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A preliminary analysis of the goat lactoferrin encoding gene

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RIASSUNTO – Analisi preliminare del gene che codifica la lattoferrina caprina. È stato sequenziato il tratto di DNA comprendente gli ultimi 30 nucleotidi del 2° introne e i primi 52 nucleotidi del 6° introne (per un totale di 2824 bp) del gene della lattoferrina (Lf) di 3 capre appartenenti rispettivamente alla razza Saanen, Maltese e ad una popolazione autoctona allevata in provincia di Catanzaro (Italia). Il confronto tra le sequenze ottenute ha evidenziato 8 siti polimorfici (6 transversioni e 2 transizioni realizzatesi a livello intronico), mentre il confronto con le sequenze dei cDNA della Lf caprina depositate in Banca Dati mostra 5 sostituzioni nucleotidiche responsabili di tre sostituzioni aminoacidiche. In generale, il gene Lf caprino presenta una struttura simile a quella dell'omologo gene nella specie bovina, fatta eccezione per l'inserzione in quest'ultima specie di una sequenza di origine retroposonica (Bov A) a livello del 4° introne.

KEY WORDS: Capra hircus, lactoferrin, milk protein, gene.

INTRODUCTION – Lactoferrin (Lf) is a glycoprotein with a molecular weight of ~80 kDa. It has been isolated from milk and identified in various different mammalian secretions. Several physiological functions have been ascribed to Lf, including regulation of cellular growth and differentiation (Iyer and Lonnerdal, 1993), intestinal iron homeostasis (Levay and Viljoen, 1995), host defense against microbial infection and inflammation (Baveye $et\ al.\ 1999$), regulation of myelopoiesis (Brock, 2002) and protection against cancer (Ward $et\ al.\ 2002$). The coding sequence of bovine lactoferrin gene is spread over 17 exons and 16 introns, spanning 34.5 kb of genomic DNA. The bovine and caprine lactoferrin gene was mapped to chromosome 22 (Schwerin $et\ al.\ 1994$). To date, only the nucleotide sequence of the goat $Lf\ cDNA$, 2333 bp in length, is known which contains 75 bp of 5' UTR (Un Traslated Region), the ORF (Open Reading Frame) coding 690 aminoacid of mature protein and the whole 3' UTR (Le Provost $et\ al.\ 1994$); Lee $et\ al.\ 1997$). The leader peptide from 19 full-length is codified by the last 14 codons of the exon 1 and the first five of the exon 2. The aim of this study was to carry out a preliminary analysis of the goat lactoferrin encoding gene, focusing on the region spanning from the 3^{rd} to the 6^{th} exon.

MATERIAL AND METHODS – For this study three different genetic types of goats (Saanen, Maltese and an autocton population reared in province of Catanzaro, Italy) were used. Genomic DNA was isolated from leucocytes. DNA regions spanning from the 2nd exon to 7th exon of the goat *Lf* gene were amplified using a Gene Amp PCR System 2400 (Perkin Elmer). Primers for amplification and sequencing were designed by means of DNASIS-Pro software (Hitachi), using the Lf cDNA sequences (EMBL n° U53857): Lf2F (5'-GCCCCGAG-GAAAAACGT-3'), Lf5R (5'-TTGAAGGCACCAGAATAAC-3'); Lf5F (5'-TGTGGCTAGATTCTTCTC-3'), Lf6R (5'-CAAACACTGTCGTCTCC-3'); Lf6F (5'-GGAGACGACAGTGTTTG-3'), Lf7R (5'-AACAGCATGAGAAGGGA-3'). A typical 50 μl of reaction mix comprised: 100 ng of genomic DNA, 3 mM MgCl₂, 200 nmol of each primer,

dNTPs each at 400 μ M, Buffer 1X and 2.5 U of Taq DNA Polymerase (Promega), 0.04% BSA. The thermal profile consisting of a total of 31 cycles involving 1 cycle at 97°C for 2 min, 46-62°C for 45 sec and 72°C for 2 min followed by 30 cycles at 94°C for 45 sec, 46-62°C for 45 sec and 72°C for 2 min, ending with a final extention at 72°C for 10 min. PCR products were analysed by electrophoresis using a 1.5% agarose gels (Biorad). Nucleotide sequencing was carried out according to the dideoxynucleotide chain-termination technique

RESULTS AND CONCLUSION – We amplified by means of PCR and sequenced the DNA region spanning the last 30 nucleotides of the 2^{nd} intron to the first 52 nt of the 6^{th} intron (2824 bp) of the goat Lf encoding gene. A comparison of the same sequenced regions for the three samples has shown a similarity of 99.3%, with a 73.8% homology with the corresponding bovine sequence. At a preliminary analysis, the goat Lf encoding gene shares a similar organization with the known bovine counterpart (Seyfert $et\ al.$, 1994) and its architecture seems to be extremely split, according to the structure of the α s1 and α s2 caseins encoding genes (CSN1S1 and CSN1S2, respectively) (Groenen $et\ al.$, 1993; Ramunno $et\ al.$, 2004). A comparison of the intronic sequenced regions has shown for the three goats the presence of 8 intronic polymorphic sites: 6 transversions and 2 transitions (Table 1).

Table 1. Intronic differences found in *Lf* gene of the three examined goats. Numbers are relative to nucleotide position in corresponding introns.

Intr	on Position	Saane	n Maltes	e Autocton
3	195	С	С	G
	1181	С	С	G
	1197	С	С	G
	1198	С	С	G
5	1257	G	С	G
	1420	С	С	Т
	1424	С	Т	С
	1611	С	Α	С

The mutations are probably not responsible for any difference in Lf gene expression since they do not affect splicing sites, which follow the 5' GT/3' AG splice rule. At the 3^{rd} intron, between nucleotides + 249 and + 260, a microsatellite sequence was evidenced; it is a $(CT)^6$ repetition that hasn't shown polymorphism in the three samples. Concerning the exonic regions, we sequenced the exons 3 (109 bp), 4 (183 bp), 5 (148 bp) and 6 (56 bp). A comparison of the sequenced exonic region with the published sequence of the French goat cDNA (Le Provost $et\ al.$, 1994) showed five nucleotide differences which result in three aminoacid substitutions. All the exonic differences are already known and it differenced the French from the Korean goat (Lee $et\ al.$, 1997) (Table 2).

Table 2. Exonic differences of the goat *Lf* gene evidenced by the comparison of the obtained sequences with those of the cDNA deposited in EMBL (n° X78902°; U53857°). Numbers are according to the published sequences.

Exon	Position	Italiar	Italian goats		French goat ^a		n goat⁵	
		DNA	Protein	DNA	Protein	DNA	Protein	
3	56	C T G	Leu	C G G	Arg	C T G	Leu	
	54	C AG	Gln	A AG	Lys	C AG	Gln	
4	74	GG C	Gln	GG T	Gln	GG C	Gln	
	144-145	TTC	Phe	CCC	Pro	TTC	Phe	

Finally the analysis of the partial goat Lf gene, from the 1st nt of the 3rd exon to the last nt of the 6th exon, evidenced a size ratio of exon vs. intron DNA of 1:4.15 vs. 1:4.45 in bovine counterpart. The different ratio observed between the two species is consequence of an artiodactyla retroposon (inverted Bov A element) located between nt 191 and nt 326 of intron 4 of bovine sequence (EMBL n° L19986). It is flanked by a 13 bp direct repeats (GGTAGCTGAGTCT), present as a single copy in the goat. Ramunno et~al. (2004) recently showed that the goat CSN1S1 gene is characterized by seven interspersed repeated elements vs. ten in the bovine one. Probably this extra retroposon element in the bovine Lf sequence is a rather young insertion and adds a further proof of the ancestral origin of the goat species as regards the bovine one, confirming that the retroposon insertions are a powerfull marker for phylogenetic studies of ruminants.

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REFERENCES - Baveye, S., Elass, E., Mazurier, J., Spik., G., Legrand, D., 1999. Lactoferrin: a multifunctional glycoprotein involved in the modulation of the inflammatory process. Clin. Chem. Lab. Med. 37:281-286. Brock, J.H., 2002. The physiology of lactoferrin. Biochem. Cell Biol. 80:1-6. Groenen, M.A.M., Dijkhof, R.J.A., Verstege, A.J.M., Van der Poel, J.J., 1993. The complete sequence of the gene encoding bovine αs2-casein. Gene 123:187-193. Iver, S., Lonnerdal, B., 1993.Lactoferrin, lactoferrin receptors and iron metabolism. Eur. J. Clin. Nutr. 47:232-241. Lee, T.H., Shimazaki, K., Yu, S.L., Nam, M.S., Lee, K.K., Yu, D.Y., 1997. Polymorphic sequence of Korean Native goat lactoferrin exhibiting greater antibacterial activity. Anim. Genet. 28:367-369. Le Provost, F., Nocart, M., Guerin, G., Martin, P., 1994. Characterization of the goat lactoferrin cDNA: assignment of the relevant locus to bovine U12 synteny group. Biochem. Biophys. Res. Commun. 203:1324-1332. Levay, P.F., Viljoen, M., 1995. Lactoferrin: a general review. Haematologica 80:252-267. Ramunno, L., Cosenza, G., Rando, A., Illario, R., Gallo, D., Di Berardino, D., Masina, P., 2004. The goat as1-casein gene:gene structure and promoter analysis, Gene, 9; 334:105-11. Schwerin, M., Solinas Toldo, S., Eggen, A., Brunner, R., Seyfert, H.M., Fries, R., 1994. The bovine lactoferrin gene (LTF) maps to chromosome 22 and syntenic group U12. Mamm. Genome 5:486-489. Seyfert, H.M., Tuckoricz, A., Interthal, H, Koczan, D., Hobom, G., 1994. Structure of the bovine lactoferrin-encoding gene and its promoter. Gene 143(2):265-269. Ward, P.P., Uribe-Luna, S., Conneely, O.M., 2002. Lactoferrin and host defense. Biochem. Cell. Biol. 80:95-102.