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## Sustainable Management of Plant Diseases

### This is the author's manuscript

*Original Citation:*

*Availability:*

This version is available <http://hdl.handle.net/2318/1724338> since 2020-01-21T17:11:21Z

*Publisher:*

Springer Nature

*Published version:*

DOI:10.1007/978-3-030-23169-9\_11

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1 **Sustainable Management of Plant Diseases**

2

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11

12 **Abstract**

13 The disease management strategy represents an important contribution to the sustainability of the  
14 farming systems. Plant disease management attempts to maintain disease levels below economic  
15 thresholds because complete elimination of disease is unnecessary and may result in unacceptable  
16 costs, labour and environmental impacts. Integrated disease management intends to manage plant  
17 diseases by assembling complementary approaches, depending on the pathosystem involved, the  
18 geographical location and the pedoclimatic conditions. The current chapter provides several examples  
19 of sustainable disease management, with particular reference to the control of soilborne diseases of  
20 vegetable and ornamentals crops. Healthy soils are fundamental to sustainable disease management.  
21 Most practices designed to improve soil health, such as organic matter supplementation also help to  
22 suppress the disease development. The use of healthy or treated propagation material is an effective  
23 tool to prevent native or alien pathogens. Chemical control with fumigants and fungicides should be  
24 considered when other approaches do not achieve the required pathogen control. Rapid and reliable  
25 diagnostic methods allow a rational and efficient choice of the management options. Decision support  
26 systems should be developed through forecasting models. The choice of the appropriate plant disease  
27 management strategy should not only integrate the impact on the soil and crop health, but also on the  
28 agricultural and non-agricultural environments, the natural resources, and human health. Economic,  
29 social, legislative and political issues should be considered together with regional, national and  
30 international regulations.

31

32 **Keywords:** biocontrol agents, chemical control, diagnostics, induced resistance, integrated disease  
33 management, plant pathogen, seed health, soilborne disease, soil health.

34

## 35 **Introduction**

36 An important goal of sustainable agriculture is the development of integrated farming systems with  
37 reduced use of natural resources (water, soil, energy), as well as of chemical fertilizers and pesticides.  
38 Sustainable farming systems should maintain and possibly enhance the quantity and quality of crop  
39 production, improve the farmer's income, and balance the economic, environmental and social  
40 consequences of human interventions. An important contribution to the sustainability of the farming  
41 systems is the choice of the disease management strategy. In fact, despite the use of pesticides, 20–  
42 30% of production is estimated to be lost due to plant diseases every year (Oerke 2006). Such figures  
43 would be even higher without any intervention for reducing losses caused by plant diseases (Esker et  
44 al. 2012). Crop losses due to plant diseases affect the potential production in industrialized countries,  
45 but in developing countries they are even costly in terms of food security, foreign exchange  
46 requirements for food imports, and income losses to farmers (Oerke et al., 1994).  
47 Plant diseases result from complex interactions among host, pathogen and the environment. The  
48 disease triangle represents the main elements required for plant diseases: a susceptible host plant, a  
49 virulent pathogen able to cause disease, and a favourable environment. Moreover, time can influence  
50 a disease, so the disease triangle could become a tridimensional disease pyramid, by including this  
51 element. Other elements important for some disease could be vectors and human activities, which  
52 modify the interaction through agricultural practices, genetic resistance and fungicide application  
53 (Burdon et al. 2009).

54

## 55 **Plant disease management**

56 Plant disease management attempts to maintain disease levels below economic thresholds because  
57 complete elimination of disease is unnecessary and may result in unacceptable costs, labour and  
58 environmental impacts. Plant disease management faces significant challenges due to increasing  
59 demands for safe and diversified food (Flood 2010); reducing the production potential due to land  
60 competition in fertile areas; depletion of natural resources; reduction of biodiversity in the agro-  
61 ecosystems; and increased risk of disease epidemics due to agricultural intensification, monoculture,  
62 and climate change (Dun-chun et al. 2016). The pathogen spread is facilitated by human  
63 transportation, but there is an increasing evidence that global warming can drive pathogen movement  
64 towards the pole, by altering their latitudinal range (Bebber et al. 2013).  
65 In the late 1960s and 1970s, the commercialization of a large number of broad-spectrum pesticides  
66 of novel structure and mostly with systemic activity marked an era characterized by intensified  
67 agricultural production. After some years of intensive chemical control, new pathogens became  
68 dominant once their competitors were eliminated and fungicide resistance developed (Delp and

69 Dekker, 1985). To address these problems, growers intensified the use of fungicides, which increased  
70 production costs and increased the risk of fungicide residues on crops (Oliver and Hewitt, 2014).  
71 Integrated disease management intends to manage plant diseases by assembling complementary  
72 approaches, depending on the pathosystem involved, the geographical location and the pedoclimatic  
73 conditions. As stated by the European Directive on the Sustainable Use of Pesticides, integrated pest  
74 management carefully considers all available plant protection methods and subsequent integration of  
75 appropriate measures that discourage the development of pathogen populations and keep the use of  
76 fungicides and other forms of intervention to economically and ecologically justified levels, by  
77 minimising the risks for human health and the environment. Plant pathogens are difficult to control  
78 partly due to their spatial-temporal dynamics and rapid evolution (Strange and Scott 2005), associated  
79 with high genetic diversity and short generation times that favour their ability to overcome effective  
80 disease control approaches. Integrated disease management emphasises the growth of a healthy crop  
81 with the least possible disruption of the agro-ecosystems and encourages natural disease control  
82 mechanisms.

83

#### 84 **Sustainable plant disease management**

85 Sustainable management of plant diseases aims to create environments adverse for the pathogens and  
86 suitable for healthy plants, by ensuring high yield through the efficient use of natural resources (Zhan  
87 et al. 2015). An agroecological approach should be used for the management of diseases, leading to  
88 solutions serving the public good by simultaneously fostering agrifood system productivity and  
89 resilience, reducing energy consumption and supporting bioenergy production, as well as conserving  
90 water resources (Kremen and Miles 2012). Agroecology is the science of applying ecological  
91 concepts and principles to the design and management of sustainable food systems (Gliessman 2014).  
92 In addition, economic and societal impacts should be evaluated for each plant disease management  
93 scheme. An agroecological system approach to plant disease management consists of four pillars: (i)  
94 prevention of pathogen introduction and spread in the cropping system; (ii) reduction of pathogen  
95 populations to levels which can be controlled through natural mechanisms; (iii) introduction of  
96 practices into the cropping system designed to promote beneficial microbiota; and (iv) reduction of  
97 fungicide use through the adoption of integrated disease management (Chellemi et al. 2016). To  
98 achieve the goal of sustainable plant disease management, multidisciplinary collaboration between  
99 disciplines, such as plant pathology, plant breeding, agronomy, horticulture, agricultural entomology,  
100 soil science, environmental science, economics and social sciences is needed. Agroecology, besides  
101 being multidisciplinary, is also transdisciplinary, as it incorporates elements of practice and collective

102 action, which enable the scaling of agricultural practices from individual farms to larger landscape-  
103 level (DeLonge and Basche 2017).

104 The current chapter provides several examples of sustainable disease management, with particular  
105 reference to the control of soilborne diseases of vegetable and ornamentals crops. Soilborne  
106 pathogens can cause heavy losses in vegetable production, by affecting both yield and quality.  
107 Soilborne pathogens can occur from the initial nurse stage, to the harvest. In vegetable production,  
108 crop rotations are minimal and soilborne pathogen propagules may accumulate in the soil, which is  
109 the primary inoculum. Soilborne pathogens are particularly favoured in vegetables, which are an  
110 intensive and dynamic system, characterized by a wide range of crop species and varieties, a  
111 continuous introduction of innovative technologies and the use of intensive cultivation techniques.  
112 For the above-mentioned reasons, the management of soilborne diseases in vegetable production  
113 represents a very interesting case study, both in terms of phytopathological issues and innovative  
114 strategies adopted for their control (Colla et al. 2012).

115

## 116 **MAINTAINING HEALTHY SOILS**

117 Healthy soils are fundamental to sustainable disease management, as they affect the density of  
118 pathogens, particularly of the soilborne ones (Janvier et al. 2007), the structure of beneficial  
119 microbiota, and the availability of organic and inorganic nutrition for plants (Larkin 2015; van  
120 Bruggen et al. 2016). Agricultural management strategies can have a major impact on soil quality  
121 with consequent effects on disease incidence. Soil organic matter, one of the primary indicators of  
122 soil health, is fundamental to the long-term sustainability of agroecosystems. Managing soil health is  
123 a matter of maintaining a suitable habitat for the soil (micro)-organisms. The aim of the practices  
124 adopted is to achieve the resilience (the capacity to self-organize into desirable steady states) and  
125 homeostasis (the maintenance of desirable steady states) of the soil microbiota. In most cases, regular  
126 additions of organic matter are necessary to replenish soil resources and improve soil health.

127

### 128 **Suppressive soils**

129 Suppressive soils are those where the disease development is naturally controlled, even in the  
130 presence of a virulent pathogen, a susceptible plant host, and with environmental conditions  
131 conducive for the development of the disease. Soil suppressiveness is a complex system of biotic and  
132 abiotic factors, such as soil structure, nutrient and water availability, microbiota (including pathogens  
133 and symbionts), and plant genotype. Natural soils have a general disease suppression compared to the  
134 same pasteurised soils, and it is directly related to the microbial activity (Schlatter et al. 2017). In  
135 cropping systems, a specific suppression is present when a group of microorganisms, selected for

136 their antagonistic activity, is directly responsible for disease suppression. Soil bacteria and fungi, as  
137 *Pseudomonas* spp. and *Alcaligenes* spp. in the USA (Kloepper et al. 1980; Yuen et al. 1985) and  
138 *Fusarium* spp. in France and Italy (Janvier et al. 2007; Garibaldi et al. 1987), have been shown to be  
139 involved in *Fusarium* wilt suppression. Antagonistic *Fusarium* spp., isolated from the rhizosphere of  
140 carnation grown in suppressive soils, showed high rhizosphere competence. When applied to soil and  
141 substrates they controlled *Fusarium* wilts on different crops, such as tomato, basil, carnation,  
142 cyclamen, and bulb crops (Gullino and Garibaldi 2007). Soils suppressive to *Rhizoctonia solani* are  
143 correlated with the presence of large amounts of *Trichoderma* spp. (Chet 1987).

144

### 145 **Soil management for disease suppression**

146 Organic matter can be added through agronomic practices, such as crop residues, rotations, and cover  
147 crops.

148 Crop rotations are one of the most interesting agronomic practices, as they are able to combine the  
149 optimal use of nutrients with the reduction of soilborne pathogens. The evolution of agriculture has  
150 led to the abandonment of rotations in favour of monoculture, with consequent negative plant disease  
151 profile. Monoculture, in fact, leads to the progressive soil accumulation of propagules of plant  
152 pathogens to unacceptable levels, which force the adoption of disinfestation practices. Some  
153 pathogens (*Fusarium* spp., *Verticillium* spp., or *Rhizoctonia* spp.), which show high competitiveness  
154 at saprophytic level or differentiate survival structures, tend to accumulate in the soil. The mechanism  
155 underlying the beneficial effects of rotation is starving the pathogen when the susceptible host is not  
156 cultivated. This occurs in the case of organisms with narrow host spectrum, modest saprophyte  
157 capacity, and lack of survival structures. The level of specialization of the parasite is important: crop  
158 rotation has higher effect on species-specific pathogens (i.e. *formae speciales* of *Fusarium*  
159 *oxysporum*) than on the polyphagous ones (*Sclerotinia sclerotiorum*, *Verticillium dahliae*). Crop  
160 rotation can also include the alternation of cultivars of the same species with different levels of  
161 pathogen susceptibility.

162 Crop rotations are associated with increasing soil microbial activity and diversity, due to the  
163 cultivation of different plant species in the soil (Garbeva et al. 2004; Welbaum et al. 2004). Crop  
164 rotations that maximize diversity of plant and root systems (mixing legumes, cereals, solanaceous,  
165 cucurbits, brassica, etc.) may significantly modify soil microbiota and their disease suppression  
166 potential.

167 Cover crops are grown primarily to cover the soil, to protect it from erosion and nutrient losses when  
168 production crops are not present. Benefits of cover crops may include disease control (Larkin, 2015).

169 Green manuring is the incorporation of fresh plant material to enrich the soil organic matter. Green  
170 manuring results in higher organic matter inputs than traditional crop rotations or cover crops,  
171 producing improvements in soil fertility, structure, and microbiota, with an effect on disease  
172 suppression (Collins et al. 2006; Stark et al. 2007). Most practices designed to improve soil health,  
173 such as organic matter supplementation also help to suppress the disease development (Welbaum et  
174 al. 2004; Bonilla et al. 2012; Page et al. 2013).

175

### 176 **Suppressive substrates**

177 Suppressiveness has been found for several substrates used in horticulture. Sphagnum peat mixes can  
178 naturally suppress soilborne pathogens, but few weeks after potting, they become conducive to  
179 diseases (Hoitink and Boehm 1999). Peat mixes well tolerate the introduction of biocontrol agents or  
180 the addition of composts (Hoitink and Locke 2012). When hardwood bark is used, improved plant  
181 vigour and disease suppressiveness, from richer microbiota, are observed in potted plants (Hoitink  
182 and Boehm 1999).

183 Increasing the use of compost as a potting substrate would contribute to waste recycling and reduction  
184 of chemical fertilizers. Compost is interesting as a peat substitute, for the lower production cost and  
185 for the increasing concern about the environmental impact of peat extraction (Silva et al. 2007). Some  
186 composts, particularly those amended with composted bark, suppress most soilborne plant pathogens  
187 (Hoitink and Boehm 1999; Noble and Coventry 2005; Termorshuizen et al. 2006). Composts were  
188 demonstrated to be more suppressive than crop residues and peat (Bonanomi et al. 2007). Low  
189 amounts of compost in growing media avoid the lower growth and the phytotoxicity caused by high  
190 pH and electrical conductivity (Sullivan and Miller 2001). Composts originating from green wastes  
191 or municipal biowastes, blended with a peat substrate effectively reduced *Fusarium* wilt on basil,  
192 *Pythium ultimum* on cucumber, *Phytophthora nicotianae* on tomato and *Phytophthora capsici* on  
193 pepper (Pugliese et al., 2014). On the contrary, saline composts were reported to enhance *Pythium*  
194 and *Phytophthora* diseases, while high nitrogen composts could enhance *Fusarium* wilts (Hoitink et  
195 al. 2001). The efficacy of compost for disease control depends on the raw materials from which the  
196 compost was prepared, the composting process used, and the compost maturity and quality  
197 (Termorshuizen et al. 2006). Of particular interest is the use of disease suppressive composts, thanks  
198 to the introduction of selected antagonists: their use is particularly interesting in the case of nurseries  
199 (Garibaldi 1988; Hadar 2011; Hoitink and Fahy 1986). In other cases, composts have been identified  
200 as a potential source of antagonistic microorganisms (Pugliese et al. 2008). In some cases, it is  
201 interesting to combine the use of compost with that of resistant rootstocks (Pugliese et al. 2014).

202 Although interesting for field crops and vegetables, the use of organic amendments for disease control  
203 is still not widespread, due to many factors such as the lack of standardization, the inconsistency in  
204 their efficacy, and the complexity of their use.

205

## 206 **Soilless media**

207 Soilless cultivation is realized in inert or cation exchange capacity substrates (rock wool, perlite,  
208 peat), used as a mechanical support for the plant, replacing the soil. Soilless cultivation requires a  
209 continuous feeding of the plants with a complete nutrient solution. This technique offers numerous  
210 advantages, such as better control of soilborne pathogens and more effective planning of crop cycles.  
211 Soilless cultivation could permit the production cycle completely free of pathogens. It also permits  
212 eradication of the soilborne pathogens in the recirculating nutrient solutions (Van Os et al. 2012).  
213 Soilless cultivation allows excluding soilborne pathogens: the possibility of contact between the host  
214 and pathogen is avoided by growing the plant in a pathogen-free environment (Postma 2004,  
215 Garibaldi and Gullino 2010). Soilless systems, while they strongly limit some pathogens, they could  
216 favour pathogens that find favourable conditions for their diffusion in the nutrient solution. *Pythium*  
217 and *Phytophthora* are the most frequent pathogen genera in the root system of soilless vegetables and  
218 ornamentals. Many pathogens (*Pythium aphanidermatum*, *P. myriotylum*, *Phytophthora cryptogea*,  
219 *P. nicotianae*) found in hydroponics are the same present in normal soil conditions, while others affect  
220 plant hosts which are resistant when grown in soil. *Phytophthora cryptogea* in soilless systems  
221 becomes strongly virulent on lettuce. *Pythium dissotocum* becomes extremely virulent in soilless  
222 cultivation of spinach and lettuce. Other pathogens are specific for soilless crops, such as *Plasmopara*  
223 *radicis-lactucae*, reported on lettuce roots.

224 Among the potential sources of pathogen infection in soilless crops, there are the substrates; perlite,  
225 vermiculite, rock wool, polyurethane, and polystyrene are generally considered sterile, but organic  
226 materials, such as peat, coconut fibre or non-composted bark, represent the main source of infection  
227 of *Pythium* spp., *Fusarium* spp., *Olpidium* spp. and *Thielaviopsis* spp. (Van Os 2010). On the other  
228 hand, the cultivation substrate could show a natural suppressiveness, depending both on chemical and  
229 microbiological factors. By comparing different substrates, there are substantial differences in the  
230 microflora established, which generate a different degree of suppressiveness.

231 In closed systems, higher electrical conductivity of the nutrient solution, amendment with potassium  
232 silicate, and their combination were effective against powdery mildews, downy mildews, leaf spots,  
233 and *Fusarium* wilts (Gullino et al. 2015). Silicon provided partial control of powdery mildews on  
234 greenhouse crops and soilborne diseases on turfgrass (Bélanger et al. 1995; Brecht et al. 2004; Uriarte



235 et al. 2004): in addition to the deposition of amorphous silica in the cell wall, there is an increased  
236 lignin production, which could limit the pathogen penetration in the plant cell (Gullino et al. 2015).  
237 Soilless systems also permit microbial optimization, thanks to the application of microorganisms able  
238 to colonize the plant rhizosphere. Slow sand filtration combined with the application of different  
239 antagonistic strains of *Fusarium* spp. and *Trichoderma* spp. was effective against *Phytophthora*  
240 *cryptogea* in gerbera (Garibaldi et al. 2004a).  
241 Pathogen diffusion in soilless cropping systems can be greatly reduced by adopting proper  
242 disinfection methods for the recirculating solution, such as slow sand filtration (Van Os, 2010).  
243 Moreover, preventative methods to increase the plant resistance to diseases and the use of diagnostic  
244 tools constitute an integrated approach for soilless systems (Van Os et al. 2012).

245

### 246 **Organic amendments**

247 Organic amendments include manure, crop residues, compost, and organic fertilisers. The application  
248 of organic amendments is commonly adopted in traditional agricultural systems to provide nutrients  
249 to the crop and to improve the soil fertility and structure (Bailey and Lazarovits 2003; Bonanomi et  
250 al. 2007; Bonilla et al. 2012). Suppressiveness has been found for organic amendments used in  
251 agriculture. Several chemical and physical changes in the soil are due to the incorporation of  
252 amendments and result in control of soilborne pathogens, with reduced application of chemicals  
253 (Pugliese et al. 2015).

254 A proper nutritional status makes plants more easily able to react to any kind of stress. High nitrogen  
255 fertilization, by favouring the vegetative growth of the host and the tissue turgidity, is conducive to  
256 the pathogen attack. Generally, adequate potassium fertilization makes the host resistant to several  
257 parasites.

258 Soil amendments can be useful to modify the soil pH. For example, pH values above 7 reduce the  
259 incidence of *Plasmodiophora brassicae* on cabbage (Webster and Dixon 1991), though at these pH  
260 values the occurrence of *Erwinia carotovora* increases (Bain et al. 1996). Alkaline soils are conducive  
261 to the spread of the scab of potatoes, as *Streptomyces scabies* usually develops between pH 5.2 and  
262 8.0 (Hooker 1981). It is, however, difficult to generalize and to choose a unique intervention practice.  
263 For example, on carnation, soil pH reduction reduces the attacks of *Phytophthora nicotianae* (Spencer  
264 and Benson 1981) and increases the wilts caused by *Fusarium oxysporum* (Jones et al. 1993).

265 When added to soil, amendments, such as cow or poultry manure and brassica residues, are subjected  
266 to microbial degradation that releases toxic and volatile compounds directly affecting soilborne  
267 pathogens or indirectly increasing microbial soil suppressiveness. Organic amendments can promote

268 the re-establishment of a more balanced and suppressive microflora. Furthermore, the development  
269 of plant disease is reduced thanks to the extended root systems growing in a rich soil (Chellemi 2010).  
270 Composts and Brassica pellets are considered among the most promising organic amendments. A  
271 growing interest is directed to the use of isothiocyanate precursors, contained in selected brassicaceae  
272 (*Brassica juncea* and *B. carinata*), used as alternating species and then applied as green manure or as  
273 flour or pellets (Larkin and Griffin 2007). The use of Brassica species as green manure is a type of  
274 biofumigation that involves the release of volatile compounds able to control a wide array of soilborne  
275 pathogens (Larkin and Griffin 2007). Biofumigation, however, provides results that are not always  
276 univocal: promising efficacy was obtained against *Colletotrichum coccodes* on tomato, *Fusarium*  
277 *oxysporum* f. sp. on cucumber, *Verticillium dahliae* on eggplant grafted onto *Solanum torvum*, and  
278 *Fusarium* wilt of lettuce, rocket and basil (Garibaldi et al. 2010; Gilardi et al. 2014). Partial or negative  
279 results have been observed in other crops, such as *Brassica* spp., where the inoculum of soilborne  
280 pathogens could be favoured (Lu et al. 2010). The combination of green manure with soil solarisation  
281 is also very effective and reduces the period of soil mulching with plastic films.  
282 Organic amendments for disease control are not yet widespread, due to lack of standardisation of  
283 production parameter, inconsistent efficacy and difficult application. Control of soilborne diseases  
284 with organic amendments must be considered a component of a system approach, where the impact  
285 of crop production practices on resident soil microflora is addressed.

286

### 287 **Soil solarisation**

288 Solarization is the soil covering with plastic film during the summer. The method has been widely  
289 exploited in warm and temperate countries (Katan and DeVay 1991). Farmers are generally sceptical  
290 about its adoption, as it requires soil free of cultivation for at least 4 weeks. An integration strategy,  
291 often adopted to increase soil solarization efficacy, is its combination with biocontrol agents, to  
292 reduce the solarisation period and to permit its use in marginal areas (Minuto et al. 2005). The  
293 combination of soil solarization and *Streptomyces griseoviridis* is effective against fusarium and  
294 verticillium wilts and corky root, and it increases the range of pathogens controlled with respect to  
295 the single treatments. Significant increases in yield and fruit weight were observed, confirming the  
296 potential additive effect caused by biocontrol agent and solarization in terms of yield increase.

297

## 298 **PLANTING MATERIAL**

299

### 300 **Healthy propagation material**

301 Considering the losses caused by most emerging pathogens, the first preventative strategy that should  
302 be considered by seed producers and farmers is the use of healthy seeds and propagation material.  
303 The use of healthy or treated propagation material is an effective tool to prevent native or alien  
304 pathogens from being introduced in the agricultural environment. It is estimated that almost 800  
305 fungi, over 150 viruses, 100 bacteria and 20 phytopathogenic nematodes are transmitted through  
306 propagation material. To avoid this risk, programs have been activated for the most important crops  
307 aimed at certifying the health of the seed or propagation material. This requires specific phytosanitary  
308 assays, which consist in estimating the possible presence of the pathogen using different biological  
309 and molecular methods.

310 The control of propagation material is important for clonal species (carnation, geranium, strawberry)  
311 for which the use of uncontrolled material could facilitate disease outbreaks. The importance of the  
312 use of healthy or treated material is particularly evident in the case of pathogens (viruses, bacteria)  
313 with few or ineffective control strategies (Gullino and Munkvold 2014). On strawberry, the use of  
314 certified propagation material, obtained by thermotherapy, meristem cultivation and subsequent  
315 indexing is a consolidated practice.

316 Another important aspect is seed health. Stock seeds should be produced in locations with low disease  
317 risk, characterized by low humidity and dry summer climate, to reduce fungal or bacterial epidemics  
318 (Munkvold 2009). The choice of proper geographical areas, possibly isolating seed and seedling  
319 production from the environment, and the application of good agricultural practices are critical for  
320 producing high-quality, pathogen-free seed.

321 As it is unrealistic to pursue an absolute seed health of the seed, a certain tolerance is admitted. Very  
322 common is the diffusion of fungal and bacterial seedborne pathogens on vegetables (Koch and  
323 Roberts 2014). The production of virus-free seed must follow appropriate production and certification  
324 schemes, which involve the controlled cultivation of the mother plants and diagnostic tests both on  
325 the mother plants and the seed produced (Gullino and Bonants 2014).

326 To reduce the risks of fungal and bacterial seedborne diseases, it is recommended that stock seeds  
327 undergo precautionary chemical or physical treatments. Chemical seed treatments have successfully  
328 been applied to vegetable seeds and are in commercial use for a wide range of crops against different  
329 seedborne pathogens (Munkvold 2009). Several surface disinfectants (bleach, hydrogen peroxide,  
330 ethanol) can be applied to remove pathogen inoculum from seed coats (Mancini and Romanazzi  
331 2014). Chemical treatments are effective but they can also negatively affect germination and cause  
332 phytotoxicity (Axelrood et al. 1995; du Toit 2004), besides having negative effects on human health  
333 and the environment (Lamichhane et al. 2016). Alternative strategies for the control of seedborne  
334 pathogens include physical seed treatments, treatments with natural compounds, antagonistic

335 microorganisms, and resistance inducers. Physical strategies include mechanical (sorting and  
336 brushing), heat, ultrasonic, radiations (with microwaves resulting in elevated temperatures), UV-C  
337 light, and redox treatments (cold plasma and electrons (Spadaro et al. 2017). Thermal treatments with  
338 hot water, aerated steam or dry heat can be very effective, but they need to be optimised for the  
339 pathosystems, due to the different temperature and time required (Koch and Roberts 2014). Although  
340 alternative seed treatments have been intensively investigated, there are few examples of commercial  
341 application (Koch and Roberts 2014; Gullino et al. 2014).

342 Seed treatments can also be an effective means to increase seedling emergence, particularly when  
343 done on seeds of low vigour and when the seed coat has been damaged (Mancini and Romanazzi  
344 2014). In general, the use of healthy or disinfected seed is a very useful practice for plant disease  
345 management.

346

### 347 **Resistant varieties and grafting**

348 Host resistance, which is the use of resistant and/or tolerant plant varieties, is one of the most effective  
349 strategies against pathogens. Varieties, which are resistant or at least tolerant to one or more  
350 pathogens, are available for many crops and the industry is investing on research in this field.  
351 Resistant cultivars of lettuce are able to control Fusarium wilt. Lettuce varieties that are resistant, or  
352 at least tolerant, to race 1 of Fusarium wilt are available (Garibaldi et al. 2004b; Gilardi et al. 2014),  
353 but their use is complicated by the presence of different races of the pathogen. Seed breeding  
354 companies are currently working hard in order to develop planting material resistant to the recently  
355 detected race 4 (Gilardi et al. 2017a).

356 Host resistance, and the integration of such varieties with other management strategies is fundamental  
357 within the framework of IPM, but few research focused on the integration of plant resistance with  
358 other IPM strategies (Stout and Davis 2009). Moreover, the breeding approach used to date to develop  
359 resistant and/or tolerant crop varieties should be revised, as most crop cultivars bred to date are based  
360 on a market-driven approach focused on high yield and remunerative crop varieties. This trend has  
361 facilitated the adoption of short rotations or monoculture practices and ignored the potential that  
362 minor side crops may have for IPM. The limited range of available minor crop varieties is one obstacle  
363 to crop diversification, thereby confining certain beneficial practices such as multiple cropping or  
364 intercropping. Sustainable disease management should develop crop breeding based on the  
365 competitiveness of crops and their adaptation to diversified cropping systems (Lamichhane et al.  
366 2017).

367 Grafting is used to reduce susceptibility against pests, root rots and wilts, and to increase yield  
368 (Rouphael et al. 2010). In spite of disadvantages associated with grafting, including the additional

369 cost and physiological disorders due to incompatibility between rootstocks and scions, the use of  
370 resistant rootstock strongly increased, mainly for vegetable crops. In spite of disadvantages associated  
371 with grafting, including the additional cost and physiological disorders due to incompatibility  
372 between rootstocks and scions, the use of resistant rootstock, despite its high cost, strongly increased.  
373 Grafting on resistant rootstock is becoming popular on pepper and some of the commercially  
374 available rootstock provide a good control of *Phytophthora* blight (Gilardi et al. 2013). In the case of  
375 *P. capsici* on bell pepper, due to the lack of commercial cultivars with resistance, growers are  
376 interested in grafting. Grafted plants are popular in the case of tomato, to control soilborne pests and  
377 pathogens and to increase yield (Chellemi 2002; Lee and Oda 2003; Gilardi et al. 2013). Grafting  
378 susceptible crops onto resistant rootstocks is interesting also for cucumber (*Cucurbita vicifolia* as  
379 rootstock resistant to *Fusarium* wilt) and melon (*Benincasa cerifera* resistant to *Fusarium* wilt) (King  
380 et al. 2008).

381

## 382 **CHEMICAL AND BIOLOGICAL CONTROL METHODS**

383

### 384 **Chemical control: fumigants and fungicides**

385 Chemical control with fumigants and fungicides is an inseparable component of plant disease  
386 management, and it should be considered when other approaches can not achieve the required level  
387 of pathogen population density reduction.

388 Soil disinfestation with fumigants is becoming very difficult due to the loss of registered fumigants  
389 due to recent regulation strongly limiting their availability (Colla et al. 2014). Among the fumigants  
390 available, dimethyl disulphide, metham sodium, and dazomet provide significant control of *Fusarium*  
391 wilt of lettuce (Gilardi et al. 2017b). Covering the soil with low-density polyethylene film (LPDE)  
392 permits the reduction of fumigant dosage, with interesting results, both under greenhouse conditions  
393 and in the open field. Combination of fumigants with alternative methods, notably solarization, are  
394 promising. The combination of solarisation for two weeks and fumigation with reduced dosage of  
395 fumigants was effective, and allowed a shortening of solarization, permitting a reduction in the non-  
396 cultivation period (Gullino et al. 2003).

397 Fungicides are not used to control soilborne pathogens in open field, , because of their relative high  
398 cost, but they could be used for seed dressing, in nursery to protect the plantlets from damping off  
399 and other soilborne diseases, and in potted plants.

400 Mechanisms of action and risk of pathogen resistance development should be considered, when  
401 selecting the active ingredient (Siegwart et al. 2015). Diversity of fungicides, concerning their  
402 chemistry and mode of action, is essential to ensure effective crop protection, to control new threats

403 and to manage fungicide resistance (Leadbeater and Gisi, 2010). Overuse of many organic fungicides  
404 can result in resistant fungal populations, so it is important to use fungicides as part of an overall  
405 resistance management plan. In the case of *Pythium* damping off, control is mainly accomplished by  
406 treatments with fungicides, such as strobilurins and phenylamides. However, *Pythium* spp. can  
407 develop resistance to common fungicides, such as azoxystrobin or mefenoxam. This further suggests  
408 the necessity of using other fungicides and alternative means for damping off control, and an accurate  
409 identification of *Pythium* spp. before choosing the appropriate control strategy (Matic et al. 2018).  
410 The use of fungicides in integrated disease management is not aimed at eradicating the disease but to  
411 reduce it at ecological and economical thresholds.

412

### 413 **Induced resistance**

414 Plants have constitutive and induces responses to defend themselves against pathogens. Two main  
415 types of induced resistance are known: systemic acquired resistance (SAR) and induced systemic  
416 resistance (ISR) (Vallad and Goodman, 2004). SAR elicits the death of one or a few cells, known as  
417 the hypersensitive response (HR) and the production of pathogenicity-related (PR) proteins, such as  
418 glucanases, chitinases and thaumatin-like proteins (Shoresh et al. 2010). New growth occurs  
419 following HR and salicylic acid plays a role in triggering the signal. SAR is often related to the  
420 induction via aerial plant parts and it usually takes a certain amount of time to be fully expressed in  
421 plants. ISR is often triggered by rhizosphere bacteria in the soil, it involves jasmonic acid and  
422 ethylene, but not salicylic acid and PR-proteins.

423 Induced resistance, mostly SAR, can be triggered by a variety of natural and chemical compounds  
424 (Walters et al., 2005). The increasing interest in their use depends on their broad spectrum of activity,  
425 and on the possibility of reducing the number of fungicide sprays (Walters et al. 2013).

426 Very interesting results have been observed against *Fusarium* wilt of lettuce and crown and root rot  
427 of zucchini, caused by *Phytophthora capsici*, using resistant inducers, based on either phosphites or  
428 acibenzolar-S-methyl, applied as pre-plant treatment in the nursery. Phosphite-based products also  
429 show a very positive effect on plant biomass (Gilardi et al. 2015; 2016). The benefits of preventive  
430 and repeated treatments with silicates to reduce the attacks of *P. aphanidermatum* (Heine et al. 2007)  
431 and *Fusarium oxysporum* f. sp. *radicis-lycopersici* on tomato (Huang et al. 2011) were demonstrated.  
432 Also commercial biocontrol agents (BCAs) were able to reduce *Fusarium* wilt of lettuce, particularly  
433 when their application starts at nursery (Gilardi et al. 2016) while they were not effective against  
434 crown and root rot on zucchini (Gilardi et al. 2015). BCAs can also be effectively applied, alone or  
435 combined with heat treatments, for seed dressing, in the case of seed-transmitted pathogens, such as  
436 *F. lactucae* (Lopez-Reyes et al. 2016). The efficacy of resistance inducers is seldom complete, as it

437 is generally influenced by several factors (target pathogen, plant genotype, phenotype, environmental  
438 conditions, application timing, and formulation) (Walters et al. 2013).

439

#### 440 **Biocontrol agents**

441 Many laboratories around the world have developed their own microorganisms and this allowed the  
442 collection of important contributions about the biology of pathogens and antagonists. Biocontrol  
443 agents may act in various ways but have specific modes of action, including antibiosis, competition,  
444 mycoparasitism and induced resistance.

445 Among the antagonists studied, saprophytic *Fusarium oxysporum*, often isolated from *Fusarium*  
446 suppressive soils, have been widely exploited for their activity against several *Fusarium* wilts  
447 (Garibaldi et al. 1994; Spadaro and Gullino 2005; Gullino et al. 2015). The good antagonistic attitude  
448 of strains belonging to *Trichoderma* spp. has been proved against *Fusarium* wilts in vegetables and  
449 ornamental crops (Harman 2006; Gilardi et al. 2016). Plant growth-promoting rhizobacteria, such as  
450 *Pseudomonas* spp. and *Bacillus* spp., can induce host systemic resistance against several diseases  
451 (Clematis et al. 2009; Lopez et al. 2014).

452 However, in spite of the initial great optimism and extensive research efforts, progress in achieving  
453 commercial, large-scale usage of biological control has been slow. When trials move towards the  
454 farm scale, many antagonists show inconsistent efficacy and lack reliability (Mathre et al. 1999).

455 Biofungicides still face significant constraints, but there are many possibilities for combining various  
456 biocontrol agents, with each other, or with agronomical, physical or chemical control methods  
457 (Spadaro and Gullino 2005). In particular, by combining different methods of control, the aim is to  
458 obtain a synergistic rather than additive effect. For that reason, a complete comprehension of the  
459 mechanism of control is needed. Combining a biocontrol agent with a fungicide improves the  
460 biofungicide efficacy and enables the reduction of the fungicide dosage. Moreover, the combination  
461 of control methods provides a wider spectrum of control, which is needed to replace fumigants.

462

### 463 **ADDITIONAL TOOLS FOR SUSTAINABLE DISEASE MANAGEMENT**

464

#### 465 **Diagnostics**

466 Rapid and reliable diagnostic methods allow a rational and efficient choice of the management  
467 options. The easy spread of fungal spores, virus and bacteria combined with the intense trading  
468 globalization are key factors to allow the movement of pathogens around the world, which can  
469 become invasive in new areas and even cause the total destruction of the crop. Traditional detection  
470 methods based on visual assessment of plant symptoms, isolation, culturing in selective media, and

471 direct microscopic observation of pathogens are frequently laborious, time-consuming and require  
472 extensive knowledge of classical taxonomy. For many diseases, the observation under microscope or  
473 stereoscopic microscope is used to determine the causal agent, taking into consideration pathogenicity  
474 tests and morphological features such as size and shape of the propagules and colony characteristics,  
475 such as colour. However, many microorganisms (including viruses) can produce the same symptoms  
476 in the plant, making difficult the correct identification of the causal agent. As many plant pathogens  
477 remain latent in the planting material, and may be present in very low numbers, high sensitivity,  
478 specificity, and reliability methods are required. The impossibility or difficulty of culturing some  
479 species *in vitro* and the inability for accurate quantification of the pathogen are other limitations.  
480 Early detection of pathogens in seeds and plant materials is of key importance to avoid further  
481 spreading and introduction of new pathogens into growing areas where they are not present yet. These  
482 limitations have led to the development of molecular approaches with improved accuracy and  
483 reliability. Molecular techniques are faster, more specific, sensitive, and accurate than traditional  
484 techniques and they can identify non-culturable microorganisms and facilitate early disease  
485 management decisions.

486 The combination of traditional and molecular techniques permits to characterize, detect, identify and  
487 quantify different pathogens. In the case of fungal pathogens, the Internal Transcribed Spacer region  
488 (rDNA ITS) has been selected by the Consortium for the Barcode of Life (CBOL) as the primary  
489 fungal barcode for species identification (Begerow et al. 2010). Other genomic regions are interesting  
490 for the fungal identification at species level, or even at subspecies level (Srinivasan et al., 2010). The  
491 16S rRNA has been selected as universal barcode for bacteria identification (Weisburg et al. 1991).  
492 An early pathogen detection represent the best preventative measure in several pathosystems, as in  
493 the case *formae speciales* and races of *Fusarium oxysporum* from seeds, plants and soil samples  
494 (Pasquali et al. 2007; Mbofung and Pryor 2010; Thomas et al. 2017; Gilardi et al. 2017a).

495 Loop-mediated isothermal amplification (LAMP) is a DNA amplification method that can be used to  
496 amplify nucleic acid in a target specific way without the need for thermal cycling (Notomi et al.  
497 2000). LAMP is particularly promising for plant pathogen detection, as it is easier and quicker to  
498 perform than PCR, it can be performed on hand-held platforms, and it is well suited for in field use.  
499 The LAMP method has been demonstrated for the detection of bacteria (Hodgetts et al. 2015), fungi  
500 (Franco Ortega et al. 2018), phytoplasmas (Hodgetts et al. 2011) and viruses (Tomlinson et al. 2013).  
501 The limit of detection of pathogens, by comparing the molecular techniques, can reach nanograms of  
502 DNA for PCR, picograms of DNA for biosensors, and femtograms of DNA for qPCR and digital  
503 PCR. NGS technologies are having an enormous impact on biological sciences, allowing the  
504 determination of genome variation within a species or a population. Comparative analysis of the



505 genome sequences allows the identification of highly conserved gene families, conserved regulatory  
506 elements, repeated elements, uncultured pathogens, new species, symbionts, etc., on which new  
507 markers could be designed. On the other side, the use of field techniques, such as LAMP and portable  
508 platforms, is a promising tool to early and quickly detect pests and a useful decision support system  
509 for appropriate pest and disease management. The choice of the diagnostic technique depends on the  
510 balance between the reliability and the cost per sample. Microbiological techniques are generally  
511 cheap, but time-consuming, while molecular technologies have a higher cost, which is  
512 counterbalanced by the higher performance. PCR, qPCR and LAMP have a progressively lower cost  
513 per sample in the order of 2-10 € sample, while NGS are more expensive and they are not yet used  
514 for routine analysis (Spadaro et al. 2018). The development of new instruments and platforms and  
515 the continuous increase of bioinformatics-data have allowed the use of bioinformatics-based  
516 techniques such as metagenomics, comparative genomics and genome sequencing as routine analysis  
517 tools. The dramatic decrease of the cost of the new sequencing technologies permits to foresee a  
518 higher adoption rate in diagnostic laboratories in the near future.

519

## 520 **Forecasting models**

521 Research tried to develop disease predictions models, also called forecasts or warnings, to help the  
522 farmers determine whether and when preventive management measures are needed. Plant disease  
523 models are simplifications of the relationships between pathogens, crops, and the environment that  
524 cause epidemics to develop over time and space. Plant disease models produce predictions about  
525 epidemics or single epidemic components that can be used as risk indicators. Such models also  
526 produce predictions about plant disease epidemics that allows growers to respond in timely and  
527 efficient ways by adjusting crop management practices. A prediction of low disease risk may result  
528 in reduced fungicide treatments with positive economic and environmental effects (Rossi et al., 2010).  
529 Disease prediction is most useful for economically important, sporadic diseases for which effective  
530 management measures are available. It is also important that growers or technicians be able to operate  
531 the prediction system themselves, or that there is a good communication tool between those who  
532 monitor and those who manage the disease.

533

## 534 **Decision support systems**

535 Decision support systems (DSS) should be developed through forecasting models, results of the early  
536 detection tools, as well as pathway, establishment and spread models. Data from various sources are  
537 interpolated using spatial statistics methods, making the DSS able to provide prediction data with  
538 high accuracy at field and site-specific scale. The DSS should have a user-friendly interface, having

539 Geographic Information System (GIS)/mapping functionalities to project the pathogen occurrence.  
540 They also could provide alerts when a new pathogen has been identified and could provide  
541 recommendations for treatment applications (ideal timing and dosage, optimal sprayer calibration,  
542 real-time indicator for tractor speed).

543 Recently developed DSSs are characterised by holistic treatment of crop management problems  
544 (including pests, diseases, fertilisation, canopy management and irrigation); conversion of complex  
545 decision processes into simple and easy-to-understand ‘decision supports’; easy and rapid access  
546 through the Internet; two-way communication between users and providers that make it possible to  
547 consider context-specific information (Rossi et al. 2012).

548

## 549 **Conclusions**

550 Attempts to control soilborne pathogen populations include the use of pesticides, genetic resistance,  
551 crop rotations and a variety of cultural practices, aimed at reducing plant infections. Since these  
552 measures not always provide adequate disease control, fumigants and fungicides are sometimes  
553 needed, as part of an integrated disease management (Ojiambo et al. 2017). Adopting preventative  
554 and combined methods of disease management has become the choice for the control of soilborne  
555 pathogens on economically important crops.

556 The management of soilborne pathogen represents a real challenge and it is a bright example of how  
557 complicated can be a sustainable approaches to plant disease management.

558 The implementation of the concepts of soil health and soil health management into agricultural  
559 production is essential for sustainable crop production and environmental quality (Larkin 2015). The  
560 choice of the appropriate plant disease management strategy should not only integrate the impact on  
561 the soil and crop health, but also on the agricultural and non-agricultural environments, the natural  
562 resources, and human health. Economic, social, legislative and political issues should be considered  
563 together with regional, national and international regulations.

564 New disease outbreaks emerge and will emerge, requiring continuous changes to the disease  
565 management system and reprioritization of goals and objectives. Globalization of trade, new  
566 consumption habits, shifts in diets, and climate change are among the factors influencing the  
567 occurrence, frequency and severity of new plant diseases, with an important impact on decision-  
568 making tools for the related disease management measures that should be adopted.

569 Effort for a continuous monitoring and disease surveillance is necessary. Strategies to produce healthy  
570 seeds and seed treatment methods need to be investigated and made available to seed companies and  
571 growers.

572 Plant disease management should be adapted to the geographical areas, to the crops and to the  
573 pathogens. Future plant disease management should continue to strengthen food security for a stable  
574 society, but also safeguard the health of associated ecosystems and reduce dependency on natural  
575 resources.

576

### 577 **Acknowledgements**

578 This research has received funding from the European Union's Horizon 2020 research and innovation  
579 programme under grants agreements No. 634179. Effective Management of Pests and Harmful Alien  
580 Species - Integrated Solutions (EMPHASIS) and EUCLID EU-CHINA Lever for IPM  
581 Demonstration, No. 633999 (EUCLID).

582

583

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