Leafhopper feeding behaviour on three grapevine cultivars with different susceptibilities to Flavescence dorée

Scaphoideus titanus Ball feeding behaviour on three grapevine cultivars with different susceptibilities to Flavescence dorée

Authors

Matteo Ripamonti[1,2], Federico Maron[1a], Daniele Cornara[3b], Cristina Marzachì[2], Alberto Fereres[3], Domenico Bosco[1*]

Affiliation

[1] Department of Agriculture, Forest and Food Sciences (DiSAFA), University of Turin, Grugliasco (TO), Italy; [2] Institute for Sustainable Plant Protection - National Research Council of Italy, IPSP-CNR, Torino, Italy; [3] Instituto de Ciencias Agrarias, Consejo Superior de Investigaciones Cientificas, ICA-CSIC, Madrid, Spain

[a] Present address: Vit.En. Centro di Saggio, Calosso (AT), Italy.

[b] Present address: University of Berkeley, Department of Environmental Science, Policy, and Management, Berkeley, CA, USA.

* Corresponding author: domenico.bosco@unito.it

Abstract

Scaphoideus titanus (Ball) is a grapevine-feeder leafhopper, and the most important vector of Flavescence dorée of grapevine (FD), a disease associated with phytoplasmas belonging to ribosomal subgroups 16Sr-V–C and –D. FD is a major constraint to viticulture in several European countries and, so far, its control has relied on roguing of infected plants and insecticide applications against the vector. Detailed knowledge on different levels of the multifaceted phytoplasma-plant-vector relationship is required to envisage and explore more sustainable ways to control the disease spread. In the present work, *S. titanus* feeding behaviour was described on three grapevine cultivars: Barbera (susceptible to FD), Brachetto, and Moscato (tolerant to FD) using the Electrical Penetration Graph (EPG) technique. Interestingly, no differences were highlighted in the non-phloem probing phases, thus suggesting that the tested cultivars have no major differences in the biochemical composition or structure of the leaf cuticle, epidermis or mesophyll, that can affect the first feeding phases. On the contrary, the results showed significant differences in leafhopper feeding behaviour in terms of the duration of the phloem feeding phase, longer on Barbera and shorter on Brachetto and Moscato, and of the frequency of interruption-salivation events inside the phloem, higher on Brachetto and Moscato. These

findings indicate a preference for the Barbera variety, that appears a more suitable hosts for the leafhopper. *Scaphoideus titanus* feeding behaviour on Barbera correlates with an enhanced FDp transmission efficiency, thus explaining, at least in part, the higher susceptibility of this variety to FD. The mechanisms for the non-preference for Brachetto and Moscato are discussed, and a possible antixenosis is hypothesized. We propose that breeding for resistance against FD should take into account both plant traits associated with the response to the phytoplasmas and to the vector.

Keywords

EPG; Electrical Penetration Graph; leafhopper vector; *Vitis vinifera*; cultivar

1. Introduction

The leafhopper *Scaphoideus titanus* Ball is the main vector of phytoplasmas associated with the Flavescence dorée of grapevine (FD), a disease spread in most European viticultural countries (EFSA, 2020) that causes severe reduction of yield and quality of grapes, requires roguing of infected plants and leads to uneven-aged vineyards (Morone et al., 2007). FD is associated with phytoplasmas belonging to the 16SrV group, subgroups –C and –D (Davis and Dally, 2001; Lee et al., 2004; Martini et al., 2002), and it causes severe losses to European viticulture (EFSA, 2016). Although different insect species are competent for the transmission of FD phytoplasmas (FDp), *S. titanus* is by far the most important vector, being strictly associated with *Vitis* plants and thus sustaining both primary (from wild grapevines outside the vineyards to cultivated vines) and secondary (from vine to vine within the vineyard) disease spread (Maggi et al., 2017; Ripamonti et al., 2020). Control of FD relies on prophylactic measures, such as the use of healthy propagation material, as well as on compulsory measures in infected vineyards, namely roguing of infected plants, and insecticide treatments against the vector (Bosco and Mori, 2013). However, the large-scale application of insecticides is a concern to human health and environment, priming cascade ecosystem effects (Desneux et al., 2007) with a strong negative impact on pollinators (Tosi et al., 2018). For this reason, recent studies focused on identifying sources of resistance to FDp phytoplasmas within the grapevine germoplasm (Eveillard et al., 2016; Ripamonti et al., 2021), that would represent the best strategy to minimise damage and limit FD spread and insecticide applications. Grapevine tolerance to FDp may be due to a direct response of the plant against the pathogen or mediated by some resistance against the vector, or by a combination of the

two. Resistance against insects occurs when plant structural or chemical traits deter herbivore feeding and thus minimize the amount of herbivore damage experienced by the plant, while tolerance occurs when plant traits reduce the negative effects of herbivore damage on crop yield (Mitchell et al., 2016). As an example, it was demonstrated that resistant tea cultivars sustained lower phloem ingestions for *Empoasca vitis* (Miao et al., 2014). Moscato and Brachetto are grapevine varieties tolerant to FD, as demonstrated by Ripamonti et al. (2021) using transmission experiments with *S. titanus* under controlled conditions (Ripamonti et al., 2021). The reduced *S. titanus* survival on Moscato observed by the above mentioned authors, suggest that vector-host interaction could be the pivotal factor underlying Moscato tolerance to FD. *S. titanus* is monophagous on *Vitis* species, mainly *Vitis vinifera* and naturalized rootstocks of *V. riparia* in Europe, while in North America, *V. labrusca* and *V. riparia* are reported as the preferred host plants (Chuche and Thiéry, 2014). Although the species is regarded as monophagous, it shows a good level of plasticity and can feed on plant species of different families, e.g. Vitaceae, Fabaceae, Ranunculaceae (Caudwell et al., 1970; Trivellone et al., 2013). Plant resistance against sap-sucking insects can be conveniently investigated by Electrical Penetration Graph (EPG), that describes the nutrition pattern of a sucking insect on a given plant genotype, by identifying possible altered nutrition on non-suitable genotypes (Backus et al., 2020; Lucini et al., 2021).

Here we expand the first findings on *S. titanus* behaviour on grapevines, by analyzing the vector probing behavior on three varieties with a different degree of susceptibility to FD: one susceptible, Barbera, and two tolerant, Moscato and Brachetto, through the Electrical Penetration Graph (EPG) (Backus and Bennett, 2009; McLean and Kinsey, 1964; Tjallingii, 1978).

EPG is a powerful tool to describe pierce-sucking insects' probing behaviour, previously applied to describe *S. titanus* feeding behaviour on Cabernet-Sauvignon cuttings (Chuche et al., 2017a, 2017b). EPG studies on different plant cultivars/genotypes provide precious information for the epidemiology of vector-borne plant pathogens, also permitting the identification of traits making a *Vitis* genotype unsuitable for the vector. A number of EPG studies aimed at identifying plant resistance to insect vectors have been performed on planthoppers (Kimmins, 1989), whiteflies (Jiang et al., 2001; Rodríguez-López et al., 2011) and aphids (Caillaud et al., 1995a, 1995b; Garzo et al., 2018; Sauge et al., 1998). Among these latter, EPG was applied to identify resistance factors involved in virus transmission inhibition (Chen et al., 1997) as well as the presence of antixenosis (Kordan et al., 2019).

Besides those on *S. titanus* (Chuche et al., 2017a, 2017b), few EPG studies have been conducted on Deltocephalinae leafhoppers (Carpane et al., 2011; Kawabe and McLean, 1980; Lett et al., 2001; Stafford and Walker, 2009; Trębicki et al., 2012), and very few of these are relevant to phytoplasmas/mollicutes transmission (Carpane et al., 2011; Chuche et al., 2017a).

The aim of this study was to compare *S. titanus* probing behaviour on three different grapevine cultivars characterised by different susceptibilities to FD, in order to better characterize the mechanisms underlying varietal tolerance/susceptibility to this phytoplasma disease.

2. Material & Methods

2.1. *S. titanus* collection and rearing

To establish a *S. titanus* laboratory colony, in January/February 2019 two-year-old grapevine canes with eggs were collected in vineyards of the Piemonte Region. The selected sites were known to host a high population of the leafhopper in the previous summer, as estimated by yellow sticky traps captures of adults. The collected canes were stored in a cold room at 6±1°C, covered with plastic film to avoid egg desiccation, until use. When needed, grapevine canes were transferred into an insect-proof greenhouse at 24 ± 2°C and maintained damp by daily water spraying. After four weeks, canes were isolated in a cage together with a three-week-old broadbean plant as a food source for the nymphs. After egg hatching, the broadbean plant was replaced every three weeks. Nymphs were reared under controlled conditions inside a greenhouse chamber, T = 24 ± 2°C, with no humidity and photoperiod control, from the beginning of April to the end of September 2019. As FDp is not transovarically transmitted, and all the plants used for the rearing and the experiments were phytoplasma-free, all *S. titanus* used in the experiments were phytoplasma-free. For the EPG experiments, adults emerged from 7-21 days were used (modified from Chuche et al., 2017a), since in this time frame they were sexually mature, highly active and not subjected to high mortality (Bocca et al., 2020; Mazzoni et al., 2009).

2.2. Plant rearing

The test plants were obtained from phytoplasma-free *V. vinifera* cuttings of three different cultivars, Barbera N. - Clone I-AT 84, Brachetto N. - Clone I-CVT 20 and Moscato Bianco B. - Clone I-CVT 190 as described in Ripamonti et al. (2021). Grapevine cuttings were

grown in a greenhouse at 24 ± 2°C, with no humidity and photoperiod control, inside 0.9 L pots (2:2:1 topsoil, clay, perlite), and watered once a week. Cuttings were used when three- to five-months old, and periodically pruned in order to keep them within 80 cm height . Broadbean plants used for *S. titanus* rearing were seedlings maintained in a growth chamber (24 ± 2°C, with no humidity and photoperiod control) in 2.4 L topsoil, five per pot, and watered twice a week.

2.3. EPG setup and data analysis

Selected adults were collected and anesthetised with carbon dioxide for 5 seconds in a glass tube, then immobilised at the edge of a cut pipette tip connected to a vacuum pump under a stereomicroscope. A small drop of water-based silver glue (EPG Systems, Wageningen, The Netherlands) was placed on the pronotum of the insect, then a gold wire of 18 µm (previously attached with solvent-based silver glue (Ted Pella Inc., USA) to a 3 cm copper wire in turn attached to a brass nail with melted stain) was positioned on the dried drop, and covered with another small drop of silver glue. Before the EPG assay, insects were starved for a 30-minute period, during which they were attached to the electrode and hanged, inserting the nail in a polystyrene base.

The substrate voltage probe was inserted in well damped soil of a potted grapevine cutting, and *S. titanus*, attached to the assembled electrode, was connected to a probe and positioned onto the abaxial surface of a leaf. The feeding behaviour was then monitored for 8 hours with a Giga-8dd DC-EPG amplifier (EPG Systems, Wageningen, The Netherlands), inside a Faraday cage to isolate the system from external electrical noise. Input resistance used was 1 giga Ohm, output set at 75x gain and plant voltage adjusted so that the EPG signal fitted into +5V and -5V. All recordings were done between June and August 2019, and started between 11:00 and 11:30 a.m. every day.

A total of 153 recordings were done, each day a total of 6 recording were run. Each single recording was represented by a different plant-insect combination, one male or one female on one grapevine plant. Potted plants of the three varieties were randomly arranged in the Faraday cage for every recording and discarded after use. In case of falling from the leaf, the insect was repositioned. At the end of the recording, dead insects were noted and excluded from further analyses.

2.4. EPG acquisition and marking of EPG files

Recordings were acquired and marked using Stylet+ software (v01.30, Electrical Penetration Graph Data Acquisition and Analysis, EPG Systems, Wageningen, The Netherlands). Waveform marking was conducted accordingly to Chuche et al. (2017a) and Stafford & Walker (2009), focusing on the following waveforms: np (non-probing activity), pathway-phase (phase "C"), active ingestion (phase "G") of mesophyll (<60 seconds) or xylem sap (>100 seconds) (see Stafford & Walker, 2009), passive ingestion of phloem sap (phase E), interruptions during ingestion (phase N of Chuche et al., 2017a). For more details, see Supplementary File S1.

Once marked, all the recordings were singly selected for the successive analysis. In particular, recordings with electrical noises, bad electric connections, or when insects fell from the plant for more than 20% of the recording time, were discarded from further analysis.

2.5. Statistical analysis

All the statistical analyses were conducted on R software v4.0.3 (R Core Team., 2020). Selected recordings were analysed through a package of the software R ad-hoc produced for the analysis on EPG recordings, called Rwaves (Chiapello, https://github.com/mchiapello/Rwaves). Rwaves conducts summary statistics on the input recordings on a set of variables of EPG analysis (Table 1), producing a table including the values of all the variables for all the input recordings. The resulting table was composed as follows: every row corresponded to a single recording (represented by the unique combination of one leafhopper and one grapevine plant), while every column represented a single EPG variable. Once obtained, the table was subjected to modifications to enhance readability (packages dplyr, tidyr, stringr: (Wickham, 2019, 2020; Wickham et al., 2020), and descriptive statistics were run (Tables 3, 4, 5, 6 and Supplementary File S2). Univariate analyses were conducted starting from Generalised Linear Model (GLM) of different families specific for the nature of the variable: quasi-Poisson or negative-binomial for counts, Gamma or inverse-Gaussian for positive continuous variables, beta-regression for proportions (packages stats, betareg, MASS: Cribari-Neto and Zeileis, 2010; Venables and Ripley, 2002). Goodness-of-fit for every model was evaluated plotting half-normal plots with simulated envelope against deviance residuals, with 95% confidence level (hnp package: Moral et al., 2017). Homoscedasticity for every model was evaluated through Levene's test (car package: Fox and Weisberg, 2019). In case of rejection of the null hypothesis, heteroscedasticity-consistent standard errors (sandwich package: Zeileis et

al., 2020) were calculated and considered for pairwise-comparisons. Comparisons among groups were conducted with least-square means method and Tukey method for p-value adjustment, at 0.05 significance level and 95% confidence intervals (packages emmeans and multcomp: Hothorn, Bretz, & Westfall, 2008; Lenth, 2020). Cultivar, Sex, and their reciprocal interaction were selected as explanatory variables. If no significant effects were found for Sex and Cultivar × Sex, the GLM was run with Cultivar as the only explanatory variable. GLMs summaries were reported in Supplementary File S3 using package jtools (Long, 2020). Packages ggplot2 (Wickham, 2016) and ggpubr (Kassambara, 2020) were used to produce Figure 1, and Supplementary File S4 and S5.

A multivariate Canonical Correspondence Analysis (CCA, Legendre & Legendre, 2012) was conducted through the vegan (Oksanen et al., 2019) and ggordiplots packages (Quensen, 2018), considering all the variables except multi-collinear ones, that were excluded from the analysis, based on a correlation coefficient higher than 0.95 (usdm package: Naimi et al., 2014), in order to strengthen the predictor value of the model. Starting from 25 variables, 5 variables were found to have collinearity problem, and were thus excluded from further analyses. The remaining variables were standardised (Hellinger method, Legendre & Gallagher, 2001) and subjected to CCA, with Cultivar, Sex and their interaction as explanatory variables. The CCA result was confirmed through a permutational Multivariate Analysis of Variance (perMANOVA; Anderson, 2001).

The complete R code will be made publicly available on GitHub (https://github.com/matteo-rpm).

Table 1. EPG variable selected for the study.

Variable	Abbreviation from Sarria et al., 2009 (implemented in Rwaves)	Abbreviation from Backus et al., 2007	Type (NS: non-sequential; S: sequential)
"Number of non-probing periods"	n_np	NWEi np	NS
"Total duration of non-probing periods [s]"	s_np	WDi np	NS
"Time from 1st np to 1st probe [s]"	s_npto1stprobe	-	S
"Duration of the 2nd non-probing period [s]"	s_2np	-	S
"Number of probes"	n_Pr	NPi	NS
"Total probing time [s]"	s_Pr	PDi	NS
"Total duration of pathway phase [s]"	s_C	WDi C	NS
"Number of active ingestion phases"	n_G	NWEi G	NS
"Total duration of active ingestion [s]"	s_G	WDi G	NS
"Number of phloem ingestions"	n_E2	NWEi E2	NS

"Number of sustained (> 600 s) phloem ingestion"	n_sE2	NWEi sE2	NS
"Total duration of phloem ingestions [s]"	s_E2	WDi E2	NS
"Mean duration of a single event of phloem ingestion [s]"	mean_E2	WDEi E2	NS
"Duration of the longest phloem ingestion [s]"	s_longestE2	-	NS
"Total duration of non-phloematic phases [s]"	s_notE	WDi C-G	NS
"Time from 1st probe to 1st phloem ingestion [s]"	t_1E2.exp	-	S
"Time from 1st probe to 1st sustained (> 600 s) phloem ingestion [s]"	t_1sE2.exp	-	S
"Time of 1st sustained phloem phase [s]"	t_1st_sE2	-	S
"Percentage of probing time spent in phloem ingestion [%]"	percprobtime_E2	-	NS
"Percentage of probing time spent in pathway-phase [%]"	percprobtime_C	-	NS
"Percentage of probing time spent in active ingestion [%]"	percprobtime_G	-	NS
"Potential E2 index [%]"	E2index	-	S
"Mean frequency of Np interruptions during phloem phase [mHz]"	mean_fr_Ninterrup	-	NS
"Percentage of time spent in Np interruption during phloem phase [%]"	percNinterrup_E2	-	NS
"Number of Np interruptions during phloem ingestion"	n_Ninterrupt_E2	NWEi Np	NS

3. Results

Number of recordings obtained from male and female *S. titanus* adults on the three grapevine varieties are summarised in Table 2. In particular, 51-cultivar specific recordings were acquired, of which a fraction was selected for further analysis (31 for Barbera, 32 for Brachetto, 37 for Moscato), as described in Material and Methods section.

Table 2. Number of total and selected recordings of *S. titanus* feeding behaviour on three grapevine cultivars.

Cultivar	Total recordings (females, males)	Selected recordings (females, males)
Barbera	51 (25, 26)	31 (18, 13)
Brachetto	51 (25, 26)	32 (13, 19)
Moscato	51 (25, 26)	37 (16, 21)

No significant differences in acquiring successful EPG signals among cultivars, possibly caused by human errors, were found (Pearson's Chi-squared test, X-squared = 0.36811, df = 2, p-value = 0.8319). From now on, when referring to recordings, only the selected ones will be considered, unless otherwise stated.

Irrespective of the cultivar, most of the insects started probing within the first minute from their access to the leaf (median ± SE = 41 ± 21 s). Waveforms were graphically summarised in a temporal progress representation (Figure 1). An overall larger area of phloem phase was found for leafhoppers feeding on the Barbera variety.

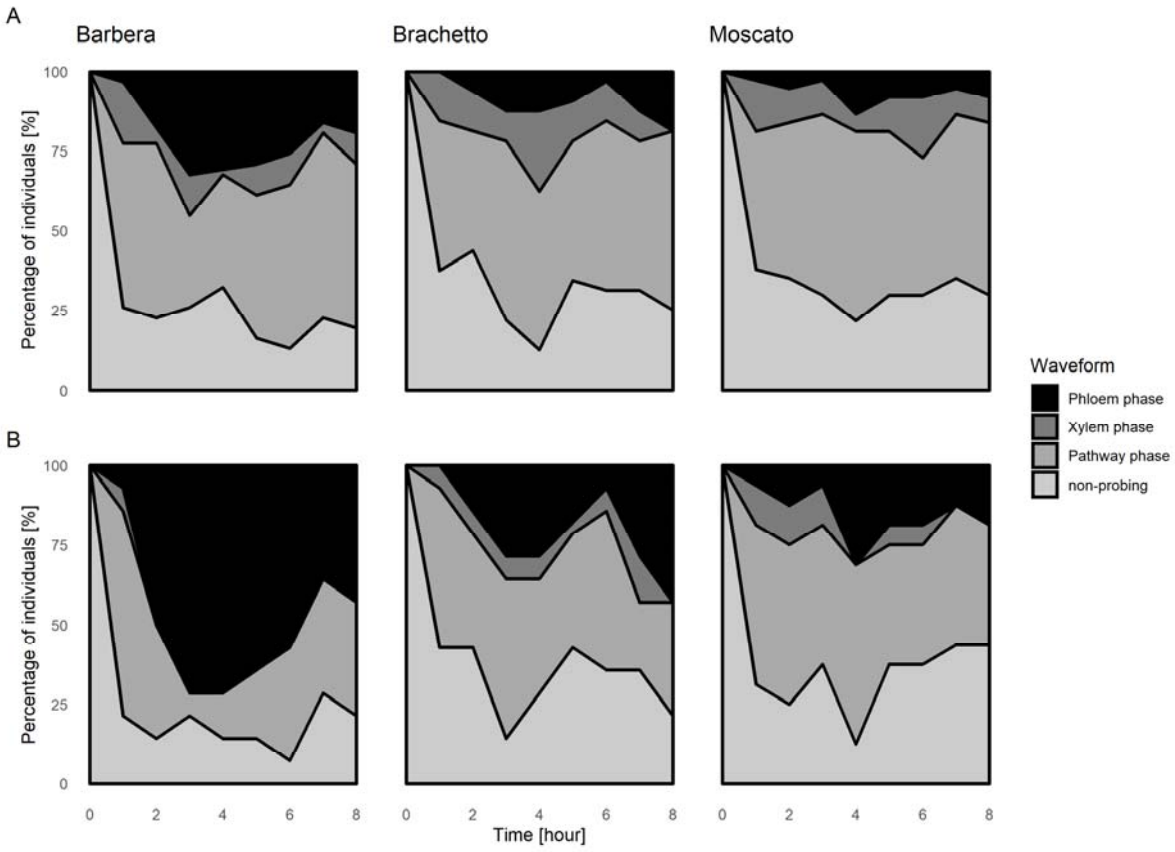


Figure 1. Temporal progress of *S. titanus* stylets activities on three grapevine cultivars during the 8-h EPG recording. Probing behaviours were represented as percentages of leafhoppers in a given phase (non-probing, pathway phase, active ingestion of xylem sap, passive ingestion of phloem sap) at 1 h intervals, starting from hour 0 (start of the recording) to hour 8 (end of the recording). a) Graphs produced considering all recordings; b) graphs produced considering only recordings where a phloem phase was present. The total number of recordings used to produce Figure 1 are reported in the third column of Supplementary File S2 (all recordings) and Table 3 (phloem recordings).

Values of non-phloem variables of all selected recordings are reported in Supplementary File S2. No differences were identified in the variables among the three grapevine varieties. The proportion of recordings with phloem phases were not significantly different among cultivars (Supplementary File S6, Pearson's Chi-squared test, X-squared = 0.026378, df = 2, p-value = 0.9869), as almost half of the recordings (45% for Barbera, 44% for Brachetto, 43% for Moscato) showed phloem phases, irrespective of the cultivar (Supplementary File S6). No differences were highlighted among cultivars for all the non-phloem variables (Supplementary File S5), when considering the recordings without a phloem-phase. Further, the non-phloem variables were analysed for recordings with phloem phases (Table 3). Number of events and their duration for the non-phloem phases did not differ among groups (Table 3). Interestingly, the total time spent by the insect with stylets inserted in the plant tissues ("Total probing time") were similar among the three *Vitis* genotypes. Some differences were found for the related variables "Number of non-probing periods", and "Number of probes", as higher values were recorded for both variables on Brachetto, compared to Barbera. On Barbera, females showed fewer "Number of active ingestion (from mesophyll or xylem) phases" than males. No differences were observed between sexes on the other varieties. No significant differences among cultivars were found for the "Number of phloem ingestions", or for the "sustained" (longer than 10 minutes) ones (Table 4). Although the "Mean duration of a single event of phloem ingestion" did not differ significantly among cultivars, a longer duration of phloem ingestion events on Barbera was evident. Indeed, significant differences were found for "Total duration of phloem ingestions", "Duration of the longest phloem ingestion", and "Time from first probe to first sustained phloem ingestion" between Barbera and the other two grapevine varieties (Table 4). "Time from first probe to first phloem ingestion" was shorter on Barbera compared to Moscato, with an intermediate duration recorded on Brachetto (Table 5). This also suggests a preference of *S. titanus* for Barbera. For the "Total duration of non-phloematic phases", for which an effect for the leafhopper sex was found, a difference was recorded between *S. titanus* feeding on Barbera and on Moscato, at least for females. *Scaphoideus titanus* also spent a higher percentage of time in the phloem ingestion phase on Barbera, compared to Brachetto and Moscato varieties and, consequently, less time in pathway- and active ingestion phases (Table 5). Since the presence of "Np" (typical interruption between two different passive ingestion phases) in phloem phases has been repeatedly recorded (Chuche et al., 2017a; Supplementary File S1 of the present work), three variables were introduced for their description in the present

276 work and are reported in Table 6: “Mean frequency of Np interruptions during phloem
277 phase”, “Percentage of time spent in Np interruption during phloem phase”, and “Number
278 of Np interruptions during phloem ingestion”. The second and third variables showed
279 significant differences between leafhoppers feeding on Barbera and those feeding on the
280 other varieties, underlying different phloem feeding behaviour on the former variety.

Table 3. Median ± SE of non-phloem variables related to recordings presenting phloem phases. Every row reports a single combination of grapevine Cultivar and leafhopper Sex. Every column reports a specific variable. Comparisons between rows were done with a specific GLM family for every variable: quasi-Poisson or negative-binomial for counts, Gamma or inverse-Gaussian for continuous time variables, beta-regression for proportions. Cultivar, Sex and their interaction (Cultivar × Sex) effects for every variable were evaluated. In case of no effect for Sex and Cultivar × Sex, the GLM was run with only Cultivar as explanatory variable (indicated in the tables with the * sign after the specific variable name). In case of effect for Sex or Cultivar × Sex, GLM was run with all the three explanatory variables (indicated in the tables with the ** sign after the specific variable name). Post-hoc comparisons were conducted with least-square means method and Tukey method for p-value adjustment, at significance level as 0.05 and 95% confidence intervals, and represented by letters for every specific group. GLMs specific details (family, coefficients, standard errors, AIC, BIC, R2) are reported in Supplementary File S3a-b.

Cultivar	Sex	n	Number of non-probing periods *	Total duration of non-probing periods [min] *	Time from 1st np to 1st probe [s] *	Duration of the 2nd non-probing period [s] *	Number of probes *	Total probing time [min] *	Total duration of pathway phase [min] **	Number of active ingestion phases **	Total duration of active ingestion [min] *
Barbera	female	8	9.5 ± 1.7 a	80.8 ± 22.8 a	33.5 ± 41.5 a	108 ± 143.4 a	9.5 ± 1.8 a	397.2 ± 22.8 a	96.1 ± 18.5 b	8 ± 6.9 a	3.2 ± 3.8 a
Barbera	male	6	14 ± 1.9 a	32.3 ± 47.7 a	19 ± 11.8 a	22.1 ± 43.8 a	12.5 ± 1.9 a	446.6 ± 47.5 a	124.9 ± 36.6 ab	42 ± 11.1 b	21.2 ± 8 a
Brachetto	female	6	21 ± 6.1 b	164.8 ± 32.1 a	48.5 ± 216.8 a	45.9 ± 80.9 a	21 ± 6.1 b	313.9 ± 31.8 a	175.1 ± 30.4 ab	52 ± 23.3 b	34.3 ± 13.7 a
Brachetto	male	8	16 ± 2 b	174.8 ± 20.9 a	63.3 ± 80.7 a	42.5 ± 48.5 a	15.5 ± 2 b	303.9 ± 20.5 a	160.3 ± 25 ab	51 ± 15.3 b	36.9 ± 11.6 a
Moscato	female	7	16 ± 3.3 ab	105 ± 24.9 a	87.6 ± 169.5 a	40.9 ± 96 a	16 ± 3.2 ab	374.5 ± 25 a	231.5 ± 36.5 a	47 ± 9.9 b	26.3 ± 7.5 a
Moscato	male	9	18.5 ± 3.5 ab	168.8 ± 28.3 a	54.6 ± 24.2 a	22.9 ± 79.5 a	17 ± 3.6 ab	310.5 ± 28.3 a	154.9 ± 24.7 ab	33 ± 14.3 b	22.5 ± 19.5 a

Table 4. Median ± SE of phloem variables related to recordings presenting phloem phases. The Table was drawn as detailed for Table 3.

Cultivar	Sex	n	Number of phloem ingestions *	Number of sustained (> 600 s) phloem ingestion *	Total duration of phloem ingestions [min] *	Mean duration of a single event of phloem ingestion [min] *	Duration of the longest phloem ingestion [min] *	Total duration of non-phloematic phases [min] **	Time from 1st probe to 1st phloem ingestion [min] *	Time from 1st probe to 1st sustained (> 600 s) phloem ingestion [min] *	Time of 1st sustained phloem phase [min] *
Barbera	female	8	13 ± 2.7 a	2 ± 0.4 a	301.9 ± 37.7	23.4 ± 11 a	240.2 ± 38.4	107.4 ± 20.1 b	76.7 ± 9.5 b	111.7 ± 27.3 b	114.1 ± 27.1 b

					a		a				
Barbera	male	6	14 ± 1.3 a	2.5 ± 0.7 a	190.3 ± 48.4 a	12.7 ± 3.1 a	158.3 ± 41.3 a	156.2 ± 37 ab	104.4 ± 32.9 b	131.1 ± 34.2 b	133.5 ± 34.1 b
Brachetto	female	6	7 ± 3.8 a	1 ± 0.6 a	83 ± 33.2 b	5.7 ± 4 a	50.2 ± 25.6 b	208.1 ± 43.2 ab	135.4 ± 15.3 ab	156.6 ± 63.7 a	157.1 ± 63.3 a
Brachetto	male	8	18 ± 5.9 a	2 ± 0.6 a	53.9 ± 26.1 b	3.5 ± 7.9 a	38 ± 13.9 b	237.6 ± 25.6 a	167.1 ± 43.4 ab	229.2 ± 49.2 a	240.5 ± 47 a
Moscato	female	7	10.5 ± 4.5 a	1 ± 0.4 a	37.6 ± 42.7 b	3.1 ± 3.5 a	20.2 ± 30.9 c	283.4 ± 44.6 a	187.1 ± 50.8 a	267.8 ± 62.3 a	270.7 ± 65.1 a
Moscato	male	9	25.5 ± 3.8 a	1 ± 0.3 a	47 ± 11.8 b	2.3 ± 0.6 a	13 ± 7.1 c	257 ± 33.8 a	107.8 ± 54.6 a	203.9 ± 55.8 a	208 ± 55.5 a

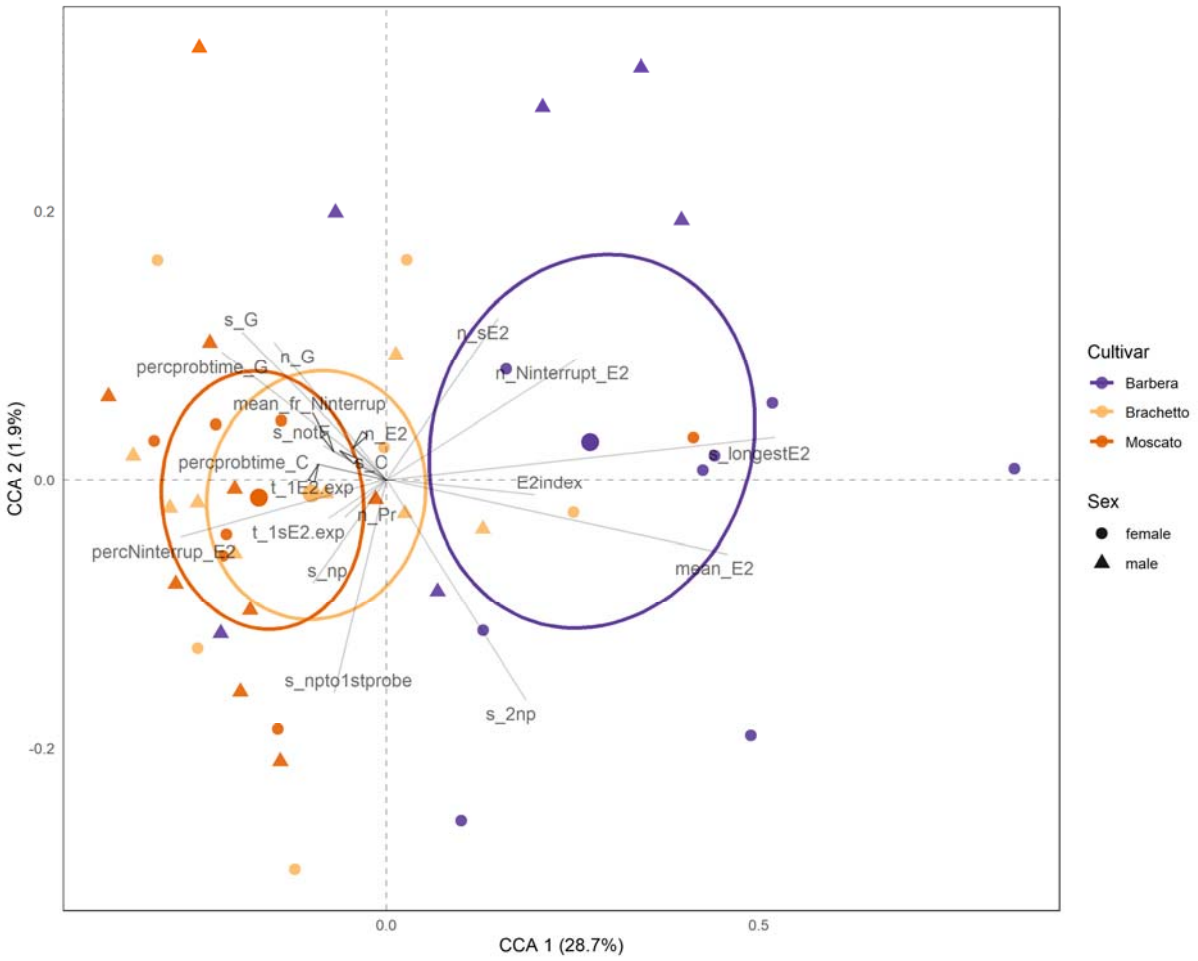
Table 5. Percentage variables related to recordings presenting phloem phases. The Table was drawn as detailed for Table 3.

Cultivar	Sex	n	Percentage of probing time spent in phloem ingestion [%] *	Percentage of probing time spent in pathway-phase [%] **	Percentage of probing time spent in active ingestion [%] **	Potential E2 index [%] *
Barbera	female	8	75 ± 6.6 b	22.4 ± 6 a	0.7 ± 1.1 a	77.7 ± 8.1 a
Barbera	male	6	51.2 ± 9.4 b	38.5 ± 9.2 ab	8.3 ± 1.7 ab	66.1 ± 16 a
Brachetto	female	6	26.9 ± 9.3 a	60 ± 7.1 bc	13.4 ± 3.2 ab	29.1 ± 7.7 a
Brachetto	male	8	14.6 ± 7.4 a	54.3 ± 5.3 bc	9.5 ± 4.2 b	48.3 ± 10.2 a
Moscato	female	7	11.6 ± 10.9 a	75.9 ± 8 c	9.9 ± 2 ab	48.6 ± 14.2 a
Moscato	male	9	11.9 ± 4.4 a	55.4 ± 4.9 bc	7.5 ± 3.9 b	32.1 ± 9.5 a

Table 6. "Np" phloem-interruptions variables related to recordings presenting phloem phases. The Table was drawn as detailed for Table 3

Cultivar	Sex	n	Mean frequency of Np interruptions during phloem phase [mHz] **	Percentage of time spent in Np interruption during phloem phase [%] *	Number of Np interruptions during phloem ingestion *
Barbera	female	8	9.4 ± 0.8 b	1.4 ± 0.7 a	148.5 ± 27.2 b
Barbera	male	6	14.6 ± 1.9 ab	3 ± 1.6 a	167.5 ± 58 b
Brachetto	female	6	16.7 ± 1 ab	6.8 ± 5.8 ab	88 ± 27.8 a
Brachetto	male	8	19 ± 3.1 a	9.7 ± 7.3 ab	38.5 ± 26.2 a
Moscato	female	7	15.5 ± 3.4 a	16.2 ± 4.6 b	44 ± 20.6 a
Moscato	male	9	19.8 ± 2 a	11.8 ± 5.3 b	53 ± 16.1 a

296 A constrained Canonical Correspondence Analysis (CCA) was conducted to explore the
297 comprehensive effect of the explanatory variables Cultivar, Sex, and their interaction with
298 *S. titanus* feeding behaviour (Figure 2). The CCA is a graphical representation of the non-
299 multi-collinear variables more related to the different groups. In particular, considering the
300 absence of effect for Sex and Cultivar × Sex (Table 7), ellipses were drawn containing
301 99% confidence intervals for the standard errors related to Cultivar variable. Again, this
302 representation highlighted the difference between *S. titanus* feeding behaviour on Barbera,
303 on one side, and on Brachetto and Moscato, on the other side. Moreover, CCA shows a
304 clear correlation between phloem variables and Barbera cultivar.



305

306 **Figure 2. Canonical Correspondence Analysis (CCA) on recordings with phloem phases.**
307 The new condensed CCA variables explained 28.7% (CCA 1, x axis) and 1.9% (CCA 2, y axis) of
308 the variability. Cultivar-specific recordings were grouped with ellipses, representing 99%
309 confidence intervals for the standard errors, and the centroid of each was represented. Every point
310 represents a single recording, colour refers to the grapevine cultivar and shape refers to the
311 leafhopper sex. Original variables were plotted and reported with their acronym (Table 1 for

312 acronym interpretation); all variables start from the intersection of the axes and are projected
313 according to their unique composition of CCA 1 and 2.

314 Results of the CCA were confirmed through a perMANOVA (Table 7), which highlighted
315 significant differences among Cultivars, while no significative differences were found for
316 Sex or the interaction of Cultivar and Sex.

317 **Table 7. perMANOVA results based on Bray-Curtis dissimilarities**, using all the non-multi-
318 collinear EPG variables (as described in Materials & Methods section). Df: degrees of freedom;
319 SumOfSqs: sequential sums of squares; F: F statistics values by permutations; Pr(>F): p-values,
320 based on 9999 permutations (the lowest possible p-value is 0.0001).

	Df	SumOfSqs	R2	F	Pr(>F)	signif
Cultivar	2	0.21459034	0.24924254	6.867267	0.0001	***
Sex	1	0.02922068	0.03393925	1.870225	0.1269	
Cultivar × Sex	2	0.02344144	0.02722678	0.750167	0.6142	
Residual	38	0.59371753	0.68959144	NA	NA	
Total	43	0.86096998	1	NA	NA	

321

322 ## 4. Discussion

323 In this work, the probing behaviour of the FD leafhopper vector *S. titanus* on grapevine
324 varieties with different susceptibility to the disease was analysed, to highlight possible
325 differences that can account for different transmission efficiencies. As phytoplasmas are
326 phloem-limited in the plant, vector acquisition and transmission abilities are related to
327 phloem feeding phases, and thus a plant genotype that does not sustain efficient phloem
328 feeding may be less prone to infection.

329 To understand if probing behaviour of *S. titanus* may contribute to explain
330 tolerance/susceptibility mechanisms of grapevine genotypes, the FD highly susceptible
331 Barbera and the FD tolerant Brachetto and Moscato varieties (Ripamonti et al., 2021) were
332 compared. Indeed, *S. titanus* showed a feeding preference for the FD highly susceptible
333 Barbera variety. To describe *S. titanus*-grapevine interaction, total probing time was
334 subdivided into different probing phases, mainly related to the inter/intra-cellular
335 movements of the stylets (pathway-phase), the active ingestion of mesophyll or xylem sap,
336 the passive ingestion of phloem-sap.

A preference of the leafhopper for Barbera was suggested at first by the overall higher proportion of *S. titanus* feeding on phloem of this variety (larger area under the phloem phase), compared to Brachetto and Moscato in the temporal progress area graph. However, it is possible that duration of phloem phases was underestimated in this study, as well as in those of Chuche et al. (2017a, 2017b), and indeed longer recording times can possibly highlight longer durations of phloem ingestion, as hypothesized for *Dalbulus maidis* (Carpane et al., 2011). Under our experimental setting, eight-hour recordings were long enough to allow 50% of the insects to reach the phloem phase, irrespective of the cultivar. This recording time was chosen for the experiments as it represents a widely used standard in EPG studies, and because in previous experiences, Chuche et al. (2017b) showed that, in average, 27% of the *S. titanus* probing time was spent in phloem feeding phases with four hour recordings. According to our results, most of the cultivar-dependent differences in *S. titanus* lies in phloem feeding behaviour. Actually, leafhoppers spent more than 50% of their probing time feeding on the Barbera phloem, while on the other two cultivars spent less than 20%. This result is in line with an enhanced possibility of acquisition and inoculation of phloem-limited agents, like FDp in the case of Barbera (Galetto et al., 2014). Although the “Potential E2 index”, a parameter regarded as a reliable indicator of phloem acceptability (Alvarez et al., 2006; Girma et al., 1992), was not significantly different among the tested cultivars, higher values were recorded for Barbera, further supporting a possible preference of the leafhopper for this cultivar. Moreover, since only half of the vectors reached the phloem phase during the 8-hour recordings, we cannot exclude that the amount of time was not sufficient to obtain a more descriptive feeding behaviour from all leafhoppers. Dramatic differences were highlighted in the “Total duration of phloem ingestions” on the different cultivars, while the “Number of phloem ingestions” and “sustained phloem ingestions” were similar. The former was actually the variable accounting for the highest differences in phloem phase among cultivars, and suggests that *S. titanus* prefers Barbera phloem to Brachetto or Moscato ones. Since no differences were recorded among cultivars in the percentage of leafhoppers reaching phloem, but “Total duration of phloem ingestion” and “Duration of the longest phloem ingestion” were higher on Barbera, it can be hypothesized that Brachetto and Moscato phloem saps contain some repellent compounds disturbing phloem feeding. During the phloem phase, the main waveforms were related to the passive ingestion of phloem and to the interruption between two different passive ingestion phases (mainly “Np”). These interruptions were already described for *S. titanus* by Chuche et al. (2017a) and for

371 *Circulifer tenellus* by Stafford and Walker (2009), and were suggested to represent
372 salivation events. Two are the main functions of saliva in piercing-sucking insects: i)
373 production of stylets sheath in the inter-cellular pathway phase (sheath saliva) or ii) dilution
374 of to-be-ingested sap and the suppression of defensive mechanism by the plant through
375 effectors (watery saliva) (Miles, 1972; Tjallingii, 2006; Will, Furch, & Zimmermann, 2013).
376 According to Chuche et al. (2017a) and Stafford & Walker (2009), the "Np" interruptions
377 found during *S. titanus* ingestion of phloem sap correspond to watery-salivation events.
378 This type of salivation is related to the inoculation of persistent-propagative agents from
379 the insect salivary glands into the plants tissues (Hogenhout et al., 2008). Therefore, the
380 greater number of interruption-salivation events on Barbera, that are a reflection of the
381 longer phloem phase, can explain, at least in part, the high susceptibility to FDp of this
382 cultivar. Phytoplasma spread can be regarded as a function of insect acquisition efficiency,
383 which is directly related to the duration of the phloem feeding phase, and of the inoculation
384 efficiency, which is putatively related to the absolute number of watery salivation events,
385 these latter also occuring during phloem feeding phase. According to this hypothesis, on
386 Barbera, the vector acquires and transmits efficiently, because it feeds longer in the
387 phloem and produces a higher number of salivation events compared to Brachetto and
388 Moscato. Indeed, Galetto et al. (2016) demonstrated that FDp acquisition by *S. titanus*
389 depends on the grapevine variety, with high efficiency from the most susceptible ones.
390 Also, on Brachetto and Moscato a high frequency of Np interruptions events were
391 recorded, but phloem phase was much shorter, leading to a lower absolute number of
392 salivation events. It is worth noting that, when the three grapevine varieties were exposed
393 to equally infected leafhoppers, Brachetto and Moscato showed a strong tolerance against
394 the infection (Ripamonti et al., 2021). This is a clear indication that either the inoculation,
395 more than acquisition, has a major impact on transmission efficiency, or plant genotype
396 account for different susceptibilities. The high frequency of Np interruptions on the tolerant
397 varieties can be explained by the presence of repellent compounds in the phloem saps.
398 Brachetto and Moscato are aromatic varieties (Pollon et al., 2019) and they are genetically
399 related (Raimondi et al., 2020). Their leaves contain high quantities of terpenoids (Mazza
400 et al., 2003), and this class of compounds can be transferred through the plant via the
401 phloem flux (Zhang et al., 2016) like other defence compounds (Will et al., 2013). Hence, it
402 can be speculated that *S. titanus* disturbed behaviour may be associated with the
403 presence of aromatic compounds, that act as repellents in Brachetto and Moscato
404 phloems. Repellent compounds can therefore act as antixenotic compounds. Antixenosis,

405 defined as the modification of herbivore behaviour by plant factors, which results in the
406 inability of a plant to serve as a host (Kogan and Ortman, 1978; Kordan et al., 2019), is a
407 well-known factor determining host plant resistance. Terpenoids and other volatile
408 compounds have well-known antixenotic activities in different plant-insect interactions
409 (Chand et al., 2017; Koul, 2008; Messchendorp et al., 1998). Antixenosis may represents
410 a valuable factor to be considered in the development of grapevine resistance against *S.*
411 *titanus*, *de facto* causing a reduction in the inoculation efficiency of FDp. Indeed,
412 leafhopper survival is reduced following a 7 day exposure to Moscato compared to
413 Barbera (Ripamonti et al., 2021). Further research is needed to clarify possible Moscato
414 antibiosis effect on *S. titanus*.

415 In our study, the leafhoppers started probing within the first minute, regardless of the
416 grapevine variety. No evident differences were highlighted in the non-phloem related
417 variables, as well as on total probing time. These results suggest that tested cultivars have
418 no major differences in the biochemical composition or structure of the leaf cuticle,
419 epidermis or mesophyll, that can impact the first feeding behaviour phases. Grapevine
420 trichomes are of the non-glandular type, subdivided in prostrate or erect (Gago et al.,
421 2016). Interestingly, Barbera has a highly dense trichomes surface in the abaxial leaf
422 blade (OIV, 2007), suggesting a possible repellence towards piercing-sucking insects
423 (Smith and Chuang, 2014). Nevertheless, Barbera was the most suitable variety for *S.*
424 *titanus* among those tested. For leafhoppers, data on trichome density acceptability are
425 available mainly for species of the Empoascini tribe, that are mostly insensitive to trichome
426 density on leaves. This is the case of *Empoasca vitis* on grapevine (Pavan and Picotti,
427 2009), *E. terminalis* on soybean (Nasruddin et al., 2014), and *E. fabae* on potato (Kaplan
428 et al., 2009). On the other hand, *E. fabae* and *Amrasca devastans* tend to avoid high
429 trichome density when feeding on edamame (*Glycine max* (L.)) and cotton, respectively
430 (Menger et al., 2018; Murugesan and Kavitha, 2010). As for *S. titanus,* it can be concluded
431 that a dense abaxially pubescence does not hamper nutrition on grapevine.

432 This work failed to identify clear differences in feeding behaviour of males and females.
433 Although small differences between sexes were recorded for some variables, no
434 differences were highlighted in the multivariate analysis conducted through CCA followed
435 by perMANOVA. On the contrary, Chuche et al. (2017b) reported that males feed more in
436 the phloem, compared to females. Following the analysis of our EPG recordings, we
437 conclude that no clear differences in feeding behaviour can be identified. Although

unlikely, we cannot exclude that the different grapevine varieties used in the studies may explain for this difference.

Future research should focus on antixenotic compounds in *V. vinifera* genotypes, and their role in vector-associated resistance to FD. On the other hand, plant secondary metabolites involved in defense mechanisms against pathogens, such as polyphenols, particularly vein flavonols and flavanonols (Kedrina-Okutan et al., 2019, 2018) may play a role in plant resistance towards the phytoplasma. All these grapevine genetic traits should be regarded as a natural resource to be exploited to obtain tolerant genotypes for a more sustainable viticulture.

5. Conclusions

The results of the present work indicate that Barbera variety is a better food source than Brachetto and Moscato for *S. titanus*. Indeed, the leafhopper showed longer phloem ingestion, with an absolute higher number of watery-salivation events, on grapevines of the Barbera cv. This latter feature is consistent with the high susceptibility of Barbera to FDp, as watery salivation have been associated with the inoculation of persistent-propagative agents from the insect salivary glands into the plants tissues. When feeding on Brachetto and Moscato, *S. titanus* showed reduced phloem nutrition, possibly due to antixenotic factors such as terpenoids, given the aromatic nature of the two varieties.

Acknowledgments

This work was funded by the project “Towards resistance to Flavescence dorée of grapevine, Flavoscreen”, Fondazione Compagnia di San Paolo and University of Torino. Authors wish to thank Nicola Bodino for revision of statistical analyses, Marco Chiapello for the fundamental help in the development of the Rwaves package, Stefano Raimondi for suggestions on ampelography, and Giovanni Marchisio for providing grapevine cuttings and dormant wood for *S. titanus* eggs.

Author contributions

Conceptualization: DB, CM, MR. Data curation: MR. Formal analysis: MR. Funding acquisition: DB, CM. Investigation: MR, FM. Methodology: AF, DC, DB, MR. Software: MR.

469 Project administration: DB. Supervision: AF, DB, DC. Visualization: MR. Writing - original
470 draft: MR. Writing - review & editing: DB, CM, AF, DC.

References

Alvarez, A.E., Tjallingii, W.F., Garzo, E., Vleeshouwers, V., Dicke, M., Vosman, B., 2006. Location of resistance factors in the leaves of potato and wild tuber-bearing Solanum species to the aphid Myzus persicae. Entomol. Exp. Appl. 121, 145–157. https://doi.org/10.1111/j.1570-8703.2006.00464.x

Anderson, M.J., 2001. A new method for non-parametric multivariate analysis of variance: NON-PARAMETRIC MANOVA FOR ECOLOGY. Austral Ecol. 26, 32–46. https://doi.org/10.1111/j.1442-9993.2001.01070.pp.x

Backus, E.A., Bennett, W.H., 2009. The AC–DC Correlation Monitor: New EPG design with flexible input resistors to detect both R and emf components for any piercing–sucking hemipteran. J. Insect Physiol. 16. https://doi.org/10.1016/j.jinsphys.2009.05.007

Backus, E.A., Cline, A.R., Ellerseick, M.R., Serrano, M.S., 2007. *Lygus hesperus* (Hemiptera: Miridae) Feeding on Cotton: New Methods and Parameters for Analysis of Nonsequential Electrical Penetration Graph Data. Ann. Entomol. Soc. Am. 100, 296–310. https://doi.org/10.1603/0013-8746(2007)100[296:LHHMFO]2.0.CO;2

Backus, E.A., Guedes, R.N.C., Reif, K.E., 2020. AC–DC electropenetrography: fundamentals, controversies, and perspectives for arthropod pest management. Pest Manag. Sci. https://doi.org/10.1002/ps.6087

Bocca, F.M., Picciau, L., Alma, A., 2020. New insights on Scaphoideus titanus biology and their implications for integrated pest management. Entomol. Gen. https://doi.org/10.1127/entomologia/2020/0977

Bosco, D., Mori, N., 2013. Flavescence dorée vector control in Italy. Phytopathogenic Mollicutes 3, 40–43.

Caillaud, C.M., Di Pietro, J.P., Chaubet, B., Pierre, J.S., 1995a. Application of discriminant analysis to electrical penetration graphs of the aphid Sitobion avenae feeding on resistant and susceptible wheat. J. Appl. Entomol. 119, 103–106. https://doi.org/10.1111/j.1439-0418.1995.tb01252.x

Caillaud, C.M., Pierre, J.S., Chaubet, B., Di Pietro, J.P., 1995b. Analysis of wheat resistance to the cereal aphid Sitobion avenae using electrical penetration graphs and flow charts combined with correspondence analysis. Entomol. Exp. Appl. 75, 9–18. https://doi.org/10.1111/j.1570-7458.1995.tb01904.x

Carpane, P., Wayadande, A., Backus, E., Dolezal, W., Fletcher, J., 2011. Characterization and correlation of new electrical penetration graph waveforms for the corn leafhopper (Hemiptera: Cicadellidae). Ann. Entomol. Soc. Am. 104, 515–525. https://doi.org/10.1603/AN10052

Caudwell, A., Kuszala, C., Bachelier, J.C., Larrue, J., 1970. Transmission de la Flavescence dorée de la vigne aux plantes herbacées par l'allongement du temps d'utilisation de la cicadelle Scaphoideus littoralis Ball et l'étude de sa survie sur un grand nombre d'espèces végétales. 2, 415–428.

Chand, R.R., Jokhan, A.D., Gopalan, R.D., 2017. A mini-review of essential oils in the South Pacific and their insecticidal properties. Adv. Hortic. Sci. 295-310 Pages. https://doi.org/10.13128/AHS-20833

Chen, J.Q., Martin, B., Rahbé, Y., Fereres, A., 1997. Early intracellular punctures by two aphid species on near-isogenic melon lines with and without the virus aphid transmission (Vat) resistance gene. J. Plant Pathol. 103, 521–536.

Chuche, J., Backus, E.A., Thiéry, D., Sauvion, N., 2017a. First finding of a dual-meaning X wave for phloem and xylem fluid ingestion: Characterization of Scaphoideus titanus (Hemiptera: Cicadellidae) EPG waveforms. J. Insect Physiol. 102, 50–61. https://doi.org/10.1016/j.jinsphys.2017.01.013

Chuche, J., Sauvion, N., Thiéry, D., 2017b. Mixed xylem and phloem sap ingestion in sheath-feeders as normal dietary behavior: Evidence from the leafhopper Scaphoideus titanus. J. Insect Physiol. 102, 62–72. https://doi.org/10.1016/j.jinsphys.2017.01.014

Chuche, J., Thiéry, D., 2014. Biology and ecology of the Flavescence dorée vector Scaphoideus titanus: A review. Agron. Sustain. Dev. 34, 381–403. https://doi.org/10.1007/s13593-014-0208-7

Cribari-Neto, F., Zeileis, A., 2010. Beta Regression in R. J. Stat. Softw. 34, 1–24. https://doi.org/10.18637/jss.v034.i02

Davis, R.E., Dally, E.L., 2001. Revised Subgroup Classification of Group 16SrV Phytoplasmas and Placement of *Flavescence Dorée* -Associated Phytoplasmas in Two Distinct Subgroups. Plant Dis. 85, 790–797. https://doi.org/10.1094/PDIS.2001.85.7.790

Desneux, N., Decourtye, A., Delpuech, J.-M., 2007. The Sublethal Effects of Pesticides on Beneficial Arthropods. Annu. Rev. Entomol. 52, 81–106. https://doi.org/10.1146/annurev.ento.52.110405.091440

EFSA Panel on Plant Health (PLH), Jeger, M., Bragard, C., Caffier, D., Candresse, T., Chatzivassiliou, E., Dehnen-Schmutz, K., Gilioli, G., Jaques Miret, J.A., MacLeod, A., Navajas Navarro, M., Niere, B., Parnell, S., Potting, R., Rafoss, T., Rossi, V., Urek, G., Van Bruggen, A., Van Der Werf, W., West, J., Winter, S., Bosco, D., Foissac, X., Strauss, G., Hollo, G., Mosbach-Schulz, O., Grégoire, J., 2016. Risk to plant health of Flavescence dorée for the EU territory. EFSA J. 14. https://doi.org/10.2903/j.efsa.2016.4603

European Food Safety Authority (EFSA), Tramontini, S., Delbianco, A., Vos, S., 2020. Pest survey card on flavescence dorée phytoplasma and its vector Scaphoideus titanus. EFSA Support. Publ. 17. https://doi.org/10.2903/sp.efsa.2020.EN-1909

Eveillard, S., Jollard, C., Labroussaa, F., Khalil, D., Perrin, M., Desqué, D., Salar, P., Razan, F., Hévin, C., Bordenave, L., Foissac, X., Masson, J.E., Malembic-Maher, S., 2016. Contrasting Susceptibilities to Flavescence Dorée in Vitis vinifera, Rootstocks and Wild Vitis Species. Front. Plant Sci. 7. https://doi.org/10.3389/fpls.2016.01762

Fox, J., Weisberg, S., 2019. An {R} Companion to Applied Regression., Third edition. ed. Sage, Thousand Oaks CA.

Gago, P., Conéjéro, G., Martínez, M.C., Boso, S., This, P., Verdeil, J.-L., 2016. Microanatomy of leaf trichomes: opportunities for improved ampelographic discrimination of grapevine (Vitis vinifera L.) cultivars. Aust. J. Grape Wine Res. 22, 494–503. https://doi.org/10.1111/ajgw.12226

Galetto, L., Miliordos, D., Pegoraro, M., Sacco, D., Veratti, F., Marzachì, C., Bosco, D., 2016. Acquisition of Flavescence Dorée Phytoplasma by Scaphoideus titanus Ball from Different Grapevine Varieties. Int. J. Mol. Sci. 17, 1563. https://doi.org/10.3390/ijms17091563

Galetto, L., Miliordos, D., Roggia, C., Rashidi, M., C, M., Bosco, D., 2014. Acquisition capability of the grapevine Flavescence doree by the leafhopper vector Scaphoideus titanus Ball correlates with phytoplasma titre in the source plant. J. Pest Sci. 87, 671–679.

Garzo, E., Fernández-Pascual, M., Morcillo, C., Fereres, A., Gómez-Guillamón, M.L., Tjallingii, W.F., 2018. Ultrastructure of compatible and incompatible interactions in phloem sieve elements during the stylet penetration by cotton aphids in melon. Insect Sci. 25, 631–642.

Girma, M., Wilde, G.E., Reese, J., 1992. Russian Wheat Aphid (Homoptera: Aphididae) Feeding Behavior on Host and Nonhost Plants. J Econ Entomol 85, 395–401.

Hogenhout, S.A., Ammar, E.-D., Whitfield, A.E., Redinbaugh, M.G., 2008. Insect Vector Interactions with Persistently Transmitted Viruses. Annu. Rev. Phytopathol. 46, 327–359. https://doi.org/10.1146/annurev.phyto.022508.092135

Hothorn, T., Bretz, F., Westfall, P., 2008. Simultaneous Inference in General Parametric Models. Biom. J. 50, 346–363.

Jiang, Y.X., Nombela, G., Muñiz, M., 2001. Analysis by DC-EPG of the resistance to Bemisia tabaci on an Mi-tomato line. Entomol. Exp. Appl. 99, 295–302. https://doi.org/10.1046/j.1570-7458.2001.00828.x

Kaplan, I., Dively, G.P., Denno, R.F., 2009. The costs of anti-herbivore defense traits in agricultural crop plants: A case study involving leafhoppers and trichomes. Ecol. Appl. 19, 864–872. https://doi.org/10.1890/07-1566.1

Kassambara, A., 2020. ggpubr: "ggplot2" Based Publication Ready Plots.

Kawabe, S., McLean, D.L., 1980. Electronic measurement of probing activities of the green leafhopper of rice. Entomol. Exp. Appl. 27, 77–82. https://doi.org/10.1111/j.1570-7458.1980.tb02948.x

Kedrina-Okutan, O., Novello, V., Hoffmann, T., Hadersdorfer, J., Occhipinti, A., Schwab, W., Ferrandino, A., 2018. Constitutive Polyphenols in Blades and Veins of Grapevine (*Vitis vinifera* L.) Healthy Leaves. J. Agric. Food Chem. 66, 10977–10990. https://doi.org/10.1021/acs.jafc.8b03418

Kedrina-Okutan, O., Novello, V., Hoffmann, T., Hadersdorfer, J., Schneider, A., Schwab, W., Ferrandino, A., 2019. Polyphenolic diversity in Vitis sp. leaves. Sci. Hortic. 256, 108569. https://doi.org/10.1016/j.scienta.2019.108569

Kimmins, F.M., 1989. Electrical penetration graphs from Nilaparvata lugens on resistant and susceptible rice varieties. Entomol. Exp. Appl. 50, 69–79. https://doi.org/10.1111/j.1570-7458.1989.tb02317.x

Kogan, M., Ortman, E.F., 1978. Antixenosis-A New Term Proposed to Define Painter's "Nonpreference" Modality of Resistance. Bull. Entomol. Soc. Am. 24, 175–176. https://doi.org/10.1093/besa/24.2.175

Kordan, B., Stec, K., Słomiński, P., Laszczak-Dawid, A., Wróblewska-Kurdyk, A., Gabryś, B., 2019. Antixenosis Potential in Pulses Against the Pea Aphid (Hemiptera: Aphididae). J. Econ. Entomol. 112, 465–474. https://doi.org/10.1093/jee/toy349

Koul, O., 2008. Phytochemicals and Insect Control: An Antifeedant Approach. Crit. Rev. Plant Sci. 27, 1–24. https://doi.org/10.1080/07352680802053908

Lee, I.-M., Martini, M., Marcone, C., Zhu, S.F., 2004. Classification of phytoplasma strains in the elm yellows group (16SrV) and proposal of 'Candidatus Phytoplasma ulmi' for the phytoplasma associated with elm yellows. Int. J. Syst. Evol. Microbiol. 54, 337–347. https://doi.org/10.1099/ijs.0.02697-0

Legendre, P., Gallagher, E.D., 2001. Ecologically meaningful transformations for ordination of species data. Oecologia 129, 271–280.

Legendre, P., Legendre, L., 2012. Numerical Ecology, 3rd English ed. ed. Elsevier.

Lenth, R., 2020. emmeans: Estimated Marginal Means, aka Least-Squares Means.

Lett, J.-M., Granier, M., Grondin, M., Turpin, P., Molinaro, F., Chiroleu, F., Peterschmitt, M., Reynaud, B., 2001. Electrical penetration graphs from Cicadulina mbila on maize, the fine structure of its stylet pathways and consequences for virus transmission efficiency. Entomol. Exp. Appl. 101, 93–109. https://doi.org/10.1046/j.1570-7458.2001.00895.x

Long, J., 2020. jtools: Analysis and Presentation of Social Scientific Data.

Lucini, T., Panizzi, A.R., de F. Bueno, A., 2021. Evaluating resistance of the soybean block technology cultivars to the Neotropical brown stink bug, Euschistus heros (F.). J. Insect Physiol. 131, 104228. https://doi.org/10.1016/j.jinsphys.2021.104228

Maggi, F., Bosco, D., Galetto, L., Palmano, S., Marzachì, C., 2017. Space-time point pattern analysis of flavescence dorée epidemic in a grapevine field: Disease progression and recovery. Front. Plant Sci. 7. https://doi.org/10.3389/fpls.2016.01987

Martini, M., Botti, S., Marcone, C., Marzachì, C., Casati, P., Bianco, P.A., Benedetti, R., Bertaccini, A., 2002. Genetic variability among flavescence dorée phytoplasmas from different origins in Italy and France. Mol. Cell. Probes 16, 197–208. https://doi.org/10.1006/mcpr.2002.0410

Mazza, G., Cascio, P., Barbieri, E., 2003. Free and Glicosidically Bound Volatile Compounds from Vitis vinifera Cv. White Muscat Leaves and Grape Berries. Riv. Vitic. E Enol. 2–3, 57–74.

Mazzoni, V., Lucchi, A., Čokl, A., Prešern, J., Virant-Doberlet, M., 2009. Disruption of the reproductive behaviour of *Scaphoideus titanus* by playback of vibrational signals: *Disruption of mating behaviour in* Scaphoideus titanus. Entomol. Exp. Appl. 133, 174–185. https://doi.org/10.1111/j.1570-7458.2009.00911.x

McLean, D.L., Kinsey, M.G., 1964. A Technique for Electronically Recording Aphid Feeding and Salivation. Nature 202, 1358–1359. https://doi.org/10.1038/2021358a0

Menger, J., Hanson, A.A., Koch, R.L., 2018. Evaluation of insect pests on edamame varieties in Minnesota. J. Econ. Entomol. 111, 2272–2280. https://doi.org/10.1093/jee/toy204

Messchendorp, L., Schoonhoven, L.M., Loon, J.J.A. van, 1998. Terpenoid antifeedants against insects: a behavioural and sensory study. Wageningen University, Wageningen, The Netherlands.

Miao, J., Han, B.-Y., Zhang, Q.-H., 2014. Probing behavior of empoasca vitis (Homoptera: Cicadellidae) on resistant and susceptible cultivars of tea plants. J. Insect Sci. 14. https://doi.org/10.1093/jisesa/ieu085

Miles, P.W., 1972. The Saliva of Hemiptera, in: Advances in Insect Physiology. Elsevier, pp. 183–255. https://doi.org/10.1016/S0065-2806(08)60277-5

Mitchell, C., Brennan, R.M., Graham, J., Karley, A.J., 2016. Plant Defense against Herbivorous Pests: Exploiting Resistance and Tolerance Traits for Sustainable Crop Protection. Front. Plant Sci. 7. https://doi.org/10.3389/fpls.2016.01132

Moral, R.A., Hinde, J., Demétrio, C.G.B., 2017. Half-Normal Plots and Overdispersed Models in R: The hnp Package. J. Stat. Softw. 81. https://doi.org/10.18637/jss.v081.i10

Morone, C., Boveri, M., Giosuè, S., Gotta, P., Rossi, V., Scapin, I., Marzachi, C., 2007. Epidemiology of flavescence dorée in vineyards in northwestern Italy. Phytopathology 97, 1422–1427. https://doi.org/10.1094/PHYTO-97-11-1422

Naimi, B., Hamm, N.A.S., Groen, T.A., Skidmore, A.K., Toxopeus, A.G., 2014. Where is positional uncertainty a problem for species distribution modelling? Ecography 37, 191–203. https://doi.org/10.1111/j.1600-0587.2013.00205.x

Nasruddin, A., Gassa, A., Melina, M., 2014. Field evaluation of several cultivated soybean varieties against Empoasca terminalis (Hemiptera: Cicadellidae). Fla. Entomol. 97, 995–1001. https://doi.org/10.1653/024.097.0357

OIV, The International Organisation of Vine and Wine, 2007. OIV descriptor list for grape varieties and Vitis species (2nd edition). Descriptors 84-87.

Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R., O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H., 2019. vegan: Community Ecology Package. R package version 2.5-6.

Pavan, F., Picotti, P., 2009. Influence of grapevine cultivars on the leafhopper Empoasca vitis and its egg parasitoids. BioControl 54, 55–63. https://doi.org/10.1007/s10526-008-9151-3

Pollon, M., Torchio, F., Giacosa, S., Segade, S.R., Rolle, L., 2019. Use of density sorting for the selection of aromatic grape berries with different volatile profile. Food Chem. 276, 562–571. https://doi.org/10.1016/j.foodchem.2018.10.040

Quensen, J., 2018. ggordiplots: Make ggplot Versions of Vegan's Ordiplots.

R Core Team., 2020. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

Raimondi, S., Tumino, G., Ruffa, P., Boccacci, P., Gambino, G., Schneider, A., 2020. DNA-based genealogy reconstruction of Nebbiolo, Barbera and other ancient grapevine cultivars from northwestern Italy. Sci. Rep. 10, 15782. https://doi.org/10.1038/s41598-020-72799-6

Ripamonti, M., Pegoraro, M., Morabito, C., Gribaudo, I., Schubert, A., Bosco, D., Marzachì, C., 2021. Susceptibility to flavescence dorée of different *Vitis vinifera* genotypes from north-western Italy. Plant Pathol. 70, 511–520. https://doi.org/10.1111/ppa.13301

Ripamonti, M., Pegoraro, M., Rossi, M., Bodino, N., Beal, D., Panero, L., Marzachì, C., Bosco, D., 2020. Prevalence of Flavescence Dorée Phytoplasma-Infected Scaphoideus titanus in Different Vineyard Agroecosystems of Northwestern Italy. Insects 11, 301. https://doi.org/10.3390/insects11050301

Rodríguez-López, M.J., Garzo, E., Bonani, J.P., Fereres, A., Fernández-Muñoz, R., Moriones, E., 2011. Whitefly resistance traits derived from the wild tomato Solanum pimpinellifolium affect the preference and feeding behavior of Bemisia tabaci and reduce the spread of Tomato yellow leaf curl virus. Phytopathology 101, 1191–1201.

Sarria, E., Cid, M., Garzo, E., Fereres, A., 2009. Excel Workbook for automatic parameter calculation of EPG data. Comput. Electron. Agric. 67, 35–42. https://doi.org/10.1016/j.compag.2009.02.006

Sauge, M.-H., Kervella, J., Rahbé, Y., 1998. Probing behaviour of the green peach aphid Myzus persicae on resistant Prunus genotypes. Entomol. Exp. Appl. 89, 223–232. https://doi.org/10.1046/j.1570-7458.1998.00403.x

Smith, C.M., Chuang, W.-P., 2014. Plant resistance to aphid feeding: behavioral, physiological, genetic and molecular cues regulate aphid host selection and feeding: Plant resistance to aphid feeding. Pest Manag. Sci. 70, 528–540. https://doi.org/10.1002/ps.3689

Stafford, C.A., Walker, G.P., 2009. Characterization and correlation of DC electrical penetration graph waveforms with feeding behavior of beet leafhopper, Circulifer tenellus. Entomol. Exp. Appl. 130, 113–129. https://doi.org/10.1111/j.1570-7458.2008.00812.x

Tjallingii, W.F., 2006. Salivary secretions by aphids interacting with proteins of phloem wound responses. J. Exp. Bot. 57, 739–745. https://doi.org/10.1093/jxb/erj088

675 Tjallingii, W.F., 1978. ELECTRONIC RECORDING OF PENETRATION BEHAVIOUR BY APHIDS. Entomol. Exp.
676 Appl. 24, 721–730. https://doi.org/10.1111/j.1570-7458.1978.tb02836.x
677 Tosi, S., Costa, C., Vesco, U., Quaglia, G., Guido, G., 2018. A 3-year survey of Italian honey bee-collected
678 pollen reveals widespread contamination by agricultural pesticides. Sci. Total Environ. 11.
679 https://doi.org/10.1016/j.scitotenv.2017.09.226
680 Trębicki, P., Tjallingii, W.F., Harding, R.M., Rodoni, B.C., Powell, K.S., 2012. EPG monitoring of the probing
681 behaviour of the common brown leafhopper Orosius orientalis on artificial diet and selected host
682 plants. Arthropod-Plant Interact. 6, 405–415. https://doi.org/10.1007/s11829-012-9192-5
683 Trivellone, V., Jermini, M., Linder, C., Cara, C., Delabays, N., Baumgärtner, J., 2013. Rôle de la flore du
684 vignoble sur la distribution de Scaphoideus titanus. Rev Suisse Vitic Arboric Hortic 45, 222–228.
685 Venables, W.N., Ripley, B.D., 2002. Modern Applied Statistics with S 504. https://doi.org/10.1007/978-0-
686 387-21706-2
687 Wickham, H., 2020. tidyr: Tidy Messy Data.
688 Wickham, H., 2019. stringr: Simple, Consistent Wrappers for Common String Operations.
689 Wickham, H., 2016. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
690 Wickham, H., François, R., Henry, L., Müller, K., 2020. dplyr: A Grammar of Data Manipulation.
691 Will, T., Furch, A.C., Zimmermann, M.R., 2013. How phloem-feeding insects face the challenge of phloem-
692 located defenses. Front. Plant Sci. 4. https://doi.org/10.3389/fpls.2013.00336
693 Zeileis, A., Köll, S., Graham, N., 2020. Various Versatile Variances: An Object-Oriented Implementation of
694 Clustered Covariances in *R*. J. Stat. Softw. 95. https://doi.org/10.18637/jss.v095.i01
695 Zhang, P., Fuentes, S., Wang, Y., Deng, R., Krstic, M., Herderich, M., Barlow, E.W.R., Howell, K., 2016.
696 Distribution of Rotundone and Possible Translocation of Related Compounds Amongst Grapevine
697 Tissues in Vitis vinifera L. cv. Shiraz. Front. Plant Sci. 7. https://doi.org/10.3389/fpls.2016.00859
698