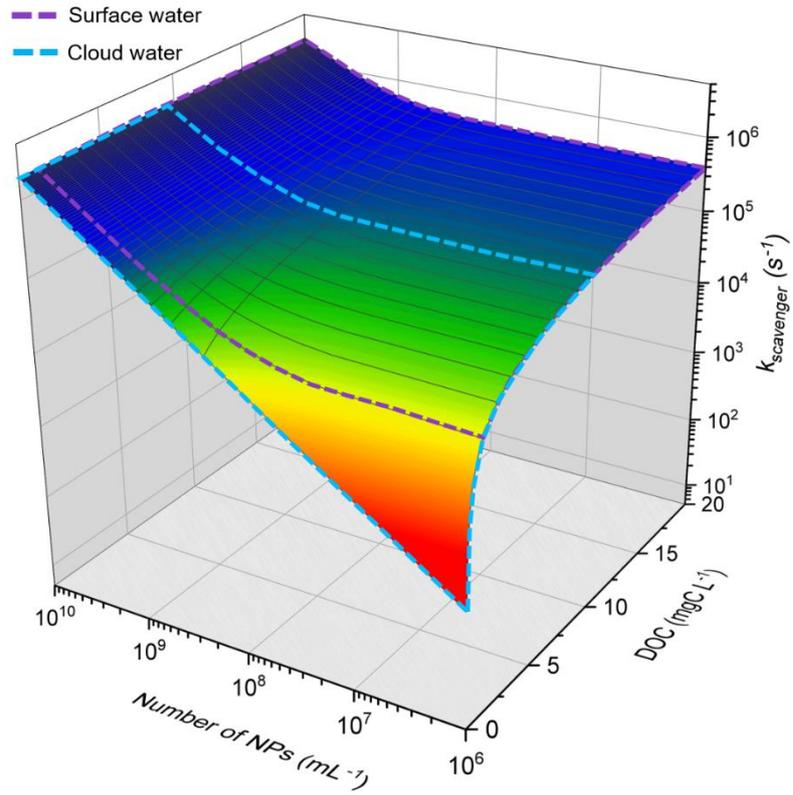
392



393

394 **Figure 3:** Proposed mechanism of degradation of PS with light and photogenerated hydroxyl radicals. Identified  
 395 compounds are framed in red.

396



397

398 **Figure 4:** Estimated rate constants ( $k_{\text{scav}}$ ) for the scavenging of  $\cdot\text{OH}$  by PS-NPs and DOC. DOC concentration is  
 399 representative of surface waters (higher than 1  $\text{mgC L}^{-1}$ ), depicted in the violet interval, and of cloud water  
 400 ( $0\text{-}10 \text{ mgC L}^{-1}$ ), in light blue.

401

402 **References**

- 403 Acampora, H., Berrow, S., Newton, S., O'Connor, I., 2017. Presence of plastic litter in pellets from  
404 Great Cormorant (*Phalacrocorax carbo*) in Ireland. *Mar. Pollut. Bull.* 117, 512–514.  
405 <https://doi.org/10.1016/j.marpolbul.2017.02.015>
- 406 Allen, S., Allen, D., Phoenix, V.R., Le Roux, G., Durántez Jiménez, P., Simonneau, A., Binet, S.,  
407 Galop, D., 2019. Atmospheric transport and deposition of microplastics in a remote mountain  
408 catchment. *Nat. Geosci.* 12, 339–344. <https://doi.org/10.1038/s41561-019-0335-5>
- 409 Anbumani, S., Kakkar, P., 2018. Ecotoxicological effects of microplastics on biota: a review.  
410 *Environ. Sci. Pollut. R.* 25, 14373–14396. <https://doi.org/10.1007/s11356-018-1999-x>
- 411 Arakaki, T., Anastasio, C., Kuroki, Y., Nakajima, H., Okada, K., Kotani, Y., Handa, D., Azechi, S.,  
412 Kimura, T., Tsuhako, A., Miyagi, Y., 2013. A general scavenging rate constant for reaction of  
413 hydroxyl radical with organic carbon in atmospheric waters. *Environ. Sci. Tech.* 47, 8196–8203.  
414 <https://doi.org/10.1021/es401927b>
- 415 Backhaus, T., Wagner, M., 2019. Microplastics in the Environment: Much Ado about Nothing? A  
416 Debate. *Global Challenges* 1900022. <https://doi.org/10.1002/gch2.201900022>
- 417 Bergmann, M., Mützel, S., Primpke, S., Tekman, M.B., Trachsel, J., Gerdt, G., 2019. White and  
418 wonderful? Microplastics prevail in snow from the Alps to the Arctic. *Sci. Adv.* 5, eaax1157.  
419 <https://doi.org/10.1126/sciadv.aax1157>
- 420 Besseling, E., Wang, B., Lüring, M., Koelmans, A.A., 2014. Nanoplastic affects growth of *S.*  
421 *obliquus* and reproduction of *D. magna*. *Environ. Sci. Technol.* 48, 12336–12343.  
422 <https://doi.org/10.1021/es503001d>
- 423 Bianco, A., Deguillaume, L., Vaïtilingom, M., Nicol, E., Baray, J.-L., Chaumerliac, N., Bridoux,  
424 M.C., 2018. Molecular characterization of cloud water samples collected at the puy de Dôme (France)  
425 by Fourier Transform Ion Cyclotron Resonance Mass Spectrometry. *Environ. Sci. Tech.*  
426 <https://doi.org/10.1021/acs.est.8b01964>
- 427 Bianco, A., Passananti, M., Perroux, H., Vyard, G., Mouchel-Vallon, C., Chaumerliac, N., Mailhot,  
428 G., Deguillaume, L., Brigante, M., 2015. A better understanding of hydroxyl radical photochemical  
429 sources in cloud waters collected at the puy de Dôme station – experimental versus modelled  
430 formation rates. *Atmos. Chem. Phys.* 15, 9191–9202. <https://doi.org/10.5194/acp-15-9191-2015>
- 431 Bittar, T.B., Berger, S.A., Birsa, L.M., Walters, T.L., Thompson, M.E., Spencer, R.G., Mann, E.L.,  
432 Stubbins, A., Frischer, M.E., Brandes, J.A., 2016. Seasonal dynamics of dissolved, particulate and  
433 microbial components of a tidal saltmarsh-dominated estuary under contrasting levels of freshwater  
434 discharge. *Estuar. Coast. Shelf. S.* 182, 72–85. <https://doi.org/10.1016/j.ecss.2016.08.046>
- 435 Bodrato, M., Vione, D., 2014. APEX (Aqueous Photochemistry of Environmentally occurring  
436 Xenobiotics): a free software tool to predict the kinetics of photochemical processes in surface waters.  
437 *Environ. Sci.: Processes Impacts* 16, 732–740. <https://doi.org/10.1039/C3EM00541K>
- 438 Buxton, G.V., Greenstock, C.L., Helman, W.P., Ross, A.B., 1988. Critical review of rate constants  
439 for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals ( $^{\bullet}\text{OH}/^{\bullet}\text{O}^-$  in aqueous  
440 solution. *J. Phys. Chem. Ref. Data* 17, 513–886. <https://doi.org/10.1063/1.555805>

- 441 Cai, L., Wang, J., Peng, J., Tan, Z., Zhan, Z., Tan, X., Chen, Q., 2017. Characteristic of microplastics  
442 in the atmospheric fallout from Dongguan city, China: preliminary research and first evidence.  
443 *Environ. Sci. Pollut. R.* 24, 24928–24935. <https://doi.org/10.1007/s11356-017-0116-x>
- 444 Calza, P., Vione, D., 2016. *Surface Water Photochemistry*, Comprehensive Series in Photochemical  
445 & Photobiological Sciences. The Royal Society of Chemistry.  
446 <https://doi.org/10.1039/9781782622154>
- 447 Chen, G., Feng, Q., Wang, J., 2020. Mini-review of microplastics in the atmosphere and their risks  
448 to humans. *Sci. Total Environ.* 703, 135504. <https://doi.org/10.1016/j.scitotenv.2019.135504>
- 449 Chen, R.F., Bissett, P., Coble, P., Conmy, R., Gardner, G.B., Moran, M.A., Wang, X., Wells, M.L.,  
450 Whelan, P., Zepp, R.G., 2004. Chromophoric dissolved organic matter (CDOM) source  
451 characterization in the Louisiana Bight. *Mar. Chem.* 89, 257–272.  
452 <https://doi.org/10.1016/j.marchem.2004.03.017>
- 453 Coble, P.G., 1996. Characterization of marine and terrestrial DOM in seawater using excitation-  
454 emission matrix spectroscopy. *Mar. Chem.* 51, 325–346. [https://doi.org/10.1016/0304-4203\(95\)00062-3](https://doi.org/10.1016/0304-4203(95)00062-3)
- 456 da Costa, J.P., Santos, P.S., Duarte, A.C., Rocha-Santos, T., 2016. (Nano) plastics in the  
457 environment—sources, fates and effects. *Sci. Total Environ.* 566, 15–26.  
458 <https://doi.org/10.1016/j.scitotenv.2016.05.041>
- 459 Davidson, M.R., Mitchell, S.A., Bradley, R.H., 2005. Surface studies of low molecular weight  
460 photolysis products from UV-ozone oxidised polystyrene. *Surf. Sci.* 581, 169–177.  
461 <https://doi.org/10.1016/j.susc.2005.02.049>
- 462 Dawson, A.L., Kawaguchi, S., King, C.K., Townsend, K.A., King, R., Huston, W.M., Nash, S.M.B.,  
463 2018. Turning microplastics into nanoplastics through digestive fragmentation by Antarctic krill. *Nat.*  
464 *Commun.* 9, 1001. <https://doi.org/10.1038/s41467-018-03465-9>
- 465 Deguillaume, L., Charbouillot, T., Joly, M., Vaïtilingom, M., Parazols, M., Marinoni, A., Amato, P.,  
466 Delort, A.-M., Vinatier, V., Flossmann, A., Chaumerliac, N., Pichon, J.M., Houdier, S., Laj, P.,  
467 Sellegri, K., Colomb, A., Brigante, M., Mailhot, G., 2014. Classification of clouds sampled at the puy  
468 de Dôme (France) based on 10 yr of monitoring of their physicochemical properties. *Atmos. Chem.*  
469 *Phys.* 14, 1485–1506. <https://doi.org/10.5194/acp-14-1485-2014>
- 470 Del Castillo, C.E., Coble, P.G., Morell, J.M., Lopez, J.M., Corredor, J.E., 1999. Analysis of the  
471 optical properties of the Orinoco River plume by absorption and fluorescence spectroscopy. *Mar.*  
472 *Chem.* 66, 35–51. [https://doi.org/10.1016/S0304-4203\(99\)00023-7](https://doi.org/10.1016/S0304-4203(99)00023-7)
- 473 Dris, R., Gasperi, J., Mirande, C., Mandin, C., Guerrouache, M., Langlois, V., Tassin, B., 2017. A  
474 first overview of textile fibers, including microplastics, in indoor and outdoor environments. *Environ.*  
475 *Pollut.* 221, 453–458. <https://doi.org/10.1016/j.envpol.2016.12.013>
- 476 Dris, R., Gasperi, J., Rocher, V., Tassin, B., 2018. Synthetic and non-synthetic anthropogenic fibers  
477 in a river under the impact of Paris Megacity: Sampling methodological aspects and flux estimations.  
478 *Sci. Total Environ.* 618, 157–164. <https://doi.org/10.1016/j.scitotenv.2017.11.009>
- 479 Dris, R., Gasperi, J., Saad, M., Mirande, C., Tassin, B., 2016. Synthetic fibers in atmospheric fallout:  
480 a source of microplastics in the environment? *Mar. Pollut. Bull.* 104, 290–293.  
481 <https://doi.org/10.1016/j.marpolbul.2016.01.006>

482 Dubinenkov, I., Flerus, R., Schmitt-Kopplin, P., Kattner, G., Koch, B.P., 2015. Origin-specific  
483 molecular signatures of dissolved organic matter in the Lena Delta. *Biogeochemistry* 123, 1–14.  
484 <https://doi.org/10.1007/s10533-014-0049-0>

485 Ervens, B., 2015. Modeling the processing of aerosol and trace gases in clouds and fogs. *Chem. Rev.*  
486 115, 4157–4198. <https://doi.org/10.1021/cr5005887>

487 Faust, B.C., 1994. Photochemistry of clouds, fogs, and aerosols. *Environ. Sci. Tech.* 28, 216A-222A.  
488 <https://doi.org/10.1021/es00054a001>

489 Faust, B.C., Allen, J.M., 1993. Aqueous-phase photochemical formation of hydroxyl radical in  
490 authentic cloudwaters and fogwaters. *Environ. Sci. Tech.* 27, 1221–1224.  
491 <https://doi.org/10.1021/es00043a024>

492 Frias, J., Nash, R., 2019. Microplastics: Finding a consensus on the definition. *Mar. Pollut. Bull.* 138,  
493 145–147. <https://doi.org/10.1016/j.marpolbul.2018.11.022>

494 Galgani, L., Engel, A., 2016. Changes in optical characteristics of surface microlayers hint to  
495 photochemically and microbially mediated DOM turnover in the upwelling region off the coast of  
496 Peru. *Biogeosciences* 13, 2453–2473. <https://doi.org/10.5194/bg-13-2453-2016>

497 Gallego-Urrea, J.A., Tuoriniemi, J., Pallander, T., Hassellöv, M., 2010. Measurements of  
498 nanoparticle number concentrations and size distributions in contrasting aquatic environments using  
499 nanoparticle tracking analysis. *Environ. Chem.* 7, 67. <https://doi.org/10.1071/EN09114>

500 Ganguly, M., Ariya, P.A., 2019. Ice nucleation of model nanoplastics and microplastics: a novel  
501 synthetic protocol and the influence of particle capping at diverse atmospheric environments. *ACS*  
502 *Earth Space Chem.* 3, 1729–1739. <https://doi.org/10.1021/acsearthspacechem.9b00132>

503 Gardette, J.-L., Mailhot, B., Lemaire, J., 1995. Photooxidation mechanisms of styrenic polymers.  
504 *Polym. Degrad. Stabil.*, 48(3), 457–470. [https://doi.org/10.1016/0141-3910\(95\)00113-Z](https://doi.org/10.1016/0141-3910(95)00113-Z)

505 Gasperi, J., Wright, S.L., Dris, R., Collard, F., Mandin, C., Guerrouache, M., Langlois, V., Kelly,  
506 F.J., Tassin, B., 2018. Microplastics in air: are we breathing it in? *Current Opinion in Environmental*  
507 *Science & Health* 1, 1–5. <https://doi.org/10.1016/j.coesh.2017.10.002>

508 Gewert, B., Plassmann, M.M., MacLeod, M., 2015. Pathways for degradation of plastic polymers  
509 floating in the marine environment. *Environ. Sci.: Processes Impacts* 17, 1513–1521.  
510 <https://doi.org/10.1039/C5EM00207A>

511 Ghigo, G., Vione, D., Berto, S., 2019. Experimental and theoretical study of the fluorescence  
512 emission of ferulic acid: Possible insights into the fluorescence properties of humic substances.  
513 *Spectrochim. Acta A* 117587. <https://doi.org/10.1016/j.saa.2019.117587>

514 Gligorovski, S., Strekowski, R., Barbati, S., Vione, D., 2015. Environmental Implications of  
515 Hydroxyl Radicals ( $\cdot\text{OH}$ ). *Chem. Rev.* 115, 13051–13092. <https://doi.org/10.1021/cr500310b>

516 Gomiero, A. (Ed.), 2019. *Plastics in the Environment*. IntechOpen.  
517 <https://doi.org/10.5772/intechopen.75849>

518 Göpferich, A., 1996. Mechanisms of polymer degradation and erosion. *Biomaterials* 17, 103–114.  
519 [https://doi.org/10.1016/0142-9612\(96\)85755-3](https://doi.org/10.1016/0142-9612(96)85755-3)

- 520 Han, M., Niu, X., Tang, M., Zhang, B.-T., Wang, G., Yue, W., Kong, X., Zhu, J., 2019. Distribution  
521 of microplastics in surface water of the lower Yellow River near estuary. *Sci. Total Environ.* 135601.  
522 <https://doi.org/10.1016/j.scitotenv.2019.135601>
- 523 Herckes, P., Lee, T., Trenary, L., Kang, G., Chang, H., Collett, J.L., 2002. Organic matter in central  
524 California radiation fogs. *Environ. Sci. Tech.* 36, 4777–4782. <https://doi.org/10.1021/es025889t>
- 525 Herckes, P., Valsaraj, K.T., Collett, J.L., 2013. A review of observations of organic matter in fogs  
526 and clouds: Origin, processing and fate. *Atmos. Res.* 132–133, 434–449.  
527 <https://doi.org/10.1016/j.atmosres.2013.06.005>
- 528 Herrmann, H., Ervens, B., Jacobi, H.-W., Wolke, R., Nowacki, P., Zellner, R., 2000. CAPRAM2.3:  
529 A chemical aqueous phase radical mechanism for tropospheric chemistry. *J. Atmos. Chem.* 36, 231–  
530 284. <https://doi.org/10.1023/A:1006318622743>
- 531 Holbrook, R.D., Yen, J.H., Grizzard, T.J., 2006. Characterizing natural organic material from the  
532 Occoquan Watershed (Northern Virginia, US) using fluorescence spectroscopy and PARAFAC. *Sci.*  
533 *Total Environ.* 361, 249–266. <https://doi.org/10.1016/j.scitotenv.2005.11.020>
- 534 Jeong, C.-B., Kang, H.-M., Lee, Y.H., Kim, M.-S., Lee, Jin-Sol, Seo, J.S., Wang, M., Lee, Jae-Seong,  
535 2018. Nanoplastic ingestion enhances toxicity of persistent organic pollutants (POPs) in the  
536 Monogonont Rotifer *Brachionus koreanus* via multixenobiotic resistance (MXR) disruption. *Environ.*  
537 *Sci. Technol.* 52, 11411–11418. <https://doi.org/10.1021/acs.est.8b03211>
- 538 Kangasluoma, J., Attoui, M., Korhonen, F., Ahonen, L., Siivola, E., Petäjä, T., 2016. Characterization  
539 of a Herrmann-type high-resolution differential mobility analyzer. *Aerosol Sci. Tech.* 50, 222–229.  
540 <https://doi.org/10.1080/02786826.2016.1142065>
- 541 Klein, M., Fischer, E.K., 2019. Microplastic abundance in atmospheric deposition within the  
542 Metropolitan area of Hamburg, Germany. *Sci. Total Environ.* 685, 96–103.  
543 <https://doi.org/10.1016/j.scitotenv.2019.05.405>
- 544 Koelmans, A.A., Mohamed Nor, N.H., Hermsen, E., Kooi, M., Mintenig, S.M., De France, J., 2019.  
545 Microplastics in freshwaters and drinking water: Critical review and assessment of data quality.  
546 *Water Res.* 155, 410–422. <https://doi.org/10.1016/j.watres.2019.02.054>
- 547 Lallement, A., Vinatier, V., Brigante, M., Deguillaume, L., Delort, A.M., Mailhot, G., 2018. First  
548 evaluation of the effect of microorganisms on steady state hydroxyl radical concentrations in  
549 atmospheric waters. *Chemosphere* 212, 715–722.  
550 <https://doi.org/10.1016/j.chemosphere.2018.08.128>
- 551 Lambert, S., Sinclair, C.J., Bradley, E.L., Boxall, A.B., 2013. Effects of environmental conditions on  
552 latex degradation in aquatic systems. *Sci. Total Environ.* 447, 225–234.  
553 <https://doi.org/10.1016/j.scitotenv.2012.12.067>
- 554 Lebreton, L., Slat, B., Ferrari, F., Sainte-Rose, B., Aitken, J., Marthouse, R., Hajbane, S., Cunsolo,  
555 S., Schwarz, A., Levivier, A., 2018. Evidence that the Great Pacific Garbage Patch is rapidly  
556 accumulating plastic. *Sci. Rep.* 8, 4666. <https://doi.org/10.1038/s41598-018-22939-w>
- 557 Lee, W.S., Cho, H.-J., Kim, E., Huh, Y.H., Kim, H.-J., Kim, B., Kang, T., Lee, J.-S., Jeong, J., 2019.  
558 Bioaccumulation of polystyrene nanoplastics and their effect on the toxicity of Au ions in zebrafish  
559 embryos. *Nanoscale* 11, 3173–3185. <https://doi.org/10.1039/C8NR09321K>

- 560 Lehner, R., Weder, C., Petri-Fink, A., Rothen-Rutishauser, B., 2019. Emergence of nanoplastic in the  
561 environment and possible impact on human health. *Environ. Sci. Technol.* 53, 1748–1765.  
562 <https://doi.org/10.1021/acs.est.8b05512>
- 563 Li, C., Busquets, R., Campos, L.C., 2019. Assessment of microplastics in freshwater systems: A  
564 review. *Sci. Total Environ.* 135578. <https://doi.org/10.1016/j.scitotenv.2019.135578>
- 565 Li, Y., Barth, M.C., Patton, E.G., Steiner, A.L., 2017. Impact of In-Cloud Aqueous Processes on the  
566 Chemistry and Transport of Biogenic Volatile Organic Compounds: Impact of In-Cloud Processes  
567 on BVOC. *J. Geophys. Res. Atmos.* 122, 11,131–11,153. <https://doi.org/10.1002/2017JD026688>
- 568 Liu, K., Wu, T., Wang, X., Song, Z., Zong, C., Wei, N., Li, D., 2019. Consistent Transport of  
569 Terrestrial Microplastics to the Ocean through Atmosphere. *Environ. Sci. Technol.* 53, 10612–10619.  
570 <https://doi.org/10.1021/acs.est.9b03427>
- 571 Massicotte, P., Asmala, E., Stedmon, C., Markager, S., 2017. Global distribution of dissolved organic  
572 matter along the aquatic continuum: Across rivers, lakes and oceans. *Sci. Total Environ.* 609, 180–  
573 191. <https://doi.org/10.1016/j.scitotenv.2017.07.076>
- 574 Mattsson, K., Jovic, S., Doverbratt, I., Hansson, L.-A., 2018. Nanoplastics in the aquatic environment,  
575 in: *Microplastic Contamination in Aquatic Environments*. Elsevier, pp. 379–399.  
576 <https://doi.org/10.1016/B978-0-12-813747-5.00013-8>
- 577 Mintenig, S.M., Bäuerlein, P.S., Koelmans, A.A., Dekker, S.C., van Wezel, A.P., 2018. Closing the  
578 gap between small and smaller: towards a framework to analyse nano- and microplastics in aqueous  
579 environmental samples. *Environ. Sci.: Nano* 5, 1640–1649. <https://doi.org/10.1039/C8EN00186C>
- 580 Mopper, K., Zhou, X., 1990. Hydroxyl radical photoproduction in the sea and its potential impact on  
581 marine processes. *Science* 250, 661–664. <https://doi.org/10.1126/science.250.4981.661>
- 582 Mouchel-Vallon, C., Deguillaume, L., Monod, A., Perroux, H., Rose, C., Ghigo, G., Long, Y.,  
583 Leriche, M., Aumont, B., Patryl, L., Armand, P., Chaumerliac, N., 2017. CLEPS 1.0: A new protocol  
584 for cloud aqueous phase oxidation of VOC mechanisms. *Geosci. Model Dev.* 10, 1339–1362.  
585 <https://doi.org/10.5194/gmd-10-1339-2017>
- 586 Muller, C.L., Baker, A., Hutchinson, R., Fairchild, I.J., Kidd, C., 2008. Analysis of rainwater  
587 dissolved organic carbon compounds using fluorescence spectrophotometry. *Atmos. Environ.* 42,  
588 8036–8045. <https://doi.org/10.1016/j.atmosenv.2008.06.042>
- 589 Neta, P., Dorfman, L.M., 1968. Pulse radiolysis studies. XIII. Rate constants for the reaction of  
590 hydroxyl radicals with aromatic compounds in aqueous solutions, *Radiation Chem.* 81, 222–230.  
591 <https://doi.org/10.1021/ba-1968-0081.ch015>
- 592 O’Brine, T., Thompson, R.C., 2010. Degradation of plastic carrier bags in the marine environment.  
593 *Mar. Pollut. Bull.* 60, 2279–2283. <https://doi.org/10.1016/j.marpolbul.2010.08.005>
- 594 Olasehinde, E.F., 2012. Determination of hydroxyl radical in Seto inland sea and its potential to  
595 degrade irgarol. *IOSRJAC* 1, 07–14. <https://doi.org/10.9790/5736-0150714>
- 596 Passananti, M., Zapadinsky, E., Zanca, T., Kangasluoma, J., Myllys, N., Rissanen, M.P., Kurtén, T.,  
597 Ehn, M., Attoui, M., Vehkamäki, H., 2019. How well can we predict cluster fragmentation inside a  
598 mass spectrometer? *Chem. Commun.* 55, 5946–5949. <https://doi.org/10.1039/C9CC02896J>

- 599 PlasticsEurope, 2019. Plastics the facts - 2018. Retrieved from  
600 <https://www.plasticseurope.org/en/resources/publications/619-plastics-facts-2018>
- 601 Scheurer, M., Bigalke, M., 2018. Microplastics in Swiss floodplain soils. *Environ. Sci. Technol.* 52,  
602 3591–3598. <https://doi.org/10.1021/acs.est.7b06003>
- 603 Schwaferts, C., Niessner, R., Elsner, M., Ivleva, N.P., 2019. Methods for the analysis of  
604 submicrometer- and nanoplastic particles in the environment. *TrAC Trend Anal. Chem.* 112, 52–65.  
605 <https://doi.org/10.1016/j.trac.2018.12.014>
- 606 Singh, B., Sharma, N., 2008. Mechanistic implications of plastic degradation. *Polym. Degrad. Stabil.*  
607 93, 561–584. <https://doi.org/10.1016/j.polymdegradstab.2007.11.008>
- 608 Stedmon, C.A., Cory, R.M., 2014. Biological origins and fate of fluorescent dissolved organic matter  
609 in aquatic environments. *Aquatic organic matter fluorescence* 278–299.  
610 <https://doi.org/10.1017/CBO9781139045452.013>
- 611 Ter Halle, A., Jeanneau, L., Martignac, M., Jardé, E., Pedrono, B., Brach, L., Gigault, J., 2017.  
612 Nanoplastic in the North Atlantic Subtropical Gyre. *Environ. Sci. Technol.* 51, 23, 13689-13697.
- 613 Todd, P.A., Ong, X., Chou, L.M., 2010. Impacts of pollution on marine life in Southeast Asia.  
614 *Biodivers. Conserv.* 19, 1063–1082. <https://doi.org/10.1007/s10531-010-9778-0>
- 615 van Pinxteren, D., Fomba, K.W., Mertes, S., Müller, K., Spindler, G., Schneider, J., Lee, T., Collett,  
616 J.L., Herrmann, H., 2016. Cloud water composition during HCCT-2010: Scavenging efficiencies,  
617 solute concentrations, and droplet size dependence of inorganic ions and dissolved organic carbon.  
618 *Atmos. Chem. Phys.* 16, 3185–3205. <https://doi.org/10.5194/acp-16-3185-2016>
- 619 Vicente, J.S., Gejo, J.L., Rothenbacher, S., Sarojiniamma, S., Gogritchiani, E., Wörner, M., Kasper,  
620 G., Braun, A.M., 2009. Oxidation of polystyrene aerosols by VUV-photolysis and/or ozone.  
621 *Photochem. Photobiol. Sci.* 8, 944. <https://doi.org/10.1039/b902749a>
- 622 Vione, D., Bagnus, D., Maurino, V., Minero, C., 2010. Quantification of singlet oxygen and hydroxyl  
623 radicals upon UV irradiation of surface water. *Environ. Chem. Lett.* 8, 193–198.  
624 <https://doi.org/10.1007/s10311-009-0208-z>
- 625 Weir, N.A., 1978. Reactions of hydroxyl radicals with polystyrene. *Eur. Polym. J.* 14, 9–14.  
626 [https://doi.org/10.1016/0014-3057\(78\)90144-1](https://doi.org/10.1016/0014-3057(78)90144-1)
- 627 Wetzel, R.G., 2001. *Limnology: lake and river ecosystems.* gulf professional publishing.
- 628 Xu, S., Ma, J., Ji, R., Pan, K., Miao, A.-J., 2020. Microplastics in aquatic environments: Occurrence,  
629 accumulation, and biological effects. *Sci. Total Environ.* 703, 134699.  
630 <https://doi.org/10.1016/j.scitotenv.2019.134699>
- 631 Yang, R., He, Q., Wang, C., Sun, S., 2018. Surface modification of polystyrene microsphere using  
632 ozone treatment. *Ferroelectrics* 530, 130–135. <https://doi.org/10.1080/00150193.2018.1453200>
- 633 Yonkos, L.T., Friedel, E.A., Perez-Reyes, A.C., Ghosal, S., Arthur, C.D., 2014. Microplastics in four  
634 estuarine rivers in the Chesapeake Bay, U.S.A. *Environ. Sci. Technol.* 48, 14195–14202.  
635 <https://doi.org/10.1021/es5036317>
- 636 Yousif, E., Haddad, R., 2013. Photodegradation and photostabilization of polymers, especially  
637 polystyrene: review. *SpringerPlus* 2, 398. <https://doi.org/10.1186/2193-1801-2-398>

- 638 Zhang, D., Dougal, S., Yeganeh, M., 2000. Effects of UV irradiation and plasma treatment on a  
639 polystyrene surface studied by IR– visible sum frequency generation spectroscopy. *Langmuir* 16,  
640 4528–4532. <https://doi.org/10.1021/la991353i>
- 641 Zhou, X., Mopper, K., 1990. Determination of photochemically produced hydroxyl radicals in  
642 seawater and freshwater. *Mar. Chem.* 30, 71–88. [https://doi.org/10.1016/0304-4203\(90\)90062-H](https://doi.org/10.1016/0304-4203(90)90062-H)