

ADOPTED: 26 November 2020

doi: 10.2903/j.efsa.2020.6355

List of non-EU phytoplasmas of tuber-forming *Solanum* spp.

EFSA Panel on Plant Health (PLH),
Claude Bragard, Katharina Dehnen-Schmutz, Paolo Gonthier, Josep Anton Jaques Miret,
Annemarie Fejer Justesen, Alan MacLeod, Christer Sven Magnusson, Panagiotis Milonas,
Juan A Navas-Cortes, Stephen Parnell, Roel Potting, Philippe Lucien Reignault,
Hans-Hermann Thulke, Wopke Van der Werf, Antonio Vicent Civera, Jonathan Yuen,
Lucia Zappalà, Domenico Bosco, Michela Chiumenti, Francesco Di Serio, Luciana Galetto,
Cristina Marzachi, Marco Pautasso and Marie-Agnès Jacques

Abstract

Following a request from the European Commission, the EFSA Panel on Plant Health prepared a list of non-EU phytoplasmas of tuber-forming *Solanum* spp. A systematic literature review and search of databases identified 12 phytoplasmas infecting *S. tuberosum*. These phytoplasmas were assigned to three categories. The first group (a) consists of seven non-EU phytoplasmas, known to occur only outside the EU ('*Candidatus* Phytoplasma americanum', '*Ca. P. australiense*', '*Ca. P. fragariae*'-related strain (YN-169, YN-10G) and '*Ca. P. hispanicum*') or having only limited presence in the EU ('*Ca. P. aurantifolia*'-related strains, '*Ca. P. pruni*'-related strains and '*Ca. P. trifolii*'). The second group (b) consists of three phytoplasmas originally described or reported from the EU. The third group (c) consists of two phytoplasmas with substantial presence in the EU, whose presence in *S. tuberosum* is not fully supported by the available literature. Phytoplasmas of categories (b) and (c) were excluded at this stage from further categorisation efforts. Three phytoplasmas from category (a) ('*Ca. P. australiense*', '*Ca. P. hispanicum*' and '*Ca. P. trifolii*') were excluded from further categorisation, as a pest categorisation has already been performed by EFSA. Comments provided by the EU Member States were integrated in the opinion. The main uncertainties of this listing concern: the taxonomy, the geographic distribution and prevalence and host range. The following phytoplasmas considered as non-EU and whose presence in *S. tuberosum* is fully supported by literature (category (a)) are categorised by the Panel in a separate opinion: '*Ca. P. americanum*', '*Ca. P. fragariae*'-related strain (YN-169, YN-10G), '*Ca. P. aurantifolia*'-related strains and '*Ca. P. pruni*'-related strains.

© 2020 European Food Safety Authority. *EFSA Journal* published by John Wiley and Sons Ltd on behalf of European Food Safety Authority.

Keywords: Clover yellow edge, pest risk, plant health, plant pest, potato hair sprouts, Potato purple top, quarantine

Requestor: European Commission

Question number: EFSA-Q-2020-00732

Correspondence: alpha@efsa.europa.eu

Panel members: Claude Bragard, Katharina Dehnen-Schmutz, Francesco Di Serio, Paolo Gonthier, Marie-Agnès Jacques, Josep Anton Jaques Miret, Annemarie Fejer Justesen, Alan MacLeod, Christer Sven Magnusson, Panagiotis Milonas, Juan A Navas-Cortes, Stephen Parnell, Roel Potting, Philippe L Reignault, Hans-Hermann Thulke, Wopke Van der Werf, Antonio Vicent Civera, Jonathan Yuen and Lucia Zappalà.

Acknowledgements: This document was prepared in cooperation with the Istituto per la Protezione Sostenibile delle Piante, Consiglio Nazionale delle Ricerche (Italy) under the tasking grant (GP/EFSA/ALPHA/2017/02). The Panel thanks for the information provided to this scientific output: Franco Finelli (Phytosanitary Service, Italy). The Panel acknowledges Jan van der Wolf for his review of a previous draft and all European competent institutions, Member State bodies and other organisations that provided data for this scientific output.

Competing interests: In line with EFSA's policy on declarations of interest, Panel member Francesco Di Serio did not participate in the adoption of this scientific output.

Suggested citation: EFSA PLH Panel (EFSA Panel on Plant Health), Bragard C, Dehnen-Schmutz K, Gonthier P, Jaques Miret JA, Justesen AF, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Van der Werf W, Vicent Civera A, Yuen J, Zappalà L, Bosco D, Chiumenti M, Di Serio F, Galetto L, Marzachi C, Pautasso M and Jacques M-A, 2020. Scientific Opinion on the list of non-EU phytoplasmas of tuber-forming *Solanum* spp. EFSA Journal 2020;18(12):6355, 22 pp. <https://doi.org/10.2903/j.efsa.2020.6355>

ISSN: 1831-4732

© 2020 European Food Safety Authority. *EFSA Journal* published by John Wiley and Sons Ltd on behalf of European Food Safety Authority.

This is an open access article under the terms of the [Creative Commons Attribution-NoDerivs](https://creativecommons.org/licenses/by/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited and no modifications or adaptations are made.



The EFSA Journal is a publication of the European Food Safety Authority, an agency of the European Union.



Table of contents

Abstract.....	1
1. Introduction.....	4
1.1. Background and Terms of Reference as provided by the requestor.....	4
1.1.1. Background.....	4
1.1.2. Terms of Reference.....	4
1.1.2.1. Terms of Reference: Appendix 1.....	5
1.1.2.2. Terms of Reference: Appendix 2.....	6
1.1.2.3. Terms of Reference: Appendix 3.....	7
1.1.3. Interpretation of the Terms of Reference.....	8
2. Data and methodologies.....	9
2.1. Data.....	9
2.1.1. Literature search.....	9
2.1.2. Database search.....	9
2.2. Methodology.....	9
3. Listing of phytoplasmas.....	10
3.1. Phytoplasmas considered as non-EU.....	10
3.2. Phytoplasmas excluded from further categorisation in the frame of the present mandate.....	10
3.3. Uncertainties.....	10
4. Conclusions.....	10
References.....	11
Abbreviations.....	14
Appendix A – Non-EU phytoplasmas of <i>Solanum tuberosum</i>	15
Appendix B – Phytoplasmas of <i>Solanum tuberosum</i> excluded from further categorisation as they have substantial presence in the EU or are originally described or reported from the EU.....	19
Appendix C – Phytoplasmas of <i>Solanum tuberosum</i> excluded from further categorisation as their presence in the species is not fully supported by available literature.....	21
Annex A – List of phytoplasmas considered in the opinion.....	22

1. Introduction

1.1. Background and Terms of Reference as provided by the requestor

1.1.1. Background

Council Directive 2000/29/EC¹ on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community establishes the present European Union plant health regime. The Directive lays down the phytosanitary provisions and the control checks to be carried out at the place of origin on plants and plant products destined for the Union or to be moved within the Union. In the Directive's 2000/29/EC annexes, the list of harmful organisms (pests) whose introduction into or spread within the Union is prohibited, is detailed together with specific requirements for import or internal movement.

Following the evaluation of the plant health regime, the new basic plant health law, Regulation (EU) 2016/2031² on protective measures against pests of plants, was adopted on 26 October 2016 and will apply from 14 December 2019 onwards, repealing Directive 2000/29/EC. In line with the principles of the above mentioned legislation and the follow-up work of the secondary legislation for the listing of EU regulated pests, EFSA is requested to provide pest categorisations of the harmful organisms included in the annexes of Directive 2000/29/EC, in the cases where recent pest risk assessment/ pest categorisation is not available.

1.1.2. Terms of Reference

EFSA is requested, pursuant to Article 22(5.b) and Article 29(1) of Regulation (EC) No 178/2002³, to provide scientific opinion in the field of plant health.

EFSA is requested to prepare and deliver a pest categorisation (step 1 analysis) for each of the regulated pests included in the appendices of the annex to this mandate. The methodology and template of pest categorisation have already been developed in past mandates for the organisms listed in Annex II Part A Section II of Directive 2000/29/EC. The same methodology and outcome is expected for this work as well.

The list of the harmful organisms included in the annex to this mandate comprises 133 harmful organisms or groups. A pest categorisation is expected for these 133 pests or groups and the delivery of the work would be stepwise at regular intervals through the year as detailed below. First priority covers the harmful organisms included in Appendix 1, comprising pests from Annex II Part A Section I and Annex II Part B of Directive 2000/29/EC. The delivery of all pest categorisations for the pests included in Appendix 1 is June 2018. The second priority is the pests included in Appendix 2, comprising the group of *Cicadellidae* (non-EU) known to be vector of Pierce's disease (caused by *Xylella fastidiosa*), the group of *Tephritidae* (non-EU), the group of potato viruses and virus-like organisms, the group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L. and the group of *Margarodes* (non-EU species). The delivery of all pest categorisations for the pests included in Appendix 2 is end 2019. The pests included in Appendix 3 cover pests of Annex I part A section I and all pests categorisations should be delivered by end 2020.

For the above-mentioned groups, each covering a large number of pests, the pest categorisation will be performed for the group and not the individual harmful organisms listed under "such as" notation in the Annexes of the Directive 2000/29/EC. The criteria to be taken particularly under consideration for these cases, is the analysis of host pest combination, investigation of pathways, the damages occurring and the relevant impact.

Finally, as indicated in the text above, all references to 'non-European' should be avoided and replaced by 'non-EU' and refer to all territories with exception of the Union territories as defined in Article 1 point 3 of Regulation (EU) 2016/2031.

¹ Council Directive 2000/29/EC of 8 May 2000 on protective measures against the introduction into the Community of organisms harmful to plants or plant products and against their spread within the Community. OJ L 169/1, 10.7.2000, p. 1–112.

² Regulation (EU) 2016/2031 of the European Parliament of the Council of 26 October 2016 on protective measures against pests of plants. OJ L 317, 23.11.2016, p. 4–104.

³ Regulation (EC) No 178/2002 of the European Parliament and of the Council of 28 January 2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. OJ L 31/1, 1.2.2002, p. 1–24.

1.1.2.1. Terms of Reference: Appendix 1

List of harmful organisms for which a pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

Annex IIAI

(a) Insects, mites and nematodes, at all stages of their development

<i>Aleurocanthus</i> spp.	<i>Numonia pyrivorella</i> (Matsumura)
<i>Anthonomus bisignifer</i> (Schenkling)	<i>Oligonychus perditus</i> Pritchard and Baker
<i>Anthonomus signatus</i> (Say)	<i>Pissodes</i> spp. (non-EU)
<i>Aschistonyx eppoi</i> Inouye	<i>Scirtothrips aurantii</i> Faure
<i>Carposina niponensis</i> Walsingham	<i>Scirtothrips citri</i> (Moultext)
<i>Enarmonia packardi</i> (Zeller)	<i>Scolytidae</i> spp. (non-EU)
<i>Enarmonia prunivora</i> Walsh	<i>Scrobipalopsis solanivora</i> Povolny
<i>Grapholita inopinata</i> Heinrich	<i>Tachypterellus quadrigibbus</i> Say
<i>Hishomonus phycitis</i>	<i>Toxoptera citricida</i> Kirk.
<i>Leucaspis japonica</i> Ckll.	<i>Unaspis citri</i> Comstock
<i>Listronotus bonariensis</i> (Kuschel)	

(b) Bacteria

Citrus variegated chlorosis	<i>Xanthomonas campestris</i> pv. <i>oryzae</i> (Ishiyama)
<i>Erwinia stewartii</i> (Smith) Dye	Dye and pv. <i>oryzicola</i> (Fang. et al.) Dye

(c) Fungi

<i>Alternaria alternata</i> (Fr.) Keissler (non-EU pathogenic isolates)	<i>Elsinoe</i> spp. Bitanc. and Jenk. Mendes
<i>Anisogramma anomala</i> (Peck) E. Müller	<i>Fusarium oxysporum</i> f. sp. <i>albedinis</i> (Kilian and Maire) Gordon
<i>Apiosporina morbosa</i> (Schwein.) v. Arx	<i>Guignardia piricola</i> (Nosa) Yamamoto
<i>Ceratocystis virescens</i> (Davidson) Moreau	<i>Puccinia pittieriana</i> Hennings
<i>Cercoseptoria pini-densiflorae</i> (Hori and Nambu) Deighton	<i>Stegophora ulmea</i> (Schweinitz: Fries) Sydow & Sydow
<i>Cercospora angolensis</i> Carv. and Mendes	<i>Venturia nashicola</i> Tanaka and Yamamoto

(d) Virus and virus-like organisms

Beet curly top virus (non-EU isolates)	Citrus tristeza virus (non-EU isolates)
Black raspberry latent virus	Leprosis
Blight and blight-like	Little cherry pathogen (non- EU isolates)
Cadang-Cadang viroid	Naturally spreading psorosis
Palm lethal yellowing mycoplasma	Tatter leaf virus
Satsuma dwarf virus	Witches' broom (MLO)

Annex IIB

(a) Insect mites and nematodes, at all stages of their development

<i>Anthonomus grandis</i> (Boh.)	<i>Ips cembrae</i> Heer
<i>Cephalcia lariciphila</i> (Klug)	<i>Ips duplicatus</i> Sahlberg
<i>Dendroctonus micans</i> Kugelan	<i>Ips sexdentatus</i> Börner
<i>Gilpinia hercyniae</i> (Hartig)	<i>Ips typographus</i> Heer
<i>Gonipterus scutellatus</i> Gyll.	<i>Sternochetus mangiferae</i> Fabricius
<i>Ips amitinus</i> Eichhof	

(b) Bacteria

Curtobacterium flaccumfaciens pv. *flaccumfaciens*
(Hedges) Collins and Jones

(c) Fungi

Glomerella gossypii Edgerton

Hypoxyton mammatum (Wahl.) J. Miller

Gremmeniella abietina (Lag.) Morelet

1.1.2.2. Terms of Reference: Appendix 2

List of harmful organisms for which a pest categorisation is requested per group. The list below follows the categorisation included in the annexes of Directive 2000/29/EC.

Annex IAI**(a) Insects, mites and nematodes, at all stages of their development**

Group of Cicadellidae (non-EU) known to be vector of Pierce's disease (caused by *Xylella fastidiosa*), such as:

- | | |
|--|---|
| 1) <i>Carneocephala fulgida</i> Nottingham | 3) <i>Graphocephala atropunctata</i> (Signoret) |
| 2) <i>Draeculacephala minerva</i> Ball | |

Group of Tephritidae (non-EU) such as:

- | | |
|--|---|
| 1) <i>Anastrepha fraterculus</i> (Wiedemann) | 12) <i>Pardalaspis cyanescens</i> Bezzi |
| 2) <i>Anastrepha ludens</i> (Loew) | 13) <i>Pardalaspis quinaria</i> Bezzi |
| 3) <i>Anastrepha obliqua</i> Macquart | 14) <i>Pterandrus rosa</i> (Karsch) |
| 4) <i>Anastrepha suspensa</i> (Loew) | 15) <i>Rhacochlaena japonica</i> Ito |
| 5) <i>Dacus ciliatus</i> Loew | 16) <i>Rhagoletis completa</i> Cresson |
| 6) <i>Dacus curcurbitae</i> Coquillet | 17) <i>Rhagoletis fausta</i> (Osten-Sacken) |
| 7) <i>Dacus dorsalis</i> Hendel | 18) <i>Rhagoletis indifferens</i> Curran |
| 8) <i>Dacus tryoni</i> (Froggatt) | 19) <i>Rhagoletis mendax</i> Curran |
| 9) <i>Dacus tsuneonis</i> Miyake | 20) <i>Rhagoletis pomonella</i> Walsh |
| 10) <i>Dacus zonatus</i> Saund. | 21) <i>Rhagoletis suavis</i> (Loew) |
| 11) <i>Epochra canadensis</i> (Loew) | |

(c) Viruses and virus-like organisms

Group of potato viruses and virus-like organisms such as:

- | | |
|----------------------------------|--|
| 1) Andean potato latent virus | 5) Potato virus T |
| 2) Andean potato mottle virus | 6) non-EU isolates of potato viruses A, M, S, V, X and Y (including Yo, Yn and Yc) and Potato leafroll virus |
| 3) Arracacha virus B, oca strain | |
| 4) Potato black ringspot virus | |

Group of viruses and virus-like organisms of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L., such as:

- | | |
|--------------------------------------|--|
| 1) Blueberry leaf mottle virus | 8) Peach yellows mycoplasma |
| 2) Cherry rasp leaf virus (American) | 9) Plum line pattern virus (American) |
| 3) Peach mosaic virus (American) | 10) Raspberry leaf curl virus (American) |
| 4) Peach phony rickettsia | 11) Strawberry witches' broom mycoplasma |
| 5) Peach rosette mosaic virus | 12) Non-EU viruses and virus-like organisms of <i>Cydonia</i> Mill., <i>Fragaria</i> L., <i>Malus</i> Mill., <i>Prunus</i> L., <i>Pyrus</i> L., <i>Ribes</i> L., <i>Rubus</i> L. and <i>Vitis</i> L. |
| 6) Peach rosette mycoplasma | |
| 7) Peach X-disease mycoplasma | |

Annex IIAI

(a) Insects, mites and nematodes, at all stages of their development

Group of *Margarodes* (non-EU species) such as:

- 1) *Margarodes vitis* (Phillipi)
- 2) *Margarodes vredendalensis* de Klerk
- 3) *Margarodes prieskaensis* Jakubski

1.1.2.3. Terms of Reference: Appendix 3

List of harmful organisms for which a pest categorisation is requested. The list below follows the annexes of Directive 2000/29/EC.

Annex IAI

(a) Insects, mites and nematodes, at all stages of their development

<i>Acleris</i> spp. (non-EU)	<i>Longidorus diadecturus</i> Eveleigh and Allen
<i>Amauromyza maculosa</i> (Malloch)	<i>Monochamus</i> spp. (non-EU)
<i>Anomala orientalis</i> Waterhouse	<i>Myndus crudus</i> Van Duzee
<i>Arrhenodes minutus</i> Drury	<i>Nacobbus aberrans</i> (Thorne) Thorne and Allen
<i>Choristoneura</i> spp. (non-EU)	<i>Naupactus leucoloma</i> Boheman
<i>Conotrachelus nenuphar</i> (Herbst)	<i>Premnotrypes</i> spp. (non-EU)
<i>Dendrolimus sibiricus</i> Tschetverikov	<i>Pseudopityophthorus minutissimus</i> (Zimmermann)
<i>Diabrotica barberi</i> Smith and Lawrence	<i>Pseudopityophthorus pruinus</i> (Eichhoff)
<i>Diabrotica undecimpunctata howardi</i> Barber	<i>Scaphoideus luteolus</i> (Van Duzee)
<i>Diabrotica undecimpunctata undecimpunctata</i> Mannerheim	<i>Spodoptera eridania</i> (Cramer)
<i>Diabrotica virgifera zea</i> Krysan & Smith	<i>Spodoptera frugiperda</i> (Smith)
<i>Diaphorina citri</i> Kuway	<i>Spodoptera litura</i> (Fabricus)
<i>Heliothis zea</i> (Boddie)	<i>Thrips palmi</i> Karny
<i>Hirschmanniella</i> spp., other than <i>Hirschmanniella gracilis</i> (de Man) Luc and Goodey	<i>Xiphinema americanum</i> Cobb sensu lato (non-EU populations)
<i>Liriomyza sativae</i> Blanchard	<i>Xiphinema californicum</i> Lamberti and Bleve-Zacheo

(b) Fungi

<i>Ceratocystis fagacearum</i> (Bretz) Hunt	<i>Mycosphaerella larici-leptolepis</i> Ito et al.
<i>Chrysomyxa arctostaphyli</i> Dietel	<i>Mycosphaerella populorum</i> G. E. Thompson
<i>Cronartium</i> spp. (non-EU)	<i>Phoma andina</i> Turkensteen
<i>Endocronartium</i> spp. (non-EU)	<i>Phyllosticta solitaria</i> Ell. and Ev.
<i>Guignardia laricina</i> (Saw.) Yamamoto and Ito	<i>Septoria lycopersici</i> Speg. var. <i>malagutii</i> Ciccarone and Boerema
<i>Gymnosporangium</i> spp. (non-EU)	<i>Thecaphora solani</i> Barrus
<i>Inonotus weirii</i> (Murril) Kotlaba and Pouzar	<i>Trechispora brinkmannii</i> (Bresad.) Rogers
<i>Melampsora farlowii</i> (Arthur) Davis	

(c) Viruses and virus-like organisms

Tobacco ringspot virus	Pepper mild tigré virus
Tomato ringspot virus	Squash leaf curl virus
Bean golden mosaic virus	Euphorbia mosaic virus
Cowpea mild mottle virus	Florida tomato virus
Lettuce infectious yellows virus	

(d) Parasitic plants

Arceuthobium spp. (non-EU)

Annex I A II**(a) Insects, mites and nematodes, at all stages of their development**

Meloidogyne fallax Karssen

Rhizoecus hibisci Kawai and Takagi

Popillia japonica Newman

(b) Bacteria

Clavibacter michiganensis (Smith) Davis et al. ssp. *Ralstonia solanacearum* (Smith) Yabuuchi et al. *sepedonicus* (Spieckermann and Kotthoff) Davis et al.

(c) Fungi

Melampsora medusae Thümen

Synchytrium endobioticum (Schilbersky) Percival

Annex I B**(a) Insects, mites and nematodes, at all stages of their development**

Leptinotarsa decemlineata Say

Liriomyza bryoniae (Kaltenbach)

(b) Viruses and virus-like organisms

Beet necrotic yellow vein virus

1.1.3. Interpretation of the Terms of Reference

This opinion provides a list of non-EU phytoplasmas of tuber-forming *Solanum* spp., for which the EFSA Plant Health Panel (from now on: "the Panel") then conducted a pest categorisation in a separate opinion (EFSA PLH Panel et al., 2020b). This list is based on information collected from databases up to January 2020, as well as information received from EU Member States (MS) during the period April-June 2020.

The search conducted for this list made it clear that the only tuber-forming species of *Solanum* genus reported to be infected by phytoplasmas is *S. tuberosum*.

Non-EU phytoplasmas of *S. tuberosum* are pests listed in the Appendices to the Terms of Reference (ToR) to be subject to pest categorisation to determine whether they fulfil the criteria of quarantine pests or those of regulated non-quarantine pests for the area of the EU excluding Ceuta, Melilla and the outermost regions of MS referred to in Article 355(1) of the Treaty on the Functioning of the European Union (TFEU), other than Madeira and the Azores.

As a first step toward this goal, the Panel prepared a list of phytoplasmas infecting *S. tuberosum*. In the process, three groups of phytoplasmas were distinguished:

- a) non-EU phytoplasmas with presence in *S. tuberosum* fully supported by literature,
- b) phytoplasmas (affecting *S. tuberosum*) with widespread presence in the EU (known to occur in several MS, frequently reported in the EU, widespread in some MS) or originally described or reported from the EU, and
- c) phytoplasmas of category (b) but with presence in *S. tuberosum* not fully supported by the literature.

A non-EU phytoplasma is defined by its geographical origin outside of the EU. Therefore, phytoplasmas not reported from the EU and occurring only outside of the EU are considered as non-EU phytoplasmas. Likewise, phytoplasmas occurring outside the EU and having only a limited presence in the EU (reported in only one or few MSs, with restricted distribution) are also considered as non-EU phytoplasmas.

This opinion provides the methodology and results for this classification, thus preparing the ground for the pest categorisation linked to the present mandate (EFSA PLH Panel et al. 2020b). This means that the Panel then performed a pest categorisation for the non-EU phytoplasmas with confirmed

ability to infect *S. tuberosum*. The phytoplasmas with uncertain ability to infect *S. tuberosum* and the phytoplasmas with significant presence in the EU or originally described or reported from the EU are excluded from further categorisation efforts, unless this will be requested by the risk managers in the future.

In this opinion, to capture the broadest possible range of phytoplasmas, even the poorly characterised ones for which very partial molecular or biological data are available, were considered. As in some cases there is uncertainty about the 'Ca. P. species definition', related strains were considered if they infect *S. tuberosum*. Instead, phytoplasma-like diseases of unknown aetiology or caused by viruses and formerly associated to mycoplasma-like organisms (MLO) or by other graft-transmissible bacteria are not addressed in this opinion.

2. Data and methodologies

2.1. Data

2.1.1. Literature search

The literature considered to generate the list of phytoplasmas infecting *S. tuberosum* (see Section 1.1.3) and to fill in the extraction tables on their distribution (see Appendices A–C and Annex A) was obtained from expert knowledge and extensive literature searches performed in Web of Science (WoS, last access January 2020). The search in WoS was performed using as keywords: phytoplasma/mycoplasma/witch/spiroplasma combined with the scientific name of the genus OR the common name of the crop. Therefore, the search in WoS was performed according to the following strategy:

TOPIC:((Phytoplasma* OR mycoplasma* OR witch* OR spiroplasma*) AND (Solanum OR potato*))

All the references were screened by title, abstract and, if needed, full paper with the specific objective of selecting those providing additional information regarding distribution and host range of the phytoplasmas included in the list or not yet included.

Information on phytoplasma taxonomy was gathered from either the original reference to species description or IRPCM (International Research Programme on Comparative Mycoplasmaology) Phytoplasma/Spiroplasma Working Team–Phytoplasma Taxonomy Group (IRPCM, 2004).

Further references and data were obtained from experts, EU National Plant Protection Organisations and from citations within primary references.

2.1.2. Database search

Data on *S. tuberosum* as natural host and distribution of the phytoplasmas were retrieved from the EPPO Global Database (GD) (EPPO, 2020), the Centre for Agriculture and Biosciences International (CABI) Crop Protection Compendium (CABI, 2020) and relevant publications.

GenBank accessions referring to phytoplasmas were added.

2.2. Methodology

A preliminary list of phytoplasmas infecting *S. tuberosum* (see Section 1.1.3) was generated by screening for phytoplasma diseases of the species present in the EPPO Lists A1 and A2. Further, all phytoplasma diseases listed in the EPPO GD were also screened for their association with *S. tuberosum*. Finally, the relevant phytoplasmas resulting from the literature search in WoS (as previously described) were included in the list.

The collected information was used to fill an extraction table (Annex A) with data regarding the taxonomy, geographical distribution of each *S. tuberosum*-infecting phytoplasma and key references and sources used to obtain that information. Taxonomy and distribution are reported in the table using the following scheme:

- the taxonomy was reported according to the 'Ca. P. species' description, when available. Although phytoplasmas have not yet been cultivated *in vitro*, phylogenetic analyses based on various conserved genes have shown that they represent a distinct, monophyletic clade within the class Mollicutes. Phytoplasmas are therefore accommodated within the 'Candidatus Phytoplasma' genus. Within this genus, several subtaxa have been described to accommodate organisms sharing less than 97.5% similarity among their 16S rRNA gene sequences. Additional species are described to accommodate organisms that, despite their 16S rRNA gene sequence being > 97.5% similar to those of other 'Ca. Phytoplasma' species,

are characterised by distinctive biological, phytopathological and genetic properties. Conversely, some organisms, despite their 16S rRNA gene sequence being < 97.5% similar to that of any other 'Ca. Phytoplasma' species, are not presently described as *Candidatus* species, due to their poor overall characterisation (IRPCM, 2004). When a phytoplasma has not been classified yet, information on a tentative classification was included based on the original literature source in which the pathogen was reported; to facilitate data retrieval from the literature and available databases, also the 16S rRNA group and subgroups were reported.

- data on distribution and on *S. tuberosum* as natural host of phytoplasmas were first searched in EPPO (2020) and in CABI (2020). Whenever conclusive information was not identified in the two databases or the information retrieved was at odds with expert knowledge, or in the absence of any information, extensive literature searches according to the protocol reported in Section 2.1 were performed.

Because only the non-EU phytoplasmas were subject of further categorisation efforts in the frame of the present mandate, it was decided to have consultation phases with EU Member States (MS) so that they could provide additional input if necessary. The information provided by EU MS was then considered by the Panel to determine the non-EU phytoplasmas that were further categorised (Section 3.1). The phytoplasmas excluded from this group are referred to here as phytoplasmas excluded from further categorisation in the frame of the present mandate (Section 3.2).

3. Listing of phytoplasmas

3.1. Phytoplasmas considered as non-EU

The phytoplasmas considered as non-EU (Appendix A) belong to two subcategories:

- Phytoplasmas not known to be present in the EU ('Ca. P. americanum', 'Ca. P. australiense', 'Ca. P. fragariae'-related strain (YN-169, YN-10G), and 'Ca. P. hispanicum')
- Phytoplasmas known to be present outside the EU and with only limited presence (i.e. reported in only one or few MSs or known to have a restricted distribution) in the EU ('Ca. P. aurantifolia'-related strains, 'Ca. P. pruni'-related strains and 'Ca. P. trifolii').

These phytoplasmas are categorised in EFSA PLH Panel et al., (2020b), with the exception of 'Ca. P. australiense', 'Ca. P. hispanicum' and 'Ca. P. trifolii', for which a pest categorisation is already available (EFSA PLH Panel et al., 2020a).

3.2. Phytoplasmas excluded from further categorisation in the frame of the present mandate

The phytoplasmas excluded from further categorisation in the frame of the present mandate are listed in Appendices B and C. Phytoplasmas listed in Appendix B are originally described or reported from the EU. For the phytoplasmas listed in Appendix C, the ability to infect the host plants is not conclusively supported by the available literature.

3.3. Uncertainties

Uncertainties potentially affecting the current list of non-EU potato phytoplasmas include:

- The geographic distribution and prevalence of the phytoplasmas.
- The taxonomy and biological status of poorly characterised phytoplasmas.
- The ability to infect *S. tuberosum* for some phytoplasmas.

4. Conclusions

The Panel was requested by the European Commission to produce a categorisation of 133 harmful organisms or groups listed in annexes of Directive 2000/29/EC. One of the groups for which a categorisation was needed is non-EU phytoplasmas of tuber-forming *Solanum* spp. As a first step, a systematic approach identified 12 phytoplasmas reported to naturally infect *S. tuberosum* (Annex A).

Among these phytoplasmas, based on information on distribution and prevalence both inside and outside the EU, the Panel identified seven non-EU phytoplasmas, known to occur only outside the EU

or having only a limited presence in the EU (Appendix A). These phytoplasmas are categorised in EFSA PLH Panel et al. (2020b), with the exception of '*Ca. P. australiense*', '*Ca. P. hispanicum*' and '*Ca. P. trifolii*', for which a pest categorisation is already available (EFSA PLH Panel et al., 2020a).

The remaining five phytoplasmas (which have a substantial presence in the EU or are originally described or reported from the EU (Appendix B, three phytoplasmas), or whose ability to infect *S. tuberosum* is not fully confirmed by available literature (Appendix C, two phytoplasmas)) were not categorised within the current mandate. However, the European Commission may request EFSA to categorise some or all the phytoplasmas excluded from the present exercise.

The main uncertainties of this listing concern the taxonomy, geographic distribution and prevalence and the ability to infect *S. tuberosum* for some phytoplasmas.

References

- Alfaro-Fernandez A, Verdeguer M, Rodriguez-Leon F, Ibanez I, Hernandez D, Teresani GR, Bertolini E, Cambra M and Font MI, 2017. Search for reservoirs of '*Candidatus Liberibacter solanacearum*' and mollicutes in weeds associated with carrot and celery crops. *European Journal of Plant Pathology*, 147, 15–20. <https://doi.org/10.1007/s10658-016-0984-9>
- Arnaud G, Malembic-Maher S, Salar P, Bonnet P, Maixner M, Marcone C, Boudon-Padieu E and Foissac X, 2007. Multilocus sequence typing confirms the close genetic interrelatedness of three distinct flavescence doree phytoplasma strain clusters and group 16SrV phytoplasmas infecting grapevine and alder in Europe. *Applied and Environmental Microbiology*, 73, 4001–4010.
- Arocha Y, Antesana O, Montellano E, Franco P, Plata G and Jones P, 2007. '*Candidatus Phytoplasma lycopersici*', a phytoplasma associated with 'hoja de perejil' disease in Bolivia. *International Journal of Systematic and Evolutionary Microbiology*, 57, 1704–1710. <https://doi.org/10.1099/ijs.0.64851-0>
- Bertaccini A, Bellardi MG, Botti S, Paltrinieri S and Restuccia P, 2006. Phytoplasma infection in *Asclepias physocarpa*. *Acta Horticulturae*, 722, 349–354.
- Borroto Fernandez EG, Calari A, Hanzer V, Katinger H, Bertaccini A and Laimer M, 2007. Phytoplasma infected plants in Austrian forests: role as a reservoir? *Bulletin of Insectology*, 60, 391.
- CABI, 2020. Crop Protection Compendium. Available online: <https://www.cabi.org/> [Accessed: November 2020].
- Castillo Carrillo C, Paltrinieri S, Bustamante JB and Bertaccini A, 2018. Detection and molecular characterization of a 16SrI-F phytoplasma in potato showing purple top disease in Ecuador. *Australasian Plant Pathology*, 47, 311–315. <https://doi.org/10.1007/s13313-018-0557-9>
- Castro S and Romero J, 2002. The association of clover proliferation phytoplasma with stolbur disease of pepper in Spain. *Journal of Phytopathology*, 150, 25–29.
- Cheng MY, Dong JH, Lee IM, Bottner-Parker KD, Zhao Y, Davis RE, Laski PJ, Zhang ZK and McBeath JH, 2015. Group 16SrXII phytoplasma strains, including subgroup 16SrXII-E ('*Candidatus Phytoplasma fragariae*') and a new subgroup, 16SrXII-I, are associated with diseased potatoes (*Solanum tuberosum*) in the Yunnan and Inner Mongolia regions of China. *European Journal of Plant Pathology*, 142, 305–318. <https://doi.org/10.1007/s10658-015-0616-9>
- Cheng M, Dong J, Han C, Zhang Z and McBeath JH, 2019. First Report of Phytoplasma '*Candidatus Phytoplasma aurantifolia*' associated with purple top diseased potatoes (*Solanum tuberosum*) in Guangdong province, China. *Plant Disease*, 103, 1015. <https://doi.org/10.1094/pdis-04-18-0701-pdn>
- Davino S, Calari A, Davino M, Tessitori M, Bertaccini A and Bellardi MG, 2007. Virescence of ten weeks stock associated to phytoplasma infection in Sicily. *Bulletin of Insectology*, 60, 279–280.
- Davis RE, Dally EL, Gundersen DE, Lee IM and Habili N, 1997. "*Candidatus phytoplasma australiense*", a new phytoplasma taxon associated with Australian grapevine yellows. *International Journal of Systematic Bacteriology*, 47, 262–269.
- Davis RE, Zhao Y, Dally EL, Lee IM, Jomantiene R and Douglas SM, 2013. '*Candidatus Phytoplasma pruni*', a novel taxon associated with X-disease of stone fruits, *Prunus* spp.: multilocus characterization based on 16S rRNA, secY, and ribosomal protein genes. *International Journal of Systematic and Evolutionary Microbiology*, 63, 766. <https://doi.org/10.1099/ijs.0.041202-0>
- Davis RE, Harrison NA, Zhao Y, Wei W and Dally EL, 2016. '*Candidatus Phytoplasma hispanicum*', a novel taxon associated with Mexican periwinkle virescence disease of *Catharanthus roseus*. *International Journal of Systematic and Evolutionary Microbiology*, 66, 3463–3467.
- Dong J, Zhang D, Wang D, McBeath JH and Zhang Z, 2011. Potato virus and phytoplasma diseases in Yunnan, China. *Phytopathology*, 101, S44.
- EFSA PLH Panel (EFSA Panel on Plant Health), Bragard C, Dehnen-Schmutz K, Gonthier P, Miret JAJ, Fejer Justesen A, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Van der Werf W, Civera AV, Yuen J, Zappalà L, Bosco D, Chiumenti M, Di Serio F, Galetto L, Marzachi C, Pautasso M and Jacques M-A, 2020a. Pest categorisation of the non-EU phytoplasmas of *Cydonia* Mill., *Fragaria* L., *Malus* Mill., *Prunus* L., *Pyrus* L., *Ribes* L., *Rubus* L. and *Vitis* L. *EFSA Journal* 2020;18(1):5929, 97 pp. <https://doi.org/10.2903/j.efsa.2020.5929>

- EFSA PLH Panel (EFSA Panel on Plant Health), Bragard C, Dehnen-Schmutz K, Gonthier P, Miret JAJ, Fejer Justesen A, MacLeod A, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke H-H, Van der Werf W, Civera AV, Yuen J, Zappalà L, Bosco D, Chiumenti M, Di Serio F, Galetto L, Marzachi C, Pautasso M and Jacques M-A, 2020b. Pest categorisation of the non-EU phytoplasmas of tuber-forming *Solanum* spp. EFSA Journal 2020;18(1):6356, 59 pp. <https://doi.org/10.2903/j.efsa.2020.6356>
- EPPO (European and Mediterranean Plant Protection Organization), 2020. EPPO Global Database. Available online: <https://gd.eppo.int> [Accessed: November 2020]
- Faggioli F, Pasquini G, Lumia V, Campobasso G, Widmer TL and Quimby PC, 2004. Molecular identification of a new member of the clover proliferation Phytoplasma group (16SrVI) associated with *Centaurea solstitialis* virescence in Italy. European Journal of Plant Pathology, 110, 353–360.
- Fahmeed F, Rosete YA, Perez KA, Boa E and Lucas J, 2009. First report of '*Candidatus* Phytoplasma asteris' (Group 16SrI) infecting fruits and vegetables in Islamabad, Pakistan. Journal of Phytopathology, 157, 639–641. <https://doi.org/10.1111/j.1439-0434.2009.01549.x>
- Firrao G, Carraro L, Gobbi E and Locci R, 1996. Molecular characterization of a phytoplasma causing phyllody in clover and other herbaceous hosts in northern Italy. European Journal of Plant Pathology, 102, 817–822.
- Franova J, Spak J and Simkova M, 2013. First report of a 16SrIII-B subgroup phytoplasma associated with leaf reddening, virescence and phyllody of purple coneflower. European Journal of Plant Pathology, 136, 7–12.
- Girsova NV, Bottner-Parker KD, Bogoutdinov DZ, Meshkov YI, Mozhaeva KA, Kastalyeva TB and Lee IM, 2016. Diverse phytoplasmas associated with potato stolbur and other related potato diseases in Russia. European Journal of Plant Pathology, 145, 139–153. <https://doi.org/10.1007/s10658-015-0824-3>
- Granata G, Paltrinieri S, Botti S and Bertaccini A, 2006. Aetiology of *Opuntia ficus-indica* malformations and stunting disease. Annals of Applied Biology, 149, 317–325.
- Hiruki C and Wang K, 2004. Clover proliferation phytoplasma: '*Candidatus* Phytoplasma trifolii'. International Journal of Systematic and Evolutionary Microbiology, 54, 1349–1353.
- Hodgetts J, Chuquillangui C, Muller G, Arocha Y, Gamarra D, Pinillos O, Velit E, Lozada P, Boa E, Boonham N, Mumford R, Barker I and Dickinson M, 2009. Surveys reveal the occurrence of phytoplasmas in plants at different geographical locations in Peru. Annals of Applied Biology, 155, 15–27. <https://doi.org/10.1111/j.1744-7348.2009.00316.x>
- Hosseini P, Bahar N, Madani G and Zirak L, 2011. Molecular characterization of phytoplasmas associated with potato purple top disease in Iran. Journal of Phytopathology, 159, 241–246. <https://doi.org/10.1111/j.1439-0434.2010.01757.x>
- IRPCM (International Research Programme on Comparative Mycoplasmaology), 2004. '*Candidatus* Phytoplasma', a taxon for the wall-less, non-helical prokaryotes that colonize plant phloem and insects. International Journal of Systematic and Evolutionary Microbiology, 54, 1243–1255.
- Jomantiene R, Davis RE, Antoniuk L and Staniulis J, 2000. First report of phytoplasmas in soybean, alfalfa, and *Lupinus* sp. in Lithuania. Plant Disease, 84, 198.
- Jones P and Arocha Y, 2006. A natural infection of Hebe is associated with an isolate of '*Candidatus* Phytoplasma asteris' causing a yellowing and little-leaf disease in the UK. Plant Pathology, 55, 821.
- Lee IM, Gundersen-Rindal DE, Davis RE, Bottner KD, Marcone C and Seemuller E, 2004a. '*Candidatus* Phytoplasma asteris', a novel phytoplasma taxon associated with aster yellows and related diseases. International Journal of Systematic and Evolutionary Microbiology, 54, 1037–1048.
- Lee IM, Martini M, Marcone C and Zhu SF, 2004b. Classification of phytoplasma strains in the elm yellows group (16SrV) and proposal of '*Candidatus* Phytoplasma ulmi' for the phytoplasma associated with elm yellows. International Journal of Systematic and Evolutionary Microbiology, 54, 337–347. <https://doi.org/10.1099/ijs.0.02697-0>
- Lee IM, Bottner KD, Secor G and Rivera-Varas V, 2006. '*Candidatus* Phytoplasma americanum' a phytoplasma associated with a potato purple top wilt disease complex. International Journal of Systematic and Evolutionary Microbiology, 56, 1593–1597.
- Leyva-Lopez NE, Ochoa-Sanchez JC, Leal-Klevezas DS and Martinez-Soriano JP, 2002. Multiple phytoplasmas associated with potato diseases in Mexico. Canadian Journal of Microbiology, 48, 1062–1068. <https://doi.org/10.1139/w02-109>
- Lindner K, Haase NU, Roman M and Seemuller E, 2011. Impact of stolbur phytoplasmas on potato tuber texture and sugar content of selected potato cultivars. Potato Research, 54, 267–282. <https://doi.org/10.1007/s11540-011-9192-3>
- Longoria-Espinoza RM, Douriet-Gamez NR, Lopez-Meyer M, Quiroz-Figueroa F, Bueno-Ibarra M, Mendez-Lozano J, Santos-Cervantes ME, Felix-Gastelum R, Chavez-Medina JA and Leyva-Lopez NE, 2013. Differentially regulated genes in *Solanum tuberosum* in response to "Mexican potato purple top phytoplasma" infection. Physiological and Molecular Plant Pathology, 81, 33–44. <https://doi.org/10.1016/j.pmp.2012.10.001>
- Mehle N, Mermal S, Vidmar S, Marn MV, Dreo T and Dermastia M, 2018. First report of carrot infection with phytoplasmas in Slovenia. pp. 2–3.
- Mejia JF, Contaldo N, Paltrinieri S, Pardo JM, Rios CA, Alvarez E and Bertaccini A, 2011. Molecular detection and identification of group 16SrV and 16SrXII phytoplasmas associated with potatoes in Colombia. Bulletin of Insectology, 64, S97–S98.

- Nisbet C, Ross S, Monger WA, Highet F and Jeffries C, 2014. First report of '*Candidatus* Phytoplasma asteris' in commercial carrots in the United Kingdom. *New Disease Reports*, 30, 16.
- Omar AF, Aljmhah KA, Alshim AS and Perez-Lopez E, 2018. Potato purple top disease associated with the novel subgroup 16SrII-X phytoplasma. *International Journal of Systematic and Evolutionary Microbiology*, 68, 3678–3682. <https://doi.org/10.1099/ijsem.0.003033>
- Palermo S, Elekes M, Botti S, Ember I, Alma A, Orosz A, Bertaccini A and Kolber M, 2004. Presence of stolbur phytoplasma in Cixiidae in Hungarian vineyards. *Vitis*, 43, 201–203.
- Paltrinieri S and Bertaccini A, 2007. Detection of phytoplasmas in plantlets grown from different batches of seed-potatoes. *Bulletin of Insectology*, 60, 379–380.
- Paltrinieri S, Bertaccini A and Lugaresi C, 2008. Phytoplasmas in declining cherry plants. *Acta Horticulturae*, 781, 409–416.
- Parrella G, Paltrinieri S, Botti S and Bertaccini A, 2008. Molecular identification of phytoplasmas from virescent *Ranunculus* plants and from leafhoppers in Southern Italian crops. *Journal of Plant Pathology*, 90, 537–543.
- Pribylova J, Petrzik K and Spak J, 2009. The first detection of '*Candidatus* Phytoplasma trifolii' in *Rhododendron hybridum*. *European Journal of Plant Pathology*, 124, 181–185. <https://doi.org/10.1007/s10658-008-9391-1>
- Prota VA, Garau R, Paltrinieri S, Botti S, Nahdi S, Calari A, Sechi A and Bertaccini A, 2007. Molecular identification of phytoplasmas infecting myrtle plantations in Sardinia (Italy). *Bulletin of Insectology*, 60, 383–384.
- Quaglino F, Zhao Y, Casati P, Bulgari D, Bianco PA, Wei W and Davis RE, 2013. '*Candidatus* Phytoplasma solani', a novel taxon associated with stolbur-and bois noir-related diseases of plants. *International Journal of Systematic and Evolutionary Microbiology*, 63, 2879–2894.
- Radisek S, Ferant N, Jakse J and Javornik B, 2009. Identification of a phytoplasma from the aster yellows group infecting purple coneflower (*Echinacea purpurea*) in Slovenia. *Plant Pathology*, 58, 392. <https://doi.org/10.1111/j.1365-3059.2008.02005.x>
- Reeder R and Arocha Y, 2008. '*Candidatus* phytoplasma asteris' identified in *Senecio jacobaea* in the United Kingdom. *Plant Pathology*, 57, 769. <https://doi.org/10.1111/j.1365-3059.2008.01849.x>
- Romanazzi G, D'Ascenzo D and Murolo S, 2009. *Tussilago farfara*: a new natural host of stolbur phytoplasma. *Plant Pathology*, 58, 392. <https://doi.org/10.1111/j.1365-3059.2008.01994.x>
- Salem NM, Tahzima R, Abdeen AO, Bianco PA, Massart S, Goedeffroit T and De Jonghe K, 2019. First report of '*Candidatus* Phytoplasma aurantifolia'-related strains infecting potato (*Solanum tuberosum*) in Jordan. *Plant Disease*, 103, 1406. <https://doi.org/10.1094/pdis-04-18-0705-pdn>
- Samuitienė M, Jomantiene R, Valiūnas D, Navalinskienė M and Davis RE, 2007. Phytoplasma strains detected in ornamental plants in Lithuania. *Bulletin of Insectology*, 60, 137–138.
- Santos-Cervantes ME, Chavez-Medina JA, Acosta-Pardini J, Flores-Zamora GL, Mendez-Lozano J and Leyva-Lopez NE, 2010. Genetic diversity and geographical distribution of phytoplasmas associated with potato purple top disease in Mexico. *Plant Disease*, 94, 388–395. <https://doi.org/10.1094/pdis-94-4-0388>
- Seemuller E and Schneider B, 2004. '*Candidatus* Phytoplasma mali', '*Candidatus* Phytoplasma pyri' and '*Candidatus* Phytoplasma prunorum', the causal agents of apple proliferation, pear decline and European stone fruit yellows, respectively. *International Journal of Systematic and Evolutionary Microbiology*, 54, 1217–1226.
- Staniulis JB, Davis RE, Jomantiene R, Kalvelyte A and Dally EL, 2000. Single and mixed phytoplasma infections in phyllody- and dwarf-diseased clover plants in Lithuania. *Plant Disease*, 84, 1061–1066.
- Tiwari AK, Khan MS, Iqbal A, Chun SC and Priya M, 2013. Molecular identification of '*Candidatus* Phytoplasma asteris' (16SRI-B) associated with the little leaf disease of potato in India. *Journal of Plant Pathology*, 95, 662.
- Tolu G, Botti S, Garau R, Prota VA, Sechi A, Prota U and Bertaccini A, 2006. Identification of a 16SrII-E phytoplasma in *Calendula arvensis*, *Solanum nigrum*, and *Chenopodium* spp. *Plant Disease*, 90, 325–330.
- Urbonaite IL, Jomantiene R, Valiunas D and Davis RE, 2016. First report of '*Candidatus* Phytoplasma asteris' subgroup 16SrI-A associated with a disease of potato (*Solanum tuberosum*) in Lithuania. *Plant Disease*, 100, 207. <https://doi.org/10.1094/pdis-05-15-0575-pdn>
- Valiunas D, Staniulis J and Davis RE, 2006. '*Candidatus* Phytoplasma fragariae', a novel phytoplasma taxon discovered in yellows diseased strawberry, *Fragaria x ananassa*. *International Journal of Systematic and Evolutionary Microbiology*, 56, 277–281.
- Valiunas D, Samuitiene M, Rasomavicius V, Navalinskiene M, Staniulis J and Davis RE, 2007. Subgroup 16SrIII-F phytoplasma strains in an invasive plant, *Heracleum sosnowskyi*, and an ornamental, *Dictamnus albus*. *Journal of Plant Pathology*, 89, 137–140.
- White DT, Blackall LL, Scott PT and Walsh KB, 1998. Phylogenetic positions of phytoplasmas associated with dieback, yellow crinkle and mosaic diseases of papaya, and their proposed inclusion in '*Candidatus* Phytoplasma australiense' and a new taxon, '*Candidatus* Phytoplasma australasia'. *International Journal of Systematic Bacteriology*, 48, 941–951.
- Zambon Y, Canel A, Bertaccini A and Contaldo N, 2018. Molecular diversity of phytoplasmas associated with grapevine yellows disease in North-Eastern Italy. *Phytopathology*, 108, 206–214. <https://doi.org/10.1094/phyto-07-17-0253-r>

Abbreviations

Ca. P.	<i>Candidatus</i> Phytoplasma
CYE	Clover yellow edge
EPPO	European and Mediterranean Plant Protection Organization
GD	Global Database
IRPCM	International Research Programme on Comparative Mycoplasmaology
MS	Member State
PCR	Polymerase Chain Reaction
PHS	Potato hair sprouts
PHYPA	<i>Candidatus</i> Phytoplasma australasia
PHYPAE	<i>Candidatus</i> Phytoplasma americanum
PHYPAS	<i>Candidatus</i> Phytoplasma asteris
PHYPAU	<i>Candidatus</i> Phytoplasma australiense
PHYCFG	<i>Candidatus</i> Phytoplasma fragariae
PHYCMA	<i>Candidatus</i> Phytoplasma mali
PHYCTR	<i>Candidatus</i> Phytoplasma trifolii
PHYC07	<i>Candidatus</i> Phytoplasma hispanicum
PHYC19	Clover yellow edge phytoplasma
PHYC74	Alder yellows phytoplasma
PLH	Plant Health
PPT	Potato purple top
RFLP	Restriction Fragment Length Polymorphism
TFEU	Treaty on the Functioning of the European Union
ToR	Terms of Reference
WoS	Web of Science

Appendix A – Non-EU phytoplasmas of *Solanum tuberosum*

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	Reasoning for considering non-EU	Uncertainties	References
1	<i>Candidatus Phytoplasma americanum</i>	–	PHYPAE	XVIII	Not reported to be present in the EU	–	Species description: (Lee et al., 2006); <i>S. tuberosum</i> : (EPPO, 2020)
2	<i>Candidatus Phytoplasma aurantifolia</i>	GD32; St_JO_10, 14, 17; PPT-SA; Rus-343F; PPT-GTO29, PPT-GTO30, PPT-SINTV; Potato Huayao Survey 2; Potato hair sprouts, PHS	PHYPAE, PHYP01, PHYP39	II	Italian reports refer to few infected individuals; present in Greece, Portugal; present in EU neighbouring Countries	Cheng et al. (2019) (despite identification of the phytoplasma as belonging to 16SrII by sequencing identity, in silico RFLP, neighbour-joining phylogenetic, in the text it is named as 'Ca. P. australiense'); Omar et al. (2018), Girsova et al. (2016), Hodgetts et al. (2009), Leyva-Lopez et al. (2002) (unclear subgroup assignment); Paltrinieri and Bertaccini (2007) (12 nested PCR-positive plants over 600 asymptomatic seed potato plants in Italy, no accession numbers available); Parrella et al. (2008) (one batch of 10 <i>Empoasca decipiens</i> in Italy); Tolu et al. (2006) (3 plants from 3 species in Italy); Prota et al. (2007) (less than 20 <i>Myrtus communis</i> plants and possibly in mixed infection in Italy); Granata et al. (2006) (in two <i>Opuntia ficus-indica</i> plants in Italy); Davino et al., 2007 (in one <i>Matthiola incana</i> plant in Italy)	Species description: (White et al., 1998; IRPCM, 2004); Strain descriptions and <i>S. tuberosum</i> : 'GD32 (Cheng et al., 2019); St_JO_10, 14, 17 (Salem et al., 2019); PPT-SA (Omar et al., 2018); Rus-343F (Girsova et al., 2016); PPT-GTO29, PPT-GTO30, PPT-SINTV (Santos-Cervantes et al., 2010); Potato Huayao Survey 2 (Hodgetts et al., 2009); Potato hair sprouts, PHS (Leyva-Lopez et al., 2002) <i>Solanum tuberosum</i> in Italy (Paltrinieri and Bertaccini, 2007); <i>Empoasca decipiens</i> in Italy (Parrella et al., 2008); <i>Calendula arvensis</i> , <i>Solanum nigrum</i> , and <i>Chenopodium</i> spp. in Italy (Tolu et al., 2006); <i>Matthiola incana</i> in Italy (Davino et al., 2007)

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	Reasoning for considering non-EU	Uncertainties	References
3	<i>Candidatus Phytoplasma australiense</i>	–	PHYPAU	XII-B	Not reported to be present in the EU	–	Species description: (Davis et al., 1997); <i>S. tuberosum</i> : (EPPO, 2020)
4	<i>Candidatus Phytoplasma fragariae</i>	YN-169, YN-10G		XII	Not reported to be present in the EU	Cheng et al., 2015 (several strains ascribed to 16SrXII-I, YN-169, but not identical to each other, plus other 16SrXII strains not assigned to any subgroup, YN-10G)	Species description: (Cheng et al., 2015); <i>S. tuberosum</i> : (Dong et al., 2011; Cheng et al., 2015)
5	<i>Candidatus Phytoplasma hispanicum</i>	–	PHYP07	XIII	Not reported to be present in the EU	Strawberry multiplier disease phytoplasma (STRAWB1) [PHYP75] is classified as RNQP (Annex IV; updated 2019). The phytoplasma is a strain of <i>Ca. P. hispanicum</i> , and the latter is not known to be present in the EU (EFSA PLH Panel et al., 2020a)	Species description: (Davis et al., 2016); <i>S. tuberosum</i> : (Santos-Cervantes et al., 2010)

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	Reasoning for considering non-EU	Uncertainties	References
6	<i>Candidatus Phytoplasma pruni</i>	Clover yellow edge, CYE (Girsova et al., 2016); Potato purple top, AKpot7, MT117, AKpot6 (Davis et al., 2013); Potato purple top, PPT-COAFP, PPT-GTOP (Santos-Cervantes et al., 2010)	PHYP19 (CYE)	III-B (CYE); III-F (AKpot7); III-M (MT117); III-N (AKpot6); III-U (PPT-COAFP, PPT-GTOP)	In the EU reported in four MSs: Czech Republic (two reports), Italy (three reports), Hungary (one report), Lithuania (four reports)	The pest was reported: in eight symptomatic <i>Echinacea purpurea</i> (Franova et al., 2013) and eight <i>Trifolium</i> spp. plants in the Czech Republic (Franova et al., 2004); in less than 50 symptomatic weed samples (<i>Leucanthemum vulgare</i> , <i>Taraxacum officinale</i> and <i>Crepis biennis</i>) (Firrao et al., 1996), in three <i>Prunus</i> spp. (cherry) plants (Paltrinieri et al., 2008) and in an undefined number (few samples) of <i>Asclepias physocarpa</i> plants (Bertaccini et al., 2006) in Italy; in an undefined number of <i>Cirsium arvense</i> and <i>Convolvulus arvensis</i> (Palermo et al., 2004) in Hungary; in two <i>Trifolium</i> spp. plants and in mixed infections (Staniulis et al., 2000), in an undefined number of <i>Gaillardia</i> sp., <i>Dictamnus albus</i> (Samuitienė et al., 2007), <i>Heracleum sosnowskyi</i> , <i>Dictamnus albus</i> (Valiunas et al., 2007), <i>Glycine max</i> and <i>Lupinus</i> spp. (Jomantiene et al., 2000), in Lithuania	Species description: (Davis et al., 2013); Strain descriptions and <i>S. tuberosum</i> : CYE (Girsova et al., 2016); AKpot7, MT117, AKpot6 (Davis et al., 2013); PPT-COAFP, PPT-GTOP (Santos-Cervantes et al., 2010); CYE in Lithuania (Staniulis et al., 2000)

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	Reasoning for considering non-EU	Uncertainties	References
7	<i>Candidatus Phytoplasma trifolii</i>	–	PHYPTR	VI-A	Reports from EU MS refer to few infected plants, ranging from 1 to 28	Reports from EU MS refer to few infected plants (Castro and Romero, 2002; Faggioli et al., 2004; Borroto Fernandez et al., 2007; Pribylova et al., 2009; Alfaro-Fernandez et al., 2017; Zambon et al., 2018); unclear subgroup assignation (Girsova et al., 2016)	Species description: (Hiruki and Wang, 2004); <i>S. tuberosum</i> : (EPPO, 2020); <i>Vitis</i> in Italy (Zambon et al., 2018); <i>Centaurea solstitialis</i> in Italy (Faggioli et al., 2004); <i>Amaranthus blitoides</i> and <i>Setaria adhaerens</i> in Spain (Alfaro-Fernandez et al., 2017); <i>Capsicum annuum</i> in Spain (Castro and Romero, 2002); <i>Rhododendron</i> spp. in Czech Republic (Pribylova et al., 2009); <i>Vaccinium myrtillus</i> in Austria (Borroto Fernandez et al., 2007)

(1): Reference isolate of 'Candidatus Phytoplasma species' is indicated by '-'.

Appendix B – Phytoplasmas of *Solanum tuberosum* excluded from further categorisation as they have substantial presence in the EU or are originally described or reported from the EU

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	EU MS in which the pathogen has been reported	Non-EU European and neighbouring countries	Reasoning for not considering as non-EU	Uncertainties	References
8	<i>Candidatus Phytoplasma asteris</i>	–	PHYPAS	I	Germany, Hungary, Italy (Present widespread); Czech Republic, Spain (Present, restricted distribution); Belgium Denmark, France, Romania (Present, no details); Lithuania; Slovenia ⁽²⁾	Russia (Present, restricted distribution; Belarus (Present, no details); UK ⁽²⁾	Reported in the EU (several MS)	–	Species description: (Lee et al., 2004a); <i>S. tuberosum</i> : (Lee et al., 2006; Arocha et al., 2007; Fahmeed et al., 2009; Hodgetts et al., 2009; Dong et al., 2011; Hosseini et al., 2011; Longoria-Espinoza et al., 2013; Tiwari et al., 2013; Girsova et al., 2016; Castillo Carrillo et al., 2018); <i>S. tuberosum</i> in Italy: (Paltrinieri and Bertaccini, 2007); <i>S. tuberosum</i> in Lithuania (Urbonaite et al., 2016); UK (Jones and Arocha, 2006; Reeder and Arocha, 2008; Nisbet et al., 2014), Slovenia: (Radisek et al., 2009; Romanazzi et al., 2009; Mehle et al., 2018)
9	<i>Candidatus Phytoplasma fragariae</i>	–	PHYPPFG	XII-E	Slovenia (EPPO report 2018/085); Belgium ⁽²⁾	UK (EPPO report 2015/031)	Originally described in the EU	-	Species description: (Valiunas et al., 2006)

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	EU MS in which the pathogen has been reported	Non-EU European and neighbouring countries	Reasoning for not considering as non-EU	Uncertainties	References
10	<i>Candidatus Phytoplasma solani</i>	–	PHPSO	XII-A	Italy (Present, widespread); Bulgaria, Croatia, France, Germany, Greece, Hungary, Slovakia, Slovenia, Spain, (Present, restricted distribution); Austria, Czech Republic, Poland (Present, few occurrences); Romania; Belgium ⁽²⁾ ; Portugal ⁽²⁾	Macedonia, Montenegro (Present, widespread); Russia, Serbia, Switzerland, Turkey (Present, restricted distribution); Albania, Armenia, Azerbaijan, Bosnia and Herzegovina, Georgia, Ukraine (Present, no details)	Originally described in the EU (several MS)	–	Species description: (Quaglino et al., 2013); <i>S. tuberosum</i> : (EPPO, 2020); <i>S. tuberosum</i> in Romania: (Lindner et al., 2011)

(1): Reference isolate of 'Candidatus Phytoplasma species' is indicated by '–'.

(2): Information provided by MS during commenting phase.

Appendix C – Phytoplasmas of *Solanum tuberosum* excluded from further categorisation as their presence in the species is not fully supported by available literature

ID	Phytoplasma name	Related strain name ⁽¹⁾	Abbreviation (EPPO code)	16S rRNA	EU MS in which the pathogen has been reported	Non-EU European and neighbouring countries	Reasoning for not considering as non-EU	Uncertainties	References
11	<i>Candidatus Phytoplasma mali</i>	–	PHYPPMA	X	Czech Republic, Germany, Hungary, Italy, Slovakia, Slovenia (Present widespread); Austria, Belgium, Bulgaria, Croatia, Finland, France, Greece, Spain (Present, restricted distribution); Poland, Romania (Present, no details); Lithuania, Netherlands (Present, few occurrences)	Switzerland (Present widespread); Belarus, Norway, Serbia (Present, restricted distribution); Albania, Bosnia and Herzegovina, Moldova, Russia, Turkey, Ukraine (Present, no details)	Originally described in the EU (several MS)	Only one nested PCR positive plant from 600 seed potato asymptomatic plants, probably in mixed infections and no accession number of the isolate available (Paltrinieri and Bertaccini, 2007)	Species description: (Seemuller and Schneider, 2004); <i>S. tuberosum</i> in Italy: (Paltrinieri and Bertaccini, 2007)
12	<i>Unclassified</i>	Potato Colombia M/V	PHYPP74	V-C	France	–	Reported in the EU	Only one report from 8 potato plants, 4 in mixed infections with <i>Ca. P. solani</i> ; no accession number of the isolate available; taxonomic status uncertain within the 16SrV-C subgroup (Mejia et al., 2011)	Species description: (Lee et al., 2004b); Strain description and <i>S. tuberosum</i> : (Mejia et al., 2011); <i>Alnus</i> in France: (Arnaud et al., 2007)

(1): Reference isolate of '*Candidatus Phytoplasma species*' is indicated by '–'.

Annex A – List of phytoplasmas considered in the opinion

See Excel file in Supplementary Information online.