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(Article begins on next page)

**Mixed culture of *Lactococcus lactis* and *Kluyveromyces marxianus* isolated from kefir grains for pollutants load removal from Jebel Chakir leachate**

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**Keywords:** ammonium nitrogen removal, bioremediation, landfill leachate, microorganisms, organic materials removal

**Abstract**

The wastewater from the dumping site usually contains high pollutant levels. Biological process and physico-chemical treatments are among several technologies for wastewater treatment. Using microorganisms in the treatment of landfill leachate is an emerging research issue. Furthermore, bioremediation is a feasible approach for pollutants removal from landfill leachate which would provide an efficient way to resolve the issue of landfill leachate. In this study, the performance of yeast and bacteria isolated from kefir grains was assessed for landfill leachate treatment. Kefir grains microbial composition was evaluated by molecular approaches (Rep-PCR and 16S rRNA gene sequencing). The obtained outcomes denoted that high concentrations of lactic acid bacteria and yeast populations (over 10<sup>7</sup> CFU/ml) were found in the kefir grains and were essentially composed of *Lactococcus lactis*, *Lactobaccillus kefirien*, *Bacillus* sp., *L. lactis*, and *Kluyveromyces marxianus*. The co-culture with 1% of inoculum size was demonstrated as the most efficient in the degradation of different contaminants. The overall abatement rate of chemical oxygen demand (COD), ammonium nitrogen (NH<sub>4</sub><sup>+</sup> -N), and salinity were 75.8%,

85.9%, and 75.13%, respectively. The bioremediation process resulted in up of 75% removal efficiency of Ni and Cd, and a 73.45%, 68.53%, and a 58.17% removal rates of Cu, Pb, and Fe, respectively. The research findings indicate the performance of *L. lactis* and *K. marxianus* co-culture isolated from kefir grains for the bioremediation of LFL.

### **Practitioner Points**

-Isolation and identification of microorganisms from kefir grains was carried out. • Biological treatment of LFL using monoculture of (*Lactococcus lactis*; *Kluyveromyces marxianus*) and co-culture (5% of *L. lactis* and 5% *K. marxianus*) has been performed.

-Biological treatment using co-culture strain is an effective approach to remove organic matter,  $\text{NH}_4^+ - \text{N}$  and heavy metals.

### **Introduction**

Industrialization, urbanization increase, and technological advancements have induced a rapid growth in the municipal solid waste (MSW) production. Throughout the world, appropriate management of MSW is becoming one of the most challenging environmental problems. For decades, landfilling has been commonly applied as an ultimate disposal practice for MSW (Klauson et al., 2015; Oulego, Collado, Laca, & Díaz, 2016). However, the percolation and filtration of rainwater into the waste layers can produce important amounts of landfill leachate (LFL) (He et al., 2016; Zhang et al., 2013). This waste-water is a complex mixture of several pollutants like organic compounds, ammonia–nitrogen, inorganic salts (e.g., chloride, sulfate, sodium, etc) and heavy metals (e.g., copper, iron, lead, manganese, etc) (Vaverková et al., 2018; Xie et al., 2012). Various factors influence the quality of leachate such as the waste age, the climatic conditions, the waste composition as well as the depth of the landfill site (Ghani, Yusoff, Zaman, Zamri, & Andas, 2017; Mandal, Dubey, & Gupta, 2017). Due to its complex composition, landfill leachate must be properly treated to remove organic materials and ammonium–nitrogen ( $\text{NH}_4^+ - \text{N}$ ) before its discharge into the environment. Therefore, it is necessary to select sustainable processes

to manage and treat this particular effluent. Accordingly, several studies have been focused on LFL treatment using different approaches such as electro-coagulation; nanofiltration; oxidation and photocatalysis (Kamaruddin, Yusoff, Aziz, & Hung, 2015). However, the high costs and the results of secondary pollutants in some cases are the major disadvantages of these processes. So far, biological processes have gained an interest for the LFL treatment since they have been considered as the most environmentally friendly processes (Klauck et al., 2017). The presence of microorganisms with important biodegradation potentials and resistance to different contaminants could be a potential problem-solving of LFL treatment (Wang et al., 2018). As reported in the literature, several investigations have been demonstrated that some microorganisms are able to degrade dissolved organic matter contained in the wastewater (Sosa et al., 2017; Wang et al., 2018; Westlund & Yargeau, 2017). However, it is worthy to highlight that a few researches have focused on  $\text{NH}_4^+ - \text{N}$  abatement rate of LFL using bioremediation process (Cherni et al., 2020; Elleuch et al., 2020). In fact, several microorganisms have been tested for the assimilation of different heavy metals (Abbas & Badr, 2015; Mohd et al., 2017). Others have described the efficiency of polycultures (consortium) in wastewater treatments, including biomass production and pollutants removal (Ayed, Abid, & Hamdi, 2019; Gonçalves, Pires, & Simões, 2016). The use of microbial consortium for contaminants removal can be very beneficial since combining microorganisms was found to lead to the improvement of a robust biological system that can operate under different stress conditions which can enhance pollutants uptake loads (Ayed, Asses, Chammem, & Hamdi, 2016; El ouaer, 2020). In the same vein, Kumari, Ghosh, and Thakur (2016) demonstrated the efficiency of LFL treatment using a consortium of microalgae and bacteria. Bacto-algal mixed culture proved efficiency in organic matters degradation and heavy metals biosorption. Furthermore, a study achieved by Zhang, Vahala, Wang, and Smets (2016) describes communities and their biological activity in LFL treatment. It has been reported that the major factor that affecting the bioremediation performance is the capacity of the added culture to display its activities and survive in different physiological conditions (Song, Wang, Yue, & Li, 2013; Westlund & Yargeau, 2017). Highly adaptive bacteria exceeded an important removal rate of pollutant substances. Consequently, a complex symbiotic microbial consortium of several yeasts and bacteria would be interesting mixture to overcome

76 stressing LFL culture conditions. In this context, the present work supposes that kefir grains (KGs) as a microbial  
77 consortium constituted mainly of some bacterial species such as lactobaccili, lactococci, and leuco- nostoc and  
78 yeast species such as Kluyveromyces, Candida, and Saccharomyces growing in ecological niche (Bengoa,  
79 Iraporda, Garrote, & Abraham, 2019; Richard, 2016) could be promote for the removal of pollutants. Thus, the  
80 aim of this research is to assess the performance of Lactococcus lactis and K. marxianus monoculture and co-  
81 culture in the biodegradation of landfill leachate.

## 83 **Materials and methods**

85 Isolation and identification of microorganisms Microorganisms were isolated from Tunisian kefir product.  
86 Seventeen gram kefir samples were aseptically taken and homogenized with sterilized Ringer's solution. The  
87 samples were homogenized for 3 min in a stomacher. The serial decimal dilutions were prepared in Ringer water  
88 and plated for bacterial and yeast counts. Bacteria strains were grown on MRS agar plate's counts (Man, Rogosa,  
89 Sharpe, Heywood, Lancashire, UK) agar supplemented with 0.025 g/ml of Delvocid (Sigma) and incubated at 30°C  
90 for 24 hr, whereas yeasts and molds were grown from W.L agar plate's counts (Wallerstein Laboratory Nutrient  
91 Agar) supplemented with 0.05 g/ml of Tetracycline (Sigma) at 25°C for 48 hr. Yeast and bacteria strains were ran-  
92 domly picked, subjected to Gram staining (for bacteria strains), purified and growth in YPD (dextrose [2%],  
93 bacteriological peptone [1%], yeast extract [1%]), and MRS broth, respectively. Purified strains were maintained  
94 at -20°C with 30% (v/v) of glycerol until the use in the bioremediation experiments. All purified isolates were  
95 subjected to DNA extraction (Cocolin et al., 2004). Afterward, amplification of the FD1-RD1 region of 16s rRNA  
96 (Weisburg, Barns, Pelletier, & Lane, 1991) and ITS-5.8S rDNA region (Korabečná, Liška, & Fajfrlik, 2003) was  
97 carried out for bacteria and yeast isolates, respectively. Bacteria and yeasts were identified by alignment of the  
98 sequenced amplicon with Basic Local Alignment Search Tool (BLAST; <https://blast.ncbi.nlm.nih.gov/Blast.cgi>).

## 100 **LFL characterization**

101

102 **LFL sampling.** Leachate samples were collected from Jebel Chakir landfill. It is located in the southwest side of  
103 Tunis City and has started operating in 1999. The site occupies 47 ha over a reserved total area of 124 ha  
104 (ANGed & GIZ, 2014; Ismail et al., 2011). In this study, raw leachate samples were collected from the collection  
105 systems at the Jebel Chakir landfill site in 20 L plastic barrels, transported to the laboratory and stored at the  
106 refrigerator before being used and analyzed.

107

108 **LFL physicochemical analysis.** The performance of biological LFL treatment is evaluated by measuring the  
109 decrease of the organic matter, ammonium, and heavy metals. The analyses were assessed on raw and treated  
110 leachate. The initial pH of the leachate was modified to the desired value using 1 M hydrochloric acid. PH, TDS,  
111 and EC were measured by a multi parameter type «Consort C 860». COD, BOD5, and  $\text{NH}_4^+ - \text{N}$  were  
112 determined according to Rodier and Legube (2009). Total Kjeldahl nitrogen was measured according to Rodier  
113 and Legube (2009). The concentrations of heavy metals were determined using flame atomic absorption  
114 method (Analytic Jena AG Spectrometer AAS vario 6). The bacterial cell biomass was detected by optical  
115 density of samples at 600 nm. The LFL characteristics are showed in Table 1.

116

#### 117 **Main physicochemical characteristics of Jebel Chakir LFL.**

118 Physicochemical proprieties of Jebel Chakir LFL were determined according to the following parameters: COD,  
119 pH, salinity, electrical conductivity (EC), and heavy metals (Table 1). The raw LFL showed a dark brown color,  
120 physical, and chemical parameters presented an alkaline pH of 7.73 and high conductivity (20.6 ms/cm) as well  
121 as considerable levels of COD (26.200 mg O<sub>2</sub>/L) that can be attributed to the high initial organic matter in the  
122 leachate. The relatively high levels of salinity (3.62 g/L) demonstrated the presence of inorganic contents in the  
123 studied leachate. It was noticed that leachate samples contain significant amounts of toxic heavy metals such  
124 as Ni (3.52 mg/L), Cu (1.62 mg/L), Cd (2.73 mg/L), Pb (1.78 mg/L), and Fe (9.23 mg/L). Similar results were  
125 reported by Ellouze, Aloui, and Sayadi (2008), the leachate presented an important quantities of organic

126 matter, nitrogen, and toxic heavy metals especially Fe (20.6 mg/L). The relatively high levels of contaminants  
127 including organics, ammonia, inorganic substances, and toxic metals confirmed the high organic load of the  
128 dumped garbage in Jebel Chakir LFL.

## 129

### 130 **Bioremediation process for landfill leachate treatment Inoculum preparation.**

131 The isolation and identification of bacteria and yeast strains were done as described previously in Section  
132 Isolation and identification of microorganisms. The yeast strain was inoculated to 50 ml of synthetic nutrient  
133 broth medium YPD (dextrose [2%], bacteriological peptone [1%], yeast extract [1%]) and incubated at 30°C for  
134 48 hr with 150 rpm agitation speed. For the preparation of bacteria inoculum, the strain was inoculated to 50  
135 ml MRS broth medium and incubated at 37°C for 24 hr with 150 rpm agitation speed. Then, the microbial  
136 inoculums were used for bioremediation process.

137 Experimental set-up. Bioremediation was performed at initial pH of 5. The process was achieved in batch with  
138 Erlenmeyer flask (50 ml) containing 20 ml of the wastewater. A set of experiments were carried out in  
139 duplicate. For the first set of experiments, the selected monoculture of (*L. lactis*; *K. marxianus*) was added  
140 separately in the test samples at different inoculum sizes (1%, 3%, and 5% [v/v]) and incubated in the orbital  
141 shaker with a rotation speed of 150 rpm at room temperature for 10 days. For the second set of experiments,  
142 three inoculum sizes (1%, 3%, and 5% [v/v]) were tested. A bacteria and yeast co-culture was prepared by  
143 taking 5% of *L. lactis* and 5% *K. marxianus*, respectively. Bacteria and yeast inoculums were added separately in  
144 the leachate samples and incubated in the orbital shaker with a rotation speed of 150 rpm at room  
145 temperature for 10 days. A blank experiment which consisted of raw leachate was run in the same condition of  
146 the test samples.

147

## 148 **Results and discussion**

149

### 150 **Strain selection**

151 Kefir grains were analyzed microbiologically to identify the predominant microorganisms. A total of 54 bacteria  
152 and 27 yeasts were subjected to Rep-PCR fingerprinting technique to estimate bacteria and yeast diversity of  
153 kefir grains. Then, 11 bacteria and 4 yeasts were chosen as representative of each sub-cluster obtained (70% of  
154 similarity) using the Pearson correlation. The sequences were aligned to the query sequences of the GenBank  
155 16S rRNA and ITS-5.8S rDNA sequences database, resulting in identities of known sequences of 99%–98%, as  
156 shown in Table S1 (Supporting Information). *Lactococcus lactis* and *K. marxianus* are among predominant  
157 microbial populations from the kefir grains. Figure 1 presented the obtained strains after visualization under  
158 ultraviolet light and the resulting profiles were determined by digital image capturing using a CCD UVI pro  
159 Platinum 1.1. Recently, Elleuch et al. (2020) reported the cost effectiveness of kefir grains as a biological  
160 pretreatment for landfill leachate. Overall, TOC, COD,  $\text{NH}_4^+ - \text{N}$ , and  $\text{PO}_3^{3-} - \text{P}$  decreased, respectively, by 93%,  
161 83.33 %, 70%, and 88.25% with respect to the raw effluent (24,000 mg/L), thus reflecting the resistance of these  
162 grains to the toxicity of leachate (Elleuch et al., 2020). Same conclusions were reached by Mohd et al. (2017) and  
163 Wang et al. (2018) showing the efficiency of these genera of lactic acid bacteria and the yeast *Kluyveromyces*  
164 *marxianus* for the removal of organic matter and toxic substances from wastewaters. In this context, lactic acid  
165 bacteria and yeast strains isolated from KGs could be potential for the removal of organic matter and toxic  
166 pollutants from LFL. In addition, Milanowski et al. (2017) worked on the biosorption of silver using *L. lactis* strains.  
167 It was found that the lactic acid bacteria was able to grow and absorbed about 70%–96% of silver from 1 ppm  
168 solution. Yadav et al. (2014) proved the performance of *K. marxianus* in the biodegradation of cheese whey (78%  
169 of COD removal after 30 hr of incubation).

## 170

### 171 **Bioremediation treatment of LFL using the selected bacterial strain and yeast**

172 The progress in bioremediation rates depends, to a great extent, on the ability of the introduced microorganisms  
173 to survive and display their activities in difficult conditions (Bardi et al., 2017; Tigini, Prigione, & Varese, 2014).  
174 In recent years, various research studies on the detoxification and treatment of wastewaters using lactic acid  
175 bacteria (LAB) and yeasts have been carried out world- wide (Reis et al., 2017; Yi et al., 2017; Zhang et al., 2016).



176 In this work, the bioremediation was monitored to provide an insight into its efficiency in reducing the  
177 contaminant load in the leachate using the selected strains. During the bioremediation process, the effects of  
178 the treatment duration (each day) and the inoculum size (1%, 3%, 5% [v/v]) of *L. lactis* and *K. marxianus* were  
179 studied.

180

#### 181 **Change in organic matter and $\text{NH}_4^+ - \text{N}$ during**

182 bioremediation. Despite the modification of the samples, the LFL was too toxic to allow the growth and the  
183 metabolic activity of the introduced microorganisms *L. lactis* and *K. marxianus* were used to assess the  
184 performance of bioremediation treatment of LFL. Furthermore, pollutants removal efficiency of LFL using the  
185 mixture of *L. lactis* and *K. marxianus* was studied to evaluate if there was any synergy or inhibition effects of  
186 these two microorganisms on the pollution removal in the bioremediation treatment process. To our knowledge,  
187 the present work is the first investigating the ability of *L. lactis* and *K. marxianus* to degrade several pollutants  
188 from LFL. The variation of COD using the two microorganisms and consortium is reported in Figure 2.

189 It is clearly seen that the consortium response seems to be better compared to both monoculture of *L. lactis* and  
190 *K. marxianus*. The co-culture exhibited an appreciable COD reduction in a shorter degradation time only after 3  
191 days. The COD reduction was comparatively higher with the samples at 1% of inoculum size. Furthermore, the  
192 findings showed that the maximum COD removal rate increased greatly (75.8%) using the co-culture compared  
193 with those of *L. lactis* (52.3%) and *K. marxianus* (56.2%), which suggested no competition or inhibition between  
194 the two selected strains. It was possible due to the co-culture synergy effect on increasing the growth abil- ity,  
195 biomass production, and enzyme activity. This outcome is promising because it proved not only the compatibility  
196 of yeast and bacteria populations but their complementarity. In fact, an important COD concentration might  
197 accelerate the growth of heterotrophic bacteria, which would consume oxygen and nutrients rapidly (Patureau  
198 et al., 2001). In addition, some researchers have noted the benefits of applying mixed cul- ture over single strains  
199 cultures (Alcántara et al., 2015; Wilkie & Mulbry, 2002). The findings of our work seem to be more interesting  
200 than those described in Razarinah, Zalina, and Abdullah (2014). As results of experiments, maximum simul-

201 taneous COD and BOD5 removal were achieved 89.14% and 2.11%, respectively, after 28 days of incubation using  
202 immo- bilized *Trametes menziesii*. Recently, Er, Seow, Lim, Ibrahim, and Sarip (2018) tested *Brevibacillus*  
203 *panacihumi* strain ZB1 for the removal of toxic compounds from LFL. As a result, COD and ammonia–nitrogen  
204 degradation were attained ~40% and ~50%, respectively, after 42 days of incubation. On the other hand, the  
205 variation trends of COD removal rate using *L. lactis* and *K. marxianus* were similar with 3% and 5% of inoculum  
206 size, as shown in Figure 2. The most important COD removal rate using *L. lactis* and *K. marxianus* with 3%  
207 inoculum size were 30.1% and 36.3%, respectively. The abatement rate of COD using *L. lactis* and *K. marxianus*  
208 with 5% of inoculum size were 31.2% and 29.6%, respectively. The mixture of *L. lactis* and *K. marxianus* with 3%  
209 of inoculum size increased the maximum COD removal rate to 38.2% and slightly decreased to 28.1% with 5% of  
210 inoculum size after 10 days of incubation for LFL treatment. As far as the control samples, no important COD  
211 degradation was observed. According to the literature, several studies reported that the inoculum size is a  
212 relevant factor to improve the biodegradation of wastewater (Bohutskyi et al., 2016; Elleuch et al., 2020). Thus,  
213 the inoculation of 1% of consortium had almost a positive effect on the degradation rate during the biological  
214 process. However, the addition of 3% and 5% of consortium was unfavorable for the degradation and pollutants  
215 removal. In the same vein, previous studies revealed the benefits of nutrients to accelerate and/ or facilitate the  
216 biodegradation of contaminants. This is principally explained by the biostimulation of microorganisms through  
217 the supply of nutrients such as carbon, nitrogen, and phosphorus (Dadrasnia, Azirun, & Ismail, 2017). For that  
218 the possible interpretation was that the inefficiency of 3% and 5% could be attributed to the low nutrients  
219 availability for their stimulation, which could have decreased the biodegradation process. Therefore, future  
220 studies should focus on improving the proprieties of substrates, inoculums, and the environmental conditions.  
221 In this research, the second result was  $\text{NH}_4^+ -\text{N}$  reduc- tion in leachate. As shown in Figure 3.  $\text{NH}_4^+ -\text{N}$  level in  
222 the studied leachate was 780 mg/L. According to the literature, the high amount of  $\text{NH}_4^+ -\text{N}$  was probably due  
223 to the fermentation and hydrolysis of the nitrogenous fragments of biodegrad- able refuse. The obtained results  
224 proved the strains co-culture capacity to survive under a considerable  $\text{NH}_4^+ -\text{N}$  amount. Bioremediation  
225 presented a great effectiveness in  $\text{NH}_4^+ -\text{N}$  reduction for all tested treatments. The reduction rate of  $\text{NH}_4^+ -\text{N}$

226 remarkably increased after one week and improved until the last day of the study. The maximum  $\text{NH}_4^+ - \text{N}$   
227 reduction of leachate with 1% of co-culture addition was approximately 85.9%. The highest  $\text{NH}_4^+ - \text{N}$  removal  
228 rate using co-culture with 3% and 5% of inoculum size was 23.06% and 33.29%, respectively. The same yields  
229 were obtained for the samples treated by *L. lactis* and *K. marxianus*. Compared with the control, amending the  
230 leachate with 1% of *L. lactis* and *K. marxianus* pure cultures increased the  $\text{NH}_4^+ - \text{N}$  removal rate about 48.6%  
231 and 37%, respectively. Although 21.05% and 26.47%  $\text{NH}_4^+ - \text{N}$  removal rates have been recorded using *K.*  
232 *marxianus* at inoculation sizes of 3% and 5%, respectively. The maximum  $\text{NH}_4^+ - \text{N}$  reduction of LFL without  
233 culture addition was approximately 15.4%. This result could be attributed to the effect of enzyme production  
234 and biomass activity of the microorganism (Mohd et al., 2017). In view of that, our outcomes showed a significant  
235 degradation of organic matter compared to other studies which highlighted the performance of the  
236 consortium used in this work. For example, Raposo, Oliveira, Castro, Bandarra, and Morais (2010) reported 13%–  
237 15% of COD removal using a consortium of *Chlorella vulgaris* and brewery wastewater native microalgal-bacterial  
238 consortia after 20 days of treatment. Our findings indicated that the co-culture possessed pollutions removal  
239 abilities for LFL. Moreover, the above results suggested that the inoculum size has a great effect on the organic  
240 matter and  $\text{NH}_4^+ - \text{N}$  degradation. 1% of inoculum size was found to be more efficient in the removal of  
241 pollutants. For this reason, in the subsequent analyses using *L. lactis*, *K. marxianus*, and the consortium, only the  
242 inoculation size of 1% will be considered.

243

#### 244 **Change in salinity during bioremediation.**

245 Salinity assessment values in the LFL ranged from 3 to 5 g/L. Such variation could probably be due to the possible  
246 precipitation of salts with other LFL compounds while it was kept under refrigeration. As illustrated in Figure 4,  
247 the salinity of different leachate samples increased rapidly at the beginning, then, it stabilized gradually over the  
248 experimentation time. The evolution trends of leachate salinity inoculated by 1% *L. lactis*, *K. marxianus*, and  
249 consortium were similar. The first observation could show the ability of different strains not only to survive but  
250 also to grow in a stress environment with a high concentration of salts. This finding confirms that *L. lactis*, *K.*

251 marxianus, and consortium are resistant strains to high salts contents in LFL. This property was also reported by  
252 Huang, Liu, Liang, and Mao (2014) and Tekarslan-Sahin, Alkim, and Sezgin (2018) who proved the ability and  
253 tolerance of lactic acid bacteria and yeasts to survive at higher salinity value. In fact, the salinity was increased  
254 from 3.62 to 0.8 g/L in leachate inoculated with consortium. Also, when *L. lactis* and *K. marxianus* were  
255 inoculated, the salinity was reduced to 1.5 and 1.2 g/L, respectively. Thus, the salinity increase is attributed to  
256 the growth and the accumulation in cell walls as well. Numerous research studies have proved that yeast cells  
257 exposed to high salt contents show dehydration, physiological and biochemical variations, and gene modification  
258 (Mage & Siderius, 2002). Applying detoxification mechanisms and ion transport, the cells demonstrate an  
259 important tolerance to Na<sup>+</sup> stress through osmotic regulation by adsorbing Na<sup>+</sup> salts ability and tolerance of  
260 lactic acid bacteria and yeasts to survive at higher salinity value. In fact, the salinity was increased from 3.62 to  
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263 accumulation in cell walls as well. Numerous research studies have proved that yeast cells exposed to high salt  
264 contents show dehydration, physiological and biochemical variations, and gene modification (Mage & Siderius,  
265 2002). Applying detoxification mechanisms and ion transport, the cells demonstrate an important tolerance to  
266 Na<sup>+</sup> stress through osmotic regulation by adsorbing Na<sup>+</sup> salts inside the cell (Dhar, Sägesser, Weikert, Yuan, &  
267 Wagner, 2011; François, Walther, & Parrou, 2012).

268

#### 269 **Change in heavy metals during bioremediation.**

270 Recently, the biosorption of heavy metals by a variety of biomasses including bacteria, fungi and algae has been  
271 demonstrated as important economical and effective alternatives (Mehta & Gaur, 2005; Romera, Gonzalez,  
272 Ballester, Blazquez, & Munoz, 2006). Since the area of biosorption is huge, our study was restricted to the toxic  
273 metals such as cadmium, nickel, copper, zinc, and iron biosorption using 1% of *L. lactis*, *K. marxianus*, and the co-  
274 culture. As reported in Table 2, the maximum recorded removal rates of Ni, Cr, Cd, Pb, and Fe were 81.53%,  
275 73.45%, 79.48%, 68.53%, and 58.17%, respectively, with the co-culture inoculation size of 1% (v/v). By comparing

the samples treated with 1% (v/v) of *L. lactis* and with 1% (v/v) of *K. marxianus*, it can be found that *L. lactis* showed higher ability in the removal of Ni (39.77%) and Cd (62.63%). According to the literature, several studies have been performed to explore LAB and yeasts in the wastewaters treatment (Reis et al., 2017; Zhang et al., 2016). These outcomes were in agreement with Han et al. (2006) findings using beer yeast for the removal of Cu<sup>2+</sup> and Pb<sup>2+</sup> from wastewater. The experimental results exhibited the capacity of yeast to consume Cu<sup>2+</sup> and Pb<sup>2+</sup> and therefore, to reduce these toxic metals level in the culture medium. Otherwise, Bhakta, Ohnishi, Munekage, Iwasaki, and Wei (2012) tested the performance of eleven LAB strains isolated from mud and sludge in heavy metals removal rates from wastewater. They reported that *Lactobacillus reuteri* showed the highest Cd<sup>2+</sup> (25%) and Pb<sup>2+</sup> (59%) removal capacities. In another research carried by Schut, Zauner, Hampel, König, and Claus (2011), it was indicated that *Lactobacillus* species may have a great application in the reduction of Cu<sup>2+</sup>.

287

## 288 **Conclusion**

In the present research, the bacteria and yeast isolates from kefir product were applied for landfill leachate treatment. The identification of the isolated microorganisms is presented to be *L. lactis* and *K. marxianus*. A consortium was constructed from bacteria and yeast mixed culture. As far as COD, NH<sub>4</sub><sup>+</sup> -N, and heavy metals removals are concerned; the results demonstrated that isolated bacteria and yeast strains have the ability to reduce the COD value up to 50% and NH<sub>4</sub><sup>+</sup> -N value up to 35%. Furthermore, the Addition of 1% (v/v) of inoculum showed the best biodegradation rate compared to 3% and 5% (v/v). However, a co-culture would prove to be more effective and beneficial compared to single strain. Significant results were obtained in co-culture (1% (v/v) of inoculum size) which reduces the COD (75.8%), NH<sub>4</sub><sup>+</sup> -N (85.9%), and salinity (75.13%). Also, the results proved that applying a bacto-yeast co-culture to Jebel Chakir leachate is a suitable treatment to remove high quantities of heavy metals like Ni (81.53%), Cu (73.45%), Cd (79.48%), Pb (68.53%), and Fe (58.17%). The LFL bioremediation process could be promising and considered as an effective green technology in the removal of organic compounds from LFL.

301

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306

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489 **Legend of figures:**

490

491 **Fig. 1** Representative polymorphic profiles of LAB and yeast colonies isolated from the Kefir grains: M, molecular  
492 marker-1,500 bp.

493

494 **Fig. 2** COD removal during the bioremediation treatment of LFL using single and co-cultures with different  
495 inoculum size: 1% (a), 3% (b), and 5% (c).

496

497 **Fig. 3**  $\text{NH}_4^+ - \text{N}$  removal during the bioremediation treatment of LFL using single and co-cultures with different  
498 inoculum size: 1% (a), 3% (b), and 5% (c).

499

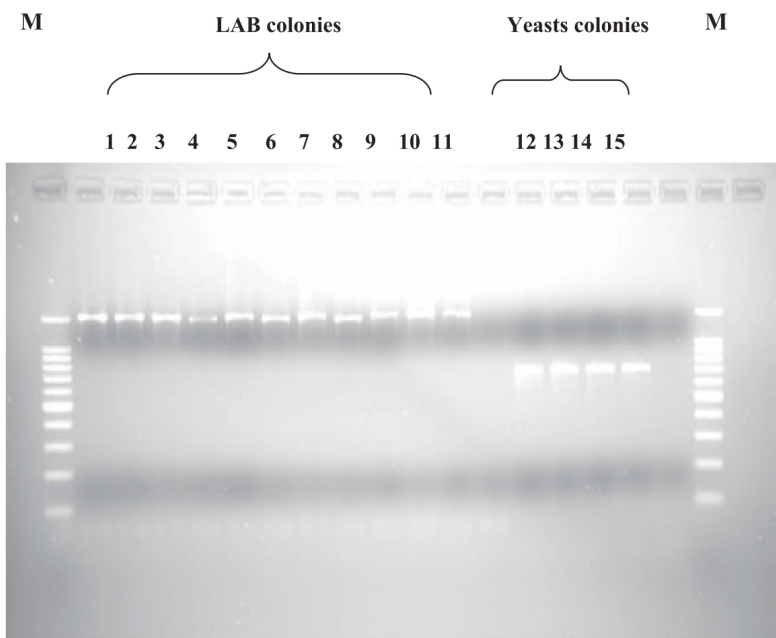
500 **Fig. 4** Salinity removal during the bioremediation treatment of LFL using 1% of single and co-cultures.

501 ability

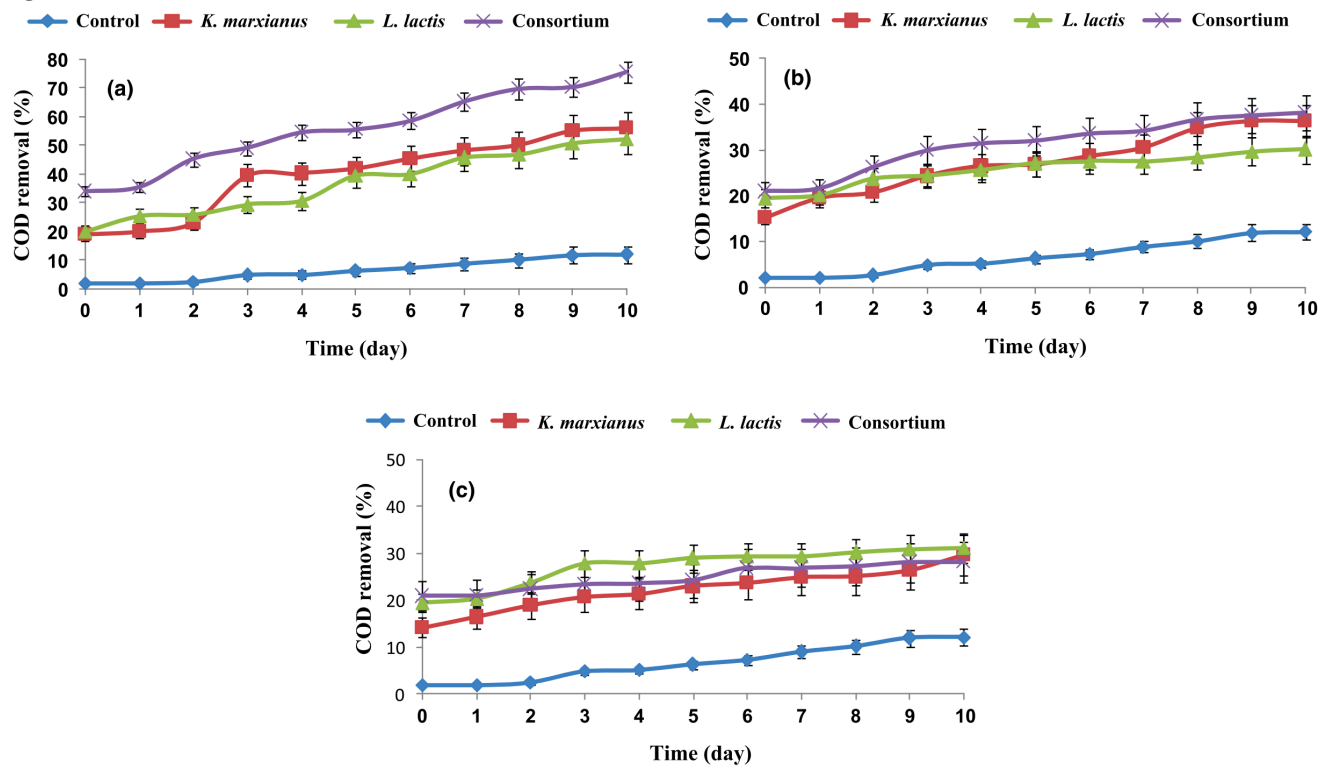
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504 **Figure 1**



**Figure 2**

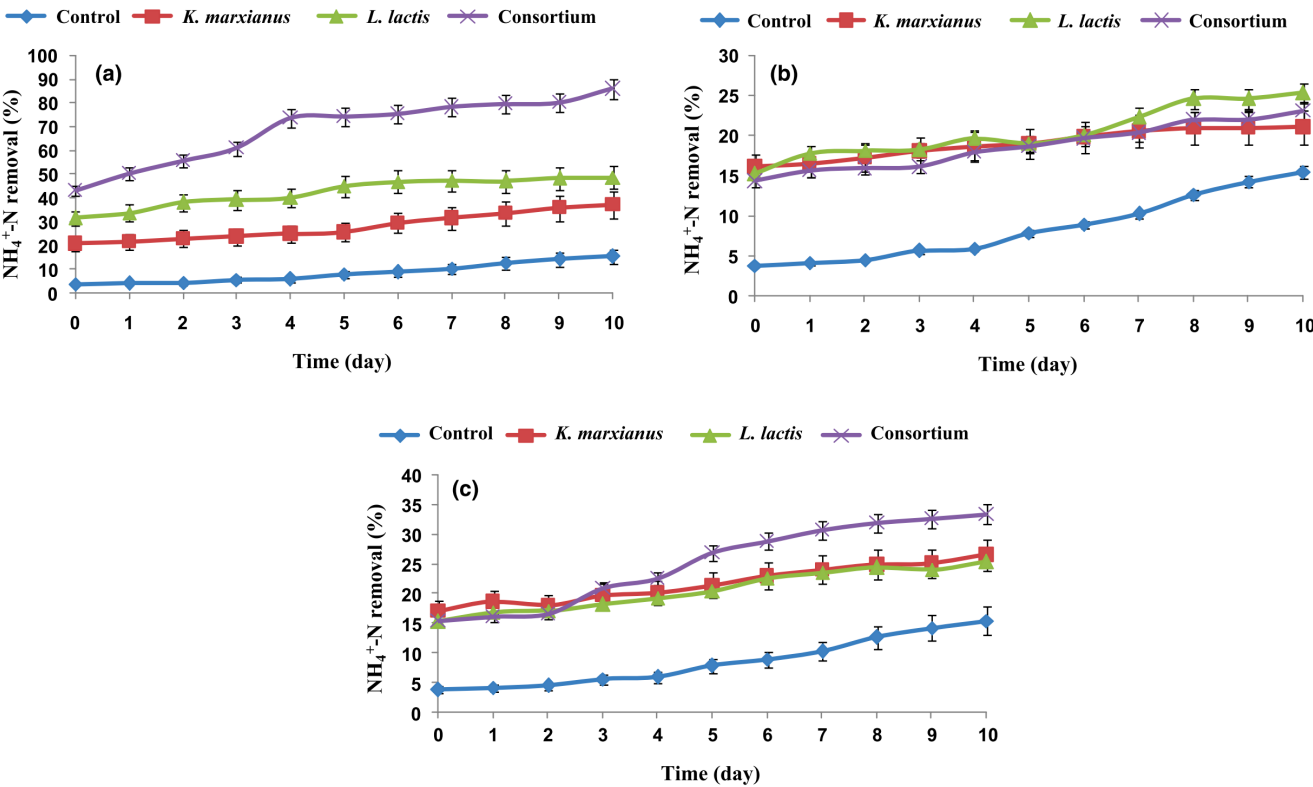




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Figure 3



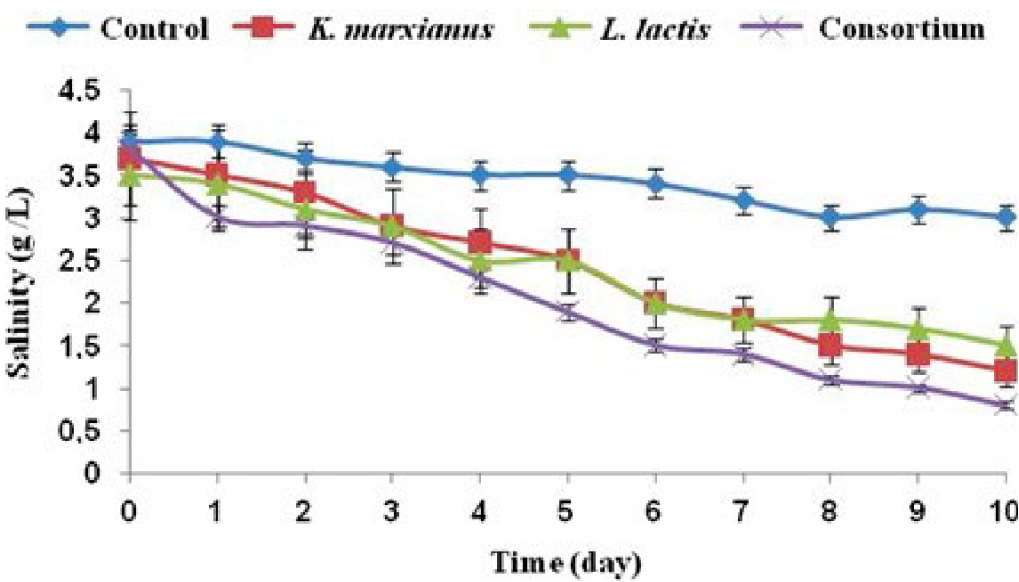
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Figure 4



517

518 **Tables**  
519

520 **Table 1** Representative polymorphic profiles of LAB and yeast colonies isolated from the Kefir grains: M,  
521 molecular marker-1,500 bp.

522

523 **Table 2** Heavy metals removal performance using single and co- culture after 10 days of treatment

524

PARAMETER	UNIT	VALUE
pH	–	7.73
COD	mg O <sub>2</sub> /L	26,200
conductivity	mS/cm	20.6
Salinity	g/L	3.62
TKN	mg/L	1,640
TDS	g/L	3.4
NH <sub>4</sub> <sup>+</sup> – N	mg/L	780
NO <sub>3</sub> <sup>-</sup>	mg/L	7.326
NO <sub>2</sub> <sup>-</sup>	mg/L	3.178
PO <sub>4</sub> <sup>2-</sup>	mg/L	28.292
Mg <sup>2+</sup>	mg/L	15.6
Ca <sup>2+</sup>	mg/L	12.3
Ni <sup>2+</sup>	mg/L	3.52
Cu <sup>2+</sup>	mg/L	1.62
Cd <sup>2+</sup>	mg/L	2.73
Pb <sup>2+</sup>	mg/L	1.78
Fe	mg/L	9.23

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STRAINS	REMOVAL RATE AFTER 10 DAYS OF TREATMENT (%)				
	NI	CU	CD	PB	FE
1% of <i>K. marxianus</i>	14.20	42.59	54.94	56.17	38.46
1% of <i>L. lactis</i>	39.77	37.03	62.63	14.04	32.50
1% of co-culture	81.53	73.45	79.48	68.53	58.17

531