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**Horses infected by Piroplasms different from *Babesia caballi* and  
*Theileria equi*: species identification and risk factors analysis in Italy**

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## 16    **Abstract**

17    Equine Piroplasmosis (EP) caused by *Theileria equi* and *Babesia caballi* is a disease affecting the  
18    health and the international movement of horses. In order to assess prevalence of Piroplasmid  
19    infection in the Northwestern part of Italy and to evaluate the associated risk factors, whole blood  
20    was collected from 135 horses from 7 different stables across the study area. PCR and sequencing  
21    were used to assess prevalence of infection and to identify detected Piroplasms to species level. A  
22    total of 23 horses (P=17.04%; CI95%: 10.70-23.38%) was found to be infected with Piroplasms and  
23    *T. equi* was the most prevalent species, found in 18 animals (P=13.33%; CI95%: 7.60%-19.07%).  
24    Although *B. caballi* was never detected, the presence of parasites belonging to the genus *Babesia*  
25    was confirmed by sequencing in 5 horses, 3 of which were infected with *B. canis* (P=2.22%; CI95%  
26    0.76%-6.33%), and 2 with *B. capreoli* (P=1.48%; CI95% 0.41%-5.24%). The natural reservoir hosts of  
27    *B. canis* and *B. capreoli* are the domestic dog and roe deer *Capreolus capreolus* respectively. These  
28    findings pose attention to the need of considering in future epidemiological and clinical studies,  
29    other Apicomplexan species as able to infect horses.

30    **Key words:** Equine Piroplasmosis, *Theileria equi*, *Babesia caballi*, *Babesia canis*, *Babesia capreoli*,  
31    Italy

## 32    **Introduction:**

33    Equine piroplasmosis (EP) is a disease that affects Equids, caused by Apicomplexan parasites  
34    *Theileria equi* and *Babesia caballi*. In endemic areas, infection occurs with varying degrees of  
35    severity from subclinical to life-threatening (Guidi et al., 2014). In Italy *Ixodes ricinus*, *Dermacentor*  
36    *marginatus* (Iori et al., 2010), *Rhipicephalus sanguineus*, *Rhipicephalus annulatus*, *Rhipicephalus*  
37    *bursa* (Scoles and Ueti, 2015) are the competent vectors of *T. equi* and *B. caballi*. *I. ricinus* has a  
38    telotrophic behavior with a host range that includes many species of birds and small to large  
39    mammals, including horses. In Italy, the abundance of *I. ricinus* and the probability of acquiring *I.*

40 *ricinus*-vectored diseases have been directly related to the abundance of Roe deer *Capreolus*  
41 *capreolus* (Rizzoli et al., 2009). Roe deer which is the main reservoir host of *Babesia capreoli*  
42 (Maladrin et al., 2010) has been expanding its presence to new areas and it increased in number  
43 throughout its entire presence area in Northern Italy (Carnevali et al., 2009) which led to a  
44 concomitant expansion of tick presence area and abundance. (Vor et al., 2010). *R. sanguineus* has  
45 the domestic dog as main host, but it can be found on a range of wild and domestic animals,  
46 horses included (Salman and Tarres-Call, 2013). In the study area *B. capreoli* is the most prevalent  
47 Piroplasmid species, infecting 43.46% of free-ranging Roe deer, Red deer and Alpine chamois  
48 (Zanet et al., 2014). Canine Piroplasmosis is also endemic in Italy as in most European countries, *B.*  
49 *canis* was molecularly identified in 2.3% of asymptomatic dogs (Cassini et al., 2009). Host-tick-  
50 pathogen relationship undergoes constant changes, mainly due to environmental, climatic and  
51 anthropogenic alterations. Recently, a horse with suspected EP was found infected with *Babesia*  
52 *canis* (Criado-Fornelio et al., 2003), while *Theileria annae*, *Theileria sergenti* and *Theileria buffeli*  
53 were isolated from horses in Italy (Moretti et al., 2010). In this context, even the epidemiology of  
54 the long known EP needs to be re-evaluated and the goal of this work is indeed to evaluate the  
55 prevalence of Piroplasms infection in horses from Northwestern Italy, to identify the species of  
56 Piroplasms involved and individual and environmental factors that might influence their infective  
57 ability.

58 **Materials and methods:**

59 Seven stables were randomly selected in the Piedmont Region, Northwestern Italy. Whole blood  
60 was collected from 135 horses and stored at -20°C until further analysis. Individuals younger than  
61 2 years were not included in the study. Total genomic DNA was extracted using PureLink Genomic  
62 DNA Mini Kit (Invitrogen, USA) following manufacturer's instructions. Direct molecular detection  
63 of *Babesia* spp./*Theileria* spp. DNA was carried out on all samples using a semi-nested PCR

64 protocol targeting the V4 hyper-variable region of the 18S rDNA as specified elsewhere (Zanet et  
 65 al., 2014). In specimens where the detection of unexpected Piroplasmid species would be  
 66 considered as a possible mistake in sample handling, we used specific PCR protocols, as reported  
 67 by their respective authors, to confirm the presence of horse DNA (primers ATPase8 and ATPase6;  
 68 Kesmen et al., 2007) and to exclude the presence of roe deer (primers 12SCC-FW and 12SCERV-  
 69 REV; Fajardo et al., 2007) and dog DNA (primers CAN-F and CAN-R; Criado-Fornelio et al., 2003). To  
 70 confirm Piroplasmid species identification , the entire 18S rRNA gene was amplified as reported by  
 71 Maladrin et al. (2010). Positive amplicons were purified using QIAQuick PCR purification kit  
 72 (QIAGEN) and directly sequenced on both DNA strands (Macrogen, The Netherlands). The  
 73 resulting sequences were compared with homologous sequences available in Genbank using the  
 74 Basic Local Alignment Search Tool (BLAST). Multiple sequence alignments were constructed using  
 75 the Clustal W algorithm and a Maximum Likelihood (ML) phylogenetic analysis was performed in  
 76 MEGA 6 using the Kimura 2-parameter model with Gamma distributed rates (Tamura et a., 2013).  
 77 Accuracy of inferred topologies were assessed via bootstrap analysis.

78 For each horse, a questionnaire was used to collect information on individual and environmental  
 79 factors that might influence their exposure to Apicomplexan parasites. The information included in  
 80 the questionnaire were: breed (Italian Saddle, Trotter, English Pureblood, mixed breed), age class,  
 81 (2-10 years, 11-20 years, >20 years), gender, activity and movements, treatment against  
 82 ectoparasites, recovering of ticks, frequency and type of deworming, province of origin, as well as  
 83 type of housing (box, paddock, both box and paddock). To identify variables associated with  
 84 *Babesia/Theileria* sp. infection we used generalized linear mixed models with PCR result was the  
 85 dichotomous response variable and stable of origin as random effect . Also we derived from a  
 86 Geographic Information System (GIS) the area occupied by pastures, forest, urbanized and  
 87 agricultural activities within 5 km from the sampled stable (area of influence of management

88 activities of the stable). The normalized area of each land-use class was included in the model.  
89 Variance inflation Factor (VIF) was used to test and avoid multicollinearity among predictors (Zuur  
90 et al., 2009). Best model selection was performed using AIC (Akaike information Criterion), while  
91 the goodness-of-fit of the final model was assessed by computing the area under the curve (AUC)  
92 of the receiver operating characteristic plots.

### 93 **Results:**

94 Among the 135 horses included in the study, 23 tested positive to semi-nested PCR (P=17.04%,  
95 IC95% 10.70-23.38). Among these, 18 horses (P=13.33%, IC95%7.60-19.07) were found to be  
96 infected with *T. equi*. *Babesia* sp. infection was detected in the remaining 5 positive horses (P=  
97 3.70%, IC95% 0.5-6.89). Detailed prevalence data are summarized in Table 1.

98 Sequencing of the entire 18S rRNA gene allowed to identify the *Babesia* isolates as belonging to *B.*  
99 *canis* (n=3, P=2.22%, IC95% 0.76-6.33; GenBank accession numbers: KX839230- KX839232) and to  
100 *Babesia capreoli* (n=2, P=1.48%, IC95% 0.41-5.24; GenBank accession numbers: KX839233  
101 KX839234). Statistical support to species identification was given through ML phylogenetic analysis  
102 (Fig.1). The PCR protocols used to confirm the presence of horse DNA (all samples tested positive)  
103 and the absence of dog/roe deer DNA (all samples tested negative) excluded the possibility of a  
104 wrong diagnosis due to mishandling of the samples. The presence of *T. equi* DNA was detected  
105 and confirmed by sequencing in 18 horses.

106 The risk factor analysis was carried out on the basis of data collected using the questionnaire and  
107 on data deriving from the GIS analysis. The covariates retained by VIF analysis and therefore used  
108 to train the model were: sex, age class, housing, activity, movements, province of origin and the  
109 GIS environmental parameters. The covariates included in the model with the highest predicting  
110 capability (AUC=0.76) are: activity (daily recreational walks in forests and bush) which was  
111 positively associated to higher risk of infection (p<0.01; OR= 2.3, IC95%= 1.2-3.4), as well as age

112 (animals from 2 to 10 years resulting more affected by Piroplasmid infection,  $p < 0.05$ ; OR= 1.9,  
113 IC95%= 1.2-2.5). Regular deworming was instead negatively associated to infection ( $p < 0.05$ ;  
114 OR=0.25, IC95%= 0.1-0.75).

#### 115 **Discussion:**

116 EP in horses is traditionally associated to infection with *T. equi* and with *B. caballi*. *T. equi* is  
117 worldwide more prevalent than *B. caballi*. (Bruning, 1996; Friedhoff et al., 1990). In Italy, *T. equi*  
118 prevalence of infection ranges from 8.2% (Grandi et al., 2011) to 82.8% (Zobba et al., 2008), while  
119 *B. caballi* reaches 31.5% (Torina et al., 2007). Our results fall within the range of infection already  
120 reported for the country, even though to our knowledge, this is the first study dealing with horses  
121 in Northwestern Italy. The most prevalent Piroplasm was *T. equi* while no *B. caballi* was detected  
122 in any of the sampled horses. Atypical *Babesia* species: *B. canis* and *B. capreoli* were detected with  
123 a prevalence of 3.70%. These species have been associated to infection in the domestic dog and  
124 Roe deer respectively (Criado Fornelio et al., 2003; Zanet et al., 2014). Two of the 3 isolates of *B.*  
125 *canis* detected in the current study, are identical to each other (GenBank Accession n.: **KX839230**,  
126 **KX839231**) and match perfectly with the *B. canis* (GenBank Accession n.: **AY150060**) detected by  
127 Criado Fornelio et al. (2003) in a horse with suspected EP and with isolates deposited in GenBank  
128 (Accession numbers: **EU165369**, **AY962187**, **AY703073**, **AY321119**, **KC902833**, **KC593879**) which  
129 were isolated from dogs, foxes or *R. sanguineus* ticks from Continental Europe (Poland, The  
130 Netherlands, France) and Siberia. For what concerns *B. capreoli*, the two positive horses were  
131 found infected with identical (100% identity, 100% query coverage) strains, that also perfectly  
132 matched the parasites isolated in the same area, from roe deer (GenBank Accession n.: **KF773723**)  
133 and red deer (GenBank Accession n.: **KF773718**) (Zanet et al., 2014). Risk factor analysis evidenced  
134 how young animals (from 2 to 10 years old) and recreational horses that are ridden in the  
135 countryside, are more exposed to infection with *Theileria/Babesia*. Higher rates of infections in

136 young animals are a common finding in endemic areas, where animals are exposed to EP causal  
137 agents at young age and eventually clear the infection in the following years (Scoles and Ueti,  
138 2015; Farkas et al., 2013). As previously reported (Peckle et al., 2013), our data evidence how horse  
139 activities that increase the exposure of animals to infected ticks enhance the risk of infection. In  
140 the present study recreational horses were significantly more exposed to Piroplasms infection  
141 than racing/breeding horses. Regular deworming was identified as protective factor, as horses that  
142 are regularly dewormed have statistically lower prevalence of infection. As confirmed by other  
143 studies (Guidi et al., 2014, Moretti et al. 2010; Santos et al. 2011; Vieira et al. in 2013) deworming  
144 like vaccination, and flies control, are indicators of good herd management practices and may  
145 reflect a reduced exposure to tick infestation and thus to a lower risk of infection. Among the risk  
146 factors that were taken into account none of the geographic covariates were able to efficiently  
147 discriminate the risk of infection. The number of horses infected with *B. canis* or *B. capreoli* was  
148 not sufficient to make inference on significant risk factors specific for these atypical *Babesia*  
149 species. **Conclusions**

150 EP is a disease of clinical and economical relevance worldwide. Despite the efforts made to control  
151 the geographic expansion of the disease, the epidemiology of Piroplasms infection is constantly  
152 evolving. The results of this work evidence how horses can be infected with Piroplasms species  
153 different from *T. equi* and *B. caballi*. Piroplasmosis, as well as numerous vector-borne diseases  
154 have been experiencing both a territorial expansion and drastic changes in host-vector  
155 relationship. Economic, social and bioclimatic changes are causing an always increasing contact  
156 among wildlife, humans and domestic animals (Daszak et al., 2001) and the role of wildlife as  
157 source of diseases is to be specially monitored (Daszak et al., 2000). Further effort is required to  
158 understand the clinical impact of atypical *Babesia* infections in horses, but clinicians, official  
159 veterinary authorities and research institutions should account for the possibility of horses being



160 infected with species of Piroplasms different from *B. caballi* and *T. equi* and should adapt  
161 diagnostic tools accordingly.

## 162 **Authors' contributions**

163 SZ performed the statistical analysis, molecular testing and prepared the manuscript, BM collected  
164 blood samples, administered the questionnaires and performed analysis, AT performed the  
165 molecular analysis, IT collected blood samples and connected to owners of the horses, EF designed  
166 and coordinated the study. All authors read and approved the final version of the manuscript.

## 167 **Conflict of interests statement**

168 The authors declare that they have no competing interests.

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## 175 **References**

- 176 Bruning, A. 1996. Equine piroplasmosis an update on diagnosis, treatment and prevention. Br. Vet.  
177 J. 152, 139–51.
- 178 Carnevali, L., Pedrotti, L., Riga, F., Toso, S., 2009. Banca Dati Ungulati: Status, distribuzione,  
179 consistenza, gestione e prelievo venatorio delle popolazioni di Ungulati in Italia. Ed: ISPRA -  
180 Biologia e conservazione della fauna, 117.
- 181 Cassini, R., Zanutto, S., Frangipane di Regalbono, A., Gabrielli, S., Calderini, P. Moretti, A., Tampieri,  
182 M.P., Pietrobelli, M., 2009. Canine piroplasmosis in Italy: epidemiological aspects in vertebrate and  
183 invertebrate hosts. Vet. Parasitol., 165, 30–35.
- 184 Criado-Fornelio, A., Martinez-Marcos, A., Buling-Sarana, A., Barba-Carretero. J.C., 2003. Molecular  
185 studies on *Babesia*, *Theileria* and *Hepatozoon* in southern Europe. Part I. Epizootiological aspects.  
186 Vet. Parasitol., 113, 189–201.
- 187 Daszak, P., Andrew, A., Hyatt, A.D., Cunningham, A.A., 2000. Emerging infectious diseases of  
188 wildlife-- threats to biodiversity and human health. Science 287 (5452), 443-449.

189 Daszak, P., Cunningham, A.A., Hyatt A.D., 2001. Anthropogenic environmental change and the  
190 emergence of infectious diseases in wildlife. *Acta Trop.* 78, 103-116.

191 Fajardo V., Gonzalez I., Lopez-Calleja I., Martin I., Rojas M., Hernandez P.E., Garcia T., Martin R.,  
192 2007. Identification of meats from red deer (*Cervus elaphus*), fallow deer (*Dama dama*), and roe  
193 deer (*Capreolus capreolus*) using polymerase chain reaction targeting specific sequences from the  
194 mitochondrial 12S rRNA gene. *Meat Sci* 76, 234–240.

195 Farkas, R., Tánczos, B., Gyurkovszky, M., Földvári, G., Solymosi, N., Edelhofer, R., Hornok, S., 2013.  
196 Serological and molecular detection of *Theileria equi* infection in horses in Hungary. *Vet. Parasitol.*  
197 192 (1–3), 143–148.

198 Friedhoff, K.T., Tenter, A.M., Muller, I. 1990. Haemoparasites of equines: impact on international  
199 trade of horses. *Rev. Sci. Tech.* 9, 1187–94.

200 Grandi, G., Molinari, G., Tittarelli, M., Sasser, D., Kramer L.H., 2011. Prevalence of *Theileria equi*  
201 and *Babesia caballi* infection in Horses from Northern Italy . *Vector-borne zoonot.*, 11(7), 955-956.

202 Guidi, E., Pradier, S., Lebert, I., Leblond, A., 2014. Piroplasmosis in an endemic area: analysis of the  
203 risk factors and their implications in the control of Theileriosis and Babesiosis in horses. *Parasitol*  
204 *Res* 114(1), 71-83.

205 Iori, A., Gabrielli, S., Calderini, P., Moretti, A., Pietrobelli, M., Tampieri, M.P., Galuppi, R., Cancrini,  
206 G., 2010. Tick reservoirs for piroplasms in central and northern Italy. *Vet Parasitol* 170, 291–296.

207 Kesmen Z., Sahin F., Yetim H., 2007. PCR assay for the identification of animal species in cooked  
208 sausages. *Meat Sci* 77(4), 649–653.

209 Maladrin, L., Jouglin, M., Sun, Y., Brisseau, N., Chauvin, A., 2010. Redescription of *Babesia capreoli*  
210 (Enigk and Friedhoff, 1962) from roe deer (*Capreolus capreolus*): isolation, cultivation, host  
211 specificity, molecular characterization and differentiation from *Babesia divergens*. *Int J Parasitol*,  
212 40, 277–284.

213 Moretti, A., Tampieri, M.P., Gabrielli, S., Moretti, I., Torina, A., Scoccia, E., Torina A., Moretti,  
214 I., Gabrielli, S., Tampieri, M.P., Pietrobelli, M., 2010. Prevalence and diagnosis of *Babesia* and  
215 *Theileria* infections in horses in Italy: a preliminary study. *Vet J* 184(3), 346–350.

216 Peckle, M., Pires, M.S., dos Santos, T.M., Roier, E.C.R., da Silva, C.B., Vilela, J.A.R., Santos, H.A.,  
217 Massard, C.L., 2013. Molecular epidemiology of *Theileria equi* in horses and their association with  
218 possible tick vectors in the state of Rio de Janeiro, Brazil. *Parasitol Res* 112, 2017–2025.

219 Rizzoli, A.P., Hauffe, H.C., Tagliapietra, V., Neteler, M., Rosa, R., 2009. Forest Structure and Roe  
220 Deer Abundance Predict Tick-borne Encephalitis Risk in Italy. *PLoS ONE* 4(2), e4336.  
221 doi:10.1371/journal.pone.0004336.

222 Salman, M., and Tarres-Call, J., 2013. Ticks and Tick-borne diseases. Geographical distribution and  
223 control strategies in the Euro-Asia region, first ed. CAB International, Boston, USA.

224 Santos, T.M., Machado, R.Z., Baldani, C.D., Almeida, F.Q., Morales, L.M., Vilela, J.A. Brito Moraes,  
225 L.M.,; Almeida, F.Q., Baldani, C.D., Machado, R.Z., Massard, C.L., 2011. Factors associated to

226 *Theileria equi* in equids of two microregions from Rio de Janeiro, Brazil. Rev Bras Parasitol Vet  
 227 20(3), 235–241.

228 Scoles G.A. and Ueti M.W. 2015. Vector Ecology of Equine Piroplasmosis. Annu Rev Entomol 60,  
 229 561–80.

230 Tamura K., Stecher G., Peterson D., Filipski A., and Kumar S., 2013. MEGA6: Molecular Evolutionary  
 231 Genetics Analysis version 6.0. Molec. Biol. Evol. 30, 2725-2729.

232 Torina, A., Vicente, J., Alongi, A., Scimeca S., Turlá, R., Nicosia, S., Di Marco, V., Caracappa, S., De La  
 233 Fuente, J., 2007. Observed prevalence of tick-borne pathogens in domestic animals in Sicily, Italy  
 234 during 2003–2005. Zoonoses Public Hlth 54(1), 8–15.

235 Vieira, R.F., Biondo, A.W., Nascimento, D.D., Vieira, T.S., Finger, M.A., Sicupira, P.M., Dutra, L.H.,  
 236 Deconto, I., Barros-Filho, I.R., Dornbusch, P.T., Biondo, A.W., Vidotto, O., 2013.  
 237 Seroepidemiological survey of *Theileria equi* and *Babesia caballi* in horses from a rural and from  
 238 urban areas of Parana State, southern Brazil. Ticks Tick Borne Dis 4(6), 537–541.

239 Vor, T., Kiffner, C., Hagedorn, P., Niedrig, M., Ruhe, F., 2010. Tick burden on European roe deer  
 240 (*Capreolus capreolus*). Exp Appl Acarol 51, 405–417.

241 Zanet, S., Trisciuglio, A., Bottero, E., Fernández de Mera, I.G., Gortazar, C., Carpignano, M.G.,  
 242 Ferroglio, E., 2014. Piroplasmosis in wildlife: *Babesia* and *Theileria* affecting free-ranging ungulates  
 243 and carnivores in the Italian Alps. Parasites Vector 7,70.

244 Zobba, R., Ardu, M., Niccolini, S., Chessa, B., Manna, L., Cocco, R., Pinna, P., 2008.  
 245 Clinical and laboratory findings in equine piroplasmosis. J Equine Vet Sci 28(5)  
 246 doi:10.1016/j.jevs.2008.03.005.

247 Zuur, A. F., Ieno, E. N., Walker, N., Saveliev, A. A. & Smith, G. M., 2009. Mixed effects models and  
 248 extensions in ecology with R, 1st edition. Springer, New York.

249 **Captions to figure:**

250 Fig. 1 Molecular Phylogenetic analysis by Maximum Likelihood method based on the Kimura 2-  
 251 parameter model of 18S rRNA gene of 15 *Babesia* spp. and *Theileria* spp. sequences. Isolates from  
 252 this study are indicated with a black arrow.