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Seep deposits from northern Istria, Croatia: a first glimpse into the Eocene seep fauna of the Tethys region

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Abstract – Three isolated limestone deposits and their fauna are described from a middle Eocene Flysch succession in northwestern Istria, Croatia. The limestones are identified as ancient methane-seep deposits based on fabrics and characteristic mineral phases, δ¹³C_{carbonate} values as low as −42.2‰, and ¹³C-depleted lipid biomarkers indicative for methane-oxidising archaea. The faint bedding of the largest seep deposit, the great dominance of authigenic micrite over early diagenetic fibrous cement, as well as biomarker patterns indicate that seepage was diffusive rather than advective. Apart from methanotrophic archaea, aerobic methanotrophic bacteria were present at the Eocene seeps as revealed by ¹³C-depleted lanostanes and hopanoids. The observed corrosion surfaces in the limestones probably reflect
carbonate dissolution caused by aerobic methanotrophy. The macrofauna consists mainly of chemosymbiotic bivalves such as solemyids (*Acharax*), thyasirids (*Thyasira*), and lucinids (*Amanocina*). The middle Eocene marks the rise of the modern seep fauna, but so far the fossil record of seeps of this age is restricted to the North Pacific region. The taxa found at Buje originated during the Cretaceous, whereas taxa typical of the modern seep fauna such as bathymodiolin mussels and vesicomyid clams are absent. Although this is only a first palaeontological glimpse into the biogeography during the rise of the modern seep fauna, it agrees with biogeographic investigations based on the modern vent fauna indicating that the dominant taxa of the modern seep fauna first appeared in the Pacific Ocean.

1. Introduction

Authigenic carbonate rocks forming where methane or oil effuse from the sediments into the bottom waters act as an archive of life in chemosynthesis-based ecosystems at marine seeps (Peckmann & Thiel, 2004; Campbell 2006). The key biogeochemical process at seeps is the anaerobic oxidation of methane (Boetius *et al.* 2000). It results in carbonate precipitation forming seep limestones even way below the carbonate compensation depth (e.g. Ritger *et al.* 1997; Greinert, Bohrmann & Elvert, 2002) and the production of hydrogen sulphide that sustains benthic sulphide-oxidizing bacteria and thiotrophic bacteria in the tissues of chemosymbiotic metazoans (Sibuet & Olu, 1998). A growing number of Phanerozoic seep deposits has been described to date (Campbell, 2006; Teichert & van de Schootbrugge, 2013, and references therein). Their fossil inventory revealed a successive colonisation of seep environments by different groups of metazoans in the course of Earth history, commonly followed by the sooner or later disappearance of these groups of highly specialized taxa. Methane-seep faunas were first discovered in the early 1980s in the Gulf of Mexico and are now recognized at most continental margins (Paull *et al.* 1984; Baker *et al.* 2010). Their highly specialized taxa are closely related to those at deep-sea hydrothermal vents and many
rely on chemotrophic symbionts for nutrition (Paull et al. 1985). Although the rise of the modern, mollusc-dominated vent and seep fauna began during the Cretaceous age, the main players at present-day vents and seeps appeared in the early Cenozoic (Campbell & Bottjer, 1995; Kiel, 2010; Kiel & Little, 2006; Vrijenhoek, 2013). Biogeographically, however, the Cenozoic fossil record of methane seeps is highly skewed toward the active continental margins of the Pacific Ocean where uplift of deep-water sediments is frequent (Goedert & Squires, 1990; Majima, Nobuhara & Kitazaki 2005; Campbell et al. 2008). In contrast, fossil occurrences in the Atlantic realm are restricted to the Caribbean region (Gill et al. 2005; Kiel & Peckmann, 2007) and the Mediterranean basin (Taviani, 1994).

Here we evaluate the fauna of middle Eocene seep deposits from the northern Mediterranean basin (Istria, Croatia; Venturini et al. 1998) in the light of the early evolution of the modern vent and seep fauna, establish the biogeochemical processes that lead to the formation of the seep deposits, describe processes that imprinted their lithology, and reconstruct the composition of fluids and the mode of seepage.

2. Geological setting and material

The Istria peninsula, shared by Croatia, Slovenia, and Italy, is bordered to the northeastern Adriatic Sea. During the Eocene, Istria was a part of the Dinaric foreland zone that experienced a strong subsidence in response to the formation of an orogenic wedge (e.g. Živkovic & Babić, 2003). The study area (Fig. 1a, b), located in the Croatian part of northwestern Istria, is characterised by a regional WNW-ESE-oriented anticlinal structure, commonly referred to as the Buje anticline or Buje Karst, whose origin is related to the formation of the Dinarides (Matičec, 1994). At the southern margin of the Buje anticline the foreland sequence is composed by more than 150 m of Lutetian lacustrine to shallow-marine foraminiferal limestones (Drobne & Pavlovec, 1991) and by at least 350 m of Lutetian to Priabonian turbidite deposits (referred to as Flysch Units; Marinčić et al. 1996; Pavšič &
Peckmann, 1996; Živkovic & Babić, 2003) that transgressively overlie an Aptian to Cenomanian sequence of shallow marine carbonates (Venturini et al. 1998). The Flysch deposits, in which the studied limestones are enclosed, consist of interbedded siliciclastic sandstones and marlstones as well as rare carbonate megabeds with basal breccias, representing calciturbidites (Venturini et al. 1998). The occurrence of turbidites indicates deposition by gravity flows in a deep-sea environment. The majority of the fine-grained marlstones, on the other hand, represents hemipelagic background sedimentation in a basinal setting (Pavšič & Peckmann, 1996). The occurrence of ichnogenera including *Paleodictyon*, as well as foraminifers and ostracods suggests deposition between 700 and 1200 m water depth (Gohrbandt et al. 1960; Pavšič & Peckmann, 1996).

The exotic blocks of limestone occurring in the vicinity of the town of Buje (Fig. 1b; 45°24′31″N, 13°40′01″E) have first been described by Venturini et al. (1998). The deposits studied here correspond to the “nearby Buje petrol station” section of Venturini et al. (1998; their Figures 4 and 5). In the captions of their Figures 10, 11, 13, and 14 as well as Table 1 Venturini et al. (1998) refer to this locality as “Buje”. The other two outcrops described by Venturini et al. (1998) were no longer accessible during field work in 2011. In the “nearby Buje petrol station” outcrop three limestone bodies are exposed in a road section in the eastern outskirts of Buje (Fig. 2, 3). These deposits are enclosed in a sequence of fine-grained marls intercalated with few decimetre-thick sandstone beds. The lowermost deposit (Buje 1) is about 4 m thick and laterally extends for approximately 20 metres in outcrop, the Buje 2 and 3 deposits are approximately 5 m and 2 m in width and 2 m and 1 m in height, respectively.

### 3. Methods

Sampling of the carbonate deposits (Buje 1, 2, and 3) has been carried out in spring 2011. Selected samples were prepared for palaeontologic, petrographic, and geochemical investigations. All fossil specimens are deposited in the Geowissenschaftliches Museum.
Georg-August-University Göttingen, Germany (GZG). Thin sections (15 x 10 cm and 10 x 7.5 cm) were studied with transmitted light and cathodoluminescence microscopy using a CITL 8200MK3, operating at about 17 kV and 400 mA. Thin sections were further analysed for their UV-fluorescence on a Nikon microscope with a UV-2A filter block, using ultraviolet light (illumination source 450-490 nm). Scanning electron microscopy and qualitative element recognition were performed with a Cambridge Instruments Stereoscan 360 scanning electron microscope equipped with an energy-dispersive Link System Oxford Instruments microprobe.

For stable isotope analyses mineral phases were drilled from the surface of slabs with a hand-held micro drill. Measurements of carbon and oxygen isotopes were performed with a Finnigan MAT 251 mass spectrometer using the “Kiel” carbonate device type “Bremen” against natural carbon dioxide from Burgbohl (Rheinland, Germany). A Solnhofen limestone was used as standard, which was calibrated against the international standard NBS 19. Values are reported in the δ-notation relative to Vienna Pee Dee Belemnite (VPDB) standard. Long time standard deviation (1σ) for this measurement was 0.05‰ for δ^{13}C and 0.07‰ for δ^{18}O values.

Lipid biomarkers were extracted from two carbonate blocks (Buje 1 and 2 deposits), yielding almost identical patterns. Samples were prepared and decalcified as described in Birgel et al. (2006a). After saponification with 6% KOH in methanol, the samples were extracted with a microwave extraction system (CEM Discovery) at 80°C and up to 250 W with dichloromethane/methanol (3:1) three times. The resulting extracts were separated into four fractions by column chromatography (500 mg DSC-NH$_2$ cartridges, Supelco) as described in Birgel et al. (2008). Carboxylic acids were measured as their methyl ester (ME) derivatives. All fractions were measured using an Agilent 7890 A GC system coupled to an Agilent 5975 C inert MSD spectrometer. The GC-MS system was equipped with a 30 m HP-5 MS UI fused silica capillary column (0.25 mm i.d., 0.25 μm film thickness). The carrier gas was He. The gas chromatography (GC) temperature program used for both fractions was as
follows: 60 °C (1 min); from 60 to 150°C at 10°C/min then to 320°C at 4°C/min; 25 min isothermal. Identification of compounds was based on GC retention times and comparison with published mass spectra. No separation of crocetane and phytane was achieved with the used column. The relative abundance of these compounds was assessed by the different fragmentation patterns, especially by the change of relative abundances of the masses 169 (characteristic for crocetane) and 183 (characteristic for phytane) within the mixed crocetane/phytane peak. Compound-specific carbon isotope analyses were carried out with a Thermo Fisher Trace GC Ultra connected via a thermo Fisher GC Isolink interface to a Thermo Fisher Delta V Advantage spectrometer. GC conditions were identical to those described above. Carbon isotopes are expressed as δ^{13}C values relative to the VPDB standard. The carbon isotope measurements were corrected for the addition of ME-derivatives. Several pulses of carbon dioxide with known δ^{13}C values at the beginning and the end of the runs were used for calibration. Instrument precision was checked using a mixture of n-alkanes (C_{14} to C_{40}) with known isotopic composition. The analytical standard deviation was <0.7‰.

4. Results

4.a. Fauna

Microfossils are abundant in the studied carbonate rocks, for the most part being represented by benthic (Bolivina sp., Stilostomella spp., Uvigerina spp., and Heterolepa spp.) and planktic (Turborotalia sp., Acarinina sp. and Hantkenina sp.) foraminifera. The occurrence of Hantkenina sp. agrees with an Upper Lutetian-Bartonian age (cf. Pavšič & Peckmann, 1996). Macrofossils were found only sporadically in the Buje 1 deposit and were almost absent in the Buje 2 and Buje 3 deposits. Most common is a lucinid bivalve that includes also the largest shell, followed by a thyasirid, and a solemyid bivalve. In addition to these bivalves, a few callianassid claws and other crustacean fragments were found. The bivalves include: (1) two specimens of a solemyid, the larger one 32 mm long and 10 mm high with the anterior
end missing. It shows an elongate S-shaped band extending from the posteroventral corner of
the anterior adductor muscle scar to the dorsal shell margin, had an external ligament, and is
therefore referred to as *Acharax* (Fig. 4a-c). (2) Two specimens of a *Nucula*; the larger one is
20 mm long and 15 mm high, and although the taxodont hinge is missing in these specimens,
they have the general shape of a *Nucula* and show the radial striation and crenulate ventral
margin common to this genus (Fig. 4d). (3) Four specimens belong to *Thyasira* due to their
general shape and strong posterior sulcus (Fig. 4e); the largest is 40 mm long. The
“undetermined Veneroida (?Kelliidae)” figured by Venturini *et al.* (1998, p. 225, Fig. 11) may
also belong to this *Thyasira* species. (4) Seven specimens and fragments of an oval lucinid
bivalve with an edentulous, narrow hinge without triangular excavation below the umbo, and
a maximum length of 52 mm (Fig. 4f-j) belong to the genus *Amanocina*. The lucinid is most
likely the same species as the “*?Lucina*” figured by Venturini *et al.* (1998, p. 225, Fig. 10).

4.b. Petrography and stable isotopes

The lithology of the three Buje carbonate deposits (Buje 1 to 3) is quite similar. The
limestones consist of fossiliferous and bioturbated mudstone and wackestone (Fig. 5). The
matrix is made up of dark brown micrite, revealing a bright autofluorescence (Fig. 6a, b).
Terrigenous particles are angular, including abundant quartz and rare feldspar grains as well
as lithic clasts. Apart from detrital grains, the micritic matrix contains abundant biogenic
detritus, mostly tests of foraminifera (Fig. 6c). Some mm to cm wide, irregular cavities occur;
the cavities are interpreted to result from bioturbation, representing successively filled
burrows. Some cavities show geopetal infill (Fig. 6d). The cavities are filled by sediment and
authigenic phases including peloids, homogenous micrite, laminated micrite, a phase referred
to as cauliflower micrite, and different generations of carbonate cements (Fig. 6d-f). Peloidal
fabrics are particular abundant (Fig. 6e). They consist of ovoidal peloids, showing an intense
fluorescence, surrounded by a non-fluorescent calcite microspar. On the basis of shape and
composition, peloids are interpreted to represent faecal pellets. Banding in the authigenic,

laminated micrites is sub-parallel to cavity walls (Fig. 6e). In places the laminated micrite is

broken to pieces, forming fragments surrounded by calcite cement.

The cauliflower micrite is an obviously authigenic variety of micrite found in some of the
cavities. It is represented by aggregates of mottled, microcrystalline calcite (Fig. 7a, b). Its
aggregates exhibit a domal, grooved shape, resembling cauliflower. Micron-sized irregular
pores, filled by calcite microspar, are present within these domes, generating a sponge-like
texture (Fig. 7c). The fluorescent cauliflower micrite (Fig. 7d) is commonly covered by a
circumgranular calcite cement (Fig. 7b, c). Remaining porosity in the cavities was

subsequently filled by two main generations of cement, (1) banded and botryoidal aggregates
of fibrous aragonite cement, mostly recrystallized to calcite, and (2) a drusy mosaic of equant
calcite cement (Fig. 6c-f). Carbonate cements are overall not abundant, being restricted to the
cavities believed to result from bioturbation.

The micritic matrix of the Buje deposits records episodes of carbonate corrosion. The

surfaces of the affected aggregates of micrite are highly irregular, and commonly covered by a
black rim of an opaque mineral up to a few tens of μm in thickness (Fig. 8a, b). Backscatter
and EDS observations revealed that these rims consist of scattered bright grains (Fig. 8c)
characterized by high contents of iron and manganese.

The volumetrically dominant micrite of the Buje carbonates has been analysed for its

stable carbon and oxygen isotope composition; the amount of banded and botryoidal cement
was not sufficient to allow for isotope analysis. The $\delta^{13}C$ values of micrite range from $-42.2$
to $-22.7\%$, the corresponding $\delta^{18}O$ values range from $-3.9$ to $0.0\%$ (Fig. 9). The Buje 1

deposit revealed the most negative $\delta^{13}C$ and $\delta^{18}O$ values, as low as $-42.2$ and $-3.9\%$,
respectively, with most $\delta^{13}C$ values falling between $-35.2$ and $-30.2\%$. Buje 2 and Buje 3
deposits show overall similar isotope values with less $^{13}C$ and $^{18}O$ depletion compared to the
Buje 1 deposit.
4.c. Biomarkers

Hydrocarbons, carboxylic acids, and alcohols were analysed. However, lipid biomarkers in
the alcohol fraction are only poorly preserved, and are thus not useful for the interpretation of
the depositional environment. The major group of compounds in the hydrocarbon fraction are
isoprenoid hydrocarbons (Fig. 10a). Among them are the head-to-tail linked isoprenoid
phytane (approximately 60% of the combined peak) and the tail-to-tail linked isoprenoid
crocetane (approximately 40%); their combined peak is the highest peak in this fraction. The
next abundant isoprenoids are the tail-to-tail linked isoprenoid pentamethylicosane (PMI) and
the head-to-head linked isoprenoid biphytane (bp-0). Other, minor constituents are
monocyclic biphytane (bp-1) with one cyclopentane ring and the tail-to-tail linked isoprenoid
squalane, as well as the head-to-tail linked isoprenoid pristane. Other than isoprenoids, few
straight-chain n-alkanes are present. Their overall distribution is patchy with the exception of
n-C\textsubscript{23}, resembling the inventory of modern and ancient, non-oil stained seep carbonates and
sediments (e.g. Thiel et al. 2001; Peckmann et al. 2007; Chevalier et al. 2013). Apart from
aliphatic lipid biomarkers, few cyclic compounds, mainly steranes and one hopanoid were
found. Among steroids, most abundant are C\textsubscript{28} and C\textsubscript{29} steranes. Other detected steroids are
lanostanes, which have been described in some seep carbonates (Birgel & Peckmann, 2008).
The most abundant cyclic terpenoid found is the hopanoid hop-17(21)-ene.

The isoprenoids have the most negative $\delta^{13}$C values with $-111\%$ and $-109\%$ for PMI
and bp-0, respectively. The head-to-tail linked isoprenoid pristane ($-60\%$) and the n-alkane
n-C\textsubscript{23} ($-66\%$) revealed intermediate values (Fig. 9), whereas other short-chain n-alkanes are
significantly less $^{13}$C-depleted ($-34\%$). The $\delta^{13}$C values of steranes fall in the same range as
short-chain and long-chain n-alkanes. Lanostanes are more $^{13}$C-depleted with an average
value of $-47\%$. Hop-17(21)-ene is more $^{13}$C-depleted ($-64\%$) than the lanostanes.
The carboxylic acid fraction is predominated by n-fatty acids ranging from C_{14} to C_{28} (Fig. 10b). The fatty acids are characterized by an overall even-over-odd predominance. Highest contents were found for short-chain n-C_{16} fatty acid. Other abundant compounds are n-C_{16} and C_{18} fatty acids with one double bond. Apart from n-fatty acids, terminally-branched fatty acids are abundant, especially those comprising 15 carbons. Other compounds in the carboxylic acid fraction are phytanoic acid and PMI acid. Phytanoic acid co-elutes with a C_{18:1} fatty acid. Only one hopanoic acid, 17β(H),21β(H)-bishomohopanoic acid, was identified.

The strongest $^{13}$C depletions in the carboxylic acids were found for the isoprenoid PMI acid ($-107\%$). Although combined with the isotopic signature of the co-eluting n-C_{18:1} fatty acid, phytanoic acid is still considerably $^{13}$C-depleted ($-75\%$). Other compounds with significant depletion in $^{13}$C are the terminally-branched iso- and anteiso-C_{15} fatty acids with $\delta^{13}$C values of $-68\%$ and $-82\%$, respectively, as well as 17β(H),21β(H)-bishomohopanoic acid ($-70\%$). Short-chain n-fatty acids yielded values of around $-50\%$, whereas the long-chain fatty acids revealed higher values (average $-31\%$).

5. Discussion

5.a. Biogeographic and evolutionary aspects

Methane seepage and associated faunal communities in the Mediterranean realm are known from the late Mesozoic when large lucinid bivalves and rhyynchonellide brachiopods inhabited cold seeps along the northern shore of the Tethys Ocean (Gaillard, Rio & Rolin, 1992; Campbell & Bottjer, 1995; Peckmann et al. 1999; Kiel, 2013) and from the Miocene onward, largely along the Apennine chain in Italy (Ricci Lucchi & Vai, 1994; Taviani, 2011). These Neogene seep deposits are generally referred to as ‘Calcari a Lucina’ (Clari et al. 1988; Taviani, 1994). Among them, the Miocene deposits contain essentially a modern seep fauna
consisting of large bathymodiolin, vesicomyid, and lucinid bivalves, while the few Pliocene
examples appear to have a reduced character of the modern Mediterranean Sea seep fauna
(Table 1; Taviani, 2014). Many of the taxa that inhabit vents and seeps today originated in the
early Cenozoic (Kiel & Little, 2006; Amano & Kiel, 2007; Kiel & Amano, 2013; Vrijenhoek,
2013). The middle Eocene Buje deposits can thus provide insights into the early evolution of
the seep fauna and its biogeography.

The only seep deposits coeval with the Buje seeps are those of the middle Eocene
Humptulips Formation in western Washington State, USA, and thus from the Pacific realm
(Goedert & Squires, 1990). They share the common solemyids, the large thyasirids, and the
edentulous lucinids, although the latter are represented by different genera in the two regions
seep deposits differ, however, by the presence of large, high spired gastropods (Goedert &
Kaler, 1996; Kiel, 2008) and vesicomyid bivalves (Squires & Goedert, 1991; Amano & Kiel,
2007), which appear to be absent from the Buje deposits. The Humptulips limestones also
include the earliest bathymodiolin mussels discovered so far (Kiel & Amano, 2013). From
one of the seep deposits at Buje, Venturini *et al.* (1998) reported several specimens of the
mytilid ‘*Modiolus*’ that could potentially represent an as-yet unidentified bathymodiolin
mussel; unfortunately that particular deposit was no longer accessible during our field work
and the identity of this mussel remains elusive. The fauna of the Buje seep deposits is only a
first glimpse into the Eocene seep fauna of the central Tethys Ocean and is unlikely to
represent the full diversity of the regional pool of seep-inhabiting taxa. However, if taken at
face value, the absence of the main modern taxa (bathymodiolins and vesicomyids) from Buje
at a time when these taxa were present at Pacific seeps is in agreement with molecular
phylogenetic analyses (*Lorion et al.* 2013; *Roterman et al.* 2013; *Stiller et al.* 2013) and
quantitative biogeographic analyses (*Bachraty et al.* 2009; *Moalic et al.* 2012), which indicate
a Pacific origin of the modern vent and seep fauna.
Compared to the ‘Calcari a Lucina’ seep deposits in the Italian Miocene (Fig.1a; Clari et al. 1994; Taviani, 1994) and the modern Mediterranean seep fauna (Olu-Le Roy et al. 2004; Ritt et al. 2010; Taviani et al. 2013), the middle Eocene seep fauna at Buje shows clear differences (Table 1). Solemyids are rare in the Neogene to modern seeps in the Mediterranean Sea (Taviani et al. 2011; Rodrigues, Duperron & Gaudron 2011) in contrast to Buje, where they are common. Also the large Thyasira is a distinctive feature of the Buje seeps, while thyasirids are absent from the ‘Calcari a Lucina’ deposits (Taviani, 2011; S. Kiel, own observation), and in the modern Mediterranean seep fauna they are represented only by a small (~10 mm) species (Olu-Le Roy et al. 2004). The lucinids at the Miocene to modern Mediterranean seeps clearly belong to different genera than the lucinid at Buje (Olu-Le Roy et al. 2004; Taviani, 2011; Kiel & Taviani, unpub. data), which belongs to the widespread Early Cretaceous to Oligocene genus Amanocina.

5.b. Microbial activity steering carbonate formation and destruction

The Buje carbonate deposits show several petrographical and geochemical lines of evidence that agree with a microbial origin sustained by hydrocarbon seepage. Not only the negative δ13C values as low as −42‰ agree with methane seeping (cf. Paul et al. 1992; Peckmann & Thiel, 2004), but also microfabrics, such as peloidal and clotted micrite, laminated micrite, and banded and botryoidal cement filling cavities are typical of seep carbonates (e.g. Peckmann & Thiel, 2004). Finally, lipid biomarkers characteristic for methane seepage are found in the Buje deposits, confirming their microbial origin resulting from methane oxidation. Among the observed compounds, the most 13C-depleted acyclic isoprenoids such as mixed phytane/crocetane (−98‰), PMI (−111‰), and acyclic biphytane (−109‰) are molecular fossils of methanotrophic archaea (e.g. Elvert, Suess & Whiticar, 1999; Peckmann & Thiel, 2004; Birgel et al. 2006a; Peckmann, Birgel & Kiel, 2009). These biomarkers are accompanied by molecular fossils of sulphate-reducing bacteria, such as iso- and anteiso-C15.
fatty acids (Elvert et al. 2003; Birgel et al. 2006b). As commonly observed in seep deposits, the lipids of the sulphate-reducing bacteria involved in anaerobic oxidation of methane are less $^{13}$C-depleted ($-82\%$ for anteiso-$C_{15}$ FA) than the lipids of methanotrophic archaea (e.g. Peckmann & Thiel, 2004).

At first glance, the petrographical characteristics and stable isotope and lipid biomarker patterns of the Buje deposits are not much different from other ancient Mediterranean seep deposits (e.g. Peckmann et al. 2004; Clari et al. 2009; Natalicchio et al. 2013). However, the Buje seep deposits show some peculiarities, as for example the occurrence of cauliflower micrite. These dome-shaped precipitates are made up of fluorescent clotted micrite and formed in-situ within cavities, properties that typify the products of organomineralisation (cf. Reitner et al. 1995; Dupraz et al. 2009). Two possible modes of formation are envisaged, (1) mineralised microbial mats or (2) sponges. (1) Mineralized biofilms have already been documented in Eocene seep deposits from western Washington State (Peckmann et al. 2003) and in Miocene seep deposits from the Italian Apennine (Peckmann et al. 1999). The cauliflower shape, representing a domal, accretionary mode of growth on a mm to cm scale in a cryptic environment is different from previous reports of much thinner mineralised biofilms within cracks of preexisting seep carbonate. Based on the larger size of the Buje cauliflower micrite and its domal growth habit along with its intense autofluorescence it seems feasible that this micrite resulted from the mineralisation of microbial mats that performed anaerobic oxidation of methane. The validity of this scenario is enforced by the presence of subsurface microbial mats of anaerobic oxidation of methane-performing prokaryotes at active seeps in the Black Sea (Treude et al. 2005). (2) Alternatively, the domal growth, clotted microfabric, and reticulate porosity of the cauliflower micrite resembles the outcome of sponge taphonomy (e.g. Delecate et al. 2001). Because no spicules have been observed, it is unlikely that cauliflower micrite represents fossils of spicular sponges. Even in case of siliceous spicules, the spicules would have been probably preserved in the authigenic seep carbonate. Where
sponges have been reported in ancient seep deposits, their overall preservation including spicules was good in case of Mesozoic examples (Peckmann et al. 1999) and excellent in case of Cenozoic examples (Goedert & Squires, 1990; Rigby & Goedert, 1996). If the sponge interpretation is correct, the sponges were probably non-spicular, belonging to a group informally referred to as keratose demosponges (J. Reitner, pers. comm.). Despite of lacking spicules, the taphonomy of keratose sponges results in micritic carbonate fabrics that can still be recognized in Phanerozoic rocks (Luo & Reitner, 2014). Seep-dwelling sponges have been reported from a number of modern sites (Olu-Le Roy et al. 2004, and references therein). Some demosponges have even been shown to contain endosymbiotic methanotrophic bacteria (Vacelet et al. 1996; Olu-Le Roy et al. 2004; Baco et al. 2010).

The abundant irregular corrosion surfaces partially covered by iron and manganese precipitates indicate dissolution of carbonate. Such dissolution features coupled with iron and manganese enrichment have commonly been interpreted as the product of microbially-driven corrosion, as for example reported for reef carbonates (Reitner et al. 2000; Tribollet et al. 2011). Analogous features have also been observed in ancient (Campbell et al. 2002; Peckmann et al. 2003; Birgel et al. 2006b) and modern (Matsumoto, 1990; Himmler et al. 2011) seep carbonates and were interpreted as biologically-induced corrosion features as well. Matsumoto (1990) was the first to suggest that carbonate corrosion at seeps is driven by bacterial aerobic methane oxidation and sulphide oxidation. Both processes have the potential to lower the pH and may thus promote carbonate dissolution (Himmler et al. 2011; Tribollet et al. 2011). Molecular fossils of sulphide-oxidizing bacteria cannot be easily identified in ancient rocks, since these lipids are of low specificity and prone to degradation (cf. Arning et al. 2008). In contrast, the former presence of aerobic methanotrophs at seeps can be constrained by lipid biomarkers including lanostanes and some hopanoids (Peckmann et al. 1999; 2004; Birgel & Peckmann, 2008; Sandy et al. 2012). The low $\delta^{13}C$ values of lanostanes and hopanoids in the Buje limestones agree with aerobic methanotrophs as source organisms,
although other sources cannot be excluded in case of the $^{13}$C-depleted hopanoids (cf. Blumenberg et al. 2006; Eickhoff et al. 2013). The potential of aerobic methanotrophs to cause carbonate dissolution has recently been proven in laboratory experiments (Krause et al. 2014). Based on the confirmation that this mechanism is indeed capable of inducing carbonate dissolution and the detection of molecular fossils of aerobic methanotrophs, carbonate corrosion archived in the Buje seep limestones is best explained by aerobic methanotrophy.

5.c. Constraints on fluid flow

The occurrence of both anaerobic oxidation of methane – as revealed by $^{13}$C-depleted biomarkers and $^{13}$C-depleted authigenic carbonates – and aerobic oxidation of methane – as revealed by $^{13}$C-depleted biomarkers and carbonate corrosion – indicates discontinuous oxygenation conditions in the subsurface close to the seafloor at the Buje seep sites. The precipitation of the $^{13}$C-depleted micrite driven by anaerobic oxidation of methane occurred in anoxic environments within the pore space of the detrital background sediment, leading to the occlusion of the sedimentary matrix. After the pore space was successively filled by micrite, carbonate precipitation was largely restricted to some cavities resulting from preceding bioturbation, and allowing for the formation of fibrous, banded and botryoidal aragonite cement and clotted micrite. Based on the evidence for carbonate corrosion and the preservation of diagnostic biomarkers, at least some of the aerobic methanotrophic bacteria most probably lived in oxic sediments, rendering unlikely that these biomarkers were exclusively sourced from bacteria dwelling in the water column above the seeps.

A set of observations indicates that the mode of seepage was diffusive rather than advective. The Buje seep limestones largely consist of authigenic micrite cementing background sediments. Such a pattern with the dominance of micrite over early diagenetic aragonite cements is typical for diffusive seepage (e.g. Peckmann, Birgel & Kiel, 2009; Haas et al. 2010). Similarly, the faint stratification apparent in the Buje 1 deposit is an additional
argument in favour of this interpretation. Similarly, the circumstance that biphytane occurs in
much higher contents than crocetane agrees with the dominance of archaea of the so-called
ANME-1 group (Blumenberg et al. 2004; Niemann & Elvert, 2008; Rossell et al. 2011),
another observation in favour of diffusive seepage (Nauhaus et al. 2005; Peckmann, Birgel &
Kiel, 2009). ANME-1 archaea, like ANME-2 archaea, are commonly associated with
sulphate-reducing bacteria of the Desulfosarcina/Desulfococcus branch of the
Deltaproteobacteria (Knittel & Boetius, 2009). The bacterial partners of the ANME-1 archaea
can be discerned from those of ANME-2 archaea by a much higher proportion of ai-C_{15} fatty
acid (Blumenberg et al. 2004; Niemann & Elvert, 2008), a compound that is particularly
abundant in the Buje limestones (see Fig. 10b). All these observations argue in favour of
diffusive seepage. It should, however, be kept in mind that other factors than just seepage
activity can influence the distribution of ANME-1 versus ANME-2 archaea and the
abundance of aerobic methanotrophs as well. An obvious factor for example is temperature,
whereby higher temperatures are known for favour ANME-1 over ANME-2 archaea
(Nauhaus et al. 2005).

It is interesting to note that some Cretaceous seep deposits for which diffusive seepage
has been envisaged contain biomarkers of aerobic methanotrophs as well (Peckmann, Birgel
& Kiel, 2009; Sandy et al. 2012), although the majority of seep deposits lacks these
compounds (e.g. Peckmann & Thiel, 2004). Because the sulphate-methane transition zone
(SMTZ) tends to be situated deeper within the sediments at sites of diffusive seepage than at
sites of advective seepage (e.g. Sahling et al. 2002; Luff & Wallmann, 2003), we suggest that
the preservation of lipids of aerobic methanotrophs is favoured in limestones forming at seeps
typified by diffusive seepage – this is not meant to say that aerobic methanotrophs are
necessarily more abundant at diffusive seeps. With aerobic methanotrophy being able to
extend to greater sediment depth at diffusive seeps, the likelihood probably increases that the
lipids of aerobic methanotrophs become engulfed in authigenic seep carbonates at a later stage
upon dilatation of the zone of anaerobic oxidation of methane. If seepage continues for
extended periods of time – as envisaged for the thick Buje 1 deposit – the prolonged
formation of methane-derived carbonates, thus, assures the preservation of process markers of
those biogeochemical processes that occurred in close proximity of the strata affected by
anaerobic oxidation of methane. This effect will be intensified upon variations of seepage
intensity that allow for vertical displacement of the SMTZ (cf. Feng, Chen & Peckmann,
2009). An upward movement of the SMTZ caused by an increase of seepage intensity and
accompanied by a shift of carbonate formation to shallower depth will particularly favour the
preservation of the lipids of aerobic methanotrophs.

6. Conclusions

The fossil record and molecular age estimates indicate that the dominant taxa of the modern
vent and seep fauna appeared during the Eocene. The fossil record of seep communities of
this age, however, is highly skewed toward the Pacific region and thus macrofauna of the
Buje seep deposits provides a first glimpse into the seep fauna of the Tethyan region. The
absence of the main modern taxa (bathymodiolin mussels and vesicomyid clams) from the
Buje seeps agrees with other lines of evidence suggesting that the modern vent and seep fauna
originated in the Pacific Ocean. The Buje seep fauna also indicates a dynamic evolution of
seep faunas in the Tethyan/Mediterranean basin: it resembles Cretaceous to early Palaeogene
seep faunas from other parts of the world, whereas the late Miocene ‘Calcari a Lucina’ fauna
in Italy resembles other Miocene to modern seep faunas worldwide, and the Pliocene seep
faunas from northern Italy have the somewhat restricted character of Mediterranean seep
fauna today that probably resulted from the extinction of the more ‘oceanic’ Miocene seep
faunas during the Messinian salinity crisis.

The Buje seep deposits formed as a consequence of anaerobic oxidation of methane as
revealed by the presence of $^{13}$C-depleted biomarkers of methanotrophic archaea and
associated sulphate-reducing bacteria. Apart from these anaerobic prokaryotes, aerobic
methanotrophic bacteria lived at the middle Eocene seeps. Their metabolism apparently led to
a local decrease of pore water pH values, which resulted in the dissolution of carbonate
minerals. The large size of the Buje 1 deposit suggests that seepage activity was long-lasting.

(1) Its faint stratification, (2) the dominance of authigenic micrite over early diagenetic
fibrous cement, (3) biomarker patterns of the prokaryotes performing anaerobic oxidation of
methane, and (4) possibly the preservation of the lipids of aerobic methanotrophs indicate that
seepage activity was mostly diffusive rather than advective.

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characterization of cold seep faunal communities on the New Zealand Hikurangi

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Figure and table captions:

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Figure 7. Petrography of cauliflower micrite. m – matrix micrite; pm – peloidal micrite; ccc – circumgranular calcite cement; cm – cauliflower micrite; ec – equant calcite cement. (a) Domal and grooved cauliflower micrite that grew on peloidal micrite and was postdated by circumgranular calcite and equant calcite cement, plane-polarized light. (b) Detail of (a). (c) Close up view of the cauliflower micrite with internal reticulate porosity filled by microspar (arrows); crossed-polarized light. (d) The cauliflower micrite exhibits an intense autofluorescence; fluorescence image.

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Table 1. Schematic overview of the Palaeogene and Neogene seep deposits across the Mediterranean

<table>
<thead>
<tr>
<th>Locality</th>
<th>Age</th>
<th>Type of seep</th>
<th>Fossil assemblage</th>
<th>δ¹³C [‰VPDB]</th>
<th>δ¹⁸O [‰VPDB]</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Emilian Apennine (Italy)</td>
<td>Early Pliocene</td>
<td>Fossiliferous limestones, conduits</td>
<td>Solemyids and lucinids</td>
<td>-25 to -17</td>
<td>-3 to +3</td>
<td>Taviani et al. 1997; Barbieri and Cavalazzi, 2005</td>
</tr>
<tr>
<td>Tortona Apennine (Piedmont, Italy)</td>
<td>Late Miocene</td>
<td>Lucina and brecciated limestones, carbonate beds with veins, conduits</td>
<td>Lucinids, tubeworms, bacterial biofilms</td>
<td>-56 to -6</td>
<td>-6 to +7</td>
<td>Dela Pierre et al. 2010; Martini et al. 2010; Natalicchio et al. 2012, 2013</td>
</tr>
<tr>
<td>Maiella, central Apennine (Italy)</td>
<td>Late Miocene</td>
<td>Brecciated limestones</td>
<td>absent</td>
<td>-40 to +4</td>
<td>-9 to +4</td>
<td>Iadanza et al. 2013</td>
</tr>
<tr>
<td>Monferrato (Piedmont, Italy)</td>
<td>Middle and Late Miocene</td>
<td>Lucina and brecciated limestones, macroconcretions with veins, conduits</td>
<td>Lucinids, tubeworms, bacterial biofilms</td>
<td>-45 to -9</td>
<td>-1 to 8</td>
<td>Clari et al. 1988, 1994, 2009; Peckmann et al. 1999</td>
</tr>
<tr>
<td>Sicily (Italy)</td>
<td>Middle and Late Miocene</td>
<td>Lucina and brecciated limestones</td>
<td>Lucinids (?)</td>
<td>-49 to -29</td>
<td>+3 to +9</td>
<td>Ricci Lucchi and Vai, 1994</td>
</tr>
<tr>
<td>Tuscan-Romagna Apennine (Italy)</td>
<td>Early and Late Miocene</td>
<td>Fossiliferous and brecciated limestones</td>
<td>Solemyids, lucinids, bathymodiolins, and vesicomyids</td>
<td>-58 to -16</td>
<td>-5 to +5</td>
<td>Taviani et al. 1997; Conti and Fontana, 1999, 2005; Lucente and Taviani, 2005</td>
</tr>
<tr>
<td>Buje (Croatia)</td>
<td>Middle Eocene</td>
<td>Fossiliferous limestones</td>
<td>Solemyids (Acharax), thyasirids (Thyasira), lucins (Amanocina), nuculids, Callianassa</td>
<td>-42 to -23</td>
<td>-4 to 0</td>
<td>Venturini et al. 1998; this study</td>
</tr>
</tbody>
</table>
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127x96mm (300 x 300 DPI)
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182x416mm (300 x 300 DPI)
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146x267mm (600 x 600 DPI)
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202x241mm (600 x 600 DPI)