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This is the author's manuscript

Original Citation:

Availability:

This version is available <http://hdl.handle.net/2318/1749824> since 2022-07-01T14:50:18Z

Published version:

DOI:10.1158/1078-0432.CCR-20-0951

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(Article begins on next page)

This is the author's final version of the contribution published as:

D'Agostino M, Zaccaria GM, Ziccheddu B, Rustad EH, Genuardi E, Capra A, Oliva S, Auclair D, Yesil J, Colucci P, Keats JJ, Gambella M, Bringham S, Larocca A, Boccadoro M, Bolli N, Maura F, Gay F. Early Relapse Risk in Patients with Newly Diagnosed Multiple Myeloma Characterized by Next-generation Sequencing. *Clin Cancer Res.* 2020 Sep 15;26(18):4832-4841. doi: 10.1158/1078-0432.CCR-20-0951. Epub 2020 Jul 2. PMID: 32616499.

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The publisher's version is available at:

<https://aacrjournals.org/clincancerres/article/26/18/4832/9542/Early-Relapse-Risk-in-Patients-with-Newly>

<https://doi.org/10.1158/1078-0432.ccr-20-0951>

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Research Article

Early Relapse Risk in Newly Diagnosed Multiple Myeloma Patients Characterized by Next-Generation Sequencing

Running title: Early relapse risk by NGS in NDMM patients

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Word count - abstract: 244

Word count - text: 4041 (Revised vsn.)

Number of figures: 4

Number of tables: 2

Number of references: 38

Supplementary Appendix: 1 file.

Authorship contributions

Conceptualization: MD, GMZ, SO, DA, JY, JK, MG, MB, NB, FM, and FG

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All the authors approved the final version of the manuscript and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Acknowledgements

F. Maura is supported by the Memorial Sloan Kettering Cancer Center NCI Core Grant (P30 CA 008748).

The CoMMpass study is sponsored by the Multiple Myeloma Research Foundation (MMRF), which had no role in the data interpretation, writing of the report or publication of this contribution. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit this manuscript for publication, together with the other authors.

We thank all the patients who participated in the study, the nurses Luisella D'Ambrosio and Tiziana De Lazzer, the data managers Debora Caldarazzo and Elena Tigano, and Ugo Panzani from the Torino site.

Competing interests

MD has served on the advisory board for GSK.

SO has received honoraria from Amgen, Celgene, and Janssen; has served on the advisory boards for Adaptive Biotechnologies, Janssen, Amgen, and Takeda.

DA is currently employed by the Multiple Myeloma Research Foundation, Norwalk, US-CT.

JY is currently employed by the Multiple Myeloma Research Foundation, Norwalk, US-CT.

JK is currently employed by the Translational Genomics Research Institute (TGen), US-AZ.

SB has received honoraria from Bristol-Myers Squibb, Celgene, Amgen and Janssen; has served on the advisory boards for Amgen, Karyopharm, Janssen and Celgene; has received consultancy fees from Takeda and Janssen.

AL has received honoraria from Amgen, Bristol-Myers Squibb, Celgene, Janssen, and GSK; has served on the advisory boards for Bristol-Myers Squibb, Celgene, Janssen, and Takeda.

MB has received honoraria from Sanofi, Celgene, Amgen, Janssen, Novartis, Bristol-Myers Squibb, and AbbVie; has received research funding from Sanofi, Celgene, Amgen, Janssen, Novartis, Bristol-Myers Squibb, and Mundipharma.

NB has received honoraria from Celgene and Janssen in the last three years, but he has no conflict with regards to the data presented.

FG has received honoraria from Amgen, Celgene, Janssen, Takeda, and Bristol-Myers Squibb; has served on the advisory boards for Amgen, Celgene, Janssen, Takeda, Bristol-Myers Squibb, Roche, AbbVie, Adaptive, and Seattle Genetics.

The other authors declare no competing financial interests.

Statement of translational relevance

Duration of first remission is an important factor for the survival of patients with multiple myeloma (MM). Conventional baseline risk stratification is not always able to predict a short duration of first remission and poor survival.

In this study, we demonstrated the independent detrimental effect of early relapse (ER) within 18 months from the start of treatment on the survival of newly-diagnosed MM patients. Exploiting the molecular characterization through next-generation sequencing (NGS) of this large cohort of patients, we found additional risk factors increasing the risk of ER, whereas treatment intensification with carfilzomib-based induction, autologous stem-cell transplantation and continuous combination therapy may mitigate the risk of ER.

We demonstrated that patients relapsing within 18 months from the start of treatment represent an unmet clinical need and may deserve dedicated trials. NGS may help to better identify patients at risk. Treatment intensification may reduce early progressive disease in patients at risk.

Abstract

Introduction. Duration of first remission is important for the survival of multiple myeloma (MM) patients.

Methods. From the CoMMpass study (NCT01454297), 926 newly diagnosed MM patients, characterized by next-generation sequencing, were analyzed to evaluate those who experienced early progressive disease (PD) (time to progression, TTP \leq 18 months).

Results. After a median follow-up of 39 months, early PD was detected in 191/926 (20.6%) patients, 228/926 (24.6%) patients had late PD (TTP $>$ 18 months), while 507/926 (54.8%) did not have PD at the current follow-up. Compared to Late PD patients, Early PD patients had a lower at least very good partial response rate (47% vs 82%, $p<0.001$) and more frequently acquired double refractoriness to immunomodulatory drugs (IMiDs) and proteasome inhibitors (PIs) (21% vs 8%, $p<0.001$). Early PD patients were at higher risk of death compared to Late PD and No PD patients (HR 3.65, 95% CI 2.7-4.93, $p<0.001$), showing a dismal median overall survival (32.8 months). In a multivariate logistic regression model, independent factors increasing the Early PD risk were *TP53* mutation (OR 3.78, $p<0.001$), high LDH levels (OR 3.15, $p=0.006$), λ -chain translocation (OR 2.25, $p=0.033$) and *IGLL5* mutation (OR 2.15, $p=0.007$). Carfilzomib-based induction (OR 0.15, $p=0.014$), autologous stem-cell transplantation (OR 0.27, $p<0.001$) and continuous therapy with PIs and IMiDs (OR 0.34, $p=0.024$) mitigated the risk of early PD.

Conclusion. Early PD identifies a high-risk MM population. Further research is needed to better identify baseline features predicting early PD and the optimal treatment approaches for patients at risk.

Introduction

The expected survival of newly diagnosed multiple myeloma (NDMM) patients is currently improving and approaching 8 years, thanks to the use of novel agents and better supportive care (1). Nevertheless, MM still remains largely incurable and about 12000 MM patients in the United States and 30000 MM patients in Europe die each year, with the main cause of death being the development of refractory disease to the currently available drugs (2–4).

Relapse is caused by MM cell clones with an increasing degree of drug refractoriness and genetic complexity eventually leading to shorter remissions (5). Since the longest remission period is usually induced by upfront treatment, the duration of first remission is one of the most important factors impacting patient prognosis (6).

This can become particularly important as a dynamic prognostic marker, if we consider the complexity associated with the evaluation of baseline prognostic features. The most widely used staging system is the Revised International Staging System (R-ISS), which is based on clinical and biological standard features (ISS, chromosomal abnormalities and lactate dehydrogenase [LDH] levels) (7). Many efforts aimed at improving the baseline stratification, including the use of gene expression profiles (GEP) and next-generation sequencing (NGS) (8–10). Of note, according to R-ISS, only 10% of patients are at high risk of progression and/or death and, according to the NGS-based “double-hit” classification, only 6.1% of patients are at high risk of progression and/or death, but the overall rate of patients who relapse or die within two years from diagnosis is about 20% (11,12). This highlights the importance of dynamic prognostic evaluation and the need for an improved baseline risk stratification. The identification and treatment of high-risk MM patients currently represent unmet medical needs. Our aims were (1) to characterize patients with early progressive disease (Early PD; time-to-progression [TTP] ≤ 18 months) after first-line treatment including

immunomodulatory drugs (IMiDs) and/or 1st-2nd generation proteasome inhibitors (PIs) incorporating baseline clinical and next-generation sequencing (NGS) molecular features; (2) to address the role of different upfront therapies in reducing the risk of early PD.

Methods

Patients and treatment

Data from patients enrolled in the prospective observational Multiple Myeloma Research Foundation (MMRF) CoMMpass study (NCT01454297) were included in this analysis. Ethics committees or institutional review boards at the study sites approved the study, which was conducted in accordance with the Declaration of Helsinki. All patients provided written informed consent.

Main inclusion criteria were: symptomatic NDMM, measurable disease and upfront systemic therapy including an IMiD and/or a PI. CoMMpass data were generated as part of the MMRF Personalized Medicine Initiatives (<https://research.themmr.org> and www.themmr.org).

Data from patients receiving treatment in the context of clinical trials as well as with real world regimens were included. Therapy (source file “mmrf_commpass_IA14_stand_alone_treatment_regimen” available upon request on <https://research.themmr.org>) was reviewed and classified according to: type of induction treatment (bortezomib-dexamethasone/bortezomib+chemotherapy triplets/lenalidomide-dexamethasone/bortezomib-lenalidomide-dexamethasone/carfilzomib-based/other), autologous stem-cell transplantation (ASCT; Yes/No), and type of continuous treatment (CT) (IMiDs CT/PIs CT/IMiDs+PIs CT/Fixed-duration therapy [FDT]). FDT was defined as ≤ 1 year of upfront treatment (13). The definition of variables is detailed in *Tables S1-S2*. Patients were considered evaluable for the ASCT vs no ASCT analysis if they were alive and relapse-free after

induction treatment and if the date of ASCT was available. Patients receiving ASCT before PD but after 18 months from the start of treatment (cut-off for the early relapse evaluation) were considered not evaluable. Patients were considered evaluable for the CT analysis if they were alive and relapse-free after 1 year from the start of treatment, if the follow-up was >1 year, and if details of treatment administered after the 1-year timepoint were available.

The Interim Analysis (IA)¹⁴ release of CoMMpass was analyzed. Updated time-to-event endpoints for CoMMpass patients co-enrolled in the NCT02203643 trial were used (data cut-off: 30/05/2018; the treatment schedule is reported in the Supplementary Appendix).

Next-generation sequencing

Baseline bone marrow CD138+ cells were obtained before the initiation of systemic therapy (within 30 days before first-line treatment). Available data on samples at relapse, a pre-planned objective within the CoMMpass study, were also evaluated. Long-insert whole genome sequencing (WGS) and whole exome sequencing (WES) were performed by the Translational Genomics Institute (TGen). Somatic tumor alterations were defined comparing tumor cells with patient-specific paired normal cells. Details on the definition of the risk factors explored in this work are provided in previous CoMMpass publications (14–16). Cytogenetic data reported by single study centers were heterogeneous in terms of fluorescence in situ hybridization (FISH) probes utilized, number of cells counted and cell sorting techniques. To uniformly define cytogenetic abnormalities in all patients, copy number abnormalities (CNAs), immunoglobulin heavy chain (IgH) translocations and immunoglobulin lambda (IgL) translocations were defined using molecular data (Seq-FISH) (17–19). The concordance of Seq-FISH and conventional FISH in a subgroup of patients evaluated in the context of a clinical trial by a centralized laboratory showed a high degree of concordance. The presence or absence of recurrent CNAs [hyperdiploidy, deletion13q, deletion17p, gain1q (3 CSK1B copies) and

amplification(1q) (>3 CSK1B copies)], IgH translocations [t(11;14), t(4;14), t(14;16), t(14;20)] and IgL translocations were evaluated using calls on WGS long-insert data (19). The threshold for a positive detection of a CNA by Seq-FISH was 20%. Non-synonymous alterations with an allele ratio of at least 5% in the tumor sample and less than 2% in the constitutional sample occurring in a customized panel of 21 genes known to be significantly mutated in MM were also analyzed (*Table S1*) (20,21). The cancer cell fraction (CCF) of mutations of interest corrected by tumor purity and MM cell ploidy was estimated using the ABSOLUTE algorithm (22). Moreover, we evaluated the aberrant activity of APOBEC cytidine deaminases (known to be associated with high mutational burden and poor prognosis in MM) (23), using the recently developed fitting algorithm *mmsig* (*Table S1*; <https://github.com/evenrus/mmsig>) (24). APOBEC activity was defined as *high* or *low* based on its quartile distribution (4th quartile vs others) (23).

Statistical analysis

Early PD was defined as occurring in the first 18 months from the start of treatment. Patients not experiencing PD within 18 months from the start of treatment were included in the reference population. The reference population was further classified in Late PD (occurring after the first 18 months from the start of treatment) and No PD at the last follow-up. TTP was defined as the duration from start of treatment to PD; deaths from causes other than progression were censored (25).

Epanechnikov kernel smoothed estimated hazard rates were used to study the risk of PD over time.

Best response to first-line treatment and drug refractoriness after first-line treatment were evaluated according to the International Myeloma Working Group guidelines (25,26). The

comparison of best response and drug refractoriness in the Early vs Late PD groups was performed according to two-sided Fisher's exact test.

Overall survival (OS) was analyzed as time-to-event data using the Kaplan–Meier method. The Cox proportional hazards model was used to estimate the hazard ratio (HR) values and the 95% confidence intervals (CIs). In order to account for potential confounders, the comparison of Early PD vs reference population was adjusted for age, ISS, high-risk cytogenetics (27), induction treatment, ASCT, CT and clinical trial enrollment. ASCT and CT were considered as time-dependent variables.

An 18-month landmark analysis for OS was also performed, comparing OS in the Early PD vs Late PD vs No PD groups.

To identify risk factors associated with early relapse, patients that were not at risk for progression for the entire 18-month period after the start of treatment were excluded from the reference population (n=101, *Figure 1*).

Univariate analysis of factors associated with Early PD vs Late/No PD was performed using Fisher's exact test, Kruskal-Wallis test or Chi-squared test as appropriate. Starting from the variables with a p-value <0.15 in univariate analysis, the final logistic model was identified through a backward selection based on the minimization of the Akaike Information Criterion (AIC), keeping in the model the therapy-related variables. The final logistic regression model was used to estimate odds ratio (OR) for Early relapse risk, 95% CIs and p-values. A confirmatory analysis on the same patient population using death within 24 months as an endpoint was conducted (11).

All the analyses were conducted using R version 3.5.1 and bespoke code, which is available upon request.

Results

Patient characteristics

Data from 1151 patients were available in the CoMMpass IA14. Patients without whole-exome sequencing (WES) data (n=213) and PD information (n=12) were excluded from the analysis. The remaining 926 patients represented the population analyzed in the current work. Patient characteristics are shown in *Table 1*.

Median age was 63 years and most of the patients had an Eastern Cooperative Oncology Group (ECOG) performance status of 0 or 1 (39% and 44%, respectively). Baseline prognostic factors were typical of a NDMM population. 27% of patients presented with ISS stage III and 8% with high LDH levels; 13% of patients presented with del(17p), 14% with t(4;14), 5% with t(14;16), 1% with t(14;20), 27% with gain(1q) and 7% with amp(1q), while IgL translocations, a recently described marker of high-risk MM (19), were present in 10% of evaluable patients.

Genes affected by somatic non-synonymous alterations in at least 25 (3%) patients were analyzed (*Table S3*). Mutational frequency was dominated by alterations in KRAS (25%), NRAS (21.5%) and IGLL5 (16%) gene.

The most frequent induction regimen administered was bortezomib-lenalidomide-dexamethasone (VRd) (34%), followed by bortezomib+chemotherapy triplets (23%) and carfilzomib-based treatment (23%).

Patients evaluable for the ASCT vs no ASCT comparison were 833. Not evaluable patients experienced PD during induction (n=40), died for reasons other than PD (n=18), were lost to follow-up (n=14), withdrew consent (n=5), or discontinued the study for other reasons (n=6). Ten patients received ASCT after the 18-month endpoint and were considered not evaluable as

well. High-dose chemotherapy followed by ASCT was received by 53% of the evaluable patients; the median time to ASCT was 169 days (range 78-508).

Patients evaluable for CT vs FDT comparison were 609. Not evaluable patients, during the first year of treatment had PD (n= 112), died for reasons other than PD (n= 32), were lost to follow-up (n= 21), withdrew consent (n= 16) or discontinued the study for other reasons (n= 15). In 121 patients, information of drugs used during CT was lacking at the current follow-up. 74% of evaluable patients received CT (IMiDs 42%, PIs 14% and IMiDs+PIs 18%); 26% of patients received FDT. The distributions of induction treatment and ASCT in each CT subgroup are shown in *Table S4*.

Early PD population

The median follow-up of the entire population was 39 months. 191/926 (20.6%) patients experienced early PD, while the remaining 735/926 (79.4%) patients were included in the reference population (*Figure 1*).

In the Early PD group, 126/191 (66%) patients discontinued the study at the last follow-up: 75 (39%) for death due to PD, 26 (14%) for death due to other reasons, 4 (2%) due to withdrawal of consent, 3 (2%) for being lost to follow-up, and 18 (9%) for other reasons.

In the reference population, 229/735 (31%) patients discontinued the study: 39 (5%) for death due to PD, 66 (9%) for death due to other reasons, 31 (4%) due to withdrawal of consent, 39 (5%) for being lost to follow-up, and 54 (7%) for other reasons. In the same reference population, 228/926 (24.6%) patients experienced a late PD (TTP>18 months), while 507/926 (54.8%) did not experience PD at the last follow-up.

Overall response rate (ORR) was significantly lower in Early-PD patients compared to Late-PD patients (80% vs 96%, respectively, $p<0.001$). Deep responses were also different, with very

good partial response (VGPR) rates of 40% vs 57%, complete remission (CR) rates of 2% vs 18% and stringent CR rates of 5% vs 8% in Early vs Late PD groups respectively. This translated into a significantly different rate of \geq VGPR in the 2 groups (47% vs 82%, $p < 0.001$; *Table 2*).

A significantly higher proportion of patients in the Early vs the Late PD group developed a refractoriness to PIs (50% vs 18%, $p < 0.001$) and IMiDs+PIs (21% vs 8%, $p < 0.001$), while no differences were found in terms of IMiD refractoriness (42% vs 38%, $p = 0.541$; *Table 2*).

OS of Early-PD patients vs. the reference population is shown in *Figure 2*.

Early-PD patients had a significantly higher risk of death compared to the reference population (HR 4.89, 95% CI 3.72-6.43, $p < 0.001$), with 53% of patient deaths at 3 years in the early PD cohort compared with only 12% in the reference cohort. This effect was maintained after adjusting the analysis for age, baseline prognostic factors (ISS, high-risk cytogenetics(27)), treatment and clinical trial enrollment (HR 3.65, 95% CI 2.70-4.93, $p < 0.001$). Of note, 61% of early relapsing patients presented with ISS stage I or II and 74% had conventionally defined standard-risk cytogenetics (27). The median OS of early relapsing patients was 32.8 months, lower than that of high-risk population defined using baseline ISS III (median OS 54 months) or baseline high-risk cytogenetics (27) (median OS 65 months).

Early-PD patients were defined using a time-dependent endpoint (18 months); consequently, a landmark analysis of OS with a landmark point at 18 months was performed to validate our findings (*Figure 3*). At the landmark timepoint, 121 Early-PD patients and 640 patients in the reference population were evaluable. The main reasons for not being evaluable were death due to PD during the first 18 months in the Early PD population (58/191, 30%) and death due to reasons other than PD during the first 18 months in the reference population (42/735, 6%). The difference in early death rates between the 2 groups led to a possible underestimation of OS differences after the landmark timepoint. Moreover, in this OS comparison we split the reference population in Late PD and No PD patients. The 18-month landmark analysis showed

a significantly worse OS in Early-PD patients compared both to Late PD (HR 2.05, 95%, CI 1.25-3.35, $p=0.004$) and No PD patients (HR 8.05, 95%, CI 4.11-15.74, $p<0.001$).

Risk of early PD

We investigated the clinical and prognostic variables impacting the risk of early relapse. In this analysis, we excluded from the reference population the patients who were not at risk for the entire 18-month period (101/926, 11%). Excluded patients were those that in the first 18 months died without a PD ($n=42$), withdrew the consent ($n=14$), were lost to follow-up ($n=25$) or interrupted the protocol for other reasons ($n=20$).

A significantly higher proportion of patients in the Early PD group vs the reference population presented with ISS stage III (39% vs 20%), gain(1q) (26% vs 20%), IgL translocations (14% vs 6%), high APOBEC signature (30% vs 24%), high LDH (9% vs 5%), ECOG \geq 2 (23% vs 11%), *KRAS* mutation (31% vs 24%), *IGLL5* mutation (20% vs 14%) and *TP53* mutation (9% vs 3%) (*Table S5*). These variables were therefore included in multivariate analysis, together with age and treatment administered.

In multivariate analysis (*Figure 4*) *TP53* mutation (OR 3.78, $p<0.01$), high LDH levels (OR 3.15, $p<0.01$), IgL translocation (OR 2.25, $p=0.03$) and *IGLL5* mutation (OR 2.15, $p<0.01$) were significantly correlated with a higher risk of early PD. Only a trend was found for gain(1q) and amp(1q) (*Figure 4*).

Receiving ASCT (OR 0.27, $p<0.01$) and CT with IMiDs+PIs (OR 0.34, $p=0.02$) were significantly correlated with a lower risk of early PD. The effect of ASCT was confirmed in age-specific patient subgroups, showing similar ORs in patients aged \leq 65 years ($n=531$, OR 0.27 95%, CI 0.13-0.54) and aged 66-75 years ($n=222$, OR 0.30 95%, CI 0.11-0.74).

A protective effect of carfilzomib-based induction was also observed (OR 0.15, $p=0.01$). Nevertheless, most of carfilzomib-treated patients were enrolled in a clinical trial and the enrollment effect itself was a protective factor as well (OR 0.09, $p<0.01$).

To confirm our results, we performed an additional analysis using death within 24 months as an endpoint (11) (*Figure S1*). The adverse effects of TP53 mutation (OR 3.35, $p=0.02$) and IgL translocation (OR 2.34, $p=0.046$) were confirmed. Moreover, also 1q abnormalities were significantly correlated with an increased risk of death within 24 months. ASCT (OR 0.44, $p=0.02$) retained its protective effect.

TP53 mutations

In our analysis, TP53 mutation was the factor with the greatest effect size for early PD. Its association with MM patients carrying concurrent del(17p) is well known. In this cohort, 865 patients were evaluable for TP53 mutation and del(17p) (*Figure S2A*). One hundred twenty-one of 865 patients had del(17p) or TP53 mutation. Among them, 82/121 (68%) had del(17p) only, 10/121 (8%) had TP53 mutation only and 29/121 (24%) had del(17p) and TP53 mutation. Rates of early PD in each patient subgroup are shown in *Figure S2B*. Patients with del(17p) but not TP53 mutation had an early PD rate of 17.1% (comparable with the general population), while the bi-allelic group (del(17p)+TP53 mutation) and the TP53-mutation-only group showed high early PD rates (41.4% and 50%, respectively). Of note, the TP53-mutation-only group was composed by only 10 patients and the majority of TP53-mutated patients experiencing early relapse were in the del(17p)+TP53 mutation group.

The use of a higher cut-off level to define del(17p) positivity (50% instead of 20%, *Figure S2C-D*) led to a slightly higher early PD rate in del(17p)-only patients (25%). However, the bi-allelic

(del(17p)+TP53 mutation) and the TP53-mutation-only groups still showed the highest rates of early PD (40.7% and 50%, respectively).

Longitudinal analysis of mutations associated with early PD

Considering that TP53 mutation is important to confer early relapse risk, we hypothesized that TP53-mutated clones needed to be conserved at relapse. Only 6 patients with TP53 mutation at diagnosis had available molecular data at relapse, although in 6/6 cases TP53 mutation was conserved in relapse samples (*Figure S3A*). Moreover, despite the small numbers, if TP53 mutation was subclonal at diagnosis, a higher cancer cell fraction was found in paired samples at relapse. This effect was different from the IGLL5 mutations, in which subclonal cases tended to disappear at relapse (*Figure S3B*).

Discussion

MM prognosis is improving and early relapse after upfront treatment is beginning to be recognized as a high-risk feature (28). The same observation had been done for other hematologic malignancies with an expected indolent course, such as follicular lymphoma and chronic lymphocytic leukemia (29,30).

Here we proposed progression ≤ 18 months after the start of first-line treatment as a marker of high risk and demonstrated its detrimental effect on the OS of NDMM patients.

The 18-month cut-off was chosen because our time to ASCT was ~ 6 months and the majority of published studies on MM patients with early PD defined early PD as a relapse within 12 months from ASCT. Indeed, the hazards of progression in our patient population increased over time with no identified peak of risk (*Figure S4*).

We incorporated in our analysis baseline clinical and biological features to identify risk factors of early PD. The characterization by NGS of this patient cohort allowed us to simultaneously study copy number abnormalities (CNAs), translocations and mutations in genes of interest by using the same platform. This is an advantage of NGS vs conventional fluorescence in situ hybridization (FISH), which cannot detect mutations and needs specific probes to detect pre-specified translocations and CNAs. Moreover, NGS and conventional FISH showed high concordance in detecting the same CNAs and translocations, as shown in *Figure S5* and by others (17,18).

TP53 mutation, which is currently not included in the standard baseline evaluation of MM patients, was the most important factor increasing the risk of early PD emerging from our analysis. Its adverse effect was confirmed in the risk of death within 24 months from diagnosis. TP53 mutation is rare in patients at diagnosis (3.5%), but about 25% of patients with del(17p) has also TP53 mutation. As similarly observed by other groups (9), our data further supported the routine testing of TP53 mutation at least in del(17p)-positive patients. Indeed, the presence of del(17p) without TP53 mutation conferred an early PD risk that was similar to that of the overall population.

In our analysis, IgL translocation and IGLL5 mutation also emerged as risk factors of early PD. Both of them have already been associated with poor prognosis (19,31). White et al. showed that mutations in IGLL5 can be associated with translocations juxtaposing IGLL5 (31). In our analysis, IGLL5 mutations and IgL translocations showed a trend toward co-occurrence, though not statistically significant ($p=0.06$). The higher risk of early relapse observed in IgL-translocated patients, the loss of subclonal IGLL5 mutations at first relapse and the significant effect of IgL translocations but not of IGLL5 mutations in the risk of death within 24 months could suggest that IgL translocations impacted patients' prognosis more than IGLL5 mutations.

Only a trend towards a higher risk of early PD was found for gain(1q) and amp(1q). However, using death within 24 months as an endpoint, the effect of 1q abnormalities was more evident. This was possibly due to the use of a later timepoint allowing more patients to experience an event and to a possible more specific effect of 1q abnormalities on the risk of death.

In our analysis, the only clinical factor that increased the risk of early PD in multivariate analysis was baseline LDH, a well-known marker of disease aggressiveness in several hematologic diseases.

Other factors not included in the current analysis – such as circulating plasma cells (32), high-risk GEP(8,33) and MM cell-extrinsic factors (34) – could also play a role in determining the risk of early PD and should be investigated in future works. Moreover, our analysis focused on MM cells derived from a random bone marrow aspirate, and spatial heterogeneity of high-risk features could also explain some of the early PD cases (35).

ASCT and CT with IMiDs+PIs showed a protective effect against early PD in this patient population. However, the majority of patients in the analyzed cohort were real-world patients and the analysis was consequently performed as per protocol, thus leading to a risk of overestimation of effects of ASCT and CT. With these limitations, our data support the intensification of therapy in patients at risk of early relapse and underline the importance of continuous treatment with combination regimens to optimize long-term disease control (36).

Carfilzomib-based induction also showed to reduce the risk of early relapse, although it is difficult to distinguish between treatment and trial effects because the majority of carfilzomib-treated patients were included in a clinical trial, whereas this was not the case for other induction regimens.

Besides clinical trial enrollment, this patient population was heterogeneously treated and our findings on early PD risk need to be confirmed in homogeneously treated patients. For instance, among the CT subgroups, heterogeneous upfront treatments before CT were received (*Table*

S4). Nevertheless, the multivariate analysis on the risk of early PD was adjusted for induction treatment, ASCT, CT and trial enrollment effect, taking into account these differences.

The median age of the analyzed cohort was 63 years, younger than the usual median age of unselected MM patients. Elderly patients were underrepresented and the confirmation of our results in this patient population is warranted. However, other variables that are patient-related but not disease-related (e.g. frailty status) may have a major prognostic role in elderly patients (37).

Early-PD patients showed suboptimal responses and, at relapse, were more frequently refractory to PIs and double refractory to IMiDs+PIs, as compared to Late-PD patients. IMiD refractoriness was not different between Early PD and Late PD groups. This was mainly due to the widespread use of PI-containing regimens during the first 18 months of therapy. On the other hand, after the 18-month timepoint, treatment with an IMiD as single agent was widely used in our patient population. Therefore, a high percentage of PI-refractory and IMiD+PI-refractory cases were observed in the Early PD group, while IMiD-refractory cases were well represented in both the Early PD and Late PD groups.

In conclusion, early PD identifies a high-risk MM population that still represents an unmet clinical need. As compared with FISH, extended genotyping through the routine use of NGS at diagnosis is feasible and may improve the patient stratification and identify patients at risk of early PD (38). Further research is needed to better identify baseline features predicting early relapse and the optimal treatment approach. Recently, clinical trials on patients experiencing PD within 18 months from the start of treatment are beginning to emerge (e.g. NCT03601078, cohorts 2a and 2b), thus suggesting that risk-adapted treatment in this patient population could soon become a feature of MM clinical management.

References

1. Usmani SZ, Hoering A, Cavo M, Miguel JS, Goldschmidt H, Hajek R, et al. Clinical predictors of long-term survival in newly diagnosed transplant eligible multiple myeloma — an IMWG Research Project. *Blood Cancer J* [Internet]. 2018 [cited 2019 Oct 7];8:123. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/30470751>
2. Kazandjian D. Multiple myeloma epidemiology and survival: A unique malignancy. *Semin Oncol* [Internet]. 2016 [cited 2019 Sep 30];43:676–81. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/28061985>
3. Mai EK, Haas E-M, Lücke S, Löprrich M, Kunz C, Pritsch M, et al. A systematic classification of death causes in multiple myeloma. *Blood Cancer J* [Internet]. 2018 [cited 2019 Sep 30];8:30. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/29520024>
4. Ferlay J, Colombet M, Soerjomataram I, Dyba T, Randi G, Bettio M, et al. Cancer incidence and mortality patterns in Europe: Estimates for 40 countries and 25 major cancers in 2018. *Eur. J. Cancer*. Elsevier Ltd; 2018. page 356–87.
5. Keats JJ, Chesi M, Egan JB, Garbitt VM, Palmer SE, Braggio E, et al. Clonal competition with alternating dominance in multiple myeloma. *Blood* [Internet]. 2012 [cited 2019 Sep 30];120:1067–76. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/22498740>
6. Majithia N, Rajkumar S V, Lacy MQ, Buadi FK, Dispenzieri A, Gertz MA, et al. Early relapse following initial therapy for multiple myeloma predicts poor outcomes in the era of novel agents. *Leukemia* [Internet]. 2016 [cited 2019 Sep 30];30:2208–13. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/27211270>
7. Palumbo A, Avet-Loiseau H, Oliva S, Lokhorst HM, Goldschmidt H, Rosinol L, et al. Revised International Staging System for Multiple Myeloma: A Report From International Myeloma Working Group. *J Clin Oncol* [Internet]. American Society of Clinical Oncology; 2015 [cited 2017 Sep 5];33:2863–9. Available from: <http://ascopubs.org/doi/10.1200/JCO.2015.61.2267>
8. Kuiper R, Broijl A, van Duin M, van Vliet1 MH, Levin M-D, van Beers1 EH, et al. PROGNOSTIC AND PREDICTIVE PERFORMANCE OF SKY92 COMBINED WITH R-ISS IN ELDERLY MULTIPLE MYELOMA PATIENTS IN THE HOVON- 87/NMSG-18 STUDY. 17th IMW [International Myeloma Workshop] Abstract Book. Boston; 2019. page 15-16 [Abstract #OAB-013]. Available from: https://files.aievolution.com/imw1901/docs/17th_IMW_Abstract_Book_FINAL_V2.pdf
9. Walker BA, Mavrommatis K, Wardell CP, Ashby TC, Bauer M, Davies F, et al. A high-risk, Double-Hit, group of newly diagnosed myeloma identified by genomic analysis. *Leukemia* [Internet]. 2019 [cited 2019 Sep 30];33:159–70. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/29967379>
10. Bolli N, Biancon G, Moarii M, Gimondi S, Li Y, de Philippis C, et al. Analysis of the genomic landscape of multiple myeloma highlights novel prognostic markers and disease subgroups. *Leukemia*. Nature Publishing Group; 2018;32:2604–16.
11. Avet-Loiseau H. Ultra High-Risk Myeloma. *Hematology* [Internet]. 2010 [cited 2019 Sep 30];2010:489–93. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/21239841>
12. Lonial S, Boise LH, Kaufman J. How I treat high-risk myeloma. *Blood* [Internet]. 2015 [cited 2019 Sep 30];126:1536–43. Available from: <http://www.bloodjournal.org/cgi/doi/10.1182/blood-2015-06-653261>
13. Palumbo A, Gay F, Cavallo F, Di Raimondo F, Larocca A, Hardan I, et al. Continuous Therapy Versus Fixed Duration of Therapy in Patients With Newly Diagnosed Multiple Myeloma. *J Clin Oncol* [Internet]. 2015 [cited 2019 Apr 5];33:3459–66. Available from: <http://ascopubs.org/doi/10.1200/JCO.2014.60.2466>

14. Keats J, Speyer G, Christofferson A, Stephenson K, Kurdoglu A, Russell M, et al. Interim Analysis of the MMRF CoMMpass Study: Comprehensive Characterization of Multiple Myeloma Patients at Diagnosis Reveals Distinct Molecular Subtypes and Clinical Outcomes. *Clin Lymphoma Myeloma Leuk* [Internet]. Elsevier; 2015 [cited 2019 Sep 30];15:e44–5. Available from: <https://www.sciencedirect.com/science/article/pii/S2152265015006102>
15. Miller A, Asmann Y, Cattaneo L, Braggio E, Keats J, Auclair D, et al. High somatic mutation and neoantigen burden are correlated with decreased progression-free survival in multiple myeloma. *Blood Cancer J* [Internet]. Nature Publishing Group; 2017 [cited 2019 Sep 30];7:e612–e612. Available from: <http://www.nature.com/articles/bcj201794>
16. Maura F, Bolli N, Angelopoulos N, Dawson KJ, Leongamornlert D, Martincorena I, et al. Genomic landscape and chronological reconstruction of driver events in multiple myeloma. *Nat Commun*. Springer Science and Business Media LLC; 2019;10.
17. Miller C, Yesil J, Derome M, Donnelly A, Marrian J, McBride K, et al. A Comparison of Clinical FISH and Sequencing Based FISH Estimates in Multiple Myeloma: An Mmrf Commpass Analysis. *Blood* [Internet]. 2016 [cited 2019 Sep 30];128:Abstract #374 [ASH 2016 58th Meeting]. Available from: <http://www.bloodjournal.org/content/128/22/374>
18. Goldsmith SR, Fiala MA, Dukeman J, Ghobadi A, Stockerl-Goldstein K, Schroeder MA, et al. Next Generation Sequencing-based Validation of the Revised International Staging System for Multiple Myeloma: An Analysis of the MMRF CoMMpass Study. *Clin Lymphoma Myeloma Leuk* [Internet]. 2019 [cited 2019 Sep 30];19:285–9. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S2152265018315246>
19. Barwick BG, Neri P, Bahlis NJ, Nooka AK, Dhodapkar M V., Jaye DL, et al. Multiple myeloma immunoglobulin lambda translocations portend poor prognosis. *Nat Commun* [Internet]. Nature Publishing Group; 2019 [cited 2019 Sep 30];10:1911. Available from: <http://www.nature.com/articles/s41467-019-09555-6>
20. Bolli N, Avet-Loiseau H, Wedge DC, Van Loo P, Alexandrov LB, Martincorena I, et al. Heterogeneity of genomic evolution and mutational profiles in multiple myeloma. *Nat Commun* [Internet]. 2014 [cited 2019 Sep 30];5:2997. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24429703>
21. Kortum KM, Mai EK, Hanafiah NH, Shi C-X, Zhu Y-X, Bruins L, et al. Targeted sequencing of refractory myeloma reveals a high incidence of mutations in CRBN and Ras pathway genes. *Blood* [Internet]. 2016 [cited 2019 Sep 30];128:1226–33. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/27458004>
22. Carter SL, Cibulskis K, Helman E, McKenna A, Shen H, Zack T, et al. Absolute quantification of somatic DNA alterations in human cancer. *Nat Biotechnol* [Internet]. 2012 [cited 2019 Sep 30];30:413–21. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/22544022>
23. Maura F, Petljak M, Lionetti M, Cifola I, Liang W, Pinatel E, et al. Biological and prognostic impact of APOBEC-induced mutations in the spectrum of plasma cell dyscrasias and multiple myeloma cell lines. *Leukemia* [Internet]. 2018 [cited 2019 Sep 30];32:1044–8. Available from: <http://www.nature.com/articles/leu2017345>
24. Maura F, Degasperi A, Nadeu F, Leongamornlert D, Davies H, Moore L, et al. A practical guide for mutational signature analysis in hematological malignancies. *Nat Commun*. Nature Publishing Group; 2019;10.
25. Rajkumar SV, Harousseau J-L, Durie B, Anderson KC, Dimopoulos M, Kyle R, et al. Consensus recommendations for the uniform reporting of clinical trials: report of the International Myeloma Workshop Consensus Panel 1. *Blood* [Internet]. 2011 [cited

- 2017 May 9];117:4691–5. Available from:
<http://www.bloodjournal.org/cgi/doi/10.1182/blood-2010-10-299487>
26. Kumar S, Paiva B, Anderson KC, Durie B, Landgren O, Moreau P, et al. International Myeloma Working Group consensus criteria for response and minimal residual disease assessment in multiple myeloma. *Lancet Oncol* [Internet]. 2016 [cited 2017 Jul 12];17:e328–46. Available from:
<http://linkinghub.elsevier.com/retrieve/pii/S1470204516302066>
 27. Fonseca R, Bergsagel PL, Drach J, Shaughnessy J, Gutierrez N, Stewart AK, et al. International Myeloma Working Group molecular classification of multiple myeloma: spotlight review. *Leukemia* [Internet]. 2009 [cited 2019 Sep 25];23:2210–21. Available from: <http://www.nature.com/articles/leu2009174>
 28. Kumar SK, Dispenzieri A, Fraser R, Mingwei F, Akpek G, Cornell R, et al. Early relapse after autologous hematopoietic cell transplantation remains a poor prognostic factor in multiple myeloma but outcomes have improved over time. *Leukemia* [Internet]. 2018 [cited 2019 Sep 30];32:986–95. Available from:
<http://www.nature.com/articles/leu2017331>
 29. Casulo C, Byrtek M, Dawson KL, Zhou X, Farber CM, Flowers CR, et al. Early Relapse of Follicular Lymphoma After Rituximab Plus Cyclophosphamide, Doxorubicin, Vincristine, and Prednisone Defines Patients at High Risk for Death: An Analysis From the National LymphoCare Study. *J Clin Oncol* [Internet]. 2015 [cited 2019 Sep 30];33:2516–22. Available from: <http://ascopubs.org/doi/10.1200/JCO.2014.59.7534>
 30. Ahn IE, Farber CM, Davids MS, Grinblatt DL, Kay NE, Lamanna N, et al. Early progression of disease as a predictor of survival in chronic lymphocytic leukemia. *Blood Adv* [Internet]. 2017 [cited 2019 Sep 30];1:2433–43. Available from:
<http://www.ncbi.nlm.nih.gov/pubmed/29296893>
 31. White BS, Lanc I, O’Neal J, Gupta H, Fulton RS, Schmidt H, et al. A multiple myeloma-specific capture sequencing platform discovers novel translocations and frequent, risk-associated point mutations in IGLL5. *Blood Cancer J* [Internet]. 2018 [cited 2019 Sep 30];8:35. Available from: <http://www.nature.com/articles/s41408-018-0062-y>
 32. Granell M, Calvo X, Garcia-Guiñón A, Escoda L, Abella E, Martínez CM, et al. Prognostic impact of circulating plasma cells in patients with multiple myeloma: implications for plasma cell leukemia definition. *Haematologica* [Internet]. 2017 [cited 2019 Oct 7];102:1099–104. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/28255016>
 33. Zhou Y, Chen L, Barlogie B, Stephens O, Wu X, Williams DR, et al. High-risk myeloma is associated with global elevation of miRNAs and overexpression of EIF2C2/AGO2. *Proc Natl Acad Sci* [Internet]. 2010 [cited 2019 Sep 30];107:7904–9. Available from:
<http://www.ncbi.nlm.nih.gov/pubmed/20385818>
 34. Manier S, Sacco A, Leleu X, Ghobrial IM, Roccaro AM. Bone Marrow Microenvironment in Multiple Myeloma Progression. *J Biomed Biotechnol* [Internet]. 2012 [cited 2019 Sep 30];2012:1–5. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/23093834>
 35. Rasche L, Chavan SS, Stephens OW, Patel PH, Tytarenko R, Ashby C, et al. Spatial genomic heterogeneity in multiple myeloma revealed by multi-region sequencing. *Nat Commun*. 2017;8:268.
 36. D’Agostino M, De Paoli L, Conticello C, Offidani M, Ria R, Petrucci MT, et al. Continuous therapy in standard- and high-risk newly-diagnosed multiple myeloma: A pooled analysis of 2 phase III trials. *Crit Rev Oncol Hematol* [Internet]. 2018 [cited 2019 Oct 7];132:9–16. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/30447931>
 37. Bringhen S, D’Agostino M, Paris L, Ballanti S, Pescosta N, Spada S, et al. Lenalidomide-based induction and maintenance in elderly newly diagnosed multiple myeloma patients: updated results of the EMN01 randomized trial. *Haematologica* [Internet].

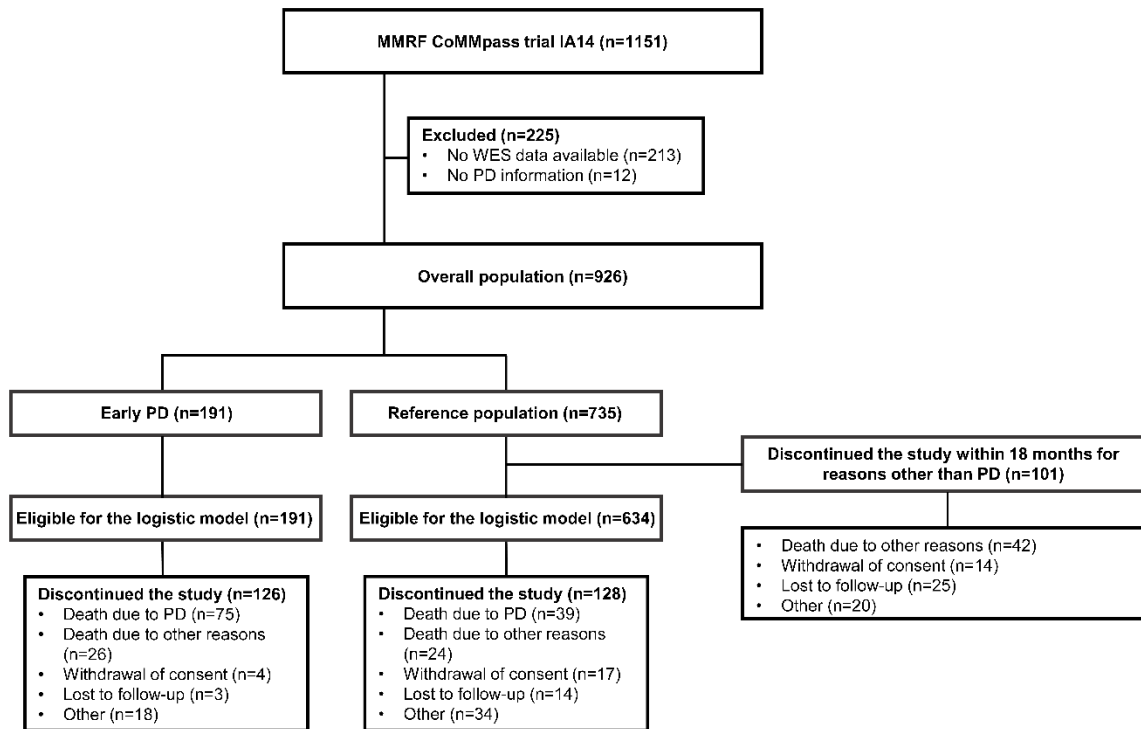
2019 [cited 2019 Oct 17];[Epub ahead of print]. Available from:
<http://www.ncbi.nlm.nih.gov/pubmed/31582542>

38. Bolli N, Li Y, Sathiaseelan V, Raine K, Jones D, Ganly P, et al. A DNA target-enrichment approach to detect mutations, copy number changes and immunoglobulin translocations in multiple myeloma. *Blood Cancer J* [Internet]. 2016 [cited 2019 Nov 11];6:e467. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/27588520>

Figures

Figure 1. Study flow

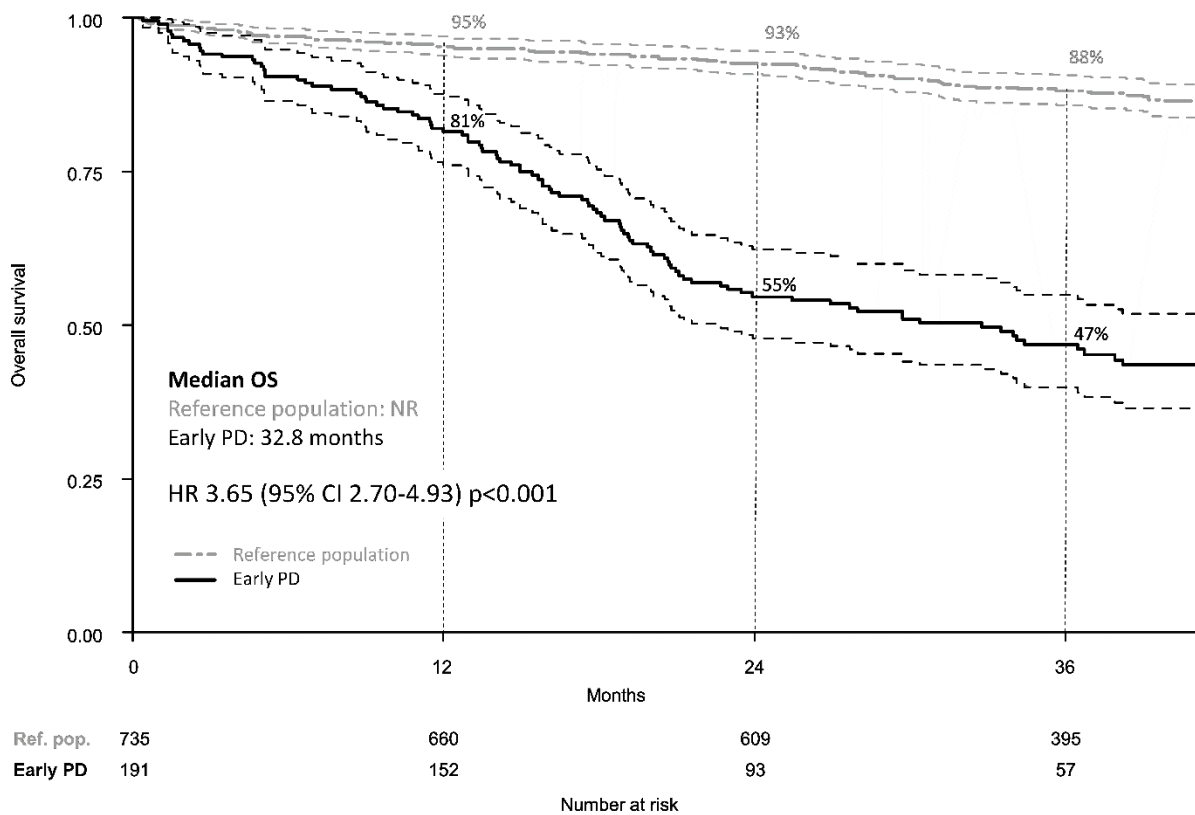
Figure 1



Abbreviations. MMRF: Multiple Myeloma Research Foundation; IA14: Interim analysis 14; WES: whole exome sequencing; PD: progressive disease; n, number.

Figure 2. Overall survival for patients with early PD versus reference population

Figure 2

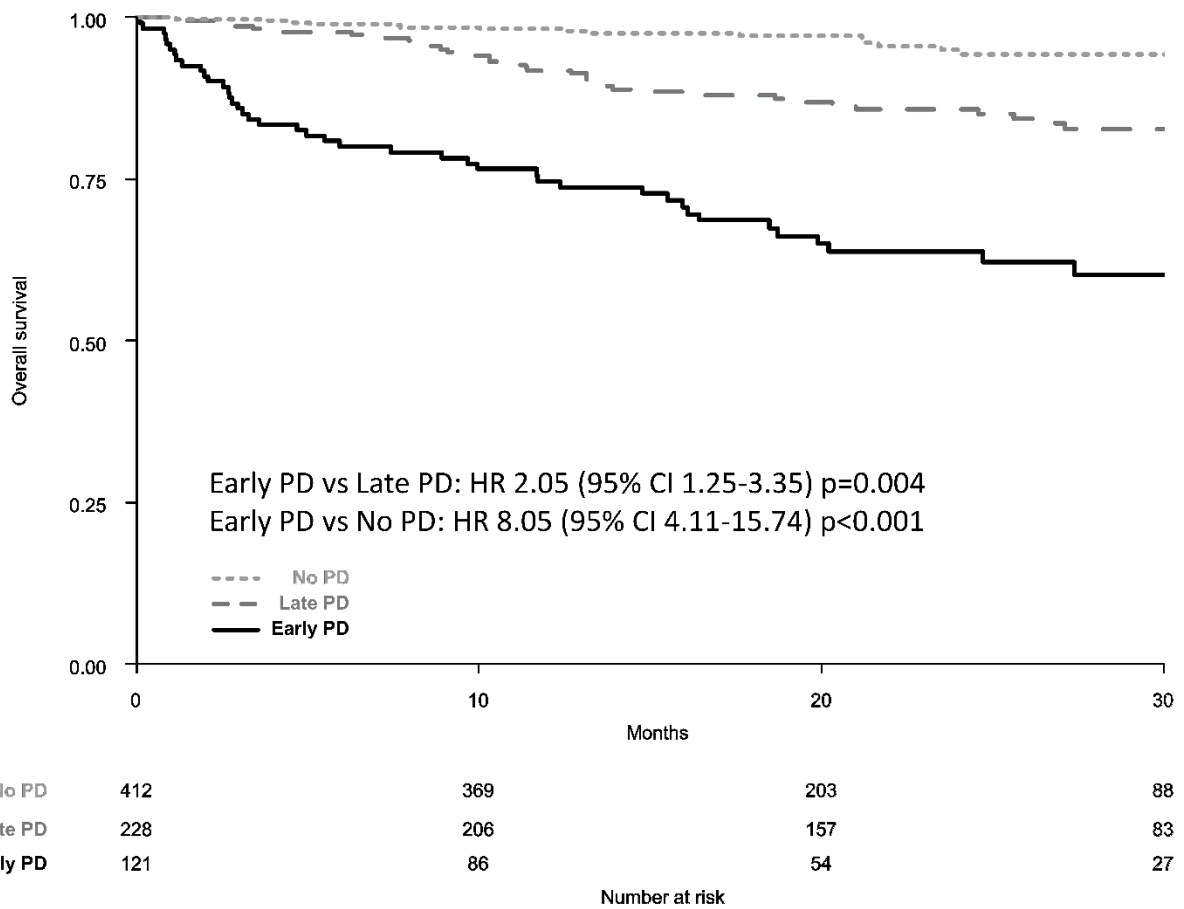


Abbreviations. OS: overall survival; PD: progressive disease; HR: hazard ratio; NR: not reached; ref. pop., reference population.

Dotted lines: 95% confidence intervals. HR adjusted for age, International Staging System (ISS) stage, high-risk cytogenetics [presence of del(17p) and/or t(4;14) and/or t(14;16], induction treatment, autologous stem-cell transplantation (ASCT), continuous therapy (CT), and clinical trial enrollment.

Figure 3. 18-month landmark analysis for OS in Early PD versus Late PD versus No PD patients

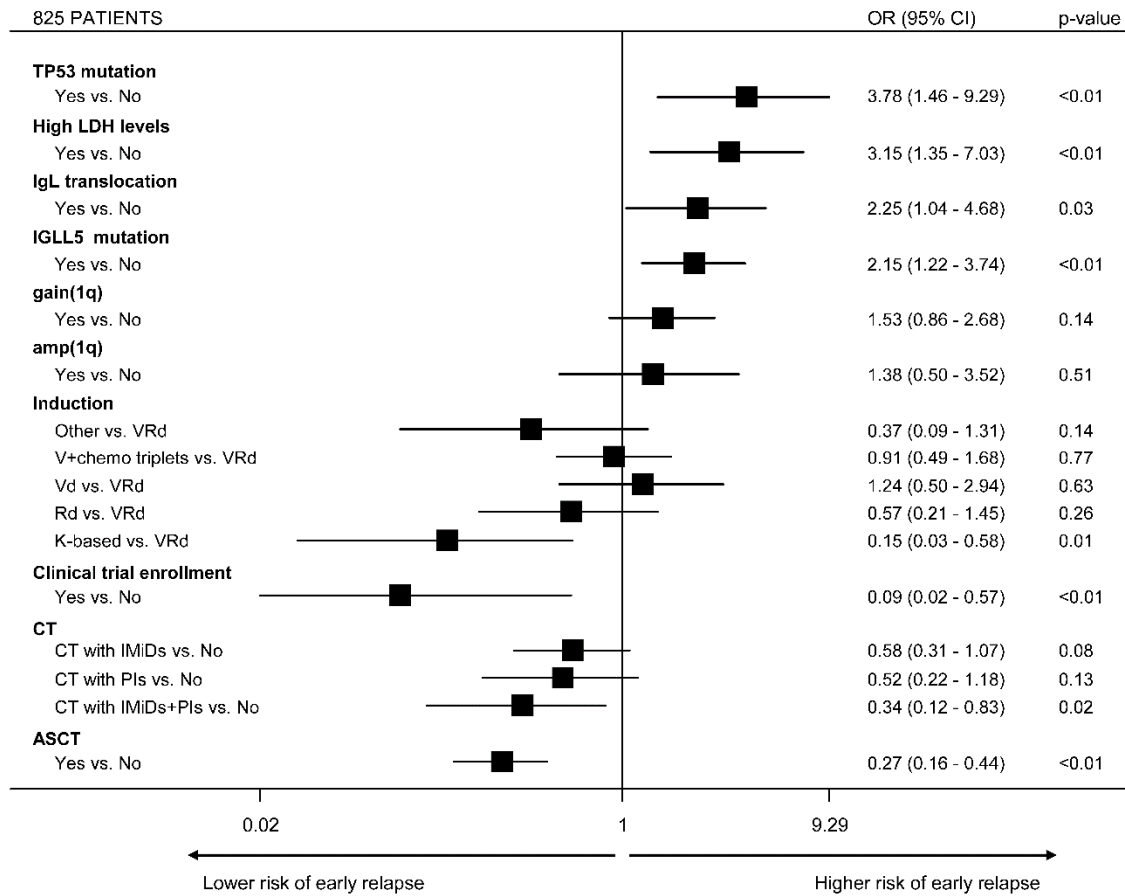
Figure 3



Abbreviations. OS: Overall survival; PD: progressive disease; HR: hazard ratio. HR adjusted for age, International Staging System (ISS) stage, high-risk cytogenetics [presence of del(17p) and/or t(4;14) and/or t(14;16)], induction treatment, autologous stem-cell transplantation (ASCT), continuous therapy (CT), and clinical trial enrollment.

Figure 4. Multivariate logistic regression model evaluating risk factors associated with early PD in the patients actually at risk for the entire 18-month period (n=825)

Figure 4



Abbreviations. PD, progressive disease; OR: odds ratio; IgL: immunoglobulin lambda chain; IGLL5, immunoglobulin lambda like polypeptide 5; LDH: lactate dehydrogenase; V: bortezomib; d: low dose dexamethasone; chemo: conventional chemotherapy; R: lenalidomide; K: carfilzomib; ASCT: autologous stem-cell transplantation; CT: continuous therapy; FDT: fixed-duration therapy; IMiDs: immunomodulatory drugs; PIs: proteasome inhibitors.

Analysis is adjusted for missing values within each variable.

Tables

Table 1. Patient characteristics

The entire cohort of patients (N=926) is shown.

Characteristic	N (%*)
Median follow-up	39 months
Median age (IQR)	63 (59-69)
Induction treatment	
VRd	319 (34%)
V+chemo triplets	216 (23%)
K-based	215 (23%)
Vd	83 (9%)
Rd	63 (7%)
Other	30 (3%)
ASCT	
Yes	440 (53%)
No	393 (47%)
Not evaluable	93
CT	
FDT	159 (26%)
IMiDs	258 (42%)
PIs	83 (14%)
IMiDs+PIs	109 (18%)
Not evaluable	317
Clinical trial enrollment	
Yes	166 (18%)
No	760 (82%)
ISS	
1	328 (37%)
2	325 (36%)
3	245 (27%)
Missing	28
CNAs	
Hyperdiploidy	499 (58%)
del(13q)	449 (52%)
del(17p)	111 (13%)
Not evaluable	61
gain(1q)	203 (27%)
amp(1q)	53 (7%)
Not evaluable	174
IgH translocations	
t(11;14)	179 (20%)
t(4;14)	123 (14%)
t(14;16)	42 (5%)
t(14;20)	12 (1%)
Not evaluable	25

IgL translocations	
Yes	77 (10%)
No	692 (90%)
Not evaluable	187
APOBEC mutational signature	
High	231 (25%)
Low	695 (75%)
Not evaluable	0
LDH	
High	60 (8%)
Normal	657 (92%)
Missing	209
ECOG	
0	329 (39%)
1	372 (44%)
≥2	141 (17%)
Missing	84

Abbreviations. IQR, interquartile range; V, bortezomib; d, low dose dexamethasone; chemo, conventional chemotherapy; R, lenalidomide; K, carfilzomib; IMiDs, immunomodulatory drugs; PIs, proteasome inhibitors; ASCT, autologous stem-cell transplantation; CT, continuous therapy; FDT, fixed-duration therapy; ISS, International Staging System; CNAs, Copy Number Abnormalities; IgH, immunoglobulin heavy chain; IgL, immunoglobulin lambda chain; LDH, lactate dehydrogenase; ECOG, Eastern Cooperative Oncology Group performance status.

*% calculated on evaluable cases within each variable.

Table 2. Best response to upfront treatment and drug refractoriness after first relapse in Early PD versus Late PD patients

	Early PD (n=191)	Late PD (n=228)	P-value
Best response to upfront treatment			
PD	9 (6%)	0	
SD	22 (14%)	8 (4%)	
PR	53 (34%)	31 (14%)	
VGPR	63 (40%)	129 (57%)	
CR	3 (2%)	40 (18%)	
sCR	8 (5%)	18 (8%)	
Not evaluable	33	2	
ORR	80%	96%	p<0.001
≥VGPR rate	47%	82%	p<0.001
Drug refractoriness after first relapse			
IMiD refractory	80 (42%)	86 (38%)	p=0.541
PI refractory	96 (50%)	41 (18%)	p<0.001
IMiD + PI double refractory	41 (21%)	18 (8%)	p<0.001

Abbreviations. PD, progressive disease; SD stable disease; PR partial response; VGPR very good partial response; CR, complete response; sCR, stringent CR; ORR, overall response rate (≥PR); n, number; IMiDs, immunomodulatory drugs; PIs, proteasome inhibitors.