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# A new genus for the eastern dwarf galagos (Primates: Galagidae)

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|----------------|----|---|
| 3<br>4         | 1  | ABSTRACT  |
| 5<br>6         | 2  | The family Galagidae (African galagos or bushbabies) comprises five genera: Euoticus Gray     |
| 7<br>8         | 3  | 1872; Galago Geoffroy Saint-Hilaire, 1796; Galagoides Smith, 1833; Otolemur Coquerel,         |
| 9<br>10        | 4  | 1859; and Sciurocheirus Gray, 1872, none of which is regarded as monotypic, but some          |
| 11<br>12       | 5  | (Euoticus, Otolemur) certainly qualify as oligotypic. We argue for the recognition of a sixth |
| 13<br>14       | 6  | genus, if the taxonomy is to reflect galagid evolution accurately. Genetic evidence has       |
| 15<br>16       | 7  | consistently demonstrated that the taxa currently referred to the genus Galagoides are not    |
| 17<br>18<br>19 | 8  | monophyletic, but form two clades (a western and an eastern clade) that do not share an       |
| 20<br>21       | 9  | exclusive common ancestor; we review 20 years of genetic studies that corroborate this        |
| 22<br>23       | 10 | conclusion. Further, we compare vocalisations emitted by small-bodied galagids with           |
| 24<br>25       | 11 | proposed phylogenetic relationships, and demonstrate congruence between these data sets.      |
| 26<br>27       | 12 | Morphological evidence, however, is not entirely congruent with genetic reconstructions;      |
| 28<br>29<br>30 | 13 | parallel dwarfing in the two clades has led to convergences in skull size and shape that have |
| 31<br>32       | 14 | complicated the classification of the smaller species. We present a craniodental morphometric |
| 33<br>34       | 15 | analysis of small-bodied galagid genera that identifies distinguishing characters for the     |
| 35<br>36<br>27 | 16 | genera, and supports our proposal that five taxa currently subsumed under Galagoides (Gs      |
| 37<br>38<br>39 | 17 | cocos, Gs granti, Gs orinus, Gs rondoensis and Gs zanzibaricus) be placed in their own        |
| 40<br>41<br>42 | 18 | genus, for which we propose the name Paragalago.  |
| 43<br>44       | 19 | ADDITIONAL KEYWORDS: Bushbaby – Biogeography – Craniodental morphometrics –                   |
| 45<br>46<br>47 | 20 | Galagoides – Paragalago – Molecular phylogeny – Vocalisations                                 |
| 48             |    |   |

| 22 | INTRODUCTION   |
|----|--|
| 23 | In A Field Guide to the Larger Mammals of Africa (Dorst & Dandelot, 1970) Pierre Dandelot          |
| 24 | illustrated five species of galagos (or bushbabies) and two species of pottos. Four of the         |
| 25 | galagid taxa were subsumed under the genus Galago: i.e. G. alleni, G. crassicaudatus, G.           |
| 26 | demidovi (sic, now referred to as G. demidoff in accordance with its initial description:          |
| 27 | Jenkins, 1987; Groves, 2001) and G. senegalensis. The needle-clawed galagos were classified        |
| 28 | in their own genus, <i>Euoticus</i> , in accordance with the taxonomies of Schwarz (1931), Simpson |
| 29 | (1945) and Hill (1953). Forty-five years later, all five taxa are regarded as distinct genera,     |
| 30 | none of which is generally viewed as monotypic, although much of the detailed research has         |
| 31 | yet to be conducted. In this contribution we discuss evidence that Galagoides, as construed        |
| 32 | by Olson (1979) on morphological grounds, does not constitute a clade in molecular                 |
| 33 | phylogenetic reconstructions (DelPero et al., 2000; Masters et al., 2007; Fabre, Rodrigues &       |
| 34 | Douzery, 2009; Springer et al., 2012; Pozzi, Disotell & Masters, 2014; Pozzi et al., 2015;         |
| 35 | Pozzi, 2016) and its members are unlikely to have shared an exclusive common ancestor.             |
| 36 | More specifically, the dwarf galagos confined to the forests of East and southern Africa           |
| 37 | require the designation of a new genus. In the subsequent text we abbreviate Galagoides as         |
| 38 | <i>Gs</i> to distinguish it from the abbreviation of <i>Galago</i> ( <i>G</i> .).                  |
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| 40 | HISTORY OF GALAGID GENERA  |
| 41 | The first galagid genus to enter the scientific literature was Galago, described by Etienne        |
| 42 | Geoffroy Saint-Hilaire (1796), and was based on a lesser galago specimen collected in              |
| 43 | Senegal, West Africa. The name was taken from the Senegalese vernacular name for these             |
| 44 | animals. The genus Galagoides was proposed by Sir Andrew Smith (1833) to distinguish the           |

dwarf (G. demidoff) and lesser (G. senegalensis) galagos from what Smith considered to be

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| 46 | the "true Galagos" among which he included species now referred to the genera Microcebus              |
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| 47 | and Otolemur. Galagoides is now restricted to the dwarf galago taxa. Otolemur was                     |
| 48 | introduced as the generic epithet to denote the greater galagos by Charles Coquerel (1859)            |
| 49 | with Otolemur agisymbanus (= garnettii) from Zanzibar as the type species. The genus                  |
| 50 | Euoticus, which distinguishes the needle-clawed species, was introduced initially by John             |
| 51 | Gray (1863) as a subgenus under the genus Otogale which also included the greater galagos.            |
| 52 | A few years later he elevated <i>Euoticus</i> to the status of a full genus (Gray, 1872). In the same |
| 53 | publication, Gray (1872) proposed Sciurocheirus as the generic designation of a squirrel              |
| 54 | galago specimen (S. alleni, sensu lato) deriving from Fernando Po [Bioko Island].                     |
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| 55 | The genus <i>Galagoides</i> was redefined on morphological grounds by Olson (1979) and                |
| 56 | employed subsequently by many authors (Honess & Bearder, 1996; Kingdon, 1997;                         |
| 57 | Wickings, Ambrose & Bearder, 1998; DelPero et al., 2000; Masters & Bragg, 2000; Masters               |
| 58 | & Brothers, 2002; Butynski et al., 2006). It comprised the true dwarf galagos (Gs demidoff,           |
| 59 | Gs thomasi), the 'Zanzibar' galagos (Gs cocos, Gs granti, Gs zanzibaricus), and the squirrel          |
| 60 | galagos (Gs alleni, s.l.). Honess & Bearder (1996) and Kingdon (1997) recognised three new            |
| 61 | dwarf galago species just before the turn of the twenty-first century: Gs orinus, originally          |
| 62 | described by Lawrence and Washburn (1936) as a subspecies of Gs demidoff from the                     |
| 63 | Uluguru Mountains, Gs rondoensis from the Rondo plateau, and Gs udzungwensis from the                 |
| 64 | Udzungwa Mountains. All three localities are in Tanzania. Gs udzungwensis has since been              |
| 65 | downgraded to a subspecies of Gs zanzibaricus confined to the Tanzanian mainland, while               |
| 66 | Gs z. zanzibaricusis reserved for the form on Zanzibar Island. Gs orinus and Gs rondoensis            |
| 67 | are now recognised as valid species.  |
| 68 | The morphological characters uniting Olson's (1979) genus <i>Galagoides</i> were not                  |

The morphological characters uniting Olson's (1979) genus *Galagoides* were not
made explicit, but from our own observations (Groves, 2001; Masters & Couette, 2015) they
include moderate basicranial flexion (i.e. stronger than in most lemuriforms and lorisids, but

not as strong as in the genera *Galago* or *Euoticus*), anterior upper premolar ( $P^2$ ) not caniniform (sometimes with a hypocone), third upper molar  $(M^3)$  > posterior upper premolar  $(P^4)$ , and premaxillaries extended into a tube that projects way beyond the lower jaw. The premaxillary tube is longest in dwarf galagos, moderate in 'Zanzibar' galagos, and least pronounced in squirrel galagos. *Euoticus*, *Galago* and *Otolemur* have no premaxillary tube, and the anterior margin of the snout is square; in the absence of a tube, the square-snouted galagos have a relictual nub on the midline beneath the nasal aperture, suggesting that extended premaxillaries may have been the ancestral condition. Among the lorisids, the two small-bodied genera, Arctocebus and Loris, both have premaxillary tubes. Galagoides is further characterized by small body size (smaller in the western clade than in the squirrel galagos or most taxa of the eastern clade), a concave nasal profile, and dark circumocular rings that range in colour from dark brown to black, separated by a grey to white nose stripe. While the deep russet colouration of squirrel galagos makes them instantly recognisable, a survey of other museum specimens designated as *Galagoides* revealed variable pelage colouration, both within and between populations. In most cases, the animals were covered in dense, soft hairs with dark-grey roots, but brown to bright russet tips on the head, dorsum and outer surfaces of the limbs. The tips of the hairs on the under surfaces are yellow-buff to white, and the animals have brown to blackish tails.

# 90 OVERVIEW OF MOLECULAR GENETIC EVIDENCE FOR RELATIONSHIPS AMONG 91 GALAGID LINEAGES

93 Genetic studies – from their earliest days – have consistently indicated that *Galagoides* is

94 polyphyletic, implying that the grouping based on morphological similarity is probably based

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| 95  | on plesiomorphic or homoplastic characters. The first such evidence came from allozymes                   |
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| 96  | (Masters et al., 1994), highly repeated DNA sequences (Crovella et al., 1994) and 12S                     |
| 97  | ribosomal mitochondrial DNA sequences (rDNA; Bayes, 1998). All of these studies                           |
| 98  | reconstructed the taxon called Galagoides alleni (s.l.) as the sister taxon to Otolemur spp.,             |
| 99  | although morphological synapomorphies for this group remain elusive. The only shared                      |
| 100 | character Masters and Brothers (2002) identified from their dataset was large, square,                    |
| 101 | bunodont molars, indicative of a predominantly frugivorous diet and potentially homoplastic.              |
| 102 | The first Zanzibar galago sequences were published by Del Pero et al. (2000). The specimen                |
| 103 | sampled was probably Gs granti, as it had been collected in northern Mozambique, but was                  |
| 104 | classified as Gs zanzibaricus on the basis of craniodental morphometrics (Masters & Bragg,                |
| 105 | 2000). Using partial sequences of three mitochondrial genes (12S and 16S rDNA and                         |
| 106 | cytochrome b), DelPero and colleagues reconstructed Gs demidoff and so-called Gs                          |
| 107 | zanzibaricus as independent lineages that showed higher levels of genetic divergence from                 |
| 108 | one another than either lineage showed from any other galagid taxon in their sample of eight              |
| 109 | taxa. This result, coupled with the alliance of Gs alleni with Otolemur, led DelPero et al.               |
| 110 | (2000) to describe the genus Galagoides as a "wastebasket taxon of plesiomorphic species".                |
| 111 | This contention has been supported by more recent and more comprehensive studies. The                     |
| 112 | squirrel galagos continue to be recovered as the sister to the Otolemur clade, and Gray's                 |
| 113 | (1872) genus, Sciurocheirus, has been resuscitated (Grubb et al., 2003; Masters et al., 2007).            |
| 114 | Despite the paucity of morphological synapomorphies for this grouping, it derives support                 |
| 115 | from the sparse fossil record. Wesselman (1984) described a fossil hypodigm from                          |
| 116 | approximately 3 Myr sediments in Ethiopia that comprises a fragmentary maxilla, an isolated               |
| 117 | M <sub>2</sub> , and an edentulous mandible. On the basis of its bunodont teeth and its intermediate size |
| 118 | between Otolemur and Sciurocheirus, he interpreted the taxon (now termed O. howelli;                      |
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Harrison, 2010) as a member of the *Sciurocheirus/Otolemur* clade, with its closest affinities
to *Otolemur*.

Following the removal of the squirrel galagos from *Galagoides*, the western and eastern dwarf galagos have continued to be reconstructed as paraphyletic or even polyphyletic in molecular analyses, indicating that the genus still includes two independent clades that did not share an exclusive common ancestor. The western clade comprises the "true" dwarf galagos, Gs demidoff and Gs thomasi, and the eastern clade includes Gs *zanzibaricus* and its allies. Using complete sequences of the cytochrome b gene, Roos et al. (2004) recovered Gs demidoff as the first galagid lineage to diverge, and Gs zanzibaricus as the sister taxon of Galago, a topology supported by Chatterjee et al. (2009) and Fabre et al. (2009). The tree of Masters et al. (2007) also depicted Galagoides as polyphyletic, but did not group Gs zanzibaricus with the genus Galago. More recently, a more comprehensive phylogenetic study of primates supported a sister taxon relationship between the Zanzibar galagos and the Otolemur/Sciurocheirus clade, with Gs demidoff and Gs thomasi again forming an independent clade (Springer *et al.*, 2012).

The disagreement among these studies regarding the phylogenetic placement of the eastern and western lineages may be related to incomplete lineage sorting (or the failure of two or more lineages in a population to coalesce, leading one of the lineages to coalesce first with a lineage from a less closely related population), or possibly past introgression events, as they were all based solely on mitochondrial sequences. To address this problem, Pozzi et al. (2014) assembled a molecular dataset including 27 independent nuclear loci and inferred phylogenetic relationships also using coalescent-based species tree methods to account for incomplete lineage sorting. Their results strongly confirmed the polyphyletic status of *Galagoides*, as well as a sister-taxon relationship between the eastern clade and the lesser galagos (Galago spp.). The largest molecular dataset compiled for galagids to date, combining

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| 144 | 53 nuclear loci and three mitochondrial genes, confirmed these results (Pozzi, 2016). Figure 1     |
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| 145 | represents our current understanding of relationships among the lorisoid primates based on         |
| 146 | both nuclear and mitochondrial sequence data, derived from the studies of Pozzi et al, (2014,      |
| 147 | 2015) and Pozzi (2016).  |
| 148 | [FIGURE 1 ABOUT HERE]  |
| 149 | Despite these large nuclear datasets, the sister-group relationships of two eastern                |
| 150 | dwarf galagos, Galagoides rondoensis and Gs orinus, remain unresolved because of limited           |
| 151 | representative specimens and genetic data; a handful of specimens is distributed across            |
| 152 | museum collections in North America and Europe. A molecular study based on complete                |
| 153 | mitochondrial cytochrome $b$ sequences strongly supported an affinity between these species        |
| 154 | and the Zanzibar galagos to the exclusion of the western Galagoides clade (Pozzi et al.,           |
| 155 | 2015), leading us to include them in the proposed new genus, which hence comprises five            |
| 156 | species distributed in forests east of the African rift and distinct from the true dwarf galagos,  |
| 157 | Gs demidoff and Gs thomasi, in the west (Figure 2).  |
| 158 | [FIGURE 2 ABOUT HERE]  |
| 159 | OVERVIEW OF EVIDENCE FROM VOCAL REPERTOIRES  |
| 160 | Vocalisations are particularly important indicators of galagid diversity because, as nocturnal     |
| 161 | animals, galagos do not rely on morphologically-encoded visual signals for the location and        |
| 162 | attraction of conspecific mates. Many galagid species and species groups have been identified      |
| 163 | on the basis of differences in loud calls (or advertisement calls), which has led to their being   |
| 164 | grouped according to call structures: crescendo callers, scaling callers, rolling callers,         |
| 165 | incremental callers and repetitive callers (Bearder et al., 1996; Kingdon, 1997; Butynski et       |
| 166 | al., 2013). Like all characters that are crucial to specific-mate recognition, specific loud calls |
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| 167 | are qualitatively different between closely related species, and the rapidity of their evolution     |
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| 168 | may obscure their phylogenetic signal at older levels of divergence (Masters, 2007). For             |
| 169 | instance, rolling and crescendo calls are polyphyletic when compared with species                    |
| 170 | relationships determined by molecular sequence data, indicating a degree of homoplasy:               |
| 171 | Galagoides granti and Gs cocos are both described as crescendo callers (Bearder et al., 1996;        |
| 172 | Kingdon, 1997; Butynski et al., 2013), whereas Gs zanzibaricus, which is reconstructed as            |
| 173 | the sister taxon to Gs cocos (see Figure 1), is a rolling caller, just like Gs rondoensis. In        |
| 174 | contrast to advertisement calls, anti-predatory calls tend to be highly conserved                    |
| 175 | phylogenetically, making them more useful as grouping criteria at deeper phylogenetic levels.        |
| 176 | Vocal homologies among the small-bodied galagos (i.e. excluding Sciurocheirus and                    |
| 177 | Otolemur spp.) can be identified in at least three call types: two anti-predatory calls (mobbing     |
| 178 | yaps and buzzy alarms; Génin et al., 2016) and the loud socio-territorial calls. These vocal         |
| 179 | homologies are congruent with phylogenetic relationships among these lineages indicated by           |
| 180 | molecular analyses, and further justify the creation of a new genus for the eastern dwarf            |
| 181 | galagos.   |
| 182 | Mobbing yaps. The mobbing yap is emitted by all small-bodied galagos under similar                   |
| 183 | contexts, and is often recorded while an animal is circling around to face the observer (FG,         |
| 184 | personal observations). It appears to be homologous to the loud call of <i>Euoticus</i> spp. (Figure |
| 185 | 3). It is a high frequency atonal call repeated at frequent intervals, often after the emission of   |
| 186 | a few buzzy alarms. The wide frequency range covered by the call that makes it sound atonal          |
| 187 | to human ears is due to very rapid modulation that is likely to be perceived by the animals.         |
| 188 | [FIGURE 3 ABOUT HERE]  |
| 189 |  |
| 190 | Buzzy alarms. Buzzy alarms are homologous in Galagoides, Galago and the eastern dwarf                |
| 191 | galagos, but have very different structures in the three groups (Figure 3). Buzzy alarms are         |
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often the first indicator of the presence of an animal that has not yet been detected visually
(FG, personal observations). Animals emit several buzzy alarms that may precede or alternate
with yaps. The call is bi-syllabic, consisting of an initial high frequency unit followed by a
highly modulated, lower frequency screech. In western *Galagoides* spp. the buzz is extremely
brief. In *Galago* spp. it is a brief, noisy, low frequency cough-like call (cho-ha). In the eastern
dwarf galago species, the buzz is a long screech peculiar to the group.

Loud calls. Loud calls are far more variable between taxa than anti-predatory calls. Each of the three small-bodied genera emits a different kind of loud call associated with different contexts, indicating that the various calls evolved different functions associated with different habitats and socio-territorial systems. On the basis of our own observations as well as those of other authors (Bearder et al., 1996; Kingdon, 1997; Butynski et al., 2013), western Galagoides spp. are crescendo callers; the crescendo either consists of a single trill (Gs *demidoff*) or a short sequence of trills (*Gs thomasi*), starting with an increase in pitch and amplitude (overtone crescendo) followed by repeated, insect-like, high frequency clicks (Figure 3). The call is typically used as a gathering call emitted when animals leave or return to their nests. Lesser galagos (*Galago* spp.) are repetitive callers. They have low frequency metronomic and tonal calls which are emitted throughout the night, indicating a territorial function. Homology between the loud calls of eastern dwarf galago species and Galagoides crescendo calls is difficult to establish, but such homology with Galago repetitive calls is clear, as they share a basic temporal structure of repeated units forming syllables.

The loud calls emitted by eastern dwarf galagos are so variable that they are difficult to characterise. The group could be called the "varied callers" or "modulated callers", as their loud calls consist of repeated, highly modulated units emitted at higher frequency than *Galago* repetitive calls. Their function is also less clear, as they are given when animals leave

| 217 | or return to their sleeping sites, as well as throughout the night when animals interact. The            |
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| 218 | calls could hence be categorised as long distance contact calls adapted to habitats that are             |
| 219 | generally drier than those of western Galagoides, but wetter and more closed than Galago                 |
| 220 | habitats (Génin et al., 2016). The specific diversity of this group still requires investigation,        |
| 221 | as only three call structures (scaling, rolling and incremental) have been described for at least        |
| 222 | five species.  |
| 223 |  |
| 224 | MORPHOLOGICAL DIFFERENTIATION: NEW ANALYSES  |
| 225 | Morphological characterization of the eastern dwarf galagos has been complicated not only                |
| 226 | by the scarcity of exemplars of some species, but also by their strong convergence with                  |
| 227 | members of the western clade. In a canonical variate morphometric analysis, the skulls of the            |
| 228 | type specimens of Gs orinus and Gs rondoensis clustered with the western clade,                          |
| 229 | contradicting their genetic affinity to Gs granti and Gs zanzibaricus (Masters & Couette,                |
| 230 | 2015). In an attempt to resolve this contradiction, we searched through museum collections in            |
| 231 | the USA and Europe, and identified seven probable Gs rondoensis specimens in addition to                 |
| 232 | the type specimen held in the Natural History Museum, London. Their identification was                   |
| 233 | based on three factors: the consistent presence of a square M <sup>3</sup> with a very small hypocone (a |
| 234 | very rare occurrence in other eastern dwarf galagos); collection locality (east of the Rift); and        |
| 235 | a disjunction between the completion of the eruption of the permanent dentition and skull                |
| 236 | maturation. In most galagid genera the attainment of adult body size occurs shortly after the            |
| 237 | complete eruption of the adult dentition. In the putative Gs rondoensis specimens we                     |
| 238 | identified, animals with adult (and often worn) dentition had unfused cranial sutures, and are           |
| 239 | likely to have continued to grow had their lives not been prematurely ended. Groves (2001)               |
| 240 | based his assessment of Gs rondoensis as the smallest living galagid on the type specimen                |
| 241 | which had a body weight of 60 g, but animals trapped in the field may be $20 - 25$ g heavier             |
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(Andrew Perkin, personal communication). The type specimen has its permanent dentition, but its morphology is juvenile, and fully grown members of this species are likely to be larger than G. orinus adults. 

Materials and methods used in the new morphometric analyses. In order to investigate the morphological distinctiveness of the eastern dwarf galagos from other small-bodied galagids, a set of 12 linear craniodental measurements was taken from 610 galago specimens including western dwarf galagos (Galagoides demidoff and Gs thomasi, n = 322; see Masters & Couette, 2015 for specimen list), lesser galagos (*Galago moholi*, n = 150) and squirrel galagos (*Sciurocheirus alleni*, s.l., n = 58), plus specimens formerly identified as *Galagoides* cocos, Gs granti, Gs orinus, Gs rondoensis, Gs udzungwensis and Gs zanzibaricus (n = 80). Accession details of these specimens are listed in Table 1 of the supplementary data, and the institutions in which mensural data were collected are listed in the Acknowledgements. We followed the methodology of Masters & Couette (2015): measurements (Figure 4, Table 1) were recorded using digital callipers, and the sample was composed only of specimens with [FIGURE 4 ABOUT HERE] fully erupted dentition.

Raw data were size-adjusted using the Burnaby (1966) procedure which consists of extracting an isometric vector from the multivariate dataset and back-projecting the values in a multivarate subspace orthogonal to this vector (Klingenberg, 1996). The geometric mean (GM) was computed using the isometric vector and served as a proxy for size. Thus size (GM) and shape (size corrected variables = shape variables) are considered independently through ANOVA (size) or MANOVA, PCA and CVA (log transformed values of shape). We performed a Between Group Principle Component Analysis (BGPCA, Mitteroecker &

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| 266  | Bookstein, 2011), which is a classic PCA based on the mean values for each group with no  |
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| 267  | regard for intragroup variation. Specimens are then back-plotted in the morphospace by  |
| 268  | multiplying the morphological data matrix (log transformed values of shape) by the  |
| 269  | coefficient of the BGPCA; the PCs are computed only on the intergroup variation rather than   |
| 270  | on a mix of intra- and intergroup variation. All statistics were performed with R 3.0.2.  |
| 271  | software (R Core Team, 2013) and the packages "candisc" (Friendly &Fox, 2015), "car" (Fox   |
| 272  | & Weisberg, 2011), "geomorph" (Adams & Otarola-Castillo, 2013) and "smatr" (Warton et   |
| 273  | <i>al.</i> , 2012).   |
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| 274  | Results of the morphometric analyses. MANOVA results attested to significant differences in   |
| 275  | cranial morphology among genera (Pillai Trace = 1.84, F = 86.09, df = 33, $p < 0.001$ ). In the   |
| 276  | BGPCA, three PCs summed up the entire variation in our sample: PC1, PC2 and PC3   |
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| 277  | accounted for 88.9%, 9.91% and 1.19%, respectively. The genera Galago, Galagoides and   |
| 277<br>278   | accounted for 88.9%, 9.91% and 1.19%, respectively. The genera <i>Galago</i> , <i>Galagoides</i> and <i>Sciurocheirus</i> were well separated in the PC1 x PC2 morphospace, with no overlap except for  |
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| 278<br>279   | <i>Sciurocheirus</i> were well separated in the PC1 x PC2 morphospace, with no overlap except for some outlier specimens (Figure 5). The presence of outliers and the degree of dispersion  |
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| 278<br>279<br>280<br>281<br>282<br>283<br>284<br>285<br>286<br>287 | <i>Sciurocheirus</i> were well separated in the PC1 x PC2 morphospace, with no overlap except for some outlier specimens (Figure 5). The presence of outliers and the degree of dispersion evident in Figure 5 may mean that some subadult skulls were included, along with their correlated allometric shape differences. The cranial morphology of <i>Galagoides</i> individuals was clearly different from that of the <i>Galago</i> specimens, and the differences constituted the major variation along PC1. On this axis all of the variables had similar loadings (values between -0.24 and 0.07) except for premaxillary length, for which the loading was very high (0.91); hence the greater part of variation along PC1 was due to differences in the length of the tip of the muzzle. PC2 separated the genus <i>Sciurocheirus</i> (positive values) and the genera <i>Galago</i> and <i>Galagoides</i> (negative values). The variation along PC2 was mainly structured by |

and Galagoides on PC1 and between Sciurocheirus and the group composed of Galago and *Galagoides* on PC2. Although the three groups were clearly distinguished on PC1, they overlapped on PC2. The putative Gs rondoensis specimens were scattered in the space between the eastern dwarf species and *Galagoides*, and their variation in body size was evident (Figure 5); despite our best efforts, it is possible our sample included representatives of more than one species. The Gs orinus specimens formed part of the main eastern dwarf cluster, although they overlapped with some Gs rondoensis specimens. [FIGURE 5 ABOUT HERE] Canonical Discriminant Analysis defined three significant canonical axes, accounting for 70.98%, 27.06% and 1.96% of interclass variation (Figure 6). The variable Premaxilla contributed the main discrimination along the first axis. The four genera were well

302 individualised on the first two axes, with high percentages of correct classification: 100% for

*Galago*, 98.14% for *Galagoides*, 91.3% for the eastern dwarf galagos and 86% for

*Sciurocheirus*. As is evident in visual comparisons of skulls, the elongation of the premaxilla

305 is greatest in *Galagoides*, smaller in the eastern dwarf galagos and smallest in *Galago* 

306 (TukeyHSD post hoc test p values<0.01 among these genera), but the difference in

307 premaxillary length between the eastern dwarf galagos and *Sciurocheirus* was not significant.

308 The variables with highest loading on the second axis were Total Skull Length, Snout Length

309 and Cheek Tooth Width. The eastern dwarf galagos differed significantly in Snout Length

310 from *Galago* and *Sciurocheirus*, but not from *Galagoides*. All of the genera differed

311 significantly in Cheek Tooth Width, with values increasing from *Galago* through *Galagoides* 

312 to the eastern dwarfs and finally *Sciurocheirus*, the large bunodont molars of which evince its

313 affinity to *Otolemur* spp. A similar trend is noticeable for Total Skull Length measurements,

314 with the smallest values in *Galagoides*, increasing in *Galago* and the eastern dwarfs, and with

*Sciurocheirus* having the longest skulls. As in the BGPCA analysis, the *Gs rondoensis* 

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specimens occupied the morphospace between the eastern dwarf galagos and *Galagoides*. while Gs orinus was more closely grouped with the eastern dwarf species. Specimens of Gs rondoensis and Gs orinus show intermediate morphology and overlap with eastern dwarf galagos and specimens of the genus Galagoides. [FIGURE 6 ABOUT HERE] An ANOVA of skull size repeats the pattern shown by Total Skull Length (p value < 0.001 and all Tukey HSD multiple comparison of means p values below 0.001). In order to test the relationship of size (GM) and shape (logged size corrected variable), we performed a multivariate regression considering the effect of size on shape, genus and the interaction between size and genus. We used a Type II ANOVA to test each term of the linear model. Size, genus and the interaction had a significant effect on shape with p values below 0.001, attesting that shape variation is explained by size variation (allometry). The common allometry, that is the proportion of shape explained by size across the entire sample, was 77.7%, but considering the allometric vectors for each genus yielded much lower values: size accounted for 15.1% of shape variation in Galago, 20% in Galagoides, 13.8% in the eastern dwarfs and 8.7% in Sciurocheirus. Pairwise comparison of multivariate allometric patterns demonstrated significant differences between the eastern dwarfs and Galago (p value of angle between allometric vectors = 0.0428), Galagoides (p = 0.019) and Sciurocheirus (p = 0.014) (Figure 7). With respect to univariate differences in allometries among genera, *Sciurocheirus* presented a very different pattern from all other genera: the allometric slopes of the variables SL, CH, FL, CTW, PW, MW, TC, and P were all significantly different. The eastern dwarf galagos differed in slope from Galago for variables SL and CTW, and from Galagoides for variables IC, CTW, TSL and P.

## [FIGURE 7 ABOUT HERE]

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| 340 | Sciurocheirus is hence clearly differentiated in both size and shape. Comparing the                               |
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| 341 | three smaller-sized genera, our multivariate analyses indicated marked differences in cranial                     |
| 342 | shape among them. From a univariate perspective, the eastern dwarf group differed mainly in                       |
| 343 | Supraoccipital Length (intermediate between Galago and Galagoides), Interorbital                                  |
| 344 | Constriction, Cheek Teeth Width and Total Skull Length (higher values in the eastern dwarf                        |
| 345 | group). These morphometric differences reinforce our proposal to distinguish the eastern                          |
| 346 | dwarf galagos from Galago and Galagoides at the generic level.  |
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| 348 | SYSTEMATICS   |
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| 349 | Paragalago gen. nov.  |
| 350 | TYPE SPECIES: Galago zanzibaricus Matschie, 1893.   |
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| 351 | INCLUDED SPECIES: Galago granti Thomas & Wroughton, 1907; Galago cocos Heller,                                    |
| 352 | 1912; Galago demidovii orinus Lawrence & Washburn, 1936; Galagoides udzungwensis                                  |
| 353 | Honess, 1996; Galagoides rondoensis Honess, 1996.   |
| 354 | DIAGNOSIS: Medium to small sized galagos (60 – 250 g), overlapping in size with Galago                            |
| 355 | spp. and notably smaller than Sciurocheirus, Euoticus and Otolemur spp. Two species                               |
| 356 | (Paragalago orinus and P. rondoensis) show convergence in shape and size with Galagoides                          |
| 357 | spp. Cranium ovoid in shape, narrowing posteriorly so that the pneumatised mastoids                               |
| 358 | protrude. Premaxillaries protracted into a short tube that extends beyond lower jaw, as in                        |
| 359 | Galagoides and Sciurocheirus; the tubular extension in Paragalago is intermediate in length                       |
| 360 | between the premaxillary tubes of the smaller and larger taxa. Anterior upper premolar $(P^2)$                    |
| 361 | double-rooted, slender but distally trenchant, as in Galagoides, not caniniform as in Euoticus                    |
| 362 | and some <i>Galago</i> spp. Upper posterior premolars (P <sup>4</sup> s) are slightly larger than upper posterior |
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equivalent in size to  $P^4$ , while in *Galago*  $M^3$  is much reduced. In *Paragalago* spp. the  $M^3$ 

365 hypocone is absent or minute, but commonly observed in *Galagoides* spp. Coronoid

366 processes delicate and curved, extending almost as far back as glenoid process, as in

*Sciurocheirus* and *Galagoides*; not flattened and foreshortened, as in *Galago* and *Euoticus*.

#### 368 Distribution east of the Great African Rift.

DESCRIPTION: The snout is longer than in *Galago*, chiefly because premaxillaries extend well beyond the lower jaw, although not to the degree seen in *Galagoides*. The facial profile is distinctly concave (Schwarz, 1931) as in *Galagoides* and *Sciurocheirus*, not straight or slightly convex as in Galago. Canines are slender. Anterior palatal foramina intrude between medial upper incisors, as in most galagid taxa. P4s fully molarised as in all crown galagids. In most *Paragalago* specimens examined, the M<sup>3</sup> had no hypocone, although a minute hypocone was present in some specimens from the Rondo Forest (probably *P. rondoensis*). The degree of basic anial flexion is moderate, as seen in *Galagoides*, *Otolemur* and Sciurocheirus, not markedly flexed as in Euoticus and in Galago. Cranial shape is oblong as in Galagoides and Sciurocheirus, not globular as in Galago and Euoticus. Postorbital bars are generally slender, lacking the flanges sometimes seen in *Euoticus*, *Galago* and even Galagoides, usually in older specimens. Lower anterior premolars  $(P_2)$  are partially procumbent, but not to the same degree as the tooth-comb, and never erect, as usually seen in male Galagoides (Masters & Couette, 2015). Parietal muscle scars/crests on either side of the medial suture outline a broad parietal plate over the orbits that narrows posteriorly. The colour of the dorsal pelage is drab-brown to cinnamon with varying degrees of 

384 The colour of the dorsal pelage is drab-brown to cinnamon with varying degrees of 385 rufous wash; outer surfaces of limbs similar to dorsum in colouration. Individual hairs are 386 slate grey near the root, contributing to the overall dark colouration. Hairs on ventrum and

inner surfaces of limbs also with grey roots but cream-buff to yellow-buff tips, and the throat may be yellowish (Groves, 2001). The ears are dark brown to black, depending on the species, and the tail varies from rufous brown to chocolate or even black. Paragalago granti and *P. orinus* have a darker tail tip. The cream to white nose stripe is emphasised by dark brown to black eve rings. Mature males of all species have unidentate penile spines (Perkin, 2007). Species of *Paragalago* show behavioural differences that distinguish them in the field from both *Galagoides* and *Galago* taxa. Eastern dwarf galagos tend to leap more often than Galagoides, but not as frequently or extensively as Galago spp. Moreover, the three genera can be readily distinguished by vocalisations that differ in structure, in context, and probably in function. NOTES: The new genus embraces several taxa originally allied with lesser or dwarf galagos, 

depending on body size. *Paragalago zanzibaricus* was described by Paul Matschie (1893) as a pale cinnamon-coloured lesser galago from western Zanzibar, although the species also occurs on the Tanzanian mainland (see Figure 2). A recent conservation risk assessment conducted by the Primate Specialist Group of the International Union for the Conservation of Nature (IUCN) considered populations on small islands to be particularly vulnerable, and deserving of subspecific recognition for the purpose of conservation monitoring. The Zanzibar population of dwarf galagos was hence designated as the subspecies P. z. zanzibaricus, while the mainland representatives of this species were classified as P. z. *udzungwensis.* Preliminary genetic studies of mainland and island populations (Pozzi, unpublished data) support their conspecific identity, but a more extensive comparison is necessary to confirm this.

*Paragalago cocos*, which is morphologically indistinguishable from *P. zanzibaricus*, was

410 described by Heller (1912) from the Kenyan mainland. *Paragalago granti*, with a type

411 locality in southern Mozambique, has the largest geographical range among representatives

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| 412 | of the genus, extending from the north-east of South Africa throughout Mozambique (and          |
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| 413 | possibly parts of Malawi, where it has been referred to under the rubric Galagoides nyasae      |
| 414 | Elliot, 1907; Grubb et al., 2003) into southern Tanzania. The type and only known skin of       |
| 415 | Galago mertensi Frade, 1924 was collected at a locality not far west of the type locality of P. |
| 416 | granti, and has, rightly or wrongly, been subsumed under this species (Schwarz, 1931).          |
| 417 | The two smallest members of the genus, P. orinus and P. rondoensis, are the most                |
| 418 | recent members of the eastern dwarf clade to have been accorded full species status.            |
| 419 | Paragalago orinus is a montane endemic and occurs within a restricted habitat at high           |
| 420 | altitude in the Udzungwa and Uluguru mountains of Tanzania. Paragalago rondoensis has a         |
| 421 | highly fragmented range in scattered lowland forest patches throughout Tanzania, but is no      |
| 422 | longer considered to be of critical conservation concern (A. Perkin, personal communication).   |
| 423 | The apparent heterochronic disjunction between the eruption of adult dentition and the          |
| 424 | cessation of growth in this species may explain why both it and the genus to which it belongs   |
| 425 | have defied characterisation for so long.   |
| 426 | Our demonstration that the eastern dwarf galagos constitute a genus entirely distinct           |
| 427 | from the western dwarf galagos reinforces the conclusions of Groves (in press) that the         |
| 428 | Eastern Arc Mountains and the Swahilian (Tanzanian/northern Mozambique) coastal forests         |
| 429 | constitute a separate subregion of the African fauna, the Zanj subregion. The Zanj              |
| 430 | mammalian fauna is unique and restricted, and deserves the highest conservation priority.       |
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| 432 | ACKNOWLEDGEMENTS  |
| 433 | Our project was funded primarily by National Research Foundation grants 93924 awarded to        |
| 434 | FG, and 92541 and 90772 awarded to JCM; grant number GB-TAF 4120 awarded by the                 |
| 435 | Synthesys Program to SC; and an ABIC grant awarded to JCM by the Royal Museum for               |
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#### 588 Figure legends

589 Figure 1: Phylogenetic relationships among galagos. The tree represents a summary of our 590 current understanding of relationships among the lorisoid primates based on both nuclear and 591 mitochondrial sequence data, derived from the studies of Pozzi et al. (2014, 2015) and Pozzi (2016). The western dwarf galago clade is identified by a red rectangle, while the eastern 592 593 clade is enclosed within a blue square. 594 Figure 2: Map showing approximate geographic ranges of the two independent dwarf galago 595 clades, Galagoides (red) and the eastern dwarf galagos (blue). The type localities of the 596 species comprising the genera are indicated by symbols. In the case of *Galagoides demidoff*, 597 the type locality is estimated from Fischer's (1806) description. Figure 3: Sonagrams of vocalisations emitted by small-bodied galagid species. Calls of 598

599 *Euoticus* (Cameroon) and *Galagoides granti* (Tshanini, South Africa) were recorded by FG.

600 All other sonograms were downloaded from the East African Primate Diversity and

601 Conservation website (http://www.wildsolutions.nl/vocal-profiles).

Figure 4: Schematic depiction of an eastern dwarf galago skull showing the 12 craniodental
measurements included in the multivariate morphometric analyses. Descriptions of the
variables are presented in Table 1.

Figure 5: Between Group Principle Component Analysis (BGPCA) calculated on the twelve
shape variables. Crosses indicate the mean values of each group that defined the principle
component axes to analyse inter-group variation. Specimen data were back-projected in this
space. Specimens of the two smallest eastern taxa, *orinus* and *rondoensis*, are indicated.

609 Figure 6: Canonical Variate Analysis of the twelve shape variables. Crosses indicate the

610 centroid of each group. The first two roots illustrate significant differences in skull shape

among genera. Specimens of the two smallest eastern taxa, orinus and rondoensis, are indicated.

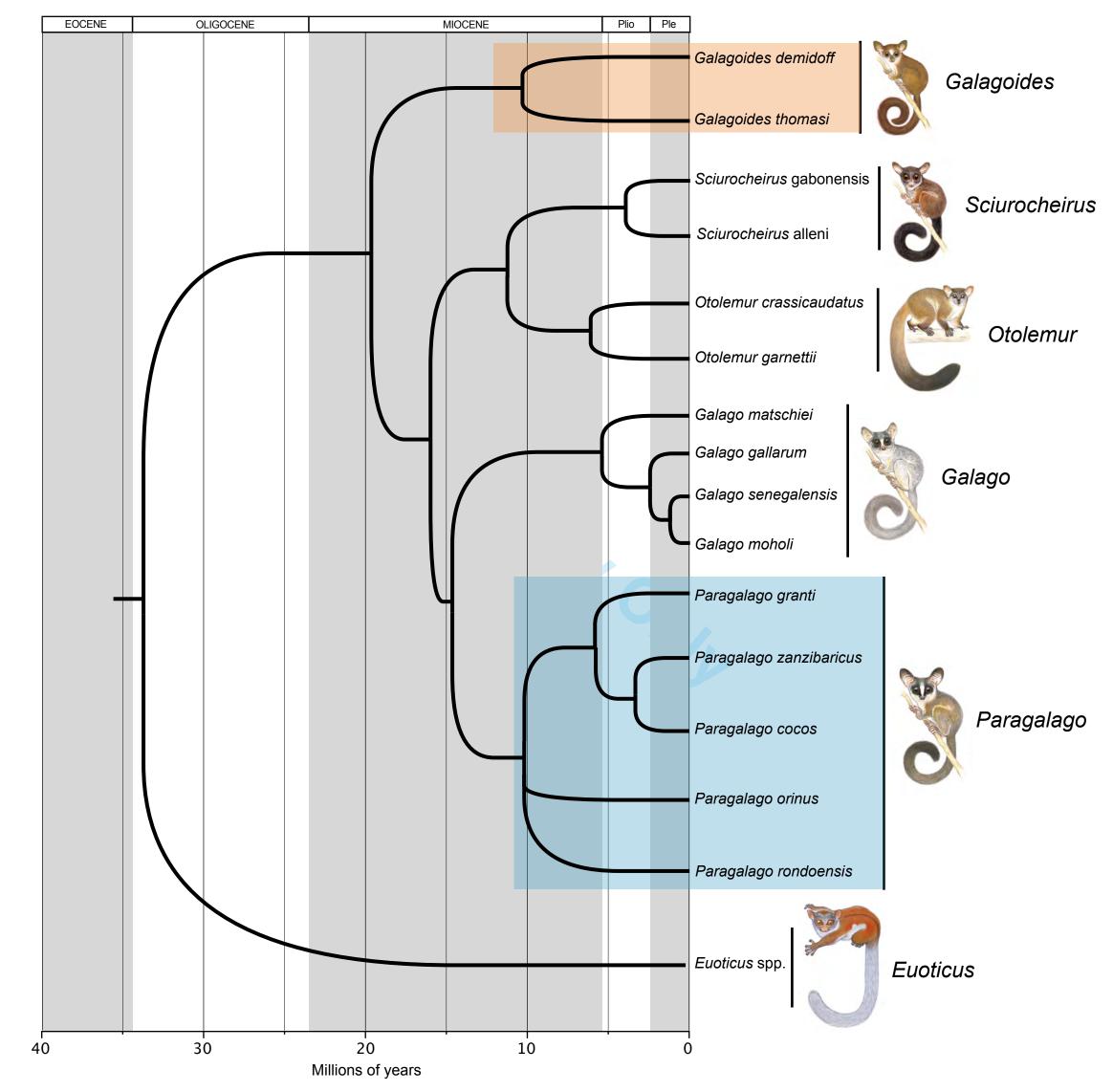
Figure 7: Allometric trajectories for each genus described by the linear regression of

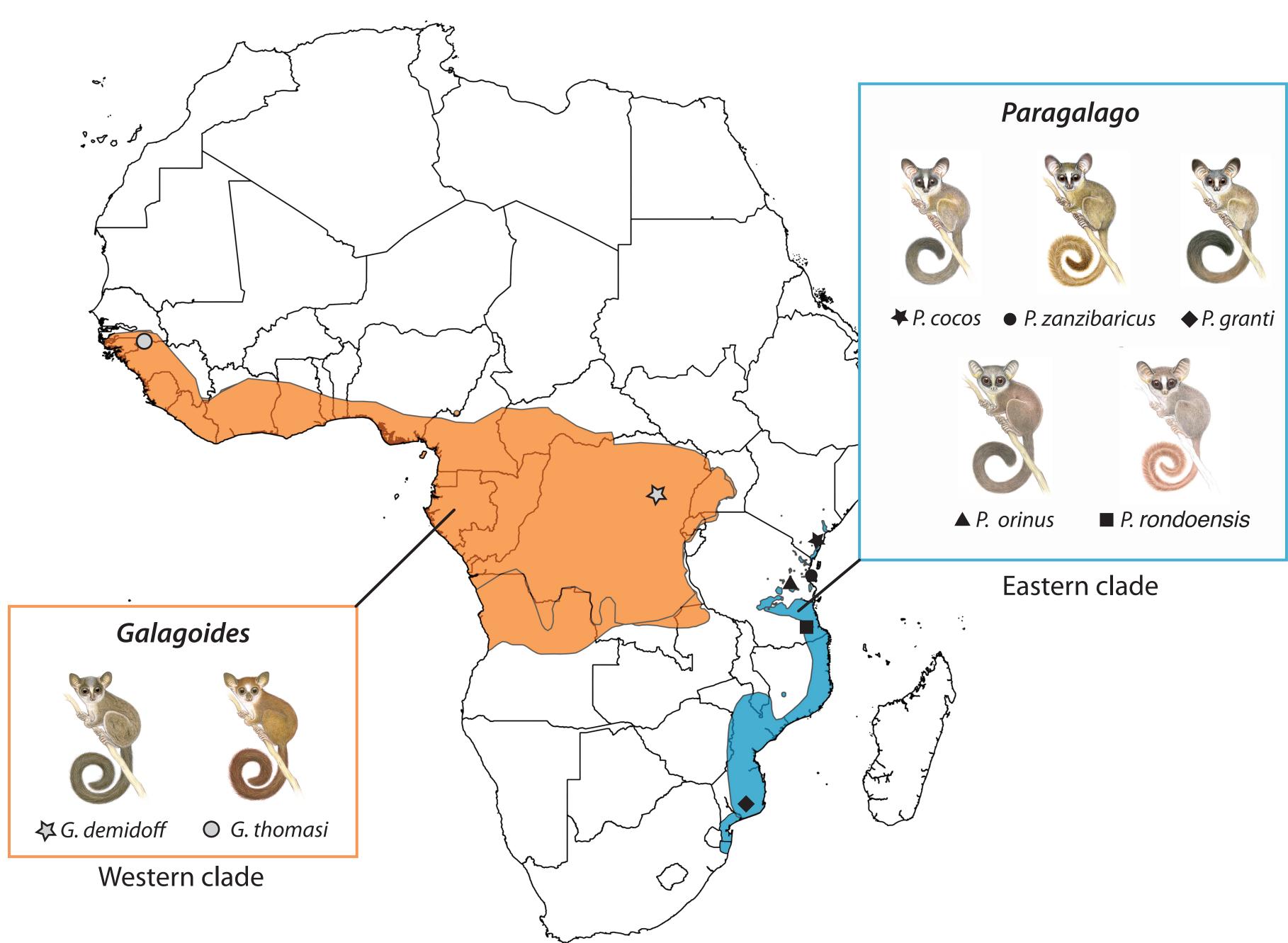
PC1against the log-transformed centroid size. The allometric patterns are different among

- genera. Unlike the other genera, the eastern dwarf galago taxa do not show significant
- allometry. Specimens of the two smallest eastern taxa, orinus and rondoensis, are indicated.

Table 1: Cranial measurements used in the study (illustrated in figure 1) 

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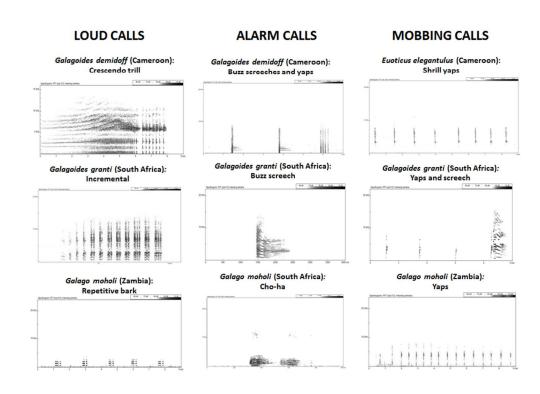
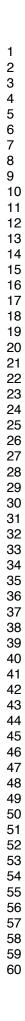


Figure 3: Sonagrams of vocalisations emitted by small-bodied galagid species. Calls of Euoticus (Cameroon) and Galagoides granti (Tshanini, South Africa) were recorded by FG. All other sonograms were downloaded from the East African Primate Diversity and Conservation website (http://www.wildsolutions.nl/vocal-profiles).



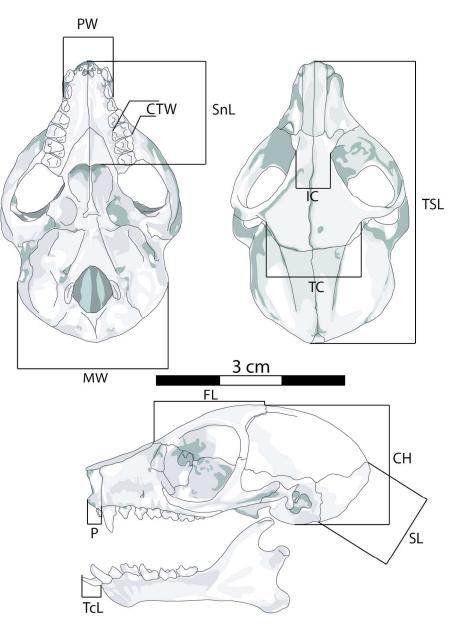


Figure 4: Schematic depiction of an eastern dwarf galago skull showing the 12 craniodental measurements included in the multivariate morphometric analyses. Descriptions of the variables are presented in Table 1.

163x220mm (300 x 300 DPI)

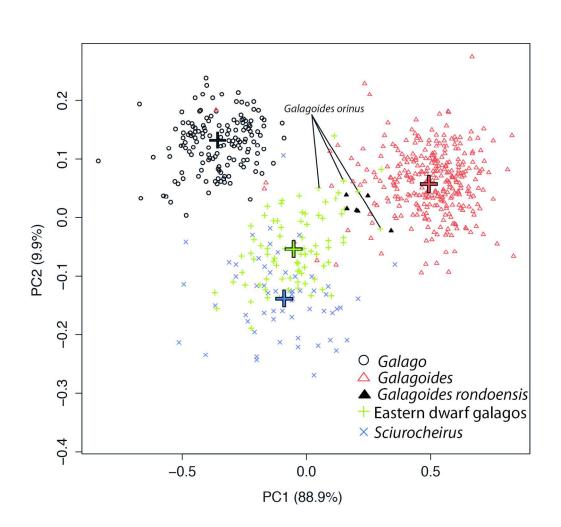


Figure 5: Between Group Principle Component Analysis (BGPCA) calculated on the twelve shape variables. Crosses indicate the mean values of each group that defined the principle component axes to analyse intergroup variation. Specimen data were back-projected in this space. Specimens of the two smallest eastern taxa, *orinus* and *rondoensis*, are indicated.

174x161mm (300 x 300 DPI)

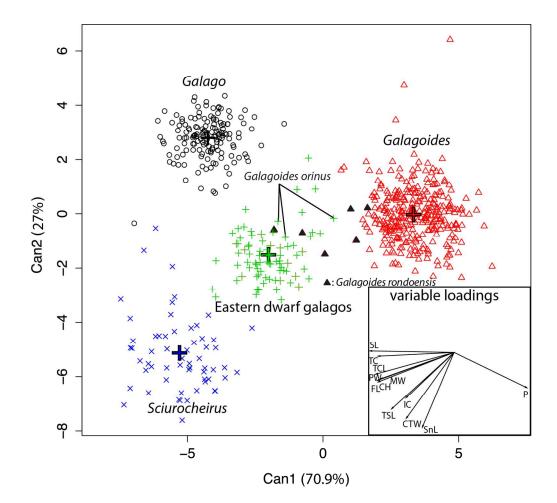


Figure 6: Canonical Variate Analysis of the twelve shape variables. Crosses indicate the centroid of each group. The first two roots illustrate significant differences in skull shape among genera. Specimens of the two smallest eastern taxa, *orinus* and *rondoensis*, are indicated.

160x152mm (300 x 300 DPI)

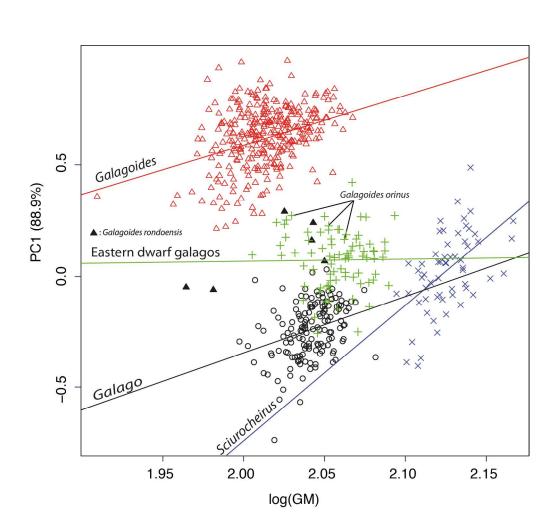


Figure 7: Allometric trajectories for each genus described by the linear regression of PC1against the logtransformed centroid size. The allometric patterns are different among genera. Unlike the other genera, the eastern dwarf galago taxa do not show significant allometry. Specimens of the two smallest eastern taxa, *orinus* and *rondoensis*, are indicated.

201x187mm (300 x 300 DPI)

### TABLE 1. Cranial measurements used in the study (illustrated in Figure 1)

| Abbreviation | Name                      | Definition   |
|--------------|---------------------------|--|
|              |                           |  |
| SL           | Supraoccipital length     | From lambda to opisthion   |
| СН           | Cranial height            | From basioccipital-basisphenoid suture to the highest point of the braincase |
| FL           | Frontal length            | From bregma to nasion  |
| IC           | Interorbital constriction | Minimal distance between the inner margins of the orbits                     |
| CTW          | Cheek teeth width         | Maximum width of right M <sup>1</sup>  |
| PW           | Palate width              | Distance between labial margins of right and left P <sup>2</sup>             |
| TSL          | Total skull length        | From prosthion to opisthocranion   |
| SnL          | Snout length              | From prosthion to nasion   |
| MW           | Mastoid width             | Distance between left and right mastoid processes                            |
| тс           | Temporal constriction     | Minimum distance between left and right fronto-temporals                     |
| Р            | Premaxilla                | Length of the premaxillary tube  |
| TCL          | Toothcomb length          | From the base to the tip of the incisors                                     |

- 6

# Supplementary Information

# Table 1. Eastern dwarf specimens included in the morphometric analysis of skulls

| Accession      |            | 0          |            |          |
|----------------|------------|------------|------------|----------|
| number         | Genus      | Species    | Subspecies |          |
| FMNH 177220    | Galagoides | granti     |            |          |
| NHM 6.11.8.5   | Galagoides | granti     |            | Paratype |
| NHM 6.11.8.6   | Galagoides | granti     |            | Paratype |
| NHM 6.11.8.7   | Galagoides | granti     |            | Туре     |
| NHM 6.11.8.9   | Galagoides | granti     |            | Paratype |
| NHM 6.11.8.10  | Galagoides | granti     |            | Paratype |
| NHM 8.1.1.12   | Galagoides | granti     |            |          |
| NHM 8.1.1.13   | Galagoides | granti     |            |          |
| NHM 8.1.1.15   | Galagoides | granti     |            |          |
| NHM 8.1.1.16   | Galagoides | granti     |            |          |
| NHM 8.1.1.129  | Galagoides | granti     |            |          |
| NHM 94.1.25.22 | Galagoides | granti     |            | Paratype |
| NMZ 28344      | Galagoides | granti     |            |          |
| NMZ 67333      | Galagoides | granti     |            |          |
| NMZ 67340      | Galagoides | granti     |            |          |
| NMZ 67341      | Galagoides | granti     |            |          |
| NMZ 67342      | Galagoides | granti     |            |          |
| NMZ 67344      | Galagoides | granti     |            |          |
| NMZ 67346      | Galagoides | granti     |            |          |
| NMZ 67347      | Galagoides | granti     |            |          |
| NMZ 67349      | Galagoides | granti     |            |          |
| NMZ 67353      | Galagoides | granti     |            |          |
| NMZ 67354      | Galagoides | granti     |            |          |
| NMZ 67355      | Galagoides | granti     |            |          |
| NMZ 67357      | Galagoides | granti     |            |          |
| NMZ 67358      | Galagoides | granti     |            |          |
| NMZ 67359      | Galagoides | granti     |            |          |
| NMZ 67361      | Galagoides | granti     |            |          |
| NMZ 67362      | Galagoides | granti     |            |          |
| USNM 352253    | Galagoides | granti     |            |          |
| USNM 352254    | Galagoides | granti     |            |          |
| USNM 181810    | Galago     | moholi     | 00000      | Туре     |
| MCZ 22453      | •          |            | cocos      |          |
| NHM 54.748     | Galagoides | orinus     |            | Туре     |
|                | Galagoides | orinus     |            |          |
| NHM 54.747     | Galagoides | orinus     |            |          |
| SFR 83201      | Galagoides | orinus     |            | -        |
| MCZ 38875      | Galagoides | rondoensis |            |          |
| SFR 94901      | Galagoides | rondoensis |            |          |
| ZMB 8903       | Galagoides | rondoensis |            |          |
| ZMB 9130       | Galagoides | rondoensis |            | _        |
| ZMB 9131       | Galagoides | rondoensis |            |          |
| NHM 64.1970    | Galagoides | rondoensis |            | Туре     |

| NHM 1995.251  | Galagoides | udzungwensis  |       | Туре |
|---------------|------------|---------------|-------|------|
| MCZ 26446     | Galagoides | zanzibaricus  |       |      |
| MCZ 26449     | Galagoides | zanizibaricus |       |      |
| MCZ 26451     | Galagoides | zanzibaricus  |       |      |
| ZMB 64201     | Galagoides | zanzibaricus  |       | Туре |
| AMNH 187362   | Galagoides | zanzibaricus  | COCOS |      |
| CMNH 57948    | Galagoides | zanzibaricus  | cocos |      |
| CMNH 98107    | Galagoides | zanzibaricus  | cocos |      |
| CMNH 98108    | Galagoides | zanzibaricus  |       |      |
| MCZ 22449     | Galagoides | zanzibaricus  |       |      |
| MCZ 22450     | Galagoides | zanzibaricus  |       |      |
| MCZ 38911     | Galagoides | zanzibaricus  |       |      |
| MCZ 38912     | Galagoides | zanzibaricus  |       |      |
| MCZ 38915     | Galagoides | zanzibaricus  |       |      |
| NMK DM 4533   | Galagoides | zanzibaricus  | cocos |      |
| NMK DM 4537   | Galagoides | zanzibaricus  | cocos |      |
| NMK DM 4547   | Galagoides | zanzibaricus  | cocos |      |
| NHM 55.330    | Galagoides | zanzibaricus  |       |      |
| NHM 64.972    | Galagoides | zanzibaricus  |       |      |
| NHM 64.974    | Galagoides | zanzibaricus  |       |      |
| NHM 64.975    | Galagoides | zanzibaricus  |       |      |
| NHM 64.976    | Galagoides | zanzibaricus  |       |      |
| NHM 64.977    | Galagoides | zanzibaricus  |       |      |
| NHM 64.978    | Galagoides | zanzibaricus  |       |      |
| NHM 64.979    | Galagoides | zanzibaricus  |       |      |
| NHM 64.980    | Galagoides | zanzibaricus  |       |      |
| NHM 20.6.10.2 | Galagoides | zanzibaricus  |       |      |
| NHM 20.6.10.3 | Galagoides | zanzibaricus  |       |      |
| NHM 54.738    | Galagoides | zanzibaricus  |       |      |
| NHM 54.739    | Galagoides | zanzibaricus  |       |      |
| NHM 54.740    | Galagoides | zanzibaricus  |       |      |
| TM 45930      | Galagoides | zanzibaricus  |       |      |
| USNM 184218   | Galagoides | zanzibaricus  | COCOS |      |
| USNM 184220   | Galagoides | zanzibaricus  | cocos |      |
| USNM 184221   | Galagoides | zanzibaricus  | COCOS |      |
| USNM 184222   | Galagoides | zanzibaricus  | COCOS |      |
| USNM 184224   | Galagoides | zanzibaricus  | COCOS |      |
| USNM 241579   | Galagoides | zanzibaricus  |       |      |

#### Key to museums:

AMNH: American Museum of Natural History, New York, USA

CMNH: Carnegie Museum of Natural History, Pittsburgh, USA

FMNH: Field Museum of Natural History, Chicago, USA

MCZ: Museum of Comparative Zoology, Harvard Museum of Natural History, Cambridge MA, USA

NHM: Natural History Museum, London, UK

NMK: National Museum of Kenya, Nairobi, Kenya

NMZ: National Museum of Natural History, Bulawayo, Zimbabwe

SFR: Senckenberg Museum of Natural History, Frankfurt, Germany

TM: Ditsong Museum of Natural History (Transvaal Museum), Pretoria, South Africa

USNM: United States National Museum of Natural History (Smithsonian Institution), Washington DC, USA

ZMB: Museum of Natural History, Berlin, Germany