

Novel functional microRNAs from virus-free and infected *Vitis vinifera* plants under water stress.

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Supplementary Methods

5'-RACE

The degradation fragments resulting from miRNA cleavage were analysed by 5'-RACE. The RNA from free and GRSPaV-infected 'Bosco' leaves was extracted using the Spectrum Plant Total RNA Kit (Sigma-Aldrich, St. Louis, MO, USA), and was treated with DNase I (Life Technologies, Carlsbad, CA, USA), incubated with a RNA adaptor (Table S1) and 5U of T4 RNA ligase (Life Technologies) following the manufacturer's instructions. The RNA was reverse-transcribed using random nonamers (Sigma-Aldrich) and M-MLV reverse transcriptase (Life Technologies). First-round and nested PCR analyses were performed using a universal forward primer and gene-specific reverse primers (Supplementary Table S2). The nested products were gel-purified, cloned into the pGEM-T Easy vector (Promega, Madison, WI, USA), and ten independent clones from each product were sequenced using a Big-Dye Terminator v1.1 Cycle Sequencing kit (Applied Biosystems, Life Technologies), and analysed using a 3130 Genetic Analyser capillary sequencer (Applied Biosystems, Life Technologies).

qRT-PCR analysis

The quantification of miRNA expression by qRT-PCR was carried out following the protocol of Shi and Chiang (2005) with some modifications. Two µg of RNA extracted from GRSPaV-free or infected leaves collected under WW, WS and SWS conditions were treated with DNaseI (Life Technologies) and polyadenylated using the Poly(A) Tailing Kit (Life Technologies) following the manufacturer's instructions. The RNA was precipitated by ethanol and reverse-transcribed with M-MLV reverse transcriptase (Life Technologies) and 0.5 µg poly(T) adaptor (Supplementary Table S2).

For the amplification of all miRNAs and housekeeping genes, a universal 3'-adaptor reverse primer was used (Supplementary Table S2) and the forward primer was designed based on the specific miRNA sequence. For some primers containing high GC content at the 3' end, one or two A residues were added to the 3'-end to facilitate binding to the target site (Supplementary Table S2).

The relative expression was calculated based on the comparative C_t ($2^{-\Delta\Delta C_t}$) method as described by Livak and Schmittgen (2001). The PCR mix (10 μ L) contained 5 μ L PowerSYBR Green master mix (Applied Biosystems, Life Technologies), 0.25 μ M of each primer and 1 μ L of cDNA diluted 1:100 or 1:250, based on initial miRNA concentrations. Cycling conditions for all primer pairs consisted of initial denaturation at 95 °C for 10 min, followed by 45 cycles at 95 °C for 15 s, 58 °C for 20 s and 60 °C for 40 s. The 5.8S rRNA and U6, which have been extensively used with the grapevine and other species as reference in qRT-PCR and northern hybridisations for sRNA analysis, were used as housekeeping genes. The geometric mean of their expression ratios was used as the normalisation factor in all samples. Accumulation of miRNAs was expressed as the mean and standard deviation calculated for three biological replicate.

Ecophysiological measurements

Transpiration rates (E) and stomatal conductance (g_s) were measured by two approaches. Firstly, we estimated E and g_s via pot weight. Daily E was calculated as the difference between two consecutive weights, and the daily grams of water loss per plant were converted into mmol H₂O m⁻² s⁻¹. The g_s was then calculated by dividing E by the environmental vapour-pressure deficit on the basis of the air temperature and relative humidity that was recorded every day during the experiment (Figure S2A). Secondly, leaf gas-exchange parameters (P_n , E, g_s , c_i) were measured by the GFS-3000 portable gas-exchange fluorescence system (Walz Heinz GmbH, Effeltrich, Germany) on two different mature leaves per plant, on each experimental day (setting: cuvette temperature 27 °C, [CO₂], photosynthetic photon flux density (PPFD) and Rh were those of the greenhouse environmental conditions).

The Ψ_{soil} was calculated following the equation obtained by pressure-plate analysis (Richards and Ogata, 1961): $\Psi_{\text{soil}} = 3e^{-10.9\theta}$, where θ represents the soil volumetric water content, calculated as described by Hochberg *et al.* (2013). The bulk density (ρ) of the peat substrate used was 0.383 kg L⁻¹. The weight of the sealed pots was measured at the end of each day (6 p.m.) using an electronic balance.

The enumeration of stomata and cells was determined on two leaves per six plants for infected and GRSPaV-free plants. Two leaf imprints were collected in the WW condition and three counts for each imprint were performed using an optical microscope with a 10-XT objective (Nikon Eclipse 55i, Tokyo, Japan).

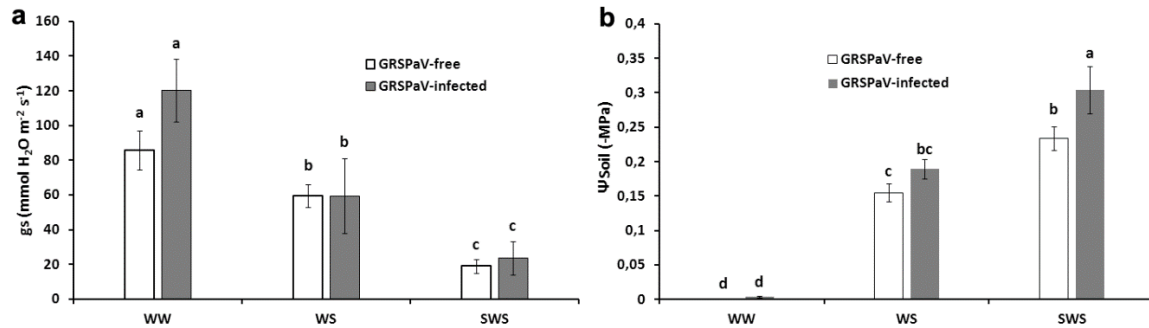
At the end of the experiments, dry weight of the root was quantified by eradicating, washing and drying the roots in oven at 70°C until reaching of constant root weigh. Roots of minor orders were separated manually from main roots after drying.

Supplementary References

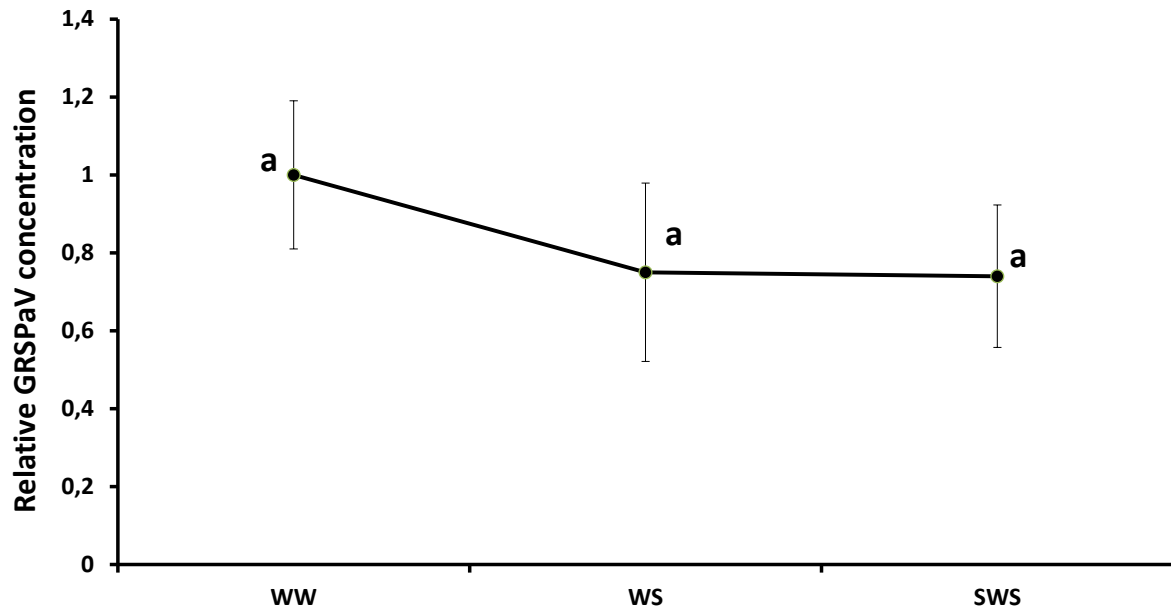
Livak, K.J. & Schmittgen, T.D. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* **25**, 402- 408 (2001).

Hochberg, U., Degu, A., Fait, A. & Rachmilevitch S. Near isohydric grapevine cultivar displays higher photosynthetic efficiency and photorespiration rates under drought stress as compared with near anisohydric grapevine cultivar. *Physiol. Plantarum* **147**, 443–452 (2013).

Supplementary Figures



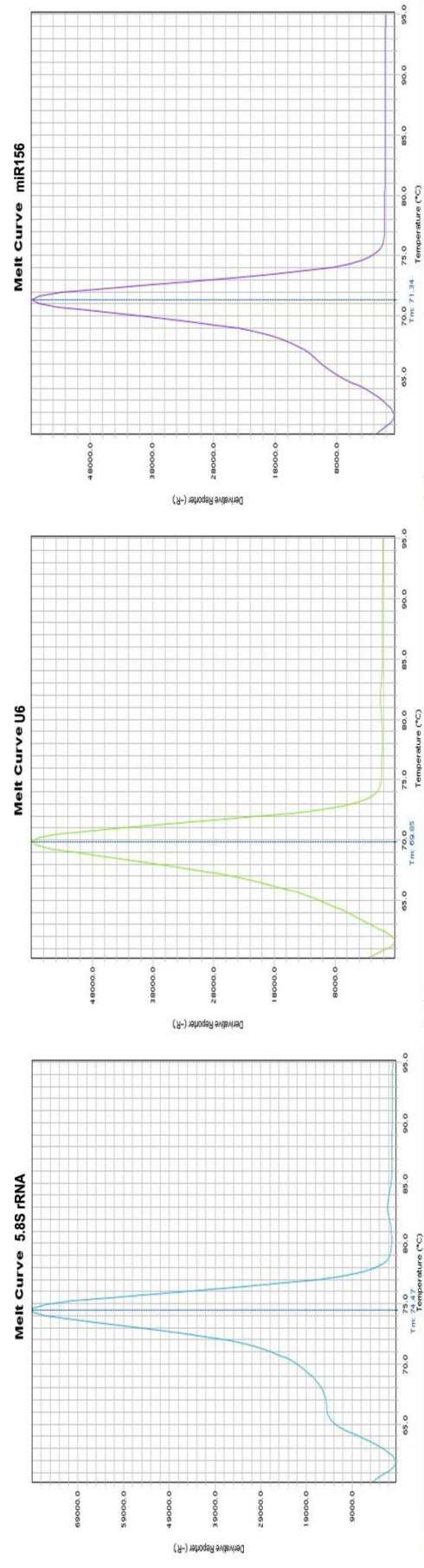
Supplementary Figure S1. a) stomatal conductance (g_s) measured and b) relative water stem potential (Ψ_{stem}) during the sampling time for small RNAs analysis. Bars represent the mean \pm standard error ($n=6$). Different letters indicate significant difference among the means ($p \leq 0.05$). WW, well watered condition; WS, water stress condition; SWS severe water stress condition.



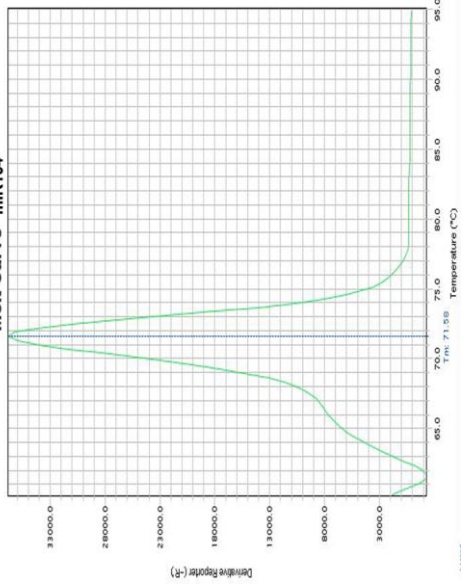
Supplementary Figure S2. Quantification of *Grapevine rupestris stem pitting-associated virus* (GRSPaV) RNA in ‘Bosco’ leaf as determined by qRT PCR. Samples were collected under well watered (WW), water stress (WS) and severe water stress (SWS) conditions. qRT-PCR signals were normalised to actin and ubiquitin levels. Data are presented as mean \pm standard deviation of six plants and three technical replicates; different letters denote significant differences at $p \leq 0.05$.

Supplementary Fig. S3: Validation of qRT-PCR for quantitation of microRNAs (miRNAs). a) dissociation curves of qRT-PCR amplification and b) qRT-PCR products resolved on ethidium bromide-stained 5% agarose gel of all miRNAs and novel miRNA candidates analysed.

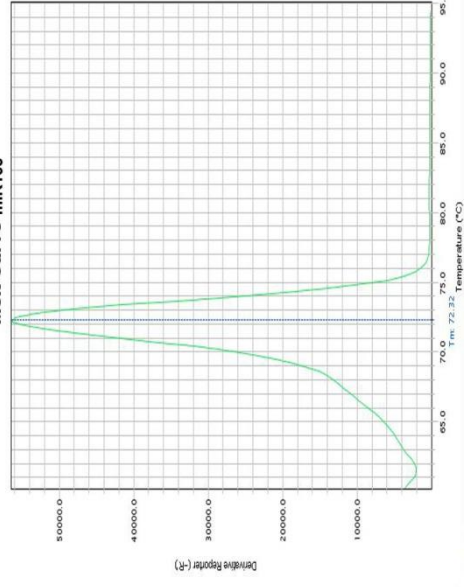
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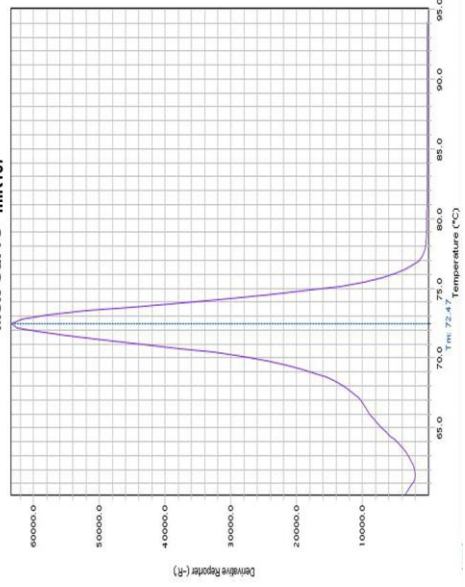
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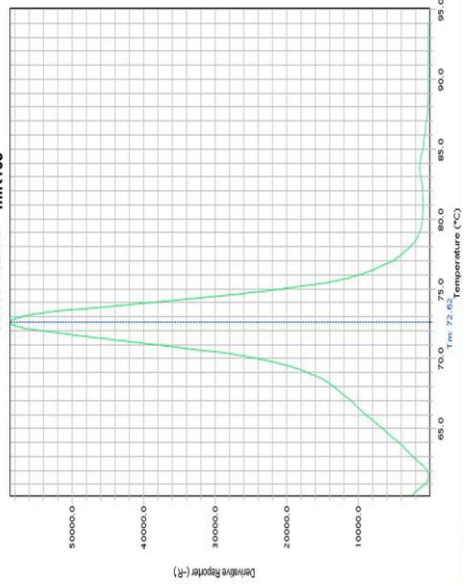
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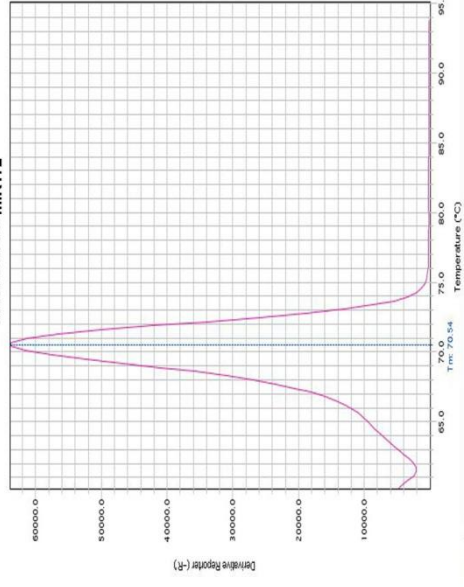
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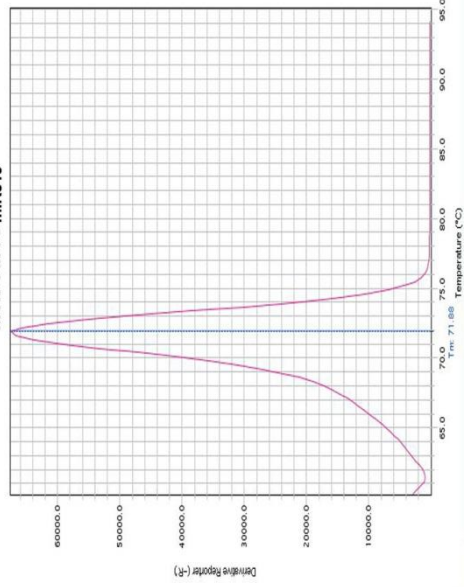
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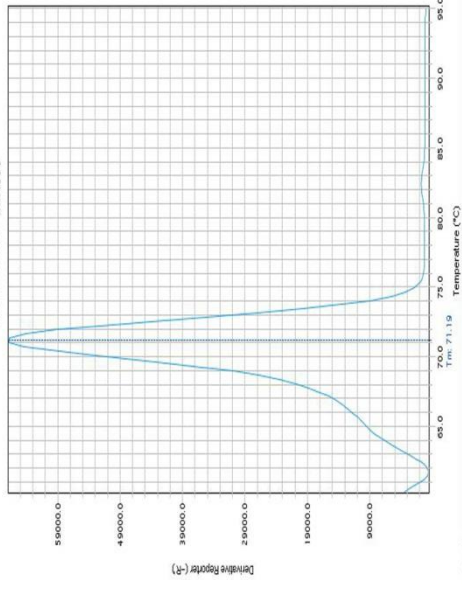
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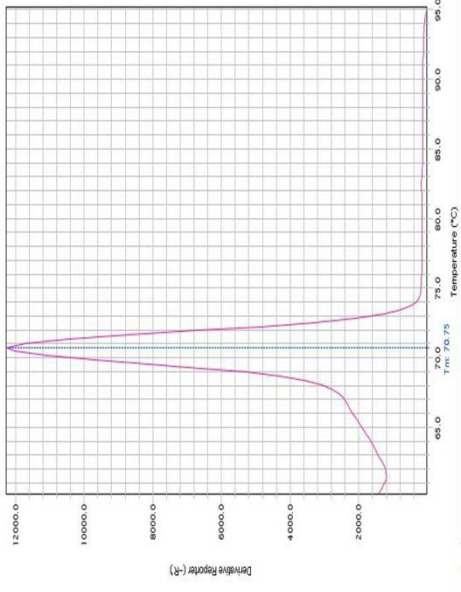
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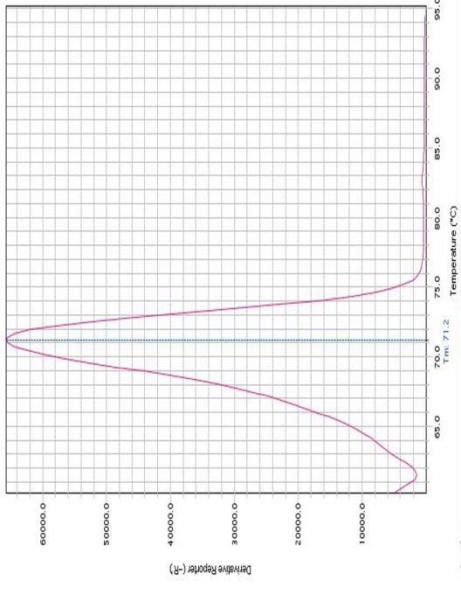
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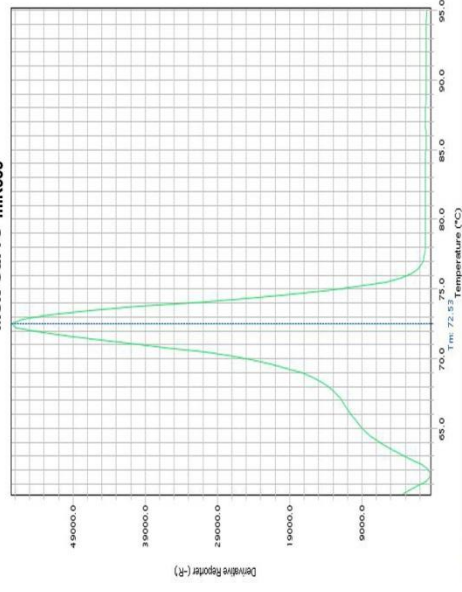
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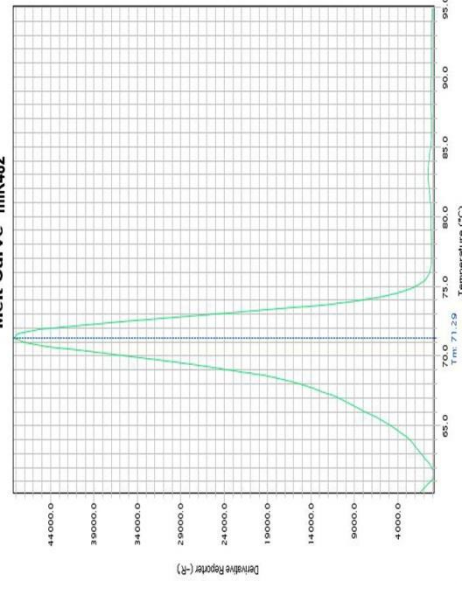
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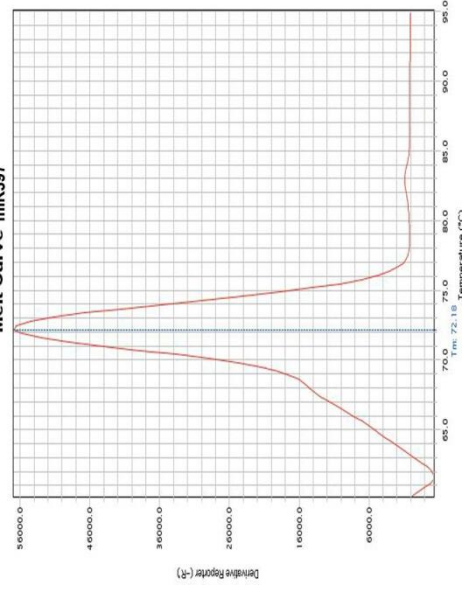
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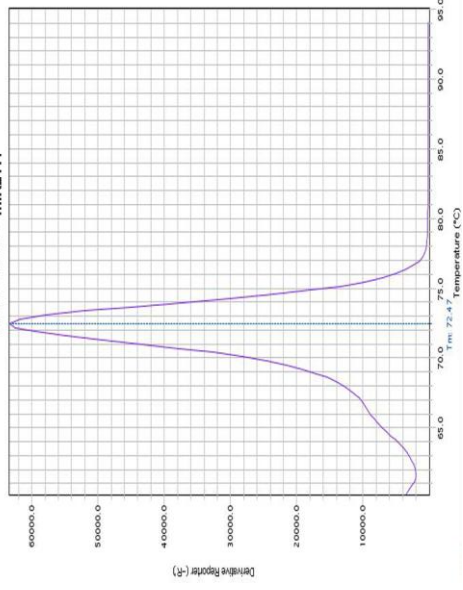
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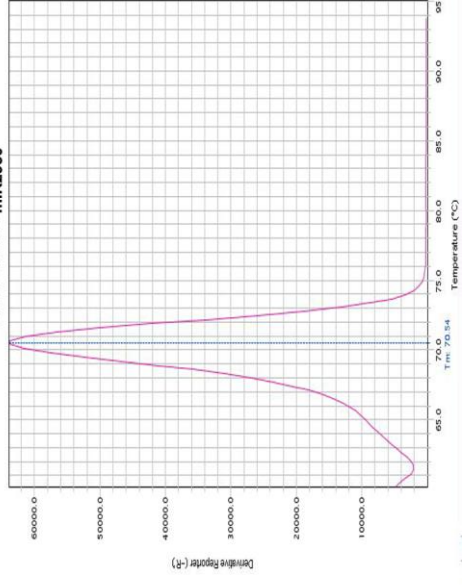
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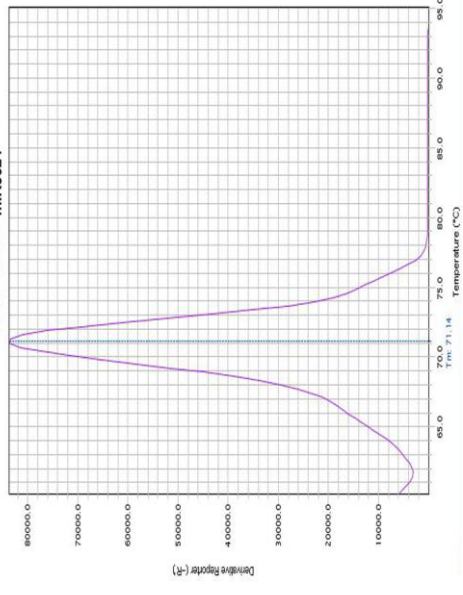
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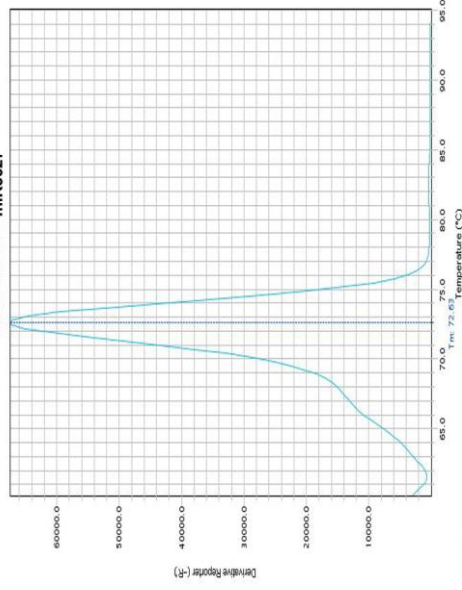
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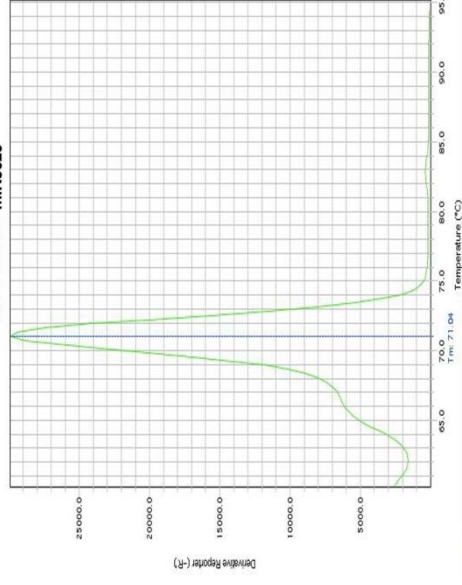
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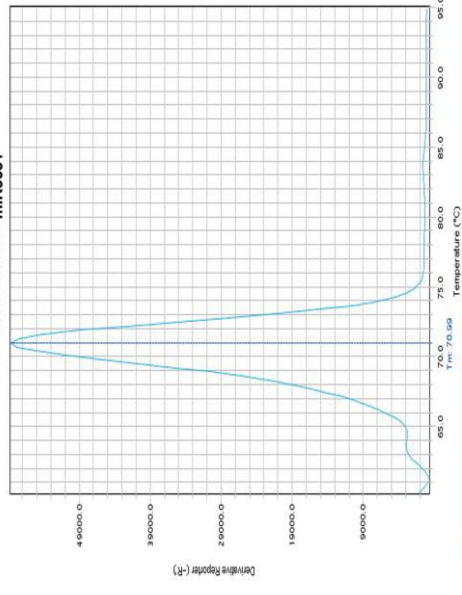
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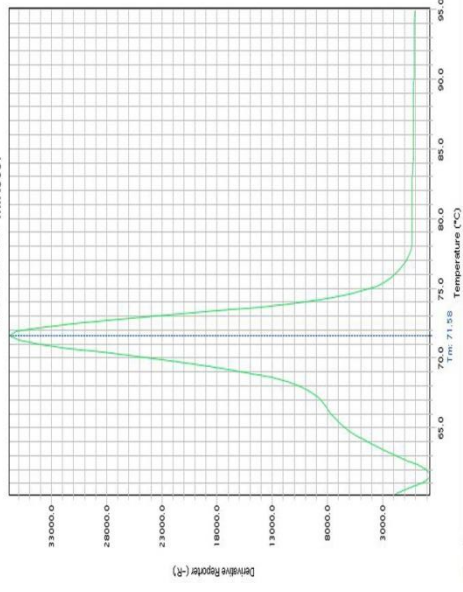
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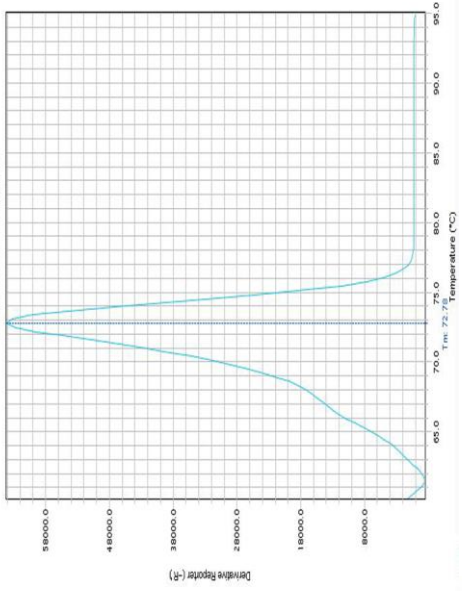
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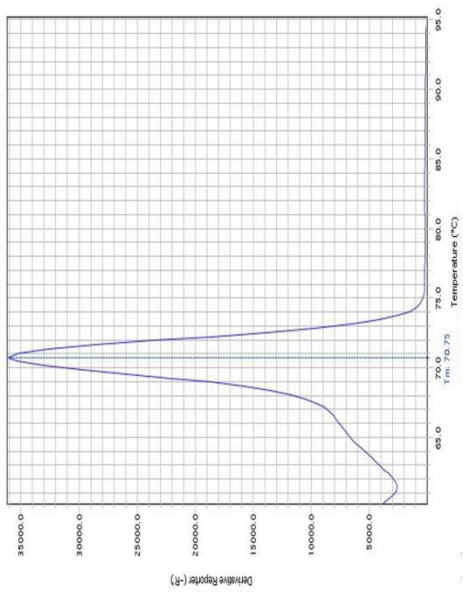
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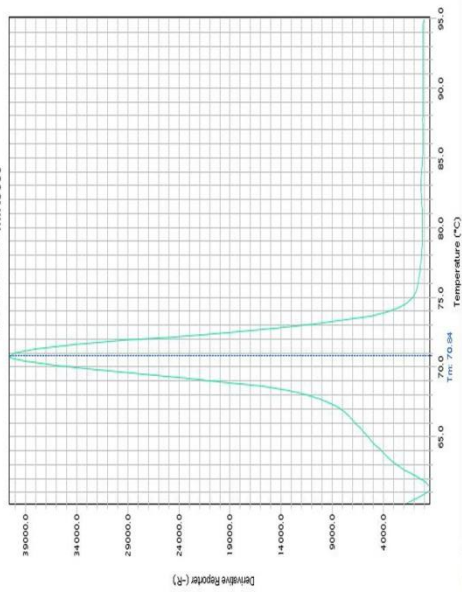
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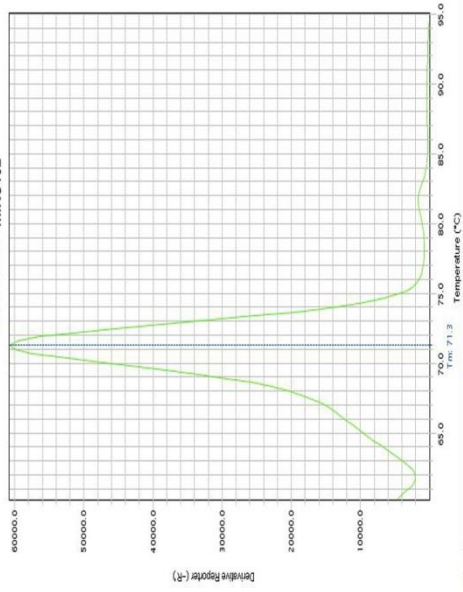
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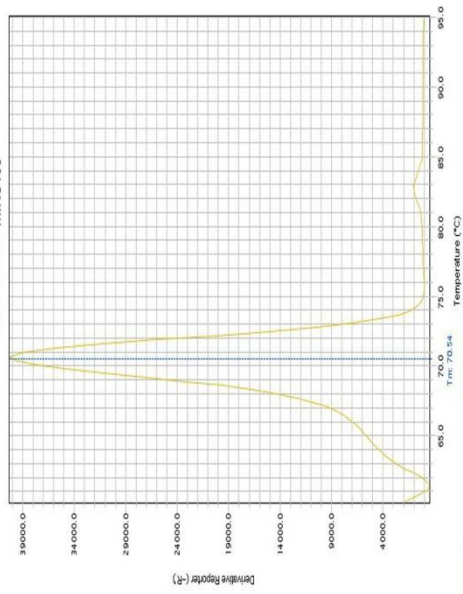
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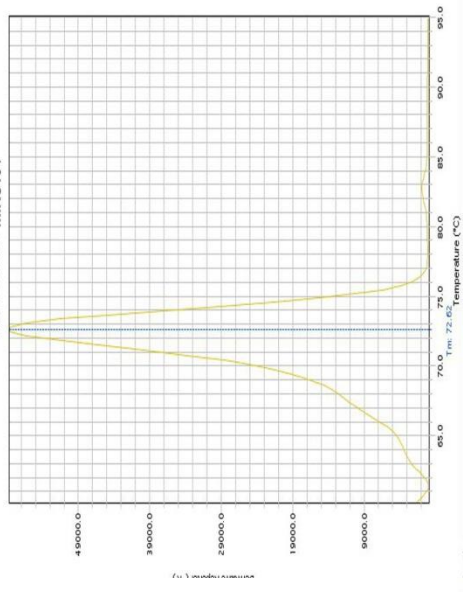
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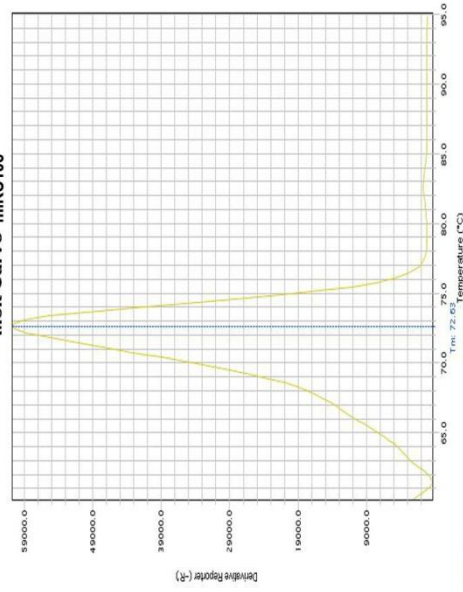
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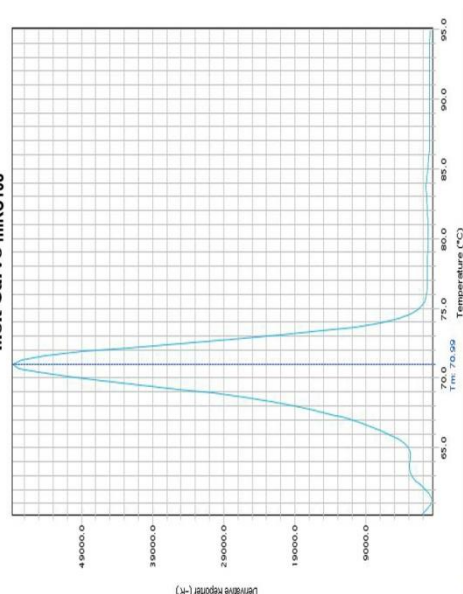
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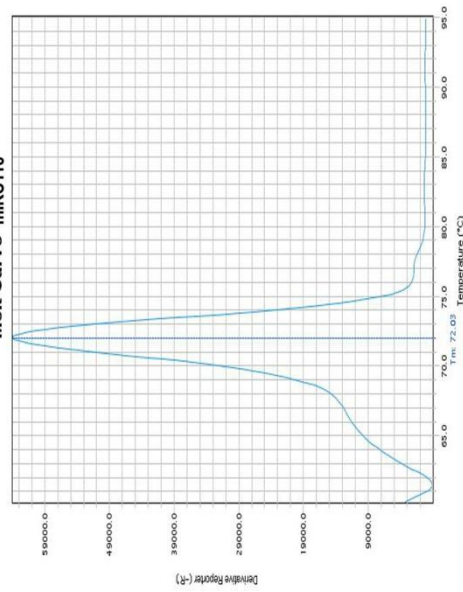
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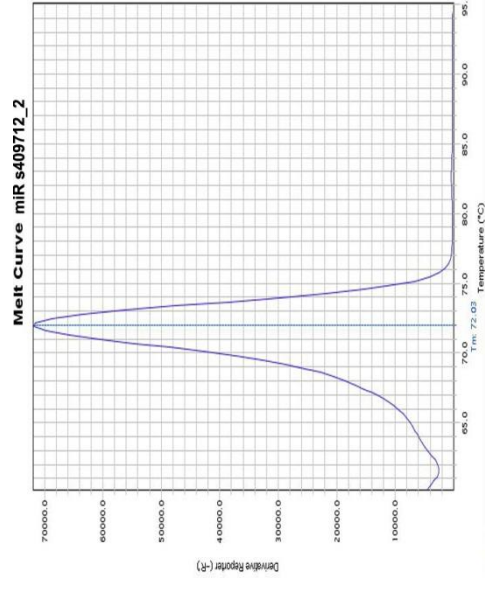
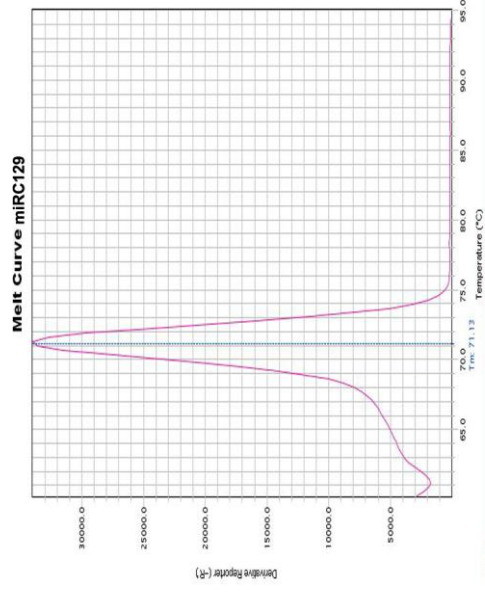
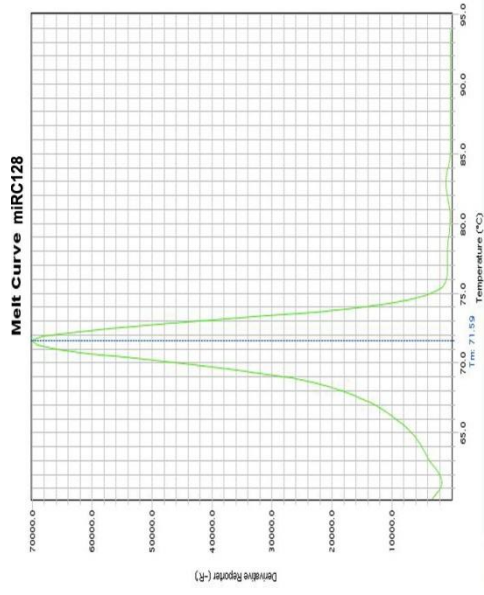
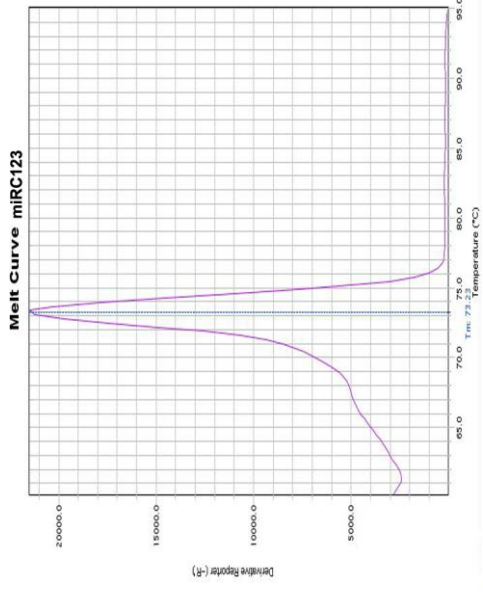
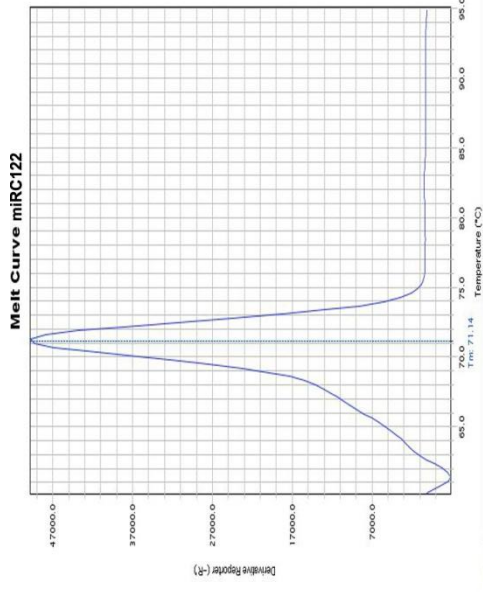
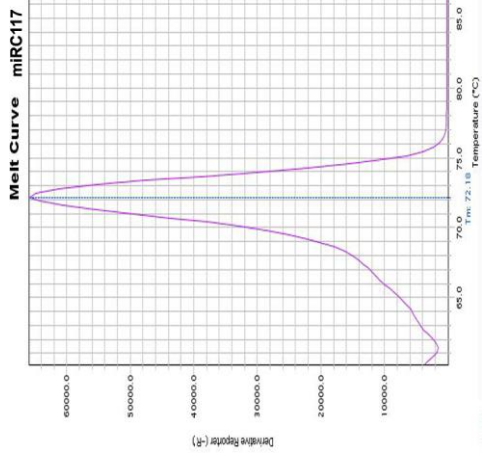


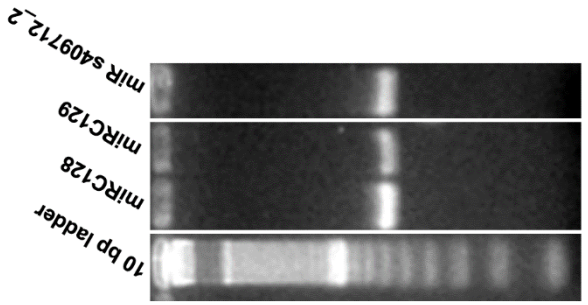
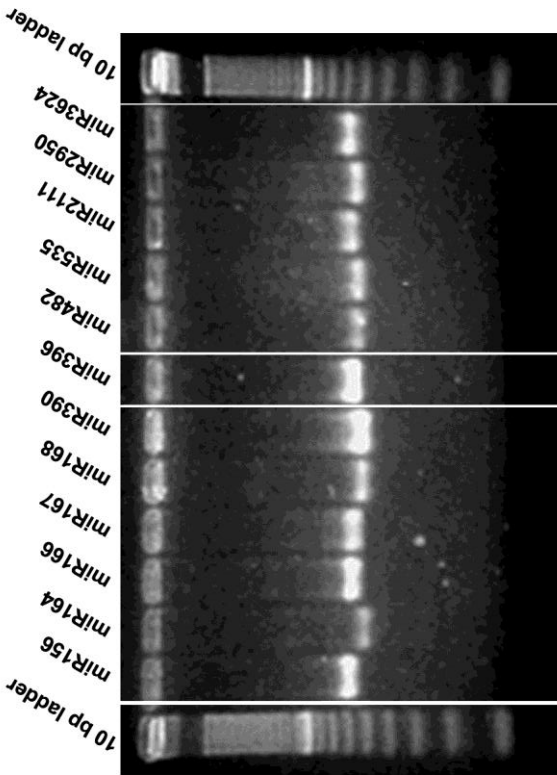
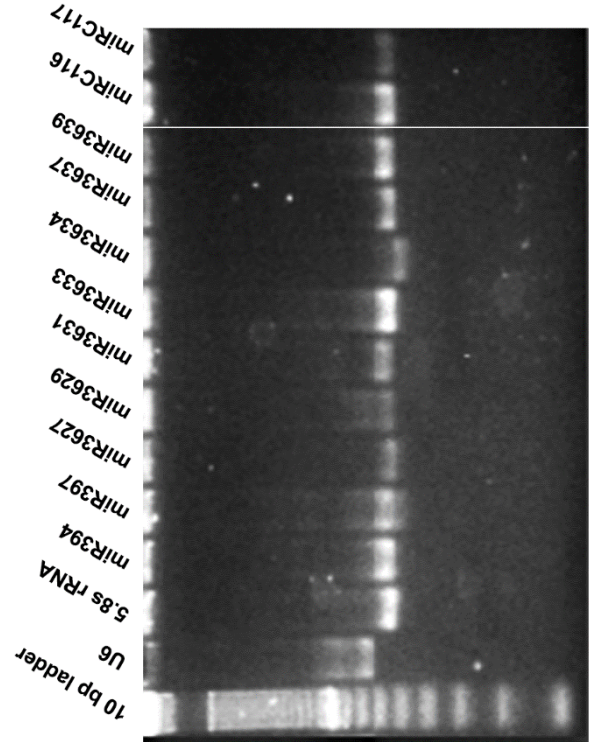
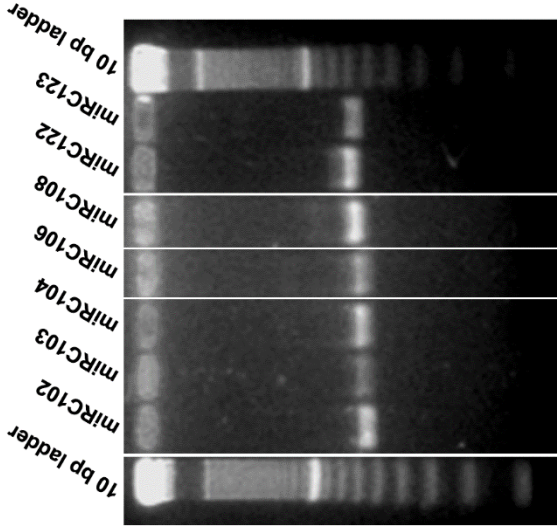
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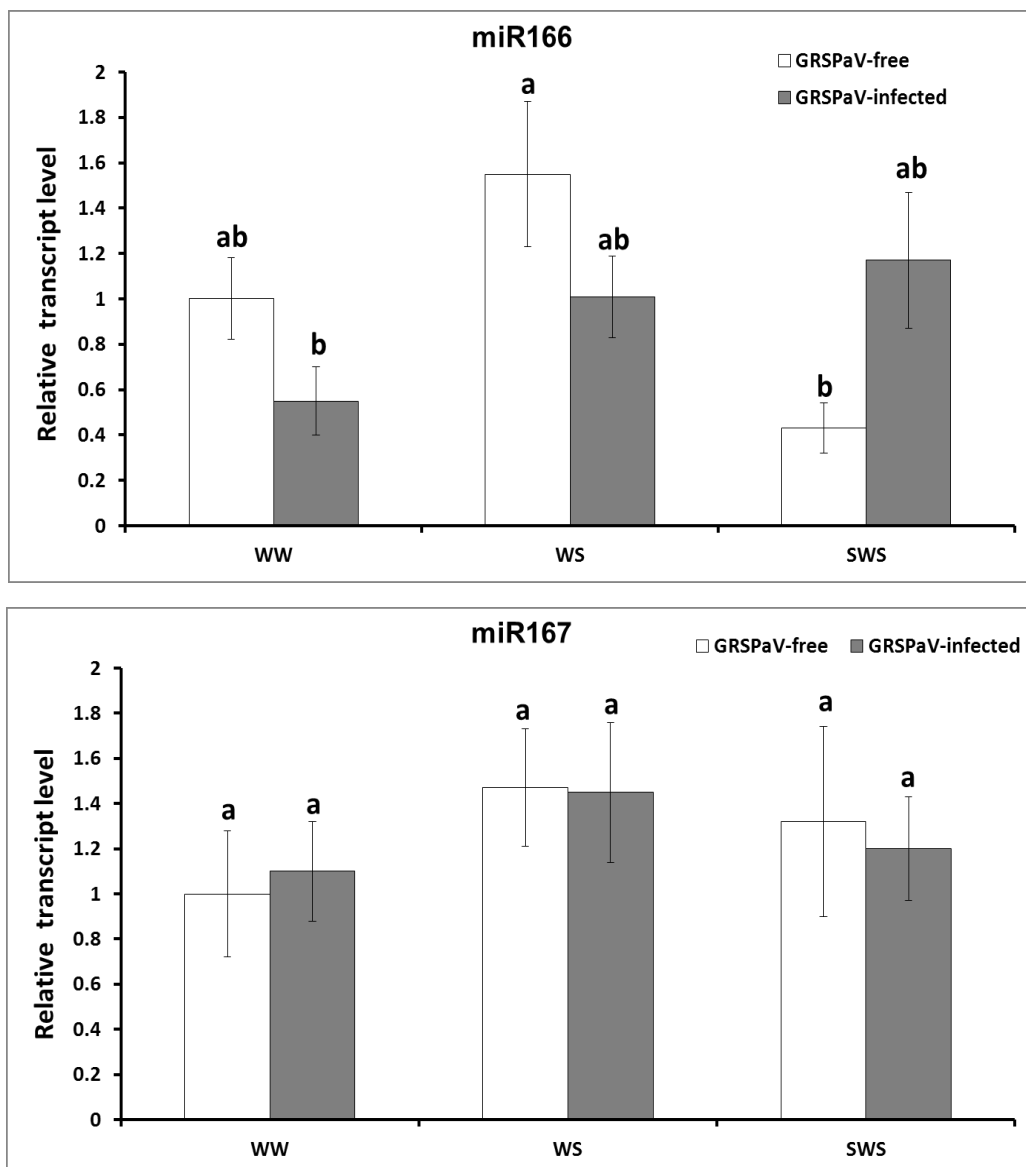


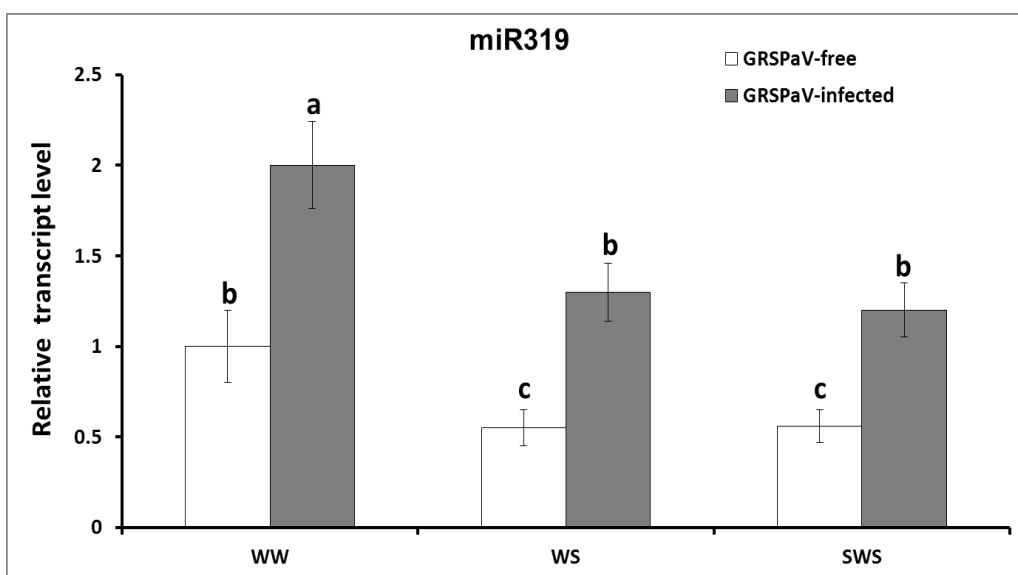
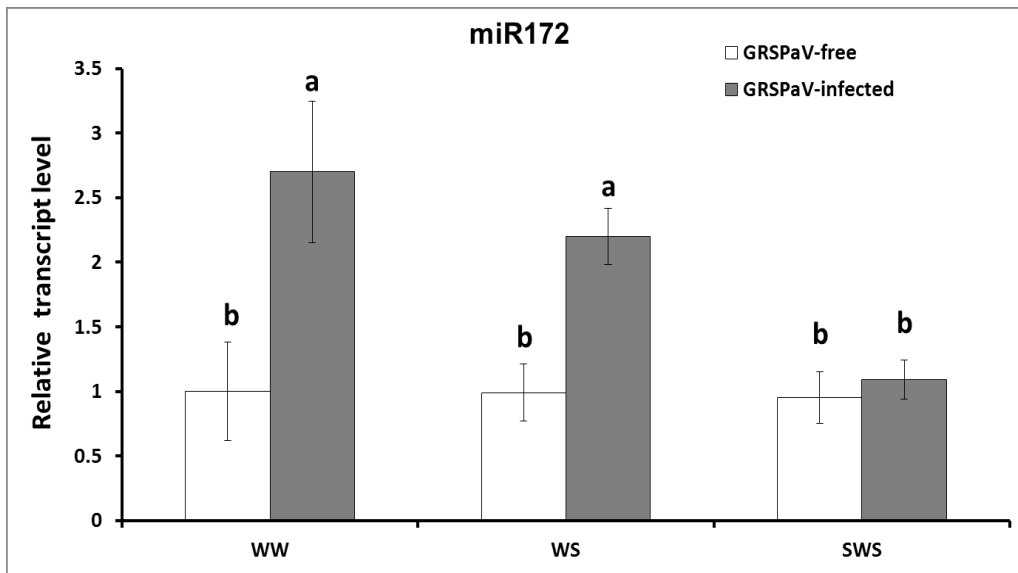
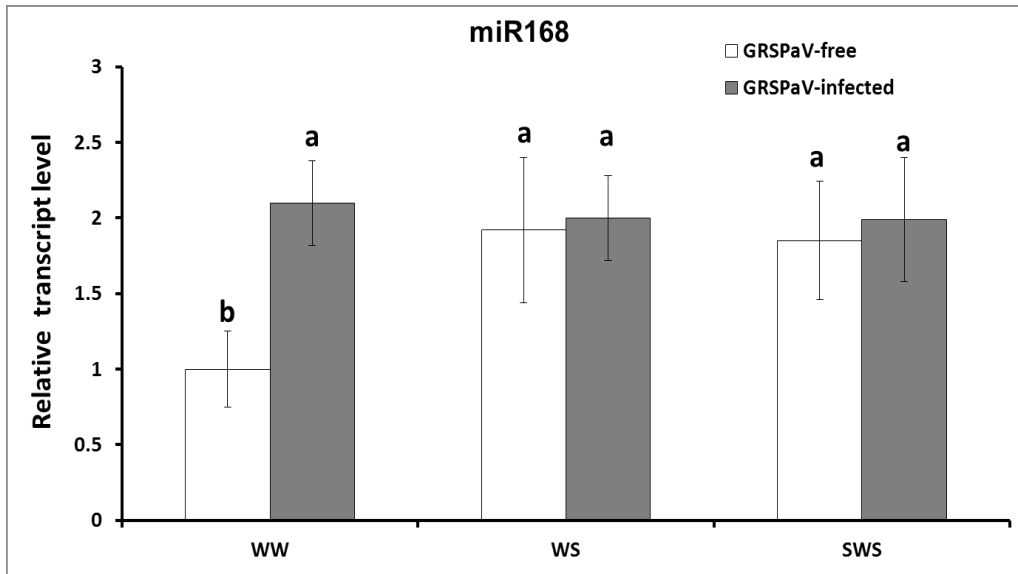


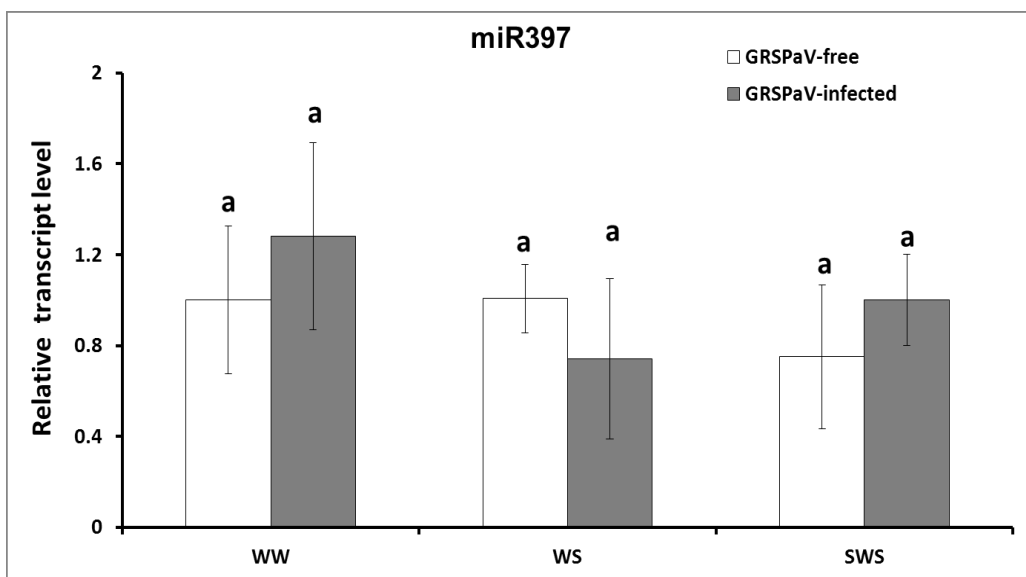
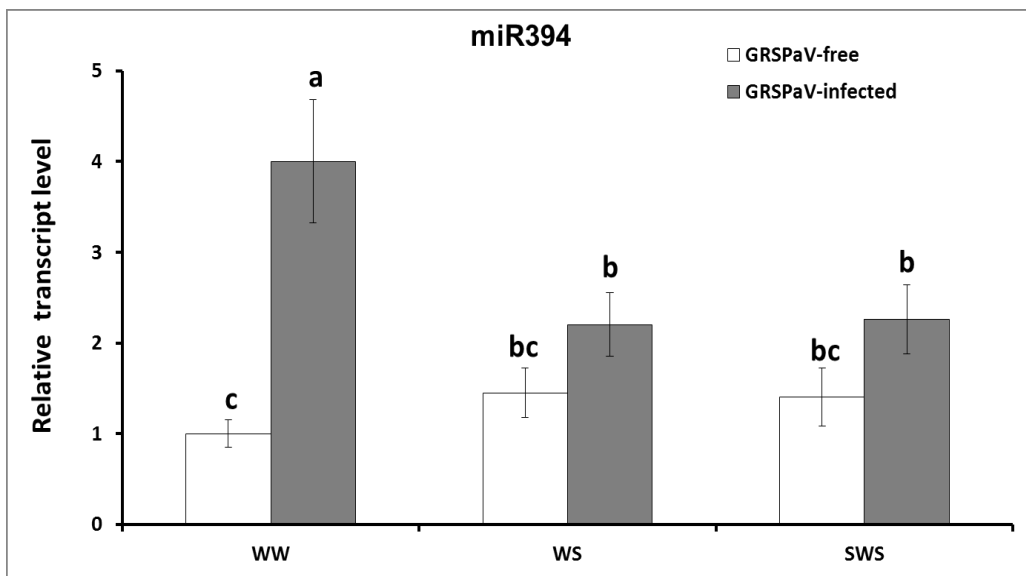
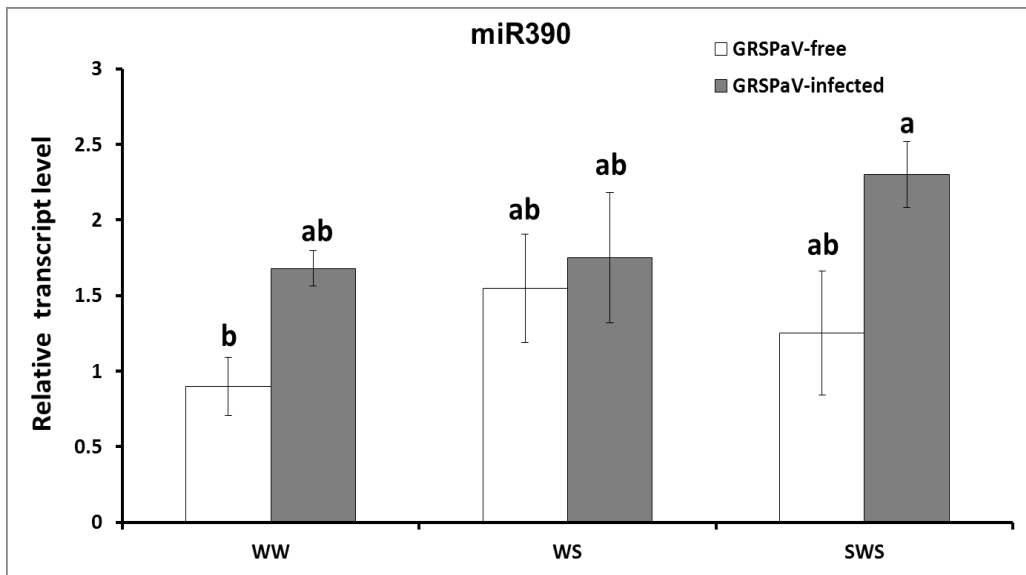


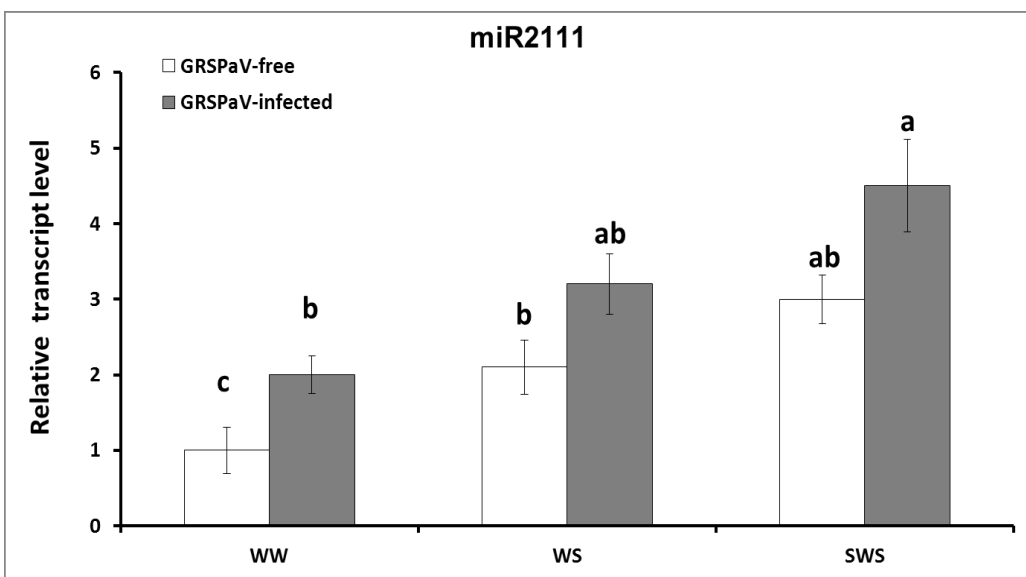
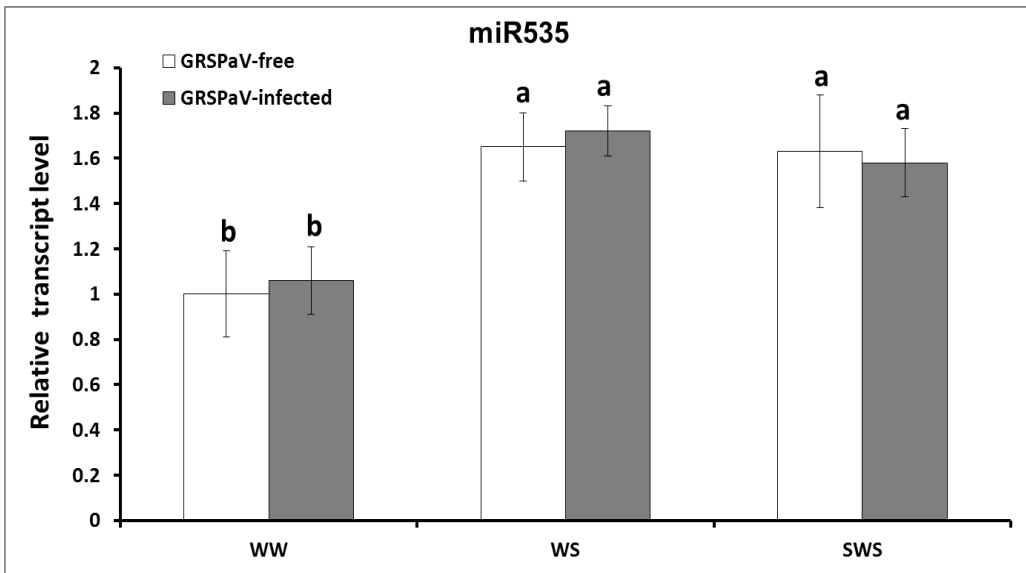
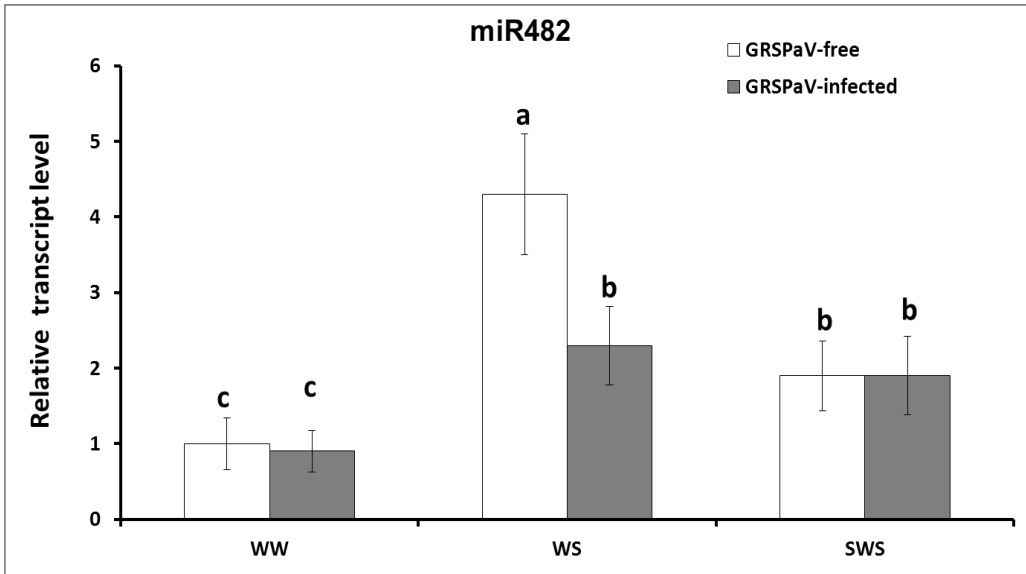
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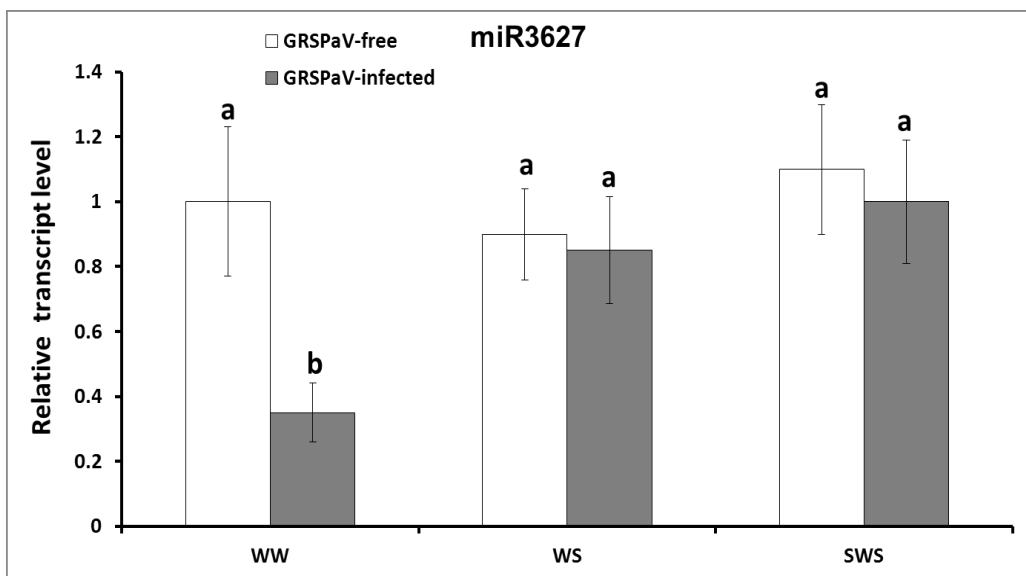
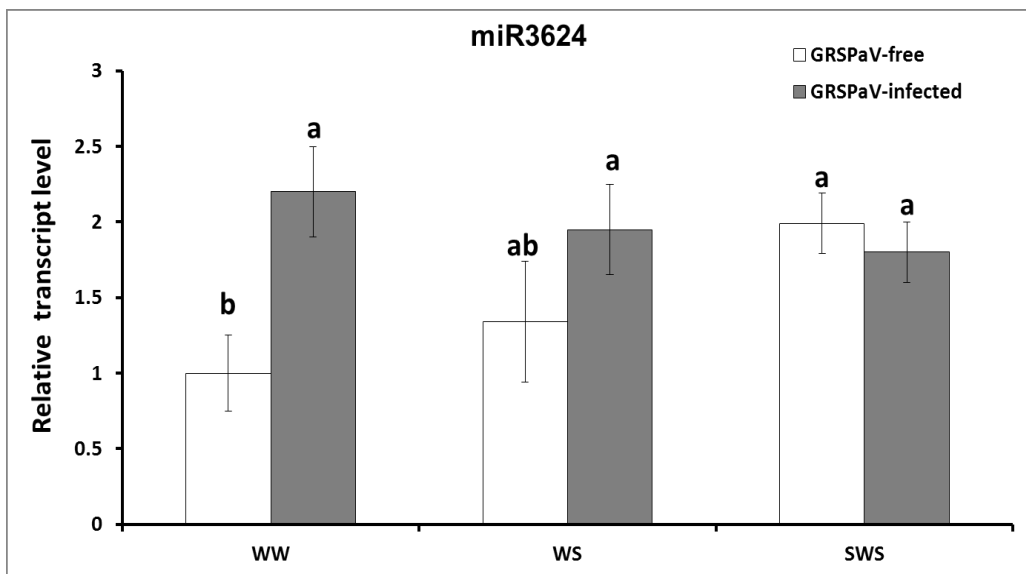
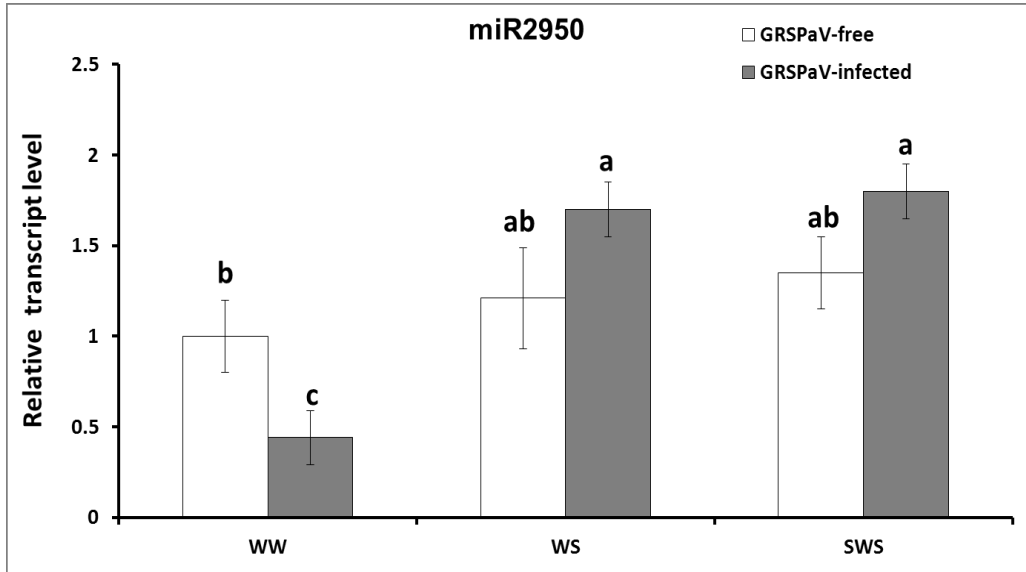
Supplementary Figure S4. Relative expression level of miR166, miR167, miR168, miR172, miR319, miR390, miR394, miR397, miR482, miR535, miR2111, miR2950, miR3624, miR3627, miR3629, miR3631, miR3633, miR3634, miR3637 and miR3639 in *Grapevine rupestris stem pitting-associated virus* (GRSPaV)-free and infected ‘Bosco’ leaf as determined by qRT-PCR. Samples were collected under: well watered (WW), water stress (WS), severe water stress (SWS) conditions. qRT-PCR signals were normalised to U6 and 5.8 rRNA. Data are presented as mean \pm standard deviation of three biological replicates; different letters denote significant differences at $p \leq 0.05$.

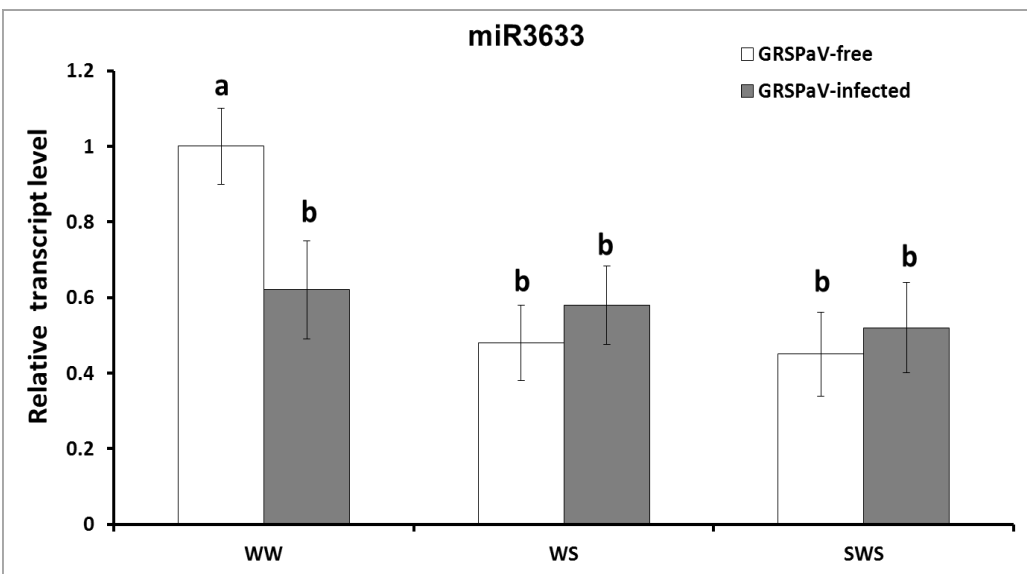
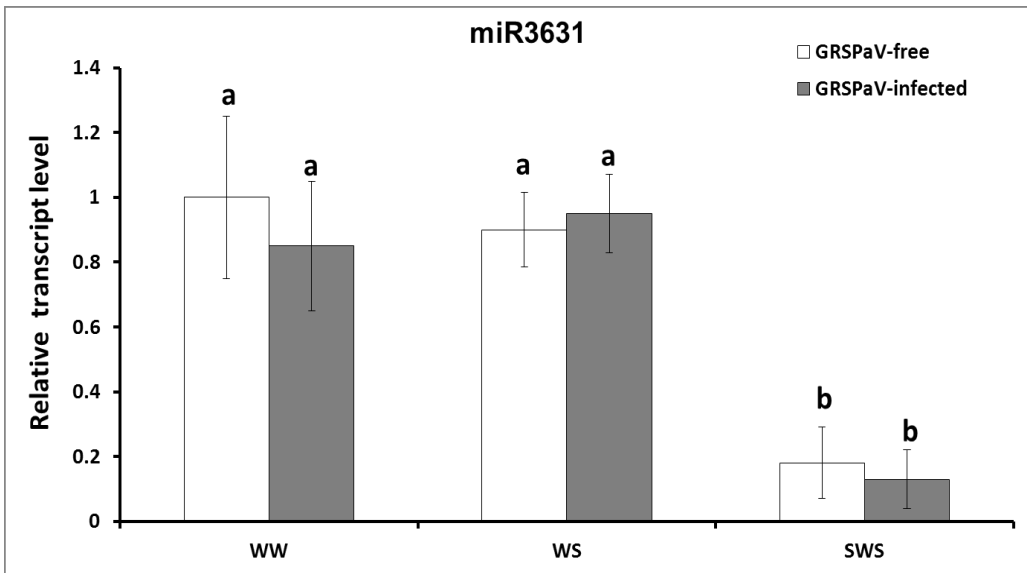
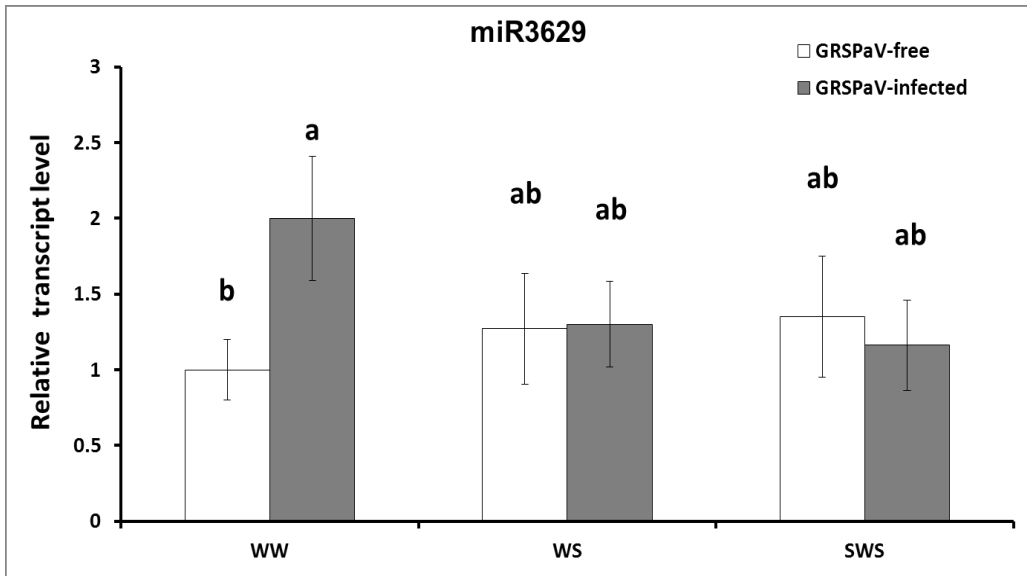


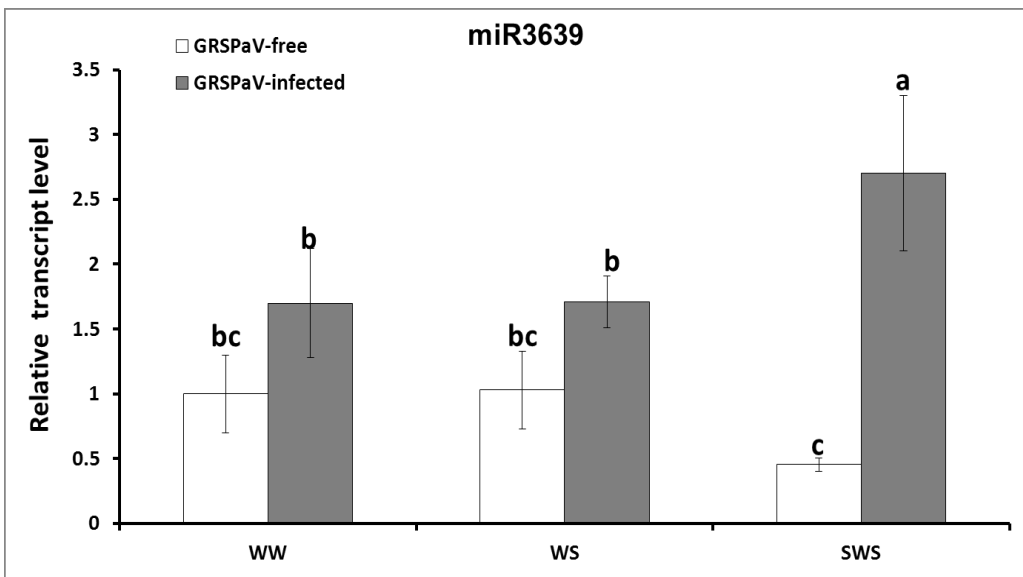
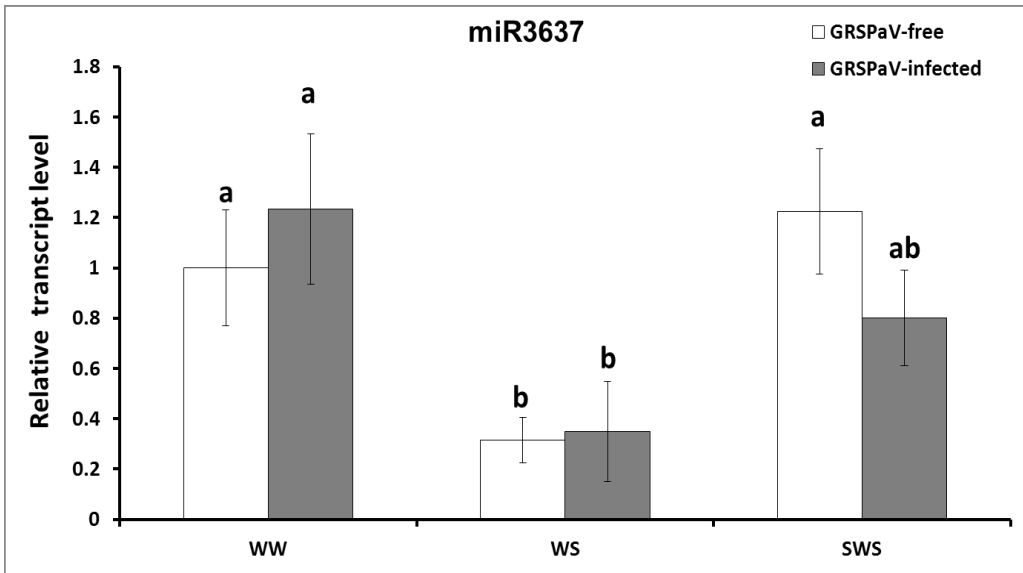
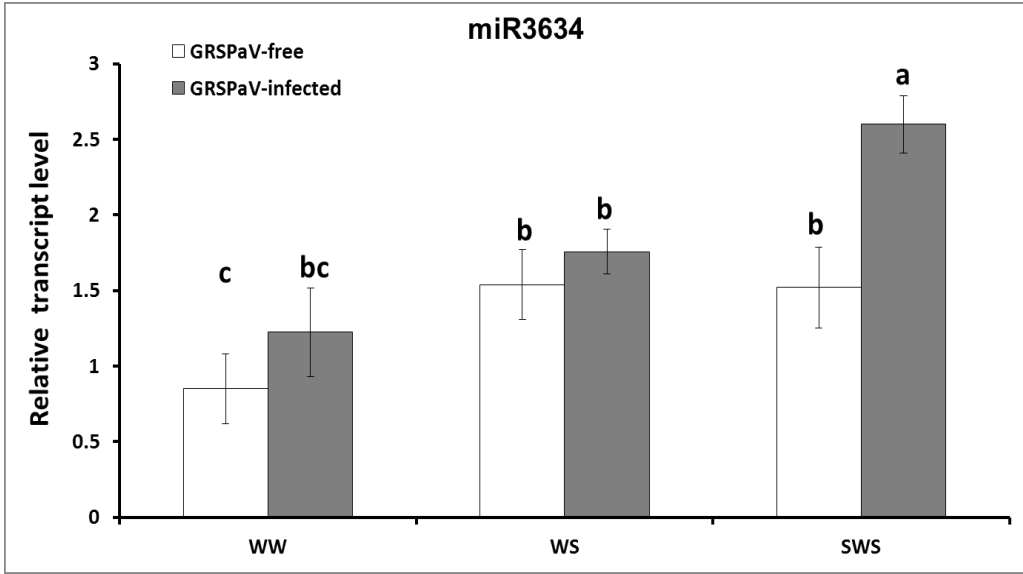












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miRC108

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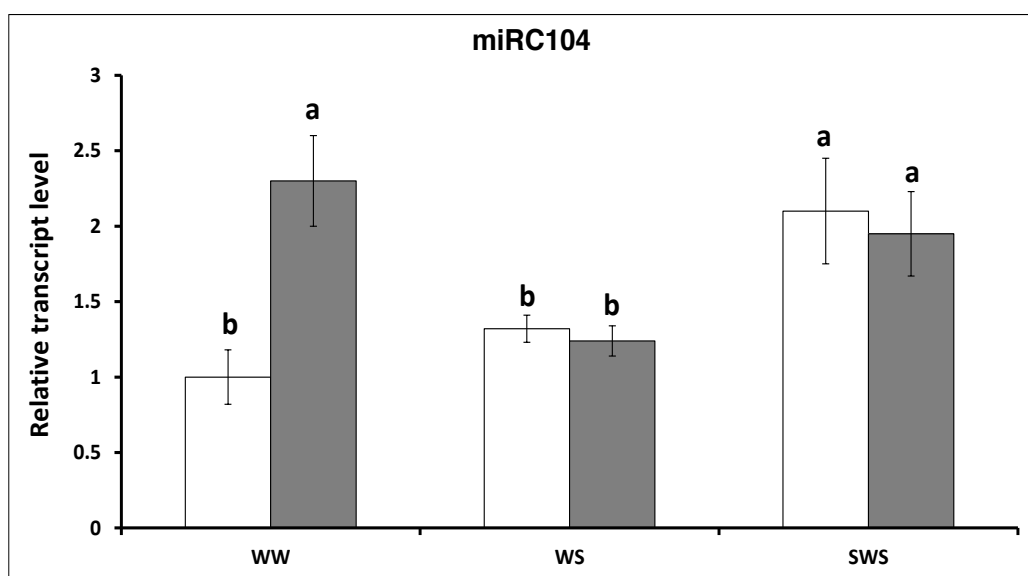
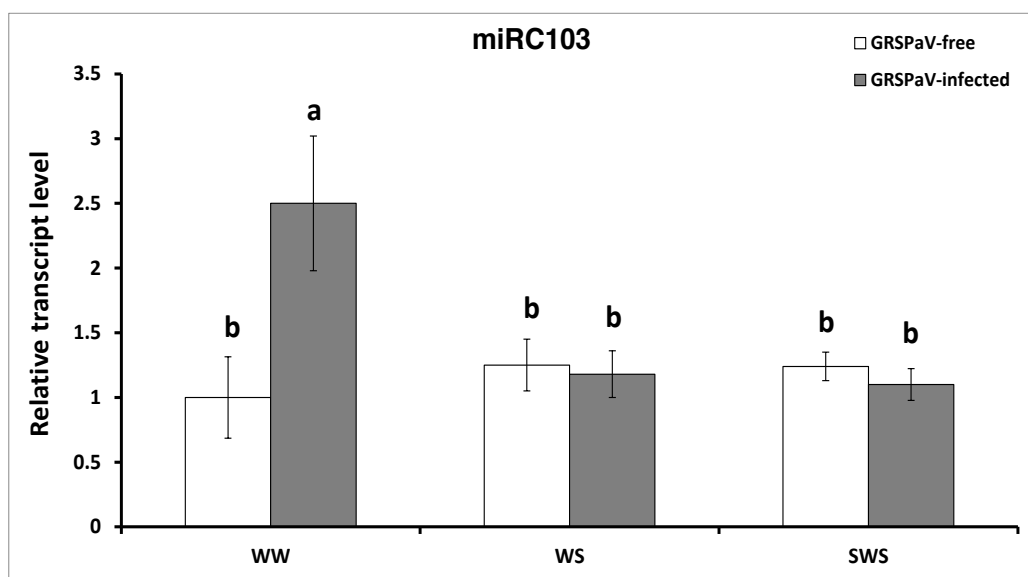
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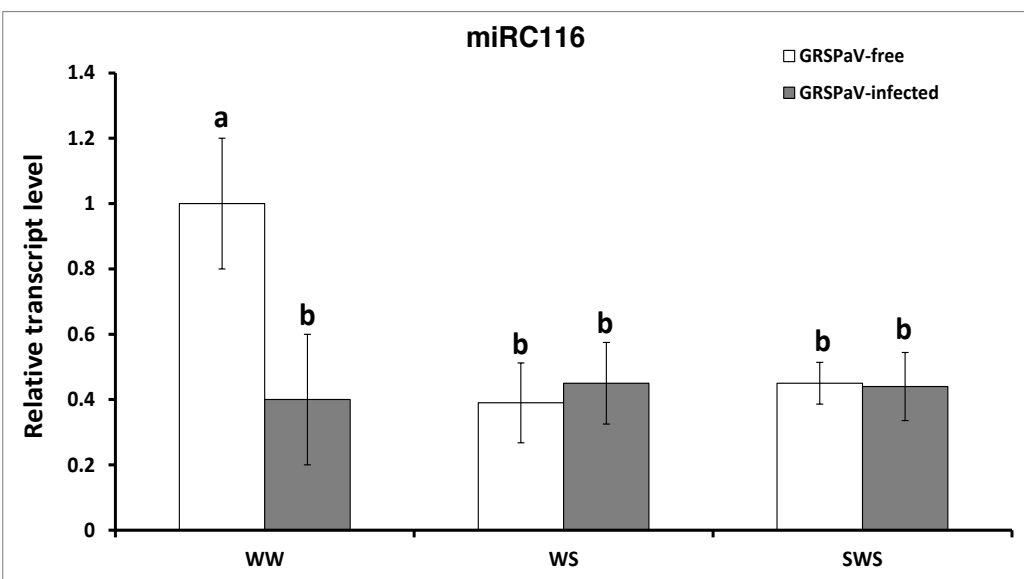
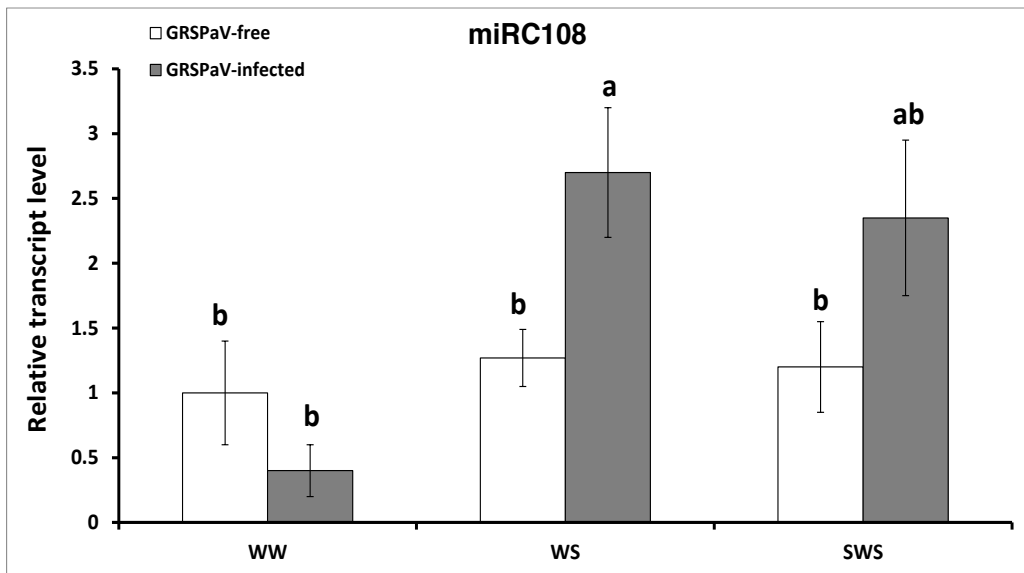
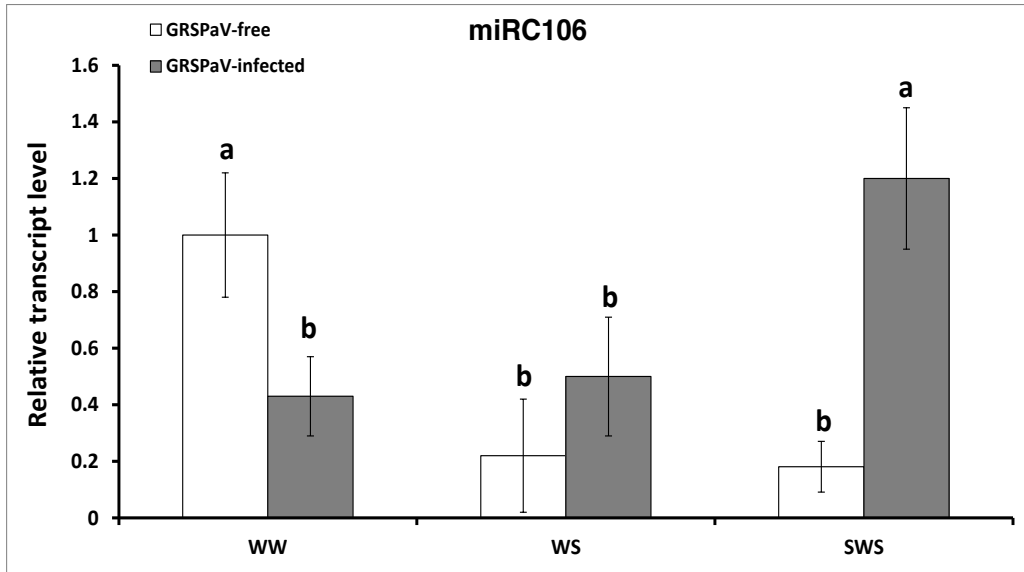
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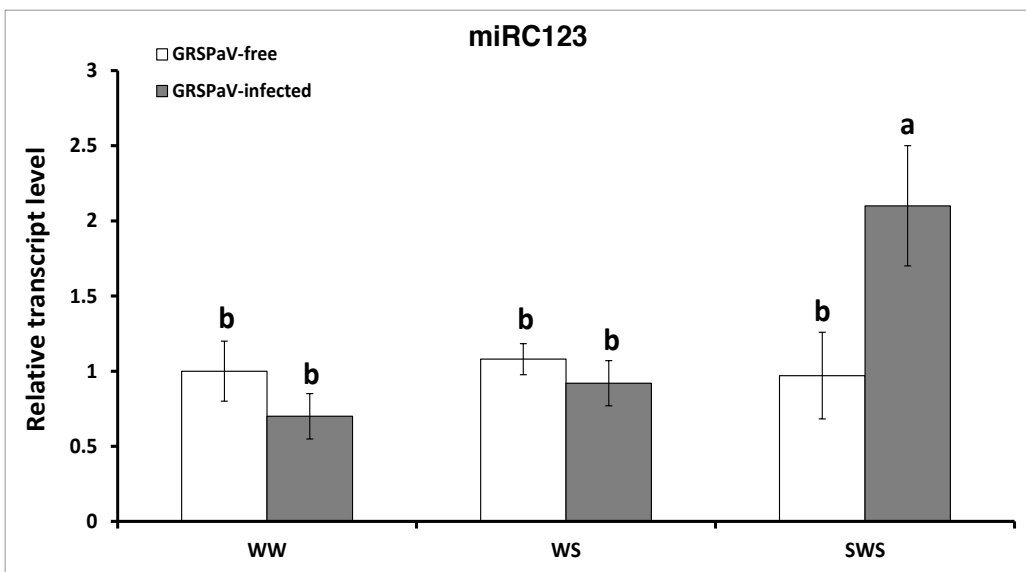
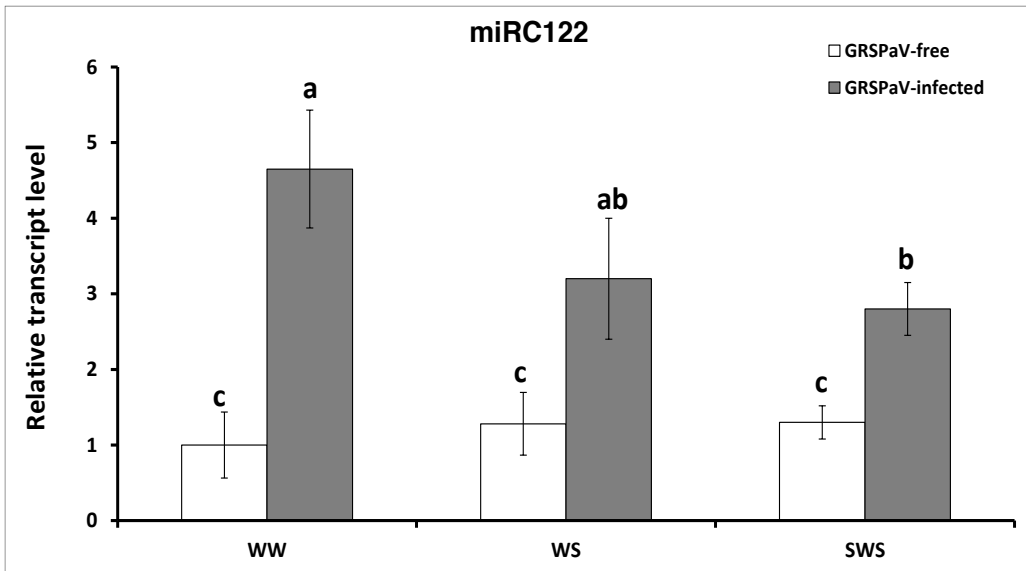
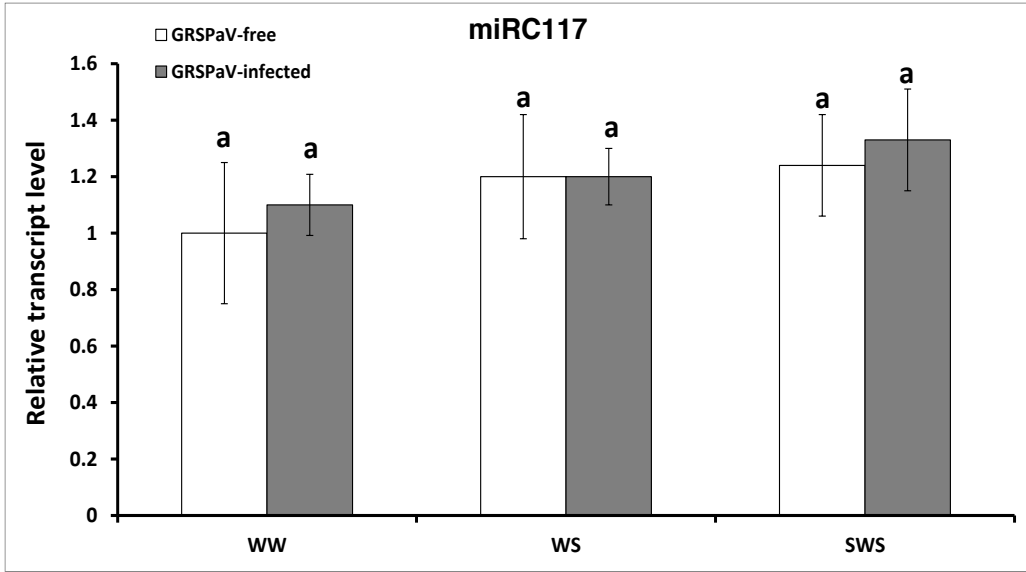
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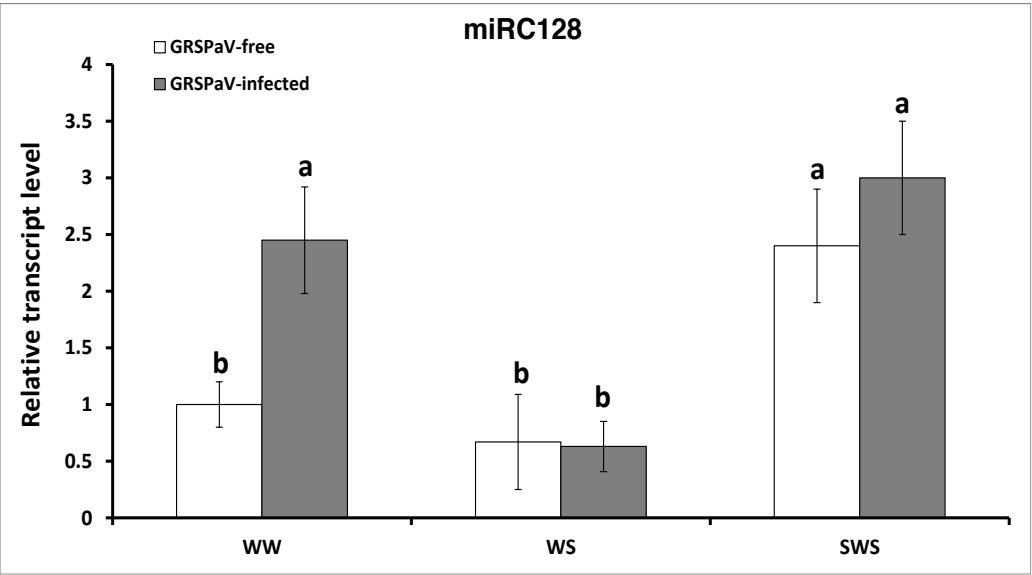
Supplementary Figure S6. BLASTN against selected mature miRNA in miRBASE.

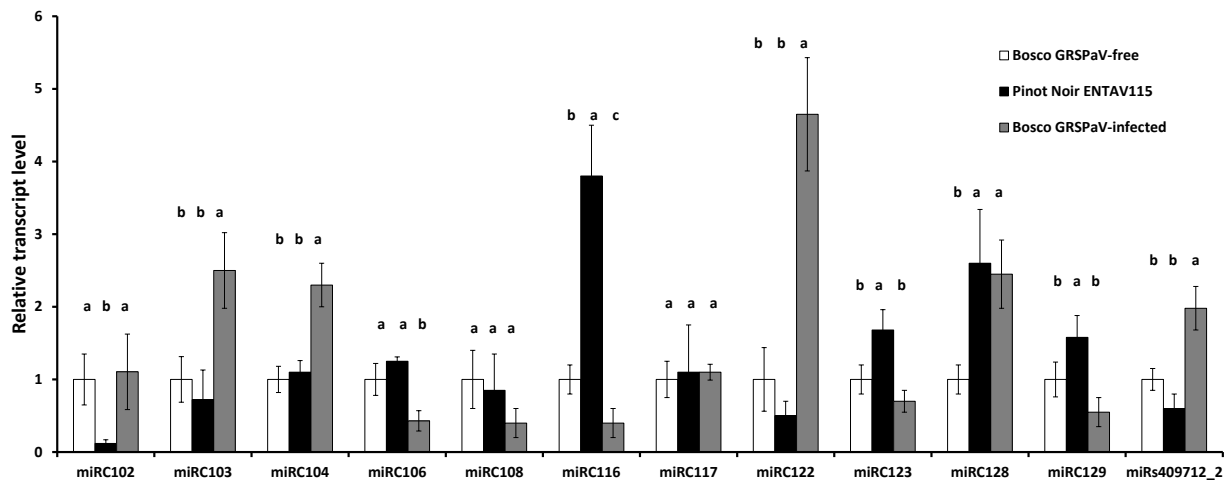
Supplementary Figure S7. Relative expression level of miRC103, miRC104, miRC106, miRC108, miRC116, miRC117, miRC122, miRC123 and miRC128 in *Grapevine rupestris stem pitting-associated virus* (GRSPaV) -free and infected ‘Bosco’ leaf as determined by qRT-PCR. Samples were collected under: well watered (WW), water stress (SW), severe water stress (SWS) conditions. qRT-PCR signals were normalised to U6 and 5.8 rRNA levels. Data are presented as mean \pm standard deviation of three biological replicates; different letters denote significant differences at $p \leq 0.05$.









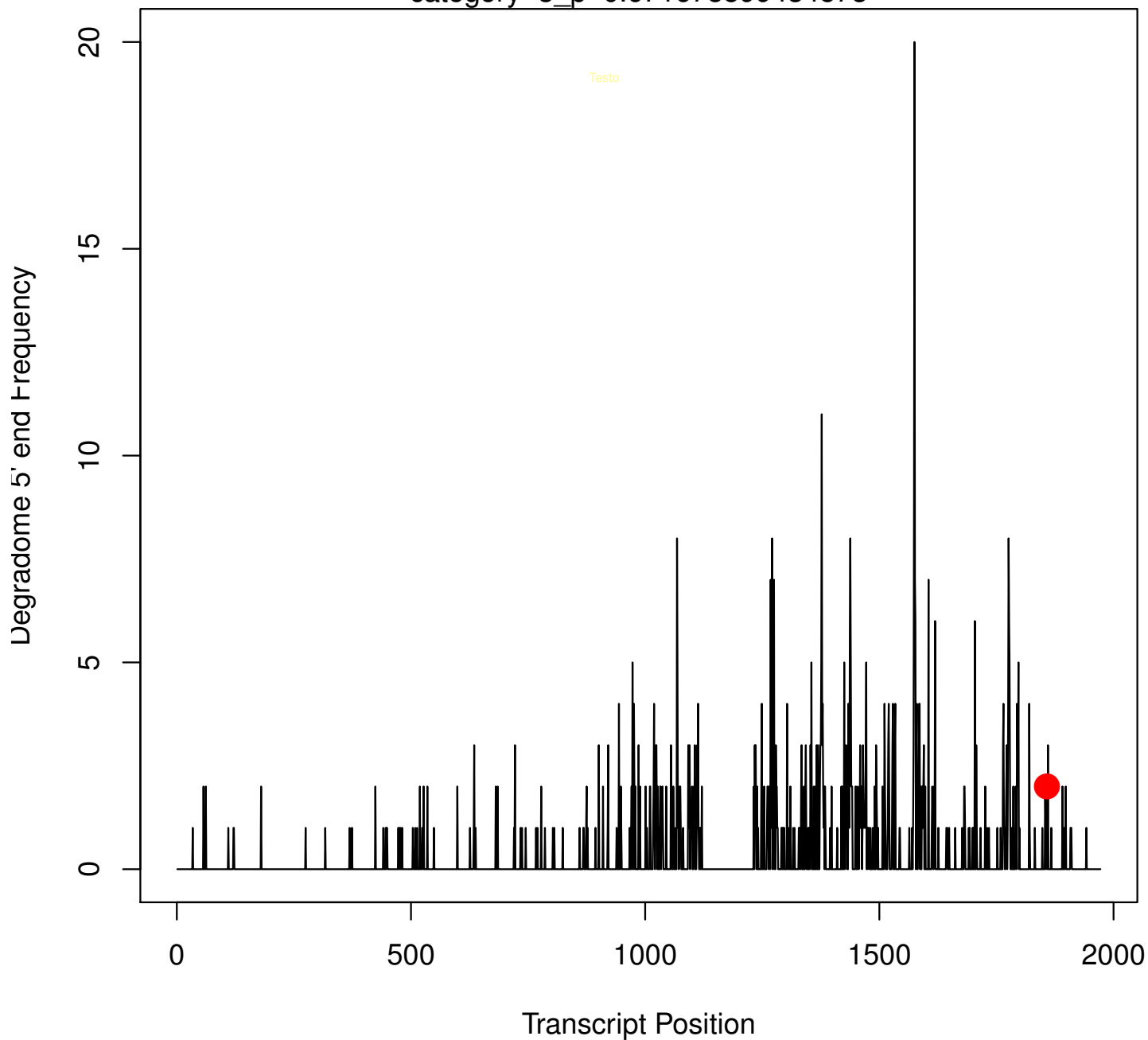


Supplementary Figure S8. Relative expression levels of all novel miRNA candidates in *Grapevine rupestris stem pitting-associated* (GRSPaV)-free, GRSPaV-infected ‘Bosco’ leaves and in ‘Pinot noir’ ENTAV115 leaves as determined by qRT-PCR. Samples were collected under well watered (WW) conditions. qRT-PCR signals were normalised to U6 and 5.8 levels. Data are presented as mean \pm standard deviation of three biological replicates; different letters denote significant differences at $p \leq 0.05$.

Supplementary Figure S9. T-plots for targets of grapevine-specific microRNAs (miRNAs).

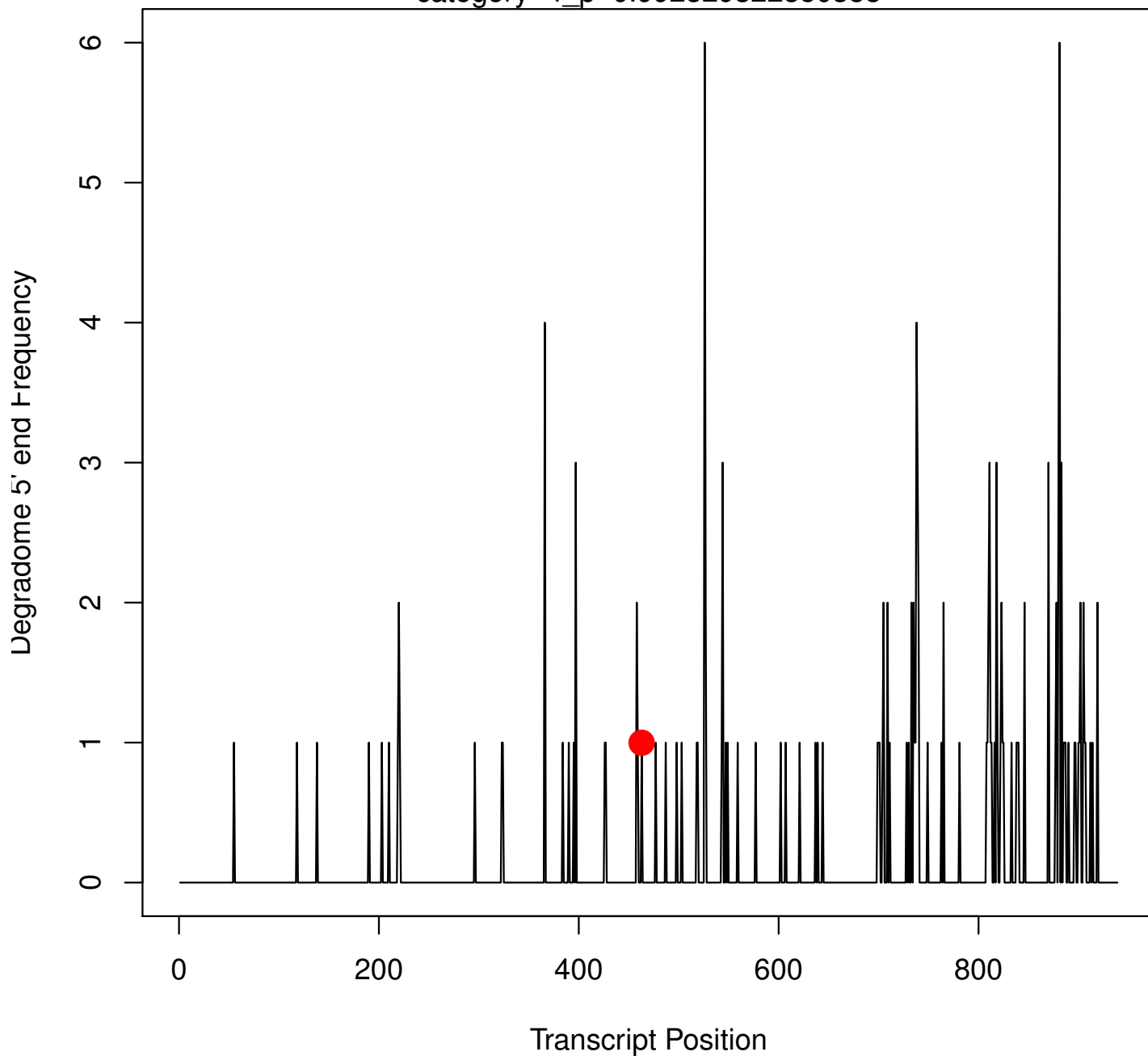
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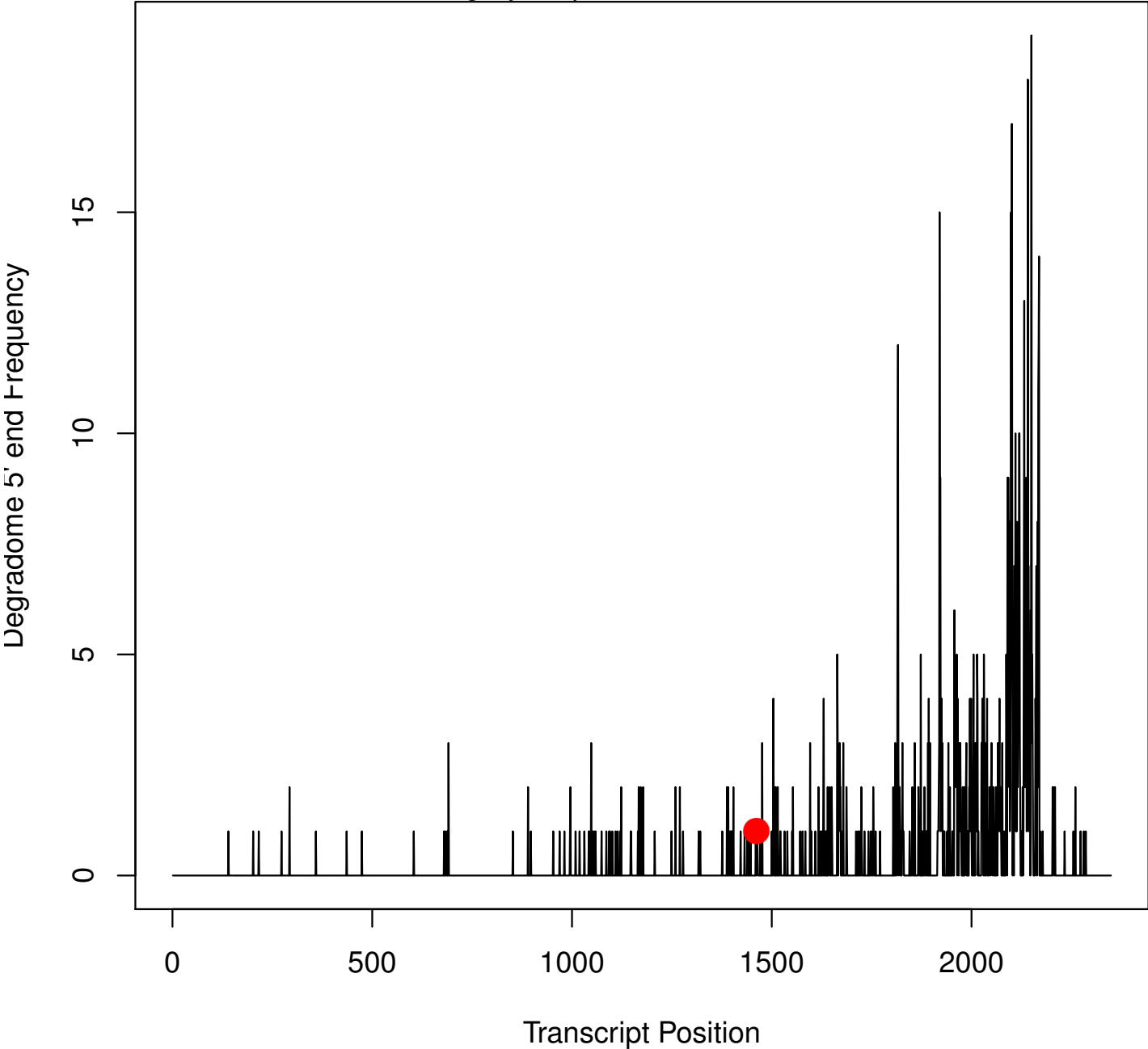
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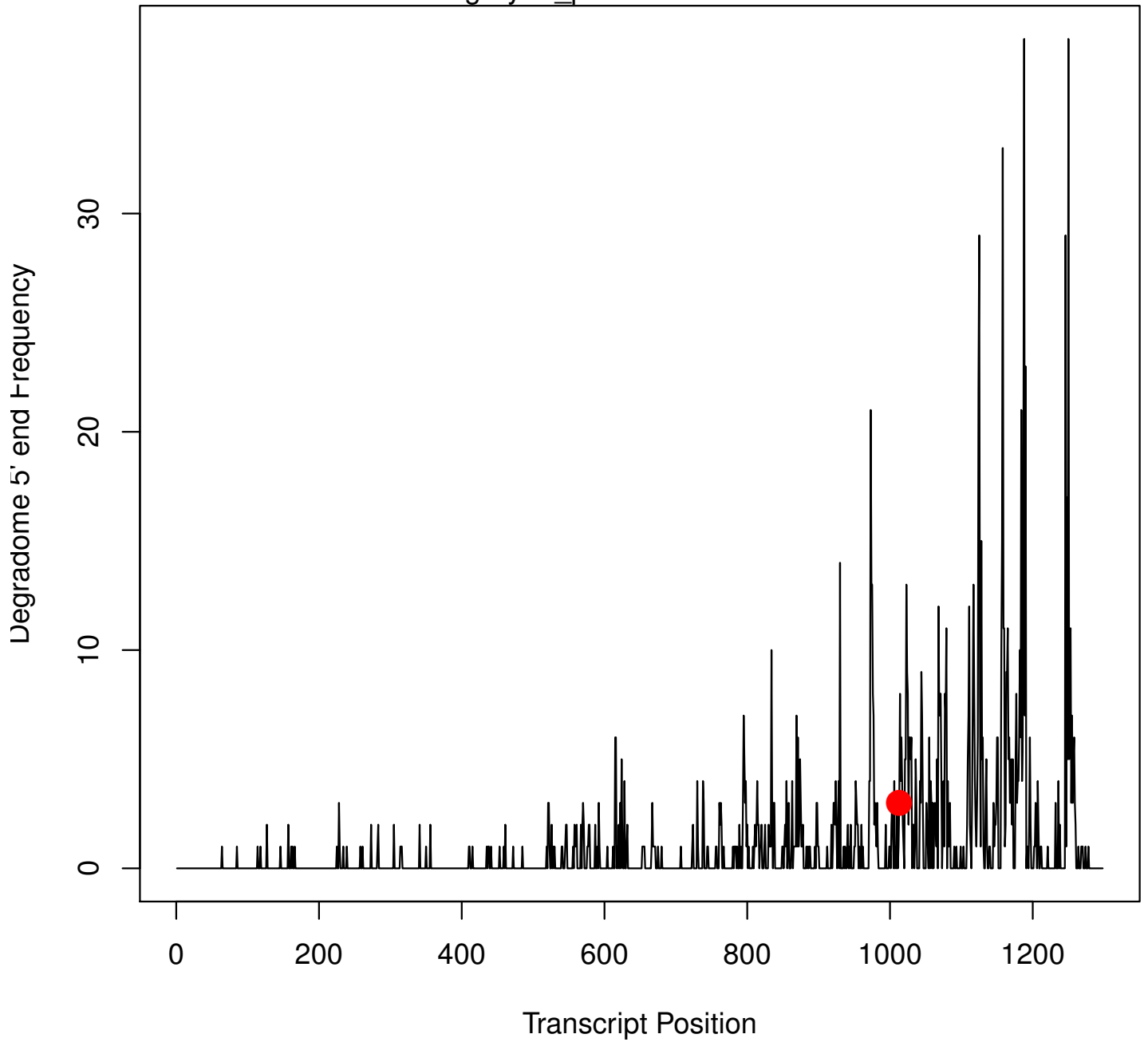
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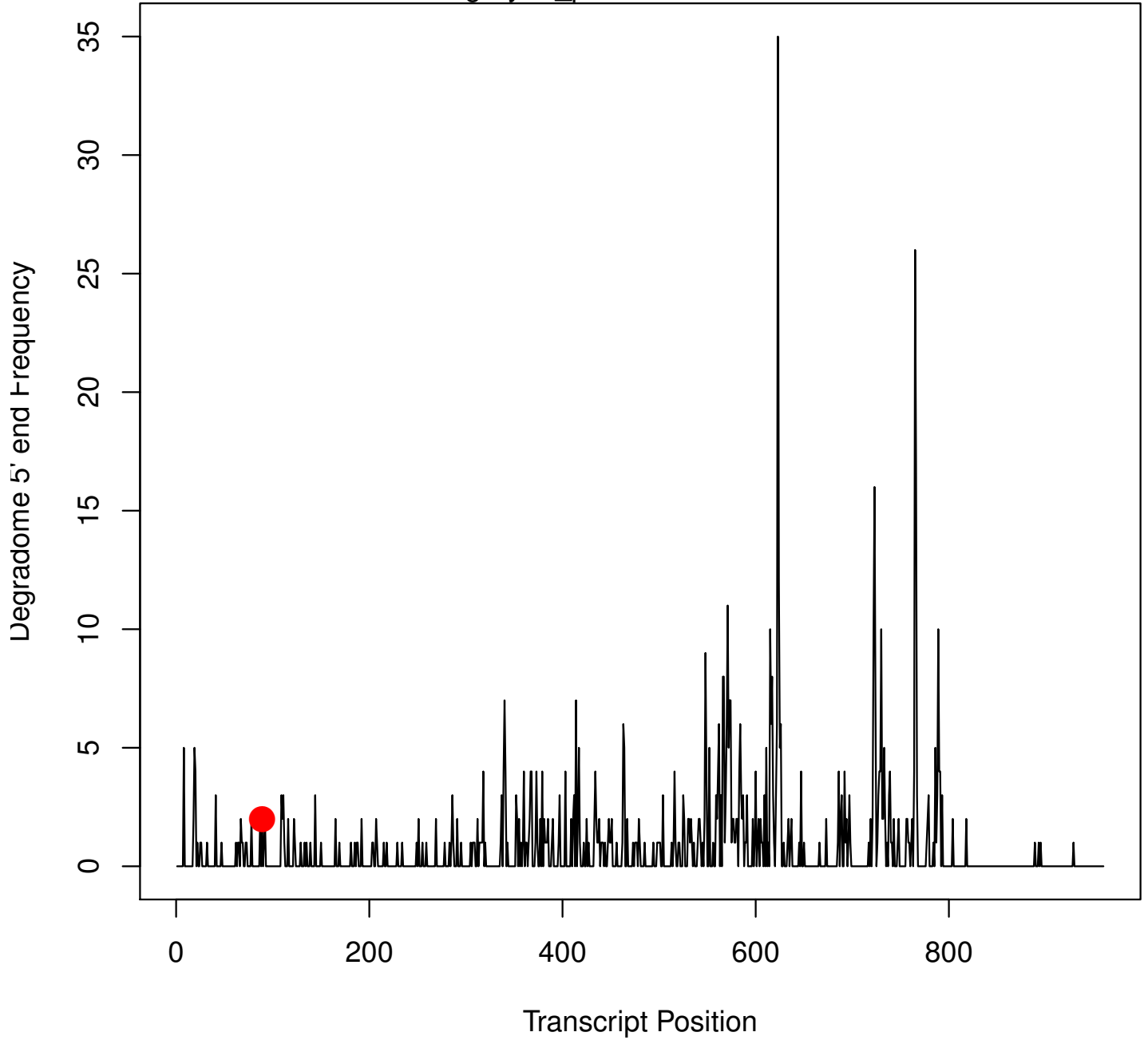
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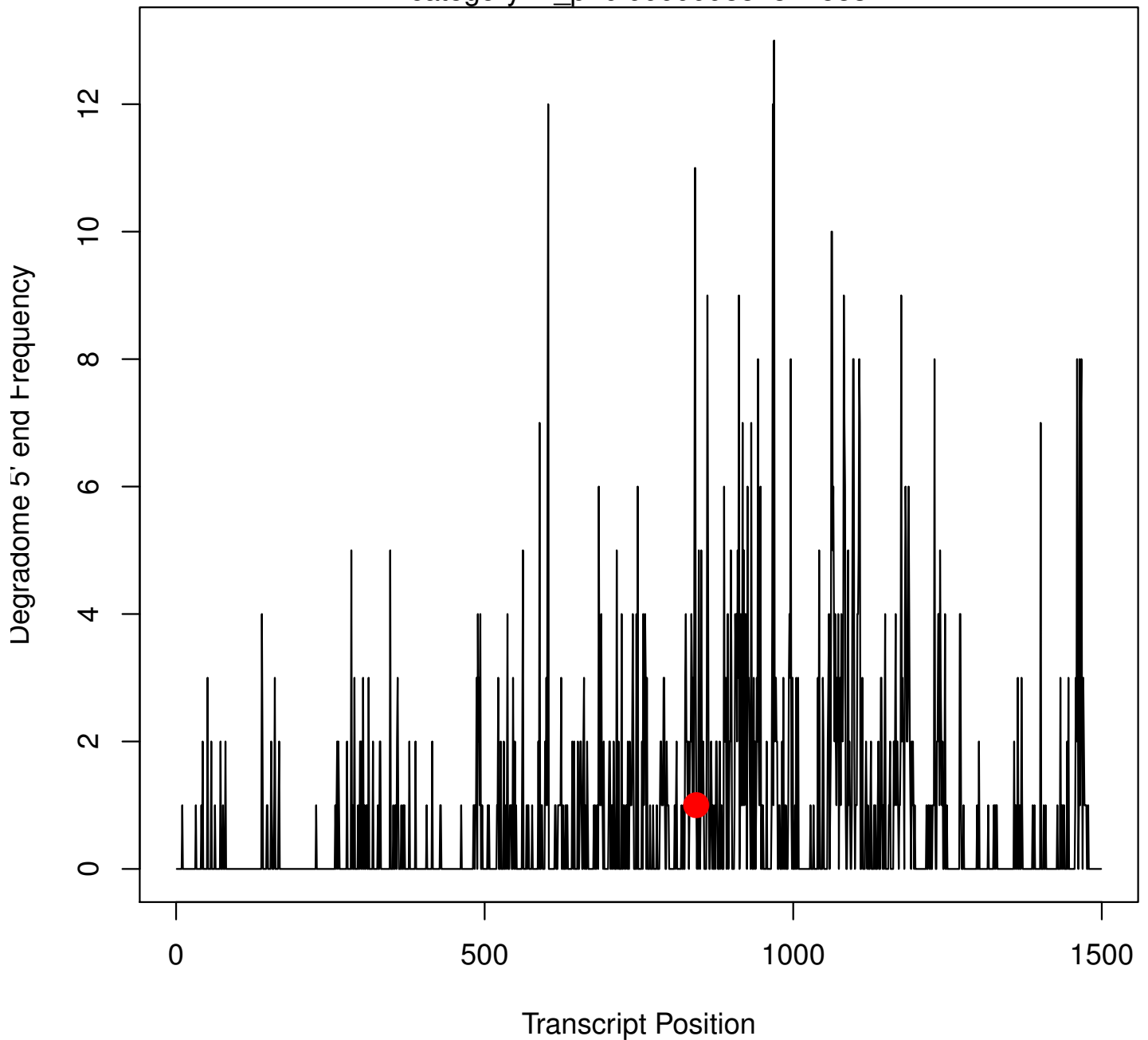
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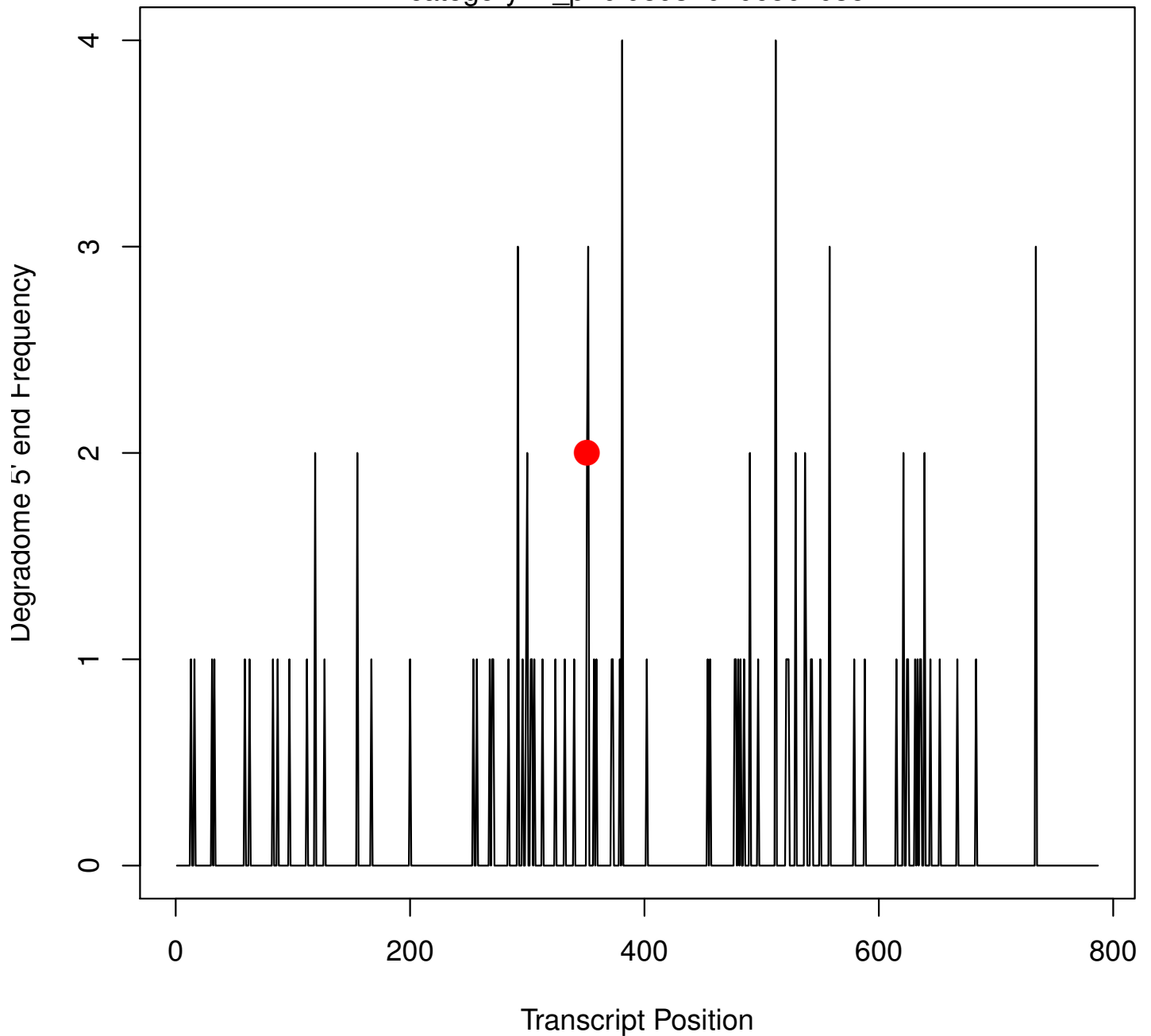
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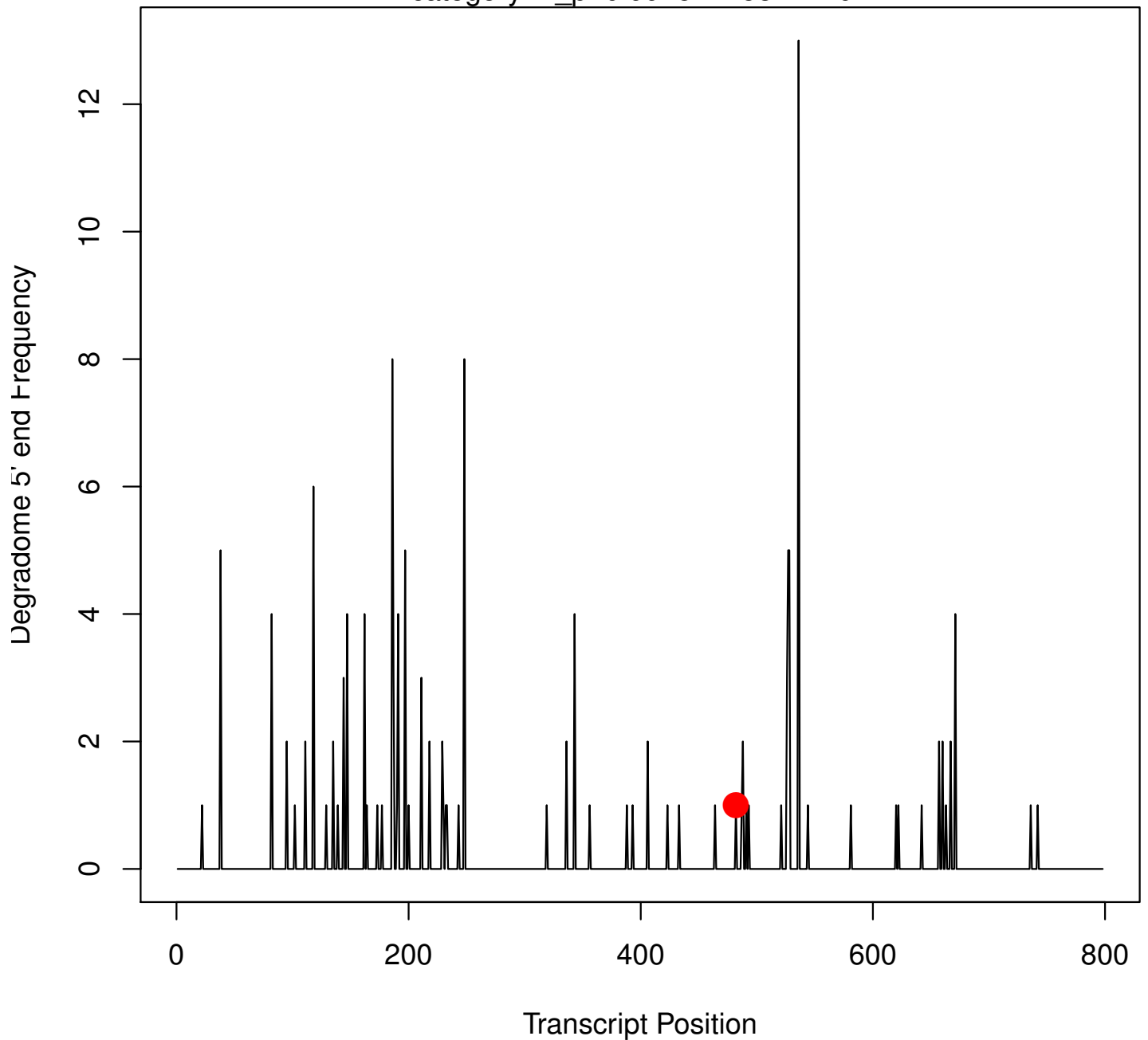
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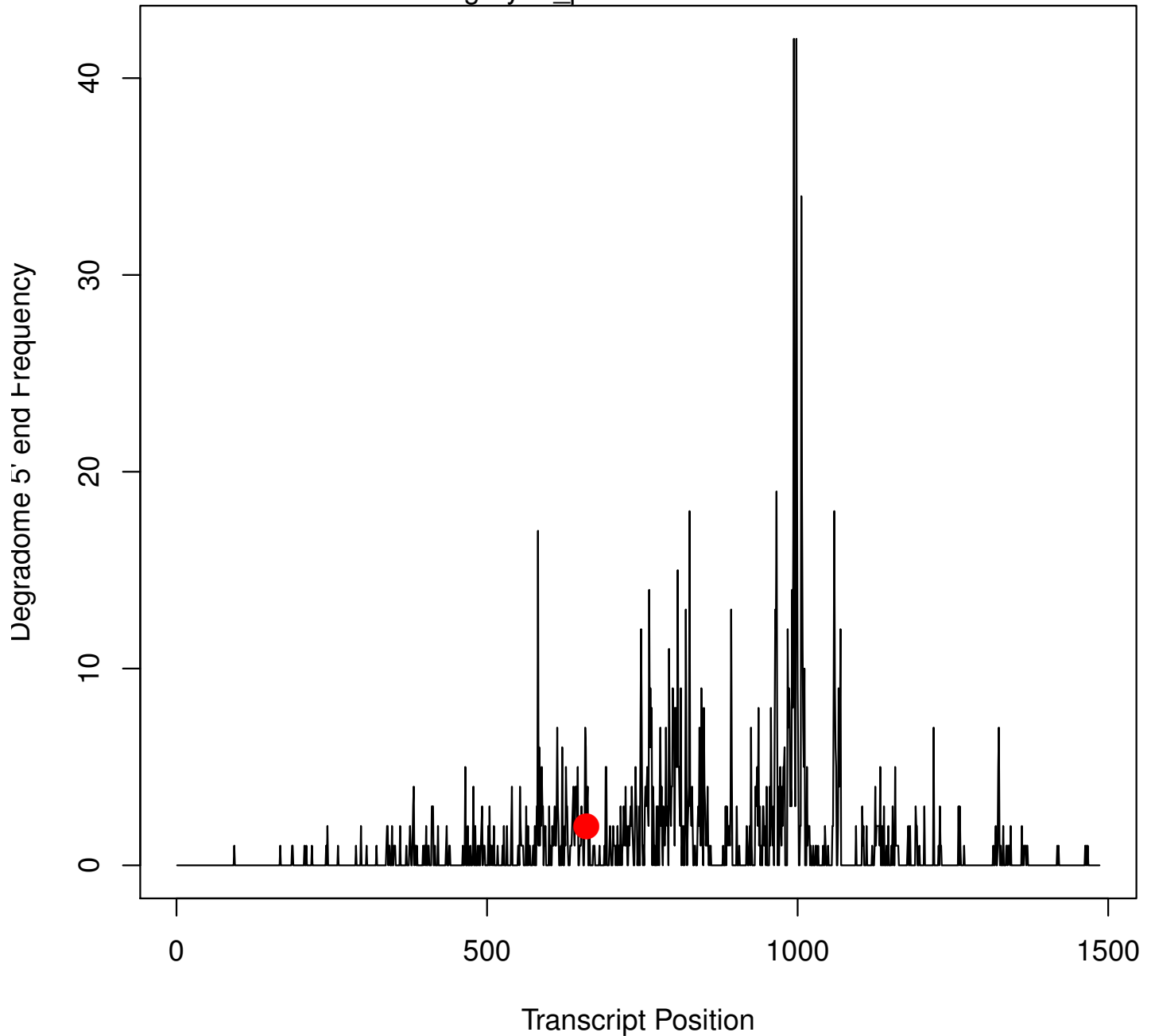
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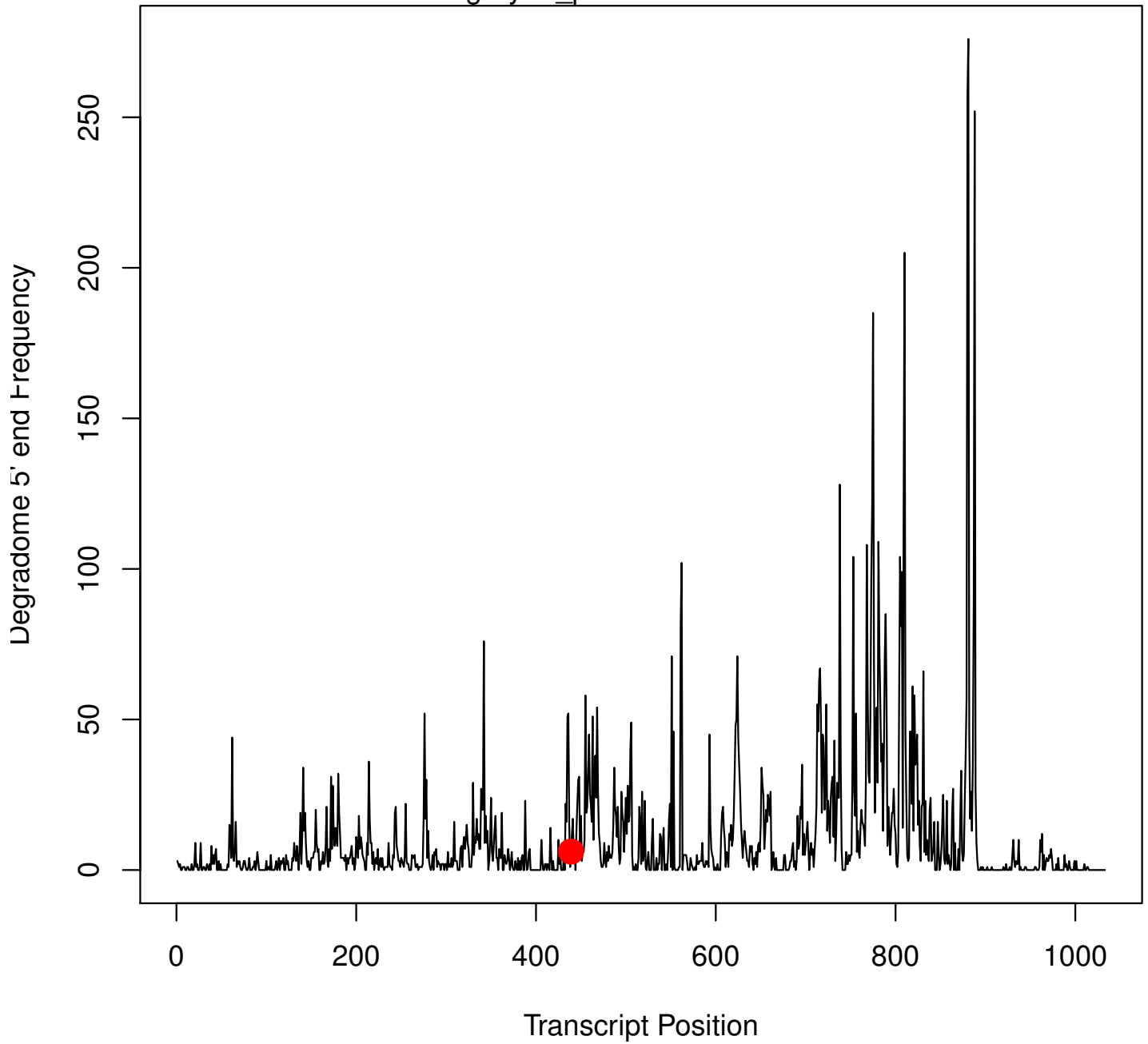
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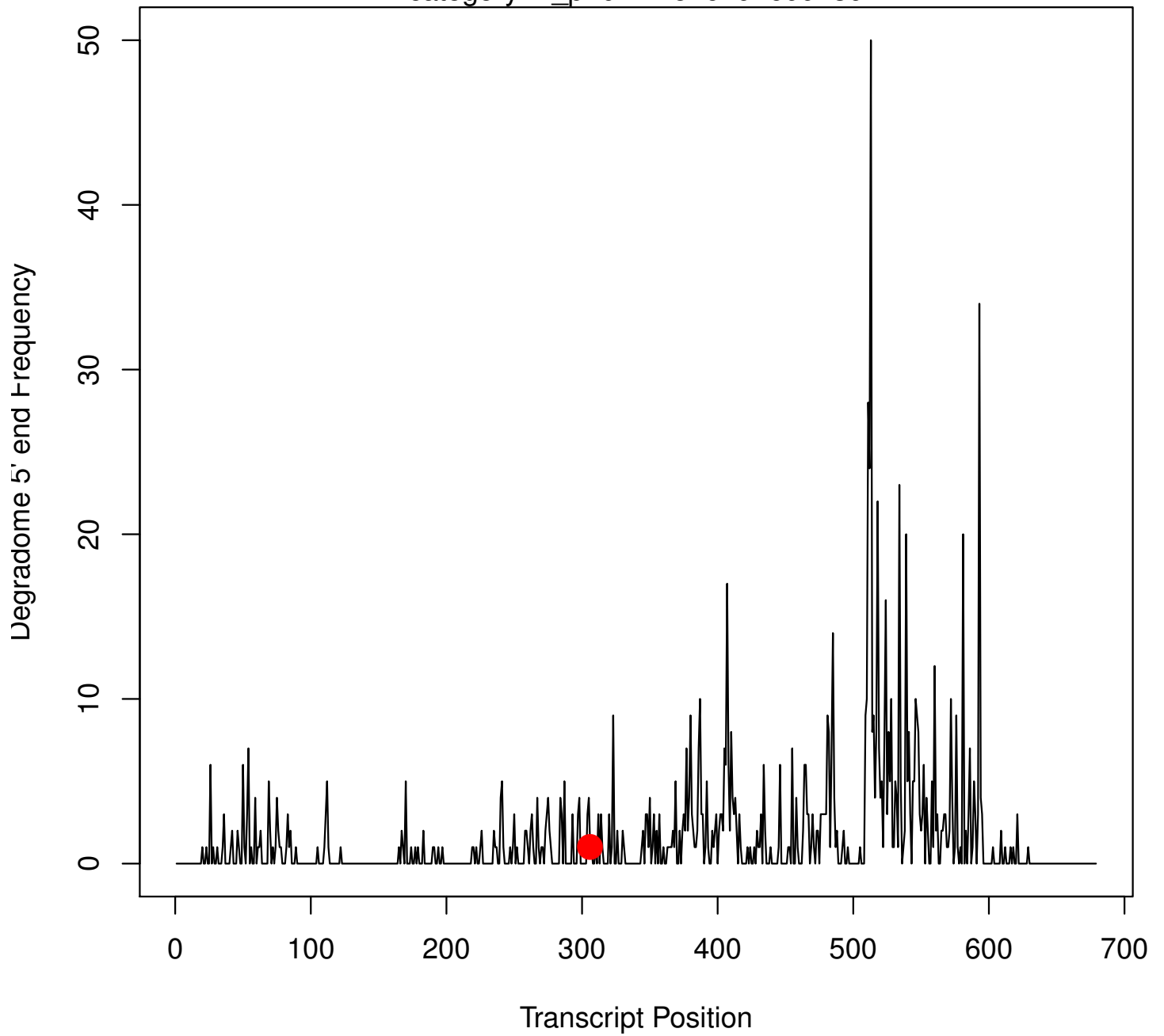
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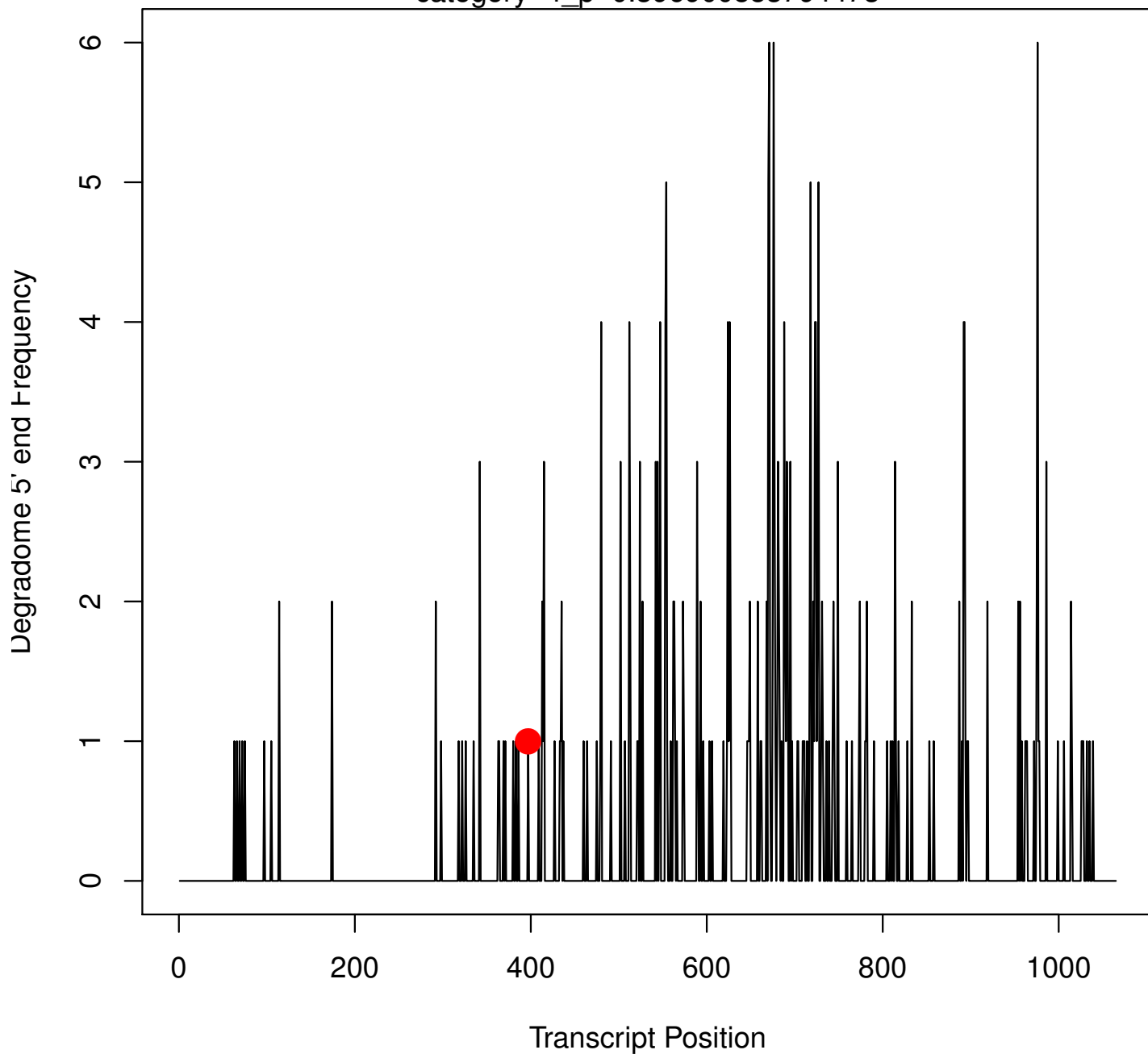
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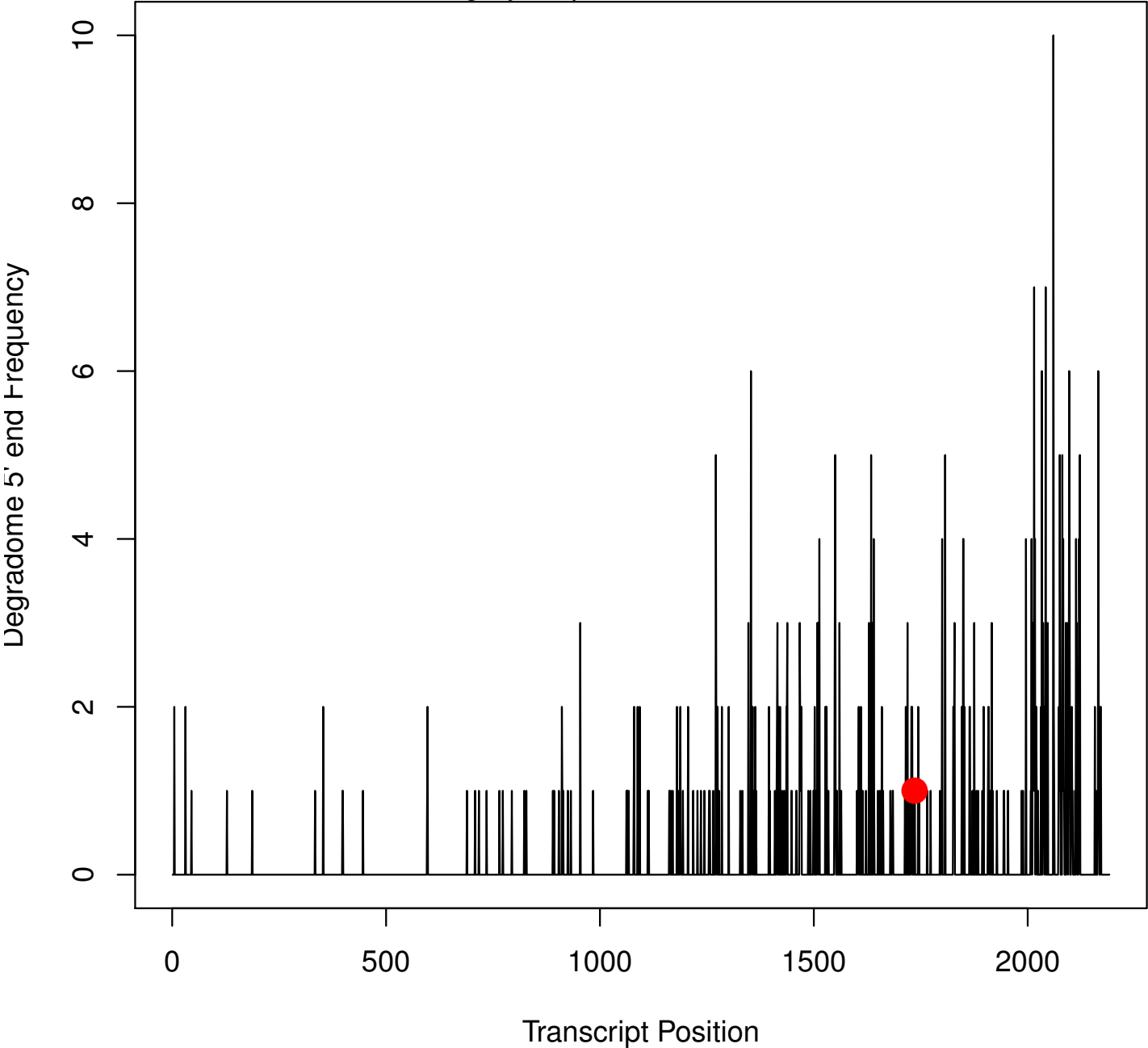
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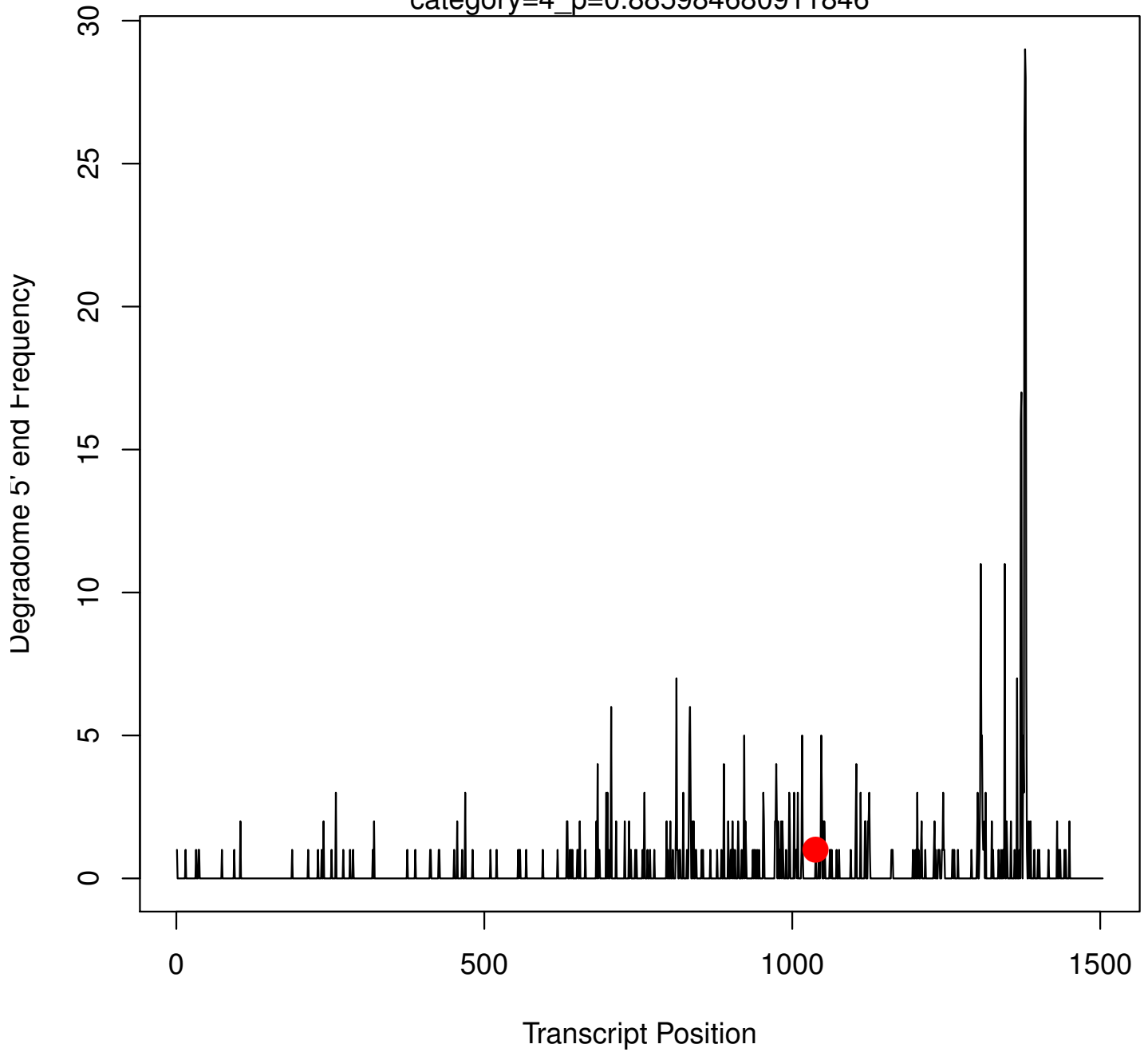
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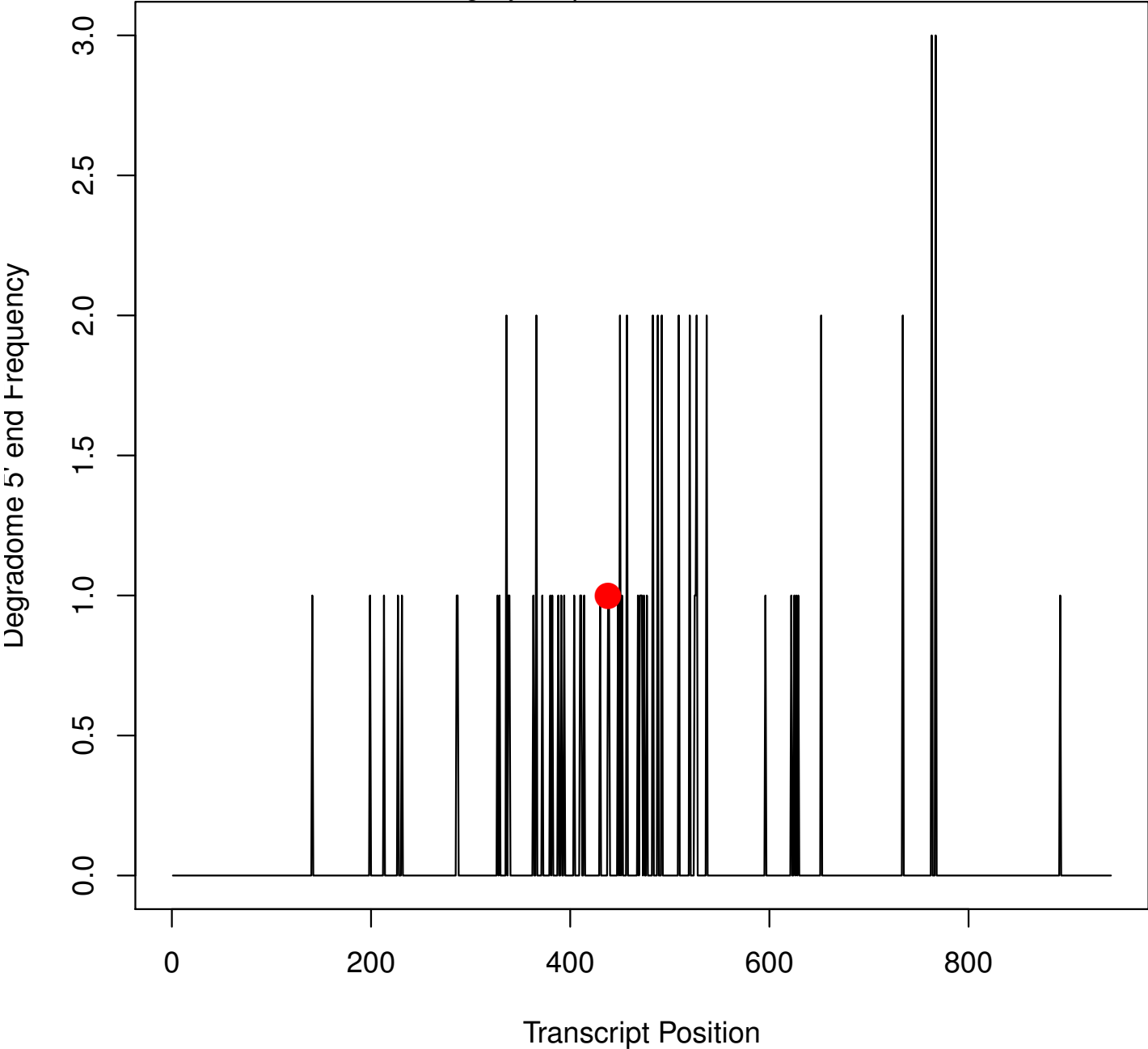
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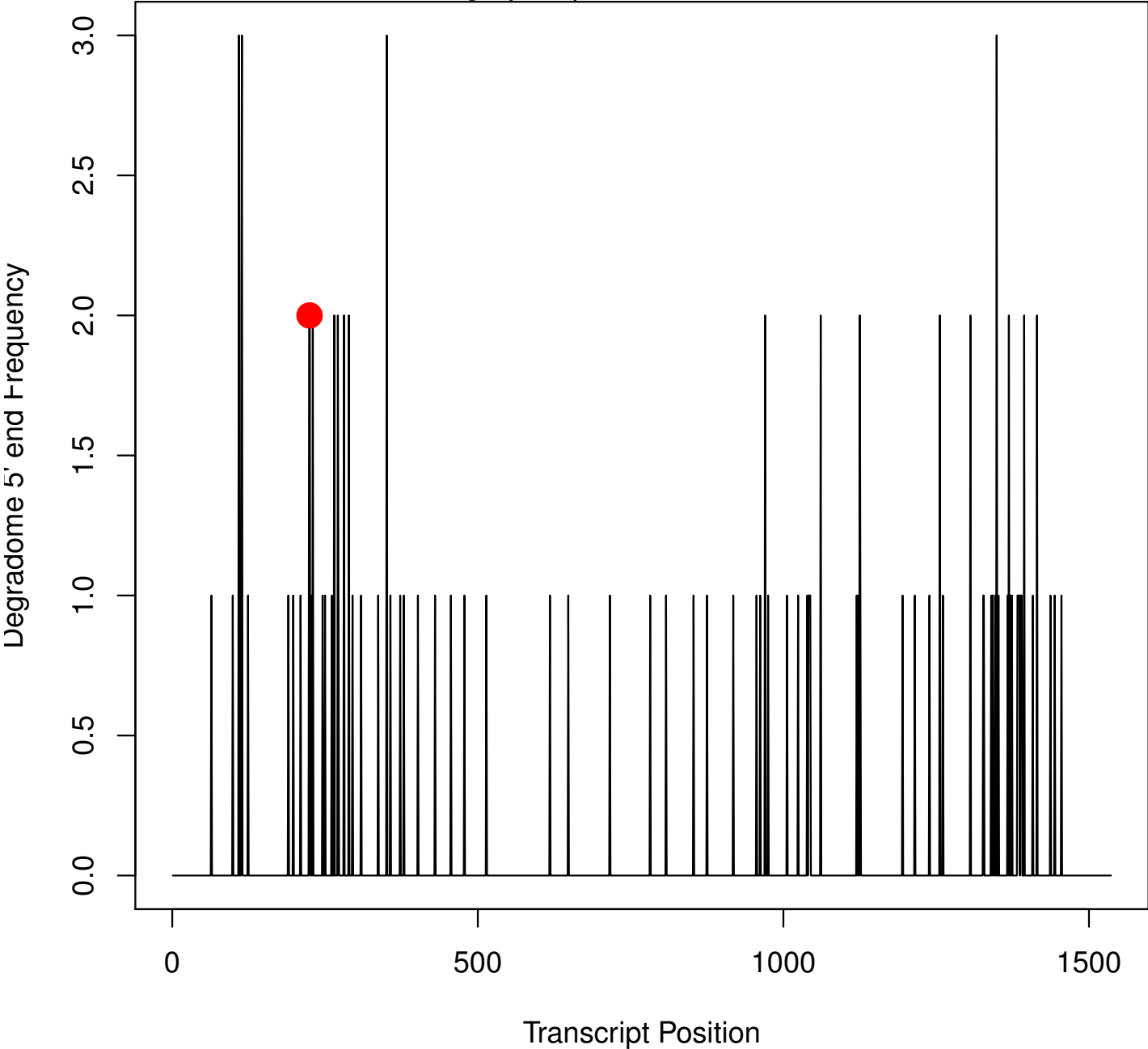
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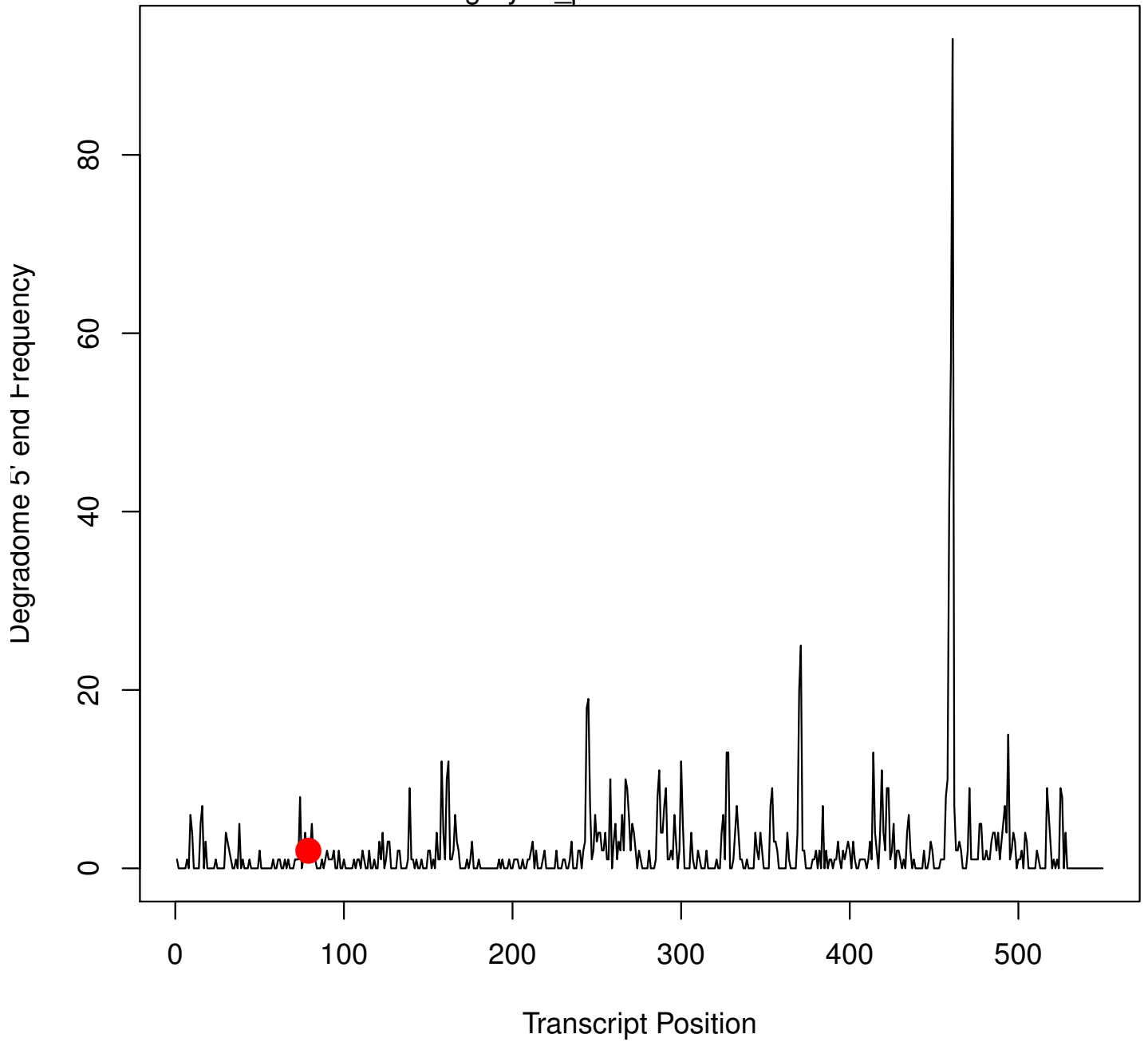
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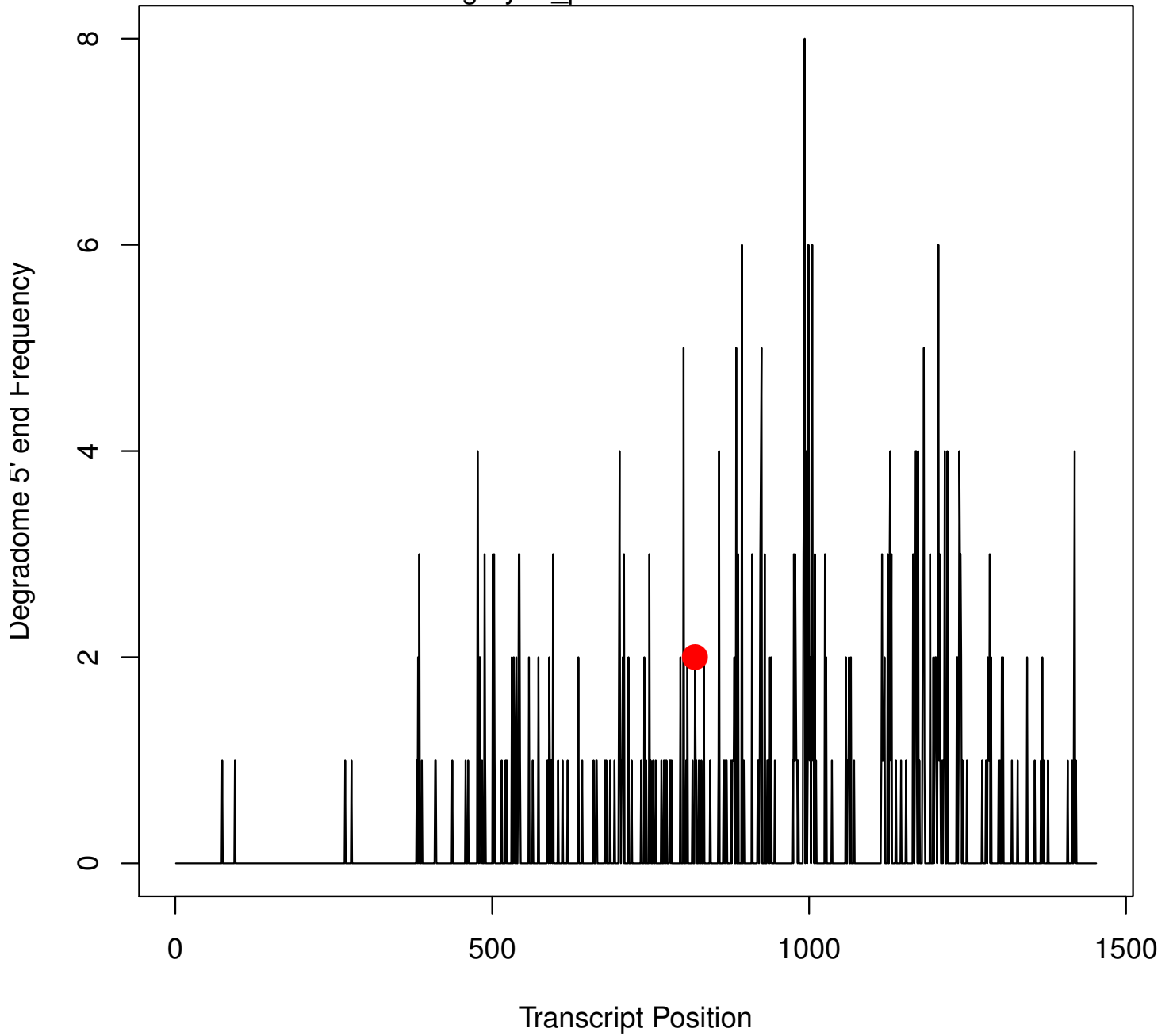
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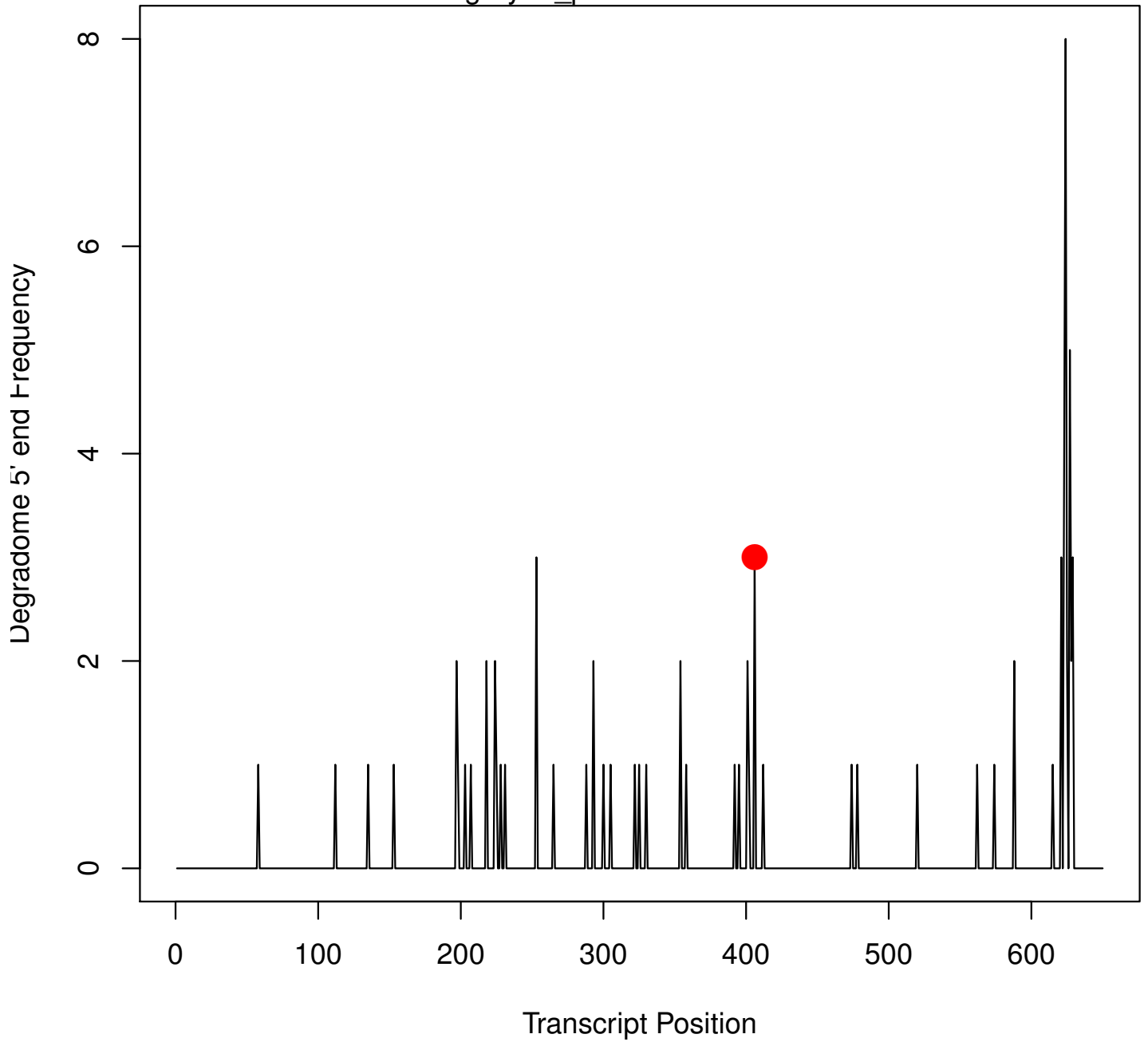
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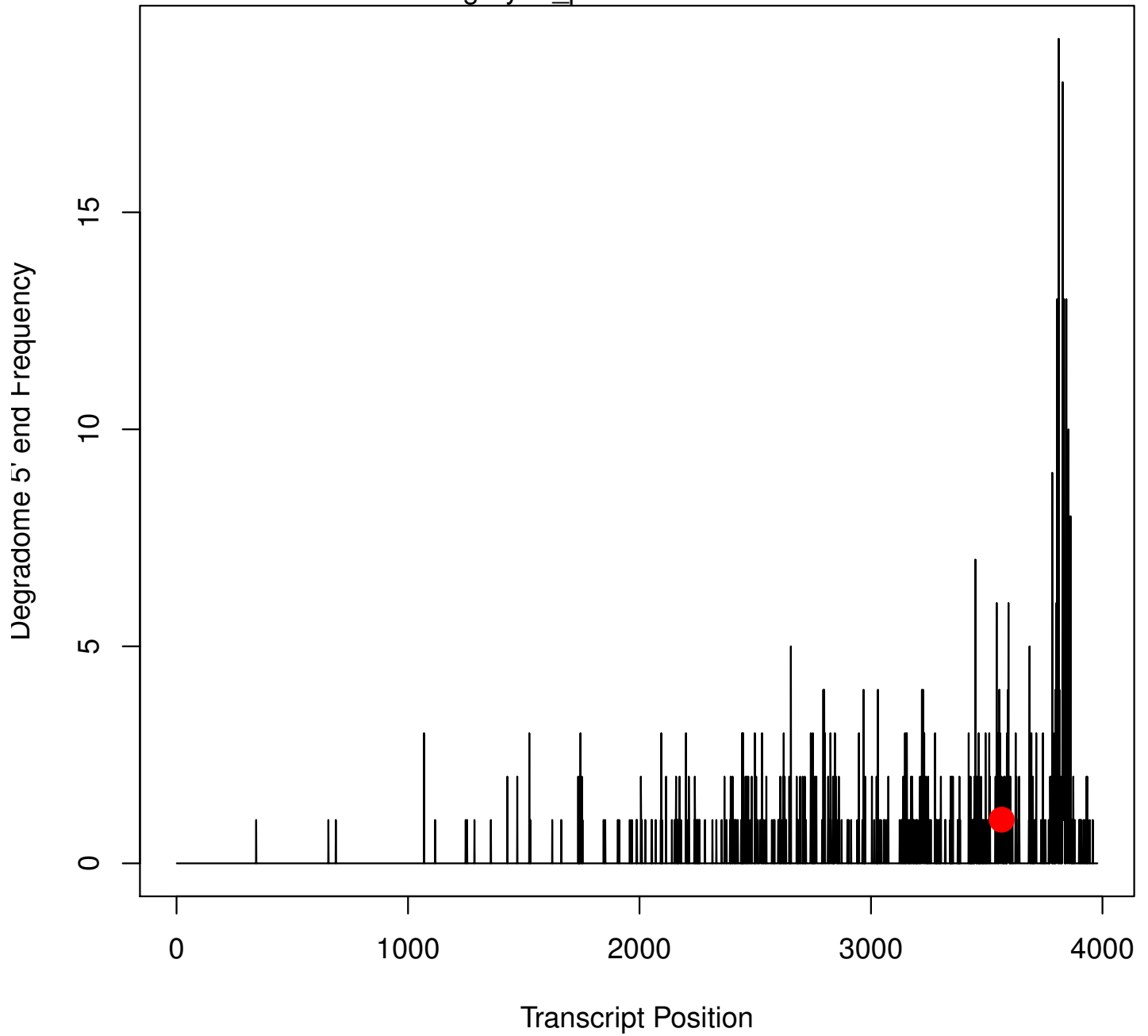
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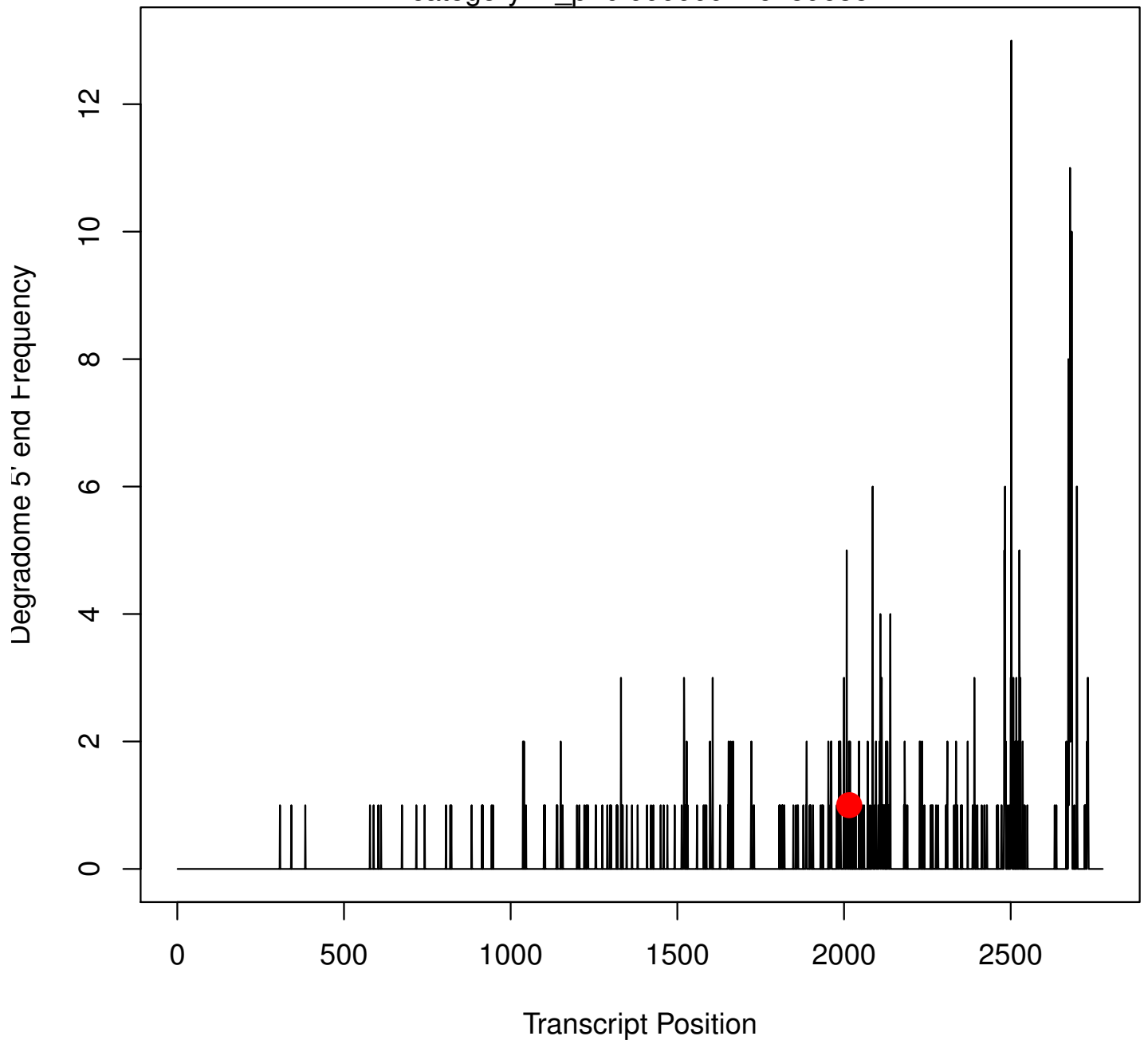
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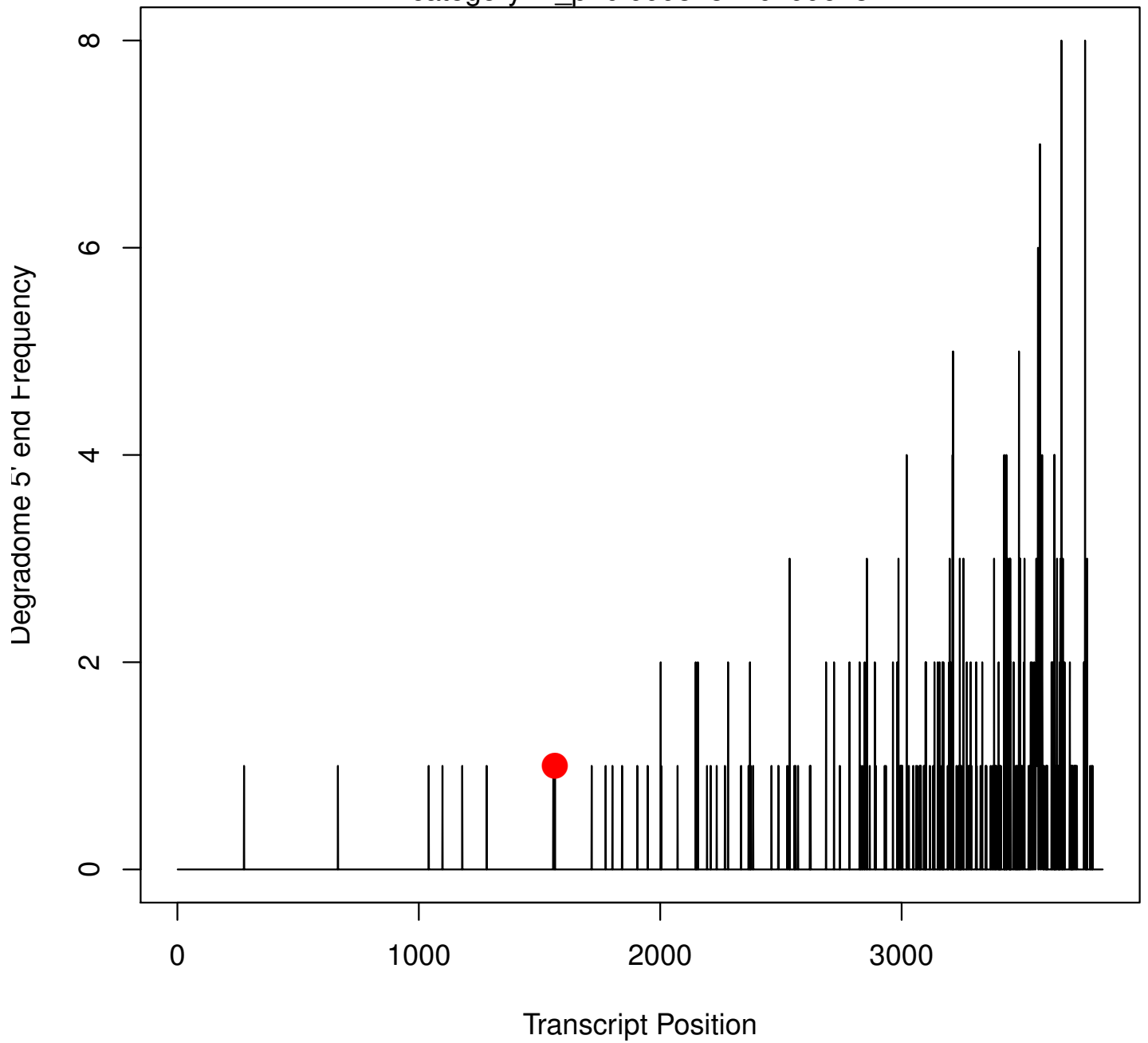
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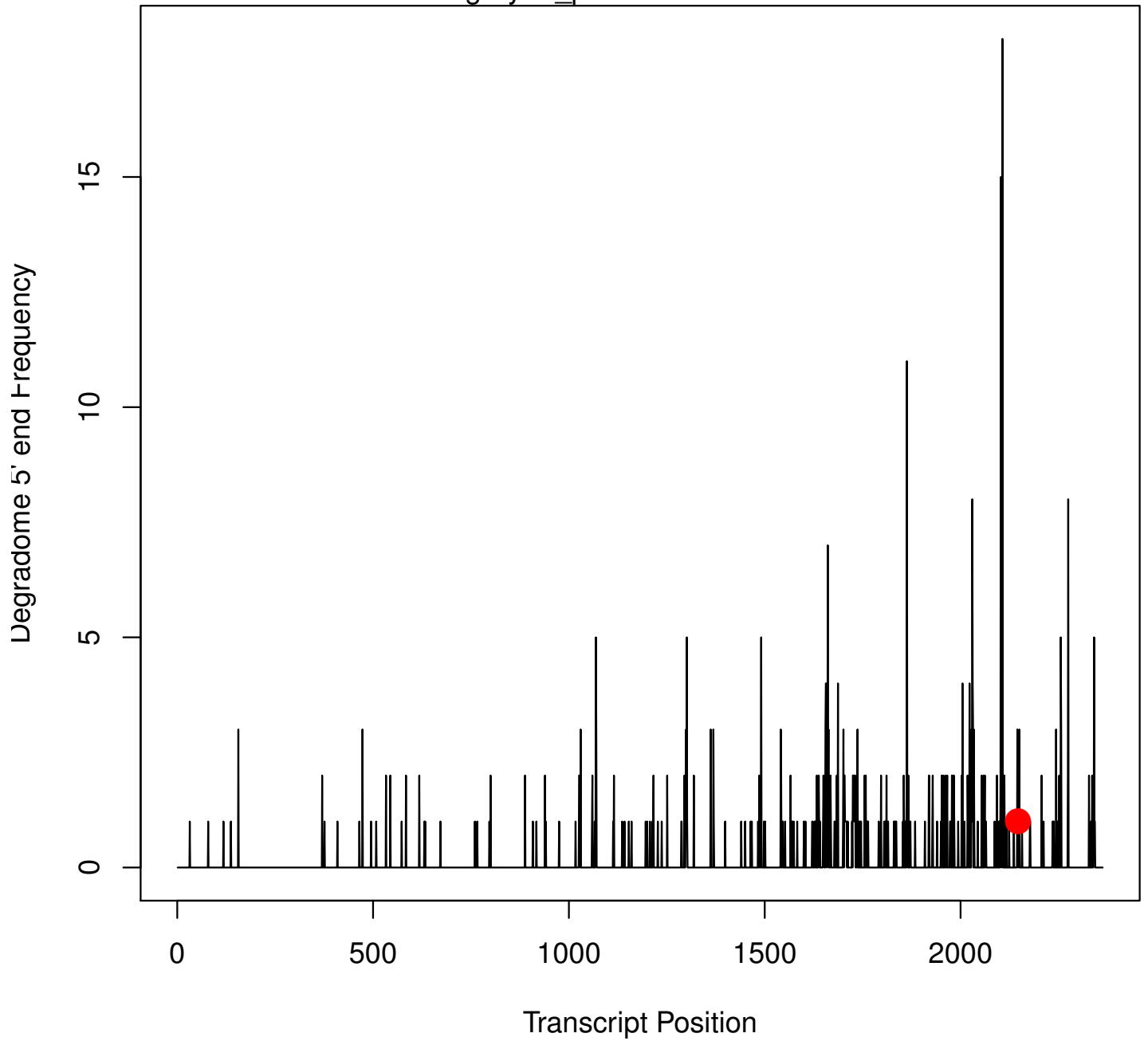
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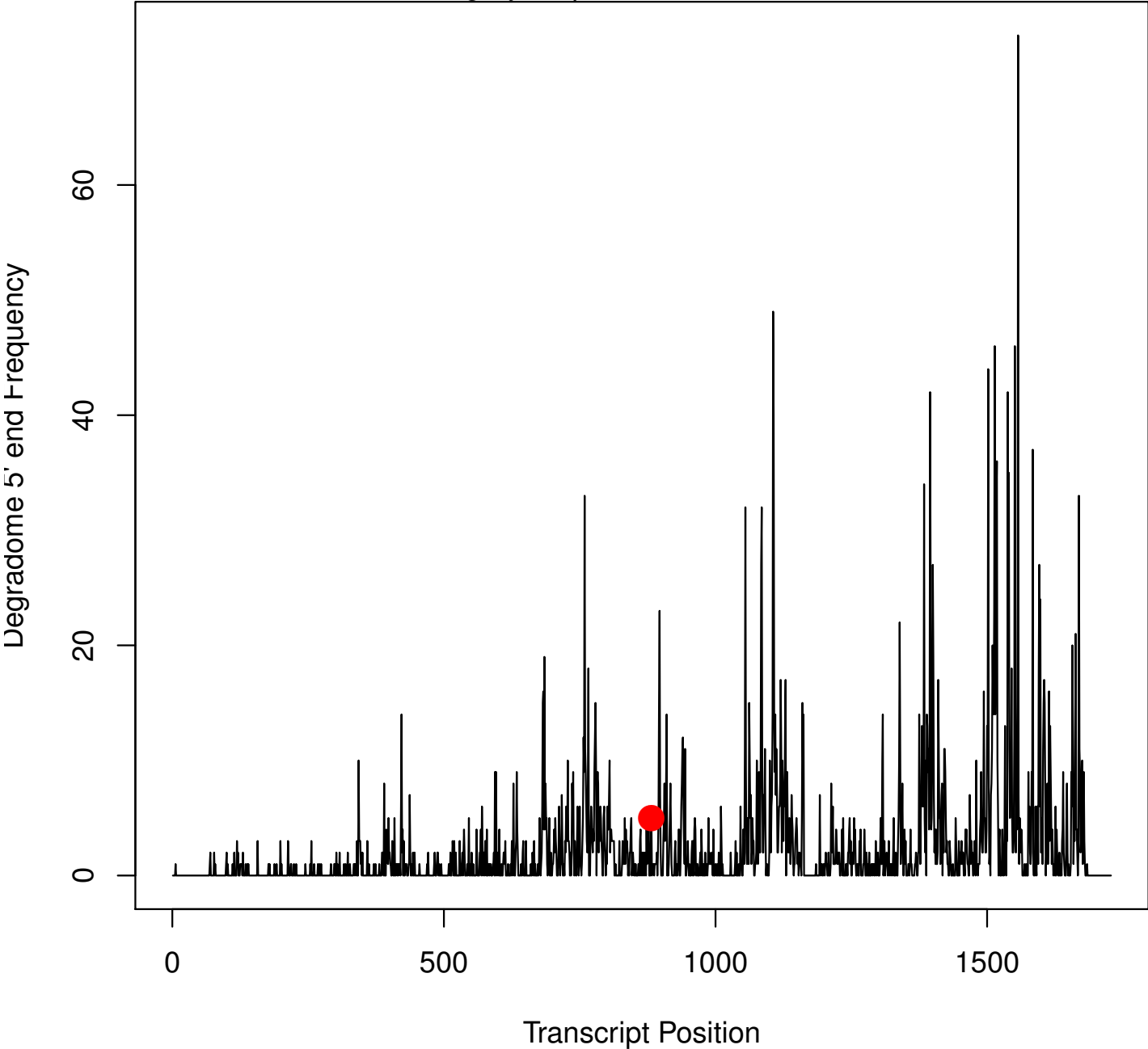
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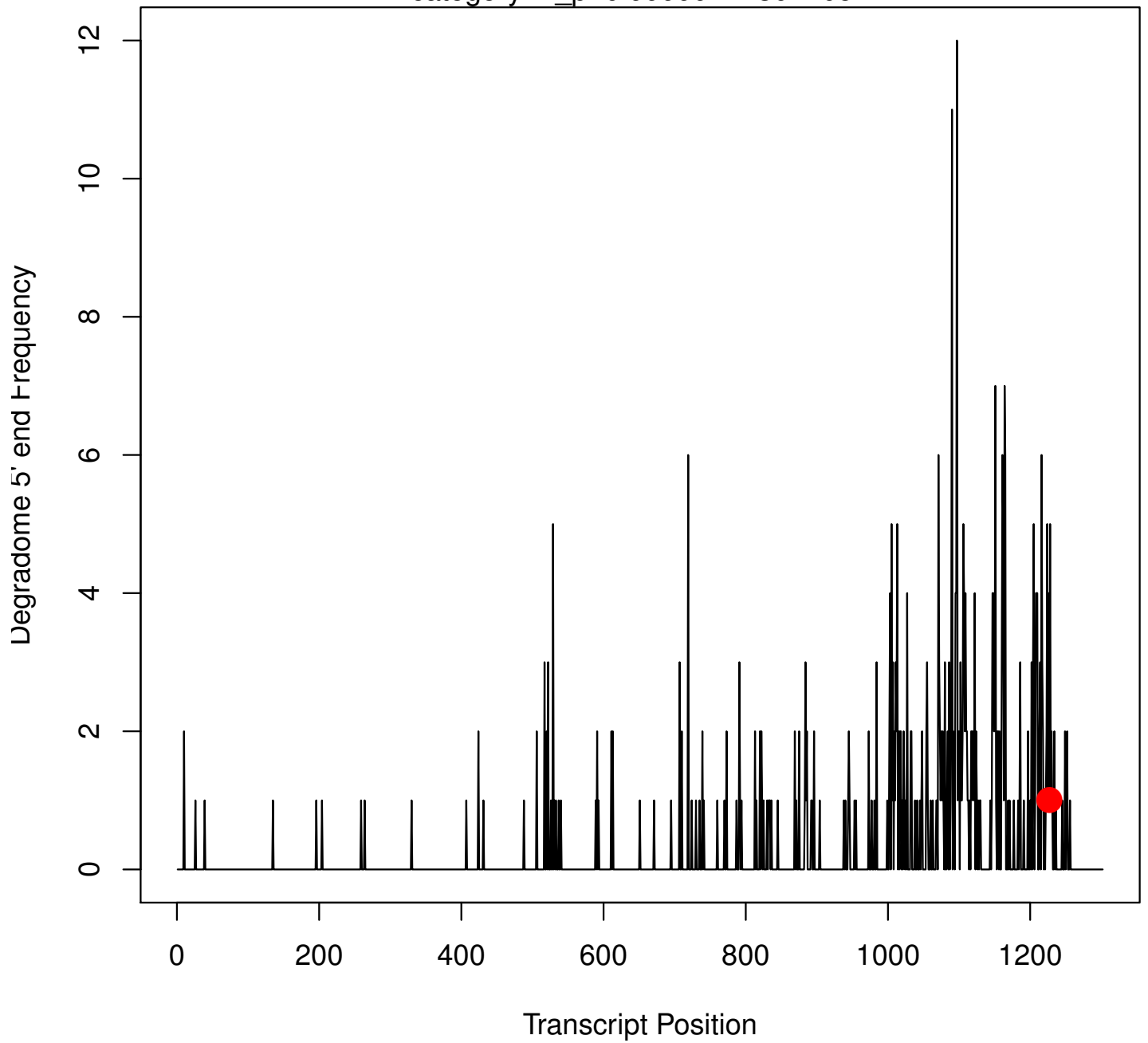
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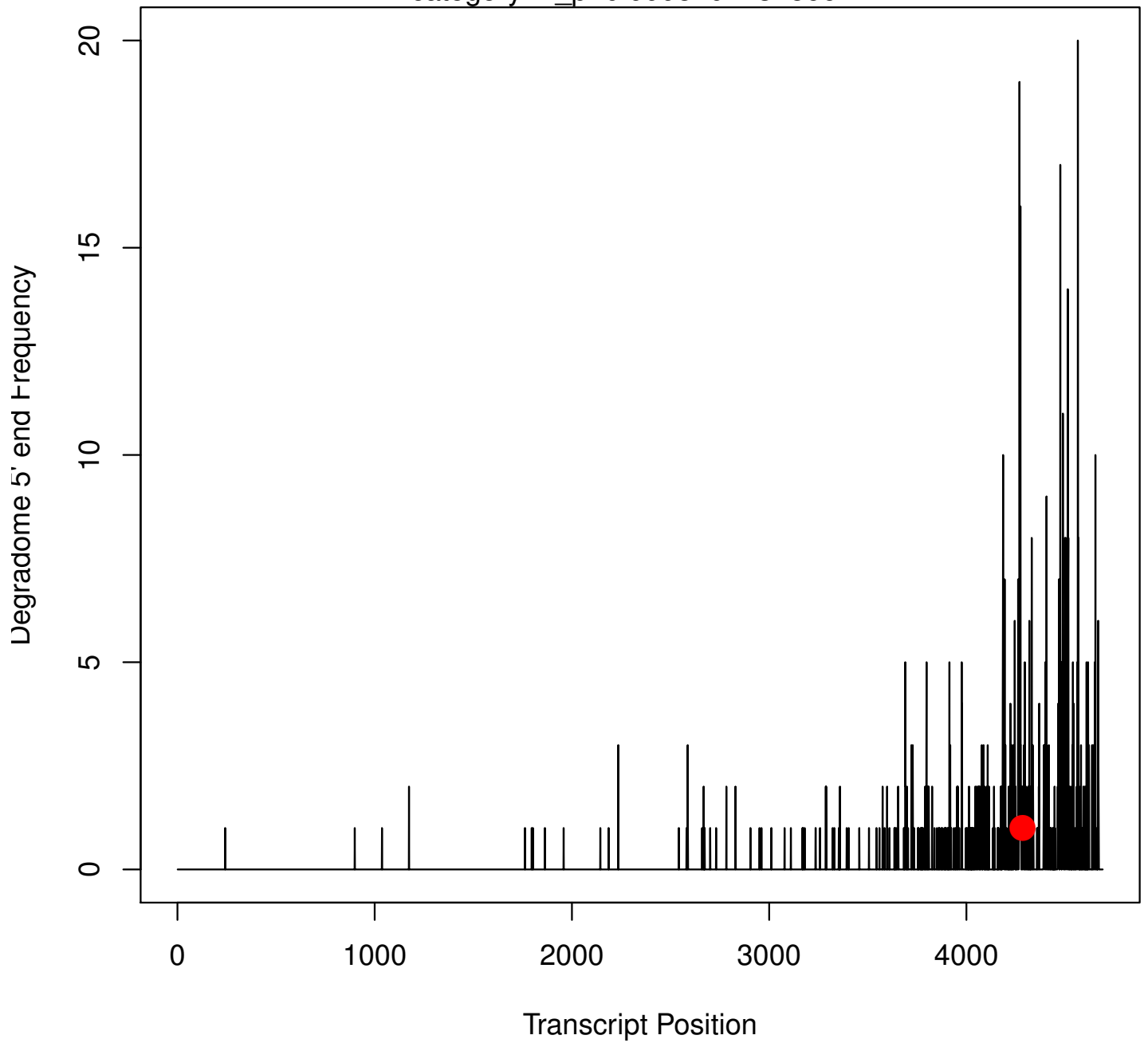
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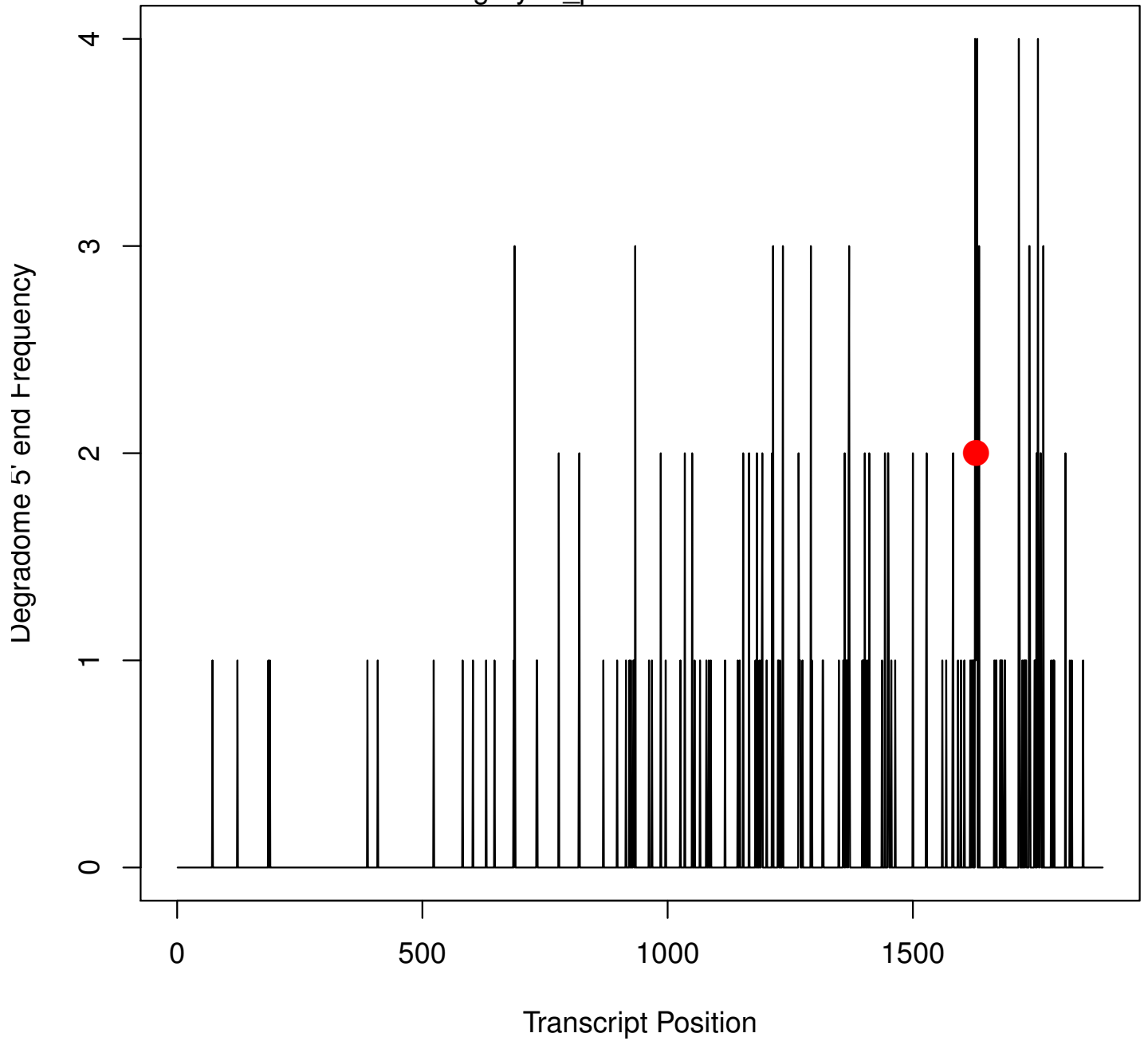
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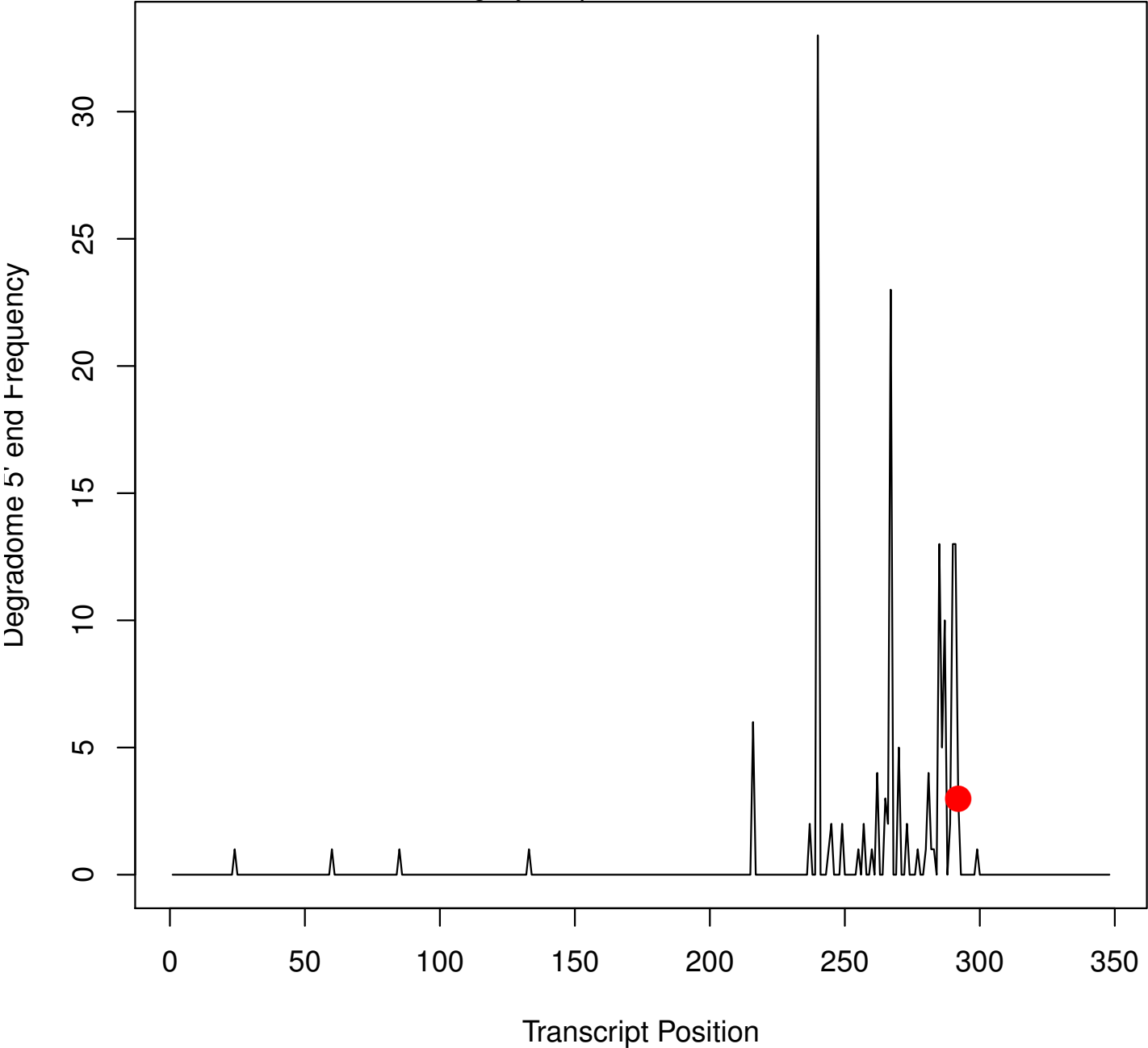
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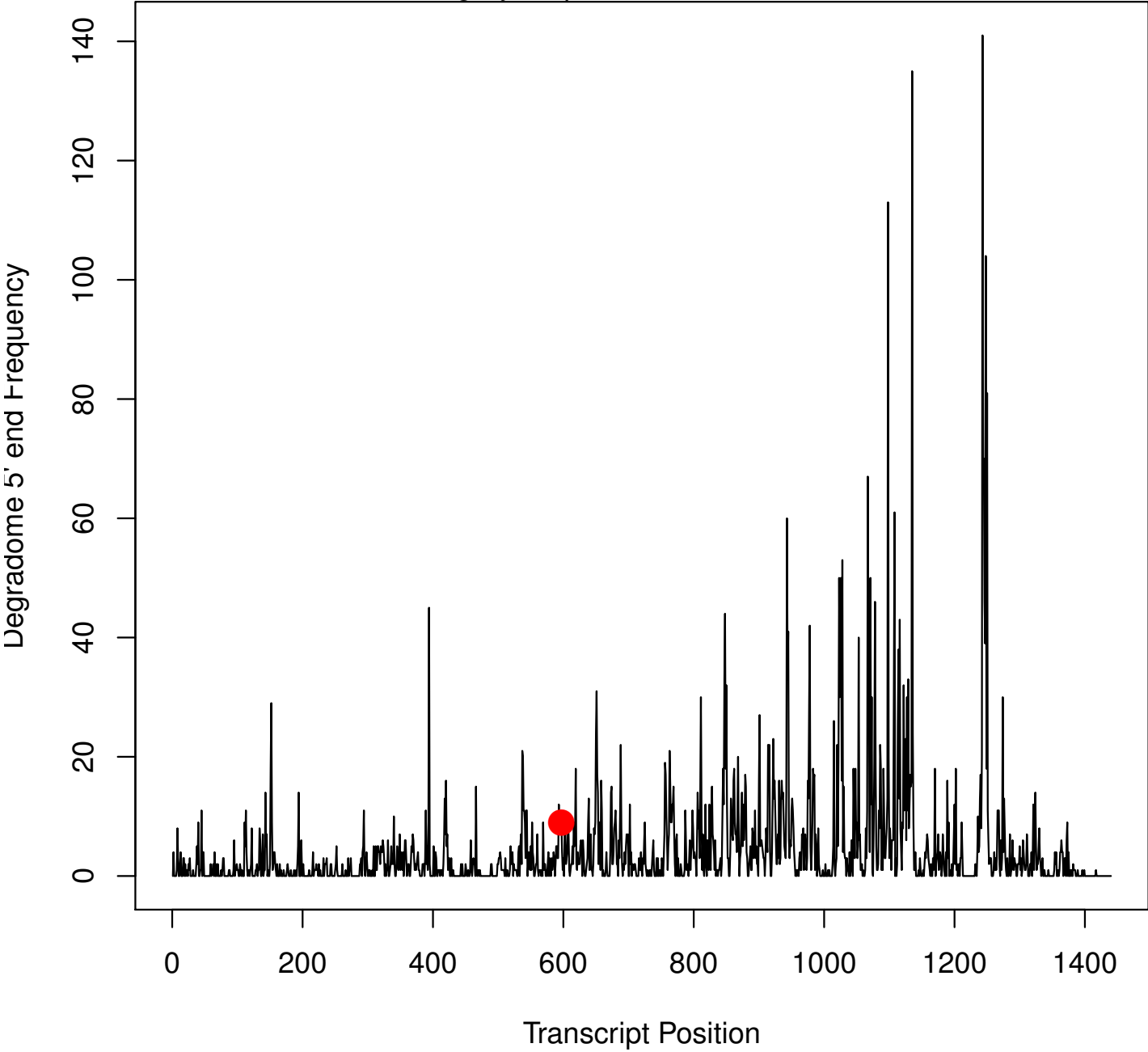
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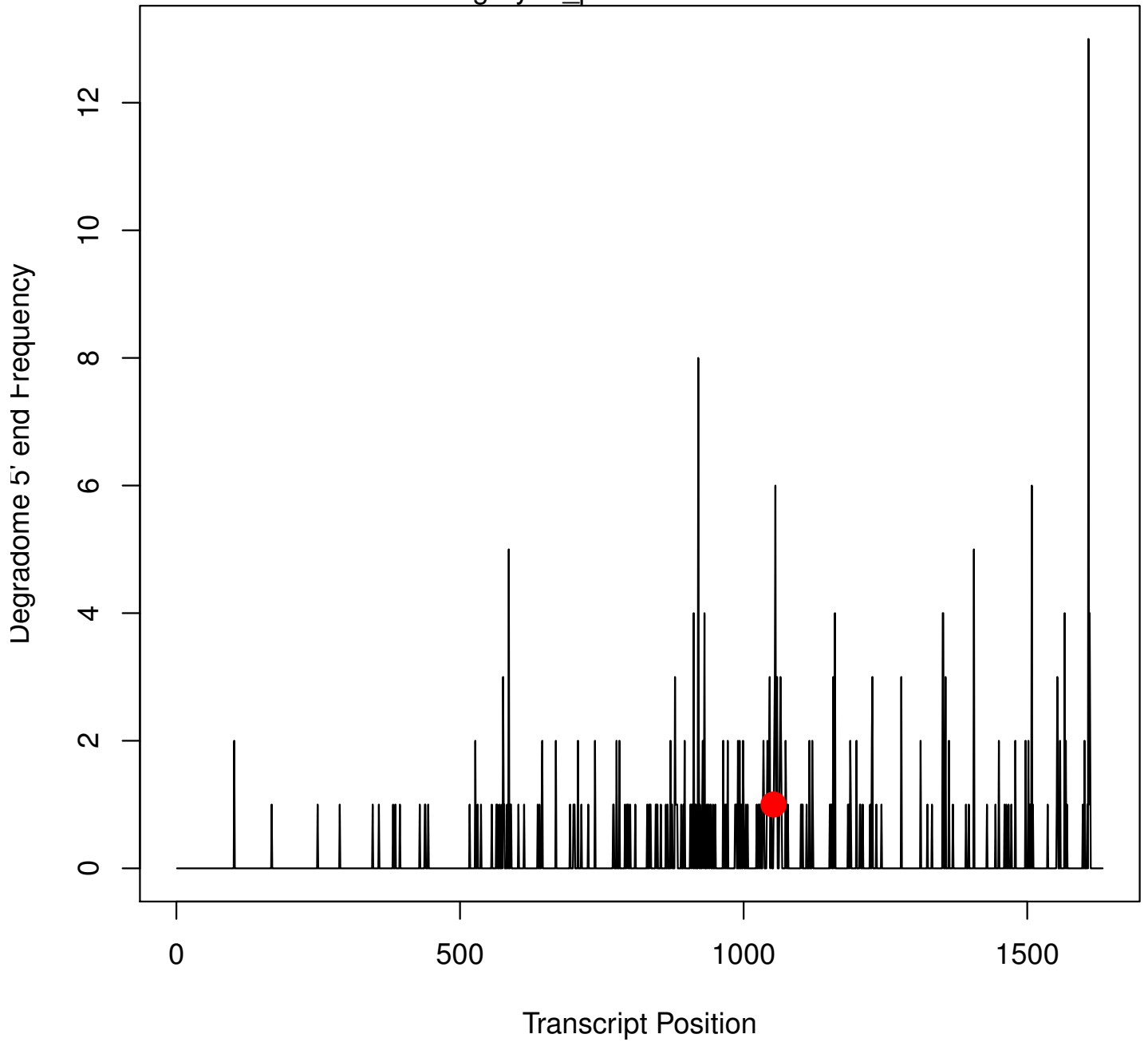
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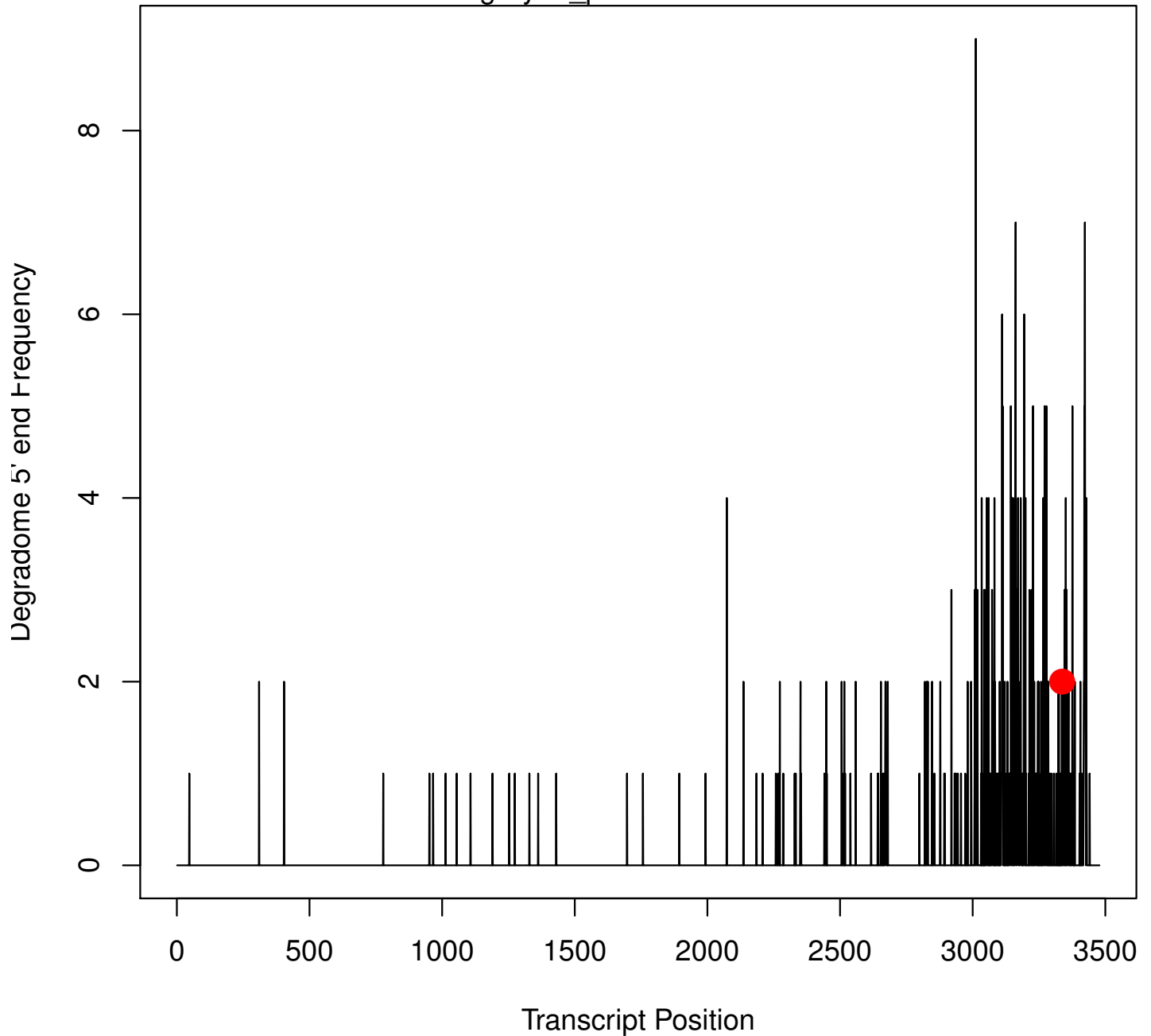
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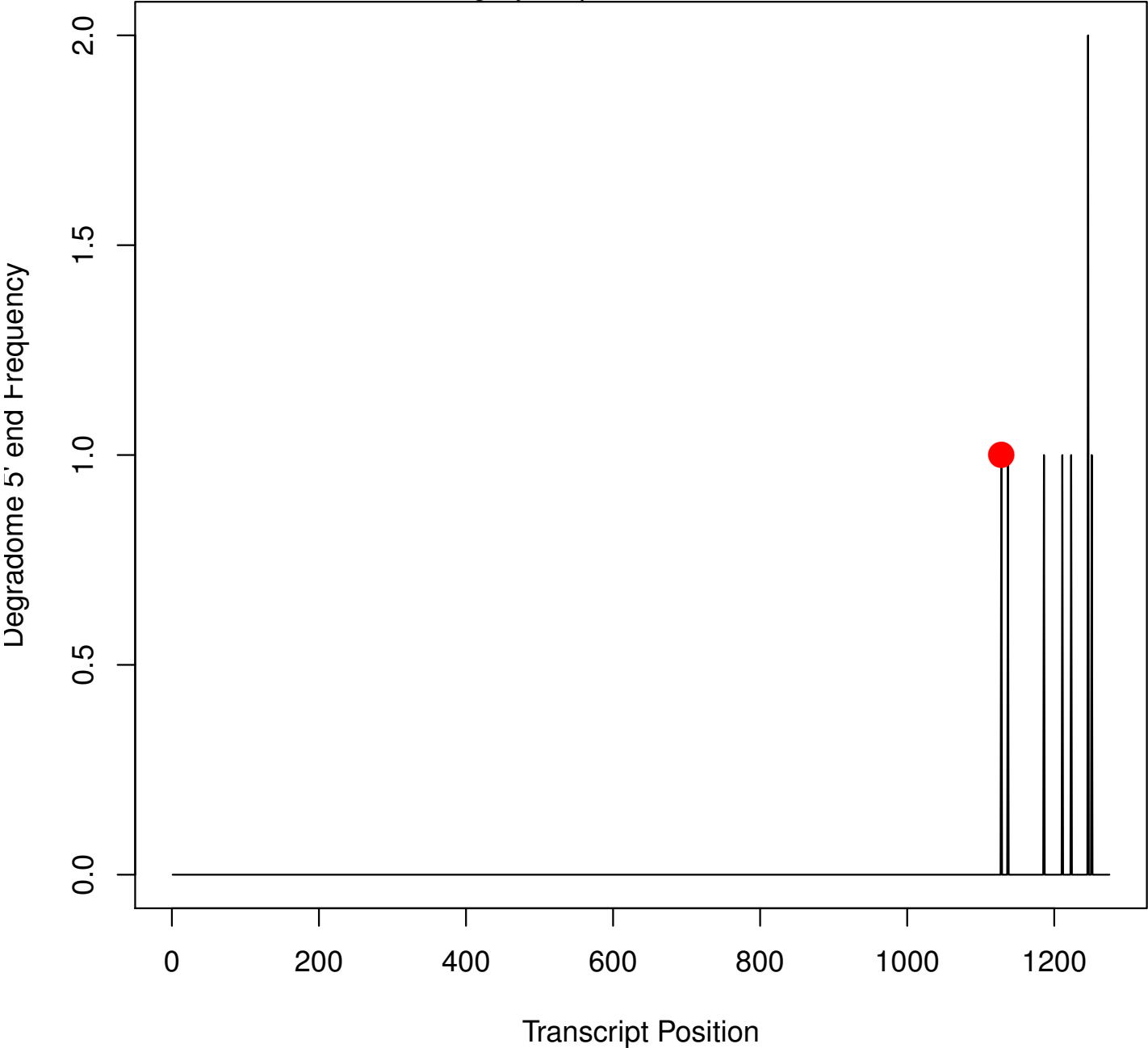
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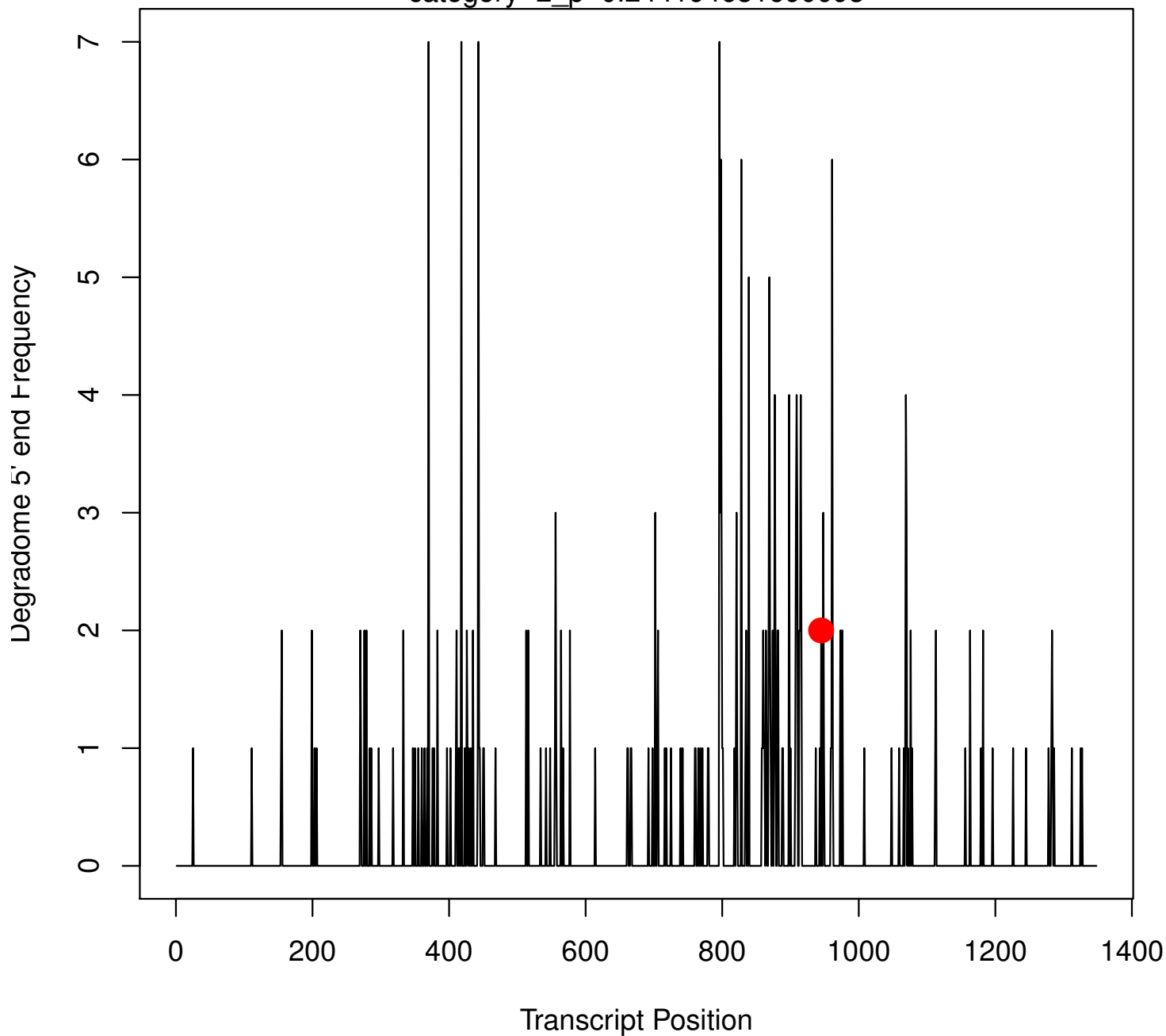
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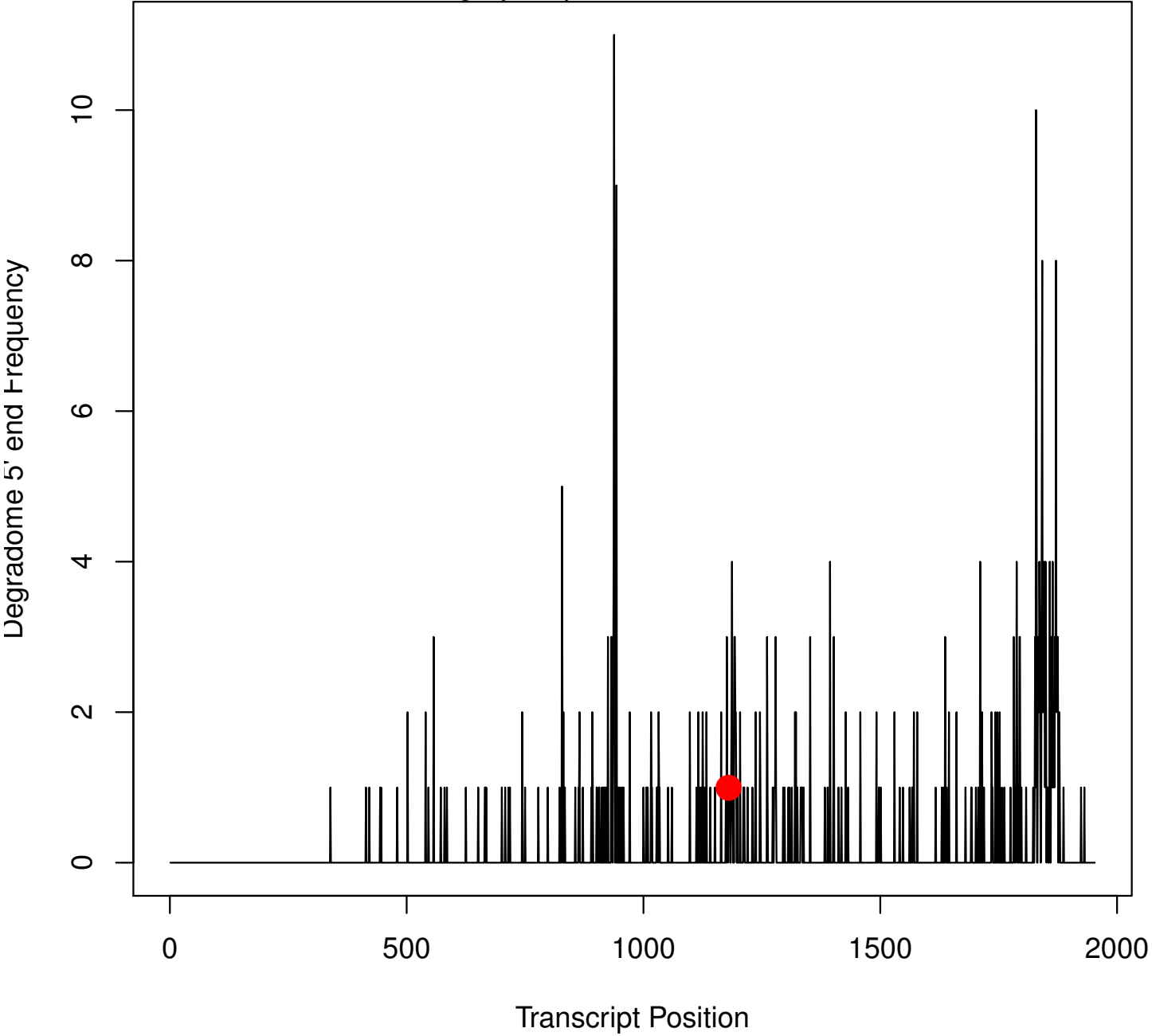
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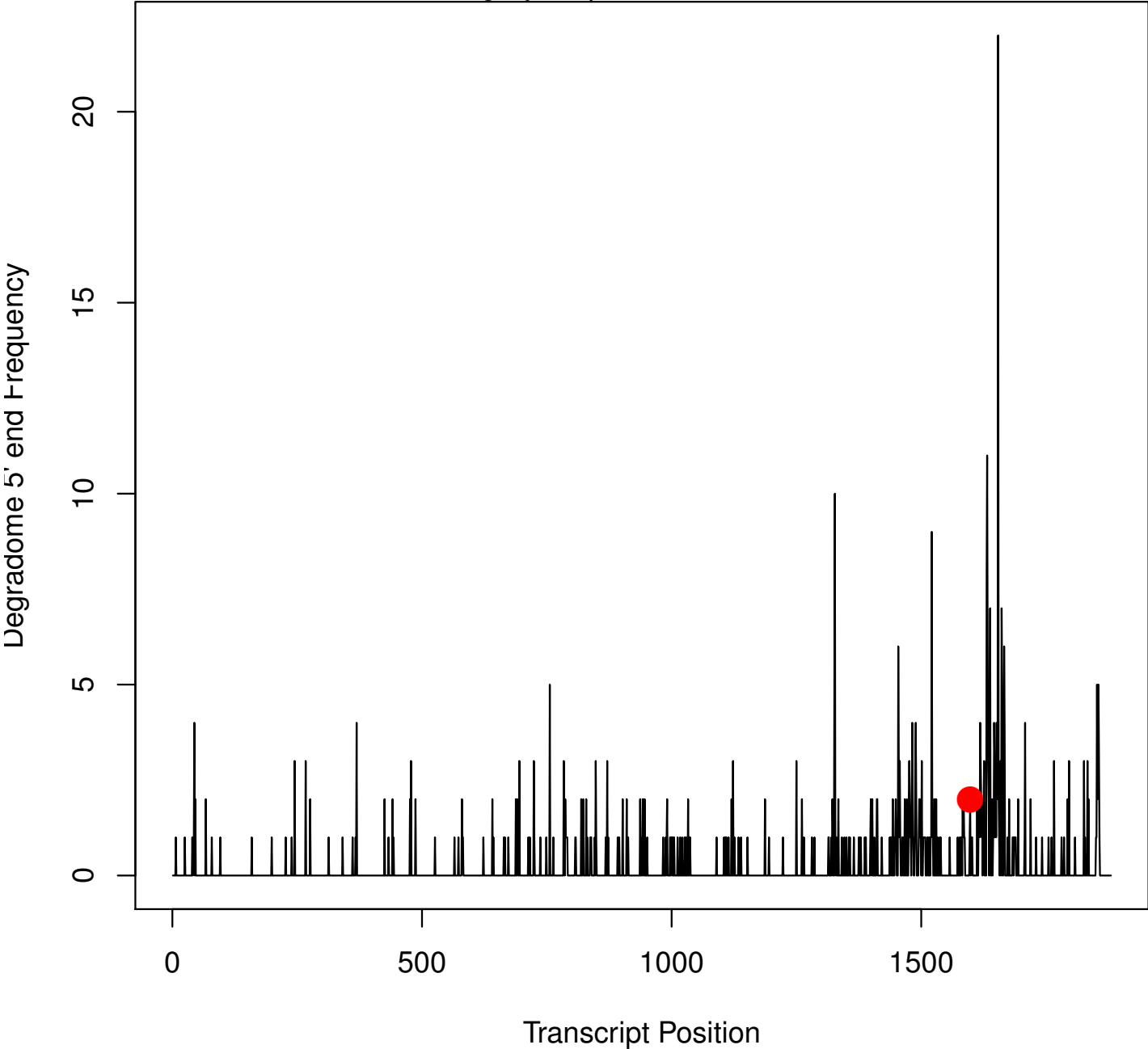
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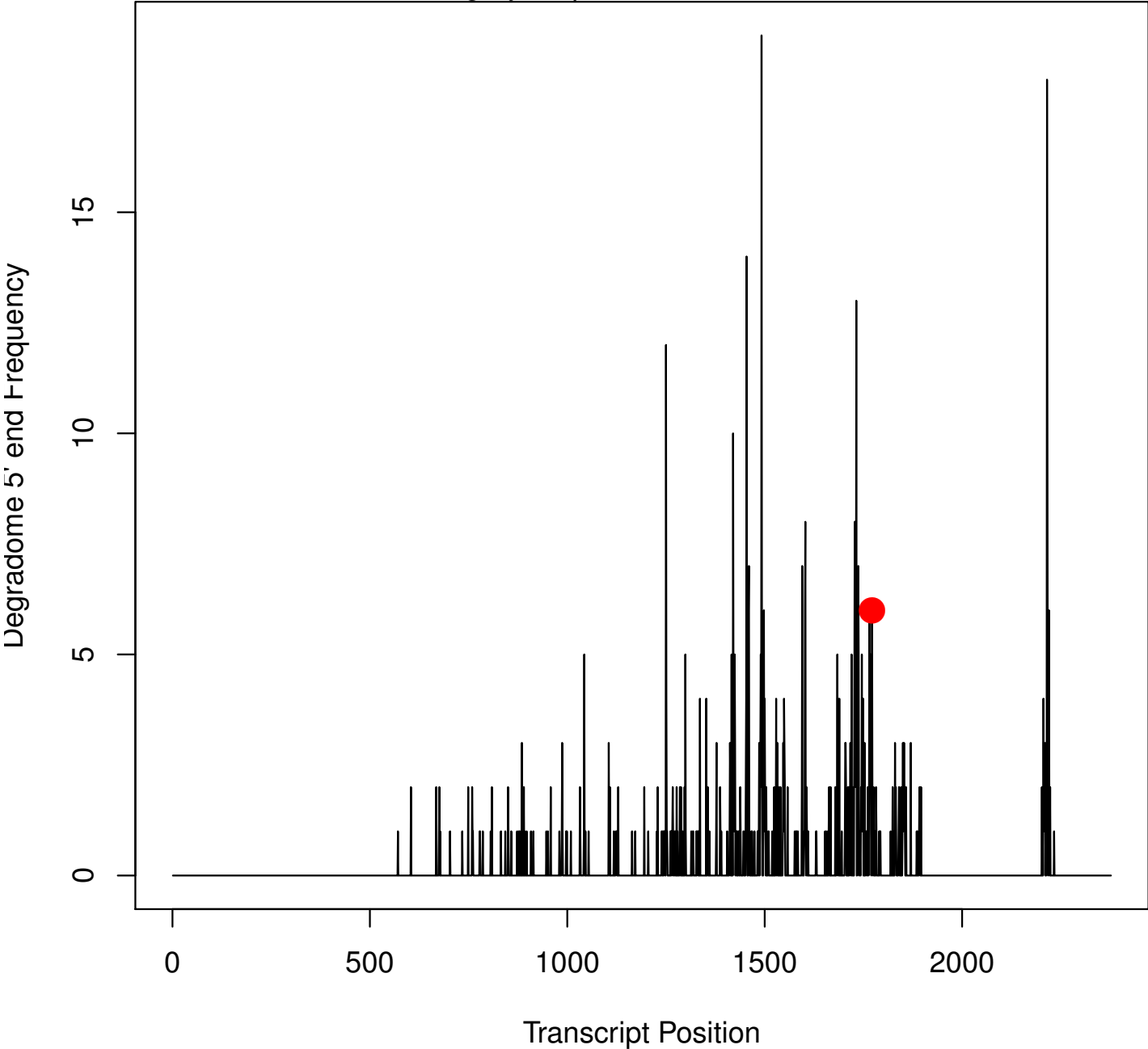
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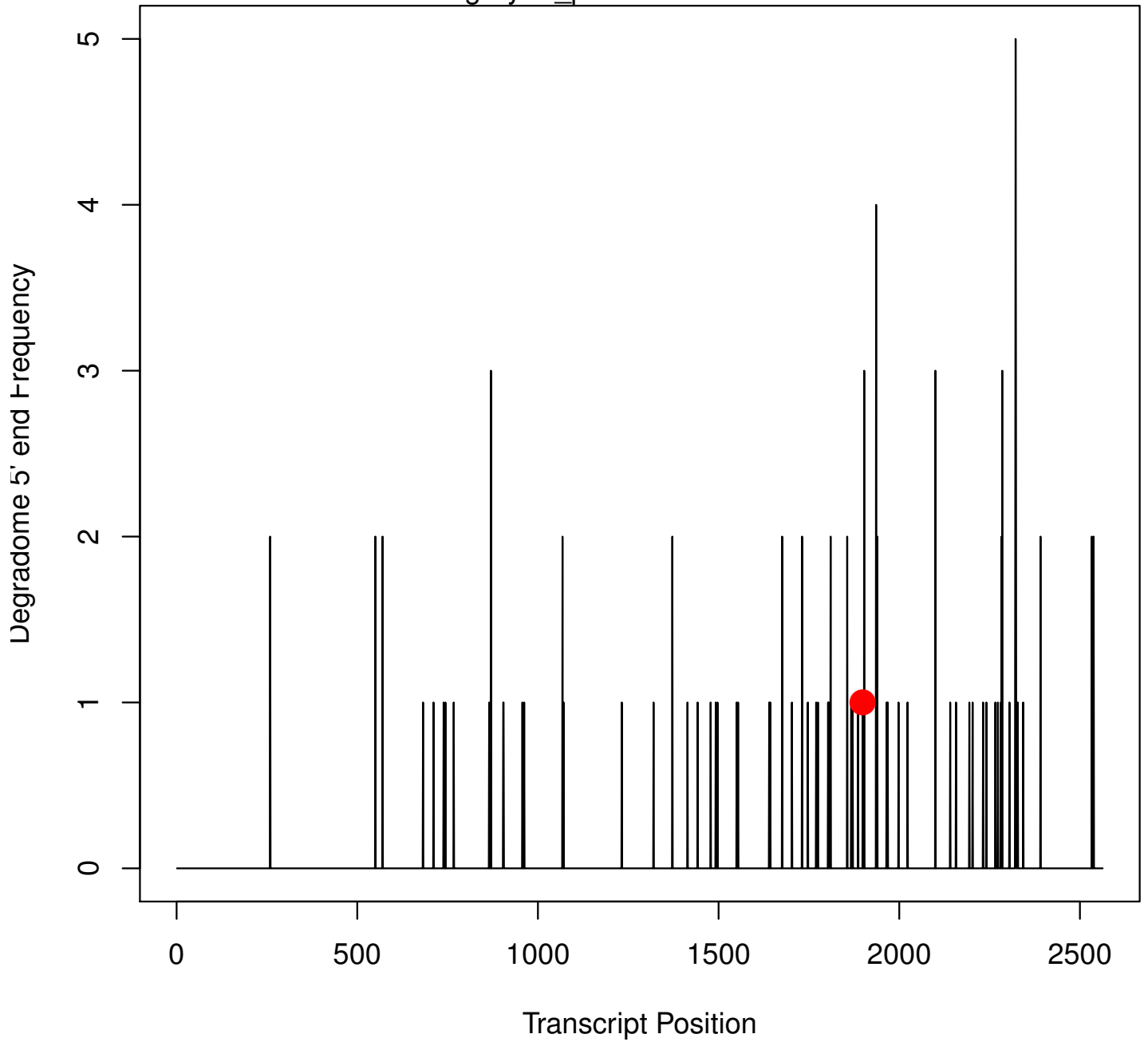
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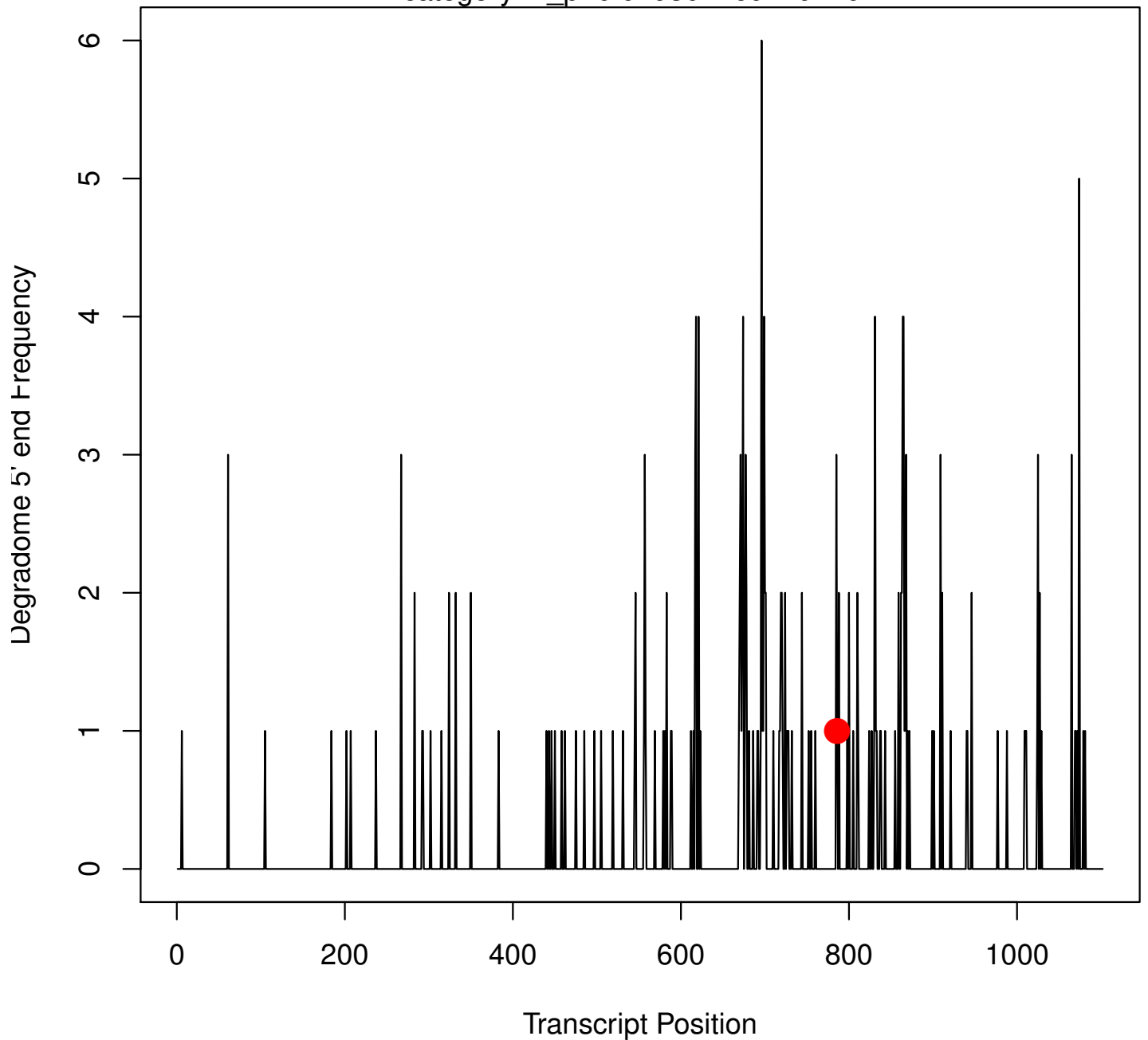
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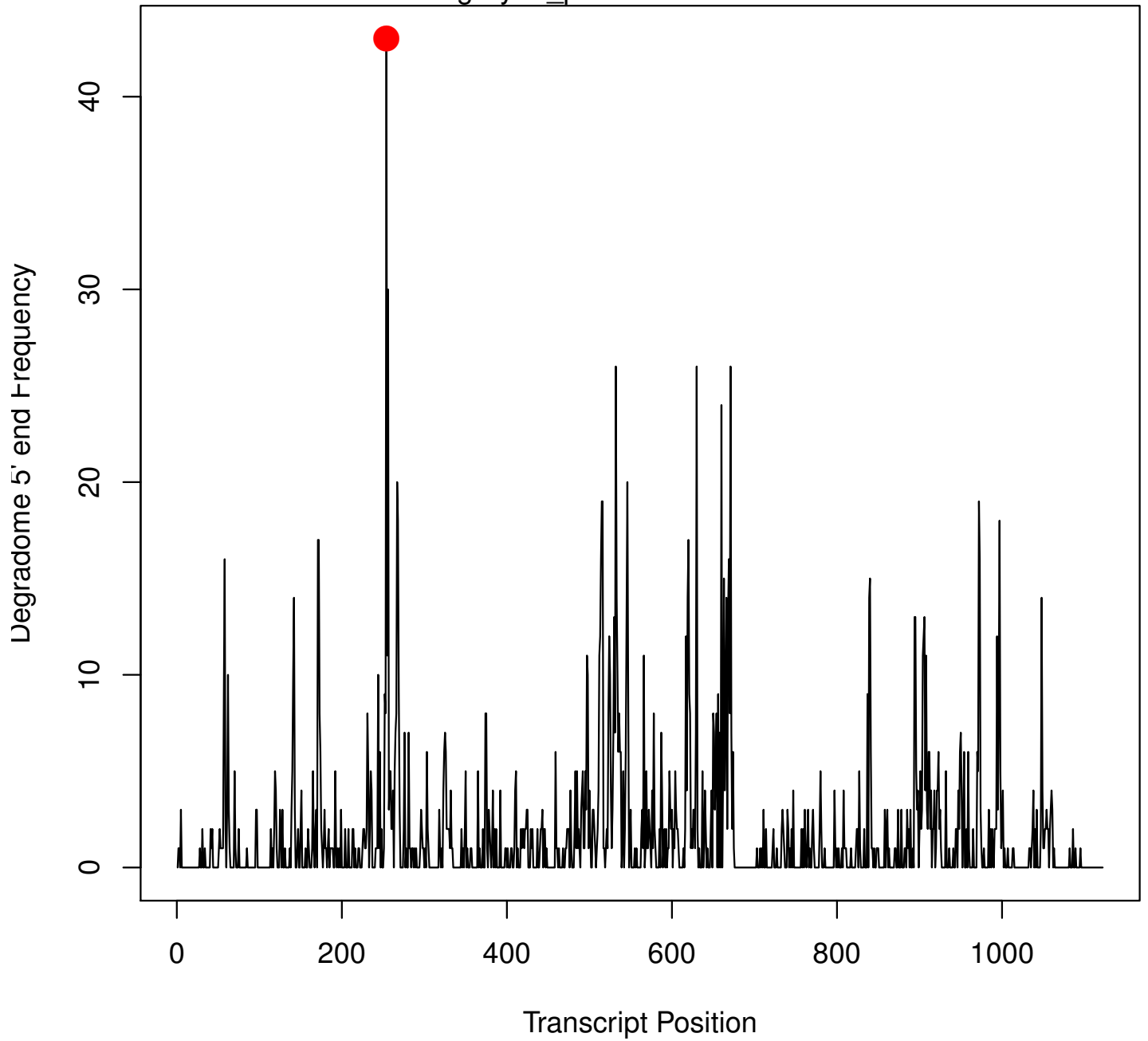
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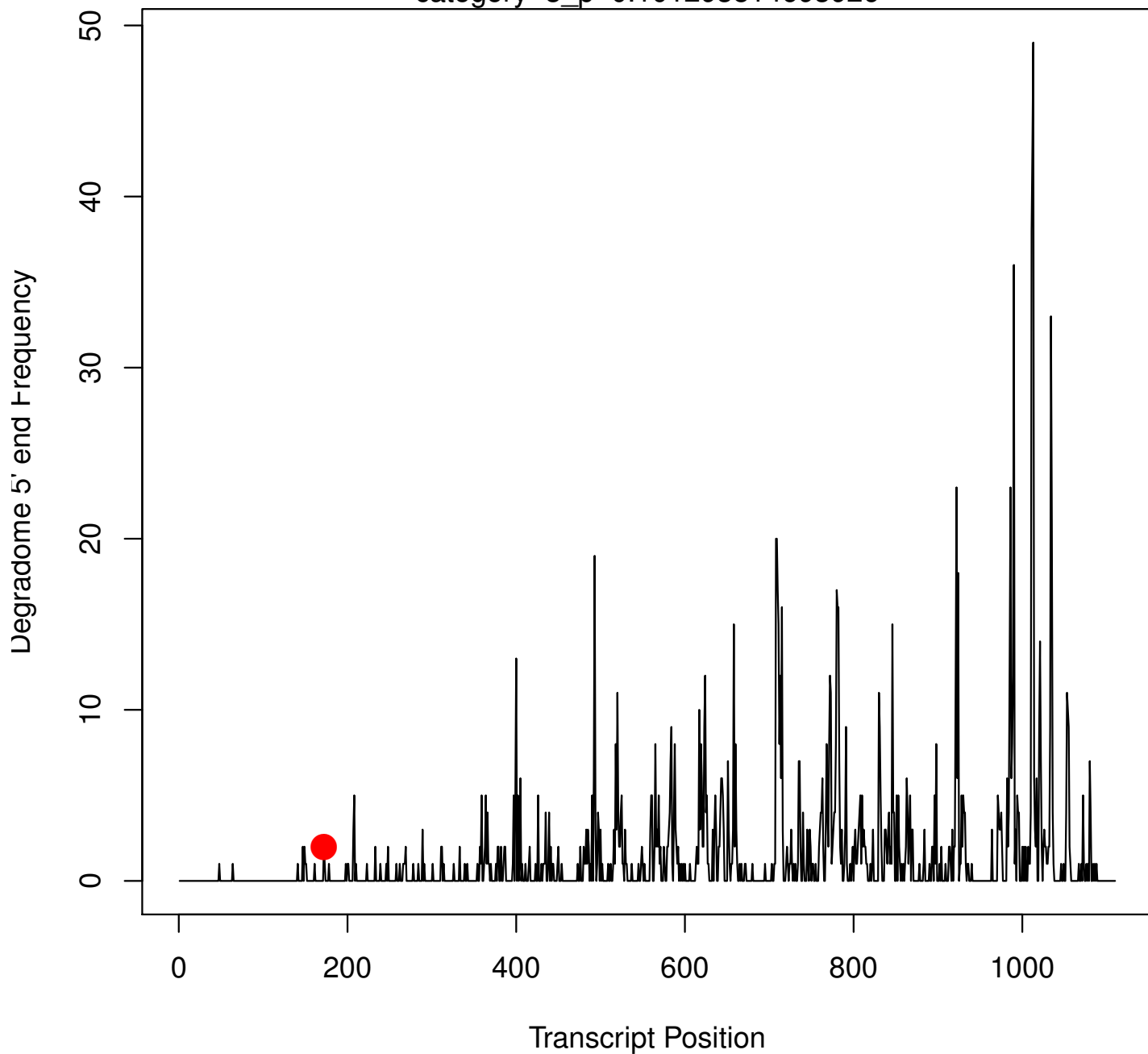
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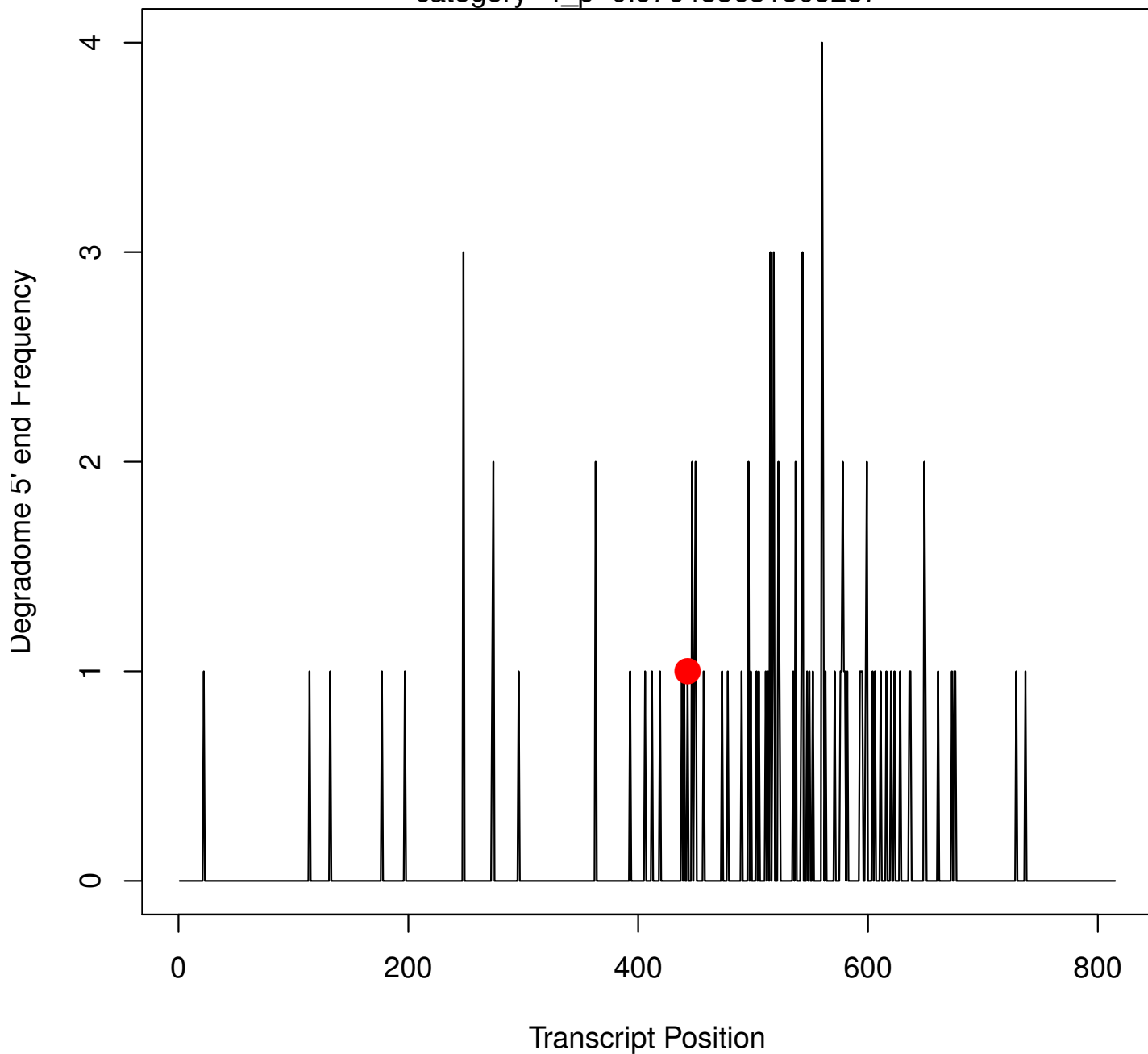
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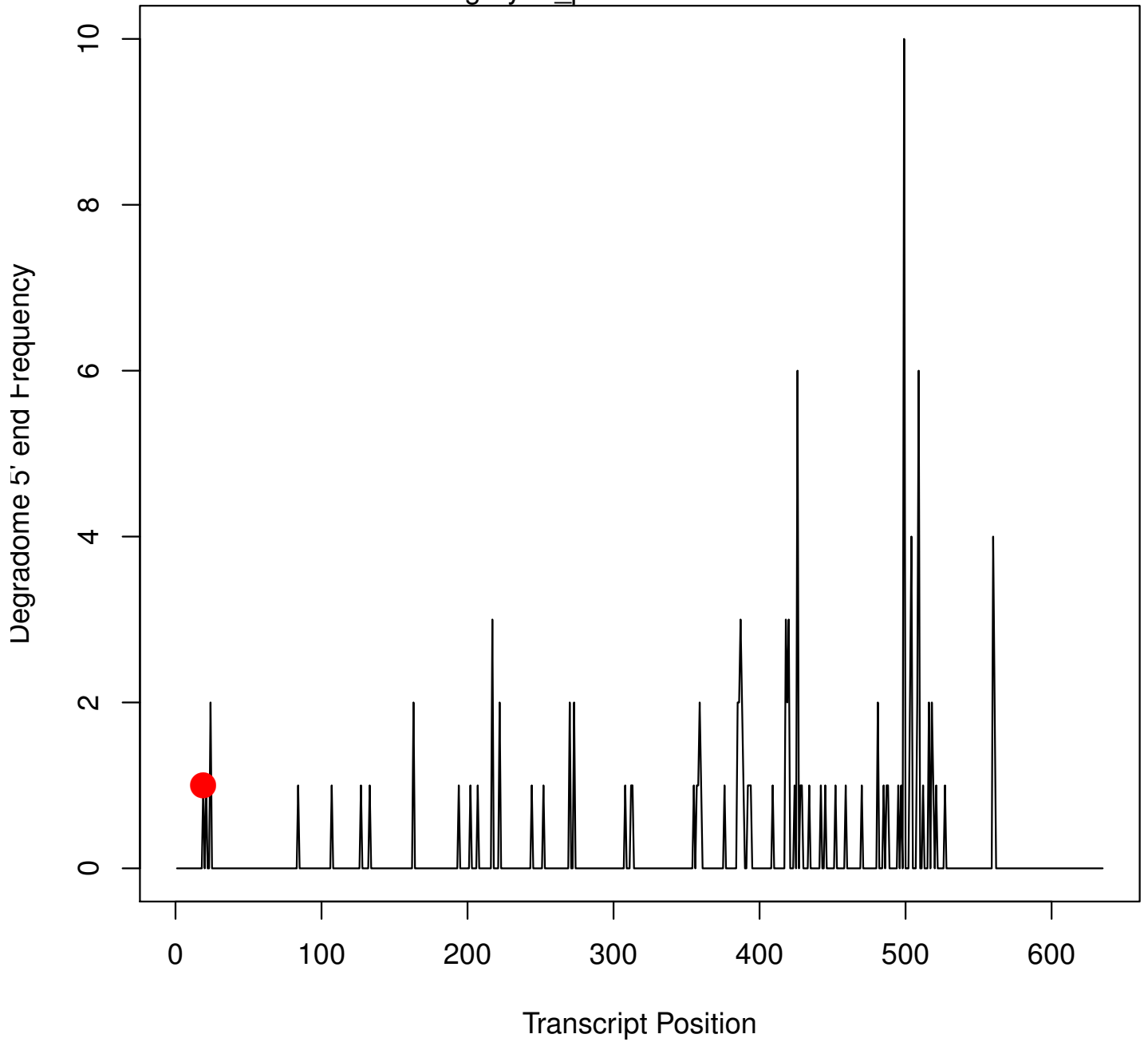
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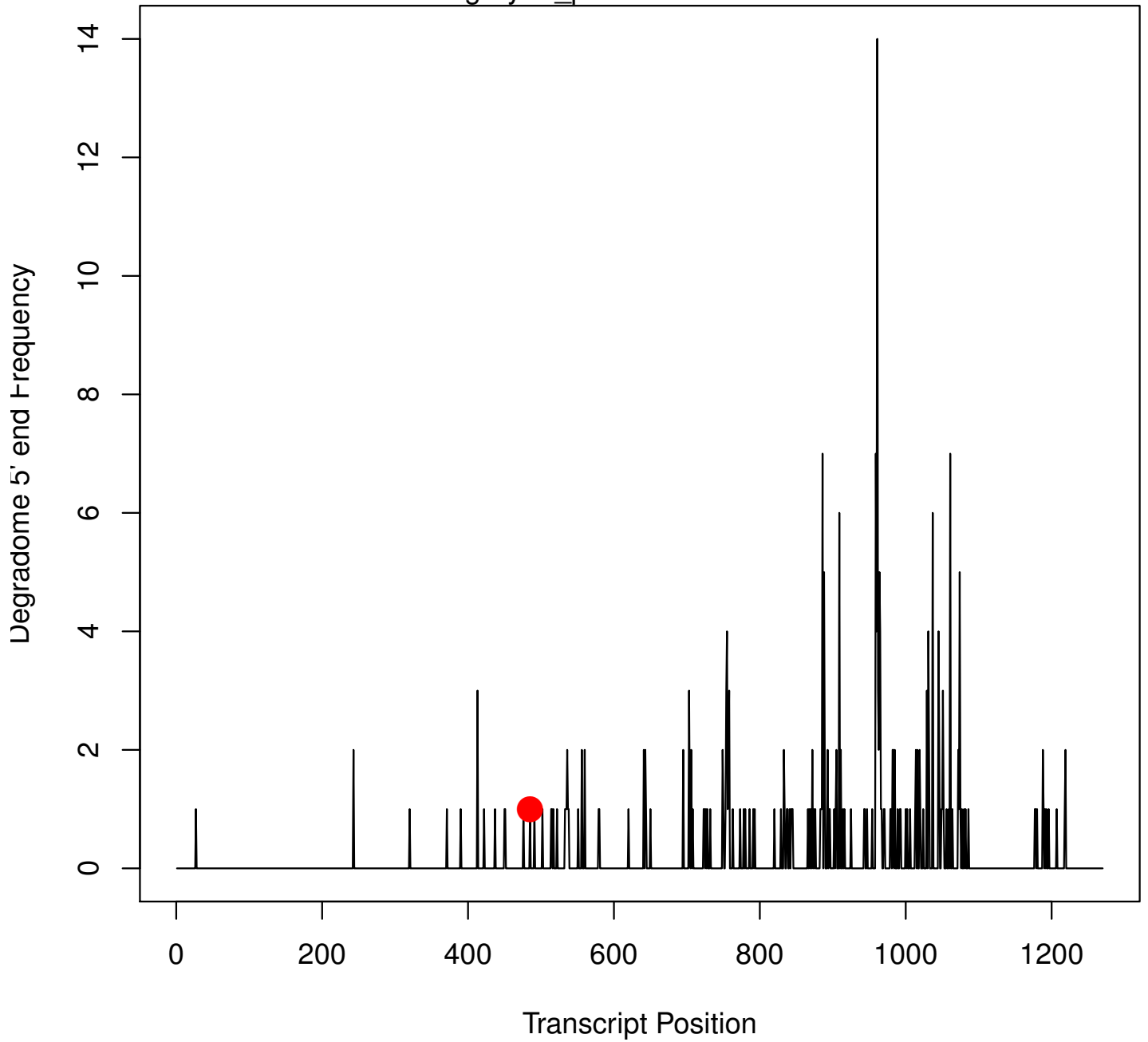
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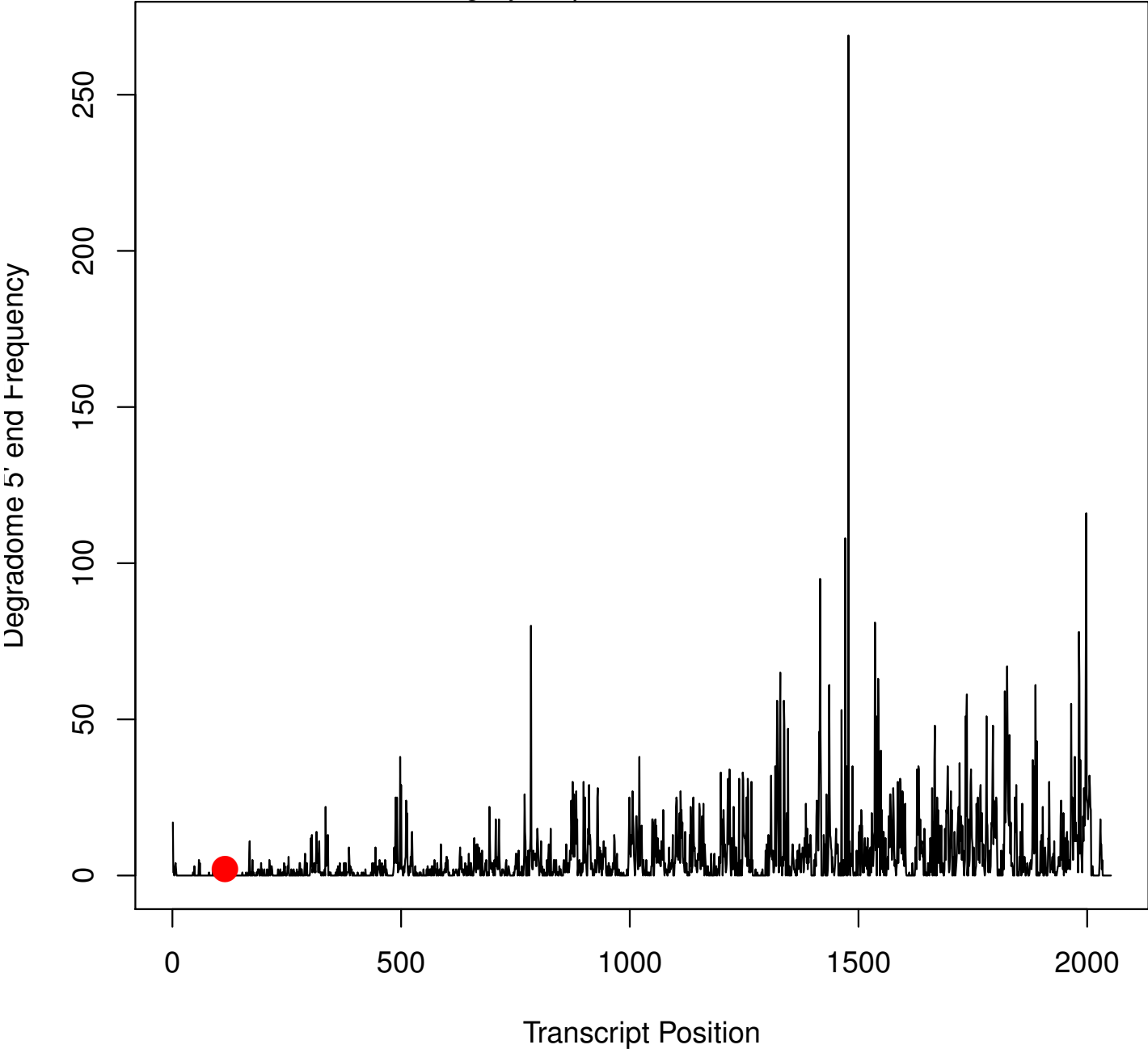
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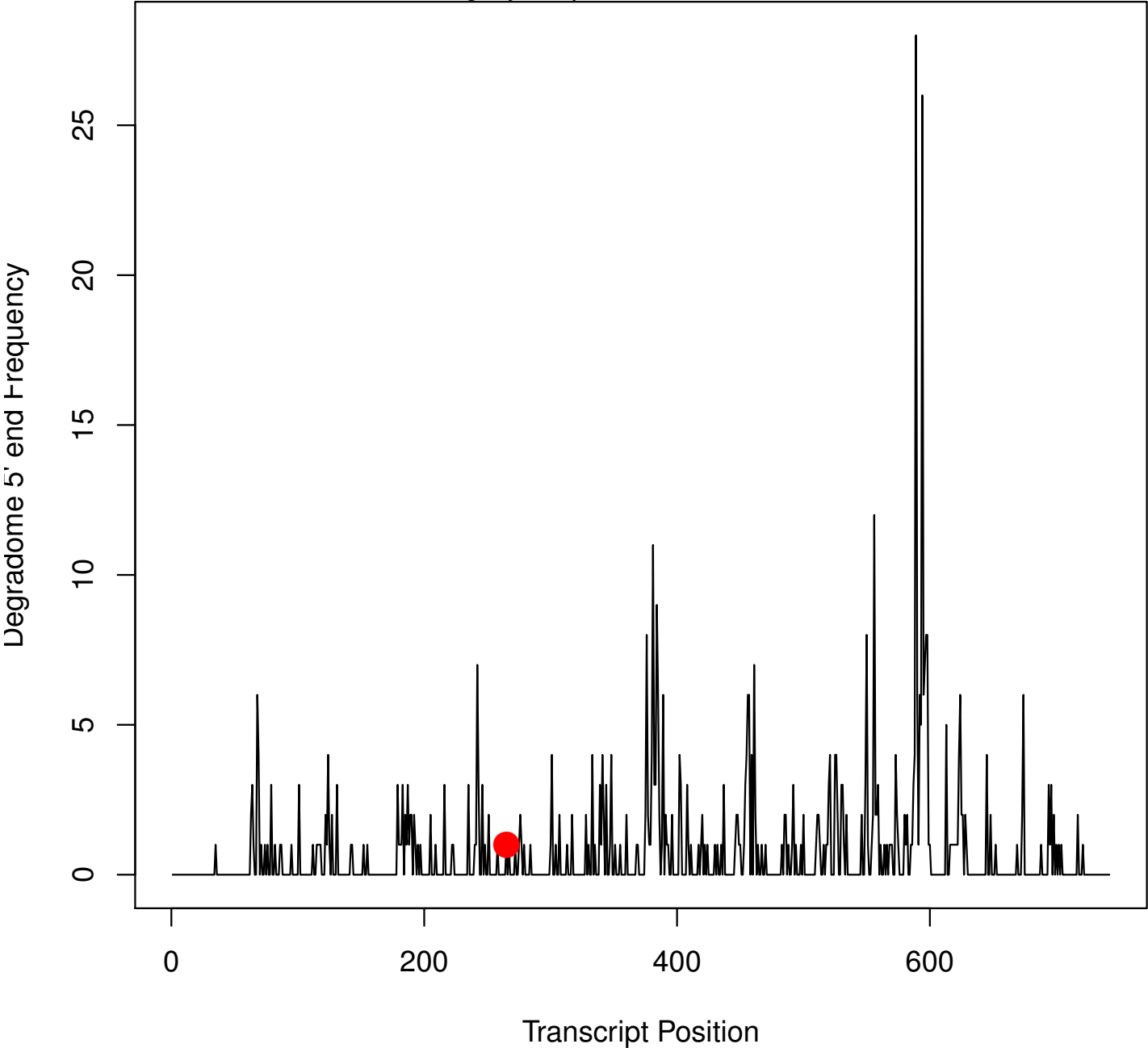
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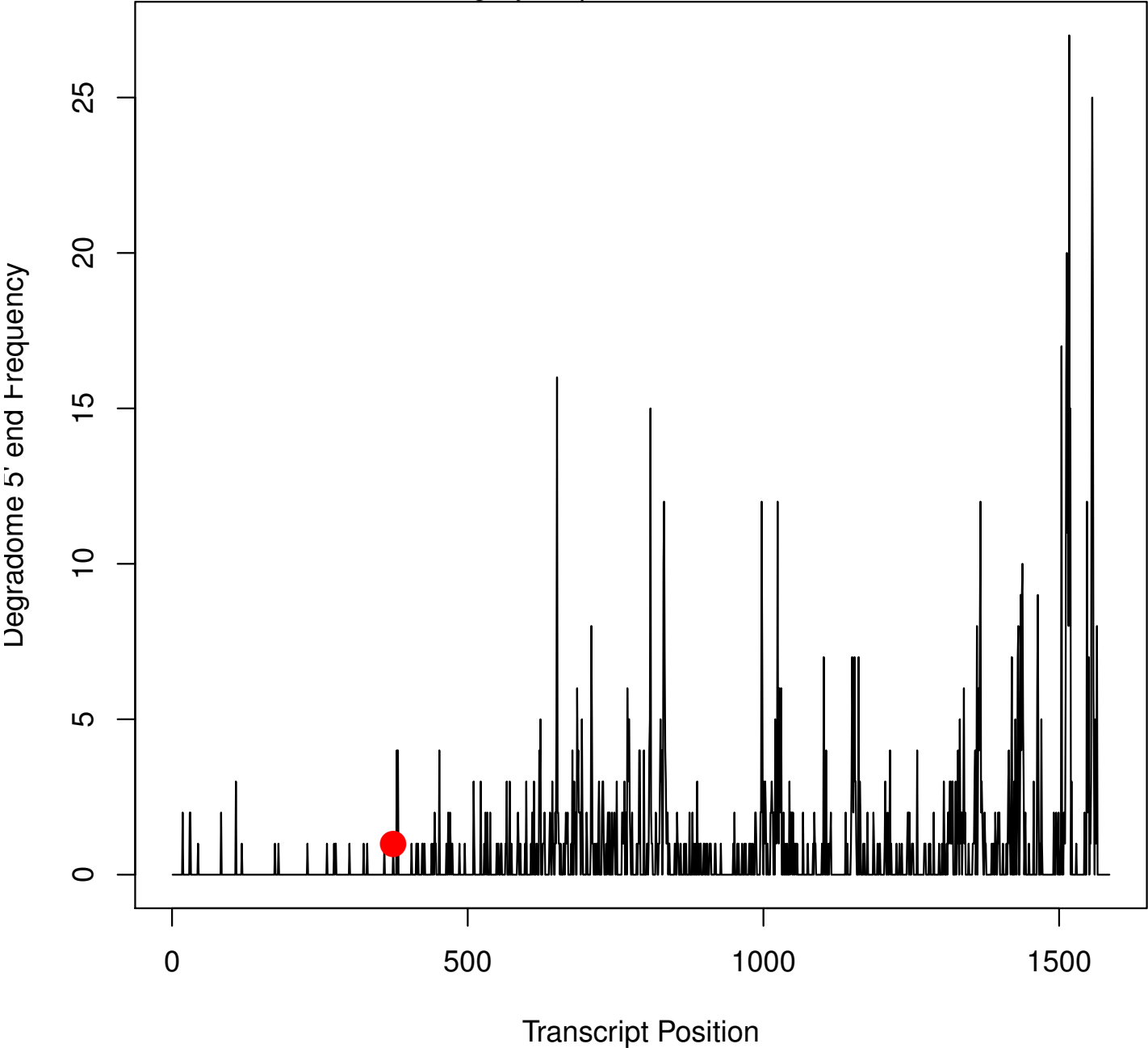
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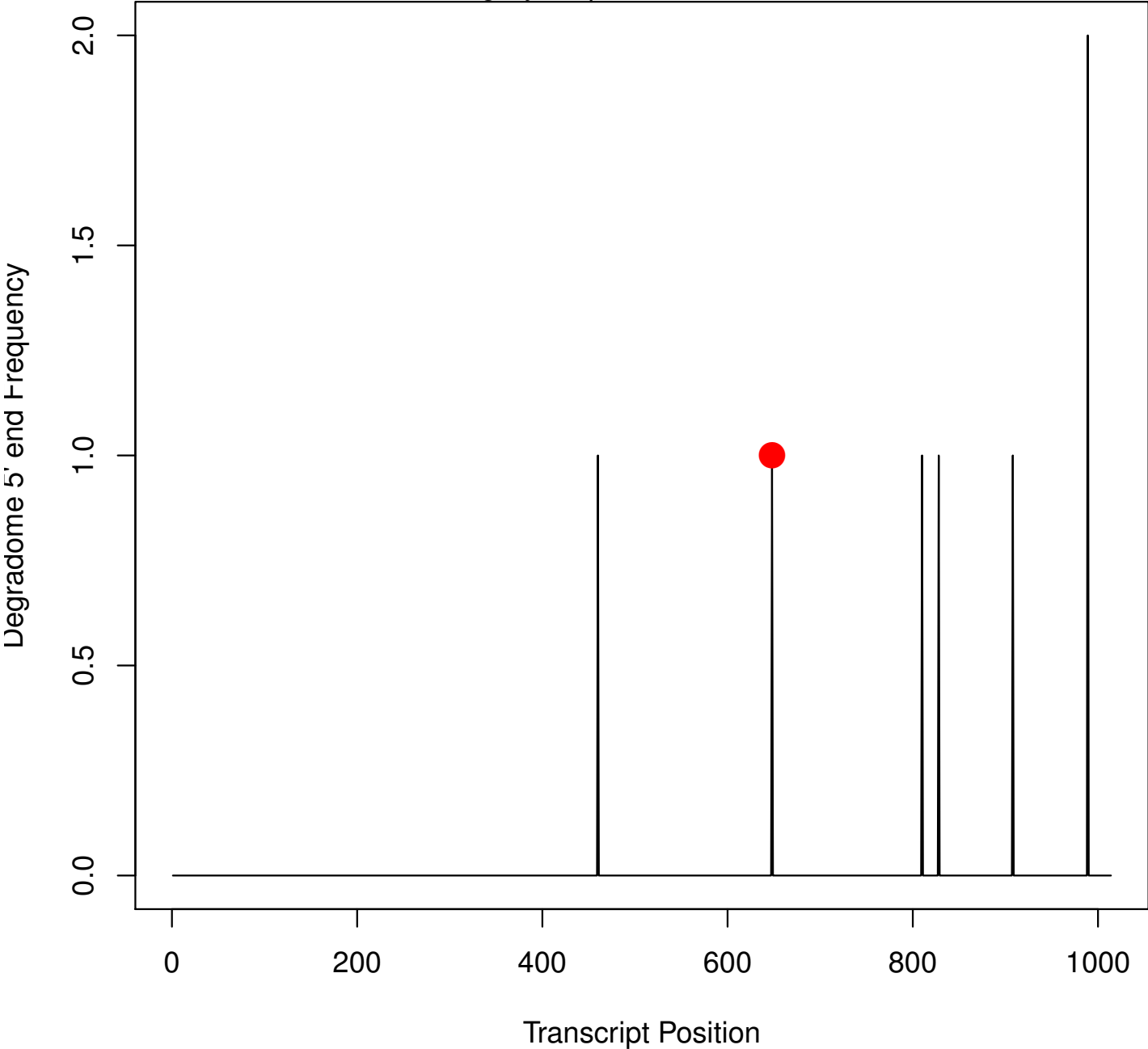
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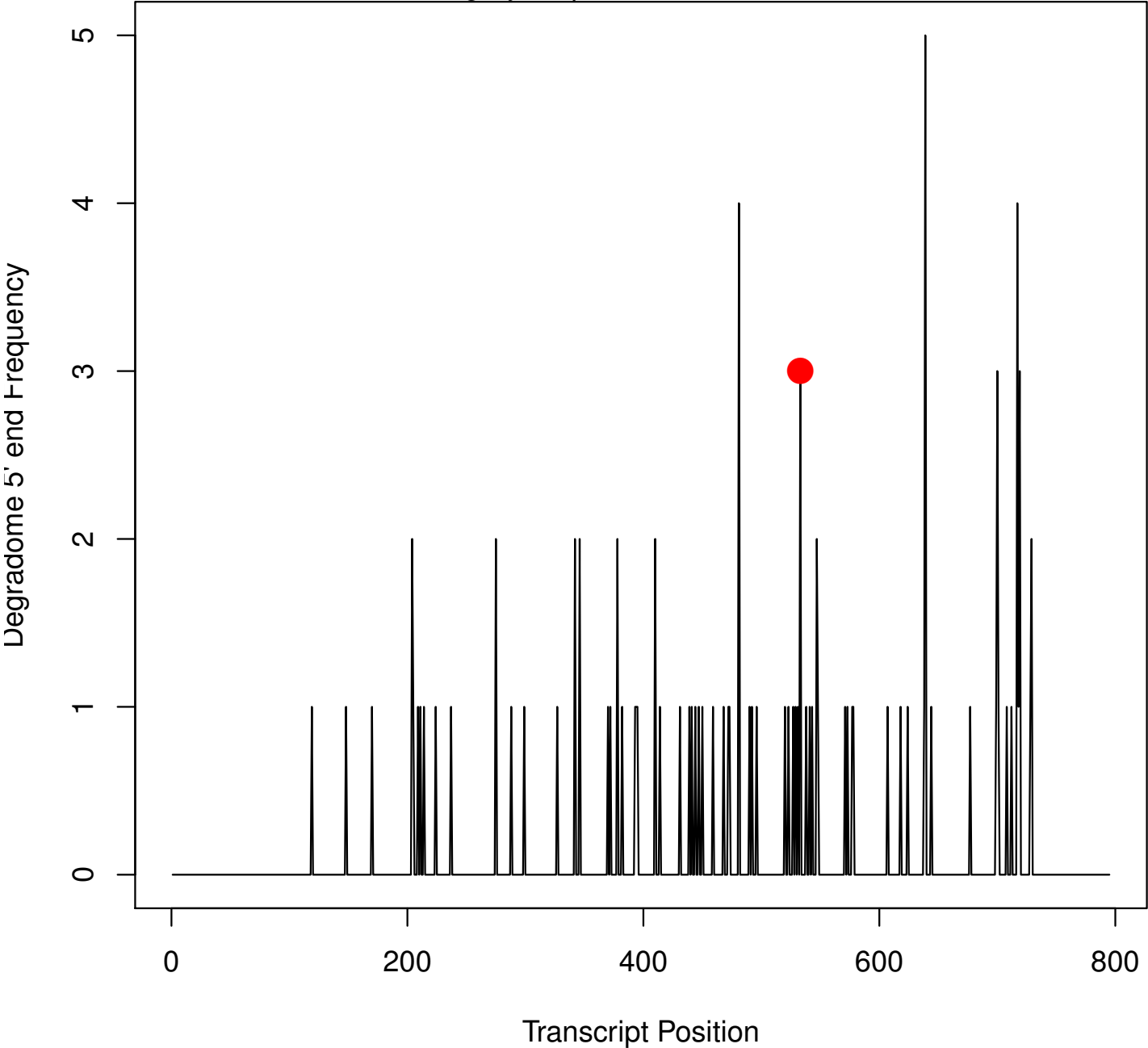
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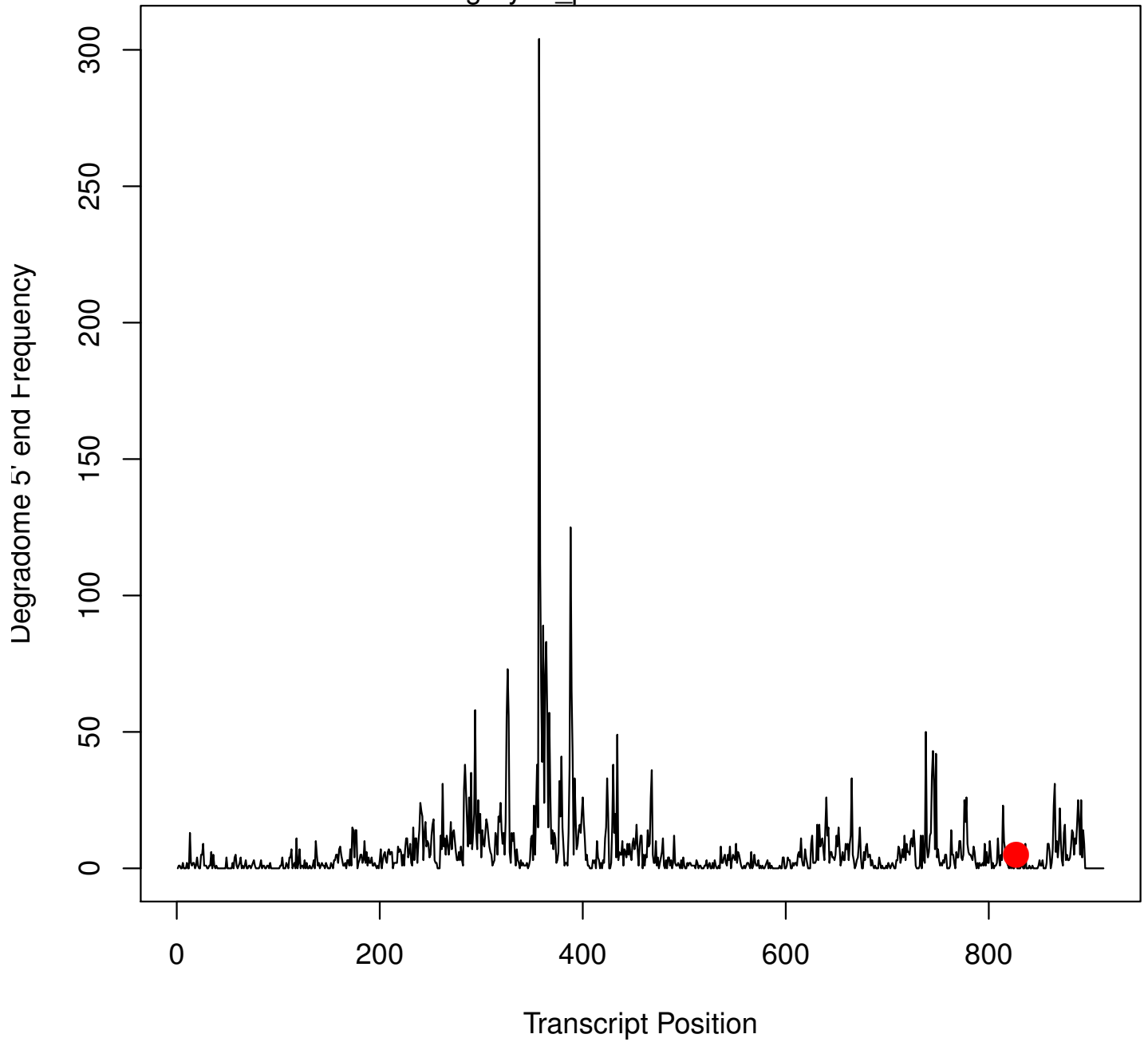
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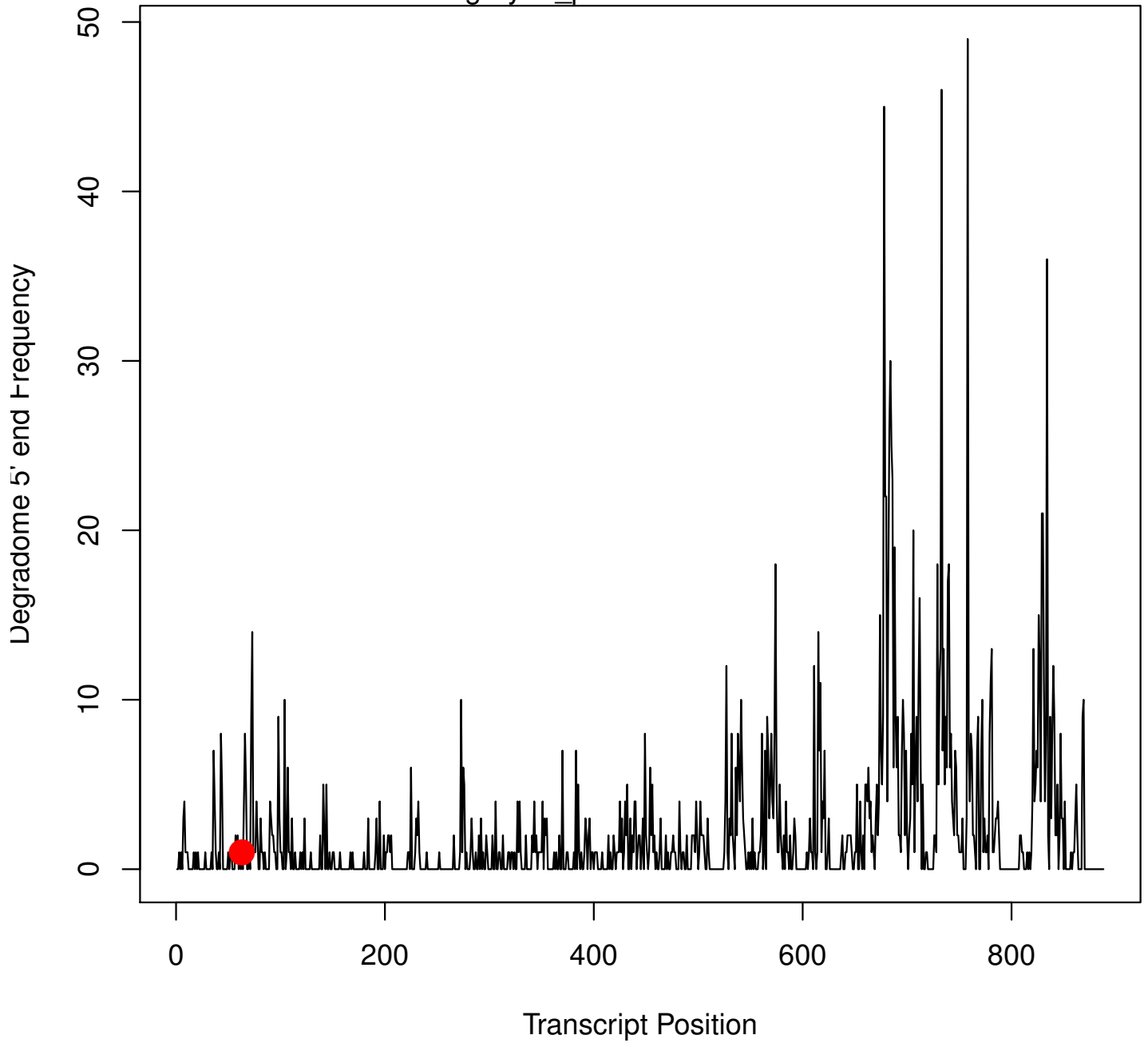
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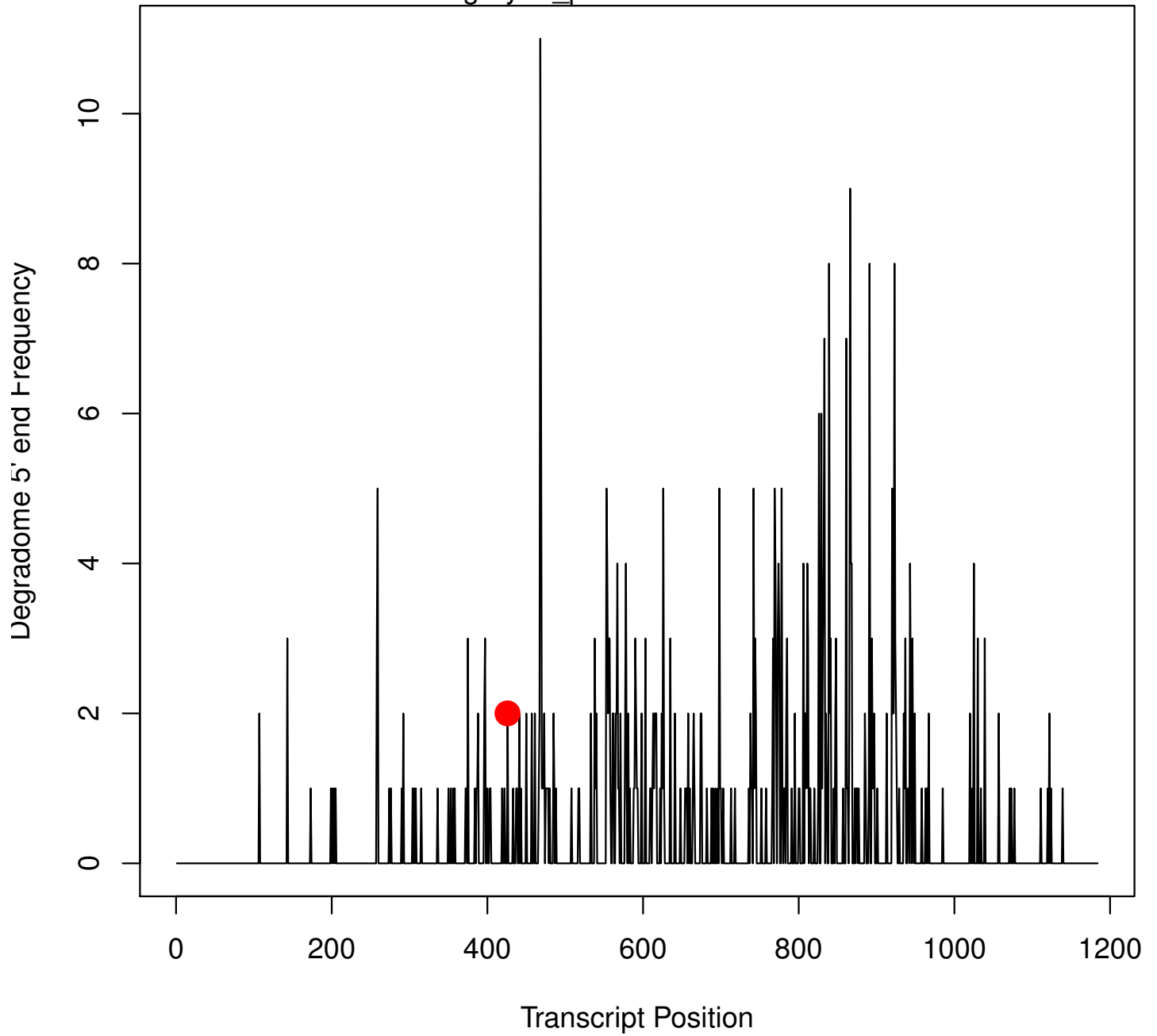
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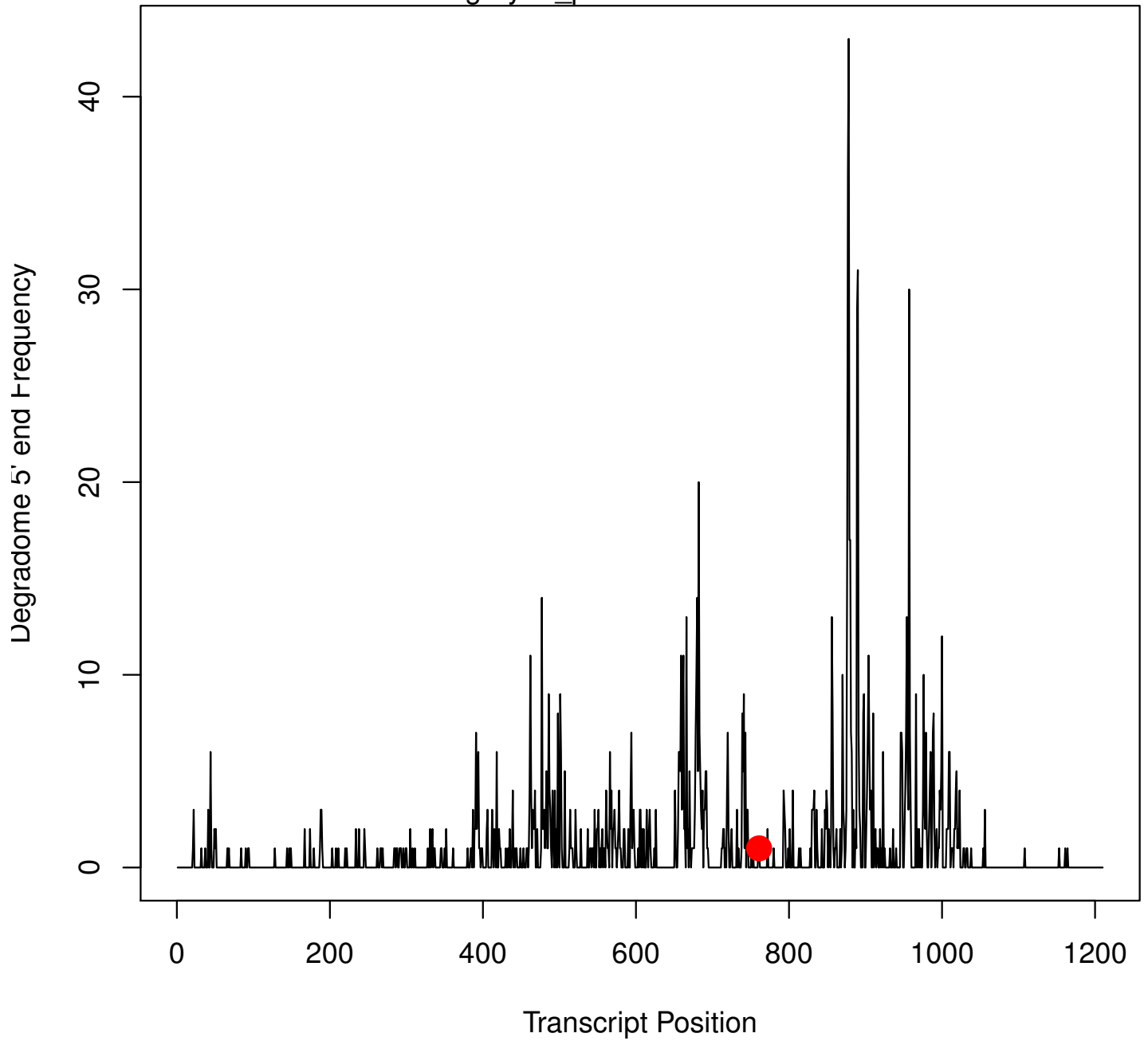
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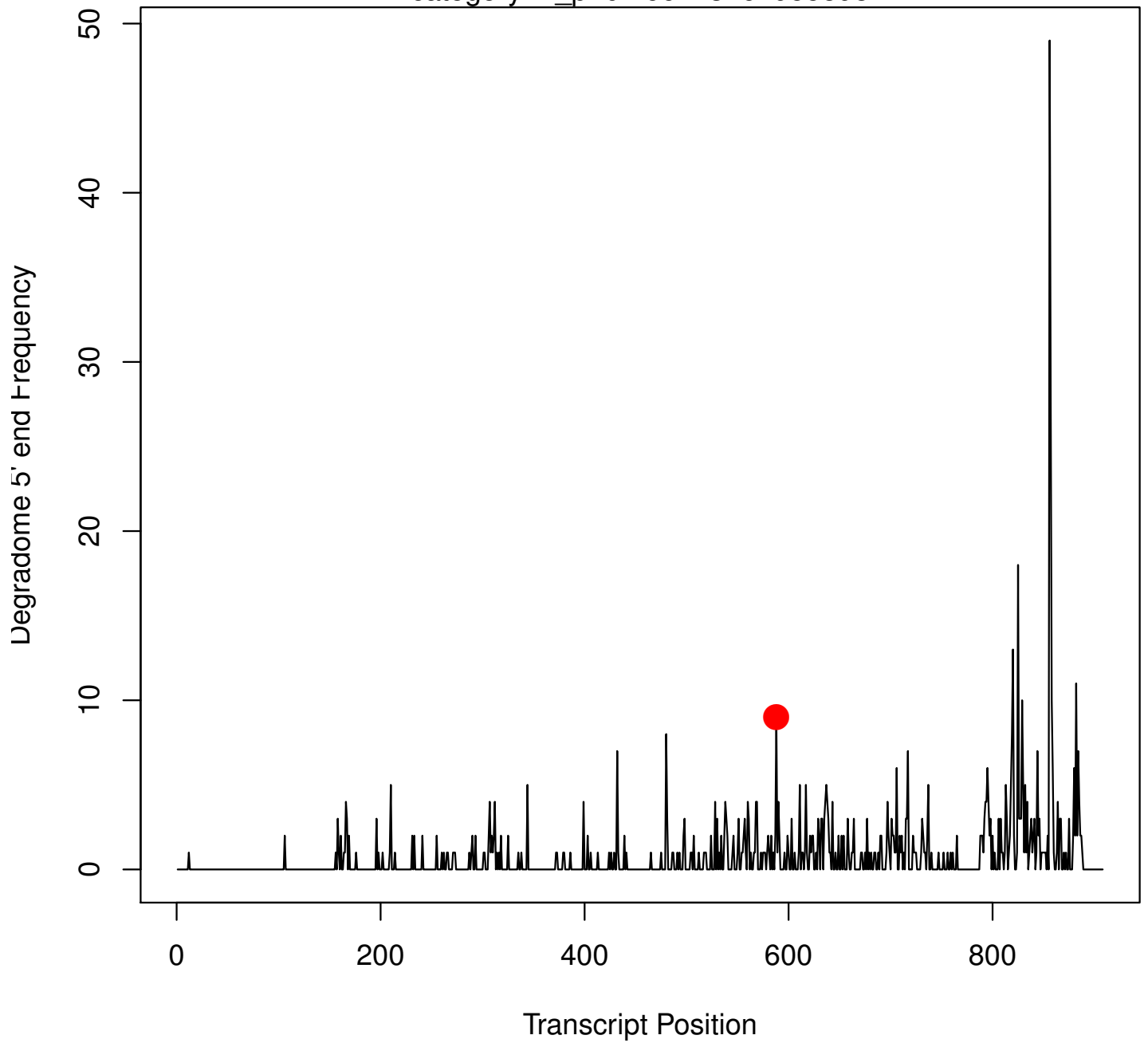
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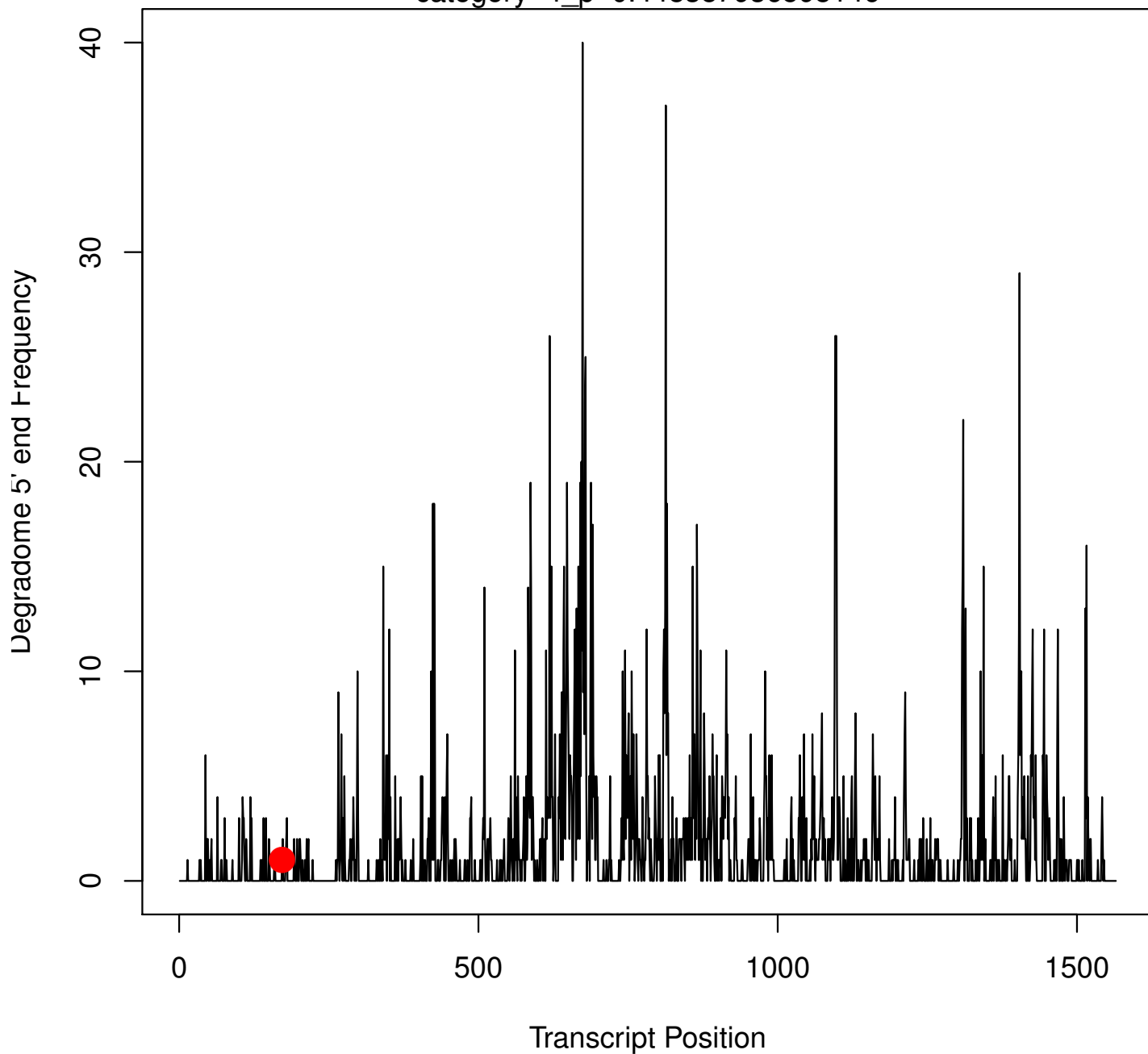
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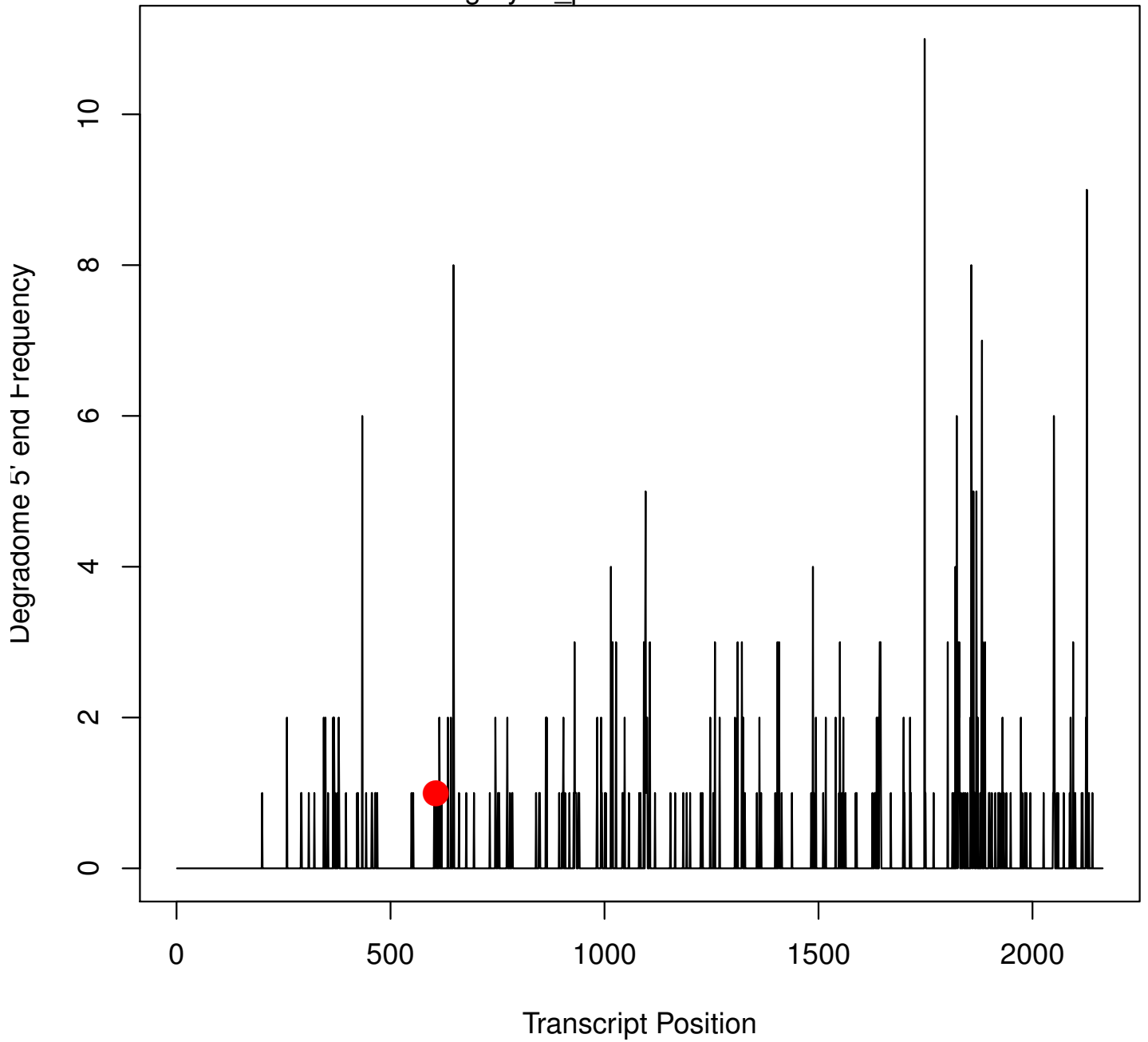
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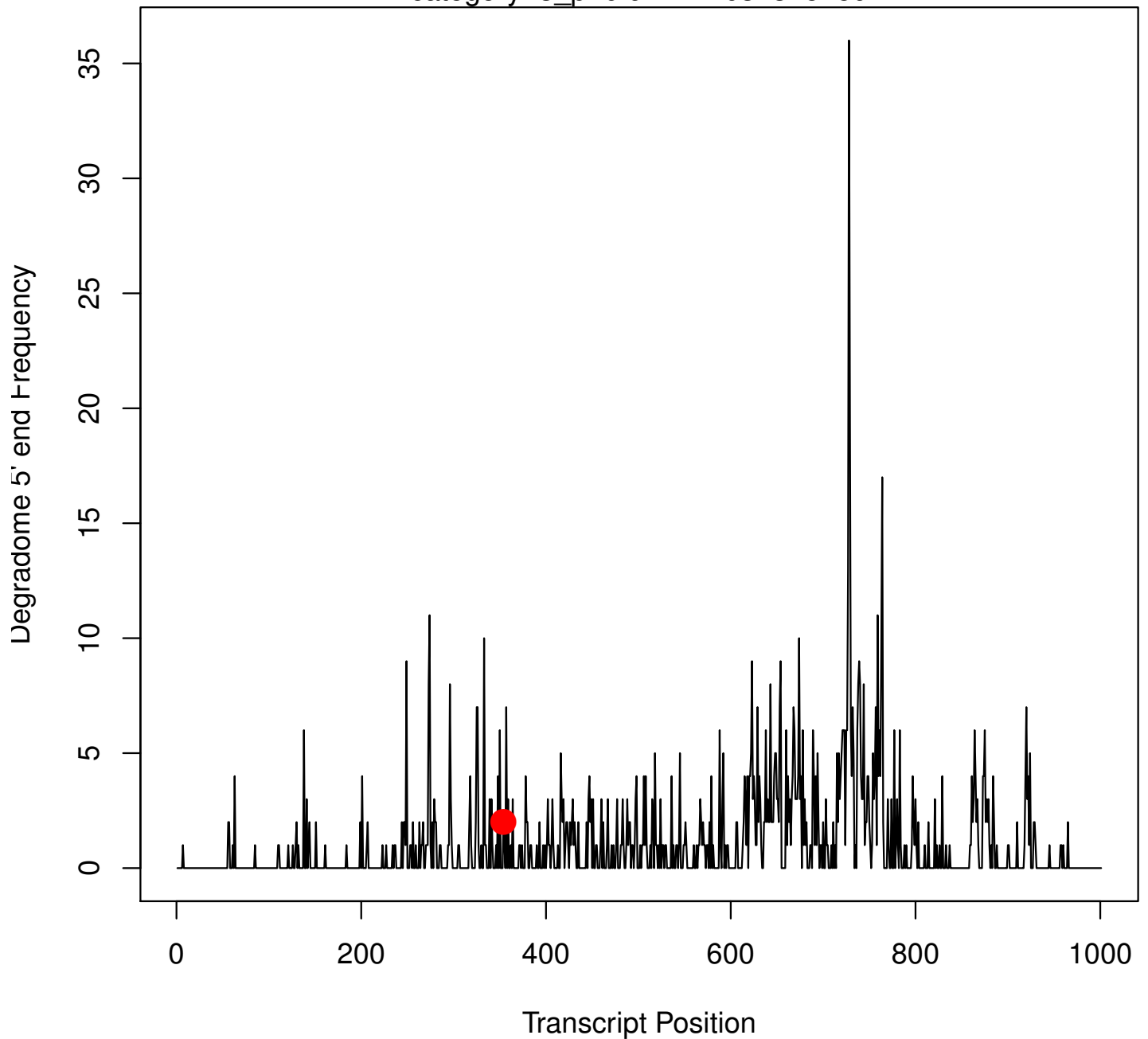
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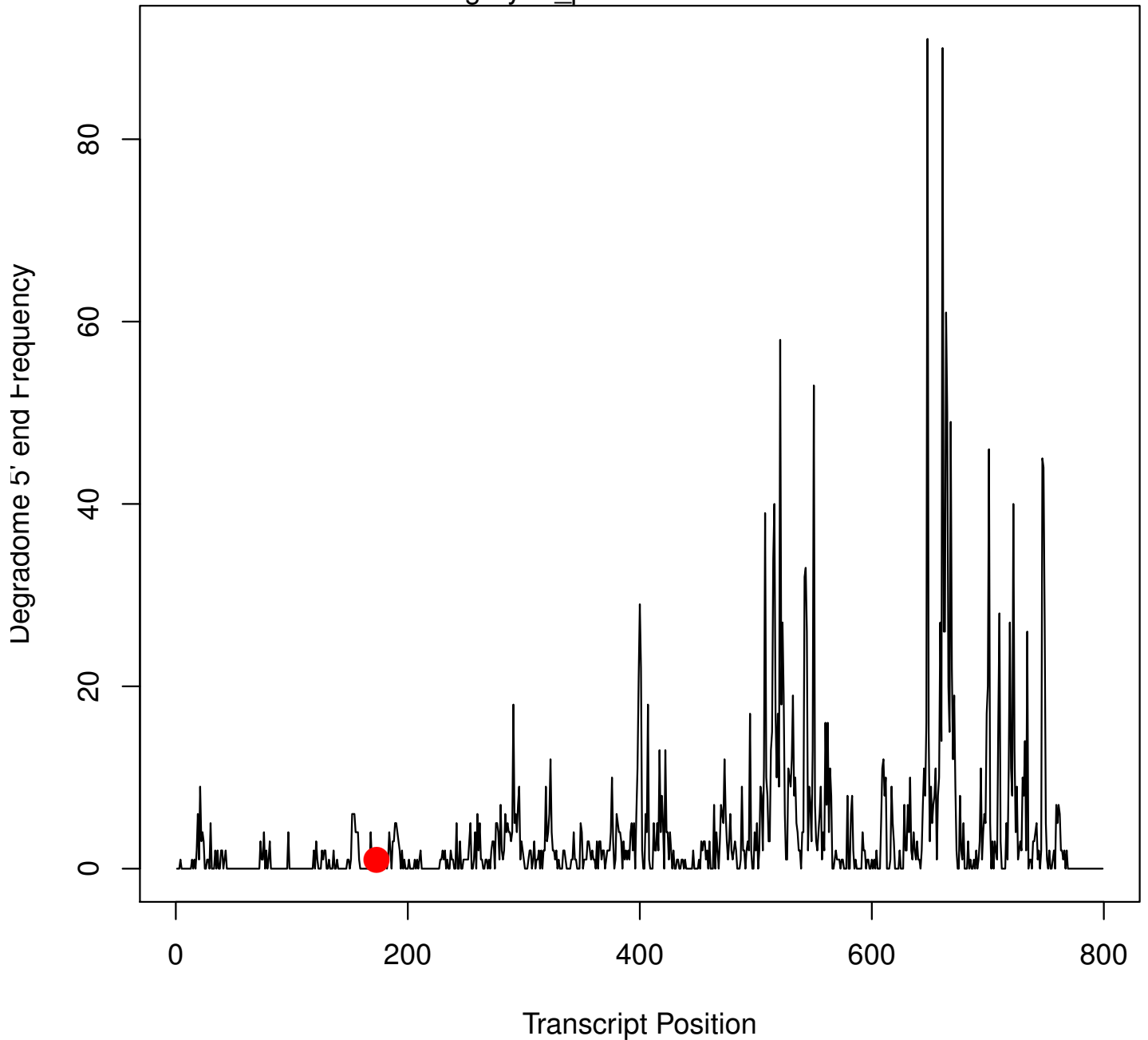
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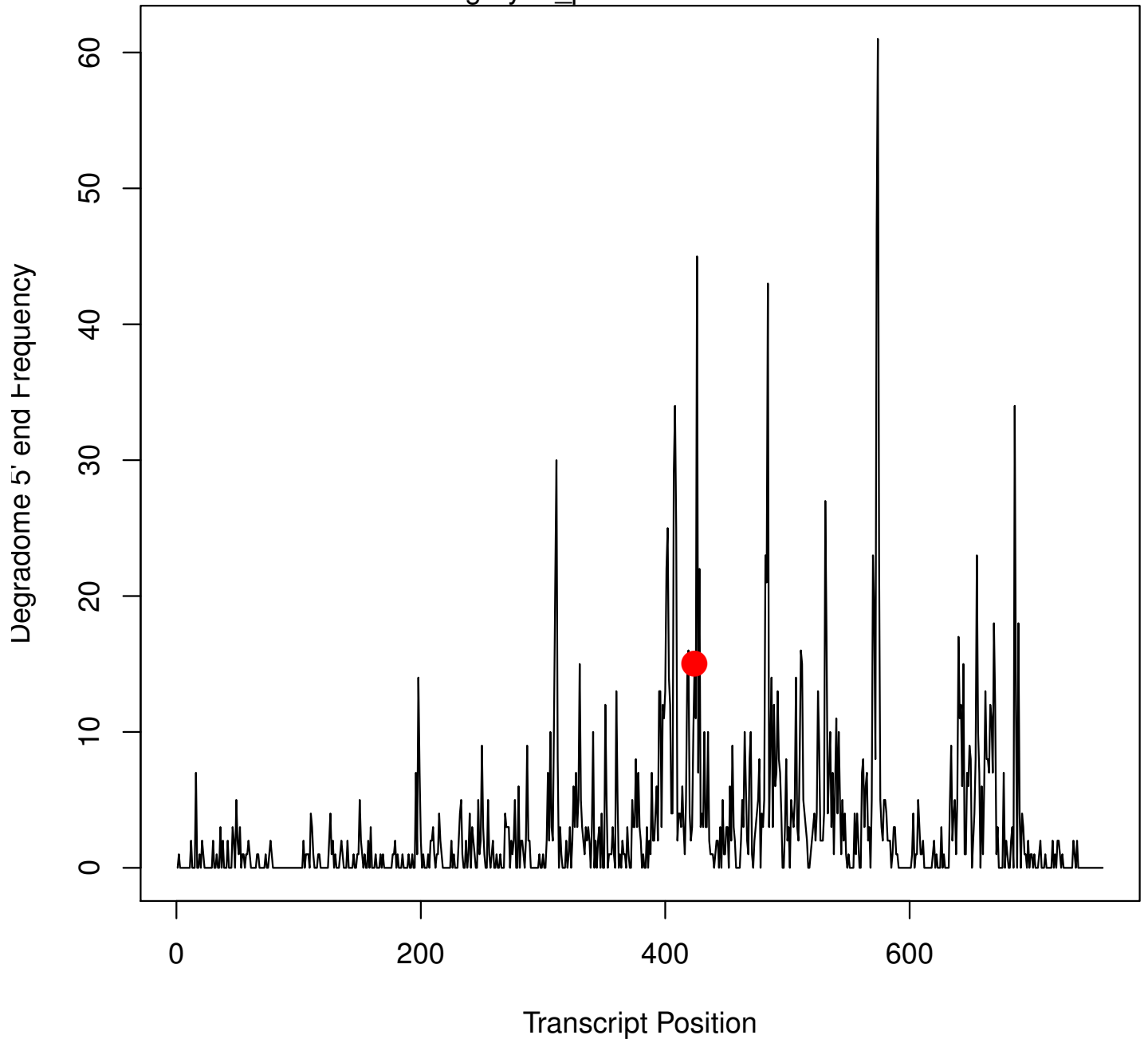
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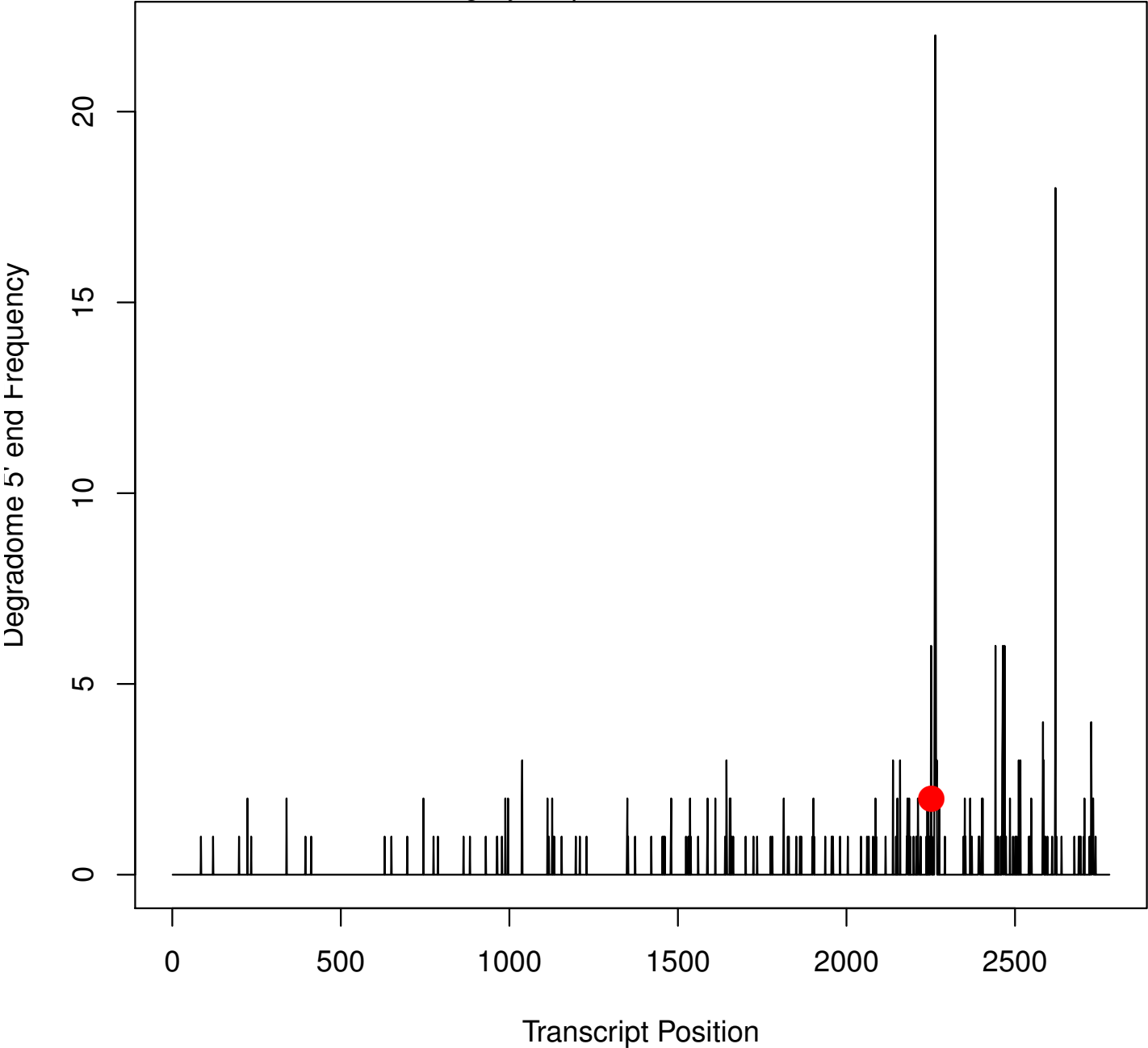
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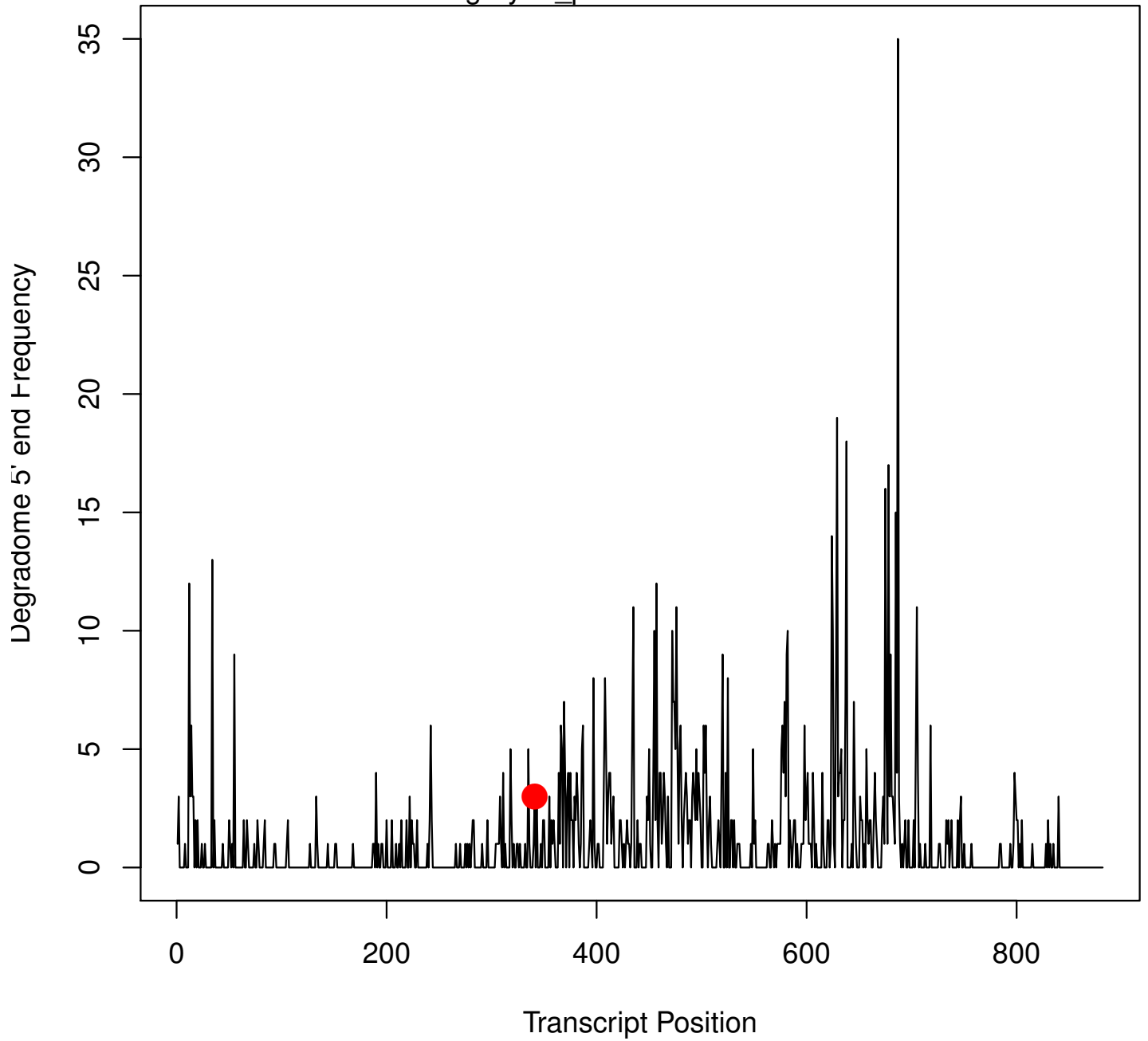
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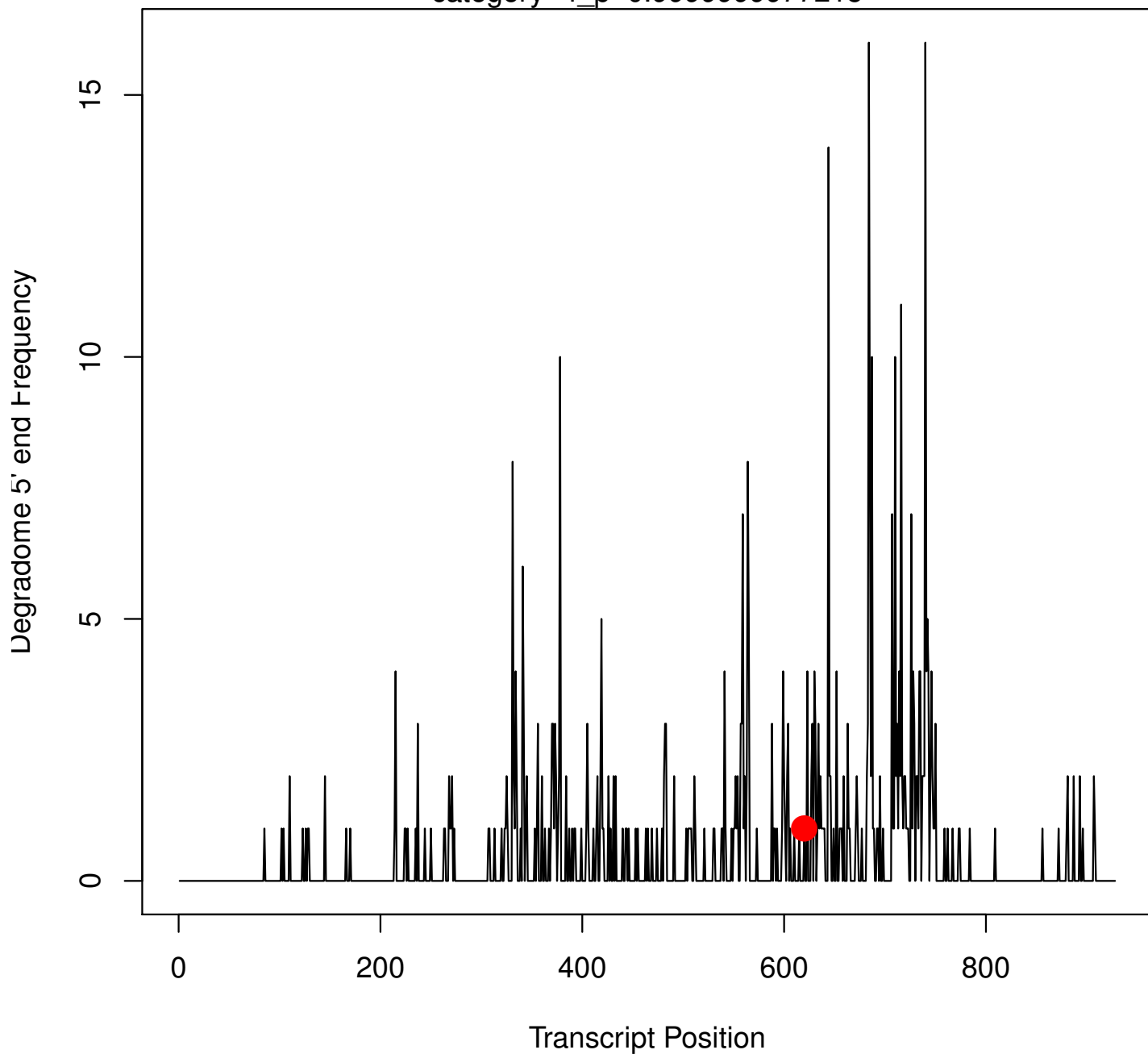
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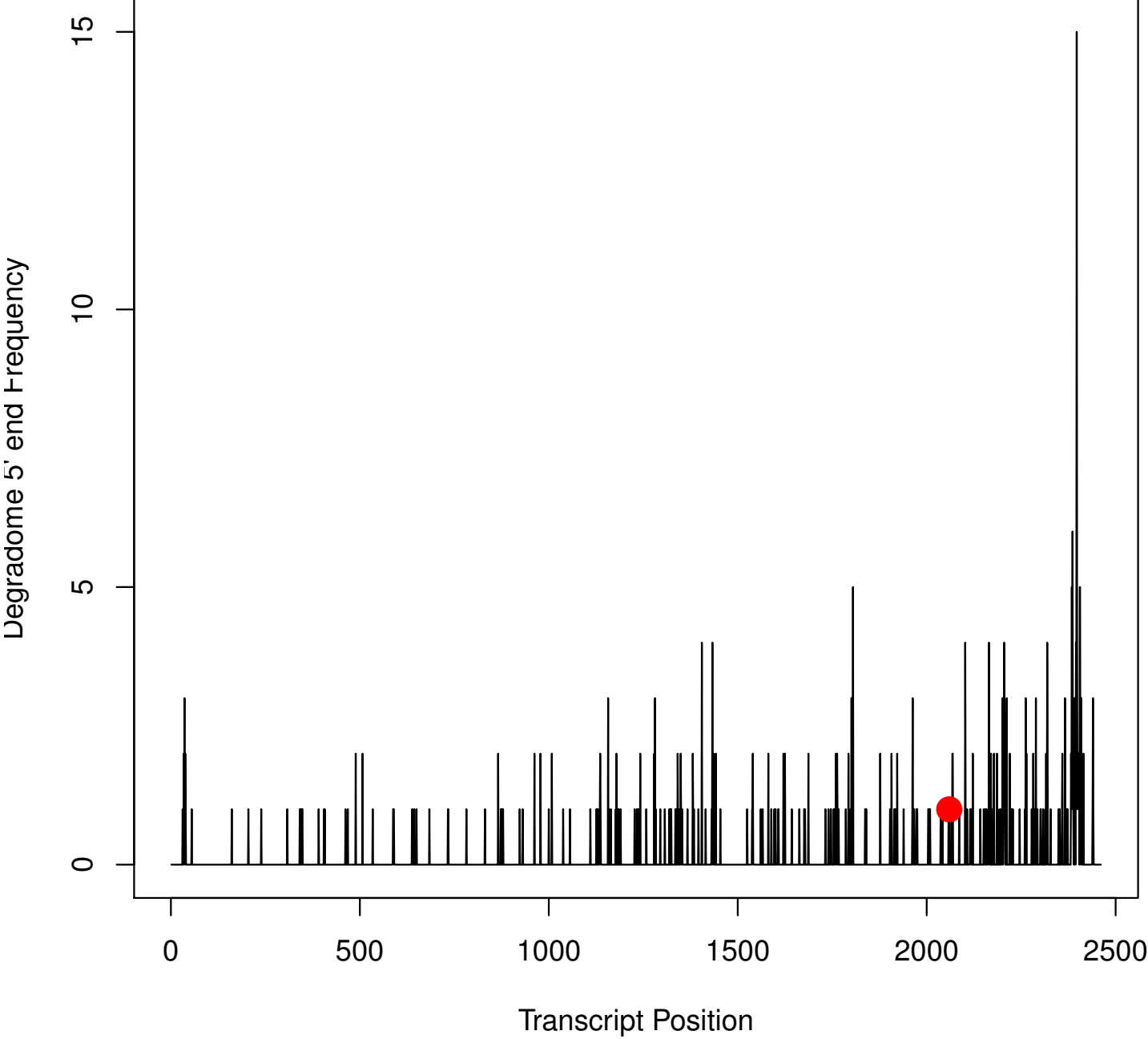
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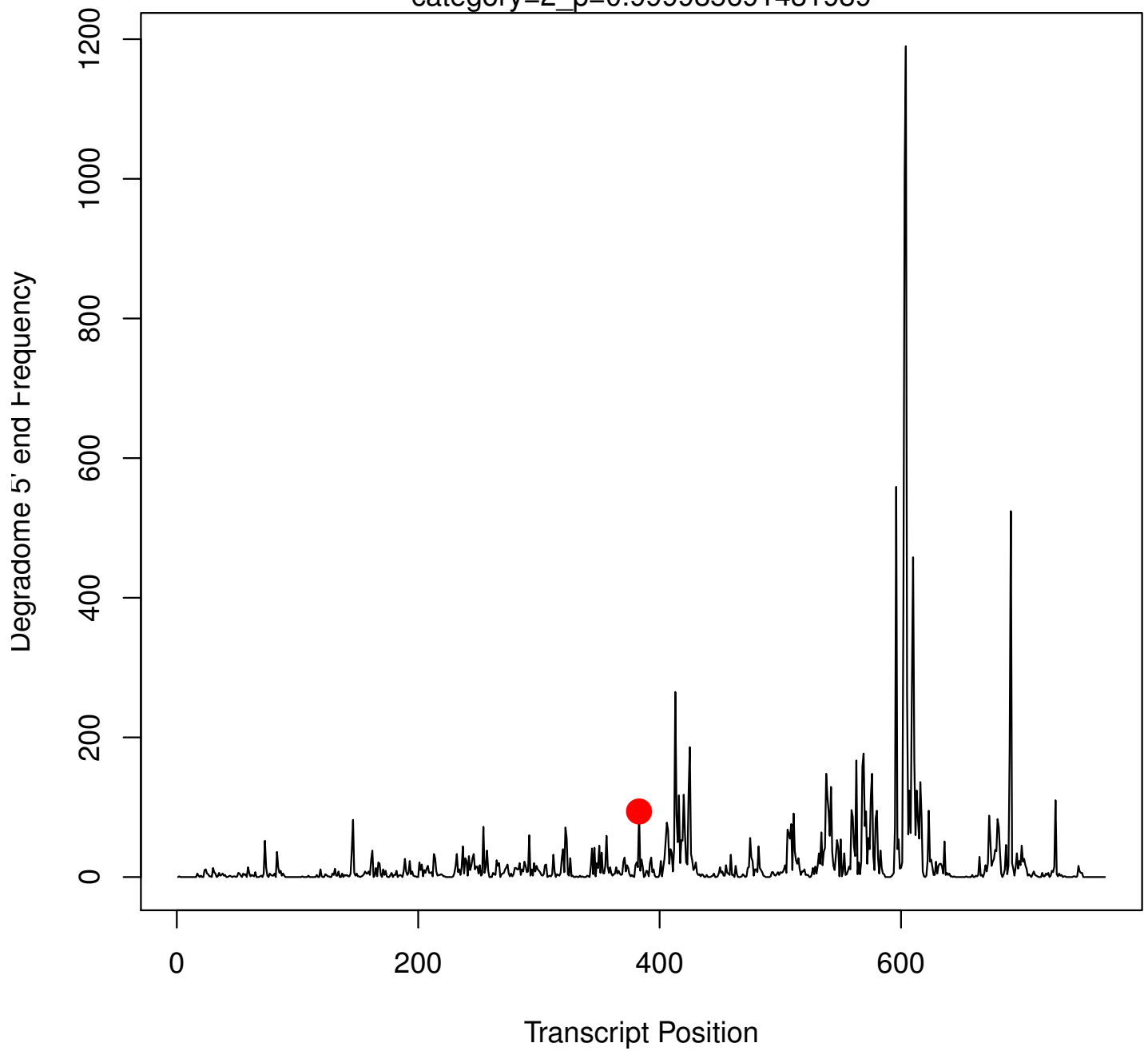
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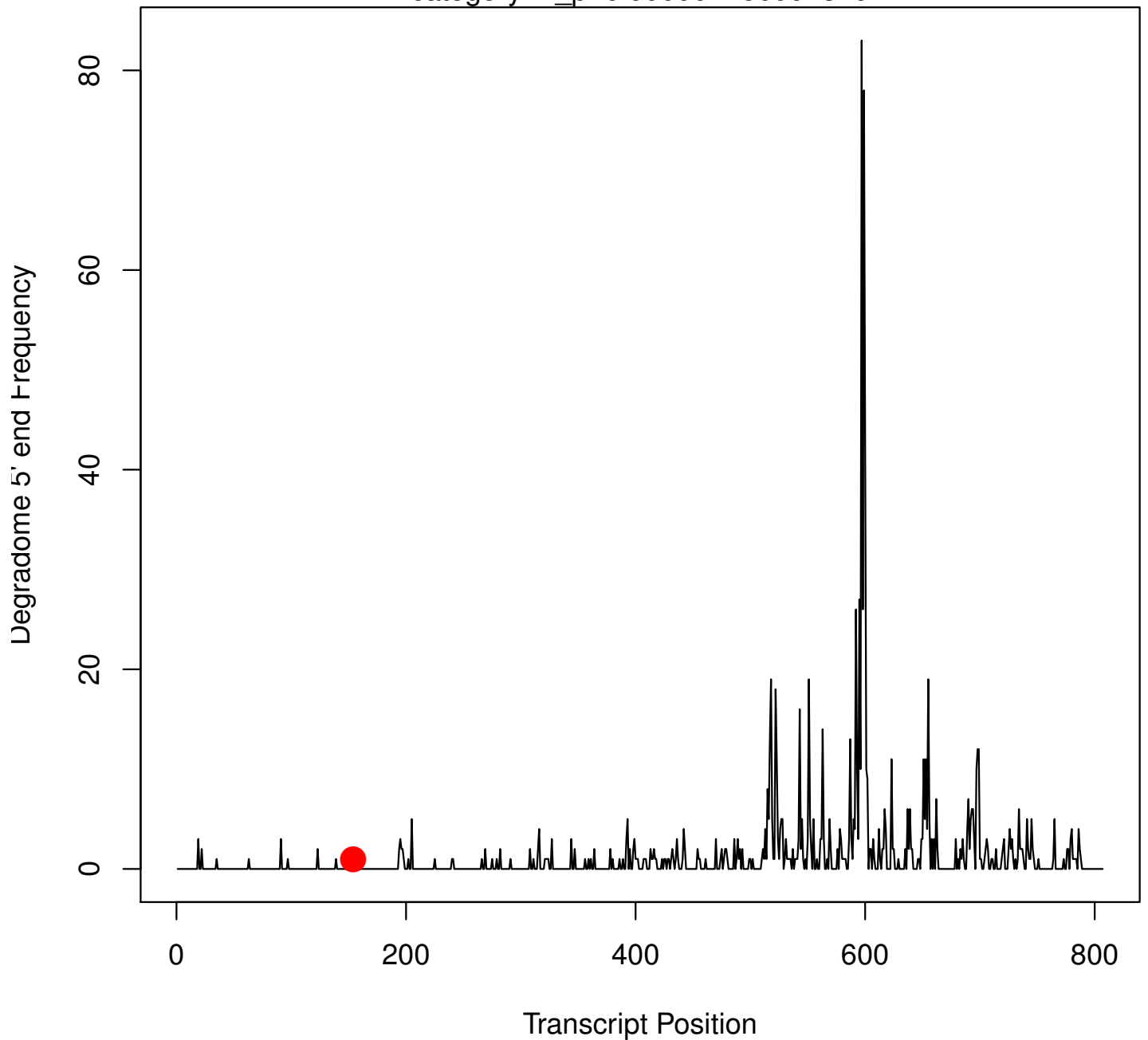
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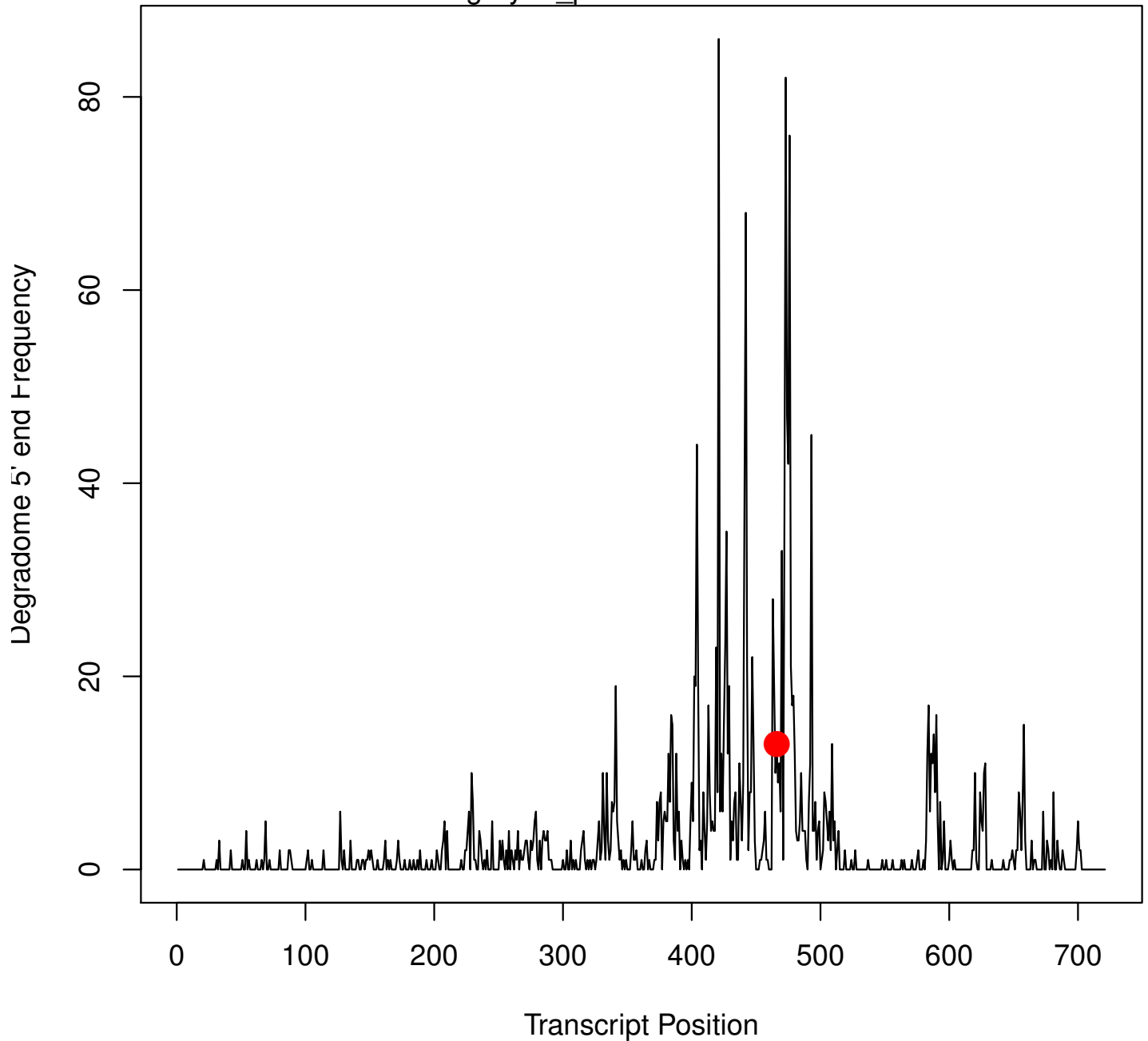
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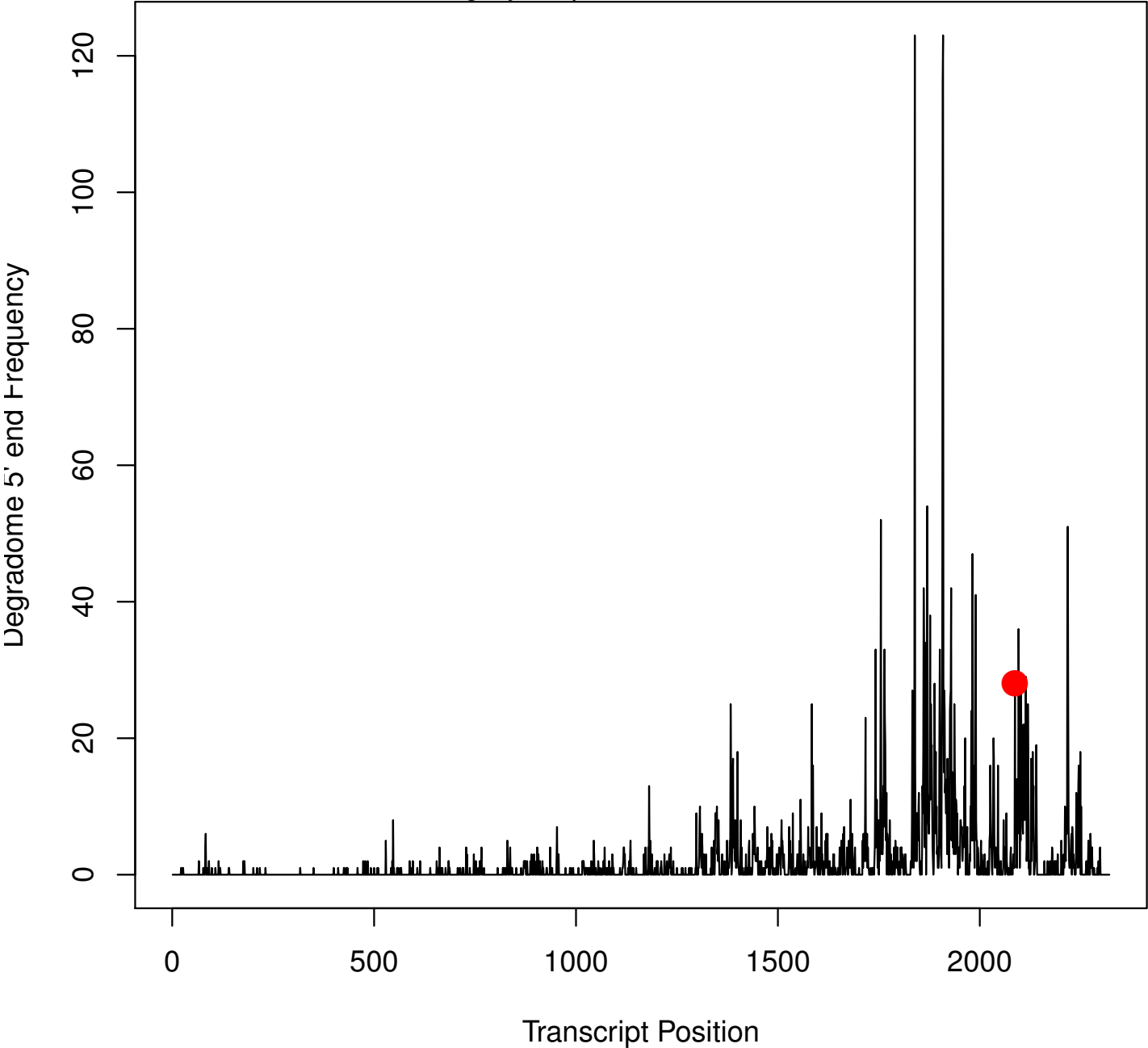
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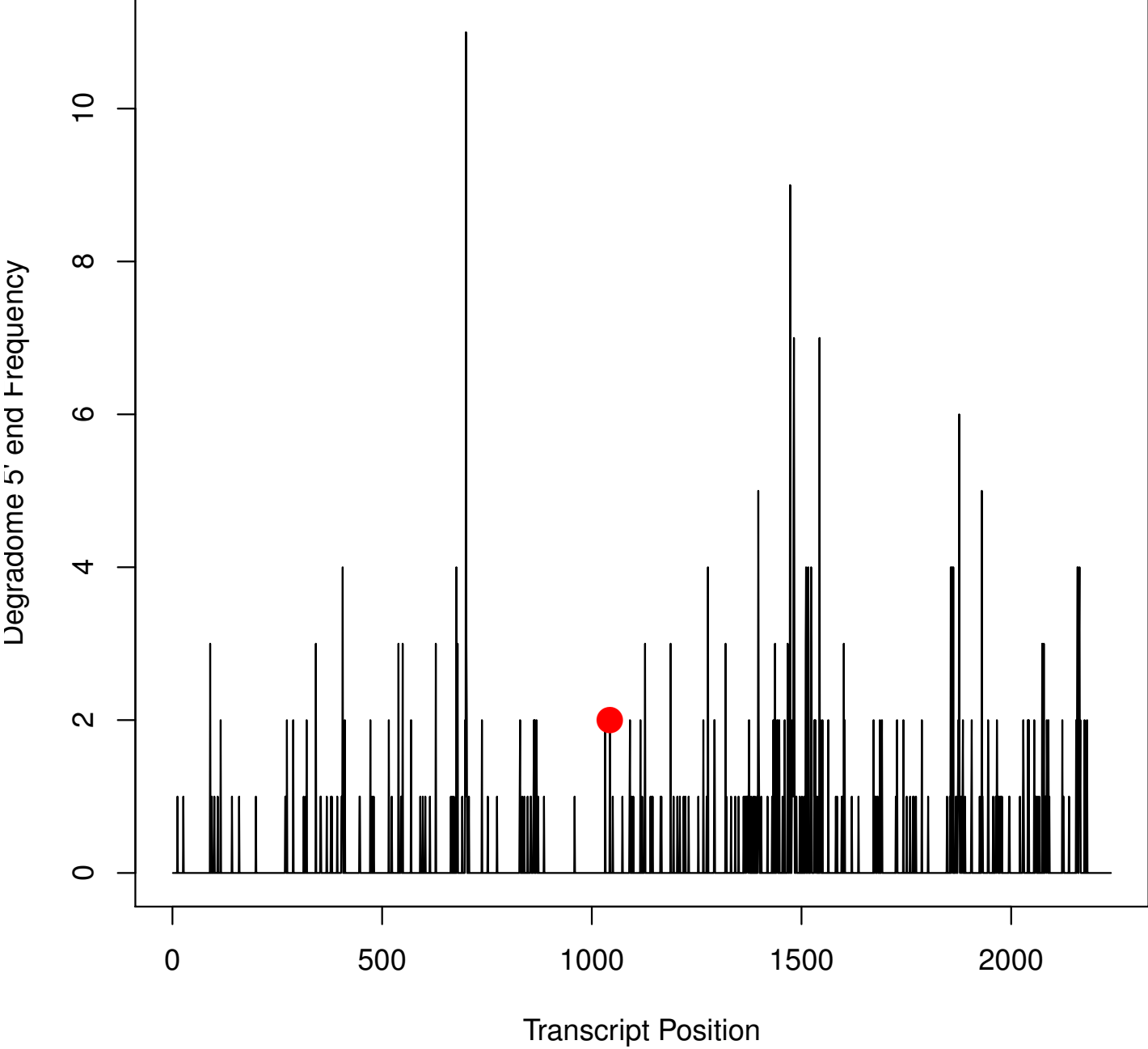
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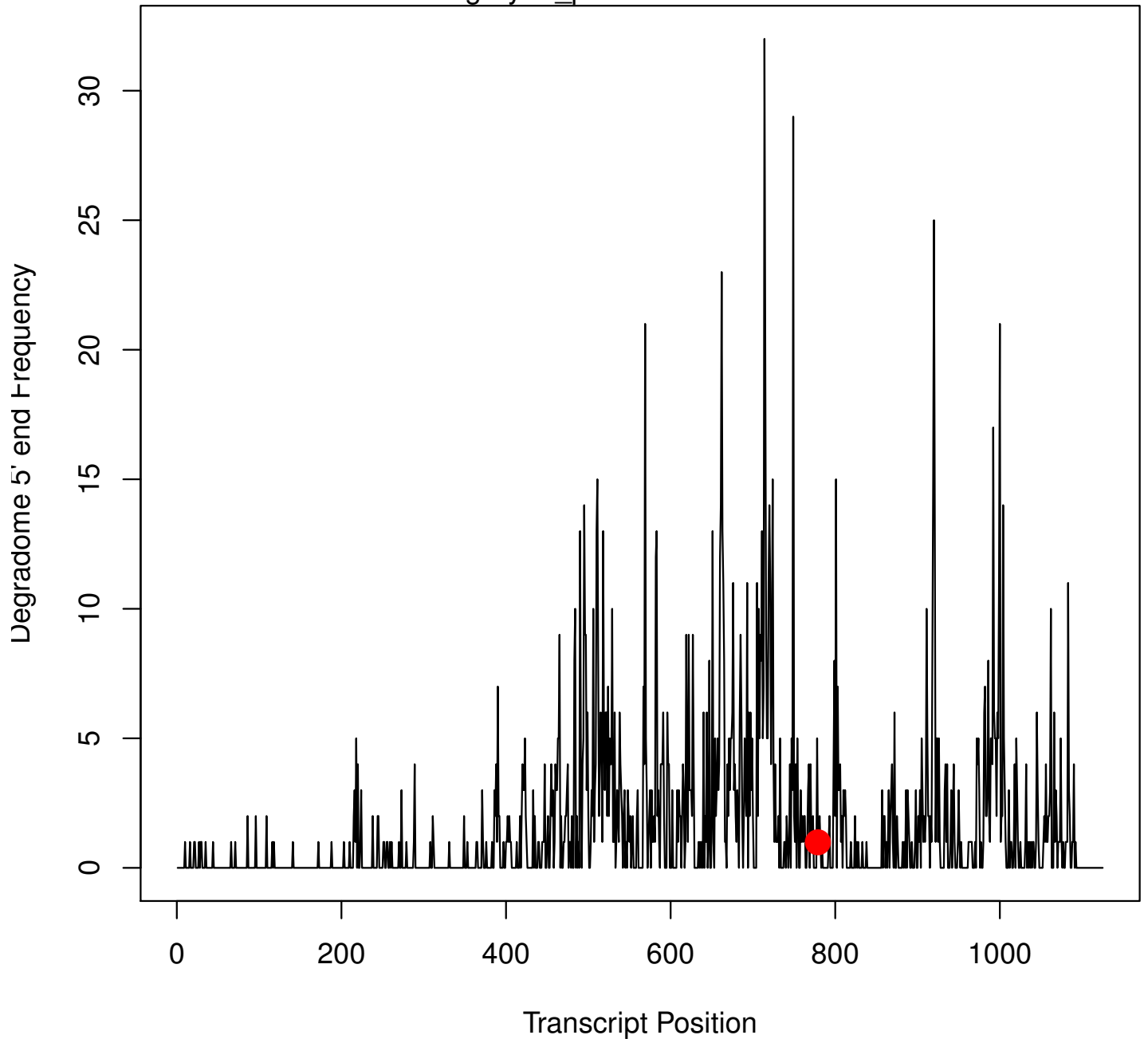
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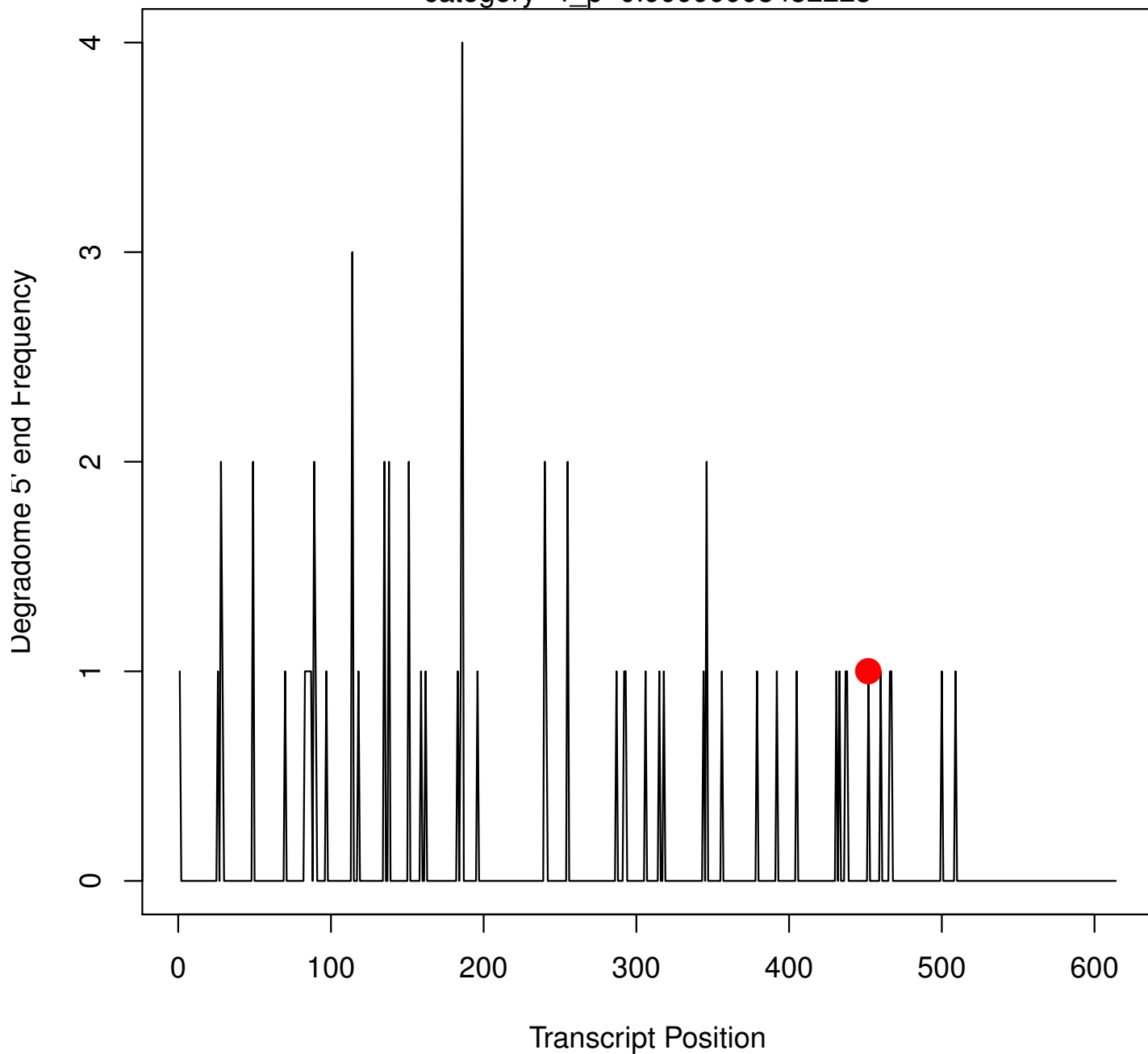
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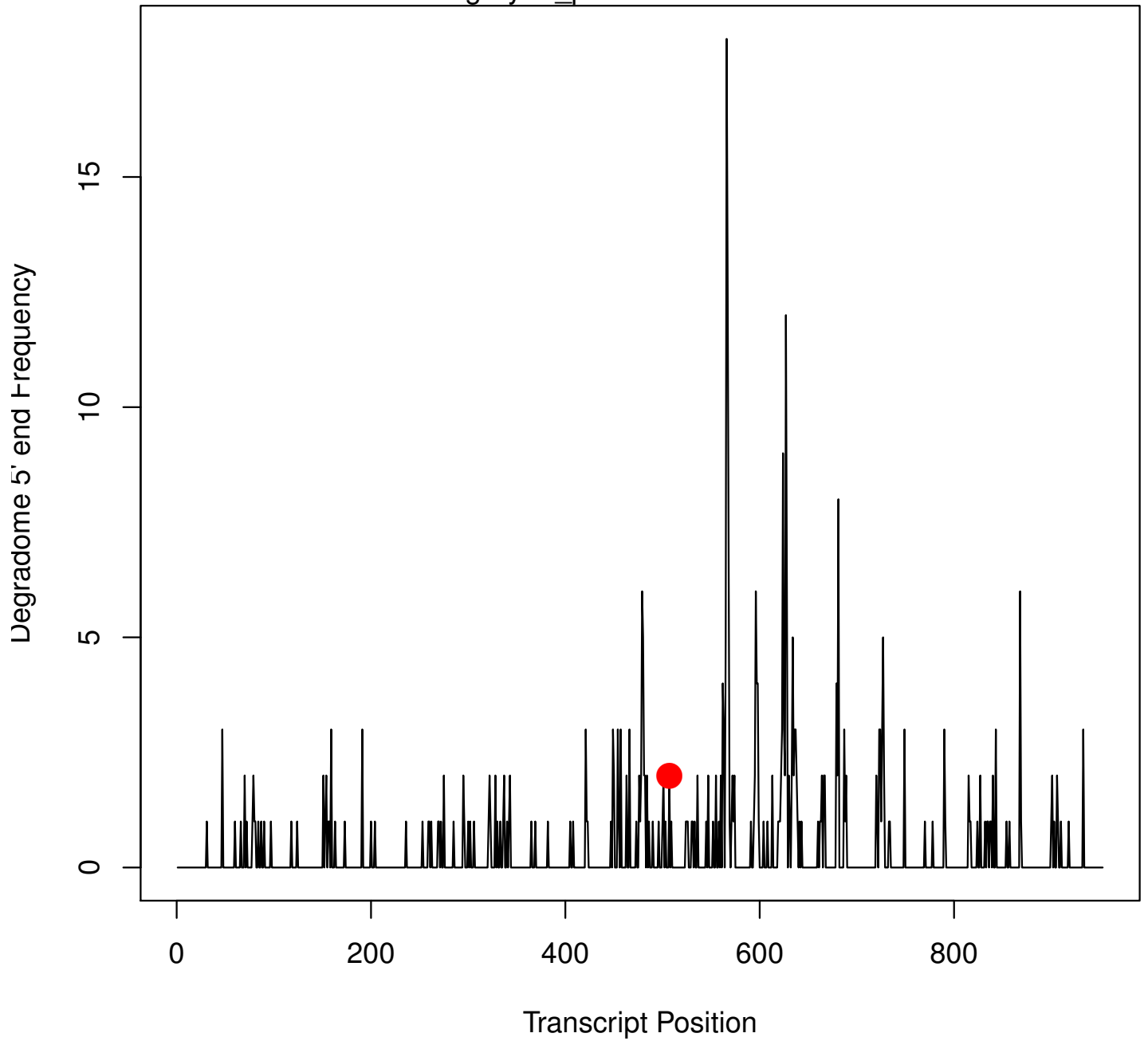
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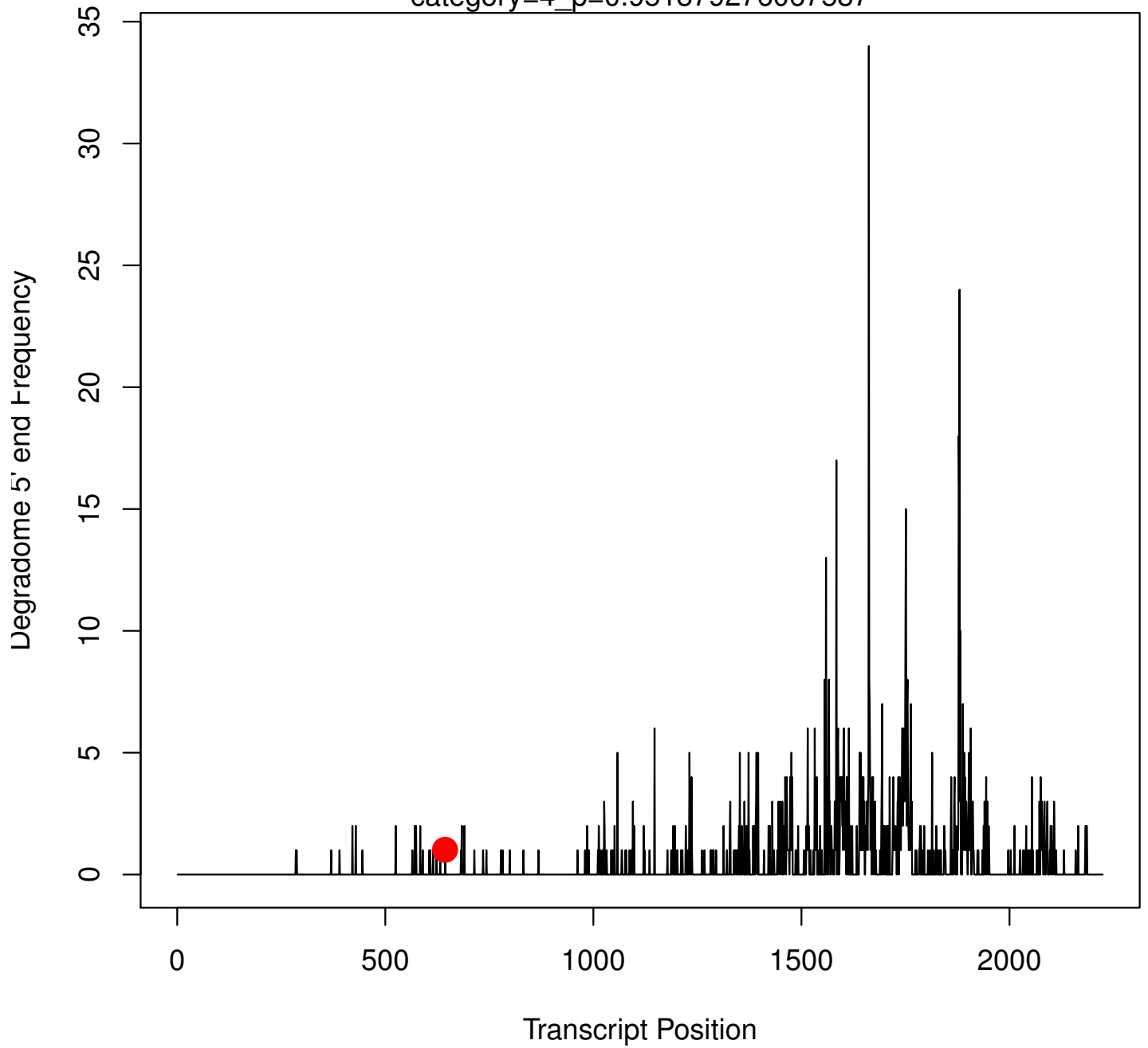
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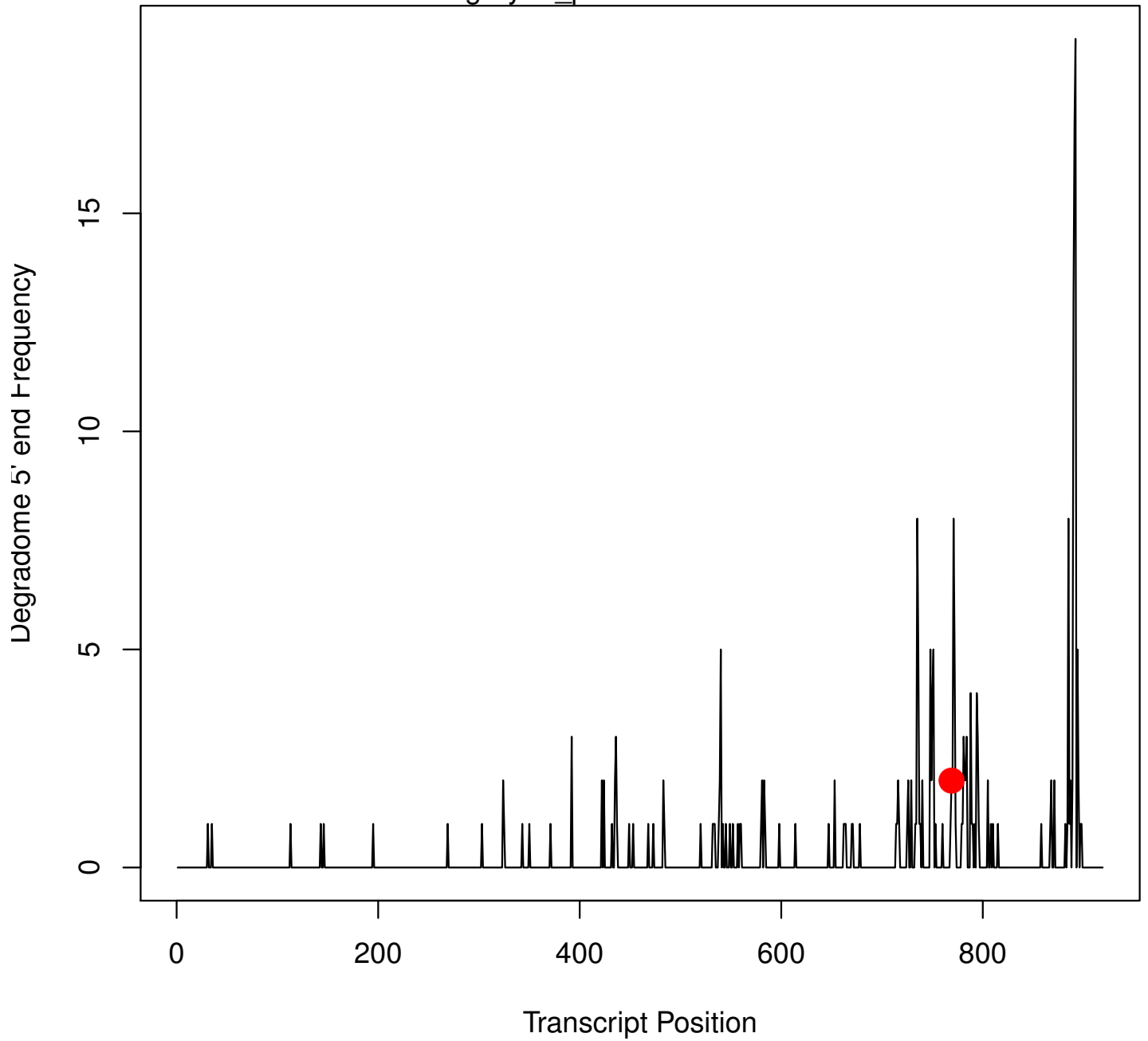
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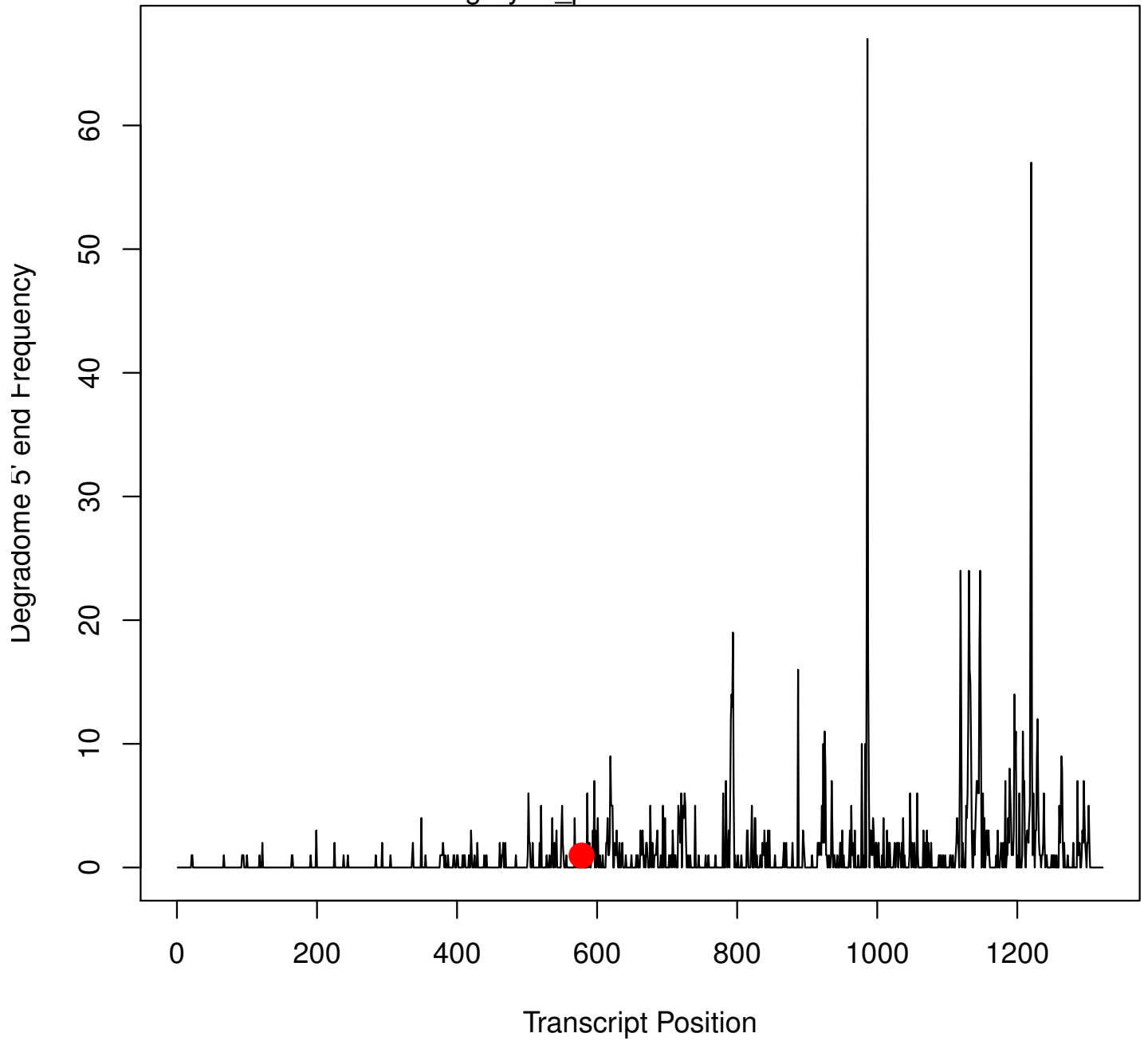
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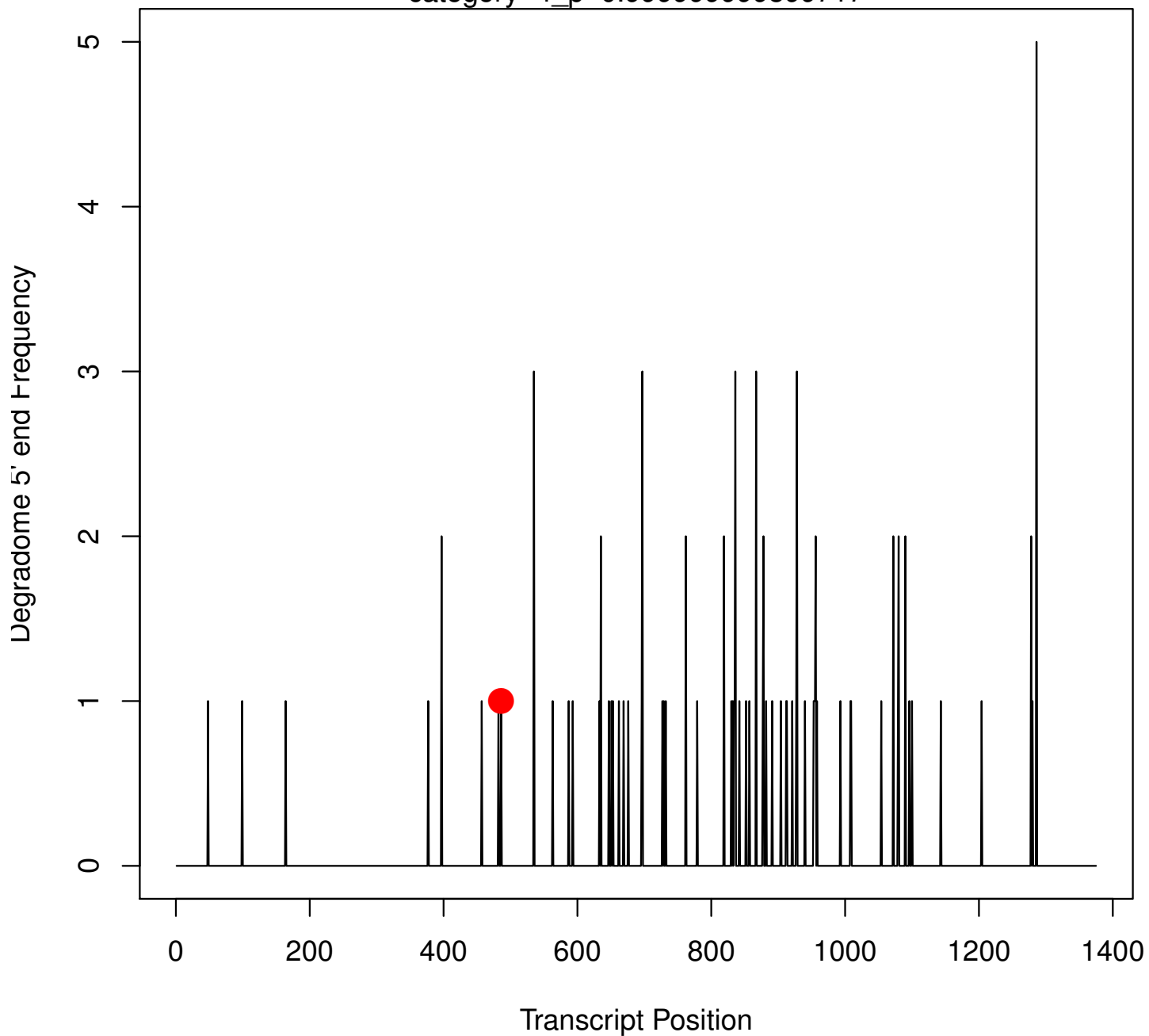
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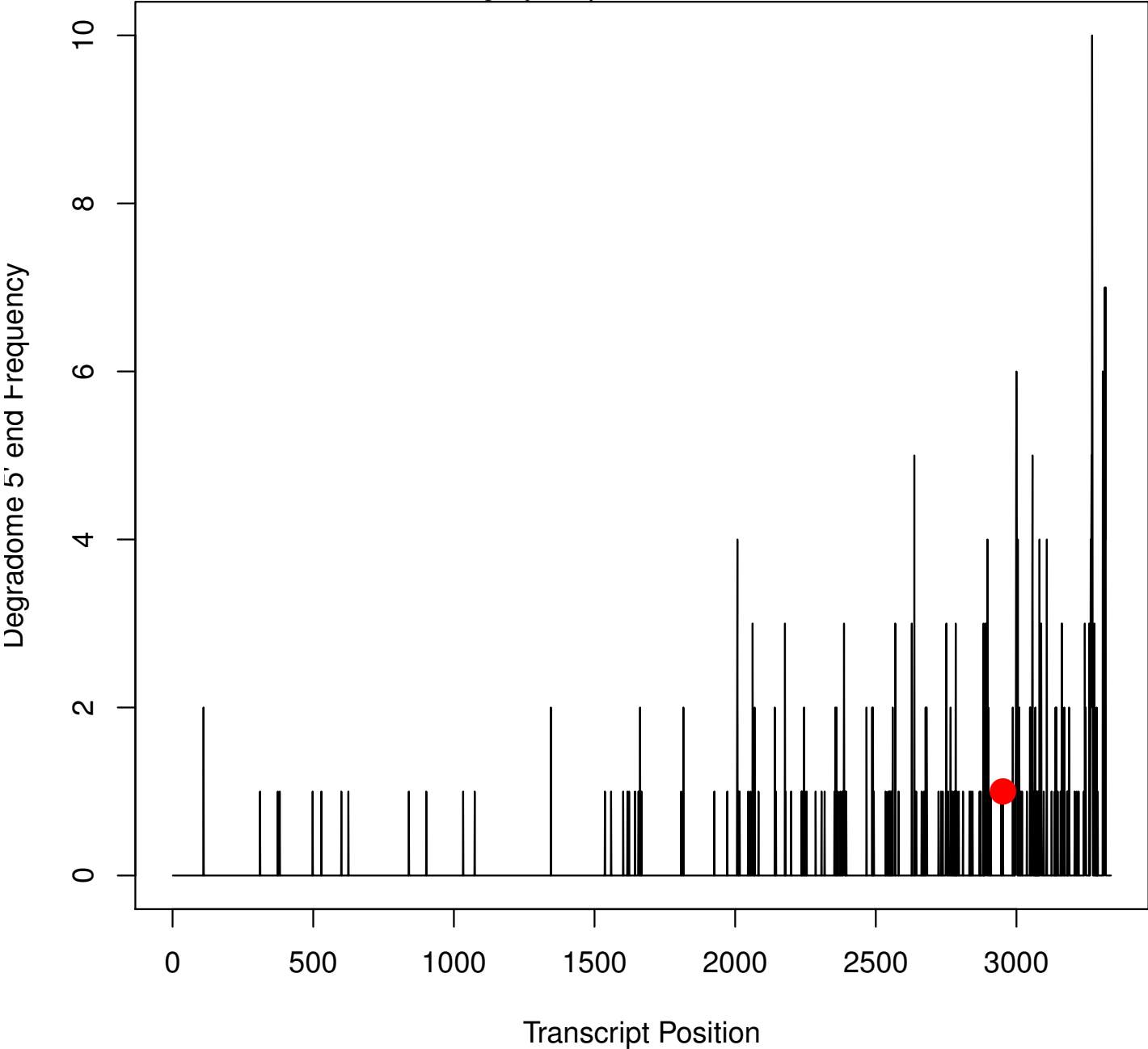
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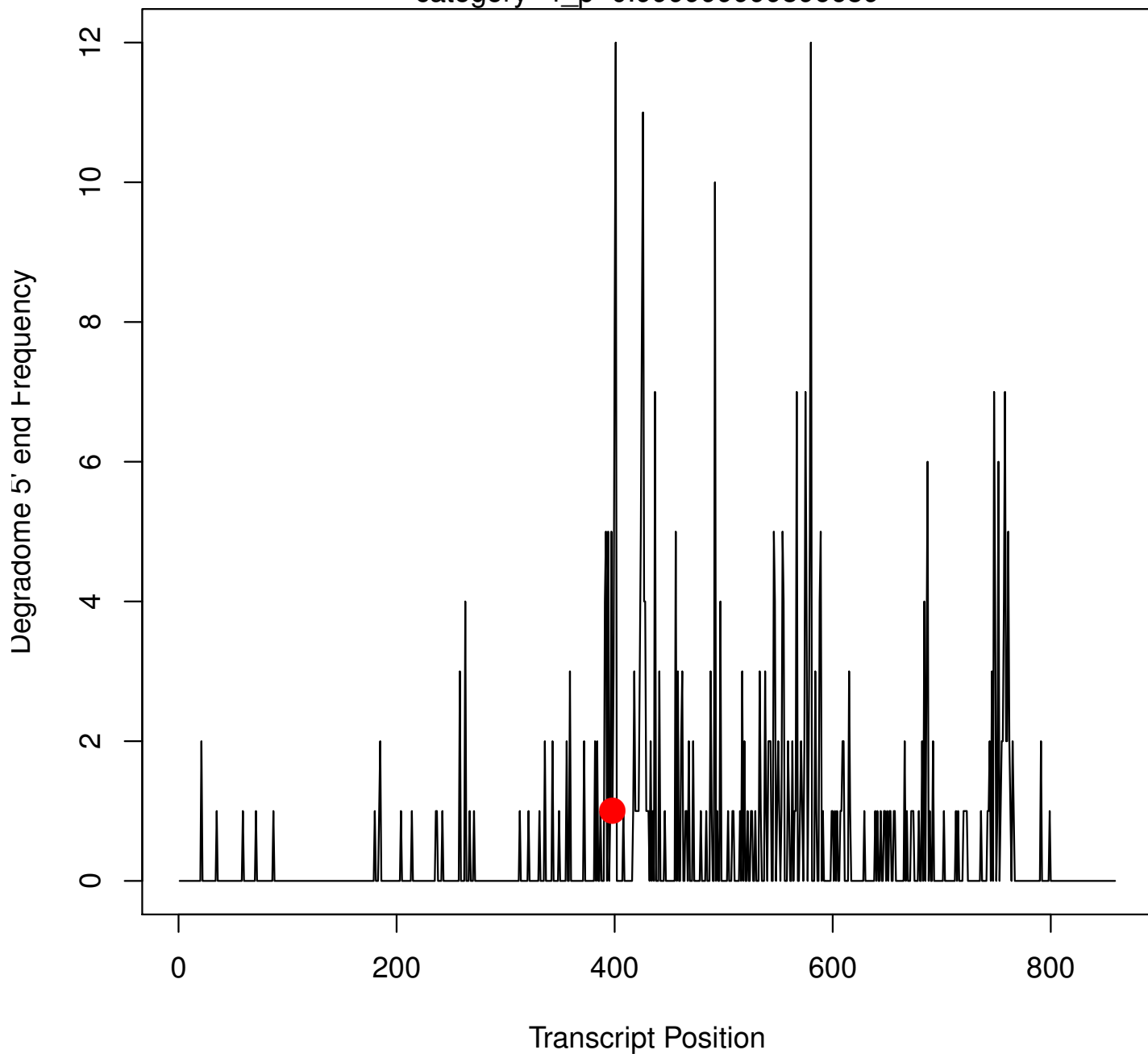
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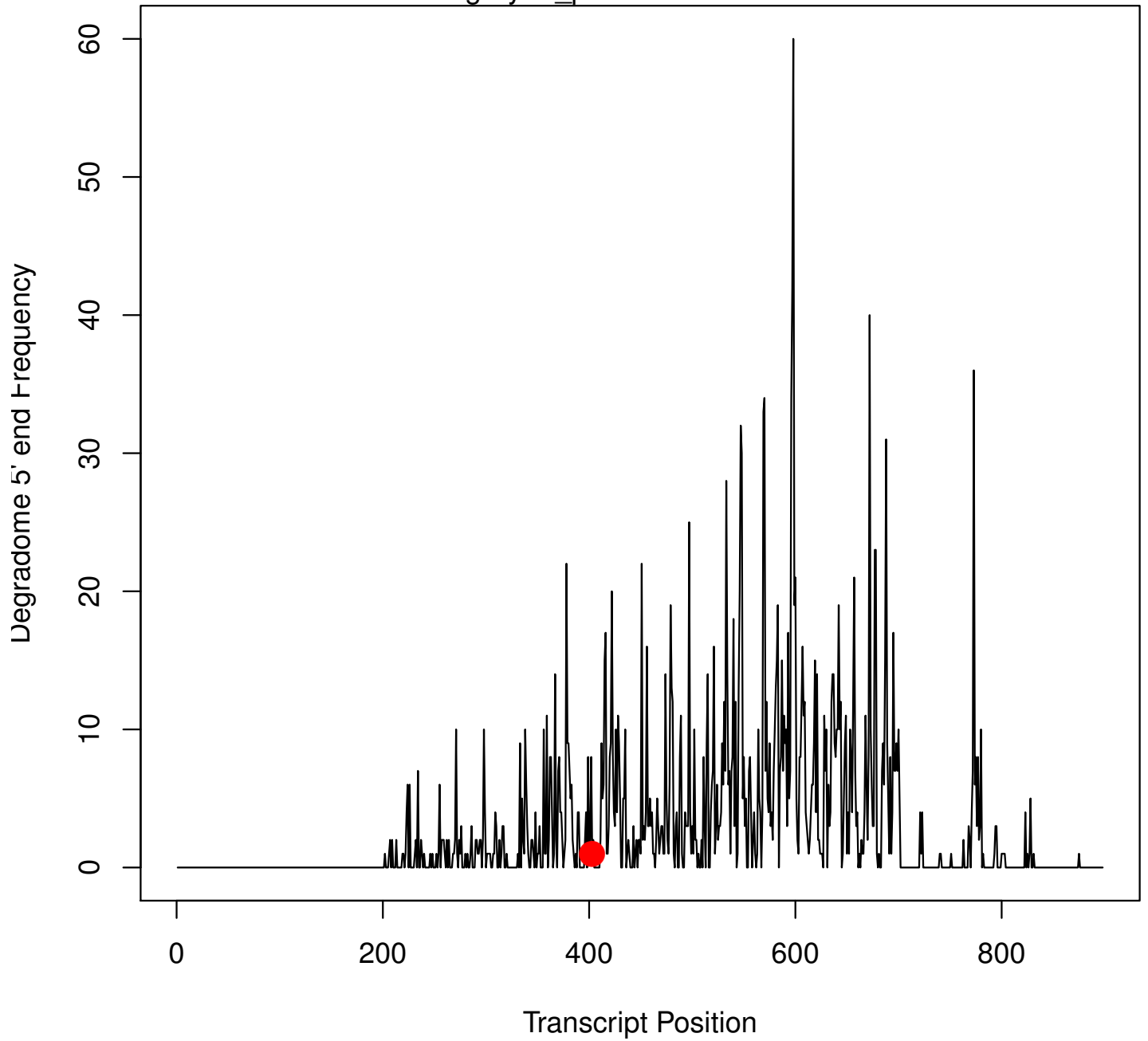
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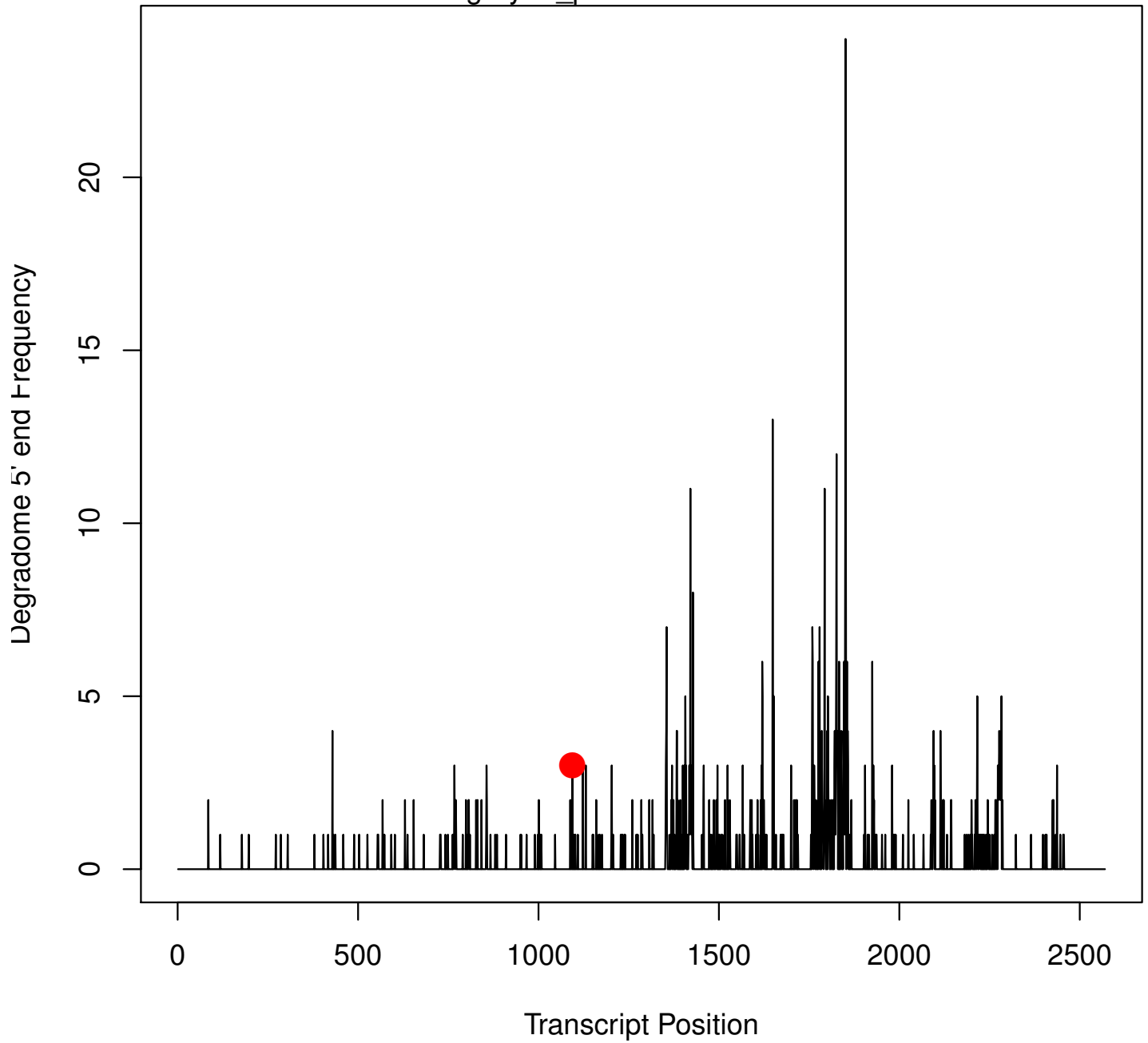
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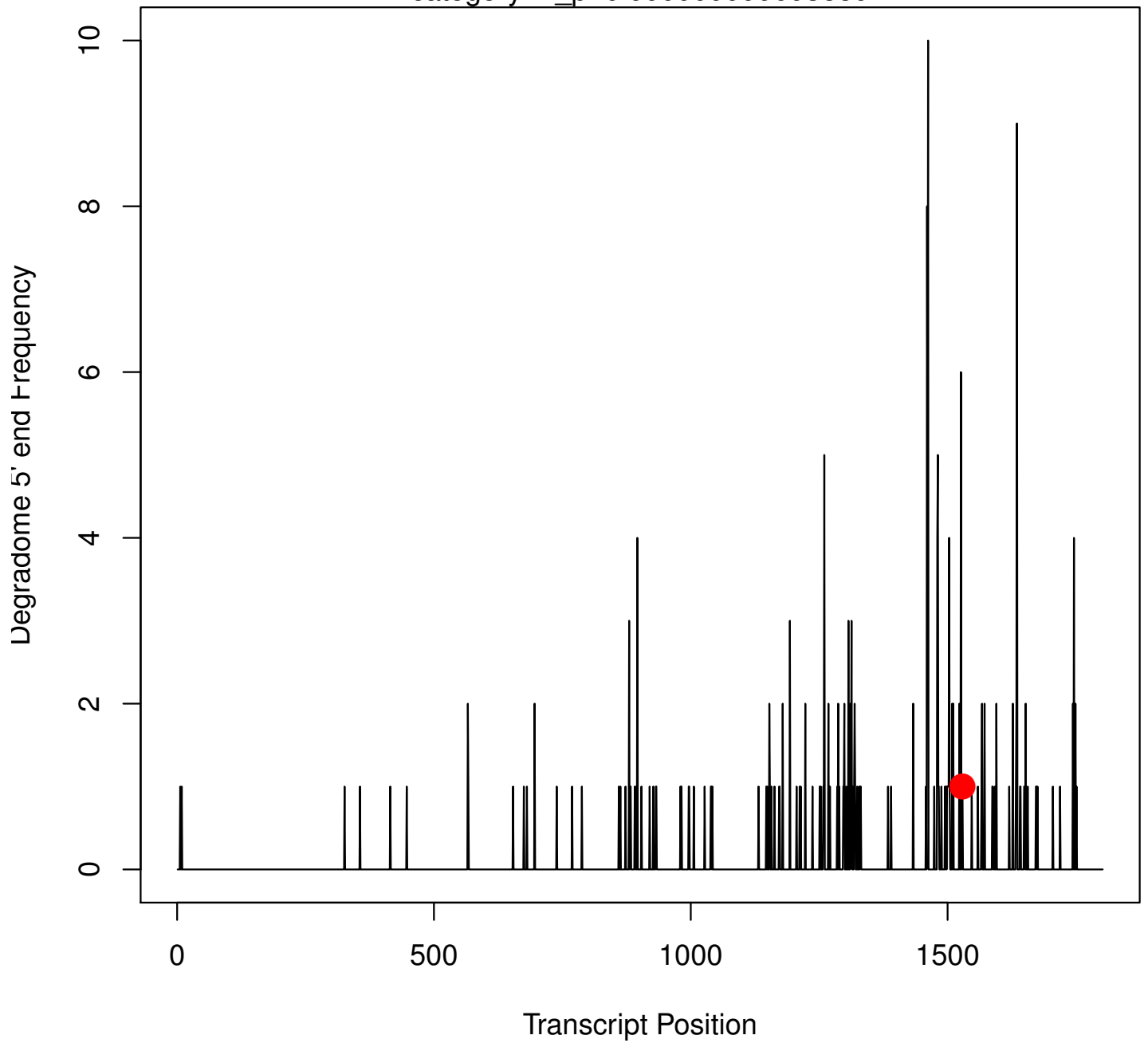
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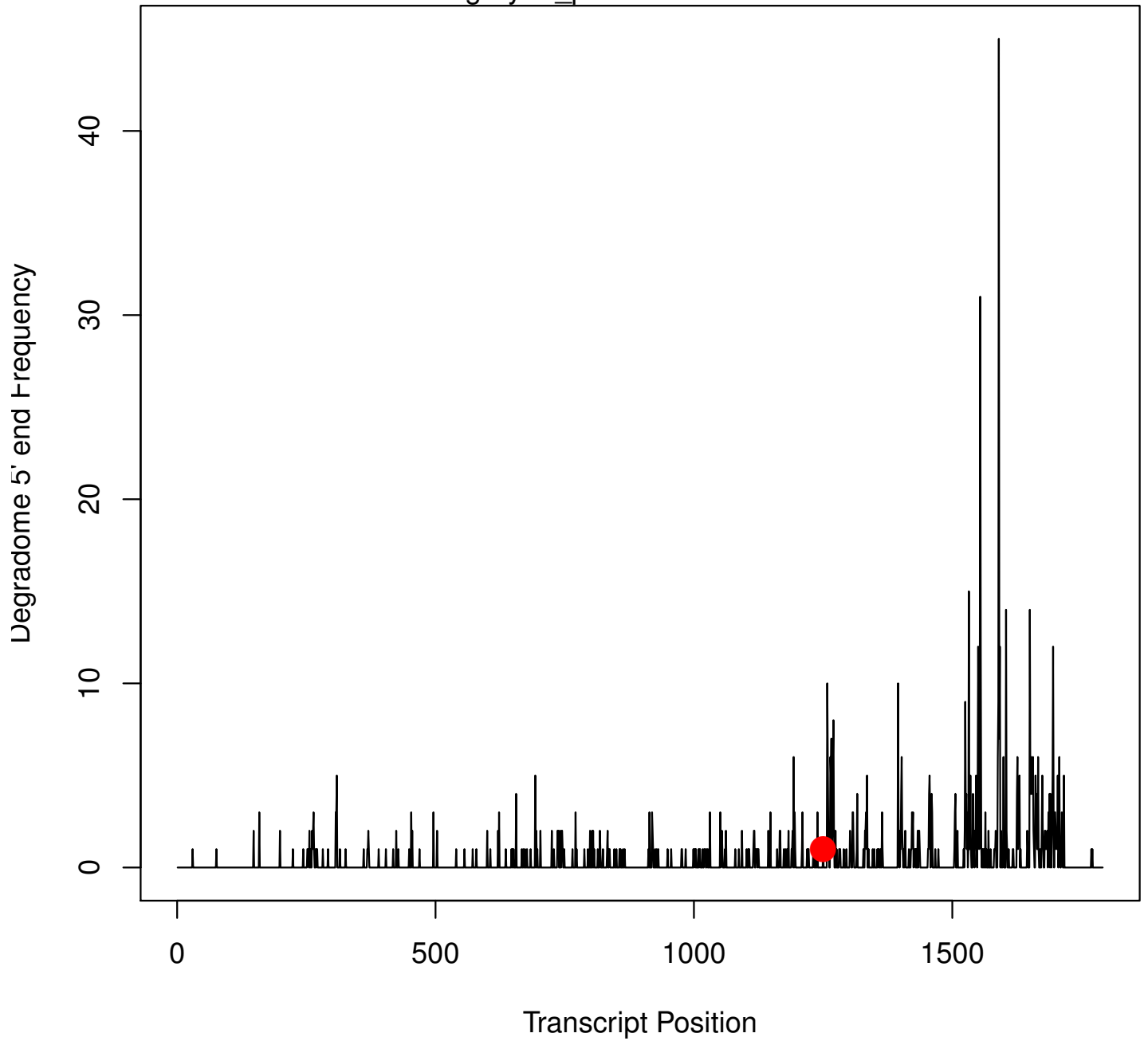
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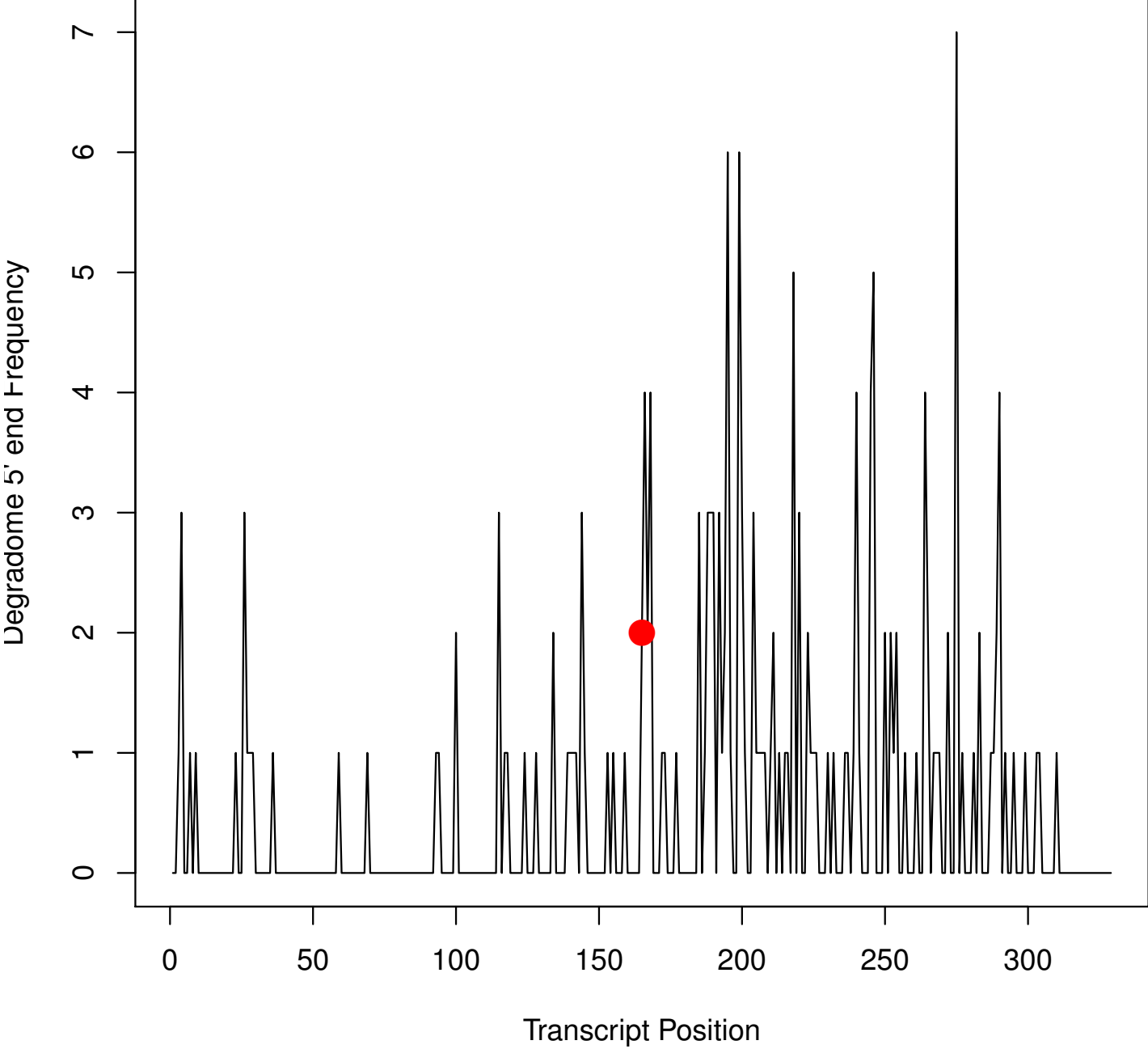
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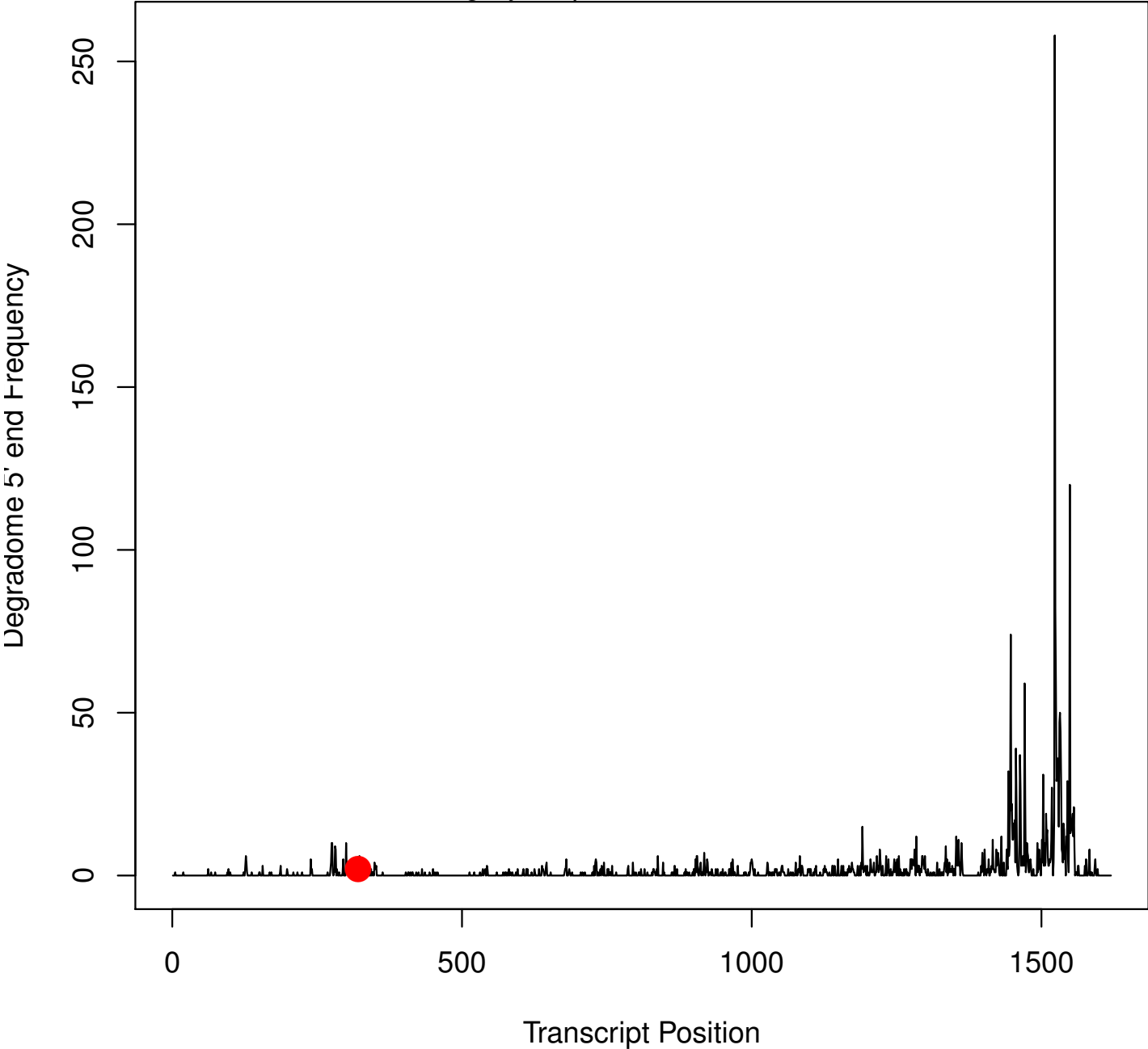
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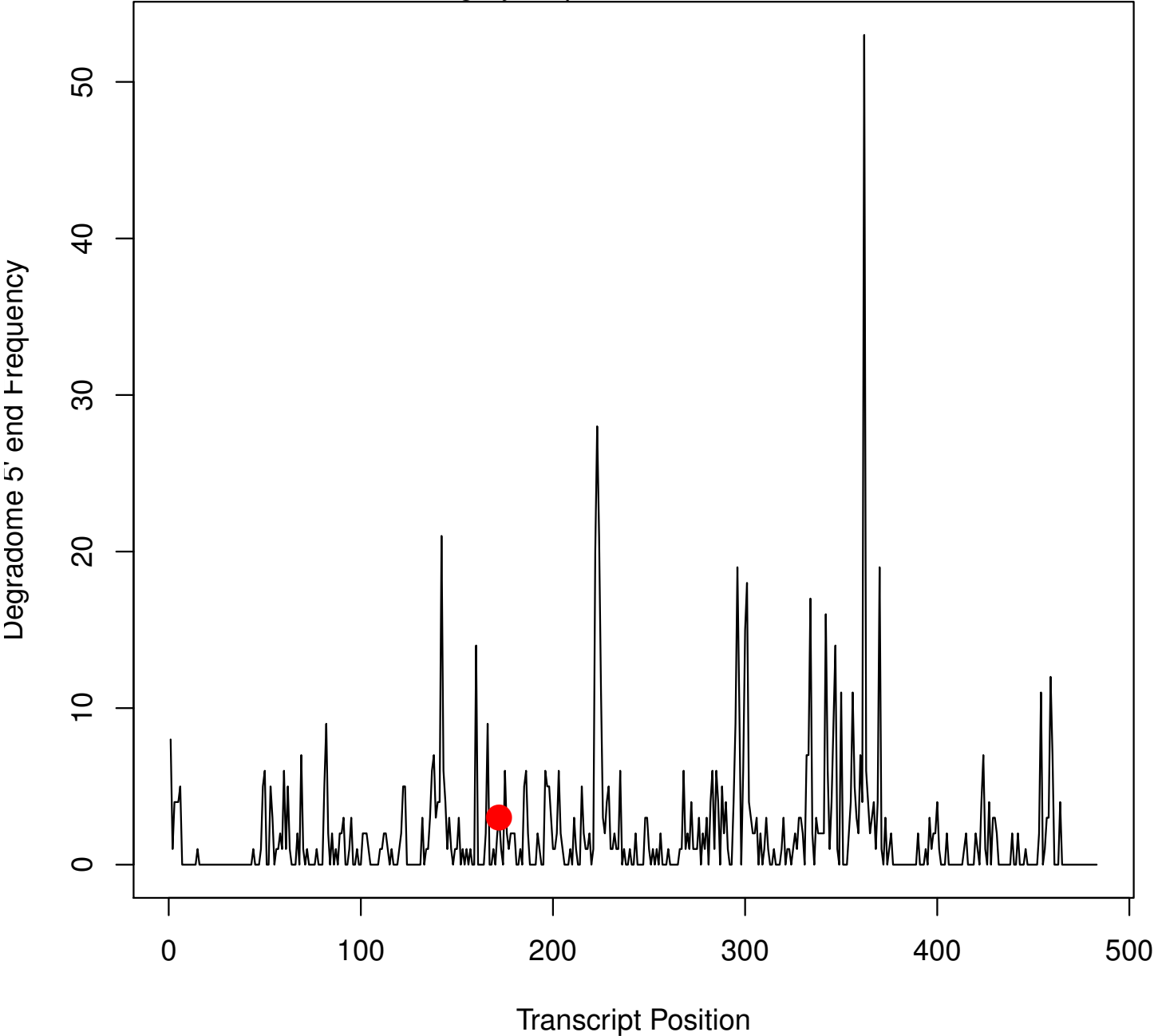
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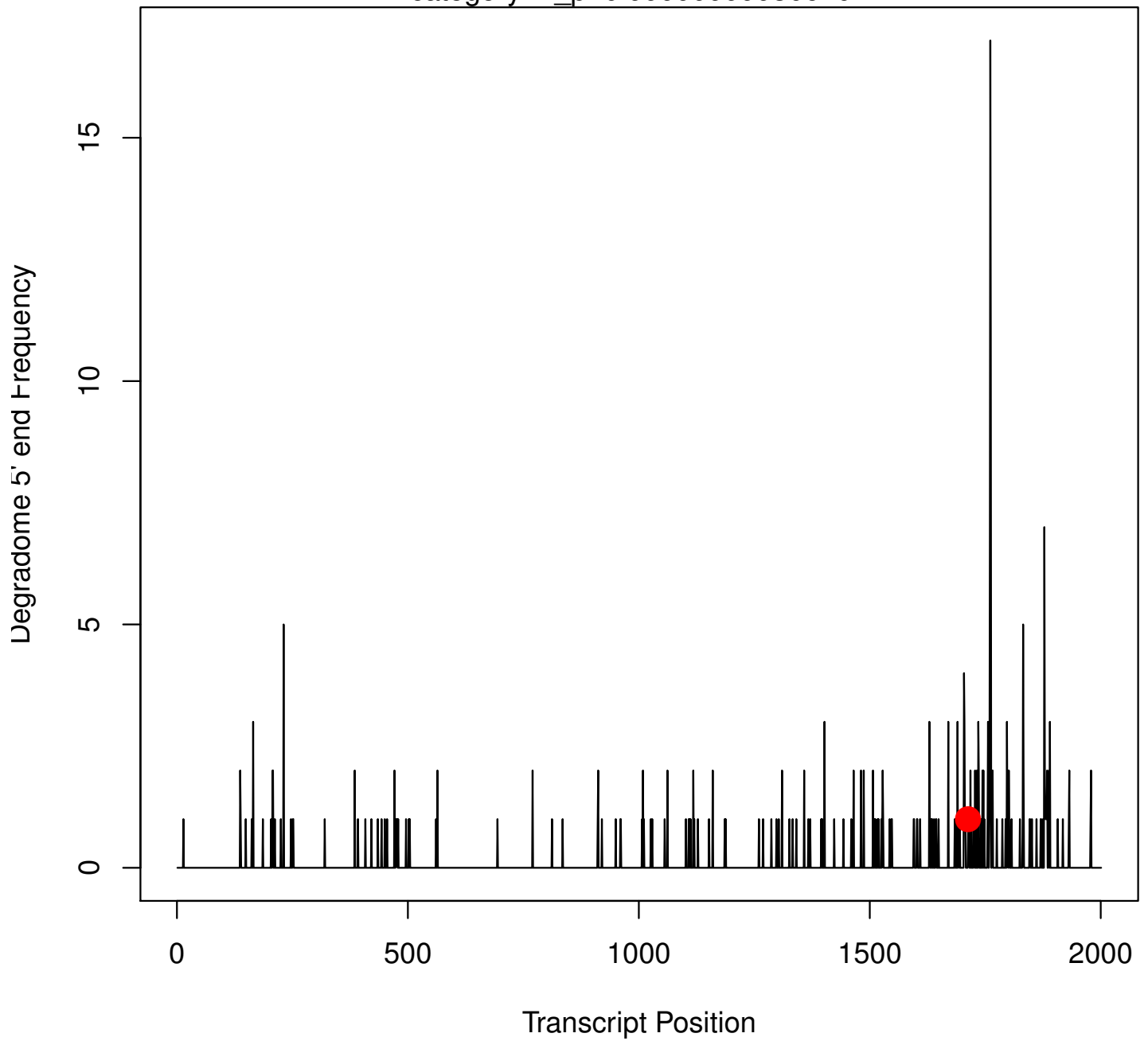
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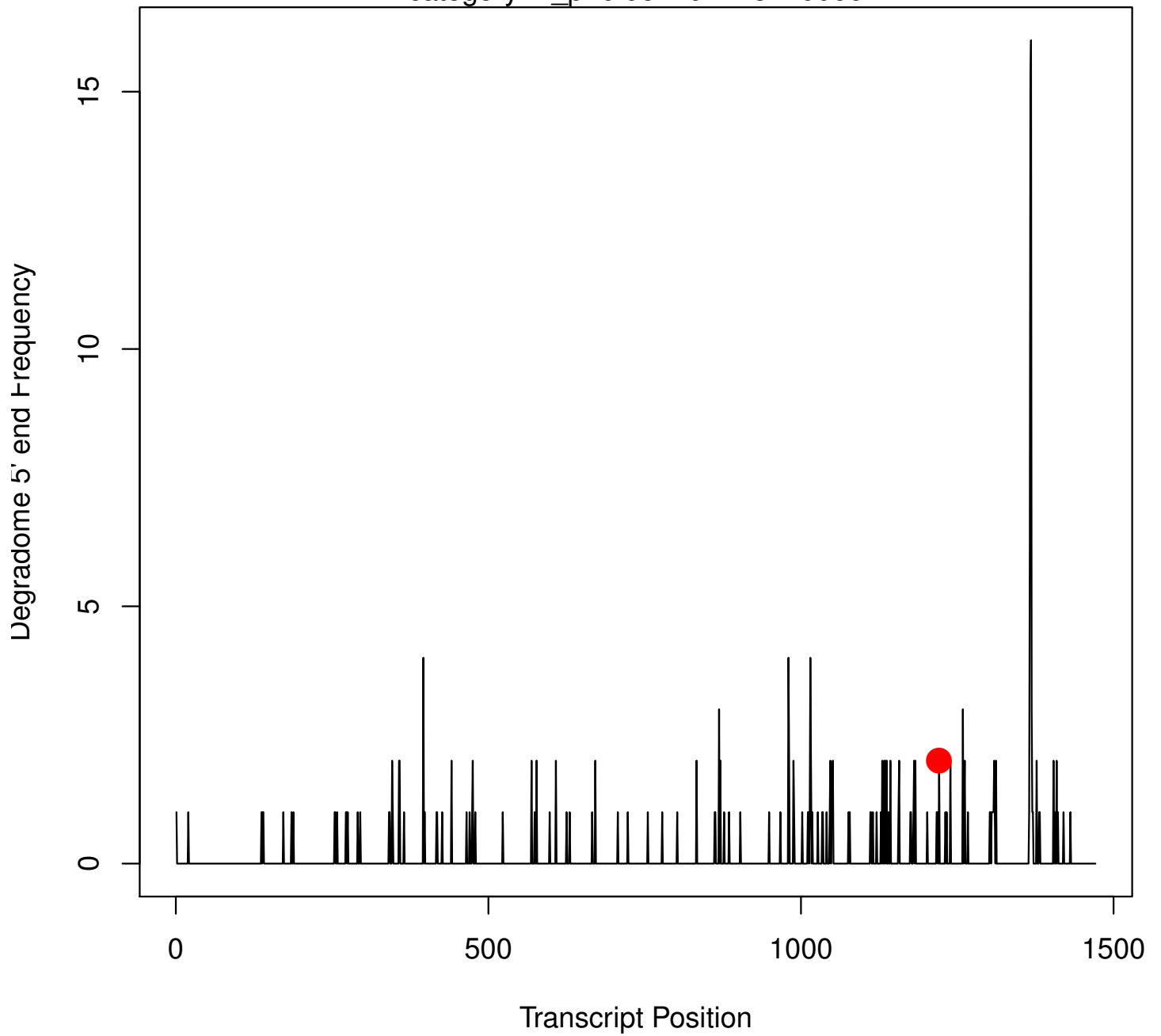
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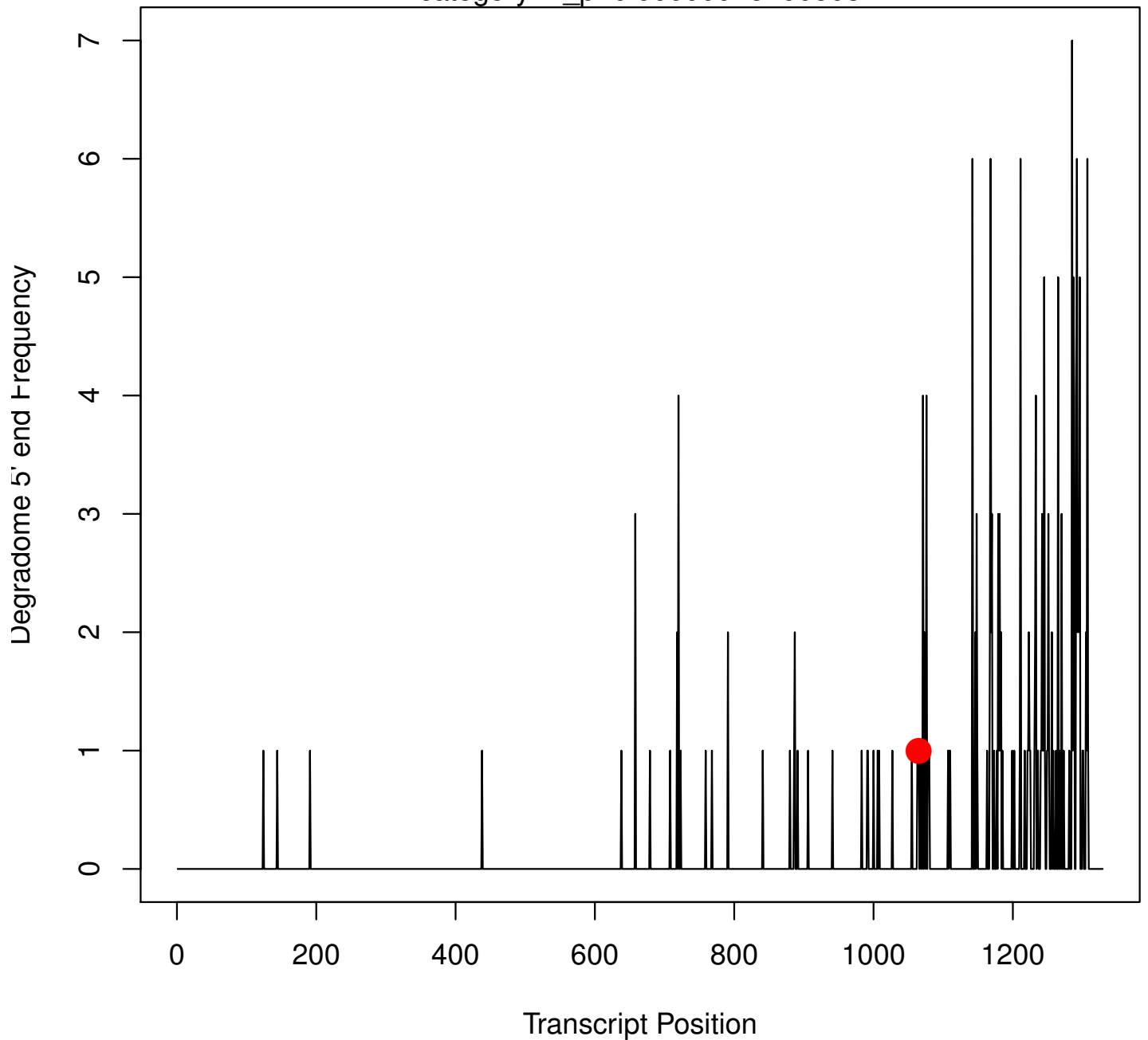
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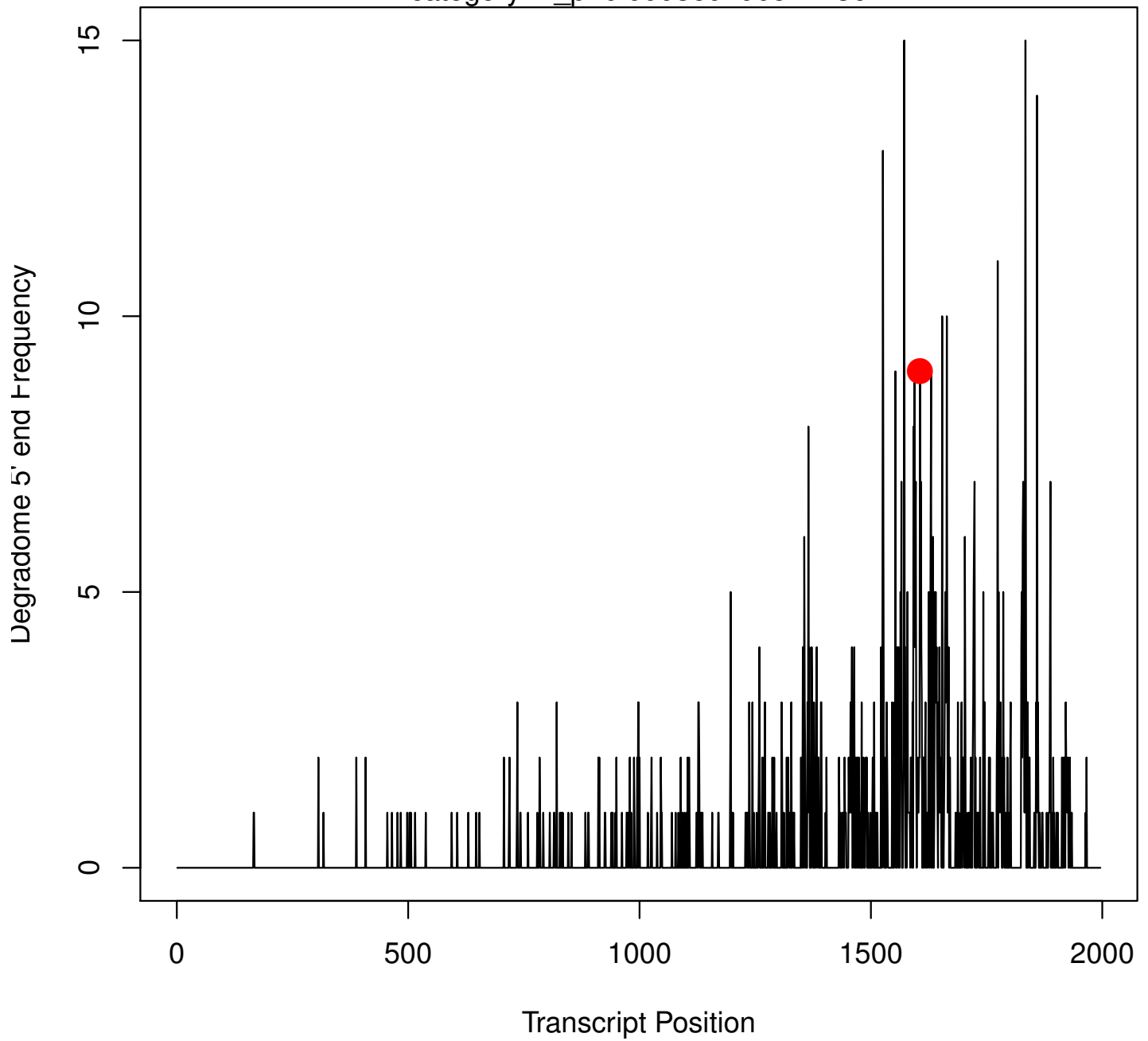
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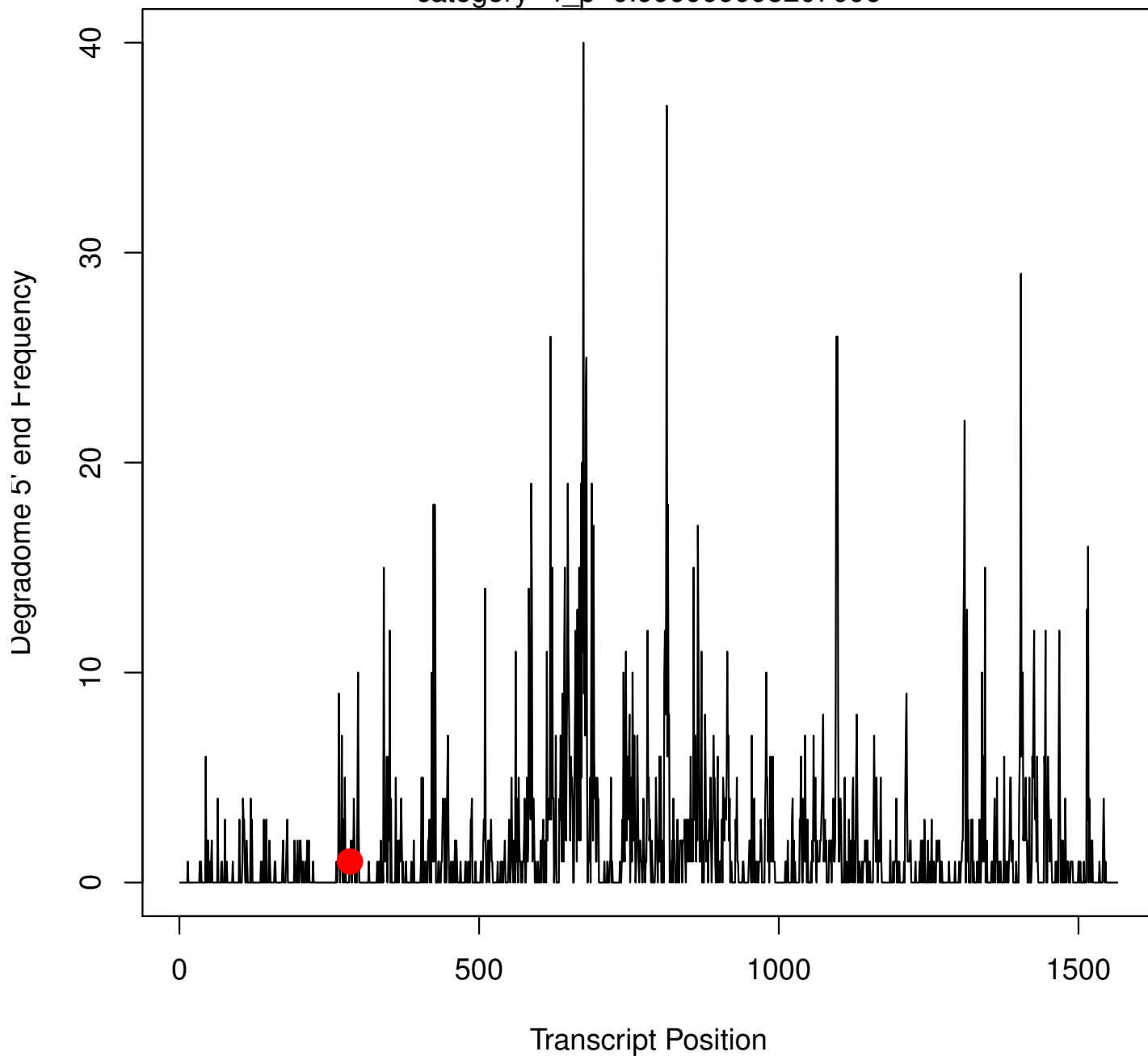
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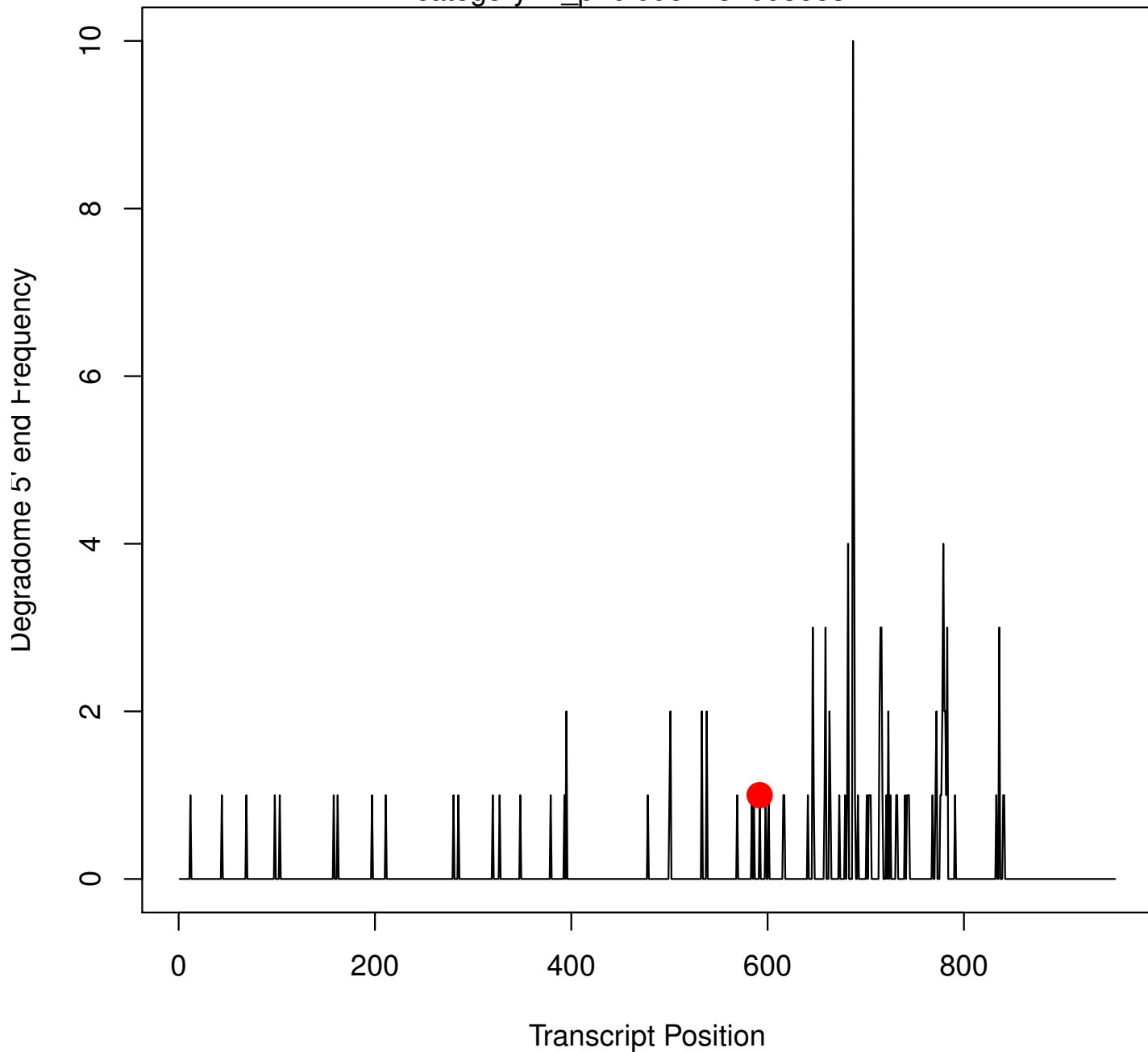
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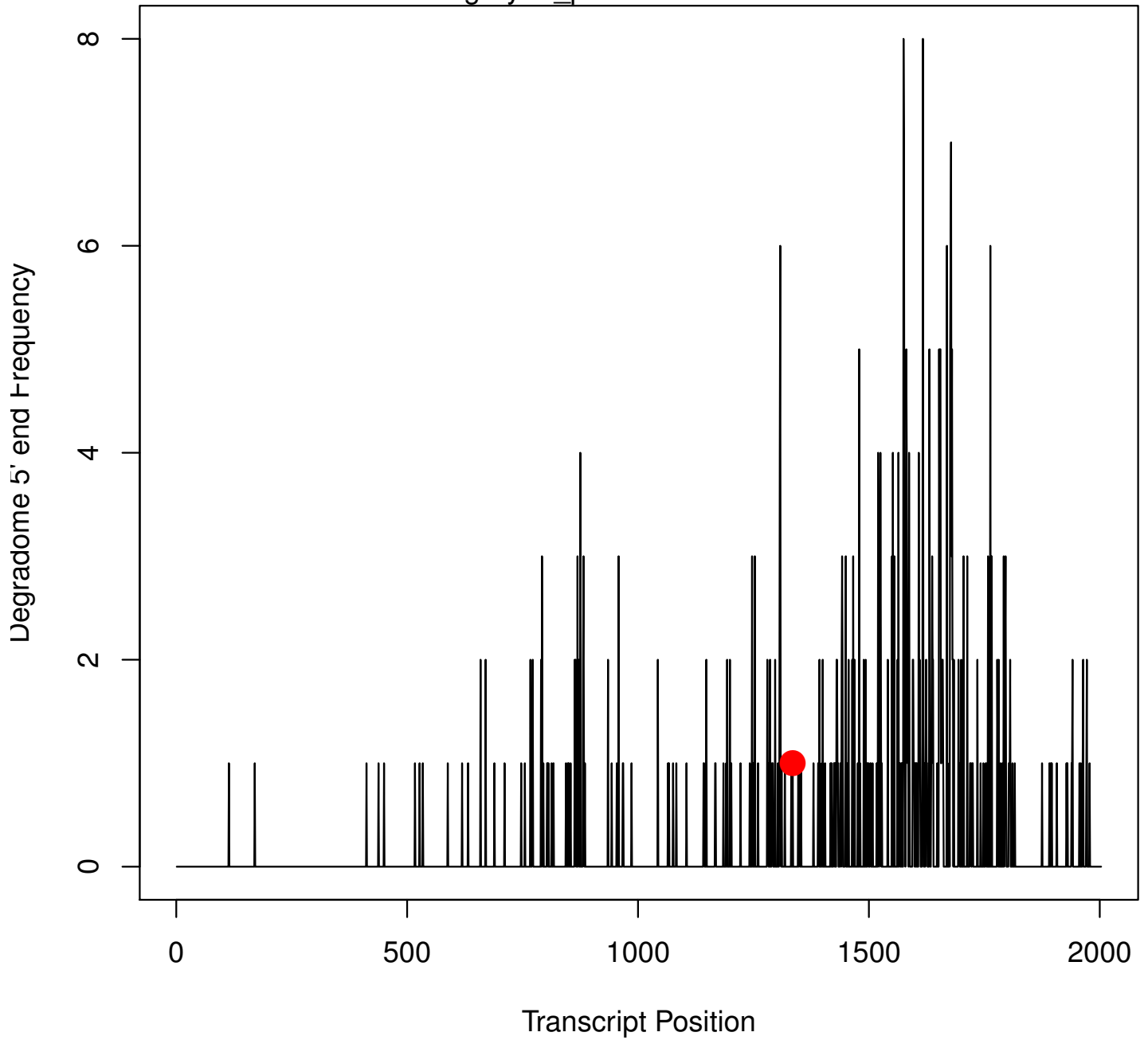
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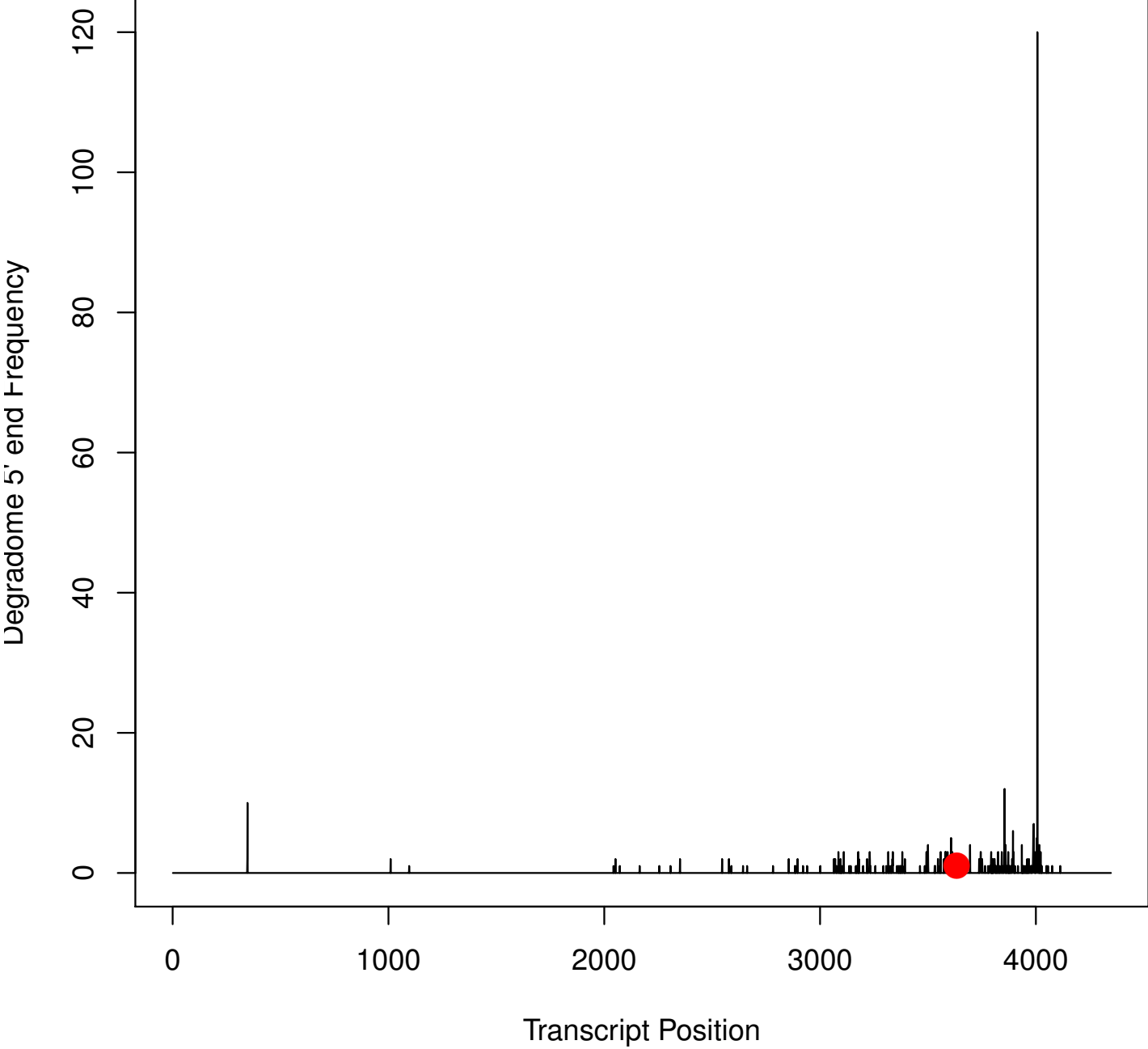
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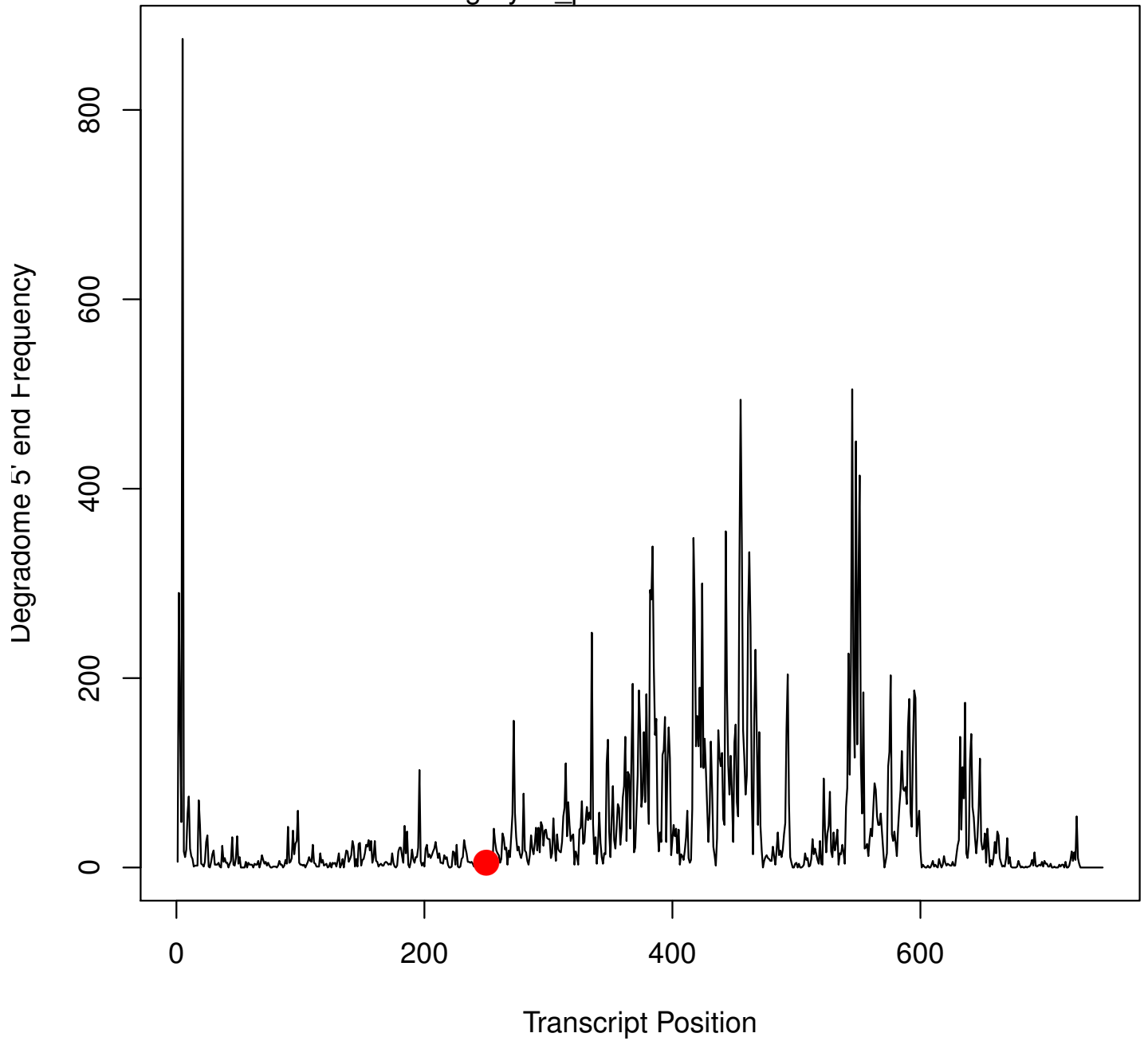
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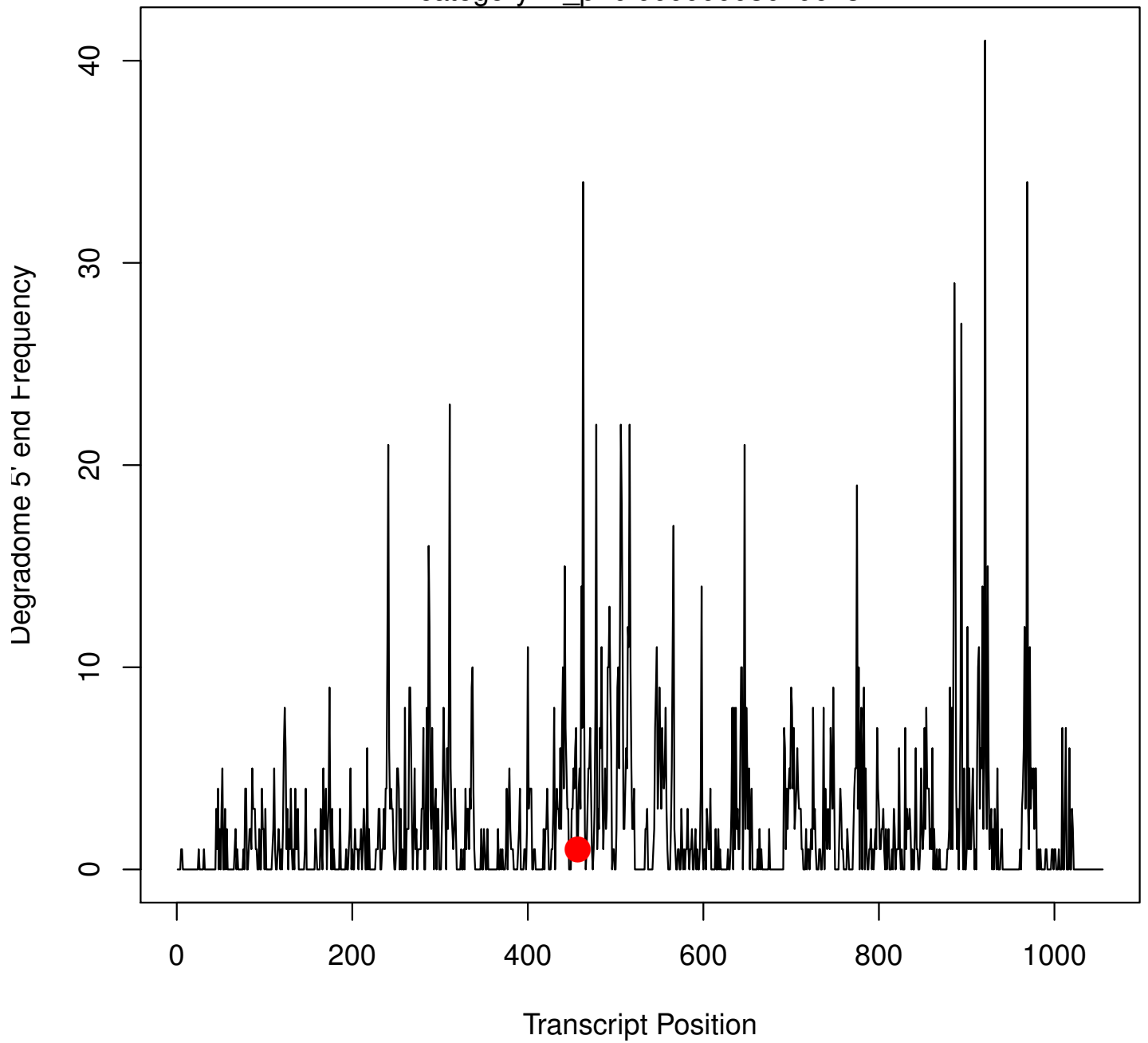
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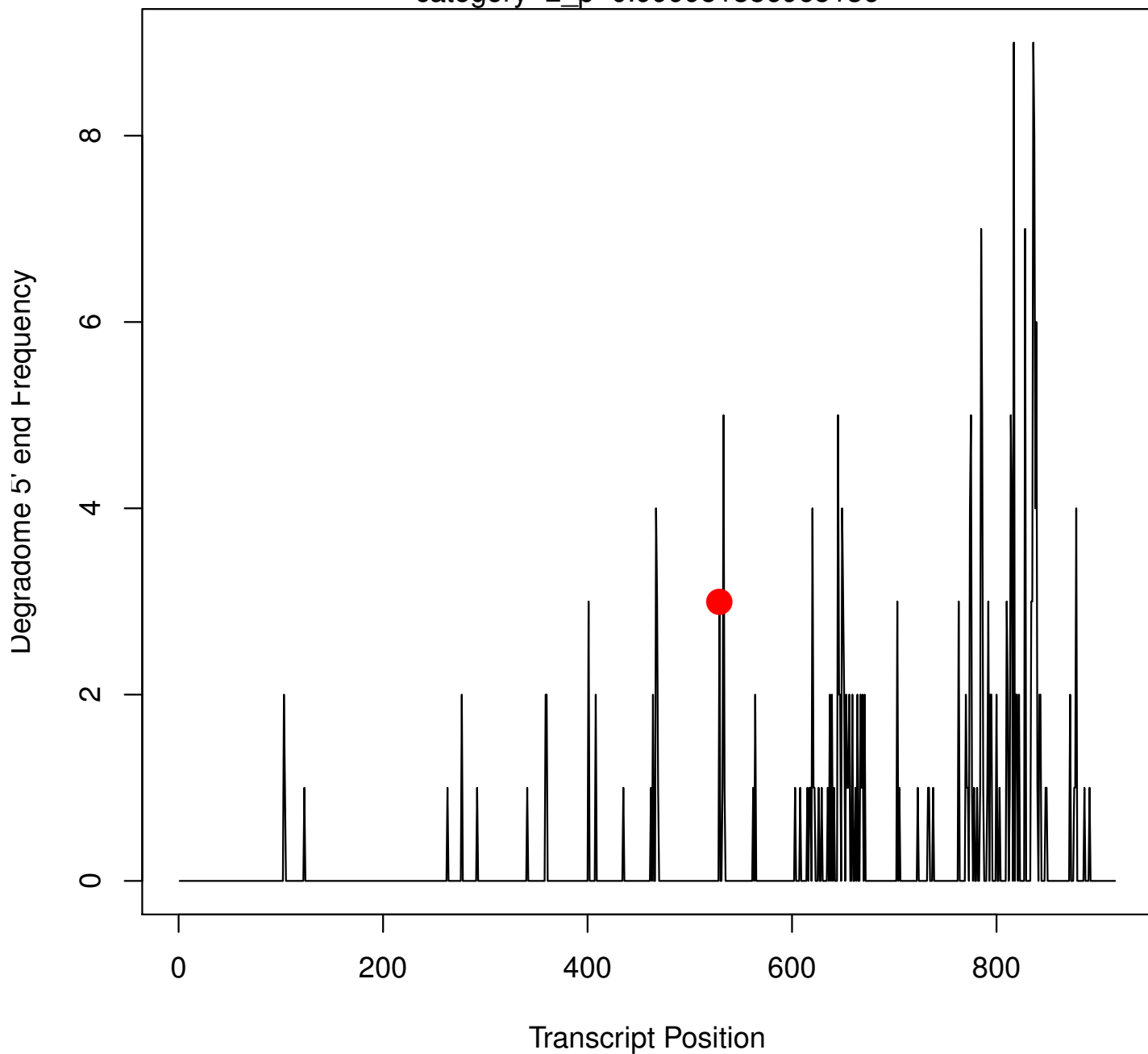
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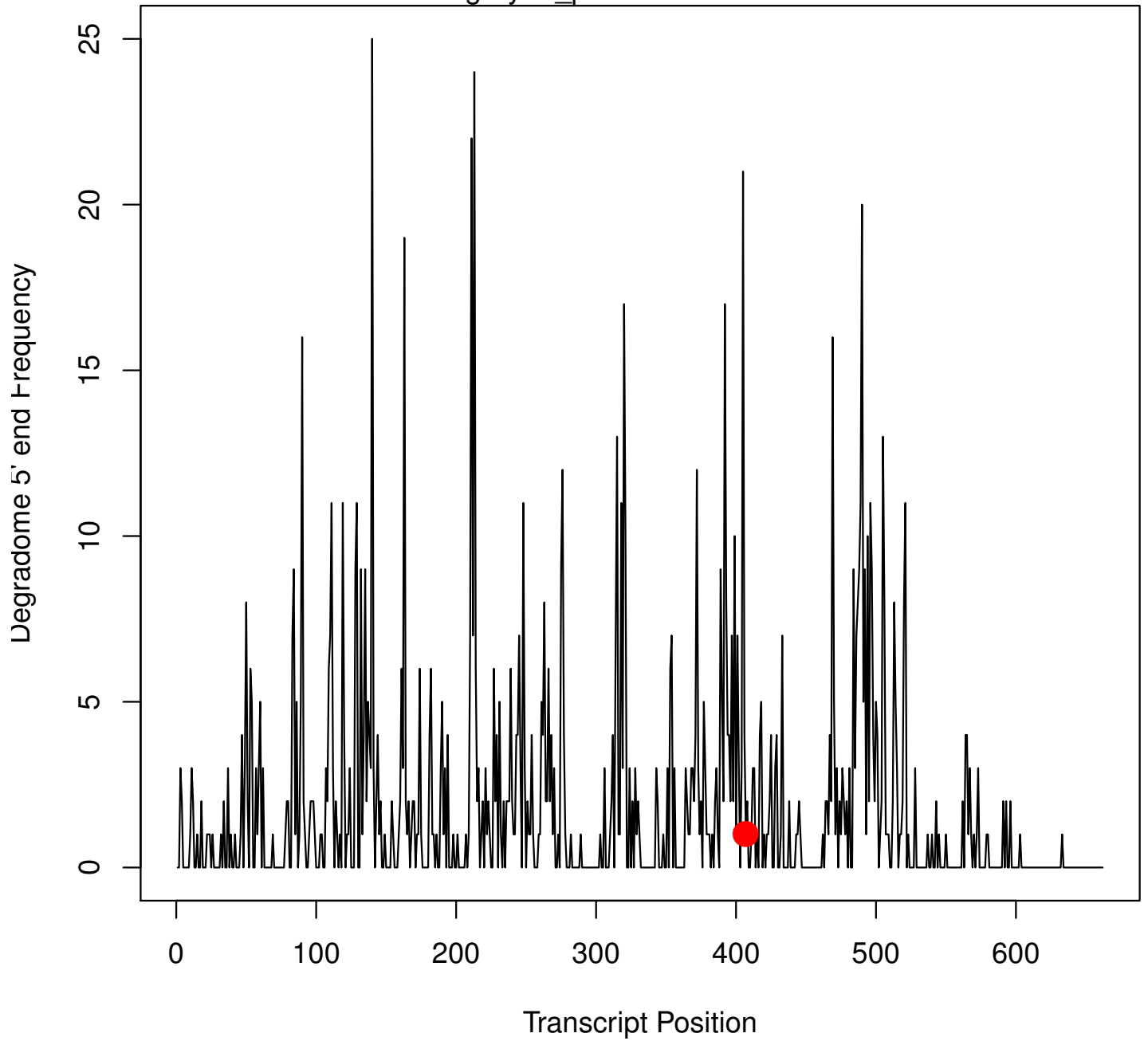
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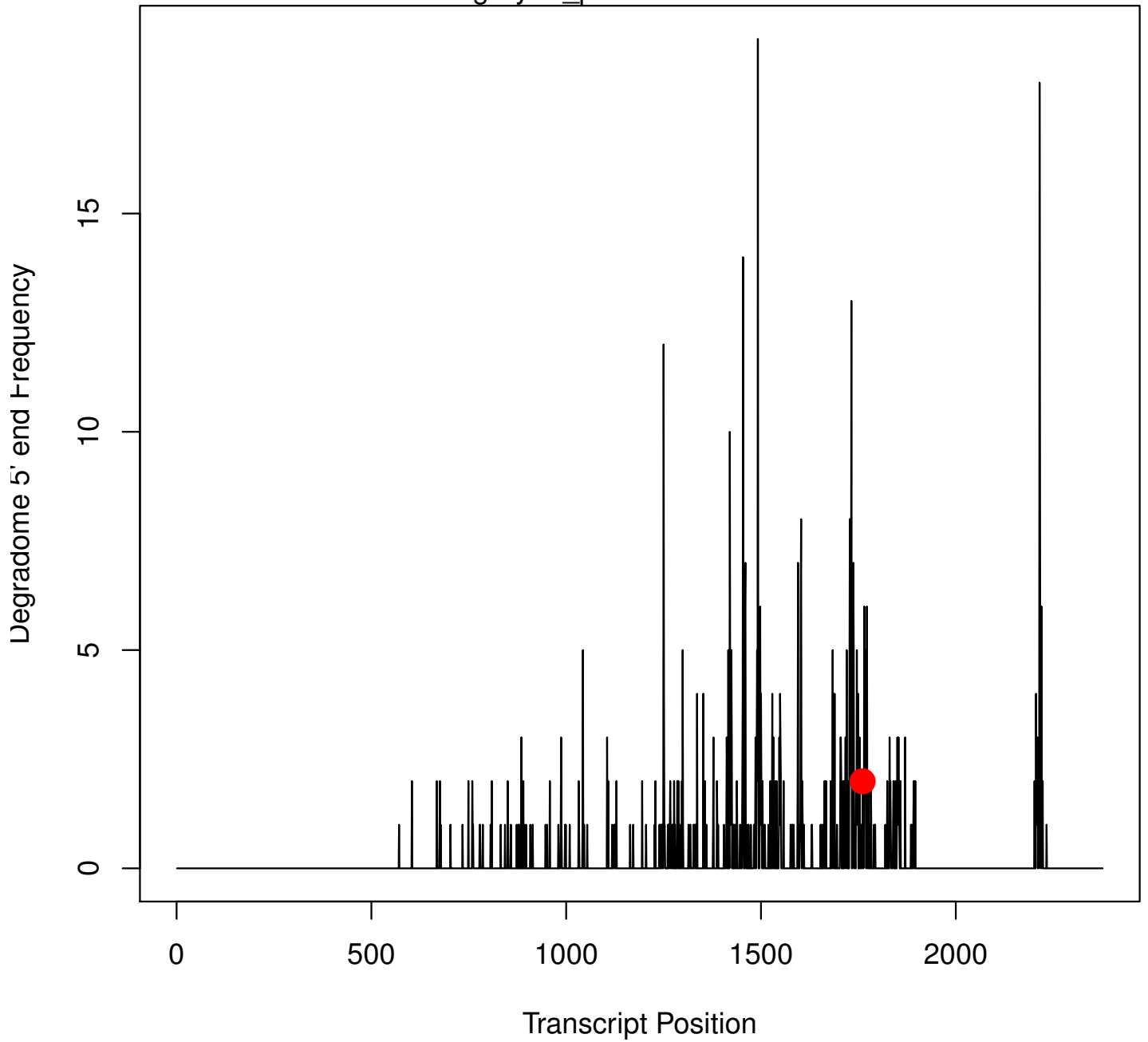
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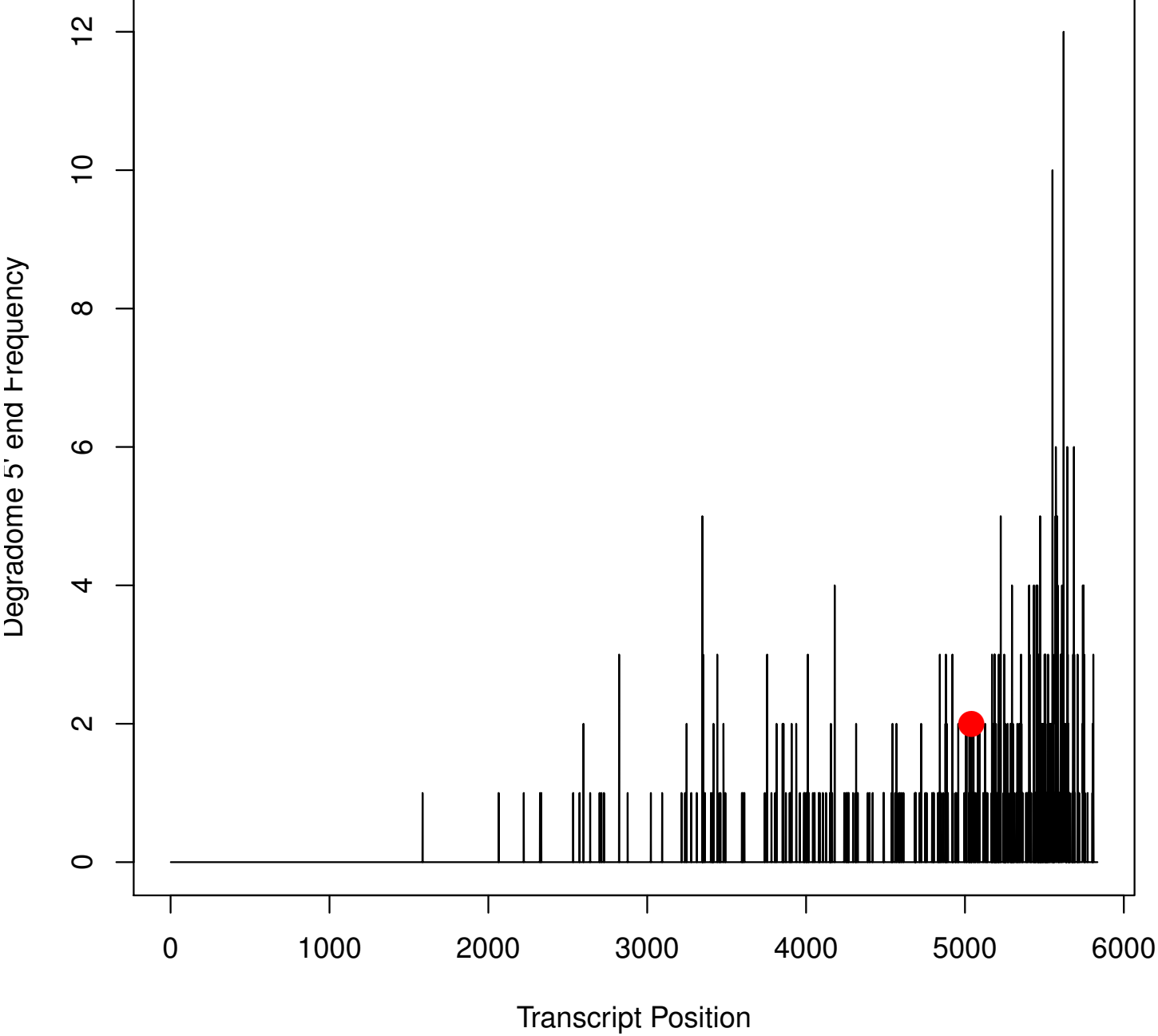
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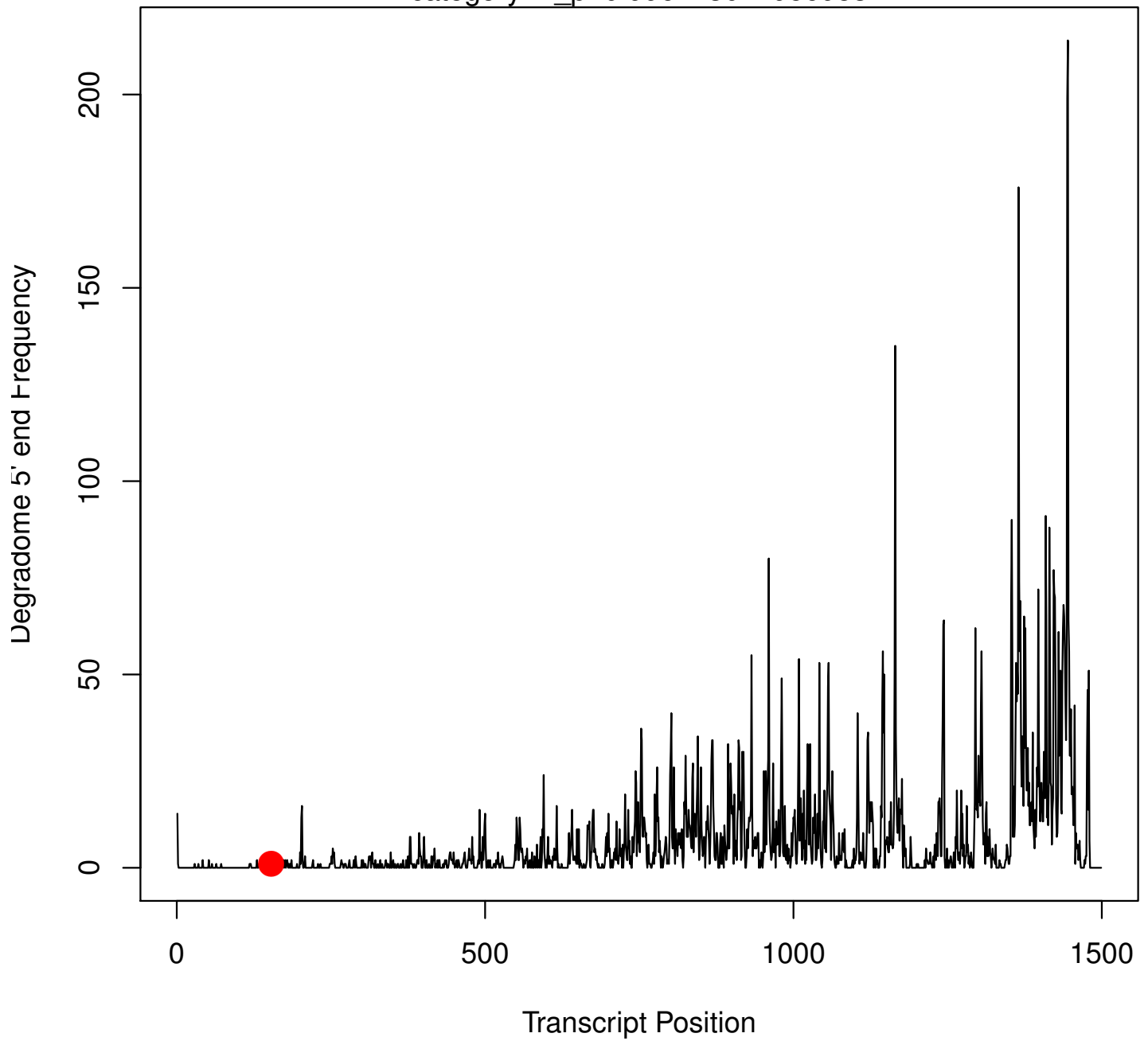
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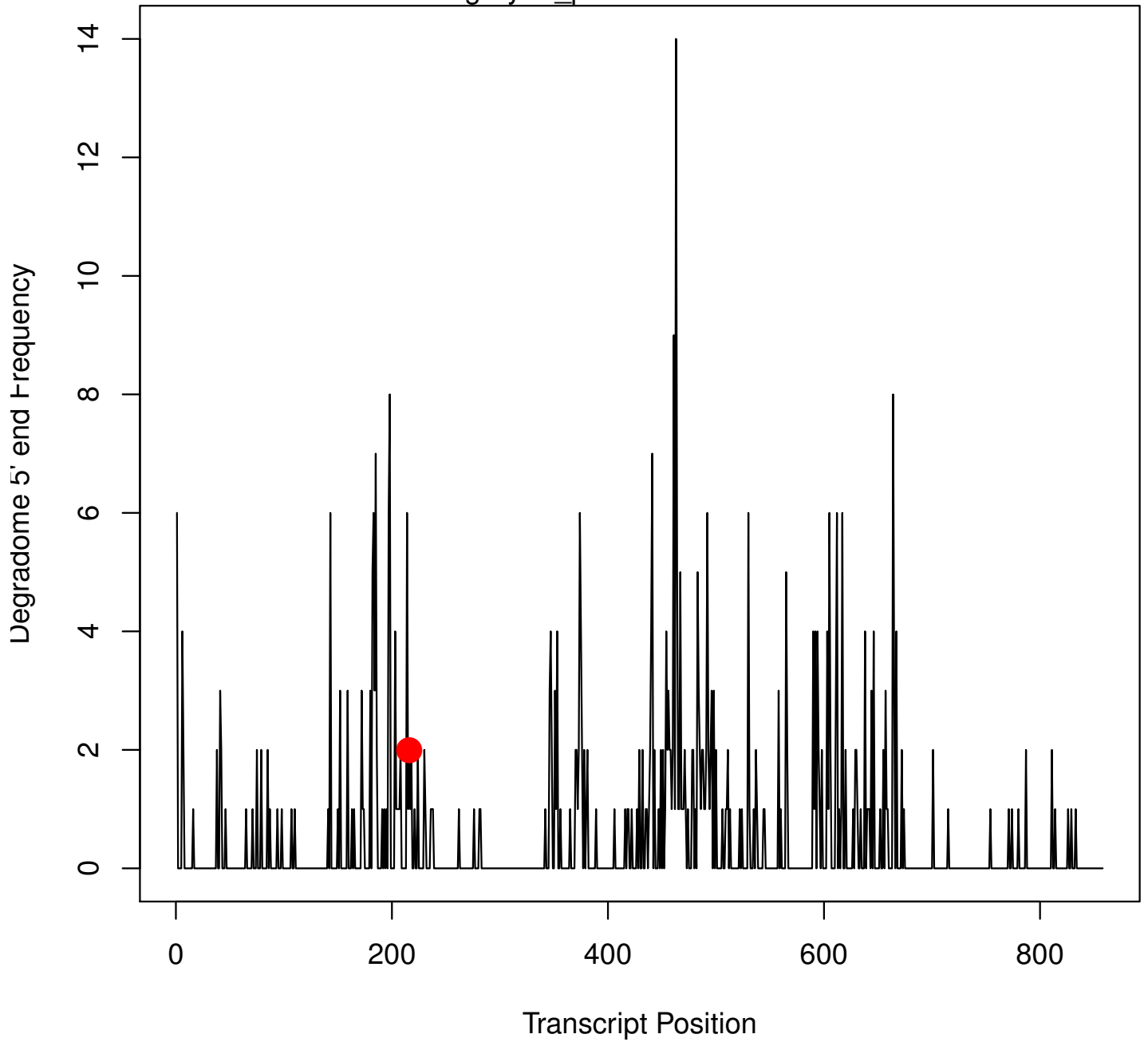
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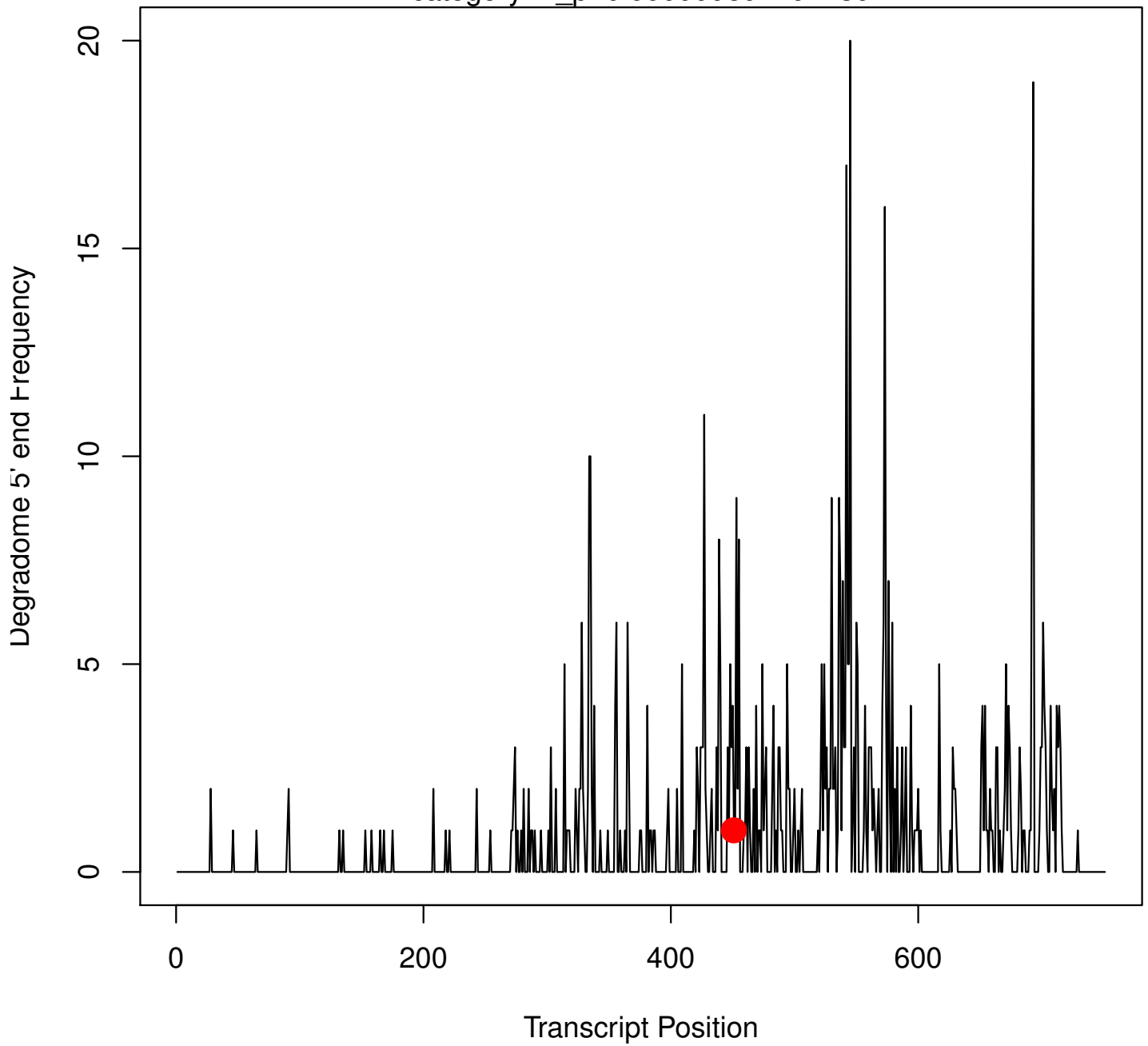
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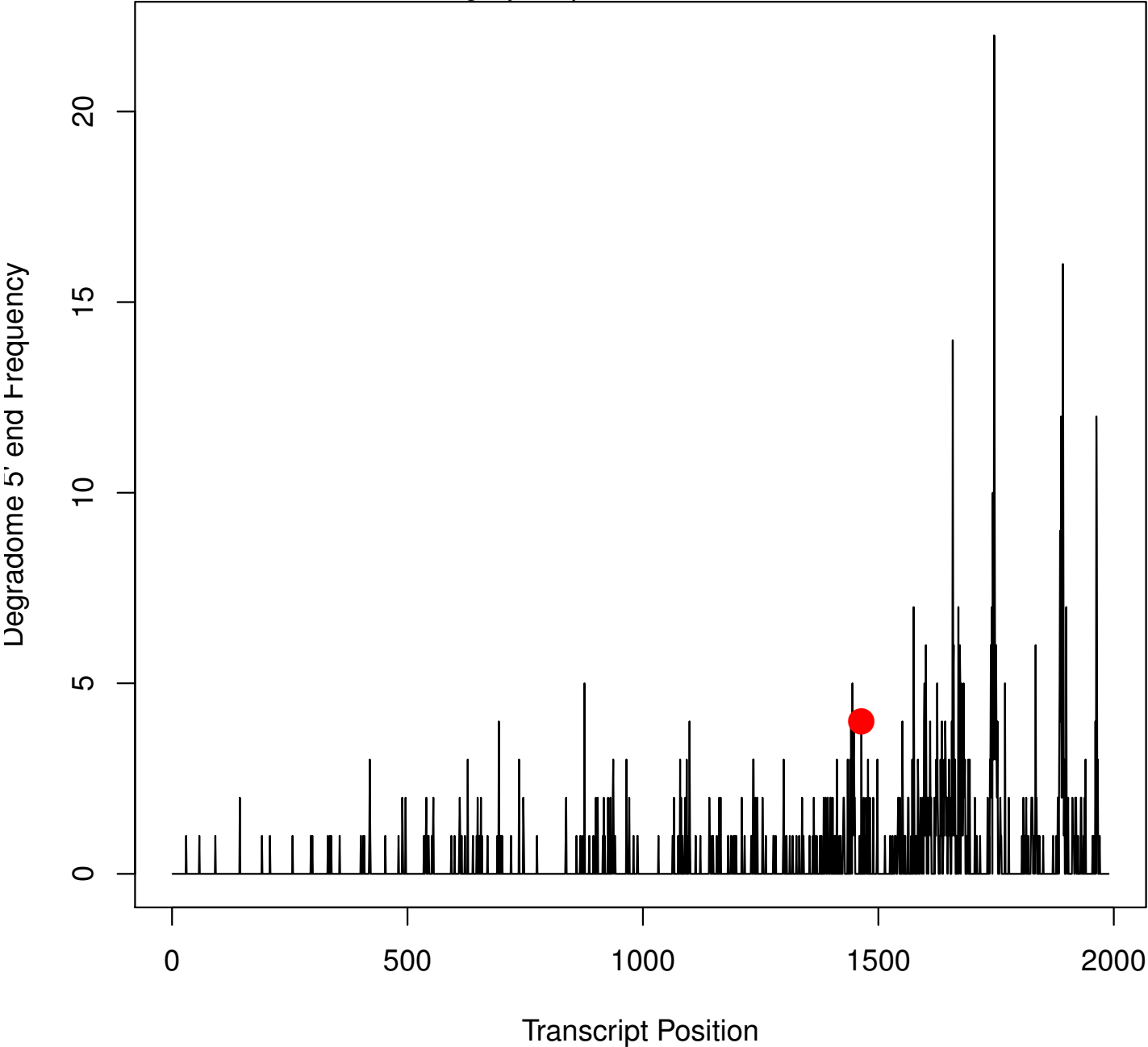
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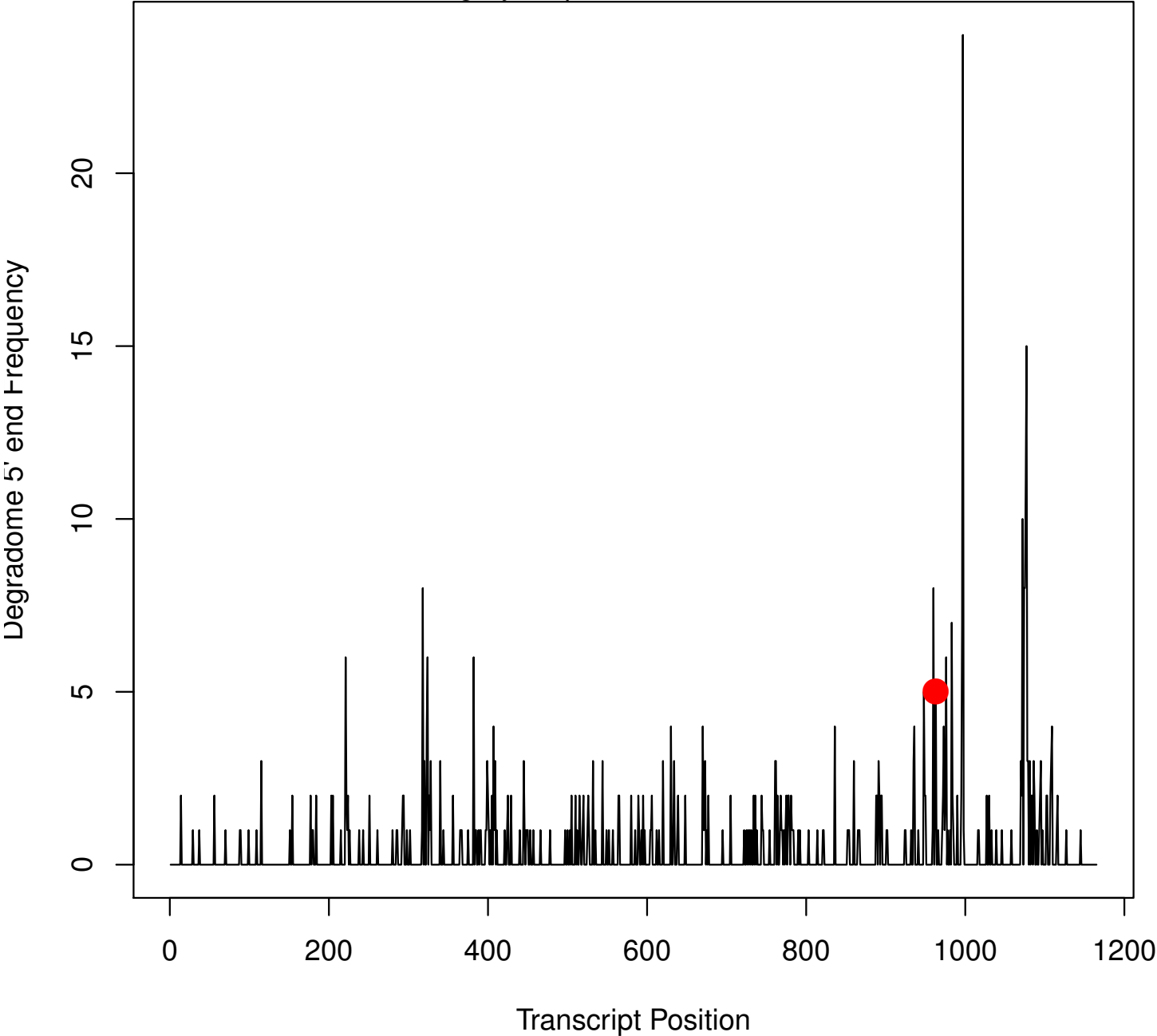
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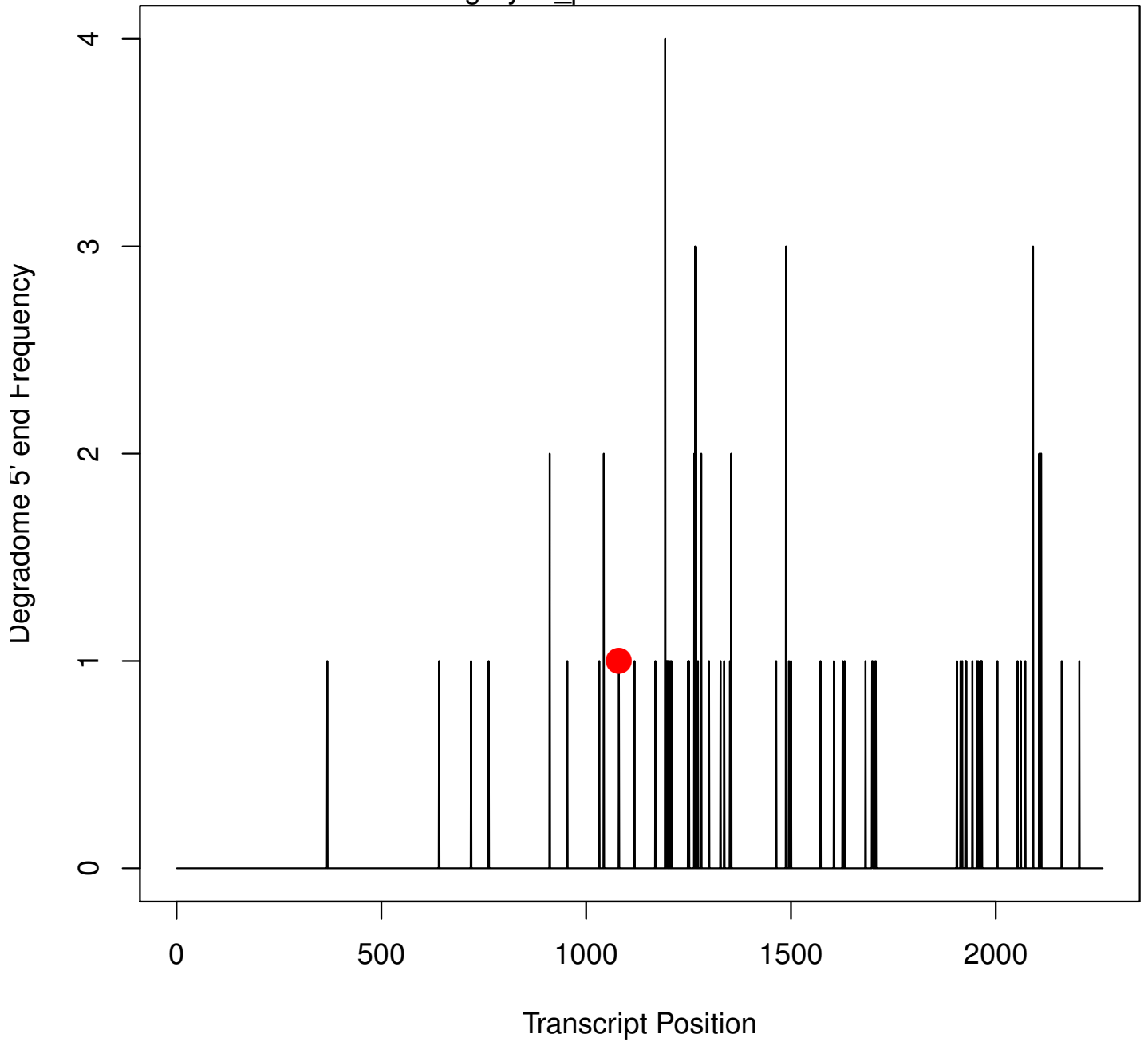
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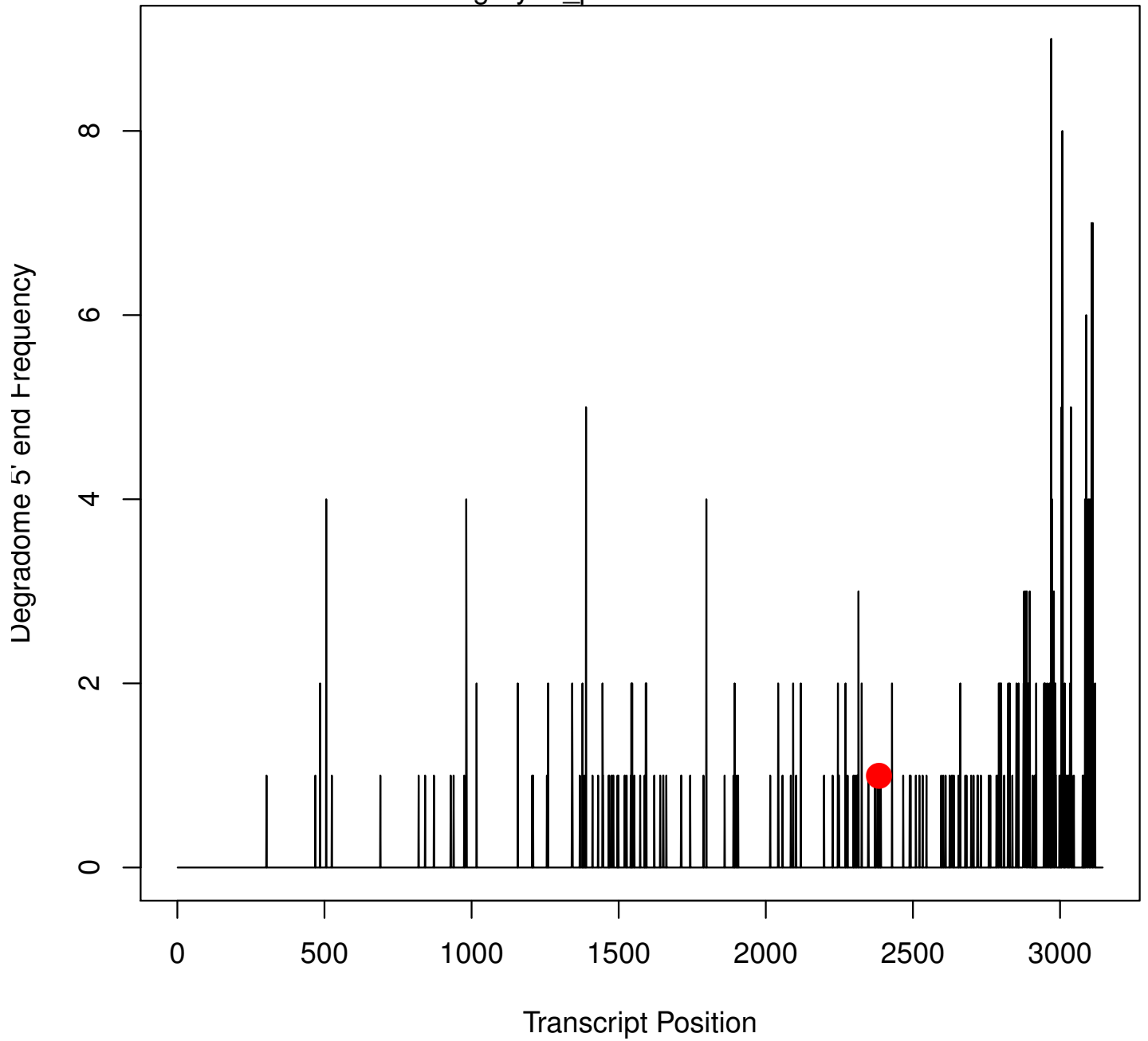
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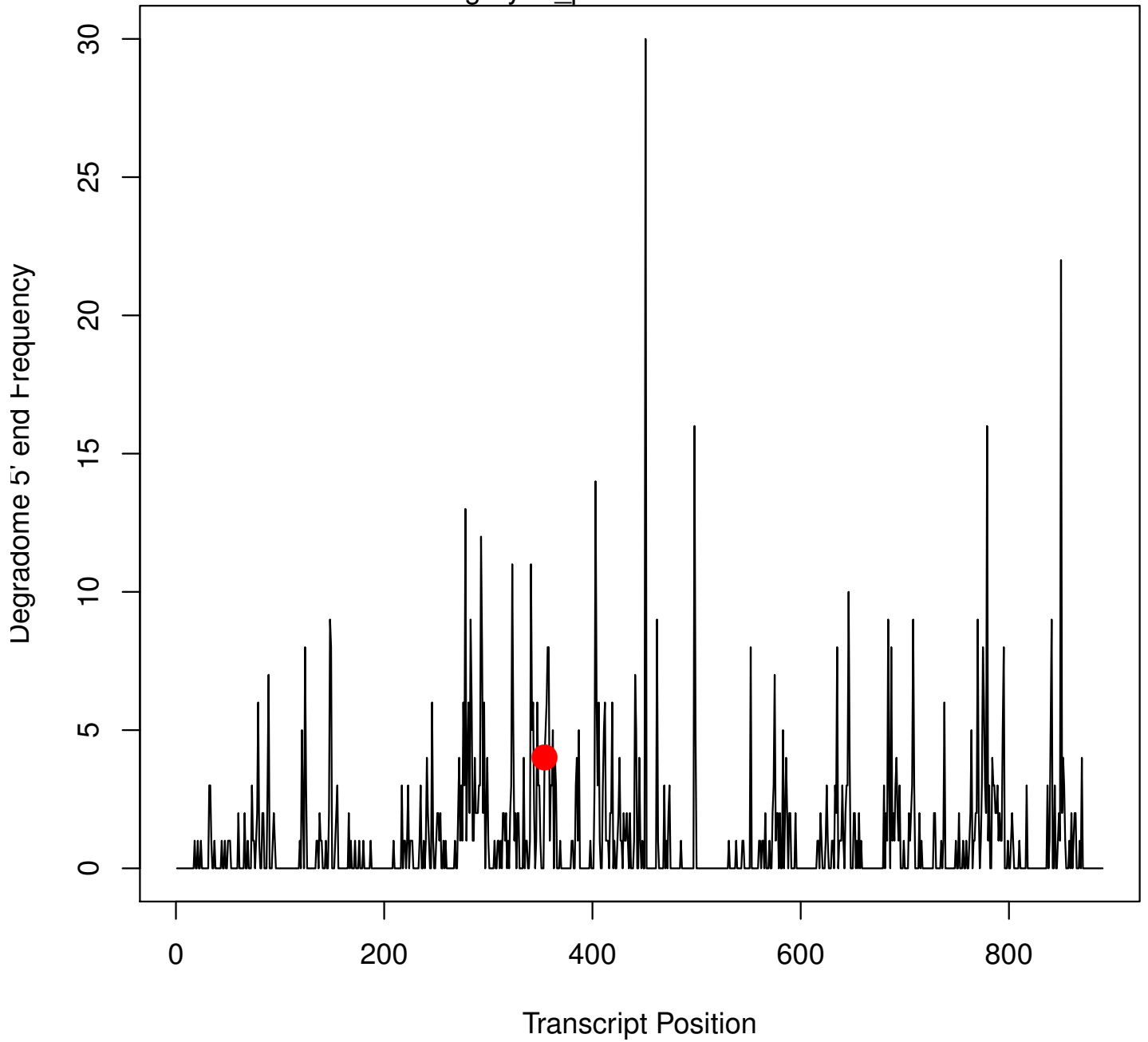
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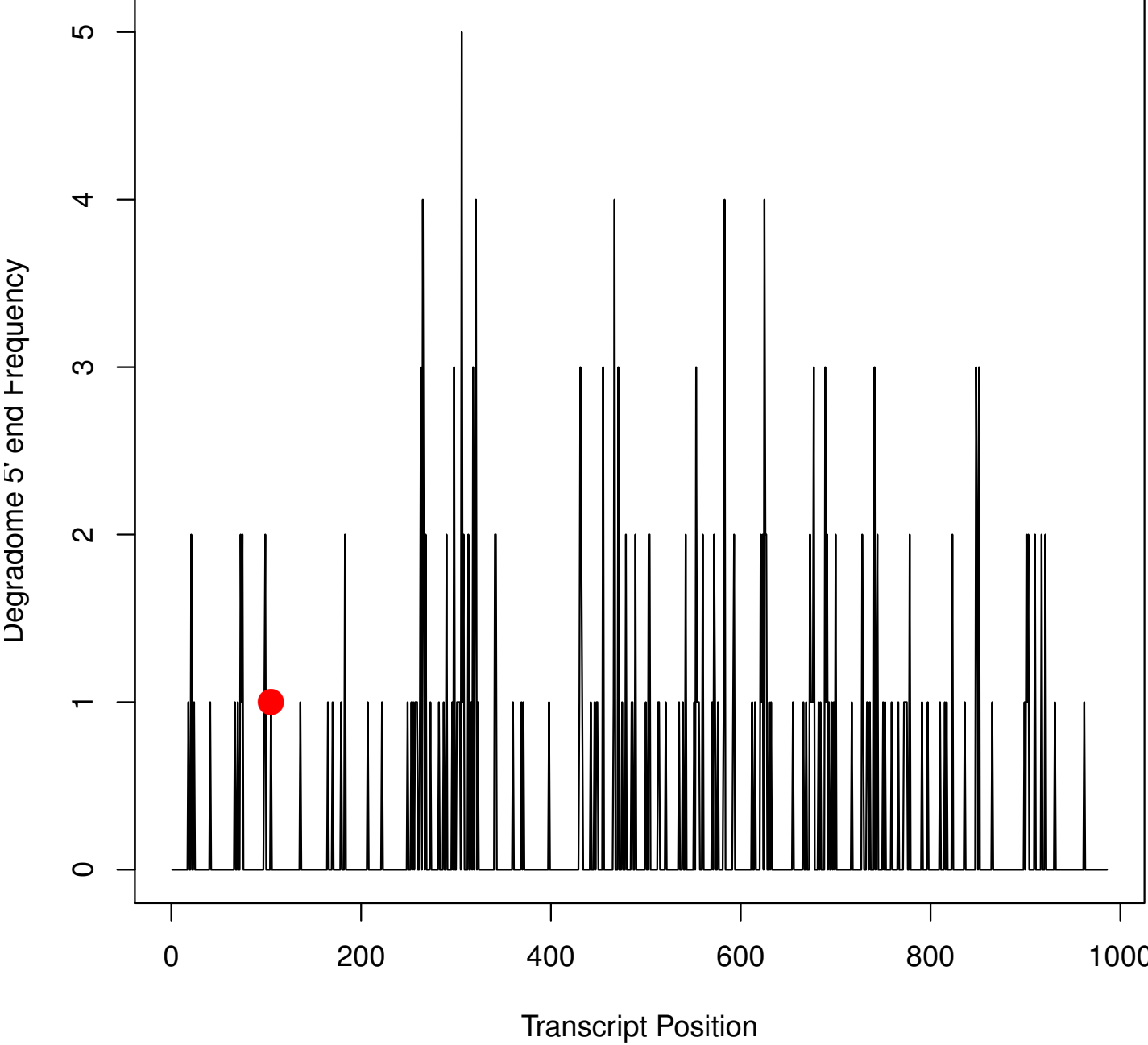
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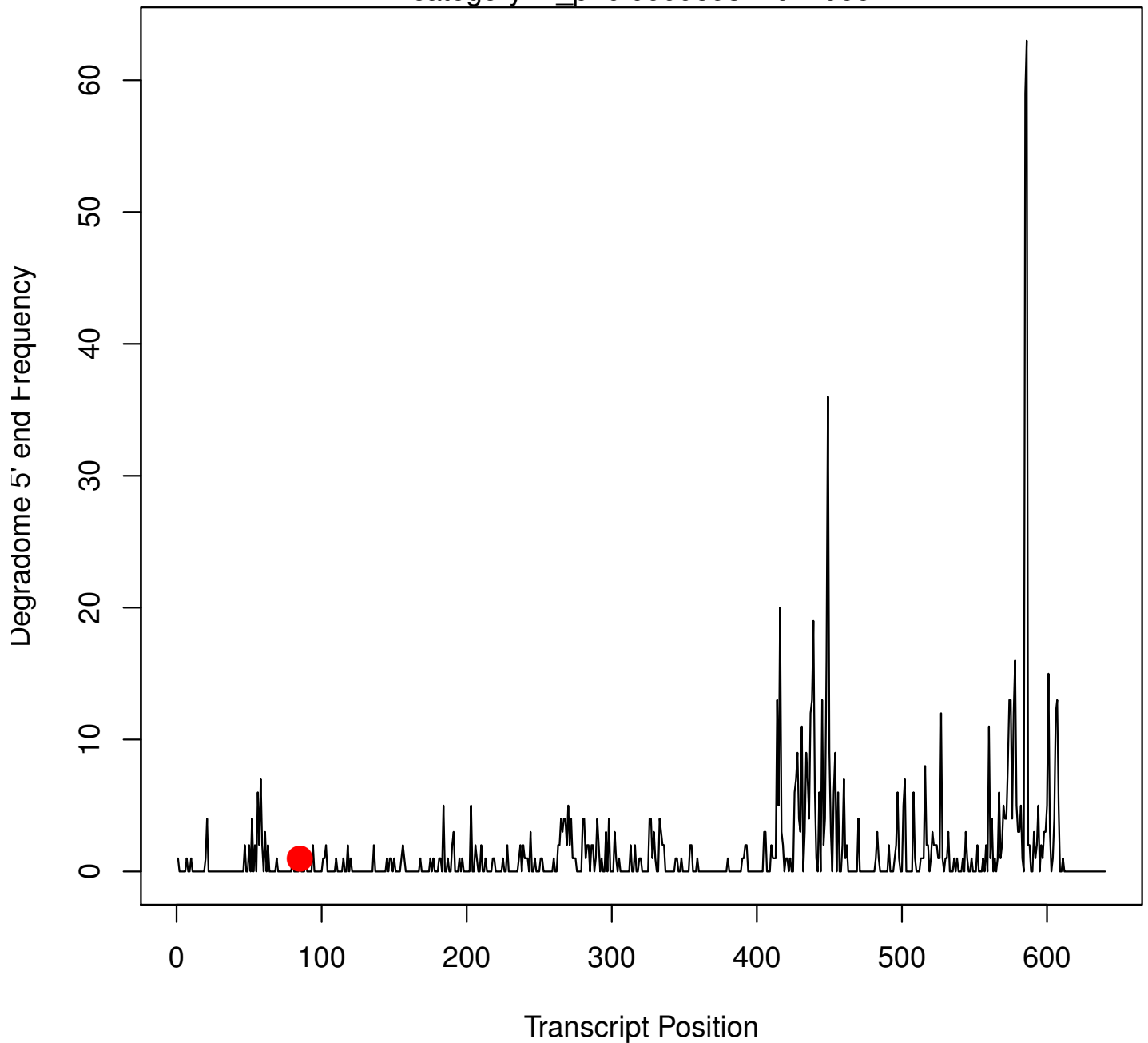
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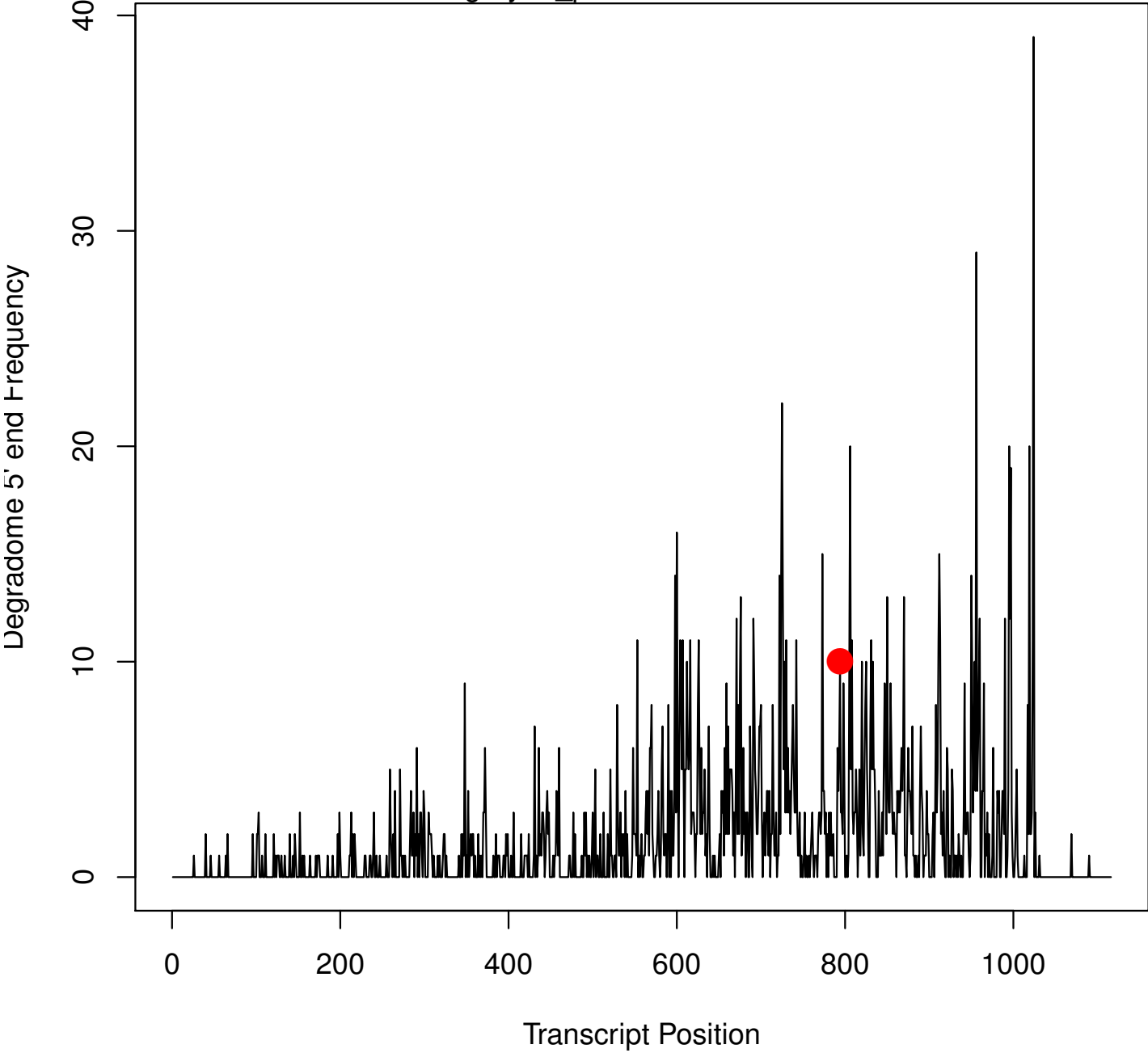
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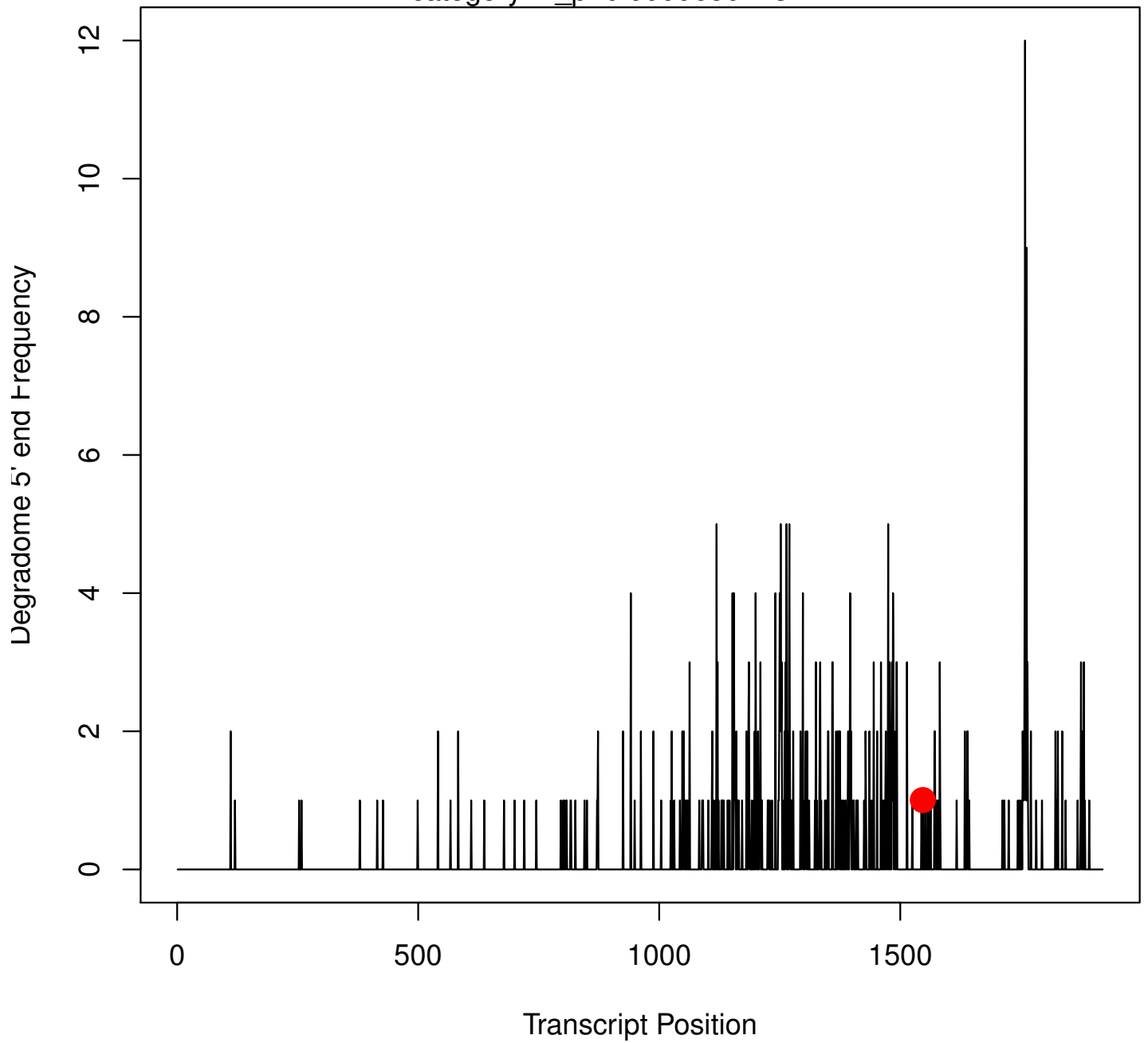
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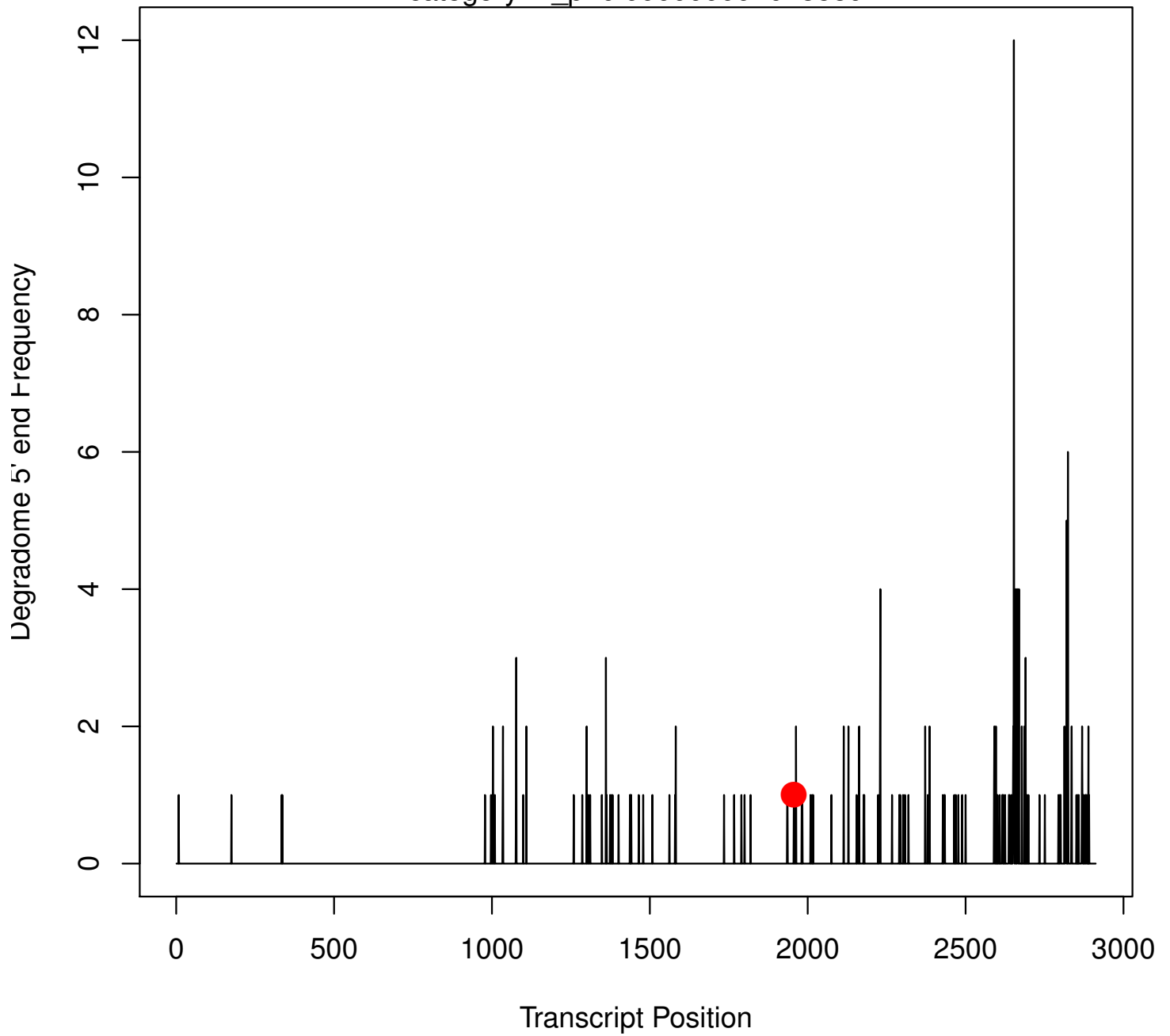
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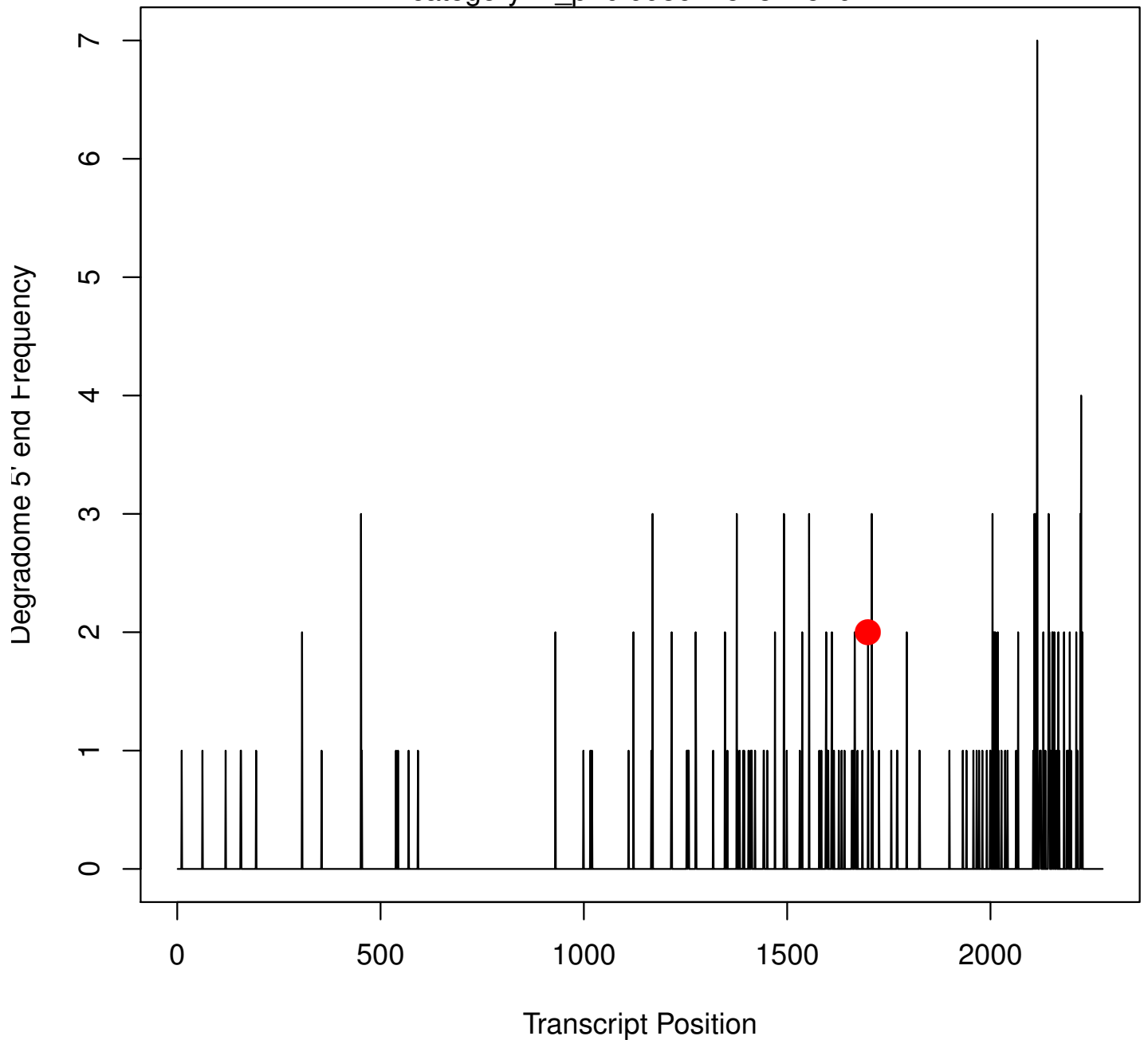
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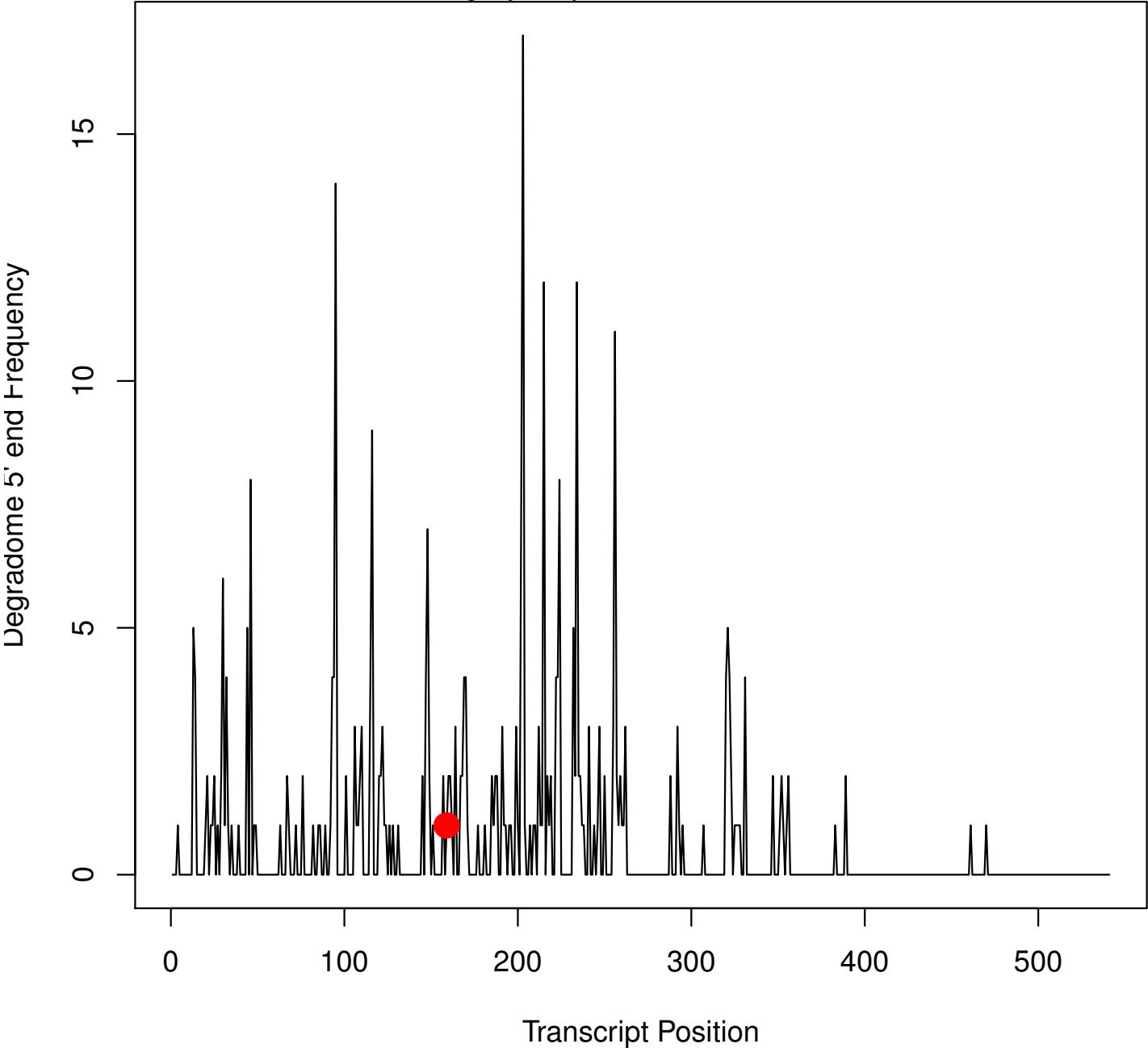
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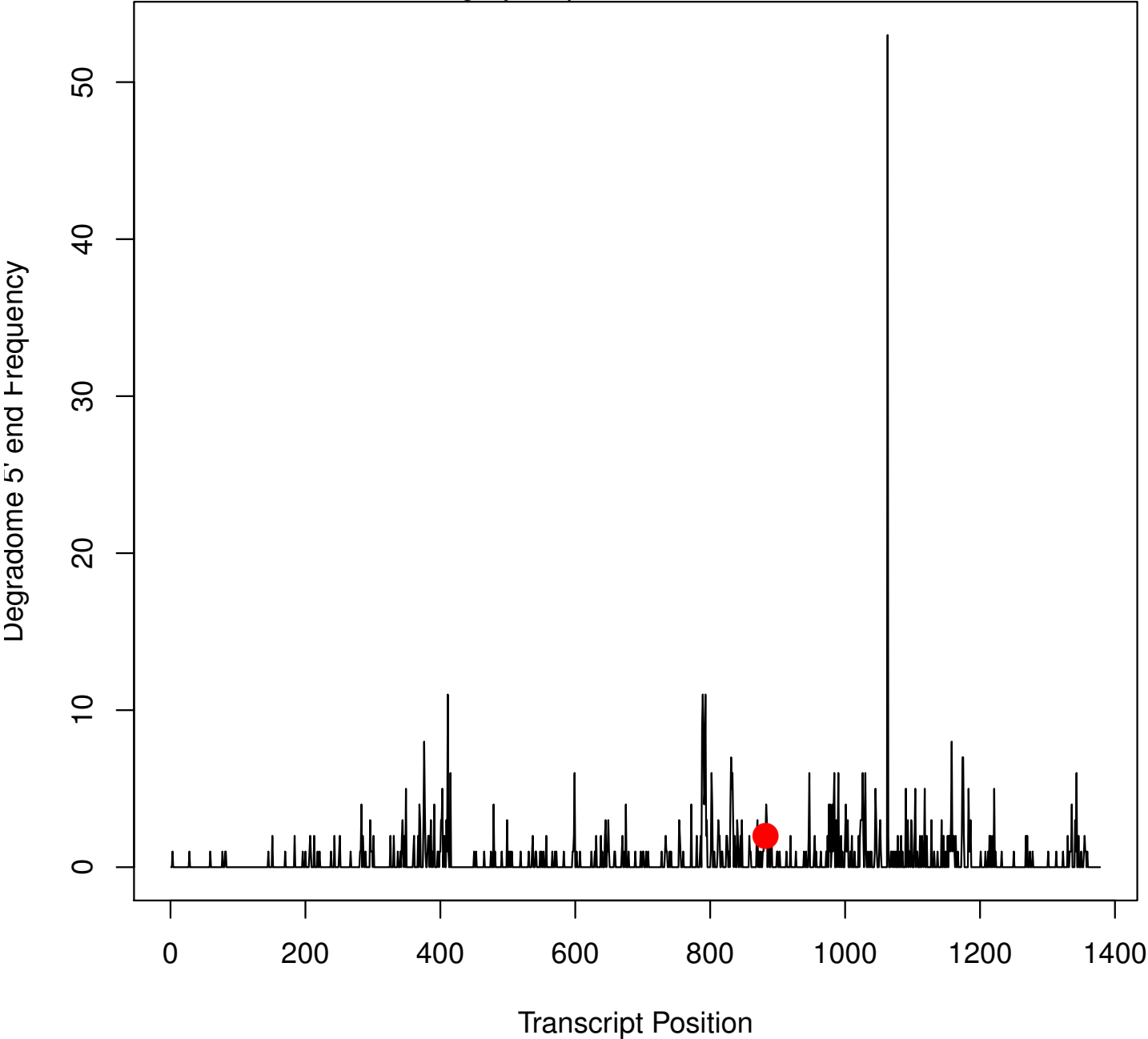
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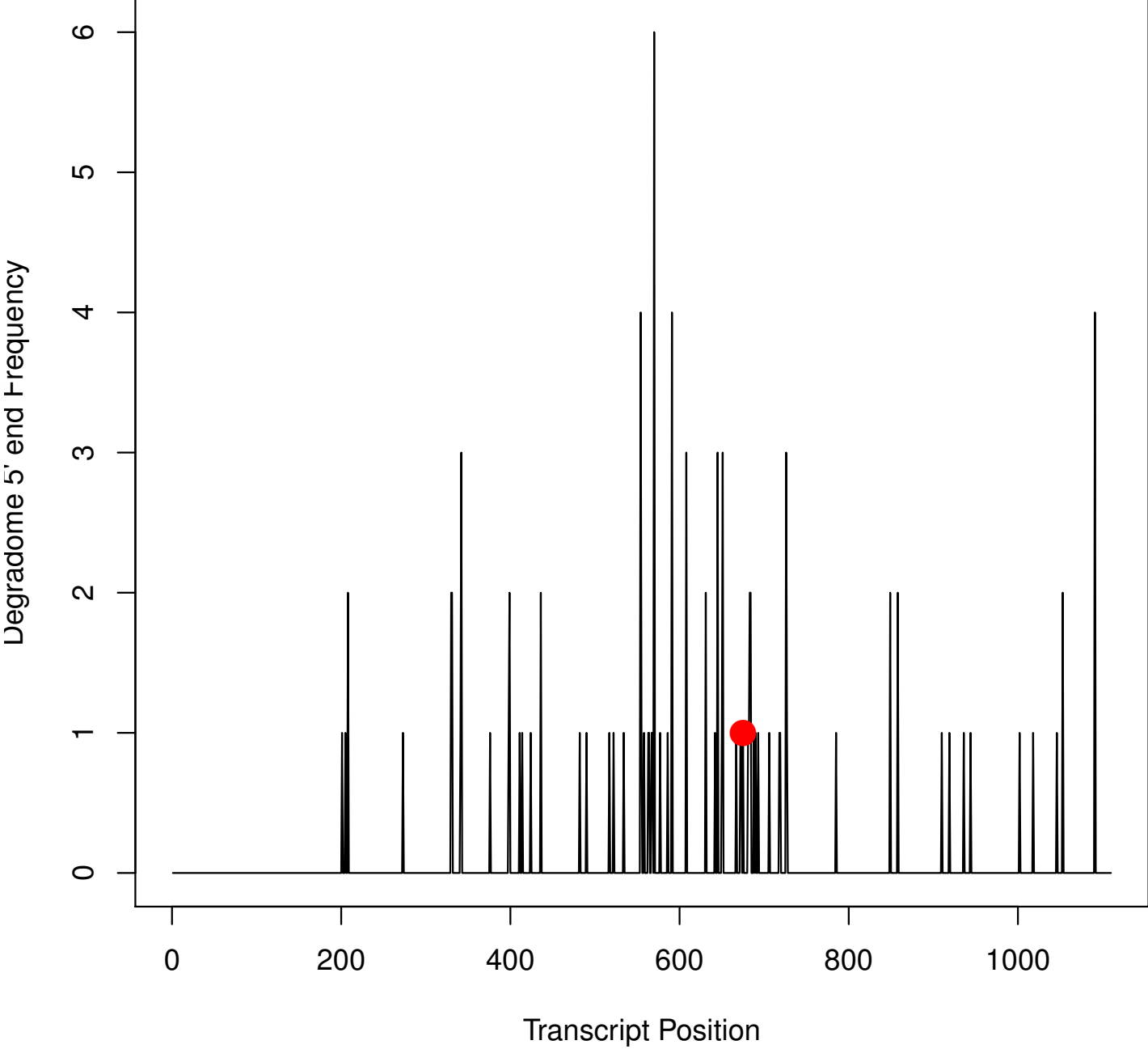
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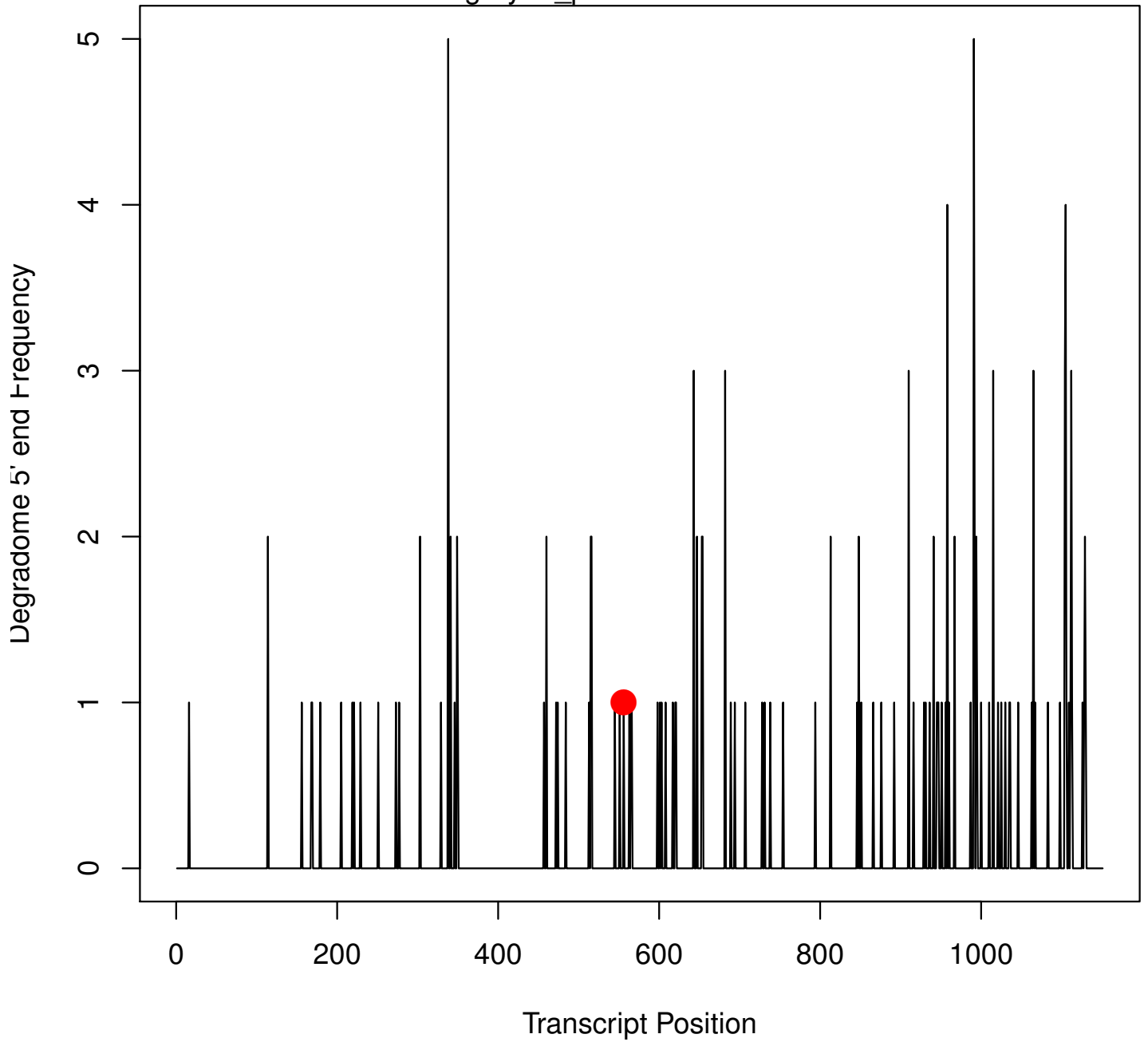
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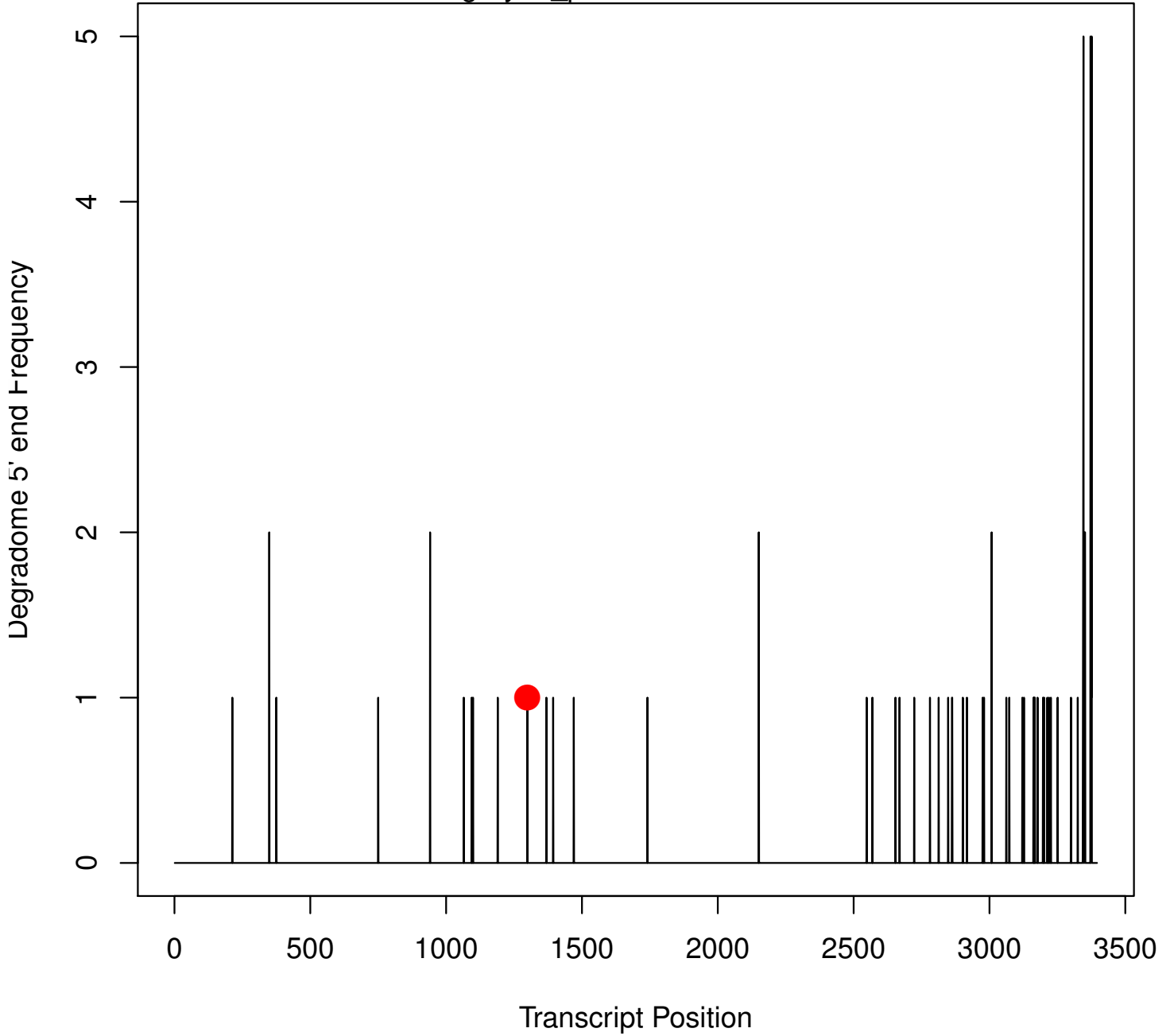
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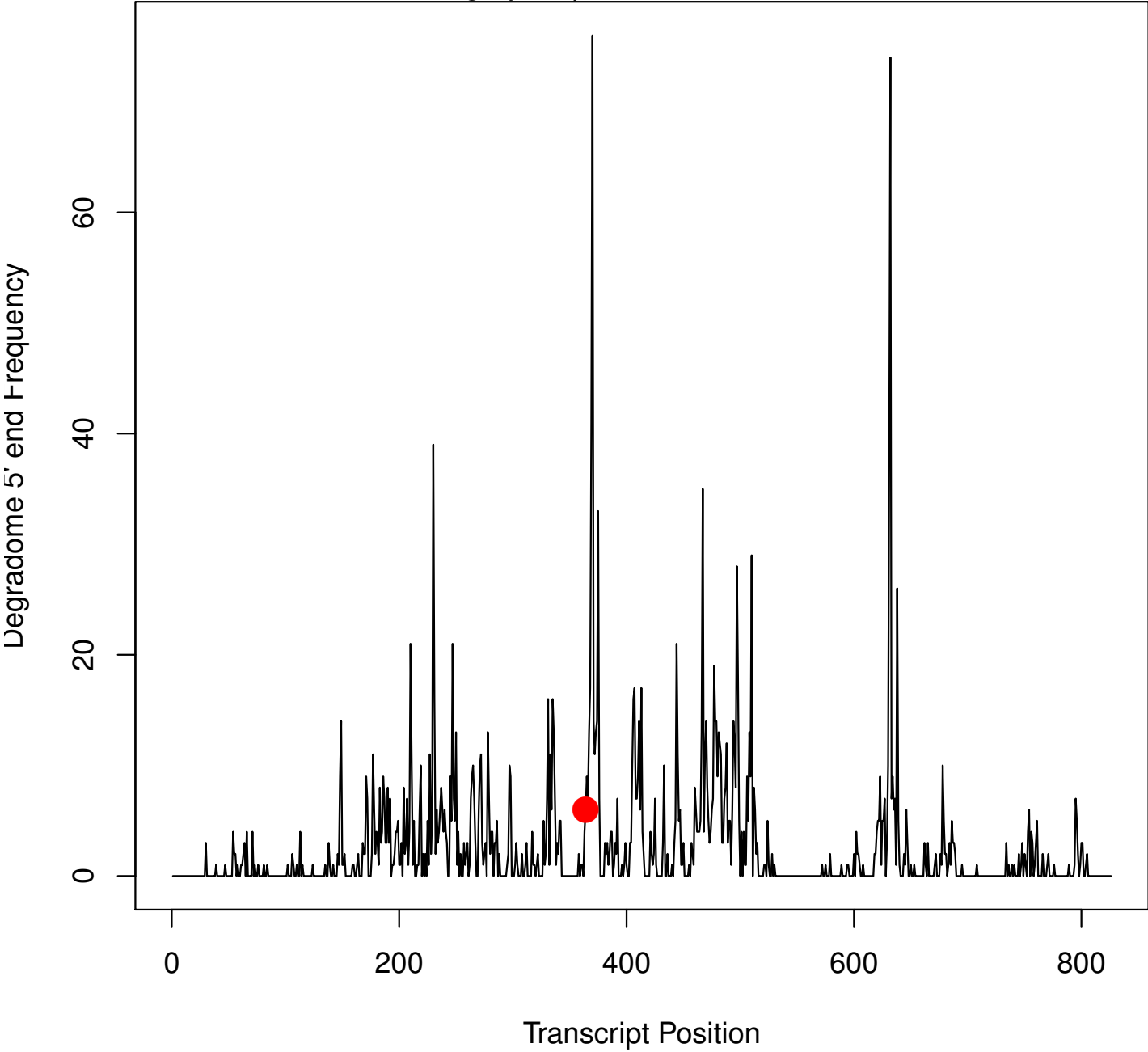
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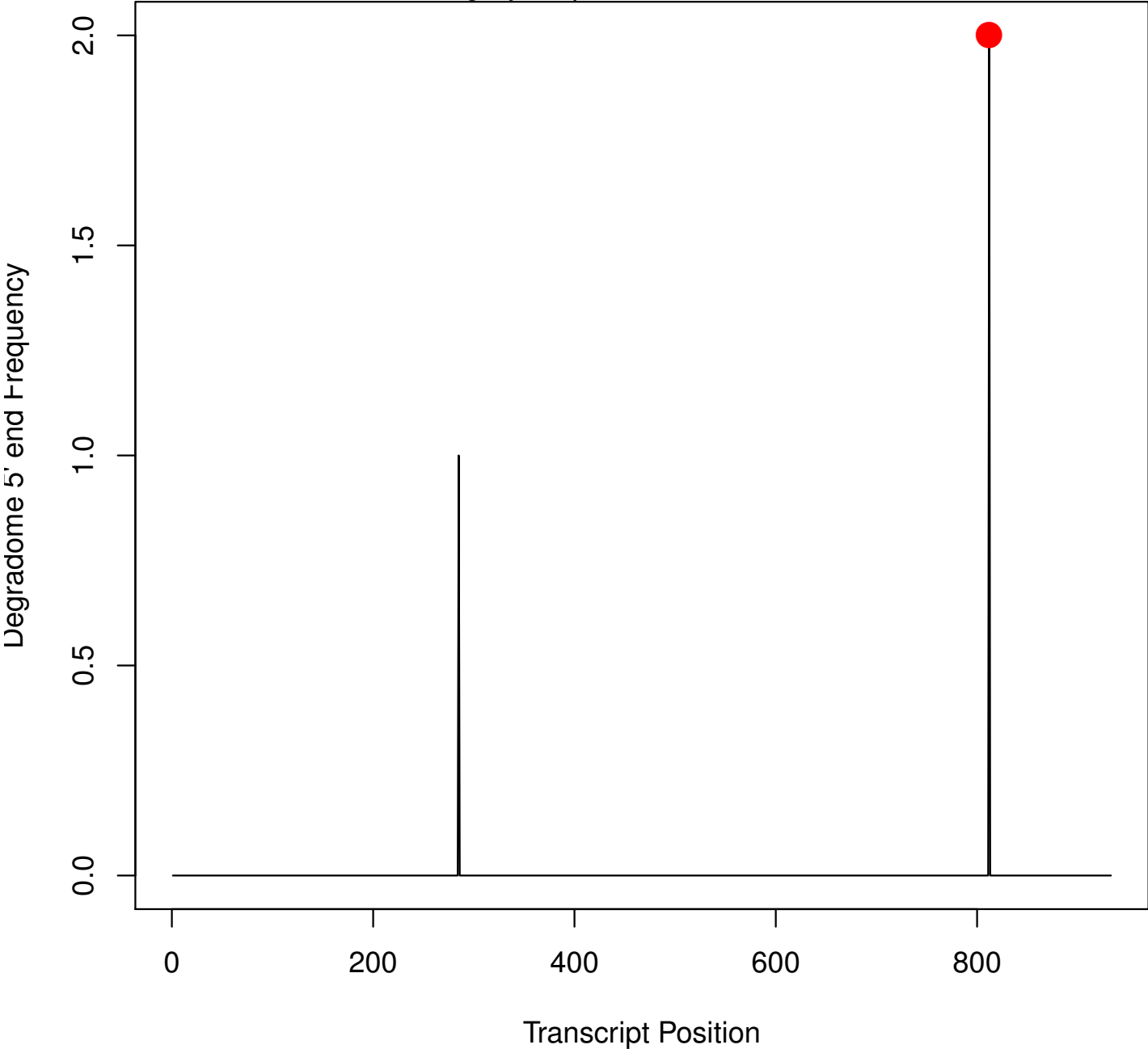
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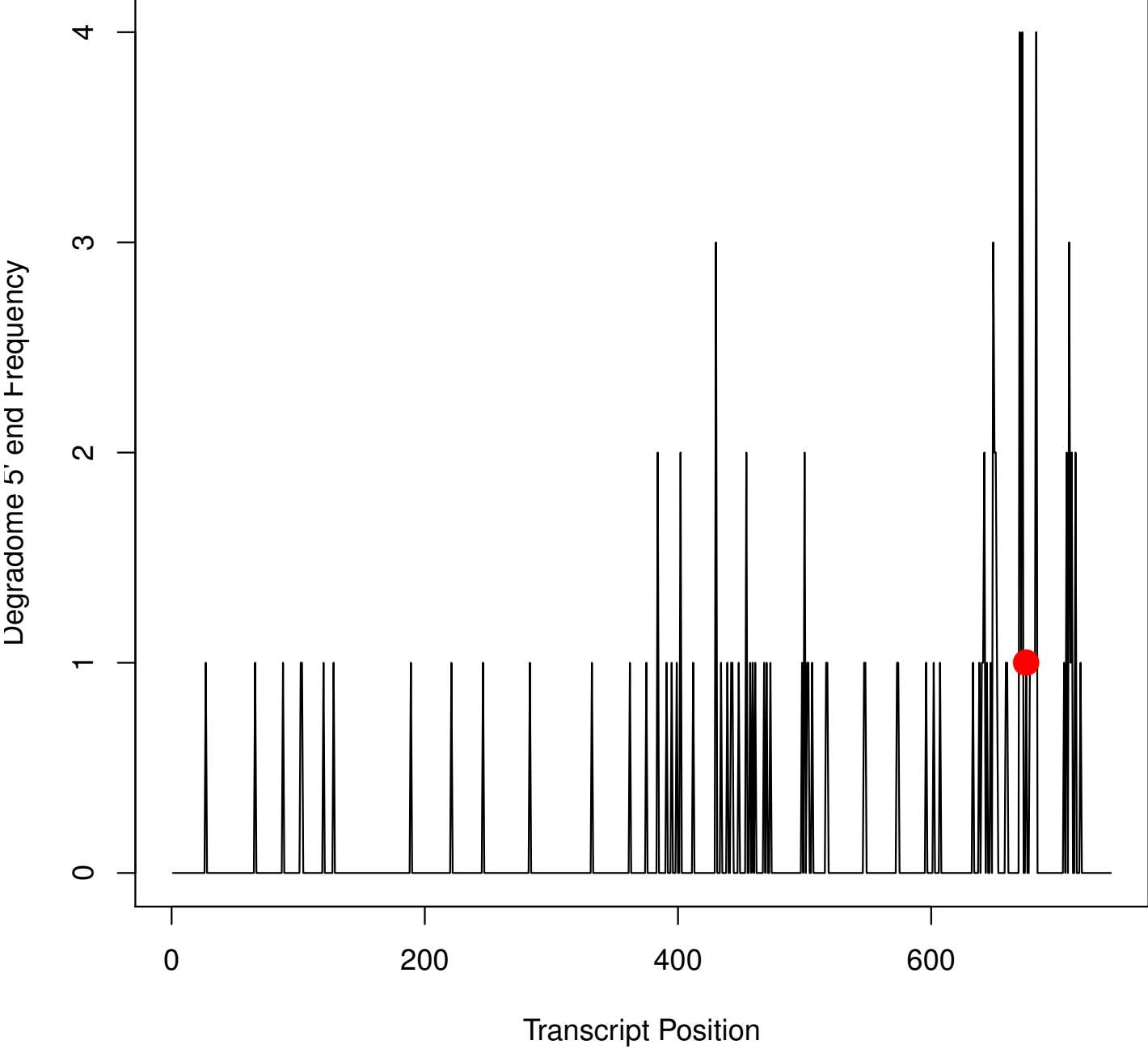
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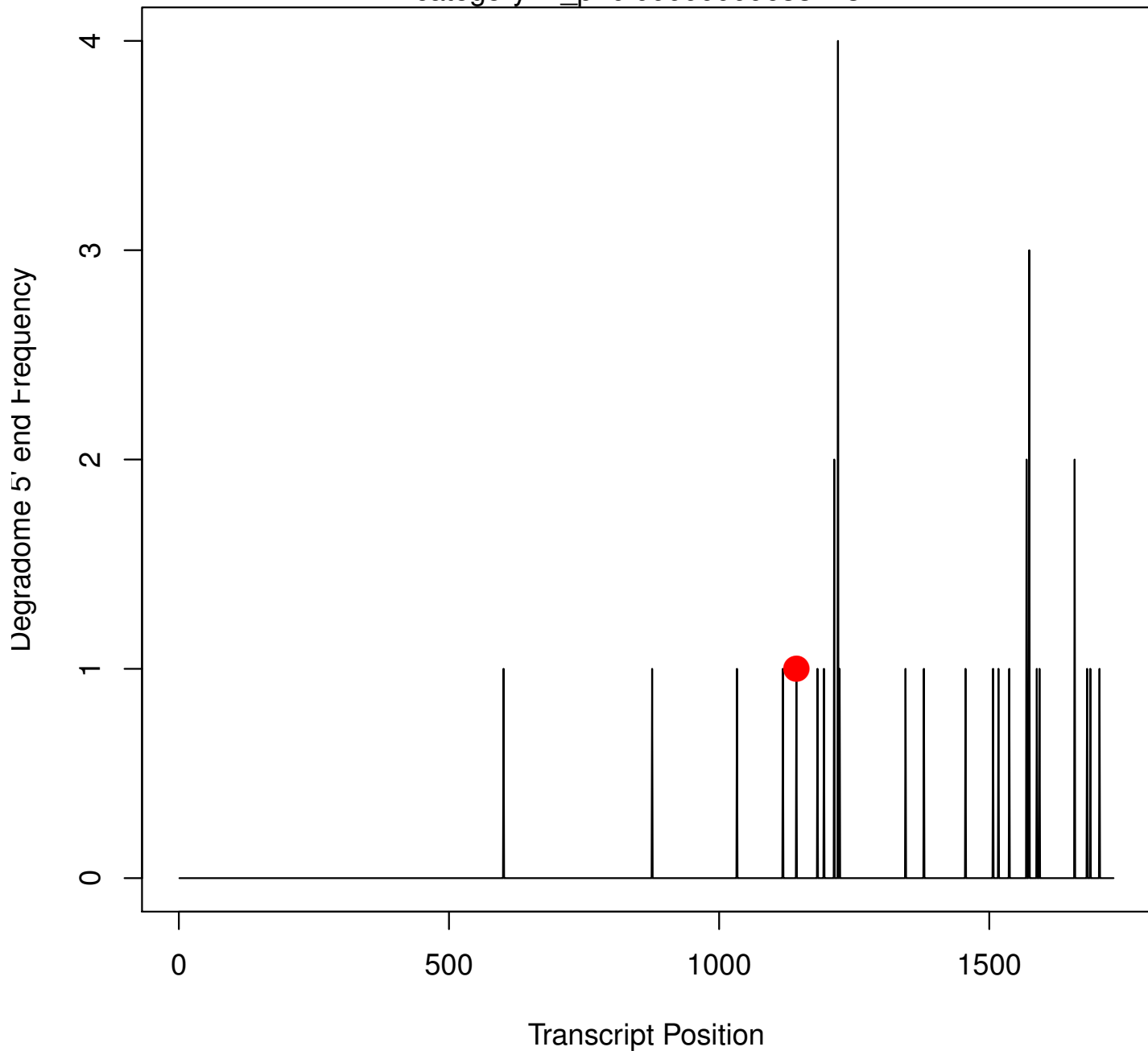
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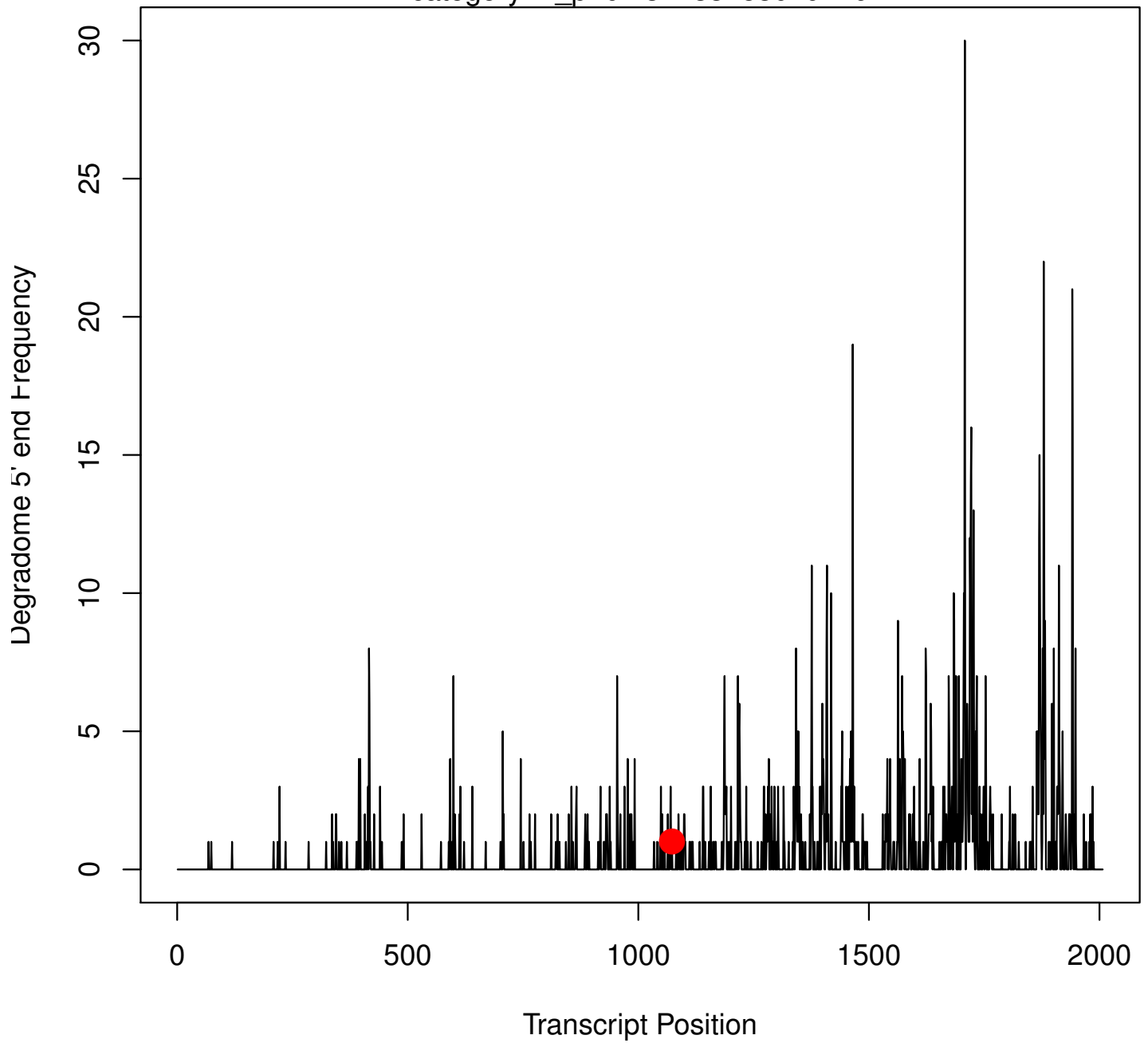
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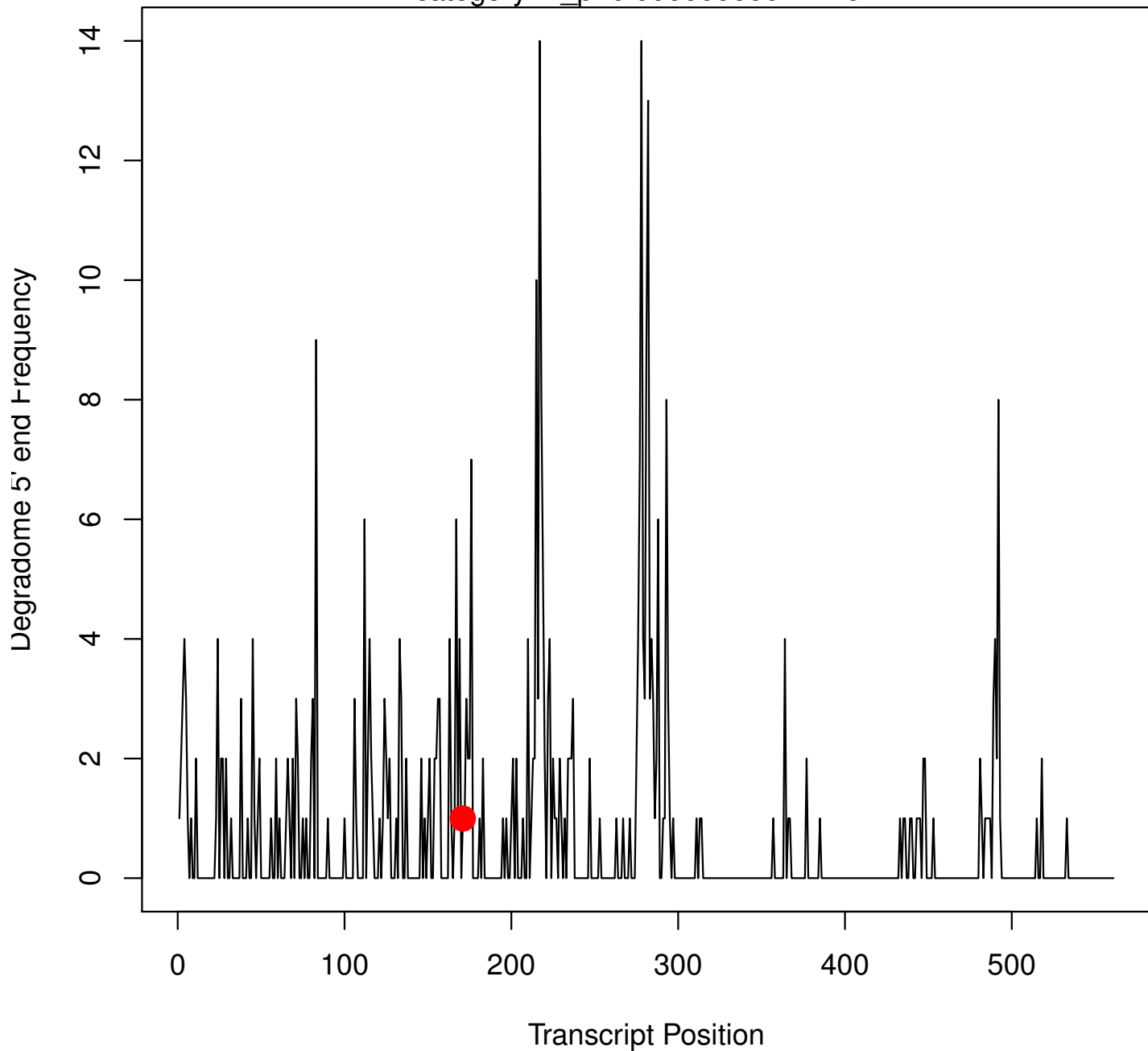
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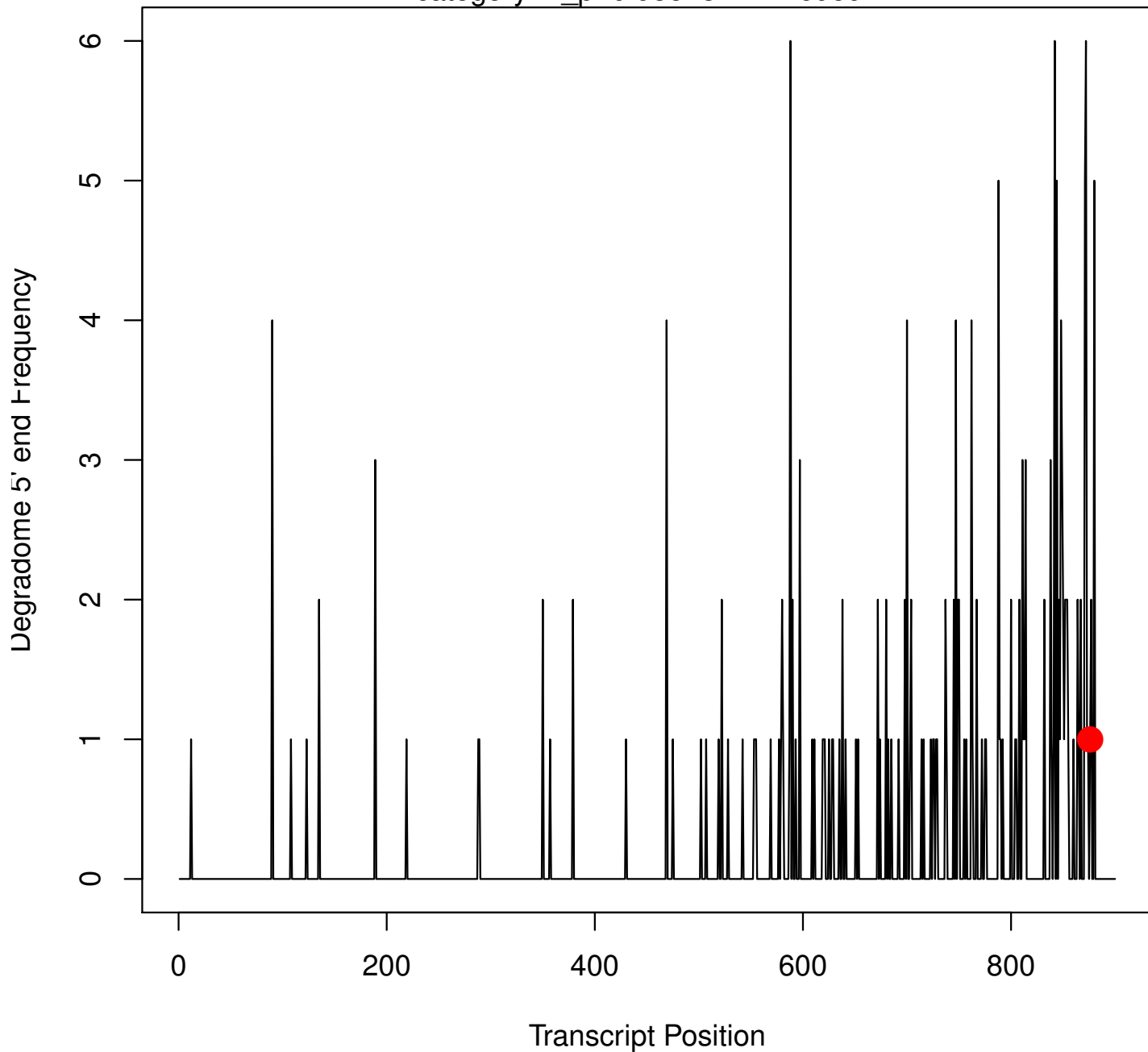
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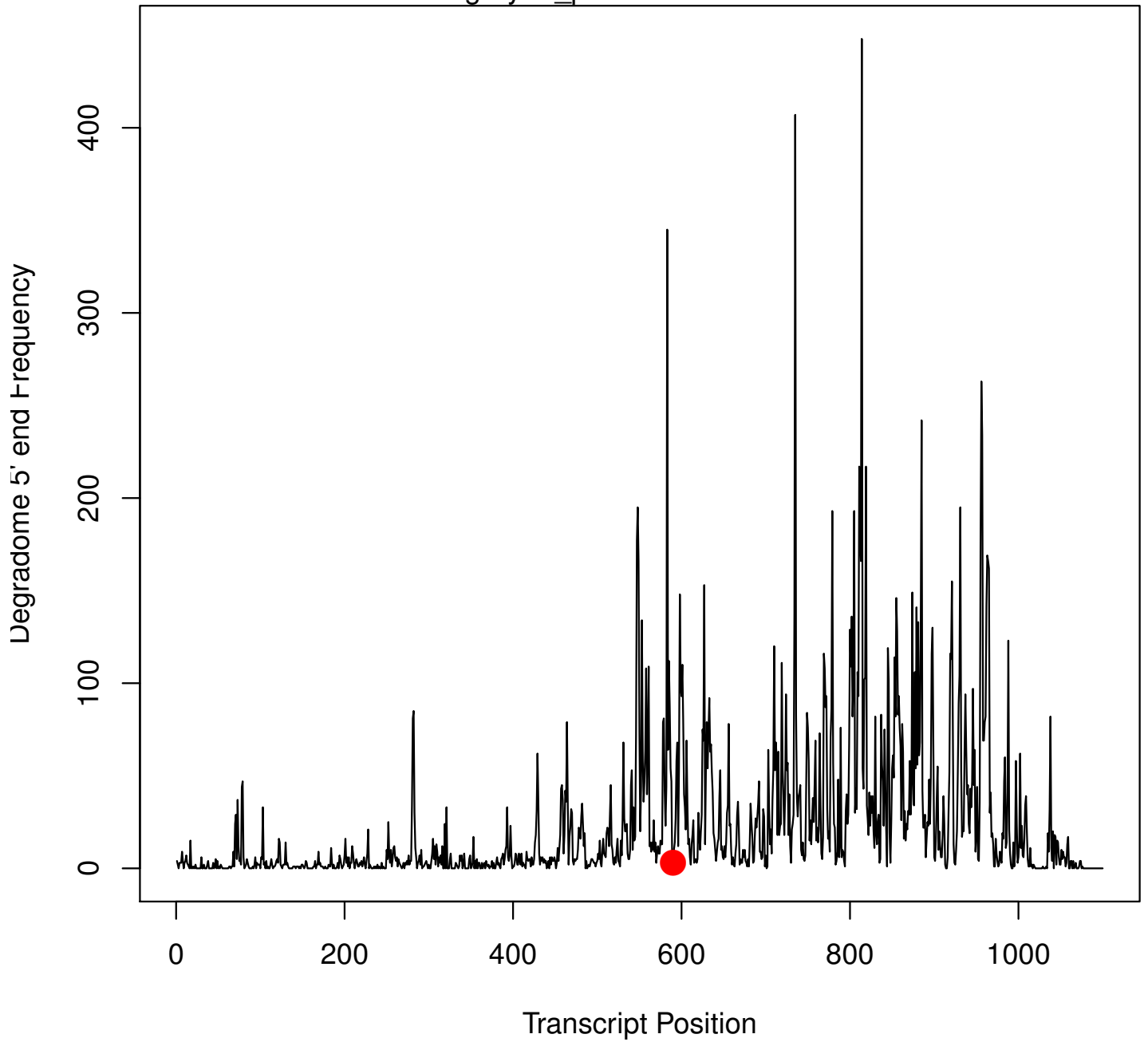
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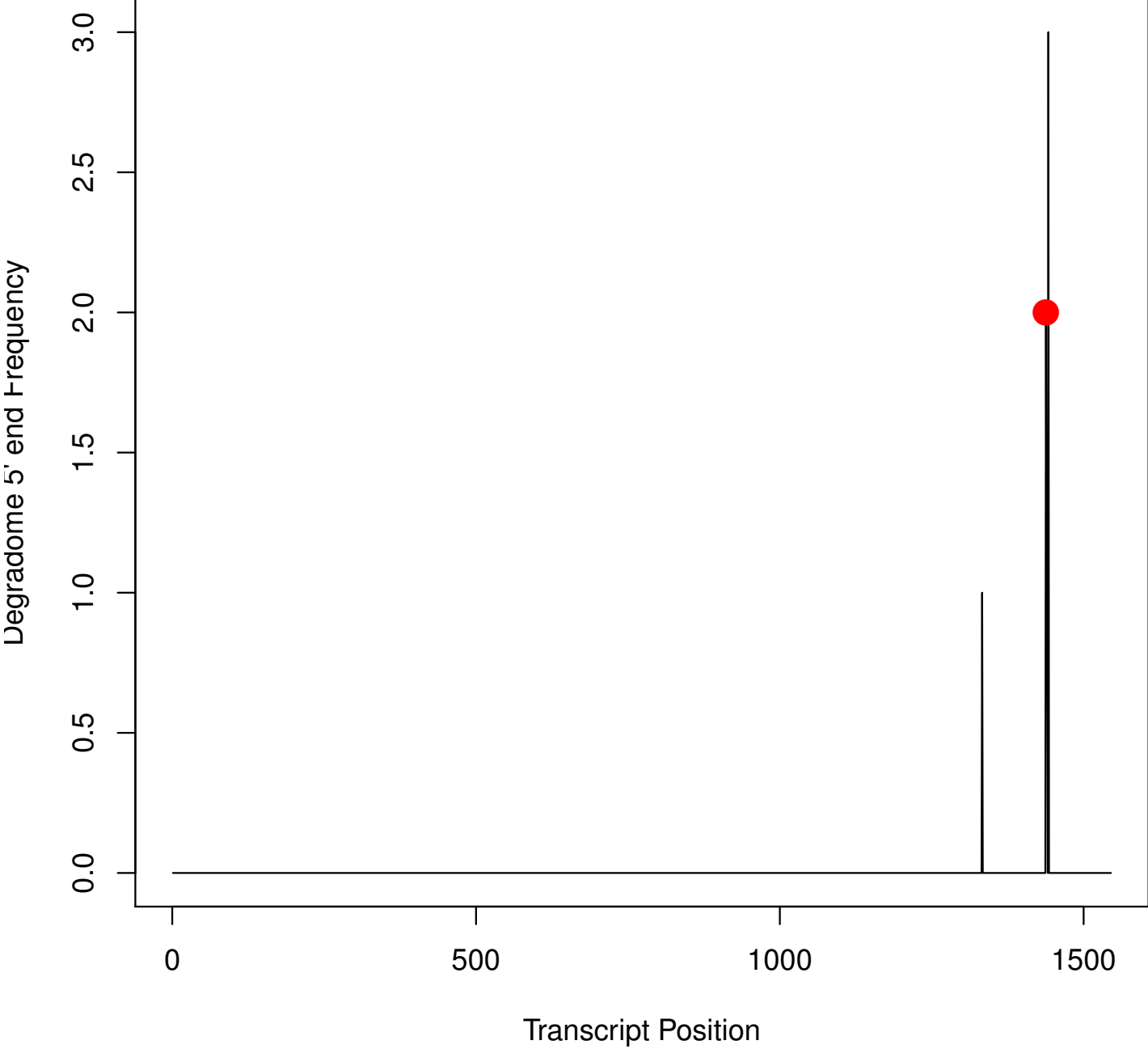
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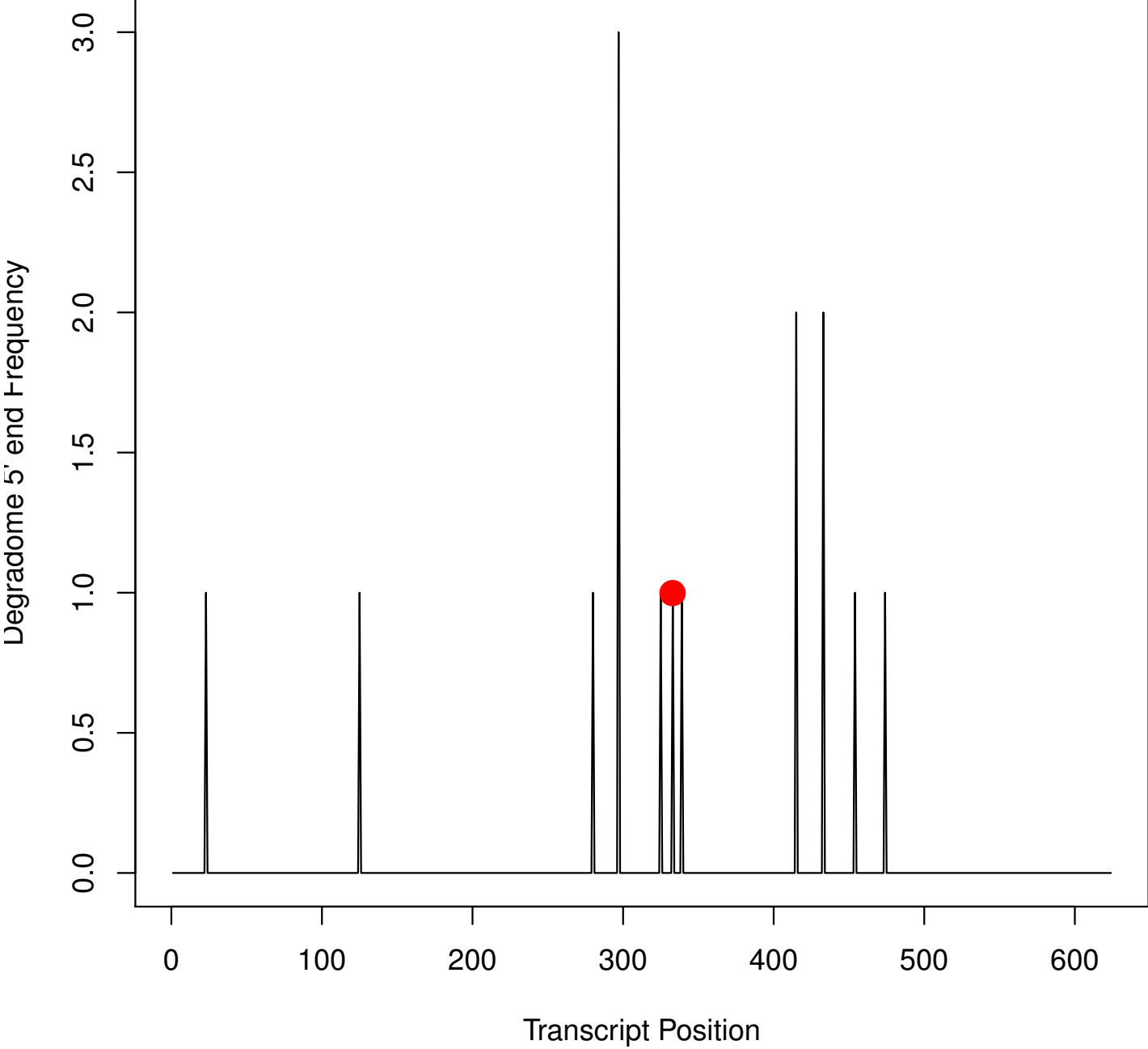
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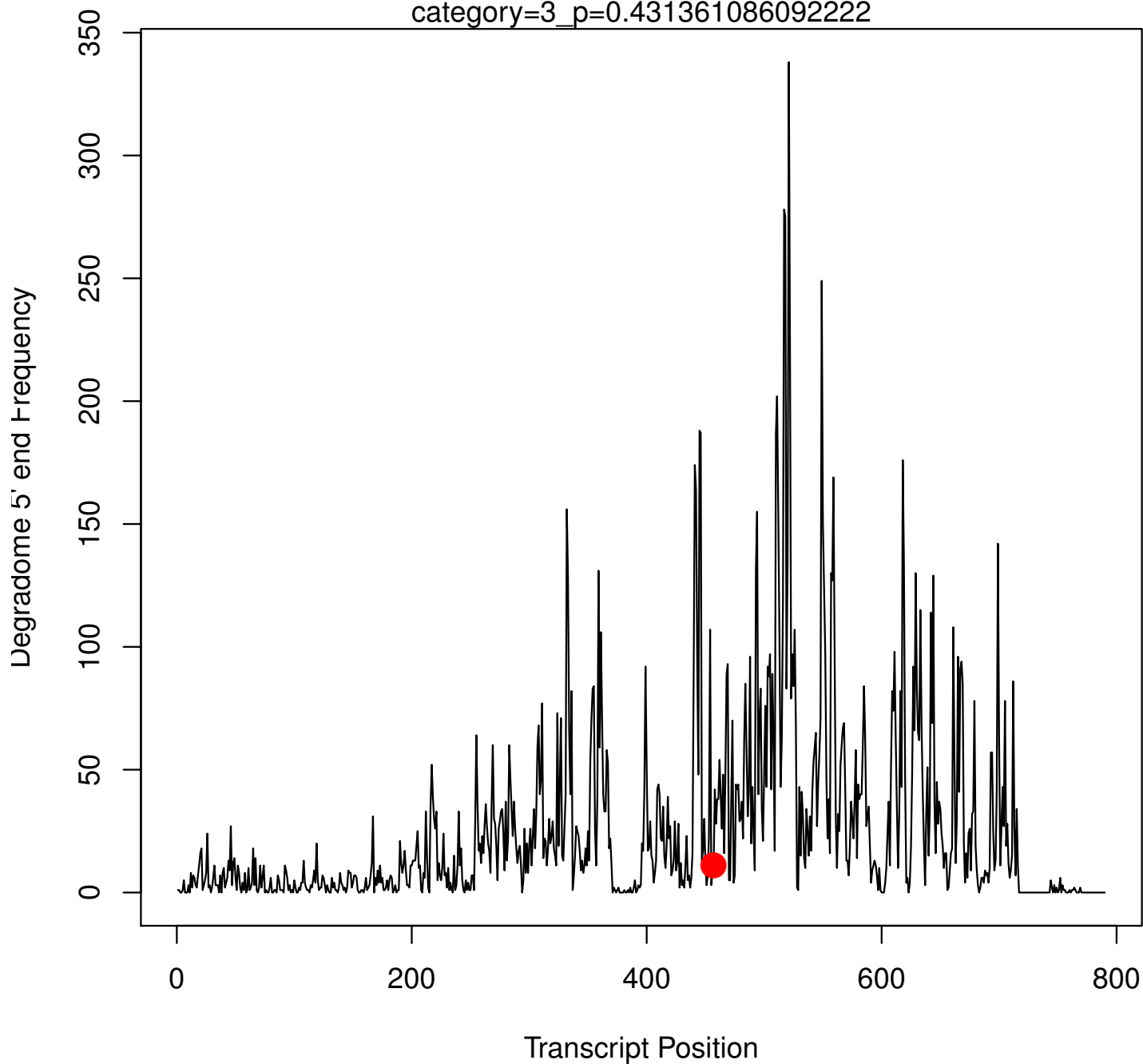
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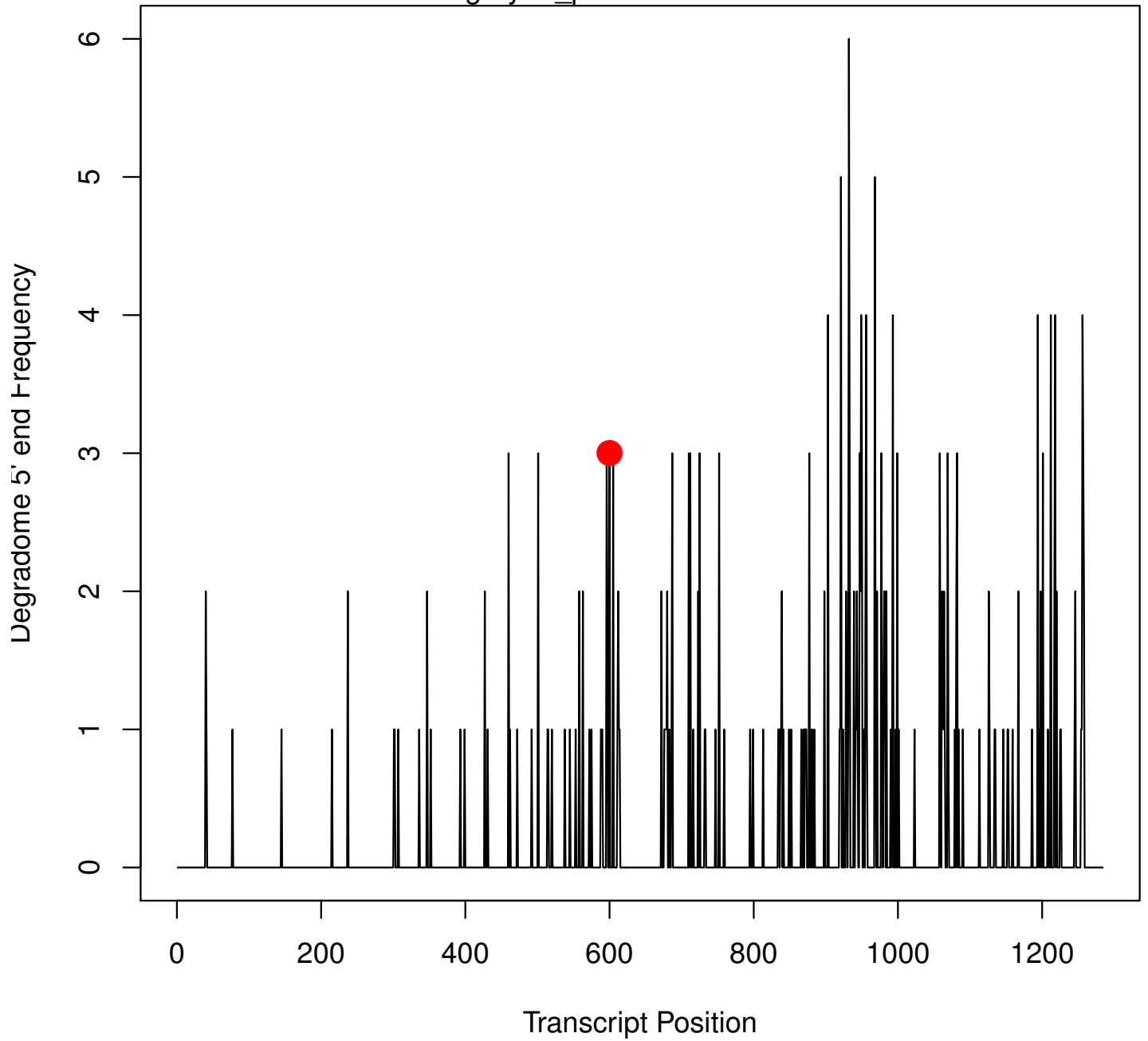
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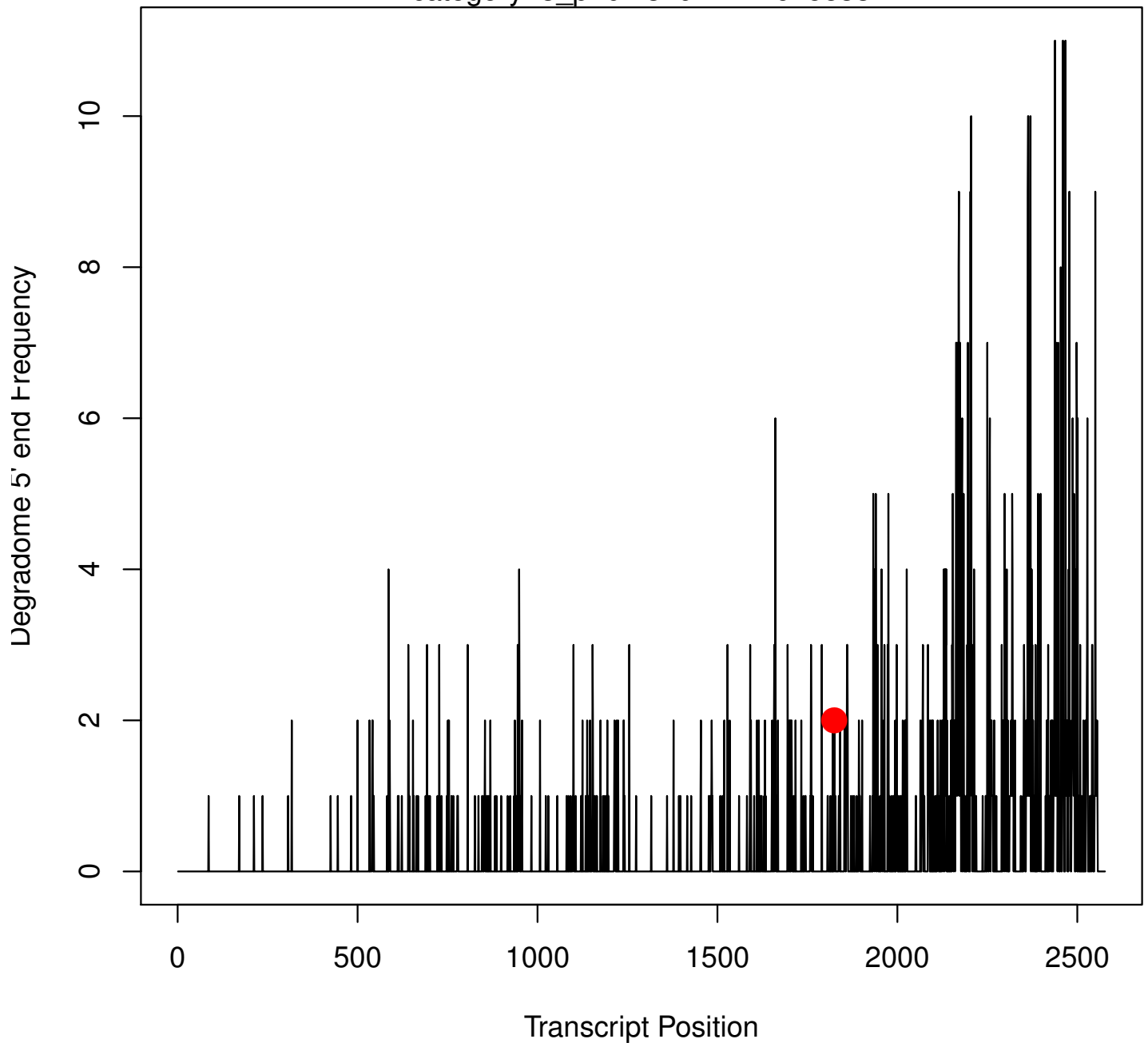
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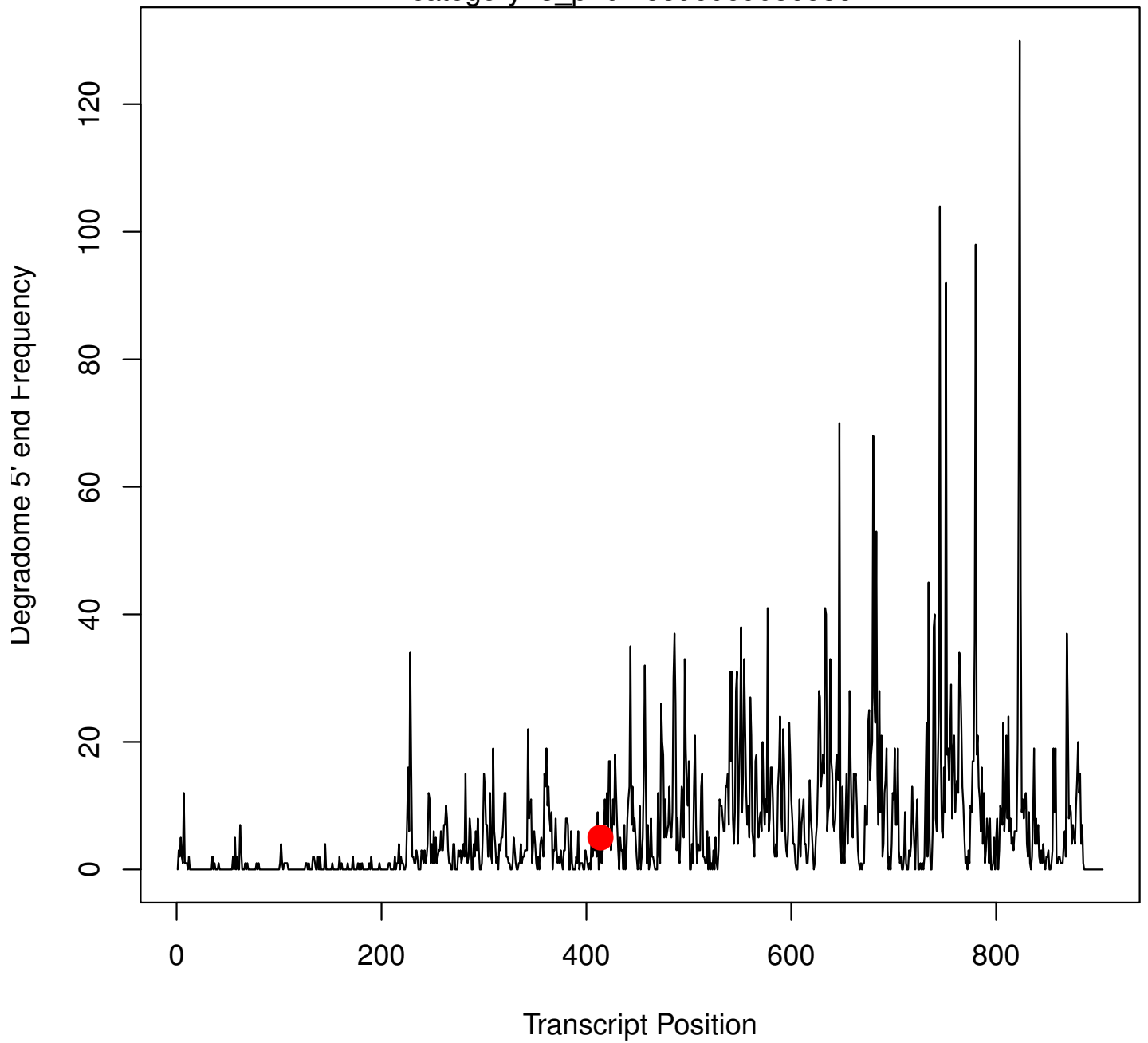
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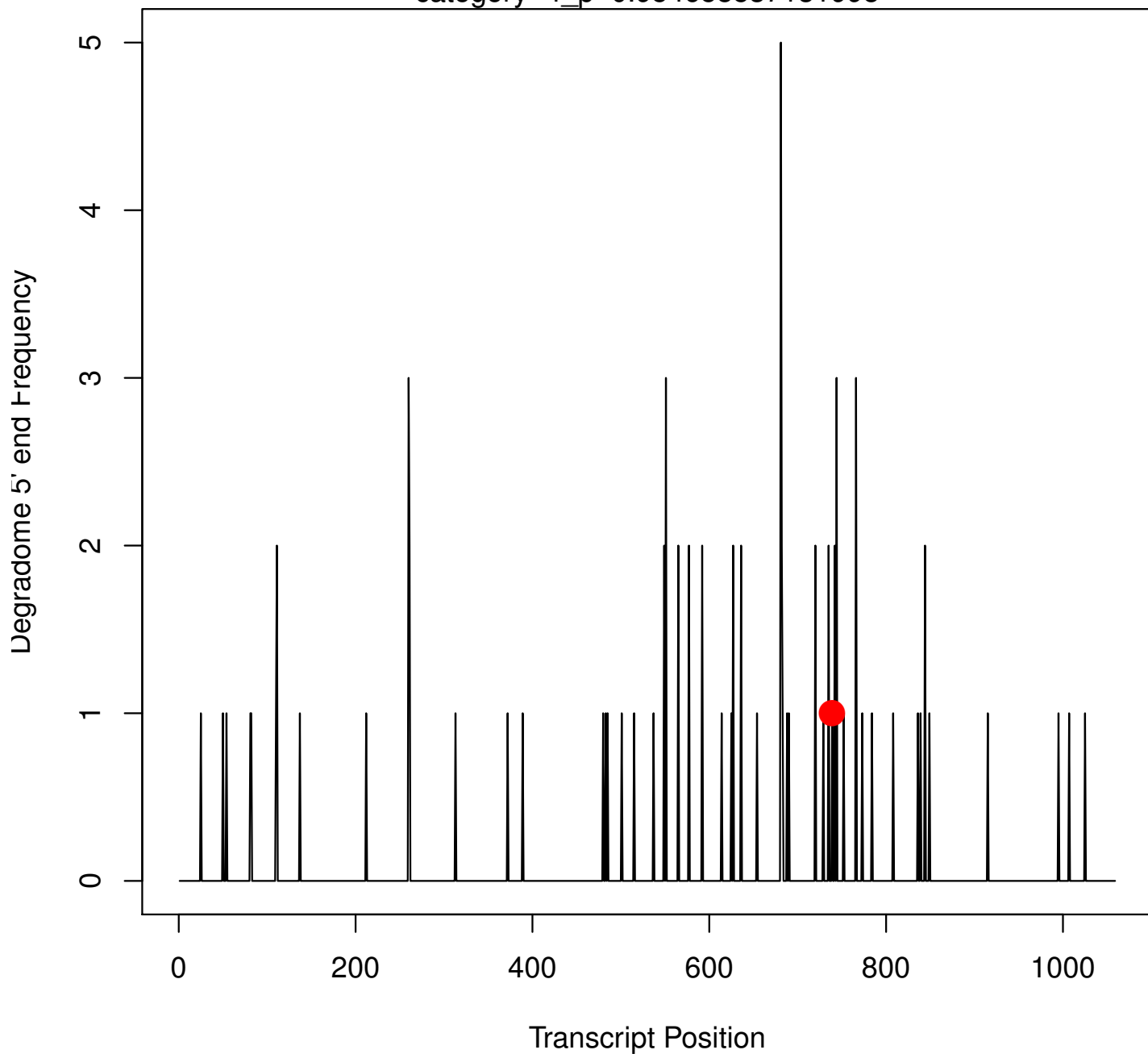
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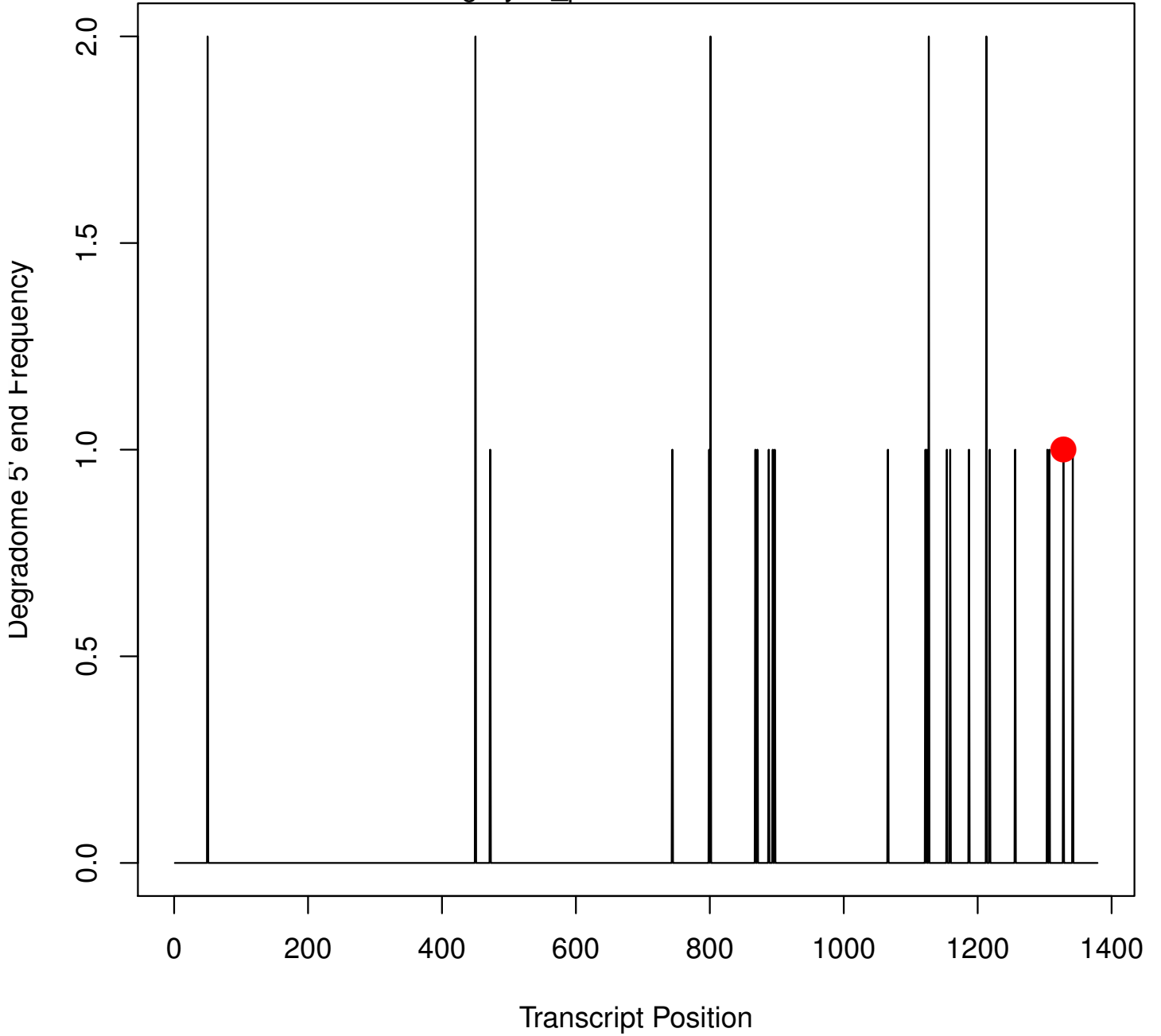
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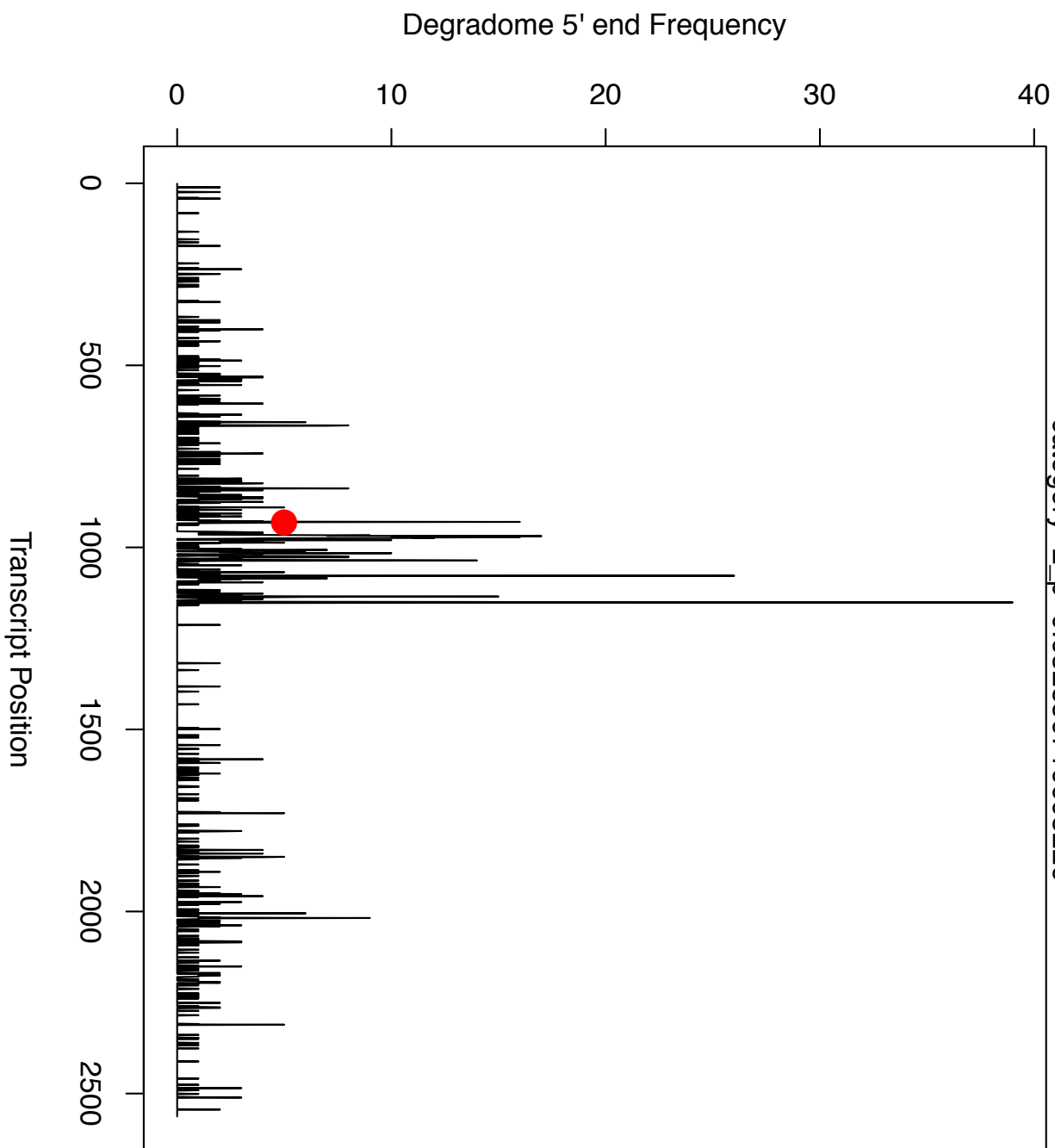
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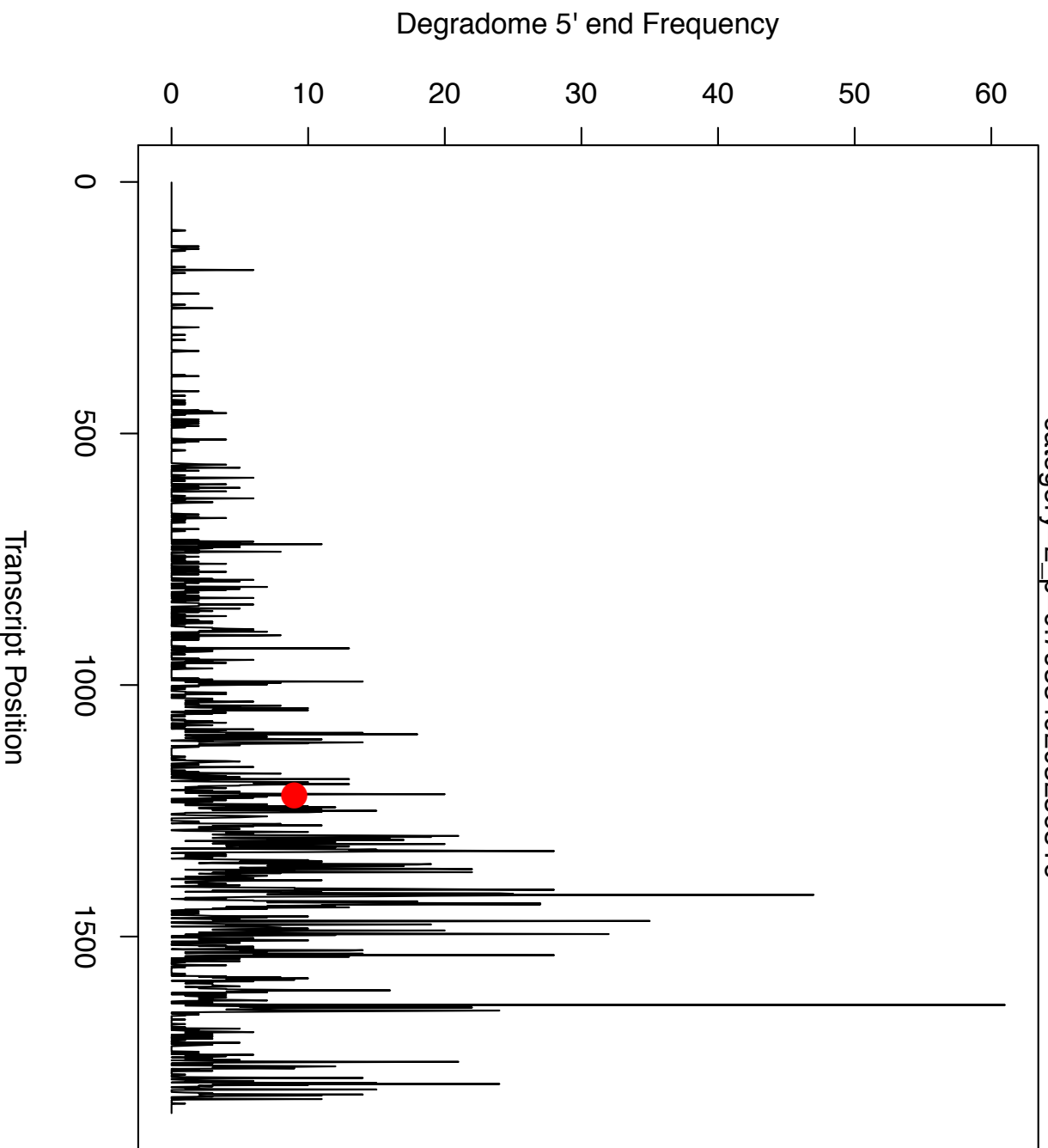
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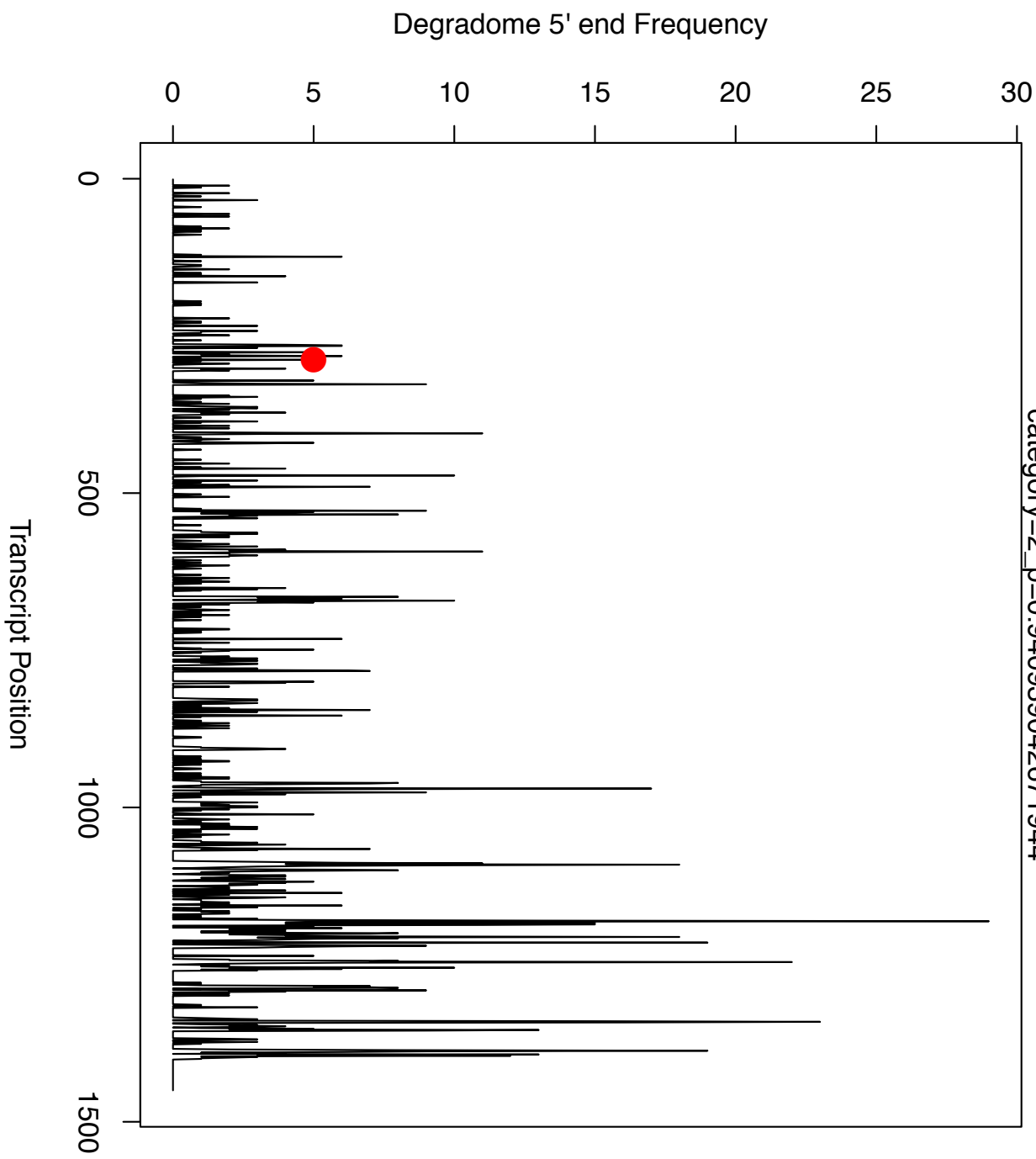
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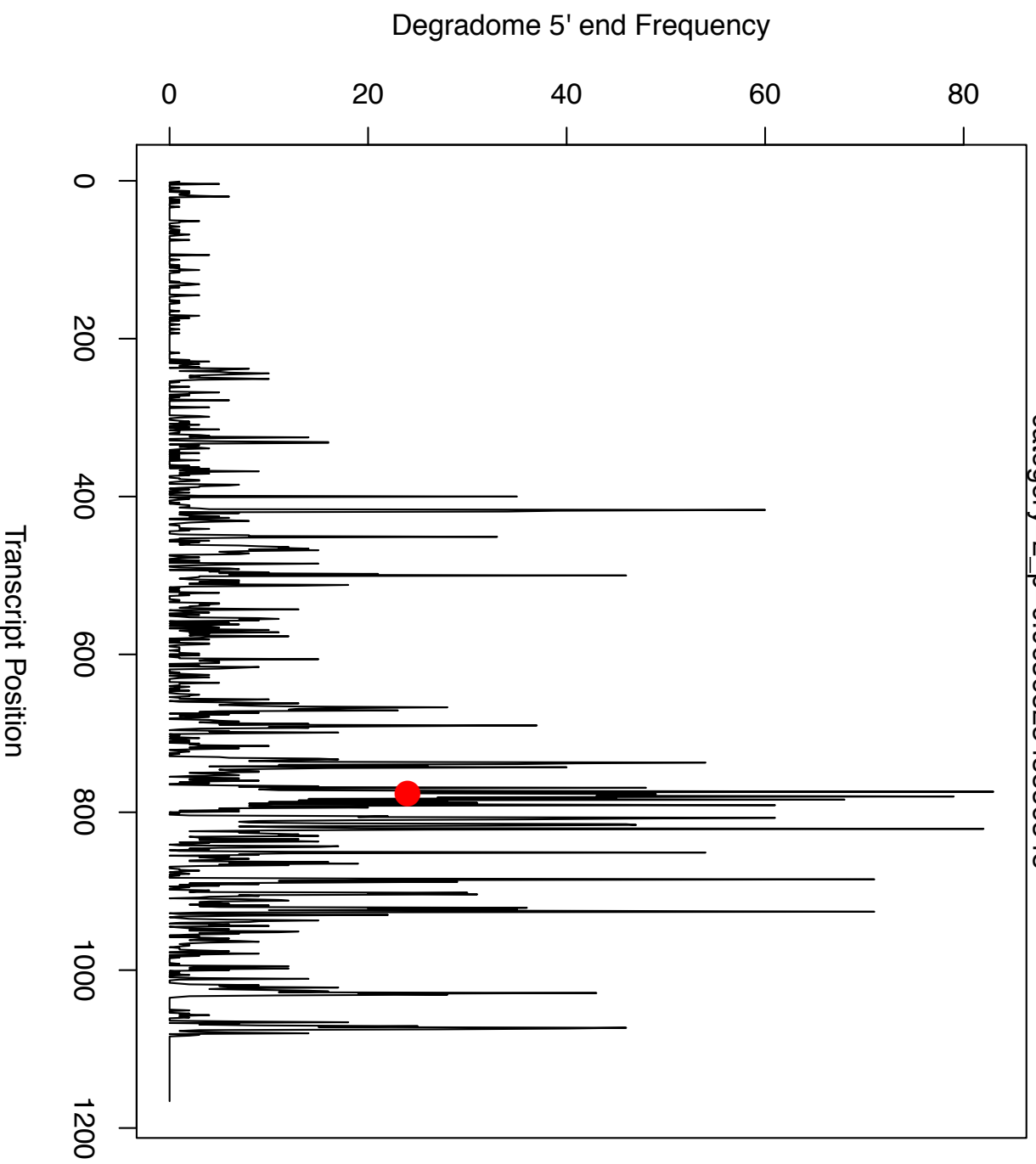
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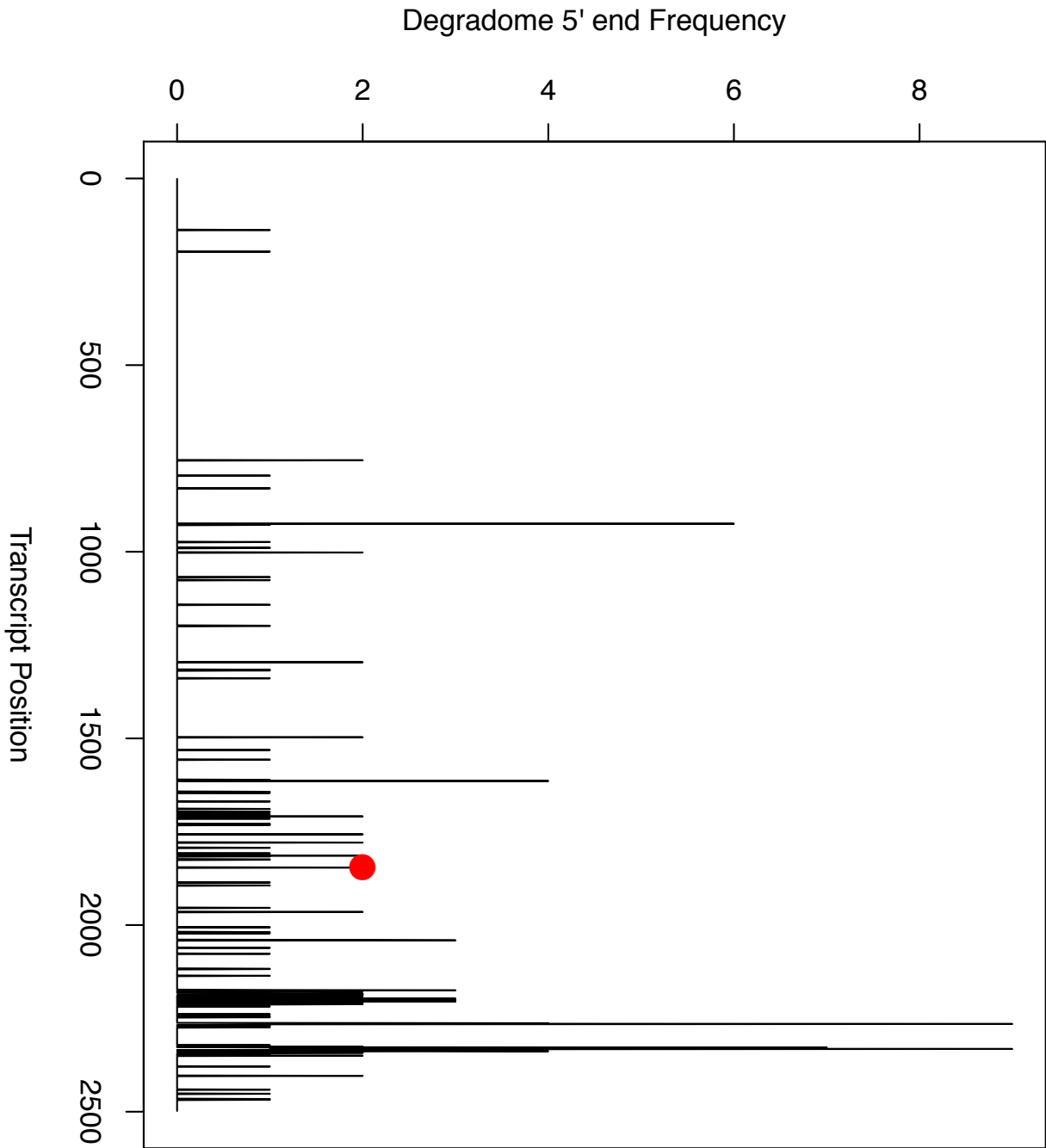
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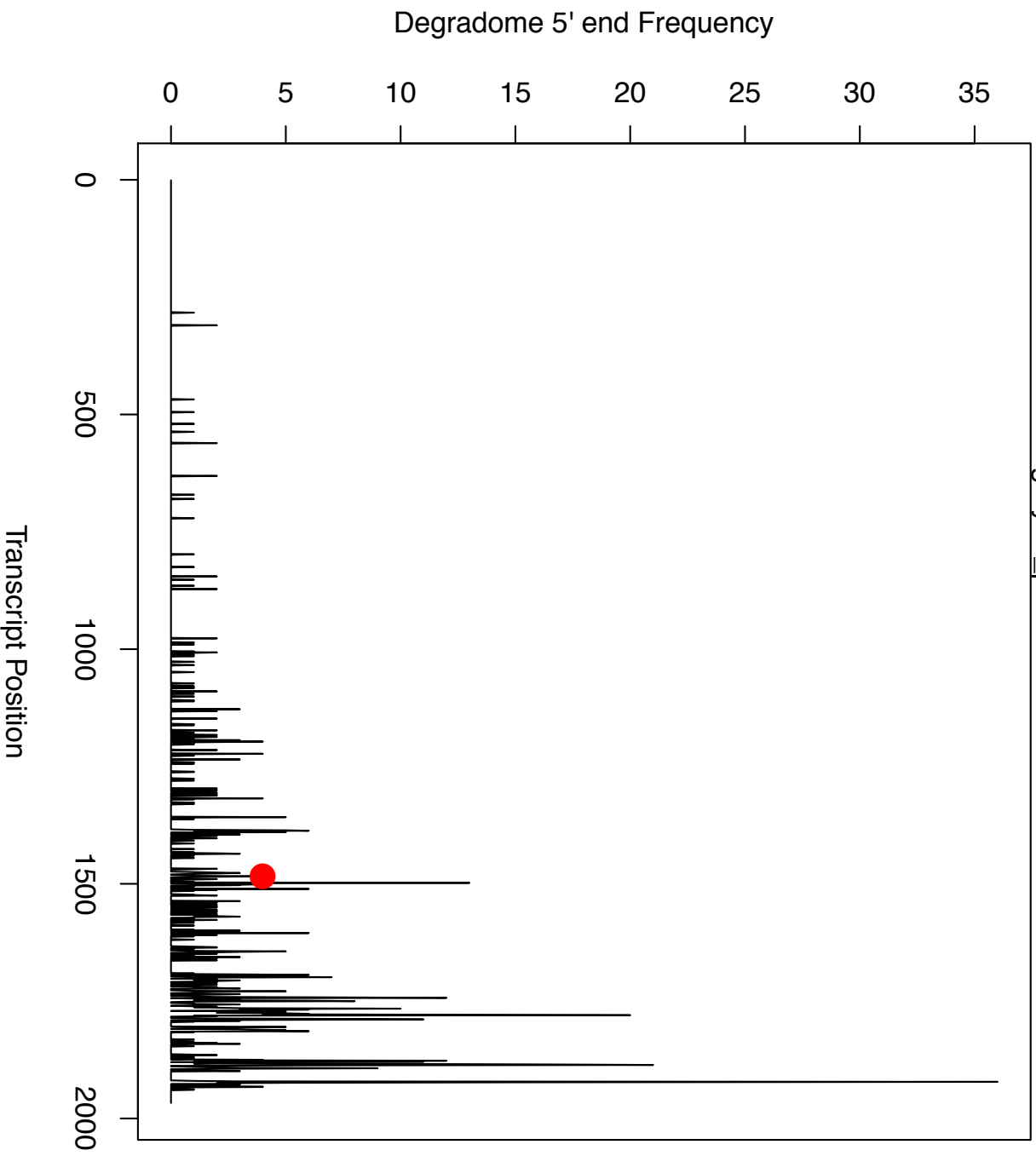
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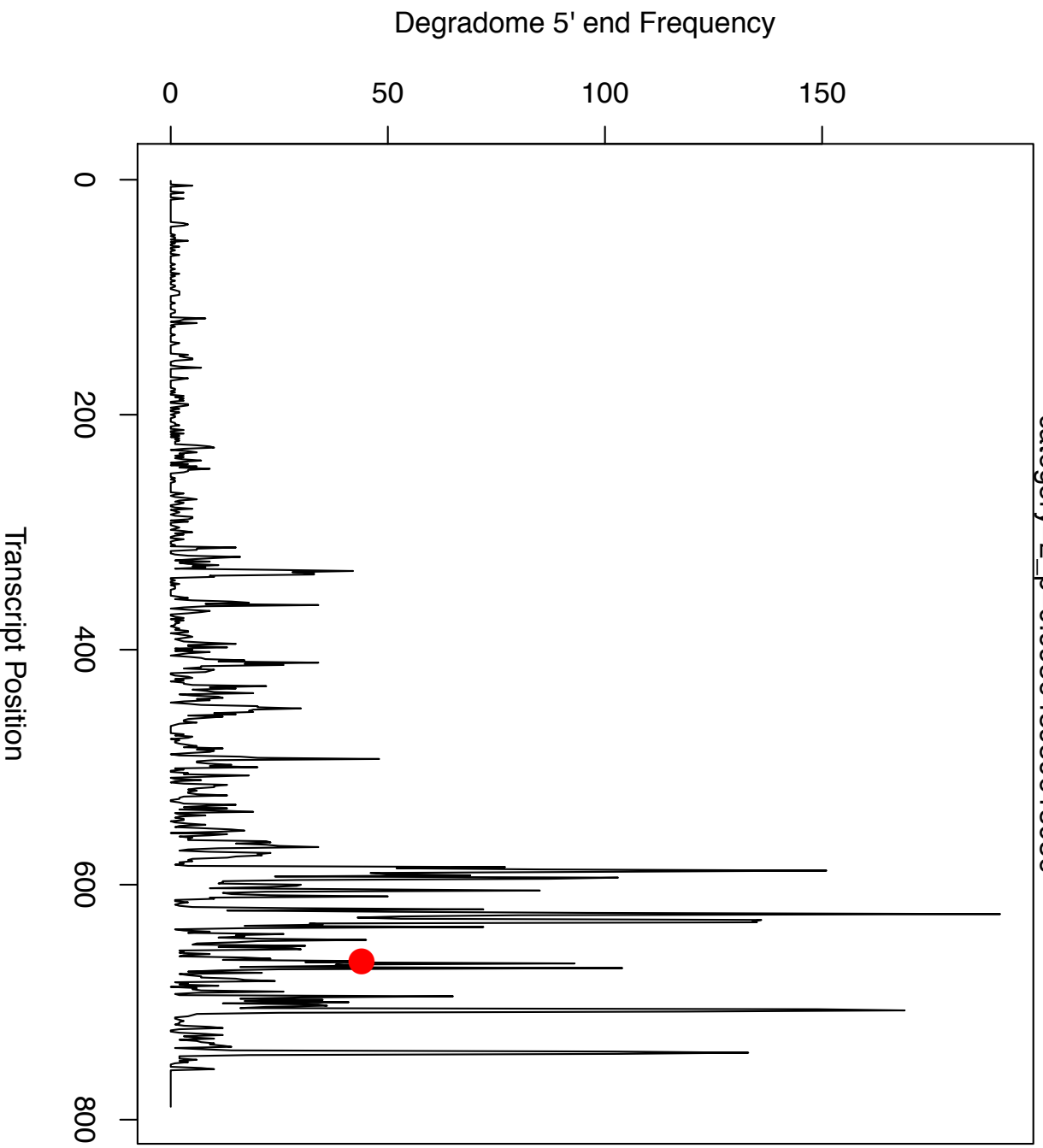
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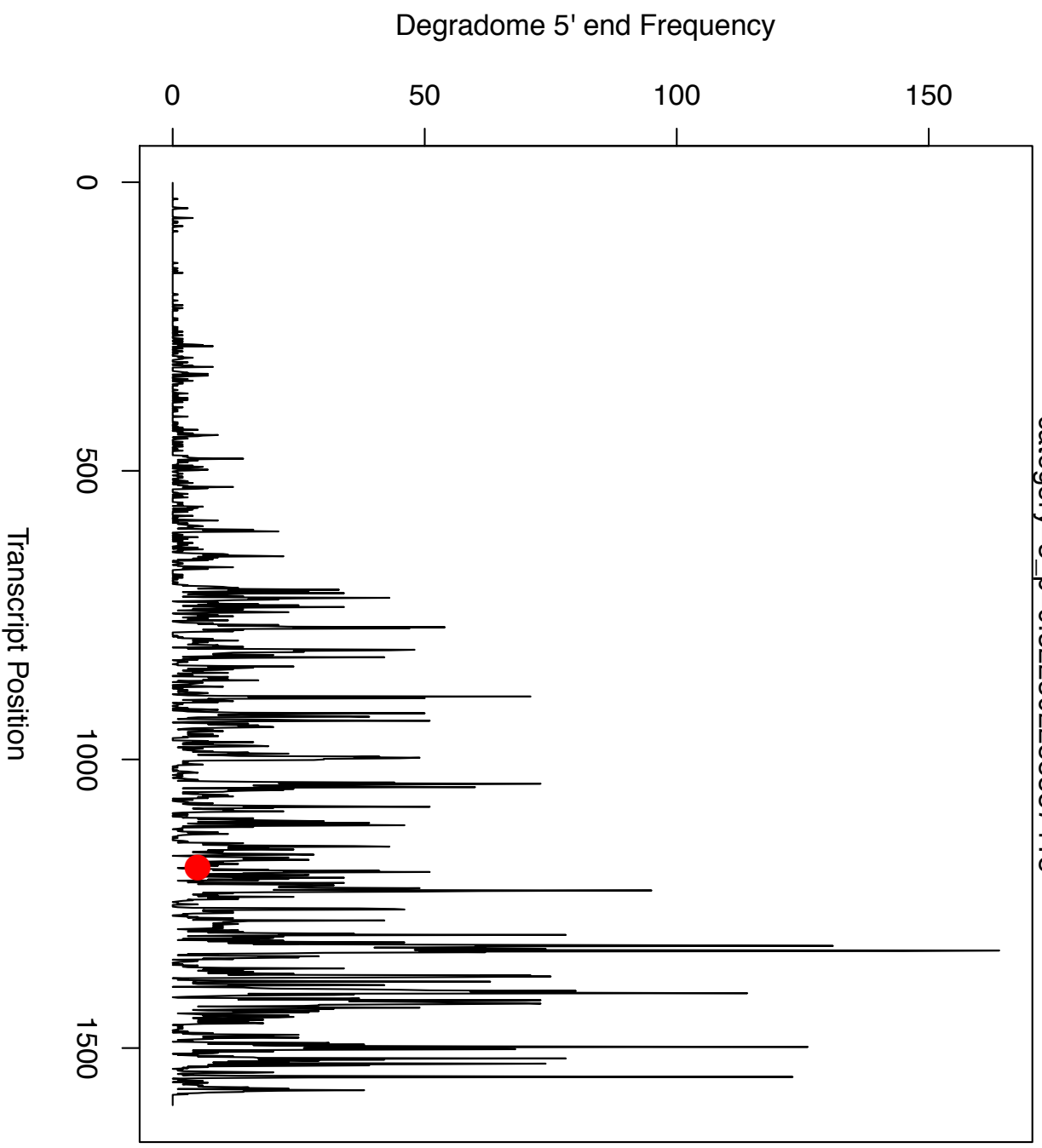
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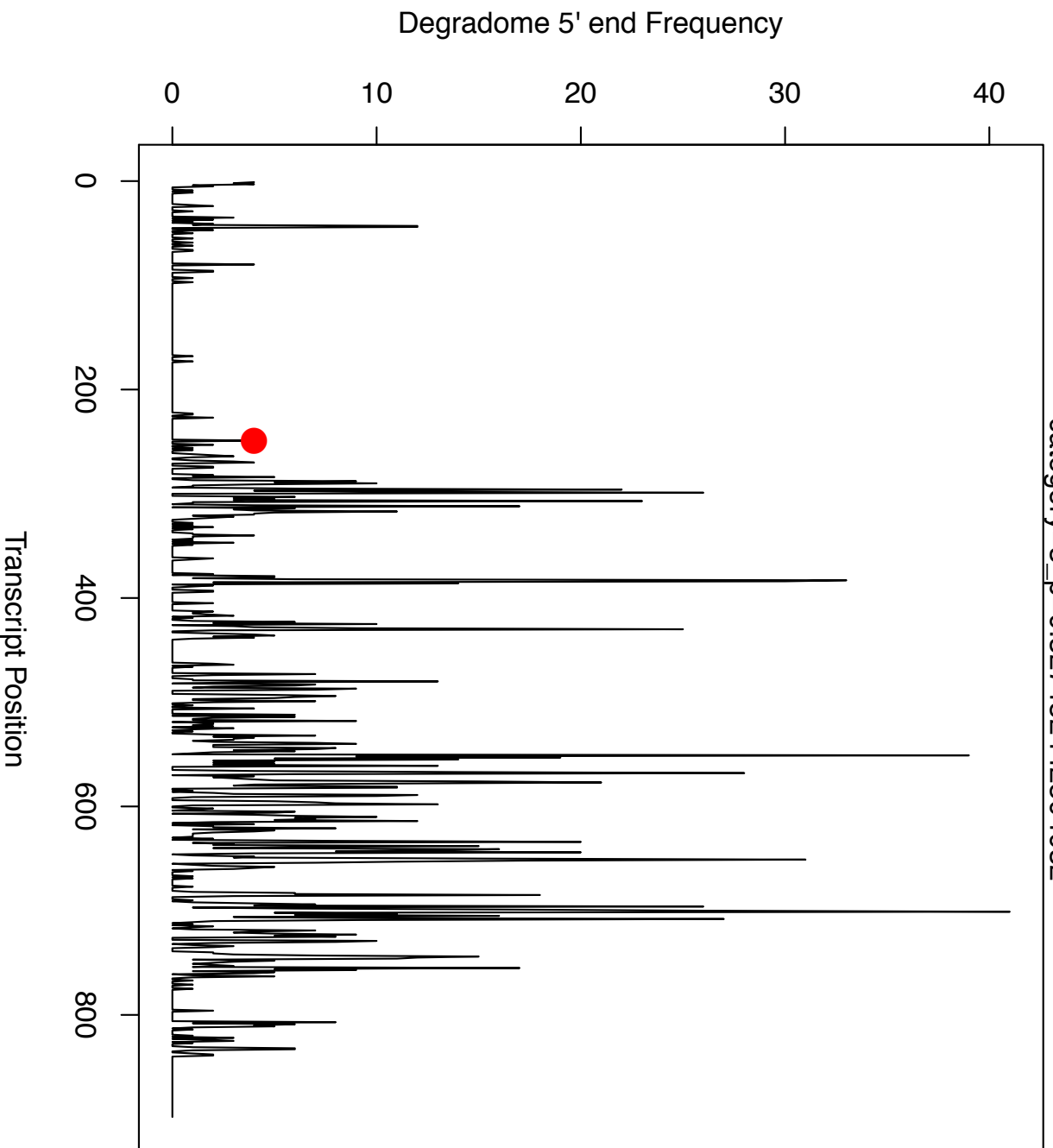
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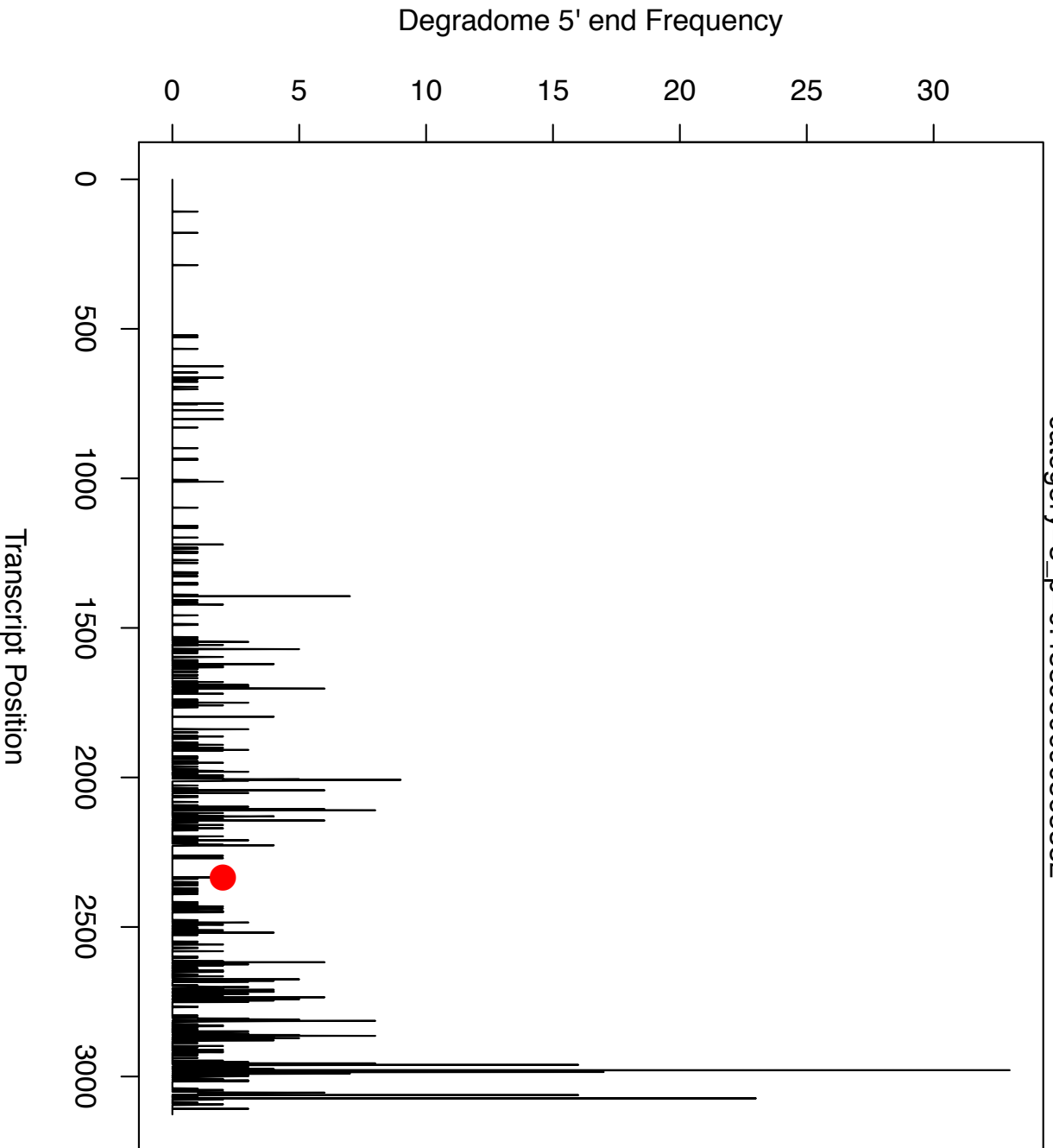
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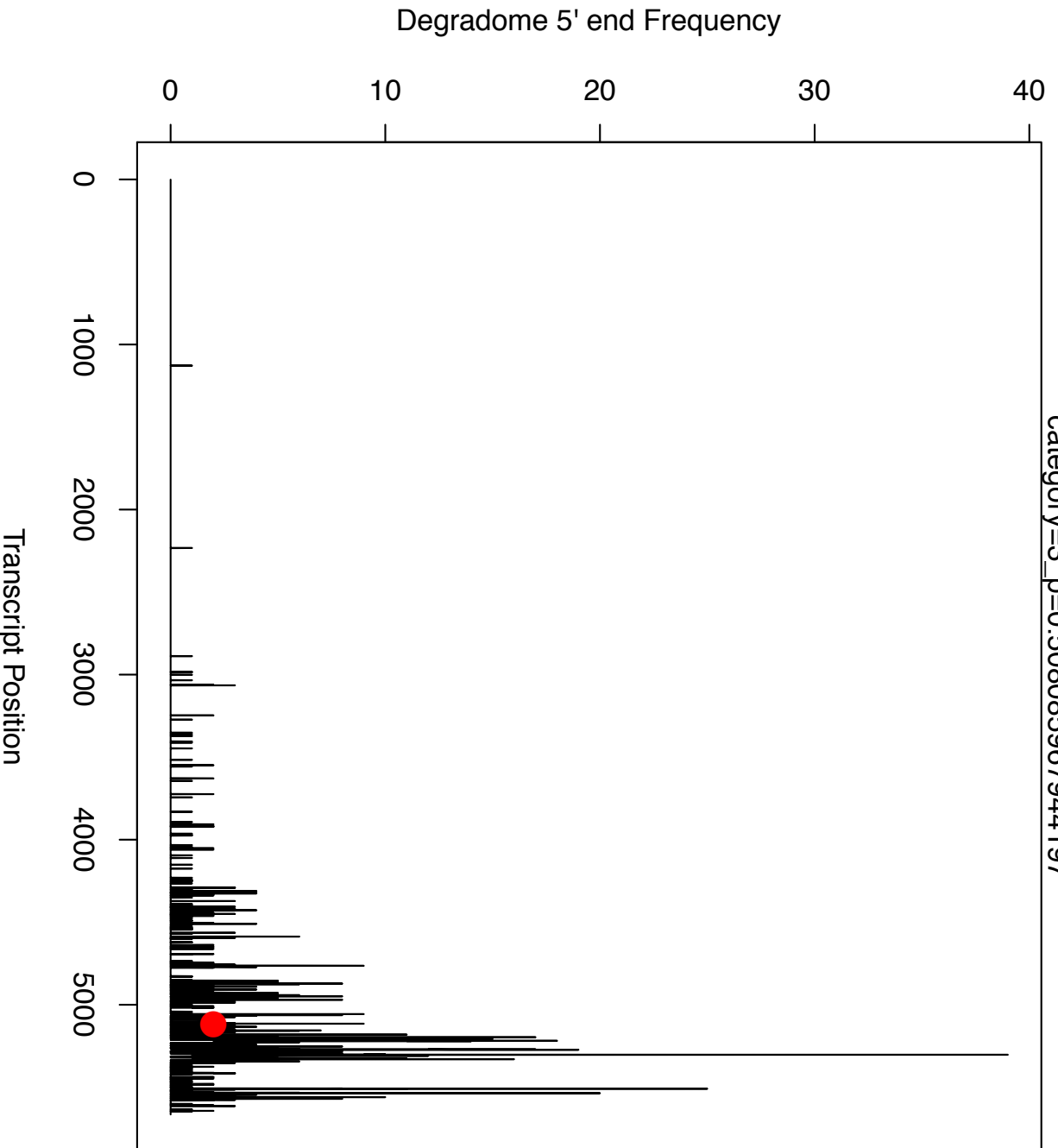
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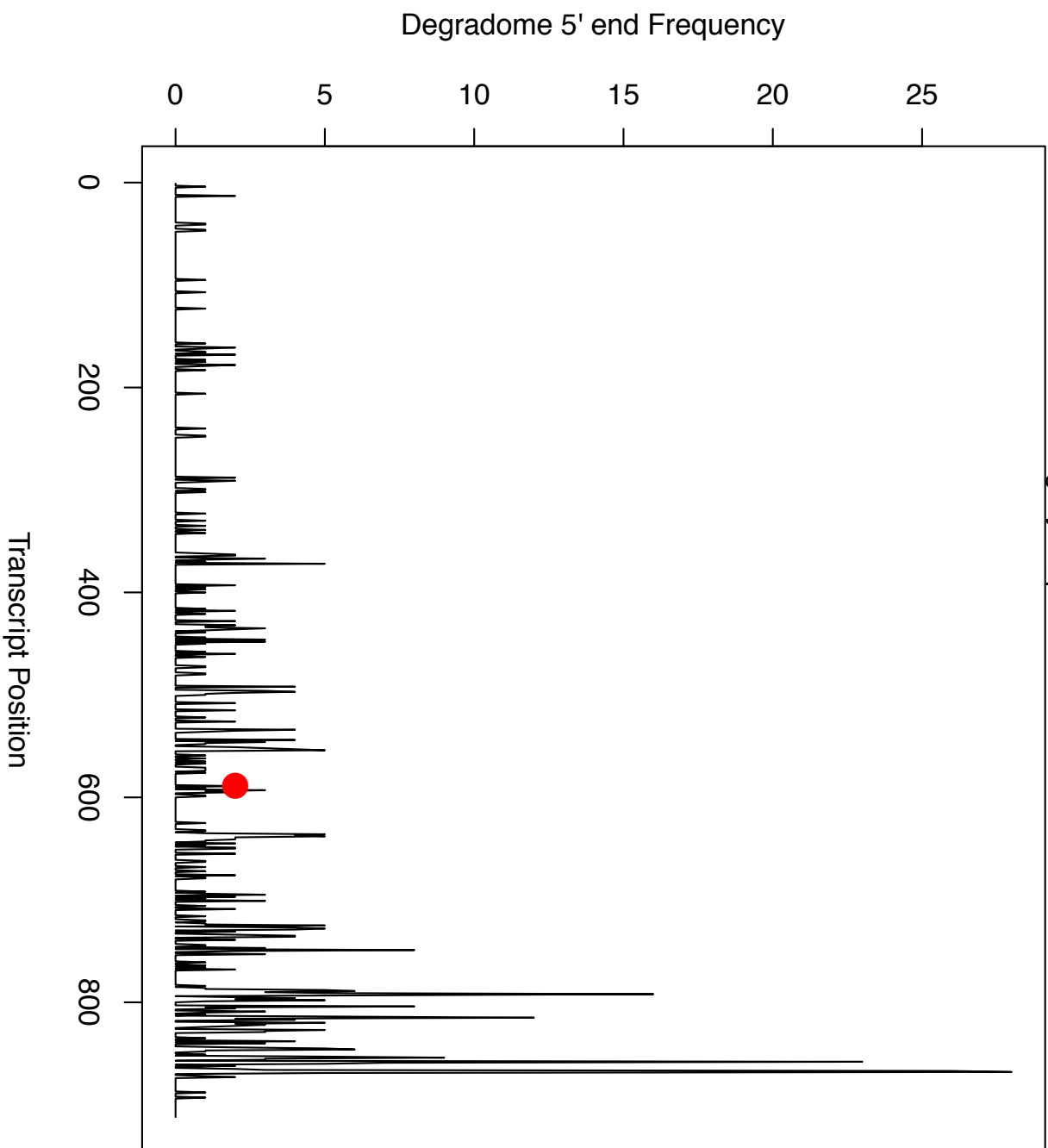
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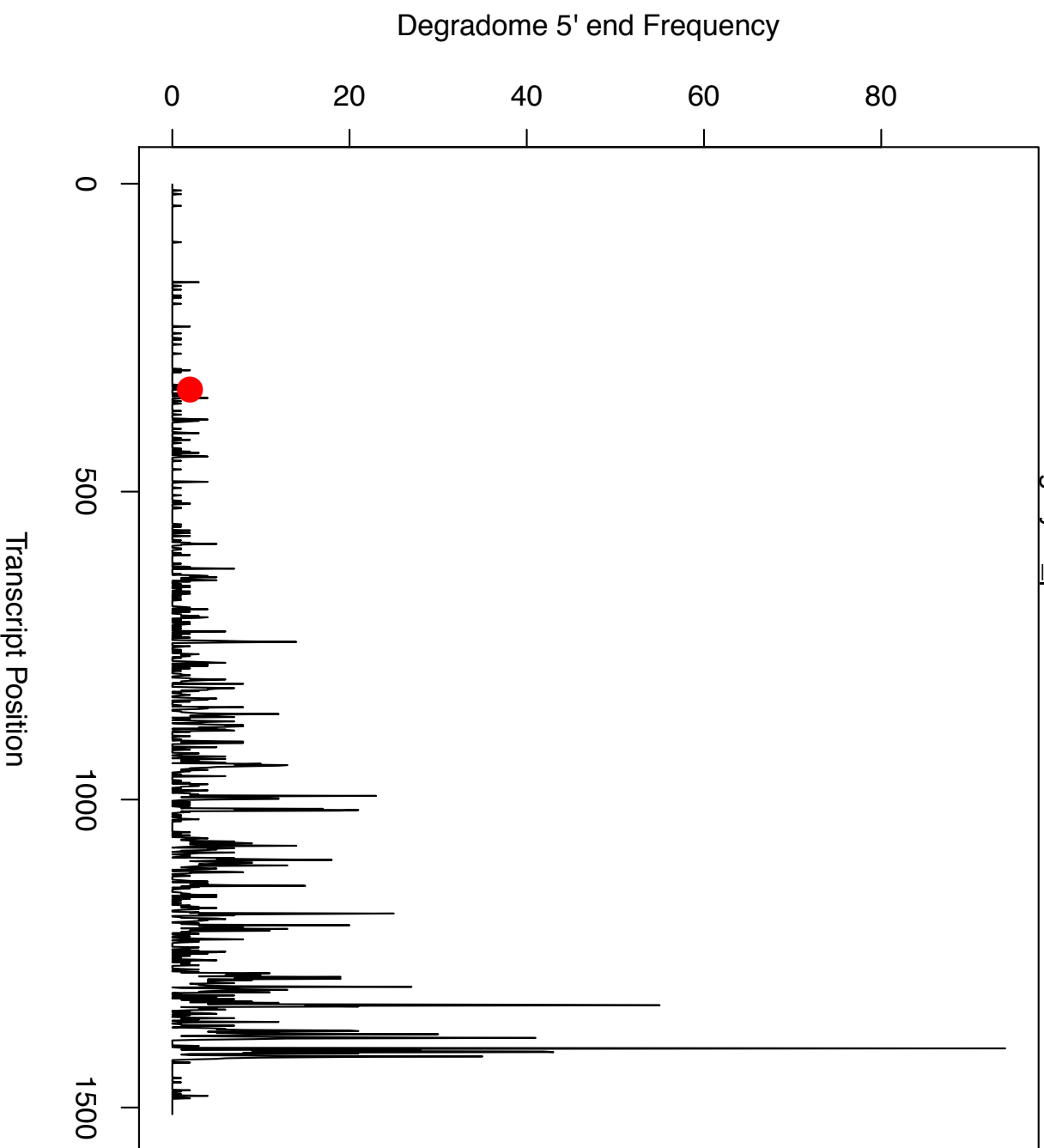
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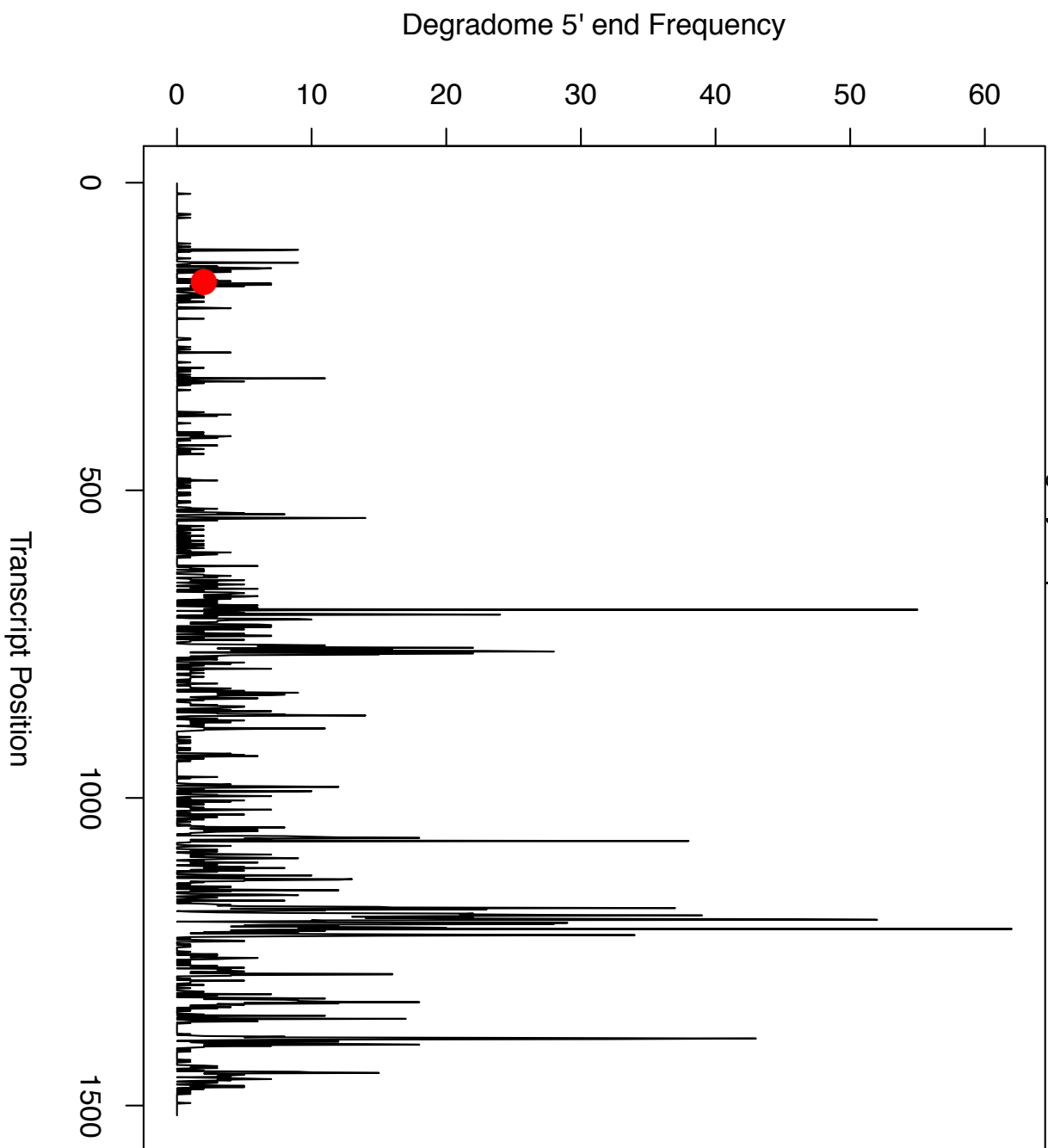
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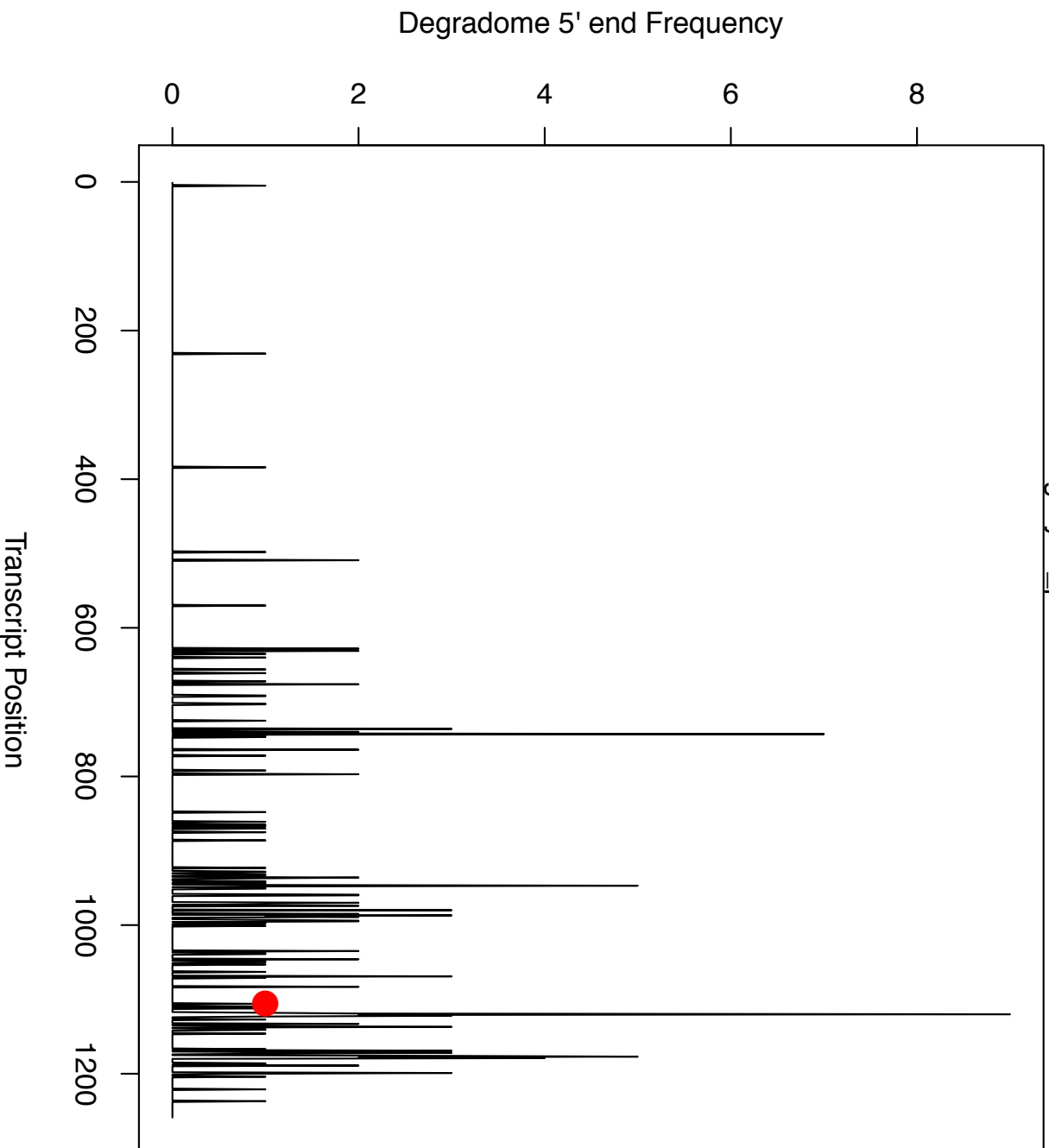
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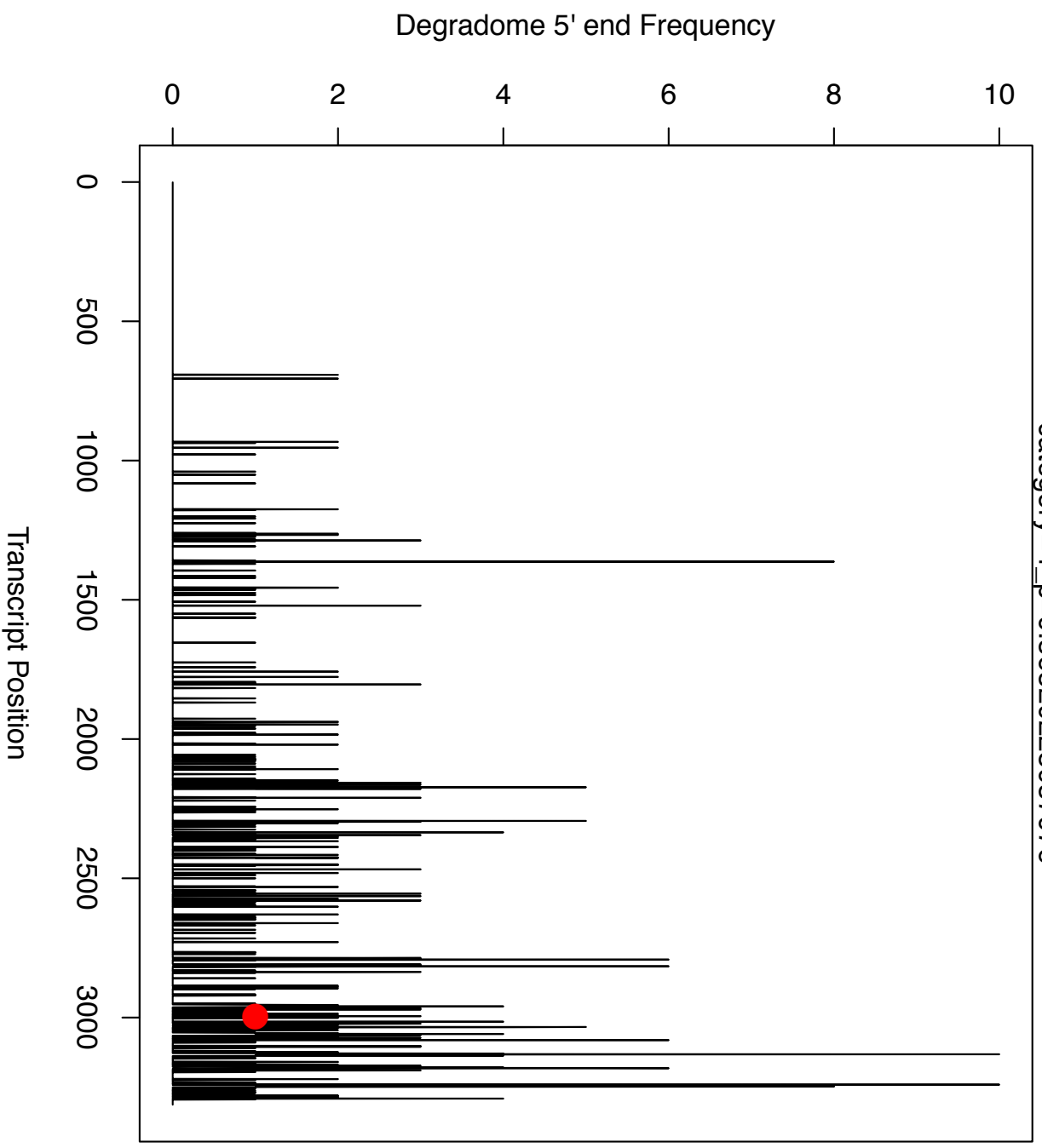
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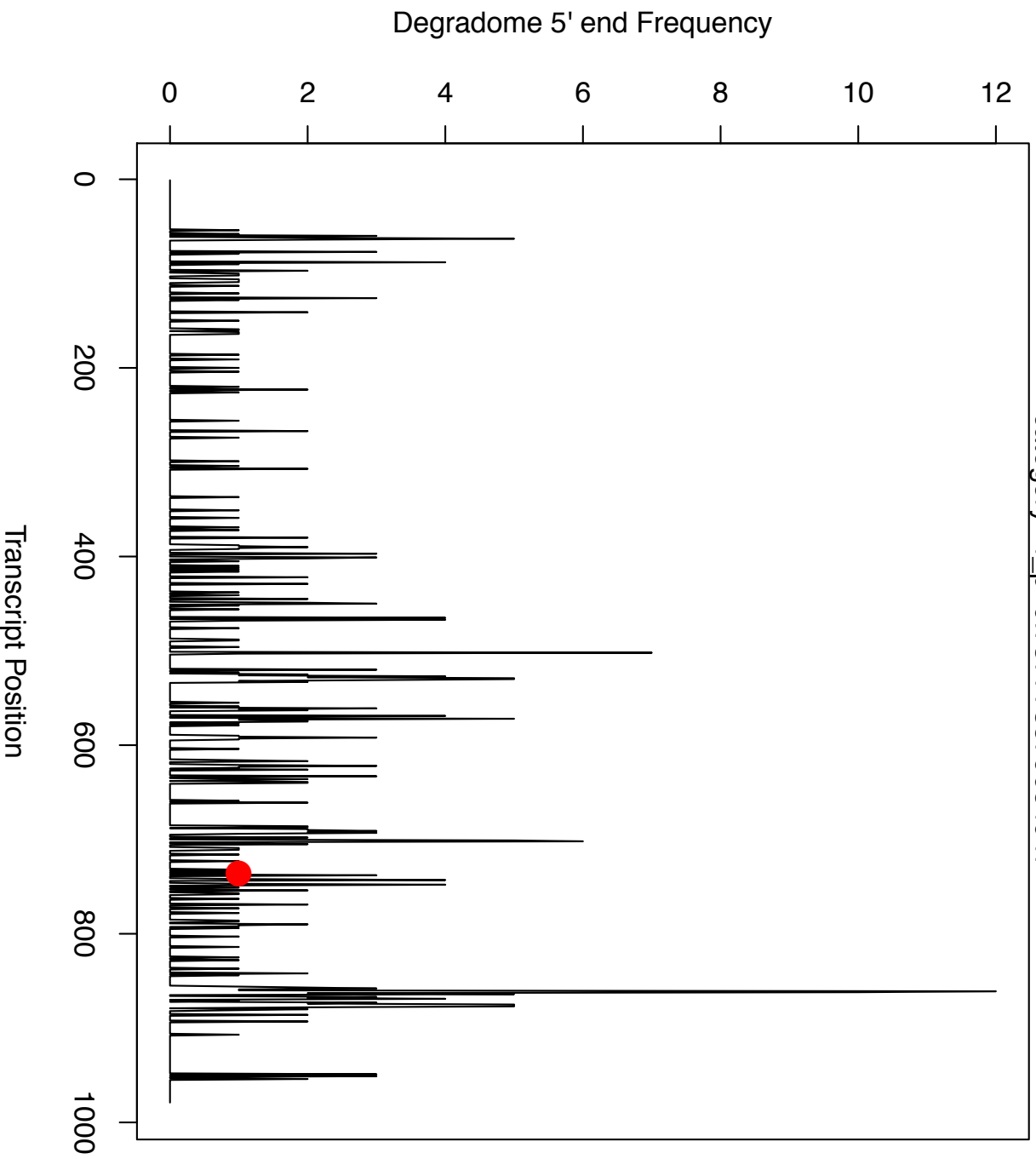
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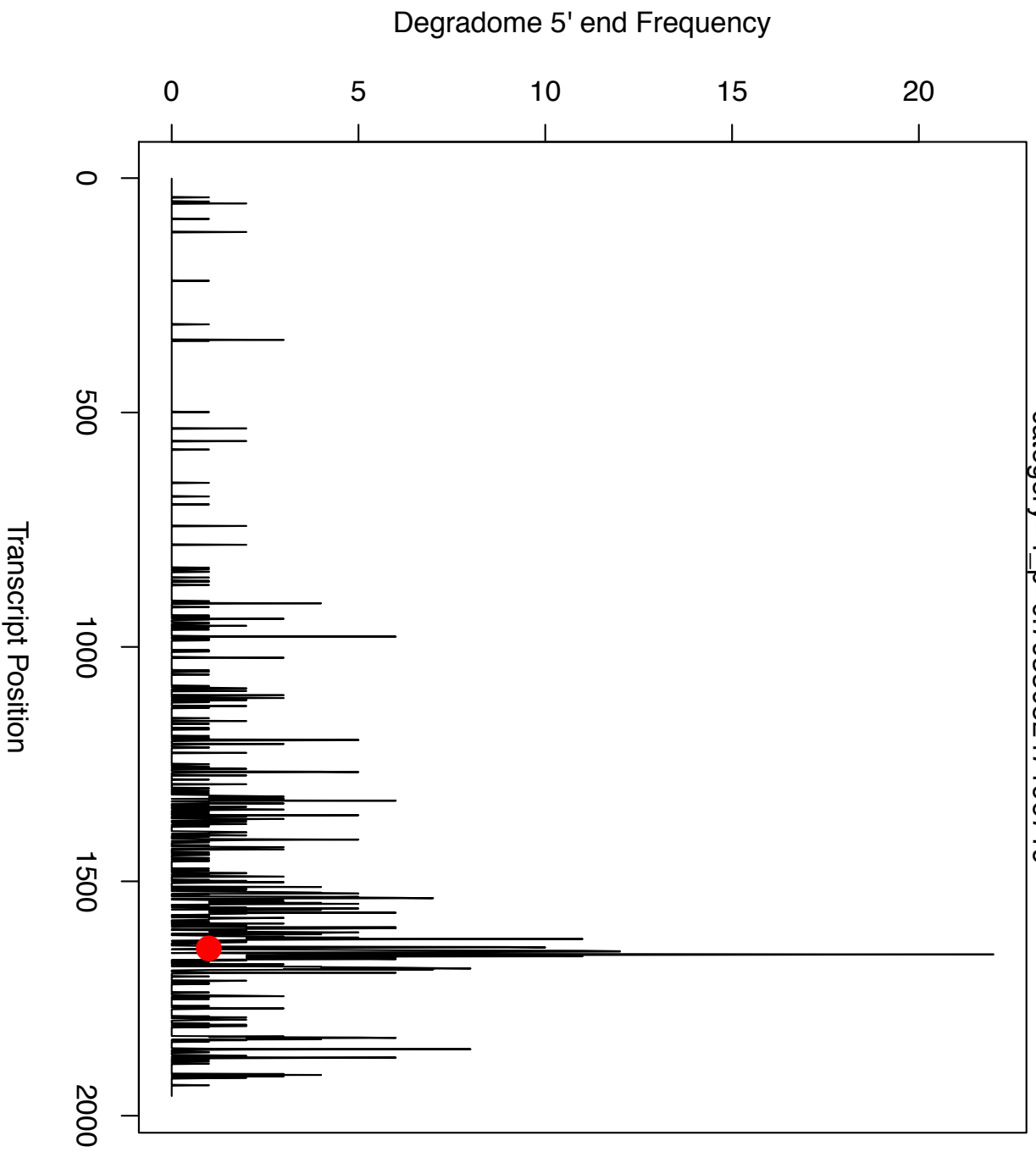
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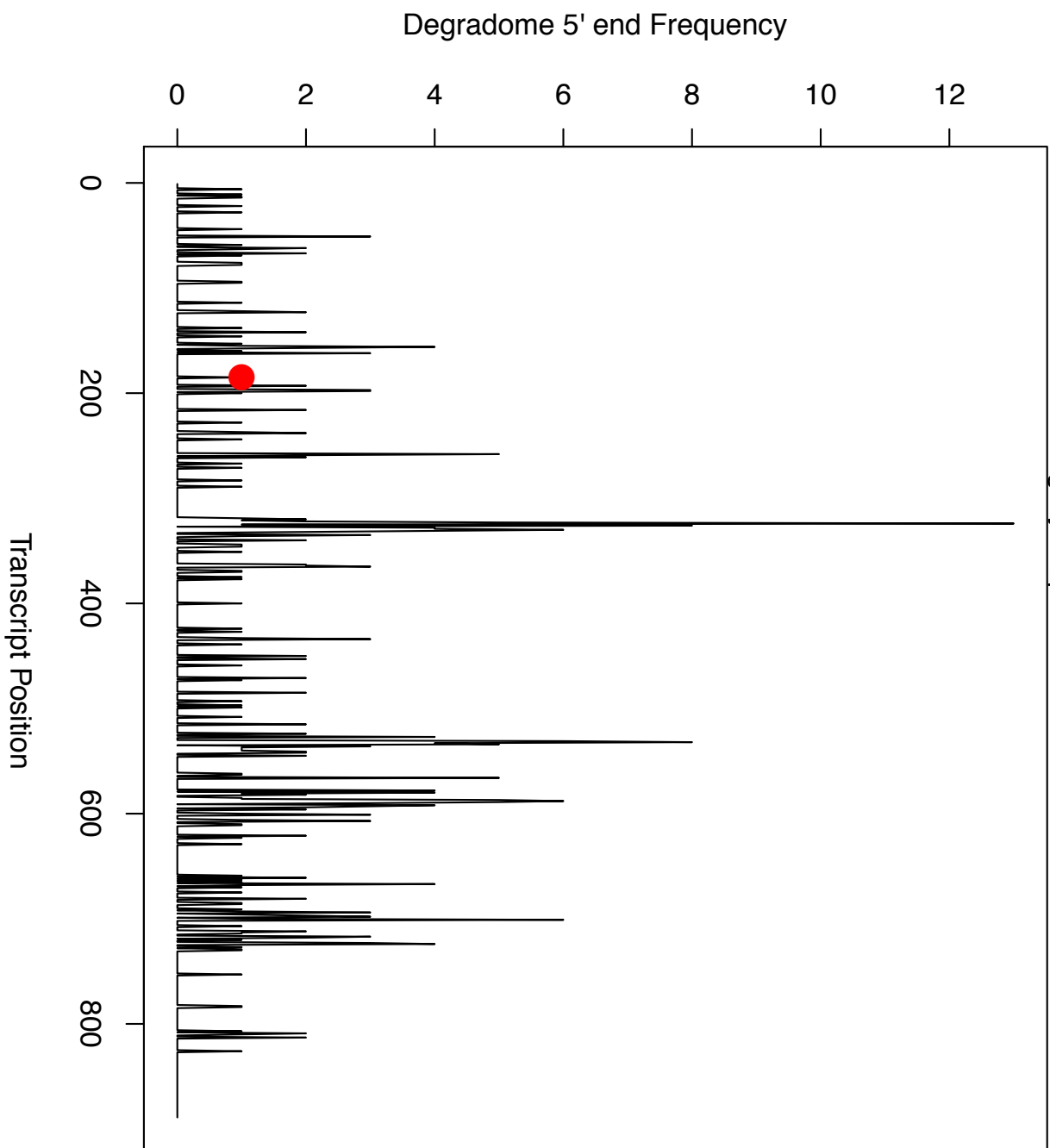
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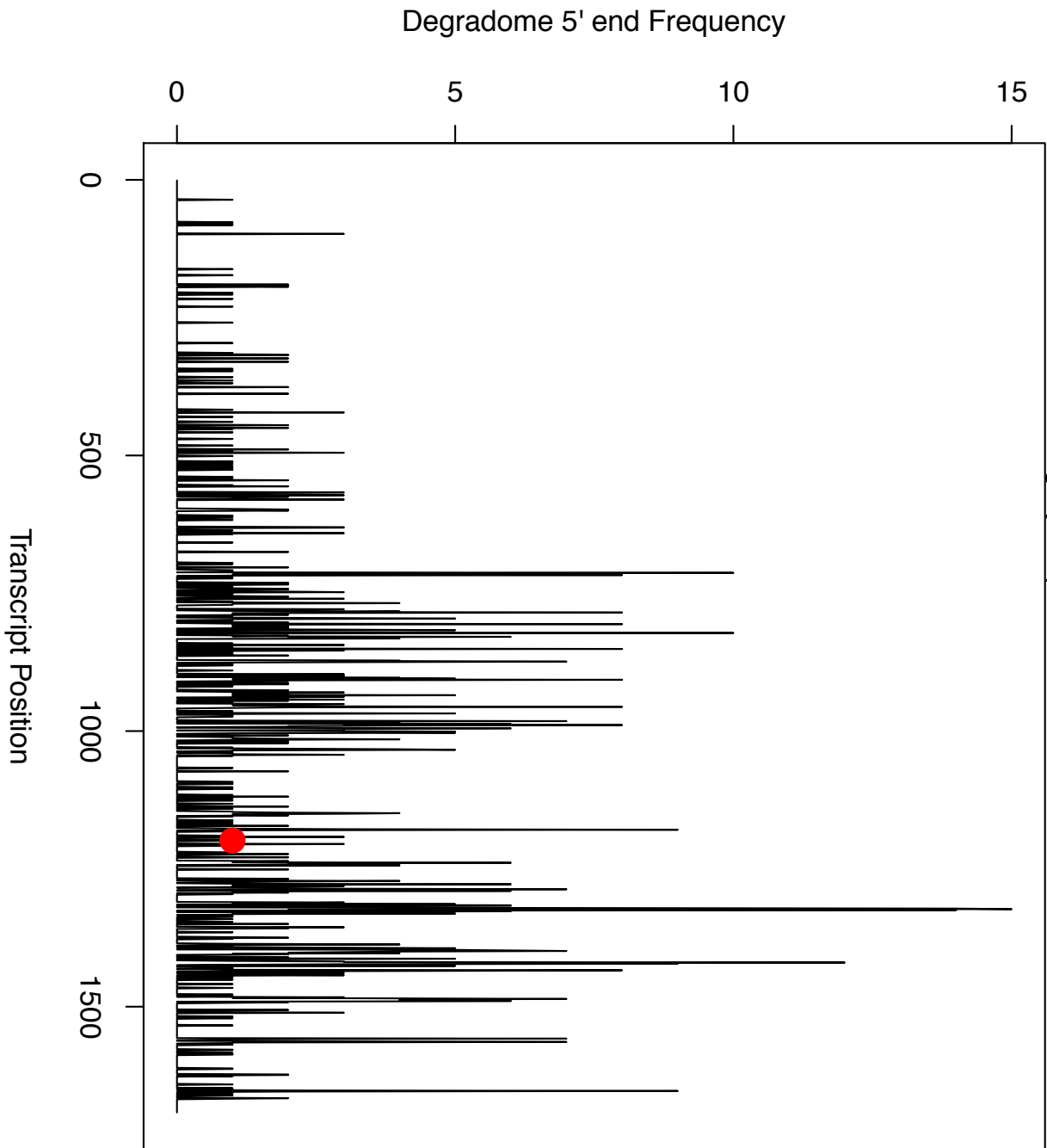
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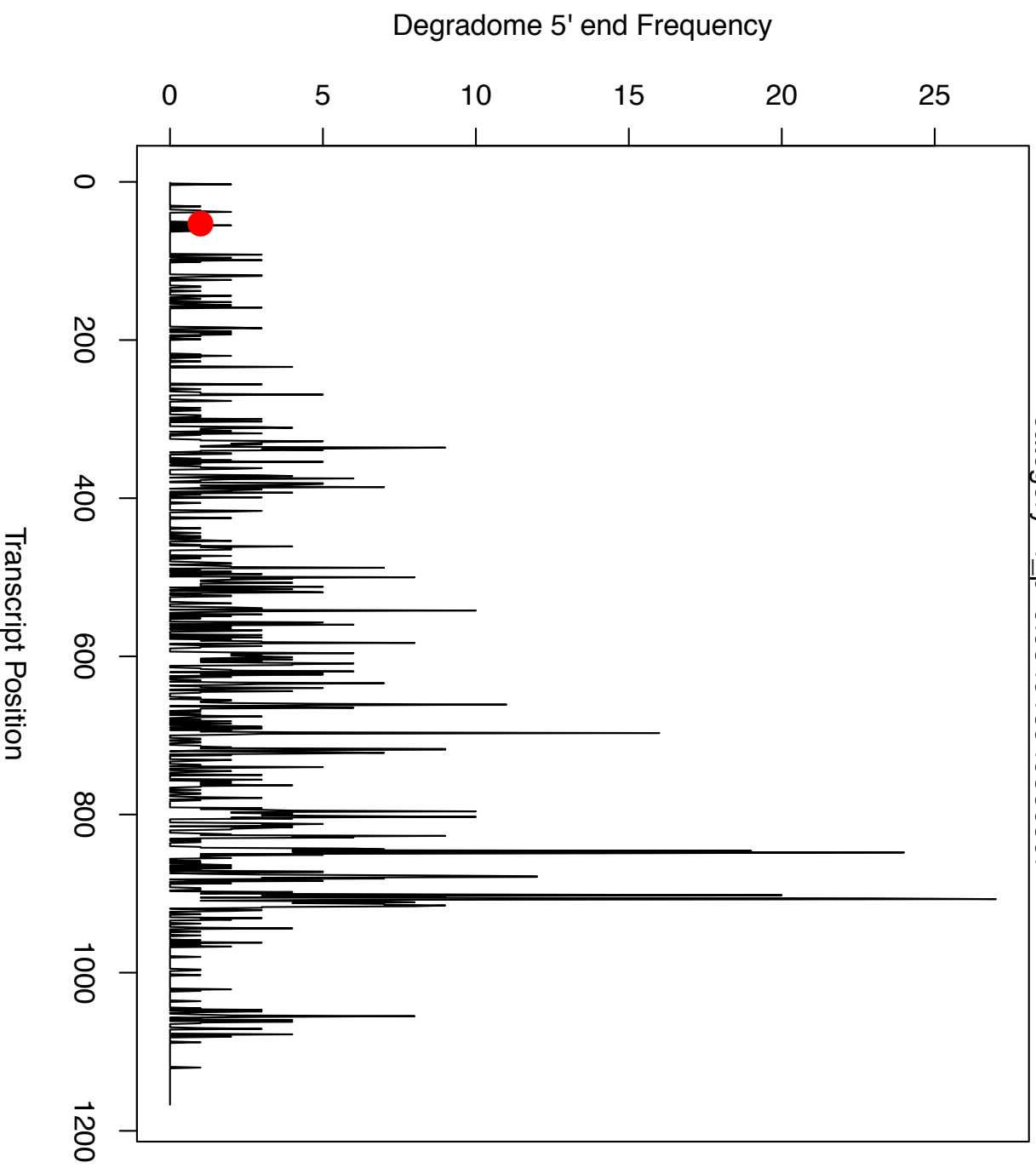
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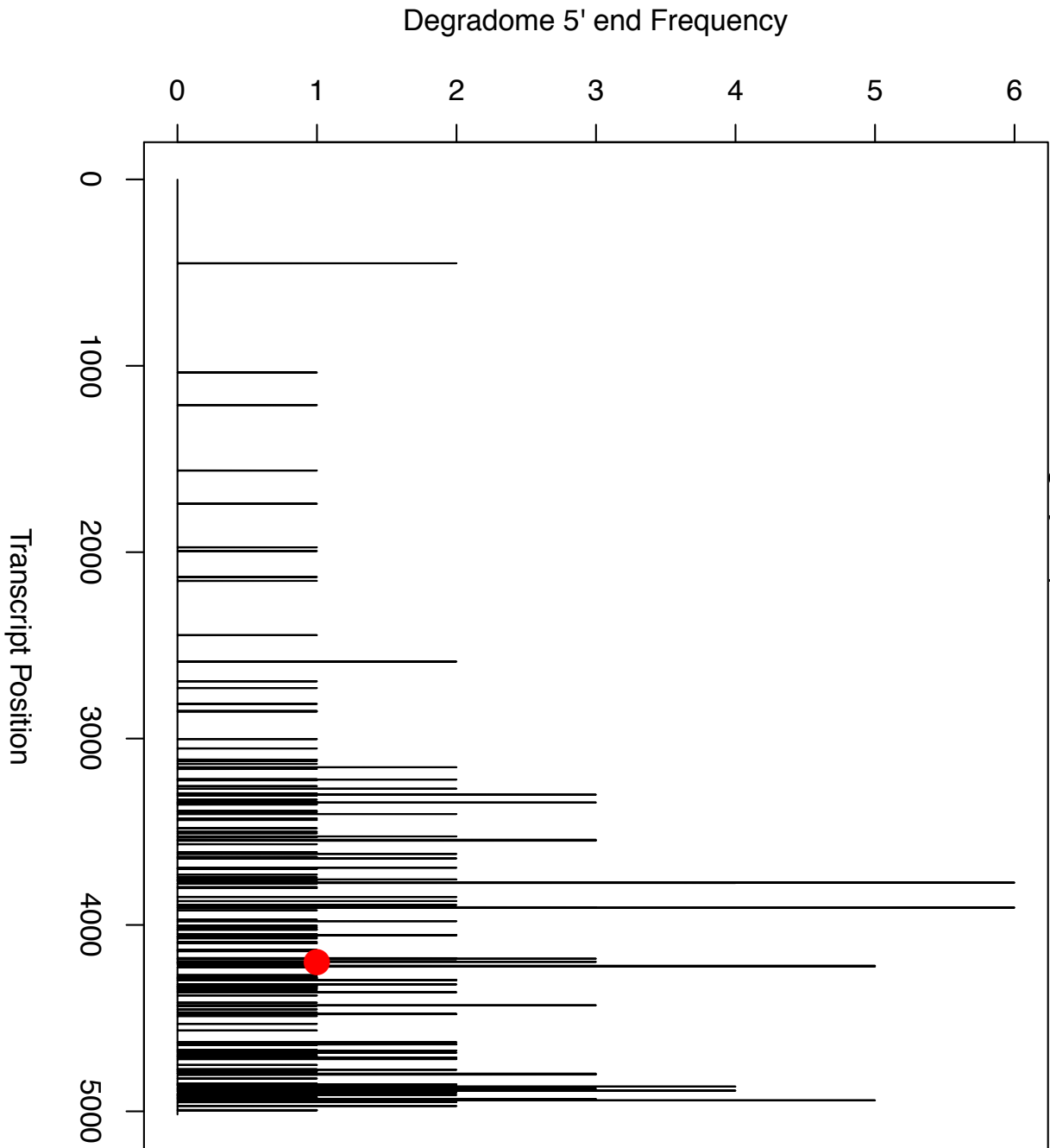
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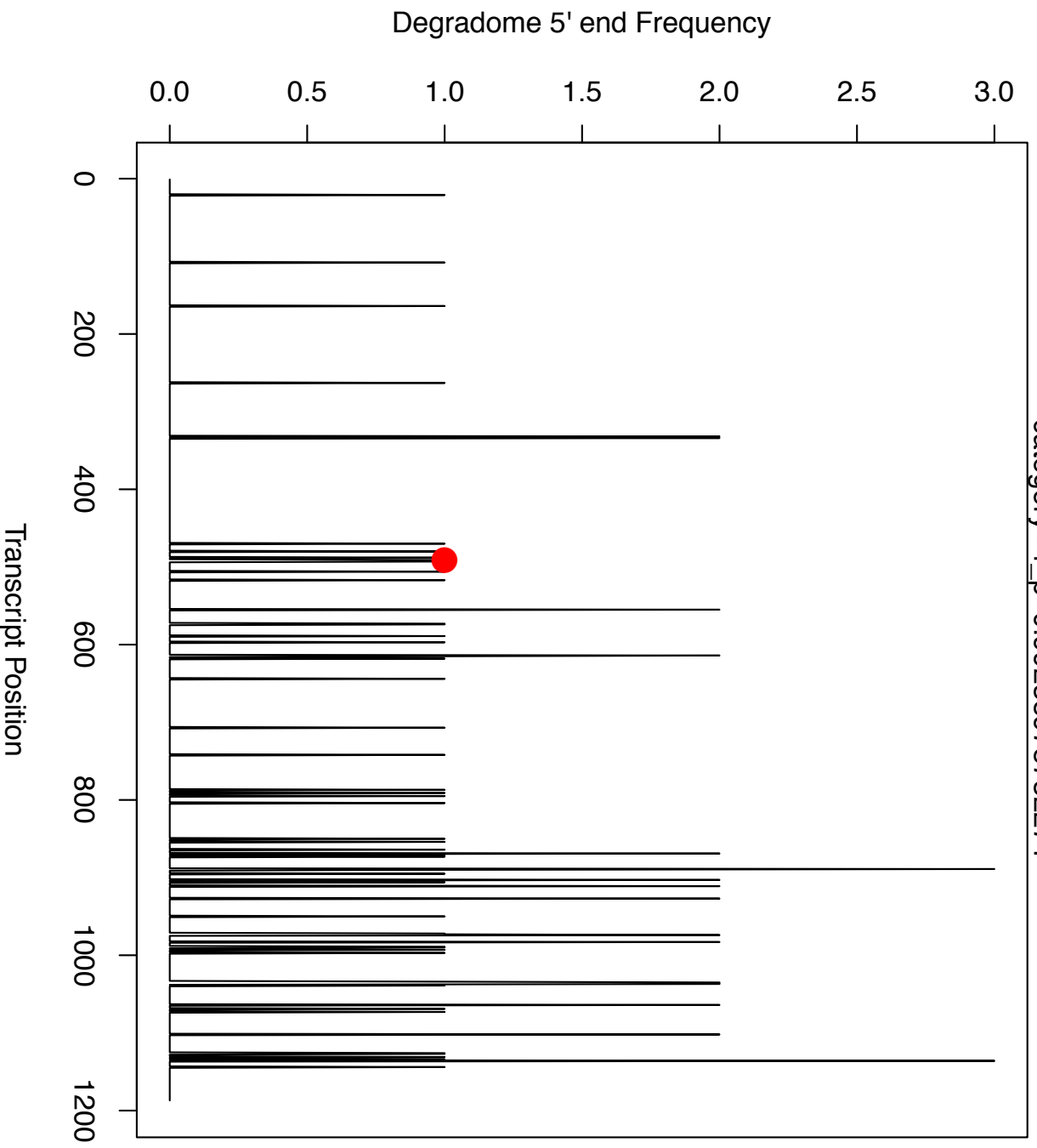
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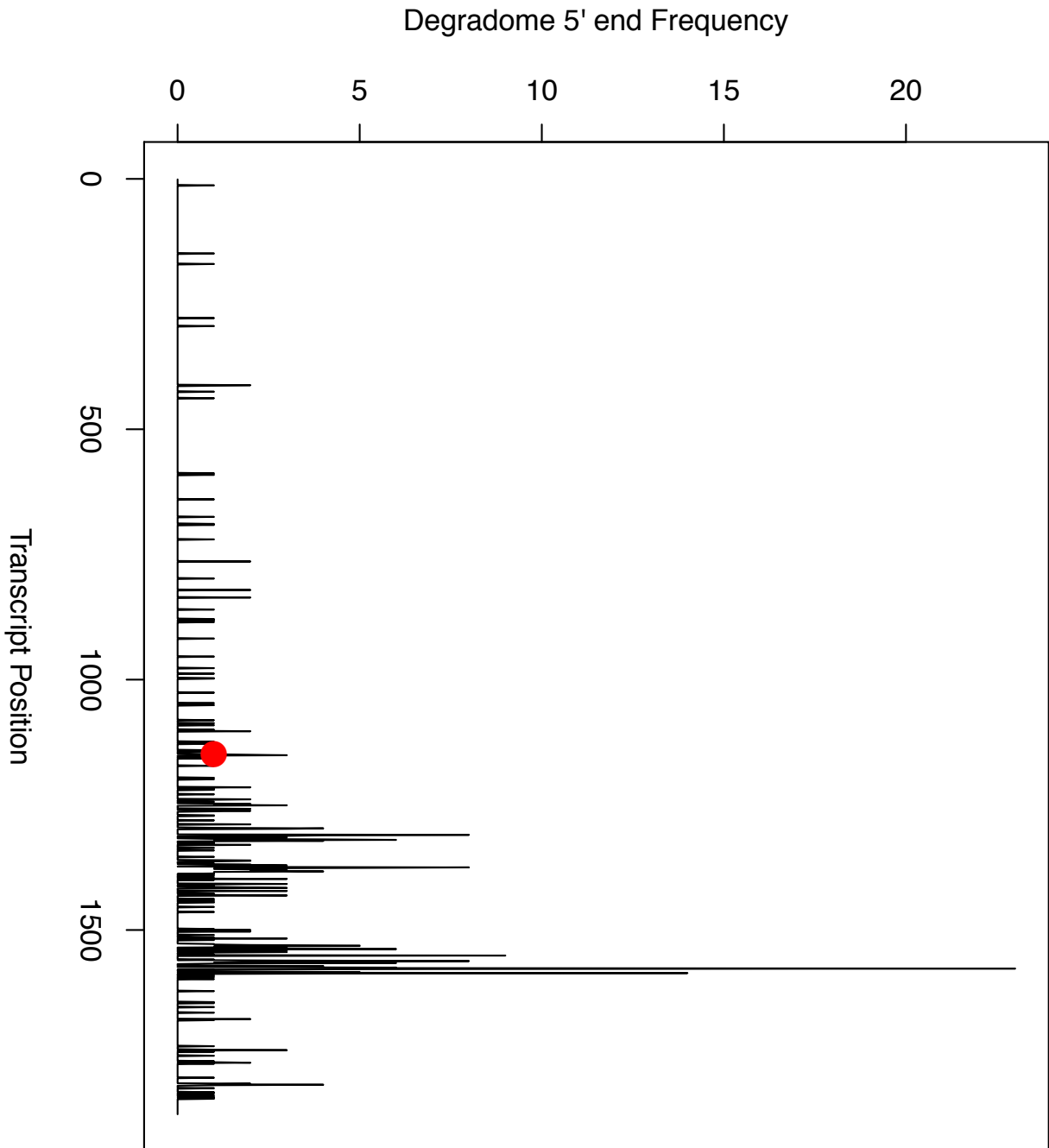
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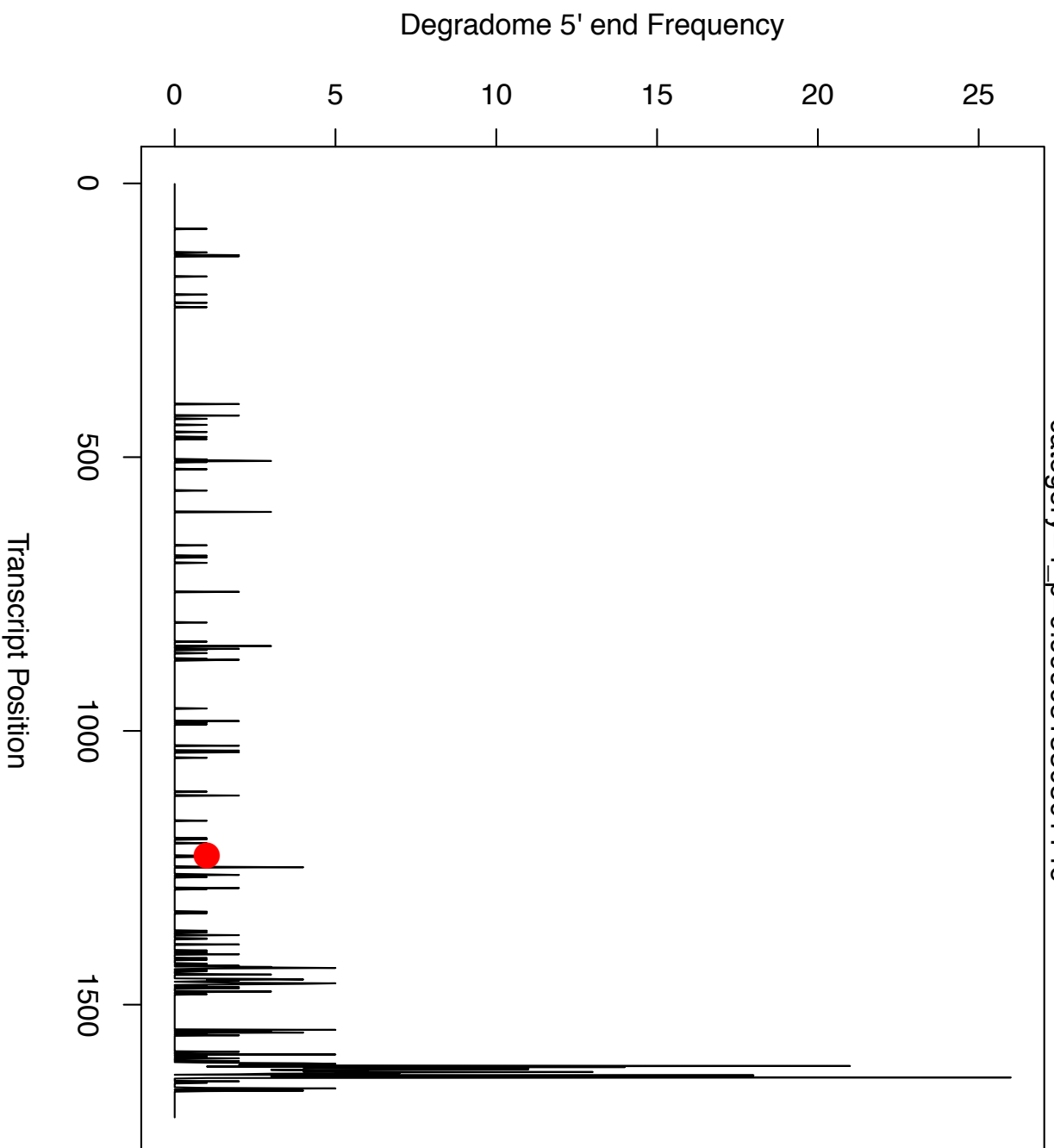
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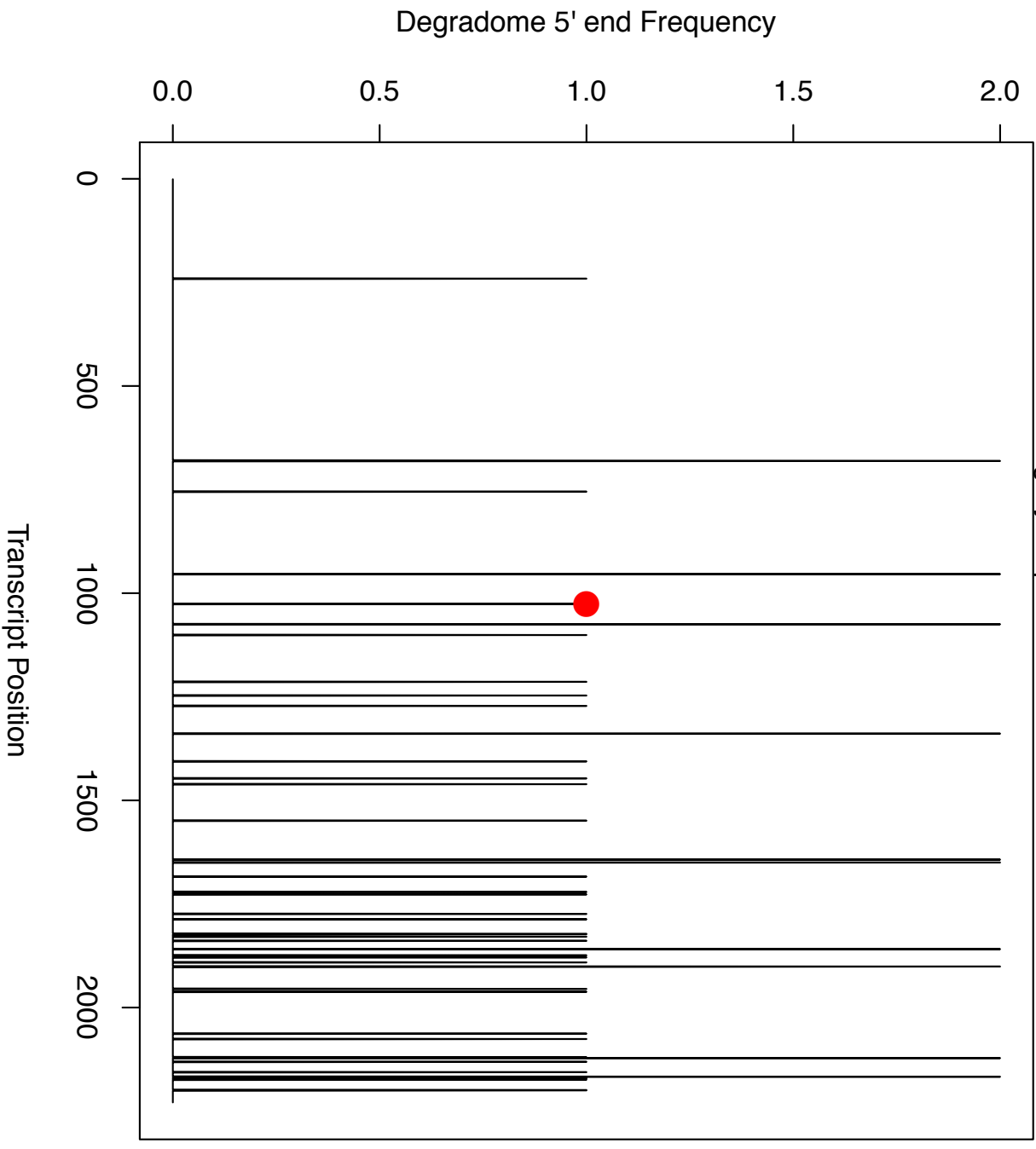
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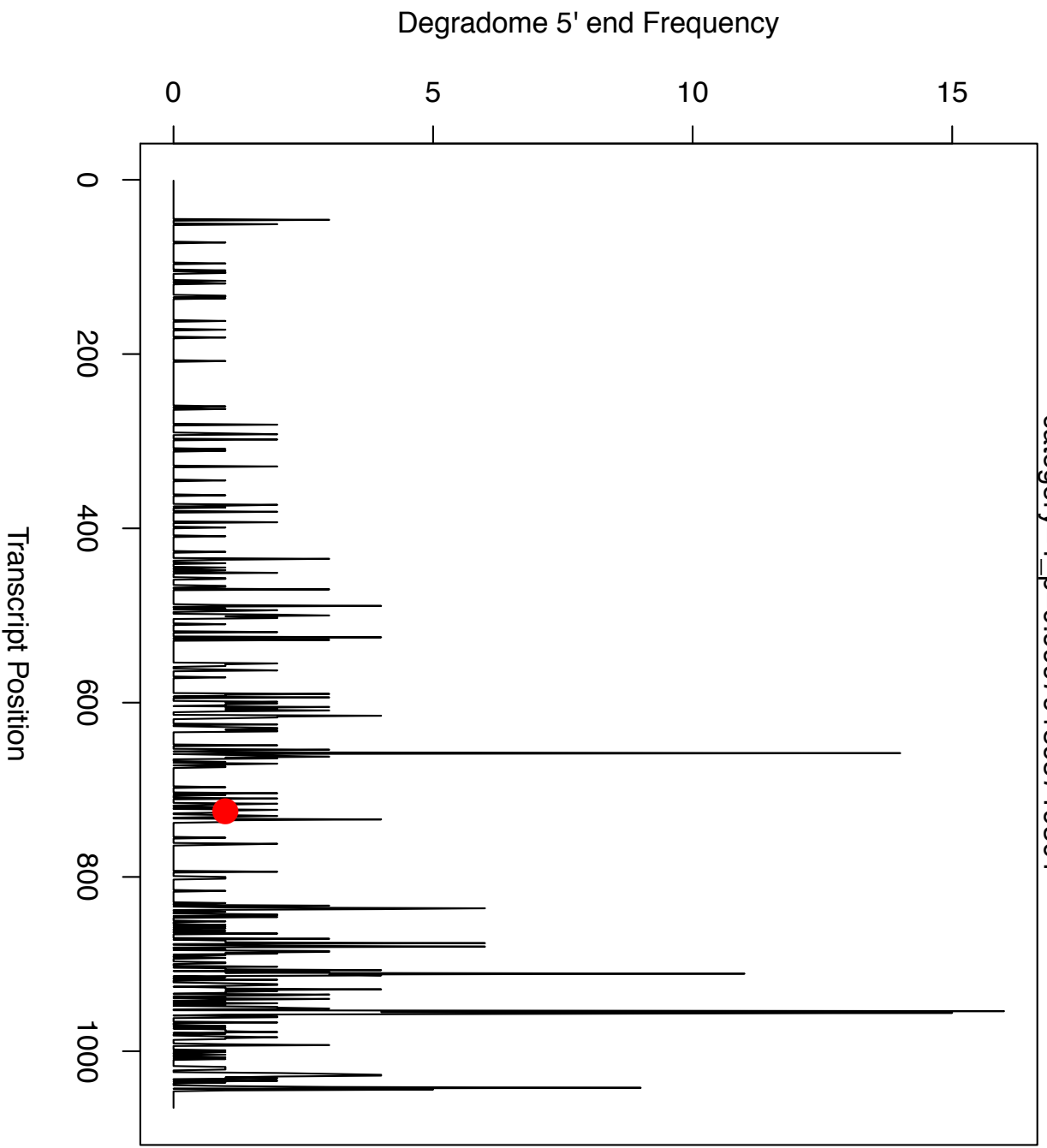
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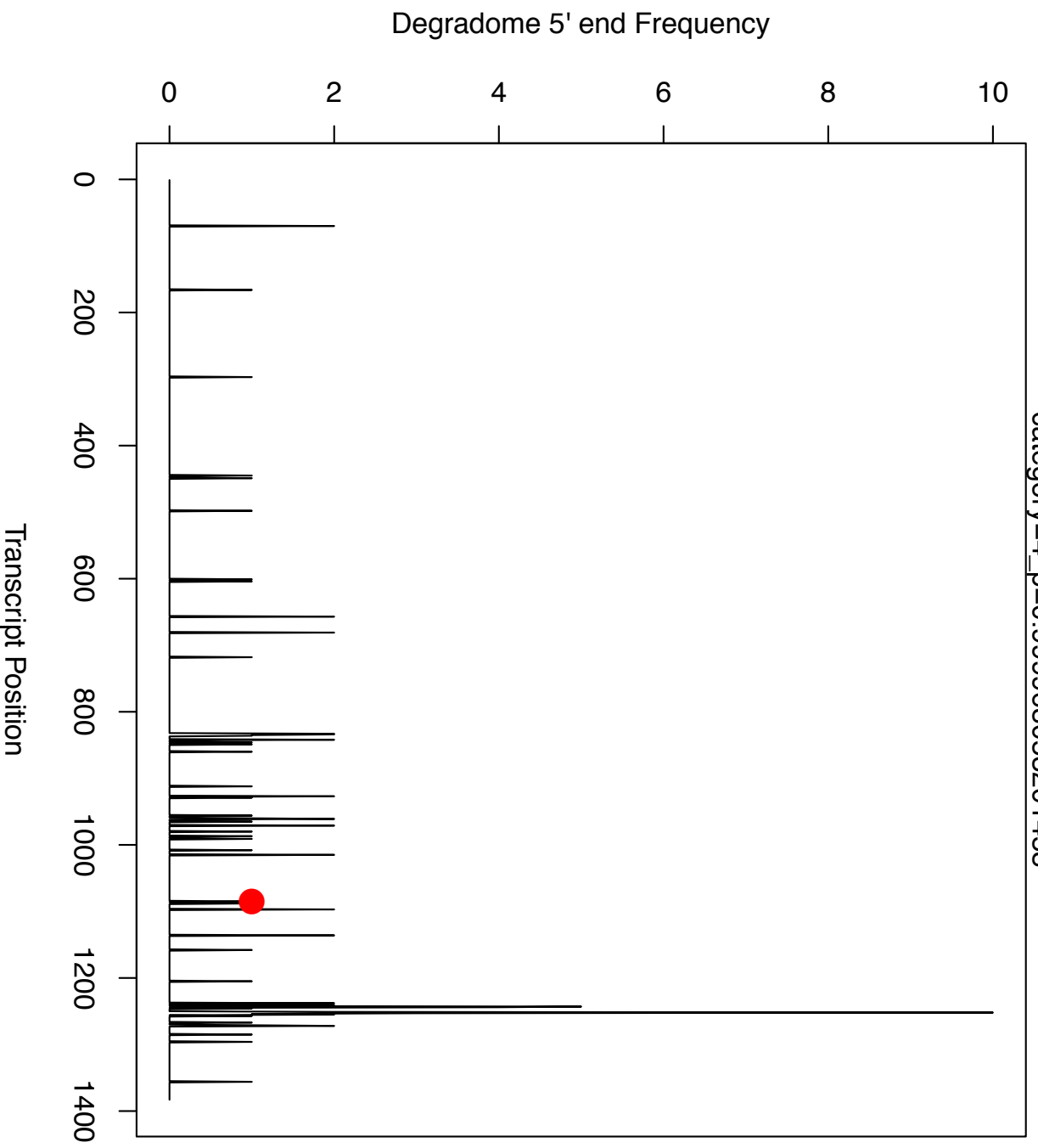
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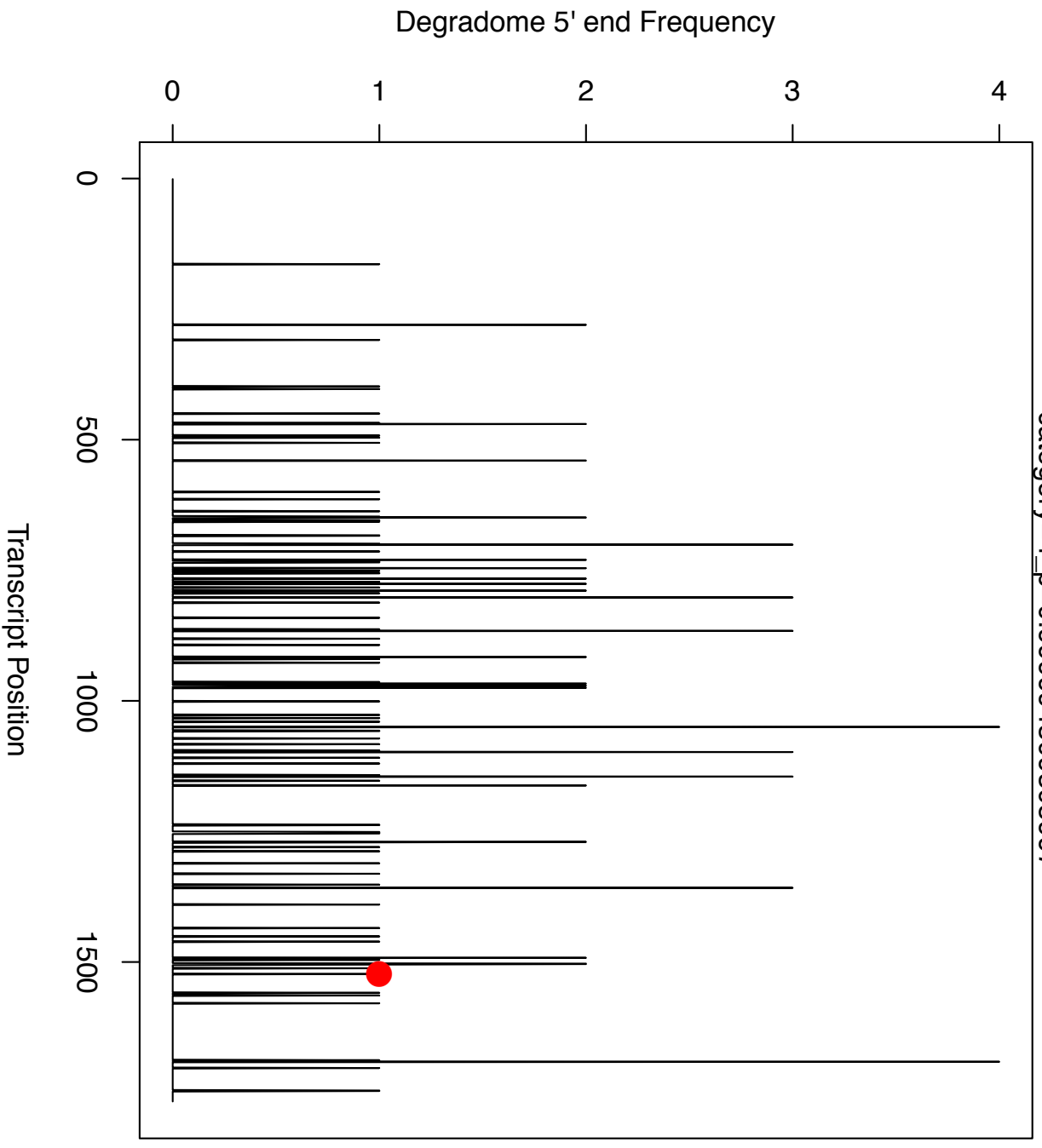
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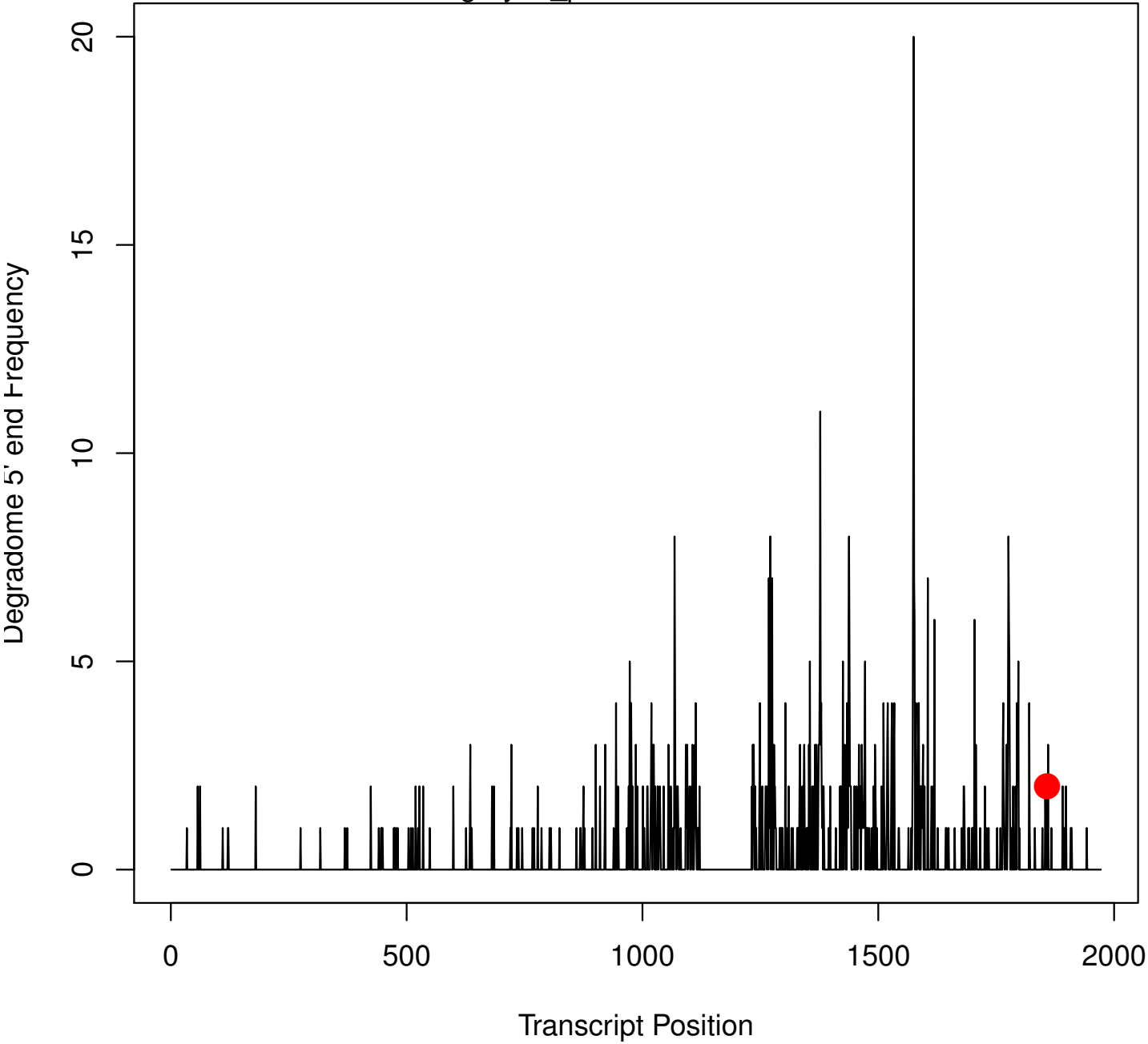
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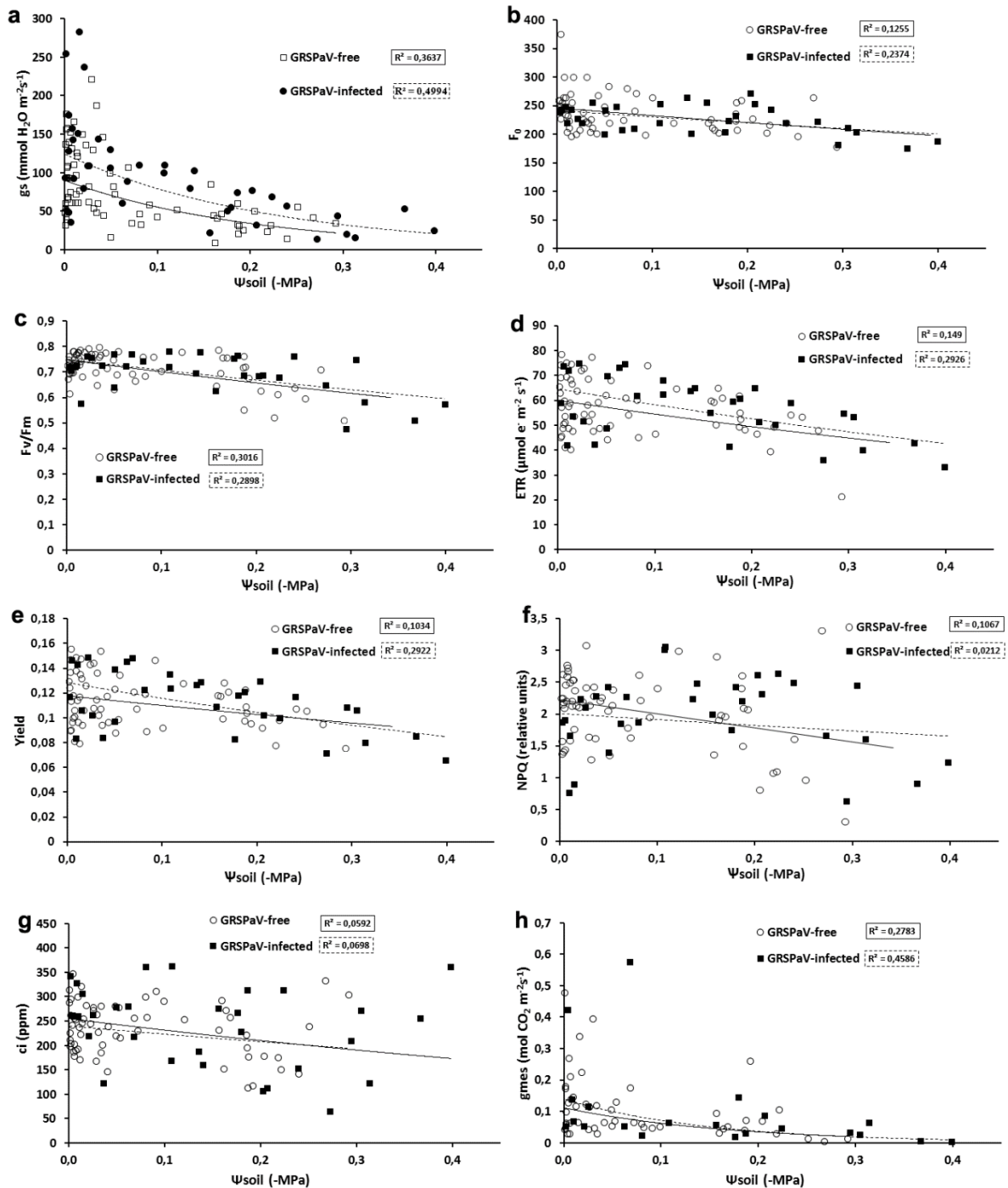
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category=3_p=0.671678599434573





Supplementary Figure S10. Relationship between soil water potential (Ψ_{soil}) and: a) estimated stomatal conductance (g_s), b) maximum fluorescence in dark adapted leaves (F_0), c) maximum quantum efficiency of PSII (F_v/F_m), d) electron transport rate (ETR), e) quantum efficiency of PSII (Yield), f) non-photochemical quenching (NPQ), g) substomatal internal carbon concentration (c_i) and h) mesophyll conductance to CO₂ (g_{mes}) in *Grapevine rupestris stem pitting-associated* (GRSPaV)-infected (filled squares, dashed trend line) and GRSPaV-free (open circles, continuous trend line) plants.