

AperTO - Archivio Istituzionale Open Access dell'Università di Torino

Disordered CYP11B2 Expression in Primary Aldosteronism

This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1796379> since 2021-08-10T14:27:43Z

Published version:

DOI:10.1055/s-0043-122238

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

Disordered CYP11B2 expression in Primary Aldosteronism

Celso E. Gomez-Sanchez^{1,2}, Maniselvan Kuppusamy^{1,2} Tracy Ann Williams^{3,4} and Martin Reincke³

Endocrine Section, G.V. (Sonny) Montgomery VA Medical Center¹ and University of Mississippi Medical Center², Jackson, MS, USA, Medizinische Klinik und Poliklinik IV, Klinikum der Ludwig-Maximilians-Universität München, Munich, Germany³, Division of Internal Medicine and Hypertension, Department of Medical Sciences, University of Turin, Turin, Italy⁴.

Address Correspondence to:

Celso E. Gomez-Sanchez, M.D.

Research Service

G.V. (Sonny) Montgomery VA Medical Center

1500 E. Woodrow Wilson Blvd

Jackson, MS 39216, USA

Tel 601 368 3844

Cgomez-sanchez@umc.edu

DISCLOSURES: Nothing to disclose.

ABSTRACT

Primary aldosteronism is the most common type of secondary hypertension affecting 6-10% of patients with primary hypertension. PA is mainly caused by unilateral hyperaldosteronism due to an aldosterone-producing adenoma, unilateral hyperplasia with or without micronodules or bilateral zona glomerulosa hyperplasias with or without macro or micronodules. The development of antibodies against the terminal enzyme of aldosterone biosynthesis (CYP11B2) has permitted the further characterization of normal adrenals and resected adrenals from patients with primary aldosteronism. Normal adrenals exhibit two different patterns of cellular expression of CYP11B2: young individuals display a relatively uniform expression of the enzyme throughout the *zona glomerulosa* while the adrenals of older individuals have dispersed CYP11B2-expressing cells but have more groups of cells called aldosterone-producing cell clusters. APAs exhibit different patterns of CYP11B2 staining that vary from uniform to homogeneous. There is also a proportion of cells within the APA that co-express different enzymes that are not normally co-expressed in normal individuals. Approximately 30% of patients with unilateral hyperaldosteronism do not have an APA, but either have an increased number of CYP11B2 expressing micronodules or hyperplasia of the zona glomerulosa.

INTRODUCTION

Primary aldosteronism (PA) is the most common form of secondary hypertension with a prevalence of 5-10% of patients with primary hypertension [1] and is associated with a significant increase in morbidity and mortality [2,3]. There are multiple forms of PA that present as sporadic cases and the two most common are aldosterone-producing adenomas (APA) and bilateral *zona glomerulosa* hyperplasia with micro- or with macronodules or with both micro- and macronodules, also called idiopathic hyperaldosteronism (IHA). Less common is unilateral *zona glomerulosa* hyperplasia with micro- and/or macronodules. Familial forms are rare and there are at least 4 different familial types of hyperaldosteronism which include Type 1 (also called glucocorticoid-suppressible aldosteronism) due to a crossover recombination of the promoter region and first exons of the *CYP11B1* gene and the late exons of the *CYP11B2* gene resulting in the production of aldosterone in the *zona fasciculata* (ZF) under ACTH control [4]. Type 2 is the most common form, but the genetic basis has not been elucidated, although there is a linkage to chromosome 7p22 in some families [5]. Type 3 is due to mutations in the *KCNJ5* gene encoding the GIRK4 potassium channel and alter the selectivity filter of the channel pore [6]. Type 4 is due to mutations in the *CACNA1H* gene encoding Cav3.2 a voltage activated calcium channel subunit [7].

The biosynthesis of aldosterone occurs in the adrenal *zona glomerulosa* (ZG) through a series of enzymatic reactions starting from cholesterol. Most of the enzymes involved in aldosterone biosynthesis are also expressed in the ZF but the terminal enzyme, CYP11B2, is only expressed in the ZG and CYP11B1 is only expressed in the ZF [8,9]. The expression and distinct distribution of these two enzymes is shared by multiple species including humans, rats, mice, hamsters and guinea pigs [8,10]. Some species, such as cows, sheep, pigs, dogs and bullfrogs, express only a single CYP11B enzyme [8,11]; despite this,, aldosterone biosynthesis is restricted to the ZG as in species with two distinctly distributed enzymes [12]. The

mechanisms by which aldosterone production is suppressed in the ZF in species with a single CYP11B enzyme is unclear. The human CYP11B2 and CYP11B1 are highly homologous at the DNA (95% in the coding region) and at the protein level (93%) [13]. The presence of CYP11B2 identifies the cells of the adrenal that produce aldosterone. In the human adrenal the ZF has two unique enzymes, the CYP17A1 and the CYP11B1 which are responsible for the synthesis of cortisol [8]. The first specific polyclonal antibodies against CYP11B1 and CYP11B2 were described by Nishimoto and were more suitable for low amplification immunohistochemistry [14]. Highly specific monoclonal antibodies were then described [15] and have been extensively used to define the immunohistochemistry of normal adrenals and of resected adrenals from patients with PA [16-19].

Zona glomerulosa in normal adrenals from rodents and humans. The rat adrenal has a clearly delineated zonation. The ZG (with CYP11B2 expression) comprises 4-6 layers of cells underneath the outer capsule that are separated from the ZF by a layer that comprises progenitor cells called the undifferentiated cell zone (without CYP11B1 or CYP11B2 expression) [20]. The number of cells in the ZG that express CYP11B2 depends on the degree and duration of stimulation by the renin-angiotensin system and on a normal salt diet about half the cells express CYP11B2 and they are scattered throughout the ZG [21]; a chronic low sodium diet increases the layers of cells and most cells express CYP11B2 [21]. The human adrenal does not have a similar clear-cut separation of the ZG and ZF and cells with CYP11B2 expression are present in scattered cells in the subcapsular region (Fig 1A) [22] and in clusters that have been called variously as aldosterone-producing cell clusters (APCC) [14,15], foci and megafoci depending on the size of the cell cluster [23] (Fig 1B). These clusters show strong, uniform immunoreactivity for CYP11B2 with a ZG morphology that extends into the ZF, with no expression of CYP11B1. Adrenals from individuals from 0-11 years show a clear layered zonation with CYP11B2 expression that occupies a significant portion of the ZG and in some

cases there is an unstained layer between ZG labeled CYP11B2 and ZF labeled CYP11B1 and no APCC are found [22]. This layered arrangement remodels with age with significant portions of the ZG displaying low CYP11B2 expression while the APCC numbers increase [22,24]. In rare cases a portion of the APCC toward the ZF show an apparent remodeling to CYP11B2 expressing cells with ZF phenotype [22]. This pattern has been found in some patients with PA [25], but no clinical data was available in this study of supposed normal individuals [22]. In addition, cells expressing CYP11B1, which define the ZF can reach the capsule in some areas [15].

Aldosterone-producing adenomas. A significant advance in the pathogenesis of APAs was the discovery of somatic mutations in the selectivity filter of the G protein activated inward rectifier potassium channel GIRK4 coded by the *KCNJ5* gene [6], which has been shown to be present in 35-70% of patients [26-28]. The higher percentage was found in individuals of east Asia [26,28,29]. Mutations in pumps including the sodium potassium ATPase alpha subunit 1 (*ATP1A1* gene), membrane calcium ATPase (*ATP2B3* gene) and the calcium channel subunit $Ca_v1.3$ (*CACNA1D* gene) were then described in other cases [7,30-32] and all together these mutations explain approximately 50-80% of cases of APA. Some cases of unilateral aldosterone hyperproduction have multiple nodules that express the CYP11B2 enzyme and can have different mutated channels or pumps within the same gland [16,33,34]. APCCs from normal adrenals have an incidence of *CACNA1D* and *ATP1A1* mutations as high as 30%, but APCCs with *KCNJ5* mutations have not been detected [17]. It is unclear if APCCs harboring the mutations can develop into an aldosterone-producing adenoma.

Immunohistochemistry in primary aldosteronism. Adrenal vein catheterization is used to determine which is the abnormal adrenal producing the excessive amount of aldosterone. In most cases, unilateral aldosterone production is produced by an APA usually greater than 0.7 cm in diameter that is visible by a computerized tomography scan. Many adrenals with a clear

adenoma frequently also have APCCs present in the hyperplastic ZG [35,36]. In 30% of cases a microadenoma which is not visible by imaging techniques [37], unilateral ZG hyperplasia with or without micro- or macro-nodules [37] and rare cases of aldosterone-producing carcinomas which are of larger size can occur [16].

Large or small APAs can have two different phenotypic cell characteristics, more common are those with clear cells containing lipid droplets similar to ZF-type cells whereas other have more compact cells similar to ZG-type cells and a mixture of both types [38]. Some studies have correlated the cell type with the somatic mutation present in the APA and those with clear ZF cells tend to have *KCNJ5* mutations while those with *ATP1A1*, *ATP2B3* and *CACNA1D* mutations tend to be of the ZG type phenotype [31,38,39]. However other studies have not confirmed these results and although many APA carrying a *KCNJ5* mutation have a ZF cell phenotype almost an equal number have a mixture of ZG and ZF type cells [27,38].

Aldosterone production in patients with larger adenomas is generally higher than in those patients with smaller adenomas [40]. In the study by Ono et al [40], the tumor area of the group of larger adenomas was 9 times greater than the group of smaller adenomas but plasma aldosterone concentrations were only 2.0-2.5 times increased in the group of patients with the larger APA. This indicated that aldosterone production per cell was much greater from smaller adenomas, a suggestion that was supported by the higher immunoreactivity of CYP11B2 observed in the smaller group of tumors[40]. The number of CYP11B2 immunoreactive cells in the larger adenomas was highly variable with some adenomas displaying a relatively uniform expression of CYP11B2 (Fig 1C) compared to a heterogeneous expression of the enzyme in others with many cells that were immunoreactive negative (Fig 1D). The immunoreactivity of other enzymes including the CYP17A1 was lower in the smaller adenomas [40]. Outcomes after adrenalectomy for patients with smaller or larger APAs were similar between the two groups [40].

Many APAs exhibit an intratumoral heterogeneity of expressed enzymes that are normally specific to a distinct zone of the adrenal. In a recent study using double and triple immunofluorescence staining of APAs with antibodies against CYP11B2, CYP11B1 and CYP17A1, Nakamura *et al* [41] demonstrated that there are cells co-expressing the CYP11B2 and CYP11B1 (2.1%), CYP11B2 and CYP17A1 (0.6%), CYP11B1 and CYP17A1 (0.6%) and a smaller number of triple immunoreactive stained cells (0.14%). However, the proportions of the different immunofluorescent mixed cells were highly variable between adenomas. The presence of cells that co-express the CYP11B2 and CYP17A1 probably explains the increased secretion of the hybrid steroids 18-hydroxycortisol and 18-oxocortisol [42].

The increased use of AVS has enabled the diagnosis of unilateral aldosterone production in image negative patients that occurs in about 30% of patients with unilateral PA [43,44]. Immunohistochemistry studies of the resected adrenals from 32 patients with PA operated due to unilateral (or in 6 cases bilateral) production of aldosterone studied using CYP11B2 staining showed that 19 of those with an adenoma showed CYP11B2 staining in the adenoma, 1 patient with an adenoma and 3 cases of bilateral production of aldosterone with a unilateral adenoma, the adenoma did not stained for the CYP11B2 but had had multiple APCCs and 2 specimens had multiple micronodules with diffuse ZG hyperplasia staining for the CYP11B2 enzyme [35]. Of the 9 patients without a tumor on CT, 6 had unilateral aldosterone production and 3 were bilateral. Of the unilateral aldosterone producers, 3 had a microadenoma and 1 had multiple micronodules staining for the CYP11B2 [35]. Of the 3 showing no tumors, but bilateral production of aldosterone 2 had multiple APCCs and 1 diffuse hyperplasia [35]. In a recent study of 25 adrenals with histopathology of cross-sectional image negative hyperaldosteronism they were classified into two types 13 had multiple adrenocortical micronodules and 12 had diffuse hyperplasia of the zona glomerulosa [16]. Somatic mutations of aldosterone-driver genes were detected in 81% of CYP11B2-positive micronodules with 65% had mutations of the

CACNA1D gene, 8% in the *KCNJ5* gene and 4% in the *ATP1A1* and *ATP2B3* genes, but no mutations were found in the CYP11B2-positive non-nodular areas [16].

The possibility that the origin of an APA is from further differentiation of an APCC was recently postulated through the finding of cases where cells from an APCC expressing the CYP11B2 changed morphologically from a compact cell phenotype characteristic of the ZG to a clear cell phenotype resembling ZF cells and these have been called possible APCC-to-APA transitional lesions some of which had *KCNJ5* mutations [25].

Patients with APA or those with unilateral production of aldosterone have been treated by unilateral adrenalectomy of the involved site with either cure or significant improvement of the hypertension and biochemical abnormalities of the PA and in fewer cases resulting in no improvement. As no standard criteria for defining surgical outcomes was accepted, a recent study aimed to create a consensus criteria for outcomes [45]. The PASO study involved an international panel of 31 experts from 28 centers using the Delphi method to reach consensus. Complete clinical success correcting the hypertension was obtained in 37% of 705 patients with wide variance (17-62%) and partial success in 47% (range of 35-66%). Complete biochemical success was seen in 94% of patients [45]. A distinction between an adenoma and a nodule is difficult histopathologically. The frequent presence of APCCs and complete contralateral suppression of aldosterone production in the contralateral adrenal led us before to postulate that many cases are of bilateral asymmetric hyperplasia with many of the ones described as an adenoma being instead a hyperplastic steroidogenically active nodule [46]. In summary, immunohistochemistry of the CYP11B2 enzyme that catalyzes last steps of aldosterone biosynthesis, has helped uncover a significant complexity in the histological features of the adrenals of patients with unilateral production of aldosterone. Whereas the normal adrenal has a very distinctive pattern of expression of steroidogenic enzymes in the different zones, many adenomas undergo a disordered expression of the various steroidogenic enzymes with the

167 appearance of hybrid cells expressing a mixture of these enzymes. The wide variation in
168 histopathological features of the adenomas and concurrent presence of APCCs raise the
169 possibility that most cases of unilateral production of aldosterone actually might represent
170 bilateral asymmetric hyperplasia with nodules frequently due to the development of somatic
171 aldosterone-driving mutations.

172

Acknowledgements:

Research reported in this publication was supported by National Heart, Lung and Blood Institute grant R01 HL27255 and the National Institute of General Medical Sciences grant U54 GM115428. This work was supported by the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (grant agreement No [694913] to MR), the Else Kröner-Fresenius Stiftung in support of the German Conns Registry-Else-Kröner Hyperaldosteronism Registry (2013_A182 and 2015_A171 to MR) and the Deutsche Forschungsgemeinschaft (RE 752/20-1 to MR; CTC/TRR and CTC/TRR, 205/1, B15 to TAW).

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Legend:

Figure 1. Immunohistochemical staining of adrenals with the CYP11B2 antibody. A. Normal adrenal from a young individual showing diffuse staining in the subcapsular area. B. Normal adrenal of an older individual showing an aldosterone-producing cell cluster. C. APA showing fairly uniform staining of the whole adenoma. C. APA showing uneven staining of the adenoma. D. Case of unilateral primary aldosteronism with multiple APCCs.

References

- 196 1. Funder JW, Carey RM, Mantero F, Murad MH, Reincke M, Shibata H, Stowasser M,
197 Young WF, Jr. The Management of Primary Aldosteronism: Case Detection, Diagnosis,
198 and Treatment: An Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol*
199 *Metab* 2016; 101: 1889-1916
- 200 2. Savard S, Amar L, Plouin PF, Steichen O. Cardiovascular Complications Associated
201 With Primary Aldosteronism: A Controlled Cross-Sectional Study. *Hypertension* 2013;
202 62: 331-336
- 203 3. Reincke M, Fischer E, Gerum S, Merkle K, Schulz S, Pallauf A, Quinkler M, Hanslik G,
204 Lang K, Hahner S, Allolio B, Meisinger C, Holle R, Beuschlein F, Bidlingmaier M, Endres
205 S. Observational Study Mortality in Treated Primary Aldosteronism: The German Conn's
206 Registry. *Hypertension* 2012; 60: 618-624
- 207 4. Lifton RP, Dluhy RG, Powers M, Rich GM, Cook S, Ulick S, Lalouel J-M. A chimaeric
208 11 β -hydroxylase/aldosterone synthase gene causes glucocorticoid-remediable
209 aldosteronism and human hypertension. *Nature* 1992; 355: 262-265
- 210 5. Carss KJ, Stowasser M, Gordon RD, O'Shaughnessy KM. Further study of chromosome
211 7p22 to identify the molecular basis of familial hyperaldosteronism type II. *J Hum*
212 *Hypertens* 2011; 25: 560-564
- 213 6. Choi M, Scholl UI, Yue P, Bjorklund P, Zhao B, Nelson-Williams C, Ji W, Cho Y, Patel A,
214 Men CJ, Lolis E, Wisgerhof MV, Geller DS, Mane S, Hellman P, Westin G, Akerstrom G,
215 Wang W, Carling T, Lifton RP. K⁺ channel mutations in adrenal aldosterone-producing
216 adenomas and hereditary hypertension. *Science* 2011; 331: 768-772
- 217 7. Scholl UI, Stolting G, Nelson-Williams C, Vichot AA, Choi M, Loring E, Prasad ML, Goh
218 G, Carling T, Juhlin CC, Quack I, Rump LC, Thiel A, Lande M, Frazier BG, Rasoulpour
219 M, Bowlin DL, Sethna CB, Trachtman H, Fahlke C, Lifton RP. Recurrent gain of function
220 mutation in calcium channel CACNA1H causes early-onset hypertension with primary
221 aldosteronism. *Elife* 2015; 4: e06315
- 222 8. Miller WL, Auchus RJ. The molecular biology, biochemistry, and physiology of human
223 steroidogenesis and its disorders. *Endocr Rev* 2011; 32: 81-151
- 224 9. Hattangady NG, Olala LO, Bollag WB, Rainey WE. Acute and chronic regulation of
225 aldosterone production. *Mol Cell Endocrinol* 2012; 350: 151-162
- 226 10. Bulow HE, Bernhardt R. Analyses of the CYP11B gene family in the guinea pig suggest
227 the existence of a primordial CYP11B gene with aldosterone synthase activity. *Eur J*
228 *Biochem* 2002; 269: 3838-3846.
- 229 11. Okamoto M, Nonaka Y, Takemori H, Doi J. Molecular identity and gene expression of
230 aldosterone synthase cytochrome P450. *Biochem Biophys Res Commun* 2005; 338:
231 325-330
- 232 12. Chavarri MR, Yamakita N, Chiou S, Gomez-Sanchez CE. Calf adrenocortical fasciculata
233 cells secrete aldosterone when placed in primary culture. *J Steroid Biochem Mol Biol*
234 1993; 45: 493-500
- 235 13. Nishimoto K, Koga M, Seki T, Oki K, Gomez-Sanchez EP, Gomez-Sanchez CE, Naruse
236 M, Sakaguchi T, Morita S, Kosaka T, Oya M, Ogishima T, Yasuda M, Suematsu M,
237 Kabe Y, Omura M, Nishikawa T, Mukai K. Immunohistochemistry of aldosterone
238 synthase leads the way to the pathogenesis of primary aldosteronism. *Mol Cell*
239 *Endocrinol* 2017; 441: 124-133
- 240 14. Nishimoto K, Nakagawa K, Li D, Kosaka T, Oya M, Mikami S, Shibata H, Itoh H, Mitani
241 F, Yamazaki T, Ogishima T, Suematsu M, Mukai K. Adrenocortical zonation in humans
242 under normal and pathological conditions. *J Clin Endocrinol Metab* 2010; 95: 2296-2305
- 243 15. Gomez-Sanchez CE, Qi X, Velarde-Miranda C, Plonczynski MW, Parker CR, Rainey W,
244 Satoh F, Maekawa T, Nakamura Y, Sasano H, Gomez-Sanchez EP. Development of
245 monoclonal antibodies against human CYP11B1 and CYP11B2. *Mol Cell Endocrinol*
246 2014; 383: 111-117

- 247 16. Yamazaki Y, Nakamura Y, Omata K, Ise K, Tezuka Y, Ono Y, Morimoto R, Nozawa Y,
248 Gomez-Sanchez CE, Tomlins SA, Rainey WE, Ito S, Satoh F, Sasano H.
249 Histopathological Classification of Cross-Sectional Image-Negative Hyperaldosteronism.
250 J Clin Endocrinol Metab 2017; 102: 1182-1192
- 251 17. Nishimoto K, Tomlins SA, Kuick R, Cani AK, Giordano TJ, Hovelson DH, Liu CJ,
252 Sanjanwala AR, Edwards MA, Gomez-Sanchez CE, Nanba K, Rainey WE. Aldosterone-
253 stimulating somatic gene mutations are common in normal adrenal glands. Proc Natl
254 Acad Sci U S A 2015; 112: E4591-4599
- 255 18. Nakamura Y, Maekawa T, Felizola SJ, Satoh F, Qi X, Velarde-Miranda C, Plonczynski
256 MW, Ise K, Kikuchi K, Rainey WE, Gomez-Sanchez EP, Gomez-Sanchez CE, Sasano
257 H. Adrenal CYP11B1/2 expression in primary aldosteronism: Immunohistochemical
258 analysis using novel monoclonal antibodies. Mol Cell Endocrinol 2014; 392: 73-79
- 259 19. Teo AE, Garg S, Johnson TI, Zhao W, Zhou J, Gomez-Sanchez CE, Gurnell M, Brown
260 MJ. Physiological and Pathological Roles in Human Adrenal of the Glomeruli-Defining
261 Matrix Protein NPNT (Nephronectin). Hypertension 2017; 69: 1207-1216
- 262 20. Mitani F. Functional zonation of the rat adrenal cortex: the development and
263 maintenance. Proc Jpn Acad Ser B Phys Biol Sci 2014; 90: 163-183
- 264 21. Romero DG, Yanes LL, de Rodriguez AF, Plonczynski MW, Welsh BL, Reckelhoff JF,
265 Gomez-Sanchez EP, Gomez-Sanchez CE. Disabled-2 is expressed in adrenal zona
266 glomerulosa and is involved in aldosterone secretion. Endocrinology 2007; 148: 2644-
267 2652
- 268 22. Nishimoto K, Seki T, Hayashi Y, Mikami S, Al-Eyd G, Nakagawa K, Morita S, Kosaka T,
269 Oya M, Mitani F, Suematsu M, Kabe Y, Mukai K. Human Adrenocortical Remodeling
270 Leading to Aldosterone-Producing Cell Cluster Generation. Int J Endocrinol 2016; 2016:
271 7834356
- 272 23. Boulkroun S, Samson-Couterie B, Dzib JF, Lefebvre H, Louiset E, Amar L, Plouin PF,
273 Lalli E, Jeunemaitre X, Benecke A, Meatchi T, Zennaro MC. Adrenal cortex remodeling
274 and functional zona glomerulosa hyperplasia in primary aldosteronism. Hypertension
275 2010; 56: 885-892
- 276 24. Nanba K, Vaidya A, Williams GH, Zheng I, Else T, Rainey WE. Age-Related
277 Autonomous Aldosteronism. Circulation 2017; 136: 347-355
- 278 25. Nishimoto K, Seki T, Kurihara I, Yokota K, Omura M, Nishikawa T, Shibata H, Kosaka T,
279 Oya M, Suematsu M, Mukai K. Case Report: Nodule Development From Subcapsular
280 Aldosterone-Producing Cell Clusters Causes Hyperaldosteronism. J Clin Endocrinol
281 Metab 2016; 101: 6-9
- 282 26. Williams TA, Monticone S, Mulatero P. KCNJ5 mutations are the most frequent genetic
283 alteration in primary aldosteronism. Hypertension 2015; 65: 507-509
- 284 27. Fernandes-Rosa FL, Williams TA, Riester A, Steichen O, Beuschlein F, Boulkroun S,
285 Strom TM, Monticone S, Amar L, Meatchi T, Mantero F, Cicala MV, Quinkler M, Fallo F,
286 Allolio B, Bernini G, Maccario M, Giacchetti G, Jeunemaitre X, Mulatero P, Reincke M,
287 Zennaro MC. Genetic spectrum and clinical correlates of somatic mutations in
288 aldosterone-producing adenoma. Hypertension 2014; 64: 354-361
- 289 28. Taguchi R, Yamada M, Nakajima Y, Satoh T, Hashimoto K, Shibusawa N, Ozawa A,
290 Okada S, Rokutanda N, Takata D, Koibuchi Y, Horiguchi J, Oyama T, Takeyoshi I, Mori
291 M. Expression and Mutations of KCNJ5 mRNA in Japanese Patients with Aldosterone-
292 Producing Adenomas. J Clin Endocrinol Metab 2012; 97: 1311-1319
- 293 29. Williams TA, Lenders JW, Burrello J, Beuschlein F, Reincke M. KCNJ5 Mutations: Sex,
294 Salt and Selection. Horm Metab Res 2015; 47: 953-958
- 295 30. Beuschlein F, Boulkroun S, Osswald A, Wieland T, Nielsen HN, Lichtenauer UD, Penton
296 D, Schack VR, Amar L, Fischer E, Walther A, Tauber P, Schwarzmayer T, Diener S, Graf
297 E, Allolio B, Samson-Couterie B, Benecke A, Quinkler M, Fallo F, Plouin PF, Mantero F,

- Meitinger T, Mulatero P, Jeunemaitre X, Warth R, Vilsen B, Zennaro MC, Strom TM, Reincke M. Somatic mutations in ATP1A1 and ATP2B3 lead to aldosterone-producing adenomas and secondary hypertension. *Nat Genet* 2013; 45: 440-444
31. Azizan EA, Poulsen H, Tuluc P, Zhou J, Clausen MV, Lieb A, Maniero C, Garg S, Bochukova EG, Zhao W, Shaikh LH, Brighton CA, Teo AE, Davenport AP, Dekkers T, Tops B, Kusters B, Ceral J, Yeo GS, Neogi SG, McFarlane I, Rosenfeld N, Marass F, Hadfield J, Margas W, Chaggar K, Solar M, Deinum J, Dolphin AC, Farooqi IS, Striessnig J, Nissen P, Brown MJ. Somatic mutations in ATP1A1 and CACNA1D underlie a common subtype of adrenal hypertension. *Nat Genet* 2013; 45: 1055-1060
32. Scholl UI, Goh G, Stolting G, de Oliveira RC, Choi M, Overton JD, Fonseca AL, Korah R, Starker LF, Kunstman JW, Prasad ML, Hartung EA, Mauras N, Benson MR, Brady T, Shapiro JR, Loring E, Nelson-Williams C, Libutti SK, Mane S, Hellman P, Westin G, Akerstrom G, Bjorklund P, Carling T, Fahlke C, Hidalgo P, Lifton RP. Somatic and germline CACNA1D calcium channel mutations in aldosterone-producing adenomas and primary aldosteronism. *Nat Genet* 2013; 45: 1050-1054
33. Nanba K, Chen AX, Omata K, Vinco M, Giordano TJ, Else T, Hammer GD, Tomlins SA, Rainey WE. Molecular Heterogeneity in Aldosterone-Producing Adenomas. *J Clin Endocrinol Metab* 2016; 101: 999-1007
34. Nanba K, Omata K, Tomlins SA, Giordano TJ, Hammer GD, Rainey WE, Else T. Double adrenocortical adenomas harboring independent KCNJ5 and PRKACA somatic mutations. *Eur J Endocrinol* 2016; 175: K1-6
35. Nanba K, Tsuiki M, Sawai K, Mukai K, Nishimoto K, Usui T, Tagami T, Okuno H, Yamamoto T, Shimatsu A, Katabami T, Okumura A, Kawa G, Tanabe A, Naruse M. Histopathological Diagnosis of Primary Aldosteronism Using CYP11B2 Immunohistochemistry. *J Clin Endocrinol Metab* 2013; 98: 1567-1574
36. Nanba AT, Nanba K, Byrd JB, Shields JJ, Giordano TJ, Miller BS, Rainey WE, Auchus RJ, Turcu AF. Discordance between Imaging and Immunohistochemistry in Unilateral Primary Aldosteronism. *Clin Endocrinol (Oxf)* 2017, DOI: 10.1111/cen.13442:
37. Young Jr WF, Stanson AW, Thompson GB, Grant CS, Farley DR, Van Heerden JA. Role for adrenal venous sampling in primary aldosteronism. *Surgery* 2004; 136: 1227-1235
38. Dekkers T, ter Meer M, Lenders JW, Hermus AR, Schultze Kool L, Langenhuijsen JF, Nishimoto K, Ogishima T, Mukai K, Azizan EA, Tops B, Deinum J, Kusters B. Adrenal nodularity and somatic mutations in primary aldosteronism: one node is the culprit? *J Clin Endocrinol Metab* 2014; 99: E1341-1351
39. Azizan EA, Lam BY, Newhouse SJ, Zhou J, Kuc RE, Clarke J, Happerfield L, Marker A, Hoffman GJ, Brown MJ. Microarray, qPCR, and KCNJ5 sequencing of aldosterone-producing adenomas reveal differences in genotype and phenotype between zona glomerulosa- and zona fasciculata-like tumors. *J Clin Endocrinol Metab* 2012; 97: E819-829
40. Ono Y, Nakamura Y, Maekawa T, Felizola SJ, Morimoto R, Iwakura Y, Kudo M, Seiji K, Takase K, Arai Y, Gomez-Sanchez CE, Ito S, Sasano H, Satoh F. Different expression of 11beta-hydroxylase and aldosterone synthase between aldosterone-producing microadenomas and macroadenomas. *Hypertension* 2014; 64: 438-444
41. Nakamura Y, Kitada M, Satoh F, Maekawa T, Morimoto R, Yamazaki Y, Ise K, Gomez-Sanchez CE, Ito S, Arai Y, Dezawa M, Sasano H. Intratumoral heterogeneity of steroidogenesis in aldosterone-producing adenoma revealed by intensive double- and triple-immunostaining for CYP11B2/B1 and CYP17. *Mol Cell Endocrinol* 2016; 422: 57-63
42. Mulatero P, di Cella SM, Monticone S, Schiavone D, Manzo M, Mengozzi G, Rabbia F, Terzolo M, Gomez-Sanchez EP, Gomez-Sanchez CE, Veglio F. 18-

- hydroxycorticosterone, 18-hydroxycortisol, and 18-oxocortisol in the diagnosis of primary aldosteronism and its subtypes. *J Clin Endocrinol Metab* 2012; 97: 881-889
43. Omura M, Sasano H, Fujiwara T, Yamaguchi K, Nishikawa T. Unique cases of unilateral hyperaldosteronemia due to multiple adrenocortical micronodules, which can only be detected by selective adrenal venous sampling. *Metabolism* 2002; 51: 350-355.
44. Omura M, Sasano H, Saito J, Yamaguchi K, Kakuta Y, Nishikawa T. Clinical characteristics of aldosterone-producing microadenoma, macroadenoma, and idiopathic hyperaldosteronism in 93 patients with primary aldosteronism. *Hypertens Res* 2006; 29: 883-889
45. Williams TA, Lenders JWM, Mulatero P, Burrello J, Rottenkolber M, Adolf C, Satoh F, Amar L, Quinkler M, Deinum J, Beuschlein F, Kitamoto KK, Pham U, Morimoto R, Umakoshi H, Prejbisz A, Kocjan T, Naruse M, Stowasser M, Nishikawa T, Young WF, Jr., Gomez-Sanchez CE, Funder JW, Reincke M. Outcomes after adrenalectomy for unilateral primary aldosteronism: an international consensus on outcome measures and analysis of remission rates in an international cohort. *Lancet Diabetes Endocrinol* 2017, DOI: 10.1016/S2213-8587(17)30135-3:
46. Gomez-Sanchez CE, Rossi GP, Fallo F, Mannelli M. Progress in primary aldosteronism: present challenges and perspectives. *Horm Metab Res* 2010; 42: 374-381

