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UNITOPATHO, A LABELED HISTOPATHOLOGICAL DATASET FOR COLORECTAL POLYPS CLASSIFICATION AND ADENOMA DYSPLASIA GRADING

Carlo Alberto Barbano* Daniele Perlo* Enzo Tartaglione*
Attilio Fiandrotti * Luca Bertero† Paola Cassoni† Marco Grangetto**

* Computer Science Department, University of Turin, 10149 Torino, Italy

† Medical Sciences Department, University of Turin, 10126, Torino, Italy

ABSTRACT

Histopathological characterization of colorectal polyps allows to tailor patients’ management and follow up with the ultimate aim of avoiding or promptly detecting an invasive carcinoma. Colorectal polyps characterization relies on the histological analysis of tissue samples to determine the polyps malignancy and dysplasia grade. Deep neural networks achieve outstanding accuracy in medical patterns recognition, however they require large sets of annotated training images. We introduce *UniToPatho*, an annotated dataset of 9536 hematoxylin and eosin (H&E) stained patches extracted from 292 whole-slide images, meant for training deep neural networks for colorectal polyps classification and adenomas grading. We present our dataset and provide insights on how to tackle the problem of automatic colorectal polyps characterization by suggesting a multi-resolution deep learning approach.

Index Terms— Deep Learning, Multi Resolution, Colorectal polyps, Colorectal Adenomas, Digital Pathology

1. INTRODUCTION

The demand for gastrointestinal histopathology is on the rise [1], fostered by the widespreading of cancer screening programs. Gastrointestinal histopathologists inspect tissue samples, collected during colonoscopies, looking for hints that can predict the insurgence of invasive carcinoma [2]. Colorectal polyps are pre-malignant lesions found in the intestinal mucosa that pathologists analyze to *i*) ascertain the polyp type (hyperplastic, adenoma) and *ii*) assess the dysplasia grade in case of adenomas. Examination of colorectal polyps represents a large share of histopathologists workload, thus methods for automating these tasks are highly sought. Despite such clinical relevance, the concordance rate even among expert pathologists, in the diagnostic assessment of colorectal polyps, is far from optimal [3, 4]. Although the distinction between non-adenomatous and adenomatous tissue is usually reliable, the inter-observer agreement between different histological types and dysplasia grades is sub-optimal. For instance, the concordance in assessing a tubulovillous polyp or low grade dysplasia ranged around 70% [3]. Deep learning-based methods have shown promising results towards automating the pathologists’ work [5]. Korbar *et al.* [6] present a patch-based framework, developed using a ResNet architecture [7], to classify different types of colorectal polyps from whole-slide images. Their work provides empirical suggestion that residual ar-

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	HP	NORM	TA		TVA		Total
			HG	LG	HG	LG	
Slides	41	21	26	146	20	38	292
$\sigma = 7000$	59	74	98	411	93	132	867
$\sigma = 800$	545	950	454	3618	916	2186	8699
Total	604	1024	552	4029	1009	2318	9536

Table 1: *UniToPatho* class distribution for whole image slides (top) and the two patch scales made available (bottom).

chitectures are better suited at this task. Wei *et al.* [8] propose an analysis model for annotated tissue samples and perform a study on the generalization of neural models with external medical institutions. Their work describes a hierarchical evaluation mechanism to extend the classification of tissue fragments to the entire slide. Song *et al.* [9] propose a patch-based fully-convolutional approach for the classification and grading of adenomas, with a strong focus on model interpretability. They also highlight how different patch sizes should be used for adenomas classification and grading. However, the scarcity of datasets large enough and suitably labeled represents a major hurdle for training deep-learning based algorithms to predict polyp type and adenoma dysplasia grade.

This work provides the following contributions towards automatic colorectal polyps characterization, in the framework of the *DeepHealth* [10] project.

First, we make available *UniToPatho* [11], a high-resolution annotated dataset of Hematoxylin and Eosin (H&E)-stained colorectal images. *UniToPatho* enables training deep neural networks to classify different colorectal polyps types and adenomas grading. We make available our annotated dataset as a collection of high-resolution patches extracted at different scales.

Second, we show that the direct application of a deep neural network fails to classify both the tissue type and adenoma dysplasia grade.

Lastly, we propose a multi-resolution deep learning approach solving the previous issues, that achieves significant accuracy in the characterization of colorectal polyps.

2. THE UNITOPATHO DATASET

UniToPatho [11] is a dataset of annotated high-resolution H&E-stained images, comprising different histological samples of colorectal polyps, collected from patients undergoing cancer screening. The dataset is a collection of the most relevant patch images extracted from 292 *whole-slide images* (simply *slides* in the follow-

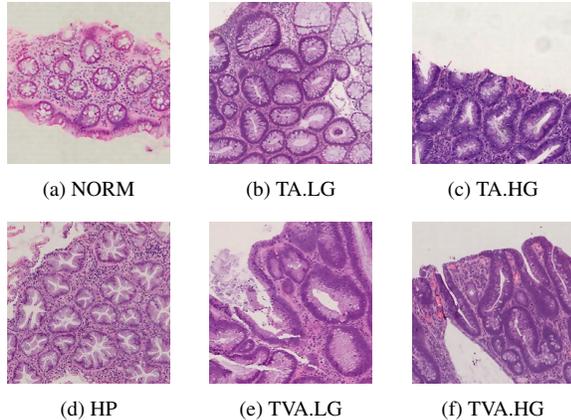


Fig. 1: Example of $800 \times 800 \mu\text{m}$ patches for the six UniToPatho colorectal polyps classes.

ing), in accordance with UniTo pathologists’ evaluation. The slides are acquired through a Hamamatsu Nanozoomer S210 scanner at $20\times$ magnification ($0.4415 \mu\text{m}/\text{px}$), as exemplified in Fig. 1. Each slide belongs to a different patient and is annotated by expert UniTo pathologists, according to six classes as follows:

- NORM** - Normal tissue
- HP** - Hyperplastic Polyp
- TA.HG** - Tubular Adenoma, High-Grade dysplasia
- TA.LG** - Tubular Adenoma, Low-Grade dysplasia
- TVA.HG** - Tubulo-Villous Adenoma, High-Grade dysplasia
- TVA.LG** - Tubulo-Villous Adenoma, Low-Grade dysplasia

Hyperplastic polyps usually exhibit no malignant potential [12], while adenomas are more likely to progress into invasive carcinomas. Tubular and tubulo-villous are common colorectal adenomas, with villous adenomas generally presenting higher malignant potential given the larger surface [12]. Adenomas are associated with a grade of dysplasia, which measures the abnormality in cellular growth and differentiation [13]. Higher grade dysplasia indicates higher malignant potential.

We split the slides into a train set and a test set with a 70% to 30% ratio, resulting in 204 slides used for training and 88 for testing. Therefore, each slide is represented either in the train set or in the test set, but not in both. Following the approach in [9, 14], we release square patches cropped from the 292 slides.¹ From each slide, we crop multiple non-overlapping square patches at different scales. We denote the side of the underlying physical area with σ , measured in μm , which we vary from 100 to 8000. The number of patches obtained for each slide hence depends on multiple factors including σ , the slide size and the polyp type. As common in similar datasets, the different classes are highly unbalanced in the dataset. We make publicly available a total of 9536 patches, 8669 of which extracted at $\sigma = 800$ (1812×1812 pixels patches) and 867 at $\sigma = 7000$ ($15,855 \times 15,855$ pixels patches). These are the sizes that will be used in Sec. 4. Tab. 1 provides a summary of the class distribution for the whole slides and the released patches.

¹The dataset is available at <https://iee-dataport.org/open-access/unitopatho>.

Type	Patch scale σ [μm]					
	100	800	1500	4000	7000	8000
BA (6-class)	0.40	0.45	0.46	0.41	0.37	0.38
NORM	0.70	0.66	0.72	0.76	0.78	0.71
HP	0.81	0.92	0.85	0.70	0.60	0.69
TA (HG+LG)	0.65	0.66	0.65	0.71	0.76	0.70
TVA (HG+LG)	0.64	0.67	0.68	0.74	0.84	0.76

Table 2: Preliminary experiments: overall BA for all of the six classes (first row) and BA for each polyp type, plus normal tissue.

3. PRELIMINARY ANALYSIS

We perform our preliminary experiments on the UniToPatho dataset with a baseline strategy owing to state-of-the-art methods [6, 8]. The method which will be described in the next section builds upon the lessons learned during these experiments.

First, we randomly color-jitter the patches described in the previous section to augment the dataset, as proposed in [8, 6]. Next, as in [6, 8], we train a deep convolutional neural network for image classification on the patches belonging to the train slides. Namely, we train an ImageNet-pretrained ResNet-18 [7] with SGD for 50 epochs, with an initial learning rate of 0.01 decayed by a factor of 0.1 every 20 epochs. The output layer is reshaped to match the UniToPatho 6-class classification problem. Each patch is downsampled to the standard 224×224 pixels ImageNet size prior being fed into the ResNet. We repeat the whole training process for different scales, choosing σ in the [100, 8000] range. For each σ , the trained network is eventually tested on the patches extracted at the same scale from the test slides².

We analyze the the results of our preliminary experiments first in terms of polyp type classification accuracy, next in terms of adenoma grade prediction for adenoma samples only. In the following, the classification accuracy will be defined in terms of Balanced Accuracy (BA) to cope with the unbalanced samples in the dataset. In Tab. 2 (first row) we show the BA achieved when attempting to discriminate all 6 classes with the baseline approach as function of the patch scale σ : it can be noted that even the best accuracy achieved at $\sigma=1500$ is quite low. Nonetheless, we conjecture that different polyp types can be better recognized at different scales: as a consequence, in Tab. 2 we also show the BA concerning the classification of each single polyp type HP, TA, TVA plus NORM. The TA and TVA classes encompass the respective low grade (LG) and high grade (HG) subclasses. Indeed, breaking down the accuracy on a per-class basis reveals that different types of polyps achieve top-classification accuracy at different scales.

Hyperplastic Polyps (HP) are best classified at a finer $800 \mu\text{m}$ scale: we hypothesize these types of benign polyps are best discriminated by looking at smaller-scale details such as gland edges [15]. Conversely, Tubular Adenomas (TA) and Tubulo-Villous Adenomas (TVA) are best classified at a coarser $7000 \mu\text{m}$ scale: we hypothesize this type of polyps is best discriminated by looking at large-scale macro structures such as entire glands shapes [16].

Coming to the problem of predicting the grade (LG or HG) of TA and TVA adenomas, we investigate whether that could be best predicted at some particular scale σ . Our experiments proved however inconclusive, i.e. the adenoma grade classification accuracy does not

²The source code along with the technical details can be found at <https://github.com/EIDOSlab/UNITOPATHO>

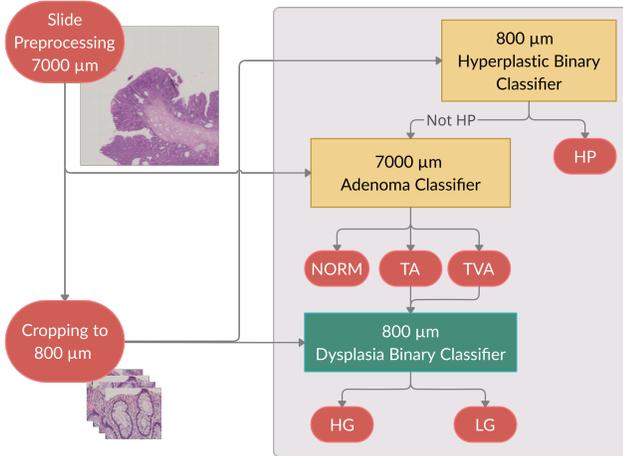


Fig. 2: The proposed multi-resolution ensemble of cascaded classifiers. The Hyperplastic and Adenoma classifiers (yellow) are trained with inputs downsampled to 224×224 pixels, while the Dysplasia classifier (green) uses full resolution images.

appear being a function of the patch scale. In fact, visual inspection of the 224×224 px downsampled patches reveals that the downsampling trashes discriminative details in the cells nuclei upon which pathologists are known to rely.

Concluding, our preliminary experiments show that *i*) different polyp types are best classified at different scales (in our case $\sigma = 800$ for HP and $\sigma = 7000$ for TA and TVA) and *ii*) adenoma grade prediction may be jeopardized if the details in cells nuclei are lost. These findings are exploited to devise our proposed method detailed in the next section.

4. PROPOSED METHOD

This section details our proposed approach towards classification of UniToPatho images: a multi-resolution ensemble of cascaded classifiers. The method relies on three cascaded ResNet-18 classifiers, having the output layer specifically adapted for each classification task. The classifiers are trained on patches extracted at either $\sigma = 800$ or $\sigma = 7000$, following the procedure described in the previous section.

The overall process of inference is depicted in Fig. 2, with the input being a single $7000 \times 7000 \mu\text{m}$ patch which is used by the three ResNet-18 classifiers mentioned above. For classifiers working at $\sigma = 800$, we crop the input image into smaller $800 \times 800 \mu\text{m}$ sub-patches which are used to generate a prediction for the entire image. Also, all of the patches are downsampled to 224×224 pixels, unless stated otherwise.

4.1. HP polyps detection

First, HP polyps are discriminated from adenomas and normal images via a binary classifier that takes as input sub-patches of size $800 \times 800 \mu\text{m}$ extracted from the larger input image. In fact, Tab. 2 shows that HP polyps are identified with top accuracy (0.92 BA) at scale $\sigma = 800$. To infer the probability of predicting HP for entire image, we compute the average predicted probability of the sub-patches of being HP. In the case the patch content is not classified as an HP polyp, the second classifier in the cascade is invoked.

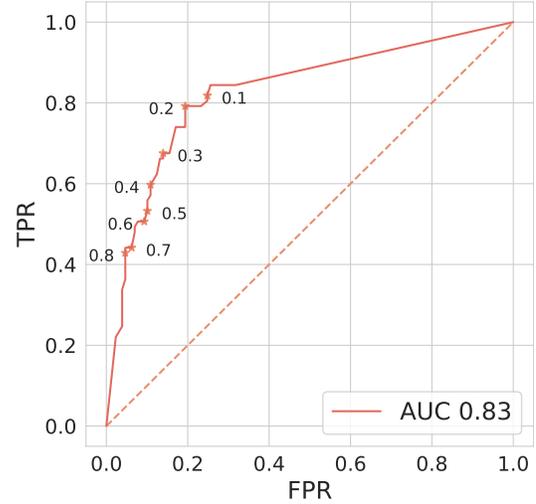


Fig. 3: ROC curve for the adenoma grade predictor as a function of the threshold T_d .

4.2. Adenoma detection

Second, TA adenomas are discriminated from TVA adenomas and from normal images via a ternary classifier taking as input the entire $7000 \times 7000 \mu\text{m}$ patch. In fact, Tab. 2 shows that TA and TVA adenomas are identified with top accuracy at this scale. In the case the patch content is classified as a TA or TVA polyp, the third and last classifier is invoked to infer its grade, otherwise the tissue will be labeled as NORM.

4.3. Dysplasia grade classification

Finally, a binary classifier is used to determine the dysplasia grade for TA and TVA adenomas. Sec. 3 suggested that downsampling the patches to 224×224 px might be detrimental towards inferring the dysplasia grade due to the loss of important features such as cells nuclei, as also observed by [9]. Thus, only for this specific classification task, we skip downsampling to prevent the loss of fine grained details. To account for the increased size of the feature space, we add an adaptive average pooling layer [17, 18] before the fully connected layer of the ResNet-18 network. We repeat the experiment of Sec. 3 sweeping the patch scale σ this time without downsampling and we find that adenoma grade classification peaks (0.81 BA) for $\sigma = 800$. As a consequence, we apply the grade classifier on sub-patches extracted from the input image at the scale $\sigma = 800$. Finally, we infer the dysplasia grade for the entire input image according to a threshold T_d : if the ratio of high grade predictions on the sub-patches is higher than T_d , the image is labeled as HG, otherwise as LG. This choice is motivated by pathologists grading guidelines, where HG can be decided on the base of small portions of tissue; clearly, setting smaller values for T_d can mimic this behaviour [12].

5. EXPERIMENTAL RESULTS

We evaluate our proposed method on the $7000 \times 7000 \mu\text{m}$ test patches of UniToPatho. As a preliminary step, we tune the classifier responsible for the dysplasia grade prediction to find a suitable T_d threshold, by analyzing its effect on the ROC curve shown in Fig. 3; as expected T_d allows one to get a good compromise between true and

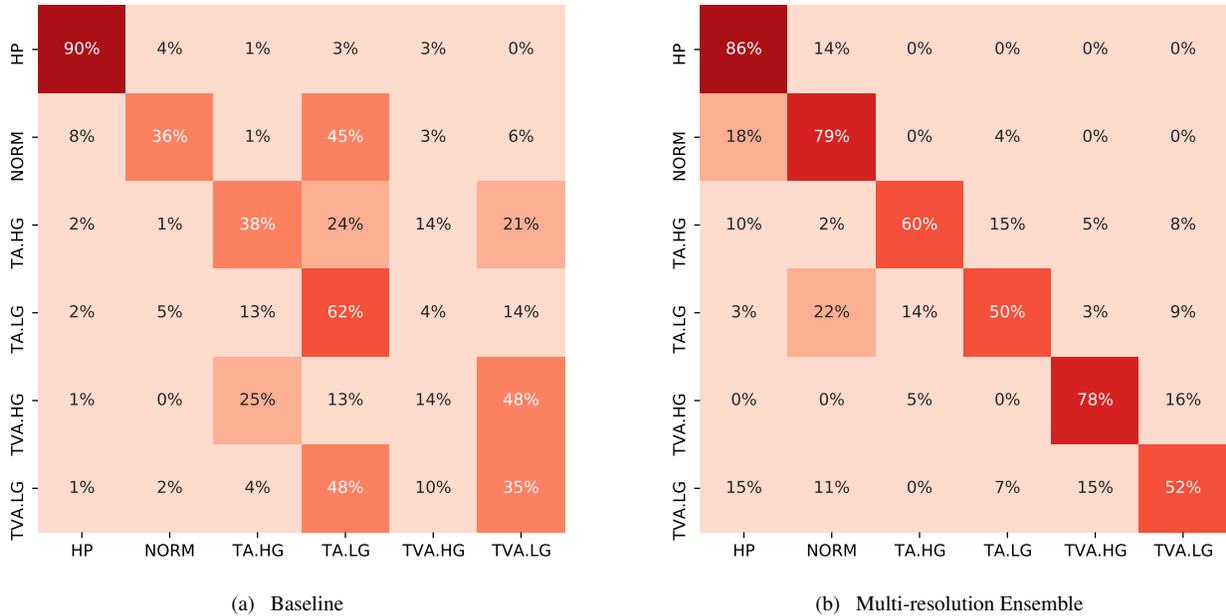


Fig. 4: Confusion matrices for the (a) baseline at $\sigma = 1500$ (b) multi-resolution ensemble, reaching a BA of 0.46 and 0.67 respectively.

	HP	NORM	TA		TVA	
			HG	LG	HG	LG
Sensitivity	0.86	0.79	0.60	0.50	0.78	0.52
Specificity	0.93	0.87	0.92	0.94	0.96	0.92
BA	0.89	0.83	0.76	0.72	0.87	0.72

Table 3: Sensitivity, Specificity and BA per class.

false positive rates yielding AUC equal to 0.83. In the following experiments we choose $T_d = 0.2$, as it strikes a favorable balance between false positives and sensitivity. Lower thresholds may be preferred, for example, in clinical applications where minimizing the false negatives rate is more important.

Fig. 4 shows the 6-class confusion matrix of the proposed method compared to the baseline approach described in Sec. 3; in this latter case we employ $\sigma = 1500$ which, as already discussed, yields the best overall BA. Looking at the diagonal, it is quite evident that the proposed method significantly improves in average accuracy, that leaps from 0.46 to 0.67 (50% relative increase). We notice how the baseline model is biased towards the lower grade classes: this represents further proof that subsampling the images results in the lack of useful features to distinguish high grade from lower grade and normal tissue. On the other hand, the multi-resolution approach shows remarkable improvements in assessing the correct grade.

Tab. 3 reports other metrics that are common in the related literature. Our proposed method achieves quite high specificity for all classes, and we also observe promising sensitivity values especially for the higher-risk TA.HG and TVA.HG classes.

Finally, we also analyze the per-type performance, as done in Sec. 3. The results are shown Tab. 4. Despite the small HP class accuracy drop - which could be due to the simple inference method - we observe an increase for all of the other tissue classes. Notably,

	σ	HP	NORM	TA	TVA
Baseline	800	0.92	0.66	0.66	0.67
Baseline	1500	0.85	0.72	0.65	0.68
Baseline	7000	0.60	0.78	0.76	0.84
Multi-resolution	-	0.89	0.83	0.81	0.87

Table 4: Comparison of the class BA between the baseline and the proposed multi-resolution approach.

we obtain a great reduction in false positive adenoma predictions, and, most importantly, a more precise distinction between TA and TVA adenomas.

6. CONCLUSIONS

This work presents UniToPatho, a histopathological dataset of colorectal polyps obtained from 292 high-resolution annotated images. UniToPatho provides annotation for hyperplastic polyps, adenomas (tubular or tubulo-villous) and their dysplasia grade (low or high). We show that a single deep neural network fails at correctly classifying the tissue type. We highlight that each of the classes discriminant features are extracted at different resolutions, making a direct classification of the tissue at a fixed scale a sub-optimal approach. This observation allowed us to design a multi-resolution deep learning strategy that, employing an ensemble of classifiers, achieves 67% accuracy. From a clinical perspective, the most relevant results are the differentiation capability between tubular and tubulo-villous adenomas and the dysplasia grading, which are the most difficult tasks for pathologists [19, 20, 21, 22]. The challenge to improve the automatic diagnostic accuracy is however still open, as UniToPatho is publicly available and still growing. Our future work will focus on collecting samples from other institutions to assess the cross-laboratory generalization capability.

7. REFERENCES

- [1] Raul S Gonzalez, “Updates and challenges in gastrointestinal pathology,” *Surgical Pathology Clinics*, vol. 13, no. 3, pp. ix, 2020.
- [2] Roisin Bevan and Matthew D Rutter, “Colorectal cancer screening—who, how, and when?,” *Clinical endoscopy*, vol. 51, no. 1, pp. 37, 2018.
- [3] Bernard Denis, Carol Peters, Catherine Chapelain, Isabelle Kleinclaus, Anne Fricker, Richard Wild, Bernard Auge, Isabelle Gendre, Philippe Perrin, Denis Chatelain, et al., “Diagnostic accuracy of community pathologists in the interpretation of colorectal polyps,” *European journal of gastroenterology & hepatology*, vol. 21, no. 10, pp. 1153–1160, 2009.
- [4] Tahmineh Mollasharifi, Mahsa Ahadi, Elena Jamali, Afshin Moradi, Parisa Asghari, Saman Maroufizadeh, and Behrang Kazeminezhad, “Interobserver agreement in assessing dysplasia in colorectal adenomatous polyps: A multicentric iranian study,” *Iranian Journal of Pathology*, pp. 167–174, 2020.
- [5] Madabhushi A Janowczyk A, “Deep learning for digital pathology image analysis: A comprehensive tutorial with selected use cases,” *Journal of pathology informatics*, pp. 7–29, 2016.
- [6] Bruno Korbar, Andrea M Olofson, Allen P Mirafior, Catherine M Nicka, Matthew A Suriawinata, Lorenzo Torresani, Arief A Suriawinata, and Saeed Hassanpour, “Deep learning for classification of colorectal polyps on whole-slide images,” *Journal of pathology informatics*, vol. 8, 2017.
- [7] Kaiming He, Xiangyu Zhang, Shaoqing Ren, and Jian Sun, “Deep residual learning for image recognition,” in *Proceedings of the IEEE conference on computer vision and pattern recognition*, 2016, pp. 770–778.
- [8] Jason W Wei, Arief A Suriawinata, Louis J Vaickus, Bing Ren, Xiaoying Liu, Mikhail Lisovsky, Naofumi Tomita, Behnaz Abdollahi, Adam S Kim, Dale C Snover, et al., “Evaluation of a deep neural network for automated classification of colorectal polyps on histopathologic slides,” *JAMA Network Open*, vol. 3, no. 4, pp. e203398–e203398, 2020.
- [9] Zhigang Song, Chunkai Yu, Shuangmei Zou, Wenmiao Wang, Yong Huang, Xiaohui Ding, Jinhong Liu, Liwei Shao, Jing Yuan, Xiangnan Gou, et al., “Automatic deep learning-based colorectal adenoma detection system and its similarities with pathologists,” *BMJ open*, vol. 10, no. 9, pp. e036423, 2020.
- [10] DeepHealth, “Deep-learning and hpc to boost biomedical applications for health,” 2019.
- [11] Luca Bertero; Carlo Alberto Barbano; Daniele Perlo; Enzo Tartaglione; Paola Cassoni; Marco Grangetto; Attilio Fian-drotti; Alessandro Gambella; Luca Cavallo, “Unitopatho,” 2021.
- [12] J. Tseung, “Robbins and cotran pathologic basis of disease: 7th edition,” *Pathology*, vol. 37, pp. 190, 2005.
- [13] National Cancer Institute, “Nci dictionary of cancer terms - dysplasia,” in <https://www.cancer.gov/publications/dictionaries/cancer-terms/def/dysplasia>.
- [14] Jakob Nikolas Kather, Niels Halama, and Alexander Marx, “100,000 histological images of human colorectal cancer and healthy tissue,” Apr. 2018.
- [15] Mehran Taherian, Saran Lotfollahzadeh, Parnaz Daneshpajouhnejad, and Komal Arora, *Tubular Adenoma*, StatPearls Publishing, Treasure Island (FL), 2020.
- [16] Mehtat Unlu, Evren Uzun, Goksel Bengi, Ozgul Sagol, and Sulen Sarioglu, “Molecular characteristics of colorectal hyperplastic polyp subgroups,” *The Turkish Journal of Gastroenterology*, vol. 31, 05 2020.
- [17] Kaiming He, Xiangyu Zhang, Shaoqing Ren, and Jian Sun, “Spatial pyramid pooling in deep convolutional networks for visual recognition,” *IEEE transactions on pattern analysis and machine intelligence*, vol. 37, no. 9, pp. 1904–1916, 2015.
- [18] Gerard Jacques van Wyk and Anna Sergeevna Bosman, “Evolutionary neural architecture search for image restoration,” in *2019 International Joint Conference on Neural Networks (IJCNN)*. IEEE, 2019, pp. 1–8.
- [19] Samir Gupta, David Lieberman, Joseph C Anderson, Carol A Burke, Jason A Dominitz, Tonya Kaltenbach, Douglas J Robertson, Aasma Shaikat, Sapna Syngal, and Douglas K Rex, “Recommendations for follow-up after colonoscopy and polypectomy: a consensus update by the us multi-society task force on colorectal cancer,” *Gastrointestinal endoscopy*, vol. 91, no. 3, pp. 463–485, 2020.
- [20] Cesare Hassan, Giulio Antonelli, Jean-Marc Dumonceau, Jaroslaw Regula, Michael Bretthauer, Stanislas Chaussade, Evelien Dekker, Monika Ferlitsch, Antonio Gimeno-Garcia, Rodrigo Jover, et al., “Post-polypectomy colonoscopy surveillance: European society of gastrointestinal endoscopy (esge) guideline—update 2020,” *Endoscopy*, vol. 52, no. 08, pp. 687–700, 2020.
- [21] Takahisa Matsuda, Han-Mo Chiu, Yasushi Sano, Takahiro Fujii, Akiko Ono, and Yutaka Saito, “Surveillance colonoscopy after endoscopic treatment for colorectal neoplasia: from the standpoint of the asia–pacific region,” *Digestive Endoscopy*, vol. 28, no. 3, pp. 342–347, 2016.
- [22] Matthew D Rutter, James East, Colin J Rees, Neil Cripps, James Docherty, Sunil Dolwani, Philip V Kaye, Kevin J Monahan, Marco R Novelli, Andrew Plumb, et al., “British society of gastroenterology/association of coloproctology of great britain and ireland/public health england post-polypectomy and post-colorectal cancer resection surveillance guidelines,” *Gut*, vol. 69, no. 2, pp. 201–223, 2020.